Serum degradation analysis by MALDI/ToF: a new method and tool

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Introduction

One of the potential risks in a biomarker discovery study is that samples with different preservation time and conditions are compared. For example, pathological serum samples collected over several years and cryo-preserved in biobanks could be compared with control samples recently obtained from healthy donors. The statistical analysis aimed at the identification of differences between the two groups could identify as biomarkers of a pathological condition signals whose difference could only depend on the degradation process of the pathological samples. Pre-analytical variables, like patient conditions, venipuncture details, etc..., may also alter the analysis of blood derived samples. Here, we propose a simple method, mass spectrometry based, able to assess the integrity of serum samples

by evaluating their absolute and relative fibrinopeptide contents.

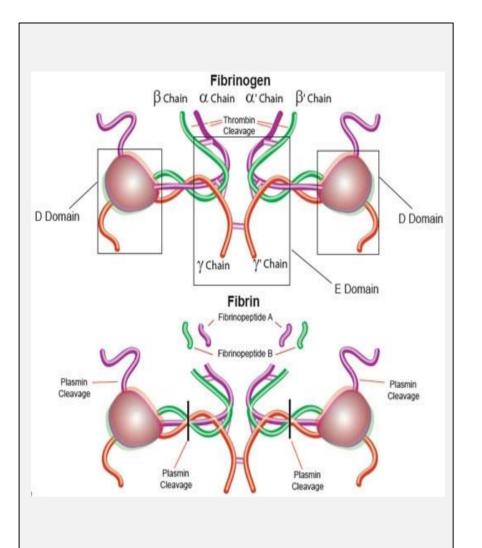


Figure 1: mechanism of fibrinogen activation. (From Sigma-Aldrich Co. LLC.)

FpA degradation

The peptidomic profile of a fresh serum sample is mainly characterized by signals due to fibrinopeptides, generated during the coagulation process (fig. 1), mainly fibrinopeptide A (fpA) and its degradation products (table 1).

A recent study showed a gradual degradation of the fpA molecule, as a result of the preservation process [1]. In the sera stored for 18 months at -80°C, a significant percentage decrease of the most conserved molecules of fpA was observed along with the percentage increase of the most degraded forms. Figure 2 shows the MALDI/ToF spectra of a fresh (panel A) and a poorly preserved (panel B) serum sample. Moreover, also the total amount of fpA, in all its forms, in the cryo-preserved samples is clearly lower than the one calculated for fresh sera (fig. 3). These data are summarized in figure 4 where the percentage of total fpA is plotted against the percentage of high MW fpA for each sample, and in figure 5 which shows the differences, in terms of percentage, between fresh and cryopreserved samples for each fpA form.

The high susceptibility of fpA to degradation suggests a possible use of this molecule as quality indicator of the cryo-preserved serum samples.

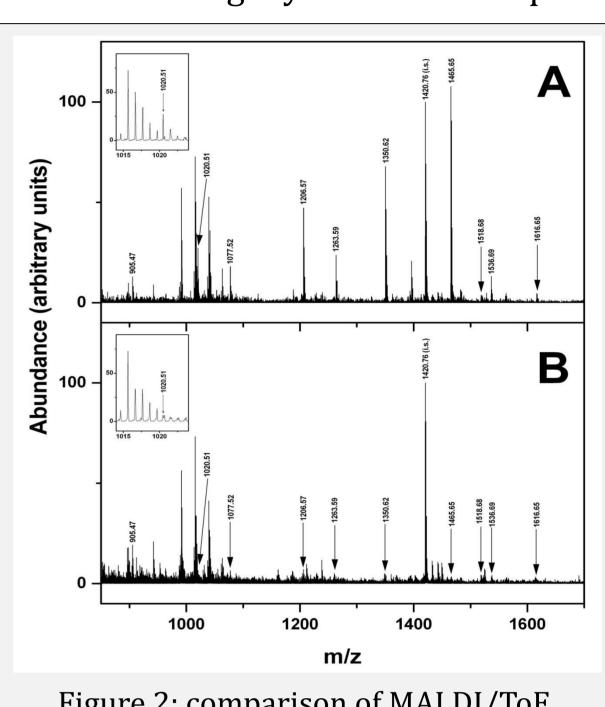


Figure 2: comparison of MALDI/ToF spectra from fresh and cryopreserved samples

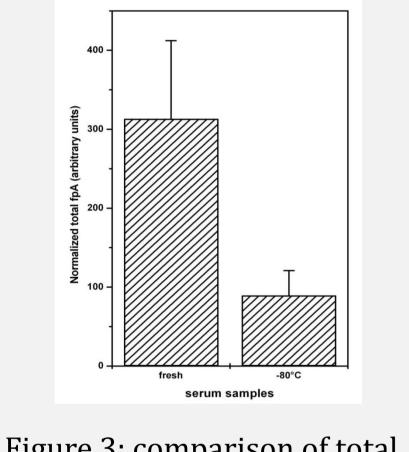


Figure 3: comparison of total fpA abundance in fresh and cryopreserved samples

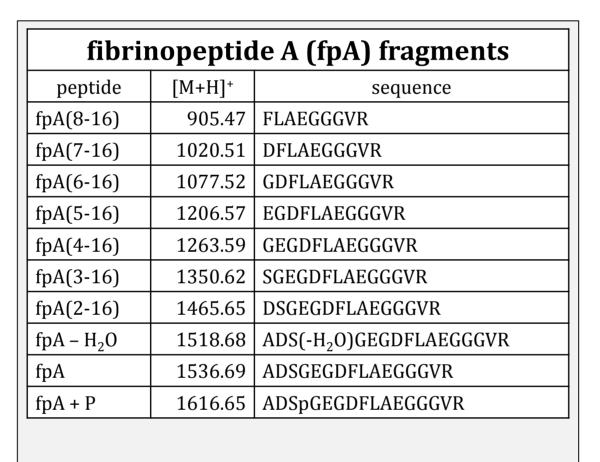


Table 1: fpA peptides

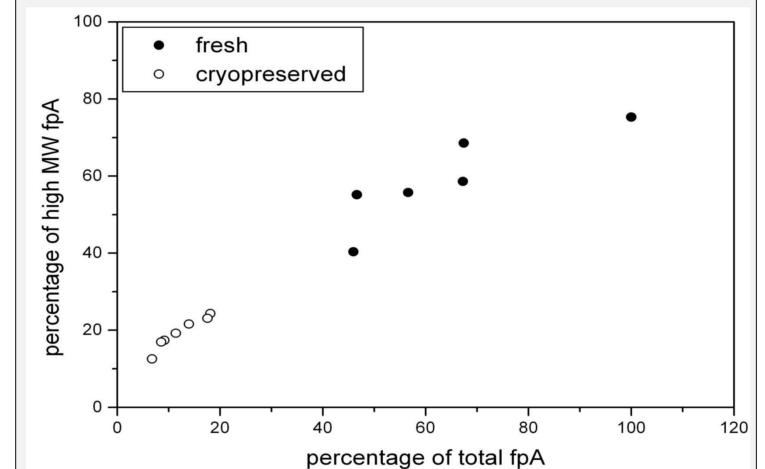


Figure 4: the percentage of total fpA plotted against the percentage of high MW fpA for each sample

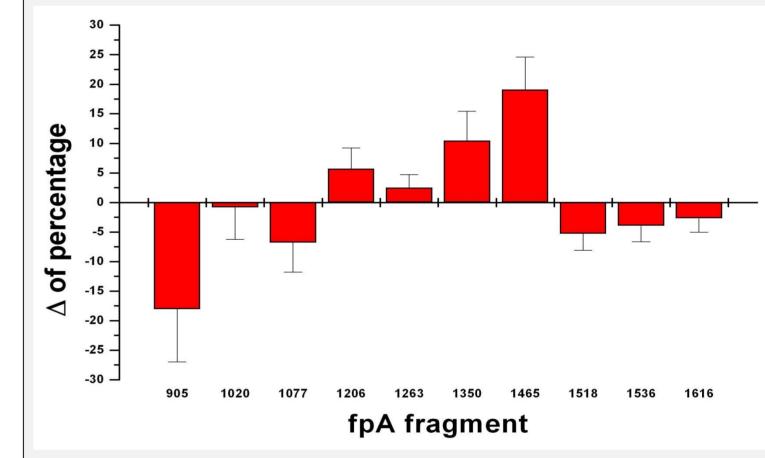


Figure 5: differences of percentage between fresh and cryopreserved samples for each fpA form

Web based degradation analysis

We developed a web tool to assess the integrity of serum samples by evaluating their fibrinopeptide contents. The tool was built in a LAMP (Linux, Apache, MySQL, PHP) environment and is available online to all interested researchers.

The tool processes input spectra to extract peak lists which are then elaborated for noise reduction. FpA peptides are identified and compared in order to compute both their total abundance and the percent contribution of each peptide. If a reference spectrum from a sample of good quality is provided, total abundances are expressed as percent of its total abundance. A qualitative score is assigned to spectra by taking into account both their total abundance and the ratio between abundances of more and less degraded forms.

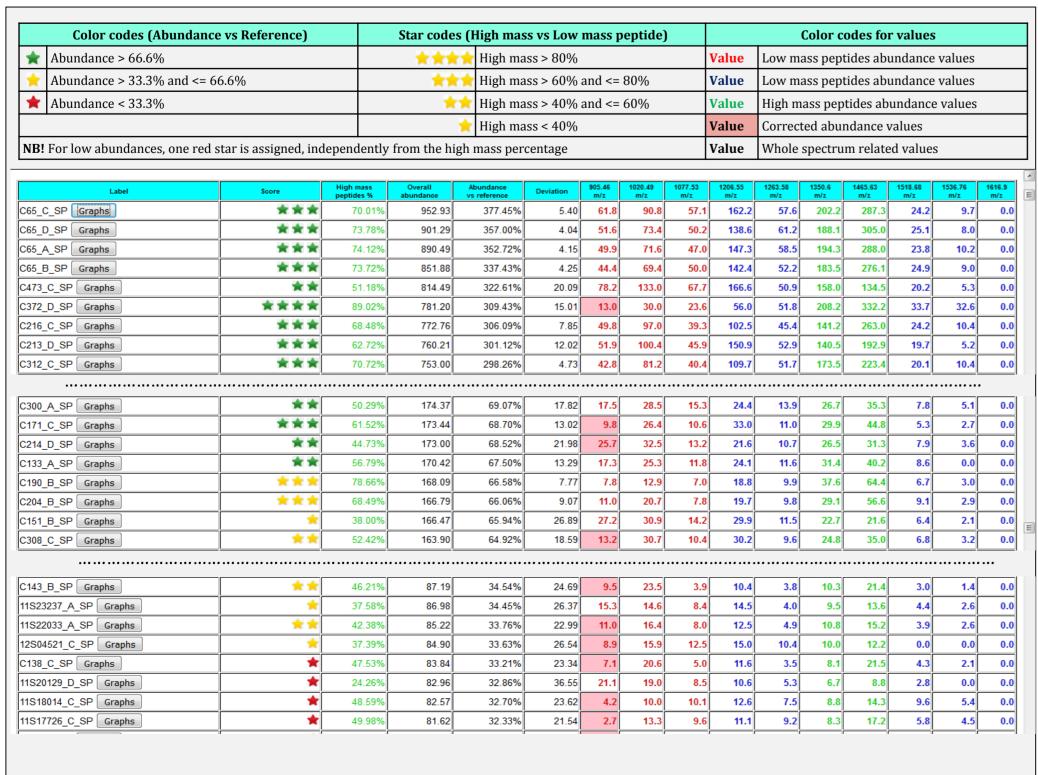


Figure 7: list of spectra, ordered by decreasing total abundance, the assigned score and their features

Input

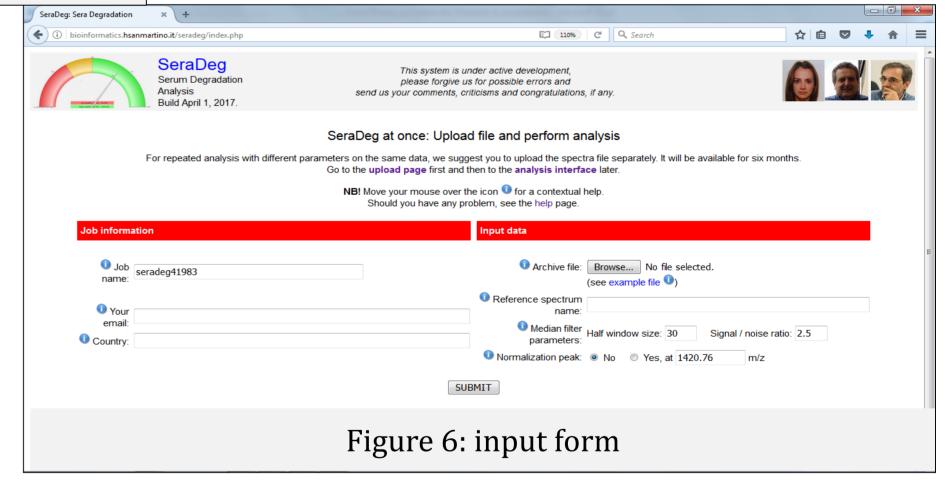
The input consists in a compressed archive of all spectra which must be formatted as CVS files including m/z – intensity pairs.

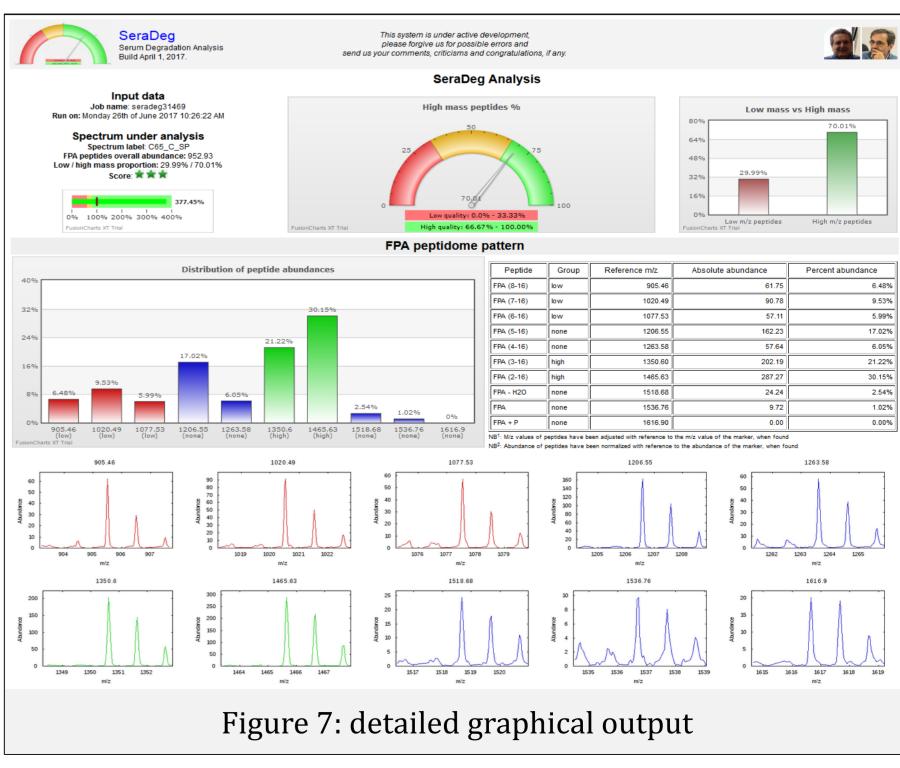
Archives may be uploaded and analysed at a later time. See fig. 6.

Output

The output consists in the list of spectra, ordered by decreasing total abundance, the assigned score and their features. See fig. 7.

For each spectrum, a detailed output, including a graphical representation of all features taken into analysis in the determination of the quality of the samples, may be requested. See fig. 8.





References

1. Mangerini R, Romano P et al. Anal Biochem 2011, 417:174-81