



# Health Economic Evaluation Using Markov Models in R for Microsoft Excel Users: A Tutorial

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## Abstract

A health economic evaluation (HEE) is a comparative analysis of alternative courses of action in terms of both costs and consequences. A cost-effectiveness analysis is a type of HEE that compares an intervention to one or more alternatives by estimating how much it costs to gain an additional unit of health outcome. Cost-effectiveness analyses are commonly performed using Microsoft (MS) Excel. However, there is current interest in using other software that is better suited to more complex problems, methods, and data, as well as improved reproducibility and transparency. That is, it is increasingly important to be able to repeat an analysis of a particular data set and obtain the same results, and access the analysis and results in a clear and comprehensive openly available form. In this tutorial we provide a step-by-step guide on how to implement a mainstay model of HEE, namely a Markov model, in the statistical programming language R. The adoption of R for the purpose of cost-effectiveness analysis is highly dependent on the ability of the health economic modeller to understand, learn, and apply programming-type skills. R is likely to be less familiar than MS Excel for many modellers and so coding a cost-effectiveness model in R can be a large jump. We describe the technical details from the perspective of a MS Excel user to help bridge the gap between software and reduce the learning curve by providing for the first-time side-by-side comparisons of the Markov model example in MS Excel and R.

## Key Points

In health economics modelling, larger and more complex problems, methods, and data have made MS Excel less fit for purpose. Alternative software that is better suited, including programming languages such as R, has become more attractive.

However, the lack of familiarity with R might be an impediment to a move away from MS Excel.

Using our technical details, guidance and templates will help bridge the gap when transitioning between software and will reduce the potentially steep learning curve.

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## 1 Introduction

A health economic evaluation (HEE) is a comparative analysis of alternative courses of action in terms of both their costs and consequences [1]. A cost-effectiveness analysis is a

type of HEE that examines and compares both the costs and health outcomes of one intervention to one or more alternatives by estimating the costs of gaining an additional unit of a health outcome [2].

Cost-effectiveness analyses are often performed using Microsoft (MS) Excel [3, 4], but there are other bespoke HEE or general software available to perform this task, such as TreeAge (TreeAge Software, Inc, Williamstown, MA), WinBUGS [5] or statistical programming languages, such as R [6]. R is increasingly being used for cost-effectiveness modelling [7, 8] along with its popular Integrated Development Environment (IDE) RStudio [9]. As a result, specific HEE tools are being written in R [10]. An overview of R functionality that is applicable to HEE is given by Jalal (2017) [11] and several tutorials already exist to help implement common HEE models in R [12, 13].

However, the adoption of R for the purpose of cost-effectiveness analysis is highly dependent on the health economic modeller's ability to understand, learn, and apply programming-type skills [8]. For individuals who are normally used to working with MS Excel, coding a cost-effectiveness model in R can be a large jump [14].

This tutorial is designed for those who are familiar with HEEs built in MS Excel but are unfamiliar with using R. The aim of this tutorial is to expose information previously presented at workshops provided by the authors on this topic, while highlighting the benefits of this type of modelling using R. This tutorial will use a Markov model example from MS Excel to demonstrate how to implement it in R.

The reader will be guided through the necessary steps for coding in R, comparing with the equivalent MS Excel model. Where possible, base R is used throughout, as previously, dependency on packages (such as *heemod* [15]) has been shown to lead to slightly different results when compared directly to equivalent models built in MS Excel [8]. Additionally, writing out necessary functions in base R allows for transparent step-by-step comparisons with MS Excel models and for greater flexibility in adapting the code for future use.

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#### Tips for those new to R coding

*Setting up and the basics* R is a free, open-access software, downloadable from <https://cran.r-project.org>. There are many introductory tutorials for R available, for example see some of the introductory basic R manuals available under Documentation Manuals. (<https://cran.r-project.org/manuals.html>)

*Health economics* There are many different packages in R - while this tutorial uses base R, some useful packages for HEE can be found here: [https://github.com/n8thangreen/health\\_economics\\_R\\_packages](https://github.com/n8thangreen/health_economics_R_packages); <https://hermes-sheprd.netlify.app/>

The standard R assignment operator `<-` will be used throughout but `=` would be equivalent.

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R has a comprehensive library of functions for generating random numbers from various statistical distributions. A full list of distributions can be seen by typing `?distribution` in the console. When users generate a "random number" in R, they are actually generating pseudorandom numbers. These numbers are generated in a sequence with an algorithm that requires a seed to be set to initialize. The `set.seed()` function can be used when running simulations to ensure all results, figures, etc. are reproducible.

We will make use of existing and user-defined functions in R. MS Excel has many existing functions which can be called from within a cell. The syntax for creating these is similar in R and Visual Basic for Applications (VBA) in MS Excel. For an introduction in R see Golemund. [16]

*Version control and collaborating* Code sharing and saving work online is recognised good practice for R users and helps maintain version control. GitHub is a popular provider of internet hosting for software development and version control.

When repositories on GitHub are an open source, they will commonly have a license with regard to how others can use, change, and distribute the software. [17]

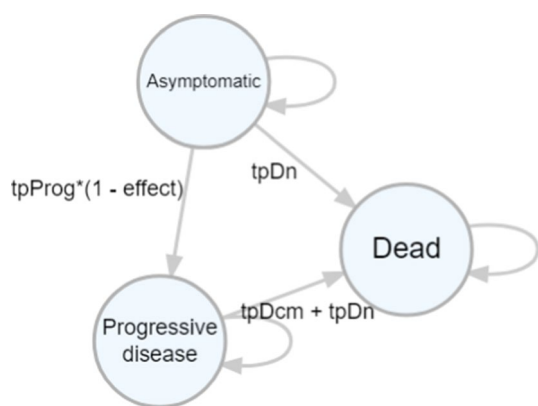
---

## 2 A Time-dependent Markov Model

A Markov model is arguably the most commonly used approach in HEEs. A Markov model is useful for repeated events over time, such as modelling disease progression. It can be thought of as a special form of a more general state transition model. Each state is mutually exclusive, discrete and may be absorbing or transient [18]. Of particular focus for the following example, transition probabilities in a Markov model can either be fixed (time homogeneous) or depend on time (time inhomogeneous). Furthermore, we will consider a "clock forward" model, which means that time refers to time since entering the initial state. Conversely, in a "clock reset" (a semi-Markov) model, time refers to time since entering the current state, meaning that time resets to 0 each time a patient enters a new state. General details of Markov chains can be found elsewhere [19].

### 2.1 Real-World Example

This tutorial focuses on providing a practical walk through of the MS Excel cost-effectiveness model from Briggs et al. [20, 21] while comparing with an R model equivalent. Code for the R model used in this tutorial can be found on GitHub at the following link [https://github.com/Excel-R-tutorials/Markov\\_Intro](https://github.com/Excel-R-tutorials/Markov_Intro). The corresponding original MS Excel model is available on open-access and can be downloaded from the following link <https://doi.org/10.17037/DATA.00002980> [20]. The model outlined in the MS Excel spreadsheet has been used to illustrate the principles of cost-effectiveness Markov modelling and probabilistic sensitivity analysis (PSA) [21, 22]. It has been made available for non-commercial teaching purposes only.



**Fig. 1** Markov model diagram created using R. Nodes represent health states and directed edges represent transitions between states. Transition probabilities are shown on the edges. tpProg probability of disease progression, tpDn probability of all-cause death, tpDcm excess probability of death, effect effectiveness of drug treatment

To more easily demonstrate the translation from the MS Excel model to R we have made some small changes to how the formulae in the original spreadsheets were written. These changes do not affect the results of the model. The GitHub repository contains both versions and the original version is in the ‘original files’ folder. From now on reference to the MS Excel model will refer to the latest version. In particular, in the updated version we adopt the @ notation, which substitutes the value into a cell from another cell on the same line from a named column. The Markov model workbook was used with MS Excel 365. Note that earlier versions of MS Excel did not explicitly use the @ notation, and so this can be omitted, or prior to this MS Excel may not even have this array functionality. The MS Excel model will be discussed further in Sect. 2.2.

In this tutorial a three-state model is used to describe how patients transition between health states with a chronic disease. Patients begin in an asymptomatic health state, which means that the patient has developed the chronic disease but has no symptoms. Patients can stay in the asymptomatic health state or move to a progressive health state, which means that the patient is experiencing symptoms of the disease. Patients can move from the asymptomatic health state to the dead state at the same rate as the general population without the disease. Patients can stay in the progressive health state or move from the progressive health state to dead, but at an increased risk of death. The dead state is an absorbing state as a patient cannot change from being dead. Briggs et al. [21] describe the model in more detail. Figure 1 shows an example of the model diagram produced using R. Arrows represent possible transitions between health states (represented by circles) after each model cycle. Backward bending arrows returning to the state they left show that it is possible to remain in the same state as the previous cycle. Let

us define the variables used in the model. In terms of transition probabilities, tpProg is the probability of disease progression, tpDn is the probability of all-cause death, tpDcm is the excess probability of death, and effect is the proportion reduction of the transition probability from asymptomatic to progressive disease due to the drug treatment. The cost of one cycle in the Asymptomatic and Progressive disease state are cAsymp and cProg, respectively. The unit cost of the drug is cDrug and the one-off cost of a death is cDeath. In terms of Quality-of-Life weight, in the Asymptomatic and Progressive disease states these are uAsymp and uProg, respectively. The values will be described in the next sections as well as a demonstration of how to fully implement this model in R. For reference, a table of all input values is given in the Supplementary Material.

### Markov model diagrams in R

There are many packages available in R for plotting edge-node type graphs, including igraph [29], diagram [30] and DiagrammeR [31]. More specific packages such as markovchain [32] and heemod [15] are used for Markov chain modelling. They build on these base packages and do the work of translating the model object into the graph.

In MS Excel, the model figure is usually created by hand and copied and pasted into a sheet as a picture. An alternative in R is to use packages to create a plot directly from the model code. This has the added advantage of literate programming, where the documentation and code are in the same place and the plot is always up-to-date with any changes to the model. Example R code to make the model diagram using DiagrammeR is available in Online Resource 1 and this tutorial paper GitHub repository.

## 2.2 MS Excel Implementation

The MS Excel cost-effectiveness model from Briggs et al. [20, 21] consists of several separate tabs each containing input parameters, computation, or output plots and tables. In particular, the Parameters and Analysis tab contains the cost, utility, death rate and other input point values and distribution parameters. The Markov model tab calculates for both treatment scenarios all of the state population counts, summary statistics, and total costs and quality-adjusted life years (QALYs) in adjacent columns. Rows correspond to a single cycle and each simulation total is calculated at the bottom of the sheet. The Scenario Summary and Cohort tabs give the output tables and plots, respectively.

We can make some direct comparisons between MS Excel and R. In MS Excel, values are entered in a cell, which can be named so that it can be referred to elsewhere in the workbook by its name as well as its cell location. In R, the equivalent is to define a variable and assign it a value. The name of the variable can subsequently be used in the code in place of explicitly embedding the value directly (called “hard-coding”). Similarly, groups of values can be referred

State populations      Outcome statistics

Time ↓

Cycle	rDeath	Asymp	Prog	Death	LE	QALE	LYs	QALYs	StateCost	TransCost	TotalCost
0	0	1000.0	0.0	0.0							
1	0.0138	976.2	10.0	13.8	986.2	934.9	930.4	882.0	488774	0	488774
2	0.0138	943.2	27.9	28.9	971.1	917.0	864.3	816.1	494180	1415	495595
3	0.0138	901.9	51.6	46.5	953.5	895.5	800.6	751.9	508632	3723	512355
4	0.0138	853.4	79.2	67.4	932.6	870.1	738.7	689.2	526261	6500	532761
5	0.0138	798.9	108.9	92.1	907.9	840.7	678.4	628.2	542688	9414	552102
6	0.0138	740.0	139.0	121.0	879.0	807.2	619.6	569.1	554831	12209	567041
7	0.0138	678.0	168.0	154.0	846.0	770.1	562.6	512.2	560718	14700	575419
8	0.0138	614.4	194.8	190.9	809.1	729.7	507.7	457.8	559306	16764	576070
9	0.0138	550.6	218.1	231.3	768.7	686.7	455.0	406.4	550311	18329	568640
10	0.0138	487.9	237.5	274.6	725.4	641.6	405.1	358.3	534043	19368	553411
11	0.0379	415.8	246.5	337.7	662.3	579.9	348.9	305.5	499111	19891	519002
12	0.0379	350.1	250.1	399.8	600.2	520.2	298.3	258.5	459869	19480	479349
13	0.0379	291.3	248.6	460.0	540.0	463.2	253.2	217.2	417980	18643	436624
14	0.0379	239.5	242.7	517.8	482.2	409.5	213.3	181.1	374993	17484	392477
15	0.0379	194.5	233.0	572.5	427.5	359.5	178.4	150.0	332266	16101	348367
16	0.0379	156.0	220.4	623.6	376.4	313.5	148.2	123.4	290928	14584	305512

(a)

#	A	B	C	D	E	F	G	H	I
1	Cycle	age	rDeath	Asymp	Asymp_lag	Prog	Prog_lag	Death	Death_lag
2	0	55	0	=n_cohort		0		0	
3	1	56	0.0138	=n_cohort-@ Prog-@ Death	=D2	=@Prog_lag*(1-tpDcm-@rDeath)+@Asymp_lag*tpProg@ncycle	=F2	=@Death_lag+@Prog_lag*tpDcm+(@Asymp_lag+@ Prog_lag)*@rDeath	=H2
4	2	57	0.0138	=n_cohort-@ Prog-@ Death	=D3	=@Prog_lag*(1-tpDcm-@rDeath)+@Asymp_lag*tpProg@ncycle	=F3	=@Death_lag+@Prog_lag*tpDcm+(@Asymp_lag+@ Prog_lag)*@rDeath	=H3
5	3	58	0.0138	=n_cohort-@ Prog-@ Death	=D4	=@Prog_lag*(1-tpDcm-@rDeath)+@Asymp_lag*tpProg@ncycle	=F4	=@Death_lag+@Prog_lag*tpDcm+(@Asymp_lag+@ Prog_lag)*@rDeath	=H4
6	4	59	0.0138	=n_cohort-@ Prog-@ Death	=D5	=@Prog_lag*(1-tpDcm-@rDeath)+@Asymp_lag*tpProg@ncycle	=F5	=@Death_lag+@Prog_lag*tpDcm+(@Asymp_lag+@ Prog_lag)*@rDeath	=H5
7	5	60	0.0138	=n_cohort-@ Prog-@ Death	=D6	=@Prog_lag*(1-tpDcm-@rDeath)+@Asymp_lag*tpProg@ncycle	=F6	=@Death_lag+@Prog_lag*tpDcm+(@Asymp_lag+@ Prog_lag)*@rDeath	=H6

(b)

**Fig. 2 a** MS Excel model cycles and population flow, costs and QALYs for cohort without drug treatment, showing values. *LE* life expectancy, *LY* life-year *QALE* quality-adjusted life expectancy, *QALY* quality-adjusted life-year, *rDeath* probability of death each year, *StateCost* total state occupancy cost each year, *TransCost* total

state transition cost each year, *TotalCost* sum of *StateCost* and *TransCost*. **b** MS Excel model cycles and population flow, showing formulae. *rDeath* probability of death each year, *tpDcm* excess probability of death, *tpProg* probability of disease progression

to by name in MS Excel by selecting a range of cells and naming these. These can then be handled as a single object e.g., as an input to a function. In R there are several ways of defining a group of values like this but for the purposes of this tutorial we shall use the base R matrix and array data structures, which are two-dimensional and n-dimensional rectangular objects, respectively. See [26] for more details.

### 2.3 Translating an MS Excel Model Using Loops

In this section we provide a demonstration on how to translate a MS Excel Markov model to an equivalent R model by defining the use of for-loops, which will be implemented in this tutorial. A for-loop is a type of control flow statement, which repeatedly executes a section of code for a defined number of iterations. The focus will be on the calculation of the state populations over time. Part of the MS Excel Markov model (Markov Model tab) is shown in Fig. 2a. Each row represents a cycle (cycle length is one year) so that discrete time steps go from top to bottom and the columns contain the state populations, time-dependent probabilities and (cumulative) functions of these, such as total QALYs and total costs. The counts are shown to one decimal place to

emphasize that fractions of the starting state population are simulated. In Fig. 2a at the start of the simulation there are 1000 patients in the asymptomatic health state (Asymp) and 0 in both the progressive health state (Prog) and Dead state. Probabilities, defined within the “Parameters and Analysis” sheet, are used to calculate the movements between health states. The all-cause mortality was dependent on age and we assumed that all individuals in the cohort had the same age at the start of the simulation. Values were taken from standard life-tables and aggregated into 10-year intervals. For example, by cycle 15, in row 20, the 1000 patients are distributed amongst the states—195 patients in the Asymp state, 233 in the Prog state and 572 are Dead.

Figure 2b outlines the corresponding formulae behind the same model. The Asymp and Prog health state formulas are shown for simplicity, but the same pattern applies to the rest of the sheet. Recall that the @ notation is used in this model, which substitutes the value on the same line from the named column.

Notice that besides the initial row, the formulae in the Asymp and Prog columns are the same for all rows. When this pattern occurs in code it can be simplified and rewritten as a loop. Figure 3 is an example of some pseudo-code



**Define constants:**

```
n_cohort = 1000
n_cycle = 45
tpProg = 0.01
tpDcm = 0.15
```

**Define starting populations at  $t=0$  i.e. first row:**

```
@Asymp = n_cohort
@Prog = 0
@Death = 0
```

**Iterate over cycles i.e. row by row:**

```
loop from t = 1 to n_cycle
  @Prog = @Prog_lag*(1 - tpDcm - @rDeath) + @Asymp_lag*tpProg*n_cycle
  @Death = @Death_lag + @Prog_lag*tpDcm + (@Asymp_lag + @Prog_lag)*@rDeath
  @Asymp = n_cohort - @Prog - @Death
End
```

**Fig. 3** Pseudo-code showing how the MS Excel formulae can be represented programmatically as a loop. *rDeath* probability of death each year, *tpDcm* excess probability of death, *tpProg* probability of disease progression

```
loop from t = 1 to n_cycle
  (@Asymp, @Prog, @Death) = (@Asymp, @Prog, @Death) %*%
  [ C   tpProg*n_cycle   @rDeath
    0   C                 tpDcm + @rDeath
    0   0                 1 ]
End
```

**Fig. 4** Pseudo-code showing an approach with matrix multiplication within a loop for the MS Excel population movement. *C* 1 minus the sum of all other probabilities from a state, *rDeath* probability of death

each year, *tpDcm* excess probability of death, *tpProg* probability of disease progression

(non-R code), which takes the MS Excel formulae and uses it in a for loop.

The pseudo-code in Fig. 3 outlines how the cohort begins with 1000 patients (*n\_cohort*), for 45 cycles (*n\_cycle*), and two probabilities (*tpProg* and *tpDcm*) influencing the movement between health states. Once the starting populations are defined (1000 patients in the Asymp health state and 0 in both the Prog and Dead states), it is then possible to loop over each cycle using the updated information from the cycle before. To loop from cycle 1 to 45, the formulae for each health state is used. Further, it is possible to rewrite the pseudo-code of the loop using a transition matrix and using matrix multiplication (Fig. 4) equivalently and succinctly. This mathematical formulation is common in Markov models.

The command `%*%` is called an infix operator and is used for matrix multiplication in R. The 1 by 3 vector (`@Asymp`, `@Prog`, `@Death`) is the number of patients in the three health states. The 3 by 3 matrix represents the possible transitions between health states. For example, the top right entry is the value *tpDn*, which describes the movement between Asymp to Death. The values *C* in the matrix

represents 1 minus the probabilities in the remainder of the corresponding row. This idea of using loops is the basis for all the following R code throughout the rest of this tutorial.

## 2.4 Setting up the R Model

The MS Excel model simulates a cohort of 1000 patients that receives a drug and compares it to a cohort of 1000 patients that does not receive a drug to assess how patients move throughout the health states over time with a chronic disease. To set up the MS Excel model in R, some definitions are needed. Figure 5 outlines the R code used to define the number of treatments and states (prefixed with *n\_*) and their names (suffixed with *\_names*). The number of cycles in the R model starts at 1 (not 0 as in spreadsheet models) and so the number of cycles is 46 rather than 45 as the cycle length is 1 year. The initial age of all patients in the cohort begins at 55. The same variable names have been used as the MS Excel model where feasible to help with comparisons. The probabilities of death each year by age taken from standard life-tables where 0.0138 for patients aged 55–65 years,

**Fig. 5** Defining the treatments, names, states, cycles and starting age in R

```
t_names <- c("without_drug", "with_drug")
n_treatments <- length(t_names)

s_names <- c("Asymptomatic_disease", "Progressive_disease", "Dead")
n_states <- length(s_names)

n_cohort <- 1000
n_cycles <- 46
Initial_age <- 55
```

**Fig. 6** Defining the costs, utilities, discount rates and transition probabilities in R

```
cAsymp <- 500; cDeath <- 1000; cProg <- 3000
cDrug <- 1000
uAsymp <- 0.95; uProg <- 0.75
oDr <- 0.06; cDr <- 0.06
tpDcm <- 0.15; tpProg <- 0.01
```

**Fig. 7 a** Creating the matrix object in R for the transition cost matrix. **b** The matrix defining the cost of transitioning into a state created in R

```
trans_c_matrix <-
  matrix(c(0, 0, 0,
           0, 0, cDeath,
           0, 0, 0),
         byrow = TRUE,
         nrow = n_states,
         dimnames = list(from = s_names,
                          to = s_names))
```

(a)

```
> trans_c_matrix
```

from	Asymptomatic_disease	Progressive_disease	Dead
Asymptomatic_disease	0	0	0
Progressive_disease	0	0	1000
Dead	0	0	0

(b)

0.0379 for 66–75 years, 0.0912 for 76–85 years and 0.1958 for patients aged > 85 years.

The unit costs and unit utilities associated with each of the health states as well as the cost of the drug need to be defined at the start and shown in Fig. 6. The utility of being in the Dead state is 0 so does not need to be defined here. Costs begin with a ‘c’ and utilities with a ‘u’. The discount rate for costs and outcomes are also included at a rate of 6%. Other discount rates may be preferred in other analyses but we remain consistent with Briggs (1998). The excess probability of dying from the disease in a single cycle (tpDcm) is now defined. The all-cause mortality tpDm is not defined here since it is a time-dependent variable and will be defined later.

This process of defining treatment names, states and cycles is similar to that in MS Excel. Using R has more flexibility for the structure of the objects we use for the

computation and for saving the results, and not just a flat, 2D matrix. We shall define the same structure for the matrices for the cost of transition to a health state, and cost and QALYs accrued being in health state. Figure 7a shows the matrix for the cost of transitioning to a state, trans\_c\_matrix. All costs are zero except for the cost of £1000 for transitioning to the Dead state from the Progressive disease state. These costs are the same for both cohorts.

The first argument defines a vector of values for each health state similar to a column of values in the parameters sheet in MS Excel. The argument byrow = TRUE makes sure that the costs from the vector fill up the rows of the matrix from left to right before moving to the next row. Therefore, the resulting matrix will look the same as to how we have arranged the vector, shown in Fig. 7b.

In a similar way, Fig. 8a describes the cost of being in a health state. The structure of state\_c\_matrix is treatment by

**Fig. 8** **a** Creating a matrix for the costs of being in each health state in R. **b** The matrix of costs of being in each health state for each treatment

```
state_c_matrix <-
  matrix(c(cAsymp, cProg, 0,
           cAsymp + cDrug, cProg, 0),
        byrow = TRUE,
        nrow = n_treatments,
        dimnames = list(t_names,
                        s_names))
```

**(a)**

```
> state_c_matrix
```

	Asymptomatic_disease	Progressive_disease	Dead
without_drug	500	3000	0
with_drug	1500	3000	0

**(b)**

**Fig. 9** **a** Creating a 3-dimensional matrix to contain transition probabilities in R. **b** The probability transition matrix created in R. The output shows two “slices” through the array, one without drug treatment and one with drug treatment

```
p_matrix <- array(data = c(1 - tpProg - tpDn, 0, 0,
                           tpProg, 1 - tpDcm - tpDn, 0,
                           tpDn, tpDcm + tpDn, 1,
                           1 - tpProg*(1-effect) - tpDn, 0, 0,
                           tpProg*(1-effect), 1 - tpDcm - tpDn, 0,
                           tpDn, tpDcm + tpDn, 1),
                  dim = c(n_states, n_states, n_treatments),
                  dimnames = list(from = s_names,
                                  to = s_names,
                                  t_names))
```

**(a)**

```
> p_matrix
, , = without_drug
```

from	to	Asymptomatic_disease	Progressive_disease	Dead
Asymptomatic_disease		0.9521	0.0100	0.0379
Progressive_disease		0.0000	0.8121	0.1879
Dead		0.0000	0.0000	1.0000

```
, , = with_drug
```

from	to	Asymptomatic_disease	Progressive_disease	Dead
Asymptomatic_disease		0.9571	0.0050	0.0379
Progressive_disease		0.0000	0.8121	0.1879
Dead		0.0000	0.0000	1.0000

**(b)**

state and is assigned non-zero costs to being in the Asymp and Prog health states. Again, the argument `byrow = TRUE` ensures that the without-drug cohort costs are along the first row and the with-drug cost along the second row (Fig. 8b).

An input matrix for the QALYs accrued from being in a health state is created the same way (Online Resource 2). The probability of dying from the disease in a single cycle (`tpDcm`) was defined above. Space is also needed to define the transition probabilities between health states. A

3-dimensional matrix, called an “array” in R, is created similar to `state_c_matrix` but with another dimension for the movements between health states (Fig. 9a). We will show how to implement a time-dependent transition probability array in the following section but we show the simpler, time-independent example here for completeness. The printed output is shown in Fig. 9b—it shows probabilities for the transitions between each health state dependent on the cohort being with or without a drug.

```

cycle_empty_array ←
  array(NA,
        dim = c(n_treatments, n_cycles),
        dimnames = list(treatment = t_names,
                          cycle = NULL))

```

(a)

```

> cycle_empty_array
      cycle
treatment [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10] [,11] [,12] [,13] [,14] [,15] [,16] [,17]
without_drug NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA
with_drug    NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA  NA

      cycle
treatment [,18] [,19] [,20] [,21] [,22] [,23] [,24] [,25] [,26] [,27] [,28] [,29] [,30] [,31] [,32] [,33]
without_drug NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA
with_drug    NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA

      cycle
treatment [,34] [,35] [,36] [,37] [,38] [,39] [,40] [,41] [,42] [,43] [,44] [,45] [,46]
without_drug NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA
with_drug    NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA   NA

```

(b)

**Fig. 10** **a** Creating empty space for each treatment and cycle in R. **b** The empty array for 46 cycles and both treatments

## 2.5 Time-dependent Components

One addition in R relative to MS Excel is that we should prepare for an analysis by creating objects with empty space for when calculations are performed. It is like creating the cells in MS Excel. Of course, in MS Excel this step is unnecessary since the spreadsheet cells are in some sense already initiated and available to assign values to. By creating matrices and arrays in R, empty space for population counts, costs and utilities in each health state for patients with or without the drug is specified. Appending values to other objects at run time in R is possible but ill-advised because it is slower and more error prone. The labelling of cells and groups of cells in MS Excel could be thought of as a similar process as creating a matrix in R. A population matrix (pop) and transition matrix (trans) are created in the same way so there is blank space for each health state, with or without a drug, for each of the 46 cycles. Figure 10a, b below show this R code for a generic array (cycle\_empty\_array); see Online Resource 2 for how this applies to transition arrays.

Transition probabilities in a Markov model can either be time-independent, such as tpDcm, or time-dependent (like @rDeath in the MS Excel model). In this model, the transition probabilities from Asymp to Dead are time dependent, as they depend on the age of the cohort. Therefore, the transition probabilities depend on the cycle which is why we need to have an extra cycle dimension in the empty array for probabilities, costs and QALYs.

To account for the time dependency, the representation by a hard-coded array in MS Excel was replaced with a function

in R named p\_matrix\_cycle, which is called at each cycle iteration, similar to the logic in Sect. 2.2. By simply replacing a fixed valued array with a function, this decouples the calculation of the transition matrix and the higher-level cost-effectiveness calculations. This makes changes and testing to either part easier and more reliable. This function is defined and broken into three components as follows. Figure 11a outlines the first part of the function, which defined the fixed transition probabilities tpProg and tpDcm as input arguments to the function with default values as well as the proportion reduction of the transition probability from asymptomatic to progressive disease due to the drug (effect = 0.5). Note that, strictly speaking, in R this is an array data structure but we have named the data structure p\_matrix to emphasise that it provides what is known in Markov modelling as the transition probability matrix. The time-dependent probabilities are those that depend on the age of the cohort. A look-up function is used to describe the transition probability from Asymp to Dead using 6 age-group categories as in the MS Excel model.

Figure 11b, c outline the specific calculations for the transition probabilities including those dependent on age linked to the age-specific values, as well as the fixed values. This p\_matrix\_cycle function can now be used when calculating the costs and QALYs over time as it will calculate the corresponding p\_matrix at each cycle. The filled array of transition probabilities (p\_matrix), matrices representing state costs and QALYs for the different health states (state\_c\_matrix, state\_q\_matrix) and transition costs and QALYs (trans\_c\_matrix, trans\_q\_matrix) have been constructed



```
p_matrix_cycle <- function(p_matrix, age, cycle,
                           tpProg = 0.01,
                           tpDcm = 0.15,
                           effect = 0.5) {
  tpDn_lookup <-
    c("(34,44]" = 0.0017,
      "(44,54]" = 0.0044,
      "(54,64]" = 0.0138,
      "(64,74]" = 0.0379,
      "(74,84]" = 0.0912,
      "(84,100]" = 0.1958)

  age_grp <- cut(age, breaks = c(34,44,54,64,74,84,100))
  tpDn <- tpDn_lookup[age_grp]
```

(a)

```
p_matrix["Asymptomatic_disease", "Progressive_disease", "without_drug"] <- tpProg*cycle
p_matrix["Asymptomatic_disease", "Dead", "without_drug"] <- tpDn
p_matrix["Asymptomatic_disease", "Asymptomatic_disease", "without_drug"] <- 1 - tpProg*cycle - tpDn
p_matrix["Progressive_disease", "Dead", "without_drug"] <- tpDcm + tpDn
p_matrix["Progressive_disease", "Progressive_disease", "without_drug"] <- 1 - tpDcm - tpDn
p_matrix["Dead", "Dead", "without_drug"] <- 1
```

(b)

```
p_matrix["Asymptomatic_disease", "Progressive_disease", "with_drug"] <- tpProg*(1 - effect)*cycle
p_matrix["Asymptomatic_disease", "Dead", "with_drug"] <- tpDn
p_matrix["Asymptomatic_disease", "Asymptomatic_disease", "with_drug"] <-
  1 - tpProg*(1 - effect)*cycle - tpDn
p_matrix["Progressive_disease", "Dead", "with_drug"] <- tpDcm + tpDn
p_matrix["Progressive_disease", "Progressive_disease", "with_drug"] <- 1 - tpDcm - tpDn
p_matrix["Dead", "Dead", "with_drug"] <- 1

return(p_matrix)
}
```

(c)

**Fig. 11** **a** The first part of the probability transition matrix function `p_matrix_cycle`, which assigns age-dependent probabilities in R. **b** The second part of the function `p_matrix_cycle` calculating transition probabilities for the cohort without a drug, where the array dimen-

sions are called in R by array [row, columns, dimension]. **c** The third and last part of the function `p_matrix_cycle` calculating transition probabilities for the cohort with a drug

based on the parameters defined in the beginning of the model code. Now, we run the model by combining these objects, much like calculations performed across multiple MS Excel sheets.

## 2.6 Running the Model

To run the model and combine all the information and code from the previous sections, an algorithm will be created. The steps in an algorithm must be ‘flattened-out’ in MS Excel. That is, one must copy and paste the same formula across several cells in order to repeat a calculation for different input values. Using the for-loop means that we do not have to do this.

A loop is first created using the `p_matrix` function. Figure 12 gives the R code used with the starting age of the cohort, and subsequently updating the age and the corresponding pop and trans matrices. This loop repeats from

cycle number 2 to cycle 46 (`n_cycles`), as cycle 1 was already defined above, equivalent to cycle 0 rows in MS Excel models. There is also an element `i` in this loop, which we shall see is from another encapsulating loop for the two treatments, which incorporates this loop. Recall the matrix multiplication operator `%*%`, used here to calculate the state population at the next time step from the current time population (pop) and the number of individuals who transition between states (trans). Notice the connection with the discussion in Sect. 2.2. This is the equivalent R code to the MS Excel pseudocode of Fig. 4.

If we execute this code snippet for a single treatment group of without drug ( $i = 1$ ) and an initial age fixed at age = 50, then the first 10-time steps of the pop array are shown in Fig. 13.

To complete the simulation code, we now also present the outer loop which uses the state population counts to calculate the cost-effectiveness analysis statistics in Fig. 14. To

```

for (j in 2:n_cycles) {
  p_matrix <- p_matrix_cycle(p_matrix, age, j - 1)

  pop[, cycle = j, treatment = i] <-
    pop[, cycle = j - 1, treatment = i] %%% p_matrix[, , treatment = i]

  trans[, cycle = j, treatment = i] <-
    pop[, cycle = j - 1, treatment = i] %%% (trans_c_matrix * p_matrix[, , treatment = i])

  age <- age + 1
}

```

**Fig. 12** The first, inner loop in running the model calculating population counts at each cycle in R

```

> pop
, , treatment = without_drug

state      cycle
[,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10]
Asymptomatic_disease 1000 976.2 943.20444 901.89209 853.37029 798.92527 739.9646 677.9556 614.3633 550.5924
Progressive_disease   0  10.0  27.88600  51.61441  79.23565 108.92537 139.0189 168.0451 194.7558 218.1475
Dead                  0   13.8  28.90956  46.49351  67.39406  92.14937 121.0165 153.9993 190.8809 231.2601

```

**Fig. 13** The first 10-time steps of the pop array for the without drug group in R

simplify this code, we substitute where the code snippet in Fig. 12 would be with a function call to `sim_pop()` which contains equivalent code (full code for running the model is given in the Supplementary Material). The cost, QALYs, life-expectancy (LE), life-years (LYs) and quality-adjusted life-expectancy (QALE) at each cycle are calculated as the `p_matrix`, which is updated at each iteration of the loop. The discount rate is also incorporated into the model here easily as each cycle's costs and QALYs will depend on the cycle number. These repeated steps are performed for each of the two treatments (1:n treatments) and the total costs and QALYs over the lifetime of the model can then be calculated for each treatment.

## 2.7 Results

Displaying the results after running the model is very easy in R. As the incremental cost-effectiveness ratio (ICER) requires the incremental costs and incremental QALYs, Fig. 15 gives the two lines of R code needed to subtract the total costs of the without-drug cohort from the with-drug cohort. These results will therefore assume that the strategy where the cohorts are without a drug is the standard of care or base-case analysis. Swapping with and without the drug will change this around. It is important to note that the calculation of the ICER is relevant here as there are two strategies being compared. If there are more than two strategies, more than one ICER may be required to be calculated. In the case of more than two strategies and one is dominated, an ICER would not be relevant for that strategy compared to the others. The model results are also presented

in a cost-effectiveness table in the Supplementary Material with the R code required to create it.

The code to plot the results on a cost-effectiveness plane is given in Fig. 16. The diagonal line indicates the willingness to pay threshold of £20,000 per QALY and the ICER is indicated by the black point. We can see that the drug treatment is cost-effective against the no drug option. The x-axis and y-axis limits are set with `xlim` and `ylim` arguments. These can be omitted to allow R to calculate the axes limits if desired. Note that the plot in Fig. 17 is a graph created using base R. There are many packages that can display more sophisticated plots, e.g., with a range of colours and other details, which are not covered in this tutorial, including BCCA [10].

## 3 Probabilistic Sensitivity Analysis (PSA)

The formulation in the previous section can be extended to include uncertainty about one or more of the parameters. Briggs (2000) [18] describe this for the current model by repeating the analytical solution of the model employing different values for the underlying parameters sampled from specified ranges and distributions. The MS Excel implementation of the example in [18] is also provided in the GitHub repository for this tutorial paper. Sensitivity analyses can be performed one at a time (one-way) or for multiple parameters simultaneously (multi-way). This section presents a multi-way probabilistic sensitivity analysis.

In MS Excel random numbers can be generated most simply for a uniform distribution using the `RAND()` function.

```

for (i in 1:n_treatments) {

  # simulate state populations
  sim_res ←
    sim_pop(n_cycles, Initial_age,
            trans_c_matrix,
            p_matrix, pop, trans, i)

  trans[, , i] ← sim_res$trans
  pop[, , i] ← sim_res$pop

  cycle_state_costs[i, ] ←
    (state_c_matrix[treatment = i, ] %*% pop[, , treatment = i]) * 1/(1 + cDr)^(1:n_cycles - 1)

  # discounting at _previous_ cycle
  cycle_trans_costs[i, ] ←
    (c(1,1,1) %*% trans[, , treatment = i]) * 1/(1 + cDr)^(1:n_cycles - 2)

  cycle_costs[i, ] ← cycle_state_costs[i, ] + cycle_trans_costs[i, ]

  # life expectancy
  LE[i, ] ← c(1,1,0) %*% pop[, , treatment = i]

  # life-years
  LYs[i, ] ← LE[i, ] * 1/(1 + oDr)^(1:n_cycles - 1)

  # quality-adjusted life expectancy
  cycle_QALE[i, ] ←
    state_q_matrix[treatment = i, ] %*% pop[, , treatment = i]

  # quality-adjusted life-years
  cycle_QALYs[i, ] ← cycle_QALE[i, ] * 1/(1 + oDr)^(1:n_cycles - 1)

  total_costs[i] ← sum(cycle_costs[treatment = i, -1])
  total_QALYs[i] ← sum(cycle_QALYs[treatment = i, -1])
}

```

**Fig. 14** The simulation code including the second, outer loop of the main model which calculates cost-effectiveness analysis summary statistics in R

**Fig. 15** The R code to calculate the ICER. *ICER* incremental cost-effectiveness ratio

```

c_incr ← total_costs["with_drug"] - total_costs["without_drug"]
q_incr ← total_QALYs["with_drug"] - total_QALYs["without_drug"]

ICER ← c_incr/q_incr

```

**Fig. 16** R code to plot the cost-effectiveness plane

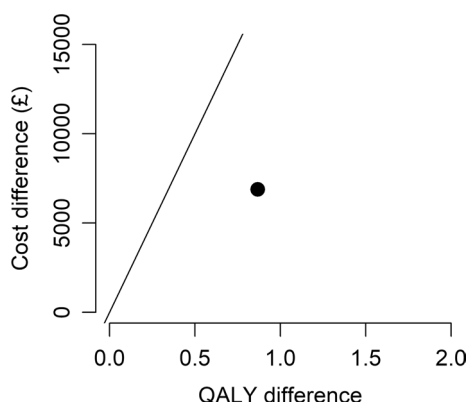
```

wtp ← 20000
plot(x = q_incr/n_cohort, y = c_incr/n_cohort,
     xlim = c(0, 2),
     ylim = c(0, 15e3),
     pch = 16, cex = 1.5,
     xlab = "QALY difference",
     ylab = paste0("Cost difference (", enc2utf8("\u00A3"), ")"),
     frame.plot = FALSE)
abline(a = 0, b = wtp) # willingness-to-pay threshold

```

Samples from some other distribution can be made using an inverse cumulative density transformation e.g., for the gamma distribution with `GAMMA.INV(p,a,b)`. However, a full PSA is arguably not practical without resorting to

additional Visual Basic for Applications (VBA) coding due to the number of simulations run. We recommend that all calculations should remain in the spreadsheet and accessible to the user without the need to understand or interpret



**Fig. 17** Results of the analysis per person on a cost-effectiveness plane comparing drug treatment against no drug cohorts, with an ICER of £7931/QALY represented by the black point. The willingness to pay threshold for £20,000 is represented by the diagonal line. *ICER* incremental cost-effectiveness ratio, *QALY* quality-adjusted life-years

VBA coding, which is only there to facilitate the repeated application of the results. At the point where the user needs to write substantial code inside of MS Excel then there is an especially strong argument to transfer to a software that is more specifically designed for this purpose, such as R.

Performing a PSA analysis can be done by inputting random draws from the unit costs and QALY distributions as inputs to the existing model function. In R, there are numerous ways of implementing a PSA. Following from the R code presented in the previous sections, we can wrap this model code in a function, e.g., called `ce_markov()`, which we can then repeatedly call with different parameter values. To this we will need to pass the starting conditions: population (`start_pop`), age (`init_age`) and number of cycles (`n_cycle`) (in our case, if not defined then age and number of cycles are assigned default values). We will also need the probability transition matrix (`p_matrix`), state cost and QALY matrices (`state_c_matrix`, `state_q_matrix`, `trans_c_matrix`).

In extension to the first analysis, the unit values have distributions rather than point values. To sample from a base R distribution the function name syntax is a short-form version of the distribution name preceded by an ‘r’ (for random or realisation). For example, to sample a single value from a standard normal distribution then call `rnorm(1)`. All of the random numbers could be sampled before running the model, which would allow us to save them to use again and improve run time because this would only be performed once outside of the main loop. Alternatively, we can sample the random variables at run time, within the Markov model function. This is arguably neater and if we wish to replicate a particular run, in the same way as pre-running the sampling, then the random seed can be set beforehand with `set.seed()`. We will demonstrate how to implement a simple version

```
cAsymp <- function() rnorm(1, 500, 127.55)
cDeath <- function() rnorm(1, 1000, 255.11)
cDrug <- function() rnorm(1, 1000, 102.04)
cProg <- function() rnorm(1, 3000, 510.21)
effect <- function() rnorm(1, 0.5, 0.051)
tpDcm <- function() rbeta(1, 29, 167)
tpProg <- function() rbeta(1, 15, 1506)
uAsymp <- function() rbeta(1, 69, 4)
uProg <- function() rbeta(1, 24, 8)
```

**Fig. 18** R code for random sampling input values in probability sensitivity analysis (PSA)

when sampling at runtime. We will not assume any correlation between parameters but if multivariate sampling is required then this may be another reason to use R. Because we will want to sample repeatedly, rather than just once at the start of the simulation, we can wrap the random sampling statements in a function so that they are called newly every time the Markov model is run. We use the same names as we used for the point values in the previous analysis (see Fig. 18).

Similarly, rather than using fixed `state_c_matrix`, `trans_c_matrix` and `state_q_matrix`, if we define these as functions, we can sample newly their component values each time they are called. In practice, the code looks the same as previously but now the unit values are function calls so are followed by open and closed brackets. The example for `state_c_matrix` is shown in Fig. 19 and the full code is given in Supplementary Material.

To finally obtain the PSA output, loop over `ce_markov()` remembering to record the cost and QALYs outputs each time (see Fig. 20).

The output of this gives the cost-effectiveness plane in Fig. 21. Code to produce this plot is given in the Supplementary Material.

## 4 Discussion

This tutorial paper has detailed a step-by-step tutorial to implement a Markov model in R for MS Excel users. This was based on a widely used original MS Excel model so that users can move more easily from using one software to the other by following the steps. Both the R code and the MS Excel workbook are freely available online at [https://github.com/Excel-R-tutorials/Markov\\_Intro](https://github.com/Excel-R-tutorials/Markov_Intro).

The gap between MS Excel and R can be bridged by not “throwing the baby out with the bathwater” when difficult modelling approaches and assumptions arise for beginner users of R. Through practical experience and discussions, the authors have learned that there are several drawbacks to creating HEE models in MS Excel, and to promote the use of R these need to be addressed. Many individuals are keen



**Fig. 19** R code for state cost matrix in probability sensitivity analysis (PSA)

```
state_c_matrix <- function() {
  matrix(c(cAsymp(), cProg(), 0, # without drug
          cAsymp() + cDrug(), cProg(), 0), # with drug
        byrow = TRUE,
        nrow = n_treatments,
        dimnames = list(t_names,
                        s_names))
}
```

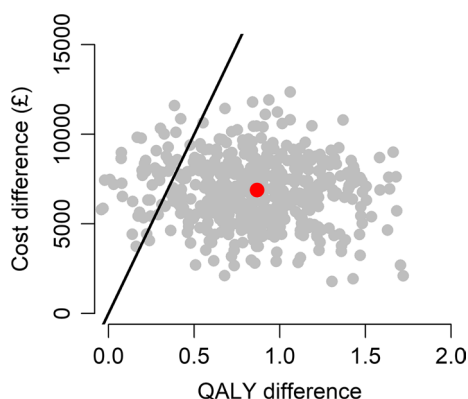
**Fig. 20** R code for full analysis model in probability sensitivity analysis (PSA)

```
n_trials <- 500

costs <- matrix(NA, nrow = n_trials, ncol = n_treatments,
               dimnames = list(NULL, t_names))
qalys <- matrix(NA, nrow = n_trials, ncol = n_treatments,
               dimnames = list(NULL, t_names))

for (i in 1:n_trials) {
  ce_res <- ce_markov(start_pop = c(n_cohort, 0, 0),
                     p_matrix,
                     state_c_matrix(),
                     trans_c_matrix(),
                     state_q_matrix())

  costs[i, ] <- ce_res$total_costs
  qalys[i, ] <- ce_res$total_QALYs
}
```



**Fig. 21** PSA cost-effectiveness plane per person comparing drug treatment against no drug cohorts. The diagonal line indicates the willingness to pay threshold of £20,000 per QALY. The red point represents the ICER. *ICER* incremental cost-effectiveness ratio, *PSA* probability sensitivity analysis, *QALY* quality-adjusted life-years

to learn new tools if they feel it is worthwhile to commit the extra time and effort and talking their “Excel language” is an important part of this. It is also important to note that performing a probabilistic sensitivity analysis in MS Excel requires the use of VBA code, and so these general programming skills will be highly beneficial when moving to R.

Several packages have been written specifically for HEE modelling in R such as *heemod* [15], *hesim* [24] and *BCEA*

[10]. The increasing number of available HEE R packages can be considered in light of an ‘open source modelling’ movement in the health economics community (see the R for Health Technology Assessment group [<https://r-hta.org/>] and the International Society for Pharmacoeconomics and Outcomes Research [ISPOR] Open Source Models Special Interest Group (<https://www.ispor.org/member-groups/special-interest-groups/open-source-models>)). These packages make it easier and more robust to implement and analyse a wide variety of models in R. In many practical cases, these packages may be a preferred solution to coding up a HEE model from scratch. They are open source, meaning that the code can be inspected, modified, and enhanced by anyone. They are also used by hundreds or thousands of people so errors can be identified and corrected that might be missed in someone's de novo code. However, we wish to highlight that a rationale for using standard MS Excel for modelling, in contrast to add-ins with additional functionality (such as @Risk) is to help the modeller to understand and question the assumptions and principles of constructing a model rather than simply how to operate a piece of software. We propose that the same principle applies to this tutorial. Further, building models in third-party software will never be as transparent or bespoke as a model written by the user in the original language.

Special Interest Group previous papers have highlighted the increased speed and flexibility in using R over other

software for HEE [11–13]. Additionally, utilising R allows for the fairly easy conversion of R code into more user-friendly models through RShiny apps [3, 8]. The intention of this tutorial paper was not to show the “best” way of creating cost-effectiveness models in R, but the code has been deliberately simplified so that the barrier to entry for people new to R is lowered. Using explicit loops rather than using the mapping functions (e.g., the base R apply family of functions) show an easier and more explicit flow of the programme and reasoning. In R there are often numerous ways of solving the same problem, depending on personal preference and experience (see base R vs tidyverse discussions [23]). This could include use of a completely different programming paradigm, such as object-oriented programming [25]. We created code that is intuitive and simple and not necessarily the shortest, fastest or most elegant. Additionally, for improved reproducibility in R, analyses can be documented with literate programming notebooks such as R Markdown [26] or Quarto [27]. These interleave the code and accompanying text to provide details about procedures and data so the same analysis could be repeated. Through open-source, documented, clean code we hope to promote the increased transparency and re-use of future health economic models.

Once users who are new to R have grasped the basic implementation of HEEs in R, the next stage is to move on to other R tutorials [12, 13] and develop more general R skills [25, 28]. This can allow integration of statistical analyses and economic analyses in the same software. In addition, further HEE analyses such as Value of Information, which is difficult to perform in MS Excel, can be performed relatively easily in R to provide a more robust HEE in addition to all of the other benefits of performing a HEE in R.

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## Declarations

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**Competing interests** The authors declare they have no conflicts of interest.

**Consent to participate** Not applicable.

**Ethics approval** Not applicable.

**Consent for publication** Not applicable.

**Data availability** All data used for this tutorial paper are available at <https://github.com/Excel-R-tutorials/Markov-model-introduction>

**Code availability** Code for the R model used in this tutorial can be found on GitHub at the following link [https://github.com/Excel-R-tutorials/Markov\\_Intro](https://github.com/Excel-R-tutorials/Markov_Intro). The corresponding original MS Excel model is available open-access and can be downloaded from the following link <https://doi.org/10.17037/DATA.00002980> [20].

**Authorship statement** N. Green: Conceptualisation, writing original draft, validation, writing review and editing. F. Lamrock: Conceptualisation, writing original draft, validation, writing review and editing. N. Naylor: validation, writing review and editing. J. Williams: validation, writing review and editing. A. Briggs: Resources, writing review and editing.

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