## Summarising Data in R

Summarising Data

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### Summarising Data

Individual data points are some abstraction away from reality, essentially removing some detail.

- Even with trial-level data, we will **summarise responses** in terms of reaction times or correct responses.
- This removes a lot of the details of the real experience, but allows us to **see patterns**.
- Similarly, we can summarise data from many participants in a number of ways to help communicate a pattern or idea.

We can present summaries of data as a **plot**, as a **table of descriptive statistics**, and make inferences and decisions based on summaries of the data by **modelling our data**.

### **Descriptive Statistics**

## [1] 413

We often summarise data in terms of measures of **central tendency** and **dispersion**.

- Central tendency: What is the average score? This can be determined in a few ways.
  - **Mean**: used to describe data that are normally distributed. Can be misleading when outliers are present.
  - **Median**: used to describe data that are skewed in some way away from a normal distribution. Suppresses the impact of outliers.

```
noskew <- c(10, 20, 15, 20, 22)
mean(noskew)

## [1] 17.4

skew <- c(10, 20, 15, 20, 2000)
mean(skew)

## [1] 20

## [1] 20

## [1] 20
```

### **Descriptive Statistics**

- **Dispersion**: The spread of scores (e.g. for individuals) around an average.
  - **Standard Deviation**: the spread of the data from the sample mean. With a normal distribution approximately 1, 2, and 3 *SD*s capture approximately 68, 95, and 99.7% of the data.
  - **Interquartile Range**: Rank orders data into four equal parts; the first region or quartile (i.e. 25%), the median (i.e. 50%), and the middle part of the third quartile (i.e. 75%). Used for non-normal data.

```
# low variability in scores = low SD
low_dispersion <- c(10, 12, 8, 9, 11, 10, 10.5, 9, 11.5)
sd(low_dispersion)

## [1] 1.293681

# lots of variability in scores = high SD
high_dispersion <- c(10, 1000, 89, -400, 90, 880, 0)
sd(high_dispersion)</pre>
```

## Why Make these in R?

- Scales up easily to new data: If you write this code to analyse data from 10 participants, you can instantly rerun it for millions.
- Easier to spot mistakes when you're writing the recipe.
- Easier to fix mistakes with a minor modification instead of repeating every step of the analysis by hand.
- **Repeatability**: If you do a task once in R, you can copy and paste code for a new study. This saves a lot of time in the long run.
- **Reproducibility**: Allows others (and future you) to check work and inspect methods at every step. You'll make science more reliable!

### **Summarise**

Let's take a look at the inbuilt starwars data set in the tidyverse pacakage. There's a lot of data here! How might we summarise this?

```
data(starwars) # load the data from within the package
glimpse(starwars) # view it
```

```
## Rows: 87
## Columns: 14
## $ name
                                            <chr> "Luke Skywalker", "C-3PO", "R2-D2", "Darth Vader", "Leia Or...
## $ height
                                            <int> 172, 167, 96, 202, 150, 178, 165, 97, 183, 182, 188, 180, 2...
## $ mass
                                            <dbl> 77.0, 75.0, 32.0, 136.0, 49.0, 120.0, 75.0, 32.0, 84.0, 77...
## $ hair_color <chr> "blond", NA, NA, "none", "brown", "brown, grey", "brown", N...
## $ skin_color <chr> "fair", "gold", "white, blue", "white", "light", "light", "...
## $ eye_color <chr> "blue", "yellow", "red", "yellow", "brown", "blue", "blue", "
## $ birth year <dbl> 19.0, 112.0, 33.0, 41.9, 19.0, 52.0, 47.0, NA, 24.0, 57.0, ...
## $ sex
                                            <chr> "male", "none", "male", "female", "male", "female", "female", "
                                            <chr> "masculine", "masculine", "masculine", "masculine", "femini...
## $ gender
## $ homeworld
                                            <chr> "Tatooine", "Tatooine", "Naboo", "Tatooine", "Alderaan", "T...
                                            <chr> "Human", "Droid", "Droid", "Human", "
## $ species
## $ films
                                            <list> <"The Empire Strikes Back", "Revenge of the Sith", "Return...</pre>
## $ vehicles
                                            <list> <"Snowspeeder", "Imperial Speeder Bike">, <>, <>, <>, "Imp...
                                            <list> <"X-wing", "Imperial shuttle">, <>, <>, "TIE Advanced x1",...
## $ starships
```

### **Summarise**

summarise() collapses across all observations in your data set to produce a single row of data.

- summarise() takes two main arguments:
  - 1: what is the **data** set you want to summarise?
  - 2: additional information specifying **how you want to summarise it**.

Let's summarise the mean height of the Star Wars characters:

```
summarise(starwars, mean_height = mean(height))

## # A tibble: 1 x 1

## mean_height

## <dbl>
## 1 NA
```

Oops, we got NA! Why? R by default produces NA for any summary of data containing NA values. (You can't average something that isn't there.)

#### Summarise

What should we do? We can make a new data set without NAs, or tell mean() to ignore them.

```
summarise(starwars, mean_height = mean(height, na.rm = TRUE))

## # A tibble: 1 x 1

## mean_height

## <dbl>
## 1 174.
```

We set the argument **na.rm** in mean() to **TRUE**, meaning "Should R remove NAs? TRUE (yes)".

## Many Summaries

- What if we want to get **different types of summaries**? Imagine we want the count, mean, and standard deviation of height?
- We also can tell R to generate these summaries on the data by first **dropping any observations with missing values (NA)** in height.

```
summarise(
  drop_na(starwars, height),
  n = n(),
  sd = sd(height),
  mean_height = mean(height)
)
```

- How might we interpret this? We have 81 measured heights of characters. The mean height is 174.36cm, while the standard deviation is 34.77.
- What's the SD mean? With normally distributed data 95% of values fall within  $\pm$  1.96 SD of the mean!

## Grouping

## 2 Besalisk

## # ... with 35 more rows

## 3 Cerean

198

198

- As we can see, the data set contains **very many species** of creates from the Star Wars universe. What if we want summaries of all of them?
- We can add an argument to our data in summarise() called group\_by().
- Within group\_by(), we specify our data but also give a column by which to **group the summaries**.

## **Getting Complicated**

We can combine tidyverse functions together. We could filter before summarising the data, and even make multiple grouped summaries.

```
summarise(
  group_by(filter(starwars, mass < 400), species),
  mean_height = mean(height, na.rm = TRUE),
  sd_height = sd(height, na.rm = TRUE),
  n = n()
)</pre>
## # A tibble: 31 x 4
```

### Interim Recap

#### We've learned...

- Why we summarise data and recapped on basic descriptive statistics.
- Why we might use R to summarise our data.
- How to pass **optional arguments** to functions (e.g. na.rm in mean() and sd()).
- How to make **separate summaries per group** using group\_by().
- How to combine tidyverse functions together to get the summaries we need.

# Analysing Data

### Some Background

Data analysis is surprisingly one of the easiest parts of working with R.

- Once your data is in the correct (long) format, analysis using any test is highly consistent.
- We rely on a formula interface like this:

```
DV ~ IV
```

- Our dependent variable/predicted variable goes to the left of the ~ (tilde), while our independent variables or predictors go to the right.
- After this we specify our data:

```
DV ~ IV, data
```

We then apply a function to our formula which is the name of our test. There's some minor options we can choose within tests, but that's pretty much it!

### **Correlations**

#### The Data

Let's check out the **starwars** data set again. We'll use this for our tests.

```
starwars <- starwars %>% filter(mass < 500)
```

We will use the height and mass columns, looking at whether mass is associated with height.

```
## Rows: 58
## Columns: 14
                                                   <chr> "Luke Skywalker", "C-3PO", "R2-D2", "Darth Vader", "Leia Or...
## $ name
## $ height
                                                   <int> 172, 167, 96, 202, 150, 178, 165, 97, 183, 182, 188, 228, 1...
## $ mass
                                                    <dbl> 77.0, 75.0, 32.0, 136.0, 49.0, 120.0, 75.0, 32.0, 84.0, 77...
## $ hair_color <chr> "blond", NA, NA, "none", "brown", "brown, grey", "brown", N...
## $ skin_color <chr> "fair", "gold", "white, blue", "white", "light", "...
## $ eye_color <chr> "blue", "yellow", "red", "yellow", "brown", "blue", "blue", "
## $ birth year <dbl> 19.0, 112.0, 33.0, 41.9, 19.0, 52.0, 47.0, NA, 24.0, 57.0, ...
## $ sex
                                                   <chr> "male", "none", "male", "female", "male", "female", "female", "
                                                    <chr> "masculine", "masculine", "masculine", "masculine", "femini...
## $ gender
## $ homeworld
                                                   <chr> "Tatooine", "Tatooine", "Naboo", "Tatooine", "Alderaan", "T...
                                                    <chr> "Human", "Droid", "Droid", "Human", "Human
## $ species
```

### Correlation

- Here, we aren't predicting any one variable from the other, so both variables go to the right of the tilde.
- We add multiple variables with a +.
- We choose the type of correlation we want (e.g. Pearson, Spearman) with the method.

```
cor.test(~ height + mass, starwars, method = "pearson")

##

## Pearson's product-moment correlation

##

## data: height and mass

## t = 8.7853, df = 56, p-value = 4.018e-12

## alternative hypothesis: true correlation is not equal to 0

## 95 percent confidence interval:

## 0.6260700 0.8520232

## sample estimates:

## cor

## 0.7612612
```

### Tests of Difference

## 5 S003 control daylater

## 6 S003 control monthlater

- We'll use some different data here on out.
- Let's assume this data looks at giving people a placebo or drug, and tests the effect of that drug at two different time points.
- We care about improvements in reaction times.

393.

368.

```
mixed_data <- read_csv(here("data", "mixed_factorial.csv"))</pre>
head(mixed data)
## # A tibble: 6 x 4
     id
           drug
##
                   time
                                  rt
##
     <chr> <chr>
                   <chr>
                               <dbl>
## 1 S001 control daylater
                                431.
          control monthlater
## 2 S001
                                421.
## 3 S002 control daylater
                                372.
## 4 S002 control monthlater
                                350.
```

### t-tests

### One-sample t-test

- We have only one variable here, so we don't even need a formula.
- We compare the mean of this variable against a specified baseline mean (here 400).

```
##
## One Sample t-test
##
## data: mixed_data$rt
## t = -9.5311, df = 479, p-value < 2.2e-16
## alternative hypothesis: true mean is not equal to 400
## 95 percent confidence interval:
## 376.3267 384.4193
## sample estimates:
## mean of x
## 380.373</pre>
```

### t-tests

### Independent-samples t-test

- Do reaction times vary depending on the drug given to participants?
- We test reaction times predicted by drug, with a regular t-test where variances are assumed to be equal (var.equal = TRUE).

```
t.test(rt ~ drug, mixed data, var.equal = TRUE)
##
##
      Two Sample t-test
##
## data: rt by drug
## t = 3.732, df = 478, p-value = 0.0002128
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
    7,18135 23,15251
##
## sample estimates:
    mean in group control mean in group treatment
##
##
                  387,9564
                                          372.7895
```

### t-tests

#### Paired t-test

- Do reaction times vary over time (i.e. practice)?
- We test reaction times predicted by the time of testing. This is a paired test (paired = TRUE) and a regular t-test where variances are assumed to be equal (var.equal = TRUE).

```
t.test(rt ~ time, mixed_data, paired = TRUE, var.equal = TRUE)

##

## Paired t-test

##

## data: rt by time

## t = 18.348, df = 239, p-value < 2.2e-16

## alternative hypothesis: true difference in means is not equal to 0

## 95 percent confidence interval:

## 49.10644 60.91970

## sample estimates:

## mean of the differences

## 55.01307</pre>
```

### One-way ANOVA

#### Between-subjects

- What if we had **more than two groups** for the drug condition? We use an ANOVA.
- We simply change the test function to aov() (Analysis Of Variance)
- We need to summarise the model results here to get a regular ANOVA output.

### One-way ANOVA

## Residuals 239 257842

## ---

#### Within-subjects

• What if we have more than two groups and a within-subjects design?

1079

## Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.05 '.' 0.1 ' ' 1

• We do the same as before, but need to add an **Error term** to the formula. This states that we adjust our errors to account for the fact scores in each group belong to the same participant (i.e. **id** in our data).

### Two-Way ANOVA

#### Mixed

```
summary(aov(rt ~ time * drug + Error(id), mixed_data))
##
## Error: id
##
        Df Sum Sq Mean Sq F value Pr(>F)
## drug 1 27604 27604 20.13 1.13e-05 ***
## Residuals 238 326365 1371
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Error: Within
           Df Sum Sq Mean Sq F value Pr(>F)
##
## time 1 363173 363173 806.3 <2e-16 ***
## time:drug 1 150636 150636 334.4 <2e-16 ***
## Residuals 238 107206 450
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

## Throw Away the Alphabet Soup

All of the statistical tests you know (e.g. *t*-tests, ANOVA, chi-square) are just extensions of the **general linear model**. This is the most important thing you can learn to use in statistics.

Learn the mean and *variance* of some measurement by using an additive combination of other measurements.

- The **geocentric model of applied statistics**: used wisely, can be useful. But we shouldn't read too much into the numbers produced. They're almost certainly wrong because we can't (and shouldn't) model all sources of variance.
- Predict a **linear relationship** between one or more variable(s) and a continuous (e.g. scale) dependent variable.
- Predictor variables can be continuous or categorical.

### **Linear Regression**

Takes the general form:

$$Y = \alpha + \beta X + e$$

- Outcome Y = intercept + (slope  $\times$  X) + residual error
- **Residuals** e = distance of observed values from predicted values
- *Note*: We do not fit a perfect model, hence the error term. This is a good thing, otherwise we are probably **overfitting** to our data; relying too much on our observed sample to draw infferences.

### **Linear Regression**

Takes the general form:

$$Y = \alpha + \beta X + e$$

- The **intercept**,  $\alpha$ , is usually the point on the y-axis at the lowest value of X (usually 0).
- The **slope**,  $\beta$ , corresponds to how much Y increases by for every increment in X.
- The **error**, *e*, corresponds to a constant by which to add to our estimates accounting for additional variation from other sources that we do not model.

### **Linear Regression**

starlm <- lm(height ~ mass, starwars)</pre>

Fit the model predicting height from weight from the starwars data.

$$Y = lpha + eta X + e$$
  $height = intercept + slope imes mass + error$ 

```
summary(starlm)
##
## Call:
## lm(formula = height ~ mass, data = starwars)
##
## Residuals:
      Min
               10 Median 30
##
                                     Max
## -53.369 -6.816 2.042 13.851 44.719
##
  Coefficients:
##
              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 103.5133 8.5937 12.045 < 2e-16 ***
               0.9327 0.1062 8.785 4.02e-12 ***
## mass
## ---
```

## Comparing tests we know...

#### Correlation

```
broom::tidy(cor.test(~ height + mass, starwars, method = "pearson"))
## # A tibble: 1 x 8
##
    estimate statistic p.value parameter conf.low conf.high method alternative
               <dbl>
##
  <dbl>
                       <dbl>
                                <int>
                                       <dbl> <dbl> <chr> <dbl> <chr>
## 1 0.761 8.79 4.02e-12
                                  56
                                       0.626
                                                0.852 Pearson'... two.sided
broom::tidy(lm(height ~ mass, starwars, method = "pearson"))
## Warning in lm(height ~ mass, starwars, method = "pearson"): method = 'pearson'
## is not supported. Using 'qr'
## # A tibble: 2 x 5
              estimate std.error statistic p.value
## term
                      <chr>
              <dbl>
##
## 1 (Intercept) 104. 8.59 12.0 3.53e-17
         0.933 0.106 8.79 4.02e-12
## 2 mass
```

## Comparing tests we know...

#### t-tests

```
broom::tidy(t.test(rt ~ drug, mixed_data, var.equal = TRUE))
## # A tibble: 1 x 10
    estimate estimate1 estimate2 statistic p.value parameter conf.low conf.high
                        <dbl>
                              <dbl>
                                                          <dbl>
##
   <dbl>
               <dbl>
                                         <dbl>
                                                  <dbl>
                                                                   <dbl>
## 1 15.2 388. 373. 3.73 0.000213
                                                   478
                                                        7.18
                                                                23.2
## # ... with 2 more variables: method <chr>, alternative <chr>
broom::tidy(summary(lm(rt ~ drug, mixed_data)))
## # A tibble: 2 x 5
         estimate std.error statistic p.value
##
    term
##
    <chr>
                   <dbl>
                            <dbl>
                                     <dbl>
                                             <dbl>
## 1 (Intercept)
                   388. 2.87 135. 0
## 2 drugtreatment
                -15.2 4.06 -3.73 0.000213
```

• *t* statistics match exactly.

### Comparing tests we know...

#### **ANOVA**

```
summary(aov(rt ~ drug, mixed_data))
       Df Sum Sq Mean Sq F value Pr(>F)
##
## drug 1 27604
                      27604 13.93 0.000213 ***
## Residuals 478 947379
                     1982
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
broom::tidy(lm(rt ~ drug, mixed_data))
## # A tibble: 2 x 5
        estimate std.error statistic p.value
## term
##
   <chr>
           <dbl>
                          <dbl> <dbl>
                                          <dbl>
## 1 (Intercept) 388. 2.87 135. 0
## 2 drugtreatment -15.2 4.06 -3.73 0.000213
```

• *t* to *F* is just *t* squared. So, 3.732 squared = 13.93...

## Bye!



Effect sizes are easily handled by the {effectsize} package. Super-easy ANOVAs are done using the {afex} package.