

Worker queens? Behavioral flexibility of queens in the little fire ant Wasmannia auropunctata

Yarira Ortiz-Alvarado^{1*}, Bert Rivera-Marchand²

¹University of Puerto Rico, Puerto Rico, ²InterAmerican University of Puerto Rico, Puerto Rico

Submitted to Journal:

Frontiers in Ecology and Evolution

Specialty Section:

Behavioral and Evolutionary Ecology

ISSN:

2296-701X

Article type:

Original Research Article

Received on:

16 Apr 2020

Accepted on:

02 Jul 2020

Provisional PDF published on:

02 Jul 2020

Frontiers website link:

www.frontiersin.org

Citation

Ortiz-alvarado Y and Rivera-marchand B(2020) Worker queens? Behavioral flexibility of queens in the little fire ant Wasmannia auropunctata. *Front. Ecol. Evol.* 8:241. doi:10.3389/fevo.2020.00241

Copyright statement:

© 2020 Ortiz-alvarado and Rivera-marchand. This is an open-access article distributed under the terms of the <u>Creative Commons Attribution License (CC BY)</u>. The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

This Provisional PDF corresponds to the article as it appeared upon acceptance, after peer-review. Fully formatted PDF and full text (HTML) versions will be made available soon.

Worker queens? Behavioral flexibility of queens in the little fire ant Wasmannia 1 2 auropunctata Yarira Ortiz-Alvarado^{1*} and Bert Rivera-Marchand² 3 4 ¹University of Puerto Rico, Department of Biology, Rio Piedras, Puerto Rico ²Inter American University, Department of Natural Sciences and Mathematics, Bayamon, Puerto 5 6 Rico 7 *Correspondence: Yarira Ortiz-Alvarado, PhD. 8 9 y.ortizal@gmail.com urn:lsid:zoobank.org:act:8A719B0D-0E2C-4661-AA0F-A05367A1D837 10 Keywords: behavior, flexibility, invasive, ant, queen, juvenile hormone, vitellogenin 11 12 **Abstract** Many species of social Hymenoptera demonstrate behavioral flexibility, where older workers 13 that typically forage can revert to younger worker tasks, such as nursing, when these are absent. 14 15 This flexibility is typical of the sterile worker class, yet rare in queens. In the little fire ant (Wasmannia auropunctata), queens have been reported to perform only egg laying. We 16 examined behavior of gueens of W. auropunctata after demographic manipulation. When half of 17 the workers were removed from the colony, queens were observed caring for eggs, larvae and 18 pupae as well as eating outside of the nest, like forager workers. We examined the relationship 19 20 between these atypical queen behaviors and their juvenile hormone binding protein (JHbp) and vitellogenin (Vg) expression via QRT-PCR method. JHbp and Vg expression decreased when 21 queens were performing worker tasks, resembling the expected expression pattern of typical 22 sterile workers. Flexibility in queen behaviors in the little fire ant may be an important adaptation 23 to changing environments. As a significant invasive species, such adaptation may increase the 24 probability of colony survival during propagation. Our results not only present new insights in 25 behavioral flexibility in social insects, but also increases our understanding of the success of this 26 significant invasive species. 27 1. Introduction 28 Eusocial insects are characterized by having reproductive division of labor (Wilson 1971). 29 Within the colony one or more individuals carry out egg laving while sterile workers perform 30 nest related tasks including queen and brood care (i.e. nursing), defense and foraging (Gordon 31 1996). Division of labor in workers may be associated with age or morphological differences 32 (Hölldobler & Wilson 1990). Nevertheless, there are species where workers show behavioral 33 flexibility, performing tasks that are not typical of their age or morphology. For example, in 34 honey bees (Apis mellifera), nurses are known to forage precociously when foragers are absent, 35 while forgers may revert to nursing according to the needs of the colony (Robinson 1992). 36 Similar behaviors has been observed in the eusocial wasp *Polybia occidentalis* (O'Donnell

- 38 2001). Workers of the ant *Pheidole dentata* are found to increase their behavioral repertory as
- 39 they age, where older workers perform tasks typical to younger individuals, yet the young
- 40 workers are not proficient in older worker tasks (Calabi & Traniello 1989; Seid & Traniello
- 41 2006; Mertil & Traniello 2009). Although behavioral flexibility in workers is an important
- adaptation that increases the chances of colony survival when the worker population decreases, it
- has not been reported in queens (Rüppell et al. 2002).
- Queens produce eggs throughout most of their life. In species where the colony goes through a
- 45 founding stage or independent colony foundation, queens perform worker type tasks until
- workers are reared. For example, in the red imported fire ant, *Solenopsis invicta*, the founding
- 47 queen starts with nest construction, lays and tends the first batch of eggs until these emerge as
- 48 adult workers, which take over worker duties. From this point on queens are known to only lay
- 49 eggs (Tschinkel 2006). Contrastingly, in species that reproduce through colony budding or
- 50 dependent colony foundation, where one or more queens depart from the main colony with a
- 51 group of workers, the queens do not experience a founding stage (Keller 1991; Peeters & Ito
- 52 2001). Since these queens keep a group of workers at all times it is not expected that they
- 53 perform worker-like tasks.
- We examined queen behavior in *Wasmannia auropunctata*, the little fire ant, a native of South
- America and an aggressive invasive species on all other continents except Antarctica (Wetterer
- & Porter 2003; Le Breton et al. 2003; Mikheyev et al. 2008). Its colonies are composed of 200-
- 57 500 monomorphic workers and one to twelve larger queens, and reproduce by colony fission
- 58 (Wetterer & Porter 2003; Foucaud et al. 2006; Mikheyev et al. 2009). Older workers, which
- 59 typically carry out foraging duties, demonstrate behavioral flexibility by performing nursing
- duties when young workers (i.e. nurses) are absent (Rivera-Marchand & Fernández-Casas
- 61 unpublished). The first objective of this study was to determine if queens of W. auropunctata
- 62 demonstrate behavioral flexibility, performing worker tasks when necessary. Since colonies of
- the little fire ant do not experience a solitary founding stage (Wetterer & Porter 2003; Foucaud et
- al. 2006; Mikheyev et al. 2009), queens typically have no need to perform worker tasks. We did
- not expect queens to perform worker duties until we had observed queens manipulating eggs.
- Based on these preliminary observations we expected that in the absence of workers, queens of
- W. auropunctata should have the behavioral flexibility to perform worker tasks.
- Reproductive division of labor in eusocial Hymenoptera (ants, bees and wasps), is under
- endocrine control (e.g. JH and Vg; Bloch et al. 2002; Amsalem et al. 2014). Juvenile hormone
- 70 (JH) and vitellogenin (Vg) have important roles in regulating insect physiology (Dolezal et al.
- 71 2009; Dolezal et al. 2012) such as development, reproduction, and behavior (Robinson & Vargo
- 72 1997; Dong et al. 2009; Azevedo et al. 2016). JH, considered a master hormone, has been found
- to control behavioral development in honey bees (Robinson & Vargo 1997; Sullivan et al. 2000).
- 74 It influences physiology in queens and guarding behavior in workers of primitive eusocial wasp
- 75 Polistes canadensis (Giray et al. 2004). JH also affects queen maturation and reproduction in the
- 76 invasive ant S. invicta where, high levels of JH induces alates to begin oogenesis (Vargo &
- Laurel 1994; Brent & Vargo 2003; Lu et al. 2009). Vg is a yolk precursor protein; its production
- 78 is typically used to produce egg yolk by oviparous animals (Amdam et al. 2003), but it may also

- affect behavior (Nelson et al. 2007). In honey bees Vg has a role in the reproductive division of
- 80 labor where concentrations are correlated with the hierarchy of the hive and reproductive
- division of labor (Corona et al. 2007; Nelson et al. 2007). Similar trends of Vg expression are
- seen in ants of S. inivicta (Lewis et al. 2001; Lu et al. 2009) and Pogonomyrmex spp. (Corona et
- al. 2013; Libbrecht et al. 2013). Therefore, the second objective of this study was to measure
- gene expression of JH and Vg in relation to the tasks performed by queens. We expected egg
- laying queens to have higher levels of JH and Vg than worker-like queens.

2. Materials and Methods

2.1 Samples

86

87

101

- Nests (N=20) of the little fire ant W. auropunctata were collected from dry twigs and leaf litter in
- 89 the northern region of the Caribbean island of Puerto Rico. They were housed in artificial nests
- 90 which consisted of plastic boxes (25cm x 13cm x 7.5cm) coated with Fluon (Northern Products
- 91 Inc., Alsip, IL.) on the sides. The boxes' lids were perforated with a pin to allow air exchange.
- The nesting area within the nest box consisted of a 3cm² piece of thin (less than 1cm thick) wood
- elevated 0.5 cm by a strip of clay placed along the edges of the wood. Each nest was kept at
- 94 25°C, a relative humidity (RH) between 80% and 85%, and 12 hour light cycles. Nests were fed
- 95 daily with 0.5 g of feeding mixture containing agar, eggs, honey and vitamin supplements
- 96 (Hölldobler & Wilson 1994) placed in a feeding arena within the box at approximately 10 cm
- 97 from the nesting area. Nests used in the study had multiple queens, eggs, larvae and pupae. We
- 98 marked 52 queens on the thorax or abdomen using unique color combinations of nail polish.
- 99 Queens are easily distinguishable from workers by being three to four times larger (workers 1.2-
- 1.5 mm long, queens 4.5-5 mm; Wetterer & Porter 2003).

2.2 Behavioral flexibility of queens assays

- Six nests (N=6) with a total of 19 queens were used for behavioral flexibility assays. Colonies
- were kept in the artificial nests three days prior to the start of experiment and fed daily. During
- the experiment observation period, queens were observed for 10 minutes daily for a total of
- nineteen days; food was removed after each observation period. The 10 minutes observation
- period was determined after observing that the proportion of behaviors does not vary
- significantly in 10, 15, 20 and 30 minute intervals. Observations were made for different tasks,
- including egg laying (a typical behavior), nursing (i.e. manipulating brood) and foraging (i.e.
- queens seen feeding in the designated arena), the latter two being non-typical behaviors. During
- the 10 minutes observational period, we tabulated by number of events, i.e., number of eggs laid,
- number of times brood was manipulated, and number of times gueens walked to the feeding
- arena and was seen feeding. Control observation periods were performed for a period of five
- days. After the first five days of observations, worker population per nest was estimated via nest
- pictures. We then randomly culled approximately 50% of workers from the nest to simulate
- natural events in the wild. Preliminary observations (Rivera-Marchand & Fernández-Casas
- unpublished) indicated that the worker caste is equally divided between nurses and foragers.
- Daily 10 minutes observations continued for fourteen days (Table 1). The remaining nests were
- used to measure *JHbp* and *Vg* gene expression.

2.3 Bioinformatic analysis and primer design

- Primers were designed for gene sequences related to JH and Vg expression. Since JH is a
- terpenoid, its gene expression levels were determined indirectly by measuring Juvenile hormone
- binding protein (JHbp), an associated protein. JHbp is directly correlated with the onset of JH
- production in the hemolymph (Kramer et al. 1976; Shemshedini & Wilson 1990) because it
- prevents the absorption and enzymatic hydrolysis of JH, thereby maintaining a steady reservoir
- of the hormone in the hemolymph. As a consequence, free JH is virtually absent (Roe
- 26 &Venkatesh 1990; Tan 2007). Also, JHbp/JH interaction is specific and of high affinity (KD=10
- ⁹M), more than 99% of JH is bound to JHbp (De Kort & Granger 1996; Tan 2007). Other studies
- have further suggested this direct involvement as well (Prestwich et al. 1996; Hagai et al. 2007).
- Sequences for JHbp and Vg of W. auropunctata were obtained from NCBI Gene Bank. Vg
- sequences (XM_011697672.1, XM_011697673.1) were aligned using MAFFT (Multiple
- sequence alignment tool: Katoh et al. 2009). Primers (Table 2) were designed using primer3
- from NCBI (Ye et al. 2012) with the obtained consensus sequence for Vg and the JHbp sequence
- 133 (XM_011708554). Actin and GAPDH (Glyceraldehyde-3-Phosphate Dehydrogenase) were used
- as housekeeping genes (Wong & Medrano 2005; Scharlaken et al. 2008).

2.4 RNA extraction, cDNA and qPCR of JHbp and Vg.

- Fourteen nests (N=14), different from the ones used in the first behavioral assay, were used with
- a total of 33 queens. Nests were randomly assigned to control or experimental groups (nest with
- workers removed), behavioral assays were repeated. Seven days after worker removal, queens
- were collected by tasks; from control nests n=15 and from experimental nests n=18 and placed in
- a microtube with 20 μL of RNAlater reagent (Qiagen Valencia, CA) stored at -80°C for later
- 141 RNA extraction. Afterwards, samples were placed in a sterilized microtube and mechanically
- homogenized. RNA extraction was performed using the RNeasy Mini Kit (Qiagen). Extracted
- RNA was quantified for each sample in $\mu g/\mu L$ units using a Nanophotometer (Implen, Westlake
- Village, CA). RNA was normalized to a concentration (10 μg/μL) in a final volume of 20 μL and
- treated with DNase 1, following BioLabs (Ipswich, MA) protocol to remove any DNA
- contamination. cDNA was synthesized from the normalized RNA using iScript Reverse
- 147 Transcription Supermix for RT-qPCR (Bio-Rad Hercules, CA.) following the manufacturer's
- protocol with 10µL of RNA as a template. cDNA synthesis was verified in an electrophoresis
- 149 1% ETBR-gel.

119

- 150 qPCR was performed using the MJ Mini-Opticon Real-Time PCR (Bio-Rad) following the
- standard protocol of forty cycles; denature at 95°C for 10s, annealing at 56°C for 30s and
- elongation at 72°C for 15s x40, with post-amplification melt curve analysis. As a standard for
- quantification purposes, actin and GAPDH were used as reference genes (Wong & Medrano
- 2005; Scharlaken et al. 2008). Primer efficiency was calculated using the standard curve analysis
- method where 1 µl of each cDNA sample were pooled and serial diluted in five points at 1:10.
- Reactions were prepared with 2 µL of first strand cDNA as a template in a master mix of 1 µL of
- 157 forward and reverse primers per gene at [10 nM] and 5 µL of iTaq Universal SYBR Green
- Supermix (BioRad) in a final volume of 10 μL. Relative gene expression was calculated using
- the geometric mean analysis method (Vandesompele et al. 2002), using the following equation:

Relative gene expression = $\frac{(E_{GOI})^{\Delta CtGOI}}{GeoMean}[(E_{REF})^{\Delta CtREF}],$ 160 E= primer efficiency, GOI= gene of interest, GeoMean= geometric mean and REF= reference 161 162 gene. Δ Ct was calculated using the average Ct values of the control group for each gene (calibrator Ct). The relative expression values presented are relative to the control group. 163 164 2.5 Statistical analysis Behavior Analysis. For each task, relative probability was calculated by the number of queens 165 performing a task with the total number of queens. Differences in relative probability between 166 before and after worker removal were calculated with a Wilcoxon signed-rank test. To compare 167 frequency of queen behaviors, frequency of typical and non-typical tasks were calculated by 168 counting the number of events by queens before and after manipulation. Frequencies were 169 compared using a Friedman test and Dunn's test as a Post-Hoc method. 170 171 Gene Expression. In order to compare relative expression between control and experimental samples, a Wilcoxon signed-rank test was used to measure differences in expression of JHbp and 172 Vg. Here experimental samples were considered queens performing both nursing and/or 173 foraging. To verify relative gene expression of JHbp and Vg differences among all tasks (egg 174 laying, nursing and foraging), a Kruskal-Wallis rank sum test was used with Dunn's test as a 175 Post-Hoc method. 176 Data was analyzed using the statistical program R (R Core Team 2014) v. 3.5.2 (2018-12-20) 177 and the package agricolae (Statistical Procedures for Agricultural Research) v. 1.3-1. Graphs 178 were done in Graph Pad Prism 6.0, (GraphPad software, La Jolla California USA). 179 180 3. Results 3.1 Behavior Analysis 181 In the first three days of observations queens laid eggs while workers performed typical tasks. 182 After worker removal queens performed worker tasks for nine consecutive days, which 183 coincided with pupae emergence. During the period after worker removal in which queens 184 behaved as workers, egg laying decreased significantly (Figure 1A: W = 62.5, p-value = 0.01) 185 while nursing and foraging behaviors increased significantly (Figure 1B: W = 2.5, p-value = 186 0.002; Figure 1C: W = 15, p-value = 0.04). Of note, during the behavior observation period, 187 some of the queens in the nests remained idle. 188 Throughout the experiment when queens performed non-typical tasks, egg laying decreased until 189 new workers emerged (Figure 2). After new workers emerged, queens returned to egg laying and 190 stopped performing worker tasks. Post hoc comparisons indicate that egg laying frequency from 191 day four through ten are significantly lower (p-value <0.05) than egg laying before worker 192 removal. As egg laying decreased, the frequency of nursing behavior by queens increased 193 significantly (p-value <0.05) from the fourth through tenth day of observations. While 194 performing nursing, queens were seen manipulating eggs, larvae and pupae. Queens (N=6) were 195 also seen foraging during days six through nine. Only on day seven was foraging frequency 196 significantly higher than the rest of the experiment. On day eleven the queens returned to egg 197

laying and by day fourteen queen behaviors resembled the behaviors during control period (p-value >0.05), thus showing that the nests endured the experimental period and returned to a typical behavioral pattern.

3.2 Gene Expression

There's a significant difference between the control and the experimental samples for *JHbp* and Vg expression. Queens from experimental nests performing worker related tasks have a lower relative gene expression of *JHbp* and Vg related to control nest queens (Figure 3A: W = 270, p-value = <0.001; Figure 3C: W = 270, p-value = <0.001). When relative gene expression of *JHbp* and Vg were compared between tasks, it shows a difference between egg laying compared to nursing and foraging for both *JHbp* and Vg; however, there is no difference in relative gene expression of *JHbp* and Vg in queens performing nursing or foraging tasks (Figure 3B:, Kruskal-Wallis chi-squared = 24.21, df = 2, p-value = <0.001; Figure 3D; Kruskal-Wallis chi-squared = 25.19, df = 2, p-value = <0.001).

4. Discussion

198

199 200

201

202

203204

205

206

207

208

209 210

- Although behavioral flexibility is known to be a trait of workers in many species of social
- 213 Hymenoptera (Gordon 1991; Robinson 1992; Giray et al. 2004; Seid & Traniello 2006), queens
- of the little fire ant also demonstrate behavioral flexibility. When worker population decreases
- 215 they perform worker tasks, principally nursing and, with less frequency, foraging (Figure 2).
- 216 Although both behaviors were not initially expected, foraging in particular was a surprising
- outcome. Both egg laying and nursing are tasks done within the nest, so we hypothesize that the
- reason queens shift primarily to nursing is that it is less risky. Queens are risk aversive, and tend
- 219 to perform nursing rather than foraging because the former is a less risky worker task. To our
- knowledge, this is the first time queens have been observed demonstrating behavioral flexibility
- after the founding stage. Given that W. auropunctata queens never experience a solitary
- founding stage (Foucaud et al. 2006; Mikheyev et al. 2009) and thus under typical conditions
- never perform worker-like duties, our results are even more remarkable.
- During the experiment, there was an apparent transition phase, where queens gradually decreased
- their egg laying while increasing worker tasks. It is likely that queens may sense the shortage of
- workers due to a decrease in contact with workers. Studies have shown that ants typically
- communicate by cuticle hydrocarbons which are perceived by the olfactory organs (Vander Meer
- 228 et al. 1989; Saïd et al. 2005; Ichinose & Lenoir 2009; Bos et al. 2010), hence a decrease in the
- amount of cuticle hydrocarbons perceived might be an indicator to queens that the worker
- population has decreased. Maximum worker behavior frequencies were observed on the eighth
- day of the experiment, coinciding with minimum egg laying frequencies. We observed individual
- 232 queens performing both nursing and foraging tasks. After the ninth day an increase in egg laying
- and a decrease in nursing were observed. From day six to nine, we observed queens eating at the
- feeding arena. Queens seem to decrease investments in reproduction in order to invest in brood
- care. As adult workers emerge and take over brood care tasks, queens begin investing in egg
- laying again. Since energy expenditures due to reproduction tend to be high in social insects
- 237 (Oster & Wilson 1978), queens of the little fire ant may not be able to invest in both egg laying

- and brood care. It is possible that these queens are diverting energy typically used for egg
- production to carry other nest duties. In our study queens performing worker tasks were not
- observed laying eggs. Moreover, studies on the reproductive biology of W. auropunctata have
- shown that all queens within nests produce viable eggs (de Ulloa 2003). Therefore, we worked
- under the assumption that queens in the experiment were inseminated.
- 243 The changes in behaviors of queens were associated with changes in the expression of *JHbp*
- gene and Vg. When queens are performing typical tasks, the levels of JHbp and Vg tend to be
- 245 higher, than when a queen displays worker behaviors. This suggests regulation by molecular and
- physiological mechanisms on behaviors, such as regulation by hormones/protein through gene
- expression. In honey bees, queens tend to have low levels of JH (Fahrbach et al. 1985; Robinson
- et al. 1991), the opposite is seen in the little fire ant queens, where queens performing egg laying
- had higher gene expression of JHbp gene than those performing worker related tasks (Figure
- 250 3A). When studying Vg, this protein has been found to have an effect on reproductive division of
- labor on honey bees, where queens have higher levels of Vg which decrease in nurses and
- foragers (Nelson et al. 2007; Page & Amdam 2007). This is not an exception in other insects
- such as S. invicta and P. canadensis (Brent & Vargo 2003; Sumner et al. 2006). The same
- pattern has been found in W. auropunctata queens, where queens have higher expression levels
- of Vg when laying eggs compared to the expression levels of Vg in the performance of worker
- related tasks (Figure 3B). Furthermore, studies have shown precocious foraging in workers
- induced by downregulation Vg using RNAi (Nelson et al. 2007; Antonio et al. 2008), suggesting
- 258 changes in behavior are mediated by Vg.
- 259 The relationship between JH and Vg has been studied before (Robinson & Vargo 1997; Lewis et
- al. 2001; Barchuk et al. 2002; Brent & Vargo 2003). In most insects, there is a positive
- relationship between JH and Vg; increasing levels of JH cause an increase in Vg synthesis
- 262 (Barchuk et al. 2002; Toth & Robinson 2007). In S. invicta queens, high levels of JH and Vg are
- 263 correlated. JH has been found to be an important promoter of vitellogenic oogenesis (Brent &
- Vargo 2003; Lu et al. 2009) therefore, a decrease of JH seems to interrupt egg production. Our
- results show a similar relation with JH (*JHbp*) and Vg as queens' reduction in Vg expression
- seems to correspond to a reduction in JHbp gene expression.
- Behavioral flexibility in social insects ensures survival, particularly where environmental
- 268 changes may affect colony demography. The adaptive value of behavioral flexibility in workers
- has been evidenced in honey bees (Robinson 1992; Scheiner et al. 2004), different *Pheidole*
- species (Seid & Traniello 2006; Mertil & Traniello 2009) and the harvester ant (*Pogonomyrmex*
- barbatus; Gordon 1991 & 2002), where increased needs for one task due to environmental
- changes may lead to a behavioral response. Flexibility in queen behaviors in the little fire ant
- 273 may be an important adaptation to changing environments. As a native to the tropics, this ant
- 274 may face frequent disturbances that may deplete worker population. Moreover, the colonies of
- 275 this ant tend to move their nests often (Wetterer & Porter 2003). During the process of moving,
- workers such as foragers may be left behind as the nest is moved. The probability of colony
- survival may increase with queens performing worker duties as an adaptation for the population
- 278 reduction that might occur during propagation. Evidence of this increased probability of survival

- can be seen in the final phase of the experiment where queens returned to their typical behaviors
- as the worker population increased. The results of this study not only present new insights in
- behavioral flexibility in social insects, but also increases our understanding of the success of this
- important invasive species. The little fire ant, is an important invasive species and agricultural
- pest that has colonized many areas around the world. Various ecological and reproductive
- adaptations are associated to its success and the unique queen behavior of this study adds to our
- 285 knowledge of the suite of adaptations allowing this ant to be a successful invader.

5. Acknowledgements

286

293

298

301

304

305

306

307

308

309

310

311

312

313

- We would like to thank Rafael Fernández-Casas, Carlos Ortiz-Alvarado, Emily Díaz-Iglesias,
- 288 Luis Marrero-Ramos, Angel Rivera-Colón and Yoselyn Rodríguez-Cruz aka the Wasmannia
- 289 *Team*, for their effort in nest collection, establishment and extractions. We also thank Timothy
- 290 Hendricks for the laboratory facilities, Rafael Canales-Pastrana and Bárbara González for their
- 291 help in data analysis. Lastly we thank James D. Ackerman for helping with the revisions of
- 292 previous versions of the manuscript.

6. Author Contributions Statement

- The experiments presented in this article were conceived and designed by YOA and BRM. All of
- the experiments were performed in Puerto Rico by YOA. Data analysis on behavior and gene
- expression was performed by YOA under the supervision of BR. The writing of this manuscript,
- preparation of the figures and editing was performed by YOA and BRM.

7. Conflict of Interest Statement

- 299 The authors declare that the research was conducted in the absence of any commercial or
- financial relationships that could be construed as a potential conflict of interest.

8. Data Availability

The original contributions presented in this study are publicly available. This data can be found here: https://doi.org/10.5061/dryad.j6q573nb2

9. References

- 1. Amdam, G. V., Norberg, K., Hagen, A., & Omholt, S. W. (2003). Social exploitation of vitellogenin. *Proceedings of the National Academy of Sciences*, 100(4), 1799-1802.
- 2. Amsalem, E., Malka, O., Grozinger, C., & Hefetz, A. (2014). Exploring the role of juvenile hormone and vitellogenin in reproduction and social behavior in bumble bees. *BMC Evolutionary Biology*, 14(1), 45.
- 3. Antonio, D. S. M., Guidugli-Lazzarini, K. R., Do Nascimento, A. M., Simões, Z. L. P., & Hartfelder, K. (2008). RNAi-mediated silencing of vitellogenin gene function turns honeybee (*Apis mellifera*) workers into extremely precocious foragers. *Naturwissenschaften*, 95(10), 953-961.
- Azevedo, D. O., de Paula, S. O., Zanuncio, J. C., Martinez, L. C., & Serrão, J. E. (2016).
 Juvenile hormone downregulates vitellogenin production in *Ectatomma tuberculatum*

- 316 (Hymenoptera: Formicidae) sterile workers. *Journal of Experimental Biology*, 219(1), 103-108.
- 5. Barchuk, A. R., Maleszka, R., & Simões, Z. L. P. (2004). *Apis mellifera* ultraspiracle: cDNA sequence and rapid up-regulation by juvenile hormone. *Insect Molecular Biology*, 13(5), 459-467.
- 6. Bloch, G., Wheeler, D. E., & Robinson, G. E. (2002). Endocrine influences on the organization of insect societies. In Hormones, brain and behavior (pp. 195-235).

 Academic Press.
- 7. Bos, N., Guerrieri, F. J., & d'Ettorre, P. (2010). Significance of chemical recognition cues is context dependent in ants. *Animal Behaviour*, 80(5), 839-844.

326

327 328

329

330

334

335

336

337

341

342

343

344

345

346

- 8. Brent, C. S., & Vargo, E. L. (2003). Changes in juvenile hormone biosynthetic rate and whole body content in maturing virgin queens of *Solenopsis invicta*. *Journal of Insect Physiology*, 49(10), 967-974.
- 9. Calabi, P., & Traniello, J. F. (1989). Behavioral flexibility in age castes of the ant *Pheidole dentata*. *Journal of Insect Behavior*, 2(5), 663-677.
- 10. Corona, M., Libbrecht, R., Wurm, Y., Riba-Grognuz, O., Studer, R. A., & Keller, L. (2013). Vitellogenin underwent subfunctionalization to acquire caste and behavioral specific expression in the harvester ant *Pogonomyrmex barbatus*. *PLoS Genetics*, 9(8).
 - 11. Corona, M., Velarde, R. A., Remolina, S., Moran-Lauter, A., Wang, Y., Hughes, K. A., & Robinson, G. E. (2007). Vitellogenin, juvenile hormone, insulin signaling, and queen honey bee longevity. *Proceedings of the National Academy of Sciences*, 104(17), 7128-7133.
- 12. [Dataset] Ortiz-Alvarado, Y. & Rivera-Marchand, B. (2020), Data Repository_Ortiz Alvarado_Rivera-Marchand 2020, Dryad, Dataset,
 https://doi.org/10.5061/dryad.j6q573nb2
 - 13. De Kort, C. A. D., & Granger, N. A. (1996). Regulation of JH titers: the relevance of degradative enzymes and binding proteins. *Archives of Insect Biochemistry and Physiology: Published in Collaboration with the Entomological Society of America*, 33(1), 1-26.
 - 14. de Ulloa, P. C. (2003). Biologia reproductiva de *Wasmannia auropunctata* (R.)(H ymenoptera: formicidae). *Revista de la Academia Colombiana de Ciencias Exactas, Físicas y Naturales*, 27(104), 441-448.
- 15. Dolezal, A. G., Brent, C. S., Gadau, J., Hölldobler, B., & Amdam, G. V. (2009).
 Endocrine physiology of the division of labour in *Pogonomyrmex californicus* founding queens. Animal *Behaviour*, 77(5), 1005-1010.
- 16. Dolezal, A. G., Brent, C. S., Hölldobler, B., & Amdam, G. V. (2012). Worker division of labor and endocrine physiology are associated in the harvester ant, *Pogonomyrmex* 353 *californicus*. *Journal of Experimental Biology*, 215(3), 454-460.
- 17. Dong, S. Z., Ye, G. Y., Guo, J. Y., & Hu, C. (2009). Roles of ecdysteroid and juvenile hormone in vitellogenesis in an endoparasitic wasp, *Pteromalus puparum* (Hymenoptera: Pteromalidae). *General and Comparative Endocrinology*, 160(1), 102-108.

18. Fahrbach, S. E., Giray, T., & Robinson, G. E. (1995). Volume changes in the mushroom bodies of adult honey bee queens. *Neurobiology of Learning and Memory*, 63(2), 181-191.

- 19. Foucaud, J., Jourdan, H., Breton, J. L., Loiseau, A., Konghouleux, D., & Estoup, A. (2006). Rare sexual reproduction events in the clonal reproduction system of introduced populations of the little fire ant. *Evolution*, 60(8), 1646-1657.
 - 20. Giray, T., Giovanetti, M., & West-Eberhard, M. J. (2005). Juvenile hormone, reproduction, and worker behavior in the neotropical social wasp *Polistes canadensis*. *Proceedings of the National Academy of Sciences*, 102(9), 3330-3335.
 - 21. Gordon, D. M. (1991). Variation and change in behavioral ecology. *Ecology*, 72(4), 1196-1203.
 - 22. Gordon, D. M. (1996). The organization of work in social insect colonies. *Nature*, 380(6570), 121-124.
- 23. Gordon, D. M. (2002). The regulation of foraging activity in red harvester ant colonies. *The American Naturalist*, 159(5), 509-518.
 - 24. Hagai, T., Cohen, M., & Bloch, G. (2007). Genes encoding putative Takeout/juvenile hormone binding proteins in the honeybee (Apis mellifera) and modulation by age and juvenile hormone of the takeout-like gene GB19811. *Insect Biochemistry and Molecular Biology*, *37*(7), 689-701.
- 25. Hölldobler B & Wilson EO (1990) The Ants. Harvard University Press Cambridge Mass. 734pp
 - 26. Hölldobler B & Wilson EO (1994) Journey to the ants. Harvard University Press Cambridge Mass. 228pp
 - 27. Ichinose, K., & Lenoir, A. (2009). Ontogeny of hydrocarbon profiles in the ant *Aphaenogaster senilis* and effects of social isolation. *Comptes Rendus Biologies*, 332(8), 697-703.
 - 28. Katoh, K., Asimenos, G., & Toh, H. (2009). Multiple alignment of DNA sequences with MAFFT. *In Bioinformatics for DNA sequence analysis* (pp. 39-64). Humana Press.
 - 29. Keller, L. (1991). Queen number, mode of colony founding, and queen reproductive success in ants (Hymenoptera Formicidae). *Ethology Ecology & Evolution*, 3(4), 307-316.
 - 30. Kramer, K. J., Dunn, P. E., Peterson, R. C., & Law, J. H. (1976). Interaction of juvenile hormone with binding proteins in insect hemolymph. *In The Juvenile Hormones* (pp. 327-341). Springer, Boston, MA.
 - 31. Le Breton, J., Chazeau, J., & Jourdan, H. (2003). Immediate impacts of invasion by *Wasmannia auropunctata* (Hymenoptera: Formicidae) on native litter ant fauna in a New Caledonian rainforest. *Austral Ecology*, 28(2), 204-209.
- 32. Lewis, D. K., Campbell, J. Q., Sowa, S. M., Chen, M. E., Vinson, S. B., & Keeley, L. L. (2001). Characterization of vitellogenin in the red imported fire ant, *Solenopsis invicta* (Hymenoptera: Apocrita: Formicidae). *Journal of Insect Physiology*, 47(6), 543-551.
- 33. Libbrecht, R., Corona, M., Wende, F., Azevedo, D. O., Serrão, J. E., & Keller, L. (2013).
 Interplay between insulin signaling, juvenile hormone, and vitellogenin regulates

maternal effects on polyphenism in ants. *Proceedings of the National Academy of Sciences*, 110(27), 11050-11055.

- 34. Lu, H. L., Vinson, S. B., & Pietrantonio, P. V. (2009). Oocyte membrane localization of vitellogenin receptor coincides with queen flying age, and receptor silencing by RNAi disrupts egg formation in fire ant virgin queens. *The FEBS Journal*, 276(11), 3110-3123.
 - 35. Mertl, A. L., & Traniello, J. F. (2009). Behavioral evolution in the major worker subcaste of twig-nesting *Pheidole* (Hymenoptera: Formicidae): does morphological specialization influence task plasticity?. *Behavioral Ecology and Sociobiology*, 63(10), 1411-1426.
 - 36. Mikheyev, A. S., Tchingnoumba, L., Henderson, A., & Alonso, A. (2008). Effect of propagule pressure on the establishment and spread of the little fire ant *Wasmannia auropunctata* in a Gabonese oilfield. *Diversity and Distributions*, 14(2), 301-306.
 - 37. Mikheyev, A. S., Bresson, S., & Conant, P. (2009). Single-queen introductions characterize regional and local invasions by the facultatively clonal little fire ant *Wasmannia auropunctata*. *Molecular Ecology*, 18(14), 2937-2944.
 - 38. Nelson, C. M., Ihle, K. E., Fondrk, M. K., Page Jr, R. E., & Amdam, G. V. (2007). The gene vitellogenin has multiple coordinating effects on social organization. *PLoS Biology*, 5(3), e62.
 - 39. O'donnell, S. (2001). Worker age, ovary development, and temporal polyethism in the swarm-founding wasp *Polybia occidentalis* (Hymenoptera: Vespidae). *Journal of Insect Behavior*, 14(2), 201-213.
 - 40. Oster GF & Wilson EO (1978) Social Insects. Princeton Univ. Press. 352pp
 - 41. Page Jr, R. E., & Amdam, G. V. (2007). The making of a social insect: developmental architectures of social design. *Bioessays*, 29(4), 334-343.
 - 42. Peeters, C., & Ito, F. (2001). Colony dispersal and the evolution of queen morphology in social Hymenoptera. *Annual review of entomology*, 46(1), 601-630.
 - 43. Prestwich, G. D., Wojtasek, H., Lentz, A. J., & Rabinovich, J. M. (1996). Biochemistry of proteins that bind and metabolize juvenile hormones. *Archives of Insect Biochemistry and Physiology: Published in Collaboration with the Entomological Society of America*, 32(3-4), 407-419.
 - 44. R Core Team (2014). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL http://www.R-project.org/.
 - 45. Robinson, G. E. (1992). Regulation of division of labor in insect societies. *Annual Review of Entomology*, 37(1), 637-665.
 - 46. Robinson, G. E., Strambi, C., Strambi, A., & Feldlaufer, M. F. (1991). Comparison of juvenile hormone and ecdysteroid haemolymph titres in adult worker and queen honey bees (*Apis mellifera*). *Journal of Insect Physiology*, 37(12), 929-935.
 - 47. Robinson, G. E., & Vargo, E. L. (1997). Juvenile hormone in adult eusocial Hymenoptera: gonadotropin and behavioral pacemaker. Archives of Insect Biochemistry and Physiology: *Published in Collaboration with the Entomological Society of America*, 35(4), 559-583.
- 48. Roe, R. M., & Venkatesh, K. (1990). Metabolism of juvenile hormones: degradation and titer regulation. *Morphogenetic Hormones of Arthropods*, *1*, 126-179.

49. Rüppell, O., Schäffler, L., & Hölldobler, B. (2002). Lack of plasticity in the behavior of queens of the ant *Leptothorax rugatulus* Emery (Formicidae: Hymenoptera). *Journal of insect Behavior*, 15(3), 447-454.

- 50. Scharlaken, B., de Graaf, D. C., Goossens, K., Brunain, M., Peelman, L. J., & Jacobs, F. J. (2008). Reference gene selection for insect expression studies using quantitative real-time PCR: The head of the honeybee, *Apis mellifera*, after a bacterial challenge. *Journal of Insect Science*, 8(1), 33.
 - 51. Saïd, I., Gaertner, C., Renou, M., & Rivault, C. (2005). Perception of cuticular hydrocarbons by the olfactory organs in *Periplaneta americana* (L.)(Insecta: Dictyoptera). *Journal of Insect Physiology*, 51(12), 1384-1389.
- 52. Scheiner, R., Page, R. E., & Erber, J. (2004). Sucrose responsiveness and behavioral plasticity in honey bees (*Apis mellifera*). *Apidologie*, 35(2), 133-142.
- 53. Seid, M. A., & Traniello, J. F. (2006). Age-related repertoire expansion and division of labor in *Pheidole dentata* (Hymenoptera: Formicidae): a new perspective on temporal polyethism and behavioral plasticity in ants. *Behavioral Ecology and Sociobiology*, 60(5), 631-644.
- 54. Shemshedini, L., & Wilson, T. G. (1990). Resistance to juvenile hormone and an insect growth regulator in *Drosophila* is associated with an altered cytosolic juvenile hormone-binding protein. *Proceedings of the National Academy of Sciences*, 87(6), 2072-2076.
- 55. Sullivan, J. P., Jassim, O., Fahrbach, S. E., & Robinson, G. E. (2000). Juvenile hormone paces behavioral development in the adult worker honey bee. *Hormones and Behavior*, 37(1), 1-14.
- 56. Sumner, S., Pereboom, J. J., & Jordan, W. C. (2006). Differential gene expression and phenotypic plasticity in behavioural castes of the primitively eusocial wasp, *Polistes canadensis*. *Proceedings of the Royal Society B: Biological Sciences*, 273(1582), 19-26.
- 57. Tan, S. H. (2007). Stable expression and purification of Juvenile Hormone Binding Protein from *Drosophila melanogaster* Schneider line-2 cells (Doctoral dissertation).
- 58. Toth, A. L., & Robinson, G. E. (2007). Evo-devo and the evolution of social behavior. *Trends in Genetics*, 23(7), 334-341.
- 59. Tschinkel W R (2006) The Fire Ants. Harvard University Press Cambridge Mass. 670pp
- 60. Vander Meer, R. K., Saliwanchik, D., & Lavine, B. (1989). Temporal changes in colony cuticular hydrocarbon patterns of *Solenopsis invicta*. *Journal of Chemical Ecology*, 15(7), 2115-2125.
 - 61. Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., & Speleman, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome biology*, 3(7), research0034-1.
- 478 62. Vargo, E. L., & Laurel, M. (1994). Studies on the mode of action of a queen primer 479 pheromone of the fire ant *Solenopsis invicta*. *Journal of Insect Physiology*, 40(7), 601-480 610.
- 481 63. Wetterer, J. K., & Porter, S. D. (2003). The little fire ant, *Wasmannia auropunctata:* distribution, impact and control.
- 483 64. Wilson, E.O (1971) The Insect Societies. Harvard Univ. Press Cambridge Mass. 548pp

- 484 65. Wong, M. L., & Medrano, J. F. (2005). Real-time PCR for mRNA quantitation.
 485 *Biotechniques*, 39(1), 75-85.
 - 66. Ye, J., Coulouris, G., Zaretskaya, I., Cutcutache, I., Rozen, S., & Madden, T. L. (2012). Primer-BLAST: a tool to design target-specific primers for polymerase chain reaction. *BMC Bioinformatics*, 13(1), 134.

Table 1. Observation table. Number of total observations during behavioral assays and extrapolated to 12 hrs periods. Behaviors observed and tabulated were the number of events counted during the observation period.

Table 1: Behavior flexibility assay observations			
Behaviors Observed	# Extrapolated Observations		
Egg Laying	2,204		
Nursing	1,064		
Foraging	54		
*Observations 10 min twice a day and extrapolated to 12hr			
periods for 19 days			

Table 2. Primer table. List of target genes including housekeeping genes primers, their accession number in NCBI and amplification length. Actin and GAPDH primer sequences were taken from Scharlaken et al. 2008.

Gene	Acc. Number	Strand	Primer sequence	Amplification (bp)
JHbp	XM_011708554	FW	TGTTGGTGCCCATCGCTAAT	140
		RV	GTCCAGCTTTATCGTCAACTTCG	
Vg	XM_011697672.1,	FW	GCCACAACTGATCACAGCCA	218
	XM_011697673.1	RV	GGACCGTCCGGTAATGTAGT	
		RV	GAGTTTGCTGCTGTGTTC	
Actin	AB023025	FW	TGCCAACACTGTCCTTTCTG	155
		RV	AGAATTGACCCACCAATCCA	
GAPDH	XM_393605	FW	GATGCACCCATGTTTGTTTG	203
		RV	TTTGCAGAAGGTGCATCAAC	

Figure 1. Relative probability of tasks in egg laying behavior (**A**), nursing behavior (**B**) and foraging behavior (**C**). (**A**). Egg Laying Behavior. Queen egg laying behaviors before and after workers were removed, task decreased significantly, W = 62.5, p-value = 0.01, Mean_{typical}=0.75, SE_{typical}=0.07, Mean_{WR}=0.36, SE_{WR}=0.08. (**B**). Nursing Behavior. Queen nursing behaviors before and after workers were removed, task increased significantly, W = 2.5, p-value = 0.002, Mean_{typical}=0.00, SE_{typical}=0.00, Mean_{WR}=0.55, SE_{WR}=0.05. (**C**). Foraging Behavior. Queen foraging behaviors before and after workers were removed, task increased significantly, W = 15, p-value = 0.04, Mean_{typical}=0.00, SE_{typical}=0.00, Mean_{WR}=0.07, SE_{WR}=0.02. n=19 queens. Behaviors observed tabulated as number of events occurred during the observation period.

Figure 2. Daily behavior rate. Task frequency of Queens (n=19) during the experiment. The shaded area represents tasks during the control period (five days). Workers were removed on day five after final control observations (C5). Egg laying behavior decreased significantly at day

510 four; Chi-square = 118.524, df = 18, p-value = <0.001, just as nursing behavior increased significantly; Chi-square = 14.58, df = 18, p-value = <0.001, foraging increased significantly, at 511 512 day seven; Chi-square = 37.71, df = 18, p-value = <0.001. On day twelve the population began to grow as workers emerged. At day thirteen queens increased egg laying and decreased nursing, 513 completing a cycle. First three days show no significant difference with the last three days; Dunn 514 p-value = >0.05. Behaviors were observed tabulated as number of events occurred during the 515 observation period.

516

517 Figure 3. Relative gene expression of JHbp (A&B) and Vg (C&D) in control and experimental nests. Results show differences in gene expression of JHbp and Vg between control 518 or egg laying queens and experimental or nursing and foraging queens. Expression was measured 519 in relative expression to endogenous genes Actin and GAPDH by geometric mean analysis 520 method. n=33 queens; control queens n=15, experimental queens n=18. (A). JHbp expression 521 decreases from control to experimental queens, W = 270, p-value = <0.001, Mean_{control}=1.30, 522 SE_{exp}=0.26. (**B**). Comparing JHbp expression by tasks shows difference between egg laying 523 compared to nursing and foraging, there's no difference in expression between nursing or 524 foraging queens Kruskal-Wallis chi-squared = 24.21, df = 2, p-value = <0.001, Mean_{egg} 525 laying=1.30, SE_{egg} laying=0.26, Mean_{nursing}=0.20, SE_{nursing}=0.002, Mean_{foraging}=0.10, SE_{egg} 526 laying=0.003. (C&D). Similar pattern as JHbp is seen with Vg expression, decreases in 527 experimental queens (W = 270, p-value = <0.001; Kruskal-Wallis chi-squared = 25.19, df = 2, p-528 value = <0.001, Meancontrol=1.22, SE_{exp}=0.22, Meanegg laying=1.22, SE_{egg laying}=0.22, 529 530 Mean_{nursing}=0.10, SE_{nursing}=0.02, Mean_{foraging}=0.05, SE_{egg laving}=0.01; **C&D** respectively.





