**Methods**

Field Collection

*Sarracenia purpurea* leaf water samples were collected from 97 plants across three sites in the Apalachicola National Forest in northern Florida in July of 2024. These sites were chosen for the presence of known *S. purpurea* populations. Fifty samples were collected from Site 1 (Crystal Bog), 38 samples were collected from Site 2 (Pleaphase Savanna), and eight samples were collected from Site 3 (Twisted Cyprus Bog). At each site, leaves were chosen haphazardly for sampling with a minimum of ~1 m distance between each leaf. No two leaves were sampled from the same plant. Samples were collected using clean plastic transfer pipettes with ~5 cm of the tip cut away to avoid getting clogged with leaf water detritus. Before collection, the leaf water was mixed to suspend detritus by repeatedly pipetting up and down. Leaf contents were transferred to 50 ml conical tubes. The entire volume of leaf water was collected when possible; if the leaf volume exceeded 50 ml, only 50 ml was collected. Samples were then transported in Styrofoam trays to Forida State University.

Within 24 h of collection, samples were visually inspected for the presence of late-instar mosquito larvae (*Wyeomyia smithii*), which if present were removed using a plastic transfer pipette. The excess leaf water discarded during the process of mosquito larvae removal was filtered using a xx µm mesh sieve and frozen for later use as a seed community of bacteria to help mimic the conditions of natural pitcher plant leaf water.

Sample tube lids were sealed with parafilm and transported via commercial flight in a checked bag to California State University, Northridge. Sample tubes were then stored in a growth chamber (growth chamber company) with a 12/12 h light/dark cycle, at 25C during the day and 18C at night. Using custom Palmer cell-style counting slides, 100 µl samples from each tube were scored for the presence of *Habrotrocha rosa*, other species of rotifers, and ciliates. Samples were monitored daily for the presence of developing mosquito larvae, which were removed promptly upon detection.

Rotifer Isolation and Rearing

In order to test for diversity in response to temperature, clonal cultures of *H. rosa* were first established by isolating individuals from the field-collected leaf water samples. To be selected for use in isolations, leaf water samples had to meet two criteria: the absence of an overwhelming abundance of ciliates, and an estimated rotifer density of >2 individuals per 100 µl. Meeting these criteria were seven samples from Site 1 (Crystal Bog), six samples from Site 2 (Pleaphase Savanna), and one sample from Site 3 (Twisted Cyprus Bog).

To isolate rotifers, 0.5 ml of leaf water was transferred into a 12-well tissue culture plate. Individual rotifers were transferred with the smallest volume of leaf water possible using a clean, pulled Pasteur pipette into another well filled with 0.5 ml of sterile spring water. This washing step was repeated a second time into another well of sterile spring water, and a third time into an unoccupied well in a 24-well tissue culture plate that contained 0.3 ml of rotifer media (Plasota and Plasota, 1980) and xx mg of fish food flakes to serve as a basal resource for bacteria (TetraColor Tropical Fish Flakes, Tetra).

Preliminary experiments using this isolation method indicate that the likelihood of a haphazardly-chosen individual of *H. rosa* proliferating into a culture of clones is only xx%. This is consistent with expectations, however, because the reproductive period of *H. rosa* lasts only 5-7 d. With a lifespan of 12-24 d, some individuals have been observed to live for nearly two weeks after losing the ability to reproduce (Ellison and Bledzki, xxxx). Therefore, in order to reach the goal of five clonal cultures per leaf, 32 individuals were isolated from each selected leaf water sample, for a total of 448 isolated individuals of *H. rosa* across 20 24-well tissue culture plates.

Temperature Response Experiment