

SARS-CoV-2 and COVID-19: An Evolving Review of Diagnostics and Therapeutics

This manuscript ([permalink](#)) was automatically generated from [greenelab/covid19-review@ca5421d](#) on May 9, 2022. It is also available as a [PDF](#). Snapshots of individual sections have been published [[1](#),[2](#),[3](#),[4](#)] or posted as preprints [[5](#)].

ⓘ This in progress manuscript is not intended for the general public.

This is a review paper that is authored by scientists for an audience of scientists to discuss research that is in progress. If you are interested in guidelines on testing, therapies, or other issues related to your health, you should not use this document. Instead, you should collect information from your local health department, the [CDC's guidance](#), or your own government.

Authors

- **Halie M. Rando**

 [0000-0001-7688-1770](#) ·  [rando2](#) ·  [tamefoxtime](#)

Department of Systems Pharmacology and Translational Therapeutics, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America; Department of Biochemistry and Molecular Genetics, University of Colorado School of Medicine, Aurora, Colorado, United States of America; Center for Health AI, University of Colorado School of Medicine, Aurora, Colorado, United States of America · Funded by the Gordon and Betty Moore Foundation (GBMF 4552); the National Human Genome Research Institute (R01 HG010067)

- **Casey S. Greene**

 [0000-0001-8713-9213](#) ·  [cgreene](#) ·  [GreeneScientist](#)

Department of Systems Pharmacology and Translational Therapeutics, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America; Childhood Cancer Data Lab, Alex's Lemonade Stand Foundation, Philadelphia, Pennsylvania, United States of America; Department of Biochemistry and Molecular Genetics, University of Colorado School of Medicine, Aurora, Colorado, United States of America; Center for Health AI, University of Colorado School of Medicine, Aurora, Colorado, United States of America · Funded by the Gordon and Betty Moore Foundation (GBMF 4552); the National Human Genome Research Institute (R01 HG010067)

- **Michael P. Robson**

 [0000-0002-4859-0033](#) ·  [mprobson](#)

Department of Computing Sciences, Villanova University, Villanova, Pennsylvania, United States of America

- **Simina M. Boca**

 [0000-0002-1400-3398](#) ·  [SiminaB](#)

Innovation Center for Biomedical Informatics, Georgetown University Medical Center,
Washington, District of Columbia, United States of America

- **Nils Wellhausen**

 [0000-0001-8955-7582](#) ·  [nilswellhausen](#)

Department of Systems Pharmacology and Translational Therapeutics, University of
Pennsylvania, Philadelphia, Pennsylvania, United States of America

- **Ronan Lordan**

 [0000-0001-9668-3368](#) ·  [RLordan](#) ·  [el_ronan](#)

Institute for Translational Medicine and Therapeutics, Perelman School of Medicine,
University of Pennsylvania, Philadelphia, PA 19104-5158, USA; Department of Medicine,
Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA;
Department of Systems Pharmacology and Translational Therapeutics, Perelman School
of Medicine, University of Pennsylvania; Philadelphia, PA 19104, USA

- **Christian Brueffer**

 [0000-0002-3826-0989](#) ·  [cbrueffer](#) ·  [cbrueffer](#)

Department of Clinical Sciences, Lund University, Lund, Sweden

- **Sandipan Ray**

 [0000-0002-9960-5768](#) ·  [rays1987](#)

Department of Biotechnology, Indian Institute of Technology Hyderabad, Kandi,
Sangareddy 502285, Telangana, India

- **Lucy D'Agostino McGowan**

 [0000-0001-7297-9359](#) ·  [LucyMcGowan](#) ·  [LucyStats](#)

Department of Mathematics and Statistics, Wake Forest University, Winston-Salem, North
Carolina, United States of America

- **Anthony Gitter**

 [0000-0002-5324-9833](#) ·  [agitter](#) ·  [anthonygitter](#)

Department of Biostatistics and Medical Informatics, University of Wisconsin-Madison,
Madison, Wisconsin, United States of America; Morgridge Institute for Research,
Madison, Wisconsin, United States of America · Funded by John W. and Jeanne M. Rowe
Center for Research in Virology

- **Anna Ada Dattoli**

 [0000-0003-1462-831X](#) ·  [aadattoli](#) ·  [aadattoli](#)

Department of Pathology and Laboratory Medicine, The Children's Hospital of
Philadelphia, Philadelphia, PA, USA; Department of Systems Pharmacology &
Translational Therapeutics, Perelman School of Medicine, University of Pennsylvania,
Philadelphia, PA, USA

- **Ryan Velazquez**

 [0000-0002-3655-3403](#) ·  [rdvelazquez](#)

Azimuth1, McLean, Virginia, United States of America

- **John P. Barton**

 [0000-0003-1467-421X](#) ·  [johnbarton](#) ·  [_jpbarton](#)

Department of Physics and Astronomy, University of California-Riverside, Riverside,

California, United States of America

- **Jeffrey M. Field**

 [0000-0001-7161-7284](#) ·  [Jeff-Field](#)

Department of Systems Pharmacology and Translational Therapeutics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

- **Bharath Ramsundar**

 [0000-0001-8450-4262](#) ·  [rbharath](#) ·  [rbhar90](#)

The DeepChem Project, <https://deepchem.io/>

- **Adam L. MacLean**

 [0000-0003-0689-7907](#) ·  [adamlmaclean](#) ·  [adamlmaclean](#)

Department of Quantitative and Computational Biology, University of Southern California, Los Angeles, California, United States of America

- **Alexandra J. Lee**

 [0000-0002-0208-3730](#) ·  [ajlee21](#)

Department of Systems Pharmacology and Translational Therapeutics, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America · Funded by the Gordon and Betty Moore Foundation (GBMF 4552)

- **Immunology Institute of the Icahn School of Medicine**

·  [ismms-himc](#)

Immunology Institute of the Icahn School of Medicine

- **Fengling Hu**

 [0000-0003-1081-5038](#) ·  [hufengling](#) ·  [hufengling](#)

Department of Biostatistics, Epidemiology and Informatics, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America

- **Nafisa M. Jadavji**

 [0000-0002-3557-7307](#) ·  [nafisajadavji](#) ·  [nafisajadavji](#)

Biomedical Science, Midwestern University, Glendale, AZ, United States of America; Department of Neuroscience, Carleton University, Ottawa, Ontario, Canada · Funded by the American Heart Association (20AIREA35050015)

- **Elizabeth Sell**

 [0000-0002-9658-1107](#) ·  [esell17](#)

Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America

- **Vincent Rubinetti**

 [0000-0002-4655-3773](#) ·  [vincerubinetti](#)

Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America; Center for Health AI, University of Colorado School of Medicine, Aurora, Colorado, United States of America

- **Jinhui Wang**

 [0000-0002-5796-8130](#) ·  [jinhui2](#)

Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America

- **Diane N. Rafizadeh**
 [0000-0002-2838-067X](#) ·  [dianerafi](#)
Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America; Department of Chemistry, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America · Funded by NIH Medical Scientist Training Program T32 GM07170
- **Ashwin N. Skelly**
 [0000-0002-1565-3376](#) ·  [anskelly](#)
Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America; Institute for Immunology, University of Pennsylvania Perelman School of Medicine, Philadelphia, United States of America · Funded by NIH Medical Scientist Training Program T32 GM07170
- **Marouen Ben Guebila**
 [0000-0001-5934-966X](#) ·  [marouenbg](#) ·  [marouenbg](#)
Department of Biostatistics, Harvard School of Public Health, Boston, Massachusetts, United States of America
- **Likhitha Kolla**
 [0000-0002-1169-906X](#) ·  [likhithakolla](#) ·  [lkolla2018](#)
Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America · Funded by NIH Medical Scientist Training Program T32 GM07170
- **David Manheim**
 [0000-0001-8599-8380](#) ·  [davidmanheim](#) ·  [davidmanheim](#)
1DaySooner, Delaware, United States of America; Risk and Health Communication Research Center, School of Public Health, University of Haifa, Haifa, Israel; Technion, Israel Institute of Technology, Haifa, Israel · Funded by Center for Effective Altruism, Long Term Future Fund
- **Soumita Ghosh**
 [0000-0002-2783-2750](#) ·  [soumitagh](#)
Institute of Translational Medicine and Therapeutics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America
- **James Brian Byrd**
 [0000-0002-0509-3520](#) ·  [byrdjb](#) ·  [thebyrdrlab](#)
University of Michigan School of Medicine, Ann Arbor, Michigan, United States of America · Funded by NIH K23HL128909; FastGrants
- **YoSon Park**
 [0000-0002-0465-4744](#) ·  [ypar](#) ·  [yoson](#)
Department of Systems Pharmacology and Translational Therapeutics, University of Pennsylvania, Philadelphia, Pennsylvania, United States of America · Funded by NHGRI R01 HG10067
- **Vikas Bansal**
 [0000-0002-0944-7226](#) ·  [bansalvi](#) ·  [VikasBansal1989](#)
Biomedical Data Science and Machine Learning Group, German Center for Neurodegenerative Diseases, Tübingen 72076, Germany

- **Stephen Capone**
ID [0000-0001-7231-1535](#) ·  [scapone01](#)
St. George's University School of Medicine, St. George's, Grenada
- **John J. Dziak**
ID [0000-0003-0762-5495](#) ·  [dziakj1](#)
Edna Bennett Pierce Prevention Research Center, The Pennsylvania State University, University Park, PA, United States of America
- **Yuchen Sun**
·  [kevinsunofficial](#)
Department of Computer Science, University of Virginia, Charlottesville, VA, United States of America
- **Yanjun Qi**
ID [0000-0002-5796-7453](#) ·  [qiyanjun](#)
Department of Computer Science, University of Virginia, Charlottesville, VA, United States of America
- **Lamonica Shinholster**
ID [0000-0001-6285-005X](#) ·  [LSH2126](#)
Mercer University, Macon, GA, United States of America · Funded by the Center for Global Genomics and Health Equity at the University of Pennsylvania
- **Temitayo Lukan**
·  [tlukan](#)
University of Pennsylvania, Philadelphia, PA, United States of America
- **Sergey Knyazev**
ID [0000-0003-0385-1831](#) ·  [Sergey-Knyazev](#) ·  [SeKnyaz](#)
Georgia State University, Atlanta, GA, United States of America
- **Dimitri Perrin**
ID [0000-0002-4007-5256](#) ·  [SystemsResearch](#) ·  [dperrin](#)
School of Computer Science, Queensland University of Technology, Brisbane, Australia; Centre for Data Science, Queensland University of Technology, Brisbane, Australia
- **Serghei Mangul**
ID [0000-0003-4770-3443](#) ·  [smangul1](#) ·  [serghei_mangul](#)
Department of Clinical Pharmacy, School of Pharmacy, University of Southern California, Los Angeles, CA, United States of America
- **Shikta Das**
ID [0000-0002-8291-2788](#) ·  [shiktadas](#) ·  [shikta_das](#)
C4X Discovery, London, United Kingdom; Medical Research Council LHA, Institute of Cardiovascular Studies, University College London, London, United Kingdom
- **Gregory L Szeto**
ID [0000-0001-7604-1333](#) ·  [gregszetoAI](#) ·  [greg_szeto](#)
Allen Institute for Immunology, Seattle, WA, United States of America

- **Tiago Lubiana**
 [0000-0003-2473-2313](#) ·  [lubianat](#) ·  [lubianat](#)
Department of Clinical and Toxicological Analyses, School of Pharmaceutical Sciences,
University of São Paulo, São Paulo, Brazil
- **David Mai**
 [0000-0002-9238-0164](#) ·  [davemai](#) ·  [lococyte](#)
Department of Bioengineering, University of Pennsylvania, Philadelphia, PA, USA; Center
for Cellular Immunotherapies, Perelman School of Medicine, and Parker Institute for
Cancer Immunotherapy at University of Pennsylvania, Philadelphia, PA, USA
- **COVID-19 Review Consortium**
- **Rishi Raj Goel**
 [0000-0003-1715-5191](#) ·  [rishirajgoel](#) ·  [rishirajgoel](#)
Institute for Immunology, University of Pennsylvania, Philadelphia, PA, United States of
America
- **Joel D Boerckel**
 [0000-0003-3126-3025](#) ·  [jboerckel](#) ·  [jboerckel](#)
Department of Orthopaedic Surgery, Perelman School of Medicine, University of
Pennsylvania, Philadelphia, PA, United States of America; Department of Bioengineering,
University of Pennsylvania, Philadelphia, PA, United States of America
- **Amruta Naik**
 [0000-0003-0673-2643](#) ·  [NAIKA86](#)
Children's Hospital of Philadelphia, Philadelphia, PA, United States of America
- **Yusha Sun**
 [0000-0003-4835-3000](#) ·  [yusha-sun](#)
Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania,
United States of America
- **Daniel S. Himmelstein**
 [0000-0002-3012-7446](#) ·  [dhimmel](#) ·  [dhimmel](#)
Department of Systems Pharmacology and Translational Therapeutics, University of
Pennsylvania, Philadelphia, Pennsylvania, United States of America; Related Sciences ·
Funded by GBMF4552
- **Jeremy P. Kamil**
 [0000-0001-8422-7656](#)
Department of Microbiology and Immunology, Louisiana State University Health Sciences
Center Shreveport, Shreveport, Louisiana, USA
- **Jesse G. Meyer**
 [0000-0003-2753-3926](#) ·  [jessegmeyerlab](#)
Department of Biochemistry, Medical College of Wisconsin, Milwaukee, Wisconsin, United
States of America · Funded by National Institute of General Medical Sciences (R35
GM142502)

- **Ariel I. Mundo**

 [0000-0002-6014-4538](#) ·  [aimundo](#)

Department of Biomedical Engineering, University of Arkansas, Fayetteville, Arkansas,
USA

COVID-19 Review Consortium: Vikas Bansal, John P. Barton, Simina M. Boca, Joel D Boerckel, Christian Brueffer, James Brian Byrd, Stephen Capone, Shikta Das, Anna Ada Dattoli, John J. Dziak, Jeffrey M. Field, Soumita Ghosh, Anthony Gitter, Rishi Raj Goel, Casey S. Greene, Marouen Ben Guebila, Daniel S. Himmelstein, Fengling Hu, Nafisa M. Jadavji, Jeremy P. Kamil, Sergey Knyazev, Likhitha Kolla, Alexandra J. Lee, Ronan Lordan, Tiago Lubiana, Temitayo Lukau, Adam L. MacLean, David Mai, Serghei Mangul, David Manheim, Lucy D'Agostino McGowan, Jesse G. Meyer, Ariel I. Mundo, Amruta Naik, YoSon Park, Dimitri Perrin, Yanjun Qi, Diane N. Rafizadeh, Bharath Ramsundar, Halie M. Rando, Sandipan Ray, Michael P. Robson, Vincent Rubinetti, Elizabeth Sell, Lamonica Shinholster, Ashwin N. Skelly, Yuchen Sun, Yusha Sun, Gregory L Szeto, Ryan Velazquez, Jinhui Wang, Nils Wellhausen

Authors are ordered arbitrarily.

1 Pathogenesis, Symptomatology, and Transmission of SARS-CoV-2 through Analysis of Viral Genomics and Structure

1.1 Abstract

The novel coronavirus SARS-CoV-2, which emerged in late 2019, has since spread around the world and infected hundreds of millions of people with coronavirus disease 2019 (COVID-19). While this viral species was unknown prior to January 2020, its similarity to other coronaviruses that infect humans has allowed for rapid insight into the mechanisms that it uses to infect human hosts, as well as the ways in which the human immune system can respond. Here, we contextualize SARS-CoV-2 among other coronaviruses and identify what is known and what can be inferred about its behavior once inside a human host. Because the genomic content of coronaviruses, which specifies the virus's structure, is highly conserved, early genomic analysis provided a significant head start in predicting viral pathogenesis and in understanding potential differences among variants. The pathogenesis of the virus offers insights into symptomatology, transmission, and individual susceptibility. Additionally, prior research into interactions between the human immune system and coronaviruses has identified how these viruses can evade the immune system's protective mechanisms. We also explore systems-level research into the regulatory and proteomic effects of SARS-CoV-2 infection and the immune response. Understanding the structure and behavior of the virus serves to contextualize the many facets of the COVID-19 pandemic and can influence efforts to control the virus and treat the disease.

1.2 Importance

COVID-19 involves a number of organ systems and can present with a wide range of symptoms. From how the virus infects cells to how it spreads between people, the available research suggests that these patterns are very similar to those seen in the closely related viruses SARS-CoV-1 and possibly MERS-CoV. Understanding the pathogenesis of the SARS-CoV-2 virus also contextualizes how the different biological systems affected by COVID-19 connect. Exploring the structure, phylogeny, and pathogenesis of the virus therefore helps to guide interpretation of the broader impacts of the virus on the human body and on human populations. For this reason, an in-depth exploration of viral mechanisms is critical to a robust understanding of SARS-CoV-2 and, potentially, future emergent HCoV.

1.3 Introduction

The current coronavirus disease 2019 (COVID-19) pandemic, caused by the *Severe acute respiratory syndrome-related coronavirus 2* (SARS-CoV-2) virus, represents an acute global health crisis. Symptoms of the disease can range from mild to severe or fatal [6] and can affect a variety of organs and systems [7]. Outcomes of infection can include acute respiratory distress (ARDS) and acute lung injury, as well as damage to other organ systems [7,8]. Understanding the progression of the disease, including these diverse symptoms, depends on understanding how the virus interacts with the host. Additionally, the fundamental biology of the virus can provide insights into how it is transmitted among people, which can, in turn, inform efforts to control its spread. As a result, a thorough understanding of the pathogenesis of SARS-CoV-2 is a critical foundation on which to build an understanding of COVID-19 and the pandemic as a whole.

The rapid identification and release of the genomic sequence of the virus in January 2020 [9] provided early insight into the virus in a comparative genomic context. The viral genomic sequence clusters with known coronaviruses (order *Nidovirales*, family *Coronaviridae*, subfamily *Orthocoronavirinae*). Phylogenetic analysis of the coronaviruses reveals four major subclades, each corresponding to a genus: the alpha, beta, gamma, and delta coronaviruses. Among them, alpha and beta coronaviruses infect mammalian species, gamma coronaviruses infect avian species, and delta coronaviruses infect both mammalian and avian species [10]. The novel virus now known as SARS-CoV-2 was identified as a beta coronavirus belonging to the B lineage based on phylogenetic analysis of a polymerase chain reaction (PCR) amplicon fragment from five patients along with the full genomic sequence [11]. This lineage also includes the *Severe acute respiratory syndrome-related coronavirus* (SARS-CoV-1) that caused the 2002-2003 outbreak of Severe Acute Respiratory Syndrome (SARS) in humans [11]. (Note that these subclades are not to be confused with variants of concern within SARS-CoV-2 labeled with Greek letters; i.e., the Delta variant of SARS-CoV-2 is still a beta coronavirus.)

Because viral structure and mechanisms of pathogenicity are highly conserved within the order, this phylogenetic analysis provided a basis for forming hypotheses about how the virus interacts with hosts, including which

tissues, organs, and systems would be most susceptible to SARS-CoV-2 infection. Coronaviruses that infect humans (HCoV) are not common, but prior research into other HCoV such as SARS-CoV-1 and *Middle East respiratory syndrome-related coronavirus* (MERS-CoV), as well as other viruses infecting humans such as a variety of influenza species, established a strong foundation that accelerated the pace of SARS-CoV-2 research.

Coronaviruses are large viruses that can be identified by their distinctive “crown-like” shape (Figure 1). Their spherical virions are made from lipid envelopes ranging from 100 to 160 nanometers in which peplomers (protruding structures) of two to three spike (S) glycoproteins are anchored, creating the crown [12,13]. These spikes, which are critical to both viral pathogenesis and to the response by the host immune response, have been visualized using cryo-electron microscopy [14]. Because they induce the human immune response, they are also the target of many proposed therapeutic agents [2,3]. Viral pathogenesis is typically broken down into three major components: entry, replication, and spread [15]. However, in order to draw a more complete picture of pathogenesis, it is also necessary to examine how infection manifests clinically, identify systems-level interactions between the virus and the human body, and consider the possible effects of variation or evolutionary change on pathogenesis and virulence. Thus, clinical medicine and traditional biology are both important pieces of the puzzle of SARS-CoV-2 presentation and pathogenesis.

1.4 Coronavirus Structure and Pathogenesis

1.4.1 Structure of Coronaviruses

Genome structure is highly conserved among coronaviruses, meaning that the relationship between the SARS-CoV-2 genome and its pathogenesis can be inferred from prior research in related viral species. The genomes of viruses in the *Nidovirales* order share several fundamental characteristics. They are non-segmented, which means the viral genome is a single continuous strand of RNA, and are enveloped, which means that the genome and capsid are encased by a lipid bilayer. Coronaviruses have large positive-sense RNA (ssRNA+) genomes ranging from 27 to 32 kilobases in length [16,17]. The SARS-CoV-2 genome lies in the middle of this range at 29,903 bp [17]. Genome organization is highly conserved within the order [16]. There are three major genomic regions: one containing the replicase gene, one containing the genes encoding structural proteins, and interspersed accessory genes [16] (Figure 1). The replicase gene comprises about two-thirds of the genome and consists of two open reading frames that are translated with ribosomal frameshifting [16]. This polypeptide is then translated into 16 non-structural proteins (nsp), except in gammacoronaviruses where nsp1 is absent, that form the replication machinery used to synthesize viral RNA [18]. The remaining third of the genome encodes structural proteins, including the spike (S), membrane, envelope, and nucleocapsid proteins. Additional accessory genes are sometimes present between these two regions, depending on the species or strain. Much attention has been focused on the S protein, which is a critical structure involved in cell entry.

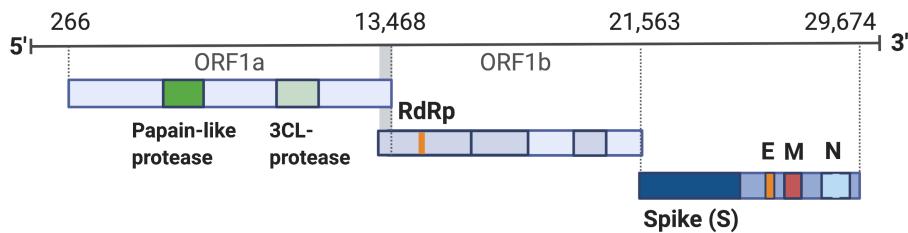
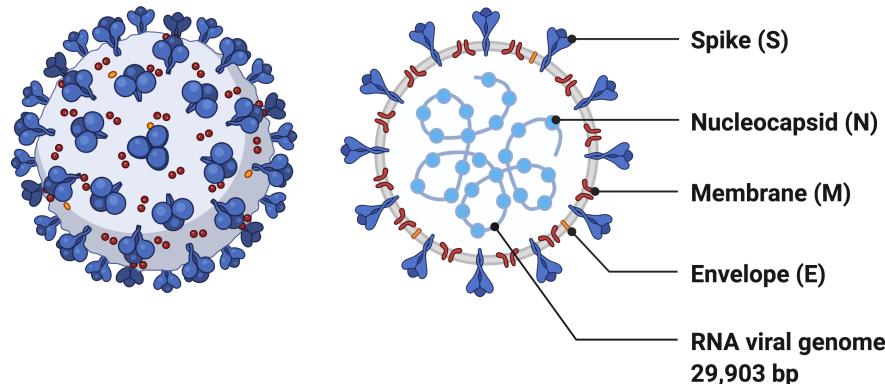
A**B**

Figure 1: Structure of SARS-CoV-2 capsid and genome. A) The genomic structure of coronaviruses is highly conserved and includes three main regions. Open reading frames (ORF) 1a and 1b contain two polyproteins that encode the non-structural proteins (nsp). The nsp include enzymes such as RNA-dependent RNA Polymerase (RdRp). The last third of the genome encodes structural proteins, including the spike (S), envelope (E), membrane (M) and nucleocapsid (N) proteins. Accessory genes can also be interspersed throughout the genome [16]. B) The physical structure of the coronavirus virion, including the components determined by the conserved structural proteins S, E, M and N. This figure was adapted from "Human Coronavirus Structure", by BioRender.com (2020), retrieved from <https://app.biorender.com/biorender-templates>.

1.4.2 Pathogenic Mechanisms of Coronaviruses

While it is possible that SARS-CoV-1 and SARS-CoV-2, like most viruses, enter cells through endocytosis, a process conserved among coronaviruses enables them to target cells for entry through fusion with the plasma membrane [19,20]. Cell entry proceeds in three steps: binding, cleavage, and fusion. First, the viral spike protein binds to a host cell via a recognized receptor or entry point. Coronaviruses can bind to a range of host receptors [21,22], with binding conserved only at the genus level [10]. Viruses in the beta coronavirus genus, to which SARS-CoV-2 belongs, are known to bind to the CEACAM1 protein, 5-N-acetyl-9-O-acetyl neuraminic acid, and to angiotensin-converting enzyme 2 (ACE2) [21]. This recognition is driven by domains in the S1 subunit [23]. SARS-CoV-2 has a high affinity for human ACE2, which is expressed in the vascular epithelium, other epithelial cells, and cardiovascular and renal tissues [24,25], as well as many others [26]. The binding process is guided by the molecular structure of the spike protein, which is structured in three segments: an ectodomain, a transmembrane anchor, and an intracellular tail [27]. The ectodomain forms the crown-like structures on the viral membrane and contains two subdomains known as the S1 and S2 subunits [28]. The S1 (N-terminal) domain forms the head of the crown and contains the receptor binding motif, and the S2 (C-terminal) domain forms the stalk that supports the head [28]. The S1 subunit guides the binding of the virus to the host cell, and the S2 subunit guides the fusion process [27].

After the binding of the S1 subunit to an entry point, the spike protein of coronaviruses is often cleaved at the S1/S2 boundary into the S1 and S2 subunits by a host protease [23,29,30]. This proteolytic priming is important because it prepares the S protein for fusion [29,30]. The two subunits remain bound by van der Waals forces, with the S1 subunit stabilizing the S2 subunit throughout the membrane fusion process [23]. Cleavage at a second site within S2 (S2') activates S for fusion by inducing conformational changes [23]. Similar to SARS-CoV-1, SARS-CoV-2 exhibits redundancy in which host proteases can cleave the S protein [31]. Both transmembrane protease serine protease-2 (TMPRSS-2) and cathepsins B/L have been shown to mediate SARS-CoV-2 S protein proteolytic priming, and small molecule inhibition of these enzymes fully inhibited viral entry *in vitro* [31,32]. Other proteases known to cleave the S1/S2 boundary in coronaviruses include TMPRSS-4, trypsin, furin, cathepsins, and human airway trypsin-like protease (HAT) [32].

Unlike in SARS-CoV-1, a second cleavage site featuring a furin-like binding motif is also present near the S1/S2 boundary in SARS-CoV-2 [33]. This site is found in HCoV belonging to the A and C lineages of beta coronavirus, including MERS-CoV, but not in the other known members of the B lineage of beta coronavirus that contains SARS-CoV-1 and SARS-CoV-2 [33]. It is associated with increased virulence in other viral species [33] and may facilitate membrane fusion of SARS-CoV-2 in the absence of other proteases that prime the S1/S2 site [34]. However, given that proteases such as HAT are likely to be present in targets like the human airway, the extent to which this site has had a real-world effect on the spread of SARS-CoV-2 was initially unclear [34]. Subsequent research has supported this site as an important contributor to pathogenesis: *in vitro* analyses have reported that it bolsters pathogenicity specifically in cell lines derived from human airway cells (Calu3 cell line) [35,36,37] and that furin inhibitors reduced pathogenic effects in VeroE6 cells [38].

Electron microscopy suggests that in some coronaviruses, including SARS-CoV-1 and MERS-CoV, a six-helix bundle separates the two subunits in the postfusion conformation, and the unusual length of this bundle facilitates membrane fusion through the release of additional energy [10]. The viral membrane can then fuse with the endosomal membrane to release the viral genome into the host cytoplasm. Once the virus enters a host cell, the replicase gene is translated and assembled into the viral replicase complex. This complex then synthesizes the double-stranded RNA (dsRNA) genome from the genomic ssRNA(+). The dsRNA genome is transcribed and replicated to create viral mRNAs and new ssRNA(+) genomes [16,39]. From there, the virus can spread into other cells. In SARS-CoV-2, the insertion of the furin-like binding site near the S1/S2 boundary is also thought to increase cell-cell adhesion, making it possible for the viral genome to spread directly from cell to cell rather than needing to propagate the virion itself [40]. In this way, the genome of SARS-CoV-2 provides insight into the pathogenic behavior of the virus.

Evidence also suggests that SARS-CoV-2 may take advantage of the specific structure of endothelial cells to enter the circulatory system. Endothelial cells are specialized epithelial cells [41] that form a barrier between the bloodstream and surrounding tissues. The endothelium facilitates nutrient,

oxygen, and cellular exchange between the blood and vascularized tissues [42]. The luminal (interior) surface of the endothelium is lined with glycocalyx, a network of both membrane-bound and soluble proteins and carbohydrates, primarily proteoglycans and glycoproteins [43,44]. The glycocalyx varies in thickness from 0.5 microns in the capillaries to 4.5 microns in the carotid arteries and forms a meshwork that localizes both endothelial- and plasma-derived signals to the inner vessel wall [43]. Heparan sulfate is the dominant proteoglycan in the glycocalyx, representing 50-90% of glycocalyx proteoglycan content [45]. The SARS-CoV-2 spike protein can bind directly to heparan sulfate, which serves in part as a scaffolding molecule to facilitate ACE2 binding and entry into endothelial cells [44]. A heparan sulfate binding site has also been identified near the ACE2 binding site on the viral receptor binding domain (RBD), and modeling has suggested that heparan sulfate binding yields an open conformation that facilitates binding to ACE2 on the cell surface [44]. Degrading or removing heparan sulfate was associated with decreased binding [44]. Heparan sulfate may also interact with the S1/S2 proteolytic cleavage site and other binding sites to promote binding affinity [46]. Notably, treatment with soluble heparan sulfate or even heparin (a commonly used anti-coagulant and vasodilator that is similar in structure to heparan sulfate [47]) potently blocked spike protein binding and viral infection [44]. This finding is particularly interesting because degradation of heparan sulfate in the glycocalyx has previously been identified as an important contributor to ARDS and sepsis [48], two common and severe outcomes of COVID-19, and suggests that heparan sulfate could be a target for pharmaceutical inhibition of cell entry by SARS-CoV-2 [49,50,51,52,53]. Together, this evidence suggests that heparan sulfate can serve as an important adhesion molecule for SARS-CoV-2 cell entry. It may represent a therapeutic target but has not been pursued as much as other candidate targets [3].

1.4.3 Immune Evasion Strategies

Research in other HCoV provides some indication of how SARS-CoV-2 infection can proceed despite human immune defenses. Infecting the epithelium can help viruses such as SARS-CoV-1 bypass the physical barriers, such as mucus, that comprise the immune system's first line of defense [54]. Once the virus infiltrates host cells, it is adept at evading detection. CD163+ and CD68+ macrophage cells are especially crucial for the establishment of SARS-CoV-1 in the body [54]. These cells most likely serve as viral reservoirs that help shield SARS-CoV-1 from the innate immune response. According to a study on the viral dissemination of SARS-CoV-1 in Chinese macaques, viral RNA could be detected in some monocytes throughout the process of differentiation into dendritic cells [54]. This lack of active viral replication allows SARS-CoV-1 to escape the innate immune response because reduced levels of detectable viral RNA allow the virus to avoid both natural killer cells and Toll-like receptors [54]. Even during replication, SARS-CoV-1 is able to mask its dsRNA genome from detection by the immune system. Although dsRNA is a pathogen-associated molecular pattern that would typically initiate a response from the innate immune system [55], *in vitro* analysis of nidoviruses including SARS-CoV-1 suggests that these viruses can induce the development of double-membrane vesicles that protect the dsRNA signature

from being detected by the host immune system [56]. This protective envelope can therefore insulate these coronaviruses from the innate immune system's detection mechanism [57].

HCoVs are also known to interfere with the host immune response, rather than just evade it. For example, the virulence of SARS-CoV-2 is increased by nsp1, which can suppress host gene expression by stalling mRNA translation and inducing endonucleolytic cleavage and mRNA degradation [58]. SARS-CoV-1 also evades the immune response by interfering with type I IFN induction signaling, which is a mechanism that leads to cellular resistance to viral infections. SARS-CoV-1 employs methods such as ubiquitination and degradation of RNA sensor adaptor molecules MAVS and TRAF3/6 [59]. Also, MERS-CoV downregulates antigen presentation via MHC class I and MHC class II, which leads to a reduction in T cell activation [59]. These evasion mechanisms, in turn, may facilitate systemic infection. Coronaviruses such as SARS-CoV-1 are also able to evade the humoral immune response through other mechanisms, such as inhibiting certain cytokine pathways or down-regulating antigen presentation by the cells [56].

1.4.4 Host Cell Susceptibility

ACE2 and TMPRSS-2 have been identified as the primary entry portal and as a critical protease, respectively, in facilitating the entry of SARS-CoV-1 and SARS-CoV-2 into a target cell [14,31,60,61,62]. This finding has led to a hypothesized role for the expression of these molecules in determining which cells, tissues, and organs are most susceptible to SARS-CoV-2 infection. ACE2 is expressed in numerous organs, such as the heart, kidney, and intestine, but it is most prominently expressed in alveolar epithelial cells; this pattern of expression is expected to contribute to the virus' association with lung pathology [24,63,64] as well as that of SARS [65]. A retrospective observational study reported indirect evidence that certain antineoplastic therapies, such as the chemotherapy drug gemcitabine, may reduce risk of SARS-CoV-2 infection in patients with cancer, possibly via decreased ACE2 expression [66]. Additionally, the addition of the furin site insertion at the S1/S2 boundary means that SARS-CoV-2 does not require TMPRSS-2 when furin, an ubiquitously expressed endoprotease [67], is present, enabling cell-cell fusion independent of TMPRSS-2 availability [68].

Clinical investigations of COVID-19 patients have detected SARS-CoV-2 transcripts in bronchoalveolar lavage fluid (BALF) (93% of specimens), sputum (72%), nasal swabs (63%), fibrobronchoscopy brush biopsies (46%), pharyngeal swabs (32%), feces (29%), and blood (1%) [69]. Two studies reported that SARS-CoV-2 could not be detected in urine specimens [69,70]; however, a third study identified four urine samples (out of 58) that were positive for SARS-CoV-2 nucleic acids [71]. Although respiratory failure remains the leading cause of death for COVID-19 patients [72], SARS-CoV-2 infection can damage many other organ systems including the heart [73], kidneys [74,75], liver [76], and gastrointestinal tract [77,78]. As it becomes clear that SARS-CoV-2 infection can damage multiple organs, the scientific community is pursuing multiple avenues of investigation in order to build a consensus about how the virus affects the human body.

1.5 Clinical Presentation of COVID-19

SARS-CoV-2 pathogenesis is closely linked with the clinical presentation of the COVID-19 disease. Reports have described diverse symptom profiles associated with COVID-19, with a great deal of variability both within and between institutions and regions. Definitions for non-severe, severe, and critical COVID-19, along with treatment recommendations, are available from the World Health Organization living guidelines [79]. A large study from Wuhan, China conducted early in the pandemic identified fever and cough as the two most common symptoms that patients reported at hospital admission [80], while a retrospective study in China described the clinical presentations of patients infected with SARS-CoV-2 as including lower respiratory tract infection with fever, dry cough, and dyspnea (shortness of breath) [81]. This study [81] noted that upper respiratory tract symptoms were less common, suggesting that the virus preferentially targets cells located in the lower respiratory tract. However, data from the New York City region [82,83] showed variable rates of fever as a presenting symptom, suggesting that symptoms may not be consistent across individuals. For example, even within New York City, one study [82] identified low oxygen saturation (<90% without the use of supplemental oxygen or ventilation support) in 20.4% of patients upon presentation, with fever being present in 30.7%, while another study [83] reported cough (79.4%), fever (77.1%), and dyspnea (56.5%) as the most common presenting symptoms; both of these studies considered only hospitalized patients. A later study reported radiographic findings such as ground-glass opacity and bilateral patchy shadowing in the lungs of many hospitalized patients, with most COVID-19 patients having lymphocytopenia, or low levels of lymphocytes (a type of white blood cell) [80]. Patients may also experience loss of smell, myalgias (muscle aches), fatigue, or headache. Gastrointestinal symptoms can also present [84], and the CDC includes nausea and vomiting, as well as congestion and runny nose, on its list of symptoms consistent with COVID-19 [6]. An analysis of an app-based survey of 500,000 individuals in the U.S. found that among those tested for SARS-CoV-2, a loss of taste or smell, fever, and a cough were significant predictors of a positive test result [85]. It is important to note that in this study, the predictive value of symptoms may be underestimated if they are not specific to COVID-19. This underestimation could occur because the outcome measured was a positive, as opposed to a negative, COVID-19 test result, meaning an association would be more easily identified for symptoms that were primarily or exclusively found with COVID-19. At the time the surveys were conducted, due to limits in U.S. testing infrastructure, respondents typically needed to have some symptoms known to be specific to COVID-19 in order to qualify for testing. Widespread testing of asymptomatic individuals may therefore provide additional insight into the range of symptoms associated with COVID-19.

Consistent with the wide range of symptoms observed and the pathogenic mechanisms described above, COVID-19 can affect a variety of systems within the body in addition to causing respiratory problems [86]. For example, COVID-19 can lead to acute kidney injury, especially in patients with severe respiratory symptoms or certain preexisting conditions [87]. Some patients are at risk for collapsing glomerulopathy [88].

COVID-19 can also cause neurological complications [89,90,91], potentially including stroke, seizures or meningitis [92,93]. One study on autopsy samples suggested that SARS-CoV-2 may be able to enter the central nervous system via the neural-mucosal interface [94]. However, a study of 41 autopsied brains [95] found no evidence that the virus can actually infect the central nervous system. Although there was viral RNA in some brain samples, it was only found in very small amounts, and no viral protein was found. The RNA may have been in the blood vessels or blood components and not in the brain tissue itself. Instead, the neuropathological effects of COVID-19 are more likely to be caused indirectly by hypoxia, coagulopathy, or inflammatory processes rather than by infection in the brain [95]. COVID-19 has been associated with an increased incidence of large vessel stroke, particularly in patients under the age of 40 [96], and other thrombotic events including pulmonary embolism and deep vein thrombosis [97]. The mechanism behind these complications has been suggested to be related to coagulopathy, with reports indicating the presence of antiphospholipid antibodies [98] and elevated levels of d-dimer and fibrinogen degradation products in deceased patients [99]. Other viral infections have been associated with coagulation defects and changes to the coagulation cascade; notably, SARS was also found to lead to disseminated intravascular coagulation and was associated with both pulmonary embolism and deep vein thrombosis [100]. The mechanism behind these insults has been suggested to be related to inflammation-induced increases in the von Willebrand factor clotting protein, leading to a pro-coagulative state [100]. Abnormal clotting (thromboinflammation or coagulopathy) has been increasingly discussed recently as a possible key mechanism in many cases of severe COVID-19, and may be associated with the high d-dimer levels often observed in severe cases [101,102,103]. This excessive clotting in lung capillaries has been suggested to be related to a dysregulated activation of the complement system, part of the innate immune system [104,105].

Finally, concerns have been raised about long-term sequelae of COVID-19. Some COVID-19 patients have reported that various somatic symptoms (such as shortness of breath, fatigue, chest pain) and psychological (depression, anxiety or mild cognitive impairment) symptoms can last for months after infection [106]. Such long-term affects occur both in adults [107] and children [108]. Sustained symptoms affecting a variety of biological systems have been reported across many studies (e.g., [106,109,110]). The phenomenon of “long COVID” is not fully understood although various possible explanations have been proposed, including damage caused by immune response to infection as well as by the infection itself, in addition to negative consequences of the experience of lengthy illness and hospitalization. However, a lack of consistency among definitions used in different studies makes it difficult to develop precise definitions or identify specific symptoms associated with long-term effects of COVID-19 [111,112]. Patient and family support groups for “long haulers” have been formed online, and patient-driven efforts to collect data about post-acute COVID-19 provide valuable sources of information (e.g., [109]). The specific relationship between viral pathogenesis and these reported sequelae remains to be uncovered, however.

1.5.1 Pediatric Presentation

The presentation of COVID-19 infection can vary greatly among pediatric patients and, in some cases, manifests in distinct ways from COVID-19 in adults. Evidence suggests that children and adolescents tend to have mostly asymptomatic infections and that those who are symptomatic typically exhibit mild illness [113,114,115,116]. One review examined symptoms reported in 17 studies of children infected with COVID-19 during the early months of the COVID-19 epidemic in China and one study from Singapore [117]. In the more than a thousand cases described, the most common reports were for mild symptoms such as fever, dry cough, fatigue, nasal congestion and/or runny nose, while three children were reported to be asymptomatic. Severe lower respiratory infection was described in only one of the pediatric cases reviewed. Gastrointestinal symptoms such as vomiting or diarrhea were occasionally reported. Radiologic findings were not always reported in the case studies reviewed, but when they were mentioned they included bronchial thickening, ground-glass opacities, and/or inflammatory lesions [117]. Neurological symptoms have also been reported [118].

These analyses indicate that most pediatric cases of COVID-19 are not severe. Indeed, it is estimated that less than 1% of pediatric cases result in critical illness [115,119], although reporting suggests that pediatric hospitalizations may be greater with the emergence of the Delta variant of concern (VOC) [120,121,122]. Serious complications and, in relatively rare cases, deaths have occurred [123]. Of particular interest, children have occasionally experienced a serious inflammatory syndrome, multisystem inflammatory syndrome in children (MIS-C), following COVID-19 infection [124]. This syndrome is similar in some respects to Kawasaki disease, including Kawasaki disease shock syndrome [125,126,127], and is thought to be a distinct clinical manifestation of SARS-CoV-2 due to its distinct cytokine profile and the presence of burr cells in peripheral blood smears [128,129]. MIS-C has been associated with heart failure in some cases [130]. A small number of case studies have identified presentations similar to MIS-C in adults associated with SARS-CoV-2 [131,132,133,134]. However, not all cases of severe COVID-19 in children are characterizable as MIS-C. A recent study [135] described demographic and clinical variables associated with MIS-C in comparison with non-MIS-C severe acute COVID-19 in young people in the United States. Efforts to characterize long-term sequelae of SARS-CoV-2 infection in children face the same challenges as in adults, but long-term effects remain a concern in pediatric patients [108,136,137], although some early studies have suggested that they may be less of a concern than in adults [138,139,140]. Research is ongoing into the differences between the pediatric and adult immune responses to SARS-CoV-2, and future research may shed light on the factors that lead to MIS-C; it is also unknown whether the relative advantages of children against severe COVID-19 will remain in the face of current and future variants [141].

1.5.2 Cytokine Release Syndrome

The inflammatory response was identified early on as a potential driver of COVID-19 outcomes due to existing research in SARS and emerging research in COVID-19. While too low of an inflammatory response is a concern because it will fail to eliminate the immune threat [142], excessive pro-inflammatory cytokine activity can cascade [143] and cause cell damage, among other problems [144]. A dysregulated immune response can cause

significant damage to the host [145,146,147] including pathogenesis associated with sepsis. Sepsis, which can lead to multi-organ failure and death [148,149], is traditionally associated with bacterial infections. However, sepsis associated with viral infections may be underidentified [150], and sepsis has emerged as a major concern associated with SARS-CoV-2 infection [151]. Hyperactivity of the pro-inflammatory response due to lung infection is commonly associated with acute lung injury and more rarely with the more severe manifestation, ARDS, which can arise from pneumonia, SARS, and COVID-19 [143,148]. Damage to the capillary endothelium can cause leaks that disrupt the balance between pro-inflammatory cytokines and their regulators [152], and heightened inflammation in the lungs can also serve as a source for systemic inflammation, or sepsis, and potentially multi-organ failure [148]. The shift from local to systemic inflammation is a phenomenon often referred to broadly as a cytokine storm [148] or, more precisely, as cytokine release syndrome [153].

Cytokine dysregulation is therefore a significant concern in the context of COVID-19. In addition to the known role of cytokines in ARDS and lung infection more broadly, immunohistological analysis at autopsy of deceased SARS patients revealed that ACE2-expressing cells that were infected by SARS-CoV-1 showed elevated expression of the cytokines IL-6, IL-1 β , and TNF- α [154]. Similarly, the introduction of the S protein from SARS-CoV-1 to mouse macrophages was found to increase production of IL-6 and TNF- α [155]. For SARS-CoV-2 infection leading to COVID-19, early reports described a cytokine storm syndrome-like response in patients with particularly severe infections [63,156,157]. Sepsis has been identified as a major contributor to COVID-19-related death. Among patients hospitalized with COVID-19 in Wuhan, China, 112 out of 191 (59%) developed sepsis, including all 54 of the non-survivors [81].

While IL-6 is sometimes used as a biomarker for cytokine storm activity in sepsis [148], the relationship between cytokine profiles and the risks associated with sepsis may be more complex. One study of patients with and at risk for ARDS, specifically those who were intubated for medical ventilation, found that shortly after the onset of ARDS, anti-inflammatory cytokine concentration in BALF increased relative to the concentration of pro-inflammatory cytokines [152]. The results suggest that an increase in pro-inflammatory cytokines such as IL-6 may signal the onset of ARDS, but recovery depends on an increased anti-inflammatory response [152]. However, patients with severe ARDS were excluded from this study. Another analysis of over 1,400 pneumonia patients in the United States reported that IL-6, tumor necrosis factor (TNF), and IL-10 were elevated at intake in patients who developed severe sepsis and/or ultimately died [158]. However, unlike the study analyzing pro- and anti-inflammatory cytokines in ARDS patients [152], this study reported that unbalanced pro-/anti-inflammatory cytokine profiles were rare. This discrepancy could be related to the fact that the sepsis study measured only three cytokines. Although IL-6 has traditionally been considered pro-inflammatory, its pleiotropic effects via both classical and trans-signaling allow it to play an integral role in both the inflammatory and anti-inflammatory responses [159], leading it to be associated with both healthy and pathological responses to viral threat [160]. While the cytokine levels observed in COVID-19 patients fall outside of the normal range, they are not as high as typically found in patients with ARDS [161]. Regardless of

variation in the anti-inflammatory response, prior work has therefore made it clear that pulmonary infection and injury are associated with systemic inflammation and with sepsis. Inflammation has received significant interest both in regards to the pathology of COVID-19 as well as potential avenues for treatment, as the relationship between the cytokine storm and the pathophysiology of COVID-19 has led to the suggestion that a number of immunomodulatory pharmaceutical interventions could hold therapeutic value for the treatment of COVID-19 [3,162].

1.6 Insights from Systems Biology

Systems biology provides a cross-disciplinary analytical paradigm through which the host response to an infection can be analyzed. This field integrates the “omics” fields (genomics, transcriptomics, proteomics, metabolomics, etc.) using bioinformatics and other computational approaches. Over the last decade, systems biology approaches have been used widely to study the pathogenesis of diverse types of life-threatening acute and chronic infectious diseases [163]. Omics-based studies have also provided meaningful information regarding host immune responses and surrogate protein markers in several viral, bacterial and protozoan infections [164]. Though the complex pathogenesis and clinical manifestations of SARS-CoV-2 infection are not yet fully understood, omics technologies offer the opportunity for discovery-driven analysis of biological changes associated with SARS-CoV-2 infection.

1.6.1 Transcriptomics

Through transcriptomic analysis, the effect of a viral infection on gene expression can be assessed. Transcriptomic analyses, whether *in vivo* or *in situ*, can potentially reveal insights into viral pathogenesis by elucidating the host response to the virus. For example, infection by some viruses, including by the coronaviruses SARS-CoV-2, SARS-CoV-1, and MERS-CoV, is associated with the upregulation of ACE2 in human embryonic kidney cells and human airway epithelial cells [63]. This finding suggests that SARS-CoV-2 facilitates the positive regulation of its own transmission between host cells [63]. The host immune response also likely plays a key role in mediating infection-associated pathologies. Therefore, transcriptomics is one critical tool for characterizing the host response in order to gain insight into viral pathogenesis. For this reason, the application of omics technologies to the process of characterizing the host response is expected to provide novel insights into how hosts respond to SARS-CoV-2 infection and how these changes might influence COVID-19 outcomes.

Several studies have examined the cellular response to SARS-CoV-2 *in vitro* in comparison to other viruses. One study [165] compared the transcriptional responses of three human cell lines to SARS-CoV-2 and to other respiratory viruses, including MERS-CoV, SARS-CoV-1, *Human parainfluenza virus 3*, *Respiratory syncytial virus*, and *Influenza A virus*. The transcriptional response differed between the SARS-CoV-1 infected cells and the cells infected by other viruses, with changes in differential expression specific to each infection type. Where SARS-CoV-2 was able to replicate efficiently, differential expression analysis revealed that the transcriptional response

was significantly different from the response to all of the other viruses tested. A unique pro-inflammatory cytokine signature associated with SARS-CoV-2 was present in cells exposed to both high and low doses of the virus, with the cytokines IL-6 and IL1RA uniquely elevated in response to SARS-CoV-2 relative to other viruses. However, one cell line showed significant IFN-I or IFN-III expression when exposed to high, but not low, doses of SARS-CoV-2, suggesting that IFN induction is dependent on the extent of exposure. These results suggest that SARS-CoV-2 induces a limited antiviral state with low IFN-I or IFN-III expression and a moderate IFN-stimulated gene response, in contrast to other viruses. Other respiratory viruses have been found to encode antagonists to the IFN response [166,167], including SARS-CoV-1 [168] and MERS-CoV [169].

The analysis of SARS-CoV-2 suggested that this transcriptional state was specific to cells expressing ACE2, as it was not observed in cells lacking expression of this protein except with ACE2 supplementation and at very high (10-fold increase) level of SARS-CoV-2 exposure [165]. In another study, direct stimulation with inflammatory cytokines such as type I interferons (e.g., IFN β) was also associated with the upregulation of ACE2 in human bronchial epithelial cells, with treated groups showing four-fold higher ACE2 expression than control groups at 18 hours post-treatment [170]. This hypothesis was further supported by studies showing that several nsps in SARS-CoV-2 suppress interferon activity [171] and that the SARS-CoV-2 *ORF3b* gene suppresses IFNB1 promoter activity (IFN-I induction) more efficiently than the SARS-CoV-1 *ORF3b* gene [172]. Taken together, these findings suggest that a unique cytokine profile is associated with the response to the SARS-CoV-2 virus, and that this response differs depending on the magnitude of exposure.

Susceptibility and IFN induction may also vary by cell type. Using poly(A) bulk RNA-seq to analyzed dynamic transcriptional responses to SARS-CoV-2 and SARS-CoV-1 revealed negligible susceptibility of cells from the H1299 line (< 0.08 viral read percentage of total reads) compared to those from the Caco-2 and Calu-3 lines (>10% of viral reads) [173]. This finding suggests that the risk of infection varies among cell types, and that cell type could influence which hosts are more or less susceptible. Based on visual inspection of microscopy images alongside transcriptional profiling, the authors also showed distinct responses among the host cell lines evaluated [173]. In contrast to Caco-2, Calu-3 cells infected with SARS-CoV-2 showed signs of impaired growth and cell death at 24 hours post infection, as well as moderate IFN induction with a strong up-regulation of IFN-stimulated genes. Interestingly, the results were similar to those reported in Calu-3 cells exposed to much higher levels of SARS-CoV-2 [165], as described above. This finding suggests that IFN induction in Calu-3 cells is not dependent on the level of exposure, in contrast to A549-ACE2 cells. The discrepancy could be explained by the observations that Calu-3 cells are highly susceptible to SARS-CoV-2 and show rapid viral replication [32], whereas A549 cells are incompatible with SARS-CoV-2 infection [174]. This discrepancy raises the concern that *in vitro* models may vary in their similarity to the human response, underscoring the importance of follow-up studies in additional models.

As a result, transcriptional analysis of patient tissue is an important application of omics technology to understanding COVID-19. Several studies have collected blood samples from COVID-19 patients and analyzed them using RNA-Seq [175,176,177,178,179,180]. Analyzing gene expression in the blood is valuable to understanding host-pathogen interactions because of the potential to identify alterations associated with the immune response and to gain insights into inflammation, among other potential insights [175]. One study compared gene expression in 39 COVID-19 inpatients admitted with community-acquired pneumonia to that of control donors using whole blood cell transcriptomes [175]. They also evaluated the effect of mild versus severe disease. A greater number of differentially expressed genes were found in severe patients compared to controls than in mild patients compared to controls. They also identified that the transcriptional profiles clustered into five groups and that the groups could not be explained by disease severity. Most severe cases fell into two clusters associated with increased inflammation and granulocyte and neutrophil activation. The presence of these clusters suggests the possibility that personalized medicine could be useful in the treatment of COVID-19 [175]. Longitudinal analysis of granulocytes from patients with mild versus severe COVID-19 revealed that granulocyte activation-associated factors differentiated the disease states, with greater numbers of differentially expressed genes early in disease course [175]. This study therefore revealed distinct patterns associated with COVID-19 and identified genes and pathways associated with each cluster.

Many other studies have also identified transcriptomic signatures associated with the immune response and inflammation. Other studies have profiled the transcriptome of BALF [177] and the nasopharynx [181]. One study used single-cell transcriptomics techniques to investigate cell types including brain and choroid plexus cells compared to healthy controls and controls with influenza; among other signals of neuroinflammation, this study reported cortical T cells only in COVID-19 patients [182]. Transcriptomic analysis can thus provide insight into the pathogenesis of SARS-CoV-2 and may also be useful in identifying candidate therapeutics [175].

1.6.2 Proteomics

Proteomics analysis offers an opportunity to characterize the response to a pathogen at a level above transcriptomics. Especially early on, this primarily involved evaluating the effect of the virus on cell lines. One early proteomics study investigated changes associated with *in vitro* SARS-CoV-2 infection using Caco-2 cells [183]. This study reported that SARS-CoV-2 induced alterations in multiple vital physiological pathways, including translation, splicing, carbon metabolism and nucleic acid metabolism in the host cells. Another area of interest is whether SARS-CoV-2 is likely to induce similar changes to other HCoV. For example, because of the high level of sequence homology between SARS-CoV-2 and SARS-CoV-1, it has been hypothesized that sera from convalescent SARS-CoV-1 patients might show some efficacy in cross-neutralizing SARS-CoV-2-S-driven entry [31]. However, despite the high level of sequence homology, certain protein structures might be immunologically distinct, which would be likely to prohibit effective cross-

neutralization across different SARS species [184]. Consequently, proteomic analyses of SARS-CoV-1 might also provide some essential information regarding the new pathogen [185,186].

Proteomics research has been able to get ahead of the timeline for development of omics-level big data sets specific to SARS-CoV-2 by adopting a comparative bioinformatics approach. Data hubs such as UniProt [187], NCBI Genome Database [188], The Immune Epitope Database and Analysis Resource [189], and The Virus Pathogen Resource [190] contain a wealth of data from studies in other viruses and even HCoV. Such databases facilitate the systems-level reconstruction of protein-protein interaction networks, providing opportunities to generate hypotheses about the mechanism of action of SARS-CoV-2 and identify potential drug targets. In an initial study [191], 26 of the 29 SARS-CoV-2 proteins were cloned and expressed in HEK293T kidney cells, allowing for the identification of 332 high-confidence human proteins interacting with them. Notably, this study suggested that SARS-CoV-2 interacts with innate immunity pathways. Ranking pathogens by the similarity between their interactomes and that of SARS-CoV-2 suggested *West Nile virus*, *Mycobacterium tuberculosis*, and *human papillomavirus* infections as the top three hits. The fact that the host-pathogen interactome of the bacterium *Mycobacterium tuberculosis* was found to be similar to that of SARS-CoV-2 suggests that changes related to lung pathology might comprise a significant contributor to these expression profiles. Additionally, it was suggested that the envelope protein, E, could disrupt host bromodomain-containing proteins, i.e., BRD2 and BRD4, that bind to histones, and the spike protein could likely intervene in viral fusion by modulating the GOLGA7-ZDHHC5 acyl-transferase complex to increase palmitoylation, which is a post-translational modification that affects how proteins interact with membranes [192].

An example of an application of this *in silico* approach comes from another study [193], which used patient-derived peripheral blood mononuclear cells to identify 251 host proteins targeted by SARS-CoV-2. This study also reported that more than 200 host proteins were disrupted following infection. In particular, a network analysis showed that nsp9 and nsp10 interacted with NF-Kappa-B-Repressing Factor, which encodes a transcriptional repressor that mediates repression of genes responsive to Nuclear Factor kappa-light-chain-enhancer of activated B-cells. These genes are important to pro-, and potentially also anti-, inflammatory signaling [194]. This finding could explain the exacerbation of the immune response that shapes the pathology and the high cytokine levels characteristic of COVID-19, possibly due to the chemotaxis of neutrophils mediated by IL-8 and IL-6. Finally, it was suggested [195] that the E protein of both SARS-CoV-1 and SARS-CoV-2 has a conserved Bcl-2 Homology 3-like motif, which could inhibit anti-apoptosis proteins, e.g., BCL2, and trigger the apoptosis of T cells. Several compounds are known to disrupt the host-pathogen protein interactome, largely through the inhibition of host proteins. Therefore, this research identifies candidate targets for intervention and suggests that drugs modulating protein-level interactions between virus and host could be relevant to treating COVID-19.

As with other approaches, analyzing the patterns found in infected versus healthy human subjects is also important. COVID-19 infection has been associated with quantitative changes in transcripts, proteins, metabolites, and lipids in patient blood samples [196]. One longitudinal study [197] compared COVID-19 patients to symptomatic controls who were PCR-negative for SARS-CoV-2. The longitudinal nature of this study allowed it to account for differences in the scale of inter- versus intraindividual changes. At the time of first sampling, common functions of proteins upregulated in COVID-19 patients relative to controls were related to immune system mediation, coagulation, lipid homeostasis, and protease inhibition. They compared these data to the patient-specific timepoints associated with the highest levels of SARS-CoV-2 antibodies and found that the actin-binding protein gelsolin, which is involved in recovery from disease, showed the steepest decline between those two time points. Immunoglobulins comprised the only proteins that were significantly different between the COVID-19 and control patients at both of these timepoints. The most significantly downregulated proteins between these time points were related to inflammation, while the most significantly upregulated proteins were immunoglobulins. Proteins related to coagulation also increased between the two timepoints. The selection of a symptomatic control cohort rather than healthy comparisons also suggests that the results are more likely to highlight the response to SARS-CoV-2 and COVID-19 specifically, rather than to disease more broadly. This study also compared the disease course in patients who ultimately survived to those who died and found that ITIH4, a protein associated with the inflammatory response to trauma, may be a biomarker useful to identifying patients at risk of death. Thus, these results indicate the value of studying patients in a longitudinal manner over the disease course. By revealing which genes are perturbed during SARS-CoV-2 infection, proteomics-based analyses can thus provide novel insights into host-virus interaction and serve to generate new avenues of investigation for therapeutics.

1.7 Viral Virulence

Like that of SARS-CoV-1, the entry of SARS-CoV-2 into host cells is mediated by interactions between the viral spike glycoprotein, S, and human ACE2 (hACE2) [23,31,198,199,200,201,202,203]. Differences in how the S proteins of the two viruses interact with hACE2 could partially account for the increased transmissibility of SARS-CoV-2. Studies have reported conflicting binding constants for the S-hACE2 interaction, though they have agreed that the SARS-CoV-2 S protein binds with equal, if not greater, affinity than the SARS-CoV-1 S protein does [14,23,201]. The C-terminal domain of the SARS-CoV-2 S protein in particular was identified as the key region of the virus that interacts with hACE2, and the crystal structure of the C-terminal domain of the SARS-CoV-2 S protein in complex with hACE2 reveals stronger interaction and a higher affinity for receptor binding than that of SARS-CoV-1 [202]. Among the 14 key binding residues identified in the SARS-CoV-1 S protein, eight are conserved in SARS-CoV-2, and the remaining six are semi-conservatively substituted, potentially explaining variation in binding affinity [23,201]. Studies of crystal structure have shown that the RBD of the SARS-CoV-2 S protein, like that of other coronaviruses, undergoes stochastic hinge-like movement that flips it from a “closed” conformation, in which key binding residues are hidden at the interface between protomers, to an “open” one

[14,23]. Spike proteins cleaved at the furin-like binding site are substantially more likely to take an open conformation (66%) than those that are uncleaved (17%) [204]. Because the RBD plays such a critical role in viral entry, blocking its interaction with ACE2 could represent a promising therapeutic approach. Nevertheless, despite the high structural homology between the SARS-CoV-2 RBD and that of SARS-CoV-1, monoclonal antibodies targeting SARS-CoV-1 RBD failed to bind to SARS-CoV-2-RBD [14]. However, in early research, sera from convalescent SARS patients were found to inhibit SARS-CoV-2 viral entry *in vitro*, albeit with lower efficiency than it inhibited SARS-CoV-1 [31].

Comparative genomic analysis reveals that several regions of the coronavirus genome are likely critical to virulence. The S1 domain of the spike protein, which contains the receptor binding motif, evolves more rapidly than the S2 domain [21,22]. However, even within the S1 domain, some regions are more conserved than others, with the receptors in S1's N-terminal domain (S1-NTD) evolving more rapidly than those in its C-terminal domain (S1-CTD) [22]. Both S1-NTD and S1-CTD are involved in receptor binding and can function as RBDs to bind proteins and sugars [21], but RBDs in the S1-NTD typically bind to sugars, while those in the S1-CTD recognize protein receptors [10]. Viral receptors show higher affinity with protein receptors than sugar receptors [10], which suggests that positive selection on or relaxed conservation of the S1-NTD might reduce the risk of a deleterious mutation that would prevent binding. The SARS-CoV-2 S protein also contains an RRAR furin recognition site at the S1/S2 junction [14,23], setting it apart from both bat coronavirus RaTG13, with which it shares 96% genome sequence identity, and SARS-CoV-1 [205]. Such furin cleavage sites are commonly found in highly virulent influenza viruses [206,207]. The furin recognition site at the S1/S2 junction is likely to increase pathogenicity via destabilization of the spike protein during fusion to ACE2 and the facilitation of cell-cell adhesion [14,23,40,204,206,207]. These factors may influence the virulence of SARS-CoV-2 relative to other beta coronaviruses. Additionally, a major concern has been the emergence of SARS-CoV-2 variants with increased virulence. The extent to which evolution within SARS-CoV-2 may affect pathogenesis is reviewed below.

1.8 Molecular Signatures, Transmission, and Variants of Concern

Genetic variation in SARS-CoV-2 has been used to elucidate patterns over time and space. Many mutations are neutral in their effect and can be used to trace transmission patterns. Such signatures within SARS-CoV-2 have provided insights during outbreak investigations [208,209,210]. Similar mutations observed in several patients may indicate that the patients belong to the same transmission group. The tracking of SARS-CoV-2 mutations is recognized as an essential tool for controlling future outbreaks and tracing the path of the spread of SARS-CoV-2. In the first months of the pandemic in early 2020, early genomic surveillance efforts in Guangdong, China revealed that local transmission rates were low and that most cases arising in the province were imported [211]. Since then, efforts have varied widely among countries: for example, the U.K. has coordinated a national database of viral genomes [212], but efforts to collect this type of data in the United States

have been more limited [213]. Studies have applied phylogenetic analyses of viral genomes to determine the source of local COVID-19 outbreaks in Connecticut (USA), [214], the New York City area (USA) [215], and Iceland [216]. There has been an ongoing effort to collect SARS-CoV-2 genomes throughout the COVID-19 outbreak, and as of summer 2021, millions of genome sequences have been collected from patients. The sequencing data can be found at GISAID [217], NCBI [218], and COVID-19 data portal [219].

Ongoing evolution can be observed in genomic data collected through molecular surveillance efforts. In some cases, mutations can produce functional changes that can impact pathogenesis. One early example is the spike protein mutation D614G, which appeared in March 2020 and became dominant worldwide by the end of May 2020 [220,221]. This variant was associated with increased infectivity and increased viral load, but not with more severe disease outcomes [220,222]. This increased virulence is likely achieved by altering the conformation of the S1 domain to facilitate binding to ACE2 [222]. Similarly, the N439K mutation within the RBD of the spike protein is likely associated with increased transmissibility and enhanced binding affinity for hACE2, although it is also not thought to affect disease outcomes [223]. In contrast, a mutation in ORF8 that was identified in Singapore in the early months of 2020 was associated with cases of COVID-19 that were less likely to require treatment with supplemental oxygen [224], and a deletion surrounding the furin site insertion at the S1/S2 boundary has been identified only rarely in clinical settings [225], suggesting that these mutations may disadvantage viral pathogenesis in human hosts. Thus, mutations have been associated with both virological and clinical differences in pathogenesis.

Several VOCs have also been identified and designated through molecular surveillance efforts [226]. The Alpha variant (lineage B.1.1.7) was first observed in the U.K. in October 2020 before it quickly spread around the world [227]. Other variants meriting further investigation have also been identified, including the Beta variant (B.1.351 lineage) first identified in South Africa and the Gamma variant (P.1 lineage) initially associated with outbreaks in Brazil. These lineages share independently acquired mutations that may affect pathogenicity [228,229,230,231,232]. For example, they are all associated with a greater binding affinity for hACE2 than that of the wildtype variant [230,233,234], but they were not found to have more efficient cell entry than the wildtype virus [235]. A fourth VOC, the Delta variant (B.1.617.2 and AY.1, AY.2, and AY.3 lineages), was identified in India in late 2020 [236]. Some of the mutations associated with this lineage may alter fusogenicity and enhance furin cleavage, among other effects associated with increased pathogenicity [237]. The changes in these VOC demonstrate how ongoing evolution in SARS-CoV-2 can drive changes in how the virus interacts with host cells.

1.9 Quantifying Viral Presence

Assessing whether a virus is present in a sample is a more complex task than it initially seems. Many diagnostic tests rely on real-time polymerase chain reaction (RT-PCR) to test for the presence versus absence of a virus [5]. They may report the cycle threshold (C_t) indicating the number of doubling cycles

required for the target (in this case, SARS-CoV-2) to become detectable. A lower C_t therefore corresponds to a higher viral load. The C_t that corresponds to a positive can vary widely, but is often around 35. This information is sufficient to answer many questions, since an amplicon must be present in order to be duplicated in RT-PCR. For example, if a patient is presenting with COVID-19 symptoms, a positive RT-PCR test can confirm the diagnosis.

However, RT-PCR analysis alone cannot provide the information needed to determine whether a virus is present at sufficient levels to be infectious [238]. Some studies have therefore taken the additional step of cultivating samples *in vitro* in order to observe whether cells become infected with SARS-CoV-2. One study collected upper respiratory tract samples from COVID-19 patients, analyzed them with RT-PCR to determine the cycle threshold, and then attempted to cultivate the SARS-CoV-2 virus in VeroE6 cells [238]. This study found that out of 246 samples, less than half (103) produced a positive culture. Moreover, at a C_t of 35, only 5 out of 60 samples grew *in vitro*. Therefore, the RT-PCR-confirmed presence of SARS-CoV-2 in a sample does not necessarily indicate that the virus is present at a high enough concentration to grow and/or spread.

1.10 Mechanisms of Transmission

When a human host is infected with a virus and is contagious, person-to-person viral transmission can occur through several possible mechanisms. When a contagious individual sneezes, coughs, or exhales, they produce respiratory droplets that can contain a large number of viral particles [239]. Viral particles can enter the body of a new host when they then come in contact with the oral, nasal, eye, or other mucus membranes [239]. The primary terms typically used to discuss the transmission of viruses via respiratory droplets are droplet, aerosol, and contact transmission [240]. The distinction between droplet and aerosol transmission is typically anchored on whether a particle containing the virus is larger or smaller than 5 micrometers (μm) [241,242]. Droplet transmission typically refers to contact with large droplets that fall quickly to the ground at close range, such as breathing in droplets produced by a sneeze [239,241]. Aerosol transmission typically refers to much smaller particles (less than 5 μm) produced by sneezing, coughing, or exhaling [239,240] that can remain suspended over a longer period of time and potentially to be moved by air currents [239]. It is also possible that viral particles deposited on surfaces via large respiratory droplets could later be aerosolized [239]. The transmission of viral particles that have settled on a surface is typically referred to as contact or fomite transmission [239,243]. Any respiratory droplets that settle on a surface could contribute to fomite transmission [239]. Droplet and contact transmission are both well-accepted modes of transmission for many viruses associated with common human illnesses, including influenza and rhinovirus [239].

The extent to which aerosol transmission contributes to the spread of respiratory viruses is more widely debated. In influenza A, for example, viral particles can be detected in aerosols produced by infected individuals, but it is not clear to what extent these particles drive the spread of influenza A

infection [239,240,244,245,246]. Regardless of its role in the spread of influenza A, however, aerosol transmission likely played a role in outbreaks such as the 1918 Spanish Influenza (H1N1) and 2009 “swine flu” (pH1N1) [246]. All three of these mechanisms have been identified as possible contributors to the transmission of HCoVs [239], including the highly pathogenic coronaviruses SARS-CoV-1 and MERS-CoV [247,248].

Transmission of SARS-CoV-1 is thought to proceed primarily through droplet transmission, but aerosol transmission is also considered possible [239,249,250], and fomite transmission may have also played an important role in some outbreaks [251]. Similarly, the primary mechanism of MERS transmission is thought to be droplets because inter-individual transmission appears to be associated with close interpersonal contact (e.g., household or healthcare settings), but aerosolized particles of the MERS virus have been reported to persist much more robustly than influenza A under a range of environmental conditions [252,253]. However, few of these analyses have sought to grow positive samples in culture and thus to confirm their potential to infect new hosts.

Contact, droplet, and aerosol transmission are therefore all worth evaluating when considering possible modes of transmission for a respiratory virus like SARS-CoV-2. The stability of the SARS-CoV-2 virus both in aerosols and on a variety of surfaces was found to be similar to that of SARS-CoV-1 [254].

Droplet-based and contact transmission were initially put forward as the greatest concern for the spread of SARS-CoV-2 [255], with droplet transmission considered the dominant mechanism driving the spread of the virus [256] because the risk of fomite transmission under real-world conditions is likely to be substantially lower than the conditions used for experimental analyses [257]. The COVID-19 pandemic has, however, exposed significant discrepancies in how terms pertaining to airborne viral particles are interpreted in different contexts [241]. The 5-μm distinction between “droplets” and “aerosols” is typical in the biological literature but is likely an artifact of historical science rather than a meaningful boundary in biology or physics [242]. Additionally, various ambient conditions such as air flow can influence how particles of different sizes fall or spread [241]. Despite initial skepticism about airborne transmission of SARS-CoV-2 through small particles [242], evidence now suggests that small particles can contribute to SARS-CoV-2 transmission [254,258,259,260]. For example, one early study detected SARS-CoV-2 viral particles in air samples taken from hospitals treating COVID-19 patients, although the infectivity of these samples was not assessed [261]. Subsequently, other studies have been successful in growing SARS-CoV-2 in culture with samples taken from the air [262,263] while others have not [264,265] (see [266] for a systematic review of available findings as of July 2020). The fact that viable SARS-CoV-2 may exist in aerosolized particles calls into question whether some axioms of COVID-19 prevention, such as 2-meter social distancing, are sufficient [242,262,267].

1.10.1 Symptoms and Viral Spread

Other aspects of pathogenesis are also important to understanding how the virus spreads, especially the relationship between symptoms, viral shedding, and contagiousness. Symptoms associated with reported cases of COVID-19 range from mild to severe [6], but some individuals who contract COVID-19 remain asymptomatic throughout the duration of the illness [268]. The

incubation period, or the time period between exposure and the onset of symptoms, has been estimated at five to eight days, with means of 4.91 (95% confidence interval (CI) 4.35-5.69) and 7.54 (95% CI 6.76-8.56) reported in two different Asian cities and a median of 5 (IQR 1 to 6) reported in a small number of patients in a Beijing hospital [269,270].

However, the exact relationship between contagiousness and viral shedding remains unclear. Estimates suggest that viral shedding can, in some cases, begin as early as 12.3 days (95% CI 5.9-17.0) before the onset of symptoms, although this was found to be very rare, with less than 0.1% of transmission events occurring 7 or more days before symptom onset [271].

Transmissibility appeared to peak around the onset of symptoms (95% CI -0.9 - 0.9 days), and only 44% (95% CI 30-57%) of transmission events were estimated to occur from presymptomatic contacts [271]. A peak in viral load corresponding to the onset of symptoms was also confirmed by another study [238]. As these trends became apparent, concerns arose due to the potential for individuals who did not yet show symptoms to transmit the virus [272]. Recovered individuals may also be able to transmit the virus after their symptoms cease. A study of the communicable period based on twenty-four individuals who tested positive for SARS-CoV-2 prior to or without developing symptoms estimated that individuals may be contagious for one to twenty-one days, but they note that this estimate may be low [268]. In an early study, viral nucleic acids were reported to remain at observable levels in the respiratory specimens of recovering hospitalized COVID-19 patients for a median of 20 days and with a maximum observed duration through 37 days, when data collection for the study ceased [81].

As more estimates of the duration of viral shedding were released, they converged around approximately three weeks from first positive PCR test and/or onset of symptoms (which, if present, are usually identified within three days of the initial PCR test). For example, in some studies, viral shedding was reported for up to 28 days following symptom onset [273] and for one to 24 days from first positive PCR test, with a median of 12 days [70]. On the other hand, almost 70% of patients were reported to still have symptoms at the time that viral shedding ceased, although all symptoms reduced in prevalence between onset and cessation of viral shedding [274]. The median time that elapsed between the onset of symptoms and cessation of viral RNA shedding was 23 days and between first positive PCR test and cessation of viral shedding was 17 days [274]. The fact that this study reported symptom onset to predate the first positive PCR test by an average of three days, however, suggests that there may be some methodological differences between it and related studies. Furthermore, an analysis of residents of a nursing home with a known SARS-CoV-2 case measured similar viral load in residents who were asymptomatic regardless of whether they later developed symptoms, and the load in the asymptomatic residents was comparable to that of residents who displayed either typical or atypical symptoms [275]. Taken together, these results suggest that the presence or absence of symptoms are not reliable predictors of viral shedding or of SARS-CoV-2 status (e.g, [276]). However, it should be noted that viral shedding is not necessarily a robust indicator of contagiousness. The risk of spreading the infection was low after ten days from the onset of symptoms, as viral

load in sputum was found to be unlikely to pose a significant risk based on efforts to culture samples *in vitro* [273]. The relationship between symptoms, detectable levels of the virus, and risk of viral spread is therefore complex.

The extent to which asymptomatic or presymptomatic individuals are able to transmit SARS-CoV-2 has been a question of high scientific and community interest. Early reports (February and March 2020) described transmission from presymptomatic SARS-CoV-2-positive individuals to close family contacts [277,278]. One of these reports [278] also included a description of an individual who tested positive for SARS-CoV-2 but never developed symptoms. Later analyses also sought to estimate the proportion of infections that could be traced back to a presymptomatic or asymptomatic individual (e.g., [279]). Estimates of the proportion of individuals with asymptomatic infections have varied widely. The proportion of asymptomatic individuals on board the Diamond Princess cruise ship, which was the site of an early COVID-19 outbreak, was estimated at 17.9% [280]. In contrast, a model using the prevalence of antibodies among residents of Wuhan, China estimated a much higher rate of asymptomatic cases, at approximately 7 in 8, or 87.5% [281]. An analysis of the populations of care homes in London found that, among the residents (median age 85), the rate of asymptomatic infection was 43.8%, and among the caretakers (median age 47), the rate was 49.1% [282]. The duration of viral shedding may also be longer in individuals with asymptomatic cases of COVID-19 compared to those who do show symptoms [283]. As a result, the potential for individuals who do not know they have COVID-19 to spread the virus raises significant concerns. In Singapore and Tianjin, two cities studied to estimate incubation period, an estimated 40-50% and 60-80% of cases, respectively, were considered to be caused by contact with asymptomatic individuals [269]. An analysis of viral spread in the Italian town of Vo', which was the site of an early COVID-19 outbreak, revealed that 42.5% of cases were asymptomatic and that the rate was similar across age groups [284]. The argument was thus made that the town's lockdown was imperative for controlling the spread of COVID-19 because it isolated asymptomatic individuals. While more models are likely to emerge to better explore the effect of asymptomatic individuals on SARS-CoV-2 transmission, these results suggest that strategies for identifying and containing asymptomatic but contagious individuals are important for managing community spread.

1.10.2 Estimating the Fatality Rate

Estimating the occurrence of asymptomatic and mild COVID-19 cases is important to identifying the mortality rate associated with COVID-19. The mortality rate of greatest interest would be the total number of fatalities as a fraction of the total number of people infected. One commonly reported metric is the case fatality rate (CFR), which compares the number of COVID-19 related deaths to the number of confirmed or suspected cases. However, in locations without universal testing protocols, it is impossible to identify all infected individuals because so many asymptomatic or mild cases go undetected. Therefore, a more informative metric is the infection fatality rate (IFR), which compares the known deaths to the estimated number of cases. It thus requires the same numerator as CFR, but divides by an approximation of the total number of cases rather than only the observed/suspected cases. IFR varies regionally, with some locations observed to have IFRs as low as

0.17% while others are as high as 1.7% [285]. Estimates of CFR at the national and continental level and IFR at the continent level is maintained by the Centre for Evidence-Based Medicine [286]. Several meta-analyses have also sought to estimate IFR at the global scale. These estimates have varied; one peer-reviewed study aggregated data from 24 other studies and estimated IFR at 0.68% (95% CI 0.53%–0.82%), but a preprint that aggregated data from 139 countries calculated a global IFR of 1.04% (95% CI 0.77%–1.38%) when false negatives were considered in the model [285,287]. A similar prevalence estimate was identified through a repeated cross-sectional serosurvey conducted in New York City that estimated the IFR as 0.97% [288].

Examination of serosurvey-based estimates of IFR identified convergence on a global IFR estimate of 0.60% (95% CI 0.42%–0.77%) [285]. All of these studies note that IFR varies widely by location, and it is also expected to vary with demographic and health-related variables such as age, sex, prevalence of comorbidities, and access to healthcare and testing [289]. Estimates of infection rates are becoming more feasible as more data becomes available for modeling and will be bolstered as serological testing becomes more common and more widely available. However, this research may be complicated due to the emergence of variants over time, as well as the varying availability and acceptance of vaccines in different communities and locations.

1.11 Dynamics of Transmission

Disease spread dynamics can be estimated using R_0 , the basic reproduction number, and R_t , the effective reproduction number. Accurate estimates of both are crucial to understanding the dynamics of infection and to predicting the effects of different interventions. R_0 is the average number of new (secondary) infections caused by one infected person, assuming a wholly susceptible population [290], and is one of the most important epidemiological parameters [291]. A simple mechanistic model used to describe infectious disease dynamics is a susceptible-infected-recovered compartmental model [292,293]. In this model, individuals move through three states: susceptible, infected, and recovered; two parameters, γ and β , specify the rate at which the infectious recover, and the infection transmission rate, respectively, and R_0 is estimated as the ratio of β and γ [291,294]. A pathogen can invade a susceptible population only if $R_0 > 1$ [291,295]. The spread of an infectious disease at a particular time t can be quantified by R_t , the effective reproduction number, which assumes that part of the population has already recovered (and thus gained immunity to reinfection) or that mitigating interventions have been put into place. For example, if only a fraction S_t of the population is still susceptible, $R_t = S_t \times R_0$. When R_t is greater than 1, an epidemic grows (i.e., the proportion of the population that is infectious increases); when R_t is less than 1, the proportion of the population that is infectious decreases. R_0 and R_t can be estimated directly from epidemiological data or inferred using susceptible-infected-recovered-type models. To capture the dynamics of SARS-CoV-2 accurately, the addition of a fourth compartment, i.e. a susceptible-exposed-infectious-recovered model, may be appropriate because such models account for the relative lengths of incubation and infectious periods [296].

Original estimates of R_0 for COVID-19 lie in the range $R_0=1.4\text{-}6.5$ [297,298,299]. Variation in R_0 is expected between different populations, and the estimated values of R_0 discussed below are for specific populations in specific environments. The different estimates of R_0 should not necessarily be interpreted as a range of estimates of the same underlying parameter. In one study of international cases, the predicted value was $R_0=1.7$ [300]. In China (both Hubei province and nationwide), the value was predicted to lie in the range $R_0=2.0\text{-}3.6$ [297,301,302]. Another estimate based on a cruise ship where an outbreak occurred predicted $R_0=2.28$ [303]. Susceptible-exposed-infectious-recovered model-derived estimates of R_0 range from 2.0 - 6.5 in China [304,305,306,307] to $R_0=4.8$ in France [308]. Using the same model as for the French population, a study estimated $R_0=2.6$ in South Korea [308], which is consistent with other studies [309]. From a meta-analysis of studies estimating R_0 , [298] the median R_0 was estimated to be 2.79 (IQR 1.16) based on twelve studies published between January 1 and February 7, 2020.

Inference of the effective reproduction number can provide insight into how populations respond to an infection and the effectiveness of interventions. In China, R_t was predicted to lie in the range 1.6-2.6 in January 2020, before travel restrictions [310]. R_t decreased from 2.35 one week before travel restrictions were imposed (January 23, 2020), to 1.05 one week after. Using their model, the authors also estimated the probability of new outbreaks occurring. Assuming individual-level variation in transmission comparable to that of MERS or SARS, the probability of a single individual exporting the virus and causing a large outbreak is 17-25%, and assuming variation like that of SARS and transmission patterns like those observed for COVID-19 in Wuhan, the probability of a large outbreak occurring after ≥ 4 infections exist at a new location is greater than 50%. An independent study came to similar conclusions, finding $R_t=2.38$ in the two-week period before January 23 with a decrease to $R_t = 1.34$ (using data from January 24 to February 3) or $R_t=0.98$ (using data from January 24 to February 8) [299]. In South Korea, R_t was inferred for February through March 2020 in two cities, Daegu (the center of the outbreak) and Seoul [309]. Metro data was also analyzed to estimate the effects of social distancing measures. R_t decreased in Daegu from around 3 to <1 over the period that social distancing measures were introduced. In Seoul, R_t decreased slightly, but remained close to 1 (and larger than R_t in Daegu). These findings indicate that social distancing measures appeared to be effective in containing the infection in Daegu, but in Seoul, R_t remained above 1, meaning secondary outbreaks remained possible. The study also shows the importance of region-specific analysis: the large decline in case load nationwide was mainly due to the Daegu region and could mask persistence of the epidemic in other regions, such as Seoul and Gyeonggi-do. In Iran, estimates of R_t declined from 4.86 in the first week to 2.1 by the fourth week after the first cases were reported [311]. In Europe, analysis of 11 countries inferred the dynamics of R_t over a time range from the beginning of the outbreak until March 28, 2020, by which point most countries had implemented major interventions (such as stay-at-home orders, public gathering bans, and school closures) [312]. Across all countries, the mean R_t before interventions began was estimated as 3.87; R_t varied considerably, from below 3 in Norway to above 4.5 in Spain. After interventions, R_t decreased by an average of 64% across all countries, with mean $R_t=1.43$. The lowest predicted value was 0.97 for Norway and the highest was 2.64 for Sweden, which could be related to the fact that Sweden

did not implement social distancing measures on the same scale as other countries. The study concludes that while large changes in R_t are observed, it is too early to tell whether the interventions put into place are sufficient to decrease R_t below 1.

Evolution within SARS-CoV-2 has also driven changes in the estimated reproduction number for different populations at different times. As of June 2021, the reproduction number had increased globally relative to 2020, and increased transmissibility over the wildtype variant was observed for the Alpha, Beta, Gamma, and Delta VOC [313]. In the U.S. between December 2020 and January 2021, B.1.1.7 (Alpha) was estimated to have an increased transmission of 35 to 45% relative to common SARS-CoV-2 variants at the time, with B.1.1.7 the dominant SARS-CoV-2 variant in some places at some timepoints [314]. This lineage was estimated to have increased transmissibility of 43 to 90% in the U.K. [315]. An estimate of the reproduction number of B.1.1.7 in the U.K. from September to December 2020 yielded 1.59 overall and between 1.56 and 1.95 in different regions of the country [232]. The Delta variant is particularly transmissible, and it has been estimated to be twice as transmissible than the wildtype variant of SARS-CoV-2 [313]. A review of the literature describing the Delta variant identified a mean estimated R_0 of 5.08 [316]. Such differences can affect fitness and therefore influence the relative contributions of different lineages to a given viral gene pool over time [317]. Therefore, the evolution of the virus can result in shifts in the reproduction rate.

More generally, population-level epidemic dynamics can be both observed and modeled [294]. Data and empirically determined biological mechanisms inform models, while models can be used to try to understand data and systems of interest or to make predictions about possible future dynamics, such as the estimation of capacity needs [318] or the comparison of predicted outcomes among prevention and control strategies [319,320]. Many current efforts to model R_t have also led to tools that assist the visualization of estimates in real time or over recent intervals [321,322]. These are valuable resources, yet it is also important to note that the estimates arise from models containing many assumptions and are dependent on the quality of the data they use, which varies widely by region.

1.12 Conclusions

The novel coronavirus SARS-CoV-2 is the third HCoV to emerge in the 21st century, and research into previous HCoVs has provided a strong foundation for characterizing the pathogenesis and transmission of SARS-CoV-2. Critical insights into how the virus interacts with human cells have been gained from previous research into HCoVs and other viral infections. With the emergence of three devastating HCoV over the past twenty years, emergent viruses are likely to represent an ongoing threat. Contextualizing SARS-CoV-2 alongside other viruses serves not only to provide insights that can be immediately useful for combating this virus itself but may also prove valuable in the face of future viral threats.

Host-pathogen interactions provide a basis not only for understanding COVID-19, but also for developing a response. As with other HCoVs, the immune response to SARS-CoV-2 is likely driven by detection of its spike protein, which allows it to enter cells through ACE2. Epithelial cells have also emerged as the major cellular target of the virus, contextualizing the respiratory and gastrointestinal symptoms that are frequently observed in COVID-19. Many of the mechanisms that facilitate the pathogenesis of SARS-CoV-2 are currently under consideration as possible targets for the treatment or prevention of COVID-19 [2,3]. Research in other viruses also provides a foundation for understanding the transmission of SARS-CoV-2 among people and can therefore inform efforts to control the virus's spread. Airborne forms of transmission (droplet and aerosol transmission) have emerged as the primary modes by which the virus spreads to new hosts. Asymptomatic transmission was also a concern in the SARS outbreak of 2002-03 and, as in the current pandemic, presented challenges for estimating rates of infection [323]. These insights are important for developing a public health response, such as the CDC's shift in its recommendations surrounding masking [324].

Even with the background obtained from research in SARS and MERS, COVID-19 has revealed itself to be a complex and difficult-to-characterize disease that has many possible presentations that vary with age. Variability in presentation, including cases with no respiratory symptoms or with no symptoms altogether, were also reported during the SARS epidemic at the beginning of the 21st century [323]. The variability of both which symptoms present and their severity have presented challenges for public health agencies seeking to provide clear recommendations regarding which symptoms indicate SARS-CoV-2 infection and should prompt isolation. Asymptomatic cases add complexity both to efforts to estimate statistics such as R_0 and R_t , which are critical to understanding the transmission of the virus, and IFR, which is an important component of understanding its impact on a given population. The development of diagnostic technologies over the course of the pandemic has facilitated more accurate identification, including of asymptomatic cases [5]. As more cases have been diagnosed, the health conditions and patient characteristics associated with more severe infection have also become more clear, although there are likely to be significant sociocultural elements that also influence these outcomes [325]. While many efforts have focused on adults, and especially older adults because of the susceptibility of this demographic, additional research is needed to understand the presentation of COVID-19 and MIS-C in pediatric patients. As more information is uncovered about the pathogenesis of HCoV and SARS-CoV-2 specifically, the diverse symptomatology of COVID-19 has and likely will continue to conform with the ever-broadening understanding of how SARS-CoV-2 functions within a human host.

While the SARS-CoV-2 virus is very similar to other HCoV in several ways, including in its genomic structure and the structure of the virus itself, there are also some differences that may account for differences in the COVID-19 pandemic compared to the SARS and MERS epidemics of the past two decades. The R_0 of SARS-CoV-2 has been estimated to be similar to SARS-CoV-1 but much higher than that of MERS-CoV [326], although a higher R_0 has been estimated for some VOC. While the structures of the viruses are very similar, evolution among these species may account for differences in their transmissibility and virulence. For example, the acquisition of a furin

cleavage site the S1/S2 boundary within the SARS-CoV-2 S protein may be associated with increased virulence. Additionally, concerns have been raised about the accumulation of mutations within the SARS-CoV-2 species itself, and whether these could influence virulence [327]. These novel variants may be resistant to vaccines and antibody treatments such as Bamlanivimab that were designed based on the wildtype spike protein [3,328]. As a consequence of reliance on targeting the SARS-CoV-2 spike protein for many therapeutic and prophylactic strategies, increased surveillance is required to rapidly identify and prevent the spread of novel SARS-CoV-2 variants with alterations to the spike protein. The coming of age of genomic technologies has made these types of analyses feasible, and genomics research characterizing changes in SARS-CoV-2 along with temporal and spatial movement is likely to provide additional insights into whether within-species evolution influences the effect of the virus on the human host. Additionally, the rapid development of sequencing technologies over the past decade has made it possible to rapidly characterize the host response to the virus. For example, proteomics analysis of patient-derived cells revealed candidate genes whose regulation is altered by SARS-CoV-2 infection, suggesting possible approaches for pharmaceutical invention and providing insight into which systems are likely to be disrupted in COVID-19 [193]. As more patient data becomes available, the biotechnological advances of the 2000s are expected to allow for more rapid identification of potential drug targets than was feasible during the SARS, or even MERS, pandemic.

Thus, the COVID-19 crisis continues to evolve, but the insights acquired over the past 20 years of HCoV research have provided a solid foundation for understanding the SARS-CoV-2 virus and the disease it causes. As the scientific community continues to respond to COVID-19 and to elucidate more of the relationships between pathogenesis, transmission, host regulatory responses, and symptomatology, this understanding will no doubt continue to evolve and to reveal additional connections among virology, pathogenesis, and health. This review represents a collaboration between scientists from diverse backgrounds to contextualize this virus at the union of many different biological disciplines [4]. At present, understanding the SARS-CoV-2 virus and its pathogenesis is critical to a holistic understanding of the COVID-19 pandemic. In the future, interdisciplinary work on SARS-CoV-2 and COVID-19 may guide a response to a new viral threat.

2 Evolutionary Perspectives on SARS-CoV-2

2.1 Abstract

2.2 Importance

2.3 Introduction

The emergence of what is now known to be the pathogen SARS-CoV-2 has dramatically reshaped modern life for the past two years. The genomic revolution provided the tools needed to understand the virus in ways that were not feasible during previous pandemics. For example, the first genome sequence of the pathogen was released on January 3, 2020, providing valuable information about the pathogen within a month and a half of the first known cases. As the pandemic has unfolded, evolutionary questions and methods of investigation have framed the scientific approach to understanding the virus. These questions have evolved along with the pandemic. Thus far, five major evolutionary questions have emerged. The first was “what is it?”, the second “where did it come from?”, the third and fourth address “whom does it affect?”, the fifth “how is it changing?” and the sixth “what is next?” Evolutionary biology provides a framework through which these questions can be evaluated and explored.

2.4 Question 1: What Is It?

What is now known as SARS-CoV-2 emerged in November 2019 as an unknown pathogen causing a cluster of pneumonia cases in Wuhan, China. The initial genome sequence, which was released in early January 2020, revealed the pathogen to be a novel coronavirus [9]. Although most coronaviruses show little transmission in humans, several human coronaviruses (HCoV) have been identified since the 1960s. Therefore, in the early days of the pandemic, many strategies to understand or manage the emergent viral threat focused on contextualizing it amongst better-studied coronaviruses.

Many people have previously been infected by an HCoV. Approximately one-third of common cold infections are thought to be caused by four seasonal HCoV: *Human coronavirus 229E* (HCoV-229E), *Human coronavirus NL63* (HCoV-NL63), *Human coronavirus OC43* (HCoV-OC43), and *Human coronavirus HKU1* (HCoV-HKU1) [329,330,331]. The first HCoV were identified in the 1960s: HCoV-229E in 1965 [332] and HCoV-OC43 in 1967 [333]. Both of these viruses typically cause cold-like symptoms, including upper and lower respiratory infections [334,335,336], but they have also been associated with gastrointestinal symptoms [337]. Two additional HCoV were subsequently identified [338,339]. In 2003, HCoV-NL63 [338] was first identified in a 7-month-old infant and then in clinical specimens collected from seven additional patients, five of whom were infants younger than 1 year old and the remainder of whom were adults. CoV-HKU1 was identified in samples collected from a 71-year-old pneumonia patient in 2004 and then found in samples collected from a second adult patient [339]. These viruses are associated with respiratory diseases of varying severity, ranging from common cold to severe pneumonia, with severe symptoms mostly observed in immunocompromised individuals [340], and also have gastrointestinal involvement in some cases [337].

In addition to these relatively mild HCoV, however, highly pathogenic human coronaviruses have been identified, including *Severe acute respiratory syndrome-related coronavirus* (SARS-CoV or SARS-CoV-1) and *Middle East respiratory syndrome-related coronavirus* (MERS-CoV) [248,329,341]. At the time that SARS-CoV-1 emerged in the early 2000s, no HCoV had been

identified in almost 40 years [248]. The first case of SARS was reported in November 2002 in the Guangdong Province of China, and over the following month, the disease spread more widely within China and then into several countries across multiple continents [248,326]. Unlike previously identified HCoV, SARS was much more severe, with an estimated death rate of 9.5% [326]. It was also highly contagious via droplet transmission, with a basic reproduction number (R_0) of 4 (i.e., each person infected was estimated to infect four other people) [326].

However, the identity of the virus behind the infection remained unknown until April of 2003, when the SARS-CoV-1 virus was identified through a worldwide scientific effort spearheaded by the WHO [248]. SARS-CoV-1 belonged to a distinct lineage from the two other HCoV known at the time [326]. By July 2003, the SARS outbreak was officially determined to be under control, with the success credited to infection management practices [248]. A decade later, a second outbreak of severe respiratory illness associated with a coronavirus emerged, this time in the Arabian Peninsula. This disease, known as Middle East respiratory syndrome (MERS), was linked to another novel coronavirus, MERS-CoV. The fatality rate associated with MERS is much higher than that of SARS, at almost 35%, but the disease is much less easily transmitted, with an R_0 of 1 [326]. Although MERS is still circulating, its low reproduction number has allowed for its spread to be contained [326]. The COVID-19 pandemic is thus associated with the seventh HCoV to be identified and the fifth since the turn of the millennium, though additional HCoVs may be in circulation but remain undetected (e.g., [342]).

Following the release of the SARS-CoV-2 genome sequence, multiple research groups sequenced the genomes of SARS-CoV-2 specimens identified in clinical samples. These samples were primarily collected from patients' lower respiratory tract, namely bronchoalveolar lavage fluid (BALF), and the upper respiratory tract, in the form of throat and nasopharyngeal swabs [17,205,343]. Integration of these sequences allowed for a more complete picture of the viral genome. Analysis of the viral genome revealed significant sequence homology with two known HCoV: the novel coronavirus shared about 79% sequence identity with SARS-CoV-1 and 50% with MERS-CoV [17]. Therefore, this early phylogenetic analysis of the novel coronavirus allow its similarity to other, known viruses to be established. SARS-CoV-1 and MERS-CoV were ultimately managed largely through infection management practices (e.g., mask wearing) and properties of the virus itself (i.e., low rate of transmission), respectively [248,326]. Research in response to prior outbreaks of HCoV-borne infections, such as SARS and MERS, provided a strong foundation for hypotheses about the pathogenesis of SARS-CoV-2 as well as potential diagnostic and therapeutic approaches, as we review elsewhere [1,3,328]. Therefore, this phylogenetic information was valuable for gaining an understanding of the pathogen and identifying strategies to manage it.

2.5 Question 2: Where Did It Come From?

Despite the high degree of similarity to SARS-CoV-1, even greater sequence identity was observed between SARS-CoV-2 and zoonotic coronaviruses. A 2001 literature review estimated that 61% of human pathogens have a

zoonotic origin [344]. A zoonotic disease, or zoonosis, arises when a pathogen can both a) infect and b) cause a disease in humans [345]. As a result, the risk of zoonotic disease increases when there is substantial interaction between humans and wildlife [345]. Many factors can influence this human/wildlife interface and therefore the risk of zoonotic transmission events [345,346].

In the SARS epidemic, SARS-CoV-1 was also thought to have emerged in a live animal market. A survey of a market in Shenzhen, China revealed that individuals from two carnivore species, namely several masked palm civets (*Paguma larvata*) and one raccoon dog (*Nyctereutes procyonoides*), were likely carriers of SARS-CoV-1, despite presenting as healthy [347]. However, further analysis suggested that these species might be only intermediate hosts who were exposed in the market setting [348]. A closely related virus was identified in Chinese horseshoe bats (*Rhinolophus sinicus*), but the sequence identity was only 88% with SARS-CoV-1 [349]. Therefore, the species of origin for SARS-CoV-1 remains unresolved.

In the case of SARS-CoV-2, early interest for the emergence of the pathogen turned to live-animal markets in Wuhan [350,351-add-to-Wuhan-riddle], where it would later emerge that many animals were sold suffering from poor health and hygiene [352]. A large percentage of early patients had visited the Huanan seafood market in Wuhan, and next-generation sequencing of samples collected from nine patients, eight of whom had visited the market, revealed extremely high sequence identity (99.98%), indicative of rapid spread [17]. The sequence of the viral pathogen collected from these patients was also compared to known zoonotic pathogens. In particular, genomic research quickly highlighted significant similarity (about 88% sequence identity) between SARS-CoV-2 and bat-derived SARS-like coronaviruses, namely bat-SL-CoVZC45 and bat-SL-CoVZXC21 [17]. Other analyses have reported even greater similarity between SARS-CoV-2 and the bat coronavirus BatCoV-RaTG13, with shared sequence identity as high as 96.2% [205,209]. Bats are well-established as a disease reservoir, including for RNA viruses [353,354,355]. This evidence therefore suggested that the virus may have emerged as a result of zoonotic transfer of a virus from bats to humans, with the wildlife trade considered a potential source of exposure.

Nevertheless, some fragments of the genome differ between SARS-CoV-2 and RATG13 by up to 17%, suggesting a complex natural selection process. Additionally, SARS-CoV-2 is closely related (91.02%) to a novel coronaviruses identified in Malayan pangolins (*Manis javanica*) infected with a respiratory disease in October 2019 [356]. Although the genome-wide sequence identity was lower between SARS-CoV-2 and this pangolin virus than BatCoV-RaTG13, its particularly high similarity in the receptor binding domain (RBD) of the spike (*S*) gene with SARS-CoV-2 drew further attention [356,357]. The SARS-CoV-2 RBD differs from the pangolin coronavirus RBD by only one amino acid change [356], and the sequence identity between the regions is 97.4% [357]. Pangolins were therefore identified as a potential intermediate host of SARS-CoV-2 between bats and humans.

However, data collected from May 2017 to November 2019 by a research team interested in tick-borne illnesses identified no bats or pangolins sold at these markets leading up to the emergence of COVID-19 [352]. Additionally,

endemic bat species are typically in hibernation at the time that SARS-CoV-2 emerged [17]. Therefore, it is possible that animals associated with these markets were infected by bats, but it is not clear whether the disease emerged in a different location and/or whether it is associated with a different species. There were 38 species observed at the market in the 2.5 years leading up to the emergence of SARS-CoV-2, indicating significant diversity in the animals with which humans were interacting [352]. As with SARS-CoV-1, the species of origin for SARS-CoV-2 therefore remains unresolved.

Genomic analyses and comparisons to other known coronaviruses suggest that SARS-CoV-2 is unlikely to have originated in a laboratory – either purposely engineered and released, or escaped – and instead evolved naturally in an animal host [358]. However, potentially due to public misunderstanding about recombination and complex evolutionary processes like coevolution, the similarity to pangolin *S* has resulted in popular conspiracy theories that the virus did not arise naturally. The similarity of *S* to that of pangolin viruses could arise from either recombination or coevolution [209,359], rather than requiring human intervention. Such suspicions may also have been fueled, in part, by the lack of well-characterized bat coronaviruses which means that SARS-CoV-2 is still relatively derived from documented coronaviruses surveyed in bats [360]. While it has been suggested that more thorough investigation of the origins of COVID-19 may have some value [361], in many cases, support for the “lab-leak” theory is politically motivated [362]. A more robust panel of zoonotic viruses against which to compare SARS-CoV-2 would allow for conclusive dismissal of these politicized claims, underscoring another potential benefit of more thorough monitoring of zoonotic diseases. More importantly, it would allow researchers to have a better understanding of and to community concerns about potential emerging viral threats.

2.6 Question 3: Which Species Are Susceptible?

Given the strong evidence for a zoonotic origin of SARS-CoV-2, another evolutionary question that received significant attention, especially early on, was whether humans could infect other species with SARS-CoV-2. In the modern age, opportunities for human-to-animal transmission events could arise in interactions with companion animals, zoo animals, house pests, hunting, urbanized wildlife, and livestock. Outbreaks of zoonotic diseases have been known to originate in environments such as zoos, farms, and petting zoos [363], indicating that disease transmission is likely to be possible in these contexts. Additionally, many coronaviruses infect animals and have been the subject of veterinary medical investigations and vaccine development efforts due to their effect on the health of companion and agricultural animals [364]. Concerns about anthropozoonotic (human-to-animal) transmission focused on a few issues. First, if animal species were susceptible to COVID-19-like infection, in addition to concerns about animal health, infections in livestock could have significant effects on food supply chains. Additionally, even if pathology in these species was limited, if they could serve as viral reservoirs, then they would pose additional risk to humans. The breadth of species susceptible to infection by a pathogen is

known as the pathogen's host range [365]. Understanding the host-pathogen relationship throughout SARS-CoV-2's host range can therefore offer valuable information for managing the spread of SARS-CoV-2.

The phylogeny of the species implicated in the origination of COVID-19 suggested that the host range of SARS-CoV-2 could encompass many species with a high level of interaction with humans. Humans last shared an ancestor with bats and pangolins almost 100 million years ago [366]. Bats belong to the order Chiroptera and pangolins to Pholidota, which both belong to the clade *Pegasoferae* [367,368,369]. They are closely related to many other species that have close relationships with humans, namely odd-toed ungulates (Euungulata) and carnivores (Carnivora) [367,368,369,370]. The part of the evolutionary tree that includes both humans and the *Pegasoferae* encompasses many species of significant social and economic importance. Therefore, concerns were raised that the species with which humans have close interactions, many of which are much more closely related to bats and pangolins than humans are, could also be infected. It seemed plausible that the host range could include both livestock, many of which are odd-toed ungulates, and companion animals, many of which are carnivores. Infection of these animals was identified as a major concern [371].

Genomic analyses seeking to identify which species were likely to be susceptible focused largely on the comparative genetics of angiotensin-converting enzyme 2 (ACE2). ACE2 is the primary protein used by SARS-CoV-2 to enter the cell (see [1]). Recognition of this protein is largely determined by domains in the S1 subunit of the RBD [23]. Alignment of the *ACE2* sequence from 19 species revealed high conservation among mammals [372]. This analysis suggested that non-human primates (three monkey and two ape species), companion animals (dogs and cats), and livestock (both odd- and even-toed ungulates) may all be susceptible to SARS-CoV-2 [372]. Similarly, another study conducted an *in silico* analysis of ACE2 protein structures and their predicted binding to SARS-CoV-2 for 410 vertebrate species [373]. The species identified as having the highest predicted binding affinities were all primates, including humans. Other taxa with high predicted affinities included other primates, rodents, even-toed ungulates (namely, several species of cetaceans and deer), and anteaters. Reindeer were the only domesticated species predicted to belong to either of these groups, but many common zoo animal species with threatened or worse IUCN risk status were identified as at risk.

Considering the evidence generated by *in silico* studies, it may not be surprising that many cases of reverse zoonotic, or anthroponotic, SARS-CoV-2 transmission have been reported. Ferrets (*Mustela furo*) as well as cats and dogs were reported to be susceptible to SARS-CoV-2 in an experimental infection study [374]. The earliest reported anthroponotic transmission events were observed in house pets, primarily cats (*Felis catus*) [375,376,377]. Similarly, cases of SARS-CoV-2 infection have been reported in dogs (*Canis familiaris*): two of fifteen dogs monitored for SARS-CoV-2 by the Hong Kong Agriculture, Fisheries, and Conservation Department during the owners' quarantine in March 2020 were found to be positive for SARS-CoV-2 [376]. Comparing estimates in studies where cats (*Felis catus*) living with SARS-CoV-2-positive humans were tested for SARS-CoV-2 suggest that 6 to 15% of house cats may become infected [375], and a large-scale study of pet

dogs and cats in Italy suggested that 4.5% of cats and 12.8% of dogs from known COVID-19-positive households had developed antibodies to the virus [378]. Some of these SARS-CoV-2-positive domestic carnivores have also shown clinical symptoms [379], and a pilot study of seven cats and three dogs found that cats, but not dogs, shed SARS-CoV-2 virus for several days after viral challenge, although none of the animals were symptomatic [380]. A few dogs and cats have reportedly died after becoming infected with SARS-CoV-2, although in most cases whether the virus is causally related to the death is unclear [381,382,383,384/?sh=4b653381275e,385,386].

Domestic pests, on the other hand, seem to be less susceptible to SARS-CoV-2. In the comparative genomic analysis of ACE2, the two rodent species analyzed, despite being the most phylogenetically similar to humans aside from the other primates, showed the most sequence divergence in ACE2 [372]. This finding was supported by experimental evidence that SARS-CoV-2 cannot use mouse (*Mus musculus*) ACE2 for cell entry [205]. In fact, research using murine models to study SARS and COVID-19 therefore uses transgenic mice designed to be sensitive to the virus (as summarized in [387]).

Similarly, SARS-CoV-2 in livestock also raised concern because of the potential effect on food supply. However, studies using *in vivo* viral challenge reported that livestock species in general do not develop clinical manifestations of SARS-CoV-2 and do not shed infectious virus [374,388]. *In vitro* exposure to SARS-CoV-2 suggested that sheep (*Ovis aries*), but not cattle (*Bos taurus*), might be susceptible to infection, but *in vivo* viral challenge suggested that sheep did not show notable susceptibility to infection [389]. Similarly, analyses of antibody response [390] suggested that sheep exposed to a high level of human interaction did not appear to have developed infections. Following viral challenge of several species, including cattle, sheep, and horses (*Equus ferus caballus*), none were found to shed culturable levels of virus [388]. As a note, despite the low risk posed by livestock themselves, the working conditions of the meat industry itself were associated with a very high risk of SARS-CoV-2 infection for workers that did cause disruptions to food supply chains [391,392].

However, one species of domesticated agricultural animal severely affected by SARS-CoV-2 was the mink (*Neovison vison*). While fur farming has declined significantly since the twentieth century, mink farming is still common in China and some European countries, and mink farms continue to exist in the United States. Mink belong to the Mustelidae family within Carnivora. SARS-CoV-2 was first reported on mink farms in the Netherlands and Denmark in 2020 [393,394]. Mink were observed to show symptoms of respiratory infection, with varied severity among individuals [393]. Dissection revealed lung pathology consistent with interstitial pneumonia [393]. An analysis of five farms in the United States reported mortality rates between 35 and 55% of adult minks [395]. Subsequently, mink farms worldwide reported outbreaks of SARS-CoV-2. Concerns were amplified when novel variants of SARS-CoV-2 were identified as having emerged on Danish mink farms and spread into the human population [394,394,396,397,398]. The fact that these variants appeared in mink populations before being observed in humans suggests that mink can indeed serve as a viral reservoir [394]. Concerns about mink-to-human transmission led to the mass destruction of domesticated mink populations in Europe [399,400]. Introgression from fur

farms into wild populations (i.e., feralization) may have also resulted in the spread of SARS-CoV-2 into wild mink populations [401,402]. Therefore, while the specific zoonotic origin of SARS-CoV-2 may still not be clear, the potential for the virus to take hold in species other than humans has been clearly demonstrated by the mink outbreak.

Finally, some species of zoo animals were also monitored to determine whether they were at risk. Several species closely related to humans (i.e., the Great Apes) are threatened with extinction and had been identified through *in silico* studies as likely to be susceptible to SARS-CoV-2 [373], and therefore the potential for a virus to target these close relatives presented a major concern. In early 2021, three gorillas (*Gorilla beringei beringei*) at the San Diego Zoo Safari Park developed respiratory symptoms that were confirmed to be associated with SARS-CoV-2 [403]. Gorillas at other zoos have also been infected [404,405,406]. Additionally, given the susceptibility of house cats, it is not so surprising that other felids are also susceptible to SARS-CoV-2. Infections of several “big cats” including Malayan tigers (*Panthera tigris jacksoni*), Amur tigers (*Panthera tigris altaica*), and African lions (*Panthera leo krugeri*) were reported at New York City’s Bronx Zoo in March 2020 [407]. In late 2020, four lions (*Panthera leo bleyenberghi*) at the Barcelona Zoo also developed respiratory symptoms that were found to be caused by SARS-CoV-2 [398]. Several captive snow leopards (*Panthera uncia*) in the United States have died from COVID-19 [408,409].

While discussions of zoonoses often focus on the risk that animal diseases carry for human populations, the COVID-19 pandemic has also underscored the risks that human diseases pose for animals. COVID-19 precautions may have reduced the spread of other respiratory illnesses to wild mountain gorilla (*Gorilla beringei beringei*) populations [410], reducing one of the most significant threats to this endangered species [411]. In the case of gorillas, the potential for cross-species application of pharmaceutical advances has also become clear: captive gorillas with COVID-19 received monoclonal antibodies [412]. Additionally, several companies are developing veterinary vaccines against SARS-CoV-2. The most visible has been Zoetis, a veterinary pharmaceutical company, that has developed vaccines that have been administered to several species, including felids in zoos, minks, and gorillas [413,414,415,416]. Russian researchers have also developed a COVID-19 vaccine for carnivores [417].

Therefore, the host range of SARS-CoV-2 is broad, including primates, bats, and carnivores. The most severe infections have been observed in humans, felids, and mustelids [413]. In the United States, as of late 2021, dogs and cats made up the majority of non-human SARS-CoV-2 infections [418], but the most severe infections were observed in felids and mustelids (in addition to humans) [413]. Interestingly, comparing ACE2 binding activity across species [419,420] revealed that it did not always align with which species known to be susceptible to SARS-CoV-2 infection, suggesting other binding sites might also be important. While the specific zoonotic origins of SARS-CoV-2 remain unknown, pharmaceutical developments in the treatment of COVID-19 have included non-human species. The complex relationship between animals, humans, and disease highlights the importance of a broad perspective on health that extends beyond a single species.

2.7 Question 4: Do Genes Influence Who is Affected?

Throughout the pandemic, many hypotheses have been raised about factors that might influence individuals' susceptibility to COVID-19 or to severe disease. Many risk factors, such as underlying health conditions, are related to the body's inflammatory response, as we review elsewhere [325]. Here, we focus narrowly on genetic bases of differences in susceptibility or outcomes. Historically, the identification of genetic risk factors for a disease typically utilized a candidate gene approach, where a gene of interest was evaluated to identify variants that showed an association with the outcome of interest. While economical in terms of sequencing, this approach is prone to spurious results when applied to complex traits [421]. Today, in the age of next-generation sequencing (NGS), alternative approaches have emerged. NGS makes it possible to conduct genome-wide scans where a large number of single-nucleotide polymorphisms (SNPs) or variants are evaluated to identify regions of the genome associated with variation in a phenotype. Genome-wide association studies (GWAS) in particular are a popular approach that employs this strategy. During COVID-19, both of these paradigms have been applied to the problem of identifying genetic correlates of disease severity.

2.7.1 Candidate-Gene Approaches

Many candidate genes have been investigated throughout the pandemic. Here, we review three examples of candidate gene studies in COVID-19. First, an early study (published in April 2020) investigated a known variant in interferon-induced transmembrane protein 3 (*IFITM3*) among hospitalized patients in Beijing [422]. This gene and variant were selected because of a prior candidate gene study by some of the same authors that found an association with influenza severity among Chinese patients during the 2009 influenza A H1N1/09 pandemic [423]. Here, they evaluated a small number (n=80) hospitalized COVID-19 patients to determine whether homozygosity for the previously identified risk allele was associated with mild versus severe disease [422]. They stated that they found an association between homozygosity for the SNP of interest and the severity of COVID-19. A follow-up study demonstrated worldwide variation in the frequency of these SNPs [424], and subsequent studies claimed to support this result by comparing the frequency of the SNP in different groups to the COVID-19 case fatality rate in those groups; they examined SNPs in several candidate genes and identified an association with another SNP in *IFITM3* [425]. However, in the original study, the population-level frequency of the risk allele was consistent with its frequency in the mild population [423]. A similar analysis examined both SNPs in Britons of different ancestral backgrounds and also reported a correlation [426]. While this gene has been investigated for functions potentially relevant to COVID-19 pathogenesis by other groups as well (e.g., [427,428]), a follow-up analysis in Germany evaluated the effect of in 239 cases and 252 controls and reported non-significant effects [426]. The narrative surrounding *IFITM3* therefore reflects a broad methodological critique about candidate gene studies, where results often fail to replicate [429]. The region associated with this gene was not identified in the large-scale GWAS conducted by the COVID-19 Host Genetics Initiative (COVID-19 HGI) [430], which is described in more detail below.

A second source of genetic variability that was hypothesized to have an effect on COVID-19 outcomes were human leukocyte antigens (HLA), or the major histocompatibility complex (MHC). Both MHC classes I and II play a critical role in both the innate and adaptive immune system because they are a pivotal component of antigen presentation. HLA classes I and II are also the most polymorphic loci in the human genome [431]. Additionally, because HLA polymorphisms are associated with geographic ancestry, study location and participant background offers important context [432]. Given the important role of the HLA complex in the immune response and the standing variation in the human population, HLA variation has been investigated for potential associations with COVID-19 outcomes.

Several approaches have been taken to evaluate a potential role of HLA in COVID-19. *In silico* analysis suggested one particular HLA locus that could affect binding of SARS-CoV-2 peptides to MHC class II [433]. Other studies evaluated outcomes using retrospective cohort analyses. An analysis of 95 South Asian COVID-19 patients found that HLA genotype was not significant in differentiating case severity when the necessary statistical corrections were applied [434]. Another study in a European population ($n = 147$) did identify HLA alleles associated with severity [435]. In St. Louis, MO (USA), another study enrolled 234 COVID-19 cases, who were genotyped for HLA alleles and compared to a control population of 20,000 individuals from the National Marrow Donor Program [436]. They compared cases and controls on the basis of four “race/ethnic” populations and reported alleles showing a statistical association within each group [436]. However, because of this stratification, two of the demographic categories had less than ten cases. Across all of these studies, there was minimal overlap in the risk alleles identified, and the small sample sizes raise concerns about the possibility for spurious hits. The hypervariability of this region means that statistical power will necessarily be reduced, with much higher recruitment needed than for studies of biallelic loci. A much larger analysis of 72,912 Israelis, 8.8% of whom tested positive for COVID-19, found no association between HLA genotype and infection or hospitalization [437]. Therefore, while MHC is functionally important to the immune response to COVID-19, it is not clear whether HLA genotypes are predictive of COVID-19 severity, and certainly such studies face exacerbated versions of the typical challenges of candidate gene studies. Because of the challenges associated with analyzing such a variable region, it was excluded from the large-scale COVID-19 HGI GWAS analysis [430].

Finally, significant attention has been paid to the question of whether ABO blood type is associated with COVID-19 outcomes. ABO blood type has been found to modulate susceptibility to other pathogens [438]. While ABO blood type is a genetic trait, it is more easily evaluated than the genetic regions discussed above because of the simple relationship between genetic variants and phenotype. The possibility for an association between blood type and COVID-19 infection was raised early in the pandemic in a preprint that reported associations in 2,173 patients in Wuhan and Shenzhen, China [439]. The protective effect of O and increased risk associated with A blood types that they reported was subsequently investigated by many studies that returned varied results (e.g., [440,441,442,443]; see [444] for a literature review). Observations of higher and lower risk, respectively, of SARS-CoV-2 infection with A and O blood types was supported by a meta-analysis [445].

While the support for the association was independent of a mechanism, a possible relationship between ACE activity and blood type has been proposed [446] as has an effect on carbohydrate-carbohydrate interactions relevant to ACE2 binding [447]. This is the only candidate gene described that has received additional support from GWAS, as is discussed below.

The COVID-19 literature related to candidate gene investigations demonstrates relatively low inter-study consistency in findings. In particular, sample size is a major challenge in designing these studies. However, for many traits, the relationships between genes and phenotypes are complex, and selecting which variants to sequence is not always straightforward. As a result, in the age of next-generation sequencing, discovery-driven studies have emerged as an alternative approach.

2.7.2 Genome-Wide Association Studies

Genome-wide association studies (GWAS) offers a discovery-driven approach that provides a different perspective than candidate gene studies. Instead of selecting a gene or variant *a priori*, in GWAS, a large number of SNPs (usually several million) are evaluated at once to identify those most likely to vary in correlation with a trait of interest. Because of the large number of statistical tests, statistical power and multiple hypothesis testing are both very important considerations in executing GWAS, which have also struggled with issues related to replicability [448]. In cases such as COVID-19 where outcomes can differ among ancestry groups (likely for non-genetic reasons, as reviewed in [325]), it is especially important that GWAS samples be selected with attention paid to ancestry, as incorrect or misleading associations can otherwise be identified with neutral markers indicative of ancestry itself [449].

Over the past two years, many GWAS have been undertaken with the aim of identifying variants associated with COVID-19 outcomes. In some cases, the results have been consistent with hypothesized genetic correlates of susceptibility to COVID-19. One study conducted a GWAS on a total of 435 COVID-19 patients from four countries and identified another HLA allele to be associated with an increased risk of intubation [450]. Other GWAS have identified an association with the ABO blood group locus. One conducted a case/control GWAS in two populations, Italians and Spaniards, with 1980 cases and 2205 controls. They reported two loci that met the genome-wide significance threshold, one on chromosome 3 and one on chromosome 9 [451]. The hit on chromosome 9 fell on the ABO locus and the alleles identified suggested a protective association with blood group O and a risk association with blood group A [451].

As the pandemic has progressed, large-scale efforts have been assembled to conduct GWAS on massive scales. In March 2020, COVID-19 HGI was established as a world-wide consortium that combines data to conduct meta-analyses [452]. One year later, COVID-19 HGI released a meta-analysis of data from 46 studies, comprising 49,562 cases and 1,770,206 controls [430]. They identified 13 loci, seven of which were significant at the genome-wide level when considering all data available, that were associated with one or more phenotypes related to COVID-19 infection or severity. Notably, strong signals were identified for both of the loci suggested by previous medium-

scale GWAS in association with COVID-19 infection [451]. Additionally, several other loci could be mapped onto hypotheses about genetic contributors to immune function, lung function and disease. This world-wide GWAS study made an effort towards strategic incorporation of genetic information from different ancestral groups. Interestingly, the risk variant on chromosome 3 is likely to be inherited from Neanderthal introgression, meaning it is likely to be more prevalent in certain populations, especially non-African populations [453,454]. The potential functional relationship between this region of the genome and COVID-19 is unknown, but phenome-wide association study has suggested blood cell traits as a potential trait regulated by this region [455].

Identifying genetic variants associated with a complex disease is always complicated. In COVID-19 studies, the results of candidate gene analyses have in general been difficult to replicate. However, large-scale collaboration on GWAS has made it possible to detect at least two loci that do appear to replicate across studies and potentially even across ancestral backgrounds.

2.8 Question 5: How is it Changing?

Evolution in SARS-CoV-2 has also been observed over a short timescale. After zoonotic transfer, SARS-CoV-2 continued evolving in the human population [208]. The SARS-CoV-2 mutation rate is moderate compared to other RNA viruses [210], which likely restricts the pace of evolution in SARS-CoV-2. Nevertheless, genomic analyses have yielded statistical evidence of ongoing evolution. Initially, two known variants of the spike protein emerged that differed by a single amino acid at position 614 (G614 and D614), and there is evidence that G614 had become more prevalent than D614 by June 2020 [220]. While there is a hypothesis that this genomic change increased the SARS-CoV-2 infectivity and virulence, this hypothesis has not yet been tested due to a lack of data [456]. Another study [210] identified 198 recurrent mutations in a dataset of 7,666 curated sequences, all of which defined non-synonymous protein-level modifications. This pattern of convergent evolution at some sites could indicate that certain mutations confer an adaptive advantage. While it is evident that SARS-CoV-2 exhibits moderate potential for ongoing and future evolution, the relationship between mutations and pathogenicity is not yet known. Additional data is needed in order to understand patterns of evolutionary change and the mechanisms by which they might affect virulence.

Several factors could promote the evolution of SARS-CoV-2, including host immunodeficiency and transient exposure to antibodies directed against SARS-CoV-2 proteins. A single case study of SARS-CoV-2 infection in an immunocompromised female with chronic lymphocytic leukemia and hypogammaglobulinemia [457] suggested that an accelerated evolution of the virus could occur in conditions of immunodeficiency. A first administration of convalescent plasma did not clear the virus, and an ensuing increase in the genomic diversity in the samples was observed, suggesting an accelerated evolution due to selection pressure. A second administration of convalescent plasma cleared the virus from the host 105 days after the initial diagnosis. However, throughout the duration of infection, the patient was asymptomatic but contagious. A second single case study in a 45-year old male with antiphospholipid syndrome [458] confirmed the earlier results,

providing evidence of persistent COVID-19 symptoms in an immunocompromised patient for 154 days following diagnosis, ultimately leading to the death of patient. The treatments administered included remdesivir and the Regeneron anti-spike protein antibody cocktail. Genomic analyses of the patient's nasopharyngeal swabs confirmed an accelerated evolution of the virus through mutations in the spike gene and the receptor-binding domain. In summary, these two case studies suggested an accelerated evolution and persistent shedding of the virus in conditions of immunodeficiency. In particular, the first case highlighted the role of convalescent plasma in creating escape variants. In fact, one study [459] exposed the SARS-CoV-2 virus to convalescent plasma *in vitro* repeatedly to see how much plasma was required to neutralize the virus. The results of the first six exposures were similar, but they reported that after the seventh exposure (on day 45), the amount of plasma required began to increase. In analyzing the viral variants present, they found that this viral escape was promoted by the sudden accumulation of mutations, especially in the receptor-binding domain (RBD) and N-terminal domain (NTD), that quickly rose in frequency. By the thirteenth exposure (day 85), the virus had evolved three mutations and could no longer be neutralized by the plasma used, even though the plasma was comprised of polyclonal serum that targeted a variety of epitopes. Taken together, these observations suggest that evolutionary analyses of SARS-CoV-2 can provide crucial information about the conditions that promote resistance in SARS-CoV-2 and the kinetics of how resistance develops, information which will be important for understanding the implications of how vaccine regimens are designed and whether/when next-generation vaccines will be needed.

When variants occur, they can rise in frequency by chance or through an adaptive process that confers a competitive advantage to the virus. Variants that had the D614G mutation in the spike glycoprotein seemed to spread faster. However, it has been suggested that the mutation rose in frequency due to early chance events rather than by adaptive events [460]. Another mutation, Y453F, that occurred in the receptor binding domain of *S*, was first detected in mink; however, the transmission to humans has been established. In mink, this mutation conferred an advantage by increasing the affinity towards ACE2 [461]. Similarly, N501Y mutation induces an increased affinity towards human ACE2 and has been involved in the dominance of B.1.1.7 by outcompeting other variants [462]. Therefore, genomic surveillance is essential to prevent the emergence of super-spreaders [463].

Emerging methods are being applied to this problem in an effort to understand which mutations are most likely to be of significant concern. Novel machine learning methods were developed to predict the mutations in the sequence that promote viral escape. While they preserve the pathogenicity of the virus, escape mutations change the virus's sequence to evade detection by the immune system. By using tools from natural language processing (NLP), viral escape was modeled as an NLP problem [464] where a modification makes a sentence grammatically correct but semantically different. Therefore, language models of viruses could predict mutations that change the presentation of the virus to the immune system but preserve its infectivity.

2.8.1 Variants of Concern and Variants under Surveillance

Viral replication naturally leads to the occurrence of mutations, and thus to genetic variation [465]. However, due to an intrinsic RNA proof-reading process in the SARS-CoV-2 virus, the pace of evolution of SARS-CoV-2 is moderate in comparison to other viruses [466]. The declaration of the first SARS-CoV-2 variant of concern (VOC) B.1.1.7 in December 2020 has attracted significant media attention. While the B.1.1.7 lineage garnered attention in November 2020, two genomes of the lineage were detected as early as September 20th, 2020 from routine genomic data sampled in Kent (U.K.) by the COVID-19 Genomics UK Consortium (COG-UK). The following day, a second B.1.1.7 genome was reported in greater London [232,460,467,468]. Since then, B.1.1.7 has spread across the UK and internationally, and it has now been detected in at least 62 countries [469], despite several countries imposing travel restrictions on travelers from the UK. Of the twenty-three mutations that define B.1.1.7 from the original strain isolated in Wuhan (lineage A), fourteen are lineage-specific and three appear to be biologically consequential mutations associated with the spike protein, namely N501Y, P681H, and 69-70del [467,468]. The latter is a 6-bp deletion that leads to the loss of two amino acids and has consequences for immune recognition; it may, in conjunction with N501Y, be responsible for the increased transmissibility of the B.1.1.7 VOC due to changes in the RBD that increase binding affinity with ACE2 [228,467]. B.1.1.7 has increased transmissibility by up to 56%, leading to an R_0 of approximately 1.4. Additionally, this VOC has been shown to be associated with increased disease severity and increased mortality [470]. Other variants also express the 69-70del mutation [471,472], and public health officials in the United States and the UK have been able to use RT-PCR-based assays (ThermoFisher TaqPath COVID-19 assay) to identify sequences with this deletion because it occurs where the qPCR probe binds [232]. In the UK, B.1.1.7 is present in more than 97% of diagnostic tests that return negative for S-gene targets and positive for the other targets; thus, the frequency of S-gene target failure can be used as a proxy for the detection of B.1.1.7 [467,473]. The FDA has highlighted that the performance of three diagnostic tests may be affected by the B.1.1.7 lineage because it could cause false negative tests [474].

While B.1.1.7 is currently the main VOC, other genetic variants also currently designated as VOCs have been detected, including B.1.351 and P.1, both of which emerged independently [475,476]. B.1.351 was first detected in October 2020 in South Africa, was later detected in the EU on December 28th, 2020 and has now spread to at least 26 countries [229,477,478]. B.1.351 contains several mutations at the RBD including K417N, E484K, and N501Y. While the biological significance of these mutations are still under investigation, it does appear that this lineage may be associated with increased transmissibility [479] due to the N501Y mutation [228,468]. Additionally, an analysis of a pseudovirus expressing the 501Y.V2 spike protein (B.1.351) showed that this variant demonstrates increased resistance to neutralization by convalescent plasma, even though total binding activity remained mostly intact [480]. Further, using a live virus neutralization assay (LVNA), it was shown that 501Y.V2 (B.1.351) is poorly neutralized by convalescent plasma obtained from individuals who responded to non-

501Y.V2 variants [481]. However, 501Y.V2 infection-elicited plasma was able to cross-neutralize earlier non-501Y.V2 variants, suggesting that vaccines targeting VOCs may be effective against other mutant lineages [481].

The P.1 variant is a sublineage of the B.1.1.28 lineage that was first detected in Japan in samples obtained from four travelers from Brazil during a screening at a Tokyo airport on January 10, 2021 [482]. Shortly thereafter, it was established that there was a concentration of cases of the P.1 variant in Manaus, Brazil. In a small number of samples (n=31) sequenced in Manaus, 42% were identified as the P.1 variant as early as mid-December, but the variant seemed to be absent in genome surveillance testing prior to December [483]. To date, at least eight countries have detected the P.1 lineage [484]. While the majority of P.1 cases detected internationally have been linked to travel originating from Brazil, the UK has also reported evidence of community transmission detected via routine community sequencing [484,485].

P.1 has eight lineage-specific mutations along with three concerning spike protein mutations in the RBD, including K417T, E484K, and N501Y [479].

There have been multiple different SARS-CoV-2 lineages detected that have mostly been of no more clinical concern than the original devastating lineage originating in Wuhan [486]. However, the spotlight has been cast on other variants of unknown clinical relevance due to the increase of cases observed that have been associated with B.1.1.7 in particular.

Although early in its ascendency, B.1.427/429 are SARS-CoV-2 variants that was detected in California, USA and also known as CAL.20C [487]. It was first detected in July 2020 but was not detected again until October 2020. In December 2020, B.1.427/429 accounted for ~24% of the total cases in Southern California and ~36% of total cases in the Los Angeles area.

B.1.427/429 have now been detected in several U.S. states and at least 38 countries worldwide [487,488]. This variant is characterized by five key lineage-specific mutations (ORF1a: I4205V, ORF1b:D1183Y, S: S13I;W152C;L452R). The latter spike mutation, L452R, is found in an area of the RBD known to resist monoclonal antibodies to the spike protein [489], and it is hypothesized that this mutation may resist polyclonal sera in convalescent patients or in individuals post-vaccination [487,490].

B.1.427/429 are now designated VOCs [476]; however, further research is still required to determine the implications of the mutations encoded in this genetic variant.

Another notable variant has recently been discovered in 35 patients in a Bavarian hospital in Germany; however, the sequencing data has not been published to date and it remains to be determined whether this variant is of any further concern [491].

There are several shared mutations and deletions between the three lineages, P.1, B.1.1.7, and B.1.315 and indeed other variants of SARS-CoV-2 that are under investigation [483]. For example, N501Y, which appears to have occurred independently in each of the three lineages.

E484K is present in both B.1.351 and P.1 [492]. The mutations N501Y and E484K are found in the RBD within the receptor-binding motif responsible for forming an interface with the ACE2 receptor, which seems to be consequential for ACE2 binding affinity [493]. Indeed, N501Y is associated with increased virulence and infectivity in mouse models [494]. E484K has

also been associated with evasion from neutralizing antibodies [459,490,495]. The del69-70 (del:11288:9) is also shared between P.1 and B.1.1.7 and happens to be a common deletion found in the N terminal mutation of the spike protein. This deletion has also been associated with several RBD mutations [228,468,496]. There is concern that mutations in the spike protein of variants may lead to clinical consequences for transmissibility, disease severity, re-infection, therapeutics, and vaccinations [459,490,497,498,499,500,501].

Vaccine producers are working to determine whether the vaccines are still effective against the novel genetic variants. Moderna recently published data for their mRNA-1273 vaccine that showed no significant impact of neutralization against the B.1.1.7 variant upon vaccination in humans and non-human primates. On the other hand, Moderna reported a reduced but significant neutralization against the B.1.351 variant upon vaccination [502]. Indeed, Pfizer–BioNTech reported that sera from twenty participants vaccinated with the BNT162b COVID-19 vaccine in previous clinical trials [503,504] elicited equivalent neutralizing titers against isogenic Y501 SARS-CoV-2 on an N501Y genetic background *in vitro* [505]. Another study has reported that the plasma neutralizing activity against SARS-CoV-2 variants encoding the combination of K417N:E484K:N501Y or E484K or N501Y was variably and significantly reduced in the sera of twenty participants who received either the Pfizer–BioNTech BNT162b (n = 6) vaccine or the Moderna’s mRNA-1273 vaccine (n = 14) [506]. In a study focusing on serum samples from a combination of convalescent individuals, those who obtained the mRNA-1273 vaccine, and those who obtained Novavax, in comparison to the D614G variant, the B.1.419 variant was 2-3 times less sensitive to neutralization while the B.1.351 variant was 9-14 times less sensitive [507]. Indeed, the E484K substitution seen in the P.1 and B.1.315 variants of the B.1.1.7 lineage are broadly reported to substantially reduce the efficacy of mRNA-based vaccines [507,508,509]. For now, the consensus appears to be that the FDA-approved vaccines still seem to be generally effective against the genetic variants of SARS-CoV-2 and their accompanying mutations, albeit with a lower neutralizing capacity [502,505,506,510], though select VOCs may present challenges. Further research is required to discern the clinical, prophylactic, and therapeutic consequences of these genetic SARS-CoV-2 variants as the pandemic evolves.

2.9 Question 6: What is Next?

The SARS-CoV-2 pandemic has presented many unprecedented scientific opportunities. The rapid identification of the genomic sequence of the virus allowed for early contextualization of SARS-CoV-2 among other known respiratory viruses, and the scientific community has continued to collect, analyze, and disseminate information about the SARS-CoV-2 virus and the associated illness, COVID-19 at previously unimaginable rates [4]. The accessibility of genome sequencing technology has allowed for deep sequencing of the virus to establish a level of viral surveillance that had never before been achieved [212,511,512]. The information obtained from genetic, bioinformatics, and evolutionary analysis has played a significant role in shaping the global pandemic response [511,513,514]. For example, wastewater surveillance has emerged as a potential epidemiological tool to monitor SARS-CoV-2 spread over large regions, complementing clinical

surveillance [515,516,517]. Humans shed SARS-CoV-2 viral RNA in feces [518] that can be detected in wastewater. Protocols have been developed to safely and reproducibly isolate and quantify SARS-CoV-2 in samples obtained from wastewater processing plants [517,519]. To date, studies show that wastewater surveillance is an effective tool to monitor SARS-CoV-2 spread over large sewersheds [515,516,517,520]. Indeed, data from a study in New York City indicated that wastewater SARS-CoV-2 detection correlated with clinical detection of infection [520]. Similar studies have been conducted in Nevada [521] and Boston [522]. To date, studies have shown that factors such as temperature, the travel time of wastewater, and diurnal variability may affect detection of SARS-CoV-2 [515,521]. Additionally, wastewater surveillance provides a tool to monitor fluctuations in the viral strains present in a community [523,524]. Due to its demonstrated utility so far, the United States CDC established the National Wastewater Surveillance System (NWSS), which has emerged as an important surveillance tool for SARS-CoV-2 spread [525].

Knowledge of the evolution of SARS-CoV-2 is imperative to managing it moving forward [511,526].

The evolutionary questions highlighted here all point back to the fact that efforts to prevent future epidemics and pandemics will benefit greatly from long-term, sustainable efforts to monitor disease. Beyond understanding the status and evolution of known pathogens via genomic surveillance, greater preparedness for novel viral threats would also result from monitoring zoonotic disease. If not addressed, economic and environmental stressors are likely to cause future zoonotic transfer of diseases in the future [527]. The COVID-19 pandemic has highlighted both the incredible insights available with modern evolutionary and genomic methodologies, but has also revealed the reluctance of political actors to commit resources to these efforts outside of periods of acute need. The One Health framework has emerged from collaborations by many prominent non-governmental organizations such as the World Health Organization to promote scientific goals supportive of pandemic preparedness [512]. Genomic surveillance of human pathogens and of pathogens at the human-wildlife interface is an important component needed to meet the goals of One Health [512]. These efforts are especially important as anthropogenic alterations to the landscape such as climate change and urbanization increase the risk of zoonotic disease transmission [528,529]. With the COVID-19 pandemic serving as a clear illustration of why this surveillance is imperative and of its feasibility, wider awareness and adoption of the One Health paradigm is the last piece needed to develop practices that will prevent the next pandemic.

3 Molecular and Serologic Diagnostic Technologies for SARS-CoV-2

3.1 Abstract

The COVID-19 pandemic has presented many challenges that have spurred biotechnological research to address specific problems. Diagnostics is one area where biotechnology has been critical. Diagnostic tests play a vital role in managing a viral threat by facilitating the detection of infected and/or recovered individuals. From the perspective of what information is provided, these tests fall into two major categories, molecular and serological.

Molecular diagnostic techniques assay whether a virus is present in a biological sample, thus making it possible to identify individuals who are currently infected. Additionally, when the immune system is exposed to a virus, it responds by producing antibodies specific to the virus. Serological tests make it possible to identify individuals who have mounted an immune response to a virus of interest and therefore facilitate the identification of individuals who have previously encountered the virus. These two categories of tests provide different perspectives valuable to understanding the spread of SARS-CoV-2. Within these categories, different biotechnological approaches offer specific advantages and disadvantages. Here we review the categories of tests developed for the detection of the SARS-CoV-2 virus or antibodies against SARS-CoV-2 and discuss the role of diagnostics in the COVID-19 pandemic.

3.2 Importance

Testing is critical to pandemic management. Among molecular tests, messaging about testing strategies has varied widely between countries, with the United States in particular emphasizing the higher sensitivity of polymerase chain reaction tests above immunoassays. However, these tests offer different advantages, and a holistic view of the testing landscape is needed to identify the information provided by each test and its relevance to addressing different questions. Another important consideration is the ease of use and ability to scale for each test, which determines how widely they can be deployed. Here we describe the different diagnostic technologies available as well as the information they provide about SARS-CoV-2 and COVID-19.

3.3 Introduction

Since the emergence of *Severe acute respiratory syndrome-like coronavirus 2* (SARS-CoV-2) in late 2019, significant international efforts have focused on managing the spread of the virus. Identifying individuals who have contracted coronavirus disease 2019 (COVID-19) and may be contagious is crucial to reducing the spread of the virus. Given the high transmissibility of SARS-CoV-2 and the potential for asymptomatic or presymptomatic individuals to be contagious [1], the development of rapid, reliable, and affordable methods to detect SARS-CoV-2 infection is and was vitally important for understanding and controlling spread. For instance, test-trace-isolate procedures were an early cornerstone of many nations' efforts to control the outbreak [530,531,532]. Such efforts depend on diagnostic testing.

The genetic sequence of the SARS-CoV-2 virus was first released by Chinese officials on January 10, 2020 [533], and the first test to detect the virus was released about 13 days later [534]. The genomic information was critical to

the development of diagnostic approaches. There are two main classes of diagnostic tests: molecular tests, which can diagnose an active infection by identifying the presence of SARS-CoV-2, and serological tests, which can assess whether an individual was infected in the past via the presence or absence of antibodies against SARS-CoV-2. Over the course of the COVID-19 pandemic, a variety of tests have emerged within these two categories.

Molecular tests detect either viral RNA or protein in a patient sample. They are essential to identifying infected individuals, which can be important for determining courses of action related to treatment, quarantine, and contact tracing. Tests for viral RNA are done by reverse transcription (RT) of viral RNA to DNA followed by DNA amplification, usually with polymerase chain reaction (PCR) [535]. Tests for viral proteins typically use an antibody pair for detection as implemented in techniques such as lateral flow tests (LFTs) and enzyme-linked immunosorbent assays (ELISAs) [536,537]. Molecular tests require the viral genome sequence in order to develop DNA primers for viral RNA detection or to express a viral protein for use as an antigen in antibody production.

Serological tests, on the other hand, detect the presence of antibodies in blood plasma samples or other biological samples, providing insight into whether an individual has acquired immunity against SARS-CoV-2. Assays that can detect the presence of antibodies in blood plasma samples include ELISA, lateral flow immunoassay, and chemiluminescence immunoassay (CLIA) [538]. To distinguish past infection from vaccination, serological tests detect antibodies that bind the nucleocapsid protein of the SARS-CoV-2 virus [539]. They are useful for collecting population-level information for epidemiological analysis, as they can be used to estimate the extent of the infection in a given area. Thus, serological tests may be useful to address population-level questions, such as the percent of cases that manifest as severe versus mild and for guiding public health and economic decisions regarding resource allocation and counter-disease measures.

Molecular and serological tests therefore offer distinct, complementary perspectives on COVID-19 infections. Some of the same technologies are useful to both strategies, and different technologies have been employed to varying extents throughout the world since the start of the COVID-19 pandemic. Two of the primary metrics used to evaluate these tests are sensitivity and specificity. Sensitivity refers to a test's ability to correctly identify a true positive; for example, a test with 50% sensitivity would identify SARS-CoV-2 in only one of every two positive samples. On the other hand, specificity refers to how well a test is able to identify a negative sample as negative. This metric can be relevant both in terms of understanding the risk of false positives and in discussing whether a test is susceptible to identifying other coronaviruses. Here, we review the different types of tests within each category that have been developed and provide perspective on their applications.

3.4 Molecular Tests to Identify SARS-CoV-2

Molecular tests are used to identify distinct genomic subsequences of a viral molecule in a sample or the presence of viral protein, and they thus can be used to diagnose an active viral infection. An important first step is identifying which biospecimens are likely to contain the virus in infected individuals and then acquiring these samples from the patient(s) to be tested. Common sampling sources for molecular tests include nasopharyngeal cavity samples, such as throat washes, throat swabs, and saliva [540], and stool samples [541]. Once a sample is acquired from a patient, molecular tests detect SARS-CoV-2 based on the presence of either viral nucleic acids or viral proteins.

3.4.1 PCR-Based Tests

When testing for RNA from viruses like SARS-CoV-2, the first step involves pre-processing in order to create complementary DNA (cDNA) from the RNA sample using RT. The second step involves the amplification of a region of interest in the cDNA using successive cycles of heating and cooling.

Depending on the application, this amplification is achieved using variations of PCR. Reverse transcription polymerase chain reaction (RT-PCR) tests determine whether a target is present by amplifying a region of interest of cDNA [542]. Some tests use the results of the PCR itself (e.g., a band on a gel) to determine whether the pathogen is present. However, this approach has not been employed widely in diagnostic testing, and instead most PCR-based tests are quantitative.

3.4.1.1 Quantitative Real-Time PCR

In contrast to RT-PCR, quantitative, real-time PCR uses fluorescent dyes that bind to the amplified DNA, thereby allowing a real time assessment of the amplification procedure [542] (in this manuscript we refer to quantitative real-time PCR as qPCR, following the Minimum Information for Publication of Quantitative Real-Time PCR Experiments guidelines [543], and when combined with reverse transcriptase steps, as is required for the evaluation of RNA, it is known as RT-qPCR.) The time resolution provided by qPCR and RT-qPCR is useful because the amount of fluorescence emitted by the sample is proportional to the amount of DNA amplified, and therefore the amount of virus present can be indirectly measured using the cycle threshold (C_t) determined by qPCR.

The first test developed and validated for the detection of SARS-CoV-2 used RT-qPCR to detect several regions of the viral genome: the *ORF1b* of the RNA-dependent RNA polymerase (RdRP), the envelope protein gene (*E*), and the nucleocapsid protein gene (*N*) [534]. The publication reporting this test was released on January 23, 2020, less than two weeks after the sequence of the virus was first reported [534]. In collaboration with several other labs in Europe and in China, the researchers confirmed the specificity of this test with respect to other coronaviruses against specimens from 297 patients infected with a broad range of respiratory agents. Specifically, this test uses two probes against RdRP, one of which is specific to SARS-CoV-2 [534]. Importantly, this assay was not found to return false positive results.

In January 2020, Chinese researchers developed a test that used RT-qPCR to identify two gene regions of the viral genome, *ORF1b* and *N* [544]. This assay was tested on samples from two COVID-19 patients and a panel of positive and negative controls consisting of RNA extracted from several cultured viruses. The assay uses the *N* gene to screen patients, while the *ORF1b* gene region is used to confirm the infection [544]. The test was designed to detect sequences conserved across sarbecoviruses, or viruses within the same subgenus as SARS-CoV-2. Considering that *Severe acute respiratory syndrome-related coronavirus 1* (SARS-CoV-1) and SARS-CoV-2 are the only sarbecoviruses currently known to infect humans, a positive test can be assumed to indicate that the patient is infected with SARS-CoV-2, although this test is not able to discriminate the genetics of viruses within the sarbecovirus clade. The fact that the targets are so conserved offers the advantage of reduced concern about sensitivity in light of the evolution of SARS-CoV-2.

qPCR tests have played an important role in diagnostics during the COVID-19 pandemic. For SARS-CoV-2, studies have typically considered a patient to be infectious if the C_t is below 33 or sometimes 35 [1,545,546]. A lower C_t corresponds to fewer qPCR cycles needed to reach a detectable level, indicating that higher amounts of virus were present in the initial reaction. Interpretations of the C_t values obtained from these tests have raised some interesting questions related to viral load and contagiousness. Lower C_t values correspond to a higher probability of a positive viral culture, but no threshold could discriminate all positive from all negative cultures [238]. Additionally, because of the variability introduced by sample collection and clinical components of testing, C_t is not a proxy for viral load [547]. Positive PCR results have also been reported for extended periods of time from symptom onset and/or the first positive PCR test [273], meaning that in some cases, a positive PCR may not indicate that someone is contagious [1].

In addition to the nuance required to interpret PCR results, there are also factors that influence their accuracy. The specificity of these tests is very high [548], meaning that a positive RT-PCR result is very likely to indicate SARS-CoV-2 infection. The weight given to these tests as an indicator of SARS-CoV-2 infection regardless of other clinical considerations is not typical [549]. In fact, while the analytical specificity of the assay is extremely high, the challenges of implementing testing can introduce variability that results in a lower clinical specificity [549]. Several factors may influence the sensitivity and specificity, with sample collection being a critically important factor in the reliability of RT-PCR results. The most reliable results were found to come from nasopharyngeal swabs and from pooled nasal and throat swabs, with lower accuracies produced by saliva or by throat or nasal swabs alone [548,550]. Differences in experimental parameters such as the use of primers more specific to SARS-CoV-2 has been found to improve sensitivity in these specimens [551]. Additionally, the impact of viral evolution on RT-PCR sensitivity is a concern [552,553]. Using a panel that includes multiple targets can mitigate these effects [554]. Additionally, a test designed to incorporate genomic differences with SARS-CoV-1 was found to offer improved sensitivity and specificity [551]. Thus, while various factors can influence the exact parameters of testing accuracy, RT-PCR is known to have very high specificity and lower, but still high, sensitivity.

3.4.1.2 Digital PCR

Digital PCR (dPCR) is a new generation of PCR technologies offering an alternative to traditional qPCR [555]. In dPCR, a sample is partitioned into thousands of compartments, such as nanodroplets (droplet dPCR or ddPCR) or nanowells, and a PCR reaction takes place in each compartment. This design allows for a digital read-out where each partition is either positive or negative for the nucleic acid sequence being tested for, allowing for absolute target quantification through Poisson statistics. While dPCR equipment is not yet as common as that for qPCR, dPCR for DNA targets generally achieves higher sensitivity than other PCR technologies while maintaining high specificity, though sensitivity is slightly lower for RNA targets [556].

High sensitivity is particularly relevant for SARS-CoV-2 detection, since low viral load in clinical samples can lead to false negatives. In one study, Suo et al. [557] performed a double-blind evaluation of ddPCR for SARS-CoV-2 detection. They evaluated on 63 samples collected from suspected positive outpatients and 14 from supposed convalescent patients. Of the 63 outpatients, only 21 (33%) were identified as positive for SARS-CoV-2 with qPCR. However, ddPCR identified 49 (78%) as positive, 10 (16%) as negative, and 4 (6%) as suspected/borderline for SARS-CoV-2 infection. While both qPCR and ddPCR were found to have very high specificity (100%), this analysis reported that the sensitivity was 40% with qPCR compared to 94% with ddPCR. Analysis of serial dilutions of a linear DNA standard suggested that ddPCR was approximately 500 times more sensitive than qPCR [557]. Thus, this study suggests that ddPCR provides an extremely sensitive molecular test that is able to detect SARS-CoV-2 even at very low viral loads.

A second study [558] confirmed that RT-ddPCR is able to detect SARS-CoV-2 at a lower threshold for viral load relative to RT-PCR. This study analyzed 196 samples, including 103 samples from suspected patients, 77 from contacts and close contacts, and 16 from suspected convalescents, using both RT-qPCR and RT-ddPCR. First, the authors evaluated samples from the 103 suspected cases. Using RT-qPCR, 29 (28%) were identified as positive, 25 (24%) as negative, and 49 (48%) as borderline, i.e., the C_t value was higher than the positive threshold of 35 but lower than the negative threshold of 40. When the 61 negative and borderline samples were reanalyzed with ddPCR, 19 (31%) of the negative and 42 (69%) of the borderline samples were identified as positive. All of the suspected cases were later confirmed to be COVID-19 through a combination of symptom development and RT-qPCR resampling, indicating that ddPCR improved the overall detection rate compared to RT-qPCR from 28.2% to 87.4%.

They repeated this analysis in patient samples from contacts and close contacts. Patients who tested negative with both methods ($n = 48$) were observed to remain healthy over a period of 14 days. Among the remaining 29 samples from contacts, RT-qPCR identified 12 as positive, 1 as negative, and 16 as borderline. All of the samples that tested positive using RT-qPCR also tested positive using ddPCR. In contrast, the negative result and all but one of the borderline results were identified as positive by RT-ddPCR, and these patients were later determined to be SARS-CoV-2 positive based on clinical evaluation and repeated molecular sampling. Similarly, in the final group, 16 convalescent patients, RT-qPCR identified 12 as positive, three as

suspect, and one as negative, but RT-dPCR identified all as positive. The evidence from this study therefore supports a lower limit of detection with ddPCR. Overall, these studies suggest that ddPCR is a promising tool for overcoming the problem of false negatives in SARS-CoV-2 RNA testing, but this method is unlikely to affect the current pandemic due to its lack of availability.

3.4.1.3 Sequencing

In some cases, the DNA amplified with PCR is sequenced. Sequencing requires an additional sample pre-processing step called library preparation. Library preparation is the process of preparing the sample for sequencing, typically by fragmenting the sequences and adding adapters [559]. In some cases, library preparation can involve other modifications of the sample, such as adding barcodes to identify a particular sample within the sequence data. Barcoding can therefore be used to pool samples from multiple sources. There are different reagents used for library preparation that are specific to identifying one or more target sections with PCR [560]. Sequential pattern matching is then used to identify unique subsequences of the virus, and if sufficient subsequences are found, the test is considered positive. Therefore, tests that use sequencing require a number of additional molecular and analytical steps relative to tests that use PCR alone.

Sequencing has been an important strategy for discovery of SARS-CoV-2 variants (e.g., see [487]). Sequencing elucidates any genetic variants located between the PCR primers. For this reason, it is critical to genomic surveillance efforts. Genomic surveillance is an important complement to epidemiological surveillance efforts [561], as described below. Through genomic surveillance, it has become possible to monitor the emergence of variants of interest and variants of concern (VOC) that may pose additional threats due to increased contagiousness, virulence, or immune escape [561,562]. Sequencing also allows for analysis of the dominant strains in an area at a given time. Worldwide, the extent of genomic surveillance varies widely, with higher-income countries typically able to sequence a higher percentage of cases [563]. Sequencing efforts are important for identifying variants containing mutations that might affect the reliability of molecular diagnostic tests, as well as mitigation measures such as therapeutics and prophylactics [552,553]. Therefore, sequencing is an important component of diagnostics: while it is not necessary for diagnosing an individual case, it is critical to monitoring trends in the variants affecting a population and to staying aware of emerging variants that may pose additional challenges.

3.4.1.4 Pooled and Automated PCR Testing

Due to limited supplies and the need for more tests, several labs have found ways to pool or otherwise strategically design tests to increase throughput. The first such result came from Yelin et al. [564], who reported that they could pool up to 32 samples in a single qPCR run. This was followed by larger-scale pooling with slightly different methods [565]. Although these approaches are also PCR based, they allow for more rapid scaling and higher efficiency for testing than the initial PCR-based methods developed.

Conceptually, pooling could also be employed in analysis with RT-qPCR [566], and this strategy has been evaluated in settings such as schools [567] and hospitals [568].

3.4.2 RT-LAMP

RT-PCR remains the gold standard for detection of SARS-CoV-2 RNA from infected patients, but the traditional method requires special equipment and reagents, including a thermocycler. Loop-mediated isothermal amplification (LAMP) is an alternative to PCR that does not require specialized equipment [569]. In this method, nucleic acids are amplified in a 25 µL reaction that is incubated and chilled on ice [569]. It uses primers designed to facilitate autocycling strand displacement DNA synthesis [569]. LAMP can be combined with reverse transcription (RT-LAMP) to enable the detection of RNA.

One study showed that RT-LAMP is effective for detection of SARS-CoV-2 with excellent specificity and sensitivity and that this method can be applied to unprocessed saliva samples [570]. This method was benchmarked against RT-PCR using 177 human nasopharyngeal RNA samples, of which 126 were COVID positive. The authors break down the sensitivity of their test according to the C_t value from RT-PCR of the same samples; RT-LAMP performs at 100% sensitivity for samples with a C_t from RT-PCR of 32 or less. The performance is worse when considering all RT-PCR positive samples (including those with C_t values between 32-40). However, there is some evidence suggesting that samples obtained from individuals that achieve C_t values >30 measured using RT-PCR tend to be less infective than those that record a C_t value <30 [571,572,573], so RT-LAMP is still a useful diagnostic tool. Various combinations of reagents are available, but one example is the WarmStart Colorimetric LAMP 2X Master Mix with a set of six primers developed previously by Zhang et al. [574]. To determine assay sensitivity, serial tenfold dilutions of *in vitro* transcribed *N*-gene RNA standard were tested using quantities from 10^5 copies down to 10 copies. The assay readout is the color of the dye changing from pink to yellow due to binding to the DNA product over 30 minutes. The RT-LAMP assay was then applied to clinical nasopharyngeal samples. For viral loads above 100 copies of genomic RNA, the RT-LAMP assay had a sensitivity of 100% and a specificity of 96.1% from purified RNA. The sensitivity of the direct assay of saliva by RT-LAMP was 85%. Sensitivity and specificity metrics were obtained by comparison with results from RT-PCR. RT-LAMP pilot studies for detection of SARS-CoV-2 were reviewed in a meta-analysis [575]. In the meta-analysis of all 2,112 samples, the cumulative sensitivity of RT-LAMP was calculated at 95.5%, and the cumulative specificity was 99.5%.

This test aims to bring the sensitivity of nucleic acid detection to the point of care or home testing setting. It could be applied for screening, diagnostics, or as a definitive test for people who are positive based on LFTs (see below). The estimated cost per test is about 2 euros when RNA extraction is included. The main strength of this test over RT-PCR is that it can be done isothermally, but the main drawback is that it is about 10-fold less sensitive than RT-PCR. The low cost, excellent sensitivity/specificity, and quick readout of RT-LAMP makes this an attractive alternative to RT-PCR. Alternative strategies like RT-LAMP are needed to bring widespread testing away from the lab and into under-resourced areas.

3.4.3 CRISPR-based Detection

Technology based on CRISPR (clustered regularly interspaced short palindromic repeats) [576] has also been instrumental in scaling up testing protocols. Two CRISPR-associated nucleases, Cas12 and Cas13, have been used for nucleic acid detection. Multiple assays exploiting these nucleases have emerged as potential diagnostic tools for the rapid detection of SARS-CoV-2 genetic material and therefore SARS-CoV-2 infection. The SHERLOCK method (Specific High-sensitivity Enzymatic Reporter unLOCKing) from Sherlock Biosciences relies on Cas13a to discriminate between inputs that differ by a single nucleotide at very low concentrations [577]. The target RNA is amplified by real-time recombinase polymerase amplification (RT-RPA) and T7 transcription, and the amplified product activates Cas13a. The nuclease then cleaves a reporter RNA, which liberates a fluorescent dye from a quencher. Several groups have used the SHERLOCK method to detect SARS-CoV-2 viral RNA. An early study reported that the method could detect 7.5 copies of viral RNA in all 10 replicates, 2.5 copies in 6 out of 10, and 1.25 copies in 2 out of 10 runs [578]. It also reported 100% specificity and sensitivity on 114 RNA samples from clinical respiratory samples (61 suspected cases, among which 52 were confirmed and nine were ruled out by metagenomic next-generation sequencing, 17 SARS-CoV-2-negative but human coronavirus (HCoV)-positive cases, and 36 samples from healthy subjects) and a reaction turnaround time of 40 minutes. A separate study screened four designs of SHERLOCK and extensively tested the best-performing assay. They determined the limit of detection to be 10 copies/ μ l using both fluorescent and lateral flow detection [579].

LFT strips are simple to use and read, but there are limitations in terms of availability and cost per test. Another group therefore proposed the CREST (Cas13-based, Rugged, Equitable, Scalable Testing) protocol, which uses a P51 cardboard fluorescence visualizer, powered by a 9-volt battery, for the detection of Cas13 activity instead of immunochromatography [580]. CREST can be run, from RNA sample to result, with no need for AC power or a dedicated facility, with minimal handling in approximately 2 hours. Testing was performed on 14 nasopharyngeal swabs. CREST picked up the same positives as the CDC-recommended TaqMan assay with the exception of one borderline sample that displayed low-quality RNA. This approach may therefore represent a rapid, accurate, and affordable procedure for detecting SARS-CoV-2.

The DETECTR (DNA Endonuclease-Targeted CRISPR Trans Reporter) method from Mammoth Biosciences involves purification of RNA extracted from patient specimens, amplification of extracted RNAs by loop-mediated amplification, and application of their Cas12-based technology. In this assay, guide RNAs (gRNAs) were designed to recognize portions of sequences corresponding to the SARS-CoV-2 genome, specifically the N2 and E regions [581]. In the presence of SARS-CoV-2 genetic material, sequence recognition by the gRNAs results in double-stranded DNA cleavage by Cas12, as well as cleavage of a single-stranded DNA molecular beacon. The cleavage of this molecular beacon acts as a colorimetric reporter that is subsequently read out in a lateral flow assay and indicates the presence of SARS-CoV-2 genetic material and therefore SARS-CoV-2 infection. The 40-minute assay is considered positive if there is detection of both the *E* and *N* genes or

presumptive positive if there is detection of either of them. The assay had 95% positive predictive agreement and 100% negative predictive agreement with the US Centers for Disease Control and Prevention SARS-CoV-2 RT-qPCR assay. The estimated limit of detection was 10 copies per μl reaction, versus 1 copy per μl reaction for the CDC assay.

These results have been confirmed by other DETECTR approaches. Using RT-RPA for amplification, another group detected 10 copies of synthetic SARS-CoV-2 RNA per μl of input within 60 minutes of RNA sample preparation in a proof-of-principle evaluation [582]. Through a similar approach, another group reported detection at 1 copy per μl [583]. The DETECTR protocol was improved by combining RT-RPA and CRISPR-based detection in a one-pot reaction that incubates at a single temperature and by using dual CRISPR RNAs, which increases sensitivity. This new assay, known as All-In-One Dual CRISPR-Cas12a, detected 4.6 copies of SARS-CoV-2 RNA per μl of input in 40 minutes [584]. Another single-tube, constant-temperature approach using Cas12b instead of Cas12a achieved a detection limit of 5 copies/ μl in 40-60 minutes [585].

It was also reported that electric field gradients can be used to control and accelerate CRISPR assays by co-focusing Cas12-gRNA, reporters, and target [586]. The authors generated an appropriate electric field gradient using a selective ionic focusing technique known as isotachophoresis (ITP) implemented on a microfluidic chip. They also used ITP for automated purification of target RNA from raw nasopharyngeal swab samples. Combining this ITP purification with loop-mediated isothermal amplification, their ITP-enhanced assay achieved detection of SARS-CoV-2 RNA (from raw sample to result) in 30 minutes.

All these methods require upstream nucleic acid amplification prior to CRISPR-based detection. They rely on type V (Cas12-based) and type IV (Cas13-based) CRISPR systems. In contrast, type III CRISPR systems have the unique property of initiating a signaling cascade, which could boost the sensitivity of direct RNA detection. In type III CRISPR systems, guide CRISPR RNAs (crRNAs) are bound by several Cas proteins [587] and can target both DNA and RNA molecules [588,589]. A study tested this hypothesis using the type III-A crRNA-guided surveillance complex from *Thermus thermophilus* [590]. The authors showed that activation of the Cas10 polymerase generates three products (cyclic nucleotides, protons, and pyrophosphates) that can all be used to detect SARS-CoV-2 RNA. Detection of viral RNA in patient samples still required an initial nucleic acid amplification step, but improvements may in the future remove that requirement.

This goal of amplification-free detection was later achieved for a Cas13a-based system [591]. This approach combined multiple CRISPR RNAs to increase Cas13a activation, which is detected by a fluorescent reporter. Importantly, because the viral RNA is detected directly, the test yields a quantitative measurement rather than a binary result. The study also shows that fluorescence can be measured in a custom-made dark box with a mobile phone camera and a low-cost laser illumination and collection optics. This approach is a truly portable assay for point-of-care diagnostics. The authors

achieved detection of 100 copies/ μ l of pre-isolated RNA in 30 minutes, and correctly identified all SARS-CoV-2-positive patient RNA samples tested in 5 minutes (n = 20).

There is an increasing body of evidence that CRISPR-based assays offer a practical solution for rapid, low-barrier testing in areas that are at greater risk of infection, such as airports and local community hospitals. In the largest study to date, DETECTR was compared to RT-qPCR on 378 patient samples [592]. The authors reported 95% reproducibility. Both techniques were equally sensitive in detecting SARS-CoV-2. Lateral flow strips showed 100% correlation to the high-throughput DETECTR assay. Importantly, DETECTR was 100% specific for SARS-CoV-2 and did not detect other human coronaviruses. A method based on a Cas9 ortholog from *Francisella novicida* known as FnCas9 achieved 100% sensitivity and 97% specificity in clinical samples, and the diagnostic kit is reported to have completed regulatory validation in India [593].

3.4.4 Immunoassays for the Detection of Antigens

Immunoassays can detect molecular indicators of SARS-CoV-2 infection, such as the proteins that act as antigens from the SARS-CoV-2 virus. They offer the advantage of generally being faster and requiring less specialized equipment than other molecular tests, especially those involving PCR. As a result, immunoassays hold particular interest for implementation at home and in situations where resources for PCR testing are limited. The trade-off is that these tests typically have a lower sensitivity, and sometimes a lower specificity, than other molecular tests. However, these tests tend to return a positive result five to 12 days after symptom onset, which may therefore correlate more closely with the timeframe during which viral replication occurs [594]. Immunoassays for the detection of the SARS-CoV-2 antigen can include LFTs and ELISA, as discussed here, as well as CLIA and chromatographic immunoassays [595], as described in the serological testing section below.

3.4.4.1 Lateral Flow Tests

LFTs provide distinct value relative to PCR tests. They can return results within 30 minutes and can be performed without specialized equipment and at low cost. They also do not require training to operate and are cheap to produce. Thus, they can be distributed widely to affected populations making them an important public health measure to curb pandemic spread. LFTs rely on the detection of viral protein with an antibody. Often this is done with an antibody sandwich format, where one antibody conjugated to a dye binds at one site on the antigen, and an immobilized antibody on the strip binds at another site [536]. This design allows the dye to accumulate to form a characteristic positive test line on the strip [536]. Outside of COVID-19 diagnostics, the applications of LFTs are broad; they are routinely used for home pregnancy tests, disease detection, and even drugs of abuse detection in urine [596].

A recent review surveyed the performance of LFTs for detection of current SARS-CoV-2 infection [597]. This review covered 24 studies that included more than 26,000 total LFTs. They reported significant heterogeneity in test

sensitivities, with estimates ranging from 37.7% to 99.2%. The estimated specificities of these tests were more homogeneous, spanning 92.4% to 100.0%.

Despite having lower sensitivity than PCR tests, LFTs occupy an important niche in the management of SARS-CoV-2. Current infection detection by LFTs enables the scale and speed of testing that is beneficial to managing viral spread. LFTs were available freely to citizens in the United Kingdom until April 1, 2022 [598] and to citizens of the United States in early 2022 [599]. These tests are particularly useful for ruling out SARS-CoV-2 infection in cases where the likelihood of infection is low (e.g., asymptomatic individuals) and positives (including false positives) can be validated with testing by alternate means [600].

3.4.4.2 Enzyme-Linked Immunosorbent Assay

ELISA is a very sensitive immunoassay that can be considered a gold standard for the detection of biological targets, including antibodies and antigenic proteins [537]. It can be used to generate either quantitative or qualitative results that can be returned within a few hours [601]. ELISA builds on the idea that antibodies and antigens bind together to form complexes [537] and utilizes an enzyme covalently linked to an antibody against the antigen to produce assay signal, usually a color change [602]. The main advantage of ELISA is that it enables signal amplification through the enzyme's activity, which increases sensitivity. With sandwich ELISA, antibodies are immobilized on a surface such as a plate, and viral protein antigens in the sample bind and are retained [603]. A second antibody is added that binds to another site on the antigen is then added, and that second antibody is covalently linked to an enzyme. A substrate for that enzyme is then added to produce signal, usually light or a color change. The exact strategy for tagging with a reporter enzyme varies among different types of ELISA [537,603]. For COVID-19 diagnostics, ELISAs have been designed to detect the antigenic Spike protein [604].

One of these assays uses two monoclonal antibodies specific to the nucleocapsid of SARS-CoV-2 to evaluate the relationship between the effect of (estimated) viral load on the ability of the assay to detect the SARS-CoV-2 antigen [605]. This study analyzed 339 naso-oropharyngeal samples that were also analyzed with RT-qPCR as a gold standard. RT-qPCR identified 147 samples as positive and 192 as negative. The authors estimated the overall sensitivity and specificity to be 61.9% and 99.0%, respectively. Sensitivity increased with higher C_t . This study also assessed the performance of the ELISA test under different conditions in order to evaluate how robust it would be to the challenges of testing in real-world settings globally. Higher sensitivity was achieved for samples that were stored under ideal conditions (immediate placement in -80° C). Therefore, while immediate access to laboratory equipment is an advantage, it is not strictly necessary for ELISA to detect the antigen.

3.4.5 Limitations of Molecular Tests

Tests that identify SARS-CoV-2 using molecular technologies will identify only individuals with current infections and are not appropriate for identifying individuals who have recovered from a previous infection. Among molecular tests, different technologies have different sensitivities and specificities. In general, specificity is high, and even then, the public health repercussions of a false positive can generally be mitigated with follow-up testing. On the other hand, a test's sensitivity, which indicates the risk of a false-negative response, can pose significant challenge to large-scale testing. False negatives are a significant concern for several reasons. Importantly, clinical reports indicate that it is imperative to exercise caution when interpreting the results of molecular tests for SARS-CoV-2 because negative results do not necessarily mean a patient is virus-free [606]. To reduce occurrence of false negatives, correct execution of the analysis is crucial [607]. Additionally, PCR-based tests can remain positive for a much longer time than the virus is likely to be actively replicating [594], raising concerns about their informativeness after the acute phase of the disease. Hence, the CDC has advised individuals who suspect they have been re-infected with SARS-CoV-2 to avoid using diagnostic tests within 90 days of receiving a previous positive test [608].

Additionally, the emerging nature of the COVID-19 pandemic has introduced some challenges related to uncertainty surrounding interactions between SARS-CoV-2 and its human hosts. For example, viral shedding kinetics are still not well understood but are expected to introduce a significant effect of timing of sample collection on test results [607]. Similarly, the type of specimen could also influence outcomes, as success in viral detection varies among clinical sample types [548,550,607]. With CRISPR-based testing strategies, the gRNA can recognize off-target interspersed sequences in the viral genome [609], potentially resulting in false positives and a loss of specificity.

There are also significant practical and logistical concerns related to the widespread deployment of molecular tests. Much of the technology used for molecular tests is expensive, and while it might be available in major hospitals and/or diagnostic centers, it is often not available to smaller facilities [610]. At times during the pandemic, the availability of supplies for testing, including swabs and testing media, has also been limited [611]. Similarly, processing times can be long, and tests might take up to 4 days to return results [610], especially during times of high demand, such as spikes in case numbers [612]. Countries have employed various and differing molecular testing strategies as a tool to reduce viral transmission, even among high-income countries [613]. The rapid development of molecular tests has provided a valuable, albeit imperfect, tool to identify active SARS-CoV-2 infections.

3.5 Serological Tests to Identify Recovered Individuals

Although several molecular diagnostic tests to detect viral genetic material have high specificity and sensitivity, they provide information only about active infection, and therefore offer just a snapshot-in-time perspective on

the spread of a disease. Most importantly, they would not work on a patient who has fully recovered from the virus at the time of sample collection. In such contexts, serological tests are informative.

Serological tests use many of the same technologies as the immunoassays used to detect the presence of an antigen but are instead used to evaluate the presence of antibodies against SARS-CoV-2 in a serum sample. These tests are particularly useful for insight into population-level dynamics and can also offer a glimpse into the development of antibodies by individual patients during the course of a disease. Immunoassays can detect antibodies produced by the adaptive immune system in response to viral threat.

Understanding the acquisition and retention of antibodies is important both to the diagnosis of prior (inactive) infections and to the development of vaccines. The two immunoglobulin classes that are most pertinent to these goals are immunoglobulin M (IgM), which are the first antibodies produced in response to an infection, and immunoglobulin G (IgG), which are the most abundant antibodies [614,615]. Serological tests detect these antibodies, offering a mechanism through which prior infection can be identified.

However, the complexity of the human immune response means that there are many facets to such analyses.

In general, SARS-CoV-2 infection will induce the immune system to produce antibodies fairly quickly. Prior research is available about the development of antibodies to SARS-CoV-1 during the course of the associated disease, severe acute respiratory syndrome (SARS). IgM and IgG antibodies were detected in the second week following SARS-CoV-1 infection. IgM titers peaked by the first month post-infection, and then declined to undetectable levels after day 180. IgG titers peaked by day 60 and persisted in all donors through the two-year duration of study [616]. Such tests can also illuminate the progression of viral disease, as IgM are the first antibodies produced by the body and indicate that the infection is active. Once the body has responded to the infection, IgG are produced and gradually replace IgM, indicating that the body has developed immunogenic memory [617]. Therefore, it was hoped that the development of assays to detect the presence of IgM and IgG antibodies against SARS-CoV-2 would allow the identification of cases from early in the infection course (via IgM) and for months or years afterwards (via IgG). Several technologies have been used to develop serological tests for COVID-19, including ELISA, lateral flow immunoassay, chemiluminescence immunoassay, and neutralizing antibody assays [618].

3.5.1 ELISA

The application of ELISA to serological testing is complementary to its use in molecular diagnostics (see above). Instead of using an enzyme-labeled antibody as a probe that binds to the target antigen, the probe is an antigen and the target is an antibody. The enzyme used for detection and signal amplification is on a secondary antibody raised generally against human IgG or IgM. In March 2020, the Krammer lab proposed an ELISA test that detects IgG and IgM that react against the receptor-binding domain (RBD) of the spike proteins (S) of the virus [619]. A subsequent ELISA test developed to detect SARS-CoV-2 IgG based on the RBD reported a specificity of over 99% and a sensitivity of up to 88.24%, which was observed in samples collected 21 to 27 days after the onset of infection (approximated with symptom onset or

positive PCR test) [620]. Earlier in the disease course, sensitivity was lower: 53.33% between days 0 and 13 and 80.47% between days 14 and 20. This study reported that their laboratory ELISA outperformed two commercial kits that also used an ELISA design [620]. Therefore, while analysis with ELISA requires laboratory support and equipment, these results do suggest that ELISA achieves relatively high sensitivity, especially in the weeks following infection. Efforts have been made to develop low-cost strategies for conducting these tests that will make them more accessible worldwide [621].

3.5.2 Chemiluminescence Immunoassay

Another early approach investigated for detection of antibodies against SARS-CoV-2 was CLIA. Like ELISA, CLIA is a type of enzyme immunoassay (EIA) [622]. While the technique varies somewhat, in one approach, a bead is coated with the antigen and then washed with the sample [623]. If the antibody is present in the sample, it will bind to the bead. Then the bead is exposed to a label, a luminescent molecule that will bind to the antigen/antibody complex and can therefore be used as an indicator [623]. One CLIA approach to identify COVID-19 used a synthetic peptide derived from the amino acid sequence of the SARS-CoV-2 S protein [624]. It was highly specific to SARS-CoV-2 and detected IgM in 57.2% and IgG in 71.4% of serum samples from 276 COVID-19 cases confirmed with RT-qPCR. IgG could be detected within two days of the onset of fever, but IgM could not be detected any earlier [624], which has been supported by other analyses as well [625]. This pattern was consistent with observations in Middle East respiratory syndrome, which is also caused by an HCoV. In comparisons of different commercial immunoassays, accuracy of CLIA tests were often roughly comparable to other EIAs [626], although one CLIA did not perform as well as several other EIAs [625,627]. The sensitivity and specificities reported vary among CLIA tests and for the detection of IgM versus IgG, but sensitivities and specificities as high as 100% have been reported among various high-throughput tests [627,628,629]. CLIA has previously been used to develop tests that can be used at point of care (e.g., [622]) which may allow for this technique to become more widely accessible in the future.

3.5.3 Lateral Flow Immunoassay

The first serological test approved for emergency use in the United States was developed by Cellex [630]. The Cellex qSARS-CoV-2 IgG/IgM Rapid Test is a chromatographic immunoassay, also known as a lateral flow immunoassay, designed to qualitatively detect IgM and IgG antibodies against SARS-CoV-2 in the plasma of patients suspected to have developed a SARS-CoV-2 infection [630]. The Cellex test cassette contains a pad of SARS-CoV-2 antigens and a nitrocellulose strip with lines for each of IgG and IgM, as well as a control (goat IgG) [630]. In a specimen that contains antibodies against the SARS-CoV-2 antigen, the antibodies will bind to the strip and be captured by the IgM and/or IgG line(s), resulting in a change of color [630]. With this particular assay, results can be read within 15 to 20 minutes [630]. Lateral flow immunoassays are often available at point of care but can have very low sensitivity [627].

3.5.4 Neutralizing Antibody Assays

Neutralizing antibody assays play a functional role in understanding immunity that distinguishes them from other serological tests. The tests described above are all binding antibody tests. On the other hand, rather than simply binding an antibody to facilitate detection, neutralizing antibody assays determine whether an antibody response is present that would prevent infection [631,632]. Therefore, these tests serve the purpose of evaluating the extent to which a sample donor has acquired immunity that will reduce susceptibility to SARS-CoV-2. As a result, neutralizing antibody assays have been used widely to characterize the duration of immunity following infection, to assess vaccine candidates, and to establish correlates of protection against infection and disease [633,634,635]. These tests are typically performed in a laboratory [631], and in SARS-CoV-2, the results of neutralizing antibody assays are often correlated with the results of binding antibody tests [631].

The gold standard for assessing the presence of neutralizing antibodies is the plaque reduction neutralization test (PRNT), but this approach does not scale well [632]. An early high-throughput neutralizing antibody assay designed against SARS-CoV-2 used a fluorescently labeled reporter virus that was incubated with different dilutions of patient serum [632]. The cells used for incubation would turn green if antibodies were not present. Essentially, this assay evaluates whether the virus is able to infect the cell in the presence of the serum. The specificity of this assay was 100%, and the correlation between the results of this assay and of PRNT was 0.85 with the results suggesting that the sensitivity of the high-throughput approach was higher than that of PRNT [632]. While this approach was performed on a plate and using cells, other methods have been developed using methods such as bead arrays [636].

3.5.5 Duration of Immune Indicators

While the adaptive immune system produces antibodies in response to SARS-CoV-2 viral challenge, these indicators of seroconversion are unlikely to remain in circulation permanently. Previously, a two-year longitudinal study following convalesced SARS patients with a mean age of 29 found that IgG antibodies were detectable in all 56 patients surveyed for at least 16 months and remained detectable in all but 4 patients (11.8%) through the full two-year study period [637]. These results suggest that immunity to SARS-CoV-1 is sustained for at least a year. Circulating antibody titers to other coronaviruses have been reported to decline significantly after one year [638]. Evidence to date suggests that sustained immunity to the SARS-CoV-2 virus remains for a shorter period of time but at least 6 to 8 months after infection [639,640,641,642]. However, this does not mean that all serological evidence of infection dissipates, but rather that the immune response becomes insufficient to neutralize the virus.

In order to study the persistence of SARS-CoV-2 antibodies, one study assessed sustained immunity using 254 blood samples from 188 COVID-19 positive patients [640]. The samples were collected at various time points between 6 and 240 days post-symptom onset; some patients were assessed longitudinally. Of the samples, 43 were collected at least 6 months after symptom onset. After one month, 98% of patients were seropositive for IgG to S. Moreover, S IgG titers were stable and heterogeneous among patients

over a period of 6 to 8 months post-symptom onset, with 90% of subjects seropositive at 6 months. Similarly, at 6 to 8 months 88% of patients were seropositive for RBD IgG, and 90% were seropositive for SARS-CoV-2 neutralizing antibodies. Another study examined 119 samples from 88 donors who had recovered from mild to severe cases of COVID-19 [642]. A relatively stable level of IgG and plasma neutralizing antibodies was identified up to 6 months post diagnosis. Significantly lower but considerable levels of anti-SARS-CoV-2 IgG antibodies were still present in 80% of samples obtained 6 to 8 months post-symptom onset.

Titers of IgM and IgG antibodies against the RBD were found to decrease from 1.3 to 6.2 months post infection in a study of 87 individuals [643]. However, the decline of IgA activity (15%) was less pronounced than that of IgM (53%) or IgG (32%). It was noted that higher levels of anti-RBD IgG and anti-N total antibodies were detected in individuals that reported persistent post-acute symptoms at both study visits. Moreover, plasma neutralizing activity decreased five-fold between 1.3 and 6.2 months in an assay of HIV-1 virus pseudotyped with SARS-CoV-2 S protein, and this neutralizing activity was directly correlated with IgG anti-RBD titers [643]. These findings are in accordance with other studies that show that the majority of seroconverters have detectable, albeit decreasing, levels of neutralizing antibodies at least 3 to 6 months post infection [644,645,646].

Determining the potency of anti-RBD antibodies early in the course of an infection may be important moving forward, as their neutralizing potency may be prognostic for disease severity and survival [647]. The duration of immunity might also vary with age [648] or ABO blood type [649]. Autopsies of lymph nodes and spleens from severe acute COVID-19 patients showed a loss of T follicular helper cells and germinal centers that may explain some of the impaired development of antibody responses [650]. Therefore, serological testing may be time-limited in its ability to detect prior infection.

Other immune indicators of prior infection have also been evaluated to see how they persist over time. SARS-CoV-2 memory CD8⁺ T cells were slightly decreased (50%) 6 months post-symptom onset. In this same subset of COVID-19 patients, 93% of subjects had detectable levels of SARS-CoV-2 memory CD4⁺ T cells, of which 42% had more than 1% SARS-CoV-2-specific CD4⁺ T cells. At 6 months, 92% of patients were positive for SARS-CoV-2 memory CD4⁺ T cells. Indeed, the abundance of S-specific memory CD4⁺ T cells over time was similar to that of SARS-CoV-2-specific CD4⁺ T cells overall [640]. T cell immunity to SARS-CoV-2 at 6 to 8 months following symptom onset has also been confirmed by other studies [642,651,652]. In another study, T cell reactivity to SARS-CoV-2 epitopes was also detected in some individuals never been exposed to SARS-CoV-2. This finding suggests the potential for cross-reactive T cell recognition between SARS-CoV-2 and pre-existing circulating HCoV that are responsible for the “common cold” [653], but further research is required. Therefore, whether T cells will provide a more stable measure through which to assess prior infection remains unknown. Notably, commercial entities have tried to develop tests specifically for T cells, some of which have been authorized by the United States Food and Drug Administration [654,655] to identify people with adaptive T cell immune responses to SARS-CoV-2, either from a previous or ongoing infection.

3.5.6 Applications of Serological Tests

In addition to the limitations posed by the fact that antibodies are not permanent indicators of prior infection, serological immunoassays carry a number of limitations that influence their utility in different situations. Importantly, false positives can occur due to cross-reactivity with other antibodies according to the clinical condition of the patient [630]. Due to the long incubation times and delayed immune responses of infected patients, serological immunoassays are insufficiently sensitive for a diagnosis in the early stages of an infection. Therefore, such tests must be used in combination with RNA detection tests if intended for diagnostic purposes [656]. False positives are particularly harmful if they are erroneously interpreted to mean that a population is more likely to have acquired immunity to a disease [657]. Similarly, while serological tests may be of interest to individuals who wish to confirm they were infected with SARS-CoV-2 in the past, their potential for false positives means that they are not currently recommended for this use. However, in the wake of vaccines becoming widely available, accurate serological tests that could be administered at point of care were investigated in the hope that they could help to prioritize vaccine recipients [658]. Another concern with serological testing is the potential for viral evolution to reduce the sensitivity of assays, especially for neutralizing antibody assays. Chen et al. performed a systematic re-analysis of published data examining the neutralizing effect of serum from vaccinated or recovered individuals on four VOC [659]. They found reduced neutralizing titers against these variants relative to the lineages used for reference. These findings suggest that such techniques will need to be modified over time as SARS-CoV-2 evolves.

These limitations make serological tests far less useful for diagnostics and for test-and-trace strategies; however, serological testing is valuable for public health monitoring at the population level. Serosurveys provide a high-level perspective of the prevalence of a disease and can provide insight into the susceptibility of a population as well as variation in severity, e.g., between geographic regions [657]. From a public health perspective, they can also provide insight into the effectiveness of mitigation efforts and to gain insight into risk factors influencing susceptibility [660]. EIA methods are high-throughput [661,662], and, as with molecular tests, additional efforts have been made to scale up the throughput of serological tests [663]. Therefore, serological tests can be useful to developing strategies for the management of viral spread.

Early in the course of the pandemic, it was also hoped that serological tests would provide information relevant to advancing economic recovery. Some infectious agents can be controlled through “herd immunity”, which is when a critical mass within the population acquires immunity through vaccination and/or infection, preventing an infectious agent from spreading widely. It was hoped that people who had recovered and developed antibodies might be able to return to work [664,665]. This strategy would have relied on recovered individuals acquiring long-term immunity, which has not been borne out [666]. Additionally, it was hoped that identifying seroconverters and specifically those who had mounted a strong immune response would reveal strong candidates for convalescent plasma donation [619]; however,

convalescent plasma has not been found to offer therapeutic benefit (reviewed in [3]). While these hopes have not been realized, serological tests have been useful for gaining a better understanding of the pandemic [660].

3.6 Possible Alternatives to Current Diagnostic Practices

One possible alternative or complement to molecular and serological testing would be diagnosing COVID-19 cases based on symptomatology. COVID-19 can present with symptoms similar to other types of pneumonia, and symptoms can vary widely among COVID-19 patients; therefore, clinical presentation is often insufficient as a sole diagnostic criterion. In addition, identifying and isolating mild or asymptomatic cases is critical to efforts to manage outbreaks. Even among mildly symptomatic patients, a predictive model based on clinical symptoms had a sensitivity of only 56% and a specificity of 91% [667]. More problematic is that clinical symptom-based tests are only able to identify already symptomatic cases, not presymptomatic or asymptomatic cases. They may still be important for clinical practice and for reducing tests needed for patients deemed unlikely to have COVID-19.

In some cases, clinical signs may also provide information that can inform diagnosis. Using computed tomography of the chest in addition to RT-qPCR testing was found to provide a higher sensitivity than either measure alone [668]. X-ray diagnostics have been reported to have high sensitivity but low specificity in some studies [669]. Other studies have shown that specificity varies between radiologists [670], though the sensitivity reported here was lower than that published in the previous paper. While preliminary machine-learning results suggested that chest X-rays might provide high sensitivity and specificity and potentially facilitate the detection of asymptomatic and presymptomatic infections (e.g., [671]), further investigation suggested that such approaches are prone to bias and are unlikely to be clinically useful [672]. Given the above, the widespread use of X-ray tests on otherwise healthy adults is likely inadvisable.

Finally, in addition to genomic and serological surveillance, other types of monitoring have proven useful in managing the pandemic [673]. One that has received significant attention is wastewater surveillance. This approach can use several of the technologies described for molecular testing, such as qPCR and dPCR, as well as *in vitro* culturing [674] and can provide insight into trends in the prevalence of SARS-CoV-2 regionally.

3.7 Strategies and Considerations for Testing

Deciding whom to test, when to test, and which test to use have proven challenging as the COVID-19 pandemic has unfolded. Early in the COVID-19 pandemic, testing was typically limited to individuals considered high risk for developing serious illness [675]. This approach often limited testing to people with severe symptoms and people showing mild symptoms that had been in contact with a person who had tested positive. Individuals who were asymptomatic (i.e., potential spreaders) and individuals who were able to

recover at home were thus often unaware of their status. Therefore, this method of testing administration misses a high proportion of infections and does not allow for test-and-trace methods to be used. For instance, a study from Imperial College estimates that in Italy, the true number of infections was around 5.9 million in a total population of ~60 million, compared to the 70,000 detected as of March 28, 2020 [312]. Another analysis, which examined New York state, indicated that as of May 2020, approximately 300,000 cases had been reported in a total population of approximately 20 million [676]. This corresponded to ~1.5% of the population, but ~12% of individuals sampled statewide were estimated as positive through antibody tests (along with indications of spatial heterogeneity at higher resolution) [676]. Technological advancements that facilitate widespread, rapid testing would therefore be valuable for accurately assessing the rate of infection and aid in controlling the virus' spread. Additionally, the trade off of accessibility, sensitivity, and time to results has raised some complex questions around which tests are best suited to certain situations. Immunoassays, including serological tests, have much higher limits of detection than PCR tests do [677].

Changes in public attitudes and the lifting of COVID-19 restrictions due to the multifactorial desire to stimulate economic activities has required a shift of testing paradigms in 2022, despite warnings from public health officials against a hard exit from public health restrictions [678,679]. An important strategy for testing moving forward is to determine when someone becomes infectious or is no longer infectious following a positive test for COVID-19. Generally, patient specimens tend to not contain culturable virus past day 5 of symptom onset [680,681]. However, due to their sensitivity to post-infectious viral RNA in specimens, PCR-based methods may mislead individuals to believe that they are still infectious several days after symptom onset [656]. Furthermore, detection of viral RNA can occur days and weeks after an active infection due to the sensitivity of PCR-based methods [546,682,683].

In contrast, LFTs were thought to have poor sensitivity and their value for identifying infections and managing the pandemic was questioned [684,685]. However, LFTs do reliably detect SARS-CoV-2 proteins when there is a high viral load, which appears to correlate with a person's infectiousness [594,686]. Therefore, LFTs are an important diagnostic tool to determine infectiousness with fast turnaround times, ease of use, and accessibility by the general public [656,687]. One study has suggested that the test sensitivity of LFTs appears to be less important than accessibility to LFTs, frequent testing, and fast reporting times for reducing the impact of viral spread [688]. While PCR-based methods are important for COVID-19 surveillance, their use is labor intensive and time consuming, and laboratories are often slow to report results, rendering such methods limited in their surveillance capacity [656].

These limitations are demonstrated by the estimated 10-fold under-reporting of cases in the United States in 2020 due to shortages in testing and slow rollout of testing and slow reporting of results [689]. However, one strategy that may balance the strengths and weaknesses of both types of tests is to corroborate a positive LFT result using a PCR-based method. Indeed, in

December 2021 sufficient surveillance and reduction of COVID-19 spread using this joint LFT-PCR strategy was demonstrated in Liverpool, U.K., where there was an estimated 21% reduction of cases [687,690].

3.8 What Lies Ahead

Diagnostic tools have played an important role during the COVID-19 pandemic. Different tests offer different advantages (Figure 2). Specifically, the results of SARS-CoV-2 diagnostic tests (typically qPCR or LFT-based tests) have been used to estimate the number of infections in the general population, thus informing public health strategies around the globe [552]. During the surges caused by the different SARS-CoV-2 variants between 2020 and 2021, government-sponsored efforts to conduct mass testing and to provide free diagnostic tests to the population were a common occurrence in many parts of the world [691,692,693]. However, recent reports indicate that such public health policies are starting to change during 2022. For example, it is known that the UK plans to dismantle its COVID-19 testing program and scale back its daily reporting requirements [694,695]. A similar approach can be seen in the US as well, where multiple state-run testing facilities are closing, despite some groups advocating to keep them open [696,697]. These ongoing changes in testing policy are likely to have a direct effect on how the pandemic is managed moving forward. SARS-CoV-2 diagnostic tests can be used effectively to slow the spread of the disease only when 1) they are used to share testing results in a timely manner so that they can reasonably be used to approximate the number of infections in the population and 2) those tests are easily accessible by the general public.

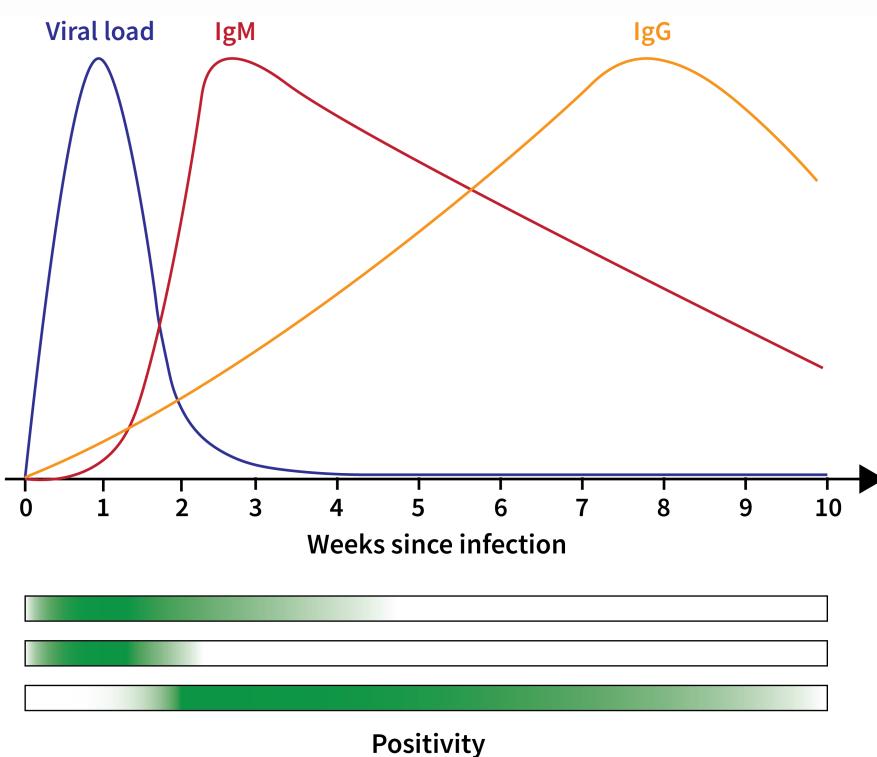


Figure 2: Summary of Diagnostic Technologies used in COVID-19 Testing. The immune response to SARS-CoV-2 means that different diagnostic approaches offer different views of COVID-19. Early in the infection course, viral load is high. This means that PCR-based testing and EIA testing for antigens are likely to return positives (as indicated by the green bars at the bottom). As viral load decreases, EIA antigen tests become negative, but PCR-based tests can still detect even very low viral loads. From a serological perspective, IgM peaks in the first few weeks following infection and then decreases, while IgG peaks much

later in the infection course. Therefore, serological tests are likely to return positives in first few months following the acute infection course. Additional detail is available above and in several analyses and reviews [1,656,683,698].

Children are one segment of the population where the importance of the two aforementioned conditions can be exemplified. This group is particularly vulnerable as there are ongoing challenges with testing in schools, increased COVID-19 mortality rates, and COVID-19-associated orphanhood. In this regard, although there is evidence of the efficacy of routine diagnostic testing to reduce the probability of having infectious students [699,700], as of March of 2022 there is an increasing number of schools that have stopped or plan to stop contact tracing efforts [701,702], in line with an announcement made by the CDC where it no longer recommended contact tracing as a strategy to contain the virus [703]. An estimated 197 children have died in the US from COVID-19 during the first three months of 2022 [704], compared to 735 deaths in the preceding 20 months of the pandemic [705], and millions of children have been orphaned as a consequence of parent or caregiver death due to COVID-19 [706]. It is likely that reducing or eliminating testing capacity in schools will directly exacerbate these negative outcomes for the remainder of 2022.

The SARS-CoV-2 diagnostic tools presented in this paper are far less useful if they are difficult to obtain, or if their limited use results in biased data that would lead to ill-informed public health strategies. Under conditions of limited supply, different strategies for testing are needed [707]. The pandemic is still an ongoing public health threat and it is worrying that active testing and tracing efforts are a low priority for public health authorities in many countries. If this trend continues, the lack of testing could result in increased morbidity and mortality and an overall failure to manage the pandemic.

4 Identification and Development of Therapeutics for COVID-19

4.1 Abstract

After emerging in China in late 2019, the novel coronavirus SARS-CoV-2 spread worldwide and as of mid-2021 remains a significant threat globally. Only a few coronaviruses are known to infect humans, and only two cause infections similar in severity to SARS-CoV-2: *Severe acute respiratory syndrome-related coronavirus*, a closely related species of SARS-CoV-2 that emerged in 2002, and *Middle East respiratory syndrome-related coronavirus*, which emerged in 2012. Unlike the current pandemic, previous epidemics were controlled rapidly through public health measures, but the body of research investigating severe acute respiratory syndrome and Middle East respiratory syndrome has proven valuable for identifying approaches to treating and preventing novel coronavirus disease 2019 (COVID-19). Building on this research, the medical and scientific communities have responded rapidly to the COVID-19 crisis to identify many candidate therapeutics. The approaches used to identify candidates fall into four main categories: adaptation of clinical approaches to diseases with related pathologies,

adaptation based on virological properties, adaptation based on host response, and data-driven identification of candidates based on physical properties or on pharmacological compendia. To date, a small number of therapeutics have already been authorized by regulatory agencies such as the Food and Drug Administration (FDA), while most remain under investigation. The scale of the COVID-19 crisis offers a rare opportunity to collect data on the effects of candidate therapeutics. This information provides insight not only into the management of coronavirus diseases, but also into the relative success of different approaches to identifying candidate therapeutics against an emerging disease.

4.2 Importance

The COVID-19 pandemic is a rapidly evolving crisis. With the worldwide scientific community shifting focus onto the SARS-CoV-2 virus and COVID-19, a large number of possible pharmaceutical approaches for treatment and prevention have been proposed. What was known about each of these potential interventions evolved rapidly throughout 2020 and 2021. This fast-paced area of research provides important insight into how the ongoing pandemic can be managed and also demonstrates the power of interdisciplinary collaboration to rapidly understand a virus and match its characteristics with existing or novel pharmaceuticals. As illustrated by the continued threat of viral epidemics during the current millennium, a rapid and strategic response to emerging viral threats can save lives. In this review, we explore how different modes of identifying candidate therapeutics have borne out during COVID-19.

4.3 Introduction

The novel coronavirus *Severe acute respiratory syndrome-related coronavirus 2* (SARS-CoV-2) emerged in late 2019 and quickly precipitated the worldwide spread of novel coronavirus disease 2019 (COVID-19). COVID-19 is associated with symptoms ranging from mild or even asymptomatic to severe, and up to 2% of patients diagnosed with COVID-19 die from COVID-19-related complications such as acute respiratory disease syndrome (ARDS) [1]. As a result, public health efforts have been critical to mitigating the spread of the virus. However, as of mid-2021, COVID-19 remains a significant worldwide concern (Figure 3), with 2021 cases in some regions surging far above the numbers reported during the initial outbreak in early 2020. While a number of vaccines have been developed and approved in different countries starting in late 2020 [328], vaccination efforts have not proceeded at the same pace throughout the world and are not yet close to ending the pandemic.

Due to the continued threat of the virus and the severity of the disease, the identification and development of therapeutic interventions have emerged as significant international priorities. Prior developments during other recent outbreaks of emerging diseases, especially those caused by human coronaviruses (HCoV), have guided biomedical research into the behavior and treatment of this novel coronavirus infection. However, previous emerging HCoV-related disease threats were controlled much more quickly

than SARS-CoV-2 through public health efforts (Figure 3). The scale of the COVID-19 pandemic has made the repurposing and development of pharmaceuticals more urgent than in previous coronavirus epidemics.

4.3.1 Lessons from Prior HCoV Outbreaks

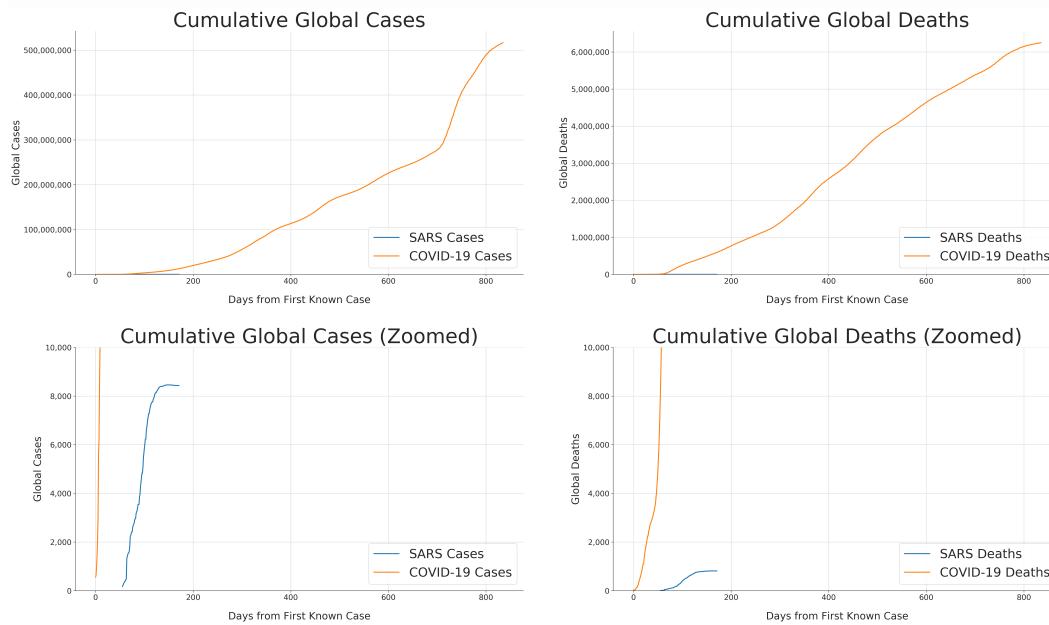


Figure 3: Cumulative global incidence of COVID-19 and SARS. As of May 6, 2022, 516,758,993 COVID-19 cases and 6,249,626 COVID-19 deaths had been reported worldwide since January 22, 2020. A total of 8,432 cases and 813 deaths were reported for SARS from March 17 to July 11, 2003. SARS-CoV-1 was officially contained on July 5, 2003, within 9 months of its appearance [708]. In contrast, SARS-CoV-2 remains a significant global threat nearly two years after its emergence. COVID-19 data are from the COVID-19 Data Repository by the Center for Systems Science and Engineering at Johns Hopkins University [709,710]. SARS data are from the WHO [711] and were obtained from a dataset on GitHub [712]. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

At first, SARS-CoV-2's rapid shift from an unknown virus to a significant worldwide threat closely paralleled the emergence of *Severe acute respiratory syndrome-related coronavirus* (SARS-CoV-1), which was responsible for the 2002-03 SARS epidemic. The first documented case of COVID-19 was reported in Wuhan, China in November 2019, and the disease quickly spread worldwide in the early months of 2020. In comparison, the first case of SARS was reported in November 2002 in the Guangdong Province of China, and it spread within China and then into several countries across continents during the first half of 2003 [248,326,708]. In fact, genome sequencing quickly revealed the virus causing COVID-19 to be a novel betacoronavirus closely related to SARS-CoV-1 [9].

While similarities between these two viruses are unsurprising given their close phylogenetic relationship, there are also some differences in how the viruses affect humans. SARS-CoV-1 infection is severe, with an estimated case fatality rate (CFR) for SARS of 9.5% [326], while estimates of the CFR associated with COVID-19 are much lower, at up to 2% [1]. SARS-CoV-1 is highly contagious and spread primarily by droplet transmission, with a basic reproduction number (R_0) of 4 (i.e., each person infected was estimated to infect four other people) [326]. There is still some controversy whether SARS-CoV-2 is primarily spread by droplets or is primarily airborne

[256,257,260,713]. Most estimates of its R_0 fall between 2.5 and 3 [1]. Therefore, SARS is thought to be a deadlier and more transmissible disease than COVID-19.

With the 17-year difference between these two outbreaks, there were major differences in the tools available to efforts to organize international responses. At the time that SARS-CoV-1 emerged, no new HCoV had been identified in almost 40 years [248]. The identity of the virus underlying the SARS disease remained unknown until April of 2003, when the SARS-CoV-1 virus was characterized through a worldwide scientific effort spearheaded by the World Health Organization (WHO) [248]. In contrast, the SARS-CoV-2 genomic sequence was released on January 3, 2020 [9], only days after the international community became aware of the novel pneumonia-like illness now known as COVID-19. While SARS-CoV-1 belonged to a distinct lineage from the two other HCoVs known at the time of its discovery [326], SARS-CoV-2 is closely related to SARS-CoV-1 and is a more distant relative of another HCoV characterized in 2012, *Middle East respiratory syndrome-related coronavirus* [17,714]. Significant efforts had been dedicated towards understanding SARS-CoV-1 and MERS-CoV and how they interact with human hosts. Therefore, SARS-CoV-2 emerged under very different circumstances than SARS-CoV-1 in terms of scientific knowledge about HCoVs and the tools available to characterize them.

Despite the apparent advantages for responding to SARS-CoV-2 infections, COVID-19 has caused many orders of magnitude more deaths than SARS did (Figure 3). The SARS outbreak was officially determined to be under control in July 2003, with the success credited to infection management practices such as mask wearing [248]. *Middle East respiratory syndrome-related coronavirus* (MERS-CoV) is still circulating and remains a concern; although the fatality rate is very high at almost 35%, the disease is much less easily transmitted, as its R_0 has been estimated to be 1 [326]. The low R_0 in combination with public health practices allowed for its spread to be contained [326]. Neither of these trajectories are comparable to that of SARS-CoV-2, which remains a serious threat worldwide over a year and a half after the first cases of COVID-19 emerged (Figure 3).

4.3.2 Potential Approaches to the Treatment of COVID-19

Therapeutic interventions can utilize two approaches: they can either mitigate the effects of an infection that harms an infected person, or they can hinder the spread of infection within a host by disrupting the viral life cycle. The goal of the former strategy is to reduce the severity and risks of an active infection, while for the latter, it is to inhibit the replication of a virus once an individual is infected, potentially freezing disease progression. Additionally, two major approaches can be used to identify interventions that might be relevant to managing an emerging disease or a novel virus: drug repurposing and drug development. Drug repurposing involves identifying an existing compound that may provide benefits in the context of interest [715]. This strategy can focus on either approved or investigational drugs, for which there may be applicable preclinical or safety information [715]. Drug development, on the other hand, provides an opportunity to identify or develop a compound specifically relevant to a particular need, but it is often

a lengthy and expensive process characterized by repeated failure [716]. Drug repurposing therefore tends to be emphasized in a situation like the COVID-19 pandemic due to the potential for a more rapid response.

Even from the early months of the pandemic, studies began releasing results from analyses of approved and investigational drugs in the context of COVID-19. The rapid timescale of this response meant that, initially, most evidence came from observational studies, which compare groups of patients who did and did not receive a treatment to determine whether it may have had an effect. This type of study can be conducted rapidly but is subject to confounding. In contrast, randomized controlled trials (RCTs) are the gold-standard method for assessing the effects of an intervention. Here, patients are prospectively and randomly assigned to treatment or control conditions, allowing for much stronger interpretations to be drawn; however, data from these trials take much longer to collect. Both approaches have proven to be important sources of information in the development of a rapid response to the COVID-19 crisis, but as the pandemic draws on and more results become available from RCTs, more definitive answers are becoming available about proposed therapeutics. Interventional clinical trials are currently investigating or have investigated a large number of possible therapeutics and combinations of therapeutics for the treatment of COVID-19 (Figure 4).

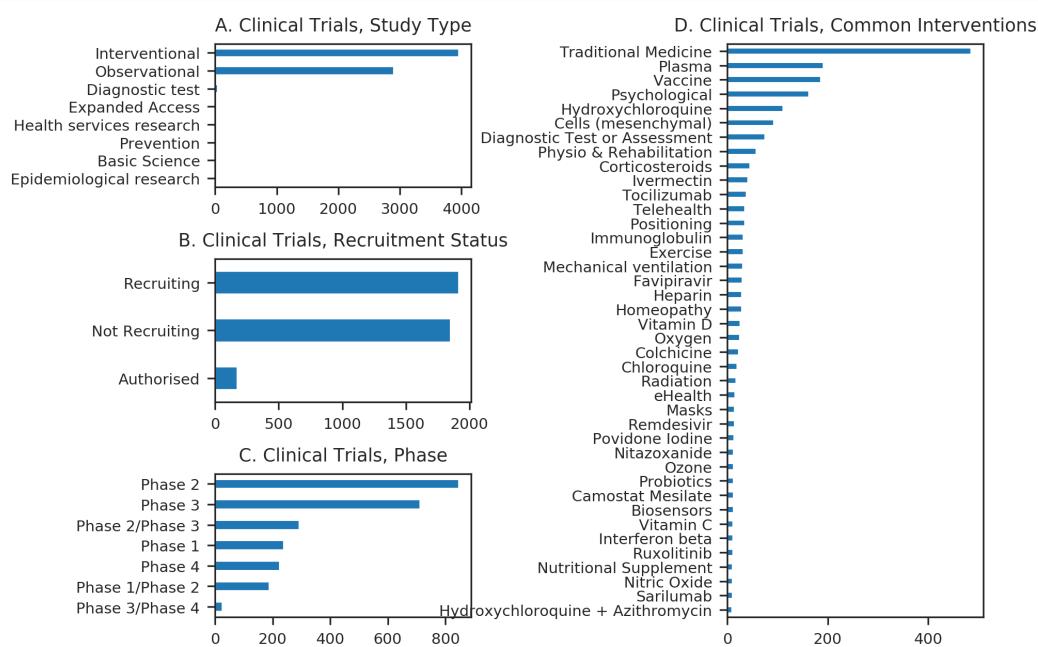


Figure 4: COVID-19 clinical trials. Trials data are from the University of Oxford Evidence-Based Medicine Data Lab's COVID-19 TrialsTracker [717]. As of December 31, 2020, there were 6,987 COVID-19 clinical trials of which 3,962 were interventional. The study types include only types used in at least five trials. Only interventional trials are analyzed in the figures depicting status, phase, and intervention. Of the interventional trials, 98 trials had reported results as of December 31, 2020. Recruitment status and trial phase are shown only for interventional trials in which the status or phase is recorded. Common interventions refers to interventions used in at least ten trials. Combinations of interventions, such as hydroxychloroquine with azithromycin, are tallied separately from the individual interventions. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

The purpose of this review is to provide an evolving resource tracking the status of efforts to repurpose and develop drugs for the treatment of COVID-19. We highlight four strategies that provide different paradigms for the

identification of potential pharmaceutical treatments. The WHO guidelines [79] and a systematic review [718] are complementary living documents that summarize COVID-19 therapeutics.

4.4 Repurposing Drugs for Symptom Management

A variety of symptom profiles with a range of severity are associated with COVID-19 [1]. In many cases, COVID-19 is not life threatening. A study of COVID-19 patients in a hospital in Berlin, Germany reported that the highest risk of death was associated with infection-related symptoms, such as sepsis, respiratory symptoms such as ARDS, and cardiovascular failure or pulmonary embolism [719]. Similarly, an analysis in Wuhan, China reported that respiratory failure (associated with ARDS) and sepsis/multi-organ failure accounted for 69.5% and 28.0% of deaths, respectively, among 82 deceased patients [720]. COVID-19 is characterized by two phases. The first is the acute response, where an adaptive immune response to the virus is established and in many cases can mitigate viral damage to organs [721]. The second phase characterizes more severe cases of COVID-19. Here, patients experience a cytokine storm, whereby excessive production of cytokines floods into circulation, leading to systemic inflammation, immune dysregulation, and multiorgan dysfunction that can cause multiorgan failure and death if untreated [722]. ARDS-associated respiratory failure can occur during this phase. Cytokine dysregulation was also identified in patients with SARS [723,724].

In the early days of the COVID-19 pandemic, physicians sought to identify potential treatments that could benefit patients, and in some cases shared their experiences and advice with the medical community on social media sites such as Twitter [725]. These on-the-ground treatment strategies could later be analyzed retrospectively in observational studies or investigated in an interventional paradigm through RCTs. Several notable cases involved the use of small-molecule drugs, which are synthesized compounds of low molecular weight, typically less than 1 kilodalton (kDa) [726]. Small-molecule pharmaceutical agents have been a backbone of drug development since the discovery of penicillin in the early twentieth century [727]. It and other antibiotics have long been among the best known applications of small molecules to therapeutics, but biotechnological developments such as the prediction of protein-protein interactions (PPIs) have facilitated advances in precise targeting of specific structures using small molecules [727]. Small molecule drugs today encompass a wide range of therapeutics beyond antibiotics, including antivirals, protein inhibitors, and many broad-spectrum pharmaceuticals.

Many treatments considered for COVID-19 have relied on a broad-spectrum approach. These treatments do not specifically target a virus or particular host receptor, but rather induce broad shifts in host biology that are hypothesized to be potential inhibitors of the virus. This approach relies on the fact that when a virus enters a host, the host becomes the virus's environment. Therefore, the state of the host can also influence the virus's ability to replicate and spread. The administration and assessment of broad-spectrum small-molecule drugs on a rapid time course was feasible because

they are often either available in hospitals, or in some cases may also be prescribed to a large number of out-patients. One of the other advantages is that these well-established compounds, if found to be beneficial, are often widely available, in contrast to boutique experimental drugs.

In some cases, prior data was available from experiments examining the response of other HCoVs or HCoV infections to a candidate drug. In addition to non-pharmaceutical interventions such as encouraging non-intubated patients to adopt a prone position [728], knowledge about interactions between HCoVs and the human body, many of which emerged from SARS and MERS research over the past two decades, led to the suggestion that a number of common drugs might benefit COVID-19 patients. However, the short duration and low case numbers of prior outbreaks were less well-suited to the large-scale study of clinical applications than the COVID-19 pandemic is. As a result, COVID-19 has presented the first opportunity to robustly evaluate treatments that were common during prior HCoV outbreaks to determine their clinical efficacy. The first year of the COVID-19 pandemic demonstrated that there are several different trajectories that these clinically suggested, widely available candidates can follow when assessed against a widespread, novel viral threat.

One approach to identifying candidate small molecule drugs was to look at the approaches used to treat SARS and MERS. Treatment of SARS and MERS patients prioritized supportive care and symptom management [326]. Among the clinical treatments for SARS and MERS that were explored, there was generally a lack of evidence indicating whether they were effective. Most of the supportive treatments for SARS were found inconclusive in meta-analysis [729], and a 2004 review reported that not enough evidence was available to make conclusions about most treatments [730]. However, one strategy adopted from prior HCoV outbreaks is currently the best-known treatment for severe cases of COVID-19. Corticosteroids represent broad-spectrum treatments and are a well-known, widely available treatment for pneumonia [731,732,733,734,735,736] that have also been debated as a possible treatment for ARDS [737,738,739,740,741,742]. Corticosteroids were also used and subsequently evaluated as possible supportive care for SARS and MERS. In general, studies and meta-analyses did not identify support for corticosteroids to prevent mortality in these HCoV infections [743,744,745]; however, one found that the effects might be masked by variability in treatment protocols, such as dosage and timing [730]. While the corticosteroids most often used to treat SARS were methylprednisolone and hydrocortisone, availability issues for these drugs at the time led to dexamethasone also being used in North America [746].

Dexamethasone (9 α -fluoro-16 α -methylprednisolone) is a synthetic corticosteroid that binds to glucocorticoid receptors [747,748]. It functions as an anti-inflammatory agent by binding to glucocorticoid receptors with higher affinity than endogenous cortisol [749]. Dexamethasone and other steroids are widely available and affordable, and they are often used to treat community-acquired pneumonia [750] as well as chronic inflammatory conditions such as asthma, allergies, and rheumatoid arthritis [751,752,753]. Immunosuppressive drugs such as steroids are typically contraindicated in the setting of infection [754], but because COVID-19 results in hyperinflammation that appears to contribute to mortality via lung damage,

immunosuppression may be a helpful approach to treatment [156]. A clinical trial that began in 2012 recently reported that dexamethasone may improve outcomes for patients with ARDS [737], but a meta-analysis of a small amount of available data about dexamethasone as a treatment for SARS suggested that it may, in fact, be associated with patient harm [755].

However, the findings in SARS may have been biased by the fact that all of the studies examined were observational and a large number of inconclusive studies were not included [756]. The questions of whether and when to counter hyperinflammation with immunosuppression in the setting of COVID-19 (as in SARS [724]) was an area of intense debate, as the risks of inhibiting antiviral immunity needed to be weighed against the beneficial anti-inflammatory effects [757]. As a result, guidelines early in the pandemic typically recommended avoiding treating COVID-19 patients with corticosteroids such as dexamethasone [755].

Despite this initial concern, dexamethasone was evaluated as a potential treatment for COVID-19 (Appendix 1). Dexamethasone treatment comprised one arm of the multi-site Randomized Evaluation of COVID-19 Therapy (RECOVERY) trial in the United Kingdom [758]. This study found that the 28-day mortality rate was lower in patients receiving dexamethasone than in those receiving standard of care (SOC). However, this finding was driven by differences in mortality among patients who were receiving mechanical ventilation or supplementary oxygen at the start of the study. The report indicated that dexamethasone reduced 28-day mortality relative to SOC in patients who were ventilated (29.3% versus 41.4%) and among those who were receiving oxygen supplementation (23.3% versus 26.2%) at randomization, but not in patients who were breathing independently (17.8% versus 14.0%). These findings also suggested that dexamethasone may have reduced progression to mechanical ventilation, especially among patients who were receiving oxygen support at randomization. Other analyses have supported the importance of disease course in determining the efficacy of dexamethasone: additional results suggest greater potential for patients who have experienced symptoms for at least seven days and patients who were not breathing independently [759]. A meta-analysis that evaluated the results of the RECOVERY trial alongside trials of other corticosteroids, such as hydrocortisone, similarly concluded that corticosteroids may be beneficial to patients with severe COVID-19 who are receiving oxygen supplementation [760]. Thus, it seems likely that dexamethasone is useful for treating inflammation associated with immunopathy or cytokine release syndrome (CRS), which is a condition caused by detrimental overactivation of the immune system [1]. In fact, corticosteroids such as dexamethasone are sometimes used to treat CRS [761]. Guidelines were quickly updated to encourage the use of dexamethasone in severe cases [762], and this affordable and widely available treatment rapidly became a valuable tool against COVID-19 [763], with demand surging within days of the preprint's release [764].

4.5 Approaches Targeting the Virus

Therapeutics that directly target the virus itself hold the potential to prevent people infected with SARS-CoV-2 from developing potentially damaging symptoms (Figure 5). Such drugs typically fall into the broad category of antivirals. Antiviral therapies hinder the spread of a virus within the host,

rather than destroying existing copies of the virus, and these drugs can vary in their specificity to a narrow or broad range of viral targets. This process requires inhibiting the replication cycle of a virus by disrupting one of six fundamental steps [765]. In the first of these steps, the virus attaches to and enters the host cell through endocytosis. Then the virus undergoes uncoating, which is classically defined as the release of viral contents into the host cell. Next, the viral genetic material enters the nucleus where it gets replicated during the biosynthesis stage. During the assembly stage, viral proteins are translated, allowing new viral particles to be assembled. In the final step new viruses are released into the extracellular environment. Although antivirals are designed to target a virus, they can also impact other processes in the host and may have unintended effects. Therefore, these therapeutics must be evaluated for both efficacy and safety. As the technology to respond to emerging viral threats has also evolved over the past two decades, a number of candidate treatments have been identified for prior viruses that may be relevant to the treatment of COVID-19.

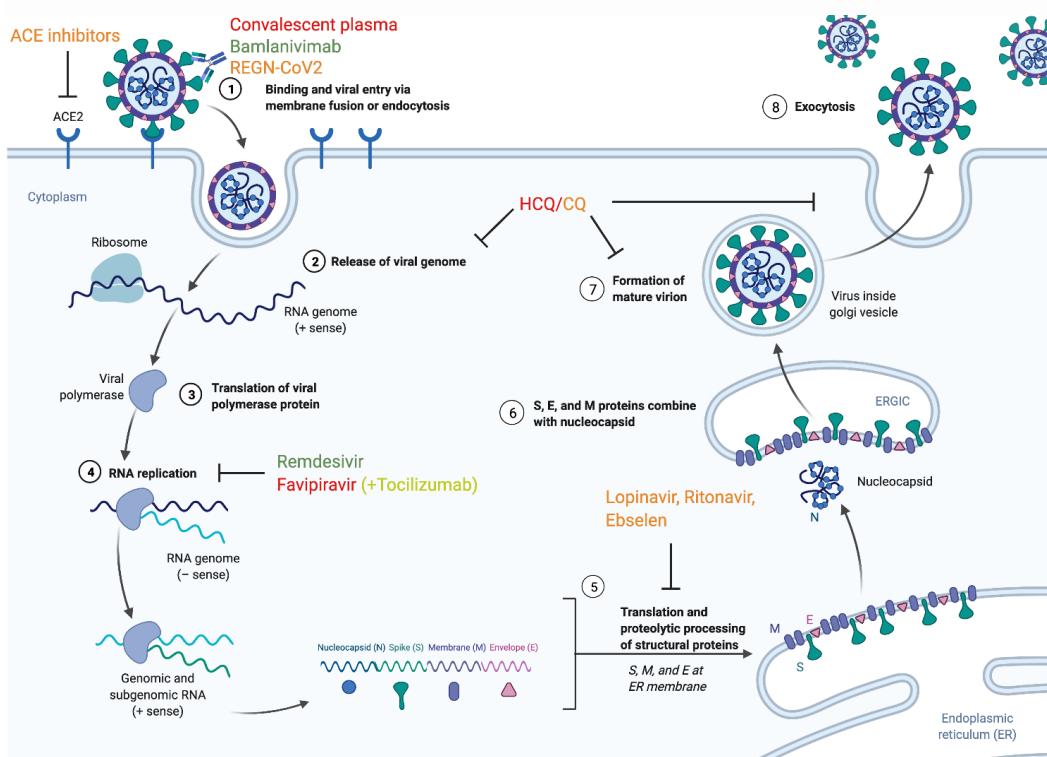


Figure 5: Mechanisms of Action for Potential Therapeutics Potential therapeutics currently being studied can target the SARS-CoV-2 virus or modify the host environment through many different mechanisms. Here, the relationships between the virus, host cells, and several therapeutics are visualized. Drug names are color-coded according to the grade assigned to them by the Center for Cytokine Storm Treatment & Laboratory's CORONA Project [766] (Green = A, Lime = B, Orange = C, and Red = D).

Many antiviral drugs are designed to inhibit the replication of viral genetic material during the biosynthesis step. Unlike DNA viruses, which can use the host enzymes to propagate themselves, RNA viruses like SARS-CoV-2 depend on their own polymerase, the RNA-dependent RNA polymerase (RdRP), for replication [767,768]. RdRP is therefore a potential target for antivirals against RNA viruses. Disruption of RdRP is the proposed mechanism underlying the treatment of SARS and MERS with ribavirin [769]. Ribavirin is an antiviral drug effective against other viral infections that was often used in combination with corticosteroids and sometimes interferon (IFN) medications to treat SARS and MERS [248]. However, analyses of its effects in

retrospective and *in vitro* analyses of SARS and the SARS-CoV-1 virus, respectively, have been inconclusive [248]. While IFNs and ribavirin have shown promise in *in vitro* analyses of MERS, their clinical effectiveness remains unknown [248]. The current COVID-19 pandemic has provided an opportunity to assess the clinical effects of these treatments. As one example, ribavarin was also used in the early days of COVID-19, but a retrospective cohort study comparing patients who did and did not receive ribivarain revealed no effect on the mortality rate [770].

Since nucleotides and nucleosides are the natural building blocks for RNA synthesis, an alternative approach has been to explore nucleoside and nucleotide analogs for their potential to inhibit viral replication. Analogs containing modifications to nucleotides or nucleosides can disrupt key processes including replication [771]. A single incorporation does not influence RNA transcription; however, multiple events of incorporation lead to the arrest of RNA synthesis [772]. One candidate antiviral considered for the treatment of COVID-19 is favipiravir (Avigan), also known as T-705, which was discovered by Toyama Chemical Co., Ltd. [773]. It was previously found to be effective at blocking viral amplification in several influenza subtypes as well as other RNA viruses, such as *Flaviviridae* and *Picornaviridae*, through a reduction in plaque formation [774] and viral replication in Madin-Darby canine kidney cells [775]. Favipiravir (6-fluoro-3-hydroxy-2-pyrazinecarboxamide) acts as a purine and purine nucleoside analogue that inhibits viral RNA polymerase in a dose-dependent manner across a range of RNA viruses, including influenza viruses [776,777,778,779,780]. Biochemical experiments showed that favipiravir was recognized as a purine nucleoside analogue and incorporated into the viral RNA template. In 2014, the drug was approved in Japan for the treatment of influenza that was resistant to conventional treatments like neuraminidase inhibitors [781]. Though initial analyses of favipiravir in observational studies of its effects on COVID-19 patients were promising, recent results of two small RCTs suggest that it is unlikely to affect COVID-19 outcomes (Appendix 1).

In contrast, another nucleoside analog, remdesivir, is one of the few treatments against COVID-19 that has received FDA approval. Remdesivir (GS-5734) is an intravenous antiviral that was proposed by Gilead Sciences as a possible treatment for Ebola virus disease. It is metabolized to GS-441524, an adenosine analog that inhibits a broad range of polymerases and then evades exonuclease repair, causing chain termination [782,783,784]. Gilead received an emergency use authorization (EUA) for remdesivir from the FDA early in the pandemic (May 2020) and was later found to reduce mortality and recovery time in a double-blind, placebo-controlled, phase III clinical trial performed at 60 trial sites, 45 of which were in the United States [785,786,787,788]. Subsequently, the WHO Solidarity trial, a large-scale, open-label trial enrolling 11,330 adult inpatients at 405 hospitals in 30 countries around the world, reported no effect of remdesivir on in-hospital mortality, duration of hospitalization, or progression to mechanical ventilation [789]. Therefore, additional clinical trials of remdesivir in different patient pools and in combination with other therapies may be needed to refine its use in the clinic and determine the forces driving these differing results. Remdesivir offers proof of principle that SARS-CoV-2 can be targeted at the level of viral replication, since remdesivir targets the viral RNA polymerase at high potency. Identification of such candidates depends on

knowledge about the virological properties of a novel threat. However, the success and relative lack of success, respectively, of remdesivir and favipiravir underscore the fact that drugs with similar mechanisms will not always produce similar results in clinical trials.

4.6 Disrupting Host-Virus Interactions

4.6.1 Interrupting Viral Colonization of Cells

Some of the most widely publicized examples of efforts to repurpose drugs for COVID-19 are broad-spectrum, small-molecule drugs where the mechanism of action made it seem that the drug might disrupt interactions between SARS-CoV-2 and human host cells (Figure 5). However, the exact outcomes of such treatments are difficult to predict *a priori*, and there are several examples where early enthusiasm was not borne out in subsequent trials. One of the most famous examples of an analysis of whether a well-known medication could provide benefits to COVID-19 patients came from the assessment of chloroquine (CQ) and hydroxychloroquine (HCQ), which are used for the treatment and prophylaxis of malaria as well as the treatment of lupus erythematosus and rheumatoid arthritis in adults [790]. These drugs are lysosomotropic agents, meaning they are weak bases that can pass through the plasma membrane. It was thought that they might provide benefits against SARS-CoV-2 by interfering with the digestion of antigens within the lysosome and inhibiting CD4 T-cell stimulation while promoting the stimulation of CD8 T-cells [791]. These compounds also have anti-inflammatory properties [791] and can decrease the production of certain key cytokines involved in the immune response, including interleukin-6 (IL-6) and inhibit the stimulation of Toll-like receptors (TLR) and TLR signaling [791].

In vitro analyses reported that CQ inhibited cell entry of SARS-CoV-1 [792] and that both CQ and HCQ inhibited viral replication within cultured cells [793], leading to early hope that it might provide similar therapeutic or protective effects in patients. However, while the first publication on the clinical application of these compounds to the inpatient treatment of COVID-19 was very positive [794], it was quickly discredited [795]. Over the following months, extensive evidence emerged demonstrating that CQ and HCQ offered no benefits for COVID-19 patients and, in fact, carried the risk of dangerous side effects (Appendix 1). The nail in the coffin came when findings from the large-scale RECOVERY trial were released on October 8, 2020. This study enrolled 11,197 hospitalized patients whose physicians believed it would not harm them to participate and used a randomized, open-label design to study the effects of HCQ compared to standard of care (SOC) at 176 hospitals in the United Kingdom [796]. Rates of COVID-19-related mortality did not differ between the control and HCQ arms, but patients receiving HCQ were slightly more likely to die due to cardiac events. Patients who received HCQ also had a longer duration of hospitalization than patients receiving usual care and were more likely to progress to mechanical ventilation or death (as a combined outcome). As a result, enrollment in the HCQ arm of the RECOVERY trial was terminated early [797]. The story of CQ/HCQ therefore illustrates how initial promising *in vitro* analyses can fail to translate to clinical usefulness.

A similar story has arisen with the broad-spectrum, small-molecule anthelmintic ivermectin, which is a synthetic analog of avermectin, a bioactive compound produced by a microorganism known as *Streptomyces avermectiniius* and *Streptomyces avermitilis* [798,799]. Avermectin disrupts the ability of parasites to avoid the host immune response by blocking glutamate-gated chloride ion channels in the peripheral nervous system from closing, leading to hyperpolarization of neuronal membranes, disruption of neural transmission, and paralysis [798,800,801]. Ivermectin has been used since the early 1980s to treat endo- and ecto-parasitic infections by helminths, insects, and arachnids in veterinary contexts [798,802] and since the late 1980s to treat human parasitic infections as well [798,800]. More recent research has indicated that ivermectin might function as a broad-spectrum antiviral by disrupting the trafficking of viral proteins by both RNA and DNA viruses [801,803,804], although most of these studies have demonstrated this effect *in vitro* [804]. The potential for antiviral effects on SARS-CoV-2 were investigated *in vitro*, and ivermectin was found to inhibit viral replication in a cell line derived from Vero cells (Vero-hSLAM) [805]. However, inhibition of viral replication was achieved at concentrations that were much higher than that explored by existing dosage guidelines [806,807], which are likely to be associated with significant side effects due to the increased potential that the compound could cross the mammalian blood-brain barrier [808,809].

Retrospective studies and small RCTs began investigating the effects of standard doses of this low-cost, widely available drug. One retrospective study reported that ivermectin reduced all-cause mortality [810] while another reported no difference in clinical outcomes or viral clearance [811]. Small RCTs enrolling less than 50 patients per arm have also reported a wide array of positive [812,813,814,815,816] and negative results [817,818]. A slightly larger RCT enrolling 115 patients in two arms reported inconclusive results [819]. Hope for the potential of ivermectin peaked with the release of a preprint reporting results of a multicenter, double-blind RCT where a four-day course of ivermectin was associated with clinical improvement and earlier viral clearance in 400 symptomatic patients and 200 close contacts [820]; however, concerns were raised about both the integrity of the data and the paper itself [821,822], and this study was removed by the preprint server Research Square [823]. A similarly sized RCT suggested no effect on the duration of symptoms among 400 patients split evenly across the intervention and control arms [824], and although meta-analyses have reported both null [825,826] and beneficial [827,828,829,830,831,832,833,834] effects of ivermectin on COVID-19 outcomes, the certainty is likely to be low [828]. These findings are potentially biased by a small number of low-quality studies, including the preprint that has been taken down [835], and the authors of one [836] have issued a notice [827] that they will revise their study with the withdrawn study removed. Thus, much like HCQ/CQ, enthusiasm for research that either has not or should not have passed peer review has led to large numbers of patients worldwide receiving treatments that might not have any effect or could even be harmful. Additionally, comments on the now-removed preprint include inquiries into how best to self-administer veterinary ivermectin as a prophylactic [823], and the FDA has posted information explaining why veterinary ivermectin should not be taken by humans concerned about COVID-19 [837]. Ivermectin is now one of several candidate therapeutics

being investigated in the large-scale TOGETHER [838] and PRINCIPLE [839] clinical trials. The TOGETHER trial, which previously demonstrated no effect of HCQ and lopinavir-ritonavir [840], released preliminary results in early August 2021 suggesting that ivermectin also has no effect on COVID-19 outcomes [841].

While CQ/HCQ and ivermectin are well-known medications that have long been prescribed in certain contexts, investigation of another well-established type of pharmaceutical was facilitated by the fact that it was already being taken by a large number of COVID-19 patients. Angiotensin-converting enzyme inhibitors (ACEIs) and angiotensin II receptor blockers (ARBs) are among today's most commonly prescribed medications, often being used to control blood pressure [842,843]. In the United States, for example, they are prescribed well over 100,000,000 times annually [844]. Prior to the COVID-19 pandemic, the relationship between ACE2, ACEIs, and SARS had been considered as possible evidence that ACE2 could serve as a therapeutic target [845], and the connection had been explored through *in vitro* and molecular docking analysis [846] but ultimately was not pursued clinically [847]. Data from some animal models suggest that several, but not all, ACEIs and several ARBs increase ACE2 expression in the cells of some organs [848], but clinical studies have not established whether plasma ACE2 expression is increased in humans treated with these medications [849]. In this case, rather than introducing ARBs/ACEIs, a number of analyses have investigated whether discontinuing use affects COVID-19 outcomes. An initial observational study of the association of exposure to ACEIs or ARBs with outcomes in COVID-19 was retracted from the *New England Journal of Medicine* [850] due to concerns related to data availability [851]. As RCTs have become available, they have demonstrated no effect of continuing versus discontinuing ARBs/ACEIs on patient outcomes [852,853] (Appendix 1). Thus, once again, despite a potential mechanistic association with the pathology of SARS-CoV-2 infection, these medications were not found to influence the trajectory of COVID-19 illness.

For medications that are widely known and common, clinical research into their efficacy against a novel threat can be developed very quickly. This feasibility can present a double-edged sword. For example, HCQ and CQ were incorporated into SOC in many countries early in the pandemic and had to be discontinued once their potential to harm COVID-19 patients became apparent [854,855]. Dexamethasone remains the major success story from this category of repurposed drugs and is likely to have saved a large number of lives since summer 2020 [763].

4.6.2 Manipulating the Host Immune Response

Treatments based on understanding a virus and/or how a virus interacts with the human immune system can fall into two categories: they can interact with the innate immune response, which is likely to be a similar response across viruses, or they can be specifically designed to imitate the adaptive immune response to a particular virus. In the latter case, conservation of structure or behavior across viruses enables exploring whether drugs developed for one virus can treat another. During the COVID-19 pandemic, a number of candidate therapeutics have been explored in these categories, with varied success.

Knowledge gained from trying to understand SARS-CoV-1 and MERS-CoV from a fundamental biological perspective and characterize how they interact with the human immune system provides a theoretical basis for identifying candidate therapies. Biologics are a particularly important class of drugs for efforts to address HCoV through this paradigm. They are produced from components of living organisms or viruses, historically primarily from animal tissues, but have become increasingly feasible to produce as recombinant technologies have advanced [856].

There are many differences on the development side between biologics and synthesized pharmaceuticals, such as small molecule drugs. Typically, biologics are orders of magnitude larger than small molecule drugs and are catabolized by the body to their amino acid components [857]. They are often heat sensitive, and their toxicity can vary, as it is not directly associated with the primary effects of the drug; in general, their physiochemical properties are much less understood compared to small molecules [857]. Biologics include significant medical breakthroughs such as insulin for the management of diabetes and vaccines, as well monoclonal antibodies (mAbs) and interferons (IFNs), which can be used to target the host immune response after infection.

mAbs have revolutionized the way we treat human diseases and have become some of the best-selling drugs in the pharmaceutical market in recent years [858]. There are currently 79 FDA approved mAbs on the market, including antibodies for viral infections (e.g. Ibalizumab for *Human immunodeficiency virus* and Palivizumab for *Respiratory syncytial virus*) [858,859]. Virus-specific neutralizing antibodies commonly target viral surface glycoproteins or host structures, thereby inhibiting viral entry through receptor binding interference [860,861]. This interference is predicted to reduce the viral load, mitigate disease, and reduce overall hospitalization. mAbs can be designed for a particular virus, and significant advances have been made in the speed at which new mAbs can be identified and produced. At the time of the SARS and MERS epidemics, interest in mAbs to reduce infection was never realized [862,863], but this allowed for mAbs to quickly be considered among the top candidates against COVID-19.

4.6.2.1 Biologics and the Innate Immune Response

Deaths from COVID-19 often occur when inflammation becomes dysregulated following an immune response to the SARS-CoV-2 virus. Therefore, one potential approach to reducing COVID-19 mortality rates is to manage the inflammatory response in severely ill patients. One candidate therapeutic identified that uses this mechanism is tocilizumab (TCZ). TCZ is a mAb that was developed to manage chronic inflammation caused by the continuous synthesis of the cytokine IL-6 [864]. IL-6 is a pro-inflammatory cytokine belonging to the interleukin family, which is comprised by immune system regulators that are primarily responsible for immune cell differentiation. Often used to treat chronic inflammatory conditions such as rheumatoid arthritis [864], TCZ has become a pharmaceutical of interest for the treatment of COVID-19 because of the role IL-6 plays in this disease. It has also been approved to treat CRS caused by CAR-T treatments [865]. While the secretion of IL-6 can be associated with chronic conditions, IL-6 is a key player in the innate immune response and is secreted by macrophages in

response to the detection of pathogen-associated molecular patterns and damage-associated molecular patterns [864]. An analysis of 191 in-patients at two Wuhan hospitals revealed that blood concentrations of IL-6 differed between patients who did and did not recover from COVID-19. Patients who ultimately died had higher IL-6 levels at admission than those who recovered [81]. Additionally, IL-6 levels remained higher throughout the course of hospitalization in the patients who ultimately died [81].

Currently, TCZ is being administered either as a monotherapy or in combination with other treatments in 73 interventional COVID-19 clinical trials (Figure 4). A number of retrospective studies have been conducted in several countries [866,867,868,869,870,871]. In general, these studies have reported a positive effect of TCZ on reducing mortality in COVID-19 patients, although due to their retrospective designs, significant limitations are present in all of them (Appendix 1). It was not until February 11, 2021 that a preprint describing preliminary results of the first RCT of TCZ was released as part of the RECOVERY trial [872]. TCZ was found to reduce 28-day mortality from 33% in patients receiving SOC alone to 29% in those receiving TCZ. Combined analysis of the RECOVERY trial data with data from smaller RCTs suggested a 13% reduction in 28-day mortality [872]. While this initial report did not include the full results expected from the RECOVERY trial, this large-scale, RCT provides strong evidence that TCZ may offer benefits for COVID-19 patients. The RECOVERY trial along with results from several other RCTs [873,874,875,876,877] were cited as support for the EUA issued for TCZ in June 2021 [878]. However, the fact that TCZ suppresses the immune response means that it does carry risks for patients, especially a potential risk of secondary infection (Appendix 1).

TCZ is just one example of a candidate drug targeting the host immune response and specifically excessive inflammation. For example, interferons (IFNs) have also been investigated; these are a family of cytokines critical to activating the innate immune response against viral infections. Synairgen has been investigating a candidate drug, SNG001, which is an IFN- β -1a formulation to be delivered to the lungs via inhalation [879] that they reported reduced progression to ventilation in a double-blind, placebo-controlled, multi-center study of 101 patients with an average age in the late 50s [880,881]. However, these findings were not supported by the large-scale WHO Solidarity trial, which reported no significant effect of IFN- β -1a on patient survival during hospitalization [789], although differences in the designs of the two studies, and specifically the severity of illness among enrolled patients, may have influenced their divergent outcomes (Appendix 1). Other biologics influencing inflammation are also being explored (Appendix 1). It is also important that studies focused on inflammation as a possible therapeutic target consider the potential differences in baseline inflammation among patients from different backgrounds, which may be caused by differing life experiences (see [325]).

4.6.2.2 Biologics and the Adaptive Immune Response

While TCZ is an example of an mAb focused on managing the innate immune response, other treatments are more specific, targeting the adaptive immune response after an infection. In some cases, treatments can utilize biologics obtained directly from recovered individuals. From the very early days of the

COVID-19 pandemic, polyclonal antibodies from convalescent plasma were investigated as a potential treatment for COVID-19 [882,883]. Convalescent plasma was used in prior epidemics including SARS, Ebola Virus Disease, and even the 1918 Spanish Influenza [882,884]. Use of convalescent plasma transfusion (CPT) over more than a century has aimed to reduce symptoms and improve mortality in infected people [884], possibly by accelerating viral clearance [882]. However, it seems unlikely that this classic treatment confers any benefit for COVID-19 patients. Several systematic reviews have investigated whether CPT reduced mortality in COVID-19 patients, and although findings from early in the pandemic (up to April 19, 2020) did support use of CPT [884], the tide has shifted as the body of available literature has grown [885]. While titer levels were suggested as a possible determining factor in the success of CPT against COVID-19 [886], the large-scale RECOVERY trial evaluated the effect of administering high-titer plasma specifically and found no effect on mortality or hospital discharge over a 28-day period [887]. These results thus suggest that, despite initial optimism and an EUA from the FDA, CPT is unlikely to be an effective therapeutic for COVID-19.

A different narrative is shaping up around the use of mAbs specifically targeting SARS-CoV-2. During the first SARS epidemic in 2002, neutralizing antibodies (nAbs) were found in SARS-CoV-1-infected patients [888,889]. Several studies following up on these findings identified various S-glycoprotein epitopes as the major targets of nAbs against SARS-CoV-1 [890]. Coronaviruses use trimeric spike (S) glycoproteins on their surface to bind to the host cell, allowing for cell entry [23,31]. Each S glycoprotein protomer is comprised of an S1 domain, also called the receptor binding domain (RBD), and an S2 domain. The S1 domain binds to the host cell while the S2 domain facilitates the fusion between the viral envelope and host cell membranes [890]. The genomic identity between the RBD of SARS-CoV-1 and SARS-CoV-2 is around 74% [891]. Due to this high degree of similarity, preexisting antibodies against SARS-CoV-1 were initially considered candidates for neutralizing activity against SARS-CoV-2. While some antibodies developed against the SARS-CoV-1 spike protein showed cross-neutralization activity with SARS-CoV-2 [892,893], others failed to bind to SARS-CoV-2 spike protein at relevant concentrations [14]. Cross-neutralizing activities were dependent on whether the epitope recognized by the antibodies were conserved between SARS-CoV-1 and SARS-CoV-2 [892].

Technological advances in antibody drug design as well as in structural biology massively accelerated the discovery of novel antibody candidates and the mechanisms by which they interact with the target structure. Within just a year of the structure of the SARS-CoV-2 spike protein being published, an impressive pipeline of monoclonal antibodies targeting SARS-CoV-2 entered clinical trials, with hundreds more candidates in preclinical stages. The first human monoclonal neutralizing antibody specifically against the SARS-CoV-2 S glycoprotein was developed using hybridoma technology [894], where antibody-producing B-cells developed by mice are inserted into myeloma cells to produce a hybrid cell line (the hybridoma) that is grown in culture. The 47D11 antibody clone was able to cross-neutralize SARS-CoV-1 and SARS-CoV-2. This antibody (now ABVV-47D11) has recently entered clinical trials in collaboration with AbbVie. Additionally, an extensive monoclonal neutralizing antibody pipeline has been developed to combat the ongoing pandemic, with

over 50 different antibodies in clinical trials [895]. Thus far, the monotherapy sotrovimab and two antibody cocktails (bamlanivimab/estesevimab and casirivimab/imdevimab) have been granted EUAs by the FDA.

One of the studied antibody cocktails consists of bamlanivimab and estesevimab. Bamlanivimab (Ly-CoV555) is a human mAb that was derived from convalescent plasma donated by a recovered COVID-19 patient, evaluated in research by the National Institute of Allergy and Infectious Diseases (NIAID), and subsequently developed by AbCellera and Eli Lilly. The neutralizing activity of bamlanivimab was initially demonstrated *in vivo* using a nonhuman primate model [896]. Based on these positive preclinical data, Eli Lilly initiated the first human clinical trial for a monoclonal antibody against SARS-CoV-2. The phase 1 trial, which was conducted in hospitalized COVID-19 patients, was completed in August 2020 [897]. Estesevimab (LY-CoV016 or JS-016) is also a monoclonal neutralizing antibody against the spike protein of SARS-CoV-2. It was initially developed by Junshi Biosciences and later licensed and developed through Eli Lilly. A phase 1 clinical trial to assess the safety of etesevimab was completed in October 2020 [898]. Etesevimab was shown to bind a different epitope on the spike protein than bamlanivimab, suggesting that the two antibodies used as a combination therapy would further enhance their clinical use compared to a monotherapy [899]. To assess the efficacy and safety of bamlanivimab alone or in combination with etesevimab for the treatment of COVID-19, a phase 2/3 trial (BLAZE-1) [900] was initiated. The interim analysis of the phase 2 portion suggested that bamlanivimab alone was able to accelerate the reduction in viral load [901]. However, more recent data suggests that only the bamlanivimab/etesevimab combination therapy is able to reduce viral load in COVID-19 patients [899]. Based on this data, the combination therapy received an EUA for COVID-19 from the FDA in February 2021 [902].

A second therapy is comprised of casirivimab and imdevimab (REGN-COV2). Casirivimab (REGN10933) and imdevimab (REGN10987) are two monoclonal antibodies against the SARS-CoV-2 spike protein. They were both developed by Regeneron in a parallel high-throughput screening (HTS) to identify neutralizing antibodies from either humanized mice or patient-derived convalescent plasma [903]. In these efforts, multiple antibodies were characterized for their ability to bind and neutralize the SARS-CoV-2 spike protein. The investigators hypothesized that an antibody cocktail, rather than each individual antibody, could increase the therapeutic efficacy while minimizing the risk for virus escape. Therefore, the authors tested pairs of individual antibodies for their ability to simultaneously bind the RBD of the spike protein. Based on this data, casirivimab and imdevimab were identified as the lead antibody pair, resulting in the initiation of two clinical trials [904,905]. Data from this phase 1-3 trial published in the *New England Journal of Medicine* shows that the REGN-COV2 antibody cocktail reduced viral load, particularly in patients with high viral load or whose endogenous immune response had not yet been initiated [906]. However, in patients who already initiated an immune response, exogenous addition of REGN-COV2 did not improve the endogenous immune response. Both doses were well tolerated with no serious events related to the antibody cocktail. Based on this data, the FDA granted an EUA for REGN-COV2 in patients with mild to

moderate COVID-19 who are at risk of developing severe disease [907]. Ongoing efforts are trying to evaluate the efficacy of REGN-COV2 to improve clinical outcomes in hospitalized patients [904].

Sotrovimab is the most recent mAb to receive an EUA. It was identified in the memory B cells of a 2003 survivor of SARS [908] and was found to be cross-reactive with SARS-CoV-2 [893]. This cross-reactivity is likely attributable to conservation within the epitope, with 17 out of 22 residues conserved between the two viruses, four conservatively substituted, and one semi-conservatively substituted [893]. In fact, these residues are highly conserved among sarbecoviruses, a clade that includes SARS-CoV-1 and SARS-CoV-2 [893]. This versatility has led to it being characterized as a “super-antibody” [909], a potent, broadly neutralizing antibody [910]. Interim analysis of data from a clinical trial [911] reported high safety and efficacy of this mAb in 583 COVID-19 patients [912]. Compared to placebo, sotrovimab was found to be 85% more effective in reducing progression to the primary endpoint, which was the proportion of patients who, within 29 days, were either hospitalized for more than 24 hours or died. Additionally, rates of adverse events were comparable, and in some cases lower, among patients receiving sotrovimab compared to patients receiving a placebo. Sotrovimab therefore represents a mAb therapeutic that is effective against SARS-CoV-2 and may also be effective against other sarbecoviruses.

Several potential limitations remain in the application of mAbs to the treatment of COVID-19. One of the biggest challenges is identifying antibodies that not only bind to their target, but also prove to be beneficial for disease management. Currently, use of mAbs is limited to people with mild to moderate disease that are not hospitalized, and it has yet to be determined whether they can be used as a successful treatment option for severe COVID-19 patients. While preventing people from developing severe illness provides significant benefits, patients with severe illness are at the greatest risk of death, and therefore therapeutics that provide benefits against severe illness are particularly desirable. It remains to be seen whether mAbs confer any benefits for patients in this category.

Another concern about therapeutics designed to amplify the response to a specific viral target is that they may need to be modified as the virus evolves. With the ongoing global spread of new SARS-CoV-2 variants, there is a growing concern that mutations in SARS-CoV-2 spike protein could escape antibody neutralization, thereby reducing the efficacy of monoclonal antibody therapeutics and vaccines. A comprehensive mutagenesis screen recently identified several amino acid substitutions in the SARS-CoV-2 spike protein that can prevent antibody neutralization [913]. While some mutations result in resistance to only one antibody, others confer broad resistance to multiple mAbs as well as polyclonal human sera, suggesting that some amino acids are “hotspots” for antibody resistance. However, it was not investigated whether the resistance mutations identified result in a fitness advantage. Accordingly, an impact on neutralizing efficiency has been reported for the B.1.1.7 (Alpha) variant first identified in the UK and the B.1.351 (Beta) variant first identified in South Africa [914,915,916]. As of June 25, 2021, the CDC recommended a pause in the use of bamlanivimab and etesevimab due to decreased efficacy against the P.1 (Gamma) and B.1.351 (Beta) variants of SARS-CoV-2 [917]. While the reported impact on antibody neutralization

needs to be confirmed *in vivo*, it suggests that some adjustments to therapeutic antibody treatments may be necessary to maintain the efficacy that was reported in previous clinical trials.

Several strategies have been employed to try to mitigate the risk of diminished antibody neutralization. Antibody cocktails such as those already holding an EUA may help overcome the risk for attenuation of the neutralizing activity of a single monoclonal antibody. These cocktails consist of antibodies that recognize different epitopes on the spike protein, decreasing the likelihood that a single amino acid change can cause resistance to all antibodies in the cocktail. However, neutralizing resistance can emerge even against an antibody cocktail if the individual antibodies target subdominant epitopes [915]. Another strategy is to develop broadly neutralizing antibodies that target structures that are highly conserved, as these are less likely to mutate [918,919] or to target epitopes that are insensitive to mutations [920]. Sotrovimab, one such “super-antibody”, is thought to be somewhat robust to neutralization escape [921] and has been found to be effective against all variants assessed as of August 12, 2021 [922]. Another antibody (ADG-2) targets a highly conserved epitope that overlaps the hACE2 binding site of all clade 1 sarbecoviruses [923]. Prophylactic administration of ADG-2 in an immunocompetent mouse model of COVID-19 resulted in protection against viral replication in the lungs and respiratory burden. Since the epitope targeted by ADG-2 represents an Achilles’ heel for clade 1 sarbecoviruses, this antibody, like sotrovimab, might be a promising candidate against all circulating variants as well as emerging SARS-related coronaviruses. To date, it has fared well against the Alpha, Beta, Gamma, and Delta variants [922].

The development of mAbs against SARS-CoV-2 has made it clear that this technology is rapidly adaptable and offers great potential for the response to emerging viral threats. However, additional investigation may be needed to adapt mAb treatments to SARS-CoV-2 as it evolves and potentially to pursue designs that confer benefits for patients at the greatest risk of death. While polyclonal antibodies from convalescent plasma have been evaluated as a treatment for COVID-19, these studies have suggested fewer potential benefits against SARS-CoV-2 than mAbs; convalescent plasma therapy has been thoroughly reviewed elsewhere [882,883]. Thus, advances in biologics for COVID-19 illustrate that an understanding of how the host and virus interact can guide therapeutic approaches. The FDA authorization of two combination mAb therapies, in particular, underscores the potential for this strategy to allow for a rapid response to a novel pathogen. Additionally, while TCZ is not yet as established, this therapy suggests that the strategy of using biologics to counteract the cytokine storm response may provide therapies for the highest-risk patients.

4.7 High-Throughput Screening for Drug Repurposing

The drug development process is slow and costly, and developing compounds specifically targeted to an emerging viral threat is not a practical short-term solution. Screening existing drug compounds for alternative indications is a popular alternative [924,925,926,927]. HTS has been a goal of

pharmaceutical development since at least the mid-1980s [928]. Traditionally, phenotypic screens were used to test which compounds would induce a desired change in *in vitro* or *in vivo* models, focusing on empirical, function-oriented exploration naïve to molecular mechanism [929,930,931]. In many cases, these screens utilize large libraries that encompass a diverse set of agents varying in many pharmacologically relevant properties (e.g., [932]). The compounds inducing a desired effect could then be followed up on. Around the turn of the millennium, advances in molecular biology allowed for HTS to shift towards screening for compounds interacting with a specific molecular target under the hypothesis that modulating that target would have a desired effect. These approaches both offer pros and cons, and today a popular view is that they are most effective in combination [929,931,933].

Today, some efforts to screen compounds for potential repurposing opportunities are experimental, but others use computational HTS approaches [924,934]. Computational drug repurposing screens can take advantage of big data in biology [715] and as a result are much more feasible today than during the height of the SARS and MERS outbreaks in the early 2000s and early 2010s, respectively. Advancements in robotics also facilitate the experimental component of HTS [926]. For viral diseases, the goal of drug repurposing is typically to identify existing drugs that have an antiviral effect likely to impede the virus of interest. While both small molecules and biologics can be candidates for repurposing, the significantly lower price of many small molecule drugs means that they are typically more appealing candidates [935].

Depending on the study design, screens vary in how closely they are tied to a hypothesis. As with the candidate therapeutics described above, high-throughput experimental or computational screens can proceed based on a hypothesis. Just as remdesivir was selected as a candidate antiviral because it is a nucleoside analog [936], so too can high-throughput screens select libraries of compounds based on a molecular hypothesis. Likewise, when the library of drugs is selected without basis in a potential mechanism, a screen can be considered hypothesis free [936]. Today, both types of analyses are common both experimentally and computationally. Both strategies have been applied to identifying candidate therapeutics against SARS-CoV-2.

4.7.1 Hypothesis-Driven Screening

Hypothesis-driven screens often select drugs likely to interact with specific viral or host targets or drugs with desired clinical effects, such as immunosuppressants. There are several properties that might identify a compound as a candidate for an emerging viral disease. Drugs that interact with a target that is shared between pathogens (i.e., a viral protease or a polymerase) or between a viral pathogen and another illness (i.e., a cancer drug with antiviral potential) are potential candidates, as are drugs that are thought to interact with additional molecular targets beyond those they were developed for [934]. Such research can be driven by *in vitro* or *in silico* experimentation. Computational analyses depend on identifying compounds that modulate pre-selected proteins in the virus or host. As a result, they build on experimental research characterizing the molecular features of the virus, host, and candidate compounds [927].

One example of the application of this approach to COVID-19 research comes from work on protease inhibitors. Studies have shown that viral proteases play an important role in the life cycle of viruses, including coronaviruses, by modulating the cleavage of viral polyprotein precursors [937]. Several FDA-approved drugs target proteases, such as lopinavir and ritonavir for HIV infection and simeprevir for hepatitis C virus infection. Serine protease inhibitors were previously suggested as possible treatments for SARS and MERS [938]. One early study [31] suggested that camostat mesylate, a protease inhibitor, could block the entry of SARS-CoV-2 into lung cells *in vitro*. Two polyproteins encoded by the SARS-CoV-2 replicase gene, pp1a and pp1ab, are critical for viral replication and transcription [939]. These polyproteins must undergo proteolytic processing, which is usually conducted by main protease (M^{Pro}), a 33.8-kDa SARS-CoV-2 protease that is therefore fundamental to viral replication and transcription. Therefore, it was hypothesized that compounds targeting M^{Pro} could be used to prevent or slow the replication of the SARS-CoV-2 virus.

Both computational and experimental approaches facilitated the identification of compounds that might inhibit SARS-CoV-2 M^{Pro} . In 2005, computer-aided design facilitated the development of a Michael acceptor inhibitor, now known as N3, to target M^{Pro} of SARS-like coronaviruses [940]. N3 binds in the substrate binding pocket of M^{Pro} in several HCoV [940,941,942,943]. The structure of N3-bound SARS-CoV-2 M^{Pro} has been solved, confirming the computational prediction that N3 would similarly bind in the substrate binding pocket of SARS-CoV-2 [939]. N3 was tested *in vitro* on SARS-CoV-2-infected Vero cells, which belong to a line of cells established from the kidney epithelial cells of an African green monkey, and was found to inhibit SARS-CoV-2 [939]. A library of approximately 10,000 compounds was screened in a fluorescence resonance energy transfer assay constructed using SARS-CoV-2 M^{Pro} expressed in *Escherichia coli* [939].

Six leads were identified in this hypothesis-driven screen. *In vitro* analysis revealed that ebselen had the strongest potency in reducing the viral load in SARS-CoV-2-infected Vero cells [939]. Ebselen is an organoselenium compound with anti-inflammatory and antioxidant properties [944]. Molecular dynamics analysis further demonstrated the potential for ebselen to bind to M^{Pro} and disrupt the protease's enzymatic functions [945]. However, ebselen is likely to be a promiscuous binder, which could diminish its therapeutic potential [939,946], and compounds with higher specificity may be needed to translate this mechanism effectively to clinical trials. In July 2020, phase II clinical trials commenced to assess the effects of SPI-1005, an investigational drug from Sound Pharmaceuticals that contains ebselen [947], on 60 adults presenting with each of moderate [948] and severe [949] COVID-19. Other M^{Pro} inhibitors are also being evaluated in clinical trials [950,951,951]. Pending the results of clinical trials, N3 remains a computationally interesting compound based on both computational and experimental data, but whether these potential effects will translate to the clinic remains unknown.

4.7.2 Hypothesis-Free Screening

Hypothesis-free screens use a discovery-driven approach, where screens are not targeted to specific viral proteins, host proteins, or desired clinical modulation. Hypothesis-free drug screening began twenty years ago with the testing of libraries of drugs experimentally. Today, like many other areas of biology, *in silico* analyses have become increasingly popular and feasible through advances in biological big data [936,952]. Many efforts have collected data about interactions between drugs and SARS-CoV-2 and about the host genomic response to SARS-CoV-2 exposure, allowing for hypothesis-free computational screens that seek to identify new candidate therapeutics. Thus, they utilize a systems biology paradigm to extrapolate the effect of a drug against a virus based on the host interactions with both the virus and the drug [927].

Resources such as the COVID-19 Drug and Gene Set Library, which at the time of its publication contained 1,620 drugs sourced from 173 experimental and computational drug sets and 18,676 human genes sourced from 444 gene sets [953], facilitate such discovery-driven approaches. Analysis of these databases indicated that some drugs had been identified as candidates across multiple independent analyses, including high-profile candidates such as CQ/HCQ and remdesivir [953]. Computational screening efforts can then mine databases and other resources to identify potential PPIs among the host, the virus, and established and/or experimental drugs [954]. Subject matter expertise from human users may be integrated to varying extents depending on the platform (e.g., [954,955]). These resources have allowed studies to identify potential therapeutics for COVID-19 without an *a priori* reason for selecting them.

One example of a hypothesis-free screen for COVID-19 drugs comes from a PPI-network-based analysis that was published early in the pandemic [191]. Here, researchers cloned the proteins expressed by SARS-CoV-2 *in vitro* and quantified 332 viral-host PPI using affinity purification mass spectrometry [191]. They identified two SARS-CoV-2 proteins (Nsp6 and Orf9c) that interacted with host Sigma-1 and Sigma-2 receptors. Sigma receptors are located in the endoplasmic reticulum of many cell types, and type 1 and 2 Sigma receptors have overlapping but distinct affinities for a variety of ligands [956]. Molecules interacting with the Sigma receptors were then analyzed and found to have an effect on viral infectivity *in vitro* [191]. A follow-up study evaluated the effect of perturbing these 332 proteins in two cell lines, A549 and Caco-2, using knockdown and knockout methods, respectively, and found that the replication of SARS-CoV-2 in cells from both lines was dependent on the expression of *SIGMAR1*, which is the gene that encodes the Sigma-1 receptor [957]. Following these results, drugs interacting with Sigma receptors were suggested as candidates for repurposing for COVID-19 (e.g., [958]). Because many well-known and affordable drugs interact with the Sigma receptors [191,959], they became a major focus of drug repurposing efforts. Some of the drugs suggested by the apparent success of Sigma receptor-targeting drugs were already being investigated at the time. HCQ, for example, forms ligands with both Sigma-1 and Sigma-2 receptors and was already being explored as a candidate therapeutic for COVID-19 [191]. Thus, this computational approach yielded interest in drugs whose antiviral activity was supported by initial *in vitro* analyses.

Follow-up research, however, called into question whether the emphasis on drugs interacting with Sigma receptors might be based on a spurious association [960]. This study built on the prior work by examining whether antiviral activity among compounds correlated with their affinity for the Sigma receptors and found that it did not. The study further demonstrated that cationic amphiphilicity was a shared property among many of the candidate drugs identified through both computational and phenotypic screens and that it was likely to be the source of many compounds' proposed antiviral activity [960]. Cationic amphiphilicity is associated with the induction of phospholipidosis, which is when phospholipids accumulate in the lysosome [961]. Phospholipidosis can disrupt viral replication by inhibiting lipid processing [962] (see discussion of HCQ in Appendix 1). However, phospholipidosis is known to translate poorly from *in vitro* models to *in vivo* models or clinical applications. Thus, this finding suggested that these screens were identifying compounds that shared a physiochemical property rather than a specific target [960]. The authors further demonstrated that antiviral activity against SARS-CoV-2 *in vitro* was correlated with the induction of phospholipidosis for drugs both with and without cationic amphiphilicity [960]. This finding supports the idea that the property of cationic amphiphilicity was being detected as a proxy for the shared effect of phospholipidosis [960]. They demonstrated that phospholipidosis-inducing drugs were not effective at preventing viral propagation *in vivo* in a murine model of COVID-19 [960]. Therefore, removing hits that induce phospholipidosis from computational and *in vitro* experimental repurposing screens (e.g., [963]) may help emphasize those that are more likely to provide clinical benefits. This work illustrates the importance of considering confounding variables in computational screens, a principle that has been incorporated into more traditional approaches to drug development [964].

One drug that acts on Sigma receptors does, however, remain a candidate for the treatment of COVID-19. Several psychotropic drugs target Sigma receptors in the central nervous system and thus attracted interest as potential COVID-19 therapeutics following the findings of two host-virus PPI studies [965]. For several of these drugs, the *in vitro* antiviral activity [957] was not correlated with their affinity for the Sigma-1 receptor [960,965] but was correlated with phospholipidosis [960]. However, fluvoxamine, a selective serotonin reuptake inhibitor that is a particularly potent Sigma-1 receptor agonist [965], has shown promise as a preventative of severe COVID-19 in a preliminary analysis of data from the large-scale TOGETHER trial [841]. As of August 6, 2021, this trial had collected data from over 1,400 patients in the fluvoxamine arm of their study, half of whom received a placebo [841]. Only 74 patients in the fluvoxamine group had progressed to hospitalization for COVID-19 compared to 107 in the placebo group, corresponding to a relative risk of 0.69; additionally, the relative risk of mortality between the two groups was calculated at 0.71. These findings support the results of small clinical trials that have found fluvoxamine to reduce clinical deterioration relative to a placebo [966,967]. However, the ongoing therapeutic potential of fluvoxamine does not contradict the finding that hypothesis-free screening hits can be driven by confounding factors. The authors point out that its relevance would not just be antiviral as it has a potential immunomodulatory mechanism [966]. It has been found to be protective against septic shock in an *in vivo* mouse model [968]. It is possible that fluvoxamine also exerts an antiviral effect [969]. Thus, Sigma-1 receptor

activity may contribute to fluvoxamine's potential effects in treating COVID-19, but is not the only mechanism by which this drug can interfere with disease progression.

4.7.3 Potential and Limitations of High-Throughput Analyses

Computational screening allows for a large number of compounds to be evaluated to identify those most likely to display a desired behavior or function. This approach can be guided by a hypothesis or can aim to discover underlying characteristics that produce new hypotheses about the relationship between a host, a virus, and candidate pharmaceuticals. The examples outlined above illustrate that HTS-based evaluations of drug repurposing can potentially provide valuable insights. Computational techniques were used to design compounds targeting M^{Pro} based on an understanding of how this protease aids viral replication, and M^{Pro} inhibitors remain promising candidates [926], although the clinical trial data is not yet available. Similarly, computational analysis correctly identified the Sigma-1 receptor as a protein of interest. Although the process of identifying which drugs might modulate the interaction led to an emphasis on candidates that ultimately have not been supported, fluvoxamine remains an appealing candidate. The difference between the preliminary evidence for fluvoxamine compared to other drugs that interact with Sigma receptors underscores a major critique of hypothesis-free HTS in particular: while these approaches allow for brute force comparison of a large number of compounds against a virus of interest, they lose the element of expertise that is associated with most successes in drug repurposing [936].

There are also practical limitations to these methods. One concern is that computational analyses inherently depend on the quality of the data being evaluated. The urgency of the COVID-19 pandemic led many research groups to pivot towards computational HTS research without familiarity with best practices in this area [926]. As a result, there is an excessive amount of information available from computational studies [970], but not all of it is high-quality. Additionally, the literature used to identify and validate targets can be difficult to reproduce [971], which may pose challenges to target-based experimental screening and to *in silico* screens. Some efforts to repurpose antivirals have focused on host, rather than viral, proteins [927], which might be expected to translate poorly *in vivo* if the targeted proteins serve essential functions in the host. Concerns about the practicality of hypothesis-free screens to gain novel insights are underscored by the fact that very few or possibly no success stories have emerged from hypothesis-free screens over the past twenty years [936]. These findings suggest that data-driven research can be an important component of the drug repurposing ecosystem, but that drug repurposing efforts that proceed without a hypothesis, an emphasis on biological mechanisms, or an understanding of confounding effects may not produce viable candidates.

4.8 Considerations in Balancing Different Approaches

The approaches described here offer a variety of advantages and limitations in responding to a novel viral threat and building on existing bodies of knowledge in different ways. Medicine, pharmacology, basic science (especially virology and immunology), and biological data science can all provide different insights and perspectives for addressing the challenging question of which existing drugs might provide benefits against an emerging viral threat. A symptom management-driven approach allows clinicians to apply experience with related diseases or related symptoms to organize a rapid response aimed at saving the lives of patients already infected with a new disease. Oftentimes, the pharmaceutical agents that are applied are small-molecule, broad-spectrum pharmaceuticals that are widely available and affordable to produce, and they may already be available for other purposes, allowing clinicians to administer them to patients quickly either with an EUA or off-label. In this vein, dexamethasone has emerged as the strongest treatment against severe COVID-19 (Table 1).

Alternatively, many efforts to repurpose drugs for COVID-19 have built on information gained through basic scientific research of HCoV. Understanding how related viruses function has allowed researchers to identify possible pharmacological strategies to disrupt pathogenesis (Figure 5). Some of the compounds identified through these methods include small-molecule antivirals, which can be boutique and experimental medications like remdesivir (Table 1). Other candidate drugs that intercept host-pathogen interactions include biologics, which imitate the function of endogenous host compounds. Most notably, several mAbs that have been developed (casirivimab, imdevimab, bamlanivimab and etesevimab) or repurposed (sotrovimab, tocilizumab) have now been granted EUAs (Table 1). Although not discussed here, several vaccine development programs have also met huge success using a range of strategies [328].

Table 1: Summary table of candidate therapeutics examined in this manuscript. “Grade” is the rating given to each treatment by the Systematic Tracker of Off-label/Repurposed Medicines Grades (STORM) maintained by the Center for Cytokine Storm Treatment & Laboratory (CSTL) at the University of Pennsylvania [766]. A grade of A indicates that a treatment is considered effective, B that all or most RCTs have shown positive results, C that RCT data are not yet available, and D that multiple RCTs have produced negative results. Treatments not in the STORM database are indicated as N/A. FDA status is also provided where available. The evidence available is based on the progression of the therapeutic through the pharmaceutical development pipeline, with RCTs as the most informative source of evidence. The effectiveness is summarized based on the current available evidence; large trials such as RECOVERY and Solidarity are weighted heavily in this summary. This table was last updated on August 20, 2021.

Treatment	Grade	Category	FDA Status	Evidence Available	Suggested Effectiveness
Dexamethasone	A	Small molecule, broad spectrum	Used off-label	RCT	Supported: RCT shows improved outcomes over SOC, especially in severe cases such as CRS

Treatment	Grade	Category	FDA Status	Evidence Available	Suggested Effectiveness
Remdesivir	A	Small molecule, antiviral, adenosine analog	Approved for COVID-19 (and EUA for combination with baricitinib)	RCT	Mixed: Conflicting evidence from large WHO-led Solidarity trial vs US-focused RCT and other studies
Tocilizumab	A	Biologic, monoclonal antibody	EUA	RCT	Mixed: It appears that TCZ may work well in combination with dexamethasone in severe cases, but not as monotherapy
Sotrovimab	N/A	Biologic, monoclonal antibody	EUA	RCT	Supported: Phase 2/3 clinical trial showed reduced hospitalization/death
Bamlanivimab and etesevimab	B & N/A	Biologic, monoclonal antibodies	EUA	RCT	Supported: Phase 2 clinical trial showed reduction in viral load, but FDA pause recommended because may be less effective against Delta variant
Casirivimab and imdevimab	N/A	Biologic, monoclonal antibodies	EUA	RCT	Supported: Reduced viral load at interim analysis
Fluvoxamine	B	Small-molecule, Sigma-1 receptor agonist	N/A	RCT	Supported: Support from two small RCTs and preliminary support from interim analysis of TOGETHER
SNG001	B	Biologic, interferon	None	RCT	Mixed: Support from initial RCT but no effect found in WHO's Solidarity trial
M ^{Pro} Protease Inhibitors	N/A	Small molecule, protease inhibitor	None	Computational prediction, <i>in vitro</i> studies	Unknown

Treatment	Grade	Category	FDA Status	Evidence Available	Suggested Effectiveness
ARBs & ACEIs	C	Small molecule, broad spectrum	None	Observational studies and some RCTs	Not supported: Observational study retracted, RCTs suggest no association
Favipiravir	D	Small molecule, antiviral, nucleoside analog	None	RCT	Not supported: RCTs do not show significant improvements for individuals taking this treatment, good safety profile
HCQ/CQ	D	Small molecule, broad spectrum	None	RCT	Not supported, possibly harmful: Non-blinded RCTs showed no improvement over SOC, safety profile may be problematic
Convalescent plasma transfusion	D	Biologic, polyclonal antibodies	EUA	RCT	Mixed: Supported in small trials but not in large-scale RECOVERY trial
Ivermectin	D	Small molecule, broad spectrum	None	RCT	Mixed: Mixed results from small RCTs, major supporting RCT now withdrawn, preliminary results of large RCT (TOGETHER) suggest no effect on emergency room visits or hospitalization for COVID-19

All of the small-molecule drugs evaluated and most of the biologics are repurposed, and thus hinge on a theoretical understanding of how the virus interacts with a human host and how pharmaceuticals can be used to modify those interactions rather than being designed specifically against SARS-CoV-2 or COVID-19. As a result, significant attention has been paid to computational approaches that automate the identification of potentially desirable interactions. However, work in COVID-19 has made it clear that relevant compounds can also be masked by confounds, and spurious associations can drive investment in candidate therapeutics that are unlikely to translate to the clinic. Such spurious hits are especially likely to impact hypothesis-free screens. However, hypothesis-free screens may still be able to contribute to the drug discovery or repurposing ecosystem, assuming the computational arm of HTS follows the same trends seen in its experimental arm. In 2011, a landmark study in drug discovery demonstrated that although more new drugs were discovered using target-based rather than phenotypic approaches, the majority of drugs with a novel molecular mechanism of action (MMOA) were identified in phenotypic screens [972]. This pattern

applied only to first-in-class drugs, with most follower drugs produced by target-based screening [930]. These findings suggest that target-based drug discovery is more successful when building on a known MMOA, and that modulating a target is most valuable when the target is part of a valuable MMOA [931]. Building on this, many within the field suggested that mechanism-informed phenotypic investigations may be the most useful approach to drug discovery [929,931,933]. As it stands, data-driven efforts to identify patterns in the results of computational screens allowed researchers to notice the shared property of cationic amphiphility among many of the hits from computational screening analyses [960]. While easier said than done, efforts to fill in the black box underlying computational HTS and recognize patterns among the identified compounds aid in moving data-oriented drug repurposing efforts in this direction.

The unpredictable nature of success and failure in drug repurposing for COVID-19 thus highlights one of the tenets of phenotypic screening: there are a lot of “unknown unknowns”, and a promising mechanism at the level of an MMOA will not necessarily propagate up to the pathway, cellular, or organismal level [929]. Despite the fact that apparently mechanistically relevant drugs may exist, identifying effective treatments for a new viral disease is extremely challenging. Targets of repurposed drugs are often non-specific, meaning that the MMOA can appear to be relevant to COVID-19 without a therapeutic or prophylactic effect being observed in clinical trials. The difference in the current status of remdesivir and favipiravir as treatments for COVID-19 (Table 1) underscores how difficult to predict whether a specific compound will produce a desired effect, even when the mechanisms are similar. Furthermore, the fact that many candidate COVID-19 therapeutics were ultimately identified because of their shared propensity to induce phospholipidosis underscores how challenging it can be to identify a mechanism *in silico* or *in vitro* that will translate to a successful treatment. While significant progress has been made thus far in the pandemic, the therapeutic landscape is likely to continue to evolve as more results become available from clinical trials and as efforts to develop novel therapeutics for COVID-19 progress.

4.9 Towards the Next HCoV Threat

Only very limited testing of candidate therapies was feasible during the SARS and MERS epidemics, and as a result, few treatments were available at the outset of the COVID-19 pandemic. Even corticosteroids, which were used to treat SARS patients, were a controversial therapeutic prior to the release of the results of the large RECOVERY trial. The scale and duration of the COVID-19 pandemic has made it possible to conduct large, rigorous RCTs such as RECOVERY, Solidarity, TOGETHER, and others. As results from these trials have continued to emerge, it has become clear that small clinical trials often produce spurious results. In the case of HCQ/CQ, the therapeutic had already attracted so much attention based on small, preliminary (and in some cases, methodologically concerning) studies that it took the results of multiple large studies before attention began to be redirected to more promising candidates [973]. In fact, most COVID-19 clinical trials lack the statistical power to reliably test their hypotheses [974,975]. In the face of an urgent crisis like COVID-19, the desire to act quickly is understandable, but it is imperative that studies maintain strict standards of scientific rigor [926,964],

especially given the potential dangers of politicization, as illustrated by HCQ/CQ [976]. Potential innovations in clinical trial structure, such as adaptable clinical trials with master protocols [977] or the sharing of data among small clinical trials [975] may help to address future crises and to bolster the results from smaller studies, respectively.

In the long-term, new drugs specific for treatment of COVID-19 may also enter development. Development of novel drugs is likely to be guided by what is known about the pathogenesis and molecular structure of SARS-CoV-2. For example, understanding the various structural components of SARS-CoV-2 may allow for the development of small molecule inhibitors of those components. Crystal structures of the SARS-CoV-2 main protease have been resolved [939,978]. Much work remains to be done to determine further crystal structures of other viral components, understand the relative utility of targeting different viral components, perform additional small molecule inhibitor screens, and determine the safety and efficacy of the potential inhibitors. While still nascent, work in this area is promising. Over the longer term, this approach and others may lead to the development of novel therapeutics specifically for COVID-19 and SARS-CoV-2. Such efforts are likely to prove valuable in managing future emergent HCoV, just as research from the SARS and MERS pandemic has provided a basis for the COVID-19 response.

5 Appendix: Identification and Development of Therapeutics for COVID-19

5.1 Dexamethasone

In order to understand how dexamethasone reduces inflammation, it is necessary to consider the stress response broadly. In response to stress, corticotropin-releasing hormone stimulates the release of neurotransmitters known as catecholamines, such as epinephrine, and steroid hormones known as glucocorticoids, such as cortisol [979,980]. While catecholamines are often associated with the fight-or-flight response, the specific role that glucocorticoids play is less clear, although they are thought to be important to restoring homeostasis [981]. Immune challenge is a stressor that is known to interact closely with the stress response. The immune system can therefore interact with the central nervous system; for example, macrophages can both respond to and produce catecholamines [979]. Additionally, the production of both catecholamines and glucocorticoids is associated with inhibition of proinflammatory cytokines such as IL-6, IL-12, and tumor necrosis factor- α (TNF- α) and the stimulation of anti-inflammatory cytokines such as IL-10, meaning that the stress response can regulate inflammatory immune activity [980]. Administration of dexamethasone has been found to correspond to dose-dependent inhibition of IL-12 production, but not to affect IL-10 [982]; the fact that this relationship could be disrupted by administration of a glucocorticoid-receptor antagonist suggests that it is regulated by the receptor itself [982]. Thus, the administration of dexamethasone for COVID-19 is likely to simulate the release of

glucocorticoids endogenously during stress, resulting in binding of the synthetic steroid to the glucocorticoid receptor and the associated inhibition of the production of proinflammatory cytokines. In this model, dexamethasone reduces inflammation by stimulating the biological mechanism that reduces inflammation following a threat such as immune challenge.

Initial support for dexamethasone as a treatment for COVID-19 came from the United Kingdom's RECOVERY trial [758], which assigned over 6,000 hospitalized COVID-19 patients to the standard of care (SOC) or treatment (dexamethasone) arms of the trial at a 2:1 ratio. At the time of randomization, some patients were ventilated (16%), others were on non-invasive oxygen (60%), and others were breathing independently (24%). Patients in the treatment arm were administered dexamethasone either orally or intravenously at 6 mg per day for up to 10 days. The primary endpoint was the patient's status at 28-days post-randomization (mortality, discharge, or continued hospitalization), and secondary outcomes analyzed included the progression to invasive mechanical ventilation over the same period. The 28-day mortality rate was found to be lower in the treatment group than in the SOC group (21.6% vs 24.6%, $p < 0.001$). However, the effect was driven by improvements in patients receiving mechanical ventilation or supplementary oxygen. One possible confounder is that patients receiving mechanical ventilation tended to be younger than patients who were not receiving respiratory support (by 10 years on average) and to have had symptoms for a longer period. However, adjusting for age did not change the conclusions, although the duration of symptoms was found to be significantly associated with the effect of dexamethasone administration. Thus, this large, randomized, and multi-site, albeit not placebo-controlled, study suggests that administration of dexamethasone to patients who are unable to breathe independently may significantly improve survival outcomes. Additionally, dexamethasone is a widely available and affordable medication, raising the hope that it could be made available to COVID-19 patients globally.

It is not surprising that administration of an immunosuppressant would be most beneficial in severe cases where the immune system was dysregulated towards inflammation. However, it is also unsurprising that care must be taken in administering an immunosuppressant to patients fighting a viral infection. In particular, the concern has been raised that treatment with dexamethasone might increase patient susceptibility to concurrent (e.g., nosocomial) infections [983]. Additionally, the drug could potentially slow viral clearance and inhibit patients' ability to develop antibodies to SARS-CoV-2 [755,983], with the lack of data about viral clearance being put forward as a major limitation of the RECOVERY trial [984]. Furthermore, dexamethasone has been associated with side effects that include psychosis, glucocorticoid-induced diabetes, and avascular necrosis [755], and the RECOVERY trial did not report outcomes with enough detail to be able to determine whether they observed similar complications. The effects of dexamethasone have also been found to differ among populations, especially in high-income versus middle- or low-income countries [985]. However, since the RECOVERY trial's results were released, strategies have been proposed for administering dexamethasone alongside more targeted treatments to minimize the

likelihood of negative side effects [983]. Given the available evidence, dexamethasone is currently the most promising treatment for severe COVID-19.

5.2 Favipiravir

The effectiveness of favipiravir for treating patients with COVID-19 is currently under investigation. Evidence for the drug inhibiting viral RNA polymerase are based on time-of-drug addition studies that found that viral loads were reduced with the addition of favipiravir in early times post-infection [776,779,780]. An open-label, nonrandomized, before-after controlled study for COVID-19 was recently conducted [986]. The study included 80 COVID-19 patients (35 treated with favipiravir, 45 control) from the isolation ward of the National Clinical Research Center for Infectious Diseases (The Third People's Hospital of Shenzhen), Shenzhen, China. The patients in the control group were treated with other antivirals, such as lopinavir and ritonavir. It should be noted that although the control patients received antivirals, two subsequent large-scale analyses, the WHO Solidarity trial and the Randomized Evaluation of COVID-19 Therapy (RECOVERY) trial, identified no effect of lopinavir or of a lopinavir-ritonavir combination, respectively, on the metrics of COVID-19-related mortality that each assessed [789,987,988]. Treatment was applied on days 2-14; treatment stopped either when viral clearance was confirmed or at day 14. The efficacy of the treatment was measured by, first, the time until viral clearance using Kaplan-Meier survival curves, and, second, the improvement rate of chest computed tomography (CT) scans on day 14 after treatment. The study found that favipiravir increased the speed of recovery, measured as viral clearance from the patient by RT-PCR, with patients receiving favipiravir recovering in four days compared to 11 days for patients receiving antivirals such as lopinavir and ritonavir. Additionally, the lung CT scans of patients treated with favipiravir showed significantly higher improvement rates (91%) on day 14 compared to control patients (62%, $p = 0.004$). However, there were adverse side effects in 4 (11%) favipiravir-treated patients and 25 (56%) control patients. The adverse side effects included diarrhea, vomiting, nausea, rash, and liver and kidney injury. Despite the study reporting clinical improvement in favipiravir-treated patients, several study design issues are problematic and lower confidence in the overall conclusions. For example, the study was neither randomized nor blinded. Moreover, the selection of patients did not take into consideration important factors such as previous clinical conditions or sex, and there was no age categorization. Additionally, it should be noted that this study was temporarily retracted and then restored without an explanation [989].

In late 2020 and early 2021, the first randomized controlled trials of favipiravir for the treatment of COVID-19 released results [990,991,992]. One study [991] was retracted in November 2021 due to concerns about the data. Of the two remaining, the first [990] used a randomized, controlled, open-label design to compare two drugs, favipiravir and baloxavir marboxil, to SOC alone. Here, SOC included antivirals such as lopinavir/ritonavir and was administered to all patients. The primary endpoint analyzed was viral clearance at day 14. The sample size for this study was very small, with 29 total patients enrolled, and no significant effect of the treatments was found for the primary or any of the secondary outcomes analyzed, which included

mortality. The second trial examined 60 patients and reported a significant effect of favipiravir on viral clearance at four days (a secondary endpoint), but not at 10 days (the primary endpoint) [992]. This study, as well as a prior study of favipiravir [993], also reported that the drug was generally well-tolerated. Thus, in combination, these small studies suggest that the effects of favipiravir as a treatment for COVID-19 cannot be determined based on the available evidence, but additionally, none raise major concerns about the safety profile of the drug.

5.3 Remdesivir

At the outset of the COVID-19 pandemic, remdesivir did not have any have any FDA-approved use. A clinical trial in the Democratic Republic of Congo found some evidence of effectiveness against ebola virus disease (EVD), but two antibody preparations were found to be more effective, and remdesivir was not pursued [994]. Remdesivir also inhibits polymerase and replication of the coronaviruses MERS-CoV and SARS-CoV-1 in cell culture assays with submicromolar IC₅₀s [995]. It has also been found to inhibit SARS-CoV-2, showing synergy with CQ *in vitro* [784].

Remdesivir was first used on some COVID-19 patients under compassionate use guidelines [998]. All were in late stages of COVID-19 infection, and initial reports were inconclusive about the drug's efficacy. Gilead Sciences, the maker of remdesivir, led a recent publication that reported outcomes for compassionate use of the drug in 61 patients hospitalized with confirmed COVID-19. Here, 200 mg of remdesivir was administered intravenously on day 1, followed by a further 100 mg/day for 9 days [788]. There were significant issues with the study design, or lack thereof. There was no randomized control group. The inclusion criteria were variable: some patients only required low doses of oxygen, while others required ventilation. The study included many sites, potentially with variable inclusion criteria and treatment protocols. The patients analyzed had mixed demographics. There was a short follow-up period of investigation. Eight patients were excluded from the analysis mainly due to missing post-baseline information; thus, their health was unaccounted for. Therefore, even though the study reported clinical improvement in 68% of the 53 patients ultimately evaluated, due to the significant issues with study design, it could not be determined whether treatment with remdesivir had an effect or whether these patients would have recovered regardless of treatment. Another study comparing 5- and 10-day treatment regimens reported similar results but was also limited because of the lack of a placebo control [999]. These studies did not alter the understanding of the efficacy of remdesivir in treating COVID-19, but the encouraging results provided motivation for placebo-controlled studies.

The double-blind placebo-controlled ACTT-1 trial [785,786] recruited 1,062 patients and randomly assigned them to placebo treatment or treatment with remdesivir. Patients were stratified for randomization based on site and the severity of disease presentation at baseline [785]. The treatment was 200 mg on day 1, followed by 100 mg on days 2 through 10. Data was analyzed from a total of 1,059 patients who completed the 29-day course of the trial, with 517 assigned to remdesivir and 508 to placebo [785]. The two groups were well matched demographically and clinically at baseline. Those who

received remdesivir had a median recovery time of 10 days, as compared with 15 days in those who received placebo (rate ratio for recovery, 1.29; 95% confidence interval (CI), 1.12 to 1.49; $p < 0.001$). The Kaplan-Meier estimates of mortality by 14 days were 6.7% with remdesivir and 11.9% with placebo, with a hazard ratio (HR) for death of 0.55 and a 95% CI of 0.36 to 0.83, and at day 29, remdesivir corresponded to 11.4% and the placebo to 15.2% (HR: 0.73; 95% CI, 0.52 to 1.03). Serious adverse events were reported in 131 of the 532 patients who received remdesivir (24.6%) and in 163 of the 516 patients in the placebo group (31.6%). This study also reported an association between remdesivir administration and both clinical improvement and a lack of progression to more invasive respiratory intervention in patients receiving non-invasive and invasive ventilation at randomization [785]. Largely on the results of this trial, the FDA reissued and expanded the EUA for remdesivir for the treatment of hospitalized COVID-19 patients ages twelve and older [1000]. Additional clinical trials [784,1001,1002,1003,1004] are currently underway to evaluate the use of remdesivir to treat COVID-19 patients at both early and late stages of infection and in combination with other drugs (Figure 4). As of October 22, 2020, remdesivir received FDA approval based on three clinical trials [1005].

However, results suggesting no effect of remdesivir on survival were reported by the WHO Solidarity trial [789]. Patients were randomized in equal proportions into four experimental conditions and a control condition, corresponding to four candidate treatments for COVID-19 and SOC, respectively; no placebo was administered. The 2,750 patients in the remdesivir group were administered 200 mg intravenously on the first day and 100 mg on each subsequent day until day 10 and assessed for in-hospital death (primary endpoint), duration of hospitalization, and progression to mechanical ventilation. There were also 2,708 control patients who would have been eligible and able to receive remdesivir were they not assigned to the control group. A total of 604 patients among these two cohorts died during initial hospitalization, with 301 in the remdesivir group and 303 in the control group. The rate ratio of death between these two groups was therefore not significant (0.95, $p = 0.50$), suggesting that the administration of remdesivir did not affect survival. The two secondary analyses similarly did not find any effect of remdesivir. Additionally, the authors compared data from their study with data from three other studies of remdesivir (including [785]) stratified by supplemental oxygen status. A meta-analysis of the four studies yielded an overall rate ratio for death of 0.91 ($p = 0.20$). These results thus do not support the previous findings that remdesivir reduced median recovery time and mortality risk in COVID-19 patients.

In response to the results of the Solidarity trial, Gilead, which manufactures remdesivir, released a statement pointing to the fact that the Solidarity trial was not placebo-controlled or double-blind and at the time of release, the statement had not been peer reviewed [1006]; these sentiments have been echoed elsewhere [1007]. Other critiques of this study have noted that antivirals are not typically targeted at patients with severe illness, and therefore remdesivir could be more beneficial for patients with mild rather than severe cases [988,1008]. However, the publication associated with the trial sponsored by Gilead did purport an effect of remdesivir on patients with severe disease, identifying an 11 versus 18 day recovery period (rate ratio for

recovery: 1.31, 95% CI 1.12 to 1.52) [785]. Additionally, a smaller analysis of 598 patients, of whom two-thirds were randomized to receive remdesivir for either 5 or 10 days, reported a small effect of treatment with remdesivir for five days relative to standard of care in patients with moderate COVID-19 [1009]. These results suggest that remdesivir could improve outcomes for patients with moderate COVID-19, but that additional information would be needed to understand the effects of different durations of treatment.

Therefore, the Solidarity trial may point to limitations in the generalizability of other research on remdesivir, especially since the broad international nature of the Solidarity clinical trial, which included countries with a wide range of economic profiles and a variety of healthcare systems, provides a much-needed global perspective in a pandemic [988]. On the other hand, only 62% of patients in the Solidarity trial were randomized on the day of admission or one day afterwards [789], and concerns have been raised that differences in disease progression could influence the effectiveness of remdesivir [988].

Despite the findings of the Solidarity trial, remdesivir remains available for the treatment of COVID-19 in many places. Remdesivir has also been investigated in combination with other drugs, such as baricitinib, which is an inhibitor of Janus kinase 1 and 2 [1010]; the FDA has issued an EUA for the combination of remdesivir and baricitinib in adult and pediatric patients [1011]. Follow-up studies are needed and, in many cases, are underway to further investigate remdesivir-related outcomes.

Similarly, the extent to which the remdesivir dosing regimen could influence outcomes continues to be under consideration. A randomized, open-label trial compared the effect of remdesivir on 397 patients with severe COVID-19 over 5 versus 10 days [787,999], complementing the study that found that a 5-day course of remdesivir improved outcomes for patients with moderate COVID-19 but a 10-day course did not [1009]. Patients in the two groups were administered 200 mg of remdesivir intravenously on the first day, followed by 100 mg on the subsequent four or nine days, respectively. The two groups differed significantly in their clinical status, with patients assigned to the 10-day group having more severe illness. This study also differed from most because it included not only adults, but also pediatric patients as young as 12 years old. It reported no significant differences across several outcomes for patients receiving a 5-day or 10-day course, when correcting for baseline clinical status. The data did suggest that the 10-day course might reduce mortality in the most severe patients at day 14, but the representation of this group in the study population was too low to justify any conclusions [999]. Thus, additional research is also required to determine whether the dosage and duration of remdesivir administration influences outcomes.

In summary, remdesivir is the first FDA approved anti-viral against SARS-CoV-2 as well as the first FDA approved COVID-19 treatment. Early investigations of this drug established proof of principle that drugs targeting the virus can benefit COVID-19 patients. Moreover, one of the most successful strategies for developing therapeutics for viral diseases is to target the viral replication machinery, which are typically virally encoded polymerases. Small molecule drugs targeting viral polymerases are the backbones of treatments for other viral diseases including human immunodeficiency virus (HIV) and herpes. Notably, the HIV and herpes polymerases are a reverse transcriptase and a DNA polymerase, respectively, whereas SARS-CoV-2 encodes an RdRP, so most of the commonly used polymerase inhibitors are not likely to be active

against SARS-CoV-2. In clinical use, polymerase inhibitors show short term benefits for HIV patients, but for long term benefits they must be part of combination regimens. They are typically combined with protease inhibitors, integrase inhibitors, and even other polymerase inhibitors. Remdesivir provides evidence that a related approach may be beneficial for the treatment of COVID-19.

5.4 Hydroxychloroquine and Chloroquine

CQ and hydroxychloroquine (HCQ) increase cellular pH by accumulating in their protonated form inside lysosomes [791,1012]. This shift in pH inhibits the breakdown of proteins and peptides by the lysosomes during the process of proteolysis [791]. Interest in CQ and HCQ for treating COVID-19 was catalyzed by a mechanism observed in *in vitro* studies of both SARS-CoV-1 and SARS-CoV-2. In one study, CQ inhibited viral entry of SARS-CoV-1 into Vero E6 cells, a cell line that was derived from Vero cells in 1968, through the elevation of endosomal pH and the terminal glycosylation of ACE2 [792]. Increased pH within the cell, as discussed above, inhibits proteolysis, and terminal glycosylation of ACE2 is thought to interfere with virus-receptor binding. An *in vitro* study of SARS-CoV-2 infection of Vero cells found both HCQ and CQ to be effective in inhibiting viral replication, with HCQ being more potent [793]. Additionally, an early case study of three COVID-19 patients reported the presence of antiphospholipid antibodies in all three patients [98]. Antiphospholipid antibodies are central to the diagnosis of the antiphospholipid syndrome, a disorder that HCQ has often been used to treat [1013,1014,1015]. Because the 90% effective concentration (EC₉₀) of CQ in Vero E6 cells (6.90 µM) can be achieved in and tolerated by rheumatoid arthritis (RA) patients, it was hypothesized that it might also be possible to achieve the effective concentration in COVID-19 patients [1016]. Additionally, clinical trials have reported HCQ to be effective in treating HIV [1017] and chronic Hepatitis C [1018]. Together, these studies triggered initial enthusiasm about the therapeutic potential for HCQ and CQ against COVID-19. HCQ/CQ has been proposed both as a treatment for COVID-19 and a prophylaxis against SARS-CoV-2 exposure, and trials often investigated these drugs in combination with azithromycin (AZ) and/or zinc supplementation. However, as more evidence has emerged, it has become clear that HCQ/CQ offer no benefits against SARS-CoV-2 or COVID-19.

5.4.1 Trials Assessing Therapeutic Administration of HCQ/CQ

The initial study evaluating HCQ as a treatment for COVID-19 patients was published on March 20, 2020 by Gautret et al. [794]. This non-randomized, non-blinded, non-placebo clinical trial compared HCQ to SOC in 42 hospitalized patients in southern France. It reported that patients who received HCQ showed higher rates of virological clearance by nasopharyngeal swab on days 3-6 when compared to SOC. This study also treated six patients with both HCQ + AZ and found this combination therapy to be more effective than HCQ alone. However, the design and analyses used showed weaknesses that severely limit interpretability of results, including the small sample size and the lack of: randomization, blinding, placebo (no "placebo pill" given to SOC group), Intention-To-Treat analysis, correction for

sequential multiple comparisons, and trial pre-registration. Furthermore, the trial arms were entirely confounded by the hospital and there were false negative outcome measurements (see [1019]). Two of these weaknesses are due to inappropriate data analysis and can therefore be corrected *post hoc* by recalculating the p-values (lack of Intention-To-Treat analysis and multiple comparisons). However, all other weaknesses are fundamental design flaws and cannot be corrected for. Thus, the conclusions cannot be generalized outside of the study. The International Society of Antimicrobial Chemotherapy, the scientific organization that publishes the journal where the article appeared, subsequently announced that the article did not meet its expected standard for publications [795], although it has not been officially retracted.

Because of the preliminary data presented in this study, HCQ treatment was subsequently explored by other researchers. About one week later, a follow-up case study reported that 11 consecutive patients were treated with HCQ + AZ using the same dosing regimen [1020]. One patient died, two were transferred to the intensive care unit (ICU), and one developed a prolonged QT interval, leading to discontinuation of HCQ + AZ administration. As in the Gautret et al. study, the outcome assessed was virological clearance at day 6 post-treatment, as measured from nasopharyngeal swabs. Of the ten living patients on day 6, eight remained positive for SARS-CoV-2 RNA. Like in the original study, interpretability was severely limited by the lack of a comparison group and the small sample size. However, these results stand in contrast to the claims by Gautret et al. that all six patients treated with HCQ + AZ tested negative for SARS-CoV-2 RNA by day 6 post-treatment. This case study illustrated the need for further investigation using robust study design to evaluate the efficacy of HCQ and/or CQ.

On April 10, 2020, a randomized, non-placebo trial of 62 COVID-19 patients at the Renmin Hospital of Wuhan University was released [1021]. This study investigated whether HCQ decreased time to fever break or time to cough relief when compared to SOC [1021]. This trial found HCQ decreased both average time to fever break and average time to cough relief, defined as mild or no cough. While this study improved on some of the methodological flaws in Gautret et al. by randomizing patients, it also had several flaws in trial design and data analysis that prevent generalization of the results. These weaknesses include the lack of placebo, lack of correction for multiple primary outcomes, inappropriate choice of outcomes, lack of sufficient detail to understand analysis, drastic disparities between pre-registration [1022] and published protocol (including differences in the inclusion and exclusion criteria, the number of experimental groups, the number of patients enrolled, and the outcome analyzed), and small sample size. The choice of outcomes may be inappropriate as both fevers and cough may break periodically without resolution of illness. Additionally, for these outcomes, the authors reported that 23 of 62 patients did not have a fever and 25 of 62 patients did not have a cough at the start of the study, but the authors failed to describe how these patients were included in a study assessing time to fever break and time to cough relief. It is important to note here that the authors claimed “neither the research performers nor the patients were aware of the treatment assignments.” This blinding seems impossible in a non-placebo trial because at the very least, providers would know whether they were administering a medication or not, and this knowledge could lead

to systematic differences in the administration of care. Correction for multiple primary outcomes can be adjusted *post hoc* by recalculating p-values, but all of the other issues were design and statistical weaknesses that cannot be corrected for. Additionally, disparities between the pre-registered and published protocols raise concerns about experimental design. The design limitations mean that the conclusions cannot be generalized outside of the study.

A second randomized trial, conducted by the Shanghai Public Health Clinical Center, analyzed whether HCQ increased rates of virological clearance at day 7 in respiratory pharyngeal swabs compared to SOC [1023]. This trial was published in Chinese along with an abstract in English, and only the English abstract was read and interpreted for this review. The trial found comparable outcomes in virological clearance rate, time to virological clearance, and time to body temperature normalization between the treatment and control groups. The small sample size is one weakness, with only 30 patients enrolled and 15 in each arm. This problem suggests the study is underpowered to detect potentially useful differences and precludes interpretation of results. Additionally, because only the abstract could be read, other design and analysis issues could be present. Thus, though these studies added randomization to their assessment of HCQ, their conclusions should be interpreted very cautiously. These two studies assessed different outcomes and reached differing conclusions about the efficacy of HCQ for treating COVID-19; the designs of both studies, especially with respect to sample size, meant that no general conclusions can be made about the efficacy of the drug.

Several widely reported studies on HCQ also have issues with data integrity and/or provenance. A Letter to the Editor published in *BioScience Trends* on March 16, 2020 claimed that numerous clinical trials have shown that HCQ is superior to control treatment in inhibiting the exacerbation of COVID-19 pneumonia [1024]. This letter has been cited by numerous primary literature, review articles, and media alike [1025,1026]. However, the letter referred to 15 pre-registration identifiers from the Chinese Clinical Trial Registry. When these identifiers are followed back to the registry, most trials claim they are not yet recruiting patients or are currently recruiting patients. For all of these 15 identifiers, no data uploads or links to publications could be located on the pre-registrations. At the very least, the lack of availability of the primary data means the claim that HCQ is efficacious against COVID-19 pneumonia cannot be verified. Similarly, a recent multinational registry analysis [1027] analyzed the efficacy of CQ and HCQ with and without a macrolide, which is a class of antibiotics that includes Azithromycin, for the treatment of COVID-19. The study observed 96,032 patients split into a control and four treatment conditions (CQ with and without a macrolide; HCQ with and without a macrolide). They concluded that treatment with CQ or HCQ was associated with increased risk of *de novo* ventricular arrhythmia during hospitalization. However, this study has since been retracted by *The Lancet* due to an inability to validate the data used [1028]. These studies demonstrate that increased skepticism in evaluation of the HCQ/CQ and COVID-19 literature may be warranted, possibly because of the significant attention HCQ and CQ have received as possible treatments for COVID-19 and the politicization of these drugs.

Despite the fact that the study suggesting that CQ/HCQ increased risk of ventricular arrhythmia in COVID-19 patients has now been retracted, previous studies have identified risks associated with HCQ/CQ. A patient with systemic lupus erythematosus developed a prolonged QT interval that was likely exacerbated by use of HCQ in combination with renal failure [1029]. A prolonged QT interval is associated with ventricular arrhythmia [1030]. Furthermore, a separate study [1031] investigated the safety associated with the use of HCQ with and without macrolides between 2000 and 2020. The study involved 900,000 cases treated with HCQ and 300,000 cases treated with HCQ + AZ. The results indicated that short-term use of HCQ was not associated with additional risk, but that HCQ + AZ was associated with an enhanced risk of cardiovascular complications (such as a 15% increased risk of chest pain, calibrated HR = 1.15, 95% CI, 1.05 to 1.26) and a two-fold increased 30-day risk of cardiovascular mortality (calibrated HR = 2.19; 95% CI, 1.22 to 3.94). Therefore, whether studies utilize HCQ alone or HCQ in combination with a macrolide may be an important consideration in assessing risk. As results from initial investigations of these drug combinations have emerged, concerns about the efficacy and risks of treating COVID-19 with HCQ and CQ have led to the removal of CQ/HCQ from SOC practices in several countries [1032,1033]. As of May 25, 2020, WHO had suspended administration of HCQ as part of the worldwide Solidarity Trial [1034], and later the final results of this large-scale trial that compared 947 patients administered HCQ to 906 controls revealed no effect on the primary outcome, mortality during hospitalization (rate ratio: 1.19; $p = 0.23$)

Additional research has emerged largely identifying HCQ/CQ to be ineffective against COVID-19 while simultaneously revealing a number of significant side effects. A randomized, open-label, non-placebo trial of 150 COVID-19 patients was conducted in parallel at 16 government-designated COVID-19 centers in China to assess the safety and efficacy of HCQ [1035]. The trial compared treatment with HCQ in conjunction with SOC to SOC alone in 150 infected patients who were assigned randomly to the two groups (75 per group). The primary endpoint of the study was the negative conversion rate of SARS-CoV-2 in 28 days, and the investigators found no difference in this parameter between the groups (estimated difference between SOC plus HCQ and SOC 4.1%; 95% CI, -10.3% to 18.5%). The secondary endpoints were an amelioration of the symptoms of the disease such as axillary temperature $\leq 36.6^{\circ}\text{C}$, $\text{SpO}_2 > 94\%$ on room air, and disappearance of symptoms like shortness of breath, cough, and sore throat. The median time to symptom alleviation was similar across different conditions (19 days in HCQ + SOC versus 21 days in SOC, $p = 0.97$). Additionally, 30% of the patients receiving SOC+HCQ reported adverse outcomes compared to 8.8% of patients receiving only SOC, with the most common adverse outcome in the SOC+HCQ group being diarrhea (10% versus 0% in the SOC group, $p = 0.004$). However, there are several factors that limit the interpretability of this study. Most of the enrolled patients had mild-to-moderate symptoms (98%), and the average age was 46. SOC in this study included the use of antivirals (Lopinavir-Ritonavir, Arbidol, Oseltamivir, Virazole, Entecavir, Ganciclovir, and Interferon alfa), which the authors note could influence the results. Thus, they note that an ideal SOC would need to exclude the use of antivirals, but that ceasing antiviral treatment raised ethical concerns at the time that the study was conducted. In this trial, the samples used to test for the presence of the SARS-CoV-2 virus were collected from the upper respiratory tract, and

the authors indicated that the use of upper respiratory samples may have introduced false negatives (e.g., [69]). Another limitation of the study that the authors acknowledge was that the HCQ treatment began, on average, at a 16-day delay from the symptom onset. The fact that this study was open-label and lacked a placebo limits interpretation, and additional analysis is required to determine whether HCQ reduces inflammatory response.

Therefore, despite some potential areas of investigation identified in *post hoc* analysis, this study cannot be interpreted as providing support for HCQ as a therapeutic against COVID-19. This study provided no support for HCQ against COVID-19, as there was no difference between the two groups in either negative seroconversion at 28 days or symptom alleviation, and in fact, more severe adverse outcomes were reported in the group receiving HCQ.

Additional evidence comes from a retrospective analysis [1036] that examined data from 368 COVID-19 patients across all United States Veteran Health Administration medical centers. The study retrospectively investigated the effect of the administration of HCQ (n=97), HCQ + AZ (n=113), and no HCQ (n=158) on 368 patients. The primary outcomes assessed were death and the need for mechanical ventilation. Standard supportive care was rendered to all patients. Due to the low representation of women (N=17) in the available data, the analysis included only men, and the median age was 65 years. The rate of death was 27.8% in the HCQ-only treatment group, 22.1% in the HCQ + AZ treatment group, and 14.1% in the no-HCQ group. These data indicated a statistically significant elevation in the risk of death for the HCQ-only group compared to the no-HCQ group (adjusted HR: 2.61, $p = 0.03$), but not for the HCQ + AZ group compared to the no-HCQ group (adjusted HR: 1.14; $p = 0.72$). Further, the risk of ventilation was similar across all three groups (adjusted HR: 1.43, $p = 0.48$ (HCQ) and 0.43, $p = 0.09$ (HCQ + AZ) compared to no HCQ). The study thus showed evidence of an association between increased mortality and HCQ in this cohort of COVID-19 patients but no change in rates of mechanical ventilation among the treatment conditions. The study had a few limitations: it was not randomized, and the baseline vital signs, laboratory tests, and prescription drug use were significantly different among the three groups. All of these factors could potentially influence treatment outcome. Furthermore, the authors acknowledge that the effect of the drugs might be different in females and pediatric subjects, since these subjects were not part of the study. The reported result that HCQ + AZ is safer than HCQ contradicts the findings of the previous large-scale analysis of twenty years of records that found HCQ + AZ to be more frequently associated with cardiac arrhythmia than HCQ alone [1031]; whether this discrepancy is caused by the pathology of COVID-19, is influenced by age or sex, or is a statistical artifact is not presently known.

Finally, findings from the RECOVERY trial were released on October 8, 2020. This study used a randomized, open-label design to study the effects of HCQ compared to SOC in 11,197 patients at 176 hospitals in the United Kingdom [796]. Patients were randomized into either the control group or one of the treatment arms, with twice as many patients enrolled in the control group as any treatment group. Of the patients eligible to receive HCQ, 1,561 were randomized into the HCQ arm, and 3,155 were randomized into the control arm. The demographics of the HCQ and control groups were similar in terms of average age (65 years), proportion female (approximately 38%), ethnic make-up (73% versus 76% white), and prevalence of pre-existing conditions

(56% versus 57% overall). In the HCQ arm of the study, patients received 800 mg at baseline and again after 6 hours, then 400 mg at 12 hours and every subsequent 12 hours. The primary outcome analyzed was all-cause mortality, and patient vital statistics were reported by physicians upon discharge or death, or else at 28 days following HCQ administration if they remained hospitalized. The secondary outcome assessed was the combined risk of progression to invasive mechanical ventilation or death within 28 days. By the advice of an external data monitoring committee, the HCQ arm of the study was reviewed early, leading to it being closed due a lack of support for HCQ as a treatment for COVID-19. COVID-19-related mortality was not affected by HCQ in the RECOVERY trial (rate ratio, 1.09; 95% CI, 0.97 to 1.23; $p = 0.15$), but cardiac events were increased in the HCQ arm (0.4 percentage points), as was the duration of hospitalization (rate ratio for discharge alive within 28 days: 0.90; 95% CI, 0.83 to 0.98) and likelihood of progression to mechanical ventilation or death (risk ratio 1.14; 95% CI, 1.03 to 1.27). This large-scale study thus builds upon studies in the United States and China to suggest that HCQ is not an effective treatment, and in fact may negatively impact COVID-19 patients due to its side effects. Therefore, though none of the studies have been blinded, examining them together makes it clear that the available evidence points to significant dangers associated with the administration of HCQ to hospitalized COVID-19 patients, without providing any support for its efficacy.

5.4.2 HCQ for the Treatment of Mild Cases

One additional possible therapeutic application of HCQ considered was the treatment of mild COVID-19 cases in otherwise healthy individuals. This possibility was assessed in a randomized, open-label, multi-center analysis conducted in Catalonia (Spain) [1037]. This analysis enrolled adults 18 and older who had been experiencing mild symptoms of COVID-19 for fewer than five days. Participants were randomized into an HCQ arm (N=136) and a control arm (N=157), and those in the treatment arm were administered 800 mg of HCQ on the first day of treatment followed by 400 mg on each of the subsequent six days. The primary outcome assessed was viral clearance at days 3 and 7 following the onset of treatment, and secondary outcomes were clinical progression and time to complete resolution of symptoms. No significant differences between the two groups were found: the difference in viral load between the HCQ and control groups was 0.01 (95% CI, -0.28 to 0.29) at day 3 and -0.07 (95% CI -0.44 to 0.29) at day 7, the relative risk of hospitalization was 0.75 (95% CI, 0.32 to 1.77), and the difference in time to complete resolution of symptoms was -2 days ($p = 0.38$). This study thus suggests that HCQ does not improve recovery from COVID-19, even in otherwise healthy adult patients with mild symptoms.

5.4.3 Prophylactic Administration of HCQ

An initial study of the possible prophylactic application of HCQ utilized a randomized, double-blind, placebo-controlled design to analyze the administration of HCQ prophylactically [1038]. Asymptomatic adults in the United States and Canada who had been exposed to SARS-CoV-2 within the past four days were enrolled in an online study to evaluate whether administration of HCQ over five days influenced the probability of developing COVID-19 symptoms over a 14-day period. Of the participants, 414 received

HCQ and 407 received a placebo. No significant difference in the rate of symptomatic illness was observed between the two groups (11.8% HCQ, 14.3% placebo, $p = 0.35$). The HCQ condition was associated with side effects, with 40.1% of patients reporting side effects compared to 16.8% in the control group ($p < 0.001$). However, likely due to the high enrollment of healthcare workers (66% of participants) and the well-known side effects associated with HCQ, a large number of participants were able to correctly identify whether they were receiving HCQ or a placebo (46.5% and 35.7%, respectively). Furthermore, due to a lack of availability of diagnostic testing, only 20 of the 107 cases were confirmed with a PCR-based test to be positive for SARS-CoV-2. The rest were categorized as “probable” or “possible” cases by a panel of four physicians who were blind to the treatment status. One possible confounder is that a patient presenting one or more symptoms, which included diarrhea, was defined as a “possible” case, but diarrhea is also a common side effect of HCQ. Additionally, four of the twenty PCR-confirmed cases did not develop symptoms until after the observation period had completed, suggesting that the 14-day trial period may not have been long enough or that some participants also encountered secondary exposure events. Finally, in addition to the young age of the participants in this study, which ranged from 32 to 51, there were possible impediments to generalization introduced by the selection process, as 2,237 patients who were eligible but had already developed symptoms by day 4 were enrolled in a separate study. It is therefore likely that asymptomatic cases were over-represented in this sample, which would not have been detected based on the diagnostic criteria used. Therefore, while this study does represent the first effort to conduct a randomized, double-blind, placebo-controlled investigation of HCQ’s effect on COVID-19 prevention after SARS-CoV-2 exposure in a large sample, the lack of PCR tests and several other design flaws significantly impede interpretation of the results. However, in line with the results from therapeutic studies, once again no evidence was found suggesting an effect of HCQ against COVID-19.

A second study [1039] examined the effect of administering HCQ to healthcare workers as a pre-exposure prophylactic. The primary outcome assessed was the conversion from SARS-CoV-2 negative to SARS-CoV-2 positive status over the 8 week study period. This study was also randomized, double-blind, and placebo-controlled, and it sought to address some of the limitations of the first prophylactic study. The goal was to enroll 200 healthcare workers, preferentially those working with COVID-19 patients, at two hospitals within the University of Pennsylvania hospital system in Philadelphia, PA. Participants were randomized 1:1 to receive either 600 mg of HCQ daily or a placebo, and their SARS-CoV-2 infection status and antibody status were assessed using RT-PCR and serological testing, respectively, at baseline, 4 weeks, and 8 weeks following the beginning of the treatment period. The statistical design of the study accounted for interim analyses at 50 and 100 participants in case efficacy or futility of HCQ for prophylaxis became clear earlier than completion of enrollment. The 139 individuals enrolled comprised a study population that was fairly young (average age 33) and made of largely of people who were white, women, and without pre-existing conditions. At the second interim analysis, more individuals in the treatment group than the control group had contracted COVID-19 (4 versus

3), causing the estimated z-score to fall below the pre-established threshold for futility. As a result, the trial was terminated early, offering additional evidence against the use of HCQ for prophylaxis.

5.4.4 Summary of HCQ/CQ Research Findings

Early *in vitro* evidence indicated that HCQ could be an effective therapeutic against SARS-CoV-2 and COVID-19, leading to significant media attention and public interest in its potential as both a therapeutic and prophylactic. Initially it was hypothesized that CQ/HCQ might be effective against SARS-CoV-2 in part because CQ and HCQ have both been found to inhibit the expression of CD154 in T-cells and to reduce TLR signaling that leads to the production of pro-inflammatory cytokines [1040]. Clinical trials for COVID-19 have more often used HCQ rather than CQ because it offers the advantages of being cheaper and having fewer side effects than CQ. However, research has not found support for a positive effect of HCQ on COVID-19 patients. Multiple clinical studies have already been carried out to assess HCQ as a therapeutic agent for COVID-19, and many more are in progress. To date, none of these studies have used randomized, double-blind, placebo-controlled designs with a large sample size, which would be the gold standard. Despite the design limitations (which would be more likely to produce false positives than false negatives), initial optimism about HCQ has largely dissipated. The most methodologically rigorous analysis of HCQ as a prophylactic [1038] found no significant differences between the treatment and control groups, and the WHO's global Solidarity trial similarly reported no effect of HCQ on mortality [789]. Thus, HCQ/CQ are not likely to be effective therapeutic or prophylactic agents against COVID-19. One case study identified drug-induced phospholipidosis as the cause of death for a COVID-19 patient treated with HCQ [962], suggesting that in some cases, the proposed mechanism of action may ultimately be harmful. Additionally, one study identified an increased risk of mortality in older men receiving HCQ, and administration of HCQ and HCQ + AZ did not decrease the use of mechanical ventilation in these patients [1036]. HCQ use for COVID-19 could also lead to shortages for anti-malarial or anti-rheumatic use, where it has documented efficacy. Despite significant early attention, these drugs appear to be ineffective against COVID-19. Several countries have now removed CQ/HCQ from their SOC for COVID-19 due to the lack of evidence of efficacy and the frequency of adverse effects.

5.5 ACE Inhibitors and Angiotensin II Receptor Blockers

Several clinical trials testing the effects of ACEIs or ARBs on COVID-19 outcomes are ongoing [1041,1042,1043,1044,1045,1046,1047]. Clinical trials are needed because the findings of the various observational studies bearing on this topic cannot be interpreted as indicating a protective effect of the drug [1048,1049]. Two analyses [1041,1047] have reported no effect of continuing or discontinuing ARBs and ACEIs on patients admitted to the hospital for COVID-19. The first, known as REPLACE COVID [852], was a randomized, open-label study that enrolled patients who were admitted to the hospital for COVID-19 and were taking an ACEI at the time of admission. They enrolled 152 patients at 20 hospitals across seven countries and

randomized them into two arms, continuation (n=75) and discontinuation (n=77). The primary outcome evaluated was a global rank score that integrated several dimensions of illness. The components of this global rank score, such as time to death and length of mechanical ventilation, were evaluated as secondary endpoints. This analysis reported no differences between the two groups in the primary or any of the secondary outcomes.

Similarly, a second study [853] used a randomized, open-label design to examine the effects of continuing versus discontinuing ARBs and ACEIs on patients hospitalized for mild to moderate COVID-19 at 29 hospitals in Brazil. This study enrolled 740 patients but had to exclude one trial site from all analyses due to the discovery of violations of Good Clinical Trial practice and data falsification. After this exclusion, 659 patients remained, with 334 randomized to discontinuation and 325 to continuation. In this study, the primary endpoint analyzed was the number of days that patients were alive and not hospitalized within 30 days of enrollment. The secondary outcomes included death (including in-hospital death separately), number of days hospitalized, and specific clinical outcomes such as heart failure or stroke. Once again, no significant differences were found between the two groups. Initial studies of randomized interventions therefore suggest that ACEIs and ARBs are unlikely to affect COVID-19 outcomes. These results are also consistent with findings from observational studies (summarized in [852]). Additional information about ACE2, observational studies of ACEIs and ARBs in COVID-19, and clinical trials on this topic have been summarized [1050]. Therefore, despite the promising potential mechanism, initial results have not provided support for ACEIs and ARBs as therapies for COVID-19.

5.6 Tocilizumab

Human IL-6 is a 26-kDa glycoprotein that consists of 184 amino acids and contains two potential N-glycosylation sites and four cysteine residues. It binds to a type I cytokine receptor (IL-6Ra or glycoprotein 80) that exists in both membrane-bound (IL-6Ra) and soluble (sIL-6Ra) forms [1051]. It is not the binding of IL-6 to the receptor that initiates pro- and/or anti-inflammatory signaling, but rather the binding of the complex to another subunit, known as IL-6R β or glycoprotein 130 (gp130) [1051,1052]. Unlike membrane-bound IL-6Ra, which is only found on hepatocytes and some types of leukocytes, gp130 is found on most cells [1053]. When IL-6 binds to sIL-6Ra, the complex can then bind to a gp130 protein on any cell [1053]. The binding of IL-6 to IL-6Ra is termed classical signaling, while its binding to sIL-6Ra is termed trans-signaling [1053,1054,1055]. These two signaling processes are thought to play different roles in health and illness. For example, trans-signaling may play a role in the proliferation of mucosal T-helper TH2 cells associated with asthma, while an earlier step in this proliferation process may be regulated by classical signaling [1053]. Similarly, IL-6 is known to play a role in Crohn's Disease via trans-, but not classical, signaling [1053]. Both classical and trans-signaling can occur through three independent pathways: the Janus-activated kinase-STAT3 pathway, the Ras/Mitogen-Activated Protein Kinases pathway and the Phosphoinositol-3 Kinase/Akt pathway [1051]. These signaling pathways are involved in a variety of different functions, including cell type differentiation, immunoglobulin synthesis, and cellular survival signaling pathways, respectively [1051]. The ultimate result of the IL-6 cascade is to direct transcriptional activity of

various promoters of pro-inflammatory cytokines, such as IL-1, TNF, and even IL-6 itself, through the activity of NF- κ B [1051]. IL-6 synthesis is tightly regulated both transcriptionally and post-transcriptionally, and it has been shown that viral proteins can enhance transcription of the IL-6 gene by strengthening the DNA-binding activity between several transcription factors and IL-6 gene-cis-regulatory elements [1056]. Therefore, drugs inhibiting the binding of IL-6 to IL-6Ra or sIL-6Ra are of interest for combating the hyperactive inflammatory response characteristic of cytokine release syndrome (CRS) and cytokine storm syndrome (CSS). TCZ is a humanized monoclonal antibody that binds both to the insoluble and soluble receptor of IL-6, providing de facto inhibition of the IL-6 immune cascade. Interest in TCZ as a possible treatment for COVID-19 was piqued by early evidence indicating that COVID-19 deaths may be induced by the hyperactive immune response, often referred to as CRS or CSS [81], as IL-6 plays a key role in this response [148]. The observation of elevated IL-6 in patients who died relative to those who recovered [81] could reflect an over-production of proinflammatory interleukins, suggesting that TCZ could potentially palliate some of the most severe symptoms of COVID-19 associated with increased cytokine production.

This early interest in TCZ as a possible treatment for COVID-19 was bolstered by a very small retrospective study in China that examined 20 patients with severe symptoms in early February 2020 and reported rapid improvement in symptoms following treatment with TCZ [871]. Subsequently, a number of retrospective studies have been conducted in several countries. Many studies use a retrospective, observational design, where they compare outcomes for COVID-19 patients who received TCZ to those who did not over a set period of time. For example, one of the largest retrospective, observational analyses released to date [866], consisting of 1,351 patients admitted to several care centers in Italy, compared the rates at which patients who received TCZ died or progressed to invasive medical ventilation over a 14-day period compared to patients receiving only SOC. Under this definition, SOC could include other drugs such as HCQ, azithromycin, lopinavir-ritonavir or darunavir-cobicistat, or heparin. While this study was not randomized, a subset of patients who were eligible to receive TCZ were unable to obtain it due to shortages; however, these groups were not directly compared in the analysis. After adjusting for variables such as age, sex, and SOFA (sequential organ failure assessment) score, they found that patients treated with TCZ were less likely to progress to invasive medical ventilation and/or death (adjusted HR = 0.61, CI 0.40-0.92, $p = 0.020$); analysis of death and ventilation separately suggests that this effect may have been driven by differences in the death rate (20% of control versus 7% of TCZ-treated patients). The study reported particular benefits for patients whose $\text{PaO}_2/\text{FiO}_2$ ratio, also known as the Horowitz Index for Lung Function, fell below a 150 mm Hg threshold. They found no differences between groups administered subcutaneous versus intravenous TCZ.

Another retrospective observational analysis of interest examined the charts of patients at a hospital in Connecticut, USA where 64% of all 239 COVID-19 patients in the study period were administered TCZ based on assignment by a standardized algorithm [867]. They found that TCZ administration was associated with more similar rates of survivorship in patients with severe versus nonsevere COVID-19 at intake, defined based on the amount of

supplemental oxygen needed. They therefore proposed that their algorithm was able to identify patients presenting with or likely to develop CRS as good candidates for TCZ. This study also reported higher survivorship in Black and Hispanic patients compared to white patients when adjusted for age. The major limitation with interpretation for these studies is that there may be clinical characteristics that influenced medical practitioners decisions to administer TCZ to some patients and not others. One interesting example therefore comes from an analysis of patients at a single hospital in Brescia, Italy, where TCZ was not available for a period of time [868]. This study compared COVID-19 patients admitted to the hospital before and after March 13, 2020, when the hospital received TCZ. Therefore, patients who would have been eligible for TCZ prior to this arbitrary date did not receive it as treatment, making this retrospective analysis something of a natural experiment. Despite this design, demographic factors did not appear to be consistent between the two groups, and the average age of the control group was older than the TCZ group. The control group also had a higher percentage of males and a higher incidence of comorbidities such as diabetes and heart disease. All the same, the multivariate HR, which adjusted for these clinical and demographic factors, found a significant difference between survival in the two groups ($HR=0.035$, $CI=0.004-0.347$, $p = 0.004$). The study reported improvement of survival outcomes after the addition of TCZ to the SOC regime, with 11 of 23 patients (47.8%) admitted prior to March 13th dying compared to 2 of 62 (3.2%) admitted afterwards ($HR=0.035$; 95% CI, 0.004 to 0.347; $p = 0.004$). They also reported a reduced progression to mechanical ventilation in the TCZ group. However, this study also holds a significant limitation: the time delay between the two groups means that knowledge about how to treat the disease likely improved over this timeframe as well. All the same, the results of these observational retrospective studies provide support for TCZ as a pharmaceutical of interest for follow-up in clinical trials.

Other retrospective analyses have utilized a case-control design to match pairs of patients with similar baseline characteristics, only one of whom received TCZ for COVID-19. In one such study, TCZ was significantly associated with a reduced risk of progression to intensive care unit (ICU) admission or death [869]. This study examined only 20 patients treated with TCZ (all but one of the patients treated with TCZ in the hospital during the study period) and compared them to 25 patients receiving SOC. For the combined primary endpoint of death and/or ICU admission, only 25% of patients receiving TCZ progressed to an endpoint compared to 72% in the SOC group ($p = 0.002$, presumably based on a chi-square test based on the information provided in the text). When the two endpoints were examined separately, progression to invasive medical ventilation remained significant (32% SOC compared to 0% TCZ, $p = 0.006$) but not for mortality (48% SOC compared to 25% TCZ, $p = 0.066$). In contrast, a study that compared 96 patients treated with TCZ to 97 patients treated with SOC only in New York City found that differences in mortality did not differ between the two groups, but that this difference did become significant when intubated patients were excluded from the analysis [870]. Taken together, these findings suggest that future clinical trials of TCZ may want to include intubation as an endpoint. However, these studies should be approached with caution, not only because of the small number of patients enrolled and the retrospective design, but also because they performed a large number of

statistical tests and did not account for multiple hypothesis testing. In general, caution must be exercised when interpreting subgroup analyses after a primary combined endpoint analysis. These last findings highlight the need to search for a balance between impairing a harmful immune response, such as the one generated during CRS/CSS, and preventing the worsening of the clinical picture of the patients by potential new viral infections. Early meta-analyses and systematic reviews have investigated the available data about TCZ for COVID-19. One meta-analysis [1057] evaluated 19 studies published or released as preprints prior to July 1, 2020 and found that the overall trends were supportive of the frequent conclusion that TCZ does improve survivorship, with a significant HR of 0.41 ($p < 0.001$). This trend improved when they excluded studies that administered a steroid alongside TCZ, with a significant HR of 0.04 ($p < 0.001$). They also found some evidence for reduced invasive ventilation or ICU admission, but only when excluding all studies except a small number whose estimates were adjusted for the possible bias introduced by the challenges of stringency during the enrollment process. A systematic analysis of sixteen case-control studies of TCZ estimated an odds ratio of mortality of 0.453 (95% CI 0.376–0.547, $p < 0.001$), suggesting possible benefits associated with TCZ treatment [1058]. Although these estimates are similar, it is important to note that they are drawing from the same literature and are therefore likely to be affected by the same potential biases in publication. A different systematic review of studies investigating TCZ treatment for COVID-19 analyzed 31 studies that had been published or released as pre-prints and reported that none carried a low risk of bias [1059]. Therefore, the present evidence is not likely to be sufficient for conclusions about the efficacy of TCZ.

On February 11, 2021, a preprint describing the first randomized control trial of TCZ was released as part of the RECOVERY trial [872]. Of the 21,550 patients enrolled in the RECOVERY trial at the time, 4,116 adults hospitalized with COVID-19 across the 131 sites in the United Kingdom were assigned to the arm of the trial evaluating the effect of TCZ. Among them, 2,022 were randomized to receive TCZ and 2,094 were randomized to SOC, with 79% of patients in each group available for analysis at the time that the initial report was released. The primary outcome measured was 28-day mortality, and TCZ was found to reduce 28-day mortality from 33% of patients receiving SOC alone to 29% of those receiving TCZ, corresponding to a rate ratio of 0.86 (95% CI 0.77-0.96; $p = 0.007$). TCZ was also significantly associated with the probability of hospital discharge within 28 days for living patients, which was 47% in the SOC group and 54% in the TCZ group (rate ratio 1.22, 95% CI 1.12-1.34, $p < 0.0001$). A potential statistical interaction between TCZ and corticosteroids was observed, with the combination providing greater mortality benefits than TCZ alone, but the authors note that caution is advisable in light of the number of statistical tests conducted. Combining the RECOVERY trial data with data from seven smaller randomized control trials indicates that TCZ is associated with a 13% reduction in 28-day mortality (rate ratio 0.87, 95% CI 0.79-0.96, $p = 0.005$) [872].

There are possible risks associated with the administration of TCZ for COVID-19. TCZ has been used for over a decade to treat RA [1060], and a recent study found the drug to be safe for pregnant and breastfeeding women [1061]. However, TCZ may increase the risk of developing infections [1060], and RA patients with chronic hepatitis B infections had a high risk of hepatitis

B virus reactivation when TCZ was administered in combination with other RA drugs [1062]. As a result, TCZ is contraindicated in patients with active infections such as tuberculosis [1063]. Previous studies have investigated, with varying results, a possible increased risk of infection in RA patients administered TCZ [1064,1065], although another study reported that the incidence rate of infections was higher in clinical practice RA patients treated with TCZ than in the rates reported by clinical trials [1066]. In the investigation of 544 Italian COVID-19 patients, the group treated with TCZ was found to be more likely to develop secondary infections, with 24% compared to 4% in the control group ($p < 0.0001$) [866]. Reactivation of hepatitis B and herpes simplex virus 1 was also reported in a small number of patients in this study, all of whom were receiving TCZ. A July 2020 case report described negative outcomes of two COVID-19 patients after receiving TCZ, including one death; however, both patients were intubated and had entered septic shock prior to receiving TCZ [1067], likely indicating a severe level of cytokine production. Additionally, D-dimer and sIL2R levels were reported by one study to increase in patients treated with TCZ, which raised concerns because of the potential association between elevated D-dimer levels and thrombosis and between sIL2R and diseases where T-cell regulation is compromised [867]. An increased risk of bacterial infection was also identified in a systematic review of the literature, based on the unadjusted estimates reported [1057]. In the RECOVERY trial, however, only three out of 2,022 participants in the group receiving TCZ developed adverse reactions determined to be associated with the intervention, and no excess deaths were reported [872]. TCZ administration to COVID-19 patients is not without risks and may introduce additional risk of developing secondary infections; however, while caution may be prudent when treating patients who have latent viral infections, the results of the RECOVERY trial indicate that adverse reactions to TCZ are very rare among COVID-19 patients broadly.

In summary, approximately 33% of hospitalized COVID-19 patients develop ARDS [1068], which is caused by an excessive early response of the immune system which can be a component of CRS/CSS [867,1063]. This overwhelming inflammation is triggered by IL-6. TCZ is an inhibitor of IL-6 and therefore may neutralize the inflammatory pathway that leads to the cytokine storm. The mechanism suggests TCZ could be beneficial for the treatment of COVID-19 patients experiencing excessive immune activity, and the RECOVERY trial reported a reduction in 28-day mortality. Interest in TCZ as a treatment for COVID-19 was also supported by two meta-analyses [1057,1069], but a third meta-analysis found that all of the available literature at that time carried a risk of bias [1059]. Additionally, different studies used different dosages, number of doses, and methods of administration. Ongoing research may be needed to optimize administration of TCZ [1070], although similar results were reported by one study for intravenous and subcutaneous administration [866]. Clinical trials that are in progress are likely to provide additional insight into the effectiveness of this drug for the treatment of COVID-19 along with how it should be administered.

5.7 Interferons

IFNs are a family of cytokines critical to activating the innate immune response against viral infections. Interferons are classified into three categories based on their receptor specificity: types I, II and III [148]. Specifically, IFNs I (IFN- α and β) and II (IFN- γ) induce the expression of antiviral proteins [1071]. Among these IFNs, IFN- β has already been found to strongly inhibit the replication of other coronaviruses, such as SARS-CoV-1, in cell culture, while IFN- α and γ were shown to be less effective in this context [1071]. There is evidence that patients with higher susceptibility to ARDS indeed show deficiency in IFN- β . For instance, infection with other coronaviruses impairs IFN- β expression and synthesis, allowing the virus to escape the innate immune response [1072]. On March 18 2020, Synairgen plc received approval to start a phase II trial for SNG001, an IFN- β -1a formulation to be delivered to the lungs via inhalation [879]. SNG001, which contains recombinant interferon beta-1a, was previously shown to be effective in reducing viral load in an *in vivo* model of swine flu and *in vitro* models of other coronavirus infections [1073]. In July 2020, a press release from Synairgen stated that SNG001 reduced progression to ventilation in a double-blind, placebo-controlled, multi-center study of 101 patients with an average age in the late 50s [880]. These results were subsequently published in November 2020 [881]. The study reports that the participants were assigned at a ratio of 1:1 to receive either SNG001 or a placebo that lacked the active compound, by inhalation for up to 14 days. The primary outcome they assessed was the change in patients' score on the WHO Ordinal Scale for Clinical Improvement (OSCI) at trial day 15 or 16. SNG001 was associated with an odds ratio of improvement on the OSCI scale of 2.32 (95% CI 1.07 – 5.04, $p = 0.033$) in the intention-to-treat analysis and 2.80 (95% CI 1.21 – 6.52, $p = 0.017$) in the per-protocol analysis, corresponding to significant improvement in the SNG001 group on the OSCI at day 15/16. Some of the secondary endpoints analyzed also showed differences: at day 28, the OR for clinical improvement on the OSCI was 3.15 (95% CI 1.39 – 7.14, $p = 0.006$), and the odds of recovery at day 15/16 and at day 28 were also significant between the two groups. Thus, this study suggested that IFN- β 1 administered via SNG001 may improve clinical outcomes.

In contrast, the WHO Solidarity trial reported no significant effect of IFN- β -1a on patient survival during hospitalization [789]. Here, the primary outcome analyzed was in-hospital mortality, and the rate ratio for the two groups was 1.16 (95% CI, 0.96 to 1.39; $p = 0.11$) administering IFN- β -1a to 2050 patients and comparing their response to 2,050 controls. However, there are a few reasons that the different findings of the two trials might not speak to the underlying efficacy of this treatment strategy. One important consideration is the stage of COVID-19 infection analyzed in each study. The Synairgen trial enrolled only patients who were not receiving invasive ventilation, corresponding to a less severe stage of disease than many patients enrolled in the SOLIDARITY trial, as well as a lower overall rate of mortality [1074]. Additionally, the methods of administration differed between the two trials, with the SOLIDARITY trial administering IFN- β -1a subcutaneously [1074]. The differences in findings between the studies suggests that the method of administration might be relevant to outcomes, with nebulized IFN- β -1a more directly targeting receptors in the lungs. A trial that analyzed the effect of subcutaneously administered IFN- β -1a on patients with ARDS between 2015 and 2017 had also reported no effect on 28-day mortality [1075], while a smaller study analyzing the effect of subcutaneous IFN administration did

find a significant improvement in 28-day mortality for COVID-19 [1076]. At present, several ongoing clinical trials are investigating the potential effects of IFN- β -1a, including in combination with therapeutics such as remdesivir [1077] and administered via inhalation [879]. Thus, as additional information becomes available, a more detailed understanding of whether and under which circumstances IFN- β -1a is beneficial to COVID-19 patients should develop.

5.8 Potential Avenues of Interest for Therapeutic Development

Given what is currently known about these therapeutics for COVID-19, a number of related therapies beyond those explored above may also prove to be of interest. For example, the demonstrated benefit of dexamethasone and the ongoing potential of tocilizumab for treatment of COVID-19 suggests that other anti-inflammatory agents might also hold value for the treatment of COVID-19. Current evidence supporting the treatment of severe COVID-19 with dexamethasone suggests that the need to curtail the cytokine storm inflammatory response transcends the risks of immunosuppression, and other anti-inflammatory agents may therefore benefit patients in this phase of the disease. While dexamethasone is considered widely available and generally affordable, the high costs of biologics such as tocilizumab therapy may present obstacles to wide-scale distribution of this drug if it proves of value. At the doses used for RA patients, the cost for tocilizumab ranges from \$179.20 to \$896 per dose for the IV form and \$355 for the pre-filled syringe [1078]. Several other anti-inflammatory agents used for the treatment of autoimmune diseases may also be able to counter the effects of the cytokine storm induced by the virus, and some of these, such as cyclosporine, are likely to be more cost-effective and readily available than biologics [1079]. While tocilizumab targets IL-6, several other inflammatory markers could be potential targets, including TNF- α . Inhibition of TNF- α by a compound such as Etanercept was previously suggested for treatment of SARS-CoV-1 [1080] and may be relevant for SARS-CoV-2 as well. Another anti-IL-6 antibody, sarilumab, is also being investigated [1081,1082]. Baricitinib and other small molecule inhibitors of the Janus-activated kinase pathway also curtail the inflammatory response and have been suggested as potential options for SARS-CoV-2 infections [1083]. Baricitinib, in particular, may be able to reduce the ability of SARS-CoV-2 to infect lung cells [1084]. Clinical trials studying baricitinib in COVID-19 have already begun in the US and in Italy [1085,1086]. Identification and targeting of further inflammatory markers that are relevant in SARS-CoV-2 infection may be of value for curtailing the inflammatory response and lung damage.

In addition to immunosuppressive treatments, which are most beneficial late in disease progression, much research is focused on identifying therapeutics for early-stage patients. For example, although studies of HCQ have not supported the early theory-driven interest in this antiviral treatment, alternative compounds with related mechanisms may still have potential. Hydroxyferroquine derivatives of HCQ have been described as a class of bioorganometallic compounds that exert antiviral effects with some

selectivity for SARS-CoV-1 *in vitro* [1087]. Future work could explore whether such compounds exert antiviral effects against SARS-CoV-2 and whether they would be safer for use in COVID-19.

Another potential approach is the development of antivirals, which could be broad-spectrum, specific to coronaviruses, or targeted to SARS-CoV-2. Development of new antivirals is complicated by the fact that none have yet been approved for human coronaviruses. Intriguing new options are emerging, however. Beta-D-N4-hydroxycytidine is an orally bioavailable ribonucleotide analog showing broad-spectrum activity against RNA viruses, which may inhibit SARS-CoV-2 replication *in vitro* and *in vivo* in mouse models of HCoVs [1088]. A range of other antivirals are also in development. Development of antivirals will be further facilitated as research reveals more information about the interaction of SARS-CoV-2 with the host cell and host cell genome, mechanisms of viral replication, mechanisms of viral assembly, and mechanisms of viral release to other cells; this can allow researchers to target specific stages and structures of the viral life cycle. Finally, antibodies against viruses, also known as antiviral monoclonal antibodies, could be an alternative as well and are described in detail in an above section. The goal of antiviral antibodies is to neutralize viruses through either cell-killing activity or blocking of viral replication [1089]. They may also engage the host immune response, encouraging the immune system to hone in on the virus. Given the cytokine storm that results from immune system activation in response to the virus, which has been implicated in worsening of the disease, a neutralizing antibody (nAb) may be preferable. Upcoming work may explore the specificity of nAbs for their target, mechanisms by which the nAbs impede the virus, and improvements to antibody structure that may enhance the ability of the antibody to block viral activity.

Some research is also investigating potential therapeutics and prophylactics that would interact with components of the innate immune response. For example, TLRs are pattern recognition receptors that recognize pathogen- and damage-associated molecular patterns and contribute to innate immune recognition and, more generally, promotion of both the innate and adaptive immune responses [145]. In mouse models, poly(I:C) and CpG, which are agonists of Toll-like receptors TLR3 and TLR9, respectively, showed protective effects when administered prior to SARS-CoV-1 infection [1090]. Therefore, TLR agonists hold some potential for broad-spectrum prophylaxis.

6 Vaccine Development Strategies for SARS-CoV-2

6.1 Abstract

Vaccines have revolutionized the relationship between people and disease. In the 21st century, several emergent viruses have emphasized the particular value of rapid and scalable vaccine development programs. During the current pandemic caused by *Severe acute respiratory syndrome coronavirus 2* (SARS-CoV-2), recent biotechnological advances in vaccine design have facilitated the development and deployment of vaccines at an unprecedented pace. The genome sequence of SARS-CoV-2 was released in

January 2020, allowing for global efforts in vaccine development to begin within two weeks of the international community becoming aware of the new viral threat. Both established vaccine platforms and more recently developed technologies have been explored against SARS-CoV-2. Although historically a slow process, vaccine development in the face of COVID-19 accelerated so much that less than a year into the pandemic, some vaccine candidates had reported interim phase III clinical trial data and were being administered in countries around the world.

In this review, we contextualize COVID-19 vaccine development in the broader vaccine landscape. We describe where these candidates currently stand in terms of efficacy, safety, and approval and discuss patterns in worldwide distribution. Vaccines have nearly 500 years of history, but the SARS-CoV-2 pandemic provides an exceptional illustration of how rapidly vaccine development technology has advanced in the last two decades in particular.

6.2 Importance

The SARS-CoV-2 pandemic has caused untold damage globally, presenting unusual opportunities and demands in vaccine development. As of May 6, 2022, SARS-CoV-2 has infected over 516,758,993 and taken the lives of 6,249,626 people globally. The development, production, and distribution of vaccines is imperative to saving lives, preventing illness, and reducing the economic and social burdens caused by the COVID-19 pandemic. Effective deployment is critical to reducing the susceptibility of worldwide populations, especially in light of emerging variants. This review provides historical context for the current state of vaccine development and highlights the main strategies utilized for COVID-19 vaccine candidates, their clinical appraisal, and their distribution. These technologies have revolutionized the timescale at which countries can mount a response to an emerging viral threat and provide potential for mitigating future threats before their damage reaches the levels caused by SARS-CoV-2. As the SARS-CoV-2 virus has evolved, they also provide insight into how these technologies have to adapt on incredibly short timescales.

6.3 Introduction

The development of vaccines is widely considered one of the most important medical advances in human history. Over the past 150 years, several new approaches to vaccine development have emerged [1091]. Today, the requirements for developing and deploying a vaccine are complex and often require coordination between government, industry, academia, and philanthropic entities [1092]. Flu-like illnesses caused by viruses that follow an annual pattern are a major target of vaccine development programs. However, vaccine development has historically been slow. The past 20 years have seen several previously unknown viruses emerge and rise rapidly to pose a global threat, challenging vaccine developers to explore approaches that would facilitate a rapid response to novel viruses.

Emergent viral threats of the 21st century include severe acute respiratory syndrome (SARS), “swine flu” (H1N1), Middle East respiratory syndrome (MERS), Ebola virus disease (EVD), and now COVID-19, all of which have underscored the importance of a rapid global response to a new infectious virus. Because vaccines fail to provide immediate prophylactic protection or treatment of ongoing infections, their application to most of these epidemics has been limited [1093]. One of the more successful recent vaccine development programs was for H1N1 influenza. This program benefited from the strong existing infrastructure for influenza vaccines along with the fact that regulatory agencies had determined that vaccines produced using egg- and cell-based platforms could be licensed under the regulations used for a strain change [1094]. Although a monovalent H1N1 vaccine was not available before the pandemic peaked in the United States and Europe, it became available soon afterward as a stand-alone vaccine that was eventually incorporated into commercially available seasonal influenza vaccines [1094]. Critiques of the production and distribution of the H1N1 vaccine have stressed the need for alternative development-and-manufacturing platforms that can be readily adapted to new pathogens.

Vaccine technologies that require only minor adjustments for novel viral threats are appealing because this modular approach would mean they could enter trials quickly in response to a new pathogen of concern. Traditional vaccine technologies were built on the principle of using either a weakened version of the virus or a fragment of the virus, while recent years have seen a paradigm shift towards reverse vaccinology. Reverse vaccinology emphasizes a hypothesis-free approach to vaccine development [1095]. This strategy was explored during development of a DNA vaccine against the Zika virus [1096]. While once again the disease was controlled before the vaccine became available [1094], the response demonstrated the potential for modular technologies to facilitate a response to emerging viral threats [1096]. The potential for such vaccines to benefit the field of oncology has encouraged vaccine developers to invest in next-generation approaches, which has spurred the diversification of vaccine development programs [1097,1098]. As a result, during the COVID-19 pandemic, these modular technologies have taken center stage in controlling a viral threat for the first time.

The pandemic caused by SARS-CoV-2 has highlighted a confluence of circumstances that positioned vaccine development as a key player in efforts to control the virus and mitigate its damage. This virus did not follow the same trajectory as other emergent viruses of recent note, such as SARS-CoV-1, MERS-CoV, and Ebola virus, none of which reached the level of pandemic. Spread of the SARS-CoV-2 virus has remained out of control in many parts of the world into 2022, especially with the emergence of novel variants exhibiting increased rates of transmission [1]. While, for a variety of reasons, SARS-CoV-2 was not controlled as rapidly as the viruses underlying prior 21st century epidemics [3], vaccine development technology had also progressed based on these and other prior viral threats to the point that a rapid international vaccine development response was possible.

Vaccines bolster the immune response to the virus at the population level, thereby driving a lower rate of infection and likely significantly reducing fatalities even for a highly infectious virus like SARS-CoV-2. The first critical

step towards developing a vaccine against SARS-CoV-2 was characterizing the viral target, which happened extremely early in the COVID-19 outbreak with the sequencing and dissemination of the viral genome in early January 2020 [533] (Figure 6). This genomic information allowed for an early identification of the sequence of the spike (S) protein (Figure 6), which is the antigen and induces an immune response [1099,1100].

The Coalition for Epidemic Preparedness Innovations (CEPI) is coordinating global health agencies and pharmaceutical companies to develop vaccines against SARS-CoV-2. As early as September 2020, there were over 180 vaccine candidates against SARS-CoV-2 in development [1101]. While little is currently known about immunity to SARS-CoV-2, vaccine developers typically tests for serum neutralizing activity, as this has been established as a biomarker for adaptive immunity in other respiratory illnesses [1102]. However, unlike in efforts to develop vaccines for prior viral threats, the duration of the COVID-19 pandemic has made it possible to test vaccine in phase III trials where the effect of the vaccine on a cohort's likelihood of contracting SARS-CoV-2 is evaluated. With vaccine candidates at all stages of development, including full approval of some vaccines, the vaccine development landscape for COVID-19 includes vaccines produced by a wide array of technologies. Examining the vaccine development programs tackling the COVID-19 pandemic alongside other 21st century efforts to control emerging viral threats demonstrates the significant biotechnological advances in this field and the importance of modular and adaptable approaches to vaccination. Here, we review the various technologies being explored for the development of SARS-CoV-2 vaccines globally.

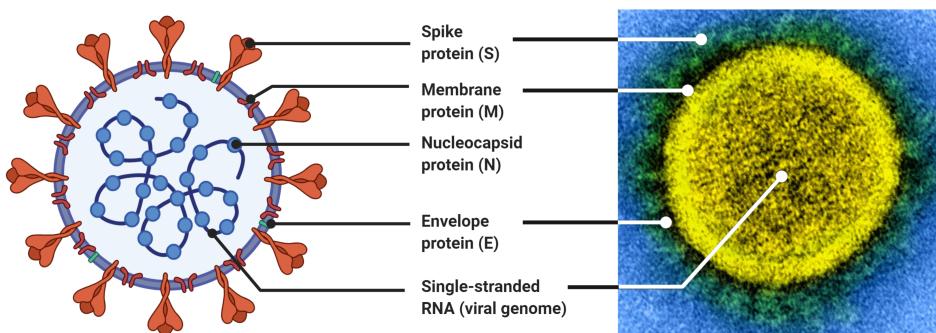


Figure 6: Structure of the SARS-CoV-2 virus. The development of vaccines depends on the immune system recognizing the virus. Here, the structure of SARS-CoV-2 is represented both in the abstract and against a visualization of the virion. The abstracted visualization was made using BioRender [1103] and the microscopy was conducted by the National Institute of Allergy and Infectious Diseases [1104].

6.4 Cross-Platform Considerations in Vaccine Development

Certain design decisions are relevant to vaccine development across multiple platforms. One applies to platforms that deliver the antigen, which in the case of SARS-CoV-2 vaccines is the Spike (S) protein. The prefusion conformation of the SARS-CoV-2 S protein, which is the structure before the virus fuses to the host cell membrane, is metastable [1105], and the release of energy during membrane fusion drives this process forward following destabilization [10,1106]. Due to the significant conformational changes that

occur during membrane fusion [23,1107,1108], S protein immunogens that are stabilized in the prefusion conformation are of particular interest, especially because a prefusion stabilized MERS-CoV S antigen was found to elicit an improved antibody response [1109]. Moreover, the prefusion conformation offers an opportunity to target S2, a region of the S protein that accumulates mutations at a slower rate [21,22,1109] (see also [1]). Vaccine developers can stabilize the prefusion conformer by selecting versions of the S protein containing mutations that lock the position [1110]. The immune response to the spike protein when it is stabilized in this conformation is improved over other S structures [1111]. Thus, vaccines that use this prefusion stabilized conformation are expected to not only offer improved immunogenicity, but also be more resilient to the accumulation of mutations in SARS-CoV-2.

Another cross-platform consideration is the use of adjuvants. Adjuvants include a variety of molecules or larger microbial-related products that affect the immune system broadly or an immune response of interest. They can either be comprised of or contain immunostimulants or immunomodulators. Adjuvants are sometimes included within vaccines in order to enhance the immune response. Different adjuvants can regulate different types of immune responses, so the type or combination of adjuvants used in a vaccine will depend on both the type of vaccine and concern related to efficacy and safety. A variety of possible mechanisms for adjuvants have been investigated [1112,1113,1114].

6.5 COVID-19 Vaccine Development Platforms

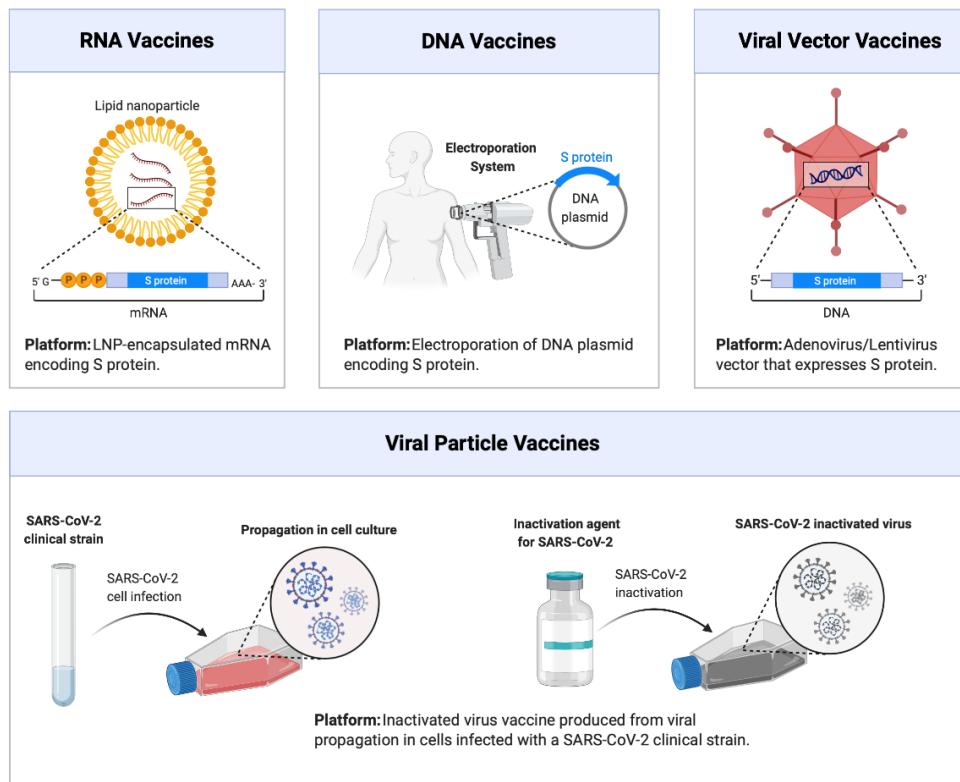


Figure 7: Vaccine Development Strategies. Several different strategies can and are being employed for the development of vaccines today. Each approach capitalizes on different features of the SARS-CoV-2 virus and delivery through a different platform. All of these approaches are being explored in the current pandemic.

The first administration of a dose of a COVID-19 vaccine to a human trial participant occurred on March 16, 2020 [1115,1116], marking an extremely rapid response to the emergence of SARS-CoV-2. Within a few weeks, at least 78 vaccine development programs were active [1116]. These programs employ a variety of technologies (Figure 2), ranging from established approaches to novel technologies that had never previously gone to market. As of May 6, 2022, 38 SARS-CoV-2 vaccines have been approved world wide and 27 are being administered throughout the world, with 12 billion doses administered across 223 countries. Many vaccines are available in only a subset of countries, and the types of vaccines available varies widely throughout the world. The status of individual vaccines continues to change and varies regionally.

6.5.1 Whole-Virus Vaccines

Whole-virus vaccines have the longest history among vaccine development approaches. Variolation, which is widely considered the first vaccination strategy in human history, is one example [1117,1118]. Famously employed against smallpox when healthy individuals were exposed to pus from an individual infected with what was believed to be either cowpox or horsepox [1117,1118,1119,1120], variolation allowed healthy individuals to be infected with a mild case of a disease. While whole-virus vaccines can confer adaptive immunity, they also face safety concerns [1119,1121,1122]. As of 2005, most vaccines still used whole-virus platforms [1123], and these technologies remain valuable tools in vaccine development today [1091]. Whole virus vaccine candidates have been developed for COVID-19 using both live attenuated viruses and inactivated whole viruses.

Table 2: Approved whole-virus vaccines [1124]

Vaccine	Company
Covaxin	Bharat Biotech
KoviVac	Chumakov Center
Turkovac	Health Institutes of Turkey
FAKHRAVAC (MIVAC)	Organization of Defensive Innovation and Research
QazVac	Research Institute for Biological Safety Problems (RIBSP)
KCONVAC	Shenzhen Kangtai Biological Products Co
COVIran Barekat	Shifa Pharmed Industrial Co
Covilo	Sinopharm (Beijing)
Inactivated (Vero Cells)	Sinopharm (Wuhan)
CoronaVac	Sinovac
VLA2001	Valneva

6.5.1.1 Live-Attenuated Virus Vaccines

Mechanism: LAV, also known as replication-competent vaccines, use a weakened, living version of a disease-causing virus or a version of a virus that is modified to induce an immune response [1101]. Whether variolation is the

first example of a live-attenuated virus (LAV) being used to induce immunity is debated [1091,1121], but subsequent efforts to incorporate attenuated viruses relied on either the identification of related viruses that were less virulent in humans (e.g., cowpox/horsepox or rotavirus vaccines) or culture of a virus *in vitro* [1091,1119]. Today, a virus can be attenuated by passaging it in a foreign host until, due to selection pressure, the virus loses its efficacy in the original host. Alternatively, selective gene deletion or codon de-optimization can be utilized to attenuate the virus [1101]. Foreign antigens can also be integrated into an attenuated viral vector [1125]. LAVs are also favored because they tend to be restricted to viral replication in the tissues around the location of inoculation [1126], and some can be administered intranasally [1101].

Prior Applications: The first deliberate (albeit pathogen-naïve) attempt to develop an attenuated viral vaccine dates back to Louis Pasteur in 1885. The next intentional LAVs were developed against the yellow fever virus in 1935 and influenza in 1936 [1126]. Today, LAVs are used globally to prevent diseases caused by viruses such as measles, rubella, polio, influenza, varicella zoster, and the yellow fever virus [1127]. There were attempts to develop LAVs against both SARS-CoV-1 and MERS-CoV [1128], but no vaccines were approved.

Application to COVID-19: LAVs have not been widely utilized against SARS-CoV-2 and COVID-19. All the same, there are at least five COVID-19 LAV candidates in the early (preclinical/phase I) stages of investigation. These candidates utilize different approaches. In one case, the vaccine delivers a noncleavable SARS-CoV-2 antigen prefusion conformation using a live-attenuated yellow fever virus [1125]. Several other candidates use codon-deoptimized SARS-CoV-2 [1129,1130,1131], leveraging the fact that different organisms display different biases in which synonymous codons are preferred to select codons that will be less optimal in the target organism without altering the amino acids encoded [1132]. Another is a chimeric vaccine that integrates genomic content from multiple viruses to create a more stable LAV [1133]. A final LAV being evaluated against COVID-19 was not specifically developed against SARS-CoV-2; instead, Bacillus Calmette-Guerin (BCG) vaccines were being investigated for the prophylaxis of COVID-19 [1134] because they are known to exert protective non-specific effects against other respiratory tract infections in *in vitro* and *in vivo* studies [1135]. However, a multicenter trial that randomly assigned participants 60 years and older to vaccination with BCG ($n = 1008$) or placebo ($n = 1006$) after 12 months of follow-up found that BCG vaccination had no effect on the incidence of SARS-CoV-2 or other respiratory infections [1136]. Despite these negative findings, BCG vaccination did induce a stronger antibody and cytokine response following COVID-19 infection.

Safety and Efficacy: Data is not yet available for human studies. In general, though safety associated with the production of LAVs was a major concern in the past, today manufacturers use safe and reliable methods to produce large quantities of vaccines once they have undergone rigorous preclinical studies and clinical trials to evaluate their safety and efficacy. However, one reason underlying the relatively slow emergence of LAV candidates against COVID-19 may be the risk presented to individuals who are immunocompromised [1137], which is an even greater concern when dealing

with a novel virus and disease. Additionally, it is generally recognized that LAVs induce an immune response similar to natural infection, and they are favored because they induce long-lasting and robust immunity that can protect from disease. This strong protective effect is induced in part by the immune response to the range of viral antigens available from LAV, which tend to be more immunogenic than those from non-replicating vaccines [1121,1128,1138]. Additional data are needed to ascertain how this technology performs in the case of SARS-CoV-2.

6.5.1.2 Inactivated Whole-Virus Vaccines

Mechanism: Inactivated whole-virus (IWV) vaccines are another well-established technology. These types of vaccines use full virus particles generally produced via cell culture that have been rendered non-infectious by chemical (i.e., formaldehyde or β -propiolactone [1139]) or physical (i.e., heat or ultraviolet radiation) means. In general, these vaccines mimic the key properties of the virus that stimulate a robust immune response, but the risk of adverse reactions is reduced because the virus is inactivated and thus unable to replicate. Though these viral particles are inactivated, they retain the capacity to prime the immune system. The size of the virus particle makes it ideal for uptake by an antigen-presenting cell (APC), which leads to the stimulation of helper T-cells [1140]. Additionally, the array of epitopes on the surface of the virus increases antibody binding efficiency [1140]. The native conformation of the surface proteins, which is also important for eliciting an immune response, is preserved using these techniques [1141]. Membrane proteins, which support B-cell responses to surface proteins, are also induced by this method [1142].

Prior Applications: IWV vaccines have been a valuable tool in efforts to control many viruses. Some targets of IWV vaccines have included influenza viruses, poliovirus, and hepatitis A virus. Inactivated vaccines are generally considered the fastest to generate once the pathogenic virus has been isolated and can be passaged in cell culture [1128], although this has not been the case for the COVID-19 pandemic. Past applications to HCoV have focused predominantly on SARS-CoV-1.

Preclinical studies have demonstrated that IWV SARS-CoV-1 vaccine candidates elicited immune responses *in vivo*. These vaccines generated neutralizing antibody titers at concentrations similar to those evoked by recombinant protein vaccines [1141,1143]. Studies in ferrets and non-human primates demonstrated that IWV vaccines can offer protection against infection due to neutralizing antibody and SARS-CoV-1-specific T cell responses [1144].

However, several attempts to develop IWV vaccines against both SARS-CoV-1 and MERS-CoV were hindered by incidences of vaccine-associated disease enhancement (VADE) in preclinical studies [1145]. In one example of a study in macaques, an inactivated SARS-CoV-1 vaccine induced even more severe lung damage than the virus due to an enhanced immune reaction [1146]. Independent studies in mice also demonstrated evidence of lung immunopathology due to VADE in response to MERS-CoV IWV vaccination [1147,1148]. The exact mechanisms responsible for VADE remain elusive due to the specificity of the virus-host interactions involved, but VADE is the

subject of investigation in preclinical SARS-CoV-2 vaccine studies to ensure the safety of any potential vaccines that may reach phase I trials and beyond [1145].

Application to COVID-19: Several whole-virus vaccines have been developed against COVID-19 and are available in countries around the world. As of May 6, 2022, 11 vaccines developed with IWV technology are being distributed in 114 countries (Figure 8). One, CoronaVac, was developed by Beijing-based biopharmaceutical company Sinovac. They inactivated a SARS-CoV-2 strain collected in China with β -propiolactone and propagated it using Vero cells [1128]. The vaccine is coupled with an aluminum adjuvant [1128]. In phase I and II clinical trials, CoronaVac elicited a strong immunogenic response in animal models and the development of neutralizing antibodies in human participants [1149,1150,1151]. Administration followed a prime-boost regimen using a 0.5 ml dose containing 3 μ g of inactivated SARS-CoV-2 virus per dose [1152]. Results from a two-dose phase III trial following a 14-day prime boost became available in late 2020 [1153], and an interim analysis identified specific IgG neutralizing antibodies against S1-RBD and a robust IFN- γ secreting T cell response was induced via immunization with CoronaVac [1154]. CoronaVac was approved for use in China and has been granted emergency use in XX countries, including Brazil, Cambodia, Chile, Colombia, Laos, Malaysia, Mexico, Turkey, Ukraine, and Uruguay [82]. In August 2021, Sinovac reported that they had produced over a billion doses of CoronaVac [1155].

Similarly, two inactivated vaccine candidates were developed following a similar approach by the state-owned China National Pharmaceutical Group Co., Ltd., more commonly known as Sinopharm CNBG. Their BBIBP-CorV vaccine was developed in Beijing using the HB02 strain of SARS-CoV-2. At their Wuhan Institute, they developed a second vaccine using the WIV04 strain of SARS-CoV-2 [1156]. The viruses were purified, propagated using Vero cells, isolated, and inactivated using β -propiolactone [1156,1157]. These vaccines are adjuvanted with aluminum hydroxide [1156,1157]. Preclinical studies indicated that the BBIBP-CorV vaccine induced sufficient neutralizing antibody titers in mice, and a prime-boost immunization scheme of 2 μ g/dose was sufficient to protect rhesus macaques from disease [1157]. For the other vaccine, neutralizing antibodies were detected in all groups 14 days after the final dose in the phase I part of the trial [1158], with similar findings reported in interim phase II data [1158].

Number of inactivated vaccines available worldwide

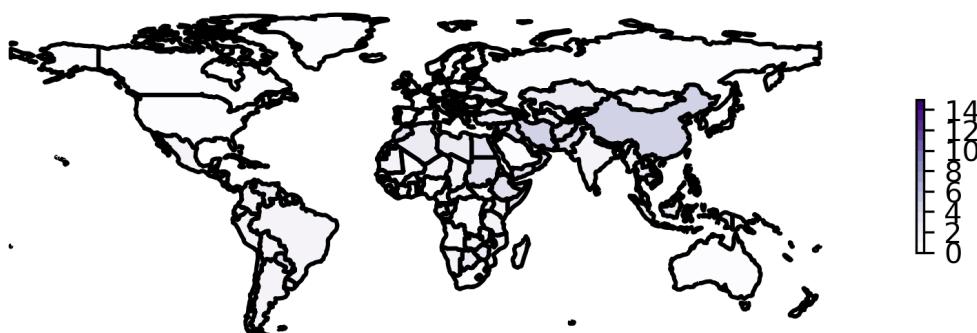


Figure 8: Worldwide availability of vaccines developed using inactivated whole viruses. This figure reflects the number of vaccines based on whole inactivated virus

technology that were available in each country as of May 6, 2022. These data are retrieved from Our World in Data <!-To Do: Cite-> and plotted using geopandas. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

Other programs have been led through industry partnerships with governmental organizations. Another IWV vaccine comes from India, where Bharat Biotech International Ltd., which is the biggest producer of vaccines globally, Bharat Biotech International Ltd., collaborated with the Indian Council of Medical Research (ICMR) - National Institute of Virology (NIV) to develop COVAXIN®, also referred to as BBV152. Preclinical studies of COVAXIN® in hamsters [1159] and macaques [1160] indicated that the vaccine induced protective responses deemed sufficient to move forward to human trials. Phase I and phase I/II studies indicated that COVAXIN® adjuvanted with alum and a Toll-like receptor 7/8 (TLR7/8) agonist was safe and immunogenic and that it induced Th1-skewed memory T-cell responses [1161,1162]. As of September 2021, COVAXIN® has been approved for emergency use in Guyana, India, Iran, Zimbabwe, and Nepal, Mauritius, Mexico, Nepal, Paraguay, and the Philippines [1163].

Trial Safety and Efficacy: In general, IWV vaccine candidates have been well-tolerated in clinical trials. Safety analysis of the CoronaVac vaccine during the phase II trial revealed that most adverse reactions were either mild (grade 1) or moderate (grade 2) in severity. In adults aged 18 to 59 years receiving a variety of dosage schedules, site injection pain was consistently the most common symptom reported [1151]. In older adults, the most common local and systemic reactions were pain at the injection site (9%) and fever (3%), respectively [1149]. In phase III trials, minimal side effects were reported [1153]. For COVAXIN, only mild to moderate side-effects reported upon immunization [1161,1162], and in phase II trials, the BBIBP-CorV vaccine appeared well-tolerated, with 23% of participants in the vaccine condition (482 total participants, 3:1, vaccine:placebo) reporting at least one adverse reaction characterized as mild to moderate [1164]. However, both CoronaVac and SinoPharm's WIV04 vaccine trials were affected by concerns about adverse events. In CoronaVac's trial of adults 18-59, 2% (n=7) of participants reported severe adverse events [1149], causing the trial to be halted for investigation [1165]. They were determined to be unrelated to the vaccine [1149]; [1165]], which is now widely distributed. Similarly, a trial of the SinoPharm WIV04 vaccine in Peru was briefly paused due to safety concerns in relation to neurological symptoms [1166], but this was later deemed unrelated to the vaccine, and the trial continued [1167].

In terms of efficacy, estimates of IWV vaccine efficacy during phase III trials varied widely, and in some cases, even estimates for a single vaccine candidate differed across analyses. In phase III trials, Sinopharm CNBG's BBIBP-CorV vaccine made from the WIV04 strain achieved an efficacy of 72.8% and was well tolerated [1168]. In July 2021, COVAXIN's overall vaccine efficacy was estimated at 77.8% for the prevention of COVID-19 based on a final enrollment of 25,798 people (~1:1 vaccine:placebo) [1169]. Sinopharm affiliates in the UAE in early December 2020 claimed the vaccine had 86% efficacy, which was later at odds with a Sinopharm Beijing affiliate that stated that the BBIBP-CorV vaccine had a 79.34% efficacy later that same month [1170]. CoronaVac demonstrated an efficacy of a little over 50% in Brazil, which was contested by Turkish officials claiming an efficacy of 91.25%, but

ultimately after multiple announcements, the efficacy debate was settled at over 50% [1171,1172]. Subsequently, an interim analysis of the phase III randomized placebo-controlled trials conducted in Turkey enrolling 10,214 participants (~2:1 vaccine:placebo) indicated efficacy of 83.5%, with minimal side effects reported [1153], and a prospective national cohort study in Chile reported an adjusted estimated effectiveness of 66% for the prevention of COVID-19 with an estimated 90% and 87% prevention of hospitalization and death, respectively [1173]. Therefore, it is difficult to ascribe a particular efficacy to these vaccines given the variation in reports.

Real-World Safety and Efficacy: One of the major limitations of IJV vaccines is their susceptibility to losing efficacy due to mutations in the epitopes of the circulating virus [1122]. This loss of specificity over time is likely to be influenced by the evolution of the virus, and specifically by the rate of evolution in the region of the genome that codes for the antigen. The beta variant appears to be more resistant to neutralizing antibodies in sera from individuals immunized with Sinovac than the alpha variant or wildtype virus, indicating that emerging variants may be of concern [1174]. In agreement with previous studies demonstrating sera from individuals vaccinated with COVAXIN® efficiently neutralized the alpha variant (B.1.1.7) and the delta variant (B.1.617.2) [1175,1176,1177], the phase III trial reported a 65.2% efficacy against the delta variant (B.1.617.2) [1169]. However, studies suggested the beta variant was more resistant (compared to the wildtype and alpha variants) to neutralizing antibodies in sera from individuals immunized with Sinovac [1174].

Indeed, another preprint determined that sera from individuals immunized with COVAXIN® had effective neutralizing antibodies against the delta variant and the so-called delta plus variant (AY.1) [1175]. Notably, a preprint reported that antisera from 12 people immunized with BBIBP-CorV exhibited neutralizing antibody capacity against the beta variant (B.1.351), wild type SARS-CoV-2 (NB02), and one of the original variants of SARS-CoV-2 (D614G) [1178]. Another preprint including sera from 282 participants used a surrogate neutralizing assay, a test that generally correlates with neutralizing antibodies, to determine that BBIBP-CorV appears to induce neutralizing antibodies against the binding of the RBD of wild type SARS-CoV-2 and the alpha, beta, and delta variants to ACE2 [1179]. Indeed, a study in *The New England Journal of Medicine* showed that the alpha variant exhibited very little resistance to neutralization by sera of those immunized with BBIBP-CorV, but the beta variant was more resistant to neutralization by almost a factor of 3 [1174]. The authors noted that no evidence of VADE was detected using this vaccine in phase II data [1158].

However, concern was raised about the efficacy of CoronaVac following reports that over 350 doctors became ill with COVID-19 in Indonesia despite being immunized with CoronaVac [1180]. In addition to concerns raised by the evolution of SARS-CoV-2, it is important to consider the duration of immunity over time. Studies are underway to determine whether a booster immunization is required for several IJV vaccines, including CoronaVac [1181] and COVAXIN [1182]. A phase I/II clinical trial of CoronaVac in an elderly cohort (adults 60 years and older) in China determined that by 6 to 8 months following the second dose, neutralizing antibody titers were detected below the seropositive cutoff [1183]. One preprint has reported that 6 months after the second vaccination, a booster dose of CoronaVac markedly

increased geometric mean titers of SARS-CoV-2 neutralizing antibodies [1184]. However, the reduction of neutralizing antibodies was ameliorated by a booster dose administered 8 months after the second CoronaVac dose.

A preprint study of healthcare workers in China has since indicated that a booster shot of BBIBP-CorV elevates B cell and T cell responses and increases neutralizing antibody titers [1185]. In May 2021, the UAE announced it would consider booster shots for all citizens who had been immunized with BBIBP-CorV, which was shortly followed by a similar announcement in Bahrain, and by August 29th, 2021, the UAE mandated booster shots for all residents who had received BBIBP-CorV [1155]. Additionally, heterogeneous vaccine boosters are also being considered in many cases. Chinese [1186] and Chilean [1187] researchers have opted to investigate options to administer different vaccines (e.g., an mRNA vaccine dose) as a booster dose to individuals who have already received two doses of CoronaVac. Another study has already determined that using the CanSino vaccine (Convidecia) instead of CoronaVac in a prime-boost vaccination regimen can induce a more robust immune response [1188]. Today, booster immunization is suggested for several whole-virus vaccines.

6.5.2 Subunit Vaccines

Table 3: Approved subunit vaccines [1124]

Vaccine	Company	Platform
Zifivax	Anhui Zhifei Longcom	protein subunit
Noora vaccine	Bagheiat-allah University of Medical Sciences	protein subunit
Corbevax	Biological E Limited	protein subunit
Abdala	Center for Genetic Engineering and Biotechnology (CIGB)	protein subunit
Soberana 02	Instituto Finlay de Vacunas Cuba	protein subunit
Soberana Plus	Instituto Finlay de Vacunas Cuba	protein subunit
Covifenz	Medicago	VLP
MVC-COV1901	Medigen	protein subunit
Recombinant SARS-CoV-2 Vaccine (CHO Cell)	National Vaccine and Serum Institute	protein subunit
Nuvaxovid	Novavax	protein subunit
Razi Cov Pars	Razi Vaccine and Serum Research Institute	protein subunit
COVOVAX (Novavax formulation)	Serum Institute of India	protein subunit

Vaccine	Company	Platform
TAK-019 (Novavax formulation)	Takeda	protein subunit
SpikoGen	Vaxine/CinnaGen Co.	protein subunit
Aurora-CoV	Vector State Research Center of Virology and Biotechnology	protein subunit
EpiVacCorona	Vector State Research Center of Virology and Biotechnology	protein subunit

Efforts to overcome the limitations of live-virus vaccines led to the development of approaches first to inactivate viruses (circa 1900), leading to I WV vaccines, and then to purify proteins from viruses cultured in eggs (circa 1920) [1091,1189]. The purification of proteins led to the emergence of subunit vaccines. Today, such approaches may use antigens isolated from the surface of the viral particle that are key targets of the immune system (protein subunit vaccines), but advances in biological engineering have also facilitated the development of approaches like viral-like particle (VLP) vaccines using nanotechnology [1190].

Mechanism: Unlike whole-virus vaccines, which introduce the whole virus, subunit vaccines stimulate the immune system by introducing one or more proteins or peptides of the virus that have been isolated. The main advantage of this platform is that subunit vaccines are considered very safe, as the antigen alone cannot cause an infection [1191]. Both protein subunit and VLP vaccines thus mimic the principle of whole virus vaccines but lack the genetic material required for replication, removing the risk of infection [1192]. Protein subunit vaccines can stimulate antibodies and CD4⁺ T-cell responses [1193,1194]. This platform is also favored for its consistency in production and defined components designed for a highly targeted immune response to a specific pathogen using synthetic immunogenic particles that can be designed to avoid allergen and reactogenic sequences [1195]. The immune response generated by protein subunit vaccines is weaker, and adjuvants are usually required to boost the response [1196] (see Appendix). These adjuvants are immunogenic substances, which include, for example, alum (aluminum hydroxide), squalene- or saponin-based adjuvants, and Freund's incomplete/complete adjuvants [1195,1197].

Prior Applications: Prior protein subunit vaccine development efforts for both SARS-CoV-1 and MERS-CoV have mostly focused on the immunogenic RBD of the S protein [1198,1199,1200]. There have been several approaches investigated in the search for a potential SARS-CoV-1 vaccine, including vaccines targeting the full-length or trimeric S protein [1201,1202], those focused on the RBD protein only [1198,1199,1200,1203] or non-RBD S protein fragments [1202,1204], as well as the N and M proteins [1205,1206,1207]; these efforts have been thoroughly reviewed elsewhere [1208]. There have been examples of success in preclinical research including candidate RBD219N-1, a 218-amino-acid residue of the SARS-CoV-1 RBD that, when adjuvanted to aluminum hydroxide, was capable of eliciting a high RBD-specific and neutralizing antibody response in both pseudovirus and live virus infections of immunized mice [1209]. Several subunit-based approaches have also been used to investigate potential vaccines against MERS. Other

strategies investigating the potential use of the full length S DNA have also been investigated in mice and rhesus macaques, which elicited immune responses [1210], but these responses were not as effective as the combination of S DNA and the S1 subunit protein together [1210,1211]. Similarly to the SARS-CoV-1 vaccine candidates, the MERS-CoV protein subunit vaccine candidates generally target the RBD [1199,1208,1212,1213,1214,1215], with some targeting the full length S protein [1109], non-RBD protein fragments such as the SP3 peptide [1216], and the recombinant N-terminal domain (rNTD) [1217]. No protein subunit vaccine for MERS-CoV has progressed beyond preclinical research to date. VLPs have been investigated for development of vaccines against MERS and SARS [1218,1219] including testing in animal models [1220,1221], but once again, only preclinical data against HCoV has been collected [1222]. However, protein subunit vaccines do play a role in public health and have contributed to vaccination against hepatitis B [1223] and pertussis [1224,1225] since the 1980s and will likely continue to contribute to public health for the foreseeable future due to ongoing research in vaccines against influenza, SARS-CoV-2, Epstein-Barr virus, dengue virus, and human papillomavirus among others [1226,1227,1228].

Application to COVID-19: The development of protein subunit vaccines against SARS-CoV-2 is a remarkable achievement given the short period of time since the emergence of SARS-CoV-2 in 2019, particularly considering these types of vaccines have not played a major role in previous pandemics. More than 20 protein subunit vaccines from companies such as Sanofi/GlaxoSmithKline, Nanogen, and the Serum Institute of India have entered clinical trials for COVID-19 since the beginning of the pandemic [1227] and 15 are being administered worldwide [1229]. VLP vaccines have not progressed as rapidly, with only 1 VLP vaccines approved [1227] as of May 6, 2022. Most of these vaccines are designed using either the full-length S protein or the RBD of the S protein specifically as an antigen, although some use several different SARS-CoV-2 proteins [1191]. As of March 30, 2022, 14 protein subunit vaccines are being distributed in 21 countries (Figure 4).

Number of protein subunit vaccines available worldwide



Figure 9: Worldwide availability of vaccines developed using protein subunit. This figure reflects the number of vaccines based on protein subunit technology that were available in each country as of May 6, 2022. These data are retrieved from Our World in Data <!--To Do: Cite--> and plotted using geopandas. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

One of the most prominent protein subunit vaccines against SARS-CoV-2 thus far is NVX-CoV2373 or Nuvaxovid, which is produced by U.S. company Novavax and partners. NVX-CoV2373 is a protein nanoparticle vaccine constructed from a mutated prefusion SARS-CoV-2 spike protein in combination with a specialized saponin-based adjuvant to elicit an immune response against SARS-CoV-2. The spike protein is recombinantly expressed in Sf9 insect cells [1230], which have previously been used for several other FDA-approved protein therapeutics [1231], and contains mutations in the furin cleavage site (682-RRAR-685 to 682-QQAQ-685) along with two proline substitutions (K986P and V987P) that improve thermostability [1230]. In preclinical mouse models, Novavax-CoV2373 elicited high anti-spike IgG titers 21 to 28 days post-vaccination that could neutralize the SARS-CoV-2 virus and protect the animals against viral challenge, with particularly strong effects when administered with the proprietary adjuvant Matrix-M™ [1230]. In a phase I/II trial, a two-dose regimen of NVX-CoV2373 was found to induce anti-spike IgG levels and neutralizing antibody-titers exceeding those observed in convalescent plasma donated by symptomatic patients [1232]. In line with the preclinical studies, the use of Matrix-M adjuvant further increased anti-spike immunoglobulin levels and induced a Th1 response. In a phase III randomized, observer-blinded, placebo-controlled clinical trial in 14,039 participants, two 5-µg doses of NVX-CoV2373 or placebo were administered 21 days apart in a 1:1 ratio from late September to late November 2020 [1233]. Novavax has since been approved in several places, including the United Kingdom [1234] and the E.U. [1235] and applied for an EUA from the FDA in early 2022 [1236]. Although Novavax promised to provide up to 1.1 billion doses to COVAX; however, there have been reports of struggle to maintain production capacity and quality, hindering its production targets [1237].

The leading example of a VLP approach applied to COVID-19 comes from Covifenz, a VLP vaccine developed by Canadian company Medicago [1238]. This vaccine was developed using plant-based VLP technology [1239] that the company had been investigating in order to develop a high-throughput quadrivalent VLP platform to provide protection against influenza [1240]. The approach utilizes *Nicotiana benthamiana*, an Australian relative of the tobacco plant, as an upstream bioreactor [1240,1241,1241]. Specifically, the S gene from SARS-CoV-2 in its prefusion conformation is inserted into a bacterial vector (*Agrobacterium tumefaciens*) that then infects the plant cells [1240,1241]. Expression of the S glycoprotein causes the production of VLPs composed of S trimers anchored in a lipid envelope that accumulate between the plasma membrane and the cell wall of the plant cell [1241]. Because these VLPs do not contain the SARS-CoV-2 genome, they offer similar advantages to while mitigating the risks of whole-virus vaccines [1240,1241].

In the Phase I study, Covifenz was administered to 180 Canadian adults 18-55 years old as two doses, 21 days apart, with three different dosages evaluated [1241]. This study reported that when the VLPs were administered with an adjuvant, the vaccine elicited a neutralizing antibody that was significantly (approximately 10 times) higher than that in convalescent sera [1241]. They also reported a cellular immune response was induced. Based on these findings, phase II and III trials began.

The findings of the phase III trial were published a few months later in the *New England Journal of Medicine* [1242]. This study examined 24,141 adults assigned to the treatment and control conditions at a 1:1 ratio between March and September of 2021. Participants were recruited from countries in North and South America, as well as the United Kingdom. Approximately 10,000 individuals in each condition completed both doses and were evaluated in the per-protocol population. Data was submitted by Medicago and GSK to Health Canada and the vaccine was approved for use in adults ages 18 to 65 in February 2022 [1243].

Plant-based expression systems such as this are relatively new [1241] but are likely to offer unparalleled feasibility at scale given the speed and low-cost associated with the platform [1244]. Additionally, it can be stored at 2 to 8°C. However, the worldwide footprint of Covifenz, and of VLP-based technologies against SARS-CoV-2 broadly, remains small, with only 1 VLP vaccine approved for distribution in 0 countries (Figure 10). Approval and administration of Covifenz in countries outside of Canada has been limited by concerns at the WHO about ties between Medicago and the tobacco industry [1238,1245]. While other species of plants have been explored as the upstream bioreactors for plant-derived VLPs, the use of the specific species of tobacco used here increased yield dramatically [1246]. Therefore, it may be feasible to identify other species of plants that can be used for future vaccines, but the selection of *N. benthamiana* was not arbitrary.

Number of VLP vaccines available worldwide



Figure 10: Worldwide availability of vaccines developed with VLPs. This figure reflects the number of vaccines based on VLP technology that were available in each country as of May 6, 2022. These data are retrieved from Our World in Data <!-To Do: Cite-> and plotted using geopandas. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

Trial Safety and Efficacy: In the phase III trial, the efficacy of Novavax was reported to be 89.7%, with a total of 10 patients developing COVID-19 in the vaccine group versus 96 in the placebo group [1233]. No hospitalizations or deaths were reported in the vaccine group. An additional phase III randomized, observer-blinded, placebo-controlled trial was conducted in the U.S. and Mexico, enrolling 29,949 participants and administering at least 1 vaccine in a 2:1 ratio from late December 2020 to late February 2021 with the same primary endpoints as the U.K. trial [1247]. A vaccine efficacy of 90.4% was reported based on 77 cases total, 63 of which occurred in the placebo group. All moderate to severe cases of COVID-19 occurred in the placebo group. Additionally, in both trials, the vaccine was found to be well-tolerated

[1233,1247]. The conclusions of both trials indicate that the NVX-CoV2373 vaccine is safe and effective against COVID-19, and those who received this vaccine through the trials are considered fully vaccinated [1248].

Similarly, Covifenz was reported to be 71% effective in preventing COVID-19 in the per-protocol analysis [1249]. Efficacy was only slightly lower in the intention-to-treat group at 69%. Prevention of moderate-to-severe disease was estimated at 78.8% in the intention-to-treat group. Over 24,000 participants were included in the safety analysis, which reported that 92.3% of vaccine recipients reported local adverse events compared to 45.5% of placebo recipients, with rates for systemic adverse events at 87.3% and 65.0%, respectively. However, the AEs reported were generally mild to moderate. Only three patients (two in the vaccine group) reported grade 4 events, all after the second dose. The most common AEs were, for local events, injection site pain and, for systemic events, headache, myalgia, fatigue, and general discomfort.

Real-World Safety and Efficacy: To date, data about the effect of viral evolution on the efficacy of subunit vaccines has been limited. *Post hoc* analysis in the phase III trial determined that the NovaVax vaccine had an efficacy of 86.3% against the Alpha variant (identified based on the presence/absence of the 69–70del polymorphism) and 96.4% against other variants [1233]. In the second phase III trial [1247], whole-genome sequencing was obtained from 61 of the 77 cases, and 79% of infections were identified as a VOC or VOI that had been characterized at the time of the study. Vaccine efficacy against cases caused by VOC, among which the Alpha variant was predominant (88.6%), was reported to be 92.6% [1247]. In late 2020, an analysis of efficacy in South African adults revealed an overall efficacy of 60.1% and a slightly lower efficacy of 50.1% against B.1.351 in particular [1250].

It has also been reported that Novavax initiated booster trials in the U.K. [1155].

Because the Covifenz results are so new (May 4, 2022), limited data is available since the publication of phase III trial results [1249]. However, it should be noted that the Covifenz trials were conducted in 2021, at a time during which the B.1.617.2 (delta) and P.1 (gamma) variants were predominant [1249]. Genomic analysis of 122 out of 176 cases (165 in the per-protocol population) revealed that none of the COVID-19 cases reported were caused by the original Wuhan strain. Instead, 45.9% of cases were identified as the delta variant, 43.4% as gamma, 4.9% as alpha, and 5.8% as VOIs. Therefore, the efficacy data from this phase III trial may be lower than it would have been if the trial had occurred earlier in the course of SARS-CoV-2's evolution given that the S glycoprotein expressed in the VLPs was isolated from a 2020 sample of SARS-CoV-2 [1249].

6.5.3 Nucleic Acid Vaccines

While traditional methods of vaccine development such as inactivated whole viruses are still used today (Figure 2), biomedical research in the 21st century has been significantly influenced by the genomic revolution, and vaccine development is no exception. The shift towards omics-based approaches to

vaccine development began to take hold with the development of the meningococcal type B vaccine using reverse vaccinology in the early 2010s [1251,1252]. In this way, the genomic revolution catalyzed a fundamental shift in the development of vaccines. These vaccine technologies could potentially provide a future approach to addressing one of the major limitations of vaccines today due to their potential to function therapeutically rather than just prophylactically [1253].

Nucleic-acid based approaches are all based on the shared underlying principle that utilizing a vector to deliver the information to produce an antigen can trigger an immune response to the antigen without introducing an infectious agent. Such approaches build on subunit vaccination strategies, where a component of a vaccine (e.g., an antigenic protein) is delivered. Platforms based on genomic sequencing began to be explored beginning in the 1980s as genetic research became increasingly feasible. Advances in genetic engineering allowed for gene sequences of specific viral antigens to be grown *in vitro* [1091]. Studies also demonstrated that model organisms could be induced to construct antigens that would trigger an immune response [1123,1254,1255]. These two developments meant that it could be possible to identify any or all of the antigens encoded by a virus's genome and train the immune response to recognize them. In nucleic-acid-based approaches, the genome of a pathogen is screened to identify potential vaccine targets [1252], and pathogens of interest are then expressed *in vitro* and tested in animal models to determine their immunogenicity [1252]. By inducing the host to express the antigen, such vaccines can activate immune pathways via both MHC I and MHC II [1256] instead of MHC II alone as with prior technologies [1255], meaning that both humoral and cellular immunity are activated [1098]. Thus, in addition to lacking an infectious agent, these approaches are likely to offer several advantages over more traditional immunization platforms because they can stimulate both B- and T-cell responses [1098,1257].

The delivery and presentation of antigens is fundamental to inducing immunity against a virus. Vaccines that deliver nucleic acids allow the introduction of foreign substances to the body to induce both humoral and cellular immune responses [1098]. Delivering a nucleic acid sequence to host cells allows the host to synthesize an antigen without exposure to a viral threat [1098]. Host-synthesized antigens can then be presented in complex with major histocompatibility complex (MHC) I and II, which can activate either T- or B-cells [1098]. While these vaccines encode specific proteins, providing many of the benefits of a protein subunit vaccine, they do not carry any risk of DNA being live, replicating, or spreading, and their manufacturing process lends itself to scalability [1098]. Here, opportunities can be framed in terms of the central dogma of genetics: instead of directly providing the proteins from the infectious agents, vaccines developers are exploring the potential for the delivery of DNA or RNA to induce the cell to produce proteins from the virus that in turn induce a host immune response.

6.5.3.1 DNA Vaccines

Nucleic acid information can be delivered antigen information to host cells using DNA. However, early attempts to use these technologies to develop vaccines revealed that DNA translated poorly to humans due to low

immunogenicity [1123,1255,1258]. Initially, concerns were raised that DNA vaccines might bind to the host genome or induce autoimmune disease [1098,1256], but pre-clinical and clinical studies have consistently disproved this hypothesis and indicated DNA vaccines to be safe [1258]. Many of the safety concerns raised about DNA vaccines were not found to be an issue during preclinical and phase I testing, although antibiotic resistance introduced during the plasmid selection process remained a concern during this initial phase of development [1098]. However, the immunogenicity of these vaccines has also not reached expectations [1098]. Despite initially disappointing immunogenicity in clinical trials [1255], a number of developments during the 2010s led to greater efficacy of DNA vaccines [1098]. However, no DNA vaccines had been approved for use in humans prior to the COVID-19 pandemic [1258,1259].

Table 4: Approved DNA vaccines [1124]

Vaccine	Company	Platform
Convidecia	CanSino	non replicating viral vector
Gam-COVID-Vac	Gamaleya	non replicating viral vector
Sputnik Light	Gamaleya	non replicating viral vector
Sputnik V	Gamaleya	non replicating viral vector
Ad26.COV2.S	Janssen (Johnson & Johnson)	non replicating viral vector
Vaxzevria	Oxford/AstraZeneca	non replicating viral vector
Covishield (Oxford/ AstraZeneca formulation)	Serum Institute of India	non replicating viral vector
ZyCoV-D	Zydus Cadila	plasmid vectored

6.5.3.2 Plasmid-Vectored DNA Vaccines

Mechanism: Many DNA vaccines use a plasmid vector-based approach, where the sequence encoding the antigen(s) against which an immune response is sought can be cultivated in a plasmid and delivered directly to an appropriate tissue [1260]. Plasmids can also be designed to act as adjuvants by encoding molecules that supplement the immune response, such as immune stimulant molecules [1256]. The DNA itself may also stimulate the innate immune response [1255]. Once the plasmid brings the DNA sequence to an APC, the host machinery can be used to construct antigen(s) from the transported genetic material, and the body can then synthesize antibodies in response [1098]. In the 1990s and 2000s, DNA vaccines delivered via plasmids sparked significant scientific interest, leading to a large number of preclinical trials [1098].

Prior Applications: Early preclinical trials primarily focused on long-standing disease threats, including viral diseases such as rabies and bacterial diseases such as malaria, and promising results led to phase I testing of the application of this technology to HIV, influenza, malaria, and other diseases of concern during this period [1098]. Although they were well-tolerated, these early attempts to develop vaccines were generally not very successful in inducing immunity to the target pathogen, with either limited T-cell or limited neutralizing antibody responses observed [1098].

Applications to COVID-19: Currently, a Phase I safety and immunogenicity clinical trial of INO-4800, a prophylactic vaccine against SARS-CoV-2, is underway [1261]. The vaccine developer Inovio Pharmaceuticals Technology is overseeing administration of INO-4800 by intradermal injection followed by electroporation with the CELLECTRA® device to healthy volunteers. Electroporation is the application of brief electric pulses to tissues in order to permeabilize cell membranes in a transient and reversible manner. It has been shown that electroporation can enhance vaccine efficacy by up to 100-fold, as measured by increases in antigen-specific antibody titers [1262]. The safety of the CELLECTRA® device has been studied for over seven years, and these studies support the further development of electroporation as a safe vaccine delivery method [1263]. The temporary formation of pores through electroporation facilitates the successful transportation of macromolecules into cells, allowing cells to robustly take up INO-4800 for the production of an antibody response. Approved by the United States (U.S.) FDA on April 6, 2020, the phase I study is enrolling up to 40 healthy adult volunteers in Philadelphia, PA at the Perelman School of Medicine and at the Center for Pharmaceutical Research in Kansas City, MO. The trial has two experimental arms corresponding to the two locations. Participants in Experimental Group 1 will receive one intradermal injection of 1.0 milligram (mg) of INO-4800 followed by electroporation using the CELLECTRA® 2000 device twice, administered at Day 0 and Week 4. Participants in Experimental Group 2 will receive two intradermal injections of 1.0 mg (total 2.0 mg per dosing visit) of INO-4800 followed by electroporation using the CELLECTRA® 2000 device, administered at Day 0 and Week 4. Safety data and the initial immune responses of participants from the trial are expected by the end of the summer of 2021. The development of a DNA vaccine against SARS-CoV-2 by Inovio could be an important step forward in the world's search for a COVID-19 vaccine. Although exciting, the cost of vaccine manufacturing and electroporation may make scaling the use of this technology for prophylactic use for the general public difficult.

6.5.3.3 Viral-Vectored DNA Vaccines

Mechanism: Plasmids are not the only vector that can be used to deliver sequences associated with viral antigens: genetic material from the target virus can also be delivered using a second virus as a vector. Viral vectors have emerged as a safe and efficient method to furnish the nucleotide sequences of an antigen to the immune system using a second virus as a vector [1264]. The genetic content of the vector virus is often altered to prevent it from replicating, but replication-competent viruses can also be used under certain circumstances [1265]. Once the plasmid or viral vector brings the DNA sequence to an APC, the host machinery can be used to construct antigen(s) from the transported genetic material, and the body can then synthesize

antibodies in response [1098]. These vaccines can be either replicating or non-replicating [1137]. One of the early viral vectors explored was adenovirus, with serotype 5 (Ad5) being particularly effective [1098]. This technology rose in popularity during the 2000s due to its being more immunogenic in humans and non-human primates than plasmid-vectored DNA vaccines [1098]. In the 2000s, interest also arose in utilizing simian adenoviruses as vectors because of the reduced risk that human vaccine recipients would have prior exposure resulting in adaptive immunity [1098,1266], and chimpanzee adenoviruses were explored as a potential vector in the development of a vaccine against *Middle East respiratory syndrome-related coronavirus* (MERS-CoV) [1267]. Today, various viral-vector platforms including poxviruses [1268,1269], adenoviruses [1270], and vesicular stomatitis viruses [1271,1272] are being developed. Viral-vector vaccines are able to induce both an antibody and cellular response; however, the response is limited due to the immunogenicity of the viral vector used [1270,1273]. An important consideration in identifying potential vectors is the immune response to the vector. Both the innate and adaptive immune responses can potentially respond to the vector, limiting the ability of the vaccine to transfer information to the immune system [1274]. Different vectors are associated with different levels of reactogenicity; for example, adenoviruses elicit a much stronger innate immune response than replication deficient adeno-associated viruses derived from parvoviruses [1274]. Additionally, using a virus circulating widely in human populations as a vector presents additional challenges because vaccine recipients may already have developed an immune response to the vector [1275].

Prior Applications: There are several viral vector vaccines that are available for veterinary use [1098,1276], but prior to the COVID-19 pandemic, only one viral vector vaccine was approved by the FDA for use in humans. This vaccine is vectored with a recombinant vesicular stomatitis virus and targeted against the ebola virus [1277]. Additionally, several phase I and phase II clinical trials for other vaccines are ongoing [1264], and the technology is currently being explored for its potential against numerous infectious diseases including malaria [1278,1279], ebola [1280,1281,1282], and human immunodeficiency virus (HIV) [1283,1284]. The threat of MERS and SARS initiated interest in the application of viral vector vaccines to human coronaviruses [1267], but efforts to apply this technology to these pathogens had not yet led to a successful vaccine candidate. In the mid-to-late 00s, adenoviral vectored vaccines against SARS were found to induce SARS-CoV-specific IgA in the lungs of mice [1285], but were later found to offer incomplete protection in ferret models [1286]. Gamaleya National Center of Epidemiology and Microbiology in Moscow sought to use an adenovirus platform for the development of vaccines for *Middle East respiratory syndrome-related coronavirus* and Ebola virus, although neither of the previous vaccines were internationally licensed [1287].

In 2017, results were published from an initial investigation of two vaccine candidates against MERS-CoV containing the MERS-CoV S gene vectored with chimpanzee adenovirus, Oxford University #1 (ChAdOx1), a replication-deficient chimpanzee adenovirus [1288]. This study reported that a candidate containing the complete spike protein sequence induced a stronger neutralizing antibody response in mice than candidates vectored with modified vaccinia virus Ankara. It was pursued in additional research, and in

the summer of 2020 results of two studies were published. The first reported that a single dose of ChAdOx1 MERS induced an immune response and inhibited viral replication in macaques [1289]. The second reported promising results from a phase I trial that administered the vaccine to adults and measured safety/tolerability and immune response (as indicated through immune assays following vaccination) [1290].

Application to COVID-19: While not all of these results were available at the time that vaccine development programs against SARS-CoV-2 began, at least three viral vector vaccines have also been developed against this hCoV. First, collaboration between AstraZeneca and researchers at the University of Oxford has successfully applied a viral vector approach to the development of a vaccine against SARS-CoV-2 using the replication-deficient ChAdOx1 vector modified to encode the spike protein of SARS-CoV-2 [1291]. In phase I and I/II trials, respectively, the immunogenic potential of vaccine candidate ChAdOx1 nCoV-19 was demonstrated through the immune challenge of two animal models, mice and rhesus macaques [1291] and patients receiving the ChAdOx1 nCoV-19 vaccine developed antibodies to the SARS-CoV-2 spike protein that peaked by day 28, with these levels remaining stable until a second observation at day 56 [1292].

Second, a viral vector approach was also applied by Gamaleya to develop Sputnik V, a replication-deficient recombinant adenovirus (rAd) vaccine that combines two adenovirus vectors, rAd26-S and rAd5-S, that express the full-length SARS-CoV-2 spike glycoprotein. The two vectors are administered intramuscularly administered sequentially, following a prime-boost regimen. Despite a lack of data from clinical trials, President Vladimir Putin of Russia announced the approval of the Sputnik V vaccines on August 11th, 2020 [1293] and it has subsequently been administered in Russia and other countries.

Third, Janssen Pharmaceuticals, Inc., a subsidiary of Johnson & Johnson, also developed a viral vector vaccine in collaboration with and funded by the United States's "Operation Warp Speed" [1294,1295]. The vaccine candidate JNJ-78436735, formerly known as Ad26.COV2-S, is a monovalent vaccine that is composed of a replication-deficient adenovirus serotype 26 (Ad26) vector expressing the stabilized prefusion S protein of SARS-CoV-2 [1111,1296]. Unlike the other two viral vector vaccines available to date, JNJ-78436735 requires only a single dose, a characteristic that is expected to aid in global deployment [1297]. JNJ-78436735 was selected from among a number of initial candidate designs [1111] and tested *in vivo* in Syrian golden hamsters and Rhesus macaques to assess safety and immunogenicity [1111,1297,1298,1299]. The JNJ-78436735 candidate was selected for its favorable immunogenicity profile and ease of manufacturability [1111,1297,1298,1299] and was found to confer protection against SARS-CoV-2 in macaques even after six months [1300]. The one- versus two-dose regimen was tested in volunteers through a phase I/IIa trial [1296], although these results are not yet available; however, the study did report that the vaccine was well-tolerated and that most participants demonstrated seroconversion in a neutralization assay 29 days after immunization [1296].

The three viral-vector vaccines described above have demonstrated the potential for this technology to facilitate a quick response to an emerging pathogen. Additionally, though the vaccines are developed using similar principles, there are some differences that might influence their efficacy as SARS-CoV-2 evolves. In the Janssen vaccine, the S protein immunogen is stabilized in its prefusion conformation, while in the Sputnik V and AstraZeneca vaccines, it is not. How these differences in design influence the efficacy of these three viral-vector vaccines over time remains to be seen.

Efficacy Estimates: The first DNA viral-vectored vaccine for which efficacy estimates became available was AstraZeneca's ChAdOx1 nCoV-19. In December 2020, preliminary results of the phase III trial were released detailing randomized control trials conducted in the U.K., Brazil, and South Africa between April and November 2020 [1099]. These trials again compared ChAdOx1 nCoV-19 to a control, but the design of each study varied; pooling data across studies indicated an overall efficacy of 70.4%.

For Sputnik V, the phase III trial indicated an overall vaccine efficacy of 91.6% for symptomatic COVID-19 [1301].

As for Janssen, the phase III trial is ongoing across several countries (Argentina, Brazil, Chile, Colombia, Mexico, Peru, South Africa, and the U.S.), but interim results were reported in a press release on January 29th, 2021 [1302,1303]. The vaccine was well-tolerated, and across all regions studied, it was found to be 66% effective after 28 days for the prevention of moderate to severe COVID-19 and to be 85% effective for the prevention of laboratory-confirmed severe COVID-19 as well as 100% protection against COVID-19-related hospitalization and death.

Distribution Status: As of May 6, 2022, 7 viral-vectored vaccines are being distributed in 201 countries (Figure 11). ChAdOx1 nCoV-19 was first approved for emergency use on December 30, 2020 in the United Kingdom [1304] and has since then been approved for emergency use in several dozen countries, in addition to receiving full approval in Brazil.

Number of non replicating viral vector vaccines available worldwide



Figure 11: Worldwide availability of vaccines developed using non-replicating viral vectors. This figure reflects the number of vaccines using non-replicating viral vectors that were available in each country as of May 6, 2022. These data are retrieved from Our World in Data <!-To Do: Cite-> and plotted using geopandas. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

As of early January, Sputnik V had been administered to as many as 1.5 million Russians [1305], and doses of Sputnik V have also been distributed to other parts of Europe, such as Belarus, Bosnia-Herzegovina, Hungary, San

Marino, Serbia, and Slovakia [1306,1307,1308], with the Czech Republic and Austria also having expressed interest in its procurement [1309]. It wasn't until February 2021, six months after its approval in Russia, that interim results of the phase III trial were released [1301].

However, two of the three vaccines have faced a number of criticisms surrounding the implementation of their clinical trials.

Variants: A range of efficacy estimates were reported for Janssen's vaccine candidate, with estimates varying from 57% in South Africa to 72% in the United States. These differences suggested that efficacy might be influenced by the prominent viral strains circulating in each country at the time of the trial, since at the time, several variants of concern were being monitored, including B.1.351, which was first identified in South Africa [229].

6.5.3.4 RNA Vaccines

Table 5: Approved RNA vaccines [1124]

Vaccine	Company
Spikevax	Moderna
Comirnaty	Pfizer/BioNTech
TAK-919 (Moderna formulation)	Takeda

Mechanism: Building on DNA vaccine technology, RNA vaccines are an even more recent advancement for vaccine development. Interest in messenger RNA (mRNA) vaccines emerged around 1990 following *in vitro* and animal model studies that demonstrated that mRNA could be transferred into cells [1310,1311]. mRNA contains the minimum information needed to create a protein [1311]. RNA vaccines are therefore nucleic-acid based modalities that code for viral antigens against which the human body elicits a humoral and cellular immune response. This approach operates one level above the DNA: instead of directly furnishing the gene sequence associated with an antigen to the host, it provides the mRNA transcribed from the DNA sequence. Some of the potential advantages of mRNA compared to DNA include safety, as it cannot be integrated by the host and the half life can be regulated, it avoids any issues of a host immune response against the vector, and it holds the potential to dramatically accelerate vaccine manufacturing and development [1311,1312].

The mRNA technology is transcribed *in vitro* and delivered to cells via lipid nanoparticles (LNP) [1313]. They are recognized by ribosomes *in vivo* and then translated and modified into functional proteins [1122]. The resulting intracellular viral proteins are displayed on surface MHC proteins, provoking a strong CD8+ T cell response as well as a CD4⁺ T cell and B cell-associated antibody responses [1122]. Naturally, mRNA is not very stable and can degrade quickly in the extracellular environment or the cytoplasm. The LNP covering protects the mRNA from enzymatic degradation outside of the cell [1314]. Codon optimization to prevent secondary structure formation and modifications of the poly-A tail as well as the 5' untranslated region to promote ribosomal complex binding can increase mRNA expression in cells.

Furthermore, purifying out double-stranded RNA and immature RNA with FPLC (fast performance liquid chromatography) and HPLC (high performance liquid chromatography) technology will improve translation of the mRNA in the cell [1122,1315].

There are three types of RNA vaccines: non-replicating, *in vivo* self-replicating, and *in vitro* dendritic cell non-replicating [1316]. Non-replicating mRNA vaccines consist of a simple open reading frame (ORF) for the viral antigen flanked by the 5' UTR and 3' poly-A tail. *In vivo* self-replicating vaccines encode a modified viral genome derived from single-stranded, positive sense RNA alphaviruses [1122,1315]. The RNA genome encodes the viral antigen along with proteins of the genome replication machinery, including an RNA polymerase. Structural proteins required for viral assembly are not included in the engineered genome [1122]. Self-replicating vaccines produce more viral antigens over a longer period of time, thereby evoking a more robust immune response [1316]. Finally, *in vitro* dendritic cell non-replicating RNA vaccines limit transfection to dendritic cells. Dendritic cells are potent antigen-presenting immune cells that easily take up mRNA and present fragments of the translated peptide on their MHC proteins, which can then interact with T cell receptors. Ultimately, primed T follicular helper cells can stimulate germinal center B cells that also present the viral antigen to produce antibodies against the virus [1317]. These cells are isolated from the patient, grown and transfected *ex vivo*, and reintroduced to the patient [1312].

Vaccines based on mRNA delivery confer many advantages over traditional viral vectored vaccines and DNA vaccines. In comparison to live attenuated viruses, mRNA vaccines are non-infectious and can be synthetically produced in an egg-free, cell-free environment, thereby reducing the risk of a detrimental immune response in the host [1318]. Unlike DNA vaccines, mRNA technologies are naturally degradable and non-integrating, and they do not need to cross the nuclear membrane in addition to the plasma membrane for their effects to be seen [1122]. Furthermore, mRNA vaccines are easily, affordably, and rapidly scalable. Although mRNA vaccines have been developed for therapeutic and prophylactic purposes, none have previously been licensed or commercialized. Nevertheless, they have shown promise in animal models and preliminary clinical trials for several indications, including rabies, coronavirus, influenza, and cytomegalovirus [1319]. Preclinical data previously identified effective antibody generation against full-length FPLC-purified influenza hemagglutinin stalk-encoding mRNA in mice, rabbits, and ferrets [1320]. Similar immunological responses for mRNA vaccines were observed in humans in Phase I and II clinical trials operated by the pharmaceutical-development companies Curevac and Moderna for rabies, flu, and zika [1315]. Positively charged bilayer LNPs carrying the mRNA attract negatively charged cell membranes, endocytose into the cytoplasm [1314], and facilitate endosomal escape. LNPs can be coated with modalities recognized and engulfed by specific cell types, and LNPs that are 150 nm or less effectively enter into lymphatic vessels [1314,1321]. Therefore, this technology holds great potential for targeted delivery of modified mRNA.

Prior Applications: mRNA vaccine technology was even slower to develop due to challenges related to the instability of mRNA molecules, the design requirements of an efficient delivery system, and the potential for mRNA to elicit either a very strong immune response or to stimulate the immune system in secondary ways [1253,1322]. As of the 2010s, mRNA was still considered a promising technology for future advances in vaccine development [1311], but prior to 2020, no mRNA vaccines had been approved for use in humans, despite significant advances in the development of this technology [1312]. Therefore, while these technologies elegantly capitalize on decades of research in vaccine development as well as the tools of the genomic revolution, it was largely unknown prior to the SARS-CoV-2 pandemic whether this potential could be realized in a real-world vaccination effort.

Application to COVID-19: Given the potential for this technology to be quickly adapted for a new pathogen, it was favored as a potential vaccine against COVID-19. In the vaccines developed under this approach, the mRNA coding for a stabilized prefusion spike protein, which is immunogenic [1323], can be furnished to the immune system in order to train its response. Two vaccine candidates in this category emerged with promising phase III results at the end of 2020. Both require two doses approximately one month apart. The first was Pfizer/BioNTech's BNT162b2, which contains the full prefusion stabilized, membrane-anchored SARS-CoV-2 spike protein in a vaccine formulation based on modified mRNA (modRNA) technology [1324,1325]. The second mRNA vaccine, mRNA-1273 developed by ModernaTX, is comprised by a conventional lipid nanoparticle encapsulated RNA encoding a full-length prefusion stabilized S protein for SARS-CoV-2 [1326]. As of May 6, 2022, 3 mRNA vaccines are available in 167 countries (Figure 12).

Number of RNA vaccines available worldwide



Figure 12: Worldwide availability of vaccines developed using mRNA. This figure reflects the number of vaccines based on mRNA technology that were available in each country as of May 6, 2022. These data are retrieved from Our World in Data <!--To Do: Cite--> and plotted using geopandas. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

Efficacy Estimates: Pfizer/BioNTech's BNT162b2 vaccine and ModernaTX's mRNA-1273 vaccine, commercially known as Comirnaty and Spikevax, are available in most countries thanks to their rapid development in 2020. In a phase II/III multinational trial, the Pfizer/BioNTech's BNT162b2 vaccine was associated with a 95% efficacy against laboratory-confirmed COVID-19 and with mild-to-moderate local and systemic effects but a low risk of serious adverse effects when the prime-boost doses were administered 21 days

apart [503]. The ModernaTX mRNA-1273 vaccine was the second mRNA vaccine to release phase III results, despite being the first mRNA vaccine to enter phase I clinical trials and publish interim results of their phase III trial a few months later. Their study reported a 94.5% vaccine efficacy in preventing symptomatic COVID-19 in adults who received the vaccine at 99 sites around the United States [1327]. Similar to BNT162b2, the mRNA-1273 vaccine was associated with mild-to-moderate adverse effects but with a low risk of serious adverse events [1327]. Extended details of the initial phase I, II, and III trials for both vaccines are documented in the appendix.

In late 2020, both vaccines received approval from the United States's Food and Drug Administration (FDA) under an emergency use authorization [1328,1329], and these vaccines have been widely distributed, primarily in North America and the European Union [1155]. As the first mRNA vaccines to make it to market, these two highly efficacious vaccines demonstrate the power of this emerging technology, which has previously attracted scientific interest because of its potential to be used to treat non-infectious as well as infectious diseases.

Between December 2020 and April 2021, one prospective cohort study obtained weekly nasal swabs from 3,975 individuals at high risk of SARS-CoV-2 exposure (health care workers, frontline workers, etc.) within the United States [1330]. Among these participants, 3,179 (80%) had received at least one dose of an mRNA vaccine and 2,686 (84%) were fully vaccinated. For each vaccinated participant (defined here as having received at least dose 1 more than 7 days ago) whose sample tested positive for SARS-CoV-2, they categorized the viral lineage(s) present in the sample as well as in samples from 3-4 unvaccinated individuals matched by site and testing date. Overall efficacy of mRNA vaccines was estimated at 91% with full vaccination, similar to the reports from the clinical trials. The occurrence of fevers was also lower in individuals who were partially or fully vaccinated, and the duration of symptoms was approximately 6 days shorter.

Just like the other available COVID-19 vaccines, the efficacy of mRNA vaccines has been challenged by the emergence of variants of concern (VOC). These VOC have gene mutations that code for an altered spike protein, so the antibodies developed resulting from the immunization with the existing vaccines may not be as efficacious, which has caused major concern [1331,1332]. Despite some reports of varying and reduced efficacy of the mRNA vaccines against the Alpha (B.1.1.7), Beta (B.1.351), and Delta (B.1.617.2) variants versus the original SARS-CoV-2 strain or the D614G variant [[1333]; [1334]; [1335]];], the greatest concern to date has been the Omicron variant (B.1.1.529), which was first identified in November 2021 [1332,1336]. As of March 2022, the Omicron variant accounts for 95% of all infections sequenced in the United States [120] and has been linked to an increased risk of SARS-CoV-2 reinfection [1331] and further infection of those who have been vaccinated with the mRNA vaccines [1337].

Some of the gene mutations carried by Omicron, of which there are between 30-37 in the spike gene (15 in the RBD), have previously been associated with increased transmission, greater affinity for ACE2, and escape from neutralizing antibodies when they have been detected in other VOC [1331,1332,1338,1339,1339,1340]. However, multiple animal study preprints

suggested that the Omicron variant may not be as severe on the respiratory system as previous SARS-CoV-2 variants as evidenced by reduced lung infectivity, reduced SARS-CoV-2 RNA detection in the lung, and reduced inflammation and pathogenicity in animals [1341,1342,1343,1344,1345].

In spite of these findings, infection rates and hospitalization rates climbed in early 2022 in many Western countries including the United States [1346,1347]. Studies have reported reduced efficacy of the mRNA vaccines based on the measurement of antibody titers. Plasma from individuals double-dosed with Pfizer/BioNTech's BNT162b2 vaccine had up to a 16-fold reduction in neutralizing capacity against the Omicron variant [1340] and a reduced efficacy (70%) [1348]. Estimates for the mRNA vaccines range from a 2-fold to over a 20-fold drop in neutralisation titers [1349], hence the push for third doses of mRNA vaccines in many Western countries. A third mRNA vaccine dose does increase antibody titers, but these levels also wane with time [1350]. Notably, immunocompromised individuals such as cancer patients seem to elicit a sufficient protective immune response against the Omicron variant when they have been boosted with a third dose of either mRNA vaccine, albeit a blunted response [1351]. While antibody titers do correlate with protection [1352,1353,1354,1355,1356], they are not the only mechanisms of immune protection; for example, T cell and non-neutralizing antibody responses may be unaffected or less affected by the new VOC and they warrant further investigation.

In countries such as Israel, a fourth dose of mRNA vaccines have been introduced in response to the Omicron variant and an initial study in healthcare workers show that the additional immunization is safe and immunogenic with antibody titers restored to peak-third dose titers. No severe illness was reported in the cohort studied (274 versus 426 age-matched controls), and vaccine efficacy against infection was reported at 30% for BNT162b2 and 11% for mRNA-1273 [1357]. Low efficacy against infection is not surprising considering the vaccines are intended to prevent against severe disease, hospitalization and death rather than infection.

Vaccine efficacy is not the only pharmacological intervention affected by VOC. Some existing therapeutics, including monoclonal antibody treatments like Bamlanivimab (AbCellera Biologics/ Eli Lilly), were ineffective against the Omicron variant. Indeed, only Sotrovimab (Vir Biotechnology/GSK) and Tixagevimab (AstraZeneca) to a much lesser extent could effectively neutralize the omicron variant out of 7 tested monoclonal antibodies [1340], which has been verified by others [1339]. The antigenic shift of the Omicron variant does raise concerns for future VOC and what effects they may have on future vaccines and therapeutics.

Serological Response/Boosters:

Variants:

6.6 Vaccine Safety Profiles

The most common adverse reactions reported across all platforms are local site injection pain, redness and swelling and systemic reactions of fever.

Tec hn olo gy	Platform	Size of Safety Population (N)	Local Adverse Reactions	Systemic Adverse Reactions	Severe Adverse Events	Notes
IW V	CoronaVac [1149]	? Older Adults	pain (9%)	fever (3%)	2%	
IW V	CoronaVac [1151]	372 adults ages 18-59	Pain (13-21%)		Up to 1	
IW V	COVAXIN					
IW V	SinoPharm [1164]	482	N/A	N/A		23% reported any AE
IW V	WIV04 [1158 , 1168]]				0.5%	40-45% of individuals reported mild to moderate side effects 7 days post immunization. Briefly halted due to a death determined to be unrelated [1167]
Pro tein Su bu nit	NVX-CoV2373 [1233]	2,310		headache, muscle pain, fatigue		
DN A	ChAdOx1 nCoV-19					
DN A	Sputnik V [1358]		pain (58%)		hypothermia (50%), headaches (42%), fatigue (28%), joint & muscle pain (24%)	
DN A	JNJ-78436735					
mR NA	BNT162b2					
mR NA	mRNA-1273 [1327]					

Several trials have faced pauses while adverse events were investigated. In most cases, these events were determined not to be related to the vaccine.

6.7 Efficacy Estimates

Efficacy estimates have been released for many vaccine candidates across a number of technology types. However, efficacy is not a static value, and real-world efficacy can vary with location and over time. Temporal shifts in efficacy have been a especially heightened topic of concern in late 2021 given the potential for the evolution of SARS-CoV-2 to influence vaccine efficacy. The original efficacy estimates are outlined in Table XX and additional considerations for each vaccine type are described in more detail below.

Technology Platform Size of Efficacy Population (N) Estimated Efficacy Source
Whole-virus CoronaVac 10,214 (~2:1 vaccine:placebo) 83.5% [1153]
Whole-virus COVAXIN 25,798 (~1:1 vaccine:placebo) 77.8% [1169]
Whole-virus BBIBP-CorV
Whole-virus Sinopharm 79% [1359]

6.8 Special Populations

CoronaVac appears to be suitable for use in immunocompromised patients such as those with autoimmune rheumatic diseases according to phase IV trials [[1360](#)]. CoronaVac was also well tolerated and induced humoral responses in phase I trials in children aged 3 to 17 years, which will now be examined in phase II and III clinical trials [[1361](#)].

6.9 Effect of Vaccines on Community Spread

The vaccine clinical trial data demonstrate a significant reduction in the likelihood of contracting symptomatic COVID-19, thereby succeeding in the primary goal of vaccination. The mRNA vaccines in particular are so effective in preventing severe disease and death that it is also worth considering whether they might reduce disease transmission, given that vaccination rates are unlikely to reach 100%. This question hinges on whether vaccinated individuals with or without symptoms of COVID-19 can still spread SARS-CoV-2. This question is made up of several components. The crux is whether vaccinated individuals with a SARS-CoV-2 infection, regardless of symptom status, are as contagious as unvaccinated, infected individuals. Additionally, as outlined above, an important qualification is that the variants of SARS-CoV-2 circulating at the time of each study must be considered in light of the effect of evolution on vaccine efficacy.

The phase 2/3 clinical trials evaluating the mRNA vaccines assessed vaccine efficacy based on COVID-19 diagnosis, thereby detecting only patients who received a diagnosis. In order to identify patients infected with SARS-CoV-2 who did not receive a diagnosis, for example, potentially those who did not develop symptoms, it would be necessary to conduct routine PCR testing even in the absence of symptoms. Prior to the development of vaccines, the evidence suggested that asymptomatic individuals could still spread SARS-

CoV-2. Investigation of viral dynamics of asymptomatic infection in early 2020 indicated that asymptomatic patients continued to shed the virus for a duration similar to that of symptomatic patients [1362] (although viral shedding should not be conflated with contagiousness without further investigation [1]). Another study found viral load to be higher in the nasopharyngeal/oropharyngeal samples of asymptomatic patients compared to symptomatic patients hospitalized due to symptoms and/or known exposure [1363]. However, the sample size in both of these studies was small, and a larger study found higher viral load in symptomatic than asymptomatic cases [1364] along with a systematic review finding a reduced probability of asymptomatic transmission [1365]. While far from conclusive, these studies suggest that asymptomatic cases still carry a risk of transmitting SARS-CoV-2.

One important consideration is therefore how likely vaccinated individuals are to develop asymptomatic SARS-CoV-2. Considering asymptomatic cases is necessary to establish a more complete picture of efficacy with respect to spread. Routine testing of healthcare workers in California who had received an mRNA vaccine revealed slightly higher rates of absolute risk for testing positive than those identified in the phase 2/3 trials, although the extent to which asymptomatic infection influenced these numbers was not investigated [1366]. Another study analyzed the results of COVID-19 screening tests administered to asymptomatic individuals prior to receiving certain medical services at the Mayo Clinic in several locations across the United States. This study found patients who had received two doses of an mRNA vaccine to be 73% less likely to have asymptomatic COVID-19 than patients who had received zero doses [1367]. Because this study began on December 17, 2020, a date selected to coincide with the first day vaccines were available at the Mayo Clinic, this number may underestimate the efficacy of the vaccines given that many people eligible for early vaccination were at increased risk for exposure (e.g., healthcare workers and residents of long-term care facilities) [1367]. In Israel, a longitudinal study of nearly 12,000 healthcare workers found that of the 5,372 fully vaccinated people with Pfizer/BioNTech BNT162b2, 8 developed symptomatic COVID-19 (BNT162b2 (.15%) and 19 developed asymptomatic COVID-19 (.35%) [1368]. While the study itself analyzed the efficacy of the vaccine based on person-days, these findings also suggest that many or even the majority of SARS-CoV-2 infections in vaccinated individuals are likely to be asymptomatic. Therefore, in addition to the symptomatic cases reported by the vaccine clinical trials, these findings suggest that asymptomatic cases can also occur in vaccinated people. In the absence of symptoms, individuals are less likely to know to self-isolate, and therefore evaluating the effect of the vaccine on viral load is critical to understanding the role vaccinated individuals can play in spreading SARS-CoV-2.

Another question of interest is therefore whether vaccinated individuals positive for SARS-CoV-2 carry a similar viral load to unvaccinated individuals. Viral load is often approximated by cycle threshold (C_t), or the cycle at which viral presence is detected during RT-qPCR, with a lower C_t corresponding to a greater viral load. A prospective cohort study that evaluated front-line workers in six U.S. states from December 2020 to April 2021 reported a 40% reduction in viral load even with just a single dose of an mRNA vaccine [1330]. The vaccine also appeared to influence the time to viral clearance: the

risk of having detectable levels of SARS-CoV-2 for more than one week was reduced by 66% in participants who had received at least one dose [1330]. However, this study compared the mean viral load across the two groups, meaning that these findings cannot be extrapolated across all points in the disease course. Similarly, between December 2020 and February 2021, positive RT-qPCR tests were analyzed for almost 5,000 Israeli patients [1369]. C_t was analyzed relative to when each patient received the first dose of the Pfizer mRNA vaccine. A sharp increase in C_t (corresponding to reduced viral load) was observed between days 11 and 12, consistent with what is known about the onset of immunity following vaccination. This pattern therefore suggested a direct effect of vaccination on viral load.

Other studies, however, have not offered support for a reduced viral load in breakthrough cases. In Singapore, which has strict protocols for screening individuals with potential COVID-19 exposure, a retrospective cohort of patients who tested positive for SARS-CoV-2 between April and June 2021 was analyzed to compare viral kinetics and symptom course between vaccinated and unvaccinated cases. Vaccinated individuals who tested positive experienced fewer symptoms than unvaccinated, SARS-CoV-2-positive individuals and were more likely to be asymptomatic [1370] (Appendix). Additionally, this study analyzed C_t over time and found that, though the median values were similar between the two groups at disease onset, viral load appeared to decrease more rapidly in vaccinated cases [1370] (Appendix). This study is likely to have evaluated a more accurate representation of all COVID-19 outcomes than has been feasible in most studies, but one limitation was that the RT-PCR reactions were conducted in many different facilities. A third study investigated viral load (as approximated by C_t) using samples processed in a single laboratory during the summer of 2021 [1371]. This study identified no significant differences in C_t between fully vaccinated and unvaccinated cases, but this study used samples sent for diagnosis and was not longitudinal. It offered the additional benefit of culturing samples to assess whether their C_t threshold was likely to represent contagiousness and found that SARS-CoV-2 could be cultured from 51 of 55 samples with C_t less than 25 (the cut-off used in many studies). Another study of samples collected at two sites in San Francisco, one of which tested only asymptomatic individuals, reported no difference in C_t between asymptomatic and symptomatic cases regardless of whether vaccination status was included in the model [1372]. Though each of these three studies offers distinct strengths and weaknesses, taken together, they suggest that viral load is likely to be similar in vaccinated and unvaccinated individuals, but that vaccinated individuals clear the virus more rapidly, meaning that the average viral load is lower over time.

Given the emergence of variants of concern, especially the Delta variant, for which breakthrough infections are more common, the potential for vaccinated individuals to spread SARS-CoV-2 is not necessarily static over time. In fact, studies reporting reduced viral load in vaccinated individuals collected samples, for the most part, prior to the emergence of the Delta variant's dominance. The emergence of this variant may partially account for why more recent studies tend to find no difference between viral load in vaccinated and unvaccinated cases.

Taken together, these findings can provide some insight into how vaccines influence community spread. While vaccinated individuals may be more likely to experience asymptomatic infection, current evidence about viral load in asymptomatic versus symptomatic cases is ambiguous. Similarly, no conclusions can be drawn about whether viral load is different in vaccinated versus unvaccinated cases. Therefore, at present, the evidence suggests that vaccinated individuals who are infected can still contribute to community spread. The one potential mitigating factor supported at present is that differences in the viral kinetics may result in vaccinated cases infecting fewer individuals over time due to a more rapid decrease in viral load [1370], although this study did not examine patterns in secondary transmission. Thus, the virological evidence suggests that public health measures such as masking and distancing remain important even in areas with high vaccination rates.

6.9.1 Other Concerns in Efficacy

Given the wide range of vaccines under development, it is possible that some vaccine products may eventually be shown to be more effective in certain subpopulations, such as children, pregnant women, immunocompromised patients, the elderly, etc.

Age distribution in clinical trials? <https://doi.org/10.1016/j.arr.2021.101455>

Concerns: diversity of volunteer pools, variants, and distribution Another benefit of vaccines is lower population size in SARS-CoV-2 = less risk of VOC emerging that are less susceptible to the vaccine

Given the apparent need for boosters, interest has also emerged in whether vaccines against SARS-CoV-2 can be administered along with annual flu vaccines. Early data came from the Novavax NVX-CoV2373 protein subunit vaccine. In a subgroup of approximately 400 patients enrolled from the U.K. phase III trial who received either NVX-CoV2373 or placebo 1:1, a concomitant dose of adjuvanted seasonal influenza vaccines (either a trivalent vaccine or a quadrivalent vaccine) was administered [1373]. This study demonstrated that both types of vaccines could be safely administered together. While no change to the immune response was noted for the influenza vaccine, a notable reduction of the antibody response for the NVX-CoV2373 was reported, but efficacy was still high at 87.5% [1373]. Novavax has since started phase I/II trials to investigate the administration of its own influenza vaccine, NanoFlu™, concomitantly with NVX-CoV2373 [1374], which appeared to be safe and effective in preclinical studies [1375].

Indeed, Chinese [1186] and Chilean [1376] researchers have opted to investigate options to administer different vaccines (e.g., an mRNA vaccine dose) as a booster dose to individuals who have already received two doses of the IIV vaccine CoronaVac. Another study has already determined that using the CanSino vaccine (Convidecia) instead of CoronaVac in a prime-boost vaccination regimen can induce a more robust immune response [1188].

6.10 Conclusions

In the early 2000s, technologies such as inactivated viral vaccines, live attenuated viral vaccines, protein subunit vaccines, and recombinant vector-based vaccines were explored for SARS [1377,1378], but the epidemic was controlled before these efforts came to fruition [1094]. DNA vaccine development efforts also began but did not proceed past animal testing [1378]. Similarly, viral vector, protein subunit, and DNA vaccines were explored for MERS-CoV, but outbreaks are sporadic and difficult to predict, making vaccine testing and the development of a vaccination strategy difficult [1379]. Likewise, the development of viral-vectorized Ebola virus vaccines was undertaken, but the pace of vaccine development was behind the spread of the virus from early on [1380]. Although a candidate Ebola vaccine V920 showed promise in preclinical and clinical testing, it did not receive breakthrough therapy designation until the summer of 2016, by which time the Ebola outbreak was winding down [1381].

7 Vaccine Development Strategies for SARS-CoV-2 Appendix

7.1 Live Attenuated Viruses

One candidate in the preclinical stage is YF-S0, a single-dose LAV developed at Katholieke Universiteit Leuven that uses live-attenuated yellow fever 17D (YF17D) as a vector for a noncleavable prefusion conformation of the SARS-CoV-2 antigen. YF-S0 induced a robust immune response in three animal models and prevented SARS-CoV-2 infection in macaques and hamsters [1125].

Other programs are exploring the development of codon deoptimized LAV [1129,1130,1131]. New York-based Codagenix and the Serum Institute of India reported a successful preclinical investigation [1131] of an intranasally administered deoptimized SARS-CoV-2 LAV known as COVI-VAC, and COVI-VAC entered phase I human trials and dosed its first participants in January 2021 [1130,1382]. It is anticipated that the COVI-VAC phase I human trials will be completed by May 2022.

Another company, Meissa Vaccines in Kansas, U.S.A., which also develops vaccines for Respiratory syncytial virus (RSV), has developed an intranasal live-attenuated chimeric vaccine. Chimeric vaccines integrate genomic content from multiple viruses to create a more stable LAV [1133]. Enrollment for phase I human trials began in March 2021 and recruitment is ongoing [1130,1383].

Finally, Bacillus Calmette-Guerin (BCG) vaccines that use LAVs are being investigated for the prophylaxis of COVID-19. The purpose of the BCG vaccine is to prevent tuberculosis, but non-specific effects against other respiratory illnesses have suggested a possible benefit against COVID-19 [1134]. Currently, investigations of BCG vaccines against COVID-19 are being sponsored by institutes in Australia in collaboration with the Bill and Melinda Gates Foundation [1384] and by Texas A&M University in collaboration with numerous other U.S. institutions [1385].

All the same, results from LAV trials in humans are largely unavailable, and no LAV vaccines are being administered. Despite the long and trusted history of LAV development, this vaccine strategy has not been favored against COVID-19. Modern, modular technologies have shown greater expediency and safety compared to the time-consuming nature of developing LAVs for a novel virus.

7.2 Sinovac's CoronaVac

Pre-clinical trials were performed using BALB/c mice and rhesus macaques [1150]. The SARS-CoV-2 strains used in this trial isolated from 11 hospitalized patients (5 from China, 3 from Italy, 1 from the UK, 1 from Spain, 1 from Switzerland). A phylogenetic analysis demonstrated that the strains were representative of the variants circulating at the time. One of the strains from China, CN2, was used as the inactivated and purified virus while the other 10 strains were used to challenge. The CN2 was grown in Vero cells. An ELISA assay was used to assess the immunogenicity of the vaccine. 10 mice were injected with the vaccine on day 0 and day 7 with varying doses (0, 1.5, 3 or 6 µg), and 10 mice were treated with physiological saline as the control. IgG developed in the serum of all vaccinated mice. Using the same setup, immunogenicity was also assessed in macaques. Four macaques were assigned to each of four groups: treatment with 3 µg at day 0, 7 and 14, treatment with a high dose of 6 µg at day 0, 7 and 14, administration of a placebo vaccine, and administration of only the adjuvant. All vaccinated macaques induced IgGs and neutralizing antibodies. After challenge with SARS-CoV-2 strain CN1, vaccinated macaques were protected compared to control macaques (placebo or adjuvant only) based on histology and viral loads collected from different regions of the lung.

Phase I/II clinical trials were conducted in adults 18-59 years old [1151] and adults over 60 years old [1149] in China. In the case of adults 18-59 years old, a single center, randomized, double-blind, placebo-controlled phase I/II trial was conducted in April 2020. Patients in this study were recruited from the community in Suining County of Jiangsu province, China. For the phase I trial, 144 (of 185 screened) participants were enrolled, with 72 enrolled in the 14-day interval cohort (i.e., treated on day 0 and day 14) and 72 in the 28-day interval cohort. This group of 72 participants was split into 2 blocks for a low-dose (3 µg) and high-dose (6 µg) vaccine. Within each block, participants were randomly assigned vaccination with CoronaVac or placebo (aluminum diluent without the virus) at a 2:1 ratio. Both the vaccine and placebo were prepared in a Good Manufacturing Practice-accredited facility of Sinovac Life Sciences (Beijing, China).

The phase II trial followed the same organization of participants, this time using 300 enrolled participants in the 14-day and another 300 enrolled in the 28-day groups. One change of note was that the vaccine was produced using a highly automated bioreactor (ReadyToProcess WAVE 25, GE, Umeå, Sweden) to increase vaccine production capacity. This change resulted in a higher intact spike protein content. The authors of this study were not aware of this antigen-level difference between the vaccine batches for the phase I/II when the ethical approval for the trials occurred.

To assess adverse responses, participants were asked to record any events up to 7 days post-treatment. The reported adverse events were graded according to the China National Medical Products Administration guidelines. In the phase I trial, the overall incidence of adverse reactions was 29-38% of patients in the 0 to 14 day group and 13-17% in the 0 to 28 day vaccination group. The most common symptom was pain at the injection site, which was reported by 17-21% of patients in the 0 to 14 day cohort and 13% in the 0 to 28 day cohort. Most adverse reactions were mild (grade 1) where patients recovered within 48 hours. A single case of acute hypersensitivity with manifestation of urticaria 48 hours following the first dose of study drug was reported in the 6 µg group. Most of the adverse reactions reported were characterized as mild and resolved within 48 hours, with a single participant experiencing a serious adverse event that could have been related to vaccination (acute hypersensitivity with manifestation of urticaria 48 hours after their first dose in the 6-µg condition). Both the 14-day and 28-day cohorts had a strong neutralizing Ab response. The neutralizing Ab response was measured using a micro cytopathogenic effect assay, which assesses the minimum dilution of neutralizing Ab to be 50% protective against structural changes in host cells in response to viral infection [1386]. Additionally IgG antibody titers against the receptor binding domain were also measured using ELISA.

Another phase I/II study was performed with older patients (older than 60 years) [1149]. The study conducted a single-center, randomized, double-blind, placebo-controlled trial. The phase I trial looked at dose escalation using 3 dosages: 1.5, 3 and 6 µg. The mean age of participants was 65.8 years (std = 4.8). Of 95 screened participants, 72 were enrolled. These 72 participants were split into low (3 µg) and high (6 µg) dose groups. Within each group, 24 participants received the treatment and 12 the placebo. A neutralizing antibody response against live SARS-CoV-2 was detected compared to baseline using the same micro cytopathogenic effect assay. This response was similar across the two dose concentrations. Additionally, they did not observe a difference in response between age groups (60–64 years, 65–69 years, and ≥70 years).

In phase II the mean age was 66.6 years (standard deviation = 4.7). 499 participants were screened and 350 were enrolled. 300 were evenly split into 1.5, 3 and 6 µg dose groups, and the remaining 50 were assigned to the placebo group. Again, they found a neutralizing antibody response in phase II. There wasn't a significant different between the response to 3 µg versus 6 µg, but the response was higher than that to 1.5 µg.

Participants were required to record adverse reaction events within the first 7 days after each dose. The safety results were combined across phase I and II. All adverse reactions were either mild (grade 1) or moderate (grade 2) in severity. The most common symptom was pain at the injection site (9%) and fever (3%). 2% (7 participants) reported severe adverse events (4 from the 1.5 µg group, 1 from the 3 µg group, 2 from the 6 µg group), though these were found to be unrelated to the vaccine.

Overall, the results from the pre-clinical and phase I/II clinical trials are promising. It was also very hopeful to see that the immune response was consistent in older adults (> 60 years). Currently, phase III trials are being

conducted in Brazil [1387]. This is a randomized, multicenter, endpoint driven, double-blind, placebo-controlled clinical trial. They are expecting 13,060 participants with 11,800 ages 18 to 59 years and 1,260 age 60+. Participants will be health care professionals. As of September 2021, CoronaVac trials are now also being held in the Philippines and Hong Kong, bringing the total number of registered phase III trials investigating the safety and efficacy of CoronaVac to 9, with emergency use approval in 40 countries [1388].

7.3 COVAXIN

In India, the COVAXIN® vaccine received emergency authorization on January 3rd, 2021, despite the lack of phase III data until March 3rd, which was communicated via press release [1389]. Following a press release of the phase III data indicating 80.6% efficacy in 25,800 participants [1389,1390], Zimbabwe authorized the use of COVAXIN® [1391]. This was followed by a detailed preprint of the double-blind, randomized, controlled phase III trial that was made available in July 2021, with a final enrollment of 25,798 people (~1:1 vaccine:placebo) [1169]. The vaccine was reported as well tolerated, with an overall vaccine efficacy of 77.8% for the prevention of COVID-19. Efficacy against severe disease and asymptomatic infection was reported as 93.4% and 63.6% respectively. Indeed, another preprint determined that sera from individuals immunized with COVAXIN® had effective neutralizing antibodies against the delta variant and the so-called delta plus variant (AY.1) [1175]. It is not yet clear what level of protection COVAXIN® offers beyond 6 to 8 months post the second vaccine; consequently, the potential requirement of a booster immunization is being explored [1182]. Furthermore, Bharat Biotech is considering other vaccine regimens such as providing one initial immunization with COVAXIN® followed by two immunizations with its intranasal vaccine (BBV154) [1392]. U.S.-based Ocugen Inc., a co-development partner of Bharat Biotech, is leading the application for an Emergency Use Authorization (EUA) for COVAXIN® intended for the U.S. market. As of September 2021 COVAXIN® has been approved for emergency use in Guyana, India, Iran, Zimbabwe, and Nepal, Mauritius, Mexico, Nepal, Paraguay, and the Philippines [1163]. It has been reported that Bharat Biotech will soon release its phase II and III pediatric trial results [1393]. However, the WHO approval of the COVAXIN® has been delayed [1394].

7.4 RNA Vaccines

RNA vaccines are nucleic-acid based modalities that code for viral antigens against which the human body elicits a humoral and cellular immune response. The mRNA technology is transcribed *in vitro* and delivered to cells via lipid nanoparticles (LNP) [1313]. They are recognized by ribosomes *in vivo* and then translated and modified into functional proteins [1122]. The resulting intracellular viral proteins are displayed on surface MHC proteins, provoking a strong CD8+ T cell response as well as a CD4+ T cell and B cell-associated antibody responses [1122]. Naturally, mRNA is not very stable and can degrade quickly in the extracellular environment or the cytoplasm. The LNP covering protects the mRNA from enzymatic degradation outside of the cell [1314]. Codon optimization to prevent secondary structure formation

and modifications of the poly-A tail as well as the 5' untranslated region to promote ribosomal complex binding can increase mRNA expression in cells. Furthermore, purifying out double-stranded RNA and immature RNA with FPLC (fast performance liquid chromatography) and HPLC (high performance liquid chromatography) technology will improve translation of the mRNA in the cell [1122,1315]. Vaccines based on mRNA delivery confer many advantages over traditional viral vectored vaccines and DNA vaccines. In comparison to live attenuated viruses, mRNA vaccines are non-infectious and can be synthetically produced in an egg-free, cell-free environment, thereby reducing the risk of a detrimental immune response in the host [1318]. Unlike DNA vaccines, mRNA technologies are naturally degradable and non-integrating, and they do not need to cross the nuclear membrane in addition to the plasma membrane for their effects to be seen [1122]. Furthermore, mRNA vaccines are easily, affordably, and rapidly scalable.

Although mRNA vaccines have been developed for therapeutic and prophylactic purposes, none have previously been licensed or commercialized. Nevertheless, they have shown promise in animal models and preliminary clinical trials for several indications, including rabies, coronavirus, influenza, and cytomegalovirus [1319]. Preclinical data previously identified effective antibody generation against full-length FPLC-purified influenza hemagglutinin stalk-encoding mRNA in mice, rabbits, and ferrets [1320]. Similar immunological responses for mRNA vaccines were observed in humans in Phase I and II clinical trials operated by the pharmaceutical-development companies Curevac and Moderna for rabies, flu, and zika [1315]. Positively charged bilayer LNPs carrying the mRNA attract negatively charged cell membranes, endocytose into the cytoplasm [1314], and facilitate endosomal escape. LNPs can be coated with modalities recognized and engulfed by specific cell types, and LNPs that are 150 nm or less effectively enter into lymphatic vessels [1314,1321]. Therefore, this technology holds great potential for targeted delivery of modified mRNA.

There are three types of RNA vaccines: non-replicating, *in vivo* self-replicating, and *in vitro* dendritic cell non-replicating [1316]. Non-replicating mRNA vaccines consist of a simple open reading frame (ORF) for the viral antigen flanked by the 5' UTR and 3' poly-A tail. *In vivo* self-replicating vaccines encode a modified viral genome derived from single-stranded, positive sense RNA alphaviruses [1122,1315]. The RNA genome encodes the viral antigen along with proteins of the genome replication machinery, including an RNA polymerase. Structural proteins required for viral assembly are not included in the engineered genome [1122]. Self-replicating vaccines produce more viral antigens over a longer period of time, thereby evoking a more robust immune response [1316]. Finally, *in vitro* dendritic cell non-replicating RNA vaccines limit transfection to dendritic cells. Dendritic cells are potent antigen-presenting immune cells that easily take up mRNA and present fragments of the translated peptide on their MHC proteins, which can then interact with T cell receptors. Ultimately, primed T follicular helper cells can stimulate germinal center B cells that also present the viral antigen to produce antibodies against the virus [1317]. These cells are isolated from the patient, grown and transfected *ex vivo*, and reintroduced to the patient [1312].

Given the potential for this technology to be quickly adapted for a new pathogen, it has held significant interest for the treatment of COVID-19. In the vaccines developed under this approach, the spike protein, which is immunogenic [1323], can be furnished to the immune system in order to train its response. The vaccine candidates developed against SARS-CoV-2 using mRNA vectors utilize similar principles and technologies, although there are slight differences in implementation among candidates such as the formulation of the platform and the specific components of the spike protein encapsulated (e.g., the full Spike protein vs. the RBD alone) [1395]. The results of the interim analyses of two mRNA vaccine candidates became available at the end of 2020 and provided strong support for this emerging approach to vaccination. Below we describe the results available as of February 2021 for two such candidates, mRNA-1273 produced by ModernaTX and BNT162b2 produced by Pfizer, Inc. and BioNTech.

7.4.1 ModernaTX mRNA Vaccine

ModernaTX's mRNA-1273 vaccine was the first COVID-19 vaccine to enter a phase I clinical trial in the United States. In this trial, Moderna spearheaded an investigation on the immunogenicity and reactogenicity of mRNA-1273, a conventional lipid nanoparticle encapsulated RNA encoding a full-length prefusion stabilized S protein for SARS-CoV-2 [1326]. An initial report described the results of enrolling forty-five participants who were administered intramuscular injections of mRNA-1273 in their deltoid muscle on day 1 and day 29, with the goal of following patients for the next twelve months [1102]. Healthy males and non-pregnant females aged 18-55 years were recruited for this study and divided into three groups receiving 25, 100, or 250 micrograms (μ g) of mRNA-1273. IgG ELISA assays on patient serology samples were used to examine the immunogenicity of the vaccine [1326]. Binding antibodies were observed at two weeks after the first dose at all concentrations. At the time point one week after the second dose was administered on day 29, the pseudotyped lentivirus reporter single-round-of-infection neutralization assay (PsVNA), which was used to assess neutralizing activity, reached a median level similar to the median observed in convalescent plasma samples. Participants reported mild and moderate systemic adverse events after the day 1 injection, and one severe local event was observed in each of the two highest dose levels. The second injection led to severe systemic adverse events for three of the participants at the highest dose levels, with one participant in the group being evaluated at an urgent care center on the day after the second dose. The reported localized adverse events from the second dose were similar to those from the first.

Several months later, a press release from ModernaTX described the results of the first interim analysis of the vaccine [1396]. On November 16, 2020, a report was released describing the initial results from Phase III testing, corresponding to the first 95 cases of COVID-19 in the 30,000 enrolled participants [1396], with additional data released to the FDA on December 17, 2020 [1397]. These results were subsequently published in a peer-reviewed journal (*The New England Journal of Medicine*) on December 30, 2020 [1327]. The first group of 30,420 study participants were randomized to receive the vaccine or a placebo at a ratio of 1:1 [1327]. Administration occurred at 99 sites within the United States in two sessions, spaced 28 days apart [1327,1398]. Patients reporting COVID-19 symptoms upon follow-up

were tested for SARS-CoV-2 using a nasopharyngeal swab that was evaluated with RT-PCR [1398]. The initial preliminary analysis reported the results of the cases observed up until a cut-off date of November 11, 2020. Of these first 95 cases reported, 90 occurred in participants receiving the placebo compared to 5 cases in the group receiving the vaccine [1396]. These results suggested the vaccine is 94.5% effective in preventing COVID-19. Additionally, eleven severe cases of COVID-19 were observed, and all eleven occurred in participants receiving the placebo. The publication reported the results through an extended cut-off date of November 21, 2020, corresponding to 196 cases [1327]. Of these, 11 occurred in the vaccine group and 185 in the placebo group, corresponding to an efficacy of 94.1%. Once again, all of the severe cases of COVID-19 observed (n=30) occurred in the placebo group, including one death. Thus, as more cases are reported, the efficacy of the vaccine has remained above 90%, and no cases of severe COVID-19 have yet been reported in participants receiving the vaccine.

These findings suggest the possibility that the vaccine might bolster immune defenses even for subjects who do still develop a SARS-CoV-2 infection. The study was designed with an explicit goal of including individuals at high risk for COVID-19, including older adults, people with underlying health conditions, and people of color [1399]. The Phase III trial population was comprised by approximately 25.3% adults over age 65 in the initial report and 24.8% in the publication [1398]. Among the cases reported by both interim analyses, 16-17% occurred in older adults [1327,1396].. Additionally, approximately 10% of participants identified a Black or African-American background and 20% identified Hispanic or Latino ethnicity [1327,1398]. Among the first 95 cases, 12.6% occurred in participants identifying a Hispanic or Latino background and 4% in participants reporting a Black or African-American background [1396]; in the publication, they indicated only that 41 of the cases reported in the placebo group and 1 case in the treatment group occurred in “communities of color”, corresponding to 21.4% of all cases [1327]. While the sample size in both analyses is small relative to the study population of over 30,000, these results suggest that the vaccine is likely to be effective in people from a variety of backgrounds. By all indications, this vaccine is likely to be highly useful in mitigating the damage of SARS-CoV-2.

In-depth safety data was released by ModernaTX as part of their application for an EUA from the FDA and summarized in the associated publication [1327,1398]. Because the detail provided in the report is greater than that provided in the publication, here we emphasize the results observed at the time of the first analysis. Overall, a large percentage of participants reported adverse effects when solicited, and these reports were higher in the vaccine group than in the placebo group (94.5% versus 59.5%, respectively, at the time of the initial analysis) [1398]. Some of these events met the criteria for grade 3 (local or systemic) or grade 4 (systemic only) toxicity [1398], but most were grade 1 or grade 2 and lasted 2-3 days [1327]. The most common local adverse reaction was pain at the injection site, reported by 83.7% of participants receiving the first dose of the vaccine and 88.4% upon receiving the second dose, compared to 19.8% and 19.8% and 17.0%, respectively, of patients in the placebo condition [1398]. Fewer than 5% of vaccine recipients reported grade 3 pain at either administration. Other frequent local reactions included erythema, swelling, and lymphadenopathy [1398]. For systemic

adverse reactions, fatigue was the most common [1398]. Among participants receiving either dose of the vaccine, 68.5% reported fatigue compared to 36.1% participants receiving the placebo [1398]. The level of fatigue experienced was usually fairly mild, with only 9.6% and 1.3% of participants in the vaccine and placebo conditions, respectively, reporting grade 3 fatigue [1398], which corresponds to significant interference with daily activity [1400]. Based on the results of the report, an EUA was issued on December 18, 2020 to allow distribution of this vaccine in the United States [1329], and it was shortly followed by an Interim Order authorizing distribution of the vaccine in Canada [1401] and a conditional marketing authorization by the European Medicines Agency to facilitate distribution in the European Union [1402].

7.4.2 Pfizer/BioNTech BNT162b2

ModernaTX was, in fact, the second company to release news of a successful interim analysis of an mRNA vaccine and receive an EUA. The first report came from Pfizer and BioNTech's mRNA vaccine BNT162b2 on November 9, 2020 [1403], and a preliminary report was published in the *New England Journal of Medicine* one month later [503]. The vaccine candidate contains the full prefusion stabilized, membrane-anchored SARS-CoV-2 spike protein in a vaccine formulation based on modified mRNA (modRNA) technology [1324,1325]. This vaccine candidate should not be confused with a similar candidate from Pfizer/BioNTech, BNT162b1, that delivered only the RBD of the spike protein [1404,1405], which was not advanced to a stage III trial because of the improved reactogenicity/immunogenicity profile of BNT162b2 [504].

During the Phase III trial of BNT162b2, 43,538 participants were enrolled 1:1 in the placebo and the vaccine candidate and received two 30- μ g doses 21 days apart [503]. Of these enrolled participants, 21,720 received BNT162b2 and 21,728 received a placebo [503]. Recruitment occurred at 135 sites across six countries: Argentina, Brazil, Germany, South Africa, Turkey, and the United States. An initial press release described the first 94 cases, which were consistent with 90% efficacy of the vaccine at 7 days following the second dose [1403]. The release of the full trial information covered a longer period and analyzed the first 170 cases occurring at least 7 days after the second dose, 8 of which occurred in patients who had received BNT162b2. The press release characterized the study population as diverse, reporting that 42% of the participants worldwide came from non-white backgrounds, including 10% Black and 26% Hispanic or Latino [1406]. Within the United States, 10% and 13% of participants, respectively, identified themselves as having Black or Hispanic/Latino backgrounds [1406]. Additionally, 41% of participants worldwide were 56 years of age or older [1406], and they reported that the efficacy of the vaccine in adults over 65 was 94% [1407]. The primary efficacy analysis of the Phase III study was concluded on November 18, 2020 [1407], and the final results indicated 94.6% efficacy of the vaccine [503].

The safety profile of the vaccine was also assessed [503]. A subset of patients were followed for reactogenicity using electronic diaries, with the data collected from these 8,183 participants comprising the solicited safety events analyzed. Much like those who received the ModernaTX vaccine candidate, a

large proportion of participants reported experiencing site injection pain within 7 days of vaccination. While percentages are broken down by age group in the publication, these proportions correspond to approximately 78% and 73% of all participants after the first and second doses, respectively, overall. Only a small percentage of these events (less than 1%) were rated as serious, with the rest being mild or moderate, and none reached grade 4. Some participants also reported redness or swelling, and the publication indicates that in most cases, such events resolved within 1 to 2 days.

Participants also experienced systemic effects, including fever (in most cases lower than 38.9°C and more common after dose 2), fatigue (25-50% of participants depending on age group and dose), headache (25-50% of participants depending on age group and dose), chills, and muscle or joint pain; more rarely, patients could experience gastrointestinal effects such as vomiting or diarrhea. As with the local events, these events were almost always grade I or II. While some events were reported by the placebo groups, these events were much rarer than in the treatment group even though compliance was similar. Based on the efficacy and safety information released, the vaccine was approved in early December by the United Kingdom's Medicines and Healthcare Products Regulatory Agency with administration outside of a clinical trial beginning on December 8, 2020 [1408,1409]. As of December 11, 2020, the United States FDA approved this vaccine under an emergency use authorization [1328].

7.5 Viral Vector Vaccines

7.5.1 ChAdOx1 nCoV-19 (AstraZeneca)

As discussed above, prior analyses of viral vector vaccines against hCoV had indicated that this approach showed potential for inducing an immune response, but little information was available about the effect on real-world immunity. In the first phase of development, a candidate ChAdOx1 nCoV-19 was evaluated through the immune challenge of two animal models, mice and rhesus macaques [1291]. Animals in the treatment condition were observed to develop neutralizing antibodies specific to SARS-CoV-2 (both macaques and mice) and to show reduced clinical scores when exposed to SARS-CoV-2 (macaques) [1291]. Next, a phase I/II trial was undertaken using a single-blind, randomized controlled design [1292]. ChAdOx1 nCoV-19 and a control, the meningococcal conjugate vaccine MenACWY, were administered intramuscularly to adults ages 18 to 55 at five sites within the United Kingdom (U.K.) at a 1:1 ratio (n=543 and n=534, respectively). All but ten participants received a single dose; this small group received a booster 28 days after their first dose of ChAdOx1 nCoV-19. Commonly reported local adverse reactions included mild-to-moderate pain and tenderness at the injection site over the course of seven days, while the most common systemic adverse reactions were fatigue and headache; some patients reported severe adverse systemic effects. The study also reported that many common reactions could be reduced through the administration of paracetamol (acetaminophen), and paracetamol was not found to reduce immunogenicity. Patients receiving the ChAdOx1 nCoV-19 vaccine developed antibodies to the SARS-CoV-2 spike protein that peaked by day 28, with these levels remaining stable until a second observation at day 56 except in the ten patients who received a booster dose at day 28, in whom they increased by

day 56. Analysis of serum indicated that participants developed antibodies to both S and the RBD, and that 100% of them achieved neutralizing titers by day 28. By day 35, the neutralization titers of vaccinated patients was comparable to that observed with plasma from convalescents. This initial study therefore suggested that the vaccine was likely to confer protection against SARS-CoV-2, although analysis of its efficacy in preventing COVID-19 was not reported.

In December 2020, preliminary results of the phase III trial were released detailing randomized control trials conducted in the U.K., Brazil, and South Africa between April and November 2020 [1099]. These trials again compared ChAdOx1 nCoV-19 to a control, but the design of each study varied. For example, in South Africa, the trial was double-blinded, whereas in the U.K. and Brazil it was single-blinded, and one of the two trials carried out in the U.K. examined two dosing regimens (low dose or standard dose, both followed by standard dose). Some of the trials used MenACWY as a control, while others used saline. Data was pooled across countries for analysis. The primary outcome assessed was symptomatic, laboratory-confirmed COVID-19. There were 131 cases observed among the 11,636 participants eligible for the primary efficacy analysis, corresponding to an overall efficacy of 70.4% (30 out of 5807 in the vaccine arm and 101 out of 5829 in the control arm); the 95.8% CI was reported as 54.8 to 80.6. However, a higher efficacy was reported in the subgroup of patients who received a low-dose followed by a standard dose (90.0%, 95% CI 67.4 to 97.0). A total of ten cases of severe COVID-19 resulting in hospitalization were observed among trial participants, and all of these occurred in patients in the control arm of the study. In line with the previously reported safety profiling for this vaccine, serious adverse events were reported to be comparable across the two arms of the study, with only three events identified as potentially associated with the vaccine itself. The U.K. Medicines and Healthcare Products Regulatory Agency (MHRA) approved ChAdOx1 nCoV-19 for emergency use on December 30, 2020 [1304]. Additional data about the efficacy of this vaccine became available in a preprint released on March 2, 2021 [1410]. This report provided data describing the efficacy of ChAdOx1 nCoV-19, along with Pfizer/BioNTech's BNT162b2, in the U.K. between December 8, 2020 and February 19, 2021 and specifically sought to evaluate the efficacy of the vaccine in the presence of a potentially more contagious variant of concern, B.1.1.7. All participants in this study were age 70 or older and the efficacy was estimated to increase from 60% at 28 days after vaccination to 73% at 35 days after vaccination, although the standard error also increased over this time. Therefore, preliminary results suggest that in a number of samples, this vaccine confers a high level of protection against SARS-CoV-2.

7.5.2 Sputnik-V (Gam-COVID-Vac and Gam-COVID-Vac-Lyo)

The vaccine Gam-COVID-Vac, nicknamed Sputnik V in reference to the space race and "V for vaccine", was developed by the Gamaleya National Center of Epidemiology and Microbiology in Moscow. Gamaleya is an organization with prior experience using the adenovirus platform for the development of vaccines for *Middle East respiratory syndrome-related coronavirus* and Ebola virus, although neither of the previous vaccines were internationally licensed [1287]. The development of Sputnik V was financed by the Russian Direct

Investment Fund (RDIF) [1293,1411]. Sputnik V is a replication-deficient recombinant adenovirus (rAd) vaccine that combines two adenovirus vectors, rAd26-S and rAd5-S, that express the full-length SARS-CoV-2 spike glycoprotein. These vectors are intramuscularly administered individually using two separate vaccines in a prime-boost regimen. The rAd26-S is administered first, followed by rAd5-S 21 days later. Both vaccines deliver 10^{11} viral particles per dose. This approach is designed to overcome any potential pre-existing immunity to adenovirus in the population [1412], as some individuals may possess immunity to Ad5 [1413]. Sputnik V is the only recombinant adenovirus vaccine to utilize two vectors. Other vaccines, such as the Oxford-AstraZeneca vaccine, utilize the chimpanzee adenovirus vector (ChAdOx1 nCoV-19) for both doses [1414]. The Sputnik V vaccines are available in both a lyophilized (Gam-COVID-Vac-Lyo) and frozen form (Gam-COVID-Vac), which are stored at 2-8°C and -18°C respectively [1358]. The lyophilized vaccine is convenient for distribution and storage, particularly to remote or disadvantaged areas [1415].

In the race to develop vaccines against SARS-CoV-2, President Vladimir Putin of Russia announced the approval of the Sputnik V vaccines on August 11th, 2020 in the absence of clinical evidence [1293]. Consequently, many international scientific agencies and public health bodies expressed concern that due diligence to the clinical trial process was subverted for the sake of expediency, leading many to question the safety and efficacy of Sputnik V [1293,1416,1417]. Despite regulatory, safety, and efficacy concerns, pre-orders for 1 billion doses of the Sputnik V were reported within days of the vaccine's approval in Russia [1293]. Almost a month later, the phase I/II trial data was published [1358].

In the phase I/II trial study conducted between late June and early August 2020, 76 participants (18-60 years old) were split into two groups of 38 participants, which were non-randomized in two hospitals in Russia. In phase I, 9 patients received rAd26 and 9 patients received rAd5-S to assess safety over 28 days. In phase II, at least 5 days after the completion of phase I, 20 patients received a prime-boost vaccination of rAd26-S on day 0 and rAd5-S on day 2, which was administered intramuscularly. The phase I/II trial reported that both vaccines were deemed safe and well tolerated. The most common adverse events reported were mild, such as pain at the injection site (58%), hypothermia (50%), headaches (42%), fatigue (28%), and joint and muscle pain (24%). Seroconversion was observed in all participants three weeks post the second vaccination (day 42), and all participants produced antibodies to the SARS-CoV-2 glycoprotein. RBD-specific IgG levels were high in both the frozen and lyophilized versions of the vaccine (14,703 and 11,143 respectively), indicating a sufficient immune response to both. Three weeks post the second vaccination, the virus-neutralizing geometric mean antibody titers were 49.25 and 45.95 from the frozen and lyophilized vaccines, respectively. At 28 days, median cell proliferation of 1.3% CD4 $^{+}$ and 1.1% CD8 $^{+}$ were reported for the lyophilized vaccine and 2.5% CD4 $^{+}$ and 1.3% CD8 $^{+}$ for the vaccine stored frozen. These results indicated that both forms of Sputnik V appeared to be safe and induce a humoral and cellular response in human subjects [1358], which may be robust enough to persist and not wane rapidly [1412].

A press release on November 11th, 2020 indicated positive results from an interim analysis of the phase III Sputnik V trials, which reported 92% efficacy in 16,000 volunteers [1418]. However, this release came only two days after both Pfizer and BioNTech reported that their vaccines had an efficacy over 90%, which led to significant skepticism of the Russian findings for a myriad of reasons including the lack of a published protocol and the “reckless” approval of the vaccine in Russia months prior to the publication of the interim results of the phase III trial [1418,1419]. In February 2021, the interim results of the phase III randomized, double-blind, placebo-controlled trial were eventually published in *The Lancet* [1301]. The participants were randomly assigned to receive either a 0.5 mL/dose of vaccine or placebo, which was comprised of the vaccine buffer composition, that was delivered intramuscularly using the same prime-boost regimen as in the phase I/II trials. From September 7th to Nov 24th, 19,866 participants completed the trial. Of the 14,964 participants who received the vaccine, 16 (0.1%) were confirmed to have COVID-19, whereas 62 of the 4,902 participants (1.3%) in the placebo group were confirmed to have COVID-19. Of these participants, no moderate or severe cases of COVID-19 were reported in the vaccine group, juxtaposed with 20 in the placebo group. However, only symptomatic individuals were confirmed for SARS-CoV-2 infection in this trial. Therefore, asymptomatic infections were not detected, thus potentially inflating the efficacy estimate. Overall, a vaccine efficacy of 91.6% (95% CI 85.6-95.2) was reported, where an efficacy of 91.8% was reported for those over 60 years old and 92.7% for those who were 51-60 years old. Indeed, 14 days after the first dose, 87.6% efficacy was achieved and the immunity required to prevent disease occurred within 18 days of vaccination. Based on these results, scientists are investigating the potential for a single dose regimen of the rAd26-S sputnik V vaccine [1420]. By the end of the trial, 7,485 participants reported adverse events, of which 94% were grade I. Of the 68 participants who experienced serious adverse events during the trial, 45 from the vaccine group and 23 from the placebo groups, none were reported to be associated with the vaccination. Likewise, 4 deaths occurred during the trial period that were not related to the vaccine [1301]. The interim findings of the phase III trial indicate that the Sputnik V vaccine regimen appears to be safe with 91.6% efficacy. Gamaleya had intended to reach a total of 40,000 participants for the completion of their phase III trial. However, the trial has stopped enrolling participants and the numbers have been cut to 31,000 as many individuals in the placebo group dropped out of the study to obtain the vaccine [1421]. Indeed, other trials involving Sputnik V are currently underway in Belarus, India, the United Arab Emirates, and Venezuela [1422].

Preliminary results of a trial of Argentinian healthcare workers in Buenos Aires who were vaccinated with the Sputnik V rAd26-R vector-based vaccine seems to support the short term safety of the first vaccination [1423]. Of the 707 vaccinated healthcare workers, 71.3% of the 96.6% of respondents reported at least one adverse event attributed to the vaccine. Of these individuals, 68% experienced joint and muscle pain, 54% had injection site pain, 11% reported redness and swelling, 40% had a fever, and 5% reported diarrhea. Only 5% of the vaccinated participants experienced serious adverse events that required medical attention, of which one was monitored as an inpatient.

Additionally, an Independent assessment of Sputnik V in a phase II clinical trial in India found the vaccine to be effective, but the data is not yet publicly available [1424]. On December 21st, 2020, Gamaleya, AstraZeneca, R-Pharm, and the Russian Direct Investment Fund agreed to assess the safety and immunogenicity of the combined use of components of the AstraZeneca and University of Oxford AZD1222 (ChAdOx1) vaccine and the rAd26-S component of the Sputnik V vaccine in clinical trials [1425]. This agreement hopes to establish scientific and business relations between the entities with an aim to co-develop a vaccine providing long-term immunization. The trial, which will begin enrollment soon, will include 100 participants in a phase II open-label study and is hoped to be complete within 6 months. Participants will first receive an intramuscular dose of AZD1222 on day 1, followed by a dose of rAd26 on day 29. Participants will be monitored from day 1 for 180 days in total. The primary outcomes measured will include incidence of serious adverse events post first dose until the end of the study. Secondary outcome measures will include incidence of local and systemic adverse events 7 days post each dose, a time course of antibody responses for the Spike protein and the presence of anti-SARS-CoV-2 neutralizing antibodies [1426].

Overall, there is hesitancy surrounding the management of the Sputnik V vaccine approval process and concerns over whether the efficacy data may be inflated due to a lack of asymptomatic testing within the trial. However, the interim results of the phase III study were promising and further trials are underway, which will likely shed light on the overall efficacy and safety of the Sputnik V vaccine regimen. There may be some advantage to the Sputnik V approach including the favorable storage conditions afforded by choice between a frozen and lyophilized vaccine. Furthermore, the producers of Gam-COVID-Vac state that they can produce the vaccine at a cost of less than \$10 per dose or less than \$20 per patient [1427].

7.5.3 Janssen's JNJ-78436735

The Johnson & Johnson (J&J) vaccine developed by Janssen Pharmaceuticals, Inc., a subsidiary of J&J, was conducted in collaboration with and funded by "Operation Warp Speed" [1294,1295]. The vaccine candidate JNJ-78436735, formerly known as Ad26.COV2-S, is a monovalent vaccine that is composed of a replication-deficient adenovirus serotype 26 (Ad26) vector expressing the stabilized pre-fusion S protein of SARS-CoV-2 [1111,1296]. The vaccine was developed using Janssen's AdVac® and PER.C6 platforms that were previously utilized to develop the European Commission-approved Ebola vaccine (Ad26 ZEBOV and MVN-BN-Filo) and their Zika, respiratory syncytial (RSV), and HIV investigational vaccine candidates [1428].

The development of a single-dose vaccine was desirable by J&J from the outset, with global deployment being a key priority [1297]. Using their AdVac® technology, the vaccine can remain stable for up to two years between -15°C and -25°C and at least three months at 2-8°C [1428]. This allows the vaccine to be distributed easily without the requirement for very low temperature storage, unlike many of the other COVID-19 vaccine candidates. J&J screened numerous potential vaccine candidates *in vitro* and in animal models using varying different designs of the S protein, heterologous signal peptides, and prefusion-stabilizing substitutions [1111].

A select few candidates were further investigated as a single dose regimen in Syrian golden hamsters, a single dose regimen in rhesus macaques, and a single- and two-dose regimen in both adult and aged rhesus macaques [1111,1297,1298,1299]. From these studies, the JNJ-78436735 candidate was selected for its favorable immunogenicity profile and ease of manufacturability [1111,1297,1298,1299]. A SARS-CoV-2 challenge study in rhesus macaques showed that vaccine doses as low as 2×10^9 viral particles/mL was sufficient to induce strong protection in bronchoalveolar lavage but that doses higher than 1.125×10^{10} were required to close achieve close to complete protection in nasal swabs [1429]. Indeed, six months post-immunization, levels of S-binding and neutralizing antibodies in rhesus macaques indicated that the JNJ-78436735 vaccine conferred durable protection against SARS-CoV-2 [1300].

Following selection of the JNJ-78436735 vaccine, J&J began phase I/Ila trials. The interim phase I/Ila data was placed on the *medRxiv* preprint server on September 25th, 2020 [1430] and was later published in the *New England Journal of Medicine* on January 13th, 2021 [1296]. The phase I/Ila multi-center, randomized, placebo-controlled trial enrolled 402 healthy participants between 18-55 years old and a further 403 healthy older participants ≥ 65 years old [1296]. Patients were administered either a placebo, a low dose (5×10^{10} viral particles per mL), or a high dose (1×10^{11} viral particles per mL) intramuscularly as part of either a single- or two-dose regimen. All patients received injections 56 days apart, but participants in the single-dose condition received the placebo at the second appointment. Those who received only one dose of either vaccine received a placebo dose at their second vaccination visit. A comparison of the single versus double dose regimen has yet to be published. The primary endpoints of both the trial were safety and reactogenicity of each dose. Fatigue, headache, myalgia, and pain at the injection site were the most frequent solicited adverse events reported by participants. Although less common, particularly for those in the elderly cohort and those on the low dose regimen, the most frequent systemic adverse effect was fever. Overall, immunization was well tolerated, particularly at the lower dose concentration. In terms of reactogenicity, over 90% of those who received either the low or high dose demonstrated seroconversion in a neutralization assay using wild-type SARS-CoV-2, 29 days after immunization [1296]. Neutralizing geometric mean ratio of antibody titers (GMT) between 224-354 were detected regardless of age. By day 57, 100% of the 18-55 year old participants had neutralizing GMT (288-488), which remained stable until day 71. In the ≥ 65 years old cohort, the incidence of seroconversion for the low- and high-dose was 96% and 88% respectively by day 29.

GMTs for the low and high doses were slightly lower for participants ≥ 65 years old (196 and 127 respectively), potentially indicating slightly lower immunogenicity. Seroconversion of the S antibodies was detected in 99% of individuals between 18-55 years old for the low and high doses (GMTs 528 and 695 respectively), with similar findings reported for the ≥ 65 years old. Indeed, both dose concentrations also induced robust Th1 cytokine-producing S-specific CD4 $^+$ T cells and CD8 $^+$ T cell responses in both age groups. The findings of the phase I/Ila study supported further investigation of a single immunization using the low dose vaccine. Therefore, 25 patients were enrolled for a second randomized double-blind, placebo-controlled

phase 1 clinical trial currently being conducted in Boston, Massachusetts for 2 years [1431]. Participants received either a single dose followed by a placebo, or a double dose of either a low dose (5×10^{10} viral particles/mL) or a high dose (1×10^{11} viral particles/mL) vaccine administered intramuscularly on day 1 or day 57. Placebo-only recipients received a placebo dose on day 1 and 57. Interim analyses conducted on day 71 indicated that binding and neutralizing antibodies developed 8 days after administration in 90% and 25% of vaccine recipients, respectively. Binding and neutralizing antibodies were detected in 100% of vaccine recipients by day 57 after a single dose immunization. Spike-specific antibodies were highly prevalent (GMT 2432 to 5729) as were neutralizing antibodies (GMT 242 to 449) in the vaccinated groups. Indeed, CD4⁺ and CD8⁺ T-cell responses were also induced, which may provide additional protection, particularly if antibodies wane or poorly respond to infection [1432].

On September 23rd, 2020, J&J launched its phase III trial ENSEMBLE and released the study protocol to the public [1428,1433]. The trial intended to enroll 60,000 volunteers to assess the safety and efficacy of the single vaccine dose versus placebo with primary endpoints of 14 and 28 days post-immunization [1428]. The trial was conducted in Argentina, Brazil, Chile, Colombia, Mexico, Peru, South Africa, and the U.S. The trial was paused briefly in October 2020 to investigate a “serious medical event”, but resumed shortly after [1434]. An interim analysis was reported via press release on January 29th, 2021 [1302,1303]. The interim data included 43,783 participants who accrued 468 symptomatic cases of COVID-19. It was reported that the JNJ-78436735 vaccine was 66% effective across all regions studied for the prevention of moderate to severe COVID-19 28 days post-vaccination in those aged 18 years and older. Notably, JNJ-78436735 was 85% effective for the prevention of laboratory-confirmed severe COVID-19 and 100% protection against COVID-19-related hospitalization and death 28 days post-vaccination across all study sites. Efficacy of the vaccine against severe COVID-19 increased over time, and there were no cases of COVID-19 reported in immunized participants after day 49. The trial also determined that the vaccine candidate has a favorable safety profile as determined by an independent Data and Safety Monitoring Board. The vaccine was well tolerated, consistent with previous vaccines produced using the AdVac® platform. Fever occurred in 9% of vaccine recipients, with grade 3 fever occurring in only 0.2% of recipients. Serious adverse events were reportedly higher in the placebo group than the vaccine group, and no anaphylaxis was reported [1303].

At the time the phase III trial was being conducted, several concerning variants, including B.1.1.7 [469] and B.1.351 [229], were spreading across the globe. In particular, B.1.351 was first identified in South Africa, which was one of the JNJ-78436735 vaccine trial sites. Therefore, the J&J investigators also analyzed the efficacy of the JNJ-78436735 vaccine associated with their various trial sites to determine any potential risk of reduced efficacy as a result of the novel variants. It was determined that JNJ-78436735 was 72% effective in the U.S., 66% effective in Latin America, and 57% effective in South Africa 28 days post-vaccination. These findings underpin the importance of monitoring for the emergence of novel SARS-CoV-2 variants and determining their effects on vaccine efficacy.

Looking forward, Janssen are also running a phase III randomized, double-blind, placebo-controlled clinical trial, Ensemble 2, which aims to assess the efficacy, safety, and immunogenicity of a two-dose regimen of JNJ-78436735 administered 57 days apart. This trial will enroll 30,000 participants ≥ 18 years old from Belgium, Colombia, France, Germany, Philippines, South Africa, Spain, U.K., and the U.S. [1435]. This trial will also include participants with and without comorbidities associated with an increased risk of COVID-19.

7.5.4 Overall Status of Viral-Vector Vaccines

The three viral-vector vaccines described above have demonstrated the potential for this technology to facilitate a quick response to an emerging pathogen. However, two of the three vaccines have faced a number of criticisms surrounding the implementation of their clinical trials. <-To Do: Suggestion to move some of the Sputnik controversy here, along with describing the issues with the AstraZeneca trial->

Additionally, though the vaccines are built using similar principles, there are some differences that might influence their efficacy as SARS-CoV-2 evolves. <-To Do: suggestion to discuss prefusion conformation (J&J) vs not (the other two)->

7.6 Sinovac's CoronaVac

The CoronaVac vaccine is being developed by Sinovac, a Beijing-based biopharmaceutical company. The vaccine is using an inactivate whole virus with the addition of an aluminum adjuvant [1436]. The vaccine is currently in Phase III clinical trials.

Pre-clinical trials were performed using BALB/c mice and rhesus macaques [1150]. The SARS-CoV-2 strains used in this trial isolated from 11 hospitalized patients (5 from China, 3 from Italy, 1 from the UK, 1 from Spain, 1 from Switzerland). A phylogenetic analysis demonstrated that the strains were representative of the current circulating variants. One of the strains, CN2, from China was used as the inactivated and purified virus while the other 10 strains were used to challenge. The CN2 was grown in Vero cells. An ELISA assay was used to assess the immunogenicity of the vaccine. 10 mice were injected with the vaccine on day 0 and day 7 with varying doses (0, 1.5, 3 or 6 μg), and 10 mice were treated with physiological saline as the control. IgG developed in the serum of all vaccinated mice. Using the same setup, immunogenicity was also assessed in macaques. Four macaques were assigned to each of four groups: treatment with 3 μg at day 0, 7 and 14, treatment with a high dose of 6 μg at day 0, 7 and 14, administration of a placebo vaccine, and administration of only the adjuvant. All vaccinated macaques induced IgGs and neutralizing antibodies. After challenge with SARS-CoV-2 strain CN1, vaccinated macaques were protected compared to control macaques (placebo or adjuvant only) based on histology and viral loads collected from different regions of the lung.

Phase I/II clinical trials were conducted in adults 18-59 years old [1151] and adults over 60 years old [1149] in China. In the case of adults 18-59 years old, a single center, randomized, double-blind, placebo-controlled phase I/II trial was conducted in April 2020. Patients in this study were recruited from the community in Suining County of Jiangsu province, China. For the phase I trial, 144 (of 185 screened) participants were enrolled, with 72 enrolled in the 14-day interval cohort (i.e., treated on day 0 and day 14) and 72 in the 28-day interval cohort. This group of 72 participants was split into 2 blocks for a low-dose (3 µg) and high-dose (6 µg) vaccine. Within each block, participants were randomly assigned vaccination with CoronaVac or placebo (aluminum diluent without the virus) at a 2:1 ratio. Both the vaccine and placebo were prepared in a Good Manufacturing Practice-accredited facility of Sinovac Life Sciences (Beijing, China).

The phase II trial followed the same organization of participants, this time using 300 enrolled participants in the 14-day and another 300 enrolled in the 28-day groups. One change of note was that the vaccine was produced using a highly automated bioreactor (ReadyToProcess WAVE 25, GE, Umeå, Sweden) to increase vaccine production capacity. This change resulted in a higher intact spike protein content. The authors of this study were not aware of this antigen-level difference between the vaccine batches for the phase I/II when the ethical approval for the trials occurred.

To assess adverse responses, participants were asked to record any events up to 7 days post-treatment. The reported adverse events were graded according to the China National Medical Products Administration guidelines. In the phase I trial, the overall incidence of adverse reactions was 29-38% of patients in the 0 to 14 day group and 13-17% in the 0 to 28 day vaccination group. The most common symptom was pain at the injection site, which was reported by 17-21% of patients in the 0 to 14 day cohort and 13% in the 0 to 28 day cohort. Most adverse reactions were mild (grade 1) where patients recovered within 48 hours. A single case of acute hypersensitivity with manifestation of urticaria 48 hours following the first dose of study drug was reported in the 6 µg group. Most adverse reactions were mild (grade 1) in severity and participants recovered within 48 hours. There was a single case, from the 6 µg group, of acute hypersensitivity with manifestation of urticaria 48 hours after the first dose. Both the 14-day and 28-day cohorts had a strong neutralizing Ab response. The neutralizing Ab response was measured using a micro cytopathogenic effect assay, which assesses the minimum dilution of neutralizing Ab to be 50% protective against structural changes in host cells in response to viral infection [1386]. Additionally IgG antibody titers against the receptor binding domain were also measured using ELISA.

Another phase I/II study was performed with older patients (older than 60 years) [1149]. The study conducted a single-center, randomized, double-blind, placebo-controlled trial. The phase I trial looked at dose escalation using 3 dosages: 1.5, 3 and 6 µg. The mean age of participants was 65.8 years (std = 4.8). Of 95 screened participants, 72 were enrolled. These 72 participants were split into low (3 µg) and high (6 µg) dose groups. Within each group, 24 participants received the treatment and 12 the placebo. A neutralizing antibody response against live SARS-CoV-2 was detected compared to baseline using the same micro cytopathogenic effect assay. This

response was similar across the two dose concentrations. Additionally, they did not observe a difference in response between age groups (60–64 years, 65–69 years, and ≥70 years).

In phase II the mean age was 66.6 years (standard deviation = 4.7). 499 participants were screened and 350 were enrolled. 300 were evenly split into 1.5, 3 and 6 µg dose groups, and the remaining 50 were assigned to the placebo group. Again, they found a neutralizing antibody response in phase II. There wasn't a significant different between the response to 3 µg versus 6 µg, but the response was higher than that to 1.5 µg.

Participants were required to record adverse reaction events within the first 7 days after each dose. The safety results were combined across phase I and II. All adverse reactions were either mild (grade 1) or moderate (grade 2) in severity. The most common symptom was pain at the injection site (9%) and fever (3%). 2% (7 participants) reported severe adverse events (4 from the 1.5 µg group, 1 from the 3 µg group, 2 from the 6 µg group), though these were found to be unrelated to the vaccine.

Overall, the results from the pre-clinical and phase I/II clinical trials are promising. It was also very hopeful to see that the immune response was consistent in older adults (> 60 years). Currently, phase III trials are being conducted in Brazil [1387]. This is a randomized, multicenter, endpoint driven, double-blind, placebo-controlled clinical trial. They are expecting 13,060 participants with 11,800 ages 18 to 59 years and 1,260 age 60+. Participants will be health care professionals.

7.6.1 Sinopharm (2 candidates)

Sinopharm Wuhan Institute also developed their SARS-CoV-2 inactivated vaccine using the WIV04 strain isolated from a patient at the Jinyintan Hospital in Wuhan, China [1156]. This vaccine was also passaged in Vero cells, inactivated using β-propiolactone, and was adjuvanted with aluminum hydroxide. This vaccine is administered intramuscularly using 5 µg of virus per dose. Preclinical data providing supporting evidence for the use of this vaccine is not available publicly. Despite the lack of publicly available preclinical results, Sinopharm Wuhan Institute initiated phase I/II trials, which reported on varying dosing and prime-boost regimens. Neutralizing antibodies were detected in all groups 14 days after the final dose in the phase I part of the trial [1158]. Similar findings were reported in the Interim phase II data [1158].

A combined phase I/II RCT of SinoPharm's BBIBP-CorV followed [1164]. In phase I, 192 participants were randomized with varying doses of 2 µg, 4 µg, or 8 µg/dose or a placebo, and they received the same as a second dose 28 days later. Approximately 29% of participants reported at least 1 adverse event, most commonly fever, and neutralizing antibody titers were reported for all doses. In the phase II trial, 482 participants were enrolled (3:1, vaccine:placebo). Participants in the vaccine condition received either a single 8 µg dose or a double immunization of a 4 µg/dose that was administered 14, 21, or 28 days post the prime dose. Participants in the placebo condition received the placebo on one of the same four schedules. The vaccine appeared well-tolerated, with 23% reporting at least one adverse reaction

characterized as mild to moderate. It was reported that all participants had a humoral immune response to the vaccines by day 42 but that the double immunization dosing regimen of 4 µg/dose achieved higher neutralizing antibody titers than a single dose of 8 µg and that the highest response was seen in the double-immunization regimen when at least 21 days separated the two doses [1164]. Similar findings were reported in another phase I/II trial published by the same authors [1168].

7.7 Protein Subunit Vaccines

Several different technologies are used to develop vaccines in this category. Proteins can be grown in yeast and then harvested, as they are culturable devoid of animal-derived growth factors. Indeed, the vaccine industry has previously mostly used *Pichia pastoris* yeast as the expression system [1128]. However, in recent years insect cells have also been utilized [1191,1230,1437]. Other protein subunit vaccines utilize virus-like particles (VLPs), which share the conformation of a virus's capsid, thereby acting as an antigen, but lack the replication machinery [1194]. VLPs are often synthesized using nanotechnology [1438].

7.7.1 Novavax NVX-CoV2373

Novavax-CoV2373 is a protein nanoparticle vaccine candidate against SARS-CoV-2. The vaccine is constructed from a mutated SARS-CoV-2 spike protein in combination with a specialized adjuvant to elicit an immune response against SARS-CoV-2. The spike protein is recombinantly expressed in Sf9 insect cells [1230], which have previously been used for several other FDA-approved protein therapeutics [1231]. The expressed spike protein contains mutations in the furin cleavage site (682-RRAR-685 to 682-QQAQ-685) to avoid cleavage of the spike protein as well as two proline substitutions (K986P and V987P) to improve thermostability [1230]. The improved stability caused by the proline substitutions is particularly critical to facilitating global distribution, particularly to regions where local refrigerator/freezer capacities are limited. Importantly, these amino acid substitutions did not affect the ability of the spike protein to bind the hACE2 receptor (the target receptor of SARS-CoV-2 spike protein). The Novavax-CoV2373 vaccine candidate uses a proprietary, saponin-based Matrix-MTM adjuvant that contains two different 40nm-sized particles formed by formulating purified saponin with cholesterol and phospholipids [1439]. In preclinical models, the use of the Matrix-M adjuvant potentiated the cellular and humoral immune responses to influenza vaccines [1439,1440,1441,1442]. Importantly, Matrix-M adjuvant-containing vaccines have shown acceptable safety profiles in human clinical trials [1443].

In preclinical mouse models, Novavax-CoV2373 elicited high anti-spike IgG titers 21-28 days post-vaccination that could neutralize the SARS-CoV-2 virus and protect the animals against virus challenge [1230]. Antibody titers were significantly elevated in groups receiving the vaccine with the Matrix-M adjuvant compared to the groups without adjuvant. Novavax-CoV2373 was able to induce a multifunctional CD4/CD8 T-cell responses and generate high frequencies of follicular helper T-cells and B-cell germinal centers after vaccination. These findings were subsequently evaluated in a baboon

primate model, in which Novavax-CoV2373 also elicited high antibody titers against the SARS-CoV-2 spike protein, as well as an antigen specific T-cell response. Based on this data Novavax initiated a Phase 1/2 clinical trial to evaluate the safety and immunogenicity of Novavax-CoV2373 with Matrix-M [1232,1444].

The phase I/II trial was a randomized, placebo-controlled study with 131 healthy adult participants in 5 treatment arms [1232]. Participants that received the recombinant SARS-CoV-2 vaccine with or without the Matrix-M adjuvant got two injections, 21 days apart. Primary outcomes that were assessed include reactogenicity, lab-values (serum chemistry and hematology), and anti-spike IgG levels. Secondary outcomes measured included virus neutralization, T-cell responses, and unsolicited adverse events. The authors reported that no serious treatment-related adverse events occurred in any of the treatment arms. Reactogenicity was mostly absent and of short duration. The two-dose vaccine regimen induced anti-spike IgG levels and neutralizing antibody-titers exceeding those in the convalescent plasma of symptomatic patients. In line with the preclinical studies, the use of Matrix-M adjuvant further increased anti-spike immunoglobulin levels and induced a Th1 response. The outcomes of this trial suggest that Novavax-CoV2373 has an acceptable safety profile and is able to induce a strong immune response with high neutralizing antibody titers. The phase II component of this phase I/II trial was recently uploaded to an open-access repository [1445]. This part of the trial was designed to identify which dosing regimen should move forward into late phase clinical trials. Both younger (18-59 years) and older patients (60-84 years) were randomly assigned to receive either 5 µg or 25 µg Novavax-CoV2373 or placebo in two doses, 21 days apart. In line with the phase I data, reactogenicity remained mild to moderate and of short duration. Both dose levels were able to induce high anti-spike IgG titers as well as neutralizing antibody responses after the second dose. Based on both safety and efficacy data, the 5 µg dosing regimen was selected as the optimal dose regimen for the ongoing phase III trial. Although the phase III trial data has not been published yet, Novavax announced an efficacy of 89.3% based on their phase 3 trial in the UK and South Africa. This trial included over 15,000 participants in the UK and 4,000 participants in South Africa with occurrence of a PCR-confirmed symptomatic case as the primary endpoint. In the first interim analysis (U.K.), 56 cases of COVID-19 were observed in the placebo group compared to 6 cases in the treatment group. Importantly, the vaccine candidate also shows significant clinical efficacy against the prevalent UK and South African variants. The company has also initiated the development of new constructs to select candidates that can be used as a booster against new strains and plans to initiate clinical trials for these new constructs in the second quarter of 2021.

The primary endpoint of the trial was the occurrence or absence of PCR-confirmed, symptomatic mild, moderate or severe COVID-19 from 7 days after the second dose onward. Side effects were monitored in 2,310 participants and were generally considered mild, with low incidences of headache, muscle pain, and fatigue.

7.7.2 Protein Subunit Vaccine Development Programs Prior to SARS-CoV-2

Another set of studies has examined the immunogenicity of a SARS-CoV-1 RBD fused with IgG1 Fc. This recombinant fusion protein could induce a robust long-lasting neutralizing antibody and cellular immune response that protected mice from SARS-CoV-1 [1128,1198,1199]. While there have been other potential protein subunit vaccines for SARS-CoV-1 investigated *in vivo* [1128,1208], none of these candidates have successfully completed clinical trials, more than likely due to the fact that the SARS-CoV-1 epidemic mostly ended by May 2004, and there was thus less of a demand or funding for SARS-CoV-1 vaccine research.

Similar vaccine candidates have emerged that target the RBD found in the S1 subunit of the trimeric MERS-CoV S protein, which binds to dipeptidyl-peptidase 4 (DPP4 also known as hCD26), the entry point through which MERS-CoV infects cells [1446,1447,1448]. After initially determining that an RBD subunit candidate (S377-588-Fc) could elicit neutralizing antibodies [1449], a study in mice determined that the administration of three sequential doses of RBD-Fc vaccine coupled with MF59, a squalene immunogenic adjuvant, induced humoral and systemic immunity in mice [1450]. Mice that had been transduced with Ad5-hCD26 and subsequently challenged with MERS-CoV five days later did not show evidence of viral infection in the lungs versus control mice at ten days post vaccination [1450]. Other variations of this vaccine approach include a stable S trimer vaccine whereby proline-substituted variants of S2 can maintain a stable prefusion conformation of the S2 domain [1128]. This approach leads to broad and potent neutralizing antibodies [1128]

7.7.3 ZF2001

Phase I/II trial

7.8 SARS-CoV-2 Evolution and Vaccine Efficacy

7.8.1 Delta Variant and C_t

One preprint [1370] analyzed a retrospective cohort of patients in Singapore who contracted COVID-19 from April to June of 2021.

This study focused on those who were confirmed or inferred to have been infected by the Delta variant of concern and its aim was to analyze virological kinetics. They identified 218 cases, 71 (33%) of whom were fully vaccinated with either the Pfizer/BioNTech or Moderna mRNA vaccines, 13 (6%) of whom had received only one dose or had received the second dose less than two weeks prior to infection, and four (2%) of whom had received a vaccine developed with another technology. Unvaccinated patients were more likely to be symptomatic or to progress to severe COVID-19 and showed more symptoms than vaccinated patients, despite the higher age of the vaccinated cohort. C_t was assessed over disease course, although the specific procedures for when additional RT-PCR was conducted is not clear; however, it is stated that the data was smoothed based on day of illness. There was no significant difference in median C_t in the initial samples taken from fully vaccinated and unvaccinated patients, but C_t increased (signifying reduced viral load) more rapidly in fully vaccinated patients. Like most analyses

analyzing C_t [1], this study does not provide the data to make conclusions about contagiousness, as the samples were not cultured. All the same, these findings do suggest that vaccinated individuals may be able to clear the infection more quickly.

A second analysis was based in a county of Wisconsin, USA during summer 2021, when the Delta variant was known to be the dominant variant in the region [1371]. According to Our World in Data, at the beginning of the study, 49.3% of residents of Dane County were fully vaccinated, with this number rising to 51.4% by the end of the study, although an earlier version of the preprint reported the vaccination rate in Dane County as 67.4%. They identified no significant differences in C_t among fully vaccinated and unvaccinated cases. The C_t thresholds reported were consistent with contagiousness as evaluated in other studies, and in the present study, SARS-CoV-2 could be cultured from 51 of 55 samples with C_t less than 25. This study was not longitudinal, but the timing of testing relative to symptom onset between symptomatic vaccinated and unvaccinated patients. The findings of this study are therefore consistent with the idea that vaccinated people are less likely to contract symptomatic or severe COVID-19, but in cases of breakthrough infection, are still likely to be able to transmit SARS-CoV-2 to others.

7.9 Complementary Approaches to Vaccine Development

A complementary approach to other vaccine development programs that is being investigated explores the potential for vaccines that are not made from the SARS-CoV-2 virus to confer what has been termed trained immunity. In a recent review [1451], trained immunity was defined as forms of memory that are temporary (e.g., months or years) and reversible. It is induced by exposure to whole-microorganism vaccines or other microbial stimuli that generates heterologous protective effects. Trained immunity can be displayed by innate immune cells or innate immune features of other cells, and it is characterized by alterations to immune responsiveness to future immune challenges due to epigenetic and metabolic mechanisms. These alterations can take the form of either an increased or decreased response to immune challenge by a pathogen. Trained immunity elicited by non-SARS-CoV-2 whole-microorganism vaccines could potentially improve SARS-CoV-2 susceptibility or severity [1452].

One type of stimulus which research indicates can induce trained immunity is bacillus Calmette-Guerin (BCG) vaccination. BCG is an attenuated form of bacteria *Mycobacterium bovis*. The vaccine is most commonly administered for the prevention of tuberculosis in humans. Clinical trials in non-SARS-CoV-2-infected adults have been designed to assess whether BCG vaccination could have prophylactic effects against SARS-CoV-2 by reducing susceptibility, preventing infection, or reducing disease severity. A number of trials are now evaluating the effects of the BCG vaccine or the related vaccine VPM1002 [1384, 1385, 1452, 1453, 1454, 1455, 1456, 1457, 1458, 1459, 1460, 1461, 1462, 1463, 1464].

The ongoing trials are using a number of different approaches. Some trials enroll healthcare workers, other trials hospitalized elderly adults without immunosuppression who get vaccinated with placebo or BCG at hospital discharge, and yet another set of trials older adults (>50 years) under chronic care for conditions like hypertension and diabetes. One set of trials, for example, uses time until first infection as the primary study endpoint; more generally, outcomes measured in some of these trials are related to incidence of disease and disease severity or symptoms. Some analyses have suggested a possible correlation at the country level between the frequency of BCG vaccination (or BCG vaccination policies) and the severity of COVID-19 [1452]. Currently it is unclear whether this correlation has any connection to trained immunity. Many possible confounding factors are also likely to vary among countries, such as age distribution, detection efficiency, stochastic epidemic dynamic effects, differences in healthcare capacity over time in relation to epidemic dynamics, and these have not been adequately accounted for in current analyses. It is unclear whether there is an effect of the timing of BCG vaccination, both during an individual's life cycle and relative to the COVID-19 pandemic. Additionally, given that severe SARS-CoV-2 may be associated with a dysregulated immune response, it is unclear what alterations to the immune response would be most likely to be protective versus pathogenic (e.g., [147,1452,1465,1466]). The article [1452] proposes that trained immunity might lead to an earlier and stronger response, which could in turn reduce viremia and the risk of later, detrimental immunopathology. While trained immunity is an interesting possible avenue to complement vaccine development efforts through the use of an existing vaccine, additional research is required to assess whether the BCG vaccine is likely to confer trained immunity in the case of SARS-CoV-2.

7.10 Viral evolution and vaccine protection

With these vaccines in place, one concern is how the virus's continued evolution will affect their efficacy. Since the start of this pandemic, we have already seen multiple variants emerge: B.1.1.7, which emerged in the UK, B.1.351, which emerged in South Africa, and P.1, which emerged in Brazil.

Viruses evolve or mutate at different rates. Mutation rate is measured as the number of substitutions per nucleotide per cell infected ($\mu_{S/n/c}$) [1467]. RNA viruses tend to have mutation rates between 10^{-6} to 10^{-4} [1467]. As a reference, influenza A virus has a mutation rate of 10^{-5} , whereas the mutation rate of SARS-CoV-2 is lower, with the mutation rate estimated at 10^{-6} [1468]. The accumulation of mutations allows the virus to escape recognition by the immune system [1469].

The efficacy of vaccines depends on their ability to train the immune system to recognize the virus. Therefore, viruses can develop resistance to vaccines through the accumulation of mutations that affect recognition. The lower mutation rate of SARS-CoV-2 suggests the possibility of SARS-CoV-2 vaccines having a more long-lasting effect compared to vaccines targeting the influenza A virus.

The current SARS-CoV-2 vaccines in distribution have been reported to provide similar efficacy against the B.1.1.7 variant compared to the variants common at the time they were developed but reduced efficacy against the B.1.351 variant [1470]. Pfizer and Moderna announced that they are working on developing a booster shot to improve efficacy against the B.1.351 variant [1471]. The WHO continues to monitor the emergence of variants and their impact on vaccine efficacy [1472]. Previous research in the computational prediction of the efficacy of vaccines targeting the influenza A virus might complement efforts to monitor these types of viral outbreaks [1473]. To adapt, future vaccines may need to account for multiple variants and strains of SARS-CoV-2, and booster shots may be required [1474].

7.11 Global Vaccine Status and Distribution

The unprecedented development of COVID-19 vaccines in under a year since the beginning of the pandemic now requires rapid global vaccine production and distribution plans. The development of vaccines is costly and complicated, but vaccine distribution can be just as challenging. Logistical considerations such as transport, storage, equipment (e.g., syringes), the workforce to administer the vaccines, and a continual supply from the manufacturers to meet global demands all must be accounted for and will vary globally due to economic, geographic, and sociopolitical reasons [1475,1476,1477]. Deciding on the prioritization and allocation of the COVID-19 vaccines is also a challenging task due to ethical and operational considerations. Various frameworks, models, and methods have been proposed to tackle these issues with many countries, regions or states as is the case in the U.S., devising their own distribution and administration plans [1478,1479,1480,1481,1482]. The majority of the distribution plans prioritize offering vaccines to key workers such as health care workers, and those who are clinically vulnerable such as the elderly, the immunocompromised, and individuals with comorbidities, before targeting the rest of the population, who are less likely to experience severe outcomes from COVID-19 [1483]. As of March 6th, 2021, approximately 319 million vaccine doses have been administered in at least 118 countries worldwide using 10 different vaccines [1484,1485]. The global vaccination rate is currently ~8.1 million doses per day, which at the current rate would take almost 4 years to vaccinate 75% of the world's population according to media estimates of a two-dose regimen [1485]. Vaccine production and distribution varies from region to region and seems to depend on the availability of the vaccines and potentially a country's resources and wealth [1486].

In North America, the majority of vaccines distributed until March 2021 have been produced by Pfizer-BioNTech and Moderna. In Canada, the vaccine approval process is conducted by Health Canada, which uses a fast-tracked process whereby vaccine producers can submit data as it becomes available to allow for rapid review. An approval may be granted following reviews of the available phase III clinical data. This is followed by a period of pharmacovigilance in the population using their post-market surveillance system, which will monitor the long-term safety and efficacy of any vaccines [1487,1488]. Health Canada has authorized the use of the Pfizer (December 9th, 2020), Moderna (December 23rd, 2020), Oxford-AstraZeneca (February 26th, 2021), and the Janssen (March 5th, 2021) vaccines, and the Novavax Inc vaccine is also under consideration [1489]. While Canada initially projected

that by the end of September 2021 a vaccine would be available for all Canadian adults, they now predict that it may be possible earlier as more vaccines have been approved and become available [1490].

In the U.S., vaccines are required to have demonstrated safety and efficacy in phase III trials before manufacturers apply for an emergency use authorization (EUA) from the FDA. If an EUA is granted, an additional evaluation of the safety and efficacy of the vaccines is conducted by the CDC's Advisory Committee on Immunization Practices (ACIP) who also provide guidance on vaccine prioritization. On December 1st, 2020, ACIP provided an interim phase 1a recommendation that healthcare workers and long-term care facility residents should be the first to be offered any vaccine approved [1491]. This was shortly followed by an EUA on December 11th, 2020 for the use of the Pfizer-BioNTech COVID vaccine [1492], which was distributed and administered to the first healthcare workers on December 14th, 2020 [1493]. Shortly thereafter, an EUA for the Moderna vaccine was issued on December 18th, 2020 [1494]. On December 20th, 2020, ACIP updated their initial recommendations to suggest that vaccinations should be offered to people aged 75 years old and older and to non-healthcare frontline workers in phase 1b [1495]. On the same date, it was recommended that phase 1c should include people aged 65-74 years old, individuals between the ages of 16-74 years old at high-risk due to health conditions, and essential workers ineligible in phase 1b [1495]. On the following day, December 21st, 2020, the first Moderna vaccines used outside of clinical trials were administered to American healthcare workers, which was the same day that President-elect Biden and Dr. Biden received their first doses of the Pfizer-BioNTech vaccine live on television to instill confidence in the approval and vaccination process [1496].

On February 27th, 2020, the FDA issued an EUA for the Janssen COVID-19 Vaccine [1497]. This was followed by an update on recommendations by ACIP for the use of the Janssen COVID-19 vaccine for those over 18 years old [1498]. The Janssen vaccine was first distributed to healthcare facilities on March 1st, 2021. On March 12, 2021, the WHO added the Janssen vaccine to the list of safe and effective emergency tools for COVID-19 [1499]. While the CDC's ACIP can provide recommendations, it is up to the public health authorities of each state, territory, and tribe to interpret the guidance and determine who will be vaccinated first [1500]. Prior to distribution of the Janssen vaccine, over 103 million doses of the Moderna and Pfizer-BioNTech vaccines were delivered across the U.S., with almost 79 million doses administered. Of the total population, 15.6% have received at least one dose and 7.9% have received a second dose of either the Moderna (~38.3 million) or the Pfizer-BioNTech (~40.2 million) vaccines by February 28th, 2021 [120/#vaccinations]. President Biden's administration has predicted that by the end of May 2021 there may be enough vaccine supply available for all adults in the U.S. [1501,1502]. However, vaccine production, approval, and distribution was not straightforward in the U.S., as information was initially sparse and the rollout of vaccines was complicated by poor planning and leadership due to political activities prior to the change of administration in January 2021 [1503]. These political complications highlight the importance of the transparent vaccine approval process conducted by the FDA [1504].

Outside the U.S., the Moderna and Pfizer-BioNTech vaccines have been administered in 29 and 69 other countries, respectively, mainly in Europe and North America [1484]. The Janssen vaccine has so far only been administered in South Africa and the U.S. [1484,1505], but it has also been approved in Bahrain, the European Union (E.U.), Iceland, Liechtenstein, and Norway [1155]. On March 11th, 2021, Johnson & Johnson received approval from the European Medicines Agency (EMA) for conditional marketing authorization of their vaccine [1506]. Notably, on March 2nd, 2021, rivals Johnson & Johnson and Merck announced that they entered an agreement to increase production of the Janssen vaccine to meet global demand [1507].

The U.K. was the first country to approve use of the Pfizer-BioNTech vaccine on December 2nd, 2020 [1508], and it was later approved by EMA on December 21st, 2020 [1509]. The U.K. was also the first to administer the Pfizer-BioNTech vaccine, making it the first COVID-19 vaccine supported by phase III data to be administered outside of clinical trials on December 8th, 2020. The Oxford-AstraZeneca vaccine, was approved by the Medicines and Healthcare Products Regulatory Agency (MHRA) in the U.K. and by EMA in the E.U. on December 30th (2020) [1510] and January 29th (2021) [1511] respectively. The Oxford-AstraZeneca vaccine was first administered in the UK on January 4th, 2021 [1512], and it is now being used in 53 countries in total, including Brazil, India, Pakistan, Mexico, and spanning most of Europe [1484]. The Moderna vaccine was authorized for use in the E.U. by EMA on January 6th, 2021 [1513] and in the U.K. by MHRA on January 8th, 2021 [1514]. As of March 5th, 2021, 22 million people in the U.K. had received at least one vaccine dose [1229].

While the Pfizer-BioNTech vaccine was the first to be distributed following phase III clinical trials, the first COVID-19 vaccine to be widely administered to people prior to the completion of phase III clinical trials was Sputnik V. Sputnik V was administered to as many as 1.5 million Russians by early January [1305] due to the establishment of mass vaccination clinics in December 2020, prior to which only approximately 100,000 Russians had already been vaccinated [1515,1516]/?sh=50650e4e62e1]. Doses of Sputnik V have also been distributed to other parts of Europe, such as Belarus, Bosnia-Herzegovina, Hungary, San Marino, Serbia, and Slovakia [1306,1307,1308], with the Czech Republic and Austria also having expressed interest in its procurement [1309]. Hungary was the first E.U. member country to approve and distribute Sputnik V outside of Russia [1309], despite the EMA stating that they had neither approved nor received a request for approval of Sputnik V [1517]. Hungary is also in talks with China to procure the Sinopharm vaccines, which have been approved by Hungarian health authorities but also have not received approval by EMA in the E.U. [1309]. In Latin America, production facilities in both Brazil and Argentina will allow for increased production capacity of Sputnik V and doses have been distributed to Mexico, Argentina, Bolivia, Nicaragua, Paraguay, and Venezuela [1518]. Guinea was the first African nation to administer Sputnik V in December 2020, and the Central African Republic, Zimbabwe, and the Ivory Coast have all registered their interest in purchasing doses of the vaccine [1518]. In the Middle East, Iran has received its first doses of Sputnik V and the United Arab Emirates is conducting phase III trials [1518]. In Asia, while China's vaccine candidates are favored, the Philippines, Nepal, and Uzbekistan have sought Sputnik V doses [1519]. In total, the RDIF claims to have received orders

totalling 1.2 billion doses by over 50 countries worldwide [1519] and at least 18 countries are currently administering Sputnik V around the globe [1484]. Sputnik V has been an attractive vaccine for many countries due to its relatively low price, high efficacy, and its favorable storage conditions. For some countries, Russia and China have also been more palatable politically than vaccine suppliers in the West [1518,1520]. For others, the delays in the distribution of the other, more-favored candidates has been a motivating factor for pursuing the Sputnik V and Chinese alternatives [1307,1520]. Additionally, Germany has stated that if Sputnik V were approved by EMA, it would be considered by the E.U. [1521]. Russia is developing other vaccine candidates and has approved a third vaccine, CoviVac, which is an inactivated vaccine produced by the Chumakov Centre in Moscow, despite the fact the clinical trials have yet to begin [1522].

In Asia, China and India are the main COVID-19 vaccination developers and providers. In India, the Covaxin vaccine produced by Bharat Biotech received emergency authorization on January 3rd, 2021, despite the lack of phase III data until March 3rd [1389]. Following the release of the phase III data indicating 81% efficacy, Zimbabwe authorized the use of Covaxin [1391]. In February, 2021, Bharat Biotech received approval from Indian officials to commence a phase I study of an intranasal chimpanzee adenovirus (ChAd) vectored SARS-CoV-2-S vaccine called BBV154 [1523]. Notably, Novavax has signed an agreement with the Serum Institute of India allowing them to produce up to 2 billion doses a year [1524]. Novavax has also signed agreements with the U.K., Canada, Australia, and South Korea [1525] and has projected that they will supply 1.1 billion doses to COVAX who will distribute the vaccines to countries with disadvantaged access to vaccine supplies [1155]. India has vaccinated approximately 24 million people [1485]. This has been achieved by mainly using the AstraZeneca-University of Oxford vaccine, known as Covishield in India, which is also produced by the Serum Institute of India, and using India's own Covaxin vaccine [1526]. India has also shipped approximately 58 million COVID-19 vaccines to 66 countries [1527]. Considering India produces approximately 60% of the world's vaccines prior to the pandemic, it is no surprise that several other vaccine candidates are under development. These include ZyCov-Di, a DNA vaccine produced by Zydus Cadila, HGCO19, India's first mRNA vaccine produced by Genova and HDT Biotech Corporation (of the U.S.), and the Bio E subunit vaccine produced by Biological E in collaboration with U.S.-based Dynavax and the Baylor College of Medicine [1526].

In China, the Sinopharm-Beijing Institute vaccine, the Sinopharm-Wuhan Institute of Biological Products vaccine, the Sinovac Biotech (CoronaVac) vaccine, and CanSino Biologics vaccine are the main vaccines being distributed. The Sinopharm-Beijing vaccine has been distributed to at least 16 countries. This vaccine is currently approved for use in Bahrain, China, and the United Arab Emirates, but has been granted emergency use in Argentina, Cambodia, Egypt, Guyana, Hungary, Iran, Iraq, Jordan, Nepal, Pakistan, Peru, Venezuela, and Zimbabwe, with limited use in both Serbia and the Seychelles [1528]. Indeed, Sinovac and Sinopharm have estimated that they will be able to produce 2 billion doses by the end of 2021, and they have been able to distribute vaccines as aid to the Philippines and Pakistan [1529]. In contrast, the Sinopharm-Wuhan vaccine, which has been approved for use in China since February 25th, 2021, has been distributed almost

exclusively within China, with limited supplies distributed to the United Arab Emirates [1530]. On the same date, the CanSino vaccine was approved for use in China and has been granted emergency use in Mexico and Pakistan, which were two participating countries in the CanSino phase III trials [1531]. However, the vaccine approval and distribution processes in China have come under increased scrutiny from other nations. China was criticized for administering vaccines to thousands of government officials and state-owned businesses in September 2020, prior to the completion of phase III clinical trials [1504]. The behavior of Chinese officials has also come into question due to misinformation campaigns questioning the safety of Western vaccine candidates such as Moderna and Pfizer-BioNTech in a way that is intended to highlight the benefits of their own vaccine candidates [1529]. Furthermore, delays in vaccine distribution have also caused issues, particularly in Turkey where 10 million doses of Sinovac were due to arrive by December 2020, but instead only 3 million were delivered in early January [1529]. Similar delays and shortages of doses promised have been reported by officials in the Philippines, Egypt, Morocco, and the United Arab Emirates [1532,1533]. This will be concerning to China who have vaccine contracts for millions of doses with Indonesia (>100 million), Brazil (100 million), Chile (60 million), Turkey (50 million), Egypt (40 million) and many others [1533]. As of September 2021, CoronaVac trials are now also being held in the Philippines and Hong Kong, bringing the total number of registered phase III trials investigating the safety and efficacy of CoronaVac to 9, with emergency use approval in 40 countries [1388]. However, concern has been raised about the efficacy of CoronaVac following reports that over 350 doctors became ill with COVID-19 in Indonesia despite being immunized with CoronaVac [1180].

Globally, North America currently leads the world vaccination rates (13.8 per 100 people) followed by Europe (8.2 per 100), South America (3.1 per 100), Asia (1.9 per 100), Africa (0.3 per 100), and Oceania (0.1 per 100) are trailing behind [1484]. Considering the wealthy nations of North America and Europe have secured most of the limited COVID-19 vaccine stocks [1534], it is likely that low- and middle-income countries will face further competition with Western countries for vaccine availability. While South Africa and Zimbabwe have their own vaccination programs, many other African nations will be reliant on the COVID-19 Vaccines Global Access (COVAX) Facility, who have promised 600 million doses to the continent [1535]. COVAX is a multilateral initiative as part of the Access to COVID-19 Tools (ACT) Accelerator coordinated by the WHO, Gavi The Vaccine Alliance, and the Coalition for Epidemic Preparedness Innovations (CEPI), the latter two of which are supported by the Bill and Melinda Gates Foundation. Their intention is to accelerate the development of COVID-19 vaccines, diagnostics, and therapeutics and to ensure the equitable distribution of vaccines to low- and middle-income countries [1536,1537]. COVAX invested in several vaccine programs to ensure they would have access to successful vaccine candidates [1538]. The COVAX plan ensured that all participating countries would be allocated vaccines in proportion to their population sizes. Once each country has received vaccine doses to account for 20% of their population, the country's risk profile will determine its place in subsequent phases of vaccine distribution. However, several limitations of this framework exist, including that the COVAX scheme seems to go against the WHO's own ethical principles of human well-being, equal respect, and global equity, and that other frameworks might have been more suitable, as is discussed elsewhere

[1539]. Furthermore, COVAX is supposed to allow poorer countries access to affordable vaccines, but the vaccines are driven by publicly traded companies that are required to make a profit [1486]. In any case, COVAX provides access to COVID-19 vaccines that may otherwise have been difficult for some countries to obtain. COVAX aims to distribute 2 billion vaccine doses globally by the end of 2021 [1540]. COVAX may also receive additional donations of doses from Western nations who purchased surplus vaccines in the race to vaccinate their populations, which will be a welcome boost to the vaccination programs of low- and middle-income countries [1541]. As of March, 2021, 9 African countries have received vaccines and at least 11 other nations have begun vaccinations via COVAX, aid from other countries, or their own agreements with producers [1535,1542]. However, much further progress is required when only 0.3 per 100 people have been vaccinated in Africa [1484].

7.12 Discussion

Additionally, major advances in vaccines using mRNA and adenoviruses that have led to three vaccines becoming available or close to becoming available in late 2020.

Though some concerns remain about the duration of sustained immunity for convalescents, vaccine development efforts are ongoing and show initial promising results. The Moderna trial, for example, reported that the neutralizing activity in participants who received two doses of the vaccine was similar to that observed in convalescent plasma.

One of the two mRNA vaccines, Pfizer and BioNTech's BNT162b2, has been issued an EUA for patients as young as 16 [1543], while ModernaTX has begun a clinical trial to assess its mRNA vaccine in adolescents ages 12 to 18 [1544].

8 Dietary Supplements and Nutraceuticals Under Investigation for COVID-19 Prevention and Treatment

8.1 Abstract

Coronavirus disease 2019 (COVID-19) has caused global disruption and a significant loss of life. Existing treatments that can be repurposed as prophylactic and therapeutic agents could reduce the pandemic's devastation. Emerging evidence of potential applications in other therapeutic contexts has led to the investigation of dietary supplements and nutraceuticals for COVID-19. Such products include vitamin C, vitamin D, omega 3 polyunsaturated fatty acids, probiotics, and zinc, all of which are currently under clinical investigation. In this review, we critically appraise the evidence surrounding dietary supplements and nutraceuticals for the prophylaxis and treatment of COVID-19. Overall, further study is required

before evidence-based recommendations can be formulated, but nutritional status plays a significant role in patient outcomes, and these products could help alleviate deficiencies. For example, evidence indicates that vitamin D deficiency may be associated with greater incidence of infection and severity of COVID-19, suggesting that vitamin D supplementation may hold prophylactic or therapeutic value. A growing number of scientific organizations are now considering recommending vitamin D supplementation to those at high risk of COVID-19. Because research in vitamin D and other nutraceuticals and supplements is preliminary, here we evaluate the extent to which these nutraceutical and dietary supplements hold potential in the COVID-19 crisis.

8.2 Importance

Sales of dietary supplements and nutraceuticals have increased during the pandemic due to their perceived “immune-boosting” effects. However, little is known about the efficacy of these dietary supplements and nutraceuticals against the novel coronavirus (SARS-CoV-2) or the disease it causes, COVID-19. This review provides a critical overview of the potential prophylactic and therapeutic value of various dietary supplements and nutraceuticals from the evidence available to date. These include vitamin C, vitamin D, and zinc, which are often perceived by the public as treating respiratory infections or supporting immune health. Consumers need to be aware of misinformation and false promises surrounding some supplements, which may be subject to limited regulation by authorities. However, considerably more research is required to determine whether dietary supplements and nutraceuticals exhibit prophylactic and therapeutic value against SARS-CoV-2 infection and COVID-19. This review provides perspective on which nutraceuticals and supplements are involved in biological processes that are relevant to recovery from or prevention of COVID-19.

8.3 Introduction

The year 2020 saw scientists and the medical community scrambling to repurpose or discover novel host-directed therapies against the coronavirus disease 2019 (COVID-19) pandemic caused by the spread of the novel *Severe acute respiratory syndrome-related coronavirus 2* (SARS-CoV-2). This rapid effort led to the identification of some promising pharmaceutical therapies for hospitalized patients, such as remdesivir and dexamethasone. Furthermore, most societies have adopted non-pharmacological preventative measures such as utilizing public health strategies that reduce the transmission of SARS-CoV-2. However, during this time, many individuals sought additional protections via the consumption of various dietary supplements and nutraceuticals that they believed to confer beneficial effects. While a patient’s nutritional status does seem to play a role in COVID-19 susceptibility and outcomes [[1545](#),[1546](#),[1547](#),[1548](#),[1549](#)], the beginning of the pandemic saw sales of vitamins and other supplements soar despite a lack of any evidence supporting their use against COVID-19. In the United States, for example, dietary supplement and nutraceutical sales have shown modest annual growth in recent years (approximately 5%, or a \$345 million increase in 2019), but during the six-week period preceding April 5, 2020, they increased by 44% (\$435 million) relative to the same period in 2019.

[1550]. While growth subsequently leveled off, sales continued to boom, with a further 16% (\$151 million) increase during the six weeks preceding May 17, 2020 relative to 2019 [1550]. In France, New Zealand, India, and China, similar trends in sales were reported [1551,1552,1553,1554]. The increase in sales was driven by a consumer perception that dietary supplements and nutraceuticals would protect consumers from infection and/or mitigate the impact of infection due to the various “immune-boosting” claims of these products [1555,1556].

Due to the significant interest from the general public in dietary additives, whether and to what extent nutraceuticals or dietary supplements can provide any prophylactic or therapeutic benefit remains a topic of interest for the scientific community. Nutraceuticals and dietary supplements are related but distinct non-pharmaceutical products. Nutraceuticals are classified as supplements with health benefits beyond their basic nutritional value [1557,1558]. The key difference between a dietary supplement and a nutraceutical is that nutraceuticals should not only supplement the diet, but also aid in the prophylaxis and/or treatment of a disorder or disease [1559]. However, dietary supplements and nutraceuticals, unlike pharmaceuticals, are not subject to the same regulatory protocols that protect consumers of medicines. Indeed, nutraceuticals do not entirely fall under the responsibility of the Food and Drug Administration (FDA), but they are monitored as dietary supplements according to the Dietary Supplement, Health and Education Act 1994 (DSHEA) [1560] and the Food and Drug Administration Modernization Act 1997 (FDAMA) [1561]. Due to increases in sales of dietary supplements and nutraceuticals, in 1996 the FDA established the Office of Dietary Supplement Programs (ODSP) to increase surveillance. Novel products or nutraceuticals must now submit a new dietary ingredient notification to the ODSP for review. There are significant concerns that these legislations do not adequately protect the consumer as they ascribe responsibility to the manufacturers to ensure the safety of the product before manufacturing or marketing [1562]. Manufacturers are not required to register or even seek approval from the FDA to produce or sell food supplements or nutraceuticals. Health or nutrient content claims for labeling purposes are approved based on an authoritative statement from the Academy of Sciences or relevant federal authorities once the FDA has been notified and on the basis that the information is known to be true and not deceptive [1562]. Therefore, there is often a gap between perceptions by the American public about a nutraceutical or dietary supplement and the actual clinical evidence surrounding its effects.

Despite differences in regulations, similar challenges exist outside of the United States. In Europe, where the safety of supplements is monitored by the European Union (EU) under Directive 2002/46/EC [1563]/?uri=celex%3A32002L0046]. However, nutraceuticals are not directly mentioned. Consequently, nutraceuticals can be generally described as either a medicinal product under Directive 2004/27/EC [1564]/?uri=CELEX:32004L0027] or as a ‘foodstuff’ under Directive 2002/46/EC of the European council. In order to synchronize the various existing legislations, Regulation EC 1924/2006 on nutrition and health claims was put into effect to assure customers of safety and efficacy of products and to deliver understandable information to consumers. However, specific legislation for nutraceuticals is still elusive. Health claims are permitted on a product label

only following compliance and authorization according to the European Food Safety Authority (EFSA) guidelines on nutrition and health claims [1565]. EFSA does not currently distinguish between food supplements and nutraceuticals for health claim applications of new products, as claim authorization is dependent on the availability of clinical data in order to substantiate efficacy [1566]. These guidelines seem to provide more protection to consumers than the FDA regulations but potentially at the cost of innovation in the sector [1567]. The situation becomes even more complicated when comparing regulations at a global level, as countries such as China and India have existing regulatory frameworks for traditional medicines and phytomedicines not commonly consumed in Western society [1568]. Currently, there is debate among scientists and regulatory authorities surrounding the development of a widespread regulatory framework to deal with the challenges of safety and health claim substantiation for nutraceuticals [1562,1566], as these products do not necessarily follow the same rigorous clinical trial frameworks used to approve the use of pharmaceuticals. Such regulatory disparities have been highlighted by the pandemic, as many individuals and companies have attempted to profit from the vulnerabilities of others by overstating claims in relation to the treatment of COVID-19 using supplements and nutraceuticals. The FDA has written several letters to prevent companies marketing or selling products based on false hyperbolic promises about preventing SARS-CoV-2 infection or treating COVID-19 [1569,1570,1571]. These letters came in response to efforts to market nutraceutical prophylactics against COVID-19, some of which charged the consumer as much as \$23,000 [1572]. There have even been some incidents highlighted in the media because of their potentially life threatening consequences; for example, the use of oleandrin was touted as a potential "cure" by individuals close to the former President of the United States despite its high toxicity [1573]. Thus, heterogeneous and at times relaxed regulatory standards have permitted high-profile cases of the sale of nutraceuticals and dietary supplements that are purported to provide protection against COVID-19, despite a lack of research into these compounds.

Notwithstanding the issues of poor safety, efficacy, and regulatory oversight, some dietary supplements and nutraceuticals have exhibited therapeutic and prophylactic potential. Some have been linked with reduced immunopathology, antiviral and anti-inflammatory activities, or even the prevention of acute respiratory distress syndrome (ARDS) [1555,1574,1575]. A host of potential candidates have been highlighted in the literature that target various aspects of the COVID-19 viral pathology, while others are thought to prime the host immune system. These candidates include vitamins and minerals along with extracts and omega-3 polyunsaturated fatty acids (n-3 PUFA) [1576]. *In vitro* and *in vivo* studies suggest that nutraceuticals containing phycocyanobilin, N-acetylcysteine, glucosamine, selenium or phase 2 inductive nutraceuticals (e.g. ferulic acid, lipoic acid, or sulforaphane) can prevent or modulate RNA virus infections via amplification of the signaling activity of mitochondrial antiviral-signaling protein (MAVS) and activation of Toll-like receptor 7 [1577]. Phase 2 inductive molecules used in the production of nutraceuticals are known to activate nuclear factor erythroid 2-related factor 2 (Nrf2), which is a protein regulator of antioxidant enzymes that leads to the induction of several antioxidant enzymes, such as gamma-glutamylcysteine synthetase. While promising, further animal and

human studies are required to assess the therapeutic potential of these various nutrients and nutraceuticals against COVID-19. For the purpose of this review, we have highlighted some of the main dietary supplements and nutraceuticals that are currently under investigation for their potential prophylactic and therapeutic applications. These include n-3 PUFA, zinc, vitamins C and D, and probiotics.

8.4 n-3 PUFA

One category of supplements that has been explored for beneficial effects against various viral infections are the n-3 PUFAs [1576], commonly referred to as omega-3 fatty acids, which include eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). EPA and DHA intake can come from a diet high in fish or through dietary supplementation with fish oils or purified oils [1578]. Other, more sustainable sources of EPA and DHA include algae [1579,1580], which can also be exploited for their rich abundance of other bioactive compounds such as angiotensin converting enzyme inhibitor peptides and antiviral agents including phycobiliproteins, sulfated polysaccharides, and calcium-spirulan [1581]. n-3 PUFAs have been investigated for many years for their therapeutic potential [1582]. Supplementation with fish oils is generally well tolerated [1582], and intake of n-3 PUFAs through dietary sources or supplementation is specifically encouraged for vulnerable groups such as pregnant and lactating women [1583,1584]. As a result, these well-established compounds have drawn significant interest for their potential immune effects and therapeutic potential.

Particular interest has arisen in n-3 PUFAs as potential therapeutics against diseases associated with inflammation. n-3 PUFAs have been found to modulate inflammation by influencing processes such as leukocyte chemotaxis, adhesion molecule expression, and the production of eicosanoids [1585,1586]. This and other evidence indicates that n-3 PUFAs may have the capacity to modulate the adaptive immune response [1558,1578,1585]; for example, they have been found to influence antigen presentation and the production of CD4(+) Th1 cells, among other relevant effects [1587]. Certainly, preliminary evidence from banked blood samples from 100 COVID-19 patients suggests that patients with a higher omega-3 index, a measure of the amount of EPA and DHA in red blood cells, had a lower risk of death due to COVID-19 [1588]. Interest has also arisen as to whether nutritional status related to n-3 PUFAs can also affect inflammation associated with severe disease, such as ARDS or sepsis [1589,1590]. ARDS and sepsis hold particular concern in the treatment of severe COVID-19; an analysis of 82 deceased COVID-19 patients in Wuhan during January to February 2020 reported that respiratory failure (associated with ARDS) was the cause of death in 69.5% of cases, and sepsis or multi-organ failure accounted for 28.0% of deaths [720]. Research in ARDS prior to current pandemic suggests that n-3 PUFAs may hold some therapeutic potential. One study randomized 16 consecutive ARDS patients to receive either a fish oil-enriched lipid emulsion or a control lipid emulsion (comprised of 100% long-chain triglycerides) under a double-blinded design [1591]. They reported a statistically significant reduction in leukotriene B4 levels in the group receiving the fish oil-enriched emulsion, suggesting that the fish oil supplementation may have reduced inflammation. However, they also

reported that most of their tests were not statistically significant, and therefore it seems that additional research using larger sample sizes is required. A recent meta-analysis of 10 randomized controlled trials (RCTs) examining the effects of n-3 PUFAs on ARDS patients did not find evidence of any effect on mortality, although the effect on secondary outcomes could not be determined due to a low quality of evidence [1592]. However, another meta-analysis that examined 24 RCTs studying the effects of n-3 fatty acids on sepsis, including ARDS-induced sepsis, did find support for an effect on mortality when n-3 fatty acids were administered via enteral nutrition, although a paucity of high-quality evidence again limited conclusions [1593]. Therefore, despite theoretical support for an immunomodulatory effect of n-3 PUFAs in COVID-19, evidence from existing RCTs is insufficient to determine whether supplementation offers an advantage in a clinical setting that would be relevant to COVID-19.

Another potential mechanism that has led to interest in n-3 PUFAs as protective against viral infections including COVID-19 is its potential as a precursor molecule for the biosynthesis of endogenous specialized proresolving mediators (SPM), such as protectins and resolvins, that actively resolve inflammation and infection [1594]. SPM have exhibited beneficial effects against a variety of lung infections, including some caused by RNA viruses [1595,1596]. Several mechanisms for SPM have been proposed, including preventing the release of pro-inflammatory cytokines and chemokines or increasing phagocytosis of cellular debris by macrophages [1597]. In influenza, SPM promote antiviral B lymphocytic activities [1598], and protectin D1 has been shown to increase survival from H1N1 viral infection in mice by affecting the viral replication machinery [1599]. It has thus been hypothesized that SPM could aid in the resolution of the cytokine storm and pulmonary inflammation associated with COVID-19 [1600,1601]. Another theory is that some comorbidities, such as obesity, could lead to deficiencies of SPM, which could in turn be related to the occurrence of adverse outcomes for COVID-19 [1602]. However, not all studies are in agreement that n-3 PUFAs or their resulting SPM are effective against infections [1603]. At a minimum, the effectiveness of n-3 PUFAs against infections would be dependent on the dosage, timing, and the specific pathogens responsible [1604]. On another level, there is still the question of whether fish oils can raise the levels of SPM levels upon ingestion and in response to acute inflammation in humans [1605]. Currently, Karolinska University Hospital is running a trial that will measure the levels of SPM as a secondary outcome following intravenous supplementation of n-3 PUFAs in hospitalized COVID-19 patients to determine whether n-3 PUFAs provides therapeutic value [1606,1607]. Therefore, while this mechanism provides theoretical support for a role for n-3 PUFAs against COVID-19, experimental support is still needed.

A third possible mechanism by which n-3 PUFAs could benefit COVID-19 patients arises from the fact that some COVID-19 patients, particularly those with comorbidities, are at a significant risk of thrombotic complications including arterial and venous thrombosis [103,1608]. Therefore, the use of prophylactic and therapeutic anticoagulants and antithrombotic agents is under consideration [1609,1610]. Considering that there is significant evidence that n-3 fatty acids and other fish oil-derived lipids possess antithrombotic properties and anti-inflammatory properties

[1578,1611,1612], they may have therapeutic value against the prothrombotic complications of COVID-19. In particular, concerns have been raised within the medical community about using investigational therapeutics on COVID-19 patients who are already on antiplatelet therapies due to pre-existing comorbidities because the introduction of such therapeutics could lead to issues with dosing and drug choice and/or negative drug-drug interactions [1609]. In such cases, dietary sources of n-3 fatty acids or other nutraceuticals with antiplatelet activities could hold particular value for reducing the risk of thrombotic complications in patients already receiving pharmaceutical antiplatelet therapies. A new clinical trial [1613] is currently recruiting COVID-19 positive patients to investigate the anti-inflammatory activity of a recently developed, highly purified nutraceutical derivative of EPA known as icosapent ethyl (VascepaTM) [1614]. Other randomized controlled trials that are in the preparatory stages intend to investigate the administration of EPA and other bioactive compounds to COVID-19 positive patients in order to observe whether anti-inflammatory effects or disease state improvements occur [1615,1616]. Finally, while there have been studies investigating the therapeutic value of n-3 fatty acids against ARDS in humans, there is still limited evidence of their effectiveness [1617]. It should be noted that the overall lack of human studies in this area means there is limited evidence as to whether these supplements could affect COVID-19 infection. Consequently, the clinical trials that are underway and those that have been proposed will provide valuable insight into whether the anti-inflammatory potential of n-3 PUFAs and their derivatives can be beneficial to the treatment of COVID-19. All the same, while the evidence is not present to draw conclusions about whether n-3 PUFAs will be useful in treating COVID-19, there is likely little harm associated with a diet rich in fish oils, and interest in n-3 PUFA supplementation by the general public is unlikely to have negative effects.

8.5 Zinc

Zinc is nutrient supplement that may exhibit some benefits against RNA viral infections. Zinc is a trace metal obtained from dietary sources or supplementation and is important for the maintenance of immune cells involved in adaptive and innate immunity [1618]. Supplements can be administered orally as a tablet or as a lozenge and are available in many forms, such as zinc picolinate, zinc acetate, and zinc citrate. Zinc is also available from dietary sources including meat, seafood, nuts, seeds, legumes, and dairy. The role of zinc in immune function has been extensively reviewed [1618]. Zinc is an important signaling molecule, and zinc levels can alter host defense systems. In inflammatory situations such as an infection, zinc can regulate leukocyte immune responses and modulate the nuclear factor kappa-light-chain-enhancer of activated B cells, thus altering cytokine production [1619,1620]. In particular, zinc supplementation can increase natural killer cell levels, which are important cells for host defense against viral infections [1618,1621]. As a result of these immune-related functions, zinc is also under consideration for possible benefits against COVID-19.

Adequate zinc intake has been associated with reduced incidence of infection [1622] and antiviral immunity [1623]. A randomized, double-blind, placebo-controlled trial that administered zinc supplementation to elderly subjects over the course of a year found that zinc supplementation decreased

susceptibility to infection and that zinc deficiency was associated with increased susceptibility to infection [1622]. Clinical trial data supports the utility of zinc to diminish the duration and severity of symptoms associated with common colds when it is provided within 24 hours of the onset of symptoms [1624,1625]. An observational study showed that COVID-19 patients had significantly lower zinc levels in comparison to healthy controls and that zinc-deficient COVID-19 patients (those with levels less than 80 µg/dl) tended to have more complications (70.4% vs 30.0%, $p = 0.009$) and potentially prolonged hospital stays (7.9 vs 5.7 days, $p = 0.048$) relative to patients who were not zinc deficient [1626]. In coronaviruses specifically, *in vitro* evidence has demonstrated that the combination of zinc (Zn^{2+}) and zinc ionophores (pyrithione) can interrupt the replication mechanisms of SARS-CoV-GFP (a fluorescently tagged SARS-CoV-1) and a variety of other RNA viruses [1627,1628]. Currently, there are over twenty clinical trials registered with the intention to use zinc in a preventative or therapeutic manner for COVID-19. However, many of these trials proposed the use of zinc in conjunction with hydroxychloroquine and azithromycin [1629,1630,1631,1632], and it is not known how the lack of evidence supporting the use of hydroxychloroquine will affect investigation of zinc. One retrospective observational study of New York University Langone hospitals in New York compared outcomes among hospitalized COVID-19 patients administered hydroxychloroquine and azithromycin with zinc sulfate ($n = 411$) versus hydroxychloroquine and azithromycin alone ($n = 521$). Notably, zinc is the only treatment that was used in this trial that is still under consideration as a therapeutic agent due to the lack of efficacy and potential adverse events associated with hydroxychloroquine and azithromycin against COVID-19 [1633,1634,1635]. While the addition of zinc sulfate did not affect the duration of hospitalization, the length of ICU stays or patient ventilation duration, univariate analyses indicated that zinc did increase the frequency of patients discharged and decreased the requirement for ventilation, referrals to the ICU, and mortality [1636]. However, a smaller retrospective study at Hoboken University Medical Center New Jersey failed to find an association between zinc supplementation and survival of hospitalized patients [1637]. Therefore, whether zinc contributes to COVID-19 recovery remains unclear. Other trials are now investigating zinc in conjunction with other supplements such as vitamin C or n-3 PUFA [1616,1638]. Though there is, overall, encouraging data for zinc supplementation against the common cold and viral infections, there is currently limited evidence to suggest zinc supplementation has any beneficial effects against the current novel COVID-19; thus, the clinical trials that are currently underway will provide vital information on the efficacious use of zinc in COVID-19 prevention and/or treatment. However, given the limited risk and the potential association between zinc deficiency and illness, maintaining a healthy diet to ensure an adequate zinc status may be advisable for individuals seeking to reduce their likelihood of infection.

8.6 Vitamin C

Vitamins B, C, D, and E have also been suggested as potential nutrient supplement interventions for COVID-19 [1576,1639]. In particular vitamin C has been proposed as a potential therapeutic agent against COVID-19 due to its long history of use against the common cold and other respiratory infections [1640,1641]. Vitamin C can be obtained via dietary sources such as

fruits and vegetables or via supplementation. Vitamin C plays a significant role in promoting immune function due to its effects on various immune cells. It affects inflammation by modulating cytokine production, decreasing histamine levels, enhancing the differentiation and proliferation of T- and B-lymphocytes, increasing antibody levels, and protecting against the negative effects of reactive oxygen species, among other effects related to COVID-19 pathology [1642,1643,1644]. Vitamin C is utilized by the body during viral infections, as evinced by lower concentrations in leukocytes and lower concentrations of urinary vitamin C. Post-infection, these levels return to baseline ranges [1645,1646,1647,1648,1649]. It has been shown that as little as 0.1 g/d of vitamin C can maintain normal plasma levels of vitamin C in healthy individuals, but higher doses of at least 1-3 g/d are required for critically ill patients in ICUs [1650]. Indeed, vitamin C deficiency appears to be common among COVID-19 patients [1651,1652]. COVID-19 is also associated with the formation of microthrombi and coagulopathy [105] that contribute to its characteristic lung pathology [1653], but these symptoms can be ameliorated by early infusions of vitamin C to inhibit endothelial surface P-selectin expression and platelet-endothelial adhesion [1654]. Intravenous vitamin C also reduced D-dimer levels in a case study of 17 COVID-19 patients [1655]. D-dimer levels are an important indicator of thrombus formation and breakdown and are notably elevated in COVID-19 patients [101,102]. There is therefore preliminary evidence suggesting that vitamin C status and vitamin C administration may be relevant to COVID-19 outcomes.

Larger-scale studies of vitamin C, however, have provided mixed results. A recent meta-analysis found consistent support for regular vitamin C supplementation reducing the duration of the common cold, but that supplementation with vitamin C (> 200 mg) failed to reduce the incidence of colds [1656]. Individual studies have found Vitamin C to reduce the susceptibility of patients to lower respiratory tract infections, such as pneumonia [1657]. Another meta-analysis demonstrated that in twelve trials, vitamin C supplementation reduced the length of stay of patients in intensive care units (ICUs) by 7.8% (95% CI: 4.2% to 11.2%; $p = 0.00003$). Furthermore, high doses (1-3 g/day) significantly reduced the length of an ICU stay by 8.6% in six trials ($p = 0.003$). Vitamin C also shortened the duration of mechanical ventilation by 18.2% in three trials in which patients required intervention for over 24 hours (95% CI 7.7% to 27%; $p = 0.001$) [1650]. Despite these findings, an RCT of 167 patients known as CITRUS ALI failed to show a benefit of a 96-hour infusion of vitamin C to treat ARDS [1658]. Clinical trials specifically investigating vitamin C in the context of COVID-19 have now begun, as highlighted by Carr et al. [1641]. These trials intend to investigate the use of intravenous vitamin C in hospitalized COVID-19 patients. The first trial to report initial results took place in Wuhan, China [1659]. These initial results indicated that the administration of 12 g/12 hr of intravenous vitamin C for 7 days in 56 critically ill COVID-19 patients resulted in a promising reduction of 28-day mortality ($p = 0.06$) in univariate survival analysis [1660]. Indeed, the same study reported a significant decrease in IL-6 levels by day 7 of vitamin C infusion ($p = 0.04$) [1661]. Additional studies that are being conducted in Canada, China, Iran, and the USA will provide additional insight into whether vitamin C supplementation affects COVID-19 outcomes on a larger scale.

Even though evidence supporting the use of vitamin C is beginning to emerge, we will not know how effective vitamin C is as a therapeutic for quite some time. Currently (as of January 2021) over fifteen trials are registered with clinicaltrials.gov that are either recruiting, active or are currently in preparation. When completed, these trials will provide crucial evidence on the efficacy of vitamin C as a therapeutic for COVID-19 infection. However, the majority of supplementation studies investigate the intravenous infusion of vitamin C in severe patients. Therefore, there is a lack of studies investigating the potential prophylactic administration of vitamin C via oral supplementation for healthy individuals or potentially asymptomatic SARS-CoV-2 positive patients. Once again, vitamin C intake is part of a healthy diet and the vitamin likely presents minimal risk, but its potential prophylactic or therapeutic effects against COVID-19 are yet to be determined. To maintain vitamin C status, it would be prudent for individuals to ensure that they consume the recommended dietary allowance of vitamin C to maintain a healthy immune system [1545]. The recommended dietary allowance according to the FDA is 75-90 mg/d, whereas EFSA recommends 110 mg/d [1662].

8.7 Vitamin D

Of all of the supplements currently under investigation, vitamin D has become a leading prophylactic and therapeutic candidate against SARS-CoV-2. Vitamin D can modulate both the adaptive and innate immune system and is associated with various aspects of immune health and antiviral defense [1663,1664,1665,1666,1667]. Vitamin D can be sourced through diet or supplementation, but it is mainly biosynthesized by the body on exposure to ultraviolet light (UVB) from sunlight. Vitamin D deficiency is associated with an increased susceptibility to infection [1668]. In particular, vitamin D deficient patients are at risk of developing acute respiratory infections [1669] and ARDS [1669]. 1,25-dihydroxyvitamin D₃ is the active form of vitamin D that is involved in adaptive and innate responses; however, due to its low concentration and a short half life of a few hours, vitamin D levels are typically measured by the longer lasting and more abundant precursor 25-hydroxyvitamin D. The vitamin D receptor is expressed in various immune cells, and vitamin D is an immunomodulator of antigen presenting cells, dendritic cells, macrophages, monocytes, and T- and B-lymphocytes [1668,1670]. Due to its potential immunomodulating properties, vitamin D supplementation may be advantageous to maintain a healthy immune system.

Early in the pandemic it was postulated that an individual's vitamin D status could significantly affect their risk of developing COVID-19 [1671]. This hypothesis was derived from the fact that the current pandemic emerged in Wuhan China during winter, when 25-hydroxyvitamin D concentrations are at their lowest due to a lack of sunlight, whereas in the Southern Hemisphere, where it was nearing the end of the summer and higher 25-hydroxyvitamin D concentrations would be higher, the number of cases was low. This led researchers to question whether there was a seasonal component to the SARS-CoV-2 pandemic and whether vitamin D levels might play a role [1671,1672,1673,1674]. Though it is assumed that COVID-19 is seasonal, multiple other factors that can affect vitamin D levels should also be considered. These factors include an individual's nutritional status, their age,

their occupation, skin pigmentation, potential comorbidities, and the variation of exposure to sunlight due to latitude amongst others. Indeed, it has been estimated that each degree of latitude north of 28 degrees corresponded to a 4.4% increase of COVID-19 mortality, indirectly linking a persons vitamin D levels via exposure to UVB light to COVID-19 mortality [1672].

As the pandemic has evolved, additional research of varying quality has investigated some of the potential links identified early in the pandemic [1671] between vitamin D and COVID-19. Indeed, studies are beginning to investigate whether there is any prophylactic and/or therapeutic relationship between vitamin D and COVID-19. A study in Switzerland demonstrated that 27 SARS-CoV-2 positive patients exhibited 25-hydroxyvitamin D plasma concentrations that were significantly lower (11.1 ng/ml) than those of SARS-CoV-2 negative patients (24.6 ng/ml; $p = 0.004$), an association that held when stratifying patients greater than 70 years old [1675]. These findings seem to be supported by a Belgian observational study of 186 SARS-CoV-2 positive patients exhibiting symptoms of pneumonia, where 25-hydroxyvitamin D plasma concentrations were measured and CT scans of the lungs were obtained upon hospitalization [1676]. A significant difference in 25-hydroxyvitamin D levels was observed between the SARS-CoV-2 patients and 2,717 season-matched hospitalized controls. It is not clear from the study which diseases caused the control subjects to be admitted at the time of their 25-hydroxyvitamin D measurement, which makes it difficult to assess the observations reported. Both female and male patients possessed lower median 25-hydroxyvitamin D concentrations than the control group as a whole (18.6 ng/ml versus 21.5 ng/ml; $p = 0.0016$) and a higher rate of vitamin D deficiency (58.6% versus 42.5%). However, when comparisons were stratified by sex, evidence of sexual dimorphism became apparent, as female patients had equivalent levels of 25-hydroxyvitamin D to females in the control group, whereas male patients were deficient in 25-hydroxyvitamin D relative to male controls (67% versus 49%; $p = 0.0006$). Notably, vitamin D deficiency was progressively lower in males with advancing radiological disease stages ($p = 0.001$). However, these studies are supported by several others that indicate that vitamin D status may be an independent risk factor for the severity of COVID-19 [1677,1678,1679,1680] and in COVID-19 patients relative to population-based controls [1681]. Indeed, serum concentrations of 25-hydroxyvitamin D above 30 ng/ml, which indicate vitamin D sufficiency, seems to be associated with a reduction in serum C-reactive protein, an inflammatory marker, along with increased lymphocyte levels, which suggests that vitamin D levels may modulate the immune response by reducing risk for cytokine storm in response to SARS-CoV-2 infection [1681]. A study in India determined that COVID-19 fatality was higher in patients with severe COVID-19 and low serum 25-hydroxyvitamin D (mean level 6.2 ng/ml; 97% vitamin D deficient) levels versus asymptomatic non-severe patients with higher levels of vitamin D (mean level 27.9 ng/ml; 33% vitamin D deficient) [1682]. In the same study, vitamin D deficiency was associated with higher levels of inflammatory markers including IL-6, ferritin, and tumor necrosis factor α . Collectively, these studies add to a multitude of observational studies reporting potential associations between low levels of 25-hydroxyvitamin D and COVID-19 incidence and severity [1675,1680,1681,1683,1684,1685,1686,1687,1688,1689].

Despite the large number of studies establishing a link between vitamin D status and COVID-19 severity, an examination of data from the UK Biobank did not support this thesis [1690,1691]. These analyses examined 25-hydroxyvitamin D concentrations alongside SARS-CoV-2 positivity and COVID-19 mortality in over 340,000 UK Biobank participants. However, these studies have caused considerable debate that will likely be settled following further studies [1692,1693]. Overall, while the evidence suggests that there is likely an association between low serum 25-hydroxyvitamin D and COVID-19 incidence, these studies must be interpreted with caution, as there is the potential for reverse causality, bias, and other confounding factors including that vitamin D deficiency is also associated with numerous pre-existing conditions and risk factors that can increase the risk for severe COVID-19 [1545,1672,1694,1695].

While these studies inform us of the potential importance of vitamin D sufficiency and the risk of SARS-CoV-2 infection and severe COVID-19, they fail to conclusively determine whether vitamin D supplementation can therapeutically affect the clinical course of COVID-19. In one study, 40 vitamin D deficient asymptomatic or mildly symptomatic participants patients were either randomized to receive 60,000 IU of cholecalciferol daily for at least 7 days ($n = 16$) or a placebo ($n = 24$) with a target serum 25-hydroxyvitamin D level >50 ng/ml. At day 7, 10 patients achieved >50 ng/ml, followed by another 2 by day 14. By the end of the study, the treatment group had a greater proportion of vitamin D-deficient participants that tested negative for SARS-CoV-2 RNA, and they had a significantly lower fibrinogen levels, potentially indicating a beneficial effect [1696]. A pilot study in Spain determined that early administration of high dose calcifediol (~21,000 IU days 1-2 and ~11,000 IU days 3-7 of hospital admission) with hydroxychloroquine and azithromycin to 50 hospitalized COVID-19 patients significantly reduced ICU admissions and may have reduced disease severity versus hydroxychloroquine and azithromycin alone [1697]. Although this study received significant criticism from the National Institute for Health and Care Excellence (NICE) in the UK [1698], an independent follow-up statistical analysis supported the findings of the study with respect to the results of cholecalciferol treatment [1699]. Another trial of 986 patients hospitalized for COVID-19 in three UK hospitals administered cholecalciferol ($\geq 280,000$ IU in a time period of 7 weeks) to 151 patients and found an association with a reduced risk of COVID-19 mortality, regardless of baseline 25-hydroxyvitamin D levels [1700]. However, a double-blind, randomized, placebo-controlled trial of 240 hospitalized COVID-19 patients in São Paulo, Brazil administered a single 200,000 IU oral dose of vitamin D. At the end of the study, there was a 24 ng/mL difference of 25-hydroxyvitamin D levels in the treatment group versus the placebo group ($p = 0.001$), and 87% of the treatment group were vitamin D sufficient versus ~11% in the placebo group. Supplementation was well tolerated. However, there was no reduction in the length of hospital stay or mortality, and no change to any other relevant secondary outcomes were reported [1701]. These early findings are thus still inconclusive with regards to the therapeutic value of vitamin D supplementation. However, other trials are underway, including one trial that is investigating the utility of vitamin D as an immune-modulating agent by monitoring whether administration of vitamin D precipitates an improvement of health status in non-severe symptomatic COVID-19 patients and whether vitamin D prevents patient deterioration [1702]. Other trials are examining various factors including

mortality, symptom recovery, severity of disease, rates of ventilation, inflammatory markers such as C-reactive protein and IL-6, blood cell counts, and the prophylactic capacity of vitamin D administration [1702,1703,1704,1705]. Concomitant administration of vitamin D with pharmaceuticals such as aspirin [1706] and bioactive molecules such as resveratrol [1707] is also under investigation.

The effectiveness of vitamin D supplementation against COVID-19 remains open for debate. All the same, there is no doubt that vitamin D deficiency is a widespread issue and should be addressed not only because of its potential link to SARS-CoV-2 incidence [1708], but also due to its importance for overall health. There is a possibility that safe exposure to sunlight could improve endogenous synthesis of vitamin D, potentially strengthening the immune system. However, sun exposure is not sufficient on its own, particularly in the winter months. Indeed, while the possible link between vitamin D status and COVID-19 is further investigated, preemptive supplementation of vitamin D and encouraging people to maintain a healthy diet for optimum vitamin D status is likely to raise serum levels of 25-hydroxyvitamin D while being unlikely to carry major health risks. These principles seem to be the basis of a number of guidelines issued by some countries and scientific organizations that have advised supplementation of vitamin D during the pandemic. The Académie Nationale de Médecine in France recommends rapid testing of 25-hydroxyvitamin D for people over 60 years old to identify those most at risk of vitamin D deficiency and advises them to obtain a bolus dose of 50,000 to 100,000 IU vitamin D to limit respiratory complications. It has also recommended that those under 60 years old should take 800 to 1,000 IU daily if they receive a SARS-CoV-2 positive test [1709]. In Slovenia, doctors have been advised to provide nursing home patients with vitamin D [1710]. Both Public Health England and Public Health Scotland have advised members of the Black, Asian, and minority ethnic communities to supplement for vitamin D in light of evidence that they may be at higher risk for vitamin D deficiency along with other COVID-19 risk factors, a trend that has also been observed in the United States [1711,1712]. However, other UK scientific bodies including the NICE recommend that individuals supplement for vitamin D as per usual UK government advice but warn that people should not supplement for vitamin D solely to prevent COVID-19. All the same, the NICE has provided guidelines for research to investigate the supplementation of vitamin D in the context of COVID-19 [1713]. Despite vitamin D deficiency being a widespread issue in the United States [1714], the National Institutes of Health have stated that there is "insufficient data to recommend either for or against the use of vitamin D for the prevention or treatment of COVID-19" [1715]. These are just some examples of how public health guidance has responded to the emerging evidence regarding vitamin D and COVID-19. Outside of official recommendations, there is also evidence that individuals may be paying increased attention to their vitamin D levels, as a survey of Polish consumers showed that 56% of respondents used vitamin D during the pandemic [1716]. However, some companies have used the emerging evidence surrounding vitamin D to sell products that claim to prevent and treat COVID-19, which in one incident required a federal court to intervene and issue an injunction barring the sale of vitamin-D-related products due to the lack of clinical data supporting these claims [1717]. It is clear that further studies and clinical trials are required to conclusively determine the prophylactic and therapeutic potential of vitamin D

supplementation against COVID-19. Until such time that sufficient evidence emerges, individuals should follow their national guidelines surrounding vitamin D intake to achieve vitamin D sufficiency.

8.8 Probiotics

Probiotics are “live microorganisms that, when administered in adequate amounts, confer a health benefit on the host” [1718]. Some studies suggest that probiotics are beneficial against common viral infections, and there is modest evidence to suggest that they can modulate the immune response [1719,1720]. As a result, it has been hypothesized that probiotics may have therapeutic value worthy of investigation against SARS-CoV-2 [1721].

Probiotics and next-generation probiotics, which are more akin to pharmacological-grade supplements, have been associated with multiple potential beneficial effects for allergies, digestive tract disorders, and even metabolic diseases through their anti-inflammatory and immunomodulatory effects [1722,1723]. However, the mechanisms by which probiotics affect these various conditions would likely differ among strains, with the ultimate effect of the probiotic depending on the heterogeneous set of bacteria present [1723]. Some of the beneficial effects of probiotics include reducing inflammation by promoting the expression of anti-inflammatory mediators, inhibiting Toll-like receptors 2 and 4, competing directly with pathogens, synthesizing antimicrobial substances or other metabolites, improving intestinal barrier function, and/or favorably altering the gut microbiota and the brain-gut axis [1723,1724,1725]. It is also thought that lactobacilli such as *Lactobacillus paracasei*, *Lactobacillus plantarum* and *Lactobacillus rhamnosus* have the capacity to bind to and inactivate some viruses via adsorptive and/or trapping mechanisms [1726]. Other probiotic lactobacilli and even non-viable bacterium-like particles have been shown to reduce both viral attachment to host cells and viral titers, along with reducing cytokine synthesis, enhancing the antiviral IFN- α response, and inducing various other antiviral mechanisms

[1726,1727,1728,1729,1730,1731,1732,1733,1734]. These antiviral and immunobiotic mechanisms and others have been reviewed in detail elsewhere [1575,1721,1735]. However, there is also a bi-directional relationship between the lungs and gut microbiota known as the gut-lung axis [1736], whereby gut microbial metabolites and endotoxins may affect the lungs via the circulatory system and the lung microbiota in return may affect the gut [1737]. Therefore, the gut-lung axis may play role in our future understanding of COVID-19 pathogenesis and become a target for probiotic treatments [1738]. Moreover, as microbial dysbiosis of the respiratory tract and gut may play a role in some viral infections, it has been suggested that SARS-CoV-2 may interact with our commensal microbiota [[1575]; [1739]; 10.3389/fmicb.2020.01840] and that the lung microbiome could play a role in developing immunity to viral infections [1740]. These postulations, if correct, could lead to the development of novel probiotic and prebiotic treatments. However, significant research is required to confirm these associations and their relevance to patient care, if any.

Probiotic therapies and prophylactics may also confer some advantages for managing symptoms of COVID-19 or risks associated with its treatment. Probiotics have tentatively been associated with the reduction of risk and duration of viral upper respiratory tract infections [1741,1742,1743]. Some

meta-analyses that have assessed the efficacy of probiotics in viral respiratory infections have reported moderate reductions in the incidence and duration of infection [1742,1744]. Indeed, randomized controlled trials have shown that administering *Bacillus subtilis* and *Enterococcus faecalis* [1745], *Lactobacillus rhamnosus GG* [1746], or *Lactobacillus casei* and *Bifidobacterium breve* with galactooligosaccharides [1747] via the nasogastric tube to ventilated patients reduced the occurrence of ventilator-associated pneumonia in comparison to the respective control groups in studies of viral infections and sepsis. These findings were also supported by a recent meta-analysis [1748]. Additionally, COVID-19 patients carry a significant risk of ventilator-associated bacterial pneumonia [1749], but it can be challenging for clinicians to diagnose this infection due to the fact that severe COVID-19 infection presents with the symptoms of pneumonia [1750]. Therefore, an effective prophylactic therapy for ventilator-associated pneumonia in severe COVID-19 patients would carry significant therapeutic value. Additionally, in recent years, probiotics have become almost synonymous with the treatment of gastrointestinal issues due to their supposed anti-inflammatory and immunomodulatory effects [1751]. Notably, gastrointestinal symptoms commonly occur in COVID-19 patients [1752], and angiotensin-converting enzyme 2, the portal by which SARS-CoV-2 enters human cells, is highly expressed in enterocytes of the ileum and colon, suggesting that these organs may be a potential route of infection [1753,1754]. Indeed, SARS-CoV-2 viral RNA has been detected in human feces [77,518], and fecal-oral transmission of the virus has not yet been ruled out [1755]. Rectal swabs of some SARS-CoV-2 positive pediatric patients persistently tested positive for several days despite negative nasopharyngeal tests, indicating the potential for fecal viral shedding [1756]. However, there is conflicting evidence for the therapeutic value of various probiotics against the incidence or severity of gastrointestinal symptoms in viral or bacterial infections such as gastroenteritis [1757,1758]. Nevertheless, it has been proposed that the administration of probiotics to COVID-19 patients and healthcare workers may prevent or ameliorate the gastrointestinal symptoms of COVID-19, a hypothesis that several clinical trials are now preparing to investigate [1759,1760]. Other studies are investigating whether probiotics may affect patient outcomes following SARS-CoV-2 infection [1761].

Generally, the efficacy of probiotic use is a controversial topic among scientists. In Europe, EFSA has banned the term probiotics on products labels, which has elicited either criticism for EFSA or support for probiotics from researchers in the field [1718,1762,1763]. This regulation is due to the hyperbolic claims placed on the labels of various probiotic products, which lack rigorous scientific data to support their efficacy. Overall, the data supporting probiotics in the treatment or prevention of many different disorders and diseases is not conclusive, as the quality of the evidence is generally considered low [1741]. However, in the case of probiotics and respiratory infections, the evidence seems to be supportive of their potential therapeutic value. Consequently, several investigations are underway to investigate the prophylactic and therapeutic potential of probiotics for COVID-19. The blind use of conventional probiotics for COVID-19 is currently cautioned against until the pathogenesis of SARS-CoV-2 can be further established [1764]. Until clinical trials investigating the prophylactic and therapeutic potential of probiotics for COVID-19 are complete, it is not

possible to provide an evidence-based recommendation for their use. Despite these concerns, complementary use of probiotics as an adjuvant therapeutic has been proposed by the Chinese National Health Commission and National Administration of Traditional Chinese Medicine [78]. While supply issues prevented the probiotics market from showing the same rapid response to the COVID-19 as some other supplements, many suppliers are reporting growth during the pandemic [1765]. Therefore, the public response once again seems to have adopted supplements promoted as bolstering the immune response despite a lack of evidence suggesting they are beneficial for preventing or mitigating COVID-19.

8.9 Discussion

In this review, we report the findings to date of analyses of several dietary supplements and nutraceuticals. While existing evidence suggests potential benefits of n-3 PUFA and probiotic supplementation for COVID-19 treatment and prophylaxis, clinical data is still lacking, although trials are underway. Both zinc and vitamin C supplementation in hospitalized patients seem to be associated with positive outcomes; however, further clinical trials are required. In any case, vitamin C and zinc intake are part of a healthy diet and likely present minimal risk when supplemented, though their potential prophylactic or therapeutic effects against COVID-19 are yet to be determined. On the other hand, mounting evidence from observational studies indicates that there is an association between vitamin D deficiency and COVID-19 incidence has also been supported by meta-analysis [1766]. Indeed, scientists are working to confirm these findings and to determine whether a patient's serum 25-hydroxyvitamin D levels are also associated with COVID-19 severity. Clinical trials are required to determine whether preemptive vitamin D supplementation may mitigate against severe COVID-19. In terms of the therapeutic potential of vitamin D, initial evidence from clinical trials is conflicting but seems to indicate that vitamin D supplementation may reduce COVID-19 severity [1697]. The various clinical trials currently underway will be imperative to provide information on the efficacious use of vitamin D supplementation for COVID-19 prevention and/or treatment.

The purported prophylactic and therapeutic benefits of dietary supplements and nutraceuticals for multiple disorders, diseases, and infections has been the subject of significant research and debate for the last few decades. Inevitably, scientists are also investigating the potential for these various products to treat or prevent COVID-19. This interest also extends to consumers, which led to a remarkable increase of sales of dietary supplements and nutraceuticals throughout the pandemic due to a desire to obtain additional protections from infection and disease. The nutraceuticals discussed in this review, namely vitamin C, vitamin D, n-3 PUFA, zinc, and probiotics, were selected because of potential biological mechanisms that could beneficially affect viral and respiratory infections and because they are currently under clinical investigation. Specifically, these compounds have all been found to influence cellular processes related to inflammation. Inflammation is particularly relevant to COVID-19 because of the negative outcomes (often death) observed in a large number of patients whose immune response becomes hyperactive in response to SARS-CoV-2, leading to severe outcomes such as ARDS and sepsis [722]. Additionally, there is a

well-established link between diet and inflammation [1767], potentially mediated in part by the microbiome [1768]. Thus, the idea that dietary modifications or supplementation could be used to modify the inflammatory response is tied to a broader view of how diet and the immune system are interconnected. The supplements and nutraceuticals discussed here therefore lie in sharp contrast to other alleged nutraceutical or dietary supplements that have attracted during the pandemic, such as colloidal silver [1769], which have no known nutritional function and can be harmful. Importantly, while little clinical evidence is available about the effects of any supplements against COVID-19, the risks associated with those discussed above are likely to be low, and in some cases, they can be obtained from dietary sources alone.

There are various other products and molecules that have garnered scientific interest and could merit further investigation. These include polyphenols, lipid extracts, and tomato-based nutraceuticals, all of which have been suggested for the potential prevention of cardiovascular complications of COVID-19 such as thrombosis [1575,1610]. Melatonin is another supplement that has been identified as a potential antiviral agent against SARS-CoV-2 using computational methods [1770], and it has also been highlighted as a potential therapeutic agent for COVID-19 due to its documented antioxidant, anti-apoptotic, immunomodulatory, and anti-inflammatory effects [1610,1771,1772]. Notably, melatonin, vitamin D and zinc have attracted public attention because they were included in the treatment plan of the former President of the United States upon his hospitalization due to COVID-19 [1773]. These are just some of the many substances and supplements that are currently under investigation but as of yet lack evidence to support their use for the prevention or treatment of COVID-19. While there is plenty of skepticism put forward by physicians and scientists surrounding the use of supplements, these statements have not stopped consumers from purchasing these products, with one study reporting that online searches for dietary supplements in Poland began trending with the start of the pandemic [1716]. Additionally, supplement usage increased between the first and second wave of the pandemic. Participants reported various reasons for their use of supplements, including to improve immunity (60%), to improve overall health (57%), and to fill nutrient gaps in their diet (53%). Other efforts to collect large datasets regarding such behavior have also sought to explore a possible association between vitamin or supplement consumption and COVID-19. An observational analysis of survey responses from 327,720 users of the COVID Symptom Study App found that the consumption of n-3 PUFA supplements, probiotics, multivitamins, and vitamin D was associated with a lower risk of SARS-CoV-2 infection in women but not men after adjusting for potential confounders [1774]. According to the authors, the sexual dimorphism observed may in part be because supplements may better support females due to known differences between the male and female immune systems, or it could be due to behavioral and health consciousness differences between the sexes [1774]. Certainly, randomized controlled trials are required to investigate these findings further.

Finally, it is known that a patient's nutritional status affects health outcomes in various infectious diseases [1549], and COVID-19 is no different [1547,1775,1776]. Some of the main risk factors for severe COVID-19, which also happen to be linked to poor nutritional status, include obesity,

hypertension, cardiovascular diseases, type II diabetes mellitus, and indeed age-related malnutrition [1545,1547,1777]. Although not the main focus of this review, it is important to consider the nutritional challenges associated with severe COVID-19 patients. Hospitalized COVID-19 patients tend to report an unusually high loss of appetite preceding admission, some suffer diarrhea and gastrointestinal symptoms that result in significantly lower food intake, and patients with poorer nutritional status were more likely to have worse outcomes and require nutrition therapy [1778]. Dysphagia also seems to be a significant problem in pediatric patients that suffered multisystem inflammatory syndrome [1779] and rehabilitating COVID-19 patients, potentially contributing to poor nutritional status [1780]. Almost two-thirds of discharged COVID-19 ICU patients exhibit significant weight loss, of which 26% had weight loss greater than 10% [1776]. As investigated in this review, hospitalized patients also tend to exhibit vitamin D deficiency or insufficiency, which may be associated with greater disease severity [1766]. Therefore, further research is required to determine how dietary supplements and nutraceuticals may contribute to the treatment of severely ill and rehabilitating patients, who often rely on enteral nutrition.

8.10 Conclusions

Despite all the potential benefits of nutraceutical and dietary supplement interventions presented, currently there is a paucity of clinical evidence to support their use for the prevention or mitigation of COVID-19 infection. Nevertheless, optimal nutritional status can prime an individual's immune system to protect against the effects of acute respiratory viral infections by supporting normal maintenance of the immune system [1545,1549]. Nutritional strategies can also play a role in the treatment of hospitalized patients, as malnutrition is a risk to COVID-19 patients [1780]. Overall, supplementation of vitamin C, vitamin D, and zinc may be an effective method of ensuring their adequate intake to maintain optimal immune function, which may also convey beneficial effects against viral infections due to their immunomodulatory effects. Individuals should pay attention to their nutritional status, particularly their intake of vitamin D, considering that vitamin D deficiency is widespread. The prevailing evidence seems to indicate an association between vitamin D deficiency with COVID-19 incidence and, potentially, severity [1672]. As a result, some international authorities have advised the general public, particularly those at high risk of infection, to consider vitamin D supplementation. However, further well-controlled clinical trials are required to confirm these observations.

Many supplements and nutraceuticals designed for various ailments that are available in the United States and beyond are not strictly regulated [1781]. Consequently, there can be safety and efficacy concerns associated with many of these products. Often, the vulnerable members of society can be exploited in this regard and, unfortunately, the COVID-19 pandemic has proven no different. As mentioned above, the FDA has issued warnings to several companies for advertising falsified claims in relation to the preventative and therapeutic capabilities of their products against COVID-19 [1782]. Further intensive investigation is required to establish the effects of these nutraceuticals, if any, against COVID-19. Until more effective therapeutics are established, the most effective mitigation strategies consist of encouraging standard public health practices such as regular hand

washing with soap, wearing a face mask, and covering a cough with your elbow [1783], along with following social distancing measures, “stay at home” guidelines, expansive testing, and contact tracing [1784,1785]. Indeed, in light of this review, it would also be pertinent to adopt a healthy diet and lifestyle following national guidelines in order to maintain optimal immune health.

Because of the broad public appeal of dietary supplements and nutraceuticals, it is important to evaluate the evidence regarding the use of such products. We will continue to update this review as more findings become available.

9 Social Factors Influencing COVID-19 Exposure and Outcomes

9.1 Social Factors Influencing COVID-19 Outcomes

In addition to understanding the fundamental biology of the SARS-CoV-2 virus and COVID-19, it is critical to consider how the broader environment can influence both COVID-19 outcomes and efforts to develop and implement treatments for the disease. The evidence clearly indicates that social environmental factors are critical determinants of individuals' and communities' risks related to COVID-19. There are distinct components to COVID-19 susceptibility, and an individual's risk can be elevated at one or all stages from exposure to recovery/mortality: an individual may be more likely to be exposed to the virus, more likely to get infected once exposed, more likely to have serious complications once infected, and be less likely to receive adequate care once they are seriously ill. The fact that differences in survival between Black and white patients were no longer significant after controlling for comorbidities and socioeconomic status (type of insurance, neighborhood deprivation score, and hospital where treatment was received) in addition to sex and age [1786] underscores the relevance of social factors to understanding mortality differences between racial and ethnic groups. Moreover, the Black patients were younger and more likely to be female than white patients, yet still had a higher mortality rate without correction for the other variables [1786]. Here, we outline a few systemic reasons that may exacerbate the COVID-19 pandemic in communities of color.

9.2 Factors Observed to be Associated with Susceptibility

As COVID-19 has spread into communities around the globe, it has become clear that the risks associated with this disease are not equally shared by all individuals or all communities. Significant disparities in outcomes have led to interest in the demographic, biomedical, and social factors that influence COVID-19 severity. Untangling the factors influencing COVID-19 susceptibility is a complex undertaking. Among patients who are admitted to the hospital, outcomes have generally been poor, with rates of admission to the intensive care unit (ICU) upwards of 15% in both Wuhan, China and Italy [80,1787,1788]. However, hospitalization rates vary by location [1789]. This

variation may be influenced by demographic (e.g., average age in the area), medical (e.g., the prevalence of comorbid conditions such as diabetes), and social (e.g., income or healthcare availability) factors that vary geographically. Additionally, some of the same factors may influence an individual's probability of exposure to SARS-CoV-2, their risk of developing a more serious case of COVID-19 that would require hospitalization, and their access to medical support. As a result, quantifying or comparing susceptibility among individuals, communities, or other groups requires consideration of a number of complex phenomena that intersect across many disciplines of research. In this section, the term "risk factor" is used to refer to variables that are statistically associated with more severe COVID-19 outcomes. Some are intrinsic characteristics that have been observed to carry an association with variation in outcomes, whereas others may be more functionally linked to the pathophysiology of COVID-19.

9.2.1 Patient Traits Associated with Increased Risk

Two traits that have been consistently associated with more severe COVID-19 outcomes are male sex and advanced age (typically defined as 60 or older, with the greatest risk among those 85 and older [1790]). In the United States, males and older individuals diagnosed with COVID-19 were found to be more likely to require hospitalization [1791,1792]. A retrospective study of hospitalized Chinese patients [81] found that a higher probability of mortality was associated with older age, and world-wide, population age structure has been found to be an important variable for explaining differences in outbreak severity [1793]. The CFR for adults over 80 has been estimated upwards of 14% or even 20% [1794]. Male sex has also been identified as a risk factor for severe COVID-19 outcomes, including death [1795,1796,1797/]. Early reports from China and Europe indicated that even though the case rates were similar across males and females, males were at elevated risk for hospital admission, ICU admission, and death [1796], although data from some US states indicates more cases among females, potentially due to gender representation in care-taking professions [1798]. In older age groups (e.g., age 60 and older), comparable absolute numbers of male and female cases actually suggests a higher rate of occurrence in males, due to increased skew in the sex ratio [1796]. Current estimates based on worldwide data suggest that, compared to females, males may be 30% more likely to be hospitalized, 80% more likely to be admitted to the ICU, and 40% more likely to die as a result of COVID-19 [1797/]. There also may be a compounding effect of advanced age and male sex, with differences time to recovery worst for males over 60 years old relative to female members of their age cohort [1799].

Both of these risk factors can be approached through the lens of biology. The biological basis for greater susceptibility with age is likely linked to the prevalence of extenuating health conditions such as heart failure or diabetes [1794]. Several hypotheses have been proposed to account for differences in severity between males and females. For example, some evidence suggests that female sex hormones may be protective [1796,1798]. ACE2 expression in the kidneys of male mice was observed to be twice as high as that of females, and a regulatory effect of estradiol on ACE2 expression was demonstrated by removing the gonads and then supplementing with estradiol [1798,1800]. Other work in mice has shown an inverse association between mortality due

to SARS-CoV-1 and estradiol, suggesting a protective role for the sex hormone [1798]. Similarly, evidence suggests that similar patterns might be found in other tissues. A preliminary analysis identified higher levels of ACE2 expression in the myocardium of male patients with aortic valve stenosis showed than female patients, although this pattern was not found in controls [1796]. Additionally, research has indicated that females respond to lower doses than males of heart medications that act on the Renin angiotensin aldosterone system (RAAS) pathway, which is shared with ACE2 [1796]. Additionally, several components of the immune response, including the inflammatory response, may differ in intensity and timing between males and females [1798,1800]. This hypothesis is supported by some preliminary evidence showing that female patients who recovered from severe COVID-19 had higher antibody titers than males [1798]. Sex steroids can also bind to immune cell receptors to influence cytokine production [1796]. Additionally, social factors may influence risks related to both age and sex: for example, older adults are more likely to live in care facilities, which have been a source for a large number of outbreaks [1801], and gender roles may also influence exposure and/or susceptibility due to differences in care-taking and/or risky behaviors (e.g., caring for elder relatives and smoking, respectively) [1796] among men and women (however, it should be noted that both transgender men and women are suspected to be at heightened risk [1802].)

9.2.2 Comorbid Health Conditions

A number of pre-existing or comorbid conditions have repeatedly been identified as risk factors for more severe COVID-19 outcomes. Several underlying health conditions were identified at high prevalence among hospitalized patients, including obesity, diabetes, hypertension, lung disease, and cardiovascular disease [1789]. Higher Sequential Organ Failure Assessment (SOFA) scores have been associated with a higher probability of mortality [81], and comorbid conditions such as cardiovascular and lung disease as well as obesity were also associated with an increased risk of hospitalization and death, even when correcting for age and sex [1795]. Diabetes may increase the risk of lengthy hospitalization [1803] or of death [1803,1804]. [1805] and [1806] discuss possible ways in which COVID-19 and diabetes may interact. Obesity also appears to be associated with higher risk of severe outcomes from SARS-CoV-2 [1807,1808]. Obesity is considered an underlying risk factor for other health problems, and the mechanism for its contributions to COVID-19 hospitalization or mortality is not yet clear [1809]. Dementia and cancer were also associated with the risk of death in an analysis of a large number (more than 20,000) COVID-19 patients in the United Kingdom [1795]. It should be noted that comorbid conditions are inextricably tied to age, as conditions tend to be accumulated over time, but that the prevalence of individual comorbidities or of population health overall can vary regionally [1810]. Several comorbidities that are highly prevalent in older adults, such as COPD, hypertension, cardiovascular disease, and diabetes, have been associated with CFRs upwards of 8% compared to an estimate of 1.4% in people without comorbidities [1794,1811]. Therefore, both age and health are important considerations when predicting the impact of COVID-19 on a population [1810]. However, other associations may exist, such as patients with sepsis having higher SOFA scores – in fact, SOFA was developed for the assessment of organ failure in the context of sepsis, and the acronym originally stood for Sepsis-Related

Organ Failure Assessment [1812,1813]. Additionally, certain conditions are likely to be more prevalent under or exacerbated by social conditions, especially poverty, as is discussed further below.

9.2.3 Ancestry

A number of studies have suggested associations between individuals' racial and ethnic backgrounds and their COVID-19 risk. In particular, Black Americans are consistently identified as carrying a higher burden of COVID-19 than white Americans [1791,1792], with differences in the rates of kidney complications from COVID-19 particularly pronounced [87]. Statistics from a number of cities indicate significant discrepancies between the proportion of COVID-19 cases and deaths in Black Americans relative to their representation in the general population [1814]. In addition to Black Americans, disproportionate harm and mortality from COVID-19 has also been noted in Latino/Hispanic Americans and in Native American and Alaskan Native communities, including the Navajo nation [1815]; [1816]; [1817]; <https://www.nytimes.com/2020/04/09/us/coronavirus-navajo-nation.html?searchResultPosition=10>; [1818]; [1819]; [1820]. In Brazil, indigenous communities likewise carry an increased burden of COVID-19 [1821]. In the United Kingdom, nonwhite ethnicity (principally Black or South Asian) was one of several factors found to be associated with a higher risk of death from COVID-19 [1822].

From a genetic standpoint, it is highly unlikely that ancestry itself predisposes individuals to contracting COVID-19 or to experiencing severe COVID-19 outcomes. Examining human genetic diversity indicates variation over a geographic continuum, and that most human genetic variation is associated with the African continent [1823]. African-Americans are also a more genetically diverse group relative to European-Americans, with a large number of rare alleles and a much smaller fraction of common alleles identified in African-Americans [1824]. Therefore, the idea that African ancestry (at the continent level) might convey some sort of genetic risk for severe COVID-19 contrasts with what is known about worldwide human genetic diversity [1825]. The possibility for genetic variants that confer some risk or some protection remains possible, but has not been widely explored, especially at a global level. Research in Beijing of a small number ($n=80$) hospitalized COVID-19 patients revealed an association between severe COVID-19 outcomes and homozygosity for an allele in the interferon-induced transmembrane protein 3 (IFITM3) gene, which was selected as a candidate because it was previously found to be associated with influenza outcomes in Chinese patients [422]. Genetic factors may also play a role in the risk of respiratory failure for COVID-19 [451,1826,1827]. However, genetic variants associated with outcomes within ancestral groups are far less surprising than genetic variants explaining outcomes between groups. Alleles in ACE2 and TMPRSS2 have been identified that vary in frequency among ancestral groups [1828], but whether these variants are associated with COVID-19 susceptibility has not been explored.

Instead, examining patterns of COVID-19 susceptibility on a global scale that suggest that social factors are of primary importance in predicting mortality. Reports from several sub-Saharan African countries have indicated that the effects of the COVID-19 pandemic have been less severe than expected

based on the outbreaks in China and Italy. In Kenya, for example, estimates of national prevalence based on testing blood donors for SARS-CoV-2 antibodies were consistent with 5% of Kenyan adults having recovered from COVID-19 [1829]. This high seroprevalence of antibodies lies in sharp contrast to the low number of COVID-19 fatalities in Kenya, which at the time was 71 out of 2093 known cases [1829]. Likewise, a serosurvey of health care workers in Blantyre City, Malawi reported an adjusted antibody prevalence of 12.3%, suggesting that the virus had been circulating more widely than thought and that the death rate was up eight times lower than models had predicted [1830]. While several possible hypotheses for the apparent reduced impact of COVID-19 on the African continent are being explored, such as young demographics in many places [1831], these reports present a stark contrast to the severity of COVID-19 in Americans and Europeans of African descent. Additionally, ethnic minorities in the United Kingdom also tend to be younger than white British living in the same areas, yet the burden of COVID-19 is still more serious for minorities, especially people of Black Caribbean ancestry, both in absolute numbers and when controlling for age and location [1832]. Furthermore, the groups in the United States and United Kingdom that have been identified as carrying elevated COVID-19 burden, namely Black American, indigenous American, and Black and South Asian British, are quite distinct in their position on the human ancestral tree. What is shared across these groups is instead a history of disenfranchisement under colonialism and ongoing systematic racism. A large analysis of over 11,000 COVID-19 patients hospitalized in 92 hospitals across U.S. states revealed that Black patients were younger, more often female, more likely to be on Medicaid, more likely to have comorbidities, and came from neighborhoods identified as more economically deprived than white patients [1786]. This study reported that when these factors were accounted for, the differences in mortality between Black and white patients were no longer significant. Thus, the current evidence suggests that the apparent correlations between ancestry and health outcomes must be examined in the appropriate social context.

9.3 Environmental Influences on Susceptibility

9.3.1 Exposure to COVID-19

Social distancing has emerged as one of the main social policies used to manage the COVID-19 epidemic in many countries. Many governments issued stay-at-home orders, especially in the initial months of the crisis. However, data clearly indicates that these orders impacted different socioeconomic groups differently. In U.S. counties with and without stay-at-home orders, smartphone tracking indicated a significant decrease in the general population's mobility in April relative to February through March of 2020 (-52.3% and -60.8%, respectively) [1833]. A linear relationship was observed between counties' reduction in mobility and their wealth and health, as measured by access to health care, food security, income, space, and other factors [1833]. Counties with greater reductions in mobility were also found to have much lower child poverty and household crowding and to be more racially segregated, and to have fewer youth and more elderly residents [1833]. Similar associations between wealth and decreased mobility were observed in cellphone GPS data from Colombia, Indonesia, and Mexico

collected between January and May 2020 [1834], as well as in a very large data set from several US cities [1835]. These disparities in mobility are likely to be related to the role that essential workers have played during the pandemic. Essential workers are disproportionately likely to be female, people of color, immigrants, and to have an income below 200% of the poverty line [1836]. Black Americans in particular are over-represented among front-line workers and in professions where social distancing is infeasible [1837]. Health care work in particular presents an increased risk of exposure to SARS-CoV-2 [1837,1838,1839,1840,1841]. In the United Kingdom, (South) Asians are more likely than their white counterparts to be medical professionals [1832/], although BAME medical professionals are still disproportionately represented in the proportion of National Health Service staff deaths [1842]. Similar trends have been reported for nurses, especially nurses of color, in the United States [1843/-/files/graphics/0920_Covid19_SinsOfOmission_Data_Report.pdf]. Furthermore, beyond the risks associated with work itself, use of public transportation may also impact COVID-19 risk [1844]. The socioeconomic and racial/ethnic gaps in who is working on the front lines of the pandemic make it clear that socioeconomic privilege is likely to decrease the probability of exposure to SARS-CoV-2.

Increased risk of exposure can also arise outside the workplace. Nursing homes and skilled nursing facilities received attention early on as high-risk locations for COVID-19 outbreaks [1845]. Prisons and detention centers also confer a high risk of exposure or infection [1846,1847]. Populations in care facilities are largely older adults, and in the United States, incarcerated people are more likely to be male and persons of color, especially Black [1848]. Additionally, multi-generational households are less common among non-Hispanic white Americans than people of other racial and ethnic backgrounds [1849], increasing the risk of exposure for more susceptible family members. Analysis suggests that household crowding may also be associated with increased risk of COVID-19 exposure [1833], and household crowding is associated with poverty [1850]. Forms of economic insecurity like housing insecurity, which is associated with poverty and more pronounced in communities subjected to racism [1851,1852], would be likely to increase household crowding and other possible sources of exposure. As a result, facets of systemic inequality such as mass incarceration of Black Americans and poverty are likely to increase the risk of exposure outside of the workplace.

9.3.2 Severity of COVID-19 Following Exposure

Following exposure to SARS-CoV-2, the likelihood that an individual develops COVID-19 and the severity of the disease presentation can be influenced by a number of social factors. As discussed above, a number of patient characteristics are associated with the likelihood of severe COVID-19 symptoms. In some cases, these trends run counter to those expected given rates of exposure: for example, although women are more likely to be exposed, men are more likely to be diagnosed with, hospitalized from, or die from COVID-19 [1798]. In the case of comorbid conditions and racial/ethnic demographics, however, social factors are highly likely to modulate or at least influence the apparent association between these traits and the

increased risk from COVID-19. In particular, the comorbidities and racial/ethnic correlates of severe COVID-19 outcomes suggest that poverty confers additional risk for COVID-19.

In order to explore the relationship between poverty and COVID-19 outcomes, it is necessary to consider how poverty impacts biology. In particular, we focus on the United States and the United Kingdom. Comorbidities that increase risk for COVID-19, including obesity, type II diabetes, hypertension, and cardiovascular disease, are known to be intercorrelated [1853]. Metabolic conditions related to heightened inflammation, like obesity, type II diabetes, and hypertension, are more strongly associated with negative COVID-19 outcomes than other comorbid conditions, such as chronic heart disease [1854]. As discussed above, dysregulated inflammation characteristic of cytokine release syndrome is one of the greatest concerns for COVID-19-related death. Therefore, it is possible that chronic inflammation characteristic of these metabolic conditions predisposes patients to COVID-19-related death [1854]. The association between these diseases and severe COVID-19 outcomes is a concern from a health equity perspective because poverty exposes people to “obesogenic” conditions [1855] and is therefore unsurprisingly associated with higher incidence of obesity and associated disorders [1856]. Furthermore, cell phone GPS data suggests that lower socioeconomic status may also be associated with decreased access to healthy food choices during the COVID-19 pandemic [1857,1858], suggesting that health-related risk factors for COVID-19 may be exacerbated as the pandemic continues [1859]. Chronic inflammation is a known outcome of chronic stress (e.g., [1860,1861,1862,1863]). Therefore, the chronic stress of poverty is likely to influence health broadly (as summarized in [1864]) and especially during the stress of the ongoing pandemic.

A preprint [1865] provided observational evidence that geographical areas in the United States that suffer from worse air pollution by fine particulate matter have also suffered more COVID-19 deaths per capita, after adjusting for demographic covariates. Although lack of individual-level exposure data and the impossibility of randomization make it difficult to elucidate the exact causal mechanism, this finding would be consistent with similar findings for all-cause mortality (e.g., [1866]). Exposure to air pollution is associated with both poverty (e.g., [1867]) and chronic inflammation [1868]. Other outcomes of environmental racism, such as the proximity of abandoned uranium mines to Navajo land, can also cause respiratory illnesses and other health issues [1820]. Similarly, preliminary findings indicate that nutritional status (e.g., vitamin D deficiency [1681]) may be associated with COVID-19 outcomes, and reduced access to grocery stores and fresh food often co-occurs with environmental racism [1820,1869]. Taken together, the evidence suggests that low-income workers who face greater exposure to SARS-CoV-2 due to their home or work conditions are also more likely to face environmental and social stressors associated with increased inflammation, and therefore with increased risk from COVID-19. In particular, structural racism can play an important role on disease severity after SARS-CoV-2 exposure, due to consequences of racism which include an increased likelihood of poverty and its associated food and housing instability. COVID-19 can thus be considered a “syndemic”, or a synergistic interaction between several epidemics [1870]. As a result, it is not surprising that people from minoritized backgrounds

and/or with certain pre-existing conditions are more likely to suffer severe effects of COVID-19, but these “risk factors” are likely to be causally linked to poverty [1871].

9.3.3 Access to Treatment

Finally, COVID-19 outcomes can be influenced by access to healthcare. Receiving care for COVID-19 can, but does not always, include receiving a positive test for the SARS-CoV-2 virus. For example, it is common to see treatment guidelines for suspected cases regardless of whether the presence of SARS-CoV-2 has been confirmed (e.g., [1872]). Whether and where a patient is diagnosed can depend on their access to testing, which can vary both between and within countries. In the United States, it is not always clear whether an individual will have access to free testing [1873,1874]. The concern has been raised that more economic privilege is likely to correspond to increased access to testing, at least within the United States [1875]. This is supported by the fact that African Americans seem to be more likely to be diagnosed in the hospital, while individuals from other groups were more likely to have been diagnosed in ambulatory settings in the community [1791]. Any delays in treatment are a cause for concern [1875], which could potentially be increased by an inability to acquire testing because in the United States, insurance coverage for care received can depend on a positive test [1876].

Another important question is whether patients with moderate to severe cases are able to access hospital facilities and treatments, to the extent that they have been identified. Early findings from China as of February 2020 suggested the COVID-19 mortality rate to be much lower in the most developed regions of the country [1877], although reported mortality is generally an estimate of CFR, which is dependent on rates of testing. Efforts to make treatment accessible for all confirmed and suspected cases of COVID-19 in China are credited with expanding care to people with fewer economic resources [1878]. In the United States, access to healthcare varies widely, with certain sectors of the workforce less likely to have health insurance; many essential workers in transportation, food service, and other frontline fields are among those likely to be uninsured or underinsured [1875]. As of 2018, Hispanic Americans of all races were much less likely to have health insurance than people from non-Hispanic backgrounds [1879]. Therefore, access to diagnostics and care prior to the development of severe COVID-19 is likely to vary depending on socioeconomic and social factors, many of which overlap with the risks of exposure and of developing more severe COVID-19 symptoms. This discrepancy ties into concerns about broad infrastructural challenges imposed by COVID-19. A major concern in many countries has been the saturation of healthcare systems due to the volume of COVID-19 hospitalizations (e.g., [318]). Similarly, there have been shortages of supplies such as ventilators that are critical to the survival of many COVID-19 patients, leading to extensive ethical discussions about how to allocate limited resources among patients [1880,1881,1882,1883]. Although it is generally considered unethical to consider demographic factors such as age, sex, race, or ethnicity while making such decisions, and ideally this information would not be shared with triage teams tasked with allocating limited resources among patients [1884], there are substantial concerns about implicit and explicit biases against older adults [1885], premature

infants [1886], and people with disabilities or comorbidities [1884,1887,1888]. Because of the greater burden of chronic disease in populations subjected to systemic racism, algorithms intended to be blind to race and ethnicity could, in fact, reinforce systemic inequalities caused by structural racism [1889,1890,1891]. Because of this inequality, it has been argued that groups facing health disparities should be prioritized by these algorithms [1892]. This approach would carry its own ethical concerns, including the fact that many resources that need to be distributed do not have well-established risks and benefits [1892].

As the pandemic has progressed, it has become clear that ICU beds and ventilators are not the only limited resources that need to be allocated, and, in fact, the survival rate for patients who receive mechanical ventilation is lower than these discussions would suggest [1893]. Allocation of interventions that may reduce suffering, including palliative care, has become critically important [1893,1894]. The ambiguities surrounding the risks and benefits associated with therapeutics that have been approved under emergency use authorizations also present ethical concerns related to the distribution of resources [1892]. For example, remdesivir, discussed above, is currently available for the treatment of COVID-19 under compassionate use guidelines and through expanded access programs, and in many cases has been donated to hospitals by Gilead [1895,1896]. Regulations guiding the distribution of drugs in situations like these typically do not address how to determine which patients receive them [1896]. Prioritizing marginalized groups for treatment with a drug like remdesivir would also be unethical because it would entail disproportionately exposing these groups to a therapeutic that may or not be beneficial [1892]. On the other hand, given that the drug is one of the most promising treatments available for many patients, using a framework that tacitly feeds into structural biases would also be unethical. At present, the report prepared for the Director of the CDC by Ethics Subcommittee of the CDC fails to address the complexity of this ethical question given the state of structural racism in the United States, instead stating that "prioritizing individuals according to their chances for short-term survival also avoids ethically irrelevant considerations, such as race or socioeconomic status" [1897]. In many cases, experimental therapeutics are made available only through participation in clinical trials [1898]. However, given the history of medical trials abusing minority communities, especially Black Americans, there is a history of unequal representation in clinical trial enrollment [1898]. As a result, the standard practice of requiring enrollment in a clinical trial in order to receive experimental treatment may also reinforce patterns established by systemic racism.

9.3.4 Access to and Representation in Clinical Trials

Experimental treatments are often made available to patients primarily or even exclusively through clinical trials. The advantage of this approach is that clinical trials are designed to collect rigorous data about the effects of a treatment on patients. The disadvantage is that access to clinical trials is not equal among all people who suffer from a disease. Two important considerations that can impact an individual's access to clinical trials are geography and social perceptions of clinical trials. For the first, the geographic distribution of trial recruitment efforts are typically bounded and

can vary widely among different locations, and for the second, the social context of medical interactions can impact strategies for and the success of outreach to different communities. Differential access to clinical trials raises concerns because it introduces biases that can influence scientific and medical research on therapeutics and prophylactics broadly. Concerns about bias in clinical trials need to address both trial recruitment and operation. In the present crisis, such biases are particularly salient because COVID-19 is a disease of global concern. Treatment is needed by people all over the world, and clinical research that characterizes treatment outcomes in a variety of populations is critically important.

Global representation in clinical trials is important to ensuring that experimental treatments are available equally to COVID-19 patients who may need them. The advantage to a patient of participation in a clinical trial is that they may receive an experimental treatment they would not have been able to access otherwise. The potential downsides of participation include that the efficacy and side effects of such treatments are often poorly characterized and that patients who enroll in clinical trials will in some cases run the risk of being assigned to a placebo condition where they do not receive the treatment but miss out on opportunities to receive other treatments. The benefits and burdens of clinical trials therefore need to be weighed carefully to ensure that they don't reinforce existing health disparities. The WHO Director-General Tedros Adhanom Ghebreyesus stated his condemnation of utilizing low and middle income countries as test subjects for clinical trials, yet having highly developed countries as the majority of clinical trial representation is also not the answer [1899]. Figure 13 showcases two choropleths detailing COVID-19 clinical trial recruitment by country. China, the United States, and France are among the countries with the most clinical trial recruiting for trials with single-country enrollment. Many countries have little to no clinical trial recruiting, with the continents of Africa and South America much less represented than Asia, Europe, and North America. Trials that recruit across multiple countries do appear to broaden geographic representation, but these trials seem to be heavily dominated by the United States and European Union.

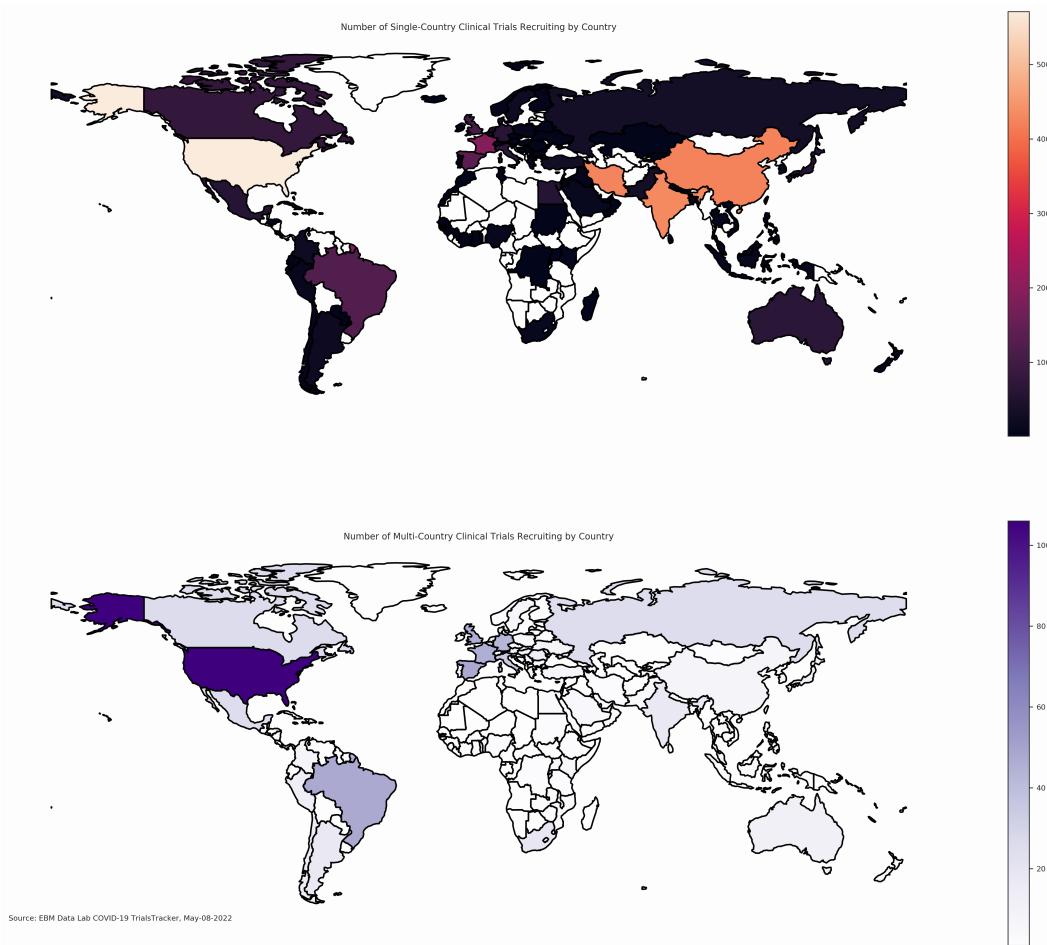


Figure 13: Geographic distribution of COVID-19 clinical trials. The density of clinical trials is reported at the country level. As of December 31, 2020, there are 6,987 trials in the University of Oxford Evidence-Based Medicine Data Lab's COVID-19 TrialsTracker [717], of which 3,962 are interventional. The top figure demonstrates the density of interventional trials recruiting only from a singular country, while the bottom shows the distribution of recruitment for interventional trials that involve more than one country.

A few different concerns arise from this skewed geographic representation in clinical trial recruitment. First, treatments such as remdesivir that are promising but primarily available to clinical trial participants are unlikely to be accessible by people in many countries. Second, it raises the concern that the findings of clinical trials will be based on participants from many of the wealthiest countries, which may lead to ambiguity in whether the findings can be extrapolated to COVID-19 patients elsewhere. Especially with the global nature of COVID-19, equitable access to therapeutics and vaccines has been a concern at the forefront of many discussions about policy (e.g., [1900]), yet data like that shown in Figure 13 demonstrates that accessibility is likely to be a significant issue. Another concern with the heterogeneous international distribution of clinical trials is that the governments of countries leading these clinical trials might prioritize their own populations once vaccines are developed, causing unequal health outcomes [1901]. Additionally, even within a single state in the United States (Maryland), geography was found to influence the likelihood of being recruited into or enrolled in a clinical trial, with patients in under-served rural areas less likely to enroll [1902]. Thus, geography both on the global and local levels may influence when treatments and vaccines are available and who is able to access them. Efforts such as the African Union's efforts to coordinate and promote vaccine development [1903] are therefore critical to promoting equity in the COVID-19 response.

Even when patients are located within the geographic recruitment area of clinical trials, however, there can still be demographic inequalities in enrollment. When efforts are made to ensure equal opportunity to participate in clinical trials, there is no significant difference in participation among racial/ethnic groups [1904]. However, within the United States, real clinical trial recruitment numbers have indicated for many years that racial minorities, especially African-Americans, tend to be under-represented (e.g., [1905,1906,1907,1908]). This trend is especially concerning given the disproportionate impact of COVID-19 on African-Americans. Early evidence suggests that the proportion of Black, Latinx, and Native American participants in clinical trials for drugs such as remdesivir is much lower than the representation of these groups among COVID-19 patients [1909].

One proposed explanation for differences among racial and ethnic groups in clinical trial enrollment refers to different experiences in healthcare settings. While some plausible reasons for the disparity in communication between physicians and patients could be a lack of awareness and education, mistrust in healthcare professionals, and a lack of health insurance [1904], a major concern is that patients from certain racial and ethnic groups are marginalized even while seeking healthcare. In the United States, many patients experience “othering” from physicians and other medical professionals due to their race or other external characteristics such as gender (e.g., [1910]). Many studies have sought to characterize implicit biases in healthcare providers and whether they affect their perceptions or treatment of patients. A systematic review that examined 37 such studies reported that most (31) identified racial and/or ethnic biases in healthcare providers in many different roles, although the evidence about whether these biases translated to different attitudes towards patients was mixed [1911], with similar findings reported by a second systematic review [1912]. However, data about real-world patient outcomes are very limited, with most studies relying on clinical vignette-based exercises [1911], and other analyses suggest that physician implicit bias could impact the patient’s perception of the negativity/positivity of the interaction regardless of the physician’s explicit behavior towards the patient [1913]. Because racism is a common factor in both, negative patient experiences with medical professionals are likely to compound other issues of systemic inequality, such as a lack of access to adequate care, a lack of insurance, or increased exposure to SARS-CoV-2 [1914]. Furthermore, the experience of being othered is not only expected to impact patients’ trust in and comfort with their provider, but also may directly impact whether or not the patient is offered the opportunity to participate in a clinical trial at all. Some studies suggest communication between physicians and patients impacts whether or not a physician offers a patient participation in a clinical trial. For example, researchers utilized a linguistic analysis to assess mean word count of phrases related to clinical trial enrollment, such as voluntary participation, clinical trial, etc. [1904]. The data indicated that the mean word count of the entire visit was 1.5 times more for white patients in comparison to Black patients. In addition, the greatest disparity between white and Black patients’ experience was the discussion of risks, with over 2 times as many risk-related words spoken with white patients than Black patients [1904]. The trends observed for other clinical trials raise the concern that COVID-19 clinical trial information may not be discussed as thoroughly or as often with Black patients compared to white patients.

These discrepancies are especially concerning given that many COVID-19 treatments are being or are considered being made available to patients prior to FDA approval through Emergency Use Authorizations. In the past, African-Americans have been over-represented relative to national demographics in use of the FDA's Exception From Informed Consent (EFIC) pathway [1915]. Through this pathway, people who are incapacitated can receive an experimental treatment even if they are not able to consent and there is not sufficient time to seek approval from an authorized representative. This pathway presents concerns, however, when it is considered in the context of a long history of systematic abuses in medical experimentation where informed consent was not obtained from people of color, such as the Tuskegee syphilis experiments [1916]. While the goal of EFIC approval is to provide treatment to patients who urgently need it, the combination of the ongoing legacy of racism in medicine renders this trend concerning. With COVID-19, efforts to prioritize people who suffer from systemic racism are often designed with the goal of righting some of these inequalities (e.g., [1917]), but particular attention to informed consent will be imperative in ensuring these trials are ethical given that the benefits and risks of emerging treatments are still poorly characterized. Making a substantial effort to run inclusive clinical trials is also important because of the possibility that racism could impact how a patient responds to a treatment. For example, as discussed above, dexamethasone has been identified as a promising treatment for patients experiencing cytokine release syndrome, but the mechanism of action is tied to the stress response. A study from 2005 reported that Black asthma patients showed reduced responsiveness to dexamethasone in comparison to white patients and suggested Black patients might therefore require higher doses of the drug [1918]. In the context of chronic stress caused by systemic racism, this result is not surprising: chronic stress is associated with dysregulated production of glucocorticoids [1919] and glucocorticoid receptor resistance [1920]. However, it underscores the critical need for treatment guidelines to take into account differences in life experience, which would be facilitated by the recruitment of patients from a wide range of backgrounds. Attention to the social aspects of clinical trial enrollment must therefore be an essential component of the medical research community's response to COVID-19.

9.4 Conclusions and Future Directions

As the COVID-19 pandemic evolves, the scientific community's response will be critical for identifying potential pharmacological and biotechnological developments that may aid in combating the virus and the disease it causes. However, this global crisis highlights the importance of mounting a response based on collaboration among a wide variety of disciplines. Understanding the basic science of the virus and its pathogenesis is imperative for identifying and envisioning possible diagnostic and therapeutic approaches; understanding how social factors can influence outcomes and shape implementation of a response is critical to disseminating any scientific advancements. Summarizing such a complex and ever-changing topic presents a number of challenges. This review represents the effort of over 50 contributors to distill and interpret the available information. However, this text represents a dynamic and evolving document, and we welcome continued contributions from all researchers who have insights into how these topics intersect. A multidisciplinary perspective is critical to

understanding this evolving crisis, and in this review we seek to use open science tools to coordinate a response among a variety of researchers. We intend to publish additional updates as the situation evolves.

10 An Open-Publishing Response to the COVID-19 Infodemic

10.1 ABSTRACT

The COVID-19 pandemic catalyzed the rapid dissemination of papers and preprints investigating the disease and its associated virus, SARS-CoV-2. The multifaceted nature of COVID-19 demands a multidisciplinary approach, but the urgency of the crisis combined with the need for social distancing measures present unique challenges to collaborative science. We applied a massive online open publishing approach to this problem using Manubot. Through GitHub, collaborators summarized and critiqued COVID-19 literature, creating a review manuscript. Manubot automatically compiled citation information for referenced preprints, journal publications, websites, and clinical trials. Continuous integration workflows retrieved up-to-date data from online sources nightly, regenerating some of the manuscript's figures and statistics. Manubot rendered the manuscript into PDF, HTML, LaTeX, and DOCX outputs, immediately updating the version available online upon the integration of new content. Through this effort, we organized over 50 scientists from a range of backgrounds who evaluated over 1,500 sources and developed seven literature reviews. While many efforts from the computational community have focused on mining COVID-19 literature, our project illustrates the power of open publishing to organize both technical and non-technical scientists to aggregate and disseminate information in response to an evolving crisis.

KEYWORDS

COVID-19, living document, open publishing, open-source, data integration, manubot

10.2 INTRODUCTION

Coronavirus Disease 2019 (COVID-19) caused a worldwide public health crisis that has reshaped many aspects of society. The scientific community has, in turn, devoted significant attention and resources towards COVID-19 and the associated virus, SARS-CoV-2, resulting in the release of data and publications at a rate and scale never previously seen for a single topic. Over 20,000 articles about COVID-19 were released in the first four months of the pandemic [1921], causing an “infodemic” [1921,1922]. The COVID-19 Open Research Dataset (CORD-19) [1923], which was developed in part with the goal of training machine learning algorithms on COVID-19-related text, illustrates the growth of related scholarly literature (Figure 14). This resource was developed by querying several sources for terms related to SARS-CoV-2 and COVID-19, as well as the coronaviruses SARS-CoV-1 and MERS-CoV and

their associated diseases [1923]. CORD-19 contained 1022888 manuscripts as of 2022-04-28. Additional curation by CoronaCentral [1924] has produced, at present, a set of over 180,000 publications particularly relevant to COVID-19 and closely related viruses. Despite many advances in understanding the virus and the disease, there are also downsides to the availability of so much information. “Excessive publication” has been recognized as a concern for over forty years [1925] and has been discussed with respect to the COVID-19 literature [970]. Any effort to synthesize, summarize, and contextualize COVID-19 research will face a vast corpus of potentially relevant material.

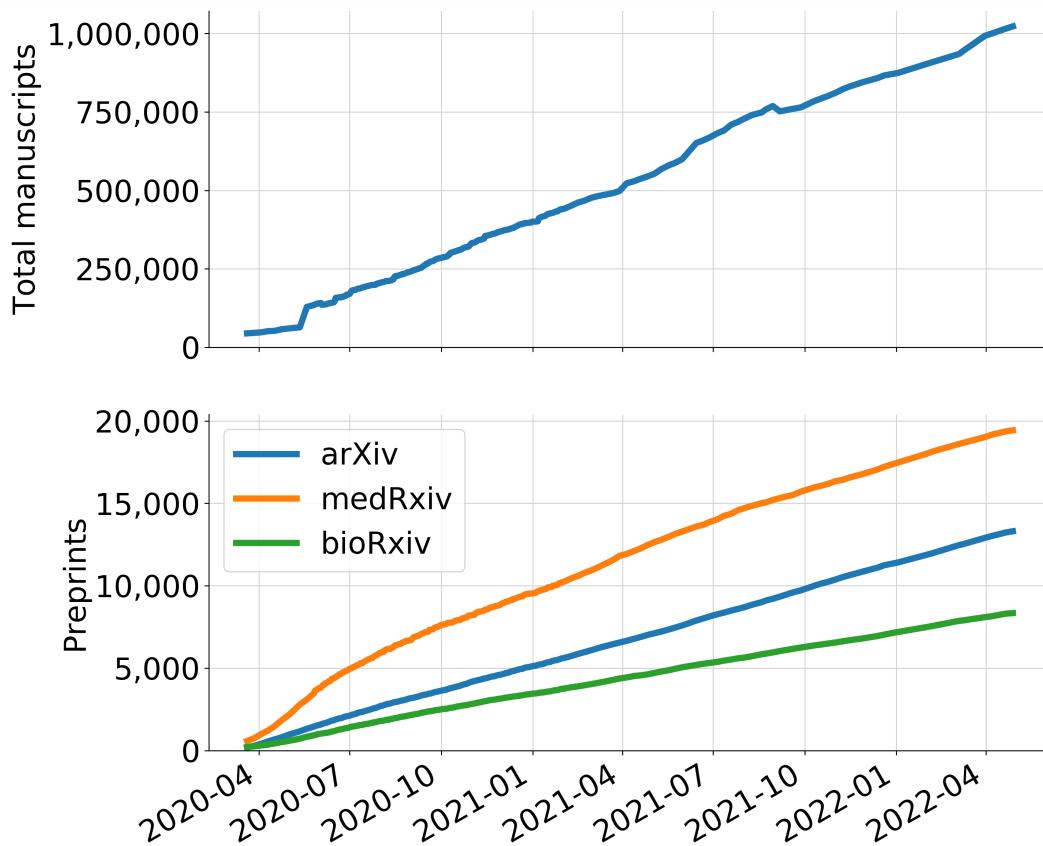


Figure 14: Growth of the CORD-19 dataset. The number of articles has proliferated, with both traditional and preprint manuscripts in the corpus. The first release (March 16, 2020) contained 28,000 documents [1923]. As of 2022-04-28, this had increased to 1022888 articles. Of these, 41079 are preprints from *arXiv*, *medRxiv*, and *bioRxiv*.

Information was released rapidly by both traditional publishers and preprint servers, and many papers faced subsequent scrutiny. The number of COVID-19 papers retracted may be higher, and potentially much higher, than is typical, although a thorough investigation of this question requires more time to elapse [1926,1927]. Many papers and preprints are also associated with corrections or expressions of concern¹ [1927]. Preprints are released prior to peer review, but some traditional publishing venues have fast-tracked COVID-19 papers through peer review, leading to questions about whether they are held to typical standards [1928]. Therefore, evaluating the COVID-19 literature requires not only digesting available information but also monitoring subsequent changes.

Because of the fast-moving nature of the topic, many efforts to summarize and synthesize the COVID-19 literature have been undertaken. These efforts include newsletters² [1929], web portals³ [1930] or the now-defunct <http://covidpreprints.com>⁴, comments on preprint servers⁵ [1931], and even

a journal⁶. However, the explosive rate of publication presents challenges for such efforts, many of which are no longer active. Similarly, many literature reviews have been written on the available COVID-19 literature [1932,1933,1934,1935,1936], but static reviews quickly become outdated as new research is released or existing research is retracted or superseded. One example is a review of topics in COVID-19 research including vaccine development [1936]. This review was published on July 10, 2020, four days before Moderna released the surprisingly promising results of their phase 1 trial [1102] that changed expectations surrounding vaccines. Therefore, the COVID-19 publishing climate presented a challenge where curation of the literature by a diverse group of experts in a format that could respond quickly to high-volume, high-velocity information was desirable.

We therefore sought to develop a platform for scientific discussion and collaboration around COVID-19 by adapting open publishing infrastructure to accommodate the scale of COVID-19 publishing. Recent advances in open publishing have created an infrastructure that facilitates distributed, version-controlled collaboration on manuscripts [1937]. Manubot [1937] is a collaborative framework developed to adapt open-source software development techniques and version control for manuscript writing. With Manubot, manuscripts are managed and maintained using GitHub, a popular, online version control interface. We selected Manubot because it offers several advantages over comparable collaborative writing platforms such as Authorea, Overleaf, Google Docs, Word Online, or wikis [1937]. Citation-by-identifier ensures consistent reference metadata standards that would be difficult to maintain manually in a manuscript with dozens of authors and over 1,500 citations. Manubot's pull request-based contribution model balances the goals of making the project open to everyone and maintaining scientific accuracy. All contributions are reviewed, discussed, and formally approved on GitHub before text updates appear in the public-facing manuscript⁷. Continuous integration (CI) seamlessly combines author-produced text and figures with automatically generated and updated statistics and figures derived from external data sources and the manuscript's own content. In addition, the authors who initially launched this project included Manubot developers who had prior successes using Manubot for massively open and traditional manuscripts.

Collaboration via massively open online papers has been identified as a strategy for promoting inclusion and interdisciplinary thought [1938]. However, the Manubot workflow can be intimidating to contributors who are not well-versed in git [1938]. The synthesis and discussion of the emerging literature by biomedical scientists and clinicians is imperative to a robust interpretation of COVID-19 research. Such efforts in biology often rely on What You See Is What You Get tools such as Google Docs, despite the significant limitations of these platforms in the face of excessive publication. We recognized that the problem of synthesizing the COVID-19 literature lent itself well to the Manubot platform, but that the potential technical expertise required to work with Manubot presented a barrier to domain experts.

Here, we describe the adaptation of Manubot to facilitate collaboration in the extreme case of the COVID-19 infodemic, with the objective of developing a centralized platform for summarizing and synthesizing a massive amount of preprints, news stories, journal publications, and data. Unlike prior

collaborations built on Manubot, most contributors to the COVID-19 collaborative literature review came from biology or medicine. The members of the COVID-19 Review Consortium consolidated information about the virus in the context of related viruses and to synthesize rapidly emerging literature. Manubot provided the infrastructure to manage contributions from the community and create a living, scholarly document integrating data from multiple sources. Its back-end allowed biomedical scientists to sort and distill informative content out of the overwhelming flood of information [1939] in order to provide a resource that would be useful to the broader scientific community. This case study demonstrates the value of open collaborative writing tools such as Manubot to emerging challenges. Because it is open source software, we were able to adapt and customize Manubot to flexibly meet the needs of COVID-19 review. Recording the evolution of information over time and assembling a resource that auto-updated in response to the evolving crisis revealed the particular value that Manubot holds for managing rapid changes in scientific thought.

10.3 METHODS

10.3.1 Contributor Recruitment and Roles

First, it was necessary to establish Manubot as a platform accessible to researchers with limited experience working version control, given that this is not typically emphasized in biology and medicine [1940,1941,1942]. Contributors were recruited primarily by word of mouth and on Twitter, and we also collaborated with existing efforts to train early-career researchers. We invited potential collaborators to contribute a short introduction on a GitHub issue in order to collect information about participants and provide an introduction to working with GitHub issues. Interested participants were encouraged to contribute in several ways. One option was to catalog articles of interest as issues. We developed a standardized set of questions for contributors to consider when evaluating an article following a framework often used for assessing medical literature. This approach emphasizes examining the methods used, assignment (whether the study was observational or randomized), assessment, results, interpretation, and how well the study extrapolates [1943]. Contributors were also invited to contribute or edit text using GitHub's pull request system. These contributions were not strictly defined and could range from minor corrections to punctuation and grammar to large-scale additions of text. Finally, a small number of contributors (the authors of this paper) contributed technical expertise, either through the development of standardized approaches to the evaluation of papers based on the MAARIE Framework [1944], the writing of code to generate manuscript figures, or the addition of features to Manubot. All of these additions were also submitted as pull requests, either to the COVID-19 review repository or to an external repository, as appropriate.

Each pull request was reviewed and approved by at least one other contributor before being merged into the main branch. We tagged potential reviewers based on the introductions they had contributed in order to encourage participation. Authorship was determined based on the Contributor Roles Taxonomy⁸. Due to the permeability of ideas among

different sections, contributors to a specific manuscript were recognized with masthead authorship, while all contributors to the project were recognized with consortium authorship on all papers. Emphasizing the use of issues and pull requests was designed to encourage authors with and without git experience to discuss papers and provide feedback (both formal and informal) on proposed text additions or changes. We also used the Gitter chat platform⁹ to promote informal questions and sharing of information among collaborators.

10.3.2 Utilization and Expansion of Manubot

Applying Manubot's existing capabilities allowed us to confront several challenges common in large-scale collaborations, such as maintaining a record of contributions that allowed us to allocate credit appropriately or to contact the original author if questions arose. Additionally, an up-to-date version of the content was available at all times online in HTML¹⁰ or PDF format¹¹. This approach also allowed us to minimize the demand on authors to curate and sync bibliographic resources. Manubot provides the functionality to create a bibliography using digital object identifiers (DOIs), website URLs, or other identifiers such as PubMed identifiers and arXiv IDs. The author can insert a citation in-line using a format such as

`[@doi:10.1371/journal.pcbi.1007128]`. Manubot then obtains reference metadata, exports the citations as Citation Style Language JSON Data Items, and renders the bibliographic information needed to generate the references section [1937]. This approach allows multiple authors to work on a piece of text without needing to make manual adjustments to the reference lists.

Due to the needs of this project, several new features were implemented in Manubot. Because of the ever-evolving nature of the COVID-19 crisis, figures and statistics in the text quickly became outdated. To address this concern, Manubot and GitHub's CI features were used to create figures that integrated online data sources and to dynamically update information, such as the current number of active COVID-19 clinical trials [3], within the text of the manuscripts (Figure 15). GitHub Actions runs a nightly workflow to update these external data and regenerate the statistics and figures for the manuscript. The workflow uses the GitHub API to detect and save the latest commit of the external data sources that are GitHub repositories¹². It then downloads versioned data from that snapshot of the external repositories and runs bash and Python scripts to calculate the desired statistics and produce the summary figures using Matplotlib [1945]. The statistics are stored in JSON files that are accessed by Manubot to populate the values of placeholder template variables dynamically every time the manuscript is built. For instance, the template variable `{{ebm_trials_results}}` in the manuscript is replaced by the actual number of clinical trials with results, 98. The template variables also include versioned URLs to the dynamically updated figures. The JSON files and figures are stored in the `external-resources` branch of the GitHub repository, providing versioned storage. The GitHub Actions workflow automatically adds and commits the new JSON files and figures to the `external-resources` branch every time it runs, and Manubot uses the latest version of these resources when it builds the manuscript. The GitHub Actions workflow file is available online¹³, as are the scripts¹⁴. The Python package versions are also available¹⁵.

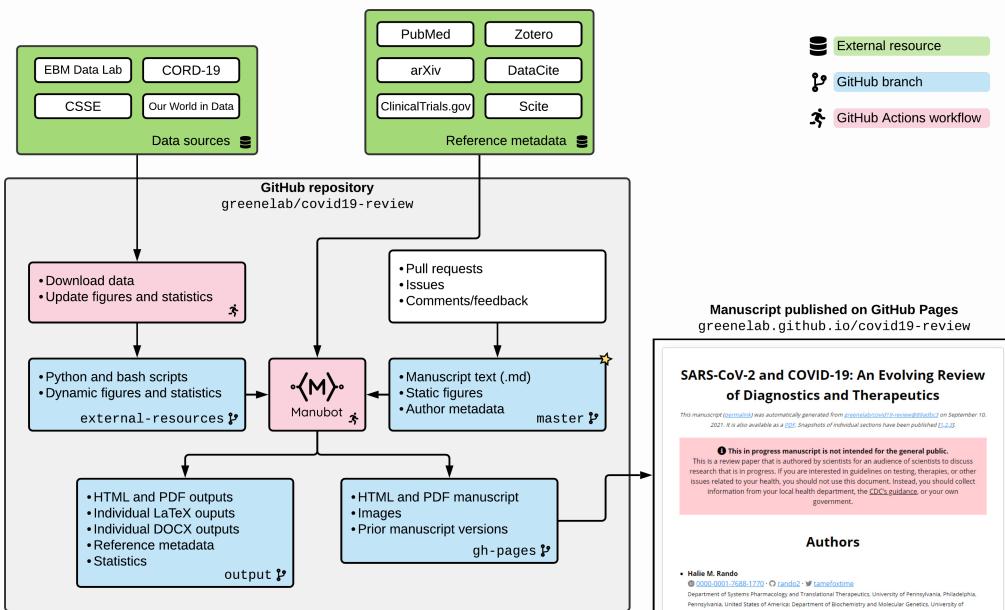


Figure 15: COVID-19 review GitHub repository organization and workflows. Manubot uses CI to combine author-contributed content with automatically updated information from outside sources. A nightly workflow updates figures and statistics derived from external resources. Authors write text and add figures to the `master` branch (starred) via GitHub pull requests. Manubot generates updated manuscript outputs for each new git commit, integrating the static text and figures with the dynamic statistics and figures and automatically-extracted citation information. GitHub Pages hosts the latest HTML and PDF versions of the manuscript along with permanent links to prior versions.

Another issue identified was the need for standardized citation to clinical trials. Other researchers identified the same need¹⁶. Trials that are registered with clinicaltrials.gov receive a unique clinical trial identifier, or "NCT ID." Because clinical trials are registered long before results are published, referencing clinical trial identifiers was a priority. Manubot uses the Zotero translation server¹⁷ to extract citation metadata for some types of citations. However, Zotero did not support clinical trial identifiers and could not extract relevant metadata from their URLs. In order to pull clinical trial metadata associated into Manubot, we added Zotero support for these identifiers. To achieve this, we query clinicaltrials.gov to retrieve XML metadata associated with each identifier using JavaScript¹⁸. This extension enables citing a trial as `@clinicaltrials:NCT04280705` instead of the URL. Then, when Manubot requests clinical trial metadata from the Zotero translation server, the response includes the trial sponsors, responsible investigators, title, and summary. Manubot now supports directly citing hundreds of registered Compact Uniform Resource Identifiers¹⁹, beyond just the `clinicaltrials` identifier.

Because of the large number of citations used in this manuscript and the fast-moving nature of COVID-19 research, keeping track of retractions, corrections, and notices of concern also became a challenge. We implemented a new Manubot plugin to support "smart citations" in the HTML build of manuscripts. The plugin uses the scite [1946] service to display a badge below any citation with a DOI. The badge contains a set of icons and numbers that indicate how many times that source has been mentioned, supported, or disputed and whether there have been any important editorial notices. We were thus able to identify references that needed to be reevaluated by an expert. This addition was invaluable given the nature of

the project, where we were disseminating rapidly evolving information of great consequence from over 1,500 different sources. The badges also allow readers to ascertain a rough approximation of the reliability of cited sources at a glance.

Because most collaborators were writing and editing text through the GitHub website rather than in a local text editor, we also needed to add spell-checking functionalities to Manubot. We integrated an existing Pandoc²⁰ spell-check extension with AppVeyor CI to automatically post spelling errors as comments in a GitHub pull request. The comment reported both unique misspelled tokens and all locations where the token was detected. Project maintainers managed a custom dictionary to allow over 1,500 scientific and technical terms that were not common English words. Spell-checking also helped standardize the writing style across dozens of authors by detecting features such as British versus American English spellings. The actual spell-checking was implemented using GNU Aspell²¹ and the Pandoc spellcheck filter²². The filter enables checking only the manuscript text, ignoring URLs and formatting.

Manubot can render a manuscript in several formats that serve different purposes. Prior to this project, Manubot could use Pandoc to convert the markdown-formatted manuscript to HTML, PDF, and DOCX formats. We expanded this functionality to export individual sections of the manuscript as separate DOCX files while still rendering the complete manuscript in HTML and PDF formats. This development was necessary because the manuscript grew so large that it needed to be split into seven separate papers for journal submission while still maintaining shared GitHub discussion across topics. When exporting an individual section, Manubot customizes the manuscript title, authors, and author contributions to pertain to that specific section. In addition, we expanded the export formats to include partial LaTeX support via Pandoc. Pandoc converts the markdown content for an individual section to TeX and the Citation Style Language JSON, which contains reference metadata generated by Manubot, to BibTeX. We customized a LaTeX template and reformatted the Manubot metadata, such as authors and their affiliations, for the LaTeX template. The exported TeX file requires manual refinement but contains all manuscript content and most of the formatting. Because LaTeX is required for manuscript submission in many fields, automating most of the process of converting markdown to a submission-friendly format expands Manubot's potential user base. Manubot users can write in the simple markdown format, render the manuscript in continuously-updated PDF or interactive HTML formats, and export the manuscript in DOCX or TeX and BibTeX for submission to traditional publishers, taking full advantage of Pandoc's powerful document conversion capabilities and Manubot's automation.

10.4 RESULTS

10.4.1 Recruitment and Manuscript Development

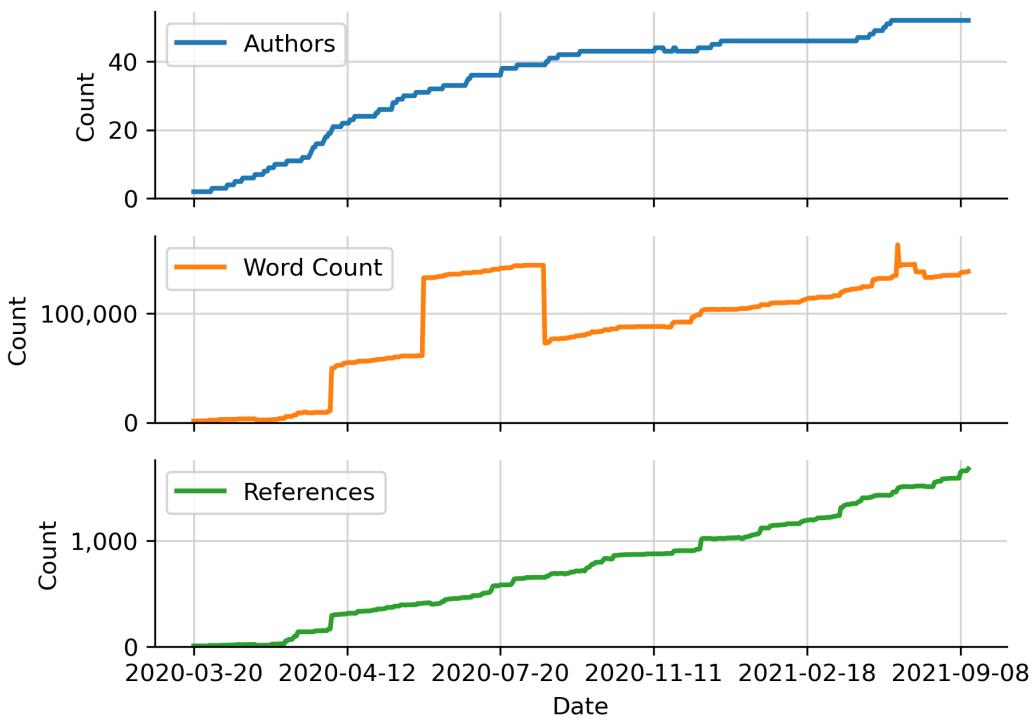


Figure 16: Project growth over time. The number of authors, word count, and number of references have all grown dramatically from when the project began on March 20, 2020. As of September 10, 2021, there were 52 authors (including consortia), 1676 references, and 138213 words. The spike in word count during summer 2020 was caused by erroneous duplication and subsequent removal of a large appendix.

Coverage by *Nature Toolbox* [1947] and an associated tweet²³ about the project on April 1, 2020 attracted the interest of the scientific community (Figure 16). Because the GitHub issues and comment systems are similar to other common web commenting systems, authors learned these tools quickly. The Gitter chat also presented a low barrier to entry. The manuscript continued to grow throughout the first year and a half of the project in both word count and the number of references (Figure 16). Though only a fraction of potential contributors contributed to the text included in the manuscripts (Figure 16), many contributors remained engaged over the long term (Figure 17). Additionally, new contributors continued to join even into the second year of the project.



Figure 17: User contributions to the manuscript text over time. The dot size indicates the number of words added or edited each month since March 2020. The figure does not depict other types of author contributions such as literature summaries, pull request review, visualization, or software.

In order to make the project more accessible, we developed resources explaining how to use GitHub's web interface to develop and edit text for Manubot assuming no prior experience working with version control. These tutorials explained how to open an issue, open a pull request, and review a pull request²⁴. Additionally, the framework for evaluating literature was converted into issue templates to simplify the review of new articles. Articles were classified as *diagnostic*, *therapeutic*, or *other*, with an associated template developed to guide the review of papers and preprints in each category. A total of 285 new paper issues had been opened as of September 13, 2021.

The manuscripts produced by the consortium (excluding this one) will be submitted to *mSystems* as part of a special issue that provides support for continuous updates as more information becomes available. One has been published and two are available as preprints. This approach allows for a version of record to be maintained alongside the most recent version, which is always available through GitHub. These manuscripts cover a wide range of topics including the fundamental biology of SARS-CoV-2 (pathogenesis [1] and evolution), biomedical advances in responding to the virus and COVID-19 (pharmaceuticals [3], nutraceuticals [2], vaccines, and diagnostic technologies), and biological and social factors influencing disease transmission and outcomes. To date, 52 authors are associated with the consortium (Figure 16).

More formal recruitment efforts to integrate with existing projects providing support for undergraduate students during COVID-19 were also successful. We incorporated summaries written by the students, post-docs, and faculty of the Immunology Institute at the Mount Sinai School of Medicine²⁵ [1931]. Additionally, two of the consortium authors were undergraduate students recruited through the American Physician Scientist Association's Virtual Summer Research Program. Thus, the consortium was successful in providing a venue for researchers across all career stages to continue investigating and publishing at a time when many biomedical researchers were unable to access their laboratory facilities.

10.4.2 Integrating Data

We integrated data into the manuscripts from several sources (Figure 15). Worldwide cases and deaths were tracked by the COVID-19 Data Repository by the Center for Systems Science and Engineering at Johns Hopkins University²⁶. The clinical trials statistics and figure were generated based on data from the University of Oxford Evidence-Based Medicine Data Lab's COVID-19 TrialsTracker [717]. Information about vaccine distribution was extracted from Our World In Data²⁷ [1948]. (Figure 14) integrates data from the CORD-19 dataset [1923].

Manubot's bibliographic management capabilities were critical because the amount of relevant literature published far outstripped what we had anticipated at the beginning of the project. As of September 10, 2021, there were 1676 references (Figure 16). The scite plugin provided a way to visually inspect the reference list to identify possible references of concern. This and the other new features required for the COVID-19 project are now included in Manubot's rootstock, which is the template GitHub repository for creating a new manuscript. Using CI, Manubot now checks that the manuscript was built correctly, runs spell-checking, and cross-references the manuscripts cited in this review. In addition, Manubot rootstock now supports citing clinical trial identifiers such as `clinicaltrials:NCT04292899` [787].

10.5 DISCUSSION

The current project was based in the GitHub repository `greenelab/covid19-review` using Manubot [1937] to continuously generate the manuscript. The Manubot framework facilitated a massive collaborative review on an urgent

topic. We demonstrated the utility of Manubot to a project where many contributors lacked expertise or even experience working with version control. This effort has produced not only seven literature reviews on topics relevant to the COVID-19 pandemic, but has also generated cyberinfrastructure for training novice users in GitHub. We also extended the functionalities of Manubot to provide more of the benefits of What You See Is What You Get platforms such as Google Docs (Table 6). Open publishing thus allowed us to harness the domain expertise of a large group of non-technical users to respond to the flood of COVID-19 publications.

Several existing and new features in Manubot aid in responding to the challenges posed by the infodemic. Manuscripts are written in markdown and can be rendered in several formats providing different advantages to users. For example, beyond building just a PDF, Manubot also renders the manuscript in HTML, DOCX, and now, LaTeX (in a more limited capacity). The interactive HTML manuscript format offers several advantages over a static PDF to harmonize available resources and address specific problems related to COVID-19. The integration of scite into the HTML build makes references more manageable by visually indicating whether their results are contested or whether they have been corrected or retracted. Cross-referencing different pieces of the manuscript, such as cited preprints with reviews stored in an appendix, is another interactive option presented by HTML. The DOCX format was preferred by most non-technical users for reviewing the final version of the manuscript and was useful for creating submissions to a biological journal. Additionally, because of the heavy emphasis on Word processing in biology, Manubot's ability to generate DOCX outputs was expanded to allow users to generate DOCX files containing only a section of the manuscript. In our case, where the full project is nearly 150,000 words, this allows individual pieces to be shared more easily. Finally, the preliminary addition of LaTeX output is useful for researchers from computational fields who submit papers in TeX format and removes the step of reformatting markdown prior to submission.

Table 6: Manubot extensions for the COVID-19 review.

Type	Description
CI	Regularly download external data sources, generate new figures and statistics, and read them when Manubot builds the latest manuscript
CI	Post spell-checking reports as pull request comments
Citations	Zotero extension to report more relevant clinical trial metadata from https://clinicaltrials.gov
Citations	Cite any Compact Uniform Resource Identifier, such as <code>clinicaltrials</code> or <code>ncbigene</code>
Citations	scite badges to track retractions, corrections, and notices of concern
Outputs	Improved support for Pandoc's LaTeX output
Outputs	Build complete manuscript alongside individual sections as standalone documents

The COVID-19 Review Consortium provided a platform for researchers to engage in scientific investigation early in the pandemic when many biological scientists were unable to access their research spaces. In turn, by seeking to adapt Manubot to allow for broader participation, we made a number of improvements that are expected to increase its appeal to researchers from all backgrounds. Manubot provided a way for contributors from a variety of backgrounds, including early-career researchers, to join a massive collaborative project while demonstrating their individual contributions to the larger work and gaining experience with version control. The licensing and infrastructure also provide the basis for individuals to adapt from this project to create their own snapshots of the COVID-19 literature that derive from, but are not wholly identical to, the primary versions of these reviews. This project suggests that massive online open publishing efforts can indeed advance scholarship through inclusion [1938], including during the extreme challenges presented by the COVID-19 pandemic.

Some challenges did arise in efforts to include an academically diverse set of authors. The barriers to entry posed by git and GitHub likely still reduced participation from individuals who might have otherwise been interested. Using pull requests as a tool for writing text is also unfamiliar to many or most scientists, and the review process can be slow, which might cause interested contributors to lose interest. Additionally, the pull request model may limit people from providing general feedback on the manuscript or a section of the manuscript, unless there is an open pull request. As a result, some feedback came through email or comments on the DOCX outputs that were then translated into issues or pull requests by the project managers. Given that our approach hinged on these version control tools, it is likely that our group of contributors was biased towards those who were interested in or experienced with computational tools. The trajectory of the pandemic itself also likely influenced participation: engagement waned over the course of the pandemic as labs opened back up and researchers were able to return to their work, and we recruited very few senior clinicians to the project, which is unsurprising given the load on medical professionals during this time. Engagement that waxes and wanes is, however, typical when writing massively open online papers [1938]. Adding features such as spell-check did improve usability, and additional features such as automatically checking the formatting of citations could further improve the usability of this tool. In the future, a formal study of participation could allow for quantification of these biases and improved efforts to foster inclusion.

Additional limitations are challenges associated with massively open online papers in general. With such a large amount of text, it is not possible to keep all sections of the manuscript up to date at all times. Readers are not able to distinguish when each section was updated. Even GitHub's blame functionality does not distinguish minor changes from substantive updates to the text. While much of the data and statistics update automatically, the text itself required updating by human experts. This asynchronicity could potentially introduce incompatibility between the figures and the surrounding text. Similarly, in line with the collaboration-related challenges of the project, some authors returned to update their text, while others did not. As a result, the lead authors of each paper often spent several weeks prior to journal submission updating the text to reflect new developments in each area. In the future, it may be possible to streamline this process

through integration with a tool such as CoronaCentral [1924] to automatically identify relevant, high-impact papers that need to be included, although expertise would still be required to incorporate them. Another challenge involves tracking preprints as they are reviewed or critiqued, revised, and potentially published. While updating the content of the manuscript would likely fall to human contributors, automatic detection of published versions of preprints [1949] could be integrated in the future. These challenges are exacerbated by the scale of the infodemic, but developing solutions would benefit future projects tracking more typical trends in publication. Similarly, outputting machine readable summaries of key information in the COVID-19 review manuscripts could reduce their contribution to the infodemic. As it stands, the integration of Compact Uniform Resource Identifier does make a step in this direction. Formal identifiers could be used to extract relationships among clinical trials, genes, publications, and other entities. Thus, the experience of using Manubot for a massive project has laid the foundation for future additions to enhance user experience and inclusivity.

10.6 CONCLUSION

With the worldwide scientific community uniting during 2020 and 2021 to investigate COVID-19 from a wide range of perspectives, findings from many disciplines are relevant on a rapid timescale to a broad scientific audience. As many other efforts have described, the publishing rate of formal manuscripts and preprints about COVID-19 has been unprecedented [1921], and efforts to review the body of COVID-19 literature are faced with an ever-expanding corpus to evaluate. In the case of the seven manuscripts produced by the COVID-19 Review Consortium, Manubot allows for continuous updating of the manuscripts as the pandemic enters its second year and the landscape shifts with the emergence of promising therapeutics and vaccines [3]. These manuscripts pull data from external sources and update information and visualizations daily using CI. By off-loading some updates to computational pipelines, domain experts can focus on the broader implications of new information as it emerges. Centralizing, summarizing, and critiquing data and literature broadly relevant to COVID-19 can expedite the interdisciplinary scientific process that is currently happening at an advanced pace. As of September 13, 2021, 2886 commits have been made to the manuscript across 575 merged pull requests. The efforts of the COVID-19 Review Consortium illustrate the value of including open source tools, including those focused on open publishing, in these efforts. By facilitating the versioning of text, such platforms also allow for documentation of the evolution of thought in an evolving area and formal analysis of a collaborative project. This application of version control holds the potential to improve scientific publishing in a range of disciplines, including those outside of traditional computational fields. While Manubot is a technologically complex tool, this project demonstrates that it can be applied to a variety of projects. Future work can address remaining limitations and continue to advance Manubot as an inclusive tool for open publishing projects.

11 Additional Items

11.1 Competing Interests

Author	Competing Interests	Last Reviewed
Halie M. Rando	None	2021-01-20
Casey S. Greene	None	2021-01-20
Michael P. Robson	None	2020-11-12
Simina M. Boca	Currently an employee at AstraZeneca, Gaithersburg, MD, USA, may own stock or stock options.	2021-07-01
Nils Wellhausen	None	2020-11-03
Ronan Lordan	None	2020-11-03
Christian Brueffer	Employee and shareholder of SAGA Diagnostics AB.	2020-11-11
Sandipan Ray	None	2020-11-11
Lucy D'Agostino McGowan	Received consulting fees from Acelity and Sanofi in the past five years	2020-11-10
Anthony Gitter	Filed a patent application with the Wisconsin Alumni Research Foundation related to classifying activated T cells	2020-11-10
Anna Ada Dattoli	None	2020-03-26
Ryan Velazquez	None	2020-11-10
John P. Barton	None	2020-11-11
Jeffrey M. Field	None	2020-11-12
Bharath Ramsundar	None	2020-11-11
Adam L. MacLean	None	2021-02-23
Alexandra J. Lee	None	2020-11-09
Immunology Institute of the Icahn School of Medicine	None	2020-04-07
Fengling Hu	None	2020-04-08
Nafisa M. Jadavji	None	2020-11-11
Elizabeth Sell	None	2020-11-11
Vincent Rubinetti	None	2021-04-29
Jinhui Wang	None	2021-01-21
Diane N. Rafizadeh	None	2020-11-11
Ashwin N. Skelly	None	2020-11-11

Author	Competing Interests	Last Reviewed
Marouen Ben Guebila	None	2021-08-02
Likhitha Kolla	None	2020-11-16
David Manheim	None	2022-03-15
Soumita Ghosh	None	2020-11-09
James Brian Byrd	Funded by FastGrants to conduct a COVID-19-related clinical trial	2020-11-12
YoSon Park	YoSon Park is affiliated with Pfizer Worldwide Research. The author has no financial interests to declare and contributed as an author prior to joining Pfizer, and the work was not part of a Pfizer collaboration nor was it funded by Pfizer.	2020-01-22
Vikas Bansal	None	2021-01-25
Stephen Capone	None	2020-11-11
John J. Dziak	None	2020-11-11
Yuchen Sun	None	2020-11-11
Yanjun Qi	None	2020-07-09
Lamonica Shinholster	None	2020-11-11
Temitayo Lukan	None	2020-11-10
Sergey Knyazev	None	2020-11-11
Dimitri Perrin	None	2020-11-11
Serghei Mangul	None	2020-11-11
Shikta Das	None	2020-08-13
Gregory L Szeto	None	2020-11-16
Tiago Lubiana	None	2020-11-11
David Mai	None	2021-01-08
COVID-19 Review Consortium	None	2021-01-16
Rishi Raj Goel	None	2021-01-20
Joel D Boerckel	None	2021-03-26
Amruta Naik	None	2021-04-05
Yusha Sun	None	2021-04-10
Daniel S. Himmelstein	None	2021-04-30
Jeremy P. Kamil	TBD	2021-04-30
Jesse G. Meyer	None	2022-01-06
Ariel I. Mundo	None	2021-12-19

11.2 Author Contributions

Author	Contributions
Halie M. Rando	D, E, Project Administration, Software, Visualization, Writing - Original Draft, Writing - Review & Editing
Casey S. Greene	Conceptualization, Project Administration, Software, Supervision, Writing - Original Draft, Writing - Review & Editing
Michael P. Robson	Methodology, Software, Supervision
Simina M. Boca	Methodology, Project Administration, Writing - Review & Editing
Nils Wellhausen	Project Administration, Visualization, Writing - Original Draft, Writing - Review & Editing
Ronan Lordan	Conceptualization, Project Administration, Writing - Original Draft, Writing - Review & Editing
Christian Brueffer	Project Administration, Writing - Original Draft, Writing - Review & Editing
Sandipan Ray	Writing - Original Draft, Writing - Review & Editing
Lucy D'Agostino McGowan	Methodology, Writing - Original Draft, Writing - Review & Editing
Anthony Gitter	Methodology, Project Administration, Software, Visualization, Writing - Original Draft, Writing - Review & Editing
Anna Ada Dattoli	Writing - Original Draft
Ryan Velazquez	Methodology, Software, Writing - Review & Editing
John P. Barton	Writing - Original Draft, Writing - Review & Editing
Jeffrey M. Field	Writing - Original Draft, Writing - Review & Editing
Bharath Ramsundar	Writing - Review & Editing
Adam L. MacLean	Writing - Original Draft, Writing - Review & Editing
Alexandra J. Lee	Writing - Original Draft, Writing - Review & Editing
Immunology Institute of the Icahn School of Medicine	Data Curation
Fengling Hu	Writing - Original Draft, Writing - Review & Editing
Nafisa M. Jadavji	Supervision, Writing - Original Draft, Writing - Review & Editing
Elizabeth Sell	Writing - Original Draft, Writing - Review & Editing

Author	Contributions
Vincent Rubinetti	Software, Visualization, Writing - Original Draft
Jinhui Wang	Writing - Original Draft, Writing - Review & Editing
Diane N. Rafizadeh	Project Administration, Writing - Original Draft, Writing - Review & Editing
Ashwin N. Skelly	Writing - Original Draft, Writing - Review & Editing
Marouen Ben Guebila	Writing - Original Draft, Writing - Review & Editing
Likhitha Kolla	Writing - Original Draft
David Manheim	Writing - Original Draft, Writing - Review & Editing
Soumita Ghosh	Writing - Original Draft
James Brian Byrd	Writing - Original Draft, Writing - Review & Editing
YoSon Park	Writing - Original Draft, Writing - Review & Editing
Vikas Bansal	Writing - Original Draft, Writing - Review & Editing
Stephen Capone	Writing - Original Draft, Writing - Review & Editing
John J. Dziak	Writing - Original Draft, Writing - Review & Editing
Yuchen Sun	Visualization
Yanjun Qi	Visualization
Lamonica Shinholster	Writing - Original Draft
Temitayo Lukan	Investigation, Writing - Original Draft
Sergey Knyazev	Writing - Original Draft, Writing - Review & Editing
Dimitri Perrin	Writing - Original Draft, Writing - Review & Editing
Serghei Mangul	Writing - Review & Editing
Shikta Das	Writing - Review & Editing
Gregory L Szeto	Writing - Review & Editing
Tiago Lubiana	Writing - Review & Editing
David Mai	Writing - Original Draft, Writing - Review & Editing
COVID-19 Review Consortium	Project Administration
Rishi Raj Goel	Writing - Original Draft, Writing - Review & Editing
Joel D Boerckel	Writing - Review & Editing
Amruta Naik	MISSING

Author	Contributions
Yusha Sun	Writing - Review & Editing
Daniel S. Himmelstein	Software
Jeremy P. Kamil	Writing - Review & Editing
Jesse G. Meyer	Writing - Original Draft, Writing - Review & Editing
Ariel I. Mundo	Writing - Original Draft, Writing - Review & Editing

11.3 Acknowledgements

We thank Nick DeVito for assistance with the Evidence-Based Medicine Data Lab COVID-19 TrialsTracker data. We thank Yael Evelyn Marshall who contributed writing (original draft) as well as reviewing and editing of pieces of the text but who did not formally approve the manuscript, as well as Ronnie Russell, who contributed text to and helped develop the structure of the manuscript early in the writing process and Matthias Fax who helped with writing and editing text related to diagnostics. We are also very grateful to James Fraser for suggestions about successes and limitations in the area of computational screening for drug repurposing. We are grateful to the following contributors for reviewing pieces of the text: Nadia Danilova, James Eberwine and Ipsita Krishnan.

11.4 References

1. **Pathogenesis, Symptomatology, and Transmission of SARS-CoV-2 through Analysis of Viral Genomics and Structure**
Halie M Rando, Adam L MacLean, Alexandra J Lee, Ronan Lordan, Sandipan Ray, Vikas Bansal, Ashwin N Skelly, Elizabeth Sell, John J Dziak, Lamonica Shinholster, ... Casey S Greene
mSystems (2021-10-26) <https://pubmed.ncbi.nlm.nih.gov/34698547/>
DOI: [10.1128/msystems.00095-21](https://doi.org/10.1128/msystems.00095-21)
2. **Dietary Supplements and Nutraceuticals under Investigation for COVID-19 Prevention and Treatment**
Ronan Lordan, Halie M Rando, COVID-19 Review Consortium, Casey S Greene
mSystems (2021-05-04) <https://pubmed.ncbi.nlm.nih.gov/33947804/>
DOI: [10.1128/msystems.00122-21](https://doi.org/10.1128/msystems.00122-21)
3. **Identification and Development of Therapeutics for COVID-19**
Halie M Rando, Nils Wellhausen, Soumita Ghosh, Alexandra J Lee, Anna Ada Dattoli, Fengling Hu, James Brian Byrd, Diane N Rafizadeh, Ronan Lordan, Yanjun Qi, ... Casey S Greene
mSystems (2021-12-21) <https://pubmed.ncbi.nlm.nih.gov/34726496/>
DOI: [10.1128/msystems.00233-21](https://doi.org/10.1128/msystems.00233-21)
4. **An Open-Publishing Response to the COVID-19 Infodemic**
Halie M Rando, Simina M Boca, Lucy D'Agostino McGowan, Daniel S Himmelstein, Michael P Robson, Vincent Rubinetti, Ryan Velazquez, Casey S Greene, Anthony Gitter
Proceedings of the Workshop on Digital Infrastructures for Scholarly Content Objects (DISCO 2021) (2021-09-30) <http://ceur-ws.org/Vol-2976/paper-2.pdf>
5. **Molecular and Serologic Diagnostic Technologies for SARS-CoV-2**
Halie M Rando, Christian Brueffer, Ronan Lordan, Anna Ada Dattoli, David Manheim, Jesse G Meyer, Ariel I Mundo, Dimitri Perrin, David Mai, Nils Wellhausen, ... Casey S Greene
arXiv (2022-04-29) <https://arxiv.org/abs/2204.12598>
6. **Coronavirus Disease 2019 (COVID-19) – Symptoms**
CDC
Centers for Disease Control and Prevention (2020-12-22)
<https://www.cdc.gov/coronavirus/2019-ncov/symptoms-testing/symptoms.html>
7. **WHO Declares COVID-19 a Pandemic**
Domenico Cucinotta, Maurizio Vanelli
Acta Bio Medica Atenei Parmensis (2020-03-19) <https://doi.org/ggq86h>
DOI: [10.23750/abm.v91i1.9397](https://doi.org/10.23750/abm.v91i1.9397) · PMID: [32191675](https://pubmed.ncbi.nlm.nih.gov/32191675/) · PMCID: [PMC7569573](https://pubmed.ncbi.nlm.nih.gov/PMC7569573/)
8. **Acute lung injury in patients with COVID-19 infection**
Liyang Li, Qihong Huang, Diane C Wang, David H Ingbar, Xiangdong Wang
Clinical and Translational Medicine (2020-03-31) <https://doi.org/ghqcrz>

9. **A Novel Coronavirus Genome Identified in a Cluster of Pneumonia Cases — Wuhan, China 2019–2020**

Wenjie Tan, Xiang Zhao, Xuejun Ma, Wenling Wang, Peihua Niu, Wenbo Xu, George F. Gao, Guizhen Wu, MHC Key Laboratory of Biosafety, National Institute for Viral Disease Control and Prevention, China CDC, Beijing, China, Center for Biosafety Mega-Science, Chinese Academy of Sciences, Beijing, China

China CDC Weekly (2020) <https://doi.org/gg8z47>

DOI: [10.46234/ccdcw2020.017](https://doi.org/10.46234/ccdcw2020.017)

10. **Structure, Function, and Evolution of Coronavirus Spike Proteins**

Fang Li

Annual Review of Virology (2016-09-29) <https://doi.org/ggr7gy>

DOI: [10.1146/annurev-virology-110615-042301](https://doi.org/10.1146/annurev-virology-110615-042301) · PMID: [27578435](https://pubmed.ncbi.nlm.nih.gov/27578435/) ·

PMCID: [PMC5457962](https://pubmed.ncbi.nlm.nih.gov/PMC5457962/)

11. **A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster**

Jasper Fuk-Woo Chan, Shuofeng Yuan, Kin-Hang Kok, Kelvin Kai-Wang To, Hin Chu, Jin Yang, Fanfan Xing, Jieling Liu, Cyril Chik-Yan Yip, Rosana Wing-Shan Poon, ... Kwok-Yung Yuen

The Lancet (2020-02) <https://doi.org/ggjs7j>

DOI: [10.1016/s0140-6736\(20\)30154-9](https://doi.org/10.1016/s0140-6736(20)30154-9) · PMID: [31986261](https://pubmed.ncbi.nlm.nih.gov/31986261/) · PMCID:

[PMC7159286](https://pubmed.ncbi.nlm.nih.gov/PMC7159286/)

12. **Fields virology**

Bernard N Fields, David M Knipe, Peter M Howley (editors)

Wolters Kluwer Health/Lippincott Williams & Wilkins (2007)

ISBN: 9780781760607

13. **Important Role for the Transmembrane Domain of Severe Acute Respiratory Syndrome Coronavirus Spike Protein during Entry**

Rene Broer, Bertrand Boson, Willy Spaan, François-Loïc Cosset, Jeroen Corver

Journal of Virology (2006-02-01) <https://doi.org/dvvg2h>

DOI: [10.1128/jvi.80.3.1302-1310.2006](https://doi.org/10.1128/jvi.80.3.1302-1310.2006) · PMID: [16415007](https://pubmed.ncbi.nlm.nih.gov/16415007/) · PMCID:

[PMC1346921](https://pubmed.ncbi.nlm.nih.gov/PMC1346921/)

14. **Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation**

Daniel Wrapp, Nianshuang Wang, Kizzmekia S Corbett, Jory A Goldsmith, Ching-Lin Hsieh, Olubukola Abiona, Barney S Graham, Jason S McLellan

Science (2020-03-13) <https://doi.org/ggmtk2>

DOI: [10.1126/science.abb2507](https://doi.org/10.1126/science.abb2507) · PMID: [32075877](https://pubmed.ncbi.nlm.nih.gov/32075877/)

15. **Medical microbiology**

Samuel Baron (editor)

University of Texas Medical Branch at Galveston (1996)

ISBN: 9780963117212

16. **Coronaviruses: An Overview of Their Replication and Pathogenesis**
Anthony R Fehr, Stanley Perlman
Methods in Molecular Biology (2015) <https://doi.org/ggpc6n>
DOI: [10.1007/978-1-4939-2438-7_1](https://doi.org/10.1007/978-1-4939-2438-7_1) · PMID: [25720466](#) · PMCID: [PMC4369385](#)
17. **Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding**
Roujian Lu, Xiang Zhao, Juan Li, Peihua Niu, Bo Yang, Honglong Wu, Wenling Wang, Hao Song, Baoying Huang, Na Zhu, ... Wenjie Tan
The Lancet (2020-02) <https://doi.org/ggjr43>
DOI: [10.1016/s0140-6736\(20\)30251-8](https://doi.org/10.1016/s0140-6736(20)30251-8)
18. **Emerging coronaviruses: Genome structure, replication, and pathogenesis**
Yu Chen, Qianyun Liu, Deyin Guo
Journal of Medical Virology (2020-02-07) <https://doi.org/ggjvwj>
DOI: [10.1002/jmv.25681](https://doi.org/10.1002/jmv.25681) · PMID: [31967327](#)
19. **SARS coronavirus entry into host cells through a novel clathrin-and caveolae-independent endocytic pathway**
Hongliang Wang, Peng Yang, Kangtai Liu, Feng Guo, Yanli Zhang, Gongyi Zhang, Chengyu Jiang
Cell Research (2008-01-29) <https://doi.org/bp9275>
DOI: [10.1038/cr.2008.15](https://doi.org/10.1038/cr.2008.15) · PMID: [18227861](#) · PMCID: [PMC7091891](#)
20. **Virus Entry by Endocytosis**
Jason Mercer, Mario Schelhaas, Ari Helenius
Annual Review of Biochemistry (2010-06-07) <https://doi.org/cw4dnb>
DOI: [10.1146/annurev-biochem-060208-104626](https://doi.org/10.1146/annurev-biochem-060208-104626) · PMID: [20196649](#)
21. **Mechanisms of Coronavirus Cell Entry Mediated by the Viral Spike Protein**
Sandrine Belouzard, Jean K Millet, Beth N Licitra, Gary R Whittaker
Viruses (2012-06-20) <https://doi.org/gbbktb>
DOI: [10.3390/v4061011](https://doi.org/10.3390/v4061011) · PMID: [22816037](#) · PMCID: [PMC3397359](#)
22. **Phylogenetic Analysis and Structural Modeling of SARS-CoV-2 Spike Protein Reveals an Evolutionary Distinct and Proteolytically Sensitive Activation Loop**
Javier A Jaimes, Nicole M André, Joshua S Chappie, Jean K Millet, Gary R Whittaker
Journal of Molecular Biology (2020-05) <https://doi.org/ggtxhr>
DOI: [10.1016/j.jmb.2020.04.009](https://doi.org/10.1016/j.jmb.2020.04.009) · PMID: [32320687](#) · PMCID: [PMC7166309](#)
23. **Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein**
Alexandra C Walls, Young-Jun Park, MAlejandra Tortorici, Abigail Wall, Andrew T McGuire, David Veesler
Cell (2020-04) <https://doi.org/dpvh>
DOI: [10.1016/j.cell.2020.02.058](https://doi.org/10.1016/j.cell.2020.02.058) · PMID: [32155444](#) · PMCID: [PMC7102599](#)

24. **Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target**
Haibo Zhang, Josef M Penninger, Yimin Li, Nanshan Zhong, Arthur S Slutsky
Intensive Care Medicine (2020-03-03) <https://doi.org/ggpx6p>
DOI: [10.1007/s00134-020-05985-9](https://doi.org/10.1007/s00134-020-05985-9) · PMID: [32125455](https://pubmed.ncbi.nlm.nih.gov/32125455/) · PMCID: [PMC7079879](https://pubmed.ncbi.nlm.nih.gov/PMC7079879/)
25. **Infection of Human Airway Epithelia by Sars Coronavirus is Associated with ACE2 Expression and Localization**
Hong Peng Jia, Dwight C Look, Melissa Hickey, Lei Shi, Lecia Pewe, Jason Netland, Michael Farzan, Christine Wohlford-Lenane, Stanley Perlman, Paul B McCray
Advances in Experimental Medicine and Biology (2006) <https://doi.org/dhh5tp>
DOI: [10.1007/978-0-387-33012-9_85](https://doi.org/10.1007/978-0-387-33012-9_85) · PMID: [17037581](https://pubmed.ncbi.nlm.nih.gov/17037581/) · PMCID: [PMC7123641](https://pubmed.ncbi.nlm.nih.gov/PMC7123641/)
26. **The protein expression profile of ACE2 in human tissues**
Feria Hikmet, Loren Méar, Åsa Edvinsson, Patrick Micke, Mathias Uhlén, Cecilia Lindskog
Molecular Systems Biology (2020-07-26) <https://doi.org/gg6mxv>
DOI: [10.1525/msb.20209610](https://doi.org/10.1525/msb.20209610) · PMID: [32715618](https://pubmed.ncbi.nlm.nih.gov/32715618/) · PMCID: [PMC7383091](https://pubmed.ncbi.nlm.nih.gov/PMC7383091/)
27. **Receptor Recognition Mechanisms of Coronaviruses: a Decade of Structural Studies**
Fang Li
Journal of Virology (2015-02-15) <https://doi.org/f633jb>
DOI: [10.1128/jvi.02615-14](https://doi.org/10.1128/jvi.02615-14) · PMID: [25428871](https://pubmed.ncbi.nlm.nih.gov/25428871/) · PMCID: [PMC4338876](https://pubmed.ncbi.nlm.nih.gov/PMC4338876/)
28. **The spike protein of SARS-CoV — a target for vaccine and therapeutic development**
Lanying Du, Yuxian He, Yusen Zhou, Shuwen Liu, Bo-Jian Zheng, Shibo Jiang
Nature Reviews Microbiology (2009-02-09) <https://doi.org/d4tq4t>
DOI: [10.1038/nrmicro2090](https://doi.org/10.1038/nrmicro2090) · PMID: [19198616](https://pubmed.ncbi.nlm.nih.gov/19198616/) · PMCID: [PMC2750777](https://pubmed.ncbi.nlm.nih.gov/PMC2750777/)
29. **Molecular Interactions in the Assembly of Coronaviruses**
Cornelis AM de Haan, Peter JM Rottier
Advances in Virus Research (2005) <https://doi.org/cf8chz>
DOI: [10.1016/s0065-3527\(05\)64006-7](https://doi.org/10.1016/s0065-3527(05)64006-7) · PMID: [16139595](https://pubmed.ncbi.nlm.nih.gov/16139595/) · PMCID: [PMC7112327](https://pubmed.ncbi.nlm.nih.gov/PMC7112327/)
30. **Coronavirus membrane fusion mechanism offers a potential target for antiviral development**
Tiffany Tang, Miya Bidon, Javier A Jaimes, Gary R Whittaker, Susan Daniel
Antiviral Research (2020-06) <https://doi.org/ggr23b>
DOI: [10.1016/j.antiviral.2020.104792](https://doi.org/10.1016/j.antiviral.2020.104792) · PMID: [32272173](https://pubmed.ncbi.nlm.nih.gov/32272173/) · PMCID: [PMC7194977](https://pubmed.ncbi.nlm.nih.gov/PMC7194977/)
31. **SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor**
Markus Hoffmann, Hannah Kleine-Weber, Simon Schroeder, Nadine Krüger, Tanja Herrler, Sandra Erichsen, Tobias S Schiergens, Georg

- Herrler, Nai-Huei Wu, Andreas Nitsche, ... Stefan Pöhlmann
Cell (2020-04) <https://doi.org/ggnq74>
DOI: [10.1016/j.cell.2020.02.052](https://doi.org/10.1016/j.cell.2020.02.052) · PMID: [32142651](#) · PMCID: [PMC7102627](#)
32. **Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV**
Xiuyuan Ou, Yan Liu, Xiaobo Lei, Pei Li, Dan Mi, Lili Ren, Li Guo, Ruixuan Guo, Ting Chen, Jiaxin Hu, ... Zhaojun Qian
Nature Communications (2020-03-27) <https://doi.org/ggqsrf>
DOI: [10.1038/s41467-020-15562-9](https://doi.org/10.1038/s41467-020-15562-9) · PMID: [32221306](#) · PMCID: [PMC7100515](#)
33. **The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade**
B Coutard, C Valle, X de Lamballerie, B Canard, NG Seidah, E Decroly
Antiviral Research (2020-04) <https://doi.org/ggpvhk>
DOI: [10.1016/j.antiviral.2020.104742](https://doi.org/10.1016/j.antiviral.2020.104742) · PMID: [32057769](#) · PMCID: [PMC7114094](#)
34. **The role of furin cleavage site in SARS-CoV-2 spike protein-mediated membrane fusion in the presence or absence of trypsin**
Shuai Xia, Qiaoshuai Lan, Shan Su, Xinling Wang, Wei Xu, Zehong Liu, Yun Zhu, Qian Wang, Lu Lu, Shibo Jiang
Signal Transduction and Targeted Therapy (2020-06-12)
<https://doi.org/gmfkwx>
DOI: [10.1038/s41392-020-0184-0](https://doi.org/10.1038/s41392-020-0184-0) · PMID: [32532959](#) · PMCID: [PMC7289711](#)
35. **Furin Cleavage Site Is Key to SARS-CoV-2 Pathogenesis**
Bryan A Johnson, Xuping Xie, Birte Kalveram, Kumari G Lokugamage, Antonio Muruato, Jing Zou, Xianwen Zhang, Terry Juelich, Jennifer K Smith, Lihong Zhang, ... Vineet D Menachery
Cold Spring Harbor Laboratory (2020-08-26) <https://doi.org/gk7dgc>
DOI: [10.1101/2020.08.26.268854](https://doi.org/10.1101/2020.08.26.268854) · PMID: [32869021](#) · PMCID: [PMC7457603](#)
36. **Loss of furin cleavage site attenuates SARS-CoV-2 pathogenesis**
Bryan A Johnson, Xuping Xie, Adam L Bailey, Birte Kalveram, Kumari G Lokugamage, Antonio Muruato, Jing Zou, Xianwen Zhang, Terry Juelich, Jennifer K Smith, ... Vineet D Menachery
Nature (2021-01-25) <https://doi.org/gmfkwz>
DOI: [10.1038/s41586-021-03237-4](https://doi.org/10.1038/s41586-021-03237-4) · PMID: [33494095](#) · PMCID: [PMC8175039](#)
37. **The furin cleavage site in the SARS-CoV-2 spike protein is required for transmission in ferrets**
Thomas P Peacock, Daniel H Goldhill, Jie Zhou, Laury Baillon, Rebecca Frise, Olivia C Swann, Ruthiran Kugathasan, Rebecca Penn, Jonathan C Brown, Raul Y Sanchez-David, ... Wendy S Barclay
Nature Microbiology (2021-04-27) <https://doi.org/gk7gn8>
DOI: [10.1038/s41564-021-00908-w](https://doi.org/10.1038/s41564-021-00908-w) · PMID: [33907312](#)
38. **Furin Inhibitors Block SARS-CoV-2 Spike Protein Cleavage to Suppress Virus Production and Cytopathic Effects**

- Ya-Wen Cheng, Tai-Ling Chao, Chiao-Ling Li, Mu-Fan Chiu, Han-Chieh Kao, Sheng-Han Wang, Yu-Hao Pang, Chih-Hui Lin, Ya-Min Tsai, Wen-Hau Lee, ... Shiou-Hwei Yeh
Cell Reports (2020-10) <https://doi.org/gjvhjf>
DOI: [10.1016/j.celrep.2020.108254](https://doi.org/10.1016/j.celrep.2020.108254) · PMID: [33007239](https://pubmed.ncbi.nlm.nih.gov/33007239/) · PMCID: [PMC7510585](https://pubmed.ncbi.nlm.nih.gov/PMC7510585/)
39. **Coronaviridae ~ ViralZone** <https://viralzone.expasy.org/30>
40. **Proteolytic Cleavage of the SARS-CoV-2 Spike Protein and the Role of the Novel S1/S2 Site**
Javier A Jaimes, Jean K Millet, Gary R Whittaker
iScience (2020-06) <https://doi.org/gg2ccm>
DOI: [10.1016/j.isci.2020.101212](https://doi.org/10.1016/j.isci.2020.101212) · PMID: [32512386](https://pubmed.ncbi.nlm.nih.gov/32512386/) · PMCID: [PMC7255728](https://pubmed.ncbi.nlm.nih.gov/PMC7255728/)
41. **Development of the Endothelium: An Emphasis on Heterogeneity**
Laura Dyer, Cam Patterson
Seminars in Thrombosis and Hemostasis (2010-04)
<https://doi.org/bbs53m>
DOI: [10.1055/s-0030-1253446](https://doi.org/10.1055/s-0030-1253446) · PMID: [20490975](https://pubmed.ncbi.nlm.nih.gov/20490975/) · PMCID: [PMC3328212](https://pubmed.ncbi.nlm.nih.gov/PMC3328212/)
42. **The vascular endothelium-pathobiologic significance.**
G Thorgeirsson, AL Robertson
The American journal of pathology (1978-12)
<https://www.ncbi.nlm.nih.gov/pubmed/362947>
PMID: [362947](https://pubmed.ncbi.nlm.nih.gov/362947/) · PMCID: [PMC2018350](https://pubmed.ncbi.nlm.nih.gov/PMC2018350/)
43. **The endothelial glycocalyx: composition, functions, and visualization**
Sietze Reitsma, Dick W Slaaf, Hans Vink, Marc AMJ van Zandvoort, Mirjam GA oude Egbrink
Pflügers Archiv - European Journal of Physiology (2007-01-26)
<https://doi.org/fsxzdn>
DOI: [10.1007/s00424-007-0212-8](https://doi.org/10.1007/s00424-007-0212-8) · PMID: [17256154](https://pubmed.ncbi.nlm.nih.gov/17256154/) · PMCID: [PMC1915585](https://pubmed.ncbi.nlm.nih.gov/PMC1915585/)
44. **SARS-CoV-2 Infection Depends on Cellular Heparan Sulfate and ACE2**
Thomas Mandel Clausen, Daniel R Sandoval, Charlotte B Spliid, Jessica Pihl, Hailee R Perrett, Chelsea D Painter, Anoop Narayanan, Sydney A Majowicz, Elizabeth M Kwong, Rachael N McVicar, ... Jeffrey D Esko
Cell (2020-11) <https://doi.org/ghj58m>
DOI: [10.1016/j.cell.2020.09.033](https://doi.org/10.1016/j.cell.2020.09.033) · PMID: [32970989](https://pubmed.ncbi.nlm.nih.gov/32970989/) · PMCID: [PMC7489987](https://pubmed.ncbi.nlm.nih.gov/PMC7489987/)
45. **Role of heparan sulfate in immune system-blood vessel interactions**
Nathan S Ihrcke, Lucile E Wrenshall, Bonnie J Lindman, Jeffrey L Platt
Immunology Today (1993-10) <https://doi.org/bp82dx>
DOI: [10.1016/0167-5699\(93\)90265-m](https://doi.org/10.1016/0167-5699(93)90265-m)
46. **Heparan sulfate proteoglycans as attachment factor for SARS-CoV-2**

Lin Liu, Pradeep Chopra, Xiuru Li, Kim M Bouwman, SMark Tompkins, Margreet A Wolfert, Robert P de Vries, Geert-Jan Boons
Cold Spring Harbor Laboratory (2020-05-10) <https://doi.org/gjgqbf>
DOI: [10.1101/2020.05.10.087288](https://doi.org/10.1101/2020.05.10.087288) · PMID: [32511404](https://pubmed.ncbi.nlm.nih.gov/32511404/) · PMCID: [PMC7263551](https://pubmed.ncbi.nlm.nih.gov/PMC7263551/)

47. **Heparin and Heparan Sulfate: Analyzing Structure and Microheterogeneity**
Zachary Shriver, Ishan Capila, Ganesh Venkataraman, Ram Sasisekharan
Heparin - A Century of Progress (2011-12-02) <https://doi.org/gjgp97>
DOI: [10.1007/978-3-642-23056-1_8](https://doi.org/10.1007/978-3-642-23056-1_8) · PMID: [22566225](https://pubmed.ncbi.nlm.nih.gov/22566225/) · PMCID: [PMC3755452](https://pubmed.ncbi.nlm.nih.gov/PMC3755452/)
48. **The Pulmonary Endothelial Glycocalyx in ARDS: A Critical Role for Heparan Sulfate**
Wells B LaRivière, Eric P Schmidt
Current Topics in Membranes (2018) <https://doi.org/gjgp98>
DOI: [10.1016/bs.ctm.2018.08.005](https://doi.org/10.1016/bs.ctm.2018.08.005) · PMID: [30360782](https://pubmed.ncbi.nlm.nih.gov/30360782/)
49. **Heparan sulfate assists SARS-CoV-2 in cell entry and can be targeted by approved drugs in vitro**
Qi Zhang, Catherine Zhengzheng Chen, Manju Swaroop, Miao Xu, Lihui Wang, Juhyoung Lee, Amy Qiu Wang, Manisha Pradhan, Natalie Hagen, Lu Chen, ... Yihong Ye
Cell Discovery (2020-11-04) <https://doi.org/gjgqbd>
DOI: [10.1038/s41421-020-00222-5](https://doi.org/10.1038/s41421-020-00222-5) · PMID: [33298900](https://pubmed.ncbi.nlm.nih.gov/33298900/) · PMCID: [PMC7610239](https://pubmed.ncbi.nlm.nih.gov/PMC7610239/)
50. **Beneficial non-anticoagulant mechanisms underlying heparin treatment of COVID-19 patients**
Baranca Buijsers, Cansu Yanginlar, Marissa L Maciej-Hulme, Quirijn de Mast, Johan van der Vlag
EBioMedicine (2020-09) <https://doi.org/gjgqbb>
DOI: [10.1016/j.ebiom.2020.102969](https://doi.org/10.1016/j.ebiom.2020.102969) · PMID: [32853989](https://pubmed.ncbi.nlm.nih.gov/32853989/) · PMCID: [PMC7445140](https://pubmed.ncbi.nlm.nih.gov/PMC7445140/)
51. **The Effect of Anticoagulation Use on Mortality in COVID-19 Infection**
Husam M Salah, Jwan A Naser, Giuseppe Calcaterra, Pier Paolo Bassareo, Jawahar L Mehta
The American Journal of Cardiology (2020-11) <https://doi.org/gjgp99>
DOI: [10.1016/j.amjcard.2020.08.005](https://doi.org/10.1016/j.amjcard.2020.08.005) · PMID: [32892991](https://pubmed.ncbi.nlm.nih.gov/32892991/) · PMCID: [PMC7428681](https://pubmed.ncbi.nlm.nih.gov/PMC7428681/)
52. **Protective Effects of Lactoferrin against SARS-CoV-2 Infection In Vitro**
Claudio Salaris, Melania Scarpa, Marina Elli, Alice Bertolini, Simone Guglielmetti, Fabrizio Pregliasco, Corrado Blandizzi, Paola Brun, Ignazio Castagliuolo
Nutrients (2021-01-23) <https://doi.org/gjgqbg>
DOI: [10.3390/nu13020328](https://doi.org/10.3390/nu13020328) · PMID: [33498631](https://pubmed.ncbi.nlm.nih.gov/33498631/) · PMCID: [PMC7911668](https://pubmed.ncbi.nlm.nih.gov/PMC7911668/)
53. **Glycocalyx as Possible Limiting Factor in COVID-19**

Patricia P Wadowski, Bernd Jilma, Christoph W Kopp, Sebastian Ertl, Thomas Gremmel, Renate Koppensteiner
Frontiers in Immunology (2021-02-22) <https://doi.org/gh4qqz>
DOI: [10.3389/fimmu.2021.607306](https://doi.org/10.3389/fimmu.2021.607306) · PMID: [33692785](#) · PMCID: [PMC7937603](#)

54. **Spatiotemporal interplay of severe acute respiratory syndrome coronavirus and respiratory mucosal cells drives viral dissemination in rhesus macaques**

L Liu, Q Wei, K Nishiura, J Peng, H Wang, C Midkiff, X Alvarez, C Qin, A Lackner, Z Chen
Mucosal Immunology (2015-12-09) <https://doi.org/f8r7dk>
DOI: [10.1038/mi.2015.127](https://doi.org/10.1038/mi.2015.127) · PMID: [26647718](#) · PMCID: [PMC4900951](#)

55. **Understanding Viral dsRNA-Mediated Innate Immune Responses at the Cellular Level Using a Rainbow Trout Model**

Sarah J Poynter, Stephanie J DeWitte-Orr
Frontiers in Immunology (2018-04-23) <https://doi.org/gdhpbs>
DOI: [10.3389/fimmu.2018.00829](https://doi.org/10.3389/fimmu.2018.00829) · PMID: [29740439](#) · PMCID: [PMC5924774](#)

56. **Ultrastructure and Origin of Membrane Vesicles Associated with the Severe Acute Respiratory Syndrome Coronavirus Replication Complex**

Eric J Snijder, Yvonne van der Meer, Jessika Zevenhoven-Dobbe, Jos JM Onderwater, Jannes van der Meulen, Henk K Koerten, AMieke Mommaas
Journal of Virology (2006-06-15) <https://doi.org/b2rh4r>
DOI: [10.1128/jvi.02501-05](https://doi.org/10.1128/jvi.02501-05) · PMID: [16731931](#) · PMCID: [PMC1472606](#)

57. **Molecular immune pathogenesis and diagnosis of COVID-19**

Xiaowei Li, Manman Geng, Yizhao Peng, Liesu Meng, Shemin Lu
Journal of Pharmaceutical Analysis (2020-04) <https://doi.org/ggppqg>
DOI: [10.1016/j.jpha.2020.03.001](https://doi.org/10.1016/j.jpha.2020.03.001) · PMID: [32282863](#) · PMCID: [PMC7104082](#)

58. **Structural basis for translational shutdown and immune evasion by the Nsp1 protein of SARS-CoV-2**

Matthias Thoms, Robert Buschauer, Michael Ameismeier, Lennart Koepke, Timo Denk, Maximilian Hirschenberger, Hanna Kratzat, Manuel Hayn, Timur Mackens-Kiani, Jingdong Cheng, ... Roland Beckmann
Science (2020-09-04) <https://doi.org/gg69nq>
DOI: [10.1126/science.abc8665](https://doi.org/10.1126/science.abc8665) · PMID: [32680882](#) · PMCID: [PMC7402621](#)

59. **Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic**

Asian Pacific Journal of Allergy and Immunology
Allergy, Asthma, and Immunology Association of Thailand (2020)
<https://doi.org/ggpvxw>
DOI: [10.12932/ap-200220-0772](https://doi.org/10.12932/ap-200220-0772) · PMID: [32105090](#)

60. **Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus**

Wenhui Li, Michael J Moore, Natalya Vasilieva, Jianhua Sui, Swee Kee Wong, Michael A Berne, Mohan Somasundaran, John L Sullivan, Katherine Luzuriaga, Thomas C Greenough, ... Michael Farzan
Nature (2003-11) <https://doi.org/bqvjhh>
DOI: [10.1038/nature02145](https://doi.org/10.1038/nature02145) · PMID: [14647384](https://pubmed.ncbi.nlm.nih.gov/14647384/) · PMCID: [PMC7095016](https://pubmed.ncbi.nlm.nih.gov/PMC7095016/)

61. **Efficient Activation of the Severe Acute Respiratory Syndrome Coronavirus Spike Protein by the Transmembrane Protease TMPRSS2**
Shutoku Matsuyama, Noriyo Nagata, Kazuya Shirato, Miyuki Kawase, Makoto Takeda, Fumihiro Taguchi
Journal of Virology (2010-12-15) <https://doi.org/d4hnfr>
DOI: [10.1128/jvi.01542-10](https://doi.org/10.1128/jvi.01542-10) · PMID: [20926566](https://pubmed.ncbi.nlm.nih.gov/20926566/) · PMCID: [PMC3004351](https://pubmed.ncbi.nlm.nih.gov/PMC3004351/)
62. **Evidence that TMPRSS2 Activates the Severe Acute Respiratory Syndrome Coronavirus Spike Protein for Membrane Fusion and Reduces Viral Control by the Humoral Immune Response**
I Glowacka, S Bertram, MA Muller, P Allen, E Soilleux, S Pfefferle, I Steffen, TS Tsegaye, Y He, K Gnirss, ... S Pohlmann
Journal of Virology (2011-02-16) <https://doi.org/bg97wb>
DOI: [10.1128/jvi.02232-10](https://doi.org/10.1128/jvi.02232-10) · PMID: [21325420](https://pubmed.ncbi.nlm.nih.gov/21325420/) · PMCID: [PMC3126222](https://pubmed.ncbi.nlm.nih.gov/PMC3126222/)
63. **Increasing host cellular receptor—angiotensin-converting enzyme 2 expression by coronavirus may facilitate 2019-nCoV (or SARS-CoV-2) infection**
Meng-Wei Zhuang, Yun Cheng, Jing Zhang, Xue-Mei Jiang, Li Wang, Jian Deng, Pei-Hui Wang
Journal of Medical Virology (2020-07-02) <https://doi.org/gg42gn>
DOI: [10.1002/jmv.26139](https://doi.org/10.1002/jmv.26139) · PMID: [32497323](https://pubmed.ncbi.nlm.nih.gov/32497323/) · PMCID: [PMC7300907](https://pubmed.ncbi.nlm.nih.gov/PMC7300907/)
64. **Physiological and pathological regulation of ACE2, the SARS-CoV-2 receptor**
Yanwei Li, Wei Zhou, Li Yang, Ran You
Pharmacological Research (2020-07) <https://doi.org/ggtxhs>
DOI: [10.1016/j.phrs.2020.104833](https://doi.org/10.1016/j.phrs.2020.104833) · PMID: [32302706](https://pubmed.ncbi.nlm.nih.gov/32302706/) · PMCID: [PMC7194807](https://pubmed.ncbi.nlm.nih.gov/PMC7194807/)
65. **Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis**
I Hamming, W Timens, MLC Bulthuis, AT Lely, GJ Navis, H van Goor
The Journal of Pathology (2004-06) <https://doi.org/bhpzc3>
DOI: [10.1002/path.1570](https://doi.org/10.1002/path.1570) · PMID: [15141377](https://pubmed.ncbi.nlm.nih.gov/15141377/)
66. **Association of Antineoplastic Therapy With Decreased SARS-CoV-2 Infection Rates in Patients With Cancer**
Michael B Foote, James Robert White, Justin Jee, Guillem Argilés, Jonathan CM Wan, Benoit Rousseau, Melissa S Pessin, Luis A Diaz Jr
JAMA Oncology (2021-11-01) <https://doi.org/gmmkfw>
DOI: [10.1001/jamaoncol.2021.3585](https://doi.org/10.1001/jamaoncol.2021.3585) · PMID: [34410305](https://pubmed.ncbi.nlm.nih.gov/34410305/) · PMCID: [PMC8377603](https://pubmed.ncbi.nlm.nih.gov/PMC8377603/)
67. **Furin at the cutting edge: From protein traffic to embryogenesis and disease**
Gary Thomas

Nature Reviews Molecular Cell Biology (2002-10)
<https://doi.org/b2n286>
DOI: [10.1038/nrm934](https://doi.org/10.1038/nrm934) · PMID: [12360192](https://pubmed.ncbi.nlm.nih.gov/12360192/) · PMCID: [PMC1964754](https://pubmed.ncbi.nlm.nih.gov/PMC1964754/)

68. **A Multibasic Cleavage Site in the Spike Protein of SARS-CoV-2 Is Essential for Infection of Human Lung Cells**

Markus Hoffmann, Hannah Kleine-Weber, Stefan Pöhlmann
Molecular Cell (2020-05) <https://doi.org/ggt624>
DOI: [10.1016/j.molcel.2020.04.022](https://doi.org/10.1016/j.molcel.2020.04.022) · PMID: [32362314](https://pubmed.ncbi.nlm.nih.gov/32362314/) · PMCID: [PMC7194065](https://pubmed.ncbi.nlm.nih.gov/PMC7194065/)

69. **Detection of SARS-CoV-2 in Different Types of Clinical Specimens**

Wenling Wang, Yanli Xu, Ruqin Gao, Roujian Lu, Kai Han, Guizhen Wu, Wenjie Tan
JAMA (2020-03-11) <https://doi.org/ggpp6h>
DOI: [10.1001/jama.2020.3786](https://doi.org/10.1001/jama.2020.3786) · PMID: [32159775](https://pubmed.ncbi.nlm.nih.gov/32159775/) · PMCID: [PMC7066521](https://pubmed.ncbi.nlm.nih.gov/PMC7066521/)

70. **Epidemiologic Features and Clinical Course of Patients Infected With SARS-CoV-2 in Singapore**

Barnaby Edward Young, Sean Wei Xiang Ong, Shirin Kalimuddin, Jenny G Low, Seow Yen Tan, Jiashen Loh, Oon-Tek Ng, Kalisvar Marimuthu, Li Wei Ang, Tze Minn Mak, ... for the Singapore 2019 Novel Coronavirus Outbreak Research Team
JAMA (2020-04-21) <https://doi.org/ggnb37>
DOI: [10.1001/jama.2020.3204](https://doi.org/10.1001/jama.2020.3204) · PMID: [32125362](https://pubmed.ncbi.nlm.nih.gov/32125362/) · PMCID: [PMC7054855](https://pubmed.ncbi.nlm.nih.gov/PMC7054855/)

71. **Persistence and clearance of viral RNA in 2019 novel coronavirus disease rehabilitation patients**

Yun Ling, Shui-Bao Xu, Yi-Xiao Lin, Di Tian, Zhao-Qin Zhu, Fa-Hui Dai, Fan Wu, Zhi-Gang Song, Wei Huang, Jun Chen, ... Hong-Zhou Lu
Chinese Medical Journal (2020-05-05) <https://doi.org/ggnz8>
DOI: [10.1097/cm9.0000000000000774](https://doi.org/10.1097/cm9.0000000000000774) · PMID: [32118639](https://pubmed.ncbi.nlm.nih.gov/32118639/) · PMCID: [PMC7147278](https://pubmed.ncbi.nlm.nih.gov/PMC7147278/)

72. **Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction**

Thomas Menter, Jasmin D Haslbauer, Ronny Nienhold, Spasenija Savic, Helmut Hopfer, Nikolaus Deigendesch, Stephan Frank, Daniel Turek, Niels Willi, Hans Pargger, ... Alexandar Tzankov
Histopathology (2020-07-05) <https://doi.org/ggwr32>
DOI: [10.1111/his.14134](https://doi.org/10.1111/his.14134) · PMID: [32364264](https://pubmed.ncbi.nlm.nih.gov/32364264/) · PMCID: [PMC7496150](https://pubmed.ncbi.nlm.nih.gov/PMC7496150/)

73. **Association of Cardiac Injury With Mortality in Hospitalized Patients With COVID-19 in Wuhan, China**

Shaobo Shi, Mu Qin, Bo Shen, Yuli Cai, Tao Liu, Fan Yang, Wei Gong, Xu Liu, Jinjun Liang, Qinyan Zhao, ... Congxin Huang
JAMA Cardiology (2020-07-01) <https://doi.org/ggq8qf>
DOI: [10.1001/jamacardio.2020.0950](https://doi.org/10.1001/jamacardio.2020.0950) · PMID: [32211816](https://pubmed.ncbi.nlm.nih.gov/32211816/) · PMCID: [PMC7097841](https://pubmed.ncbi.nlm.nih.gov/PMC7097841/)

74. **The need for urogenital tract monitoring in COVID-19**

Shangqian Wang, Xiang Zhou, Tongtong Zhang, Zengjun Wang
Nature Reviews Urology (2020-04-20) <https://doi.org/ggv4xb>

DOI: [10.1038/s41585-020-0319-7](https://doi.org/10.1038/s41585-020-0319-7) · PMID: [32313110](#) · PMCID:
[PMC7186932](#)

75. **Acute kidney injury in SARS-CoV-2 infected patients**

Vito Fanelli, Marco Fiorentino, Vincenzo Cantaluppi, Loreto Gesualdo, Giovanni Stallone, Claudio Ronco, Giuseppe Castellano
Critical Care (2020-04-16) <https://doi.org/ggy45f>
DOI: [10.1186/s13054-020-02872-z](https://doi.org/10.1186/s13054-020-02872-z) · PMID: [32299479](#) · PMCID:
[PMC7161433](#)

76. **Liver injury in COVID-19: management and challenges**

Chao Zhang, Lei Shi, Fu-Sheng Wang
The Lancet Gastroenterology & Hepatology (2020-05)
<https://doi.org/ggp6x6s>
DOI: [10.1016/s2468-1253\(20\)30057-1](https://doi.org/10.1016/s2468-1253(20)30057-1) · PMID: [32145190](#) · PMCID:
[PMC7129165](#)

77. **Evidence for Gastrointestinal Infection of SARS-CoV-2**

Fei Xiao, Meiwen Tang, Xiaobin Zheng, Ye Liu, Xiaofeng Li, Hong Shan
Gastroenterology (2020-05) <https://doi.org/ggpx27>
DOI: [10.1053/j.gastro.2020.02.055](https://doi.org/10.1053/j.gastro.2020.02.055) · PMID: [32142773](#) · PMCID:
[PMC7130181](#)

78. **2019 Novel coronavirus infection and gastrointestinal tract**

Qin Yan Gao, Ying Xuan Chen, Jing Yuan Fang
Journal of Digestive Diseases (2020-03) <https://doi.org/ggqr86>
DOI: [10.1111/1751-2980.12851](https://doi.org/10.1111/1751-2980.12851) · PMID: [32096611](#) · PMCID:
[PMC7162053](#)

79. **A living WHO guideline on drugs for covid-19**

Arnav Agarwal, Bram Rochwerg, François Lamontagne, Reed AC Siemieniuk, Thomas Agoritsas, Lisa Askie, Lyubov Lytvyn, Yee-Sin Leo, Helen Macdonald, Linan Zeng, ... Per Olav Vandvik
BMJ (2020-09-04) <https://doi.org/ghktgm>
DOI: [10.1136/bmj.m3379](https://doi.org/10.1136/bmj.m3379) · PMID: [32887691](#)

80. **Clinical Characteristics of Coronavirus Disease 2019 in China**

Wei-jie Guan, Zheng-yi Ni, Yu Hu, Wen-hua Liang, Chun-quan Ou, Jian-xing He, Lei Liu, Hong Shan, Chun-liang Lei, David SC Hui, ... Nan-shan Zhong
New England Journal of Medicine (2020-04-30) <https://doi.org/ggm6dh>
DOI: [10.1056/nejmoa2002032](https://doi.org/10.1056/nejmoa2002032) · PMID: [32109013](#) · PMCID: [PMC7092819](#)

81. **Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study**

Fei Zhou, Ting Yu, Ronghui Du, Guohui Fan, Ying Liu, Zhibo Liu, Jie Xiang, Yeming Wang, Bin Song, Xiaoying Gu, ... Bin Cao
The Lancet (2020-03) <https://doi.org/ggnxb3>
DOI: [10.1016/s0140-6736\(20\)30566-3](https://doi.org/10.1016/s0140-6736(20)30566-3)

82. **Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With COVID-19 in the New York City Area**

Safiya Richardson, Jamie S Hirsch, Mangala Narasimhan, James M Crawford, Thomas McGinn, Karina W Davidson, Douglas P Barnaby,

Lance B Becker, John D Chelico, Stuart L Cohen, ... and the Northwell COVID-19 Research Consortium
JAMA (2020-05-26) <https://doi.org/ggsrkd>
DOI: [10.1001/jama.2020.6775](https://doi.org/10.1001/jama.2020.6775) · PMID: [32320003](https://pubmed.ncbi.nlm.nih.gov/32320003/) · PMCID: [PMC7177629](https://pubmed.ncbi.nlm.nih.gov/PMC7177629/)

83. **Clinical Characteristics of Covid-19 in New York City**

Parag Goyal, Justin J Choi, Laura C Pinheiro, Edward J Schenck, Ruijun Chen, Assem Jabri, Michael J Satlin, Thomas R Campion, Musarrat Nahid, Joanna B Ringel, ... Monika M Safford
New England Journal of Medicine (2020-06-11) <https://doi.org/ggtsjc>
DOI: [10.1056/nejmc2010419](https://doi.org/10.1056/nejmc2010419) · PMID: [32302078](https://pubmed.ncbi.nlm.nih.gov/32302078/) · PMCID: [PMC7182018](https://pubmed.ncbi.nlm.nih.gov/PMC7182018/)

84. **Symptom Profiles of a Convenience Sample of Patients with COVID-19 — United States, January–April 2020**

Rachel M Burke, Marie E Killerby, Suzanne Newton, Candace E Ashworth, Abby L Berns, Skyler Brennan, Jonathan M Bressler, Erica Bye, Richard Crawford, Laurel Harduar Morano, ... Case Investigation Form Working Group
MMWR. Morbidity and Mortality Weekly Report (2020-07-17)
<https://doi.org/gg8r2m>
DOI: [10.15585/mmwr.mm6928a2](https://doi.org/10.15585/mmwr.mm6928a2) · PMID: [32673296](https://pubmed.ncbi.nlm.nih.gov/32673296/) · PMCID: [PMC7366851](https://pubmed.ncbi.nlm.nih.gov/PMC7366851/)

85. **Population-scale longitudinal mapping of COVID-19 symptoms, behaviour and testing**

William E Allen, Han Altae-Tran, James Briggs, Xin Jin, Glen McGee, Andy Shi, Rumya Raghavan, Mireille Kamariza, Nicole Nova, Albert Pereta, ... Xihong Lin
Nature Human Behaviour (2020-08-26) <https://doi.org/gg9dfq>
DOI: [10.1038/s41562-020-00944-2](https://doi.org/10.1038/s41562-020-00944-2) · PMID: [32848231](https://pubmed.ncbi.nlm.nih.gov/32848231/) · PMCID: [PMC7501153](https://pubmed.ncbi.nlm.nih.gov/PMC7501153/)

86. **Extrapulmonary manifestations of COVID-19**

Aakriti Gupta, Mahesh V Madhavan, Kartik Sehgal, Nandini Nair, Shiwani Mahajan, Tejasav S Sehrawat, Behnoor Bikdeli, Neha Ahluwalia, John C Ausiello, Elaine Y Wan, ... Donald W Landry
Nature Medicine (2020-07-10) <https://doi.org/gg4r37>
DOI: [10.1038/s41591-020-0968-3](https://doi.org/10.1038/s41591-020-0968-3) · PMID: [32651579](https://pubmed.ncbi.nlm.nih.gov/32651579/)

87. **Acute kidney injury in patients hospitalized with COVID-19**

Jamie S Hirsch, Jia H Ng, Daniel W Ross, Purva Sharma, Hitesh H Shah, Richard L Barnett, Azzour D Hazzan, Steven Fishbane, Kenar D Jhaveri, Mersema Abate, ... Jia Hwei Ng
Kidney International (2020-07) <https://doi.org/ggx24k>
DOI: [10.1016/j.kint.2020.05.006](https://doi.org/10.1016/j.kint.2020.05.006) · PMID: [32416116](https://pubmed.ncbi.nlm.nih.gov/32416116/) · PMCID: [PMC7229463](https://pubmed.ncbi.nlm.nih.gov/PMC7229463/)

88. **COVAN is the new HIVAN: the re-emergence of collapsing glomerulopathy with COVID-19**

Juan Carlos Q Velez, Tiffany Caza, Christopher P Larsen
Nature Reviews Nephrology (2020-08-04) <https://doi.org/gmmfx7>
DOI: [10.1038/s41581-020-0332-3](https://doi.org/10.1038/s41581-020-0332-3) · PMID: [32753739](https://pubmed.ncbi.nlm.nih.gov/32753739/) · PMCID: [PMC7400750](https://pubmed.ncbi.nlm.nih.gov/PMC7400750/)

89. **Nervous system involvement after infection with COVID-19 and other coronaviruses**

Yeshun Wu, Xiaolin Xu, Zijun Chen, Jiahao Duan, Kenji Hashimoto, Ling Yang, Cunming Liu, Chun Yang

Brain, Behavior, and Immunity (2020-07) <https://doi.org/ggg7s2>

DOI: [10.1016/j.bbi.2020.03.031](https://doi.org/10.1016/j.bbi.2020.03.031) · PMID: [32240762](#) · PMCID: [PMC7146689](#)

90. **Neurological associations of COVID-19**

Mark A Ellul, Laura Benjamin, Bhagteshwari Singh, Suzannah Lant, Benedict Daniel Michael, Ava Easton, Rachel Kneen, Sylviane Defres, Jim Sejvar, Tom Solomon

The Lancet Neurology (2020-09) <https://doi.org/d259>

DOI: [10.1016/s1474-4422\(20\)30221-0](https://doi.org/10.1016/s1474-4422(20)30221-0) · PMID: [32622375](#) · PMCID: [PMC7332267](#)

91. **How COVID-19 Affects the Brain**

Maura Boldrini, Peter D Canoll, Robyn S Klein

JAMA Psychiatry (2021-06-01) <https://doi.org/gjj3cd>

DOI: [10.1001/jamapsychiatry.2021.0500](https://doi.org/10.1001/jamapsychiatry.2021.0500) · PMID: [33769431](#)

92. **Update on the neurology of COVID-19**

Josef Finsterer, Claudia Stollberger

Journal of Medical Virology (2020-06-02) <https://doi.org/gg2qnn>

DOI: [10.1002/jmv.26000](https://doi.org/10.1002/jmv.26000) · PMID: [32401352](#) · PMCID: [PMC7272942](#)

93. **COVID-19: A Global Threat to the Nervous System**

Igor J Koralnik, Kenneth L Tyler

Annals of Neurology (2020-06-23) <https://doi.org/gg3hzh>

DOI: [10.1002/ana.25807](https://doi.org/10.1002/ana.25807) · PMID: [32506549](#) · PMCID: [PMC7300753](#)

94. **Olfactory transmucosal SARS-CoV-2 invasion as a port of central nervous system entry in individuals with COVID-19**

Jenny Meinhardt, Josefine Radke, Carsten Dittmayer, Jonas Franz, Carolina Thomas, Ronja Mothes, Michael Laue, Julia Schneider, Sebastian Brünink, Selina Greuel, ... Frank L Heppner

Nature Neuroscience (2020-11-30) <https://doi.org/fk46>

DOI: [10.1038/s41593-020-00758-5](https://doi.org/10.1038/s41593-020-00758-5) · PMID: [33257876](#)

95. **COVID-19 neuropathology at Columbia University Irving Medical Center/New York Presbyterian Hospital**

Kiran T Thakur, Emily Happy Miller, Michael D Glendinning, Osama Al-Dalahmah, Matei A Banu, Amelia K Boehme, Alexandra L Boubour, Samuel S Bruce, Alexander M Chong, Jan Claassen, ... Peter Canoll

Brain (2021-04-15) <https://doi.org/gk72kx>

DOI: [10.1093/brain/awab148](https://doi.org/10.1093/brain/awab148) · PMID: [33856027](#) · PMCID: [PMC8083258](#)

96. **Large-Vessel Stroke as a Presenting Feature of Covid-19 in the Young**

Thomas J Oxley, J Mocco, Shahram Majidi, Christopher P Kellner, Hazem Shoirah, IPaul Singh, Reade A De Leacy, Tomoyoshi Shigematsu, Travis R Ladner, Kurt A Yaeger, ... Johanna T Fifi

New England Journal of Medicine (2020-05-14) <https://doi.org/ggtsjg>

DOI: [10.1056/nejmc2009787](https://doi.org/10.1056/nejmc2009787) · PMID: [32343504](#) · PMCID: [PMC7207073](#)

97. **Incidence of thrombotic complications in critically ill ICU patients with COVID-19**
FA Klok, MJHA Kruip, NJM van der Meer, MS Arbous, DAMPJ Gommers, KM Kant, FHJ Kaptein, J van Paassen, MAM Stals, MV Huisman, H Endeman
Thrombosis Research (2020-07) <https://doi.org/dt2q>
DOI: [10.1016/j.thromres.2020.04.013](https://doi.org/10.1016/j.thromres.2020.04.013) · PMID: [32291094](https://pubmed.ncbi.nlm.nih.gov/32291094/) · PMCID: [PMC7146714](https://pubmed.ncbi.nlm.nih.gov/PMC7146714/)
98. **Coagulopathy and Antiphospholipid Antibodies in Patients with Covid-19**
Yan Zhang, Meng Xiao, Shulan Zhang, Peng Xia, Wei Cao, Wei Jiang, Huan Chen, Xin Ding, Hua Zhao, Hongmin Zhang, ... Shuyang Zhang
New England Journal of Medicine (2020-04-23) <https://doi.org/ggrgz7>
DOI: [10.1056/nejmc2007575](https://doi.org/10.1056/nejmc2007575) · PMID: [32268022](https://pubmed.ncbi.nlm.nih.gov/32268022/) · PMCID: [PMC7161262](https://pubmed.ncbi.nlm.nih.gov/PMC7161262/)
99. **Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia**
Ning Tang, Dengju Li, Xiong Wang, Ziyong Sun
Journal of Thrombosis and Haemostasis (2020-04)
<https://doi.org/ggqx6>
DOI: [10.1111/jth.14768](https://doi.org/10.1111/jth.14768) · PMID: [32073213](https://pubmed.ncbi.nlm.nih.gov/32073213/) · PMCID: [PMC7166509](https://pubmed.ncbi.nlm.nih.gov/PMC7166509/)
100. **Review: Viral infections and mechanisms of thrombosis and bleeding**
M Goeijenbier, M van Wissen, C van de Weg, E Jong, VEA Gerdes, JCM Meijers, DPM Brandjes, ECM van Gorp
Journal of Medical Virology (2012-10) <https://doi.org/f37tfr>
DOI: [10.1002/jmv.23354](https://doi.org/10.1002/jmv.23354) · PMID: [22930518](https://pubmed.ncbi.nlm.nih.gov/22930518/) · PMCID: [PMC7166625](https://pubmed.ncbi.nlm.nih.gov/PMC7166625/)
101. **Immune mechanisms of pulmonary intravascular coagulopathy in COVID-19 pneumonia**
Dennis McGonagle, James S O'Donnell, Kassem Sharif, Paul Emery, Charles Bridgewood
The Lancet Rheumatology (2020-07) <https://doi.org/ggvd74>
DOI: [10.1016/s2665-9913\(20\)30121-1](https://doi.org/10.1016/s2665-9913(20)30121-1) · PMID: [32835247](https://pubmed.ncbi.nlm.nih.gov/32835247/) · PMCID: [PMC7252093](https://pubmed.ncbi.nlm.nih.gov/PMC7252093/)
102. **"War to the knife" against thromboinflammation to protect endothelial function of COVID-19 patients**
Gabriele Guglielmetti, Marco Quaglia, Pier Paolo Sainaghi, Luigi Mario Castello, Rosanna Vaschetto, Mario Pirisi, Francesco Della Corte, Gian Carlo Avanzi, Piero Stratta, Vincenzo Cantaluppi
Critical Care (2020-06-19) <https://doi.org/gg35w7>
DOI: [10.1186/s13054-020-03060-9](https://doi.org/10.1186/s13054-020-03060-9) · PMID: [32560665](https://pubmed.ncbi.nlm.nih.gov/32560665/) · PMCID: [PMC7303575](https://pubmed.ncbi.nlm.nih.gov/PMC7303575/)
103. **COVID-19 update: Covid-19-associated coagulopathy**
Richard C Becker
Journal of Thrombosis and Thrombolysis (2020-05-15)
<https://doi.org/ggwpp5>
DOI: [10.1007/s11239-020-02134-3](https://doi.org/10.1007/s11239-020-02134-3) · PMID: [32415579](https://pubmed.ncbi.nlm.nih.gov/32415579/) · PMCID: [PMC7225095](https://pubmed.ncbi.nlm.nih.gov/PMC7225095/)
104. **The complement system in COVID-19: friend and foe?**

Anuja Java, Anthony J Apicelli, MKathryn Liszewski, Ariella Coler-Reilly, John P Atkinson, Alfred HJ Kim, Hrishikesh S Kulkarni
JCI Insight (2020-08-06) <https://doi.org/gg4b5b>
DOI: [10.1172/jci.insight.140711](https://doi.org/10.1172/jci.insight.140711) · PMID: [32554923](#) · PMCID: [PMC7455060](#)

105. **COVID-19, microangiopathy, hemostatic activation, and complement**
Wen-Chao Song, Garret A FitzGerald
Journal of Clinical Investigation (2020-06-22) <https://doi.org/gg4b5c>
DOI: [10.1172/jci140183](https://doi.org/10.1172/jci140183) · PMID: [32459663](#) · PMCID: [PMC7410042](#)
106. **Post-acute COVID-19 syndrome**
Ani Nalbandian, Kartik Sehgal, Aakriti Gupta, Mahesh V Madhavan, Claire McGroder, Jacob S Stevens, Joshua R Cook, Anna S Nordvig, Daniel Shalev, Tejasav S Sehrawat, ... Elaine Y Wan
Nature Medicine (2021-03-22) <https://doi.org/gjh7b4>
DOI: [10.1038/s41591-021-01283-z](https://doi.org/10.1038/s41591-021-01283-z) · PMID: [33753937](#) · PMCID: [PMC8893149](#)
107. **Six-Month Outcomes in Patients Hospitalized with Severe COVID-19**
Leora I Horwitz, Kira Garry, Alexander M Prete, Sneha Sharma, Felicia Mendoza, Tamara Kahan, Hannah Karpel, Emily Duan, Katherine A Hochman, Himali Weerahandi
Journal of General Internal Medicine (2021-08-05)
<https://doi.org/gmg6wv>
DOI: [10.1007/s11606-021-07032-9](https://doi.org/10.1007/s11606-021-07032-9) · PMID: [34355349](#) · PMCID: [PMC8341831](#)
108. **Preliminary evidence on long COVID in children**
Danilo Buonsenso, Daniel Munblit, Cristina De Rose, Dario Sinatti, Antonia Ricchiuto, Angelo Carfi, Piero Valentini
Acta Paediatrica (2021-04-18) <https://doi.org/gj9qd5>
DOI: [10.1111/apa.15870](https://doi.org/10.1111/apa.15870) · PMID: [33835507](#) · PMCID: [PMC8251440](#)
109. **Characterizing long COVID in an international cohort: 7 months of symptoms and their impact**
Hannah E Davis, Gina S Assaf, Lisa McCorkell, Hannah Wei, Ryan J Low, Yochai Re'em, Signe Redfield, Jared P Austin, Athena Akrami
EClinicalMedicine (2021-08) <https://doi.org/gmbdm7>
DOI: [10.1016/j.eclim.2021.101019](https://doi.org/10.1016/j.eclim.2021.101019) · PMID: [34308300](#) · PMCID: [PMC8280690](#)
110. **6-month consequences of COVID-19 in patients discharged from hospital: a cohort study**
Chaolin Huang, Lixue Huang, Yeming Wang, Xia Li, Lili Ren, Xiaoying Gu, Liang Kang, Li Guo, Min Liu, Xing Zhou, ... Bin Cao
The Lancet (2021-01) <https://doi.org/ghstsk>
DOI: [10.1016/s0140-6736\(20\)32656-8](https://doi.org/10.1016/s0140-6736(20)32656-8) · PMID: [33428867](#) · PMCID: [PMC7833295](#)
111. **Challenges in defining Long COVID: Striking differences across literature, Electronic Health Records, and patient-reported information**

Halie M Rando, Tellen D Bennett, James Brian Byrd, Carolyn Bramante, Tiffany J Callahan, Christopher G Chute, Hannah E Davis, Rachel Deer, Joel Gagnier, Farrukh M Koraishi, ... Melissa A Haendel
Cold Spring Harbor Laboratory (2021-03-26) <https://doi.org/gjk3ng>
DOI: [10.1101/2021.03.20.21253896](https://doi.org/10.1101/2021.03.20.21253896) · PMID: [33791733](#) · PMCID: [PMC8010765](#)

112. **Characterizing Long COVID: Deep Phenotype of a Complex Condition**

Rachel R Deer, Madeline A Rock, Nicole Vasilevsky, Leigh Carmody, Halie Rando, Alfred J Anzalone, Tiffany J Callahan, Carolyn T Bramante, Christopher G Chute, Casey S Greene, ... Peter N Robinson
Cold Spring Harbor Laboratory (2021-06-29) <https://doi.org/gk5nmb>
DOI: [10.1101/2021.06.23.21259416](https://doi.org/10.1101/2021.06.23.21259416)

113. **SARS-CoV-2 infection in primary schools in northern France: A retrospective cohort study in an area of high transmission**

Arnaud Fontanet, Rebecca Grant, Laura Tondeur, Yoann Madec, Ludivine Grzelak, Isabelle Cailleau, Marie-Noëlle Ungeheuer, Charlotte Renaudat, Sandrine Fernandes Pellerin, Lucie Kuhmel, ... Bruno Hoen
Cold Spring Harbor Laboratory (2020-06-29) <https://doi.org/gg87nn>
DOI: [10.1101/2020.06.25.20140178](https://doi.org/10.1101/2020.06.25.20140178)

114. **SARS-CoV-2 Infection in Children**

Xiaoxia Lu, Liqiong Zhang, Hui Du, Jingjing Zhang, Yuan Y Li, Jingyu Qu, Wenxin Zhang, Youjie Wang, Shuangshuang Bao, Ying Li, ... Gary WK Wong
New England Journal of Medicine (2020-04-23) <https://doi.org/ggpt2q>
DOI: [10.1056/nejmc2005073](https://doi.org/10.1056/nejmc2005073) · PMID: [32187458](#) · PMCID: [PMC7121177](#)

115. **Systematic review of COVID-19 in children shows milder cases and a better prognosis than adults**

Jonas F Ludvigsson
Acta Paediatrica (2020-04-14) <https://doi.org/gqq8wr>
DOI: [10.1111/apa.15270](https://doi.org/10.1111/apa.15270) · PMID: [32202343](#) · PMCID: [PMC7228328](#)

116. **Reopening schools during COVID-19**

Ronan Lordan, Garret A FitzGerald, Tilo Grosser
Science (2020-09-03) <https://doi.org/ghsv9p>
DOI: [10.1126/science.abe5765](https://doi.org/10.1126/science.abe5765) · PMID: [32883837](#)

117. **Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection in Children and Adolescents**

Riccardo Castagnoli, Martina Votto, Amelia Licari, Ilaria Brambilla, Raffaele Bruno, Stefano Perlini, Francesca Rovida, Fausto Baldanti, Gian Luigi Marseglia
JAMA Pediatrics (2020-09-01) <https://doi.org/dswz>
DOI: [10.1001/jamapediatrics.2020.1467](https://doi.org/10.1001/jamapediatrics.2020.1467) · PMID: [32320004](#)

118. **Neurologic and Radiographic Findings Associated With COVID-19 Infection in Children**

Omar Abdel-Mannan, Michael Eyre, Ulrike Löbel, Alasdair Bamford, Christin Eltze, Biju Hameed, Cheryl Hemingway, Yael Hacohen
JAMA Neurology (2020-11-01) <https://doi.org/gg339f>

DOI: [10.1001/jamaneurol.2020.2687](https://doi.org/10.1001/jamaneurol.2020.2687) · PMID: [32609336](#) · PMCID: [PMC7330822](#)

119. **Children with Covid-19 in Pediatric Emergency Departments in Italy**

Niccolò Parri, Matteo Lenge, Danilo Buonsenso

New England Journal of Medicine (2020-07-09) <https://doi.org/ggtp6z>

DOI: [10.1056/nejmc2007617](https://doi.org/10.1056/nejmc2007617) · PMID: [32356945](#) · PMCID: [PMC7206930](#)

120. **COVID Data Tracker**

CDC

Centers for Disease Control and Prevention (2020-03-28)

<https://covid.cdc.gov/covid-data-tracker>

121. **Delta variant: What is happening with transmission, hospital admissions, and restrictions?**

Elisabeth Mahase

BMJ (2021-06-15) <https://doi.org/gmkxxq>

DOI: [10.1136/bmj.n1513](https://doi.org/10.1136/bmj.n1513) · PMID: [34130949](#)

122. **The Delta Variant Is Sending More Children to the Hospital. Are They Sicker, Too?**

Emily Anthes

The New York Times (2021-08-09)

<https://www.nytimes.com/2021/08/09/health/coronavirus-children-delta.html>

123. **COVID-19 in 7780 pediatric patients: A systematic review**

Ansel Hoang, Kevin Chorath, Axel Moreira, Mary Evans, Finn

Burmeister-Morton, Fiona Burmeister, Rija Naqvi, Matthew Petershak, Alvaro Moreira

EClinicalMedicine (2020-07) <https://doi.org/gg4hn2>

DOI: [10.1016/j.eclinm.2020.100433](https://doi.org/10.1016/j.eclinm.2020.100433) · PMID: [32766542](#) · PMCID: [PMC7318942](#)

124. **Multi-inflammatory syndrome and Kawasaki disease in children during the COVID-19 pandemic: A nationwide register-based study and time series analysis**

Ulla Koskela, Otto Helve, Emmi Sarvikivi, Merja Helminen, Tea

Nieminen, Ville Peltola, Marjo Renko, Harri Saxén, Hanna Pasma, Tytti Pokka, ... Terhi Tapiainen

Acta Paediatrica (2021-08-04) <https://doi.org/gmdmsr>

DOI: [10.1111/apa.16051](https://doi.org/10.1111/apa.16051) · PMID: [34331326](#) · PMCID: [PMC8444808](#)

125. **Multisystem Inflammatory Syndrome in Children During the Coronavirus 2019 Pandemic: A Case Series**

Kathleen Chiotos, Hamid Bassiri, Edward M Behrens, Allison M Blatz, Joyce Chang, Caroline Diorio, Julie C Fitzgerald, Alexis Topjian, Audrey Rodom John

Journal of the Pediatric Infectious Diseases Society (2020-07)

<https://doi.org/ggx4pd>

DOI: [10.1093/jpids/piaa069](https://doi.org/10.1093/jpids/piaa069) · PMID: [32463092](#) · PMCID: [PMC7313950](#)

126. **Clinical Characteristics of 58 Children With a Pediatric Inflammatory Multisystem Syndrome Temporally Associated With**

SARS-CoV-2

Elizabeth Whittaker, Alasdair Bamford, Julia Kenny, Myrsini Kaforou, Christine E Jones, Priyen Shah, Padmanabhan Ramnarayan, Alain Fraisse, Owen Miller, Patrick Davies, ... for the PIMS-TS Study Group and EUCLIDS and PERFORM Consortia

JAMA (2020-07-21) <https://doi.org/gg2v75>

DOI: [10.1001/jama.2020.10369](https://doi.org/10.1001/jama.2020.10369) · PMID: [32511692](https://pubmed.ncbi.nlm.nih.gov/32511692/) · PMCID: [PMC7281356](https://pubmed.ncbi.nlm.nih.gov/PMC7281356/)

127. **Toxic shock-like syndrome and COVID-19: Multisystem inflammatory syndrome in children (MIS-C)**

Andrea G Greene, Mona Saleh, Eric Roseman, Richard Sinert
The American Journal of Emergency Medicine (2020-11)

<https://doi.org/gg2586>

DOI: [10.1016/j.ajem.2020.05.117](https://doi.org/10.1016/j.ajem.2020.05.117) · PMID: [32532619](https://pubmed.ncbi.nlm.nih.gov/32532619/) · PMCID: [PMC7274960](https://pubmed.ncbi.nlm.nih.gov/PMC7274960/)

128. **Multisystem inflammatory syndrome in children and COVID-19 are distinct presentations of SARS-CoV-2**

Caroline Diorio, Sarah E Henrickson, Laura A Vella, Kevin O McNerney, Julie Chase, Chakkapong Burudpakdee, Jessica H Lee, Cristina Jasen, Fran Balamuth, David M Barrett, ... Hamid Bassiri

Journal of Clinical Investigation (2020-10-05) <https://doi.org/gg7mz2>

DOI: [10.1172/jci140970](https://doi.org/10.1172/jci140970) · PMID: [32730233](https://pubmed.ncbi.nlm.nih.gov/32730233/) · PMCID: [PMC7598044](https://pubmed.ncbi.nlm.nih.gov/PMC7598044/)

129. **The Immunology of Multisystem Inflammatory Syndrome in Children with COVID-19**

Camila Rosat Consiglio, Nicola Cotugno, Fabian Sardh, Christian Pou, Donato Amodio, Lucie Rodriguez, Ziyang Tan, Sonia Zicari, Alessandra Ruggiero, Giuseppe Rubens Pascucci, ... Petter Brodin

Cell (2020-11) <https://doi.org/d8fh>

DOI: [10.1016/j.cell.2020.09.016](https://doi.org/10.1016/j.cell.2020.09.016) · PMID: [32966765](https://pubmed.ncbi.nlm.nih.gov/32966765/) · PMCID: [PMC7474869](https://pubmed.ncbi.nlm.nih.gov/PMC7474869/)

130. **Acute Heart Failure in Multisystem Inflammatory Syndrome in Children in the Context of Global SARS-CoV-2 Pandemic**

Zahra Belhadjer, Mathilde Méot, Fanny Bajolle, Diala Khraiche, Antoine Legendre, Samya Abakka, Johanne Auriau, Marion Grimaud, Mehdi Oualha, Maurice Beghetti, ... Damien Bonnet

Circulation (2020-08-04) <https://doi.org/ggwkv6>

DOI: [10.1161/circulationaha.120.048360](https://doi.org/10.1161/circulationaha.120.048360) · PMID: [32418446](https://pubmed.ncbi.nlm.nih.gov/32418446/)

131. **An adult with Kawasaki-like multisystem inflammatory syndrome associated with COVID-19**

Sheila Shaigany, Marlis Gnrke, Allison Guttmann, Hong Chong, Shane Meehan, Vanessa Raabe, Eddie Louie, Bruce Solitar, Alisa Femia

The Lancet (2020-07) <https://doi.org/gg4sd6>

DOI: [10.1016/s0140-6736\(20\)31526-9](https://doi.org/10.1016/s0140-6736(20)31526-9) · PMID: [32659211](https://pubmed.ncbi.nlm.nih.gov/32659211/) · PMCID: [PMC7351414](https://pubmed.ncbi.nlm.nih.gov/PMC7351414/)

132. **Multisystem inflammatory syndrome in an adult following the SARS-CoV-2 vaccine (MIS-V)**

Arvind Nune, Karthikeyan P Iyengar, Christopher Goddard, Ashar E Ahmed

BMJ Case Reports (2021-07) <https://doi.org/gmdmss>

133. **COVID-19 associated Kawasaki-like multisystem inflammatory disease in an adult**

Sabrina Sokolovsky, Parita Soni, Taryn Hoffman, Philip Kahn, Joshua Scheers-Masters

The American Journal of Emergency Medicine (2021-01)

<https://doi.org/gg5tf4>

DOI: [10.1016/j.ajem.2020.06.053](https://doi.org/10.1016/j.ajem.2020.06.053) · PMID: [32631771](#) · PMCID:
[PMC7315983](#)

134. **Case Report: Adult Post-COVID-19 Multisystem Inflammatory Syndrome and Thrombotic Microangiopathy**

Idris Boudhabhay, Marion Rabant, Lubka T Roumenina, Louis-Marie Coupry, Victoria Poillerat, Armance Marchal, Véronique Frémeaux-Bacchi, Khalil El Karoui, Mehran Monchi, Franck Pourcine

Frontiers in Immunology (2021-06-23) <https://doi.org/gmdmst>

DOI: [10.3389/fimmu.2021.680567](https://doi.org/10.3389/fimmu.2021.680567) · PMID: [34248962](#) · PMCID:
[PMC8260674](#)

135. **Characteristics and Outcomes of US Children and Adolescents With Multisystem Inflammatory Syndrome in Children (MIS-C) Compared With Severe Acute COVID-19**

Leora R Feldstein, Mark W Tenforde, Kevin G Friedman, Margaret Newhams, Erica Billig Rose, Heda Dapul, Vijaya L Soma, Aline B Maddux, Peter M Mourani, Cindy Bowens, ...

JAMA (2021-03-16) <https://doi.org/gh599q>

DOI: [10.1001/jama.2021.2091](https://doi.org/10.1001/jama.2021.2091) · PMID: [33625505](#) · PMCID: [PMC7905703](#)

136. **Preliminary Evidence on Long COVID in children**

Danilo Buonsenso, Daniel Munblit, Cristina De Rose, Dario Sinatti, Antonia Ricchiuto, Angelo Carfi, Piero Valentini

Cold Spring Harbor Laboratory (2021-01-26) <https://doi.org/fv9t>

DOI: [10.1101/2021.01.23.21250375](https://doi.org/10.1101/2021.01.23.21250375)

137. **Pediatric long-COVID: An overlooked phenomenon?**

Caroline LH Brackel, Coen R Lap, Emilie P Buddingh, Marlies A Houten, Linda JTM Sande, Eveline J Langereis, Michiel AGE Bannier, Marielle WH Pijnenburg, Simone Hashimoto, Suzanne WJ Terheggen-Lagro

Pediatric Pulmonology (2021-06-08) <https://doi.org/gkhcc4>

DOI: [10.1002/ppul.25521](https://doi.org/10.1002/ppul.25521) · PMID: [34102037](#) · PMCID: [PMC8242715](#)

138. **Post-acute COVID-19 outcomes in children with mild and asymptomatic disease**

Daniela Say, Nigel Crawford, Sarah McNab, Danielle Wurzel, Andrew Steer, Shidan Tosif

The Lancet Child & Adolescent Health (2021-06) <https://doi.org/gj9p7b>

DOI: [10.1016/s2352-4642\(21\)00124-3](https://doi.org/10.1016/s2352-4642(21)00124-3) · PMID: [33891880](#) · PMCID:
[PMC8057863](#)

139. **Long-term symptoms after SARS-CoV-2 infection in school children: population-based cohort with 6-months follow-up**

Thomas Radtke, Agne Ulyte, Milo A Puhan, Susi Kriemler

Cold Spring Harbor Laboratory (2021-05-18) <https://doi.org/gk78mz>

140. **Long-term Symptoms After SARS-CoV-2 Infection in Children and Adolescents**
Thomas Radtke, Agne Ulyte, Milo A Puhan, Susi Kriemler
JAMA (2021-09-07) <https://doi.org/gmgjmf>
DOI: [10.1001/jama.2021.11880](https://doi.org/10.1001/jama.2021.11880) · PMID: [34264266](https://pubmed.ncbi.nlm.nih.gov/34264266/) · PMCID: [PMC8283661](https://pubmed.ncbi.nlm.nih.gov/PMC8283661/)
141. **Kids and COVID: why young immune systems are still on top**
Smriti Mallapaty
Nature (2021-09-07) <https://doi.org/gmqjd4>
DOI: [10.1038/d41586-021-02423-8](https://doi.org/10.1038/d41586-021-02423-8) · PMID: [34493845](https://pubmed.ncbi.nlm.nih.gov/34493845/)
142. **Dynamic balance of pro- and anti-inflammatory signals controls disease and limits pathology**
Joseph M Cicchese, Stephanie Evans, Caitlin Hult, Louis R Joslyn, Timothy Wessler, Jess A Millar, Simeone Marino, Nicholas A Cifone, Joshua T Mattila, Jennifer J Linderman, Denise E Kirschner
Immunological Reviews (2018-09) <https://doi.org/gd4g4p>
DOI: [10.1111/imr.12671](https://doi.org/10.1111/imr.12671) · PMID: [30129209](https://pubmed.ncbi.nlm.nih.gov/30129209/) · PMCID: [PMC6292442](https://pubmed.ncbi.nlm.nih.gov/PMC6292442/)
143. **Cytokine Dysregulation, Inflammation and Well-Being**
Ilia J Elenkov, Domenic G Iezzoni, Adrian Daly, Alan G Harris, George P Chrousos
Neuroimmunomodulation (2005) <https://doi.org/bsn7kn>
DOI: [10.1159/000087104](https://doi.org/10.1159/000087104) · PMID: [16166805](https://pubmed.ncbi.nlm.nih.gov/16166805/)
144. **Inflammatory responses and inflammation-associated diseases in organs**
Linlin Chen, Huidan Deng, Hengmin Cui, Jing Fang, Zhicai Zuo, Junliang Deng, Yinglun Li, Xun Wang, Ling Zhao
Oncotarget (2017-12-14) <https://doi.org/ggps2p>
DOI: [10.18632/oncotarget.23208](https://doi.org/10.18632/oncotarget.23208) · PMID: [29467962](https://pubmed.ncbi.nlm.nih.gov/29467962/) · PMCID: [PMC5805548](https://pubmed.ncbi.nlm.nih.gov/PMC5805548/)
145. **Molecular biology of the cell**
Bruce Alberts (editor)
Garland Science (2002)
ISBN: 9780815332183
146. **Vander's human physiology: the mechanisms of body function**
Eric P Widmaier, Hershel Raff, Kevin T Strang
McGraw-Hill Higher Education (2008)
ISBN: 9780071283663
147. **The Innate Immune System: Fighting on the Front Lines or Fanning the Flames of COVID-19?**
Julia L McKechnie, Catherine A Blish
Cell Host & Microbe (2020-06) <https://doi.org/gg28pq>
DOI: [10.1016/j.chom.2020.05.009](https://doi.org/10.1016/j.chom.2020.05.009) · PMID: [32464098](https://pubmed.ncbi.nlm.nih.gov/32464098/) · PMCID: [PMC7237895](https://pubmed.ncbi.nlm.nih.gov/PMC7237895/)
148. **Into the Eye of the Cytokine Storm**
JR Tisoncik, MJ Korth, CP Simmons, J Farrar, TR Martin, MG Katze

149. **Inpatient care for septicemia or sepsis: a challenge for patients and hospitals.**
Margaret Jean Hall, Sonja N Williams, Carol J DeFrances, Aleksandr Golosinskiy
NCHS data brief (2011-06)
<https://www.ncbi.nlm.nih.gov/pubmed/22142805>
PMID: [22142805](https://pubmed.ncbi.nlm.nih.gov/22142805/)
150. **Respiratory viral sepsis: epidemiology, pathophysiology, diagnosis and treatment**
Xiaoying Gu, Fei Zhou, Yeming Wang, Guohui Fan, Bin Cao
European Respiratory Review (2020-07-21) <https://doi.org/gg5x69>
DOI: [10.1183/16000617.0038-2020](https://doi.org/10.1183/16000617.0038-2020) · PMID: [32699026](https://pubmed.ncbi.nlm.nih.gov/32699026/)
151. **SARS-CoV-2 and viral sepsis: observations and hypotheses**
Hui Li, Liang Liu, Dingyu Zhang, Jiuyang Xu, Huaping Dai, Nan Tang, Xiao Su, Bin Cao
The Lancet (2020-05) <https://doi.org/ggsps7>
DOI: [10.1016/s0140-6736\(20\)30920-x](https://doi.org/10.1016/s0140-6736(20)30920-x) · PMID: [32311318](https://pubmed.ncbi.nlm.nih.gov/32311318/) · PMCID: [PMC7164875](https://pubmed.ncbi.nlm.nih.gov/PMC7164875/)
152. **Cytokine Balance in the Lungs of Patients with Acute Respiratory Distress Syndrome**
WILLIAM Y PARK, RICHARD B GOODMAN, KENNETH P STEINBERG, JOHN T RUZINSKI, FRANK RADELLA, DAVID R PARK, JEROME PUGIN, SHAWN J SKERRETT, LEONARD D HUDSON, THOMAS R MARTIN
American Journal of Respiratory and Critical Care Medicine (2001-11-15) <https://doi.org/ggqfqz>
DOI: [10.1164/ajrccm.164.10.2104013](https://doi.org/10.1164/ajrccm.164.10.2104013) · PMID: [11734443](https://pubmed.ncbi.nlm.nih.gov/11734443/)
153. **Cytokine release syndrome**
Alexander Shimabukuro-Vornhagen, Philipp Gödel, Marion Subklewe, Hans Joachim Stemmler, Hans Anton Schlößer, Max Schlaak, Matthias Kochanek, Boris Böll, Michael S von Bergwelt-Baildon
Journal for ImmunoTherapy of Cancer (2018-06-15)
<https://doi.org/ghbncj>
DOI: [10.1186/s40425-018-0343-9](https://doi.org/10.1186/s40425-018-0343-9) · PMID: [29907163](https://pubmed.ncbi.nlm.nih.gov/29907163/) · PMCID: [PMC6003181](https://pubmed.ncbi.nlm.nih.gov/PMC6003181/)
154. **Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2⁺ cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS**
L He, Y Ding, Q Zhang, X Che, Y He, H Shen, H Wang, Z Li, L Zhao, J Geng, ... S Jiang
The Journal of Pathology (2006-11) <https://doi.org/bwb8ns>
DOI: [10.1002/path.2067](https://doi.org/10.1002/path.2067) · PMID: [17031779](https://pubmed.ncbi.nlm.nih.gov/17031779/)
155. **Up-regulation of IL-6 and TNF-α induced by SARS-coronavirus spike protein in murine macrophages via NF-κB pathway**
Wei Wang, Linbai Ye, Li Ye, Baozong Li, Bo Gao, Yingchun Zeng, Lingbao Kong, Xiaonan Fang, Hong Zheng, Zhenghui Wu, Yinglong She

156. **COVID-19: consider cytokine storm syndromes and immunosuppression**
Puja Mehta, Daniel F McAuley, Michael Brown, Emilie Sanchez, Rachel S Tattersall, Jessica J Manson
The Lancet (2020-03) <https://doi.org/ggnzmc>
DOI: [10.1016/s0140-6736\(20\)30628-0](https://doi.org/10.1016/s0140-6736(20)30628-0)
157. **Cytokine Storms: Understanding COVID-19**
Nilam Mangalmurti, Christopher A Hunter
Immunity (2020-07) <https://doi.org/gg4fd7>
DOI: [10.1016/j.immuni.2020.06.017](https://doi.org/10.1016/j.immuni.2020.06.017) · PMID: [32610079](https://pubmed.ncbi.nlm.nih.gov/32610079/) · PMCID: [PMC7321048](https://pubmed.ncbi.nlm.nih.gov/32610079/)
158. **Understanding the Inflammatory Cytokine Response in Pneumonia and Sepsis**
John A Kellum
Archives of Internal Medicine (2007-08-13) <https://doi.org/dbxb66>
DOI: [10.1001/archinte.167.15.1655](https://doi.org/10.1001/archinte.167.15.1655) · PMID: [17698689](https://pubmed.ncbi.nlm.nih.gov/17698689/) · PMCID: [PMC4495652](https://pubmed.ncbi.nlm.nih.gov/17698689/)
159. **The pro- and anti-inflammatory properties of the cytokine interleukin-6**
Jürgen Scheller, Athena Chalaris, Dirk Schmidt-Arras, Stefan Rose-John
Biochimica et Biophysica Acta (BBA) - Molecular Cell Research (2011-05) <https://doi.org/cvn4nr>
DOI: [10.1016/j.bbamcr.2011.01.034](https://doi.org/10.1016/j.bbamcr.2011.01.034) · PMID: [21296109](https://pubmed.ncbi.nlm.nih.gov/21296109/)
160. **The Role of Interleukin 6 During Viral Infections**
Lauro Velazquez-Salinas, Antonio Verdugo-Rodriguez, Luis L Rodriguez, Manuel V Borca
Frontiers in Microbiology (2019-05-10) <https://doi.org/ghbnck>
DOI: [10.3389/fmicb.2019.01057](https://doi.org/10.3389/fmicb.2019.01057) · PMID: [31134045](https://pubmed.ncbi.nlm.nih.gov/31134045/) · PMCID: [PMC6524401](https://pubmed.ncbi.nlm.nih.gov/31134045/)
161. **Is a “Cytokine Storm” Relevant to COVID-19?**
Pratik Sinha, Michael A Matthay, Carolyn S Calfee
JAMA Internal Medicine (2020-09-01) <https://doi.org/gg3k6r>
DOI: [10.1001/jamainternmed.2020.3313](https://doi.org/10.1001/jamainternmed.2020.3313) · PMID: [32602883](https://pubmed.ncbi.nlm.nih.gov/32602883/)
162. **Can we use interleukin-6 (IL-6) blockade for coronavirus disease 2019 (COVID-19)-induced cytokine release syndrome (CRS)?**
Bingwen Liu, Min Li, Zhiguang Zhou, Xuan Guan, Yufei Xiang
Journal of Autoimmunity (2020-07) <https://doi.org/ggr79c>
DOI: [10.1016/j.jaut.2020.102452](https://doi.org/10.1016/j.jaut.2020.102452) · PMID: [32291137](https://pubmed.ncbi.nlm.nih.gov/32291137/) · PMCID: [PMC7151347](https://pubmed.ncbi.nlm.nih.gov/32291137/)
163. **A systems approach to infectious disease**
Manon Eckhardt, Judd F Hultquist, Robyn M Kaake, Ruth Hüttenhain, Nevan J Krogan
Nature Reviews Genetics (2020-02-14) <https://doi.org/ggnv63>
DOI: [10.1038/s41576-020-0212-5](https://doi.org/10.1038/s41576-020-0212-5) · PMID: [32060427](https://pubmed.ncbi.nlm.nih.gov/32060427/)

164. **Differential expression of serum/plasma proteins in various infectious diseases: Specific or nonspecific signatures**
Sandipan Ray, Sandip K Patel, Vipin Kumar, Jagruti Damahe, Sanjeeva Srivastava
PROTEOMICS - Clinical Applications (2014-02) <https://doi.org/f2px3h>
DOI: [10.1002/prca.201300074](https://doi.org/10.1002/prca.201300074) · PMID: [24293340](https://pubmed.ncbi.nlm.nih.gov/24293340/)
165. **Imbalanced Host Response to SARS-CoV-2 Drives Development of COVID-19**
Daniel Blanco-Melo, Benjamin E Nilsson-Payant, Wen-Chun Liu, Skyler Uhl, Daisy Hoagland, Rasmus Møller, Tristan X Jordan, Kohei Oishi, Maryline Panis, David Sachs, ... Benjamin R tenOever
Cell (2020-05) <https://doi.org/ggw5tq>
DOI: [10.1016/j.cell.2020.04.026](https://doi.org/10.1016/j.cell.2020.04.026) · PMID: [32416070](https://pubmed.ncbi.nlm.nih.gov/32416070/) · PMCID: [PMC7227586](https://pubmed.ncbi.nlm.nih.gov/PMC7227586/)
166. **Viral tricks to grid-lock the type I interferon system**
Gijs A Versteeg, Adolfo García-Sastre
Current Opinion in Microbiology (2010-08) <https://doi.org/fd7c89>
DOI: [10.1016/j.mib.2010.05.009](https://doi.org/10.1016/j.mib.2010.05.009) · PMID: [20538505](https://pubmed.ncbi.nlm.nih.gov/20538505/) · PMCID: [PMC2920345](https://pubmed.ncbi.nlm.nih.gov/PMC2920345/)
167. **Inhibitors of the Interferon Response Enhance Virus Replication In Vitro**
Claire E Stewart, Richard E Randall, Catherine S Adamson
PLoS ONE (2014-11-12) <https://doi.org/gmf4jb>
DOI: [10.1371/journal.pone.0112014](https://doi.org/10.1371/journal.pone.0112014) · PMID: [25390891](https://pubmed.ncbi.nlm.nih.gov/25390891/) · PMCID: [PMC4229124](https://pubmed.ncbi.nlm.nih.gov/PMC4229124/)
168. **Severe Acute Respiratory Syndrome Coronavirus Open Reading Frame (ORF) 3b, ORF 6, and Nucleocapsid Proteins Function as Interferon Antagonists**
Sarah A Kopecky-Bromberg, Luis Martínez-Sobrido, Matthew Frieman, Ralph A Baric, Peter Palese
Journal of Virology (2007-01-15) <https://doi.org/cdc829>
DOI: [10.1128/jvi.01782-06](https://doi.org/10.1128/jvi.01782-06) · PMID: [17108024](https://pubmed.ncbi.nlm.nih.gov/17108024/) · PMCID: [PMC1797484](https://pubmed.ncbi.nlm.nih.gov/PMC1797484/)
169. **Middle East Respiratory Syndrome Coronavirus Accessory Protein 4a Is a Type I Interferon Antagonist**
Daniela Niemeyer, Thomas Zillinger, Doreen Muth, Florian Zielecki, Gabor Horvath, Tasnim Suliman, Winfried Barchet, Friedemann Weber, Christian Drosten, Marcel A Müller
Journal of Virology (2013-11-15) <https://doi.org/f5d5k7>
DOI: [10.1128/jvi.01845-13](https://doi.org/10.1128/jvi.01845-13) · PMID: [24027320](https://pubmed.ncbi.nlm.nih.gov/24027320/) · PMCID: [PMC3807936](https://pubmed.ncbi.nlm.nih.gov/PMC3807936/)
170. **Increasing Host Cellular Receptor—Angiotensin-Converting Enzyme 2 (ACE2) Expression by Coronavirus may Facilitate 2019-nCoV Infection**
Pei-Hui Wang, Yun Cheng
Cold Spring Harbor Laboratory (2020-02-27) <https://doi.org/ggscwd>
DOI: [10.1101/2020.02.24.963348](https://doi.org/10.1101/2020.02.24.963348)
171. **SARS-CoV-2 nsp13, nsp14, nsp15 and orf6 function as potent interferon antagonists**

Chun-Kit Yuen, Joy-Yan Lam, Wan-Man Wong, Long-Fung Mak, Xiaohui Wang, Hin Chu, Jian-Piao Cai, Dong-Yan Jin, Kelvin Kai-Wang To, Jasper Fuk-Woo Chan, ... Kin-Hang Kok

Emerging Microbes & Infections (2020-01-01) <https://doi.org/gg8msv>
DOI: [10.1080/22221751.2020.1780953](https://doi.org/10.1080/22221751.2020.1780953) · PMID: [32529952](#) · PMCID: [PMC7473193](#)

172. **SARS-CoV-2 ORF3b Is a Potent Interferon Antagonist Whose Activity Is Increased by a Naturally Occurring Elongation Variant**
Yoriyuki Konno, Izumi Kimura, Keiya Uriu, Masaya Fukushi, Takashi Irie, Yoshio Koyanagi, Daniel Sauter, Robert J Gifford, So Nakagawa, Kei Sato
Cell Reports (2020-09) <https://doi.org/ghvf8>
DOI: [10.1016/j.celrep.2020.108185](https://doi.org/10.1016/j.celrep.2020.108185) · PMID: [32941788](#) · PMCID: [PMC7473339](#)

173. **Bulk and single-cell gene expression profiling of SARS-CoV-2 infected human cell lines identifies molecular targets for therapeutic intervention**
Wyler Emanuel, Mösbauer Kirstin, Franke Vedran, Diag Asija, Gottula Lina Theresa, Arsie Roberto, Klironomos Filippos, Koppstein David, Ayoub Salah, Buccitelli Christopher, ... Landthaler Markus
Cold Spring Harbor Laboratory (2020-05-05) <https://doi.org/ggxd2g>
DOI: [10.1101/2020.05.05.079194](https://doi.org/10.1101/2020.05.05.079194)

174. **Isolation and characterization of SARS-CoV-2 from the first US COVID-19 patient**
Jennifer Harcourt, Azaibi Tamin, Xiaoyan Lu, Shifaq Kamili, Senthil Kumar Sakthivel, Janna Murray, Krista Queen, Ying Tao, Clinton R Paden, Jing Zhang, ... Natalie J Thornburg
Cold Spring Harbor Laboratory (2020-03-07) <https://doi.org/gg2fkm>
DOI: [10.1101/2020.03.02.972935](https://doi.org/10.1101/2020.03.02.972935) · PMID: [32511316](#) · PMCID: [PMC7239045](#)

175. **Disease severity-specific neutrophil signatures in blood transcriptomes stratify COVID-19 patients**
Anna C Aschenbrenner, Maria Mouktaroudi, Benjamin Krämer, Marie Oestreich, Nikolaos Antonakos, Melanie Nuesch-Germano, Konstantina Gkizeli, Lorenzo Bonaguro, Nico Reusch, ... Thomas Ulas
Genome Medicine (2021-01-13) <https://doi.org/gk64bx>
DOI: [10.1186/s13073-020-00823-5](https://doi.org/10.1186/s13073-020-00823-5) · PMID: [33441124](#) · PMCID: [PMC7805430](#)

176. **Longitudinal Multi-omics Analyses Identify Responses of Megakaryocytes, Erythroid Cells, and Plasmablasts as Hallmarks of Severe COVID-19**
Joana P Bernardes, Neha Mishra, Florian Tran, Thomas Bahmer, Lena Best, Johanna I Blase, Dora Bordoni, Jeanette Franzenburg, Ulf Geisen, Jonathan Josephs-Spaulding, ... John Ziebuhr
Immunity (2020-12) <https://doi.org/gh2svc>
DOI: [10.1016/j.immuni.2020.11.017](https://doi.org/10.1016/j.immuni.2020.11.017) · PMID: [33296687](#) · PMCID: [PMC7689306](#)

177. **Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients**

Yong Xiong, Yuan Liu, Liu Cao, Dehe Wang, Ming Guo, Ao Jiang, Dong Guo, Wenjia Hu, Jiayi Yang, Zhidong Tang, ... Yu Chen
Emerging Microbes & Infections (2020-01-01) <https://doi.org/gqg79z>
DOI: [10.1080/22221751.2020.1747363](https://doi.org/22221751.2020.1747363) · PMID: [32228226](#) · PMCID: [PMC7170362](#)

178. **Time-resolved systems immunology reveals a late juncture linked to fatal COVID-19**

Can Liu, Andrew J Martins, William W Lau, Nicholas Rachmaninoff, Jinguo Chen, Luisa Imberti, Darius Mostaghimi, Danielle L Fink, Peter D Burbelo, Kerry Dobbs, ... Alessandra Tucci
Cell (2021-04) <https://doi.org/gjq2k5>
DOI: [10.1016/j.cell.2021.02.018](https://doi.org/10.1016/j.cell.2021.02.018) · PMID: [33713619](#) · PMCID: [PMC7874909](#)

179. **High-Density Blood Transcriptomics Reveals Precision Immune Signatures of SARS-CoV-2 Infection in Hospitalized Individuals**

Jeremy W Prokop, Nicholas L Hartog, Dave Chesla, William Faber, Chanise P Love, Rachid Karam, Nelly Abualkheir, Benjamin Feldmann, Li Teng, Tamara McBride, ... Surender Rajasekaran
Frontiers in Immunology (2021-07-16) <https://doi.org/gmgbc>
DOI: [10.3389/fimmu.2021.694243](https://doi.org/10.3389/fimmu.2021.694243) · PMID: [34335605](#) · PMCID: [PMC8322982](#)

180. **Systems biological assessment of immunity to mild versus severe COVID-19 infection in humans**

Prabhu S Arunachalam, Florian Wimmers, Chris Ka Pun Mok, Ranawaka APM Perera, Madeleine Scott, Thomas Hagan, Natalia Sigal, Yupeng Feng, Laurel Bristow, Owen Tak-Yin Tsang, ... Bali Pulendran
Science (2020-09-04) <https://doi.org/gg7vf3>
DOI: [10.1126/science.abc6261](https://doi.org/10.1126/science.abc6261) · PMID: [32788292](#) · PMCID: [PMC7665312](#)

181. **Dynamic innate immune response determines susceptibility to SARS-CoV-2 infection and early replication kinetics**

Nagarjuna R Cheemarla, Timothy A Watkins, Valia T Mihaylova, Bao Wang, Dejian Zhao, Guilin Wang, Marie L Landry, Ellen F Foxman
Journal of Experimental Medicine (2021-06-15) <https://doi.org/gksdmr>
DOI: [10.1084/jem.20210583](https://doi.org/10.1084/jem.20210583) · PMID: [34128960](#) · PMCID: [PMC8210587](#)

182. **Dysregulation of brain and choroid plexus cell types in severe COVID-19**

Andrew C Yang, Fabian Kern, Patricia M Losada, Maayan R Agam, Christina A Maat, Georges P Schmartz, Tobias Fehlmann, Julian A Stein, Nicholas Schaum, Davis P Lee, ... Tony Wyss-Coray
Nature (2021-06-21) <https://doi.org/gmfcvz>
DOI: [10.1038/s41586-021-03710-0](https://doi.org/10.1038/s41586-021-03710-0) · PMID: [34153974](#) · PMCID: [PMC8400927](#)

183. **Proteomics of SARS-CoV-2-infected host cells reveals therapy targets**

Denisa Bojkova, Kevin Klann, Benjamin Koch, Marek Widera, David Krause, Sandra Ciesek, Jindrich Cinatl, Christian Münch
Nature (2020-05-14) <https://doi.org/dw7s>
DOI: [10.1038/s41586-020-2332-7](https://doi.org/10.1038/s41586-020-2332-7) · PMID: [32408336](#)

184. **Potent human neutralizing antibodies elicited by SARS-CoV-2 infection**
Bin Ju, Qi Zhang, Xiangyang Ge, Ruoke Wang, Jiazen Yu, Sisi Shan, Bing Zhou, Shuo Song, Xian Tang, Jinfang Yu, ... Linqi Zhang
Cold Spring Harbor Laboratory (2020-03-26) <https://doi.org/ggp7t4>
DOI: [10.1101/2020.03.21.990770](https://doi.org/10.1101/2020.03.21.990770)
185. **Plasma proteome of severe acute respiratory syndrome analyzed by two-dimensional gel electrophoresis and mass spectrometry**
J-H Chen, Y-W Chang, C-W Yao, T-S Chiueh, S-C Huang, K-Y Chien, A Chen, F-Y Chang, C-H Wong, Y-J Chen
Proceedings of the National Academy of Sciences (2004-11-30)
<https://doi.org/dtv8sx>
DOI: [10.1073/pnas.0407992101](https://doi.org/10.1073/pnas.0407992101) · PMID: [15572443](https://pubmed.ncbi.nlm.nih.gov/15572443/) · PMCID: [PMC535397](https://pubmed.ncbi.nlm.nih.gov/PMC535397/)
186. **Analysis of multimerization of the SARS coronavirus nucleocapsid protein**
Runtao He, Frederick Dobie, Melissa Ballantine, Andrew Leeson, Yan Li, Nathalie Bastien, Todd Cutts, Anton Andonov, Jingxin Cao, Timothy F Booth, ... Xuguang Li
Biochemical and Biophysical Research Communications (2004-04)
<https://doi.org/dbfwr9>
DOI: [10.1016/j.bbrc.2004.02.074](https://doi.org/10.1016/j.bbrc.2004.02.074) · PMID: [15020242](https://pubmed.ncbi.nlm.nih.gov/15020242/) · PMCID: [PMC7111152](https://pubmed.ncbi.nlm.nih.gov/PMC7111152/)
187. **UniProt: a worldwide hub of protein knowledge**
The UniProt Consortium
Nucleic Acids Research (2019-01-08) <https://doi.org/gfwqck>
DOI: [10.1093/nar/gky1049](https://doi.org/10.1093/nar/gky1049) · PMID: [30395287](https://pubmed.ncbi.nlm.nih.gov/30395287/) · PMCID: [PMC6323992](https://pubmed.ncbi.nlm.nih.gov/PMC6323992/)
188. **Home - Genome - NCBI** <https://www.ncbi.nlm.nih.gov/genome>
189. **The Immune Epitope Database (IEDB): 2018 update**
Randi Vita, Swapnil Mahajan, James A Overton, Sandeep Kumar Dhanda, Sheridan Martini, Jason R Cantrell, Daniel K Wheeler, Alessandro Sette, Bjoern Peters
Nucleic Acids Research (2019-01-08) <https://doi.org/gfhz6n>
DOI: [10.1093/nar/gky1006](https://doi.org/10.1093/nar/gky1006) · PMID: [30357391](https://pubmed.ncbi.nlm.nih.gov/30357391/) · PMCID: [PMC6324067](https://pubmed.ncbi.nlm.nih.gov/PMC6324067/)
190. **ViPR: an open bioinformatics database and analysis resource for virology research**
Brett E Pickett, Eva L Sadat, Yun Zhang, Jyothi M Noronha, RBurke Squires, Victoria Hunt, Mengya Liu, Sanjeev Kumar, Sam Zaremba, Zhiping Gu, ... Richard H Scheuermann
Nucleic Acids Research (2012-01) <https://doi.org/c3tds5>
DOI: [10.1093/nar/gkr859](https://doi.org/10.1093/nar/gkr859) · PMID: [22006842](https://pubmed.ncbi.nlm.nih.gov/22006842/) · PMCID: [PMC3245011](https://pubmed.ncbi.nlm.nih.gov/PMC3245011/)
191. **A SARS-CoV-2 protein interaction map reveals targets for drug repurposing**
David E Gordon, Gwendolyn M Jang, Mehdi Bouhaddou, Jiewei Xu, Kirsten Obernier, Kris M White, Matthew J O'Meara, Veronica V Rezelj, Jeffrey Z Guo, Danielle L Swaney, ... Nevan J Krogan
Nature (2020-04-30) <https://doi.org/ggvr6p>
DOI: [10.1038/s41586-020-2286-9](https://doi.org/10.1038/s41586-020-2286-9) · PMID: [32353859](https://pubmed.ncbi.nlm.nih.gov/32353859/) · PMCID: [PMC7431030](https://pubmed.ncbi.nlm.nih.gov/PMC7431030/)

192. **Protein Palmitoylation and Its Role in Bacterial and Viral Infections**
Justyna Sobocińska, Paula Roszczenko-Jasińska, Anna Ciesielska, Katarzyna Kwiatkowska
Frontiers in Immunology (2018-01-19) <https://doi.org/gcxpp2>
DOI: [10.3389/fimmu.2017.02003](https://doi.org/10.3389/fimmu.2017.02003) · PMID: [29403483](https://pubmed.ncbi.nlm.nih.gov/29403483/) · PMCID: [PMC5780409](https://pubmed.ncbi.nlm.nih.gov/PMC5780409/)
193. **Virus-host interactome and proteomic survey of PMBCs from COVID-19 patients reveal potential virulence factors influencing SARS-CoV-2 pathogenesis**
Jingjiao Li, Mingquan Guo, Xiaoxu Tian, Chengrong Liu, Xin Wang, Xing Yang, Ping Wu, Zixuan Xiao, Yafei Qu, Yue Yin, ... Qiming Liang
Cold Spring Harbor Laboratory (2020-04-02) <https://doi.org/ggrgbv>
DOI: [10.1101/2020.03.31.019216](https://doi.org/10.1101/2020.03.31.019216)
194. **The Nuclear Factor NF- B Pathway in Inflammation**
T Lawrence
Cold Spring Harbor Perspectives in Biology (2009-10-07)
<https://doi.org/fptfvp>
DOI: [10.1101/cshperspect.a001651](https://doi.org/10.1101/cshperspect.a001651) · PMID: [20457564](https://pubmed.ncbi.nlm.nih.gov/20457564/) · PMCID: [PMC2882124](https://pubmed.ncbi.nlm.nih.gov/PMC2882124/)
195. **The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) envelope (E) protein harbors a conserved BH3-like sequence**
Vincent Navratil, Loïc Lionnard, Sonia Longhi, JMarie Hardwick, Christophe Combet, Abdel Aouacheria
Cold Spring Harbor Laboratory (2020-06-09) <https://doi.org/ggrp43>
DOI: [10.1101/2020.04.09.033522](https://doi.org/10.1101/2020.04.09.033522)
196. **Large-Scale Multi-omic Analysis of COVID-19 Severity**
Katherine A Overmyer, Evgenia Shishkova, Ian J Miller, Joseph Balnis, Matthew N Bernstein, Trenton M Peters-Clarke, Jesse G Meyer, Qiuwen Quan, Laura K Muehlbauer, Edna A Trujillo, ... Ariel Jaitovich
Cell Systems (2021-01) <https://doi.org/gjkp3p>
DOI: [10.1016/j.cels.2020.10.003](https://doi.org/10.1016/j.cels.2020.10.003) · PMID: [33096026](https://pubmed.ncbi.nlm.nih.gov/33096026/) · PMCID: [PMC7543711](https://pubmed.ncbi.nlm.nih.gov/PMC7543711/)
197. **High-resolution serum proteome trajectories in COVID-19 reveal patient-specific seroconversion**
Philipp E Geyer, Florian M Arend, Sophia Doll, Marie-Luise Louiset, Sebastian Virreira Winter, Johannes B Müller-Reif, Furkan M Torun, Michael Weigand, Peter Eichhorn, Mathias Bruegel, ... Daniel Teupser
EMBO Molecular Medicine (2021-07-07) <https://doi.org/gmgybb>
DOI: [10.15252/emmm.202114167](https://doi.org/10.15252/emmm.202114167) · PMID: [34232570](https://pubmed.ncbi.nlm.nih.gov/34232570/) · PMCID: [PMC8687121](https://pubmed.ncbi.nlm.nih.gov/PMC8687121/)
198. **Structure of SARS Coronavirus Spike Receptor-Binding Domain Complexed with Receptor**
F Li
Science (2005-09-16) <https://doi.org/fww324>
DOI: [10.1126/science.1116480](https://doi.org/10.1126/science.1116480) · PMID: [16166518](https://pubmed.ncbi.nlm.nih.gov/16166518/)
199. **Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2**

Renhong Yan, Yuanyuan Zhang, Yaning Li, Lu Xia, Yingying Guo, Qiang Zhou
Science (2020-03-27) <https://doi.org/ggpvc8>
DOI: [10.1126/science.abb2762](https://doi.org/10.1126/science.abb2762) · PMID: [32132184](https://pubmed.ncbi.nlm.nih.gov/32132184/) · PMCID: [PMC7164635](https://pubmed.ncbi.nlm.nih.gov/PMC7164635/)

200. **Structural basis of receptor recognition by SARS-CoV-2**
Jian Shang, Gang Ye, Ke Shi, Yushun Wan, Chuming Luo, Hideki Aihara, Qibin Geng, Ashley Auerbach, Fang Li
Nature (2020-03-30) <https://doi.org/ggqspv>
DOI: [10.1038/s41586-020-2179-y](https://doi.org/10.1038/s41586-020-2179-y) · PMID: [32225175](https://pubmed.ncbi.nlm.nih.gov/32225175/)
201. **Crystal structure of the 2019-nCoV spike receptor-binding domain bound with the ACE2 receptor**
Jun Lan, Jiwan Ge, Jinfang Yu, Sisi Shan, Huan Zhou, Shilong Fan, Qi Zhang, Xuanling Shi, Qisheng Wang, Linqi Zhang, Xinquan Wang
Cold Spring Harbor Laboratory (2020-02-20) <https://doi.org/ggqzp5>
DOI: [10.1101/2020.02.19.956235](https://doi.org/10.1101/2020.02.19.956235)
202. **Structural and Functional Basis of SARS-CoV-2 Entry by Using Human ACE2**
Qihui Wang, Yanfang Zhang, Lili Wu, Sheng Niu, Chunli Song, Zengyuan Zhang, Guangwen Lu, Chengpeng Qiao, Yu Hu, Kwok-Yung Yuen, ... Jianxun Qi
Cell (2020-05) <https://doi.org/ggr2cz>
DOI: [10.1016/j.cell.2020.03.045](https://doi.org/10.1016/j.cell.2020.03.045) · PMID: [32275855](https://pubmed.ncbi.nlm.nih.gov/32275855/) · PMCID: [PMC7144619](https://pubmed.ncbi.nlm.nih.gov/PMC7144619/)
203. **Receptor Recognition by the Novel Coronavirus from Wuhan: an Analysis Based on Decade-Long Structural Studies of SARS Coronavirus**
Yushun Wan, Jian Shang, Rachel Graham, Ralph S Baric, Fang Li
Journal of Virology (2020-03-17) <https://doi.org/ggjvwn>
DOI: [10.1128/jvi.00127-20](https://doi.org/10.1128/jvi.00127-20) · PMID: [31996437](https://pubmed.ncbi.nlm.nih.gov/31996437/) · PMCID: [PMC7081895](https://pubmed.ncbi.nlm.nih.gov/PMC7081895/)
204. **SARS-CoV-2 and bat RaTG13 spike glycoprotein structures inform on virus evolution and furin-cleavage effects**
Antoni G Wrobel, Donald J Benton, Pengqi Xu, Chloë Roustan, Stephen R Martin, Peter B Rosenthal, John J Skehel, Steven J Gamblin
Nature Structural & Molecular Biology (2020-07-09)
<https://doi.org/gj2gjx>
DOI: [10.1038/s41594-020-0468-7](https://doi.org/10.1038/s41594-020-0468-7) · PMID: [32647346](https://pubmed.ncbi.nlm.nih.gov/32647346/) · PMCID: [PMC7610980](https://pubmed.ncbi.nlm.nih.gov/PMC7610980/)
205. **A pneumonia outbreak associated with a new coronavirus of probable bat origin**
Peng Zhou, Xing-Lou Yang, Xian-Guang Wang, Ben Hu, Lei Zhang, Wei Zhang, Hao-Rui Si, Yan Zhu, Bei Li, Chao-Lin Huang, ... Zheng-Li Shi
Nature (2020-02-03) <https://doi.org/ggj5cg>
DOI: [10.1038/s41586-020-2012-7](https://doi.org/10.1038/s41586-020-2012-7) · PMID: [32015507](https://pubmed.ncbi.nlm.nih.gov/32015507/) · PMCID: [PMC7095418](https://pubmed.ncbi.nlm.nih.gov/PMC7095418/)
206. **Structure of the Hemagglutinin Precursor Cleavage Site, a Determinant of Influenza Pathogenicity and the Origin of the Labile Conformation**

Jue Chen, Kon Ho Lee, David A Steinhauer, David J Stevens, John J Skehel, Don C Wiley
Cell (1998-10) <https://doi.org/bvgh5b>
DOI: [10.1016/s0092-8674\(00\)81771-7](https://doi.org/10.1016/s0092-8674(00)81771-7)

207. **Role of Hemagglutinin Cleavage for the Pathogenicity of Influenza Virus**

David A Steinhauer
Virology (1999-05) <https://doi.org/fw3jz4>
DOI: [10.1006/viro.1999.9716](https://doi.org/10.1006/viro.1999.9716) · PMID: [10329563](#)

208. **Emerging SARS-CoV-2 mutation hot spots include a novel RNA-dependent-RNA polymerase variant**

Maria Pachetti, Bruna Marini, Francesca Benedetti, Fabiola Giudici, Elisabetta Mauro, Paola Storici, Claudio Masciovecchio, Silvia Angeletti, Massimo Ciccozzi, Robert C Gallo, ... Rudy Ippodrino
Journal of Translational Medicine (2020-04-22) <https://doi.org/ggtzrr>
DOI: [10.1186/s12967-020-02344-6](https://doi.org/10.1186/s12967-020-02344-6) · PMID: [32321524](#) · PMCID: [PMC7174922](#)

209. **On the origin and continuing evolution of SARS-CoV-2**

Xiaolu Tang, Changcheng Wu, Xiang Li, Yuhe Song, Xinmin Yao, Xinkai Wu, Yuange Duan, Hong Zhang, Yirong Wang, Zhaojun Qian, ... Jian Lu
National Science Review (2020-06) <https://doi.org/ggndzn>
DOI: [10.1093/nsr/nwaa036](https://doi.org/10.1093/nsr/nwaa036) · PMCID: [PMC7107875](#)

210. **Emergence of genomic diversity and recurrent mutations in SARS-CoV-2**

Lucy van Dorp, Mislav Acman, Damien Richard, Liam P Shaw, Charlotte E Ford, Louise Ormond, Christopher J Owen, Juanita Pang, Cedric CS Tan, Florencia AT Boshier, ... François Balloux
Infection, Genetics and Evolution (2020-09) <https://doi.org/ggvz4h>
DOI: [10.1016/j.meegid.2020.104351](https://doi.org/10.1016/j.meegid.2020.104351) · PMID: [32387564](#) · PMCID: [PMC7199730](#)

211. **Genomic Epidemiology of SARS-CoV-2 in Guangdong Province, China**

Jing Lu, Louis du Plessis, Zhe Liu, Verity Hill, Min Kang, Huifang Lin, Jiufeng Sun, Sarah François, Moritz UG Kraemer, Nuno R Faria, ... Changwen Ke
Cell (2020-05) <https://doi.org/gmgb3b>
DOI: [10.1016/j.cell.2020.04.023](https://doi.org/10.1016/j.cell.2020.04.023) · PMID: [32359424](#) · PMCID: [PMC7192124](#)

212. **An integrated national scale SARS-CoV-2 genomic surveillance network**

The Lancet Microbe
Elsevier BV (2020-07) <https://doi.org/d5mg>
DOI: [10.1016/s2666-5247\(20\)30054-9](https://doi.org/10.1016/s2666-5247(20)30054-9) · PMID: [32835336](#) · PMCID: [PMC7266609](#)

213. **Cases, Data, and Surveillance**

CDC
Centers for Disease Control and Prevention (2020-02-11)
<https://www.cdc.gov/coronavirus/2019-ncov/variants/spheres.html>

214. **Coast-to-Coast Spread of SARS-CoV-2 during the Early Epidemic in the United States**

Joseph R Fauver, Mary E Petrone, Emma B Hodcroft, Kayoko Shioda, Hanna Y Ehrlich, Alexander G Watts, Chantal BF Vogels, Anderson F Brito, Tara Alpert, Anthony Muyombwe, ... Nathan D Grubaugh

Cell (2020-05) <https://doi.org/gg6r9x>

DOI: [10.1016/j.cell.2020.04.021](https://doi.org/10.1016/j.cell.2020.04.021) · PMID: [32386545](https://pubmed.ncbi.nlm.nih.gov/32386545/) · PMCID: [PMC7204677](https://pubmed.ncbi.nlm.nih.gov/PMC7204677/)

215. **Introductions and early spread of SARS-CoV-2 in the New York City area**

Ana S Gonzalez-Reiche, Matthew M Hernandez, Mitchell J Sullivan, Brianne Ciferri, Hala Alshammary, Ajay Obla, Shelcie Fabre, Giulio Kleiner, Jose Polanco, Zenab Khan, ... Harm van Bakel

Science (2020-05-29) <https://doi.org/gg5gy7>

DOI: [10.1126/science.abc1917](https://doi.org/10.1126/science.abc1917) · PMID: [32471856](https://pubmed.ncbi.nlm.nih.gov/32471856/) · PMCID: [PMC7259823](https://pubmed.ncbi.nlm.nih.gov/PMC7259823/)

216. **Spread of SARS-CoV-2 in the Icelandic Population**

Daniel F Gudbjartsson, Agnar Helgason, Hakon Jonsson, Olafur T Magnusson, Pall Melsted, Guðmundur L Norddahl, Jóna Saemundsdóttir, Asgeir Sigurdsson, Patrick Sulem, Arna B Agustsdóttir, ... Kari Stefansson

New England Journal of Medicine (2020-06-11) <https://doi.org/ggr6wx>

DOI: [10.1056/nejmoa2006100](https://doi.org/10.1056/nejmoa2006100) · PMID: [32289214](https://pubmed.ncbi.nlm.nih.gov/32289214/) · PMCID: [PMC7175425](https://pubmed.ncbi.nlm.nih.gov/PMC7175425/)

217. **GISAID - Initiative** <https://www.gisaid.org/>

218. **NCBI SARS-CoV-2 Resources** <https://www.ncbi.nlm.nih.gov/sars-cov-2/>

219. **COVID-19 Data Portal - accelerating scientific research through data** <https://www.covid19dataportal.org/>

220. **Tracking Changes in SARS-CoV-2 Spike: Evidence that D614G Increases Infectivity of the COVID-19 Virus**

Bette Korber, Will M Fischer, Sandrasegaran Gnanakaran, Hyejin Yoon, James Theiler, Werner Abfaltrerer, Nick Hengartner, Elena E Giorgi, Tanmoy Bhattacharya, Brian Foley, ... Matthew D Wyles

Cell (2020-08) <https://doi.org/gg3wqn>

DOI: [10.1016/j.cell.2020.06.043](https://doi.org/10.1016/j.cell.2020.06.043) · PMID: [32697968](https://pubmed.ncbi.nlm.nih.gov/32697968/) · PMCID: [PMC7332439](https://pubmed.ncbi.nlm.nih.gov/PMC7332439/)

221. **Structural and Functional Analysis of the D614G SARS-CoV-2 Spike Protein Variant**

Leonid Yurkovetskiy, Xue Wang, Kristen E Pascal, Christopher Tomkins-Tinch, Thomas P Nyalile, Yetao Wang, Alina Baum, William E Diehl, Ann Dauphin, Claudia Carbone, ... Jeremy Luban

Cell (2020-10) <https://doi.org/ghkt47>

DOI: [10.1016/j.cell.2020.09.032](https://doi.org/10.1016/j.cell.2020.09.032) · PMID: [32991842](https://pubmed.ncbi.nlm.nih.gov/32991842/) · PMCID: [PMC7492024](https://pubmed.ncbi.nlm.nih.gov/PMC7492024/)

222. **Spike mutation D614G alters SARS-CoV-2 fitness**

Jessica A Plante, Yang Liu, Jianying Liu, Hongjie Xia, Bryan A Johnson, Kumari G Lokugamage, Xianwen Zhang, Antonio E Muruato, Jing Zou, Camila R Fontes-Garfias, ... Pei-Yong Shi

Nature (2020-10-26) <https://doi.org/ghht2p>

DOI: [10.1038/s41586-020-2895-3](https://doi.org/10.1038/s41586-020-2895-3) · PMID: [33106671](#) · PMCID: [PMC8158177](#)

223. **Circulating SARS-CoV-2 spike N439K variants maintain fitness while evading antibody-mediated immunity**

Emma C Thomson, Laura E Rosen, James G Shepherd, Roberto Spreafico, Ana da Silva Filipe, Jason A Wojcechowskyj, Chris Davis, Luca Piccoli, David J Pascall, Josh Dillen, ... Gyorgy Snell

Cell (2021-03) <https://doi.org/gkmx9k>

DOI: [10.1016/j.cell.2021.01.037](https://doi.org/10.1016/j.cell.2021.01.037) · PMID: [33621484](#) · PMCID: [PMC7843029](#)

224. **Effects of a major deletion in the SARS-CoV-2 genome on the severity of infection and the inflammatory response: an observational cohort study**

Barnaby E Young, Siew-Wai Fong, Yi-Hao Chan, Tze-Minn Mak, Li Wei Ang, Danielle E Anderson, Cheryl Yi-Pin Lee, Siti Naqiah Amrun, Bennett Lee, Yun Shan Goh, ... Lisa FP Ng

The Lancet (2020-08) <https://doi.org/d6x7>

DOI: [10.1016/s0140-6736\(20\)31757-8](https://doi.org/10.1016/s0140-6736(20)31757-8) · PMID: [32822564](#) · PMCID: [PMC7434477](#)

225. **Identification of Common Deletions in the Spike Protein of Severe Acute Respiratory Syndrome Coronavirus 2**

Zhe Liu, Huanying Zheng, Huirang Lin, Mingyue Li, Runyu Yuan, Jinju Peng, Qianling Xiong, Jiufeng Sun, Baisheng Li, Jie Wu, ... Jing Lu

Journal of Virology (2020-08-17) <https://doi.org/gk7nmx>

DOI: [10.1128/jvi.00790-20](https://doi.org/10.1128/jvi.00790-20) · PMID: [32571797](#) · PMCID: [PMC7431800](#)

226. **The biological and clinical significance of emerging SARS-CoV-2 variants**

Kaiming Tao, Philip L Tzou, Janin Nouhin, Ravindra K Gupta, Tulio de Oliveira, Sergei L Kosakovsky Pond, Daniela Fera, Robert W Shafer

Nature Reviews Genetics (2021-09-17) <https://doi.org/gmvzrr>

DOI: [10.1038/s41576-021-00408-x](https://doi.org/10.1038/s41576-021-00408-x) · PMID: [34535792](#) · PMCID: [PMC8447121](#)

227. **SARS-CoV-2 Variant of Concern 202012/01 Has about Twofold Replicative Advantage and Acquires Concerning Mutations**

Frederic Grabowski, Grzegorz Preibisch, Stanisław Giziński, Marek Kochańczyk, Tomasz Lipniacki

Viruses (2021-03-01) <https://doi.org/gmkjw8>

DOI: [10.3390/v13030392](https://doi.org/10.3390/v13030392) · PMID: [33804556](#) · PMCID: [PMC8000749](#)

228. **Recurrent emergence and transmission of a SARS-CoV-2 Spike deletion H69/V70**

Steven Kemp, William Harvey, Rawlings Datir, Dami Collier, Isabella Ferreira, Bo Meng, Alessandro Carabelii, David L Robertson, Ravindra K Gupta, COVID-19 Genomics UK (COG-UK) consortium

Cold Spring Harbor Laboratory (2021-01-13) <https://doi.org/ghvq45>

DOI: [10.1101/2020.12.14.422555](https://doi.org/10.1101/2020.12.14.422555)

229. **B.1.351 report** https://cov-lineages.org/global_report_B.1.351.html

230. **SARS-CoV-2 B.1.1.7 and B.1.351 spike variants bind human ACE2 with increased affinity**
Muthukumar Ramanathan, Ian D Ferguson, Weili Miao, Paul A Khavari
The Lancet Infectious Diseases (2021-08) <https://doi.org/gsfg>
DOI: [10.1016/s1473-3099\(21\)00262-0](https://doi.org/s1473-3099(21)00262-0) · PMID: [34022142](https://pubmed.ncbi.nlm.nih.gov/34022142/) · PMCID: [PMC8133765](https://pubmed.ncbi.nlm.nih.gov/PMC8133765/)
231. **Evolution, correlation, structural impact and dynamics of emerging SARS-CoV-2 variants**
Austin N Spratt, Saathvik R Kannan, Lucas T Woods, Gary A Weisman, Thomas P Quinn, Christian L Lorson, Anders Sönnnerborg, Siddappa N Byrareddy, Kamal Singh
Computational and Structural Biotechnology Journal (2021) <https://doi.org/gk726x>
DOI: [10.1016/j.csbj.2021.06.037](https://doi.org/j.csbj.2021.06.037) · PMID: [34188776](https://pubmed.ncbi.nlm.nih.gov/34188776/) · PMCID: [PMC8225291](https://pubmed.ncbi.nlm.nih.gov/PMC8225291/)
232. **Transmission of SARS-CoV-2 Lineage B.1.1.7 in England: Insights from linking epidemiological and genetic data**
Erik Volz, Swapnil Mishra, Meera Chand, Jeffrey C Barrett, Robert Johnson, Lily Geidelberg, Wes R Hinsley, Daniel J Laydon, Gavin Dabrera, Áine O'Toole, ... The COVID-19 Genomics UK (COG-UK) consortium
Cold Spring Harbor Laboratory (2021-01-04) <https://doi.org/ghrqv8>
DOI: [10.1101/2020.12.30.20249034](https://doi.org/10.1101/2020.12.30.20249034)
233. **Experimental Evidence for Enhanced Receptor Binding by Rapidly Spreading SARS-CoV-2 Variants**
Charlie Laffeber, Kelly de Koning, Roland Kanaar, Joyce HG Lebbink
Journal of Molecular Biology (2021-07) <https://doi.org/gmkjw2>
DOI: [10.1016/j.jmb.2021.167058](https://doi.org/j.jmb.2021.167058) · PMID: [34023401](https://pubmed.ncbi.nlm.nih.gov/34023401/) · PMCID: [PMC8139174](https://pubmed.ncbi.nlm.nih.gov/PMC8139174/)
234. **The new SARS-CoV-2 strain shows a stronger binding affinity to ACE2 due to N501Y mutant**
Fedaa Ali, Amal Kasry, Muhammed Amin
Medicine in Drug Discovery (2021-06) <https://doi.org/gmk9wv>
DOI: [10.1016/j.medidd.2021.100086](https://doi.org/j.medidd.2021.100086) · PMID: [33681755](https://pubmed.ncbi.nlm.nih.gov/33681755/) · PMCID: [PMC7923861](https://pubmed.ncbi.nlm.nih.gov/PMC7923861/)
235. **SARS-CoV-2 variants B.1.351 and P.1 escape from neutralizing antibodies**
Markus Hoffmann, Prerna Arora, Rüdiger Groß, Alina Seidel, Bojan F Hörnich, Alexander S Hahn, Nadine Krüger, Luise Graichen, Heike Hofmann-Winkler, Amy Kempf, ... Stefan Pöhlmann
Cell (2021-04) <https://doi.org/gjnjzm>
DOI: [10.1016/j.cell.2021.03.036](https://doi.org/j.cell.2021.03.036) · PMID: [33794143](https://pubmed.ncbi.nlm.nih.gov/33794143/) · PMCID: [PMC7980144](https://pubmed.ncbi.nlm.nih.gov/PMC7980144/)
236. **Tracking SARS-CoV-2 variants**
<https://www.who.int/activities/tracking-SARS-CoV-2-variants>
237. **SARS-CoV-2 spike P681R mutation enhances and accelerates viral fusion**

Akatsuki Saito, Hesham Nasser, Keiya Uriu, Yusuke Kosugi, Takashi Irie, Kotaro Shirakawa, Kenji Sadamasu, Izumi Kimura, Jumpei Ito, Jiaqi Wu,

...

Cold Spring Harbor Laboratory (2021-06-17) <https://doi.org/gk7d6w>

DOI: [10.1101/2021.06.17.448820](https://doi.org/10.1101/2021.06.17.448820)

238. **Duration of infectiousness and correlation with RT-PCR cycle threshold values in cases of COVID-19, England, January to May 2020**

Anika Singanayagam, Monika Patel, Andre Charlett, Jamie Lopez Bernal, Vanessa Saliba, Joanna Ellis, Shamez Ladhani, Maria Zambon, Robin Gopal

Eurosurveillance (2020-08-13) <https://doi.org/gg9jt2>

DOI: [10.2807/1560-7917.es.2020.25.32.2001483](https://doi.org/10.2807/1560-7917.es.2020.25.32.2001483) · PMID: [32794447](https://pubmed.ncbi.nlm.nih.gov/32794447/) ·

PMCID: [PMC7427302](https://pubmed.ncbi.nlm.nih.gov/PMC7427302/)

239. **Viral infections acquired indoors through airborne, droplet or contact transmission.**

Giuseppina La Rosa, Marta Fratini, Simonetta Della Libera, Marcello Iaconelli, Michele Muscillo

Annali dell'Istituto superiore di sanità (2013)

<https://www.ncbi.nlm.nih.gov/pubmed/23771256>

DOI: [10.4415/ann_13_02_03](https://doi.org/10.4415/ann_13_02_03) · PMID: [23771256](https://pubmed.ncbi.nlm.nih.gov/23771256/)

240. **Controversy around airborne versus droplet transmission of respiratory viruses**

Eunice YC Shiu, Nancy HL Leung, Benjamin J Cowling

Current Opinion in Infectious Diseases (2019-08)

<https://doi.org/ggbwdb>

DOI: [10.1097/qco.0000000000000563](https://doi.org/10.1097/qco.0000000000000563)

241. **Dismantling myths on the airborne transmission of severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2)**

JW Tang, WP Bahnfleth, PM Bluysen, G Buonanno, JL Jimenez, J Kurnitski, Y Li, S Miller, C Sekhar, L Morawska, ... SJ Dancer

Journal of Hospital Infection (2021-04) <https://doi.org/ghs2qt>

DOI: [10.1016/j.jhin.2020.12.022](https://doi.org/10.1016/j.jhin.2020.12.022) · PMID: [33453351](https://pubmed.ncbi.nlm.nih.gov/33453351/) · PMCID:

[PMC7805396](https://pubmed.ncbi.nlm.nih.gov/PMC7805396/)

242. **How Did We Get Here: What Are Droplets and Aerosols and How Far Do They Go? A Historical Perspective on the Transmission of Respiratory Infectious Diseases**

K Randall, ET Ewing, L Marr, J Jimenez, L Bourouiba

SSRN (2021-04) https://papers.ssrn.com/sol3/papers.cfm?abstract_id=3829873

243. **Transmission of SARS-CoV-2: implications for infection prevention precautions** <https://www.who.int/news-room/commentaries/detail/transmission-of-sars-cov-2-implications-for-infection-prevention-precautions>

244. **Questioning Aerosol Transmission of Influenza**

Camille Lemieux, Gabrielle Brankston, Leah Gitterman, Zahir Hirji, Michael Gardam

Emerging Infectious Diseases (2007-01) <https://doi.org/c2skj8>

245. **Assessing the Dynamics and Control of Droplet- and Aerosol-Transmitted Influenza Using an Indoor Positioning System**

Timo Smieszek, Gianrocco Lazzari, Marcel Salathé

Scientific Reports (2019-02-18) <https://doi.org/ggnqbc>

DOI: [10.1038/s41598-019-38825-y](https://doi.org/10.1038/s41598-019-38825-y) · PMID: [30778136](https://pubmed.ncbi.nlm.nih.gov/30778136/) · PMCID: [PMC6379436](https://pubmed.ncbi.nlm.nih.gov/PMC6379436/)

246. **Influenza A virus transmission via respiratory aerosols or droplets as it relates to pandemic potential**

Mathilde Richard, Ron AM Fouchier

FEMS Microbiology Reviews (2016-01) <https://doi.org/f8cp4h>

DOI: [10.1093/femsre/fuv039](https://doi.org/10.1093/femsre/fuv039) · PMID: [26385895](https://pubmed.ncbi.nlm.nih.gov/26385895/) · PMCID: [PMC5006288](https://pubmed.ncbi.nlm.nih.gov/PMC5006288/)

247. **Coronavirus Pathogenesis**

Susan R Weiss, Julian L Leibowitz

Advances in Virus Research (2011) <https://doi.org/ggyvd7>

DOI: [10.1016/b978-0-12-385885-6.00009-2](https://doi.org/10.1016/b978-0-12-385885-6.00009-2) · PMID: [22094080](https://pubmed.ncbi.nlm.nih.gov/22094080/) · PMCID: [PMC7149603](https://pubmed.ncbi.nlm.nih.gov/PMC7149603/)

248. **SARS and MERS: recent insights into emerging coronaviruses**

Emmie de Wit, Neeltje van Doremalen, Darryl Falzarano, Vincent J Munster

Nature Reviews Microbiology (2016-06-27) <https://doi.org/f8v5cv>

DOI: [10.1038/nrmicro.2016.81](https://doi.org/10.1038/nrmicro.2016.81) · PMID: [27344959](https://pubmed.ncbi.nlm.nih.gov/27344959/) · PMCID: [PMC7097822](https://pubmed.ncbi.nlm.nih.gov/PMC7097822/)

249. **Role of air distribution in SARS transmission during the largest nosocomial outbreak in Hong Kong**

Y Li, X Huang, ITS Yu, TW Wong, H Qian

Indoor Air (2005-04) <https://doi.org/fgp268>

DOI: [10.1111/j.1600-0668.2004.00317.x](https://doi.org/10.1111/j.1600-0668.2004.00317.x) · PMID: [15737151](https://pubmed.ncbi.nlm.nih.gov/15737151/)

250. **Detection of Airborne Severe Acute Respiratory Syndrome (SARS) Coronavirus and Environmental Contamination in SARS Outbreak Units**

Timothy F Booth, Bill Kournikakis, Nathalie Bastien, Jim Ho, Darwyn Kobasa, Laurie Stadnyk, Yan Li, Mel Spence, Shirley Paton, Bonnie Henry, ... Frank Plummer

The Journal of Infectious Diseases (2005-05-01) <https://doi.org/b7z5g6>

DOI: [10.1086/429634](https://doi.org/10.1086/429634) · PMID: [15809906](https://pubmed.ncbi.nlm.nih.gov/15809906/) · PMCID: [PMC7202477](https://pubmed.ncbi.nlm.nih.gov/PMC7202477/)

251. **Role of fomites in SARS transmission during the largest hospital outbreak in Hong Kong**

Shenglan Xiao, Yuguo Li, Tze-wai Wong, David SC Hui

PLOS ONE (2017-07-20) <https://doi.org/gbpgv7>

DOI: [10.1371/journal.pone.0181558](https://doi.org/10.1371/journal.pone.0181558) · PMID: [28727803](https://pubmed.ncbi.nlm.nih.gov/28727803/) · PMCID: [PMC5519164](https://pubmed.ncbi.nlm.nih.gov/PMC5519164/)

252. **Stability of Middle East respiratory syndrome coronavirus (MERS-CoV) under different environmental conditions**

N van Doremalen, T Bushmaker, VJ Munster

Eurosurveillance (2013-09-19) <https://doi.org/ggnnjt>

DOI: [10.2807/1560-7917.es2013.18.38.20590](https://doi.org/10.2807/1560-7917.es2013.18.38.20590) · PMID: [24084338](https://pubmed.ncbi.nlm.nih.gov/24084338/)

253. **MERS coronavirus: diagnostics, epidemiology and transmission**
Ian M Mackay, Katherine E Arden
Virology Journal (2015-12-22) <https://doi.org/f745px>
DOI: [10.1186/s12985-015-0439-5](https://doi.org/10.1186/s12985-015-0439-5) · PMID: [26695637](#) · PMCID: [PMC4687373](#)
254. **Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1**
Neeltje van Doremale, Trenton Bushmaker, Dylan H Morris, Myndi G Holbrook, Amandine Gamble, Brandi N Williamson, Azaibi Tamin, Jennifer L Harcourt, Natalie J Thornburg, Susan I Gerber, ... Vincent J Munster
New England Journal of Medicine (2020-04-16) <https://doi.org/ggn88w>
DOI: [10.1056/nejmc2004973](https://doi.org/10.1056/nejmc2004973) · PMID: [32182409](#) · PMCID: [PMC7121658](#)
255. **Transmission routes of 2019-nCoV and controls in dental practice**
Xian Peng, Xin Xu, Yuqing Li, Lei Cheng, Xuedong Zhou, Biao Ren
International Journal of Oral Science (2020-03-03)
<https://doi.org/ggnf47>
DOI: [10.1038/s41368-020-0075-9](https://doi.org/10.1038/s41368-020-0075-9) · PMID: [32127517](#) · PMCID: [PMC7054527](#)
256. **Airborne Transmission of SARS-CoV-2**
Michael Klompas, Meghan A Baker, Chanu Rhee
JAMA (2020-08-04) <https://doi.org/gg4ttq>
DOI: [10.1001/jama.2020.12458](https://doi.org/10.1001/jama.2020.12458)
257. **Exaggerated risk of transmission of COVID-19 by fomites**
Emanuel Goldman
The Lancet Infectious Diseases (2020-08) <https://doi.org/gg6br7>
DOI: [10.1016/s1473-3099\(20\)30561-2](https://doi.org/10.1016/s1473-3099(20)30561-2) · PMID: [32628907](#) · PMCID: [PMC7333993](#)
258. **Reducing transmission of SARS-CoV-2**
Kimberly A Prather, Chia C Wang, Robert T Schooley
Science (2020-06-26) <https://doi.org/ggxp9w>
DOI: [10.1126/science.abc6197](https://doi.org/10.1126/science.abc6197) · PMID: [32461212](#)
259. **It Is Time to Address Airborne Transmission of Coronavirus Disease 2019 (COVID-19)**
Lidia Morawska, Donald K Milton
Clinical Infectious Diseases (2020-07-06) <https://doi.org/gg34zn>
DOI: [10.1093/cid/ciaa939](https://doi.org/10.1093/cid/ciaa939) · PMID: [32628269](#) · PMCID: [PMC7454469](#)
260. **Ten scientific reasons in support of airborne transmission of SARS-CoV-2**
Trisha Greenhalgh, Jose L Jimenez, Kimberly A Prather, Zeynep Tufekci, David Fisman, Robert Schooley
The Lancet (2021-05) <https://doi.org/gjgmvq>
DOI: [10.1016/s0140-6736\(21\)00869-2](https://doi.org/10.1016/s0140-6736(21)00869-2) · PMID: [33865497](#) · PMCID: [PMC8049599](#)
261. **Aerodynamic analysis of SARS-CoV-2 in two Wuhan hospitals**
Yuan Liu, Zhi Ning, Yu Chen, Ming Guo, Yingle Liu, Nirmal Kumar Gali, Li Sun, Yusen Duan, Jing Cai, Dane Westerdahl, ... Ke Lan

Nature (2020-04-27) <https://doi.org/ggtgng>
DOI: [10.1038/s41586-020-2271-3](https://doi.org/10.1038/s41586-020-2271-3) · PMID: [32340022](#)

262. **Viable SARS-CoV-2 in the air of a hospital room with COVID-19 patients**

John A Lednicky, Michael Lauzardo, ZHugh Fan, Antarpreet Jutla, Trevor B Tilly, Mayank Gangwar, Moiz Usmani, Sripriya Nannu Shankar, Karim Mohamed, Arantza Eiguren-Fernandez, ... Chang-Yu Wu
International Journal of Infectious Diseases (2020-11)
<https://doi.org/ghhkjp>
DOI: [10.1016/j.ijid.2020.09.025](https://doi.org/10.1016/j.ijid.2020.09.025) · PMID: [32949774](#) · PMCID: [PMC7493737](#)

263. **The Infectious Nature of Patient-Generated SARS-CoV-2 Aerosol**

Joshua L Santarpia, Vicki L Herrera, Danielle N Rivera, Shanna Ratnesar-Shumate, StPatrick Reid, Paul W Denton, Jacob WS Martens, Ying Fang, Nicholas Conoan, Michael V Callahan, ... John J Lowe
Cold Spring Harbor Laboratory (2020-07-20) <https://doi.org/gmkxxp>
DOI: [10.1101/2020.07.13.20041632](https://doi.org/10.1101/2020.07.13.20041632)

264. **Aerosol and surface contamination of SARS-CoV-2 observed in quarantine and isolation care**

Joshua L Santarpia, Danielle N Rivera, Vicki L Herrera, MJane Morwitzer, Hannah M Creager, George W Santarpia, Kevin K Crown, David M Brett-Major, Elizabeth R Schnaubelt, MJana Broadhurst, ... John J Lowe
Scientific Reports (2020-07-29) <https://doi.org/gg6wj8>
DOI: [10.1038/s41598-020-69286-3](https://doi.org/10.1038/s41598-020-69286-3) · PMID: [32728118](#) · PMCID: [PMC7391640](#)

265. **Investigating Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Surface and Air Contamination in an Acute Healthcare Setting During the Peak of the Coronavirus Disease 2019 (COVID-19) Pandemic in London**

Jie Zhou, Jonathan A Otter, James R Price, Cristina Cimpeanu, Daniel Meno Garcia, James Kinross, Piers R Boshier, Sam Mason, Frances Bolt, Alison H Holmes, Wendy S Barclay
Clinical Infectious Diseases (2020-07-08) <https://doi.org/gg4fwh>
DOI: [10.1093/cid/ciaa905](https://doi.org/10.1093/cid/ciaa905) · PMID: [32634826](#) · PMCID: [PMC7454437](#)

266. **Airborne transmission of SARS-CoV-2 via aerosols**

Laura Comber, Eamon O Murchu, Linda Drummond, Paul G Carty, Kieran A Walsh, Cillian F De Gascun, Máire A Connolly, Susan M Smith, Michelle O'Neill, Máirín Ryan, Patricia Harrington
Reviews in Medical Virology (2020-10-26) <https://doi.org/gk7fsw>
DOI: [10.1002/rmv.2184](https://doi.org/10.1002/rmv.2184) · PMID: [33105071](#) · PMCID: [PMC7645866](#)

267. **Turbulent Gas Clouds and Respiratory Pathogen Emissions**

Lydia Bourouiba
JAMA (2020-03-26) <https://doi.org/ggqtj4>
DOI: [10.1001/jama.2020.4756](https://doi.org/10.1001/jama.2020.4756) · PMID: [32215590](#)

268. **Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China**

Zhiliang Hu, Ci Song, Chuanjun Xu, Guangfu Jin, Yaling Chen, Xin Xu, Hongxia Ma, Wei Chen, Yuan Lin, Yishan Zheng, ... Hongbing Shen
Science China Life Sciences (2020-03-04) <https://doi.org/dqbn>
DOI: [10.1007/s11427-020-1661-4](https://doi.org/10.1007/s11427-020-1661-4) · PMID: [32146694](https://pubmed.ncbi.nlm.nih.gov/32146694/) · PMCID: [PMC7088568](https://pubmed.ncbi.nlm.nih.gov/PMC7088568/)

269. **Evidence for transmission of COVID-19 prior to symptom onset**

Lauren C Tindale, Jessica E Stockdale, Michelle Coombe, Emma S Garlock, Wing Yin Venus Lau, Manu Saraswat, Louxin Zhang, Dongxuan Chen, Jacco Wallinga, Caroline Colijn
eLife (2020-06-22) <https://doi.org/gg6dtw>
DOI: [10.7554/elife.57149](https://doi.org/10.7554/elife.57149) · PMID: [32568070](https://pubmed.ncbi.nlm.nih.gov/32568070/) · PMCID: [PMC7386904](https://pubmed.ncbi.nlm.nih.gov/PMC7386904/)

270. **Time Kinetics of Viral Clearance and Resolution of Symptoms in Novel Coronavirus Infection**

De Chang, Guoxin Mo, Xin Yuan, Yi Tao, Xiaohua Peng, Fu-Sheng Wang, Lixin Xie, Lokesh Sharma, Charles S Dela Cruz, Enqiang Qin
American Journal of Respiratory and Critical Care Medicine (2020-05-01) <https://doi.org/ggq8xs>
DOI: [10.1164/rccm.202003-0524le](https://doi.org/10.1164/rccm.202003-0524le) · PMID: [32200654](https://pubmed.ncbi.nlm.nih.gov/32200654/) · PMCID: [PMC7193851](https://pubmed.ncbi.nlm.nih.gov/PMC7193851/)

271. **Temporal dynamics in viral shedding and transmissibility of COVID-19**

Xi He, Eric HY Lau, Peng Wu, Xilong Deng, Jian Wang, Xinxin Hao, Yiu Chung Lau, Jessica Y Wong, Yujuan Guan, Xinghua Tan, ... Gabriel M Leung
Nature Medicine (2020-04-15) <https://doi.org/ggr99q>
DOI: [10.1038/s41591-020-0869-5](https://doi.org/10.1038/s41591-020-0869-5) · PMID: [32296168](https://pubmed.ncbi.nlm.nih.gov/32296168/)

272. **COVID-19 and Your Health**

CDC
Centers for Disease Control and Prevention (2020-10-28)
<https://www.cdc.gov/coronavirus/2019-ncov/prevent-getting-sick/how-covid-spreads.html>

273. **Virological assessment of hospitalized patients with COVID-2019**

Roman Wölfel, Victor M Corman, Wolfgang Guggemos, Michael Seilmäier, Sabine Zange, Marcel A Müller, Daniela Niemeyer, Terry C Jones, Patrick Vollmar, Camilla Rothe, ... Clemens Wendtner
Nature (2020-04-01) <https://doi.org/ggqr7>
DOI: [10.1038/s41586-020-2196-x](https://doi.org/10.1038/s41586-020-2196-x) · PMID: [32235945](https://pubmed.ncbi.nlm.nih.gov/32235945/)

274. **Clinical predictors and timing of cessation of viral RNA shedding in patients with COVID-19**

Cristina Corsini Campioli, Edison Cano Cevallos, Mariam Assi, Robin Patel, Matthew J Binnicker, John C O'Horo
Journal of Clinical Virology (2020-09) <https://doi.org/gg7m96>
DOI: [10.1016/j.jcv.2020.104577](https://doi.org/10.1016/j.jcv.2020.104577) · PMID: [32777762](https://pubmed.ncbi.nlm.nih.gov/32777762/) · PMCID: [PMC7405830](https://pubmed.ncbi.nlm.nih.gov/PMC7405830/)

275. **Presymptomatic SARS-CoV-2 Infections and Transmission in a Skilled Nursing Facility**

Melissa M Arons, Kelly M Hatfield, Sujan C Reddy, Anne Kimball, Allison James, Jesica R Jacobs, Joanne Taylor, Kevin Spicer, Ana C Bardossy,

Lisa P Oakley, ... John A Jernigan
New England Journal of Medicine (2020-05-28) <https://doi.org/ggszfg>
DOI: [10.1056/nejmoa2008457](https://doi.org/10.1056/nejmoa2008457) · PMID: [32329971](https://pubmed.ncbi.nlm.nih.gov/32329971/) · PMCID: [PMC7200056](https://pubmed.ncbi.nlm.nih.gov/PMC7200056/)

276. **Prevalence of SARS-CoV-2 Infection in Residents of a Large Homeless Shelter in Boston**
Travis P Baggett, Harrison Keyes, Nora Sporn, Jessie M Gaeta
JAMA (2020-06-02) <https://doi.org/ggtsh3>
DOI: [10.1001/jama.2020.6887](https://doi.org/10.1001/jama.2020.6887) · PMID: [32338732](https://pubmed.ncbi.nlm.nih.gov/32338732/) · PMCID: [PMC7186911](https://pubmed.ncbi.nlm.nih.gov/PMC7186911/)
277. **Presumed Asymptomatic Carrier Transmission of COVID-19**
Yan Bai, Lingsheng Yao, Tao Wei, Fei Tian, Dong-Yan Jin, Lijuan Chen, Meiyun Wang
JAMA (2020-04-14) <https://doi.org/ggmbs8>
DOI: [10.1001/jama.2020.2565](https://doi.org/10.1001/jama.2020.2565) · PMID: [32083643](https://pubmed.ncbi.nlm.nih.gov/32083643/) · PMCID: [PMC7042844](https://pubmed.ncbi.nlm.nih.gov/PMC7042844/)
278. **Transmission of COVID-19 in the terminal stages of the incubation period: A familial cluster**
Peng Li, Ji-Bo Fu, Ke-Feng Li, Jie-Nan Liu, Hong-Ling Wang, Lei-Jie Liu, Yan Chen, Yong-Li Zhang, She-Lan Liu, An Tang, ... Jian-Bo Yan
International Journal of Infectious Diseases (2020-07)
<https://doi.org/ggg844>
DOI: [10.1016/j.ijid.2020.03.027](https://doi.org/10.1016/j.ijid.2020.03.027) · PMID: [32194239](https://pubmed.ncbi.nlm.nih.gov/32194239/) · PMCID: [PMC7264481](https://pubmed.ncbi.nlm.nih.gov/PMC7264481/)
279. **A Cohort of SARS-CoV-2 Infected Asymptomatic and Pre-Symptomatic Contacts from COVID-19 Contact Tracing in Hubei Province, China: Short-Term Outcomes**
Peng Zhang, Fei Tian, Yuan Wan, Jing Cai, Zhengmin Qian, Ran Wu, Yunquan Zhang, Shiyu Zhang, Huan Li, Mingyan Li, ... Hualiang Lin
SSRN Electronic Journal (2020) <https://doi.org/ghf3n2>
DOI: [10.2139/ssrn.3678556](https://doi.org/10.2139/ssrn.3678556)
280. **Estimating the asymptomatic proportion of coronavirus disease 2019 (COVID-19) cases on board the Diamond Princess cruise ship, Yokohama, Japan, 2020**
Kenji Mizumoto, Katsushi Kagaya, Alexander Zarebski, Gerardo Chowell
Eurosurveillance (2020-03-12) <https://doi.org/ggn4bd>
DOI: [10.2807/1560-7917.es.2020.25.10.2000180](https://doi.org/10.2807/1560-7917.es.2020.25.10.2000180) · PMID: [32183930](https://pubmed.ncbi.nlm.nih.gov/32183930/) · PMCID: [PMC7078829](https://pubmed.ncbi.nlm.nih.gov/PMC7078829/)
281. **Estimated prevalence and viral transmissibility in subjects with asymptomatic SARS-CoV-2 infections in Wuhan, China**
Kang Zhang, Weiwei Tong, Xinghuan Wang, Johnson Yiu-Nam Lau
Precision Clinical Medicine (2020-12) <https://doi.org/ghjmks>
DOI: [10.1093/pcmedi/pbaa032](https://doi.org/10.1093/pcmedi/pbaa032) · PMCID: [PMC7499683](https://pubmed.ncbi.nlm.nih.gov/PMC7499683/)
282. **Investigation of SARS-CoV-2 outbreaks in six care homes in London, April 2020**
Shamez N Ladhani, JYimmy Chow, Roshni Janarthanan, Jonathan Fok, Emma Crawley-Boevey, Amoolya Vusirikala, Elena Fernandez, Marina Sanchez Perez, Suzanne Tang, Kate Dun-Campbell, ... Maria Zambon
EClinicalMedicine (2020-09) <https://doi.org/ghbj9v>

DOI: [10.1016/j.eclinm.2020.100533](https://doi.org/10.1016/j.eclinm.2020.100533) · PMID: [32923993](#) · PMCID: [PMC7480335](#)

283. **Clinical and immunological assessment of asymptomatic SARS-CoV-2 infections**

Quan-Xin Long, Xiao-Jun Tang, Qiu-Lin Shi, Qin Li, Hai-Jun Deng, Jun Yuan, Jie-Li Hu, Wei Xu, Yong Zhang, Fa-Jin Lv, ... Ai-Long Huang

Nature Medicine (2020-06-18) <https://doi.org/gg26dx>

DOI: [10.1038/s41591-020-0965-6](https://doi.org/10.1038/s41591-020-0965-6) · PMID: [32555424](#)

284. **Suppression of a SARS-CoV-2 outbreak in the Italian municipality of Vo'**

Enrico Lavezzo, Elisa Franchin, Constanze Ciavarella, Gina Cuomo-Dannenburg, Luisa Barzon, Claudia Del Vecchio, Lucia Rossi, Riccardo Manganelli, Arianna Loreanian, Nicolò Navarin, ... Imperial College COVID-19 Response Team

Nature (2020-06-30) <https://doi.org/gg3w87>

DOI: [10.1038/s41586-020-2488-1](https://doi.org/10.1038/s41586-020-2488-1) · PMID: [32604404](#)

285. **A systematic review and meta-analysis of published research data on COVID-19 infection fatality rates**

Gideon Meyerowitz-Katz, Lea Merone

International Journal of Infectious Diseases (2020-12)

<https://doi.org/ghgjpw>

DOI: [10.1016/j.ijid.2020.09.1464](https://doi.org/10.1016/j.ijid.2020.09.1464) · PMID: [33007452](#) · PMCID: [PMC7524446](#)

286. **Global Covid-19 Case Fatality Rates**

The Centre for Evidence-Based Medicine

<https://www.cebm.net/covid-19/global-covid-19-case-fatality-rates/>

287. **Estimating the Global Infection Fatality Rate of COVID-19**

Richard Grewelle, Giulio De Leo

Cold Spring Harbor Laboratory (2020-05-18) <https://doi.org/ghbvcj>

DOI: [10.1101/2020.05.11.20098780](https://doi.org/10.1101/2020.05.11.20098780)

288. **Repeated cross-sectional sero-monitoring of SARS-CoV-2 in New York City**

Daniel Stadlbauer, Jessica Tan, Kaijun Jiang, Matthew M Hernandez, Shelcie Fabre, Fatima Amanat, Catherine Teo, Guha Asthagiri Arunkumar, Meagan McMahon, Christina Capuano, ... Florian Krammer

Nature (2020-11-03) <https://doi.org/ghhtq9>

DOI: [10.1038/s41586-020-2912-6](https://doi.org/10.1038/s41586-020-2912-6) · PMID: [33142304](#)

289. **What do we know about the risk of dying from COVID-19?**

Our World in Data

<https://ourworldindata.org/covid-mortality-risk>

290. **The concept of R₀ in epidemic theory**

JAP Heesterbeek, K Dietz

Statistica Neerlandica (1996-03) <https://doi.org/d29ch4>

DOI: [10.1111/j.1467-9574.1996.tb01482.x](https://doi.org/10.1111/j.1467-9574.1996.tb01482.x)

291. **Modeling infectious diseases in humans and animals**

Matthew James Keeling, Pejman Rohani

Princeton University Press (2008)

ISBN: 9780691116174

292. **A contribution to the mathematical theory of epidemics**

Proceedings of the Royal Society of London. Series A, Containing Papers of a Mathematical and Physical Character

The Royal Society (1997-01) <https://doi.org/fwx2qw>

DOI: [10.1098/rspa.1927.0118](https://doi.org/rspa.1927.0118)

293. **Population biology of infectious diseases: Part I**

Roy M Anderson, Robert M May

Nature (1979-08-01) <https://doi.org/b6z9hc>

DOI: [10.1038/280361a0](https://doi.org/280361a0) · PMID: [460412](#)

294. **Modeling infectious disease dynamics**

Sarah Cobey

Science (2020-05-15) <https://doi.org/ggsztw>

DOI: [10.1126/science.abb5659](https://doi.org/science.abb5659) · PMID: [32332062](#)

295. **Theoretical ecology: principles and applications**

Robert M May, Angela R McLean (editors)

Oxford University Press (2007)

ISBN: 9780199209989

296. **The approximately universal shapes of epidemic curves in the Susceptible-Exposed-Infectious-Recovered (SEIR) model**

Kevin Heng, Christian L Althaus

Scientific Reports (2020-11-09) <https://doi.org/ghj6mh>

DOI: [10.1038/s41598-020-76563-8](https://doi.org/s41598-020-76563-8) · PMID: [33168932](#) · PMCID: [PMC7653910](#)

297. **Nowcasting and forecasting the potential domestic and international spread of the 2019-nCoV outbreak originating in Wuhan, China: a modelling study**

Joseph T Wu, Kathy Leung, Gabriel M Leung

The Lancet (2020-02) <https://doi.org/ggjvr7>

DOI: [10.1016/s0140-6736\(20\)30260-9](https://doi.org/10.1016/s0140-6736(20)30260-9)

298. **The reproductive number of COVID-19 is higher compared to SARS coronavirus**

Ying Liu, Albert A Gayle, Annelies Wilder-Smith, Joacim Rocklöv

Journal of Travel Medicine (2020-03) <https://doi.org/ggnntv>

DOI: [10.1093/jtm/taaa021](https://doi.org/jtm/taaa021) · PMID: [32052846](#) · PMCID: [PMC7074654](#)

299. **Substantial undocumented infection facilitates the rapid dissemination of novel coronavirus (SARS-CoV-2)**

Ruiyun Li, Sen Pei, Bin Chen, Yimeng Song, Tao Zhang, Wan Yang, Jeffrey Shaman

Science (2020-05-01) <https://doi.org/ggn6c2>

DOI: [10.1126/science.abb3221](https://doi.org/science.abb3221) · PMID: [32179701](#)

300. **Epidemiological parameters of coronavirus disease 2019: a pooled analysis of publicly reported individual data of 1155 cases from seven countries**

Shujuan Ma, Jiayue Zhang, Minyan Zeng, Qingping Yun, Wei Guo, Yixiang Zheng, Shi Zhao, Maggie H Wang, Zuyao Yang
Cold Spring Harbor Laboratory (2020-03-24) <https://doi.org/ggqhzz>
DOI: [10.1101/2020.03.21.20040329](https://doi.org/10.1101/2020.03.21.20040329)

301. **Early Transmissibility Assessment of a Novel Coronavirus in Wuhan, China**

Maimuna Majumder, Kenneth D Mandl
SSRN Electronic Journal (2020) <https://doi.org/ggqhz3>
DOI: [10.2139/ssrn.3524675](https://doi.org/10.2139/ssrn.3524675) · PMID: [32714102](#)

302. **Time-varying transmission dynamics of Novel Coronavirus Pneumonia in China**

Tao Liu, Jianxiong Hu, Jianpeng Xiao, Guanhao He, Min Kang, Zuhua Rong, Lifeng Lin, Haojie Zhong, Qiong Huang, Aiping Deng, ... Wenjun Ma
Cold Spring Harbor Laboratory (2020-02-13) <https://doi.org/dkx9>
DOI: [10.1101/2020.01.25.919787](https://doi.org/10.1101/2020.01.25.919787)

303. **Estimation of the reproductive number of novel coronavirus (COVID-19) and the probable outbreak size on the Diamond Princess cruise ship: A data-driven analysis**

Sheng Zhang, MengYuan Diao, Wenbo Yu, Lei Pei, Zhaofen Lin, Dechang Chen
International Journal of Infectious Diseases (2020-04)
<https://doi.org/ggp56>
DOI: [10.1016/j.ijid.2020.02.033](https://doi.org/10.1016/j.ijid.2020.02.033) · PMID: [32097725](#) · PMCID: [PMC7110591](#)

304. **Estimation of the Transmission Risk of the 2019-nCoV and Its Implication for Public Health Interventions**

Biao Tang, Xia Wang, Qian Li, Nicola Luigi Bragazzi, Sanyi Tang, Yanni Xiao, Jianhong Wu
Journal of Clinical Medicine (2020-02-07) <https://doi.org/ggmkf4>
DOI: [10.3390/jcm9020462](https://doi.org/10.3390/jcm9020462) · PMID: [32046137](#) · PMCID: [PMC7074281](#)

305. **Estimating the effective reproduction number of the 2019-nCoV in China**

Zhidong Cao, Qingpeng Zhang, Xin Lu, Dirk Pfeiffer, Zhongwei Jia, Hongbing Song, Daniel Dajun Zeng
medRxiv (2020-01)
<https://www.medrxiv.org/content/10.1101/2020.01.27.20018952v1>
DOI: [10.1101/2020.01.27.20018952](https://doi.org/10.1101/2020.01.27.20018952)

306. **Modelling the epidemic trend of the 2019 novel coronavirus outbreak in China**

Mingwang Shen, Zhihang Peng, Yanni Xiao, Lei Zhang
Cold Spring Harbor Laboratory (2020-01-25) <https://doi.org/ggqhzw>
DOI: [10.1101/2020.01.23.916726](https://doi.org/10.1101/2020.01.23.916726)

307. **Novel coronavirus 2019-nCoV: early estimation of epidemiological parameters and epidemic predictions**

Jonathan M Read, Jessica RE Bridgen, Derek AT Cummings, Antonia Ho, Chris P Jewell
Cold Spring Harbor Laboratory (2020-01-28) <https://doi.org/dkzb>

DOI: [10.1101/2020.01.23.20018549](https://doi.org/10.1101/2020.01.23.20018549)

308. **Using early data to estimate the actual infection fatality ratio from COVID-19 in France**

Lionel Roques, Etienne Klein, Julien Papaïx, Antoine Sar, Samuel Soubeyrand

Cold Spring Harbor Laboratory (2020-05-07) <https://doi.org/ggqhz2>

DOI: [10.1101/2020.03.22.20040915](https://doi.org/10.1101/2020.03.22.20040915)

309. **Potential Role of Social Distancing in Mitigating Spread of Coronavirus Disease, South Korea**

Sang Woo Park, Kaiyuan Sun, Cécile Viboud, Bryan T Grenfell, Jonathan Dushoff

Emerging Infectious Diseases (2020-11) <https://doi.org/gmv3sn>

DOI: [10.3201/eid2611.201099](https://doi.org/10.3201/eid2611.201099) · PMID: [32795385](https://pubmed.ncbi.nlm.nih.gov/32795385/) · PMCID: [PMC7588540](https://pubmed.ncbi.nlm.nih.gov/PMC7588540/)

310. **Early dynamics of transmission and control of COVID-19: a mathematical modelling study**

Adam J Kucharski, Timothy W Russell, Charlie Diamond, Yang Liu, John Edmunds, Sebastian Funk, Rosalind M Eggo, Fiona Sun, Mark Jit, James D Munday, ... Stefan Flasche

The Lancet Infectious Diseases (2020-05) <https://doi.org/ggptcf>

DOI: [10.1016/s1473-3099\(20\)30144-4](https://doi.org/10.1016/s1473-3099(20)30144-4)

311. **Estimating the reproduction number of COVID-19 in Iran using epidemic modeling**

Ebrahim Sahafizadeh, Samaneh Sartoli

Cold Spring Harbor Laboratory (2020-04-23) <https://doi.org/ggqhzhx>

DOI: [10.1101/2020.03.20.20038422](https://doi.org/10.1101/2020.03.20.20038422)

312. **Report 13: Estimating the number of infections and the impact of non-pharmaceutical interventions on COVID-19 in 11 European countries**

S Flaxman, S Mishra, A Gandy, H Unwin, H Coupland, T Mellan, H Zhu, T Berah, J Eaton, P Perez Guzman, ... S Bhatt

Imperial College London (2020-03-30) <https://doi.org/ggrbmf>

DOI: [10.2561/77731](https://doi.org/10.2561/77731)

313. **Increased transmissibility and global spread of SARS-CoV-2 variants of concern as at June 2021**

Finlay Campbell, Brett Archer, Henry Laurenson-Schafer, Yuka Jinnai, Franck Konings, Neale Batra, Boris Pavlin, Katrijn Vandemaele, Maria D Van Kerkhove, Thibaut Jombart, ... Olivier le Polain de Waroux

Eurosurveillance (2021-06-17) <https://doi.org/gmkjw6>

DOI: [10.2807/1560-7917.es.2021.26.24.2100509](https://doi.org/10.2807/1560-7917.es.2021.26.24.2100509) · PMID: [34142653](https://pubmed.ncbi.nlm.nih.gov/34142653/) ·

PMCID: [PMC8212592](https://pubmed.ncbi.nlm.nih.gov/PMC8212592/)

314. **Genomic epidemiology identifies emergence and rapid transmission of SARS-CoV-2 B.1.1.7 in the United States**

Nicole L Washington, Karthik Gangavarapu, Mark Zeller, Alexandre Bolze, Elizabeth T Cirulli, Kelly MSchiabor Barrett, Brendan B Larsen, Catelyn Anderson, Simon White, Tyler Cassens, ... Kristian G Andersen

Cold Spring Harbor Laboratory (2021-02-07) <https://doi.org/gh598v>

DOI: [10.1101/2021.02.06.21251159](https://doi.org/10.1101/2021.02.06.21251159) · PMID: [33564780](https://pubmed.ncbi.nlm.nih.gov/33564780/) · PMCID:

[PMC7872373](https://pubmed.ncbi.nlm.nih.gov/PMC7872373/)

315. **Estimated transmissibility and impact of SARS-CoV-2 lineage B.1.1.7 in England**
Nicholas G Davies, Sam Abbott, Rosanna C Barnard, Christopher I Jarvis, Adam J Kucharski, James D Munday, Carl AB Pearson, Timothy W Russell, Damien C Tully, Alex D Washburne, ...
Science (2021-04-09) <https://doi.org/gh6x68>
DOI: [10.1126/science.abg3055](https://doi.org/10.1126/science.abg3055) · PMID: [33658326](https://pubmed.ncbi.nlm.nih.gov/33658326/) · PMCID: [PMC8128288](https://pubmed.ncbi.nlm.nih.gov/PMC8128288/)
316. **The reproductive number of the Delta variant of SARS-CoV-2 is far higher compared to the ancestral SARS-CoV-2 virus**
Ying Liu, Joacim Rocklöv
Journal of Travel Medicine (2021-08-09) <https://doi.org/gmkjw4>
DOI: [10.1093/jtm/taab124](https://doi.org/10.1093/jtm/taab124) · PMID: [34369565](https://pubmed.ncbi.nlm.nih.gov/34369565/) · PMCID: [PMC8436367](https://pubmed.ncbi.nlm.nih.gov/PMC8436367/)
317. **Predicted dominance of variant Delta of SARS-CoV-2 before Tokyo Olympic Games, Japan, July 2021**
Kimihiro Ito, Chayada Piantham, Hiroshi Nishiura
Eurosurveillance (2021-07-08) <https://doi.org/gmkjw7>
DOI: [10.2807/1560-7917.es.2021.26.27.2100570](https://doi.org/10.2807/1560-7917.es.2021.26.27.2100570) · PMID: [34240695](https://pubmed.ncbi.nlm.nih.gov/34240695/) · PMCID: [PMC8268651](https://pubmed.ncbi.nlm.nih.gov/PMC8268651/)
318. **Projecting hospital utilization during the COVID-19 outbreaks in the United States**
Seyed M Moghadas, Affan Shoukat, Meagan C Fitzpatrick, Chad R Wells, Pratha Sah, Abhishek Pandey, Jeffrey D Sachs, Zheng Wang, Lauren A Meyers, Burton H Singer, Alison P Galvani
Proceedings of the National Academy of Sciences (2020-04-21) <https://doi.org/ggg7jc>
DOI: [10.1073/pnas.2004064117](https://doi.org/10.1073/pnas.2004064117) · PMID: [32245814](https://pubmed.ncbi.nlm.nih.gov/32245814/) · PMCID: [PMC7183199](https://pubmed.ncbi.nlm.nih.gov/PMC7183199/)
319. **The effect of control strategies to reduce social mixing on outcomes of the COVID-19 epidemic in Wuhan, China: a modelling study**
Kiesha Prem, Yang Liu, Timothy W Russell, Adam J Kucharski, Rosalind M Eggo, Nicholas Davies, Mark Jit, Petra Klepac, Stefan Flasche, Samuel Clifford, ... Joel Hellewell
The Lancet Public Health (2020-05) <https://doi.org/ggp3xq>
DOI: [10.1016/s2468-2667\(20\)30073-6](https://doi.org/10.1016/s2468-2667(20)30073-6) · PMID: [32220655](https://pubmed.ncbi.nlm.nih.gov/32220655/) · PMCID: [PMC7158905](https://pubmed.ncbi.nlm.nih.gov/PMC7158905/)
320. **Spread and dynamics of the COVID-19 epidemic in Italy: Effects of emergency containment measures**
Marino Gatto, Enrico Bertuzzo, Lorenzo Mari, Stefano Miccoli, Luca Carraro, Renato Casagrandi, Andrea Rinaldo
Proceedings of the National Academy of Sciences (2020-05-12) <https://doi.org/ggv4j6>
DOI: [10.1073/pnas.2004978117](https://doi.org/10.1073/pnas.2004978117) · PMID: [32327608](https://pubmed.ncbi.nlm.nih.gov/32327608/) · PMCID: [PMC7229754](https://pubmed.ncbi.nlm.nih.gov/PMC7229754/)
321. **Covid-19: Temporal variation in transmission during the COVID-19 outbreak**
EpiForecasts and the CMMID Covid working group
<https://epiforecasts.io/covid/>

322. **Rt COVID-19**
Kevin Systrom, Thomas Vladeck, Mike Krieger
<https://rt.live/>
323. **Epidemiology, transmission dynamics and control of SARS: the 2002–2003 epidemic**
Roy M Anderson, Christophe Fraser, Azra C Ghani, Christl A Donnelly, Steven Riley, Neil M Ferguson, Gabriel M Leung, TH Lam, Anthony J Hedley
Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences (2004-07-29) <https://doi.org/c2n646>
DOI: [10.1098/rstb.2004.1490](https://doi.org/rstb.2004.1490) · PMID: [15306395](#) · PMCID: [PMC1693389](#)
324. **A timeline of the CDC's advice on face masks**
Los Angeles Times
(2021-07-27) <https://www.latimes.com/science/story/2021-07-27/timeline-cdc-mask-guidance-during-covid-19-pandemic>
325. **Social Factors Influencing COVID-19 Exposure and Outcomes**
COVID-19 Review Consortium
Manubot (2021-04-30) <https://greenelab.github.io/covid19-review/v/32afa309f69f0466a91acec5d0df3151fe4d61b5/#social-factors-influencing-covid-19-exposure-and-outcomes>
326. **Three Emerging Coronaviruses in Two Decades**
Jeannette Guarner
American Journal of Clinical Pathology (2020-04)
<https://doi.org/ggppq3>
DOI: [10.1093/ajcp/aqaa029](https://doi.org/ajcp/aqaa029) · PMID: [32053148](#) · PMCID: [PMC7109697](#)
327. **Evolutionary and Genomic Analysis of SARS-CoV-2**
COVID-19 Review Consortium
Manubot (2021-03-30) <https://greenelab.github.io/covid19-review/v/910dd7b7479f5336a1c911c57446829bef015dbe/#evolutionary-and-genomic-analysis-of-sars-cov-2>
328. **Vaccine Development Strategies for SARS-CoV-2**
COVID-19 Review Consortium
Manubot (2021-02-19) <https://greenelab.github.io/covid19-review/v/d9d90fd7e88ef547fb4cb0ef73baef5fee7fb5/#vaccine-development-strategies-for-sars-cov-2>
329. **Origin and evolution of pathogenic coronaviruses**
Jie Cui, Fang Li, Zheng-Li Shi
Nature Reviews Microbiology (2018-12-10) <https://doi.org/ggh4vb>
DOI: [10.1038/s41579-018-0118-9](https://doi.org/s41579-018-0118-9) · PMID: [30531947](#) · PMCID: [PMC7097006](#)
330. **Human Coronaviruses: A Review of Virus–Host Interactions**
Yvonne Lim, Yan Ng, James Tam, Ding Liu
Diseases (2016-07-25) <https://doi.org/ggjs23>
DOI: [10.3390/diseases4030026](https://doi.org/10.3390/diseases4030026) · PMID: [28933406](#) · PMCID: [PMC5456285](#)
331. **Human coronavirus circulation in the United States 2014–2017**

Marie E Killerby, Holly M Biggs, Amber Haynes, Rebecca M Dahl,
Desiree Mustaqim, Susan I Gerber, John T Watson
Journal of Clinical Virology (2018-04) <https://doi.org/gc7sf3>
DOI: [10.1016/j.jcv.2018.01.019](https://doi.org/10.1016/j.jcv.2018.01.019) · PMID: [29427907](#) · PMCID: [PMC7106380](#)

332. **A New Virus Isolated from the Human Respiratory Tract.**
D Hamre, JJ Procknow
Experimental Biology and Medicine (1966-01-01) <https://doi.org/gg84fc>
DOI: [10.3181/00379727-121-30734](https://doi.org/10.3181/00379727-121-30734) · PMID: [4285768](#)
333. **Recovery in tracheal organ cultures of novel viruses from patients with respiratory disease.**
K McIntosh, JH Dees, WB Becker, AZ Kapikian, RM Chanock
Proceedings of the National Academy of Sciences (1967-04-01)
<https://doi.org/bhfd6w>
DOI: [10.1073/pnas.57.4.933](https://doi.org/10.1073/pnas.57.4.933) · PMID: [5231356](#) · PMCID: [PMC224637](#)
334. **:(unav)**
Krzysztof Pyrc, Maarten F Jebbink, Ben Berkhout, Lia van der Hoek
Virology Journal (2004) <https://doi.org/dnmj8m>
DOI: [10.1186/1743-422x-1-7](https://doi.org/10.1186/1743-422x-1-7) · PMID: [15548333](#) · PMCID: [PMC538260](#)
335. **Understanding Human Coronavirus HCoV-NL63.**
Sahar Abdul-Rasool, Burtram C Fielding
The open virology journal (2010-05-25)
<https://www.ncbi.nlm.nih.gov/pubmed/20700397>
DOI: [10.2174/1874357901004010076](https://doi.org/10.2174/1874357901004010076) · PMID: [20700397](#) · PMCID:
[PMC2918871](#)
336. **Coronaviruses and gastrointestinal diseases**
Xi Luo, Guan-Zhou Zhou, Yan Zhang, Li-Hua Peng, Li-Ping Zou, Yun-Sheng Yang
Military Medical Research (2020-10-14) <https://doi.org/ghqfmj>
DOI: [10.1186/s40779-020-00279-z](https://doi.org/10.1186/s40779-020-00279-z) · PMID: [33054860](#) · PMCID:
[PMC7556584](#)
337. **Human (non-severe acute respiratory syndrome) coronavirus infections in hospitalised children in France**
Astrid Vabret, Julia Dina, Stéphanie Gouarin, Joëlle Petitjean, Valérie Tripey, Jacques Brouard, François Freymuth
Journal of Paediatrics and Child Health (2008-04) <https://doi.org/cxt434>
DOI: [10.1111/j.1440-1754.2007.01246.x](https://doi.org/10.1111/j.1440-1754.2007.01246.x) · PMID: [17999671](#) · PMCID:
[PMC7166728](#)
338. **Identification of a new human coronavirus**
Lia van der Hoek, Krzysztof Pyrc, Maarten F Jebbink, Wilma Vermeulen-Oost, Ron JM Berkhout, Katja C Wolthers, Pauline ME Wertheim-van Dillen, Jos Kaandorp, Joke Spaargaren, Ben Berkhout
Nature Medicine (2004-03-21) <https://doi.org/b5wtsn>
DOI: [10.1038/nm1024](https://doi.org/10.1038/nm1024) · PMID: [15034574](#) · PMCID: [PMC7095789](#)
339. **Characterization and Complete Genome Sequence of a Novel Coronavirus, Coronavirus HKU1, from Patients with Pneumonia**
Patrick CY Woo, Susanna KP Lau, Chung-ming Chu, Kwok-hung Chan, Hoi-wah Tsui, Yi Huang, Beatrice HL Wong, Rosana WS Poon, James J

Cai, Wei-kwang Luk, ... Kwok-yung Yuen
Journal of Virology (2005-01-15) <https://doi.org/bk7m7h>
DOI: [10.1128/jvi.79.2.884-895.2005](https://doi.org/10.1128/jvi.79.2.884-895.2005) · PMID: [15613317](#) · PMCID: [PMC538593](#)

340. **Coronavirus 229E-Related Pneumonia in Immunocompromised Patients**
F Pene, A Merlat, A Vabret, F Rozenberg, A Buzyn, F Dreyfus, A Cariou, F Freymuth, P Lebon
Clinical Infectious Diseases (2003-10-01) <https://doi.org/dcjk64>
DOI: [10.1086/377612](https://doi.org/10.1086/377612) · PMID: [13130404](#) · PMCID: [PMC7107892](#)
341. **Hosts and Sources of Endemic Human Coronaviruses**
Victor M Corman, Doreen Muth, Daniela Niemeyer, Christian Drosten
Advances in Virus Research (2018) <https://doi.org/ggwx4j>
DOI: [10.1016/bs.aivir.2018.01.001](https://doi.org/10.1016/bs.aivir.2018.01.001) · PMID: [29551135](#) · PMCID: [PMC7112090](#)
342. **Novel Canine Coronavirus Isolated from a Hospitalized Patient With Pneumonia in East Malaysia**
Anastasia N Vlasova, Annika Diaz, Debasu Damtie, Leshan Xiu, Teck-Hock Toh, Jeffrey Soon-Yit Lee, Linda J Saif, Gregory C Gray
Clinical Infectious Diseases (2021-05-20) <https://doi.org/gj8zkg>
DOI: [10.1093/cid/ciab456](https://doi.org/10.1093/cid/ciab456) · PMID: [34013321](#) · PMCID: [PMC8194511](#)
343. **A new coronavirus associated with human respiratory disease in China**
Fan Wu, Su Zhao, Bin Yu, Yan-Mei Chen, Wen Wang, Zhi-Gang Song, Yi Hu, Zhao-Wu Tao, Jun-Hua Tian, Yuan-Yuan Pei, ... Yong-Zhen Zhang
Nature (2020-02-03) <https://doi.org/dk2w>
DOI: [10.1038/s41586-020-2008-3](https://doi.org/10.1038/s41586-020-2008-3) · PMID: [32015508](#) · PMCID: [PMC7094943](#)
344. **Risk factors for human disease emergence**
Louise H Taylor, Sophia M Latham, Mark EJ Woolhouse
Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences (2001-07-29) <https://doi.org/cz56wx>
DOI: [10.1098/rstb.2001.0888](https://doi.org/10.1098/rstb.2001.0888) · PMID: [11516376](#) · PMCID: [PMC1088493](#)
345. **Global Patterns of Zoonotic Disease in Mammals**
Barbara A Han, Andrew M Kramer, John M Drake
Trends in Parasitology (2016-07) <https://doi.org/ggt5f2>
DOI: [10.1016/j.pt.2016.04.007](https://doi.org/10.1016/j.pt.2016.04.007) · PMID: [27316904](#) · PMCID: [PMC4921293](#)
346. **Bushmeat Hunting, Deforestation, and Prediction of Zoonotic Disease**
Nathan D Wolfe, Peter Daszak, AMarm Kilpatrick, Donald S Burke
Emerging Infectious Diseases (2005-12) <https://doi.org/fzzcgh>
DOI: [10.3201/eid1112.040789](https://doi.org/10.3201/eid1112.040789) · PMID: [16485465](#) · PMCID: [PMC3367616](#)
347. **Isolation and Characterization of Viruses Related to the SARS Coronavirus from Animals in Southern China**
Y Guan, BJ Zheng, YQ He, XL Liu, ZX Zhuang, CL Cheung, SW Luo, PH Li, LJ Zhang, YJ Guan, ... LLM Poon
Science (2003-10-10) <https://doi.org/dn5nxb>

348. **Zoonotic origins of human coronaviruses**

Zi-Wei Ye, Shuofeng Yuan, Kit-San Yuen, Sin-Yee Fung, Chi-Ping Chan, Dong-Yan Jin

International Journal of Biological Sciences (2020)

<https://doi.org/ggqspq>

DOI: [10.7150/ijbs.45472](https://doi.org/10.7150/ijbs.45472) · PMID: [32226286](#) · PMCID: [PMC7098031](#)

349. **Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats**

Susanna KP Lau, Patrick CY Woo, Kenneth SM Li, Yi Huang, Hoi-Wah Tsoi, Beatrice HL Wong, Samson SY Wong, Suet-Yi Leung, Kwok-Hung Chan, Kwok-Yung Yuen

Proceedings of the National Academy of Sciences (2005-09-16)

<https://doi.org/b36mtm>

DOI: [10.1073/pnas.0506735102](https://doi.org/10.1073/pnas.0506735102) · PMID: [16169905](#) · PMCID: [PMC1236580](#)

350. **Mystery deepens over animal source of coronavirus**

David Cyranoski

Nature (2020-02-26) <https://doi.org/ggpc9x>

DOI: [10.1038/d41586-020-00548-w](https://doi.org/10.1038/d41586-020-00548-w) · PMID: [32127703](#)

351. **III husband and wife add to Wuhan riddle - The Standard**

<https://www.thestandard.com.hk/sections-news-print/215457/iii-husband-and-wife>

352. **Animal sales from Wuhan wet markets immediately prior to the COVID-19 pandemic**

Xiao Xiao, Chris Newman, Christina D Buesching, David W Macdonald, Zhao-Min Zhou

Scientific Reports (2021-06-07) <https://doi.org/gkf5bc>

DOI: [10.1038/s41598-021-91470-2](https://doi.org/10.1038/s41598-021-91470-2) · PMID: [34099828](#) · PMCID: [PMC8184983](#)

353. **Bats as 'special' reservoirs for emerging zoonotic pathogens**

Cara E Brook, Andrew P Dobson

Trends in Microbiology (2015-03) <https://doi.org/gdqcss>

DOI: [10.1016/j.tim.2014.12.004](https://doi.org/10.1016/j.tim.2014.12.004) · PMID: [25572882](#) · PMCID: [PMC7126622](#)

354. **Bats as reservoirs of severe emerging infectious diseases**

Hui-Ju Han, Hong-ling Wen, Chuan-Min Zhou, Fang-Fang Chen, Li-Mei Luo, Jian-wei Liu, Xue-Jie Yu

Virus Research (2015-07) <https://doi.org/f7hnds>

DOI: [10.1016/j.virusres.2015.05.006](https://doi.org/10.1016/j.virusres.2015.05.006) · PMID: [25997928](#) · PMCID: [PMC7132474](#)

355. **Bats: Important Reservoir Hosts of Emerging Viruses**

Charles H Calisher, James E Childs, Hume E Field, Kathryn V Holmes, Tony Schountz

Clinical Microbiology Reviews (2006-07) <https://doi.org/dnjg3n>

DOI: [10.1128/cmrv.00017-06](https://doi.org/10.1128/cmrv.00017-06) · PMID: [16847084](#) · PMCID: [PMC1539106](#)

356. **Probable Pangolin Origin of SARS-CoV-2 Associated with the COVID-19 Outbreak**

Tao Zhang, Qunfu Wu, Zhigang Zhang

Current Biology (2020-04) <https://doi.org/dqsk>

DOI: [10.1016/j.cub.2020.03.022](https://doi.org/j.cub.2020.03.022) · PMID: [32197085](#) · PMCID: [PMC7156161](#)

357. **Identifying SARS-CoV-2-related coronaviruses in Malayan pangolins**

Tommy Tsan-Yuk Lam, Na Jia, Ya-Wei Zhang, Marcus Ho-Hin Shum, Jia-Fu Jiang, Hua-Chen Zhu, Yi-Gang Tong, Yong-Xia Shi, Xue-Bing Ni, Yun-Shi Liao, ... Wu-Chun Cao

Nature (2020-03-26) <https://doi.org/dqjz>

DOI: [10.1038/s41586-020-2169-0](https://doi.org/s41586-020-2169-0) · PMID: [32218527](#)

358. **The proximal origin of SARS-CoV-2**

Kristian G Andersen, Andrew Rambaut, Wian Lipkin, Edward C Holmes, Robert F Garry

Nature Medicine (2020-03-17) <https://doi.org/ggn4dn>

DOI: [10.1038/s41591-020-0820-9](https://doi.org/s41591-020-0820-9) · PMCID: [PMC7095063](#)

359. **Emergence of SARS-CoV-2 through recombination and strong purifying selection**

Xiaojun Li, Elena E Giorgi, Manukumar Honnayakanahalli Marichannegowda, Brian Foley, Chuan Xiao, Xiang-Peng Kong, Yue Chen, S Gnanakaran, Bette Korber, Feng Gao

Science Advances (2020-07) <https://doi.org/gg6r93>

DOI: [10.1126/sciadv.abb9153](https://doi.org/sciadv.abb9153) · PMID: [32937441](#) · PMCID: [PMC7458444](#)

360. **Unraveling the Zoonotic Origin and Transmission of SARS-CoV-2**

Arinjay Banerjee, Andrew C Doxey, Karen Mossman, Aaron T Irving

Trends in Ecology & Evolution (2021-03) <https://doi.org/gh7wpd>

DOI: [10.1016/j.tree.2020.12.002](https://doi.org/j.tree.2020.12.002) · PMID: [33384197](#) · PMCID: [PMC7733689](#)

361. **Investigate the origins of COVID-19**

Jesse D Bloom, Yujia Alina Chan, Ralph S Baric, Pamela J Bjorkman, Sarah Cobey, Benjamin E Deverman, David N Fisman, Ravindra Gupta, Akiko Iwasaki, Marc Lipsitch, ... David A Relman

Science (2021-05-14) <https://doi.org/gcfc>

DOI: [10.1126/science.abj0016](https://doi.org/science.abj0016) · PMID: [33986172](#)

362. **Divisive COVID 'lab leak' debate prompts dire warnings from researchers**

Amy Maxmen

Nature (2021-05-27) <https://doi.org/gj7v3k>

DOI: [10.1038/d41586-021-01383-3](https://doi.org/d41586-021-01383-3) · PMID: [34045757](#)

363. **Reports of zoonotic disease outbreaks associated with animal exhibits and availability of recommendations for preventing zoonotic disease transmission from animals to people in such settings**

Jeff B Bender, Stephanie A Shulman

Journal of the American Veterinary Medical Association (2004-04)

<https://doi.org/c552gn>

364. **Coronavirus immunogens**
Linda J Saif
Veterinary Microbiology (1993-11) <https://doi.org/ckfn8b>
DOI: [10.1016/0378-1135\(93\)90030-b](https://doi.org/10.1016/0378-1135(93)90030-b) · PMID: [8116187](#) · PMCID: [PMC7117163](#)
365. **Host Range, Host-Virus Interactions, and Virus Transmission**
Gustavo Fermin
Viruses (2018) <https://doi.org/gp3hd4>
DOI: [10.1016/b978-0-12-811257-1.00005-x](https://doi.org/10.1016/b978-0-12-811257-1.00005-x) · PMCID: [PMC7173471](#)
366. **Reconstruction and evolutionary history of eutherian chromosomes**
Jaebum Kim, Marta Farré, Loretta Auvil, Boris Capitanu, Denis M Larkin, Jian Ma, Harris A Lewin
Proceedings of the National Academy of Sciences (2017-06-19) <https://doi.org/gbms9r>
DOI: [10.1073/pnas.1702012114](https://doi.org/10.1073/pnas.1702012114) · PMID: [28630326](#) · PMCID: [PMC5502614](#)
367. **Phylogenomic Analyses Elucidate the Evolutionary Relationships of Bats**
Georgia Tsagkogeorga, Joe Parker, Elia Stupka, James A Cotton, Stephen J Rossiter
Current Biology (2013-11) <https://doi.org/f5hx4v>
DOI: [10.1016/j.cub.2013.09.014](https://doi.org/10.1016/j.cub.2013.09.014) · PMID: [24184098](#)
368. **Pegasoferae, an unexpected mammalian clade revealed by tracking ancient retroposon insertions**
Hidenori Nishihara, Masami Hasegawa, Norihiro Okada
Proceedings of the National Academy of Sciences (2006-06-27) <https://doi.org/bvcdff>
DOI: [10.1073/pnas.0603797103](https://doi.org/10.1073/pnas.0603797103) · PMID: [16785431](#) · PMCID: [PMC1479866](#)
369. **Using genomic data to unravel the root of the placental mammal phylogeny**
William J Murphy, Thomas H Pringle, Tess A Crider, Mark S Springer, Webb Miller
Genome Research (2007-02-23) <https://doi.org/cf7pd6>
DOI: [10.1101/gr.5918807](https://doi.org/10.1101/gr.5918807) · PMID: [17322288](#) · PMCID: [PMC1832088](#)
370. <http://www.timetree.org/public/data/pdf/Murphy2009Chap71.pdf>
371. **A Critical Needs Assessment for Research in Companion Animals and Livestock Following the Pandemic of COVID-19 in Humans**
Tracey McNamara, Juergen A Richt, Larry Glickman
Vector-Borne and Zoonotic Diseases (2020-06-01) <https://doi.org/ggv2nt>
DOI: [10.1089/vbz.2020.2650](https://doi.org/10.1089/vbz.2020.2650) · PMID: [32374208](#) · PMCID: [PMC7249469](#)
372. **Analysis of angiotensin-converting enzyme 2 (ACE2) from different species sheds some light on cross-species receptor usage of a**

novel coronavirus 2019-nCoV

Rui Li, Songlin Qiao, Gaiping Zhang

Journal of Infection (2020-04) <https://doi.org/ggmwp8>

DOI: [10.1016/j.jinf.2020.02.013](https://doi.org/j.jinf.2020.02.013) · PMID: [32092392](https://pubmed.ncbi.nlm.nih.gov/32092392/) · PMCID:

[PMC7127620](https://pubmed.ncbi.nlm.nih.gov/PMC7127620/)

373. Broad host range of SARS-CoV-2 predicted by comparative and structural analysis of ACE2 in vertebrates

Joana Damas, Graham M Hughes, Kathleen C Keough, Corrie A Painter, Nicole S Persky, Marco Corbo, Michael Hiller, Klaus-Peter Koepfli, Andreas R Pfenning, Huabin Zhao, ... Harris A Lewin

Proceedings of the National Academy of Sciences (2020-08-21)

<https://doi.org/ghbsq7>

DOI: [10.1073/pnas.2010146117](https://doi.org/10.1073/pnas.2010146117) · PMID: [32826334](https://pubmed.ncbi.nlm.nih.gov/32826334/) · PMCID:

[PMC7486773](https://pubmed.ncbi.nlm.nih.gov/PMC7486773/)

374. Update on possible animal sources for COVID-19 in humans

Tanja Opriessnig, Yao-Wei Huang

Xenotransplantation (2020-05) <https://doi.org/gk7k8j>

DOI: [10.1111/xen.12621](https://doi.org/10.1111/xen.12621) · PMID: [32557711](https://pubmed.ncbi.nlm.nih.gov/32557711/) · PMCID: [PMC7323145](https://pubmed.ncbi.nlm.nih.gov/PMC7323145/)

375. Anthropogenic Infection of Cats during the 2020 COVID-19 Pandemic

Margaret J Hosie, Regina Hofmann-Lehmann, Katrin Hartmann, Herman Egberink, Uwe Truyen, Diane D Addie, Sándor Belák, Corine Boucraut-Baralon, Tadeusz Frymus, Albert Lloret, ... Karin Möstl

Viruses (2021-01-26) <https://doi.org/gk65t3>

DOI: [10.3390/v13020185](https://doi.org/10.3390/v13020185) · PMID: [33530620](https://pubmed.ncbi.nlm.nih.gov/33530620/) · PMCID: [PMC7911697](https://pubmed.ncbi.nlm.nih.gov/PMC7911697/)

376. Infection of dogs with SARS-CoV-2

Thomas HC Sit, Christopher J Brackman, Sin Ming Ip, Karina WS Tam, Pierra YT Law, Esther MW To, Veronica YT Yu, Leslie D Sims, Dominic NC Tsang, Daniel KW Chu, ... Malik Peiris

Nature (2020-05-14) <https://doi.org/dvt4>

DOI: [10.1038/s41586-020-2334-5](https://doi.org/10.1038/s41586-020-2334-5) · PMID: [32408337](https://pubmed.ncbi.nlm.nih.gov/32408337/) · PMCID:

[PMC7606701](https://pubmed.ncbi.nlm.nih.gov/PMC7606701/)

377. Possible Human-to-Dog Transmission of SARS-CoV-2, Italy, 2020

Nicola Decaro, Gabriele Vaccari, Alessio Lorusso, Eleonora Lorusso, Luca De Sabato, Edward I Patterson, Ilaria Di Bartolo, Grant L Hughes, Liana Teodori, Costantina Desario, ... Gabriella Elia

Emerging Infectious Diseases (2021) <https://doi.org/gp3hff>

DOI: [10.3201/eid2707.204959](https://doi.org/10.3201/eid2707.204959) · PMID: [33979566](https://pubmed.ncbi.nlm.nih.gov/33979566/) · PMCID: [PMC8237870](https://pubmed.ncbi.nlm.nih.gov/PMC8237870/)

378. Evidence of exposure to SARS-CoV-2 in cats and dogs from households in Italy

EI Patterson, G Elia, A Grassi, A Giordano, C Desario, M Medardo, SL Smith, ER Anderson, T Prince, GT Patterson, ... N Decaro

Nature Communications (2020-12) <https://doi.org/gp3hd9>

DOI: [10.1038/s41467-020-20097-0](https://doi.org/10.1038/s41467-020-20097-0) · PMID: [33277505](https://pubmed.ncbi.nlm.nih.gov/33277505/) · PMCID:

[PMC7718263](https://pubmed.ncbi.nlm.nih.gov/PMC7718263/)

379. SARS-CoV-2 in domestic cats (*Felis catus*) in the northwest of Iran: Evidence for SARS-CoV-2 circulating between human and cats

Mehdi Mohebali, Gholamreza Hassanpour, Mohammad Zainali, Mohammad Mehdi Gouya, Simin Khayatzadeh, Mehdi Parsaei, Nazila Sarafraz, Mehdi Hassanzadeh, Amrollah Azarm, Mostafa Salehi-Vaziri, ... Zabihollah Zarei
Virus Research (2022-03) <https://doi.org/gn3dz5>
DOI: [10.1016/j.virusres.2022.198673](https://doi.org/10.1016/j.virusres.2022.198673) · PMID: [34998863](#)

380. **Experimental infection of domestic dogs and cats with SARS-CoV-2: Pathogenesis, transmission, and response to reexposure in cats**
Angela M Bosco-Lauth, Aireen E Hartwig, Stephanie M Porter, Paul W Gordy, Mary Nehring, Alex D Byas, Sue VandeWoude, Izabela K Ragan, Rachel M Maison, Richard A Bowen
Proceedings of the National Academy of Sciences (2020-09-29)
<https://doi.org/ghj9d7>
DOI: [10.1073/pnas.2013102117](https://doi.org/10.1073/pnas.2013102117) · PMID: [32994343](#) · PMCID: [PMC7585007](#)
381. **Exclusive: Buddy, first dog to test positive for COVID-19 in the U.S., has died**
Animals
(2020-07-29)
<https://www.nationalgeographic.com/animals/article/first-dog-to-test-positive-for-covid-in-us-dies>
382. **Connecticut puppy that died unexpectedly tested positive for coronavirus, UConn researchers say**
Hartford Courant
<https://www.courant.com/coronavirus/hc-news-coronavirus-uconn-dog-infection-20210413-yrgb6icd6bcirk6ylwnzkxi6di-story.html>
383. **North Carolina dog that died after 'acute' illness tests positive for coronavirus**
NBC News
<https://www.nbcnews.com/news/us-news/north-carolina-dog-died-after-acute-illness-tests-positive-coronavirus-n1236477>
384. **Kitten Dies After Catching Covid As Study Uncovers More Evidence Of Human-To-Cat Transmission**
Robert Hart
Forbes <https://www.forbes.com/sites/roberthart/2021/04/23/kitten-dies-after-catching-covid-as-study-uncovers-more-evidence-of-human-to-cat-transmission/>
385. **Detection of SARS-CoV-2 in respiratory samples from cats in the UK associated with human-to-cat transmission**
Margaret J Hosie, Ilaria Epifano, Vanessa Herder, Richard J Orton, Andrew Stevenson, Natasha Johnson, Emma MacDonald, Dawn Dunbar, Michael McDonald, Fiona Howie, ...
Veterinary Record (2021-04) <https://doi.org/gk67vp>
DOI: [10.1002/vetr.247](https://doi.org/10.1002/vetr.247) · PMID: [33890314](#) · PMCID: [PMC8251078](#)
386. **Investigation into cat's death, cat had COVID**
Alan Collins
<https://www.wbrc.com>
<https://www.wbrc.com/2020/10/09/investigation-into-cats-death-cat->

[had-covid/](#)

387. **Bats, pangolins, minks and other animals - villains or victims of SARS-CoV-2?**

Beatriz do Vale, Ana Patrícia Lopes, Maria da Conceição Fontes, Mário Silvestre, Luís Cardoso, Ana Cláudia Coelho

Veterinary Research Communications (2021-01-19)

<https://doi.org/gk6ttw>

DOI: [10.1007/s11259-021-09787-2](https://doi.org/s11259-021-09787-2) · PMID: [33464439](https://pubmed.ncbi.nlm.nih.gov/33464439/) · PMCID: [PMC7813668](https://pubmed.ncbi.nlm.nih.gov/PMC7813668/)

388. **Susceptibility of livestock to SARS-CoV-2 infection**

Angela M Bosco-Lauth, Audrey Walker, Lauren Guilbert, Stephanie Porter, Airn Hartwig, Emma McVicker, Helle Bielefeldt-Ohmann, Richard A Bowen

Emerging Microbes & Infections (2021-01-01)

<https://doi.org/gp3hfb>

DOI: [10.1080/22221751.2021.2003724](https://doi.org/22221751.2021.2003724) · PMID: [34749583](https://pubmed.ncbi.nlm.nih.gov/34749583/) · PMCID: [PMC8635583](https://pubmed.ncbi.nlm.nih.gov/PMC8635583/)

389. **Susceptibility of sheep to experimental co-infection with the ancestral lineage of SARS-CoV-2 and its alpha variant**

Natasha N Gaudreault, Konner Cool, Jessie D Trujillo, Igor Morozov, David A Meekins, Chester McDowell, Dashzeveg Bold, Mariano Carossino, Velmurugan Balaraman, Dana Mitzel, ... Juergen A Richt

Emerging Microbes & Infections (2022-02-24)

<https://doi.org/gp3hfc>

DOI: [10.1080/22221751.2022.2037397](https://doi.org/22221751.2022.2037397) · PMID: [35105272](https://pubmed.ncbi.nlm.nih.gov/35105272/) · PMCID: [PMC8881078](https://pubmed.ncbi.nlm.nih.gov/PMC8881078/)

390. **Absence of SARS-CoV-2 Antibodies in Natural Environment Exposure in Sheep in Close Contact with Humans**

Sergio Villanueva-Saz, Jacobo Giner, Antonio Fernández, Delia Lacasta, Aurora Ortín, Juan José Ramos, Luis Miguel Ferrer, Marta Ruiz de Arcante, Ana Pilar Tobajas, María Dolores Pérez, ... Héctor Ruíz

Animals (2021-07-02) <https://doi.org/gp3hfg>

DOI: [10.3390/ani11071984](https://doi.org/10.3390/ani11071984) · PMID: [34359111](https://pubmed.ncbi.nlm.nih.gov/34359111/) · PMCID: [PMC8300300](https://pubmed.ncbi.nlm.nih.gov/PMC8300300/)

391. **Livestock plants and COVID-19 transmission**

Charles A Taylor, Christopher Boulos, Douglas Almond

Proceedings of the National Academy of Sciences (2020-11-19)

<https://doi.org/ghzzhs>

DOI: [10.1073/pnas.2010115117](https://doi.org/10.1073/pnas.2010115117) · PMID: [33214147](https://pubmed.ncbi.nlm.nih.gov/33214147/) · PMCID: [PMC7749337](https://pubmed.ncbi.nlm.nih.gov/PMC7749337/)

392. **COVID-19 Effects on Livestock Production: A One Welfare Issue**

Jeremy N Marchant-Forde, Laura A Boyle

Frontiers in Veterinary Science (2020-09-30) <https://doi.org/gjtrhd>

DOI: [10.3389/fvets.2020.585787](https://doi.org/10.3389/fvets.2020.585787) · PMID: [33195613](https://pubmed.ncbi.nlm.nih.gov/33195613/) · PMCID: [PMC7554581](https://pubmed.ncbi.nlm.nih.gov/PMC7554581/)

393. **SARS-CoV-2 infection in farmed minks, the Netherlands, April and May 2020**

Nadia Oreshkova, Robert Jan Molenaar, Sandra Vreman, Frank Harders, Bas B Oude Munnink, Renate W Hakze-van der Honing, Nora

Gerhards, Paulien Tolsma, Ruth Bouwstra, Reina S Sikkema, ... Arjan Stegeman
Eurosurveillance (2020-06-11) <https://doi.org/gkz252>
DOI: [10.2807/1560-7917.es.2020.25.23.2001005](https://doi.org/10.2807/1560-7917.es.2020.25.23.2001005) · PMID: [32553059](#) ·
PMCID: [PMC7403642](#)

394. **SARS-CoV-2 Transmission between Mink (<i>Neovison vison</i>) and Humans, Denmark**
Anne Sofie Hammer, Michelle Lauge Quaade, Thomas Bruun Rasmussen, Jannik Fonager, Morten Rasmussen, Karin Mundbjerg, Louise Lohse, Bertel Strandbygaard, Charlotte Sværke Jørgensen, Alonso Alfaro-Núñez, ... Anette Bøtner
Emerging Infectious Diseases (2021-02) <https://doi.org/gk7pkw>
DOI: [10.3201/eid2702.203794](https://doi.org/10.3201/eid2702.203794) · PMID: [33207152](#) · PMCID: [PMC7853580](#)
395. **An outbreak of SARS-CoV-2 with high mortality in mink (Neovison vison) on multiple Utah farms**
Chrissy D Eckstrand, Thomas J Baldwin, Kerry A Rood, Michael J Clayton, Jason K Lott, Rebecca M Wolking, Daniel S Bradway, Timothy Baszler
PLOS Pathogens (2021-11-12) <https://doi.org/gp3hfd>
DOI: [10.1371/journal.ppat.1009952](https://doi.org/10.1371/journal.ppat.1009952) · PMID: [34767598](#) · PMCID: [PMC8589170](#)
396. **Preliminary report of an outbreak of SARS-CoV-2 in mink and mink farmers associated with community spread, Denmark, June to November 2020**
Helle Daugaard Larsen, Jannik Fonager, Frederikke Kristensen Lomholt, Tine Dalby, Guido Benedetti, Brian Kristensen, Tinna Ravnholt Urth, Morten Rasmussen, Ria Lassaunière, Thomas Bruun Rasmussen, ... Kåre Mølbak
Eurosurveillance (2021-02-04) <https://doi.org/gk67zt>
DOI: [10.2807/1560-7917.es.2021.26.5.210009](https://doi.org/10.2807/1560-7917.es.2021.26.5.210009) · PMID: [33541485](#) ·
PMCID: [PMC7863232](#)
397. **SARS-CoV-2 mutations acquired in mink reduce antibody-mediated neutralization**
Markus Hoffmann, Lu Zhang, Nadine Krüger, Luise Graichen, Hannah Kleine-Weber, Heike Hofmann-Winkler, Amy Kempf, Stefan Nessler, Joachim Riggert, Martin Sebastian Winkler, ... Stefan Pöhlmann
Cell Reports (2021-04) <https://doi.org/gp3hd5>
DOI: [10.1016/j.celrep.2021.109017](https://doi.org/10.1016/j.celrep.2021.109017) · PMID: [33857422](#) · PMCID: [PMC8018833](#)
398. **Monitoring Natural SARS-CoV-2 Infection in Lions (*Panthera leo*) at the Barcelona Zoo: Viral Dynamics and Host Responses**
Hugo Fernández-Bellón, Jordi Rodon, Leira Fernández-Bastit, Vanessa Almagro, Pilar Padilla-Solé, Cristina Lorca-Oró, Rosa Valle, Núria Roca, Santina Grazioli, Tiziana Trogu, ... Júlia Vergara-Alert
Viruses (2021-08-25) <https://doi.org/gp3hfh>
DOI: [10.3390/v13091683](https://doi.org/10.3390/v13091683) · PMID: [34578266](#) · PMCID: [PMC8472846](#)
399. **Mass culling of minks to protect the COVID-19 vaccines: is it rational?**
R Frutos, CA Devaux

400. **Coronavirus rips through Dutch mink farms, triggering culls**
Martin Enserink
Science (2020-06-12) <https://doi.org/gg6n2t>
DOI: [10.1126/science.368.6496.1169](https://doi.org/10.1126/science.368.6496.1169) · PMID: [32527808](#)
401. **Wild American mink (<i>Neovison vison</i>) may pose a COVID-19 threat**
Lauren A Harrington, María Díez-León, Asunción Gómez, Andrew Harrington, David W Macdonald, Tiit Maran, Madis Põdra, Sugoto Roy
Frontiers in Ecology and the Environment (2021-06)
<https://doi.org/gkb35b>
DOI: [10.1002/fee.2344](https://doi.org/10.1002/fee.2344) · PMID: [34149325](#) · PMCID: [PMC8207089](#)
402. **First Description of SARS-CoV-2 Infection in Two Feral American Mink (Neovison vison) Caught in the Wild**
Jordi Aguiló-Gisbert, Miguel Padilla-Blanco, Victor Lizana, Elisa Maiques, Marta Muñoz-Baquero, Eva Chillida-Martínez, Jesús Cardells, Consuelo Rubio-Guerri
Animals (2021-05-16) <https://doi.org/gk63mr>
DOI: [10.3390/ani11051422](https://doi.org/10.3390/ani11051422) · PMID: [34065657](#) · PMCID: [PMC8156136](#)
403. **USDA APHIS | Confirmation of COVID-19 in Gorillas at a California Zoo** https://www.aphis.usda.gov/aphis/newsroom/stakeholder-info/sa_by_date/sa-2021/sa-01/ca-gorillas-sars-cov-2
404. **Dallas Zoo says 5 gorillas have tested positive for COVID-19**
wfaa.com
(2022-02-10) <https://www.wfaa.com/article/news/local/dallas-zoo-5-gorillas-tested-positive-covid-19/287-901a235f-b120-49ff-9fbe-1a1a6dfbadb5>
405. **Nearly all gorillas at Atlanta's zoo have contracted COVID-19**
CTVNews
(2021-09-15) <https://www.ctvnews.ca/health/coronavirus/nearly-all-gorillas-at-atlanta-s-zoo-have-contracted-covid-19-1.5586112>
406. **Kansas City Zoo gorillas recovering from COVID-19**
KMBC 9News Staff
KMBC (2021-10-05) <https://www.kmbc.com/article/kc-zoo-gorillas-recovering-from-covid/37873956>
407. **From People to <i>Panthera</i> : Natural SARS-CoV-2 Infection in Tigers and Lions at the Bronx Zoo**
Denise McAloose, Melissa Laverack, Leyi Wang, Mary Lea Killian, Leonardo C Caserta, Fangfeng Yuan, Patrick K Mitchell, Krista Queen, Matthew R Mauldin, Brittany D Cronk, ... Diego G Diel
mBio (2020-10-27) <https://doi.org/ghj9fh>
DOI: [10.1128/mbio.02220-20](https://doi.org/10.1128/mbio.02220-20) · PMID: [33051368](#) · PMCID: [PMC7554670](#)
408. **A snow leopard at Miller Park Zoo dies from COVID-induced pneumonia**

WGLT

(2022-01-06) <https://www.wglt.org/local-news/2022-01-06/a-snow-leopard-at-miller-park-zoo-is-mclean-countys-latest-death-from-covid-19>

409. **A zoo's three 'beloved' snow leopards die of covid-19**

Washington Post

<https://www.washingtonpost.com/science/2021/11/14/snow-leopard-death-covid/>

410. **When People Take Pandemic Precautions, Gorillas Breathe Easier**

Emily Anthes

The New York Times (2022-02-21)

<https://www.nytimes.com/2022/02/21/health/gorillas-respiratory-illness-colds.html>

411. **Mountain gorilla research: the risk of disease transmission relative to the benefit from the perspective of ecosystem health**

Michael R Cranfield

American Journal of Primatology (2008-08) <https://doi.org/dc7dst>

DOI: [10.1002/ajp.20564](https://doi.org/10.1002/ajp.20564) · PMID: [18506694](https://pubmed.ncbi.nlm.nih.gov/18506694/)

412. **A group of gorillas is being treated for covid. The great apes will soon get their shots, too, zoo says.**

Washington Post

<https://www.washingtonpost.com/nation/2021/09/12/zoo-atlanta-gorillas-coronavirus-vaccine/>

413. **Experimental veterinary SARS-CoV-2 vaccine cross neutralization of the Delta (B.1.617.2) variant virus in cats**

Ashley Hoyte, Mark Webster, Keith Ameiss, Douglas A Conlee, Nicole Hainer, Kendra Hutchinson, Yulia Burakova, Paul J Dominowski, Eric T Baima, Vickie L King, ... Mahesh Kumar

Veterinary Microbiology (2022-05) <https://doi.org/gp3hd8>

DOI: [10.1016/j.vetmic.2022.109395](https://doi.org/10.1016/j.vetmic.2022.109395) · PMID: [35339817](https://pubmed.ncbi.nlm.nih.gov/35339817/) · PMCID: [PMC8915440](https://pubmed.ncbi.nlm.nih.gov/PMC8915440/)

414. **COVID-19 Animal Vaccines | Zoetis** <https://www.zoetis.com/news-and-media/feature-stories/posts/zoetis-emerging-infectious-disease-capabilities-support-covid-19-solutions-for-great-apes-and-minks.aspx>

415. **SARS-CoV-2 vaccine for domestic and captive animals: An effort to counter COVID-19 pandemic at the human-animal interface**

Khan Sharun, Ruchi Tiwari, AbdulRahman A Saied, Kuldeep Dhama

Vaccine (2021-12) <https://doi.org/gp3hd7>

DOI: [10.1016/j.vaccine.2021.10.053](https://doi.org/10.1016/j.vaccine.2021.10.053) · PMID: [34782159](https://pubmed.ncbi.nlm.nih.gov/34782159/) · PMCID: [PMC8570933](https://pubmed.ncbi.nlm.nih.gov/PMC8570933/)

416. **Do we need to have a Covid vaccine for domestic animals?**

Emily Anthes

The Irish Times <https://www.irishtimes.com/life-and-style/do-we-need-to-have-a-covid-vaccine-for-domestic-animals-1.4736360>

417. **A Veterinary Vaccine for SARS-CoV-2: The First COVID-19 Vaccine for Animals**

Vivek P Chavda, Jack Feehan, Vasso Apostolopoulos
Vaccines (2021-06-10) <https://doi.org/gkhtrd>
DOI: [10.3390/vaccines9060631](https://doi.org/10.3390/vaccines9060631) · PMID: [34200587](#) · PMCID:
[PMC8228738](#)

418. **Cats and Dogs Top List of COVID-19 Infected Animals in US**
C, ice Nguyen, Robert Campos, Michael Horn • •
NBC Bay Area <https://www.nbcbayarea.com/investigations/cats-and-dogs-top-list-of-covid-19-infected-animals-in-u-s/2625085/>
419. **Functional and genetic analysis of viral receptor ACE2 orthologs reveals a broad potential host range of SARS-CoV-2**
Yinghui Liu, Gaowei Hu, Yuyan Wang, Wenlin Ren, Xiaomin Zhao, Fansen Ji, Yunkai Zhu, Fei Feng, Mingli Gong, Xiaohui Ju, ... Qiang Ding
Proceedings of the National Academy of Sciences (2021-03-03)
<https://doi.org/gk63kd>
DOI: [10.1073/pnas.2025373118](https://doi.org/10.1073/pnas.2025373118) · PMID: [33658332](#) · PMCID:
[PMC8000431](#)
420. **The SARS-CoV-2 Spike protein has a broad tropism for mammalian ACE2 proteins**
Carina Conceicao, Nazia Thakur, Stacey Human, James T Kelly, Leanne Logan, Dagmara Bialy, Sushant Bhat, Phoebe Stevenson-Leggett, Adrian K Zagrajek, Philippa Hollinghurst, ... Dalan Bailey
PLOS Biology (2020-12-21) <https://doi.org/gk6z7p>
DOI: [10.1371/journal.pbio.3001016](https://doi.org/10.1371/journal.pbio.3001016) · PMID: [33347434](#) · PMCID:
[PMC7751883](#)
421. **Spurious Genetic Associations**
Patrick F Sullivan
Biological Psychiatry (2007-05) <https://doi.org/c89p4c>
DOI: [10.1016/j.biopsych.2006.11.010](https://doi.org/10.1016/j.biopsych.2006.11.010) · PMID: [17346679](#)
422. **Interferon-Induced Transmembrane Protein 3 Genetic Variant rs12252-C Associated With Disease Severity in Coronavirus Disease 2019**
Yonghong Zhang, Ling Qin, Yan Zhao, Ping Zhang, Bin Xu, Kang Li, Lianchun Liang, Chi Zhang, Yanchao Dai, Yingmei Feng, ... Ronghua Jin
The Journal of Infectious Diseases (2020-07-01) <https://doi.org/ggv3tj>
DOI: [10.1093/infdis/jiaa224](https://doi.org/10.1093/infdis/jiaa224) · PMID: [32348495](#) · PMCID: [PMC7197559](#)
423. **Interferon-induced transmembrane protein-3 genetic variant rs12252-C is associated with severe influenza in Chinese individuals**
Yong-Hong Zhang, Yan Zhao, Ning Li, Yan-Chun Peng, Eleni Giannoulatou, Rong-Hua Jin, Hui-Ping Yan, Hao Wu, Jin-Hua Liu, Ning Liu, ... Tao Dong
Nature Communications (2013-01-29) <https://doi.org/gbcr8v>
DOI: [10.1038/ncomms2433](https://doi.org/10.1038/ncomms2433) · PMID: [23361009](#) · PMCID: [PMC3562464](#)
424. **Ethnic variation in risk genotypes based on single nucleotide polymorphisms (SNPs) of the interferon-inducible transmembrane 3 (IFITM3) gene, a susceptibility factor for pandemic 2009 H1N1 influenza A virus**
Yong-Chan Kim, Byung-Hoon Jeong

425. **Strong Correlation between the Case Fatality Rate of COVID-19 and the rs6598045 Single Nucleotide Polymorphism (SNP) of the Interferon-Induced Transmembrane Protein 3 (IFITM3) Gene at the Population-Level**
Yong-Chan Kim, Byung-Hoon Jeong
Genes (2020-12-30) <https://doi.org/gpvr6f>
DOI: [10.3390/genes12010042](https://doi.org/10.3390/genes12010042) · PMID: [33396837](https://pubmed.ncbi.nlm.nih.gov/33396837/) · PMCID: [PMC7824003](https://pubmed.ncbi.nlm.nih.gov/PMC7824003/)
426. **The frequency of combined IFITM3 haplotype involving the reference alleles of both rs12252 and rs34481144 is in line with COVID-19 standardized mortality ratio of ethnic groups in England**
Dimitris Nikoloudis, Dimitrios Kountouras, Asimina Hiona
PeerJ (2020-11-12) <https://doi.org/gk7gn9>
DOI: [10.7717/peerj.10402](https://doi.org/10.7717/peerj.10402) · PMID: [33240681](https://pubmed.ncbi.nlm.nih.gov/33240681/) · PMCID: [PMC7666821](https://pubmed.ncbi.nlm.nih.gov/PMC7666821/)
427. **Opposing activities of IFITM proteins in SARS-CoV-2 infection**
Guoli Shi, Adam D Kenney, Elena Kudryashova, Ashley Zani, Lizhi Zhang, Kin Kui Lai, Luanne Hall-Stoodley, Richard T Robinson, Dmitri S Kudryashov, Alex A Compton, Jacob S Yount
The EMBO Journal (2020-12-21) <https://doi.org/gpvr6d>
DOI: [10.15252/embj.2020106501](https://doi.org/10.15252/embj.2020106501) · PMID: [33270927](https://pubmed.ncbi.nlm.nih.gov/33270927/) · PMCID: [PMC7744865](https://pubmed.ncbi.nlm.nih.gov/PMC7744865/)
428. **Bat SARS-Like WIV1 coronavirus uses the ACE2 of multiple animal species as receptor and evades IFITM3 restriction <i>via</i> TMPRSS2 activation of membrane fusion**
Mei Zheng, Xuesen Zhao, Shuangli Zheng, Danying Chen, Pengcheng Du, Xinglin Li, Dong Jiang, Ju-Tao Guo, Hui Zeng, Hanxin Lin
Emerging Microbes & Infections (2020-01-01)
<https://doi.org/gg7yjf>
DOI: [10.1080/22221751.2020.1787797](https://doi.org/10.1080/22221751.2020.1787797) · PMID: [32602823](https://pubmed.ncbi.nlm.nih.gov/32602823/) · PMCID: [PMC7473123](https://pubmed.ncbi.nlm.nih.gov/PMC7473123/)
429. **Candidate-gene approaches for studying complex genetic traits: practical considerations**
Holly K Tabor, Neil J Risch, Richard M Myers
Nature Reviews Genetics (2002-05) <https://doi.org/dbd7md>
DOI: [10.1038/nrg796](https://doi.org/10.1038/nrg796) · PMID: [11988764](https://pubmed.ncbi.nlm.nih.gov/11988764/)
430. **Mapping the human genetic architecture of COVID-19**
, Mari EK Niemi, Juha Karjalainen, Rachel G Liao, Benjamin M Neale, Mark Daly, Andrea Ganna, Gita A Pathak, Shea J Andrews, Masahiro Kanai, ...
Nature (2021-07-08) <https://doi.org/gk5gcv>
DOI: [10.1038/s41586-021-03767-x](https://doi.org/10.1038/s41586-021-03767-x) · PMID: [34237774](https://pubmed.ncbi.nlm.nih.gov/34237774/) · PMCID: [PMC8674144](https://pubmed.ncbi.nlm.nih.gov/PMC8674144/)
431. **HLA DNA typing: past, present, and future**
H Erlich
Tissue Antigens (2012-06-01) <https://doi.org/f3z8j3>
DOI: [10.1111/j.1399-0039.2012.01881.x](https://doi.org/10.1111/j.1399-0039.2012.01881.x) · PMID: [22651253](https://pubmed.ncbi.nlm.nih.gov/22651253/)

432. **HLA Diversity in the 1000 Genomes Dataset**
Pierre-Antoine Gourraud, Pouya Khankhanian, Nezih Cereb, Soo Young Yang, Michael Feolo, Martin Maiers, John D. Rioux, Stephen Hauser, Jorge Oksenberg
PLoS ONE (2014-07-02) <https://doi.org/f6gjzn>
DOI: [10.1371/journal.pone.0097282](https://doi.org/10.1371/journal.pone.0097282) · PMID: [24988075](https://pubmed.ncbi.nlm.nih.gov/24988075/) · PMCID: [PMC4079705](https://pubmed.ncbi.nlm.nih.gov/PMC4079705/)
433. **Mortality in COVID-19 disease patients: Correlating the association of major histocompatibility complex (MHC) with severe acute respiratory syndrome 2 (SARS-CoV-2) variants**
Eric de Sousa, Dário Ligeiro, Joana R Lérias, Chao Zhang, Chiara Agrati, Mohamed Osman, Sherif A El-Kafrawy, Esam I Azhar, Giuseppe Ippolito, Fu-Sheng Wang, ... Markus Maeurer
International Journal of Infectious Diseases (2020-09)
<https://doi.org/gg5kdg>
DOI: [10.1016/j.ijid.2020.07.016](https://doi.org/10.1016/j.ijid.2020.07.016) · PMID: [32693089](https://pubmed.ncbi.nlm.nih.gov/32693089/) · PMCID: [PMC7368421](https://pubmed.ncbi.nlm.nih.gov/PMC7368421/)
434. **Association between the HLA genotype and the severity of COVID-19 infection among South Asians**
Fatmah MA Naemi, Shurooq Al-adwani, Heba Al-khatabi, Ashwaq Al-nazawi
Journal of Medical Virology (2021-04-23) <https://doi.org/gpwf57>
DOI: [10.1002/jmv.27003](https://doi.org/10.1002/jmv.27003) · PMID: [33830530](https://pubmed.ncbi.nlm.nih.gov/33830530/) · PMCID: [PMC8251353](https://pubmed.ncbi.nlm.nih.gov/PMC8251353/)
435. **The influence of HLA genotype on the severity of COVID-19 infection**
David J Langton, Stephen C Bourke, Benedicte A Lie, Gabrielle Reiff, Shonali Natu, Rebecca Darlay, John Burn, Carlos Echevarria
HLA (2021-05-04) <https://doi.org/gmb2n4>
DOI: [10.1111/tan.14284](https://doi.org/10.1111/tan.14284) · PMID: [33896121](https://pubmed.ncbi.nlm.nih.gov/33896121/) · PMCID: [PMC8251294](https://pubmed.ncbi.nlm.nih.gov/PMC8251294/)
436. **<scp>HLA</scp> genetic polymorphism in patients with Coronavirus Disease 2019 in Midwestern United States**
Emily Schindler, Marian Dribus, Brian F Duffy, Karl Hock, Christopher W Farnsworth, Loren Gragert, Chang Liu
HLA (2021-08-10) <https://doi.org/gpwf6f>
DOI: [10.1111/tan.14387](https://doi.org/10.1111/tan.14387) · PMID: [34338446](https://pubmed.ncbi.nlm.nih.gov/34338446/) · PMCID: [PMC8429120](https://pubmed.ncbi.nlm.nih.gov/PMC8429120/)
437. **MHC Haplotyping of SARS-CoV-2 Patients: HLA Subtypes Are Not Associated with the Presence and Severity of COVID-19 in the Israeli Population**
Shay Ben Shachar, Noam Barda, Sigal Manor, Sapir Israeli, Noa Dagan, Shai Carmi, Ran Balicer, Bracha Zisser, Yoram Louzoun
Journal of Clinical Immunology (2021-05-29) <https://doi.org/gj896d>
DOI: [10.1007/s10875-021-01071-x](https://doi.org/10.1007/s10875-021-01071-x) · PMID: [34050837](https://pubmed.ncbi.nlm.nih.gov/34050837/) · PMCID: [PMC8164405](https://pubmed.ncbi.nlm.nih.gov/PMC8164405/)
438. **Investigating Whether Blood Type Is Linked to COVID-19 Risk**
Rita Rubin
JAMA (2020-10-06) <https://doi.org/gpwf56>
DOI: [10.1001/jama.2020.16516](https://doi.org/10.1001/jama.2020.16516) · PMID: [32936219](https://pubmed.ncbi.nlm.nih.gov/32936219/)

439. **Relationship between the ABO Blood Group and the COVID-19 Susceptibility**

Jiao Zhao, Yan Yang, Hanping Huang, Dong Li, Dongfeng Gu, Xiangfeng Lu, Zheng Zhang, Lei Liu, Ting Liu, Yukun Liu, ... Peng George Wang
Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggpn3d>
DOI: [10.1101/2020.03.11.20031096](https://doi.org/10.1101/2020.03.11.20031096)

440. **Impact of ABO and Rhesus blood groups on COVID-19 susceptibility and severity: A case-control study**

Anthony Kerbage, Sara F Haddad, Lewis Nasr, Albert Riachy, Elio Mekhael, Nabil Nassim, Karim Hoyek, Ghassan Sleilaty, Fadi Nasr, Moussa Riachy
Journal of Medical Virology (2021-11-16) <https://doi.org/gpwf58>
DOI: [10.1002/jmv.27444](https://doi.org/10.1002/jmv.27444) · PMID: [34755349](https://pubmed.ncbi.nlm.nih.gov/34755349/) · PMCID: [PMC8662239](https://pubmed.ncbi.nlm.nih.gov/PMC8662239/)

441. **Association Between ABO Blood Group System and COVID-19 Susceptibility in Wuhan**

Qian Fan, Wei Zhang, Bo Li, De-Jia Li, Jian Zhang, Fang Zhao
Frontiers in Cellular and Infection Microbiology (2020-07-21)
<https://doi.org/gg7t8k>
DOI: [10.3389/fcimb.2020.00404](https://doi.org/10.3389/fcimb.2020.00404) · PMID: [32793517](https://pubmed.ncbi.nlm.nih.gov/32793517/) · PMCID: [PMC7385064](https://pubmed.ncbi.nlm.nih.gov/PMC7385064/)

442. **The association of ABO blood group with indices of disease severity and multiorgan dysfunction in COVID-19**

Ryan L Hoiland, Nicholas A Ferguson, Anish R Mitra, Donald EG Griesdale, Dana V Devine, Sophie Stukas, Jennifer Cooper, Sonny Thiara, Denise Foster, Luke YC Chen, ... Mypinder S Sekhon
Blood Advances (2020-10-14) <https://doi.org/ghk7dz>
DOI: [10.1182/bloodadvances.2020002623](https://doi.org/10.1182/bloodadvances.2020002623) · PMID: [33057633](https://pubmed.ncbi.nlm.nih.gov/33057633/) · PMCID: [PMC7594392](https://pubmed.ncbi.nlm.nih.gov/PMC7594392/)

443. **Association of Sociodemographic Factors and Blood Group Type With Risk of COVID-19 in a US Population**

Jeffrey L Anderson, Heidi T May, Stacey Knight, Tami L Bair, Joseph B Muhlestein, Kirk U Knowlton, Benjamin D Horne
JAMA Network Open (2021-04-05) <https://doi.org/gk6xzc>
DOI: [10.1001/jamanetworkopen.2021.7429](https://doi.org/10.1001/jamanetworkopen.2021.7429) · PMID: [33818622](https://pubmed.ncbi.nlm.nih.gov/33818622/) · PMCID: [PMC8022215](https://pubmed.ncbi.nlm.nih.gov/PMC8022215/)

444. **Association between ABO blood types and coronavirus disease 2019 (COVID-19), genetic associations, and underlying molecular mechanisms: a literature review of 23 studies**

Yujia Zhang, Rachael Garner, Sana Salehi, Marianna La Rocca, Dominique Duncan
Annals of Hematology (2021-03-08) <https://doi.org/gpwf59>
DOI: [10.1007/s00277-021-04489-w](https://doi.org/10.1007/s00277-021-04489-w) · PMID: [33686492](https://pubmed.ncbi.nlm.nih.gov/33686492/) · PMCID: [PMC7939543](https://pubmed.ncbi.nlm.nih.gov/PMC7939543/)

445. **Association between ABO blood groups and COVID-19 infection, severity and demise: A systematic review and meta-analysis**

Bing-Bing Wu, Dong-Zhou Gu, Jia-Ning Yu, Jie Yang, Wang-Qin Shen
Infection, Genetics and Evolution (2020-10) <https://doi.org/gg7n6p>
DOI: [10.1016/j.meegid.2020.104485](https://doi.org/10.1016/j.meegid.2020.104485) · PMID: [32739464](https://pubmed.ncbi.nlm.nih.gov/32739464/) · PMCID: [PMC7391292](https://pubmed.ncbi.nlm.nih.gov/PMC7391292/)

446. **ABO blood group predisposes to COVID-19 severity and cardiovascular diseases**
Xiaofeng Dai
European Journal of Preventive Cardiology (2020-04-28)
<https://doi.org/ggtv59>
DOI: [10.1177/2047487320922370](https://doi.org/10.1177/2047487320922370) · PMID: [32343152](https://pubmed.ncbi.nlm.nih.gov/32343152/) · PMCID: [PMC7717262](https://pubmed.ncbi.nlm.nih.gov/PMC7717262/)
447. **The influence of ABO blood groups on COVID-19 susceptibility and severity: A molecular hypothesis based on carbohydrate-carbohydrate interactions**
José Caetano Silva-Filho, Cynthia Germoglio Farias de Melo, Janaína Lima de Oliveira
Medical Hypotheses (2020-11) <https://doi.org/gpwf6c>
DOI: [10.1016/j.mehy.2020.110155](https://doi.org/10.1016/j.mehy.2020.110155) · PMID: [33254482](https://pubmed.ncbi.nlm.nih.gov/33254482/) · PMCID: [PMC7395945](https://pubmed.ncbi.nlm.nih.gov/PMC7395945/)
448. **Replicability and Prediction: Lessons and Challenges from GWAS**
Urko M Marigorta, Juan Antonio Rodríguez, Greg Gibson, Arcadi Navarro
Trends in Genetics (2018-07) <https://doi.org/gdqr42>
DOI: [10.1016/j.tig.2018.03.005](https://doi.org/10.1016/j.tig.2018.03.005) · PMID: [29716745](https://pubmed.ncbi.nlm.nih.gov/29716745/) · PMCID: [PMC6003860](https://pubmed.ncbi.nlm.nih.gov/PMC6003860/)
449. **New approaches to population stratification in genome-wide association studies**
Alkes L Price, Noah A Zaitlen, David Reich, Nick Patterson
Nature Reviews Genetics (2010-06-15) <https://doi.org/bw853v>
DOI: [10.1038/nrg2813](https://doi.org/10.1038/nrg2813) · PMID: [20548291](https://pubmed.ncbi.nlm.nih.gov/20548291/) · PMCID: [PMC2975875](https://pubmed.ncbi.nlm.nih.gov/PMC2975875/)
450. **Increased risk of severe clinical course of COVID-19 in carriers of HLA-C*04:01**
January Weiner 3rd, Phillip Suwalski, Manuel Holtgrewe, Alexander Rakitko, Charlotte Thibeault, Melina Müller, Dimitri Patriki, Claudia Quedenau, Ulrike Krüger, Valery Ilinsky, ... Bettina Heidecker
EClinicalMedicine (2021-10) <https://doi.org/gpwf6b>
DOI: [10.1016/j.eclim.2021.101099](https://doi.org/10.1016/j.eclim.2021.101099) · PMID: [34490415](https://pubmed.ncbi.nlm.nih.gov/34490415/) · PMCID: [PMC8410317](https://pubmed.ncbi.nlm.nih.gov/PMC8410317/)
451. **Genomewide Association Study of Severe Covid-19 with Respiratory Failure**
The Severe Covid-19 GWAS Group
New England Journal of Medicine (2020-10-15) <https://doi.org/gg2pqx>
DOI: [10.1056/nejmoa2020283](https://doi.org/10.1056/nejmoa2020283) · PMID: [32558485](https://pubmed.ncbi.nlm.nih.gov/32558485/) · PMCID: [PMC7315890](https://pubmed.ncbi.nlm.nih.gov/PMC7315890/)
452. **The COVID-19 Host Genetics Initiative, a global initiative to elucidate the role of host genetic factors in susceptibility and severity of the SARS-CoV-2 virus pandemic** *European Journal of Human Genetics* (2020-05-13) <https://doi.org/gg8f6j>
DOI: [10.1038/s41431-020-0636-6](https://doi.org/10.1038/s41431-020-0636-6) · PMID: [32404885](https://pubmed.ncbi.nlm.nih.gov/32404885/) · PMCID: [PMC7220587](https://pubmed.ncbi.nlm.nih.gov/PMC7220587/)
453. **The major genetic risk factor for severe COVID-19 is inherited from Neanderthals**
Hugo Zeberg, Svante Pääbo
Nature (2020-09-30) <https://doi.org/gh2s7v>

454. **Neanderthal Introgression at Chromosome 3p21.31 Was Under Positive Natural Selection in East Asians**
Qiliang Ding, Ya Hu, Shuhua Xu, Jiucun Wang, Li Jin
Molecular Biology and Evolution (2013-12-13) <https://doi.org/f5xpfn>
DOI: [10.1093/molbev/mst260](https://doi.org/10.1093/molbev/mst260) · PMID: [24336922](#)
455. **Altered Blood Cell Traits Underlie a Major Genetic Locus of Severe COVID-19**
Jingqi Zhou, Yitang Sun, Weishan Huang, Kaixiong Ye
The Journals of Gerontology: Series A (2021-02-02)
<https://doi.org/gpwf6d>
DOI: [10.1093/gerona/glab035](https://doi.org/10.1093/gerona/glab035) · PMID: [33530099](#) · PMCID: [PMC7929197](#)
456. **Making Sense of Mutation: What D614G Means for the COVID-19 Pandemic Remains Unclear**
Nathan D Grubaugh, William P Hanage, Angela L Rasmussen
Cell (2020-08) <https://doi.org/gg4gqt>
DOI: [10.1016/j.cell.2020.06.040](https://doi.org/10.1016/j.cell.2020.06.040) · PMID: [32697970](#) · PMCID: [PMC7332445](#)
457. **Case Study: Prolonged Infectious SARS-CoV-2 Shedding from an Asymptomatic Immunocompromised Individual with Cancer**
Victoria A Avanzato, MJeremiah Matson, Stephanie N Seifert, Rhys Pryce, Brandi N Williamson, Sarah L Anzick, Kent Barbian, Seth D Judson, Elizabeth R Fischer, Craig Martens, ... Vincent J Munster
Cell (2020-12) <https://doi.org/ghhxkp>
DOI: [10.1016/j.cell.2020.10.049](https://doi.org/10.1016/j.cell.2020.10.049) · PMID: [33248470](#) · PMCID: [PMC7640888](#)
458. **Persistence and Evolution of SARS-CoV-2 in an Immunocompromised Host**
Bina Choi, Manish C Choudhary, James Regan, Jeffrey A Sparks, Robert F Padera, Xuetong Qiu, Isaac H Solomon, Hsiao-Hsuan Kuo, Julie Boucau, Kathryn Bowman, ... Jonathan Z Li
New England Journal of Medicine (2020-12-03) <https://doi.org/fhv8>
DOI: [10.1056/nejmc2031364](https://doi.org/10.1056/nejmc2031364) · PMID: [33176080](#) · PMCID: [PMC7673303](#)
459. **SARS-CoV-2 escape <i>in vitro</i> from a highly neutralizing COVID-19 convalescent plasma**
Emanuele Andreano, Giulia Piccini, Danilo Licastro, Lorenzo Casalino, Nicole V Johnson, Ida Paciello, Simeone Dal Monego, Elisa Pantano, Noemi Manganaro, Alessandro Manenti, ... Rino Rappuoli
Cold Spring Harbor Laboratory (2020-12-28) <https://doi.org/ghs97s>
DOI: [10.1101/2020.12.28.424451](https://doi.org/10.1101/2020.12.28.424451) · PMID: [33398278](#) · PMCID: [PMC7781313](#)
460. **Genetic Variants of SARS-CoV-2—What Do They Mean?**
Adam S Lauring, Emma B Hodcroft
JAMA (2021-01-06) <https://doi.org/ghtbcr>
DOI: [10.1001/jama.2020.27124](https://doi.org/10.1001/jama.2020.27124) · PMID: [33404586](#)
461. **Transmission of SARS-CoV-2 on mink farms between humans and mink and back to humans**

Bas B Oude Munnink, Reina S Sikkema, David F Nieuwenhuijse, Robert Jan Molenaar, Emmanuelle Munger, Richard Molenkamp, Arco van der Spek, Paulien Tolsma, Ariene Rietveld, Miranda Brouwer, ... Marion PG Koopmans
Science (2021-01-08) <https://doi.org/ghssrq>
DOI: [10.1126/science.abe5901](https://doi.org/10.1126/science.abe5901) · PMID: [33172935](#)

462. **Estimated transmissibility and severity of novel SARS-CoV-2 Variant of Concern 202012/01 in England**
Nicholas G Davies, Rosanna C Barnard, Christopher I Jarvis, Adam J Kucharski, James Munday, Carl AB Pearson, Timothy W Russell, Damien C Tully, Sam Abbott, Amy Gimma, ... CMMID COVID-19 Working Group
Cold Spring Harbor Laboratory (2020-12-26) <https://doi.org/fp3v>
DOI: [10.1101/2020.12.24.20248822](https://doi.org/10.1101/2020.12.24.20248822)
463. **Establishment and lineage dynamics of the SARS-CoV-2 epidemic in the UK**
Louis du Plessis, John T McCrone, Alexander E Zarebski, Verity Hill, Christopher Ruis, Bernardo Gutierrez, Jayna Raghwani, Jordan Ashworth, Rachel Colquhoun, Thomas R Connor, ... COVID-19 Genomics UK (COG-UK) Consortium
Science (2021-01-08) <https://doi.org/ghsbdt>
DOI: [10.1126/science.abf2946](https://doi.org/10.1126/science.abf2946) · PMID: [33419936](#)
464. **Learning the language of viral evolution and escape**
Brian Hie, Ellen D Zhong, Bonnie Berger, Bryan Bryson
Science (2021-01-14) <https://doi.org/ghtbcv>
DOI: [10.1126/science.abd7331](https://doi.org/10.1126/science.abd7331) · PMID: [33446556](#)
465. **We shouldn't worry when a virus mutates during disease outbreaks**
Nathan D Grubaugh, Mary E Petrone, Edward C Holmes
Nature Microbiology (2020-02-18) <https://doi.org/ggqsbc>
DOI: [10.1038/s41564-020-0690-4](https://doi.org/10.1038/s41564-020-0690-4) · PMID: [32071422](#) · PMCID: [PMC7095397](#)
466. Zhongming Zhao, Haipeng Li, Xiaozhuang Wu, Yixi Zhong, Keqin Zhang, Ya-Ping Zhang, Eric Boerwinkle, Yun-Xin Fu
BMC Evolutionary Biology (2004) <https://doi.org/d76xw2>
DOI: [10.1186/1471-2148-4-21](https://doi.org/10.1186/1471-2148-4-21) · PMID: [15222897](#) · PMCID: [PMC446188](#)
467. **PHE document**
Ed Collington
(2020-12-21)
https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/947048/Technical_Briefing_VOC_SH_NJL2_SH2.pdf
468. **Preliminary genomic characterisation of an emergent SARS-CoV-2 lineage in the UK defined by a novel set of spike mutations**
Virological
(2020-12-18) <https://virological.org/t/preliminary-genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-the-uk-defined-by-a-novel-set-of-spike-mutations/563>

469. **B.1.1.7 report** https://cov-lineages.org/global_report_B.1.1.7.html
470. **Increased mortality in community-tested cases of SARS-CoV-2 lineage B.1.1.7**
Nicholas G Davies, Christopher I Jarvis, WJohn Edmunds, Nicholas P Jewell, Karla Diaz-Ordaz, Ruth H Keogh
Nature (2021-03-15) <https://doi.org/gjg fsm>
DOI: [10.1038/s41586-021-03426-1](https://doi.org/s41586-021-03426-1) · PMID: [33723411](#)
471. **Identification of a novel SARS-CoV-2 Spike 69-70 deletion lineage circulating in the United States**
Virological
(2020-12-31) <https://virological.org/t/identification-of-a-novel-sars-cov-2-spike-69-70-deletion-lineage-circulating-in-the-united-states/577>
472. **S gene dropout patterns in SARS-CoV-2 tests suggest spread of the H69del/V70del mutation in the US**
Nicole L Washington, Simon White, Kelly MSchiabor Barrett, Elizabeth T Cirulli, Alexandre Bolze, James T Lu
Cold Spring Harbor Laboratory (2020-12-30) <https://doi.org/ghvq46>
DOI: [10.1101/2020.12.24.20248814](https://doi.org/10.1101/2020.12.24.20248814)
473. https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/950823/Variant_of_Concern_VOC_202012_01_Technical_Briefing_3_-_England.pdf
474. **Genetic Variants of SARS-CoV-2 May Lead to False Negative Results with Molecular Tests for Detection of SARS-CoV-2 - Letter to Clinical Laboratory Staff and Health Care Providers**
Center for Devices and Radiological Health
FDA (2021-01-08) <https://www.fda.gov/medical-devices/letters-health-care-providers/genetic-variants-sars-cov-2-may-lead-false-negative-results-molecular-tests-detection-sars-cov-2>
475. **Coronavirus Disease 2019 (COVID-19)**
CDC
Centers for Disease Control and Prevention (2020-02-11)
<https://www.cdc.gov/coronavirus/2019-ncov/more/science-and-research/scientific-brief-emerging-variants.html>
476. **Coronavirus Disease 2019 (COVID-19)**
CDC
Centers for Disease Control and Prevention (2020-02-11)
<https://www.cdc.gov/coronavirus/2019-ncov/variants/variant-classifications.html>
477. **Minister Zweli Mkhize confirms 8 725 more cases of Coronavirus COVID-19 | South African Government**
<https://www.gov.za/speeches/minister-zweli-mkhize-confirms-8-725-more-cases-coronavirus-covid-19-18-dec-2020-0000>
478. **Tracking the international spread of SARS-CoV-2 lineages B.1.1.7 and B.1.351/501Y-V2**
Virological

(2021-02-04) <https://virological.org/t/tracking-the-international-spread-of-sars-cov-2-lineages-b-1-1-7-and-b-1-351-501y-v2/592>

479. **Emergence and rapid spread of a new severe acute respiratory syndrome-related coronavirus 2 (SARS-CoV-2) lineage with multiple spike mutations in South Africa**

Houriyyah Tegally, Eduan Wilkinson, Marta Giovanetti, Arash Iranzadeh, Vagner Fonseca, Jennifer Giandhari, Deelan Doolabh, Sureshnee Pillay, Emmanuel James San, Nokukhanya Msomi, ... Tulio de Oliveira
Cold Spring Harbor Laboratory (2020-12-22) <https://doi.org/fqth>
DOI: [10.1101/2020.12.21.20248640](https://doi.org/10.1101/2020.12.21.20248640)

480. **SARS-CoV-2 501Y.V2 escapes neutralization by South African COVID-19 donor plasma**

Constantinos Kurt Wibmer, Frances Ayres, Tandile Hermanus, Mashudu Madzivhandila, Prudence Kgagudi, Brent Oosthuysen, Bronwen E Lambson, Tulio de Oliveira, Marion Vermeulen, Karin van der Berg, ... Penny L Moore
Nature Medicine (2021-03-02) <https://doi.org/gh7d4s>
DOI: [10.1038/s41591-021-01285-x](https://doi.org/10.1038/s41591-021-01285-x) · PMID: [33654292](#)

481. **Escape of SARS-CoV-2 501Y.V2 from neutralization by convalescent plasma**

Sandile Cele, Inbal Gazy, Laurelle Jackson, Shi-Hsia Hwa, Houriyyah Tegally, Gila Lustig, Jennifer Giandhari, Sureshnee Pillay, Eduan Wilkinson, ...
Nature (2021-03-29) <https://doi.org/f362>
DOI: [10.1038/s41586-021-03471-w](https://doi.org/10.1038/s41586-021-03471-w) · PMID: [33780970](#)

482. **Risk of spread of new SARS-CoV-2 variants of concern in the EU/EEA - first update**

ECDC
(2021-01-21)
<https://www.ecdc.europa.eu/sites/default/files/documents/COVID-19-risk-related-to-spread-of-new-SARS-CoV-2-variants-EU-EEA-first-update.pdf>

483. **Genomic characterisation of an emergent SARS-CoV-2 lineage in Manaus: preliminary findings**

Virological
(2021-01-12) <https://virological.org/t/genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-manaus-preliminary-findings/586>

484. **P.1 report** https://cov-lineages.org/global_report_P.1.html

485. **UK detects 77 cases of South African COVID variant, nine of Brazilian**

Reuters Staff
Reuters (2021-01-24) <https://www.reuters.com/article/uk-health-coronavirus-britain-variants-idUSKBN29T07E>

486. **PANGO lineages** <https://cov-lineages.org/lineages.html>

487. **Emergence of a novel SARS-CoV-2 strain in Southern California, USA**

Wenjuan Zhang, Brian D Davis, Stephanie S Chen, Jorge MSincuir Martinez, Jasmine T Plummer, Eric Vail
Cold Spring Harbor Laboratory (2021-01-20) <https://doi.org/ghvq48>
DOI: [10.1101/2021.01.18.21249786](https://doi.org/10.1101/2021.01.18.21249786)

488. **GISAID - hCov19 Variants** <https://www.gisaid.org/hcov19-variants/>
489. **The Impact of Mutations in SARS-CoV-2 Spike on Viral Infectivity and Antigenicity**
Qianqian Li, Jiajing Wu, Jianhui Nie, Li Zhang, Huan Hao, Shuo Liu, Chenyan Zhao, Qi Zhang, Huan Liu, Lingling Nie, ... Youchun Wang
Cell (2020-09) <https://doi.org/gg4665>
DOI: [10.1016/j.cell.2020.07.012](https://doi.org/10.1016/j.cell.2020.07.012) · PMID: [32730807](#) · PMCID: [PMC7366990](#)
490. **Comprehensive mapping of mutations to the SARS-CoV-2 receptor-binding domain that affect recognition by polyclonal human serum antibodies**
Allison J Greaney, Andrea N Loes, Katharine HD Crawford, Tyler N Starr, Keara D Malone, Helen Y Chu, Jesse D Bloom
Cold Spring Harbor Laboratory (2021-01-04) <https://doi.org/ghr85d>
DOI: [10.1101/2020.12.31.425021](https://doi.org/10.1101/2020.12.31.425021)
491. **Neue Corona-Variante: 35 Fälle in Garmisch-Partenkirchen BR24**
(2021-01-18) <https://www.br.de/nachrichten/bayern/neue-coronavirus-mutation-35-faelle-in-garmisch-partenkirchen,SMQ1V6u>
492. **PANGO lineages** https://cov-lineages.org/global_report.html
493. **Deep Mutational Scanning of SARS-CoV-2 Receptor Binding Domain Reveals Constraints on Folding and ACE2 Binding**
Tyler N Starr, Allison J Greaney, Sarah K Hilton, Daniel Ellis, Katharine HD Crawford, Adam S Dingens, Mary Jane Navarro, John E Bowen, Malejandra Tortorici, Alexandra C Walls, ... Jesse D Bloom
Cell (2020-09) <https://doi.org/gg72tr>
DOI: [10.1016/j.cell.2020.08.012](https://doi.org/10.1016/j.cell.2020.08.012) · PMID: [32841599](#) · PMCID: [PMC7418704](#)
494. **Adaptation of SARS-CoV-2 in BALB/c mice for testing vaccine efficacy**
Hongjing Gu, Qi Chen, Guan Yang, Lei He, Hang Fan, Yong-Qiang Deng, Yanxiao Wang, Yue Teng, Zhongpeng Zhao, Yujun Cui, ... Yusen Zhou
Science (2020-07-30) <https://doi.org/ghc5mn>
DOI: [10.1126/science.abc4730](https://doi.org/10.1126/science.abc4730) · PMID: [32732280](#) · PMCID: [PMC7574913](#)
495. **Complete Mapping of Mutations to the SARS-CoV-2 Spike Receptor-Binding Domain that Escape Antibody Recognition**
Allison J Greaney, Tyler N Starr, Pavlo Gilchuk, Seth J Zost, Elad Binshtain, Andrea N Loes, Sarah K Hilton, John Huddleston, Rachel Egua, Katharine HD Crawford, ... Jesse D Bloom
Cell Host & Microbe (2021-01) <https://doi.org/ghvq3m>
DOI: [10.1016/j.chom.2020.11.007](https://doi.org/10.1016/j.chom.2020.11.007) · PMID: [33259788](#) · PMCID: [PMC7676316](#)

496. **Recurrent deletions in the SARS-CoV-2 spike glycoprotein drive antibody escape**
Kevin R McCarthy, Linda J Rennick, Sham Nambulli, Lindsey R Robinson-McCarthy, William G Bain, Ghady Haidar, WPaul Duprex
Cold Spring Harbor Laboratory (2021-01-19) <https://doi.org/ghvq44>
DOI: [10.1101/2020.11.19.389916](https://doi.org/10.1101/2020.11.19.389916)
497. **Viral mutations may cause another ‘very, very bad’ COVID-19 wave, scientists warn**
Kai Kupferschmidt
Science (2021-01-05) <https://doi.org/ghvq5b>
DOI: [10.1126/science.abg4312](https://doi.org/10.1126/science.abg4312)
498. **SARS-CoV-2 reinfection by the new Variant of Concern (VOC) P.1 in Amazonas, Brazil**
Virological
(2021-01-18) <https://virological.org/t/sars-cov-2-reinfection-by-the-new-variant-of-concern-voc-p-1-in-amazonas-brazil/596>
499. **Fast-spreading COVID variant can elude immune responses**
Ewen Callaway
Nature (2021-01-21) <https://doi.org/ght924>
DOI: [10.1038/d41586-021-00121-z](https://doi.org/10.1038/d41586-021-00121-z) · PMID: [33479534](#)
500. **Prospective mapping of viral mutations that escape antibodies used to treat COVID-19**
Tyler N Starr, Allison J Greaney, Amin Addetia, William W Hannon, Manish C Choudhary, Adam S Dingens, Jonathan Z Li, Jesse D Bloom
Science (2021-01-25) <https://doi.org/ghvntq>
DOI: [10.1126/science.abf9302](https://doi.org/10.1126/science.abf9302) · PMID: [33495308](#)
501. **New mutations raise specter of ‘immune escape’**
Kai Kupferschmidt
Science (2021-01-21) <https://doi.org/ght923>
DOI: [10.1126/science.371.6527.329](https://doi.org/10.1126/science.371.6527.329) · PMID: [33479129](#)
502. **mRNA-1273 vaccine induces neutralizing antibodies against spike mutants from global SARS-CoV-2 variants**
Kai Wu, Anne P Werner, Juan I Moliva, Matthew Koch, Angela Choi, Guillaume BE Stewart-Jones, Hamilton Bennett, Seyhan Boyoglu-Barnum, Wei Shi, Barney S Graham, ... Darin K Edwards
Cold Spring Harbor Laboratory (2021-01-25) <https://doi.org/fr2g>
DOI: [10.1101/2021.01.25.427948](https://doi.org/10.1101/2021.01.25.427948) · PMID: [33501442](#) · PMCID: [PMC7836112](#)
503. **Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine**
Fernando P Polack, Stephen J Thomas, Nicholas Kitchin, Judith Absalon, Alejandra Gurtman, Stephen Lockhart, John L Perez, Gonzalo Pérez Marc, Edson D Moreira, Cristiano Zerbini, ... William C Gruber
New England Journal of Medicine (2020-12-31) <https://doi.org/ghn625>
DOI: [10.1056/nejmoa2034577](https://doi.org/10.1056/nejmoa2034577) · PMID: [33301246](#) · PMCID: [PMC7745181](#)
504. **Safety and Immunogenicity of Two RNA-Based Covid-19 Vaccine Candidates**

Edward E Walsh, Robert W Frenck, Ann R Falsey, Nicholas Kitchin, Judith Absalon, Alejandra Gurtman, Stephen Lockhart, Kathleen Neuzil, Mark J Mulligan, Ruth Bailey, ... William C Gruber
New England Journal of Medicine (2020-12-17) <https://doi.org/ghjktx>
DOI: [10.1056/nejmoa2027906](https://doi.org/10.1056/nejmoa2027906) · PMID: [33053279](https://pubmed.ncbi.nlm.nih.gov/33053279/) · PMCID: [PMC7583697](https://pubmed.ncbi.nlm.nih.gov/PMC7583697/)

505. **Neutralization of N501Y mutant SARS-CoV-2 by BNT162b2 vaccine-elicited sera**

Xuping Xie, Jing Zou, Camila R Fontes-Garfias, Hongjie Xia, Kena A Swanson, Mark Cutler, David Cooper, Vineet D Menachery, Scott Weaver, Philip R Dormitzer, Pei-Yong Shi
Cold Spring Harbor Laboratory (2021-01-07) <https://doi.org/ghvq47>
DOI: [10.1101/2021.01.07.425740](https://doi.org/10.1101/2021.01.07.425740) · PMID: [33442691](https://pubmed.ncbi.nlm.nih.gov/33442691/) · PMCID: [PMC7805448](https://pubmed.ncbi.nlm.nih.gov/PMC7805448/)

506. **mRNA vaccine-elicited antibodies to SARS-CoV-2 and circulating variants**

Zijun Wang, Fabian Schmidt, Yiska Weisblum, Frauke Muecksch, Christopher O Barnes, Shlomo Finkin, Dennis Schaefer-Babajew, Melissa Cipolla, Christian Gaebler, Jenna A Lieberman, ... Michel C Nussenzweig
Cold Spring Harbor Laboratory (2021-01-30) <https://doi.org/frdn>
DOI: [10.1101/2021.01.15.426911](https://doi.org/10.1101/2021.01.15.426911) · PMID: [33501451](https://pubmed.ncbi.nlm.nih.gov/33501451/) · PMCID: [PMC7836122](https://pubmed.ncbi.nlm.nih.gov/PMC7836122/)

507. **Neutralization of SARS-CoV-2 Variants B.1.429 and B.1.351**

Xiaoying Shen, Haili Tang, Rolando Pajon, Gale Smith, Gregory M Glenn, Wei Shi, Bette Korber, David C Montefiori
New England Journal of Medicine (2021-06-17) <https://doi.org/f5kc>
DOI: [10.1056/nejmc2103740](https://doi.org/10.1056/nejmc2103740) · PMID: [33826819](https://pubmed.ncbi.nlm.nih.gov/33826819/) · PMCID: [PMC8063884](https://pubmed.ncbi.nlm.nih.gov/PMC8063884/)

508. **SARS-CoV-2 spike E484K mutation reduces antibody neutralisation**

Sonia Jangra, Chengjin Ye, Raveen Rathnasinghe, Daniel Stadlbauer, Florian Krammer, Viviana Simon, Luis Martinez-Sobrido, Adolfo García-Sastre, Michael Schotsaert, Hala Alshammary, ... Komal Srivastava
The Lancet Microbe (2021-04) <https://doi.org/f53t>
DOI: [10.1016/s2666-5247\(21\)00068-9](https://doi.org/10.1016/s2666-5247(21)00068-9) · PMID: [33846703](https://pubmed.ncbi.nlm.nih.gov/33846703/) · PMCID: [PMC8026167](https://pubmed.ncbi.nlm.nih.gov/PMC8026167/)

509. **Sensitivity of SARS-CoV-2 B.1.1.7 to mRNA vaccine-elicited antibodies**

Dami A Collier, Anna De Marco, Isabella ATM Ferreira, Bo Meng, Rawlings P Dahir, Alexandra C Walls, Steven A Kemp, Jessica Bassi, Dora Pinto, ...
Nature (2021-03-11) <https://doi.org/gjm7v5>
DOI: [10.1038/s41586-021-03412-7](https://doi.org/10.1038/s41586-021-03412-7) · PMID: [33706364](https://pubmed.ncbi.nlm.nih.gov/33706364/)

510. **A SARS-CoV-2 vaccine candidate would likely match all currently circulating variants**

Bethany Dearlove, Eric Lewitus, Hongjun Bai, Yifan Li, Daniel B Reeves, MGordon Joyce, Paul T Scott, Mihret F Amare, Sandhya Vasan, Nelson L Michael, ... Morgane Rolland
Proceedings of the National Academy of Sciences (2020-09-22) <https://doi.org/fdkz>

511. **Unlocking capacities of genomics for the COVID-19 response and future pandemics**

Sergey Knyazev, Karishma Chhugani, Varuni Sarwal, Ram Ayyala, Harman Singh, Smruthi Karthikeyan, Dhrithi Deshpande, Pelin Icer Baykal, Zoia Comarova, Angela Lu, ... Serghei Mangul
Nature Methods (2022-04) <https://doi.org/gpvs87>
DOI: [10.1038/s41592-022-01444-z](https://doi.org/10.1038/s41592-022-01444-z) · PMID: [35396471](https://pubmed.ncbi.nlm.nih.gov/35396471/)

512. **Pandemics- One Health preparedness for the next**

Frank M Aarestrup, Marc Bonten, Marion Koopmans
The Lancet Regional Health - Europe (2021-10) <https://doi.org/gp3nsz>
DOI: [10.1016/j.lanepe.2021.100210](https://doi.org/10.1016/j.lanepe.2021.100210) · PMID: [34642673](https://pubmed.ncbi.nlm.nih.gov/34642673/) · PMCID: [PMC8495373](https://pubmed.ncbi.nlm.nih.gov/PMC8495373/)

513. **The emergence, genomic diversity and global spread of SARS-CoV-2**

Juan Li, Shengjie Lai, George F Gao, Weifeng Shi
Nature (2021-12-08) <https://doi.org/gns3bb>
DOI: [10.1038/s41586-021-04188-6](https://doi.org/10.1038/s41586-021-04188-6) · PMID: [34880490](https://pubmed.ncbi.nlm.nih.gov/34880490/)

514. **Accelerating genomics-based surveillance for COVID-19 response in Africa**

Sofonias K Tessema, Seth C Inzaule, Alan Christoffels, Yenew Kebede, Tulio de Oliveira, Ahmed EOgwell Ouma, Christian T Happi, John N Nkengasong
The Lancet Microbe (2020-10) <https://doi.org/hskp>
DOI: [10.1016/s2666-5247\(20\)30117-8](https://doi.org/10.1016/s2666-5247(20)30117-8) · PMID: [32838350](https://pubmed.ncbi.nlm.nih.gov/32838350/) · PMCID: [PMC7434434](https://pubmed.ncbi.nlm.nih.gov/PMC7434434/)

515. **Computational analysis of SARS-CoV-2/COVID-19 surveillance by wastewater-based epidemiology locally and globally: Feasibility, economy, opportunities and challenges**

Olga E Hart, Rolf U Halden
Science of The Total Environment (2020-08) <https://doi.org/ds22>
DOI: [10.1016/j.scitotenv.2020.138875](https://doi.org/10.1016/j.scitotenv.2020.138875) · PMID: [32371231](https://pubmed.ncbi.nlm.nih.gov/32371231/) · PMCID: [PMC7175865](https://pubmed.ncbi.nlm.nih.gov/PMC7175865/)

516. **Implementation of environmental surveillance for SARS-CoV-2 virus to support public health decisions: Opportunities and challenges**

Gertjan Medema, Frederic Been, Leo Heijnen, Susan Petterson
Current Opinion in Environmental Science & Health (2020-10) <https://doi.org/gjpjh2>
DOI: [10.1016/j.coesh.2020.09.006](https://doi.org/10.1016/j.coesh.2020.09.006) · PMID: [33024908](https://pubmed.ncbi.nlm.nih.gov/33024908/) · PMCID: [PMC7528975](https://pubmed.ncbi.nlm.nih.gov/PMC7528975/)

517. **Surveillance of SARS-CoV-2 RNA in wastewater: Methods optimization and quality control are crucial for generating reliable public health information**

Warish Ahmed, Aaron Bivins, Paul M Bertsch, Kyle Bibby, Phil M Choi, Kata Farkas, Pradip Gyawali, Kerry A Hamilton, Eiji Haramoto, Masaaki Kitajima, ... Jochen F Mueller

518. **Prolonged presence of SARS-CoV-2 viral RNA in faecal samples**

Yongjian Wu, Cheng Guo, Lantian Tang, Zhongsi Hong, Jianhui Zhou, Xin Dong, Huan Yin, Qiang Xiao, Yanping Tang, Xiujuan Qu, ... Xi Huang
The Lancet Gastroenterology & Hepatology (2020-05)

<https://doi.org/ggg8zp>

DOI: [10.1016/s2468-1253\(20\)30083-2](https://doi.org/10.1016/s2468-1253(20)30083-2) · PMID: [32199469](https://pubmed.ncbi.nlm.nih.gov/32199469/) · PMCID:

[PMC7158584](https://pubmed.ncbi.nlm.nih.gov/PMC7158584/)

519. **Protocol for safe, affordable, and reproducible isolation and quantitation of SARS-CoV-2 RNA from wastewater**

Monica Trujillo, Kristen Cheung, Anna Gao, Irene Hoxie, Sherin Kannoly, Nanami Kubota, Kaung Myat San, Davida S Smyth, John J Dennehy

PLOS ONE (2021-09-23) <https://doi.org/gnnr74>

DOI: [10.1371/journal.pone.0257454](https://doi.org/10.1371/journal.pone.0257454) · PMID: [34555079](https://pubmed.ncbi.nlm.nih.gov/34555079/) · PMCID:

[PMC8459947](https://pubmed.ncbi.nlm.nih.gov/PMC8459947/)

520. **Monitoring SARS-CoV-2 in wastewater during New York City's second wave of COVID-19: sewershed-level trends and relationships to publicly available clinical testing data**

Catherine Hoar, Francoise Chauvin, Alexander Clare, Hope McGibbon, Esmervaldo Castro, Samantha Patinella, Dimitrios Katehis, John J Dennehy, Monica Trujillo, Davida S Smyth, Andrea I Silverman

Environmental Science: Water Research & Technology (2022)

<https://doi.org/gp32w6>

DOI: [10.1039/d1ew00747e](https://doi.org/10.1039/d1ew00747e)

521. **Early-pandemic wastewater surveillance of SARS-CoV-2 in Southern Nevada: Methodology, occurrence, and incidence/prevalence considerations**

Daniel Gerrity, Katerina Papp, Mitchell Stoker, Alan Sims, Wilbur Frehner

Water Research X (2021-01) <https://doi.org/gnnr5j>

DOI: [10.1016/j.wroa.2020.100086](https://doi.org/10.1016/j.wroa.2020.100086) · PMID: [33398255](https://pubmed.ncbi.nlm.nih.gov/33398255/) · PMCID:

[PMC7774458](https://pubmed.ncbi.nlm.nih.gov/PMC7774458/)

522. **SARS-CoV-2 Titers in Wastewater Are Higher than Expected from Clinically Confirmed Cases**

Fuqing Wu, Jianbo Zhang, Amy Xiao, Xiaoqiong Gu, Wei Lin Lee, Federica Armas, Kathryn Kauffman, William Hanage, Mariana Matus, Newsha Ghaeli, ... Eric J Alm

mSystems (2020-08-25) <https://doi.org/gg5tgt>

DOI: [10.1128/msystems.00614-20](https://doi.org/10.1128/msystems.00614-20) · PMID: [32694130](https://pubmed.ncbi.nlm.nih.gov/32694130/) · PMCID:

[PMC7566278](https://pubmed.ncbi.nlm.nih.gov/PMC7566278/)

523. **Wastewater surveillance using ddPCR reveals highly accurate tracking of Omicron variant due to altered N1 probe binding efficiency**

Melissa K Schussman, Adelaide Roguet, Angela Schmoldt, Brooke Dinan, Sandra L McLellan

524. **Quantitative detection of SARS-CoV-2 Omicron BA.1 and BA.2 variants in wastewater through allele-specific RT-qPCR**
Wei Lin Lee, Xiaoqiong Gu, Federica Armas, Fuqing Wu, Franciscus Chandra, Hongjie Chen, Amy Xiao, Mats Leifels, Feng Jun Desmond Chua, Germaine WC Kwok, ... Eric J Alm
Cold Spring Harbor Laboratory (2021-12-21) <https://doi.org/gp37v5>
DOI: [10.1101/2021.12.21.21268077](https://doi.org/10.1101/2021.12.21.21268077)
525. **National Wastewater Surveillance System**
CDC
Centers for Disease Control and Prevention (2022-03-21)
<https://www.cdc.gov/healthywater/surveillance/wastewater-surveillance/wastewater-surveillance.html>
526. **Genomic surveillance at scale is required to detect newly emerging strains at an early timepoint**
Darcy Vavrek, Lucia Speroni, Kirsten J Curnow, Michael Oberholzer, Vanessa Moeder, Phillip G Febbo
Cold Spring Harbor Laboratory (2021-01-15) <https://doi.org/gp3ns3>
DOI: [10.1101/2021.01.12.21249613](https://doi.org/10.1101/2021.01.12.21249613)
527. **Ecology and economics for pandemic prevention**
Andrew P Dobson, Stuart L Pimm, Lee Hannah, Les Kaufman, Jorge A Ahumada, Amy W Ando, Aaron Bernstein, Jonah Busch, Peter Daszak, Jens Engelmann, ... Mariana M Vale
Science (2020-07-24) <https://doi.org/gk6szk>
DOI: [10.1126/science.abc3189](https://doi.org/10.1126/science.abc3189) · PMID: [32703868](#)
528. **Climate change increases cross-species viral transmission risk**
Colin J Carlson, Gregory F Albery, Cory Merow, Christopher H Trisos, Casey M Zipfel, Evan A Eskew, Kevin J Olival, Noam Ross, Shweta Bansal
Nature (2022-04-28) <https://doi.org/hrxm>
DOI: [10.1038/s41586-022-04788-w](https://doi.org/10.1038/s41586-022-04788-w) · PMID: [35483403](#)
529. **How urbanization affects the epidemiology of emerging infectious diseases**
Carl-Johan Neiderud
Infection Ecology & Epidemiology (2015-01) <https://doi.org/gd52fb>
DOI: [10.3402/iee.v5.27060](https://doi.org/10.3402/iee.v5.27060) · PMID: [26112265](#) · PMCID: [PMC4481042](#)
530. **Aggressively find, test, trace and isolate to beat COVID-19**
Larissa M Matukas, Irfan A Dhalla, Andreas Laupacis
Canadian Medical Association Journal (2020-09-09)
<https://doi.org/gh2jvk>
DOI: [10.1503/cmaj.202120](https://doi.org/10.1503/cmaj.202120) · PMID: [32907821](#) · PMCID: [PMC7546740](#)
531. **COVID-19 in New Zealand and the impact of the national response: a descriptive epidemiological study**
Sarah Jefferies, Nigel French, Charlotte Gilkison, Giles Graham, Virginia Hope, Jonathan Marshall, Caroline McElnay, Andrea McNeill, Petra Muellner, Shevaun Paine, ... Patricia Priest

532. **Potential lessons from the Taiwan and New Zealand health responses to the COVID-19 pandemic**
Jennifer Summers, Hao-Yuan Cheng, Hsien-Ho Lin, Lucy Telfar Barnard, Amanda Kvalsvig, Nick Wilson, Michael G Baker
The Lancet Regional Health - Western Pacific (2020-11)
<https://doi.org/ghrbz4>
DOI: [10.1016/j.lanwpc.2020.100044](https://doi.org/10.1016/j.lanwpc.2020.100044) · PMID: [34013216](#) · PMCID:
[PMC7577184](#)
533. **Novel Coronavirus – China**
<https://www.who.int/emergencies/diseases-outbreak-news/item/2020-DON233>
534. **Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR**
Victor M Corman, Olfert Landt, Marco Kaiser, Richard Molenkamp, Adam Meijer, Daniel KW Chu, Tobias Bleicker, Sebastian Brünink, Julia Schneider, Marie Luisa Schmidt, ... Christian Drosten
Eurosurveillance (2020-01-23) <https://doi.org/ggjs7g>
DOI: [10.2807/1560-7917.es.2020.25.3.2000045](https://doi.org/10.2807/1560-7917.es.2020.25.3.2000045) · PMID: [31992387](#) ·
PMCID: [PMC6988269](#)
535. **Polymerase Chain Reaction**
Lilit Garibyan, Nidhi Avashia
Journal of Investigative Dermatology (2013-03) <https://doi.org/ggtkkc>
DOI: [10.1038/jid.2013.1](https://doi.org/10.1038/jid.2013.1) · PMID: [23399825](#) · PMCID: [PMC4102308](#)
536. **Development and application of lateral flow test strip technology for detection of infectious agents and chemical contaminants: a review**
Babacar Ngom, Yancheng Guo, Xiliang Wang, Dingren Bi
Analytical and Bioanalytical Chemistry (2010-04-27)
<https://doi.org/cn8cn9>
DOI: [10.1007/s00216-010-3661-4](https://doi.org/10.1007/s00216-010-3661-4) · PMID: [20422164](#)
537. **Enzyme Linked Immunosorbent Assay**
Mandy Alhajj, Aisha Farhana
StatPearls (2022-02-02) <https://pubmed.ncbi.nlm.nih.gov/32310382>
PMID: [32310382](#)
538. **Evaluation and Comparison of Serological Methods for COVID-19 Diagnosis**
Fanwu Gong, Hua-xing Wei, Qiangsheng Li, Liu Liu, Bofeng Li
Frontiers in Molecular Biosciences (2021-07-23) <https://doi.org/gnxn82>
DOI: [10.3389/fmolb.2021.682405](https://doi.org/10.3389/fmolb.2021.682405) · PMID: [34368226](#) · PMCID:
[PMC8343015](#)
539. **Sensitivity in Detection of Antibodies to Nucleocapsid and Spike Proteins of Severe Acute Respiratory Syndrome Coronavirus 2 in Patients With Coronavirus Disease 2019**

Peter D Burbelo, Francis X Riedo, Chihiro Morishima, Stephen Rawlings, Davey Smith, Sanchita Das, Jeffrey R Strich, Daniel S Chertow, Richard T Davey Jr, Jeffrey I Cohen
The Journal of Infectious Diseases (2020-05-19) <https://doi.org/ggxz2f>
DOI: [10.1093/infdis/jiaa273](https://doi.org/10.1093/infdis/jiaa273) · PMID: [32427334](https://pubmed.ncbi.nlm.nih.gov/32427334/) · PMCID: [PMC7313936](https://pubmed.ncbi.nlm.nih.gov/PMC7313936/)

540. **Temporal profiles of viral load in posterior oropharyngeal saliva samples and serum antibody responses during infection by SARS-CoV-2: an observational cohort study**
Kelvin Kai-Wang To, Owen Tak-Yin Tsang, Wai-Shing Leung, Anthony Raymond Tam, Tak-Chiu Wu, David Christopher Lung, Cyril Chik-Yan Yip, Jian-Piao Cai, Jacky Man-Chun Chan, Thomas Shiu-Hong Chik, ... Kwok-Yung Yuen
The Lancet Infectious Diseases (2020-05) <https://doi.org/ggp4qx>
DOI: [10.1016/s1473-3099\(20\)30196-1](https://doi.org/10.1016/s1473-3099(20)30196-1) · PMID: [32213337](https://pubmed.ncbi.nlm.nih.gov/32213337/) · PMCID: [PMC7158907](https://pubmed.ncbi.nlm.nih.gov/PMC7158907/)
541. **Fecal specimen diagnosis 2019 novel coronavirus-infected pneumonia**
JingCheng Zhang, SaiBin Wang, YaDong Xue
Journal of Medical Virology (2020-03-12) <https://doi.org/ggpx6d>
DOI: [10.1002/jmv.25742](https://doi.org/10.1002/jmv.25742) · PMID: [32124995](https://pubmed.ncbi.nlm.nih.gov/32124995/)
542. **A beginner's guide to RT-PCR, qPCR and RT-qPCR**
Grace Adams
The Biochemist (2020-06-15) <https://doi.org/gm8nfz>
DOI: [10.1042/bio20200034](https://doi.org/10.1042/bio20200034)
543. **The MIQE Guidelines: Minimum Information for Publication of Quantitative Real-Time PCR Experiments**
Stephen A Bustin, Vladimir Benes, Jeremy A Garson, Jan Hellemans, Jim Huggett, Mikael Kubista, Reinhold Mueller, Tania Nolan, Michael W Pfaffl, Gregory L Shipley, ... Carl T Wittwer
Clinical Chemistry (2009-04-01) <https://doi.org/fkkjq5>
DOI: [10.1373/clinchem.2008.112797](https://doi.org/10.1373/clinchem.2008.112797) · PMID: [19246619](https://pubmed.ncbi.nlm.nih.gov/19246619/)
544. **Molecular Diagnosis of a Novel Coronavirus (2019-nCoV) Causing an Outbreak of Pneumonia**
Daniel KW Chu, Yang Pan, Samuel MS Cheng, Kenrie PY Hui, Pavithra Krishnan, Yingzhi Liu, Daisy YM Ng, Carrie KC Wan, Peng Yang, Quanyi Wang, ... Leo LM Poon
Clinical Chemistry (2020-04) <https://doi.org/ggnbpp>
DOI: [10.1093/clinchem/hvaa029](https://doi.org/10.1093/clinchem/hvaa029) · PMID: [32031583](https://pubmed.ncbi.nlm.nih.gov/32031583/) · PMCID: [PMC7108203](https://pubmed.ncbi.nlm.nih.gov/PMC7108203/)
545. **Viral RNA load as determined by cell culture as a management tool for discharge of SARS-CoV-2 patients from infectious disease wards**
Bernard La Scola, Marion Le Bideau, Julien Andreani, Van Thuan Hoang, Clio Grimaldier, Philippe Colson, Philippe Gautret, Didier Raoult
European Journal of Clinical Microbiology & Infectious Diseases (2020-04-27) <https://doi.org/ghf8cj>
DOI: [10.1007/s10096-020-03913-9](https://doi.org/10.1007/s10096-020-03913-9) · PMID: [32342252](https://pubmed.ncbi.nlm.nih.gov/32342252/) · PMCID: [PMC7185831](https://pubmed.ncbi.nlm.nih.gov/PMC7185831/)

546. **Duration and key determinants of infectious virus shedding in hospitalized patients with coronavirus disease-2019 (COVID-19)**
Jeroen JA van Kampen, David AMC van de Vijver, Pieter LA Fraaij, Bart L Haagmans, Mart M Lamers, Nisreen Okba, Johannes PC van den Akker, Henrik Endeman, Diederik AMPJ Gommers, Jan J Cornelissen, ...
Annemiek A van der Eijk
Nature Communications (2021-01-11) <https://doi.org/gjm8g2>
DOI: [10.1038/s41467-020-20568-4](https://doi.org/10.1038/s41467-020-20568-4) · PMID: [33431879](https://pubmed.ncbi.nlm.nih.gov/33431879/) · PMCID: [PMC7801729](https://pubmed.ncbi.nlm.nih.gov/PMC7801729/)
547. **Ct values from SARS-CoV-2 diagnostic PCR assays should not be used as direct estimates of viral load**
Elias Dahdouh, Fernando Lázaro-Perona, María Pilar Romero-Gómez, Jesús Mingorance, Julio García-Rodríguez
Journal of Infection (2021-03) <https://doi.org/gjkr5s>
DOI: [10.1016/j.jinf.2020.10.017](https://doi.org/10.1016/j.jinf.2020.10.017) · PMID: [33131699](https://pubmed.ncbi.nlm.nih.gov/33131699/) · PMCID: [PMC7585367](https://pubmed.ncbi.nlm.nih.gov/PMC7585367/)
548. **Estimating the false-negative test probability of SARS-CoV-2 by RT-PCR**
Paul S Wikramaratna, Robert S Paton, Mahan Ghafari, José Lourenço
Eurosurveillance (2020-12-17) <https://doi.org/gmb6n3>
DOI: [10.2807/1560-7917.es.2020.25.50.2000568](https://doi.org/10.2807/1560-7917.es.2020.25.50.2000568) · PMID: [33334398](https://pubmed.ncbi.nlm.nih.gov/33334398/) · PMCID: [PMC7812420](https://pubmed.ncbi.nlm.nih.gov/PMC7812420/)
549. **Diagnosing SARS-CoV-2 infection: the danger of over-reliance on positive test results**
Andrew N Cohen, Bruce Kessel, Michael G Milgroom
Cold Spring Harbor Laboratory (2020-05-01) <https://doi.org/gh3xk7>
DOI: [10.1101/2020.04.26.20080911](https://doi.org/10.1101/2020.04.26.20080911)
550. **Diagnostic performance of different sampling approaches for SARS-CoV-2 RT-PCR testing: a systematic review and meta-analysis**
Nicole Ngai Yung Tsang, Hau Chi So, Ka Yan Ng, Benjamin J Cowling, Gabriel M Leung, Dennis Kai Ming Ip
The Lancet Infectious Diseases (2021-09) <https://doi.org/gjrfns>
DOI: [10.1016/s1473-3099\(21\)00146-8](https://doi.org/10.1016/s1473-3099(21)00146-8) · PMID: [33857405](https://pubmed.ncbi.nlm.nih.gov/33857405/) · PMCID: [PMC8041361](https://pubmed.ncbi.nlm.nih.gov/PMC8041361/)
551. **Improved Molecular Diagnosis of COVID-19 by the Novel, Highly Sensitive and Specific COVID-19-RdRp/HeI Real-Time Reverse Transcription-PCR Assay Validated In Vitro and with Clinical Specimens**
Jasper Fuk-Woo Chan, Cyril Chik-Yan Yip, Kelvin Kai-Wang To, Tommy Hing-Cheung Tang, Sally Cheuk-Ying Wong, Kit-Hang Leung, Agnes Yim-Fong Fung, Anthony Chin-Ki Ng, Zijiao Zou, Hoi-Wah Tsoi, ... Kwok-Yung Yuen
Journal of Clinical Microbiology (2020-04-23) <https://doi.org/ggpv64>
DOI: [10.1128/jcm.00310-20](https://doi.org/10.1128/jcm.00310-20) · PMID: [32132196](https://pubmed.ncbi.nlm.nih.gov/32132196/) · PMCID: [PMC7180250](https://pubmed.ncbi.nlm.nih.gov/PMC7180250/)
552. **Testing at scale during the COVID-19 pandemic**
Tim R Mercer, Marc Salit
Nature Reviews Genetics (2021-05-04) <https://doi.org/gjvw2n>
DOI: [10.1038/s41576-021-00360-w](https://doi.org/10.1038/s41576-021-00360-w) · PMID: [33948037](https://pubmed.ncbi.nlm.nih.gov/33948037/) · PMCID: [PMC8094986](https://pubmed.ncbi.nlm.nih.gov/PMC8094986/)

553. **SARS-CoV-2 samples may escape detection because of a single point mutation in the N gene**
Katharina Ziegler, Philipp Steininger, Renate Ziegler, Jörg Steinmann, Klaus Korn, Armin Ensser
Eurosurveillance (2020-10-01) <https://doi.org/ghnwss>
DOI: [10.2807/1560-7917.es.2020.25.39.2001650](https://doi.org/10.2807/1560-7917.es.2020.25.39.2001650) · PMID: [33006300](#) ·
PMCID: [PMC7531073](#)
554. **Multiple assays in a real-time RT-PCR SARS-CoV-2 panel can mitigate the risk of loss of sensitivity by new genomic variants during the COVID-19 outbreak**
Luis Peñarrubia, Maria Ruiz, Roberto Porco, Sonia N Rao, Martí Juanola-Falgarona, Davide Manissero, Marta López-Fontanals, Josep Pareja
International Journal of Infectious Diseases (2020-08)
<https://doi.org/gmb6rv>
DOI: [10.1016/j.ijid.2020.06.027](https://doi.org/10.1016/j.ijid.2020.06.027) · PMID: [32535302](#) · PMCID:
[PMC7289722](#)
555. **Droplet Digital PCR versus qPCR for gene expression analysis with low abundant targets: from variable nonsense to publication quality data**
Sean C Taylor, Genevieve Laperriere, Hugo Germain
Scientific Reports (2017-05-25) <https://doi.org/gbgsjw>
DOI: [10.1038/s41598-017-02217-x](https://doi.org/10.1038/s41598-017-02217-x) · PMID: [28546538](#) · PMCID:
[PMC5445070](#)
556. **dPCR: A Technology Review**
Phenix-Lan Quan, Martin Sauzade, Eric Brouzes
Sensors (2018-04-20) <https://doi.org/ggr39c>
DOI: [10.3390/s18041271](https://doi.org/10.3390/s18041271) · PMID: [29677144](#) · PMCID: [PMC5948698](#)
557. **ddPCR: a more accurate tool for SARS-CoV-2 detection in low viral load specimens**
Tao Suo, Xinjin Liu, Jiangpeng Feng, Ming Guo, Wenjia Hu, Dong Guo, Hafiz Ullah, Yang Yang, Qiuhan Zhang, Xin Wang, ... Yu Chen
Emerging Microbes & Infections (2020-06-07) <https://doi.org/ggx2t2>
DOI: [10.1080/22221751.2020.1772678](https://doi.org/10.1080/22221751.2020.1772678) · PMID: [32438868](#) · PMCID:
[PMC7448897](#)
558. **Highly accurate and sensitive diagnostic detection of SARS-CoV-2 by digital PCR**
Lianhua Dong, Junbo Zhou, Chunyan Niu, Quanyi Wang, Yang Pan, Sitong Sheng, Xia Wang, Yongzhuo Zhang, Jiayi Yang, Manqing Liu, ... Xiang Fang
Talanta (2021-03) <https://doi.org/gh2jy>
DOI: [10.1016/j.talanta.2020.121726](https://doi.org/10.1016/j.talanta.2020.121726) · PMID: [33379001](#) · PMCID:
[PMC7588801](#)
559. **Library preparation for next generation sequencing: A review of automation strategies**
JF Hess, TA Kohl, M Kotrová, K Rönsch, T Paprotka, V Mohr, T Hutzenlaub, M Brüggemann, R Zengerle, S Niemann, N Paust
Biotechnology Advances (2020-07) <https://doi.org/ggth2v>
DOI: [10.1016/j.biotechadv.2020.107537](https://doi.org/10.1016/j.biotechadv.2020.107537) · PMID: [32199980](#)

560. **Diagnosing COVID-19: The Disease and Tools for Detection**
Buddhisha Udugama, Pranav Kadhiiresan, Hannah N Kozlowski, Ayden Malekjahani, Matthew Osborne, Vanessa YC Li, Hongmin Chen, Samira Mubareka, Jonathan B Gubbay, Warren CW Chan
ACS Nano (2020-03-30) <https://doi.org/ggq8ds>
DOI: [10.1021/acsnano.0c02624](https://doi.org/10.1021/acsnano.0c02624) · PMID: [32223179](https://pubmed.ncbi.nlm.nih.gov/32223179/) · PMCID: [PMC7144809](https://pubmed.ncbi.nlm.nih.gov/PMC7144809/)
561. **Genomic surveillance to combat COVID-19: challenges and opportunities**
Janet D Robishaw, Scott M Alter, Joshua J Solano, Richard D Shih, David L DeMets, Dennis G Maki, Charles H Hennekens
The Lancet Microbe (2021-09) <https://doi.org/hkwr>
DOI: [10.1016/s2666-5247\(21\)00121-x](https://doi.org/10.1016/s2666-5247(21)00121-x) · PMID: [34337584](https://pubmed.ncbi.nlm.nih.gov/34337584/) · PMCID: [PMC8315763](https://pubmed.ncbi.nlm.nih.gov/PMC8315763/)
562. **Cov-Lineages** <https://cov-lineages.org/>
563. **Global disparities in SARS-CoV-2 genomic surveillance**
Anderson F Brito, Elizaveta Semenova, Gytis Dudas, Gabriel W Hassler, Chaney C Kalinich, Moritz UG Kraemer, Joses Ho, Houriiyah Tegally, George Githinji, Charles N Agoti, ...
Cold Spring Harbor Laboratory (2021-08-26) <https://doi.org/gn2th>
DOI: [10.1101/2021.08.21.21262393](https://doi.org/10.1101/2021.08.21.21262393) · PMID: [34462754](https://pubmed.ncbi.nlm.nih.gov/34462754/) · PMCID: [PMC8404891](https://pubmed.ncbi.nlm.nih.gov/PMC8404891/)
564. **Evaluation of COVID-19 RT-qPCR Test in Multi sample Pools**
Idan Yelin, Noga Aharony, Einat Shaer Tamar, Amir Argoetti, Esther Messer, Dina Berenbaum, Einat Shafran, Areen Kuzli, Nagham Gandali, Omer Shkedi, ... Roy Kishony
Clinical Infectious Diseases (2020-05-02) <https://doi.org/ggtx9r>
DOI: [10.1093/cid/ciaa531](https://doi.org/10.1093/cid/ciaa531) · PMID: [32358960](https://pubmed.ncbi.nlm.nih.gov/32358960/) · PMCID: [PMC7197588](https://pubmed.ncbi.nlm.nih.gov/PMC7197588/)
565. **Analytical Validation of a COVID-19 qRT-PCR Detection Assay Using a 384-well Format and Three Extraction Methods**
Andrew C Nelson, Benjamin Auch, Matthew Schomaker, Daryl M Gohl, Patrick Grady, Darrell Johnson, Robyn Kincaid, Kylene E Karnuth, Jerry Daniel, Jessica K Fiege, ... Sophia Yohe
Cold Spring Harbor Laboratory (2020-04-05) <https://doi.org/ggs45d>
DOI: [10.1101/2020.04.02.2022186](https://doi.org/10.1101/2020.04.02.2022186)
566. **Two-Stage Adaptive Pooling with RT-QPCR for Covid-19 Screening**
Anoosheh Heidarzadeh, Krishna Narayanan
ICASSP 2021 - 2021 IEEE International Conference on Acoustics, Speech and Signal Processing (ICASSP) (2021-06-06) <https://doi.org/gppbs9>
DOI: [10.1109/icassp39728.2021.9413685](https://doi.org/10.1109/icassp39728.2021.9413685)
567. **Pooled RT-qPCR testing for SARS-CoV-2 surveillance in schools - a cluster randomised trial**
Alexander Joachim, Felix Dewald, Isabelle Suárez, Michael Zemlin, Isabelle Lang, Regine Stutz, Anna Marthaler, Hans Martin Bosse, Nadine Lübke, Juliane Münch, ... Anna Kern
EClinicalMedicine (2021-09) <https://doi.org/gpzfjz>
DOI: [10.1016/j.eclinm.2021.101082](https://doi.org/10.1016/j.eclinm.2021.101082) · PMID: [34458708](https://pubmed.ncbi.nlm.nih.gov/34458708/) · PMCID: [PMC8384501](https://pubmed.ncbi.nlm.nih.gov/PMC8384501/)

568. **Pooling RT-qPCR testing for SARS-CoV-2 in 1000 individuals of healthy and infection-suspected patients**

Yosuke Hirotsu, Makoto Maejima, Masahiro Shibusawa, Yuki Nagakubo, Kazuhiro Hosaka, Kenji Amemiya, Hitomi Sueki, Miyoko Hayakawa, Hitoshi Mochizuki, Toshiharu Tsutsui, ... Masao Omata
Scientific Reports (2020-11-03) <https://doi.org/gpzfj4>

DOI: [10.1038/s41598-020-76043-z](https://doi.org/10.1038/s41598-020-76043-z) · PMID: [33144632](#) · PMCID: [PMC7641135](#)

569. **Loop-mediated isothermal amplification of DNA**

T Notomi

Nucleic Acids Research (2000-06-15) <https://doi.org/bx567n>

DOI: [10.1093/nar/28.12.e63](https://doi.org/10.1093/nar/28.12.e63) · PMID: [10871386](#) · PMCID: [PMC102748](#)

570. **A molecular test based on RT-LAMP for rapid, sensitive and inexpensive colorimetric detection of SARS-CoV-2 in clinical samples**

Catarina Amaral, Wilson Antunes, Elin Moe, Américo G Duarte, Luís MP Lima, Cristiana Santos, Inês L Gomes, Gonçalo S Afonso, Ricardo Vieira, Helena Sofia S Teles, ... Catarina Pimentel

Scientific Reports (2021-08-12) <https://doi.org/gnx4h3>

DOI: [10.1038/s41598-021-95799-6](https://doi.org/10.1038/s41598-021-95799-6) · PMID: [34385527](#) · PMCID: [PMC8361189](#)

571. **Can the cycle threshold (Ct) value of RT-PCR test for SARS CoV2 predict infectivity among close contacts?**

Soha Al Bayat, Jesha Mundodan, Samina Hasnain, Mohamed Sallam, Hayat Khogali, Dina Ali, Saif Alateeg, Mohamed Osama, Aiman Elberdiny, Hamad Al-Romaihi, Mohammed Hamad J Al-Thani

Journal of Infection and Public Health (2021-09) <https://doi.org/gnx4jw>

DOI: [10.1016/j.jiph.2021.08.013](https://doi.org/10.1016/j.jiph.2021.08.013) · PMID: [34416598](#) · PMCID: [PMC8362640](#)

572. **Evaluation of cycle threshold values at deisolation**

Clayton T Mowrer, Hannah Creager, Kelly Cawcutt, Justin Birge, Elizabeth Lyden, Trevor C Van Schooneveld, Mark E Rupp, Angela Hewlett

Infection Control & Hospital Epidemiology (2021-04-06)

<https://doi.org/gnx4jx>

DOI: [10.1017/ice.2021.132](https://doi.org/10.1017/ice.2021.132) · PMID: [33820588](#) · PMCID: [PMC8060537](#)

573. **To Interpret the SARS-CoV-2 Test, Consider the Cycle Threshold Value**

Michael R Tom, Michael J Mina

Clinical Infectious Diseases (2020-05-21) <https://doi.org/ggxf2s>

DOI: [10.1093/cid/ciaa619](https://doi.org/10.1093/cid/ciaa619) · PMID: [32435816](#) · PMCID: [PMC7314112](#)

574. **Rapid Molecular Detection of SARS-CoV-2 (COVID-19) Virus RNA Using Colorimetric LAMP**

Yinhua Zhang, Nelson Odiwuor, Jin Xiong, Luo Sun, Raphael Ohuru Nyaruaba, Hongping Wei, Nathan A Tanner

Cold Spring Harbor Laboratory (2020-02-29) <https://doi.org/ggx3wn>

DOI: [10.1101/2020.02.26.20028373](https://doi.org/10.1101/2020.02.26.20028373)

575. **Reverse Transcriptase Loop Mediated Isothermal Amplification (RT-LAMP) for COVID-19 diagnosis: a systematic review and meta-analysis.**
Anita Dominique Subali, Lowilius Wiyono
Pathogens and global health (2021-06-04)
<https://www.ncbi.nlm.nih.gov/pubmed/34086539>
DOI: [10.1080/20477724.2021.1933335](https://doi.org/10.1080/20477724.2021.1933335) · PMID: [34086539](https://pubmed.ncbi.nlm.nih.gov/34086539/) · PMCID: [PMC8182821](https://pubmed.ncbi.nlm.nih.gov/PMC8182821/)
576. **The CRISPR tool kit for genome editing and beyond**
Mazhar Adli
Nature Communications (2018-05-15) <https://doi.org/gdj266>
DOI: [10.1038/s41467-018-04252-2](https://doi.org/10.1038/s41467-018-04252-2) · PMID: [29765029](https://pubmed.ncbi.nlm.nih.gov/29765029/) · PMCID: [PMC5953931](https://pubmed.ncbi.nlm.nih.gov/PMC5953931/)
577. **Nucleic acid detection with CRISPR-Cas13a/C2c2**
Jonathan S Gootenberg, Omar O Abudayyeh, Jeong Wook Lee, Patrick Essletzbichler, Aaron J Dy, Julia Joung, Vanessa Verdine, Nina Donghia, Nichole M Daringer, Catherine A Freije, ... Feng Zhang
Science (2017-04-28) <https://doi.org/f93x8p>
DOI: [10.1126/science.aam9321](https://doi.org/10.1126/science.aam9321) · PMID: [28408723](https://pubmed.ncbi.nlm.nih.gov/28408723/) · PMCID: [PMC5526198](https://pubmed.ncbi.nlm.nih.gov/PMC5526198/)
578. **Development and evaluation of a rapid CRISPR-based diagnostic for COVID-19**
Tieying Hou, Weiqi Zeng, Minling Yang, Wenjing Chen, Lili Ren, Jingwen Ai, Ji Wu, Yalong Liao, Xuejing Gou, Yongjun Li, ... Teng Xu
PLOS Pathogens (2020-08-27) <https://doi.org/ghn7rp>
DOI: [10.1371/journal.ppat.1008705](https://doi.org/10.1371/journal.ppat.1008705) · PMID: [32853291](https://pubmed.ncbi.nlm.nih.gov/32853291/) · PMCID: [PMC7451577](https://pubmed.ncbi.nlm.nih.gov/PMC7451577/)
579. **CRISPR-based surveillance for COVID-19 using genomically-comprehensive machine learning design**
Hayden C Metsky, Catherine A Freije, Tinna-Solveig F Kosoko-Thoroddsen, Pardis C Sabeti, Cameron Myhrvold
Cold Spring Harbor Laboratory (2020-03-02) <https://doi.org/ggr3zf>
DOI: [10.1101/2020.02.26.967026](https://doi.org/10.1101/2020.02.26.967026)
580. **A Scalable, Easy-to-Deploy Protocol for Cas13-Based Detection of SARS-CoV-2 Genetic Material**
Jennifer N Rauch, Eric Valois, Sabrina C Solley, Friederike Braig, Ryan S Lach, Morgane Audouard, Jose Carlos Ponce-Rojas, Michael S Costello, Naomi J Baxter, Kenneth S Kosik, ... Maxwell Z Wilson
Journal of Clinical Microbiology (2021-03-19) <https://doi.org/gh3nm3>
DOI: [10.1128/jcm.02402-20](https://doi.org/10.1128/jcm.02402-20) · PMID: [33478979](https://pubmed.ncbi.nlm.nih.gov/33478979/) · PMCID: [PMC8092748](https://pubmed.ncbi.nlm.nih.gov/PMC8092748/)
581. **CRISPR-Cas12-based detection of SARS-CoV-2**
James P Broughton, Xianding Deng, Guixia Yu, Clare L Fasching, Venice Servellita, Jasmeet Singh, Xin Miao, Jessica A Streithorst, Andrea Granados, Alicia Sotomayor-Gonzalez, ... Charles Y Chiu
Nature Biotechnology (2020-04-16) <https://doi.org/ggv47f>
DOI: [10.1038/s41587-020-0513-4](https://doi.org/10.1038/s41587-020-0513-4) · PMID: [32300245](https://pubmed.ncbi.nlm.nih.gov/32300245/)
582. **An ultrasensitive, rapid, and portable coronavirus SARS-CoV-2 sequence detection method based on CRISPR-Cas12**

Curti Lucia, Pereyra-Bonnet Federico, Gimenez Carla Alejandra
Cold Spring Harbor Laboratory (2020-03-02) <https://doi.org/gg7km6>
DOI: [10.1101/2020.02.29.971127](https://doi.org/10.1101/2020.02.29.971127)

583. **Rapid detection of SARS-CoV-2 with CRISPR-Cas12a**
Dan Xiong, Wenjun Dai, Jiaoqiao Gong, Guande Li, Nansong Liu, Wei Wu, Jiaqiang Pan, Chen Chen, Yingzhen Jiao, Huina Deng, ... Guanghui Tang
PLOS Biology (2020-12-15) <https://doi.org/gh3nm6>
DOI: [10.1371/journal.pbio.3000978](https://doi.org/10.1371/journal.pbio.3000978) · PMID: [33320883](#) · PMCID: [PMC7737895](#)
584. **Ultrasensitive and visual detection of SARS-CoV-2 using all-in-one dual CRISPR-Cas12a assay**
Xiong Ding, Kun Yin, Ziyue Li, Rajesh V Lalla, Enrique Ballesteros, Maroun M Sfeir, Changchun Liu
Nature Communications (2020-09-18) <https://doi.org/ghwjb2>
DOI: [10.1038/s41467-020-18575-6](https://doi.org/10.1038/s41467-020-18575-6) · PMID: [32948757](#) · PMCID: [PMC7501862](#)
585. **SARS-CoV-2 detection with CRISPR diagnostics**
Lu Guo, Xuehan Sun, Xinge Wang, Chen Liang, Haiping Jiang, Qingqin Gao, Moyu Dai, Bin Qu, Sen Fang, Yihuan Mao, ... Wei Li
Cell Discovery (2020-05-19) <https://doi.org/ggx2wt>
DOI: [10.1038/s41421-020-0174-y](https://doi.org/10.1038/s41421-020-0174-y) · PMID: [32435508](#) · PMCID: [PMC7235268](#)
586. **Electric field-driven microfluidics for rapid CRISPR-based diagnostics and its application to detection of SARS-CoV-2**
Ashwin Ramachandran, Diego A Huyke, Eesha Sharma, Malaya K Sahoo, ChunHong Huang, Niaz Banaei, Benjamin A Pinsky, Juan G Santiago
Proceedings of the National Academy of Sciences (2020-11-04)
<https://doi.org/gh3nms>
DOI: [10.1073/pnas.2010254117](https://doi.org/10.1073/pnas.2010254117) · PMID: [33148808](#) · PMCID: [PMC7703567](#)
587. **Annotation and Classification of CRISPR-Cas Systems**
Kira S Makarova, Eugene V Koonin
Methods in Molecular Biology (2015) <https://doi.org/gppg52>
DOI: [10.1007/978-1-4939-2687-9_4](https://doi.org/10.1007/978-1-4939-2687-9_4) · PMID: [25981466](#) · PMCID: [PMC5901762](#)
588. **Type III CRISPR-Cas systems: when DNA cleavage just isn't enough**
Nora C Pyenson, Luciano A Marraffini
Current Opinion in Microbiology (2017-06) <https://doi.org/gcx7r3>
DOI: [10.1016/j.mib.2017.08.003](https://doi.org/10.1016/j.mib.2017.08.003) · PMID: [28865392](#)
589. **Type III CRISPR-Cas Immunity: Major Differences Brushed Aside**
Gintautas Tamulaitis, Česlovas Venclovas, Virginijus Siksnys
Trends in Microbiology (2017-01) <https://doi.org/f9nj96>
DOI: [10.1016/j.tim.2016.09.012](https://doi.org/10.1016/j.tim.2016.09.012) · PMID: [27773522](#)
590. **Intrinsic signal amplification by type III CRISPR-Cas systems provides a sequence-specific SARS-CoV-2 diagnostic**

Andrew Santiago-Frangos, Laina N Hall, Anna Nemudraia, Artem Nemudryi, Pushya Krishna, Tanner Wiegand, Royce A Wilkinson, Deann T Snyder, Jodi F Hedges, Calvin Cicha, ... Blake Wiedenheft
Cell Reports Medicine (2021-06) <https://doi.org/gpmv6n>
DOI: [10.1016/j.xcrm.2021.100319](https://doi.org/10.1016/j.xcrm.2021.100319) · PMID: [34075364](https://pubmed.ncbi.nlm.nih.gov/34075364/) · PMCID: [PMC8157118](https://pubmed.ncbi.nlm.nih.gov/PMC8157118/)

591. **Amplification-free detection of SARS-CoV-2 with CRISPR-Cas13a and mobile phone microscopy**
Parinaz Fozouni, Sungmin Son, María Díaz de León Derby, Gavin J Knott, Carley N Gray, Michael V D'Ambrosio, Chunyu Zhao, Neil A Switz, GRenuka Kumar, Stephanie I Stephens, ... Melanie Ott
Cell (2021-01) <https://doi.org/ghnszx>
DOI: [10.1016/j.cell.2020.12.001](https://doi.org/10.1016/j.cell.2020.12.001) · PMID: [33306959](https://pubmed.ncbi.nlm.nih.gov/33306959/) · PMCID: [PMC7834310](https://pubmed.ncbi.nlm.nih.gov/PMC7834310/)
592. **Rapid, Sensitive, and Specific Severe Acute Respiratory Syndrome Coronavirus 2 Detection: A Multicenter Comparison Between Standard Quantitative Reverse-Transcriptase Polymerase Chain Reaction and CRISPR-Based DETECTR**
Eelke Brandsma, Han JMP Verhagen, Thijs JW van de Laar, Eric CJ Claas, Marion Cornelissen, Emile van den Akker
The Journal of Infectious Diseases (2020-10-10) <https://doi.org/gh3nmt>
DOI: [10.1093/infdis/jiaa641](https://doi.org/10.1093/infdis/jiaa641) · PMID: [33535237](https://pubmed.ncbi.nlm.nih.gov/33535237/) · PMCID: [PMC7665660](https://pubmed.ncbi.nlm.nih.gov/PMC7665660/)
593. **Rapid, accurate, nucleobase detection using FnCas9**
Mohd Azhar, Rhythm Phutela, Manoj Kumar, Asgar Hussain Ansari, Riya Rauthan, Sneha Gulati, Namrata Sharma, Dipanjali Sinha, Saumya Sharma, Sunaina Singh, ...
Cold Spring Harbor Laboratory (2020-09-14) <https://doi.org/gh3nmv>
DOI: [10.1101/2020.09.13.20193581](https://doi.org/10.1101/2020.09.13.20193581)
594. **Rapid Diagnostic Testing for SARS-CoV-2**
Paul K Drain
New England Journal of Medicine (2022-01-20) <https://doi.org/gn2sfk>
DOI: [10.1056/nejmcp2117115](https://doi.org/10.1056/nejmcp2117115) · PMID: [34995029](https://pubmed.ncbi.nlm.nih.gov/34995029/) · PMCID: [PMC8820190](https://pubmed.ncbi.nlm.nih.gov/PMC8820190/)
595. **The Performance of Two Rapid Antigen Tests During Population-Level Screening for SARS-CoV-2 Infection**
Mohammad Alghounaim, Hamad Bastaki, Farah Bin Essa, Hoda Motlagh, Salman Al-Sabah
Frontiers in Medicine (2021-12-23) <https://doi.org/gpp4xw>
DOI: [10.3389/fmed.2021.797109](https://doi.org/10.3389/fmed.2021.797109) · PMID: [35004772](https://pubmed.ncbi.nlm.nih.gov/35004772/) · PMCID: [PMC8733308](https://pubmed.ncbi.nlm.nih.gov/PMC8733308/)
596. **Lateral flow (immuno)assay: its strengths, weaknesses, opportunities and threats. A literature survey**
Geertruida A Posthuma-Trumpie, Jakob Korf, Aart van Amerongen
Analytical and Bioanalytical Chemistry (2008-08-13)
<https://doi.org/bcsjdw>
DOI: [10.1007/s00216-008-2287-2](https://doi.org/10.1007/s00216-008-2287-2) · PMID: [18696055](https://pubmed.ncbi.nlm.nih.gov/18696055/)
597. **A systematic review of the sensitivity and specificity of lateral flow devices in the detection of SARS-CoV-2.**

Dylan A Mistry, Jenny Y Wang, Mika-Erik Moeser, Thomas Starkey,
Lennard YW Lee
BMC infectious diseases (2021-08-18)
<https://www.ncbi.nlm.nih.gov/pubmed/34407759>
DOI: [10.1186/s12879-021-06528-3](https://doi.org/10.1186/s12879-021-06528-3) · PMID: [34407759](https://pubmed.ncbi.nlm.nih.gov/34407759/) · PMCID:
[PMC8371300](https://pubmed.ncbi.nlm.nih.gov/PMC8371300/)

598. **Government sets out next steps for living with COVID**
GOV.UK
<https://www.gov.uk/government/news/government-sets-out-next-steps-for-living-with-covid>
599. **Fact Sheet: The Biden Administration to Begin Distributing At-Home, Rapid COVID-19 Tests to Americans for Free**
The White House
(2022-01-14) <https://www.whitehouse.gov/briefing-room/statements-releases/2022/01/14/fact-sheet-the-biden-administration-to-begin-distributing-at-home-rapid-covid-19-tests-to-americans-for-free/>
600. **Scaling up COVID-19 rapid antigen tests: promises and challenges**
Rosanna W Peeling, Piero L Olliaro, Debrah I Boeras, Noah Fongwen
The Lancet Infectious Diseases (2021-09) <https://doi.org/hk34>
DOI: [10.1016/s1473-3099\(21\)00048-7](https://doi.org/10.1016/s1473-3099(21)00048-7) · PMID: [33636148](https://pubmed.ncbi.nlm.nih.gov/33636148/) · PMCID:
[PMC7906660](https://pubmed.ncbi.nlm.nih.gov/PMC7906660/)
601. **Detection technologies and recent developments in the diagnosis of COVID-19 infection**
Praveen Rai, Ballamoole Krishna Kumar, Vijaya Kumar Deekshit,
Indrani Karunasagar, Iddya Karunasagar
Applied Microbiology and Biotechnology (2021-01)
<https://doi.org/gnntp9>
DOI: [10.1007/s00253-020-11061-5](https://doi.org/10.1007/s00253-020-11061-5) · PMID: [33394144](https://pubmed.ncbi.nlm.nih.gov/33394144/) · PMCID:
[PMC7780074](https://pubmed.ncbi.nlm.nih.gov/PMC7780074/)
602. **Enzyme Immunoassay (EIA)/Enzyme-Linked Immunosorbent Assay (ELISA)**
Rudolf M Lequin
Clinical Chemistry (2005-12-01) <https://doi.org/dts5xp>
DOI: [10.1373/clinchem.2005.051532](https://doi.org/10.1373/clinchem.2005.051532) · PMID: [16179424](https://pubmed.ncbi.nlm.nih.gov/16179424/)
603. **Enzyme-Immunoassay: A Powerful Analytical Tool**
AHWM Schuurs, BK Van Weemen
Journal of Immunoassay (1980-01) <https://doi.org/btqbvh>
DOI: [10.1080/01971528008055786](https://doi.org/10.1080/01971528008055786) · PMID: [6785317](https://pubmed.ncbi.nlm.nih.gov/6785317/)
604. **Antibody testing for COVID-19: A report from the National COVID Scientific Advisory Panel**
Emily R Adams, Mark Ainsworth, Rekha Anand, Monique I Andersson,
Kathryn Auckland, JKenneth Baillie, Eleanor Barnes, Sally Beer, John I
Bell, Tamsin Berry, ...
Wellcome Open Research (2020-06-11) <https://doi.org/gpp4wq>
DOI: [10.12688/wellcomeopenres.15927.1](https://doi.org/10.12688/wellcomeopenres.15927.1) · PMID: [33748431](https://pubmed.ncbi.nlm.nih.gov/33748431/) · PMCID:
[PMC7941096](https://pubmed.ncbi.nlm.nih.gov/PMC7941096/)

605. **Detection of SARS-CoV-2 by antigen ELISA test is highly swayed by viral load and sample storage condition**

Nihad Adnan, Shahad Saif Khandker, Ahsanul Haq, Mousumi Akter Chaity, Abdul Khalek, Anawarul Quader Nazim, Taku Kaitsuka, Kazuhito Tomizawa, Masayasu Mie, Eiry Kobatake, ... MohdRaeed Jamiruddin

Expert Review of Anti-infective Therapy (2021-09-11)

<https://doi.org/gprjx9>

DOI: [10.1080/14787210.2021.1976144](https://doi.org/10.1080/14787210.2021.1976144) · PMID: [34477019](https://pubmed.ncbi.nlm.nih.gov/34477019/) · PMCID:

[PMC8442762](https://pubmed.ncbi.nlm.nih.gov/PMC8442762/)

606. **Negative Nasopharyngeal and Oropharyngeal Swabs Do Not Rule Out COVID-19**

Poramed Winichakoon, Romanee Chaiwarith, Chalerm Liwsrisakun, Parichat Salee, Aree Goonna, Atikun Limsukon, Quanhathai Kaewpoowat

Journal of Clinical Microbiology (2020-04-23) <https://doi.org/ggpw9m>

DOI: [10.1128/jcm.00297-20](https://doi.org/10.1128/jcm.00297-20) · PMID: [32102856](https://pubmed.ncbi.nlm.nih.gov/32102856/) · PMCID: [PMC7180262](https://pubmed.ncbi.nlm.nih.gov/PMC7180262/)

607. **Coronavirus and the race to distribute reliable diagnostics**

Cormac Sheridan

Nature Biotechnology (2020-02-19) <https://doi.org/ggm4nt>

DOI: [10.1038/d41587-020-00002-2](https://doi.org/10.1038/d41587-020-00002-2) · PMID: [32265548](https://pubmed.ncbi.nlm.nih.gov/32265548/)

608. **Quarantine & Isolation**

CDC

Centers for Disease Control and Prevention (2022-03-30)

<https://www.cdc.gov/coronavirus/2019-ncov/your-health/quarantine-isolation.html>

609. **CRISPR-Cas System: An Approach With Potentials for COVID-19 Diagnosis and Therapeutics**

Prashant Kumar, Yashpal Singh Malik, Balasubramanian Ganesh, Somnath Rahangdale, Sharad Saurabh, Senthilkumar Natesan, Ashish Srivastava, Khan Sharun, MohdIqbal Yatoo, Ruchi Tiwari, ... Kuldeep Dhamo

Frontiers in Cellular and Infection Microbiology (2020-11-02)

<https://doi.org/ghz57p>

DOI: [10.3389/fcimb.2020.576875](https://doi.org/10.3389/fcimb.2020.576875) · PMID: [33251158](https://pubmed.ncbi.nlm.nih.gov/33251158/) · PMCID:

[PMC7673385](https://pubmed.ncbi.nlm.nih.gov/PMC7673385/)

610. **The standard coronavirus test, if available, works well—but can new diagnostics help in this pandemic?**

Robert Service

Science (2020-03-22) <https://doi.org/ggq9wm>

DOI: [10.1126/science.abb8400](https://doi.org/10.1126/science.abb8400)

611. **Laboratory Diagnosis of COVID-19: Current Issues and Challenges**

Yi-Wei Tang, Jonathan E Schmitz, David H Persing, Charles W Stratton

Journal of Clinical Microbiology (2020-05-26) <https://doi.org/ggq7h8>

DOI: [10.1128/jcm.00512-20](https://doi.org/10.1128/jcm.00512-20) · PMID: [32245835](https://pubmed.ncbi.nlm.nih.gov/32245835/)

612. **The Plane Is Boarding, Where Are Your Test Results?**

Lauren Sloss

The New York Times (2021-12-31)

<https://www.nytimes.com/2021/12/31/travel/covid-test-chaos.html>

613. **A Stark Contrast Between the U.S. and Europe on Tests**
Naomi Kresge
Bloomberg (2022-01-07)
<https://www.bloomberg.com/news/newsletters/2022-01-07/a-stark-contrast-between-the-u-s-and-europe-on-tests>
614. **IgG Subclasses and Allotypes: From Structure to Effector Functions**
Gestur Vidarsson, Gillian Dekkers, Theo Rispens
Frontiers in Immunology (2014-10-20) <https://doi.org/gc6vx6>
DOI: [10.3389/fimmu.2014.00520](https://doi.org/10.3389/fimmu.2014.00520) · PMID: [25368619](#) · PMCID: [PMC4202688](#)
615. **The distribution and functions of immunoglobulin isotypes**
Charles A Janeway Jr, Paul Travers, Mark Walport, Mark J Shlomchik (editors)
Immunobiology: The Immune System in Health and Disease (2001)
<https://www.ncbi.nlm.nih.gov/books/NBK27162>
ISBN: 978-0815336426
616. **Longitudinal profile of antibodies against SARS-coronavirus in SARS patients and their clinical significance**
Hongying MO, Guangqiao ZENG, Xiaolan REN, Hui LI, Changwen KE, Yaxia TAN, Chaoda CAI, Kefang LAI, Rongchang CHEN, Moira CHAN-YEUNG, Nanshan ZHONG
Respirology (2006-01) <https://doi.org/dn23vj>
DOI: [10.1111/j.1440-1843.2006.00783.x](https://doi.org/10.1111/j.1440-1843.2006.00783.x) · PMID: [16423201](#) · PMCID: [PMC7192223](#)
617. **Detection of antibodies against SARS-CoV-2 in patients with COVID-19**
Zhe Du, Fengxue Zhu, Fuzheng Guo, Bo Yang, Tianbing Wang
Journal of Medical Virology (2020-04-10) <https://doi.org/gqq7m2>
DOI: [10.1002/jmv.25820](https://doi.org/10.1002/jmv.25820) · PMID: [32243608](#)
618. **Review of Current Advances in Serologic Testing for COVID-19**
Andrea P Espejo, Yamac Akgun, Abdulaziz F Al Mana, Youley Tjendra, Nicolas C Millan, Carmen Gomez-Fernandez, Carolyn Cray
American Journal of Clinical Pathology (2020-06-25)
<https://doi.org/gkzfrn>
DOI: [10.1093/ajcp/aqaa112](https://doi.org/10.1093/ajcp/aqaa112) · PMID: [32583852](#) · PMCID: [PMC7337672](#)
619. **A serological assay to detect SARS-CoV-2 seroconversion in humans**
Fatima Amanat, Daniel Stadlbauer, Shirin Strohmeier, Thi HO Nguyen, Veronika Chromikova, Meagan McMahon, Kaijun Jiang, Guha Asthagiri Arunkumar, Denise Jurczyszak, Jose Polanco, ... Florian Krammer
Nature Medicine (2020-05-12) <https://doi.org/ggx28b>
DOI: [10.1038/s41591-020-0913-5](https://doi.org/10.1038/s41591-020-0913-5) · PMID: [32398876](#) · PMCID: [PMC8183627](#)
620. **Development of a Fast SARS-CoV-2 IgG ELISA, Based on Receptor-Binding Domain, and Its Comparative Evaluation Using Temporally Segregated Samples From RT-PCR Positive Individuals**

Farha Mehdi, Souvick Chattopadhyay, Ramachandran Thiruvengadam, Sarla Yadav, Manjit Kumar, Sangita Kumari Sinha, Sandeep Goswami, Pallavi Kshetrapal, Nitya Wadhwa, Uma Chandramouli Natchu, ...
Gaurav Batra
Frontiers in Microbiology (2021-01-20) <https://doi.org/gpp5md>
DOI: [10.3389/fmicb.2020.618097](https://doi.org/10.3389/fmicb.2020.618097) · PMID: [33552028](https://pubmed.ncbi.nlm.nih.gov/33552028/) · PMCID: [PMC7854536](https://pubmed.ncbi.nlm.nih.gov/PMC7854536/)

621. **Development and evaluation of a low cost IgG ELISA test based in RBD protein for COVID-19**

Luciana Villafaña, Lucía Gallo Vaulet, Florencia M Viere, Laura I Klepp, Marina A Forrellad, María M Bigi, María I Romano, Giovanni Magistrelli, Marcelo Rodríguez Fermepin, Fabiana Bigi
Journal of Immunological Methods (2022-01) <https://doi.org/gpp4wh>
DOI: [10.1016/j.jim.2021.113182](https://doi.org/10.1016/j.jim.2021.113182) · PMID: [34762914](https://pubmed.ncbi.nlm.nih.gov/34762914/) · PMCID: [PMC8574101](https://pubmed.ncbi.nlm.nih.gov/PMC8574101/)

622. **A sensitive and rapid chemiluminescence immunoassay for point-of-care testing (POCT) of copeptin in serum based on high-affinity monoclonal antibodies via cytokine-assisted immunization**

Yu Wang, Emmanuel Enoch Dzakah, Ye Kang, Yanxue Cai, Peidian Wu, Bo Tang, Run Li, Xiaowei He
International Journal of Nanomedicine (2019-06)
<https://doi.org/gpp5mc>
DOI: [10.2147/ijn.s200556](https://doi.org/10.2147/ijn.s200556) · PMID: [31354261](https://pubmed.ncbi.nlm.nih.gov/31354261/) · PMCID: [PMC6580123](https://pubmed.ncbi.nlm.nih.gov/PMC6580123/)

623. **Chemiluminescent immunoassay technology: what does it change in autoantibody detection?**

Luigi Cinquanta, Desré Ethel Fontana, Nicola Bizzaro
Autoimmunity Highlights (2017-06-24) <https://doi.org/gh6hcm>
DOI: [10.1007/s13317-017-0097-2](https://doi.org/10.1007/s13317-017-0097-2) · PMID: [28647912](https://pubmed.ncbi.nlm.nih.gov/28647912/) · PMCID: [PMC5483212](https://pubmed.ncbi.nlm.nih.gov/PMC5483212/)

624. **A Peptide-Based Magnetic Chemiluminescence Enzyme Immunoassay for Serological Diagnosis of Coronavirus Disease 2019**

Xue-fei Cai, Juan Chen, Jie-li Hu, Quan-xin Long, Hai-jun Deng, Ping Liu, Kai Fan, Pu Liao, Bei-zhong Liu, Gui-cheng Wu, ... De-qiang Wang
The Journal of Infectious Diseases (2020-07-15) <https://doi.org/ggv2fx>
DOI: [10.1093/infdis/jiaa243](https://doi.org/10.1093/infdis/jiaa243) · PMID: [32382737](https://pubmed.ncbi.nlm.nih.gov/32382737/) · PMCID: [PMC7239108](https://pubmed.ncbi.nlm.nih.gov/PMC7239108/)

625. **Comparison of SARS-CoV-2 serological tests with different antigen targets**

Alix T Coste, Katia Jaton, Matthaios Papadimitriou-Olivgeris, Gilbert Greub, Antony Croxatto
Journal of Clinical Virology (2021-01) <https://doi.org/gk8s5q>
DOI: [10.1016/j.jcv.2020.104690](https://doi.org/10.1016/j.jcv.2020.104690) · PMID: [33253926](https://pubmed.ncbi.nlm.nih.gov/33253926/) · PMCID: [PMC7670982](https://pubmed.ncbi.nlm.nih.gov/PMC7670982/)

626. **Assessment of SARS-CoV-2 serological tests for the diagnosis of COVID-19 through the evaluation of three immunoassays: Two automated immunoassays (Euroimmun and Abbott) and one rapid lateral flow immunoassay (NG Biotech)**

Thomas Nicol, Caroline Lefevre, Orianne Serri, Adeline Pivert, Françoise Joubaud, Vincent Dubée, Achille Kouatchet, Alexandra

Ducancelle, Françoise Lunel-Fabiani, Hélène Le Guillou-Guillemette
Journal of Clinical Virology (2020-08) <https://doi.org/gg2ks6>
DOI: [10.1016/j.jcv.2020.104511](https://doi.org/10.1016/j.jcv.2020.104511) · PMID: [32593133](#) · PMCID: [PMC7295485](#)

627. **Evaluation of Six Commercial Mid- to High-Volume Antibody and Six Point-of-Care Lateral Flow Assays for Detection of SARS-CoV-2 Antibodies**

Carmen L Charlton, Jamil N Kanji, Kam Johal, Ashley Bailey, Sabrina S Plitt, Clayton MacDonald, Andrea Kunst, Emily Buss, Laura E Burnes, Kevin Fonseca, ... Graham Tipples
Journal of Clinical Microbiology (2020-09-22) <https://doi.org/gpp5mb>
DOI: [10.1128/jcm.01361-20](https://doi.org/10.1128/jcm.01361-20) · PMID: [32665420](#) · PMCID: [PMC7512179](#)

628. **Diagnostic accuracy of an automated chemiluminescent immunoassay for anti-SARS-CoV-2 IgM and IgG antibodies: an Italian experience**

Maria Infantino, Valentina Grossi, Barbara Lari, Riccardo Bambi, Alessandro Perri, Matteo Menneschi, Giovanni Terenzi, Irene Liotti, Giovanni Ciotta, Cristina Taddei, ... Mariangela Manfredi
Journal of Medical Virology (2020-05-10) <https://doi.org/ggv4c6>
DOI: [10.1002/jmv.25932](https://doi.org/10.1002/jmv.25932) · PMID: [32330291](#) · PMCID: [PMC7264663](#)

629. **SARS-CoV-2 serology: Validation of high-throughput chemiluminescent immunoassay (CLIA) platforms and a field study in British Columbia**

Inna Sekirov, Vilte E Barakauskas, Janet Simons, Darrel Cook, Brandon Bates, Laura Burns, Shazia Masud, Marthe Charles, Meghan McLennan, Annie Mak, ... Muhammad Morshed
Journal of Clinical Virology (2021-09) <https://doi.org/gpp5kb>
DOI: [10.1016/j.jcv.2021.104914](https://doi.org/10.1016/j.jcv.2021.104914) · PMID: [34304088](#) · PMCID: [PMC8282439](#)

630. **Cellex qSARS-CoV-2 IgG/IgM Rapid Test**

Cellex
(2020-04-07) <https://www.fda.gov/media/136625/download>

631. **Evaluation of Humoral Immune Response after SARS-CoV-2 Vaccination Using Two Binding Antibody Assays and a Neutralizing Antibody Assay**

Minjeong Nam, Jong Do Seo, Hee-Won Moon, Hanah Kim, Mina Hur, Yeo-Min Yun
Microbiology Spectrum (2021-12-22) <https://doi.org/gnkz5s>
DOI: [10.1128/spectrum.01202-21](https://doi.org/10.1128/spectrum.01202-21) · PMID: [34817223](#) · PMCID: [PMC8612149](#)

632. **A high-throughput neutralizing antibody assay for COVID-19 diagnosis and vaccine evaluation**

Antonio E Muruato, Camila R Fontes-Garfias, Ping Ren, Mariano A Garcia-Blanco, Vineet D Menachery, Xuping Xie, Pei-Yong Shi
Nature Communications (2020-08-13) <https://doi.org/gjkvr9>
DOI: [10.1038/s41467-020-17892-0](https://doi.org/10.1038/s41467-020-17892-0) · PMID: [32792628](#) · PMCID: [PMC7426916](#)

633. **Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection**
David S Khoury, Deborah Cromer, Arnold Reynaldi, Timothy E Schlub, Adam K Wheatley, Jennifer A Juno, Kanta Subbarao, Stephen J Kent, James A Triccas, Miles P Davenport
Nature Medicine (2021-05-17) <https://doi.org/gj3h47>
DOI: [10.1038/s41591-021-01377-8](https://doi.org/s41591-021-01377-8) · PMID: [34002089](#)
634. **Neutralizing Antibodies Correlate with Protection from SARS-CoV-2 in Humans during a Fishery Vessel Outbreak with a High Attack Rate**
Amin Addetia, Katharine HD Crawford, Adam Dingens, Haiying Zhu, Pavitra Roychoudhury, Meei-Li Huang, Keith R Jerome, Jesse D Bloom, Alexander L Greninger
Journal of Clinical Microbiology (2020-10-21) <https://doi.org/gk7n4d>
DOI: [10.1128/jcm.02107-20](https://doi.org/10.1128/jcm.02107-20) · PMID: [32826322](#) · PMCID: [PMC7587101](#)
635. **Neutralising antibody titres as predictors of protection against SARS-CoV-2 variants and the impact of boosting: a meta-analysis**
Deborah Cromer, Megan Steain, Arnold Reynaldi, Timothy E Schlub, Adam K Wheatley, Jennifer A Juno, Stephen J Kent, James A Triccas, David S Khoury, Miles P Davenport
The Lancet Microbe (2022-01) <https://doi.org/gnx76z>
DOI: [10.1016/s2666-5247\(21\)00267-6](https://doi.org/10.1016/s2666-5247(21)00267-6) · PMID: [34806056](#) · PMCID: [PMC8592563](#)
636. **A cell-free high throughput assay for assessment of SARS-CoV-2 neutralizing antibodies**
Sara Mravinacova, Malin Jönsson, Wanda Christ, Jonas Klingström, Jamil Yousef, Cecilia Hellström, My Hedhammar, Sebastian Havervall, Charlotte Thålin, Elisa Pin, ... Sophia Hober
New Biotechnology (2022-01) <https://doi.org/gm9pkb>
DOI: [10.1016/j.nbt.2021.10.002](https://doi.org/10.1016/j.nbt.2021.10.002) · PMID: [34628049](#) · PMCID: [PMC8495044](#)
637. **Two-Year Prospective Study of the Humoral Immune Response of Patients with Severe Acute Respiratory Syndrome**
Wei Liu, Arnaud Fontanet, Pan-He Zhang, Lin Zhan, Zhong-Tao Xin, Laurence Baril, Fang Tang, Hui Lv, Wu-Chun Cao
The Journal of Infectious Diseases (2006-03-15) <https://doi.org/cmzn2k>
DOI: [10.1086/500469](https://doi.org/10.1086/500469) · PMID: [16479513](#) · PMCID: [PMC7109932](#)
638. **The time course of the immune response to experimental coronavirus infection of man**
KA Callow, HF Parry, M Sergeant, DAJ Tyrrell
Epidemiology and Infection (2009-05-15) <https://doi.org/c9pnmg>
DOI: [10.1017/s0950268800048019](https://doi.org/10.1017/s0950268800048019) · PMID: [2170159](#) · PMCID: [PMC2271881](#)
639. **Antibody Responses 8 Months after Asymptomatic or Mild SARS-CoV-2 Infection**
Pyoeng Gyun Choe, Kye-Hyung Kim, Chang Kyung Kang, Hyeyon Jeong Suh, EunKyo Kang, Sun Young Lee, Nam Joong Kim, Jongyoun Yi, Wan Beom Park, Myoung-don Oh
Emerging Infectious Diseases (2021-03) <https://doi.org/ghs9kg>

640. **Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection**

Jennifer M Dan, Jose Mateus, Yu Kato, Kathryn M Hastie, Esther Dawen Yu, Caterina E Faliti, Alba Grifoni, Sydney I Ramirez, Sonya Haupt, April Frazier, ... Shane Crotty

Science (2021-01-06) <https://doi.org/ghrv9b>

DOI: [10.1126/science.abf4063](https://doi.org/10.1126/science.abf4063) · PMID: [33408181](#)

641. **Rapid generation of durable B cell memory to SARS-CoV-2 spike and nucleocapsid proteins in COVID-19 and convalescence.**

Gemma E Hartley, Emily SJ Edwards, Pei M Aui, Nirupama Varese, Stephanie Stojanovic, James McMahon, Anton Y Peleg, Irene Boo, Heidi E Drummer, PMark Hogarth, ... Menno C van Zelm

Science immunology (2020-12-22)

<https://www.ncbi.nlm.nih.gov/pubmed/33443036>

DOI: [10.1126/sciimmunol.abf8891](https://doi.org/10.1126/sciimmunol.abf8891) · PMID: [33443036](#)

642. **Persistence of SARS-CoV-2-specific B and T cell responses in convalescent COVID-19 patients 6–8 months after the infection**

Natalia Sherina, Antonio Piralla, Likun Du, Hui Wan, Makiko Kumagai-Braesch, Juni Andréll, Sten Braesch-Andersen, Irene Cassaniti, Elena Percivalle, Antonella Sarasini, ... Qiang Pan-Hammarström

Med (2021-03) <https://doi.org/gh3xkz>

DOI: [10.1016/j.medj.2021.02.001](https://doi.org/10.1016/j.medj.2021.02.001) · PMID: [33589885](#) · PMCID:

[PMC7874960](#)

643. **Evolution of antibody immunity to SARS-CoV-2**

Christian Gaebler, Zijun Wang, Julio CC Lorenzi, Frauke Muecksch, Shlomo Finkin, Minami Tokuyama, Alice Cho, Mila Jankovic, Dennis Schaefer-Babajew, Thiago Y Oliveira, ... Michel C Nussenzweig

Nature (2021-01-18) <https://doi.org/fq6k>

DOI: [10.1038/s41586-021-03207-w](https://doi.org/10.1038/s41586-021-03207-w) · PMID: [33461210](#) · PMCID:

[PMC8221082](#)

644. **Robust neutralizing antibodies to SARS-CoV-2 infection persist for months**

Ania Wajnberg, Fatima Amanat, Adolfo Firpo, Deena R Altman, Mark J Bailey, Mayce Mansour, Meagan McMahon, Philip Meade, Damodara Rao Mendu, Kimberly Muellers, ... Carlos Cordon-Cardo

Science (2020-12-04) <https://doi.org/fgf8>

DOI: [10.1126/science.abd7728](https://doi.org/10.1126/science.abd7728) · PMID: [33115920](#) · PMCID:

[PMC7810037](#)

645. **Longitudinal observation and decline of neutralizing antibody responses in the three months following SARS-CoV-2 infection in humans**

Jeffrey Seow, Carl Graham, Blair Merrick, Sam Acors, Suzanne Pickering, Kathryn JA Steel, Oliver Hemmings, Aoife O'Byrne, Neophytos Kouphou, Rui Pedro Galao, ... Katie J Doores

Nature Microbiology (2020-10-26) <https://doi.org/fh7j>

DOI: [10.1038/s41564-020-00813-8](https://doi.org/10.1038/s41564-020-00813-8) · PMID: [33106674](#) · PMCID:

[PMC7610833](#)

646. **Evolution of immune responses to SARS-CoV-2 in mild-moderate COVID-19**

Adam K Wheatley, Jennifer A Juno, Jing J Wang, Kevin J Selva, Arnold Reynaldi, Hyon-Xhi Tan, Wen Shi Lee, Kathleen M Wragg, Hannah G Kelly, Robyn Esterbauer, ... Stephen J Kent

Nature Communications (2021-02-19) <https://doi.org/gh9vd5>

DOI: [10.1038/s41467-021-21444-5](https://doi.org/10.1038/s41467-021-21444-5) · PMID: [33608522](#) · PMCID: [PMC7896046](#)

647. **COVID-19-neutralizing antibodies predict disease severity and survival**

Wilfredo F Garcia-Beltran, Evan C Lam, Michael G Astudillo, Diane Yang, Tyler E Miller, Jared Feldman, Blake M Hauser, Timothy M Caradonna, Kiera L Clayton, Adam D Nitido, ... Alejandro B Balazs

Cell (2021-01) <https://doi.org/gh9vdx>

DOI: [10.1016/j.cell.2020.12.015](https://doi.org/10.1016/j.cell.2020.12.015) · PMID: [33412089](#) · PMCID: [PMC7837114](#)

648. **Children develop robust and sustained cross-reactive spike-specific immune responses to SARS-CoV-2 infection**

Alexander C Dowell, Megan S Butler, Elizabeth Jinks, Gokhan Tut, Tara Lancaster, Panagiota Sylla, Jusnara Begum, Rachel Bruton, Hayden Pearce, Kriti Verma, ... Shamez Ladhami

Nature Immunology (2021-12-22) <https://doi.org/gnz6x6>

DOI: [10.1038/s41590-021-01089-8](https://doi.org/10.1038/s41590-021-01089-8) · PMID: [34937928](#) · PMCID: [PMC8709786](#)

649. **ABO blood group is involved in the quality of the specific immune response anti-SARS-CoV-2**

Sergio Gil-Manso, Iria Miguens Blanco, Bruce Motyka, Anne Halpin, Rocío López-Estebar, Verónica Astrid Pérez-Fernández, Diego Carbonell, Luis Andrés López-Fernández, Lori West, Rafael Correa-Rocha, Marjorie Pion

Virulence (2021-12-30) <https://doi.org/gpphhv>

DOI: [10.1080/21505594.2021.2019959](https://doi.org/10.1080/21505594.2021.2019959) · PMID: [34967260](#)

650. **Loss of Bcl-6-Expressing T Follicular Helper Cells and Germinal Centers in COVID-19**

Naoki Kaneko, Hsiao-Hsuan Kuo, Julie Boucau, Jocelyn R Farmer, Hugues Allard-Chamard, Vinay S Mahajan, Alicja Piechocka-Trocha, Kristina Lefteri, Matthew Osborn, Julia Bals, ... Shiv Pillai

Cell (2020-10) <https://doi.org/gg9rdv>

DOI: [10.1016/j.cell.2020.08.025](https://doi.org/10.1016/j.cell.2020.08.025) · PMID: [32877699](#) · PMCID: [PMC7437499](#)

651. **Robust SARS-CoV-2-specific T-cell immunity is maintained at 6 months following primary infection**

J Zuo, A Dowell, H Pearce, K Verma, HM Long, J Begum, F Aiano, Z Amin-Chowdhury, B Hallis, L Stapley, ... P Moss

Cold Spring Harbor Laboratory (2020-11-02) <https://doi.org/ghhrps>

DOI: [10.1101/2020.11.01.362319](https://doi.org/10.1101/2020.11.01.362319)

652. **Persistent Cellular Immunity to SARS-CoV-2 Infection**

Gaëlle Breton, Pilar Mendoza, Thomas Hagglof, Thiago Y Oliveira, Dennis Schaefer-Babajew, Christian Gaebler, Martina Turroja, Arlene

Hurley, Marina Caskey, Michel C Nussenzweig
Cold Spring Harbor Laboratory (2020-12-09) <https://doi.org/ghs9kk>
DOI: [10.1101/2020.12.08.416636](https://doi.org/10.1101/2020.12.08.416636) · PMID: [33330867](https://pubmed.ncbi.nlm.nih.gov/33330867/) · PMCID: [PMC7743071](https://pubmed.ncbi.nlm.nih.gov/PMC7743071/)

653. **Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19 Disease and Unexposed Individuals**

Alba Grifoni, Daniela Weiskopf, Sydney I Ramirez, Jose Mateus, Jennifer M Dan, Carolyn Rydznski Moderbacher, Stephen A Rawlings, Aaron Sutherland, Lakshmanane Premkumar, Ramesh S Jadi, ... Alessandro Sette

Cell (2020-06) <https://doi.org/ggzxz2>

DOI: [10.1016/j.cell.2020.05.015](https://doi.org/10.1016/j.cell.2020.05.015) · PMID: [32473127](https://pubmed.ncbi.nlm.nih.gov/32473127/) · PMCID: [PMC7237901](https://pubmed.ncbi.nlm.nih.gov/PMC7237901/)

654. **COVID-19 testing turns to T cells**

Cormac Sheridan

Nature Biotechnology (2021-05) <https://doi.org/gprdmb>

DOI: [10.1038/s41587-021-00920-9](https://doi.org/10.1038/s41587-021-00920-9) · PMID: [33981082](https://pubmed.ncbi.nlm.nih.gov/33981082/)

655. **Coronavirus (COVID-19) Update: FDA Authorizes Adaptive Biotechnologies T-Detect COVID Test**

Office of the Commissioner

FDA (2021-03-09) <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-adaptive-biotechnologies-t-detect-covid-test>

656. **Rethinking Covid-19 Test Sensitivity — A Strategy for Containment**

Michael J Mina, Roy Parker, Daniel B Larremore

New England Journal of Medicine (2020-11-26) <https://doi.org/ghdg6n>

DOI: [10.1056/nejmp2025631](https://doi.org/10.1056/nejmp2025631) · PMID: [32997903](https://pubmed.ncbi.nlm.nih.gov/32997903/)

657. **The time to do serosurveys for COVID-19 is now**

Rosanna W Peeling, Piero L Olliaro

The Lancet Respiratory Medicine (2020-09) <https://doi.org/gg559s>

DOI: [10.1016/s2213-2600\(20\)30313-1](https://doi.org/10.1016/s2213-2600(20)30313-1) · PMID: [32717209](https://pubmed.ncbi.nlm.nih.gov/32717209/) · PMCID: [PMC7380934](https://pubmed.ncbi.nlm.nih.gov/PMC7380934/)

658. **Accurate point-of-care serology tests for COVID-19**

Charles F Schuler, Carmen Gherasim, Kelly O'Shea, David M Manthei, Jesse Chen, Don Giacherio, Jonathan P Troost, James L Baldwin, James R Baker

PLOS ONE (2021-03-16) <https://doi.org/gpq4mz>

DOI: [10.1371/journal.pone.0248729](https://doi.org/10.1371/journal.pone.0248729) · PMID: [33725025](https://pubmed.ncbi.nlm.nih.gov/33725025/) · PMCID: [PMC7963097](https://pubmed.ncbi.nlm.nih.gov/PMC7963097/)

659. **Neutralizing Antibodies Against Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Variants Induced by Natural Infection or Vaccination: A Systematic Review and Pooled Analysis**

Xinhua Chen, Zhiyuan Chen, Andrew S Azman, Ruijia Sun, Wanying Lu, Nan Zheng, Jiaxin Zhou, Qianhui Wu, Xiaowei Deng, Zeyao Zhao, ... Hongjie Yu

Clinical Infectious Diseases (2021-07-24) <https://doi.org/gmdht2>

DOI: [10.1093/cid/ciab646](https://doi.org/10.1093/cid/ciab646) · PMID: [34302458](https://pubmed.ncbi.nlm.nih.gov/34302458/) · PMCID: [PMC9016754](https://pubmed.ncbi.nlm.nih.gov/PMC9016754/)

660. **COVID-19 serosurveys for public health decision making**
Manoj V Murhekar, Hannah Clapham
The Lancet Global Health (2021-05) <https://doi.org/gh7598>
DOI: [10.1016/s2214-109x\(21\)00057-7](https://doi.org/10.1016/s2214-109x(21)00057-7) · PMID: [33705691](#) · PMCID: [PMC8049585](#)
661. **COVID-19 Serological Tests: How Well Do They Actually Perform?**
Abdi Ghaffari, Robyn Meurant, Ali Ardakani
Diagnostics (2020-07-04) <https://doi.org/gg4h62>
DOI: [10.3390/diagnostics10070453](https://doi.org/10.3390/diagnostics10070453) · PMID: [32635444](#) · PMCID: [PMC7400479](#)
662. **Serological tests for COVID-19**
Katherine Bond, Eloise Williams, Benjamin P Howden, Deborah A Williamson
Medical Journal of Australia (2020-09-06) <https://doi.org/gpqt86>
DOI: [10.5694/mja2.50766](https://doi.org/10.5694/mja2.50766) · PMID: [32892381](#)
663. **A high-throughput multiplexed microfluidic device for COVID-19 serology assays**
Roberto Rodriguez-Moncayo, Diana F Cedillo-Alcantar, Pablo E Guevara-Pantoja, Oriana G Chavez-Pineda, Jose A Hernandez-Ortiz, Josue U Amador-Hernandez, Gustavo Rojas-Velasco, Fausto Sanchez-Muñoz, Daniel Manzur-Sandoval, Luis D Patino-Lopez, ... Jose L Garcia-Cordero
Lab on a Chip (2021) <https://doi.org/gpqt7j>
DOI: [10.1039/d0lc01068e](https://doi.org/10.1039/d0lc01068e) · PMID: [33319882](#)
664. **A systematic review of antibody mediated immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease**
Angkana T Huang, Bernardo Garcia-Carreras, Matt DT Hitchings, Bingyi Yang, Leah Katzelnick, Susan M Rattigan, Brooke Borgert, Carlos Moreno, Benjamin D Solomon, Isabel Rodriguez-Barraquer, ... Derek AT Cummings
Cold Spring Harbor Laboratory (2020-04-17) <https://doi.org/ggsfmz>
DOI: [10.1101/2020.04.14.20065771](https://doi.org/10.1101/2020.04.14.20065771) · PMID: [32511434](#)
665. **Antibodies, Immunity, and COVID-19**
Brad Spellberg, Travis B Nielsen, Arturo Casadevall
JAMA Internal Medicine (2021-04-01) <https://doi.org/gpphhs>
DOI: [10.1001/jamainternmed.2020.7986](https://doi.org/10.1001/jamainternmed.2020.7986) · PMID: [33231673](#) · PMCID: [PMC8371694](#)
666. **Protection and waning of natural and hybrid COVID-19 immunity**
Yair Goldberg, Micha Mandel, Yinon M Bar-On, Omri Bodenheimer, Laurence Freedman, Nachman Ash, Sharon Alroy-Preis, Amit Huppert, Ron Milo
Cold Spring Harbor Laboratory (2021-12-05) <https://doi.org/g9rq>
DOI: [10.1101/2021.12.04.21267114](https://doi.org/10.1101/2021.12.04.21267114)
667. **Strong associations and moderate predictive value of early symptoms for SARS-CoV-2 test positivity among healthcare workers, the Netherlands, March 2020**

Alma Tostmann, John Bradley, Teun Bousema, Wing-Kee Yiek, Minke Holwerda, Chantal Bleeker-Rovers, Jaap ten Oever, Corianne Meijer, Janette Rahamat-Langendoen, Joost Hopman, ... Heiman Wertheim
Eurosurveillance (2020-04-23) <https://doi.org/ggthwx>
DOI: [10.2807/1560-7917.es.2020.25.16.2000508](https://doi.org/10.2807/1560-7917.es.2020.25.16.2000508) · PMID: [32347200](#) ·
PMCID: [PMC7189649](#)

668. **Application and optimization of RT-PCR in diagnosis of SARS-CoV-2 infection**

Xiaoshuai Ren, Yan Liu, Hongtao Chen, Wei Liu, Zhaowang Guo, Yaqin Zhang, Chaoqun Chen, Jianhui Zhou, Qiang Xiao, Guanmin Jiang, Hong Shan
Cold Spring Harbor Laboratory (2020-02-27) <https://doi.org/gpq4k9>
DOI: [10.1101/2020.02.25.20027755](https://doi.org/10.1101/2020.02.25.20027755)

669. **Correlation of Chest CT and RT-PCR Testing for Coronavirus Disease 2019 (COVID-19) in China: A Report of 1014 Cases**

Tao Ai, Zhenlu Yang, Hongyan Hou, Chenao Zhan, Chong Chen, Wenzhi Lv, Qian Tao, Ziyong Sun, Liming Xia
Radiology (2020-08) <https://doi.org/ggmw6p>
DOI: [10.1148/radiol.2020200642](https://doi.org/10.1148/radiol.2020200642) · PMID: [32101510](#)

670. **Performance of Radiologists in Differentiating COVID-19 from Non-COVID-19 Viral Pneumonia at Chest CT**

Harrison X Bai, Ben Hsieh, Zeng Xiong, Kasey Halsey, Ji Whae Choi, Thi My Linh Tran, Ian Pan, Lin-Bo Shi, Dong-Cui Wang, Ji Mei, ... Wei-Hua Liao
Radiology (2020-08) <https://doi.org/ggnqw4>
DOI: [10.1148/radiol.2020200823](https://doi.org/10.1148/radiol.2020200823) · PMID: [32155105](#)

671. **Covid-19: automatic detection from X-ray images utilizing transfer learning with convolutional neural networks**

Ioannis D Apostolopoulos, Tzani A Mpesiana
Physical and Engineering Sciences in Medicine (2020-04-03)
<https://doi.org/ggs448>
DOI: [10.1007/s13246-020-00865-4](https://doi.org/10.1007/s13246-020-00865-4) · PMCID: [PMC7118364](#)

672. **Common pitfalls and recommendations for using machine learning to detect and prognosticate for COVID-19 using chest radiographs and CT scans**

Michael Roberts, Derek Driggs, Matthew Thorpe, Julian Gilbey, Michael Yeung, Stephan Ursprung, Angelica I Aviles-Rivero, Christian Etmann, Cathal McCague, ... Carola-Bibiane Schönlieb
Nature Machine Intelligence (2021-03) <https://doi.org/gjkjvw>
DOI: [10.1038/s42256-021-00307-0](https://doi.org/10.1038/s42256-021-00307-0)

673. **Epidemiologic surveillance for controlling Covid-19 pandemic: types, challenges and implications**

Nahla Khamis Ibrahim
Journal of Infection and Public Health (2020-11) <https://doi.org/gk7ghp>
DOI: [10.1016/j.jiph.2020.07.019](https://doi.org/10.1016/j.jiph.2020.07.019) · PMID: [32855090](#) · PMCID:
[PMC7441991](#)

674. **Wastewater and public health: the potential of wastewater surveillance for monitoring COVID-19**

Kata Farkas, Luke S Hillary, Shelagh K Malham, James E McDonald,

David L Jones

Current Opinion in Environmental Science & Health (2020-10)

<https://doi.org/gg4tb6>

DOI: [10.1016/j.coesh.2020.06.001](https://doi.org/10.1016/j.coesh.2020.06.001) · PMID: [32835157](https://pubmed.ncbi.nlm.nih.gov/32835157/) · PMCID:

[PMC7291992](https://pubmed.ncbi.nlm.nih.gov/PMC7291992/)

675. **Healthcare Workers**

CDC

Centers for Disease Control and Prevention (2020-02-11)

<https://www.cdc.gov/coronavirus/2019-ncov/hcp/testing-overview.html>

676. **NY Forward: a guide to reopening New York & building back better** (2020-05-15)

<https://www.governor.ny.gov/sites/governor.ny.gov/files/atoms/files/NYForwardReopeningGuide.pdf>

677. **Antigen tests for COVID-19**

Yuta Kyosei, Sou Yamura, Mayuri Namba, Teruki Yoshimura, Satoshi Watabe, Etsuro Ito

Biophysics and Physicobiology (2021) <https://doi.org/gpp4wr>

DOI: [10.2142/biophysico.bppb-v18.004](https://doi.org/10.2142/biophysico.bppb-v18.004) · PMID: [33954080](https://pubmed.ncbi.nlm.nih.gov/33954080/) · PMCID: [PMC8049777](https://pubmed.ncbi.nlm.nih.gov/PMC8049777/)

678. **Covid-19: Show us evidence for lifting restrictions, doctors tell Johnson**

Adele Waters

BMJ (2022-02-15) <https://doi.org/gprm69>

DOI: [10.1136/bmj.o383](https://doi.org/10.1136/bmj.o383) · PMID: [35168994](https://pubmed.ncbi.nlm.nih.gov/35168994/)

679. **Exit strategies from lockdowns due to COVID-19: a scoping review**

Madhavi Misra, Harsha Joshi, Rakesh Sarwal, Krishna D Rao

BMC Public Health (2022-03-12) <https://doi.org/gprm7c>

DOI: [10.1186/s12889-022-12845-2](https://doi.org/10.1186/s12889-022-12845-2) · PMID: [35279102](https://pubmed.ncbi.nlm.nih.gov/35279102/) · PMCID: [PMC8917328](https://pubmed.ncbi.nlm.nih.gov/PMC8917328/)

680. **Viral Cultures for Coronavirus Disease 2019 Infectivity Assessment: A Systematic Review**

Tom Jefferson, Elisabeth A Spencer, Jon Brassey, Carl Heneghan

Clinical Infectious Diseases (2020-12-03) <https://doi.org/ghpwks>

DOI: [10.1093/cid/ciaa1764](https://doi.org/10.1093/cid/ciaa1764) · PMID: [33270107](https://pubmed.ncbi.nlm.nih.gov/33270107/) · PMCID: [PMC7799320](https://pubmed.ncbi.nlm.nih.gov/PMC7799320/)

681. **SARS-CoV-2, SARS-CoV, and MERS-CoV viral load dynamics, duration of viral shedding, and infectiousness: a systematic review and meta-analysis**

Muge Cevik, Matthew Tate, Ollie Lloyd, Alberto Enrico Maraolo, Jenna Schafers, Antonia Ho

The Lancet Microbe (2021-01) <https://doi.org/ghk47x>

DOI: [10.1016/s2666-5247\(20\)30172-5](https://doi.org/10.1016/s2666-5247(20)30172-5) · PMID: [33521734](https://pubmed.ncbi.nlm.nih.gov/33521734/) · PMCID: [PMC7837230](https://pubmed.ncbi.nlm.nih.gov/PMC7837230/)

682. **Long-term SARS-CoV-2 RNA shedding and its temporal association to IgG seropositivity**

Vineet Agarwal, AJ Venkatakrishnan, Arjun Puranik, Christian Kirkup, Agustin Lopez-Marquez, Douglas W Challener, Elitza S Theel, John C

O'Horo, Matthew J Binnicker, Walter K Kremers, ... Venky Soundararajan
Cell Death Discovery (2020-12) <https://doi.org/gprm6q>
DOI: [10.1038/s41420-020-00375-y](https://doi.org/10.1038/s41420-020-00375-y) · PMID: [33298894](#) · PMCID: [PMC7709096](#)

683. **Trajectory of Viral RNA Load Among Persons With Incident SARS-CoV-2 G614 Infection (Wuhan Strain) in Association With COVID-19 Symptom Onset and Severity**
Helen C Stankiewicz Karita, Tracy Q Dong, Christine Johnston, Kathleen M Neuzil, Michael K Paasche-Orlow, Patricia J Kissinger, Anna Bershteyn, Lorna E Thorpe, Meagan Deming, Angelica Kottkamp, ... Elizabeth R Brown
JAMA Network Open (2022-01-10) <https://doi.org/gprm58>
DOI: [10.1001/jamanetworkopen.2021.42796](https://doi.org/10.1001/jamanetworkopen.2021.42796) · PMID: [35006245](#) · PMCID: [PMC8749477](#)
684. **Covid-19: Tests on students are highly inaccurate, early findings show**
Stephen Armstrong
BMJ (2020-12-23) <https://doi.org/gprm67>
DOI: [10.1136/bmj.m4941](https://doi.org/10.1136/bmj.m4941) · PMID: [33361271](#)
685. **Covid-19: Controversial rapid test policy divides doctors and scientists**
Zosia Kmietowicz
BMJ (2021-01-12) <https://doi.org/gprm68>
DOI: [10.1136/bmj.n81](https://doi.org/10.1136/bmj.n81) · PMID: [33436413](#)
686. **Assessment of the Analytical Sensitivity of 10 Lateral Flow Devices against the SARS-CoV-2 Omicron Variant**
Joshua Deerain, Julian Druce, Thomas Tran, Mitchell Batty, Yano Yoga, Michael Fennell, Dominic E Dwyer, Jen Kok, Deborah A Williamson
Journal of Clinical Microbiology (2022-02-16) <https://doi.org/gnvpb8>
DOI: [10.1128/jcm.02479-21](https://doi.org/10.1128/jcm.02479-21) · PMID: [34936477](#) · PMCID: [PMC8849215](#)
687. **Clarifying the evidence on SARS-CoV-2 antigen rapid tests in public health responses to COVID-19**
Michael J Mina, Tim E Peto, Marta García-Fiñana, Malcolm G Semple, Iain E Buchan
The Lancet (2021-04) <https://doi.org/gnmbjd>
DOI: [10.1016/s0140-6736\(21\)00425-6](https://doi.org/10.1016/s0140-6736(21)00425-6) · PMID: [33609444](#) · PMCID: [PMC8049601](#)
688. **Test sensitivity is secondary to frequency and turnaround time for COVID-19 screening**
Daniel B Larremore, Bryan Wilder, Evan Lester, Soraya Shehata, James M Burke, James A Hay, Milind Tambe, Michael J Mina, Roy Parker
Science Advances (2021-01) <https://doi.org/ghs8s7>
DOI: [10.1126/sciadv.abd5393](https://doi.org/10.1126/sciadv.abd5393) · PMID: [33219112](#) · PMCID: [PMC7775777](#)
689. **CDC chief says coronavirus cases may be 10 times higher than reported**
Washington Post

<https://www.washingtonpost.com/health/2020/06/25/coronavirus-cases-10-times-larger/>

690. **Liverpool coronavirus (COVID-19) community testing pilot: full evaluation report summary**

GOV.UK

<https://www.gov.uk/government/publications/liverpool-coronavirus-covid-19-community-testing-pilot-full-evaluation-report-summary/liverpool-coronavirus-covid-19-community-testing-pilot-full-evaluation-report-summary>

691. **Covid-19: Government rolls out twice weekly rapid testing to all in England**

Gareth Iacobucci

BMJ (2021-04-06) <https://doi.org/gprcnx>

DOI: [10.1136/bmj.n902](https://doi.org/10.1136/bmj.n902) · PMID: [33824178](#)

692. **A comparative study of COVID-19 responses in South Korea and Japan: political nexus triad and policy responses**

M Jae Moon, Kohei Suzuki, Tae In Park, Kentaro Sakuwa

International Review of Administrative Sciences (2021-03-18)

<https://doi.org/gprcn2>

DOI: [10.1177/0020852321997552](https://doi.org/10.1177/0020852321997552) · PMCID: [PMC8685564](#)

693. **All things equal? Heterogeneity in policy effectiveness against COVID-19 spread in chile**

Magdalena Bennett

World Development (2021-01) <https://doi.org/gjggkh>

DOI: [10.1016/j.worlddev.2020.105208](https://doi.org/10.1016/j.worlddev.2020.105208) · PMID: [32994662](#) · PMCID:

[PMC7513907](#)

694. **UK ending Covid testing 'very worrying' as WHO chief warns pandemic 'isn't over'**

The Independent

(2022-03-11) <https://www.independent.co.uk/news/health/who-covid-testing-anil-soni-b2032884.html>

695. **UK scales back routine covid-19 surveillance**

Jonathan Clarke, Thomas Beaney, Azeem Majeed

BMJ (2022-03-04) <https://doi.org/gprcnz>

DOI: [10.1136/bmj.o562](https://doi.org/10.1136/bmj.o562) · PMID: [35246445](#)

696. **Local groups continue push for COVID testing and vaccinations as larger state-run sites plan to close**

KUSA.com

(2022-03-10)

<https://www.9news.com/article/news/health/coronavirus/local-groups-continue-push-for-covid-testing-and-vaccinations/73-bbcd8384-d96a-425e-aaeb-16ac9f36e581>

697. **Utah will stop daily COVID case counts, close test sites in wind-down, Gov. Cox announces**

The Salt Lake Tribune

<https://www.sltrib.com/news/2022/02/18/utah-will-stop-daily/>

698. **Safety, tolerability and viral kinetics during SARS-CoV-2 human challenge in young adults**
Ben Killingley, Alex J Mann, Mariya Kalinova, Alison Boyers, Niluka Goonawardane, Jie Zhou, Kate Lindsell, Samanjit S Hare, Jonathan Brown, Rebecca Frise, ... Christopher Chiu
Nature Medicine (2022-03-31) <https://doi.org/gptbvp>
DOI: [10.1038/s41591-022-01780-9](https://doi.org/10.1038/s41591-022-01780-9) · PMID: [35361992](#)
699. **Considerations for the Safe Operation of Schools During the Coronavirus Pandemic**
Ronan Lordan, Samantha Prior, Elizabeth Hennessy, Amruta Naik, Soumita Ghosh, Georgios K Paschos, Carsten Skarke, Kayla Barekat, Taylor Hollingsworth, Sydney Juska, ... Tilo Grosser
Frontiers in Public Health (2021-12-16) <https://doi.org/gprq6z>
DOI: [10.3389/fpubh.2021.751451](https://doi.org/10.3389/fpubh.2021.751451) · PMID: [34976917](#) · PMCID: [PMC8716382](#)
700. **SARS-CoV-2 infection and transmission in school settings during the second COVID-19 wave: a cross-sectional study, Berlin, Germany, November 2020**
Stefanie Theuring, Marlene Thielecke, Welmoed van Loon, Franziska Hommes, Claudia Hülso, Annkathrin von der Haar, Jennifer Körner, Michael Schmidt, Falko Böhringer, Marcus A Mall, ...
Eurosurveillance (2021-08-26) <https://doi.org/gprq6x>
DOI: [10.2807/1560-7917.es.2021.26.34.2100184](https://doi.org/10.2807/1560-7917.es.2021.26.34.2100184) · PMID: [34448448](#) · PMCID: [PMC8393892](#)
701. **End to quarantines, contact tracing among changes in new Oregon guidance for schools, starting March 12**
Elizabeth Miller
OPB (2022-03-02) <https://www.opb.org/article/2022/03/02/oregon-schools-guidance-mask-mandate-end/>
702. **Palm Beach County public schools to stop COVID-19 contact tracing**
WPTV
(2022-03-11) <https://www.wptv.com/news/education/palm-beach-county-public-schools-to-stop-covid-19-contact-tracing>
703. **Covid News: C.D.C. Drops Contact Tracing Recommendation**
Adeel Hassan
The New York Times (2022-03-02)
<https://www.nytimes.com/live/2022/03/02/world/covid-19-tests-cases-vaccine>
704. **Children and COVID-19: State-Level Data Report**
American Academy of Pediatrics
<http://www.aap.org/en/pages/2019-novel-coronavirus-covid-19-infections/children-and-covid-19-state-level-data-report/>
705. **A fifth of all US child Covid deaths occurred during Omicron surge**
Melody Schreiber
The Guardian (2022-03-11)
<https://www.theguardian.com/world/2022/mar/11/us-child-covid-deaths-omicron-surge>

706. **Global, regional, and national minimum estimates of children affected by COVID-19-associated orphanhood and caregiver death, by age and family circumstance up to Oct 31, 2021: an updated modelling study**
HJuliette T Unwin, Susan Hillis, Lucie Cluver, Seth Flaxman, Philip S Goldman, Alexander Butchart, Gretchen Bachman, Laura Rawlings, Christl A Donnelly, Oliver Ratmann, ... Lorraine Sherr
The Lancet Child & Adolescent Health (2022-04)
<https://doi.org/gpr264>
DOI: [10.1016/s2352-4642\(22\)00005-0](https://doi.org/s2352-4642(22)00005-0) · PMID: [35219404](https://pubmed.ncbi.nlm.nih.gov/35219404/) · PMCID: [PMC8872796](https://pubmed.ncbi.nlm.nih.gov/PMC8872796/)
707. **Five Ways that COVID-19 Diagnostics Can Save Lives: Prioritizing Uses of Tests to Maximize Cost-Effectiveness**
Tristan Reed, William Waites, David Manheim, Damien de Walque, Chiara Vallini, Roberta Gatti, Timothy B Hallett
World Bank (2021-02-23)
<https://openknowledge.worldbank.org/handle/10986/35150>
708. **A Visual Approach for the SARS (Severe Acute Respiratory Syndrome) Outbreak Data Analysis**
Jie Hua, Guohua Wang, Maolin Huang, Shuyang Hua, Shuanghe Yang
International Journal of Environmental Research and Public Health
(2020-06-03) <https://doi.org/gjqqg6z>
DOI: [10.3390/ijerph17113973](https://doi.org/10.3390/ijerph17113973) · PMID: [32503333](https://pubmed.ncbi.nlm.nih.gov/32503333/) · PMCID: [PMC7312089](https://pubmed.ncbi.nlm.nih.gov/PMC7312089/)
709. **COVID-19 Data Repository**
Center for Systems Science and Engineering at Johns Hopkins University
GitHub https://github.com/CSSEGISandData/COVID-19/tree/master/csse_covid_19_data/csse_covid_19_time_series
710. **An interactive web-based dashboard to track COVID-19 in real time**
Ensheng Dong, Hongru Du, Lauren Gardner
The Lancet Infectious Diseases (2020-05) <https://doi.org/ggnjsk>
DOI: [10.1016/s1473-3099\(20\)30120-1](https://doi.org/s1473-3099(20)30120-1) · PMID: [32087114](https://pubmed.ncbi.nlm.nih.gov/32087114/) · PMCID: [PMC7159018](https://pubmed.ncbi.nlm.nih.gov/PMC7159018/)
711. <https://www.who.int/csr/sars/country/en>
712. **GitHub - imdevskp/sars-2003-outbreak-data-webscraping-code: repository contains complete WHO data of 2003 outbreak with code used to web scrap, data mung and cleaning**
GitHub
<https://github.com/imdevskp/sars-2003-outbreak-data-webscraping-code>
713. **Covid-19 has redefined airborne transmission**
Julian W Tang, Linsey C Marr, Yuguo Li, Stephanie J Dancer
BMJ (2021-04-14) <https://doi.org/gj3jh4>
DOI: [10.1136/bmj.n913](https://doi.org/10.1136/bmj.n913) · PMID: [33853842](https://pubmed.ncbi.nlm.nih.gov/33853842/)
714. **Isolation of a Novel Coronavirus from a Man with Pneumonia in Saudi Arabia**

Ali M Zaki, Sander van Boheemen, Theo M Bestebroer, Albert DME Osterhaus, Ron AM Fouchier
New England Journal of Medicine (2012-11-08) <https://doi.org/f4czx5>
DOI: [10.1056/nejmoa1211721](https://doi.org/10.1056/nejmoa1211721) · PMID: [23075143](https://pubmed.ncbi.nlm.nih.gov/23075143/)

715. **Drug repurposing: progress, challenges and recommendations**

Sudeep Pushpakom, Francesco Iorio, Patrick A Eyers, Kjane Escott, Shirley Hopper, Andrew Wells, Andrew Doig, Tim Guilliams, Joanna Latimer, Christine McNamee, ... Munir Pirmohamed
Nature Reviews Drug Discovery (2018-10-12) <https://doi.org/gfrbsz>
DOI: [10.1038/nrd.2018.168](https://doi.org/10.1038/nrd.2018.168) · PMID: [30310233](https://pubmed.ncbi.nlm.nih.gov/30310233/)

716. **Drug discovery and development: Role of basic biological research**

Richard C Mohs, Nigel H Greig
Alzheimer's & Dementia: Translational Research & Clinical Interventions (2017-11) <https://doi.org/gf92kj>
DOI: [10.1016/j.trci.2017.10.005](https://doi.org/10.1016/j.trci.2017.10.005) · PMID: [29255791](https://pubmed.ncbi.nlm.nih.gov/29255791/) · PMCID: [PMC5725284](https://pubmed.ncbi.nlm.nih.gov/PMC5725284/)

717. **Evidence-Based Medicine Data Lab COVID-19 TrialsTracker**

Nick DeVito, Peter Inglesby
GitHub (2020-03-29)
https://github.com/ebmdata/covid_trials_tracker-covid
DOI: [10.5281/zenodo.3732709](https://zenodo.org/record/3732709)

718. **Drug treatments for covid-19: living systematic review and network meta-analysis**

Reed AC Siemieniuk, Jessica J Bartoszko, Long Ge, Dena Zeraatkar, Ariel Izcovich, Elena Kum, Hector Pardo-Hernandez, Anila Qasim, Juan Pablo Díaz Martinez, Bram Rochwerg, ... Romina Brignardello-Petersen
BMJ (2020-07-30) <https://doi.org/ghs8st>
DOI: [10.1136/bmj.m2980](https://doi.org/10.1136/bmj.m2980) · PMID: [32732190](https://pubmed.ncbi.nlm.nih.gov/32732190/) · PMCID: [PMC7390912](https://pubmed.ncbi.nlm.nih.gov/PMC7390912/)

719. **Causes of Death and Comorbidities in Patients with COVID-19**

Sefer Elezkurtaj, Selina Greuel, Jana Ihlow, Edward Michaelis, Philip Bischoff, Catarina Alisa Kunze, Bruno Valentin Sinn, Manuela Gerhold, Kathrin Hauptmann, Barbara Ingold-Heppner, ... David Horst
Cold Spring Harbor Laboratory (2020-06-17) <https://doi.org/gg926j>
DOI: [10.1101/2020.06.15.20131540](https://doi.org/10.1101/2020.06.15.20131540)

720. **Clinical characteristics of 82 cases of death from COVID-19**

Bicheng Zhang, Xiaoyang Zhou, Yanru Qiu, Yuxiao Song, Fan Feng, Jia Feng, Qibin Song, Qingzhu Jia, Jun Wang
PLOS ONE (2020-07-09) <https://doi.org/gg4sgx>
DOI: [10.1371/journal.pone.0235458](https://doi.org/10.1371/journal.pone.0235458) · PMID: [32645044](https://pubmed.ncbi.nlm.nih.gov/32645044/) · PMCID: [PMC7347130](https://pubmed.ncbi.nlm.nih.gov/PMC7347130/)

721. **COVID-19 infection: the perspectives on immune responses**

Yufang Shi, Ying Wang, Changshun Shao, Jianan Huang, Jianhe Gan, Xiaoping Huang, Enrico Bucci, Mauro Piacentini, Giuseppe Ippolito, Gerry Melino
Cell Death & Differentiation (2020-03-23) <https://doi.org/ggg8td>
DOI: [10.1038/s41418-020-0530-3](https://doi.org/10.1038/s41418-020-0530-3) · PMID: [32205856](https://pubmed.ncbi.nlm.nih.gov/32205856/) · PMCID: [PMC7091918](https://pubmed.ncbi.nlm.nih.gov/PMC7091918/)

722. **Cytokine Storm**
David C Fajgenbaum, Carl H June
New England Journal of Medicine (2020-12-03) <https://doi.org/ghnhm7>
DOI: [10.1056/nejmra2026131](https://doi.org/10.1056/nejmra2026131) · PMID: [33264547](https://pubmed.ncbi.nlm.nih.gov/33264547/) · PMCID: [PMC7727315](https://pubmed.ncbi.nlm.nih.gov/PMC7727315/)
723. **Lung pathology of fatal severe acute respiratory syndrome**
John M Nicholls, Leo LM Poon, Kam C Lee, Wai F Ng, Sik T Lai, Chung Y Leung, Chung M Chu, Pak K Hui, Kong L Mak, Wilna Lim, ... JS Malik Peiris
The Lancet (2003-05) <https://doi.org/c8mmbg>
DOI: [10.1016/s0140-6736\(03\)13413-7](https://doi.org/10.1016/s0140-6736(03)13413-7) · PMID: [12781536](https://pubmed.ncbi.nlm.nih.gov/12781536/) · PMCID: [PMC7112492](https://pubmed.ncbi.nlm.nih.gov/PMC7112492/)
724. **Pro/con clinical debate: Steroids are a key component in the treatment of SARS**
Charles D Gomersall, Marcus J Kargel, Stephen E Lapinsky
Critical Care (2004) <https://doi.org/dpj29>
DOI: [10.1186/cc2452](https://doi.org/10.1186/cc2452) · PMID: [15025770](https://pubmed.ncbi.nlm.nih.gov/15025770/) · PMCID: [PMC420028](https://pubmed.ncbi.nlm.nih.gov/PMC420028/)
725. **Content Analysis and Characterization of Medical Tweets During the Early Covid-19 Pandemic**
Ross Prager, Michael T Pratte, Rudy R Unni, Sudarshan Bala, Nicholas Ng Fat Hing, Kay Wu, Trevor A McGrath, Adam Thomas, Brent Thoma, Kwadwo Kyeremanteng
Cureus (2021-02-27) <https://doi.org/gjpccg>
DOI: [10.7759/cureus.13594](https://doi.org/10.7759/cureus.13594) · PMID: [33815994](https://pubmed.ncbi.nlm.nih.gov/33815994/) · PMCID: [PMC8007019](https://pubmed.ncbi.nlm.nih.gov/PMC8007019/)
726. **Small Molecules vs Biologics | Drug Development Differences**
Nuventra Pharma Sciences 2525 Meridian Parkway, Suite 200 Durham
PK / PD and Clinical Pharmacology Consultants (2020-05-13)
<https://www.nuventra.com/resources/blog/small-molecules-versus-biologics/>
727. **Drug Discovery: A Historical Perspective**
J Drews
Science (2000-03-17) <https://doi.org/d6bvp7>
DOI: [10.1126/science.287.5460.1960](https://doi.org/10.1126/science.287.5460.1960) · PMID: [10720314](https://pubmed.ncbi.nlm.nih.gov/10720314/)
728. **Prone Positioning in Awake, Nonintubated Patients With COVID-19 Hypoxic Respiratory Failure**
Alison E Thompson, Benjamin L Ranard, Ying Wei, Sanja Jelic
JAMA Internal Medicine (2020-11-01) <https://doi.org/gg2pq4>
DOI: [10.1001/jamainternmed.2020.3030](https://doi.org/10.1001/jamainternmed.2020.3030) · PMID: [32584946](https://pubmed.ncbi.nlm.nih.gov/32584946/) · PMCID: [PMC7301298](https://pubmed.ncbi.nlm.nih.gov/PMC7301298/)
729. **SARS: Systematic Review of Treatment Effects**
Lauren J Stockman, Richard Bellamy, Paul Garner
PLoS Medicine (2006-09-12) <https://doi.org/d7xwh2>
DOI: [10.1371/journal.pmed.0030343](https://doi.org/10.1371/journal.pmed.0030343) · PMID: [16968120](https://pubmed.ncbi.nlm.nih.gov/16968120/) · PMCID: [PMC1564166](https://pubmed.ncbi.nlm.nih.gov/PMC1564166/)
730. **Current concepts in SARS treatment**
Takeshi Fujii, Aikichi Iwamoto, Tetsuya Nakamura, Aikichi Iwamoto
Journal of Infection and Chemotherapy (2004) <https://doi.org/dpmxk2>

DOI: [10.1007/s10156-003-0296-9](https://doi.org/10.1007/s10156-003-0296-9) · PMID: [14991510](https://pubmed.ncbi.nlm.nih.gov/14991510/) · PMCID: [PMC7088022](https://pubmed.ncbi.nlm.nih.gov/PMC7088022/)

731. **Corticosteroids for pneumonia**

Anat Stern, Keren Skalsky, Tomer Avni, Elena Carrara, Leonard Leibovici, Mical Paul

Cochrane Database of Systematic Reviews (2017-12-13)

<https://doi.org/gc9cdk>

DOI: [10.1002/14651858.cd007720.pub3](https://doi.org/10.1002/14651858.cd007720.pub3) · PMID: [29236286](https://pubmed.ncbi.nlm.nih.gov/29236286/) · PMCID: [PMC6486210](https://pubmed.ncbi.nlm.nih.gov/PMC6486210/)

732. **Corticosteroids for pneumonia**

Yuanjing Chen, Ka Li, Hongshan Pu, Taixiang Wu

Cochrane Database of Systematic Reviews (2011-03-16)

<https://doi.org/cvc92x>

DOI: [10.1002/14651858.cd007720.pub2](https://doi.org/10.1002/14651858.cd007720.pub2) · PMID: [21412908](https://pubmed.ncbi.nlm.nih.gov/21412908/)

733. **Corticosteroids in severe pneumonia**

O Sibila, C Agusti, A Torres

European Respiratory Journal (2008-03-19) <https://doi.org/bmdrvg>

DOI: [10.1183/09031936.00154107](https://doi.org/10.1183/09031936.00154107) · PMID: [18669784](https://pubmed.ncbi.nlm.nih.gov/18669784/)

734. **Efficacy of Corticosteroids in the Treatment of Community-Acquired Pneumonia Requiring Hospitalization**

Katsunaka Mikami, Masaru Suzuki, Hiroshi Kitagawa, Masaki Kawakami, Nobuaki Hirota, Hiromichi Yamaguchi, Osamu Narumoto, Yoshiko Kichikawa, Makoto Kawai, Hiroyuki Tashimo, ... Yoshio Sakamoto

Lung (2007-08-21) <https://doi.org/fk5f5d>

DOI: [10.1007/s00408-007-9020-3](https://doi.org/10.1007/s00408-007-9020-3) · PMID: [17710485](https://pubmed.ncbi.nlm.nih.gov/17710485/)

735. **Corticosteroids in the Treatment of Community-Acquired Pneumonia in Adults: A Meta-Analysis**

Wei Nie, Yi Zhang, Jinwei Cheng, Qingyu Xiu

PLoS ONE (2012-10-24) <https://doi.org/gj3jh5>

DOI: [10.1371/journal.pone.0047926](https://doi.org/10.1371/journal.pone.0047926) · PMID: [23112872](https://pubmed.ncbi.nlm.nih.gov/23112872/) · PMCID: [PMC3480455](https://pubmed.ncbi.nlm.nih.gov/PMC3480455/)

736. **Effect of corticosteroids on the clinical course of community-acquired pneumonia: a randomized controlled trial**

Silvia Fernández-Serrano, Jordi Dorca, Carolina García-Vidal, Núria Fernández-Sabé, Jordi Carratalà, Ana Fernández-Agüera, Mercè Corominas, Susana Padrones, Francesc Gudiol, Frederic Manresa

Critical Care (2011) <https://doi.org/c8ksgr>

DOI: [10.1186/cc10103](https://doi.org/10.1186/cc10103) · PMID: [21406101](https://pubmed.ncbi.nlm.nih.gov/21406101/) · PMCID: [PMC3219361](https://pubmed.ncbi.nlm.nih.gov/PMC3219361/)

737. **Dexamethasone treatment for the acute respiratory distress syndrome: a multicentre, randomised controlled trial**

Jesús Villar, Carlos Ferrando, Domingo Martínez, Alfonso Ambrós, Tomás Muñoz, Juan A Soler, Gerardo Aguilar, Francisco Alba, Elena González-Higueras, Luís A Conesa, ... Jesús Villar

The Lancet Respiratory Medicine (2020-03) <https://doi.org/ggpzxc>

DOI: [10.1016/s2213-2600\(19\)30417-5](https://doi.org/10.1016/s2213-2600(19)30417-5)

738. **Corticosteroids in acute respiratory distress syndrome: a step forward, but more evidence is needed**
Kiran Reddy, Cecilia O'Kane, Daniel McAuley
The Lancet Respiratory Medicine (2020-03) <https://doi.org/gcv2>
DOI: [10.1016/s2213-2600\(20\)30048-5](https://doi.org/s2213-2600(20)30048-5)
739. **Nonventilatory Treatments for Acute Lung Injury and ARDS**
Carolyn S Calfee, Michael A Matthay
Chest (2007-03) <https://doi.org/bqzn5v>
DOI: [10.1378/chest.06-1743](https://doi.org/10.1378/chest.06-1743) · PMID: [17356114](#) · PMCID: [PMC2789489](#)
740. **Corticosteroids in ARDS**
GUmberio Meduri, Paul E Marik, Stephen M Pastores, Djillali Annane
Chest (2007-09) <https://doi.org/cjdz2d>
DOI: [10.1378/chest.07-0714](https://doi.org/10.1378/chest.07-0714) · PMID: [17873207](#)
741. **Efficacy and Safety of Corticosteroids for Persistent Acute Respiratory Distress Syndrome**
New England Journal of Medicine
N Engl J Med (2006-04-20) <https://doi.org/c3sfcb>
DOI: [10.1056/nejmoa051693](https://doi.org/10.1056/nejmoa051693) · PMID: [16625008](#)
742. **Corticosteroids in the prevention and treatment of acute respiratory distress syndrome (ARDS) in adults: meta-analysis**
John Victor Peter, Preeta John, Petra L Graham, John L Moran, Ige Abraham George, Andrew Bersten
BMJ (2008-04-23) <https://doi.org/b7qtn2>
DOI: [10.1136/bmj.39537.939039.be](https://doi.org/10.1136/bmj.39537.939039.be) · PMID: [18434379](#) · PMCID: [PMC2364864](#)
743. **Antiviral agents and corticosteroids in the treatment of severe acute respiratory syndrome (SARS)**
WC Yu
Thorax (2004-08-01) <https://doi.org/bks99t>
DOI: [10.1136/thx.2003.017665](https://doi.org/10.1136/thx.2003.017665) · PMID: [15282381](#) · PMCID: [PMC1747111](#)
744. **Corticosteroid treatment of severe acute respiratory syndrome in Hong Kong**
Loretta Yin-Chun Yam, Arthur Chun-Wing Lau, Florence Yuk-Lin Lai, Edwina Shung, Jane Chan, Vivian Wong
Journal of Infection (2007-01) <https://doi.org/dffg65>
DOI: [10.1016/j.jinf.2006.01.005](https://doi.org/10.1016/j.jinf.2006.01.005) · PMID: [16542729](#) · PMCID: [PMC7112522](#)
745. **Impact of corticosteroid therapy on outcomes of persons with SARS-CoV-2, SARS-CoV, or MERS-CoV infection: a systematic review and meta-analysis**
Huan Li, Chongxiang Chen, Fang Hu, Jiaoqiao Wang, Qingyu Zhao, Robert Peter Gale, Yang Liang
Leukemia (2020-05-05) <https://doi.org/ggv2rb>
DOI: [10.1038/s41375-020-0848-3](https://doi.org/10.1038/s41375-020-0848-3) · PMID: [32372026](#) · PMCID: [PMC7199650](#)
746. **Managing SARS amidst Uncertainty**

Richard P Wenzel, Michael B Edmond
New England Journal of Medicine (2003-05-15) <https://doi.org/ddkjnr>
DOI: [10.1056/nejmp030072](https://doi.org/10.1056/nejmp030072) · PMID: [12748313](https://pubmed.ncbi.nlm.nih.gov/12748313/)

747. **Synthesis and Pharmacology of Anti-Inflammatory Steroidal Antedrugs**

MOmar F Khan, Henry J Lee
Chemical Reviews (2008-12-10) <https://doi.org/cmkrtc>
DOI: [10.1021/cr068203e](https://doi.org/10.1021/cr068203e) · PMID: [19035773](https://pubmed.ncbi.nlm.nih.gov/19035773/) · PMCID: [PMC2650492](https://pubmed.ncbi.nlm.nih.gov/PMC2650492/)

748. **Drug vignettes: Dexamethasone**

The Centre for Evidence-Based Medicine
<https://www.cebm.net/covid-19/dexamethasone/>

749. **Pharmacology of Postoperative Nausea and Vomiting**

Eric S Zabirowicz, Tong J Gan
Elsevier BV (2019) <https://doi.org/ghfkjw>
DOI: [10.1016/b978-0-323-48110-6.00034-x](https://doi.org/10.1016/b978-0-323-48110-6.00034-x)

750. **Potential benefits of precise corticosteroids therapy for severe 2019-nCoV pneumonia**

Wei Zhou, Yisi Liu, Dongdong Tian, Cheng Wang, Sa Wang, Jing Cheng, Ming Hu, Minghao Fang, Yue Gao
Signal Transduction and Targeted Therapy (2020-02-21)
<https://doi.org/ggqr84>
DOI: [10.1038/s41392-020-0127-9](https://doi.org/10.1038/s41392-020-0127-9) · PMID: [32296012](https://pubmed.ncbi.nlm.nih.gov/32296012/) · PMCID: [PMC7035340](https://pubmed.ncbi.nlm.nih.gov/PMC7035340/)

751. **16-METHYLATED STEROIDS. I. 16 α -METHYLATED ANALOGS OF CORTISONE, A NEW GROUP OF ANTI-INFLAMMATORY STEROIDS**

Glen E Arth, David BR Johnston, John Fried, William W Spooncer, Dale R Hoff, Lewis H Sarett
Journal of the American Chemical Society (2002-05-01)
<https://doi.org/cj5c82>
DOI: [10.1021/ja01545a061](https://doi.org/10.1021/ja01545a061)

752. **Treatment of Rheumatoid Arthritis with Dexamethasone**

Abraham Cohen
JAMA (1960-10-15) <https://doi.org/csfrmhc>
DOI: [10.1001/jama.1960.03030070009002](https://doi.org/10.1001/jama.1960.03030070009002) · PMID: [13694317](https://pubmed.ncbi.nlm.nih.gov/13694317/)

753. **Dexamethasone**

DailyMed
(2007-10-25) <https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?setid=537b424a-3e07-4c81-978c-1ad99014032a>

754. **Prevention of infection caused by immunosuppressive drugs in gastroenterology**

Katarzyna Orlicka, Eleanor Barnes, Emma L Culver
Therapeutic Advances in Chronic Disease (2013-04-22)
<https://doi.org/ggrqd3>
DOI: [10.1177/2040622313485275](https://doi.org/10.1177/2040622313485275) · PMID: [23819020](https://pubmed.ncbi.nlm.nih.gov/23819020/) · PMCID: [PMC3697844](https://pubmed.ncbi.nlm.nih.gov/PMC3697844/)

755. **Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury**
Clark D Russell, Jonathan E Millar, JKenneth Baillie
The Lancet (2020-02) <https://doi.org/ggks86>
DOI: [10.1016/s0140-6736\(20\)30317-2](https://doi.org/s0140-6736(20)30317-2) · PMID: [32043983](https://pubmed.ncbi.nlm.nih.gov/32043983/) · PMCID: [PMC7134694](https://pubmed.ncbi.nlm.nih.gov/PMC7134694/)
756. **On the use of corticosteroids for 2019-nCoV pneumonia**
Lianhan Shang, Jianping Zhao, Yi Hu, Ronghui Du, Bin Cao
The Lancet (2020-02) <https://doi.org/ggg356>
DOI: [10.1016/s0140-6736\(20\)30361-5](https://doi.org/s0140-6736(20)30361-5) · PMID: [32122468](https://pubmed.ncbi.nlm.nih.gov/32122468/) · PMCID: [PMC7159292](https://pubmed.ncbi.nlm.nih.gov/PMC7159292/)
757. **Immunosuppression for hyperinflammation in COVID-19: a double-edged sword?**
Andrew I Ritchie, Aran Singanayagam
The Lancet (2020-04) <https://doi.org/ggq8hs>
DOI: [10.1016/s0140-6736\(20\)30691-7](https://doi.org/s0140-6736(20)30691-7) · PMID: [32220278](https://pubmed.ncbi.nlm.nih.gov/32220278/) · PMCID: [PMC7138169](https://pubmed.ncbi.nlm.nih.gov/PMC7138169/)
758. **Effect of Dexamethasone in Hospitalized Patients with COVID-19 – Preliminary Report**
Peter Horby, Wei Shen Lim, Jonathan Emberson, Marion Mafham, Jennifer Bell, Louise Linsell, Natalie Staplin, Christopher Brightling, Andrew Ustianowski, Einas Elmahi, ... RECOVERY Collaborative Group
Cold Spring Harbor Laboratory (2020-06-22) <https://doi.org/dz5x>
DOI: [10.1101/2020.06.22.20137273](https://doi.org/10.1101/2020.06.22.20137273)
759. **Dexamethasone in Hospitalized Patients with Covid-19 — Preliminary Report**
The RECOVERY Collaborative Group
New England Journal of Medicine (2020-07-17) <https://doi.org/gg5c8p>
DOI: [10.1056/nejmoa2021436](https://doi.org/10.1056/nejmoa2021436) · PMID: [32678530](https://pubmed.ncbi.nlm.nih.gov/32678530/) · PMCID: [PMC7383595](https://pubmed.ncbi.nlm.nih.gov/PMC7383595/)
760. **Corticosteroids for Patients With Coronavirus Disease 2019 (COVID-19) With Different Disease Severity: A Meta-Analysis of Randomized Clinical Trials**
Laura Pasin, Paolo Navalesi, Alberto Zangrillo, Artem Kuzovlev, Valery Likhvantsev, Ludhmila Abrahão Hajjar, Stefano Fresilli, Marcus Vinicius Guimaraes Lacerda, Giovanni Landoni
Journal of Cardiothoracic and Vascular Anesthesia (2021-02) <https://doi.org/ghzkp9>
DOI: [10.1053/j.jvca.2020.11.057](https://doi.org/10.1053/j.jvca.2020.11.057) · PMID: [33298370](https://pubmed.ncbi.nlm.nih.gov/33298370/) · PMCID: [PMC7698829](https://pubmed.ncbi.nlm.nih.gov/PMC7698829/)
761. **Current concepts in the diagnosis and management of cytokine release syndrome**
Daniel W Lee, Rebecca Gardner, David L Porter, Chrystal U Louis, Nabil Ahmed, Michael Jensen, Stephan A Grupp, Crystal L Mackall
Blood (2014-07-10) <https://doi.org/ggsrwk>
DOI: [10.1182/blood-2014-05-552729](https://doi.org/10.1182/blood-2014-05-552729) · PMID: [24876563](https://pubmed.ncbi.nlm.nih.gov/24876563/) · PMCID: [PMC4093680](https://pubmed.ncbi.nlm.nih.gov/PMC4093680/)
762. **Corticosteroids in COVID-19 ARDS**
Hallie C Prescott, Todd W Rice

763. **Dexamethasone: Therapeutic potential, risks, and future projection during COVID-19 pandemic**
Sobia Noreen, Irsah Maqbool, Asadullah Madni
European Journal of Pharmacology (2021-03) <https://doi.org/gj4qgn>
DOI: [10.1016/j.ejphar.2021.173854](https://doi.org/10.1016/j.ejphar.2021.173854) · PMID: [33428898](#) · PMCID: [PMC7836247](#)
764. **Covid-19: Demand for dexamethasone surges as RECOVERY trial publishes preprint**
Elisabeth Mahase
BMJ (2020-06-23) <https://doi.org/gj4qgp>
DOI: [10.1136/bmj.m2512](https://doi.org/10.1136/bmj.m2512) · PMID: [32576548](#)
765. **Introduction to modern virology**
NJ Dimmock, AJ Easton, KN Leppard
Blackwell Pub (2007)
ISBN: 9781405136457
766. **CORONA Data Viewer**
Castleman Disease Collaborative Network
<https://cdcn.org/corona-data-viewer/>
767. **Coronaviruses**
Helena Jane Maier, Erica Bickerton, Paul Britton (editors)
Methods in Molecular Biology (2015) <https://doi.org/ggqfqx>
DOI: [10.1007/978-1-4939-2438-7](https://doi.org/10.1007/978-1-4939-2438-7) · PMID: [25870870](#) · ISBN: 9781493924370
768. **The potential chemical structure of anti-SARS-CoV-2 RNA-dependent RNA polymerase**
Jrhau Lung, Yu-Shih Lin, Yao-Hsu Yang, Yu-Lun Chou, Li-Hsin Shu, Yu-Ching Cheng, Hung Te Liu, Ching-Yuan Wu
Journal of Medical Virology (2020-03-18) <https://doi.org/ggp6fm>
DOI: [10.1002/jmv.25761](https://doi.org/10.1002/jmv.25761) · PMID: [32167173](#)
769. **Broad-spectrum coronavirus antiviral drug discovery**
Allison L Totura, Sina Bavari
Expert Opinion on Drug Discovery (2019-03-08) <https://doi.org/gg74z5>
DOI: [10.1080/17460441.2019.1581171](https://doi.org/10.1080/17460441.2019.1581171) · PMID: [30849247](#) · PMCID: [PMC7103675](#)
770. **Ribavirin therapy for severe COVID-19: a retrospective cohort study**
Song Tong, Yuan Su, Yuan Yu, Chuangyan Wu, Jiuling Chen, Sihua Wang, Jinjun Jiang
International Journal of Antimicrobial Agents (2020-09) <https://doi.org/gg5w75>
DOI: [10.1016/j.ijantimicag.2020.106114](https://doi.org/10.1016/j.ijantimicag.2020.106114) · PMID: [32712334](#) · PMCID: [PMC7377772](#)
771. **The evolution of nucleoside analogue antivirals: A review for chemists and non-chemists. Part 1: Early structural modifications**

to the nucleoside scaffold

Katherine L Seley-Radtke, Mary K Yates

Antiviral Research (2018-06) <https://doi.org/gdpn35>

DOI: [10.1016/j.antiviral.2018.04.004](https://doi.org/10.1016/j.antiviral.2018.04.004) · PMID: [29649496](https://pubmed.ncbi.nlm.nih.gov/29649496/) · PMCID: [PMC6396324](https://pubmed.ncbi.nlm.nih.gov/PMC6396324/)

772. The Ambiguous Base-Pairing and High Substrate Efficiency of T-705 (Favipiravir) Ribofuranosyl 5'-Triphosphate towards Influenza A Virus Polymerase

Zhinan Jin, Lucas K Smith, Vivek K Rajwanshi, Baek Kim, Jerome Deval
PLoS ONE (2013-07-10) <https://doi.org/f5br92>

DOI: [10.1371/journal.pone.0068347](https://doi.org/10.1371/journal.pone.0068347) · PMID: [23874596](https://pubmed.ncbi.nlm.nih.gov/23874596/) · PMCID: [PMC3707847](https://pubmed.ncbi.nlm.nih.gov/PMC3707847/)

773. Favipiravir

DrugBank

(2020-06-12) <https://www.drugbank.ca/drugs/DB12466>

774. In Vitro and In Vivo Activities of Anti-Influenza Virus Compound T-705

Y Furuta, K Takahashi, Y Fukuda, M Kuno, T Kamiyama, K Kozaki, N

Nomura, H Egawa, S Minami, Y Watanabe, ... K Shiraki

Antimicrobial Agents and Chemotherapy (2002-04)

<https://doi.org/cndw7n>

DOI: [10.1128/aac.46.4.977-981.2002](https://doi.org/10.1128/aac.46.4.977-981.2002) · PMID: [11897578](https://pubmed.ncbi.nlm.nih.gov/11897578/) · PMCID: [PMC127093](https://pubmed.ncbi.nlm.nih.gov/PMC127093/)

775. Efficacy of Orally Administered T-705 on Lethal Avian Influenza A (H5N1) Virus Infections in Mice

Robert W Sidwell, Dale L Barnard, Craig W Day, Donald F Smee, Kevin W Bailey, Min-Hui Wong, John D Morrey, Yousuke Furuta

Antimicrobial Agents and Chemotherapy (2007-03)

<https://doi.org/dm9xr2>

DOI: [10.1128/aac.01051-06](https://doi.org/10.1128/aac.01051-06) · PMID: [17194832](https://pubmed.ncbi.nlm.nih.gov/17194832/) · PMCID: [PMC1803113](https://pubmed.ncbi.nlm.nih.gov/PMC1803113/)

776. Mechanism of Action of T-705 against Influenza Virus

Yousuke Furuta, Kazumi Takahashi, Masako Kuno-Maekawa, Hidehiro Sangawa, Sayuri Uehara, Kyo Kozaki, Nobuhiko Nomura, Hiroyuki Egawa, Kimiyasu Shiraki

Antimicrobial Agents and Chemotherapy (2005-03)

<https://doi.org/dgbwdh>

DOI: [10.1128/aac.49.3.981-986.2005](https://doi.org/10.1128/aac.49.3.981-986.2005) · PMID: [15728892](https://pubmed.ncbi.nlm.nih.gov/15728892/) · PMCID: [PMC549233](https://pubmed.ncbi.nlm.nih.gov/PMC549233/)

777. Activity of T-705 in a Hamster Model of Yellow Fever Virus Infection in Comparison with That of a Chemically Related Compound, T-1106

Justin G Julander, Kristiina Shafer, Donald F Smee, John D Morrey, Yousuke Furuta

Antimicrobial Agents and Chemotherapy (2009-01)

<https://doi.org;brknds>

DOI: [10.1128/aac.01074-08](https://doi.org/10.1128/aac.01074-08) · PMID: [18955536](https://pubmed.ncbi.nlm.nih.gov/18955536/) · PMCID: [PMC2612161](https://pubmed.ncbi.nlm.nih.gov/PMC2612161/)

778. In Vitro and In Vivo Activities of T-705 against Arenavirus and Bunyavirus Infections

Brian B Gowen, Min-Hui Wong, Kie-Hoon Jung, Andrew B Sanders, Michelle Mendenhall, Kevin W Bailey, Yousuke Furuta, Robert W Sidwell

Antimicrobial Agents and Chemotherapy (2007-09)

<https://doi.org/d98c87>

DOI: [10.1128/aac.00356-07](https://doi.org/10.1128/aac.00356-07) · PMID: [17606691](https://pubmed.ncbi.nlm.nih.gov/17606691/) · PMCID: [PMC2043187](https://pubmed.ncbi.nlm.nih.gov/PMC2043187/)

779. **Favipiravir (T-705) inhibits in vitro norovirus replication**

J Rocha-Pereira, D Jochmans, K Dallmeier, P Leyssen, MSJ Nascimento, J Neys

Biochemical and Biophysical Research Communications (2012-08)

<https://doi.org/f369jZ>

DOI: [10.1016/j.bbrc.2012.07.034](https://doi.org/10.1016/j.bbrc.2012.07.034) · PMID: [22809499](https://pubmed.ncbi.nlm.nih.gov/22809499/)

780. **T-705 (Favipiravir) Inhibition of Arenavirus Replication in Cell Culture**

Michelle Mendenhall, Andrew Russell, Terry Juelich, Emily L Messina, Donald F Smee, Alexander N Freiberg, Michael R Holbrook, Yousuke Furuta, Juan-Carlos de la Torre, Jack H Nunberg, Brian B Gowen

Antimicrobial Agents and Chemotherapy (2011-02)

<https://doi.org/cppwsc>

DOI: [10.1128/aac.01219-10](https://doi.org/10.1128/aac.01219-10) · PMID: [21115797](https://pubmed.ncbi.nlm.nih.gov/21115797/) · PMCID: [PMC3028760](https://pubmed.ncbi.nlm.nih.gov/PMC3028760/)

781. **Favipiravir (T-705), a broad spectrum inhibitor of viral RNA polymerase**

Yousuke FURUTA, Takashi KOMENO, Takaaki NAKAMURA

Proceedings of the Japan Academy, Series B (2017)

<https://doi.org/gbxcxw>

DOI: [10.2183/pjab.93.027](https://doi.org/10.2183/pjab.93.027) · PMID: [28769016](https://pubmed.ncbi.nlm.nih.gov/28769016/) · PMCID: [PMC5713175](https://pubmed.ncbi.nlm.nih.gov/PMC5713175/)

782. **The antiviral compound remdesivir potently inhibits RNA-dependent RNA polymerase from Middle East respiratory syndrome coronavirus**

Calvin J Gordon, Egor P Tchesnokov, Joy Y Feng, Danielle P Porter, Matthias Götte

Journal of Biological Chemistry (2020-04) <https://doi.org/ggqm6x>

DOI: [10.1074/jbc.ac120.013056](https://doi.org/10.1074/jbc.ac120.013056) · PMID: [32094225](https://pubmed.ncbi.nlm.nih.gov/32094225/)

783. **Coronavirus Susceptibility to the Antiviral Remdesivir (GS-5734) Is Mediated by the Viral Polymerase and the Proofreading Exoribonuclease**

Maria L Agostini, Erica L Andres, Amy C Sims, Rachel L Graham, Timothy P Sheahan, Xiaotao Lu, Everett Clinton Smith, James Brett Case, Joy Y Feng, Robert Jordan, ... Mark R Denison

mBio (2018-03-06) <https://doi.org/gc45v6>

DOI: [10.1128/mbio.00221-18](https://doi.org/10.1128/mbio.00221-18) · PMID: [29511076](https://pubmed.ncbi.nlm.nih.gov/29511076/) · PMCID: [PMC5844999](https://pubmed.ncbi.nlm.nih.gov/PMC5844999/)

784. **Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro**

Manli Wang, Ruiyuan Cao, Leike Zhang, Xinglou Yang, Jia Liu, Mingyue Xu, Zhengli Shi, Zhihong Hu, Wu Zhong, Gengfu Xiao

Cell Research (2020-02-04) <https://doi.org/ggkbsg>

DOI: [10.1038/s41422-020-0282-0](https://doi.org/10.1038/s41422-020-0282-0) · PMID: [32020029](https://pubmed.ncbi.nlm.nih.gov/32020029/) · PMCID: [PMC7054408](https://pubmed.ncbi.nlm.nih.gov/PMC7054408/)

785. **Remdesivir for the Treatment of Covid-19 — Final Report**
John H Beigel, Kay M Tomashek, Lori E Dodd, Aneesh K Mehta, Barry S Zingman, Andre C Kalil, Elizabeth Hohmann, Helen Y Chu, Annie Luetkemeyer, Susan Kline, ... HClifford Lane
New England Journal of Medicine (2020-11-05) <https://doi.org/dwkd>
DOI: [10.1056/nejmoa2007764](https://doi.org/10.1056/nejmoa2007764) · PMID: [32445440](https://pubmed.ncbi.nlm.nih.gov/32445440/) · PMCID: [PMC7262788](https://pubmed.ncbi.nlm.nih.gov/PMC7262788/)
786. **A Multicenter, Adaptive, Randomized Blinded Controlled Trial of the Safety and Efficacy of Investigational Therapeutics for the Treatment of COVID-19 in Hospitalized Adults**
National Institute of Allergy and Infectious Diseases (NIAID)
clinicaltrials.gov (2020-12-05)
<https://clinicaltrials.gov/ct2/show/NCT04280705>
787. **A Phase 3 Randomized Study to Evaluate the Safety and Antiviral Activity of Remdesivir (GS-5734™) in Participants With Severe COVID-19**
Gilead Sciences
clinicaltrials.gov (2020-12-15)
<https://clinicaltrials.gov/ct2/show/NCT04292899>
788. **Compassionate Use of Remdesivir for Patients with Severe Covid-19**
Jonathan Grein, Norio Ohmagari, Daniel Shin, George Diaz, Erika Asperges, Antonella Castagna, Torsten Feldt, Gary Green, Margaret L Green, François-Xavier Lescure, ... Timothy Flanigan
New England Journal of Medicine (2020-06-11) <https://doi.org/ggrm99>
DOI: [10.1056/nejmoa2007016](https://doi.org/10.1056/nejmoa2007016) · PMID: [32275812](https://pubmed.ncbi.nlm.nih.gov/32275812/) · PMCID: [PMC7169476](https://pubmed.ncbi.nlm.nih.gov/PMC7169476/)
789. **Repurposed Antiviral Drugs for Covid-19 — Interim WHO Solidarity Trial Results**
WHO Solidarity Trial Consortium
New England Journal of Medicine (2020-12-02) <https://doi.org/ghnhnw>
DOI: [10.1056/nejmoa2023184](https://doi.org/10.1056/nejmoa2023184) · PMID: [33264556](https://pubmed.ncbi.nlm.nih.gov/33264556/) · PMCID: [PMC7727327](https://pubmed.ncbi.nlm.nih.gov/PMC7727327/)
790. **PLAQUENIL - hydroxychloroquine sulfate tablet**
DailyMed
(2020-08-12) <https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?setid=34496b43-05a2-45fb-a769-52b12e099341>
791. **New concepts in antimalarial use and mode of action in dermatology**
Sunil Kalia, Jan P Dutz
Dermatologic Therapy (2007-07) <https://doi.org/fv69cb>
DOI: [10.1111/j.1529-8019.2007.00131.x](https://doi.org/10.1111/j.1529-8019.2007.00131.x) · PMID: [17970883](https://pubmed.ncbi.nlm.nih.gov/17970883/) · PMCID: [PMC7163426](https://pubmed.ncbi.nlm.nih.gov/PMC7163426/)
792. **Chloroquine is a potent inhibitor of SARS coronavirus infection and spread**
Martin J Vincent, Eric Bergeron, Suzanne Benjannet, Bobbie R Erickson, Pierre E Rollin, Thomas G Ksiazek, Nabil G Seidah, Stuart T Nichol
Virology Journal (2005) <https://doi.org/dvbds4>
DOI: [10.1186/1743-422x-2-69](https://doi.org/10.1186/1743-422x-2-69) · PMID: [16115318](https://pubmed.ncbi.nlm.nih.gov/16115318/) · PMCID: [PMC1232869](https://pubmed.ncbi.nlm.nih.gov/PMC1232869/)

793. **In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2)**
Xueting Yao, Fei Ye, Miao Zhang, Cheng Cui, Baoying Huang, Peihua Niu, Xu Liu, Li Zhao, Erdan Dong, Chunli Song, ... Dongyang Liu
Clinical Infectious Diseases (2020-08-01) <https://doi.org/ggpx7z>
DOI: [10.1093/cid/ciaa237](https://doi.org/10.1093/cid/ciaa237) · PMID: [32150618](https://pubmed.ncbi.nlm.nih.gov/32150618/) · PMCID: [PMC7108130](https://pubmed.ncbi.nlm.nih.gov/PMC7108130/)
794. **Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial**
Philippe Gautret, Jean-Christophe Lagier, Philippe Parola, Van Thuan Hoang, Line Meddeb, Morgane Mailhe, Barbara Doudier, Johan Courjon, Valérie Giordanengo, Vera Esteves Vieira, ... Didier Raoult
International Journal of Antimicrobial Agents (2020-07)
<https://doi.org/dp7d>
DOI: [10.1016/j.ijantimicag.2020.105949](https://doi.org/10.1016/j.ijantimicag.2020.105949) · PMID: [32205204](https://pubmed.ncbi.nlm.nih.gov/32205204/) · PMCID: [PMC7102549](https://pubmed.ncbi.nlm.nih.gov/PMC7102549/)
795. **Official Statement from International Society of Antimicrobial Chemotherapy**
Andreas Voss
(2020-04-03) <https://www.isac.world/news-and-publications/official-isac-statement>
796. **Effect of Hydroxychloroquine in Hospitalized Patients with Covid-19**
The RECOVERY Collaborative Group
New England Journal of Medicine (2020-11-19) <https://doi.org/ghd8c7>
DOI: [10.1056/nejmoa2022926](https://doi.org/10.1056/nejmoa2022926) · PMID: [33031652](https://pubmed.ncbi.nlm.nih.gov/33031652/) · PMCID: [PMC7556338](https://pubmed.ncbi.nlm.nih.gov/PMC7556338/)
797. **No clinical benefit from use of hydroxychloroquine in hospitalised patients with COVID-19 — RECOVERY Trial**
<https://www.recoverytrial.net/news/statement-from-the-chief-investigators-of-the-randomised-evaluation-of-covid-19-therapy-recovery-trial-on-hydroxychloroquine-5-june-2020-no-clinical-benefit-from-use-of-hydroxychloroquine-in-hospitalised-patients-with-covid-19>
798. **The life and times of ivermectin — a success story**
Satoshi Ōmura, Andy Crump
Nature Reviews Microbiology (2004-12) <https://doi.org/fftvr8>
DOI: [10.1038/nrmicro1048](https://doi.org/10.1038/nrmicro1048) · PMID: [15550944](https://pubmed.ncbi.nlm.nih.gov/15550944/)
799. **Avermectins, New Family of Potent Anthelmintic Agents: Producing Organism and Fermentation**
Richard W Burg, Brinton M Miller, Edward E Baker, Jerome Birnbaum, Sara A Currie, Robert Hartman, Yu-Lin Kong, Richard L Monaghan, George Olson, Irving Putter, ... Satoshi Ōmura
Antimicrobial Agents and Chemotherapy (1979-03)
<https://doi.org/gmd8cj>
DOI: [10.1128/aac.15.3.361](https://doi.org/10.1128/aac.15.3.361) · PMID: [464561](https://pubmed.ncbi.nlm.nih.gov/464561/) · PMCID: [PMC352666](https://pubmed.ncbi.nlm.nih.gov/PMC352666/)
800. **Ivermectin, 'Wonder drug' from Japan: the human use perspective**
Andy CRUMP, Satoshi OMURA
Proceedings of the Japan Academy, Series B (2011)
<https://doi.org/cpq4wk>

801. **Ivermectin: enigmatic multifaceted ‘wonder’ drug continues to surprise and exceed expectations**
Andy Crump
The Journal of Antibiotics (2017-02-15) <https://doi.org/gmd8cf>
DOI: [10.1038/ja.2017.11](https://doi.org/10.1038/ja.2017.11) · PMID: [28196978](https://pubmed.ncbi.nlm.nih.gov/28196978/)
802. **Avermectins, New Family of Potent Anthelmintic Agents: Efficacy of the B1a Component**
JR Egerton, DA Ostlind, LS Blair, CH Eary, D Suhayda, S Cifelli, RF Riek, WC Campbell
Antimicrobial Agents and Chemotherapy (1979-03) <https://doi.org/825>
DOI: [10.1128/aac.15.3.372](https://doi.org/10.1128/aac.15.3.372) · PMID: [464563](https://pubmed.ncbi.nlm.nih.gov/464563/) · PMCID: [PMC352668](https://pubmed.ncbi.nlm.nih.gov/PMC352668/)
803. **Ivermectin: a systematic review from antiviral effects to COVID-19 complementary regimen**
Fatemeh Heidary, Reza Gharebaghi
The Journal of Antibiotics (2020-06-12) <https://doi.org/ghcz8p>
DOI: [10.1038/s41429-020-0336-z](https://doi.org/s41429-020-0336-z) · PMID: [32533071](https://pubmed.ncbi.nlm.nih.gov/32533071/) · PMCID: [PMC7290143](https://pubmed.ncbi.nlm.nih.gov/PMC7290143/)
804. **Ivermectin, a new candidate therapeutic against SARS-CoV-2/COVID-19**
Khan Sharun, Kuldeep Dhamma, Shailesh Kumar Patel, Mamta Pathak, Ruchi Tiwari, Bhoj Raj Singh, Ranjit Sah, DKatterine Bonilla-Aldana, Alfonso J Rodriguez-Morales, Hakan Leblebicioglu
Annals of Clinical Microbiology and Antimicrobials (2020-05-30) <https://doi.org/gmhmf>
DOI: [10.1186/s12941-020-00368-w](https://doi.org/10.1186/s12941-020-00368-w) · PMID: [32473642](https://pubmed.ncbi.nlm.nih.gov/32473642/) · PMCID: [PMC7261036](https://pubmed.ncbi.nlm.nih.gov/PMC7261036/)
805. **The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 in vitro**
Leon Caly, Julian D Druce, Mike G Catton, David A Jans, Kylie M Wagstaff
Antiviral Research (2020-06) <https://doi.org/ggqvsj>
DOI: [10.1016/j.antiviral.2020.104787](https://doi.org/10.1016/j.antiviral.2020.104787) · PMID: [32251768](https://pubmed.ncbi.nlm.nih.gov/32251768/) · PMCID: [PMC7129059](https://pubmed.ncbi.nlm.nih.gov/PMC7129059/)
806. **Ivermectin and COVID-19: Keeping Rigor in Times of Urgency**
Carlos Chaccour, Felix Hammann, Santiago Ramón-García, NRegina Rabinovich
The American Journal of Tropical Medicine and Hygiene (2020-06-03) <https://doi.org/gj6kbh>
DOI: [10.4269/ajtmh.20-0271](https://doi.org/10.4269/ajtmh.20-0271) · PMID: [32314704](https://pubmed.ncbi.nlm.nih.gov/32314704/) · PMCID: [PMC7253113](https://pubmed.ncbi.nlm.nih.gov/PMC7253113/)
807. **The Approved Dose of Ivermectin Alone is not the Ideal Dose for the Treatment of COVID-19**
Virginia D Schmith, Jie (Jessie) Zhou, Lauren RL Lohmer
Clinical Pharmacology & Therapeutics (2020-06-07) <https://doi.org/ggvcz2>
DOI: [10.1002/cpt.1889](https://doi.org/10.1002/cpt.1889) · PMID: [32378737](https://pubmed.ncbi.nlm.nih.gov/32378737/) · PMCID: [PMC7267287](https://pubmed.ncbi.nlm.nih.gov/PMC7267287/)

808. **Ivermectin as a potential COVID-19 treatment from the pharmacokinetic point of view: antiviral levels are not likely attainable with known dosing regimens**
Georgi Momekov, Denitsa Momekova
Biotechnology & Biotechnological Equipment (2020-01-01)
<https://doi.org/gj6kbm>
DOI: [10.1080/13102818.2020.1775118](https://doi.org/10.1080/13102818.2020.1775118)
809. **Relative Neurotoxicity of Ivermectin and Moxidectin in Mdr1ab (-/-) Mice and Effects on Mammalian GABA(A) Channel Activity**
Cécile Ménez, Jean-François Sutra, Roger Prichard, Anne Lespine
PLoS Neglected Tropical Diseases (2012-11-01) <https://doi.org/gmhmfh>
DOI: [10.1371/journal.pntd.0001883](https://doi.org/10.1371/journal.pntd.0001883) · PMID: [23133688](#) · PMCID: [PMC3486876](#)
810. **Use of Ivermectin Is Associated With Lower Mortality in Hospitalized Patients With Coronavirus Disease 2019**
Juliana Cepelowicz Rajter, Michael S Sherman, Naaz Fatteh, Fabio Vogel, Jamie Sacks, Jean-Jacques Rajter
Chest (2021-01) <https://doi.org/gjr28f>
DOI: [10.1016/j.chest.2020.10.009](https://doi.org/10.1016/j.chest.2020.10.009) · PMID: [33065103](#) · PMCID: [PMC7550891](#)
811. **Lack of efficacy of standard doses of ivermectin in severe COVID-19 patients**
Daniel Camprubí, Alex Almuedo-Riera, Helena Martí-Soler, Alex Soriano, Juan Carlos Hurtado, Carme Subirà, Berta Grau-Pujol, Alejandro Krolewiecki, Jose Muñoz
PLOS ONE (2020-11-11) <https://doi.org/gmhmfj>
DOI: [10.1371/journal.pone.0242184](https://doi.org/10.1371/journal.pone.0242184) · PMID: [33175880](#) · PMCID: [PMC7657540](#)
812. **Ivermectin as an adjunct treatment for hospitalized adult COVID-19 patients: A randomized multi-center clinical trial**
Nematollah Gheibi, Morteza Shakhs Niae, Peyman Namdar, Abbas Allami, Leila Zolghadr, Amir Javadi, Amin Karampour, Mehran Varnaseri, Behzad Bijani, Fatemeh Cheraghi, ... Ramin Jamshidian
Asian Pacific Journal of Tropical Medicine (2021)
<https://doi.org/gmhmfjp>
DOI: [10.4103/1995-7645.318304](https://doi.org/10.4103/1995-7645.318304)
813. **Ivermectin shows clinical benefits in mild to moderate COVID19: a randomized controlled double-blind, dose-response study in Lagos**
OE Babalola, CO Bode, AA Ajayi, FM Alakaloko, IE Akase, E Otrofanowei, OB Salu, WL Adeyemo, AO Ademuyiwa, S Omilabu
QJM: An International Journal of Medicine (2021-02-18)
<https://doi.org/gmhbg5>
DOI: [10.1093/qjmed/hcab035](https://doi.org/10.1093/qjmed/hcab035) · PMID: [33599247](#) · PMCID: [PMC7928689](#)
814. **Use of ivermectin in the treatment of Covid-19: A pilot trial**
Henrique Pott-Junior, Mônica Maria Bastos Paoliello, Alice de Queiroz Constantino Miguel, Anderson Ferreira da Cunha, Caio Cesar de Melo Freire, Fábio Fernandes Neves, Lucimar Retto da Silva de Avó, Meliza

Goi Roscani, Sigrid De Sousa dos Santos, Silvana Gama Florêncio

Chachá

Toxicology Reports (2021) <https://doi.org/gmhmdc>

DOI: [10.1016/j.toxrep.2021.03.003](https://doi.org/j.toxrep.2021.03.003) · PMID: [33723507](#) · PMCID:

[PMC7942165](#)

815. **A five-day course of ivermectin for the treatment of COVID-19 may reduce the duration of illness**

Sabeena Ahmed, Mohammad Mahbubul Karim, Allen G Ross, Mohammad Sharif Hossain, John D Clemens, Mariya Kibtiya Sumiya, Ching Swe Phru, Mustafizur Rahman, Khalequ Zaman, Jyoti Somani, ... Wasif Ali Khan

International Journal of Infectious Diseases (2021-02)

<https://doi.org/gjwcdt>

DOI: [10.1016/j.ijid.2020.11.191](https://doi.org/j.ijid.2020.11.191) · PMID: [33278625](#) · PMCID:

[PMC7709596](#)

816. **Effects of Ivermectin in Patients With COVID-19: A Multicenter, Double-blind, Randomized, Controlled Clinical Trial**

Leila Shahbaznejad, Alireza Davoudi, Gohar Eslami, John S Markowitz, Mohammad Reza Navaeifar, Fatemeh Hosseinzadeh, Faeze Sadat Movahedi, Mohammad Sadegh Rezai

Clinical Therapeutics (2021-06) <https://doi.org/gmb256>

DOI: [10.1016/j.clinthera.2021.04.007](https://doi.org/j.clinthera.2021.04.007) · PMID: [34052007](#) · PMCID:

[PMC8101859](#)

817. **The effect of early treatment with ivermectin on viral load, symptoms and humoral response in patients with non-severe COVID-19: A pilot, double-blind, placebo-controlled, randomized clinical trial**

Carlos Chaccour, Aina Casellas, Andrés Blanco-Di Matteo, Iñigo Pineda, Alejandro Fernandez-Montero, Paula Ruiz-Castillo, Mary-Ann Richardson, Mariano Rodríguez-Mateos, Carlota Jordán-Iborra, Joe Brew, ... Mirian Fernández-Alonso

EClinicalMedicine (2021-02) <https://doi.org/gmf4d3>

DOI: [10.1016/j.eclim.2020.100720](https://doi.org/j.eclim.2020.100720) · PMID: [33495752](#) · PMCID:

[PMC7816625](#)

818. **Ivermectin in mild and moderate COVID-19 (RIVET-COV): a randomized, placebo-controlled trial**

Anant Mohan, Pawan Tiwari, Tejas Suri, Saurabh Mittal, Ankit Patel, Avinash Jain, Velpandian T., Ujjwal Kumar Das, Tarun K Bopanna, RM Pandey, ... Randeep Guleria

Research Square Platform LLC (2021-02-02) <https://doi.org/gmh3hq>

DOI: [10.21203/rs.3.rs-191648/v1](https://doi.org/10.21203/rs.3.rs-191648/v1)

819. **Evaluation of Ivermectin as a Potential Treatment for Mild to Moderate COVID-19: A Double-Blind Randomized Placebo Controlled Trial in Eastern India**

Ravikirti, Ranjini Roy, Chandrima Pattadar, Rishav Raj, Neeraj Agarwal, Biju Biswas, Pramod Kumar Manjhi, Deependra Kumar Rai, Shyama, Anjani Kumar, Asim Sarfaraz

Journal of Pharmacy & Pharmaceutical Sciences (2021-07-15)

<https://doi.org/gmhmfk>

DOI: [10.18433/jpps32105](https://doi.org/10.18433/jpps32105) · PMID: [34265236](#)

820. **Efficacy and Safety of Ivermectin for Treatment and prophylaxis of COVID-19 Pandemic**
Ahmed Elgazzar, Abdelaziz Eltawee, Shaimaa Abo Youssef, Basma Hany, Mohy Hafez, Hany Moussa
Research Square Platform LLC (2020-12-28) <https://doi.org/gmhmfr>
DOI: [10.21203/rs.3.rs-100956/v3](https://doi.org/10.21203/rs.3.rs-100956/v3)
821. **Why Was a Major Study on Ivermectin for COVID-19 Just Retracted?**
Jack Lawrence
Grftr News (2021-07-15) <https://grftr.news/why-was-a-major-study-on-ivermectin-for-covid-19-just-retracted/>
822. **Nick Brown's blog: Some problems in the dataset of a large study of Ivermectin for the treatment of Covid-19**
Nick Brown
Nick Brown's blog (2021-07-15)
<https://steamtraen.blogspot.com/2021/07/Some-problems-with-the-data-from-a-Covid-study.html>
823. **Efficacy and Safety of Ivermectin for Treatment and prophylaxis of COVID-19 Pandemic**
Research Square Platform LLC
(2021-07-14) <https://doi.org/gmhfm>
DOI: [10.21203/rs.3.rs-100956/v4](https://doi.org/10.21203/rs.3.rs-100956/v4)
824. **Effect of Ivermectin on Time to Resolution of Symptoms Among Adults With Mild COVID-19**
Eduardo López-Medina, Pío López, Isabel C Hurtado, Diana M Dávalos, Oscar Ramirez, Ernesto Martínez, Jesus A Díazgranados, José M Oñate, Hector Chavarriaga, Sócrates Herrera, ... Isabella Caicedo
JAMA (2021-04-13) <https://doi.org/gift3s>
DOI: [10.1001/jama.2021.3071](https://doi.org/10.1001/jama.2021.3071) · PMID: [33662102](https://pubmed.ncbi.nlm.nih.gov/33662102/) · PMCID: [PMC7934083](https://pubmed.ncbi.nlm.nih.gov/PMC7934083/)
825. **Ivermectin for the treatment of COVID-19: A systematic review and meta-analysis of randomized controlled trials**
Yuani M Roman, Paula Alejandra Burela, Vinay Pasupuleti, Alejandro Piscoya, Jose E Vidal, Adrian V Hernandez
Cold Spring Harbor Laboratory (2021-05-25) <https://doi.org/gmh3hv>
DOI: [10.1101/2021.05.21.21257595](https://doi.org/10.1101/2021.05.21.21257595)
826. **Outcomes of Ivermectin in the treatment of COVID-19: a systematic review and meta-analysis**
Alex Castañeda-Sabogal, Diego Chambergo-Michilot, Carlos J Toro-Huamanchumo, Christian Silva-Rengifo, José Gonzales-Zamora, Joshua J Barboza
Cold Spring Harbor Laboratory (2021-01-27) <https://doi.org/gmhmd>
DOI: [10.1101/2021.01.26.21250420](https://doi.org/10.1101/2021.01.26.21250420)
827. **RETRACTED: Expression of Concern: “Meta-analysis of Randomized Trials of Ivermectin to Treat SARS-CoV-2 Infection”**
Andrew Hill, Anna Garratt, Jacob Levi, Jonathan Falconer, Leah Ellis, Kaitlyn McCann, Victoria Pilkington, Ambar Qavi, Junzheng Wang, Hannah Wentzel
Open Forum Infectious Diseases (2021-08-01) <https://doi.org/gmhrz6>

828. **Ivermectin and mortality in patients with COVID-19: A systematic review, meta-analysis, and meta-regression of randomized controlled trials**

Ahmad Fariz Malvi Zamzam Zein, Catur Setiya Sulistiyyana, Wilson Matthew Raffaelo, Raymond Pranata

Diabetes & Metabolic Syndrome: Clinical Research & Reviews (2021-07)

<https://doi.org/gmh mdb>

DOI: [10.1016/j.dsx.2021.102186](https://doi.org/10.1016/j.dsx.2021.102186) · PMID: [34237554](https://pubmed.ncbi.nlm.nih.gov/34237554/) · PMCID: [PMC8236126](https://pubmed.ncbi.nlm.nih.gov/PMC8236126/)

829. **Ivermectin for Prevention and Treatment of COVID-19 Infection**

Andrew Bryant, Theresa A Lawrie, Therese Dowswell, Edmund J Fordham, Scott Mitchell, Sarah R Hill, Tony C Tham

American Journal of Therapeutics (2021-06-17) <https://doi.org/gksqvz>

DOI: [10.1097/mjt.0000000000001402](https://doi.org/10.1097/mjt.0000000000001402) · PMID: [34145166](https://pubmed.ncbi.nlm.nih.gov/34145166/) · PMCID: [PMC8248252](https://pubmed.ncbi.nlm.nih.gov/PMC8248252/)

830. **Ivermectin and outcomes from Covid-19 pneumonia: A systematic review and meta-analysis of randomized clinical trial studies**

Timotius Ivan Hariyanto, Devina Adella Halim, Jane Rosalind, Catherine Gunawan, Andree Kurniawan

Reviews in Medical Virology (2021-06-06) <https://doi.org/gmhmc9>

DOI: [10.1002/rmv.2265](https://doi.org/10.1002/rmv.2265) · PMCID: [PMC8209939](https://pubmed.ncbi.nlm.nih.gov/PMC8209939/)

831. **Ivermectin for prevention and treatment of COVID-19 infection: a systematic review and meta-analysis**

Andrew Bryant, Theresa A Lawrie, Therese Dowswell, Edmund Fordham, Mitchell Scott MD, Sarah R Hill, Tony C Tham

Center for Open Science (2021-03-11) <https://doi.org/gmhmf n>

DOI: [10.31219/osf.io/k37ft](https://doi.org/10.31219/osf.io/k37ft)

832. **Review of the Emerging Evidence Demonstrating the Efficacy of Ivermectin in the Prophylaxis and Treatment of COVID-19**

Pierre Kory, Gianfranco Umberto Meduri, Joseph Varon, Jose Iglesias, Paul E Marik

American Journal of Therapeutics (2021-04-22) <https://doi.org/gjxvpr>

DOI: [10.1097/mjt.0000000000001377](https://doi.org/10.1097/mjt.0000000000001377) · PMID: [34375047](https://pubmed.ncbi.nlm.nih.gov/34375047/) · PMCID: [PMC8088823](https://pubmed.ncbi.nlm.nih.gov/PMC8088823/)

833. **The association between the use of ivermectin and mortality in patients with COVID-19: a meta-analysis**

Chia Siang Kow, Hamid A Merchant, Zia Ul Mustafa, Syed Shahzad Hasan

Pharmacological Reports (2021-03-29) <https://doi.org/gmhrpr>

DOI: [10.1007/s43440-021-00245-z](https://doi.org/10.1007/s43440-021-00245-z) · PMID: [33779964](https://pubmed.ncbi.nlm.nih.gov/33779964/) · PMCID: [PMC8005369](https://pubmed.ncbi.nlm.nih.gov/PMC8005369/)

834. **An Updated Systematic Review and Meta-Analysis of Mortality, Need for ICU admission, Use of Mechanical Ventilation, Adverse effects and other Clinical Outcomes of Ivermectin Treatment in COVID-19 Patients**

Smruti Karale, Vikas Bansal, Janaki Makadia, Muhammad Tayyeb, Hira Khan, Shree Spandana Ghanta, Romil Singh, Aysun Tekin, Abhishek

Bhurwal, Hemant Mutneja, ... Rahul Kashyap
Cold Spring Harbor Laboratory (2021-05-04) <https://doi.org/gmhmdn>
DOI: [10.1101/2021.04.30.21256415](https://doi.org/10.1101/2021.04.30.21256415)

835. <https://gidmk.medium.com/does-ivermectin-work-for-covid-19-1166126c364a>
836. **Retracted: Meta-analysis of Randomized Trials of Ivermectin to Treat SARS-CoV-2 Infection**
Andrew Hill, Anna Garratt, Jacob Levi, Jonathan Falconer, Leah Ellis, Kaitlyn McCann, Victoria Pilkington, Ambar Qavi, Junzheng Wang, Hannah Wentzel
Open Forum Infectious Diseases (2021-07-06) <https://doi.org/gmh4jn>
DOI: [10.1093/ofid/ofab358](https://doi.org/10.1093/ofid/ofab358) · PMID: [34796244](#) · PMCID: [PMC8420640](#)
837. **FAQ: COVID-19 and Ivermectin Intended for Animals**
Center for Veterinary Medicine
FDA (2022-02-11) <https://www.fda.gov/animal-veterinary/product-safety-information/faq-covid-19-and-ivermectin-intended-animals>
838. **A Multicenter, Prospective, Adaptive, Double-blind, Randomized, Placebo-controlled Study to Evaluate the Effect of Fluvoxamine, Ivermectin, Doxasozin and Interferon Lambda 1A in Mild COVID-19 and High Risk of Complications**
Cardresearch
clinicaltrials.gov (2022-01-15)
<https://clinicaltrials.gov/ct2/show/NCT04727424>
839. **Ivermectin to be investigated in adults aged 18+ as a possible treatment for COVID-19 in the PRINCIPLE trial — PRINCIPLE Trial**
<https://www.principletrial.org/news/ivermectin-to-be-investigated-as-a-possible-treatment-for-covid-19-in-oxford2019s-principle-trial>
840. **Effect of Early Treatment With Hydroxychloroquine or Lopinavir and Ritonavir on Risk of Hospitalization Among Patients With COVID-19**
Gilmar Reis, Eduardo Augusto dos Santos Moreira Silva, Daniela Carla Medeiros Silva, Lehana Thabane, Gurmit Singh, Jay JH Park, Jamie I Forrest, Ofir Harari, Castilho Vitor Quirino dos Santos, Ana Paula Figueiredo Guimarães de Almeida, ...
JAMA Network Open (2021-04-22) <https://doi.org/gk4298>
DOI: [10.1001/jamanetworkopen.2021.6468](https://doi.org/10.1001/jamanetworkopen.2021.6468) · PMID: [33885775](#) · PMCID: [PMC8063069](#)
841. **August 6, 2021: Early Treatment of COVID-19 with Repurposed Therapies: The TOGETHER Adaptive Platform Trial (Edward Mills, PhD, FRCP)**
Rethinking Clinical Trials
(2021-08-11) <https://rethinkingclinicaltrials.org/news/august-6-2021-early-treatment-of-covid-19-with-repurposed-therapies-the-together-adaptive-platform-trial-edward-mills-phd-frcp/>
842. **Lisinopril - Drug Usage Statistics**
ClinCalc DrugStats Database
<https://clincalc.com/DrugStats/Drugs/Lisinopril>

843. **Hypertension Hot Potato — Anatomy of the Angiotensin-Receptor Blocker Recalls**
JBrian Byrd, Glenn M Chertow, Vivek Bhalla
New England Journal of Medicine (2019-04-25) <https://doi.org/ggvc7g>
DOI: [10.1056/nejmp1901657](https://doi.org/10.1056/nejmp1901657) · PMID: [30865819](https://pubmed.ncbi.nlm.nih.gov/30865819/) · PMCID: [PMC7066505](https://pubmed.ncbi.nlm.nih.gov/PMC7066505/)
844. **ACE Inhibitor and ARB Utilization and Expenditures in the Medicaid Fee-For-Service Program from 1991 to 2008**
Boyang Bian, Christina ML Kelton, Jeff J Guo, Patricia R Wigle
Journal of Managed Care Pharmacy (2010-11) <https://doi.org/gh294c>
DOI: [10.18553/jmcp.2010.16.9.671](https://doi.org/10.18553/jmcp.2010.16.9.671) · PMID: [21067253](https://pubmed.ncbi.nlm.nih.gov/21067253/)
845. **ACE2: from vasopeptidase to SARS virus receptor**
Anthony J Turner, Julian A Hiscox, Nigel M Hooper
Trends in Pharmacological Sciences (2004-06) <https://doi.org/dn77dn>
DOI: [10.1016/j.tips.2004.04.001](https://doi.org/10.1016/j.tips.2004.04.001) · PMID: [15165741](https://pubmed.ncbi.nlm.nih.gov/15165741/) · PMCID: [PMC7119032](https://pubmed.ncbi.nlm.nih.gov/PMC7119032/)
846. **Structure-Based Discovery of a Novel Angiotensin-Converting Enzyme 2 Inhibitor**
Matthew J Huentelman, Jasenka Zubcevic, Jose A Hernández Prada, Xiaodong Xiao, Dimiter S Dimitrov, Mohan K Raizada, David A Ostrov
Hypertension (2004-12) <https://doi.org/d5szrp>
DOI: [10.1161/01.hyp.0000146120.29648.36](https://doi.org/10.1161/01.hyp.0000146120.29648.36) · PMID: [15492138](https://pubmed.ncbi.nlm.nih.gov/15492138/)
847. **The Secret Life of ACE2 as a Receptor for the SARS Virus**
Dimiter S Dimitrov
Cell (2003-12) <https://doi.org/d85vmw>
DOI: [10.1016/s0092-8674\(03\)00976-0](https://doi.org/10.1016/s0092-8674(03)00976-0) · PMID: [14675530](https://pubmed.ncbi.nlm.nih.gov/14675530/) · PMCID: [PMC7133233](https://pubmed.ncbi.nlm.nih.gov/PMC7133233/)
848. **Hypertension, the renin-angiotensin system, and the risk of lower respiratory tract infections and lung injury: implications for COVID-19**
Reinhold Kreutz, Engi Abd El-Hady Algharably, Michel Azizi, Piotr Dobrowolski, Tomasz Guzik, Andrzej Januszewicz, Alexandre Persu, Aleksander Prejbisz, Thomas Günther Riemer, Ji-Guang Wang, Michel Burnier
Cardiovascular Research (2020-08-01) <https://doi.org/ggtwpj>
DOI: [10.1093/cvr/cvaa097](https://doi.org/10.1093/cvr/cvaa097) · PMID: [32293003](https://pubmed.ncbi.nlm.nih.gov/32293003/) · PMCID: [PMC7184480](https://pubmed.ncbi.nlm.nih.gov/PMC7184480/)
849. **Angiotensin converting enzyme 2 activity and human atrial fibrillation: increased plasma angiotensin converting enzyme 2 activity is associated with atrial fibrillation and more advanced left atrial structural remodelling**
Tomos E Walters, Jonathan M Kalman, Sheila K Patel, Megan Mearns, Elena Velkoska, Louise M Burrell
Europace (2016-10-12) <https://doi.org/gbt2jw>
DOI: [10.1093/europace/euw246](https://doi.org/10.1093/europace/euw246) · PMID: [27738071](https://pubmed.ncbi.nlm.nih.gov/27738071/)
850. **Cardiovascular Disease, Drug Therapy, and Mortality in Covid-19**
Mandeep R Mehra, Sapan S Desai, SreyRam Kuy, Timothy D Henry, Amit N Patel
New England Journal of Medicine (2020-06-18) <https://doi.org/ggtp6v>
DOI: [10.1056/nejmoa2007621](https://doi.org/10.1056/nejmoa2007621) · PMID: [32356626](https://pubmed.ncbi.nlm.nih.gov/32356626/) · PMCID: [PMC7206931](https://pubmed.ncbi.nlm.nih.gov/PMC7206931/)

851. **Retraction: Cardiovascular Disease, Drug Therapy, and Mortality in Covid-19. N Engl J Med. DOI: 10.1056/NEJMoa2007621.**
Mandeep R Mehra, Sapan S Desai, SreyRam Kuy, Timothy D Henry, Amit N Patel
New England Journal of Medicine (2020-06-25) <https://doi.org/ggzkpj>
DOI: [10.1056/nejmc2021225](https://doi.org/10.1056/nejmc2021225) · PMID: [32501665](https://pubmed.ncbi.nlm.nih.gov/32501665/) · PMCID: [PMC7274164](https://pubmed.ncbi.nlm.nih.gov/PMC7274164/)
852. **Continuation versus discontinuation of renin–angiotensin system inhibitors in patients admitted to hospital with COVID-19: a prospective, randomised, open-label trial**
Jordana B Cohen, Thomas C Hanff, Preethi William, Nancy Sweitzer, Nelson R Rosado-Santander, Carola Medina, Juan E Rodriguez-Mori, Nicolás Renna, Tara I Chang, Vicente Corrales-Medina, ... Julio A Chirinos
The Lancet Respiratory Medicine (2021-03) <https://doi.org/fvgt>
DOI: [10.1016/s2213-2600\(20\)30558-0](https://doi.org/10.1016/s2213-2600(20)30558-0) · PMID: [33422263](https://pubmed.ncbi.nlm.nih.gov/33422263/) · PMCID: [PMC7832152](https://pubmed.ncbi.nlm.nih.gov/PMC7832152/)
853. **Effect of Discontinuing vs Continuing Angiotensin-Converting Enzyme Inhibitors and Angiotensin II Receptor Blockers on Days Alive and Out of the Hospital in Patients Admitted With COVID-19**
Renato D Lopes, Ariane VS Macedo, Pedro GM de Barros E Silva, Renata J Moll-Bernardes, Tiago M dos Santos, Lilian Mazza, André Feldman, Guilherme D'Andréa Saba Arruda, Denílson C de Albuquerque, Angelina S Camiletti, ...
JAMA (2021-01-19) <https://doi.org/gh2tw5>
DOI: [10.1001/jama.2020.25864](https://doi.org/10.1001/jama.2020.25864) · PMID: [33464336](https://pubmed.ncbi.nlm.nih.gov/33464336/) · PMCID: [PMC7816106](https://pubmed.ncbi.nlm.nih.gov/PMC7816106/)
854. **Frequently Asked Questions on the Revocation of the Emergency Use Authorization for Hydroxychloroquine Sulfate and Chloroquine Phosphate**
U.S. Food and Drug Administration
(2020-06-19) <https://www.fda.gov/media/138946/download>
855. **COVID-19: chloroquine and hydroxychloroquine only to be used in clinical trials or emergency use programmes**
EMA
European Medicines Agency (2020-04-01)
<https://www.ema.europa.eu/en/news/covid-19-chloroquine-hydroxychloroquine-only-be-used-clinical-trials-emergency-use-programmes>
856. **Formulation and manufacturability of biologics**
Steven J Shire
Current Opinion in Biotechnology (2009-12) <https://doi.org/cjk8p6>
DOI: [10.1016/j.copbio.2009.10.006](https://doi.org/10.1016/j.copbio.2009.10.006) · PMID: [19880308](https://pubmed.ncbi.nlm.nih.gov/19880308/)
857. **Early Development of Therapeutic Biologics - Pharmacokinetics**
A Baumann
Current Drug Metabolism (2006-01-01) <https://doi.org/bhc79>
DOI: [10.2174/138920006774832604](https://doi.org/10.2174/138920006774832604) · PMID: [16454690](https://pubmed.ncbi.nlm.nih.gov/16454690/)
858. **Development of therapeutic antibodies for the treatment of diseases**

Ruei-Min Lu, Yu-Chyi Hwang, I-Ju Liu, Chi-Chiu Lee, Han-Zen Tsai, Hsin-Jung Li, Han-Chung Wu
Journal of Biomedical Science (2020-01-02) <https://doi.org/ggqbp>
DOI: [10.1186/s12929-019-0592-z](https://doi.org/10.1186/s12929-019-0592-z) · PMID: [31894001](https://pubmed.ncbi.nlm.nih.gov/31894001/) · PMCID: [PMC6939334](https://pubmed.ncbi.nlm.nih.gov/PMC6939334/)

859. **Broadly Neutralizing Antiviral Antibodies**

Davide Corti, Antonio Lanzavecchia
Annual Review of Immunology (2013-03-21) <https://doi.org/gf25g8>
DOI: [10.1146/annurev-immunol-032712-095916](https://doi.org/10.1146/annurev-immunol-032712-095916) · PMID: [23330954](https://pubmed.ncbi.nlm.nih.gov/23330954/)

860. **Ibalizumab Targeting CD4 Receptors, An Emerging Molecule in HIV Therapy**

Simona A Iacob, Diana G Iacob
Frontiers in Microbiology (2017-11-27) <https://doi.org/gcn3kh>
DOI: [10.3389/fmicb.2017.02323](https://doi.org/10.3389/fmicb.2017.02323) · PMID: [29230203](https://pubmed.ncbi.nlm.nih.gov/29230203/) · PMCID: [PMC5711820](https://pubmed.ncbi.nlm.nih.gov/PMC5711820/)

861. **Product review on the monoclonal antibody palivizumab for prevention of respiratory syncytial virus infection**

Bernhard Resch
Human Vaccines & Immunotherapeutics (2017-06-12)
<https://doi.org/ggqbps>
DOI: [10.1080/21645515.2017.1337614](https://doi.org/10.1080/21645515.2017.1337614) · PMID: [28605249](https://pubmed.ncbi.nlm.nih.gov/28605249/) · PMCID: [PMC5612471](https://pubmed.ncbi.nlm.nih.gov/PMC5612471/)

862. **Prophylaxis With a Middle East Respiratory Syndrome Coronavirus (MERS-CoV)-Specific Human Monoclonal Antibody Protects Rabbits From MERS-CoV Infection**

Katherine V Houser, Lisa Gretebeck, Tianlei Ying, Yanping Wang, Leatrice Vogel, Elaine W Lamirande, Kevin W Bock, Ian N Moore, Dimiter S Dimitrov, Kanta Subbarao
Journal of Infectious Diseases (2016-03-03) <https://doi.org/f8pm7j>
DOI: [10.1093/infdis/jiw080](https://doi.org/10.1093/infdis/jiw080) · PMID: [26941283](https://pubmed.ncbi.nlm.nih.gov/26941283/) · PMCID: [PMC4837915](https://pubmed.ncbi.nlm.nih.gov/PMC4837915/)

863. **Efficacy of antibody-based therapies against Middle East respiratory syndrome coronavirus (MERS-CoV) in common marmosets**

Neeltje van Doremalen, Darryl Falzarano, Tianlei Ying, Emmie de Wit, Trenton Bushmaker, Friederike Feldmann, Atsushi Okumura, Yanping Wang, Dana P Scott, Patrick W Hanley, ... Vincent J Munster
Antiviral Research (2017-07) <https://doi.org/gbh5c2>
DOI: [10.1016/j.antiviral.2017.03.025](https://doi.org/10.1016/j.antiviral.2017.03.025) · PMID: [28389142](https://pubmed.ncbi.nlm.nih.gov/28389142/) · PMCID: [PMC6957253](https://pubmed.ncbi.nlm.nih.gov/PMC6957253/)

864. **IL-6 in Inflammation, Immunity, and Disease**

T Tanaka, M Narazaki, T Kishimoto
Cold Spring Harbor Perspectives in Biology (2014-09-04)
<https://doi.org/gftpj5>
DOI: [10.1101/cshperspect.a016295](https://doi.org/10.1101/cshperspect.a016295) · PMID: [25190079](https://pubmed.ncbi.nlm.nih.gov/25190079/) · PMCID: [PMC4176007](https://pubmed.ncbi.nlm.nih.gov/PMC4176007/)

865. **ACTEMRA - tocilizumab**

DailyMed

(2020-12-17) [https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?
setid=2e5365ff-cb2a-4b16-b2c7-e35c6bf2de13](https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?setid=2e5365ff-cb2a-4b16-b2c7-e35c6bf2de13)

866. **Tocilizumab in patients with severe COVID-19: a retrospective cohort study**

Giovanni Guaraldi, Marianna Meschiari, Alessandro Cozzi-Lepri, Jovana Milic, Roberto Tonelli, Marianna Menozzi, Erica Franceschini, Gianluca Cuomo, Gabriella Orlando, Vanni Borghi, ... Cristina Mussini

The Lancet Rheumatology (2020-08) <https://doi.org/d2pk>

DOI: [10.1016/s2665-9913\(20\)30173-9](https://doi.org/s2665-9913(20)30173-9) · PMID: [32835257](https://pubmed.ncbi.nlm.nih.gov/32835257/) · PMCID: [PMC7314456](https://pubmed.ncbi.nlm.nih.gov/PMC7314456/)

867. **Tocilizumab Treatment for Cytokine Release Syndrome in Hospitalized Patients With Coronavirus Disease 2019**

Christina C Price, Frederick L Altice, Yu Shyr, Alan Koff, Lauren Pischel, George Goshua, Marwan M Azar, Dayna Mcmanus, Sheau-Chiann Chen, Shana E Gleeson, ... Maricar Malinis

Chest (2020-10) <https://doi.org/gg2789>

DOI: [10.1016/j.chest.2020.06.006](https://doi.org/j.chest.2020.06.006) · PMID: [32553536](https://pubmed.ncbi.nlm.nih.gov/32553536/) · PMCID: [PMC7831876](https://pubmed.ncbi.nlm.nih.gov/PMC7831876/)

868. **Impact of low dose tocilizumab on mortality rate in patients with COVID-19 related pneumonia**

Ruggero Capra, Nicola De Rossi, Flavia Mattioli, Giuseppe Romanelli, Cristina Scarpazza, Maria Pia Sormani, Stefania Cossi

European Journal of Internal Medicine (2020-06)

<https://doi.org/ggx4fm>

DOI: [10.1016/j.ejim.2020.05.009](https://doi.org/j.ejim.2020.05.009) · PMID: [32405160](https://pubmed.ncbi.nlm.nih.gov/32405160/) · PMCID: [PMC7219361](https://pubmed.ncbi.nlm.nih.gov/PMC7219361/)

869. **Tocilizumab therapy reduced intensive care unit admissions and/or mortality in COVID-19 patients**

T Klopfenstein, S Zayet, A Lohse, J-C Balblanc, J Badie, P-Y Royer, L Toko, C Mezher, NJ Kadiane-Oussou, M Bossert, ... T Conrozier

Médecine et Maladies Infectieuses (2020-08) <https://doi.org/ggvz45>

DOI: [10.1016/j.medmal.2020.05.001](https://doi.org/j.medmal.2020.05.001) · PMID: [32387320](https://pubmed.ncbi.nlm.nih.gov/32387320/) · PMCID: [PMC7202806](https://pubmed.ncbi.nlm.nih.gov/PMC7202806/)

870. **Outcomes in patients with severe COVID-19 disease treated with tocilizumab: a case-controlled study**

G Rojas-Marte, M Khalid, O Mukhtar, AT Hashmi, MA Waheed, S Ehrlich, A Aslam, S Siddiqui, C Agarwal, Y Malyshev, ... J Shani

QJM: An International Journal of Medicine (2020-08)

<https://doi.org/gg496t>

DOI: [10.1093/qjmed/hcaa206](https://doi.org/10.1093/qjmed/hcaa206) · PMID: [32569363](https://pubmed.ncbi.nlm.nih.gov/32569363/) · PMCID: [PMC7337835](https://pubmed.ncbi.nlm.nih.gov/PMC7337835/)

871. **Effective treatment of severe COVID-19 patients with tocilizumab**

Xiaoling Xu, Mingfeng Han, Tiantian Li, Wei Sun, Dongsheng Wang, Binqing Fu, Yonggang Zhou, Xiaohu Zheng, Yun Yang, Xiuyong Li, ... Haiming Wei

Proceedings of the National Academy of Sciences (2020-05-19)

<https://doi.org/ggy3r3>

DOI: [10.1073/pnas.2005615117](https://doi.org/10.1073/pnas.2005615117) · PMID: [32350134](https://pubmed.ncbi.nlm.nih.gov/32350134/) · PMCID: [PMC7245089](https://pubmed.ncbi.nlm.nih.gov/PMC7245089/)

872. **Tocilizumab in patients admitted to hospital with COVID-19 (RECOVERY): preliminary results of a randomised, controlled, open-label, platform trial**
, Peter W Horby, Guilherme Pessoa-Amorim, Leon Peto, Christopher E Brightling, Rahuldeb Sarkar, Koshy Thomas, Vandana Jeebun, Abdul Ashish, Redmond Tully, ... Martin J Landray
Cold Spring Harbor Laboratory (2021-02-11) <https://doi.org/fvqj>
DOI: [10.1101/2021.02.11.21249258](https://doi.org/10.1101/2021.02.11.21249258)
873. **Tocilizumab in Hospitalized Patients with Severe Covid-19 Pneumonia**
Ivan O Rosas, Norbert Bräu, Michael Waters, Ronaldo C Go, Bradley D Hunter, Sanjay Bhagani, Daniel Skiest, Mariam S Aziz, Nichola Cooper, Ivor S Douglas, ... Atul Malhotra
New England Journal of Medicine (2021-04-22) <https://doi.org/gh5vk5>
DOI: [10.1056/nejmoa2028700](https://doi.org/10.1056/nejmoa2028700) · PMID: [33631066](#) · PMCID: [PMC7953459](#)
874. **Tocilizumab in Patients Hospitalized with Covid-19 Pneumonia**
Carlos Salama, Jian Han, Linda Yau, William G Reiss, Benjamin Kramer, Jeffrey D Neidhart, Gerard J Criner, Emma Kaplan-Lewis, Rachel Baden, Lavannya Pandit, ... Shalini V Mohan
New England Journal of Medicine (2021-01-07) <https://doi.org/ghp8xc>
DOI: [10.1056/nejmoa2030340](https://doi.org/10.1056/nejmoa2030340) · PMID: [33332779](#) · PMCID: [PMC7781101](#)
875. <https://clinicaltrials.gov/ct2/show/NCT04409262>
876. **A Randomized, Double-Blind, Placebo-Controlled, Multicenter Study to Evaluate the Safety and Efficacy of Tocilizumab in Patients With Severe COVID-19 Pneumonia**
Hoffmann-La Roche
clinicaltrials.gov (2021-06-28)
<https://clinicaltrials.gov/ct2/show/NCT04320615>
877. **A Randomized, Double-Blind, Placebo-Controlled, Multicenter Study to Evaluate the Efficacy and Safety of Tocilizumab in Hospitalized Patients With COVID-19 Pneumonia**
Genentech, Inc.
clinicaltrials.gov (2021-09-23)
<https://clinicaltrials.gov/ct2/show/NCT04372186>
878. **Genentech tocilizumab Letter of Authority**
U.S. Food and Drug Administration
(2021-06-24) <https://www.fda.gov/media/150319/download>
879. **A Randomised Double-blind Placebo-controlled Trial to Determine the Safety and Efficacy of Inhaled SNG001 (IFN- β 1a for Nebulisation) for the Treatment of Patients With Confirmed SARS-CoV-2 Infection**
Synairgen Research Ltd.
clinicaltrials.gov (2021-03-19)
<https://clinicaltrials.gov/ct2/show/NCT04385095>
880. **Synairgen announces positive results from trial of SNG001 in hospitalised COVID-19 patients**
Synairgen plc press release

(2020-07-20)

<http://synairgen.web01.hosting.bdci.co.uk/umbraco/Surface/DownloadFile?cid=1130026e-0983-4338-b648-4ac7928b9a37>

881. **Safety and efficacy of inhaled nebulised interferon beta-1a (SNG001) for treatment of SARS-CoV-2 infection: a randomised, double-blind, placebo-controlled, phase 2 trial**

Phillip D Monk, Richard J Marsden, Victoria J Tear, Jody Brookes, Toby N Batten, Marcin Mankowski, Felicity J Gabbay, Donna E Davies, Stephen T Holgate, Ling-Pei Ho, ... Pedro MB Rodrigues

The Lancet Respiratory Medicine (2021-02) <https://doi.org/ghjzm4>

DOI: [10.1016/s2213-2600\(20\)30511-7](https://doi.org/10.1016/s2213-2600(20)30511-7) · PMID: [33189161](https://pubmed.ncbi.nlm.nih.gov/33189161/) · PMCID: [PMC7836724](https://pubmed.ncbi.nlm.nih.gov/PMC7836724/)

882. **Convalescent plasma as a potential therapy for COVID-19**

Long Chen, Jing Xiong, Lei Bao, Yuan Shi

The Lancet Infectious Diseases (2020-04) <https://doi.org/ggqr7s>

DOI: [10.1016/s1473-3099\(20\)30141-9](https://doi.org/10.1016/s1473-3099(20)30141-9) · PMID: [32113510](https://pubmed.ncbi.nlm.nih.gov/32113510/) · PMCID: [PMC7128218](https://pubmed.ncbi.nlm.nih.gov/PMC7128218/)

883. **Convalescent Plasma to Treat COVID-19**

John D Roback, Jeannette Guarner

JAMA (2020-04-28) <https://doi.org/ggqf6k>

DOI: [10.1001/jama.2020.4940](https://doi.org/10.1001/jama.2020.4940) · PMID: [32219429](https://pubmed.ncbi.nlm.nih.gov/32219429/)

884. **Convalescent plasma transfusion for the treatment of COVID-19: Systematic review**

Karthick Rajendran, Narayanasamy Krishnasamy, Jayanthi Rangarajan, Jeyalalitha Rathinam, Murugan Natarajan, Arunkumar Ramachandran

Journal of Medical Virology (2020-05-12) <https://doi.org/ggv3gx>

DOI: [10.1002/jmv.25961](https://doi.org/10.1002/jmv.25961) · PMID: [32356910](https://pubmed.ncbi.nlm.nih.gov/32356910/) · PMCID: [PMC7267113](https://pubmed.ncbi.nlm.nih.gov/PMC7267113/)

885. **Association of Convalescent Plasma Treatment With Clinical Outcomes in Patients With COVID-19**

Perrine Janiaud, Cathrine Axfors, Andreas M Schmitt, Viktoria Gloy, Fahim Ebrahimi, Matthias Hepprich, Emily R Smith, Noah A Haber, Nina Khanna, David Moher, ... Lars G Hemkens

JAMA (2021-03-23) <https://doi.org/gjjk4j>

DOI: [10.1001/jama.2021.2747](https://doi.org/10.1001/jama.2021.2747) · PMID: [33635310](https://pubmed.ncbi.nlm.nih.gov/33635310/) · PMCID: [PMC7911095](https://pubmed.ncbi.nlm.nih.gov/PMC7911095/)

886. **Convalescent Plasma Antibody Levels and the Risk of Death from Covid-19**

Michael J Joyner, Rickey E Carter, Jonathon W Senefeld, Stephen A Klassen, John R Mills, Patrick W Johnson, Elitza S Theel, Chad C Wiggins, Katelyn A Bruno, Allan M Klompas, ... Arturo Casadevall

New England Journal of Medicine (2021-01-13) <https://doi.org/ghs26g>

DOI: [10.1056/nejmoa2031893](https://doi.org/10.1056/nejmoa2031893) · PMID: [33523609](https://pubmed.ncbi.nlm.nih.gov/33523609/) · PMCID: [PMC7821984](https://pubmed.ncbi.nlm.nih.gov/PMC7821984/)

887. **Convalescent plasma in patients admitted to hospital with COVID-19 (RECOVERY): a randomised, controlled, open-label, platform trial**

, Peter W Horby, Lise Estcourt, Leon Peto, Jonathan R Emberson, Natalie Staplin, Enti Spata, Guilherme Pessoa-Amorim, Mark Campbell, Alistair Roddick, ... Martin J Landray

Cold Spring Harbor Laboratory (2021-03-10) <https://doi.org/gmcq2g>

888. **Chronological evolution of IgM, IgA, IgG and neutralisation antibodies after infection with SARS-associated coronavirus**
P-R Hsueh, L-M Huang, P-J Chen, C-L Kao, P-C Yang
Clinical Microbiology and Infection (2004-12) <https://doi.org/cwwg87>
DOI: [10.1111/j.1469-0691.2004.01009.x](https://doi.org/10.1111/j.1469-0691.2004.01009.x) · PMID: [15606632](#)
889. **Neutralizing Antibodies in Patients with Severe Acute Respiratory Syndrome-Associated Coronavirus Infection**
Nie Yuchun, Wang Guangwen, Shi Xuanling, Zhang Hong, Qiu Yan, He Zhongping, Wang Wei, Lian Gewei, Yin Xiaolei, Du Liying, ... Ding Mingxiao
The Journal of Infectious Diseases (2004-09) <https://doi.org/cgqj5b>
DOI: [10.1086/423286](https://doi.org/10.1086/423286) · PMID: [15319862](#)
890. **Potent human monoclonal antibodies against SARS CoV, Nipah and Hendra viruses**
Ponraj Prabakaran, Zhongyu Zhu, Xiaodong Xiao, Arya Biragyn, Antony S Dimitrov, Christopher C Broder, Dimiter S Dimitrov
Expert Opinion on Biological Therapy (2009-04-08)
<https://doi.org/b88kw8>
DOI: [10.1517/14712590902763755](https://doi.org/10.1517/14712590902763755) · PMID: [19216624](#) · PMCID: [PMC2705284](#)
891. **SARS-CoV-2 and SARS-CoV Spike-RBD Structure and Receptor Binding Comparison and Potential Implications on Neutralizing Antibody and Vaccine Development**
Chunyun Sun, Long Chen, Ji Yang, Chunxia Luo, Yanjing Zhang, Jing Li, Jiahui Yang, Jie Zhang, Liangzhi Xie
Cold Spring Harbor Laboratory (2020-02-20) <https://doi.org/ggq63j>
DOI: [10.1101/2020.02.16.951723](https://doi.org/10.1101/2020.02.16.951723)
892. **Fruitful Neutralizing Antibody Pipeline Brings Hope To Defeat SARS-CoV-2**
Alex Renn, Ying Fu, Xin Hu, Matthew D Hall, Anton Simeonov
Trends in Pharmacological Sciences (2020-11) <https://doi.org/gg72sv>
DOI: [10.1016/j.tips.2020.07.004](https://doi.org/10.1016/j.tips.2020.07.004) · PMID: [32829936](#) · PMCID: [PMC7572790](#)
893. **Cross-neutralization of SARS-CoV-2 by a human monoclonal SARS-CoV antibody**
Dora Pinto, Young-Jun Park, Martina Beltramello, Alexandra C Walls, MAlejandra Tortorici, Siro Bianchi, Stefano Jaconi, Katja Culap, Fabrizia Zatta, Anna De Marco, ... Davide Corti
Nature (2020-05-18) <https://doi.org/dv4x>
DOI: [10.1038/s41586-020-2349-y](https://doi.org/10.1038/s41586-020-2349-y) · PMID: [32422645](#)
894. **A human monoclonal antibody blocking SARS-CoV-2 infection**
Chunyan Wang, Wentao Li, Dubravka Drabek, Nisreen MA Okba, Rien van Haperen, Albert DME Osterhaus, Frank JM van Kuppeveld, Bart L Haagmans, Frank Grosveld, Berend-Jan Bosch
Cold Spring Harbor Laboratory (2020-03-12) <https://doi.org/ggnw4t>
DOI: [10.1101/2020.03.11.987958](https://doi.org/10.1101/2020.03.11.987958)

895. **An update to monoclonal antibody as therapeutic option against COVID-19**

Paroma Deb, MdMaruf Ahmed Molla, KM Saif-Ur-Rahman

Biosafety and Health (2021-04) <https://doi.org/gh4m7h>

DOI: [10.1016/j.bsheal.2021.02.001](https://doi.org/10.1016/j.bsheal.2021.02.001) · PMID: [33585808](#) · PMCID: [PMC7872849](#)

896. **LY-CoV555, a rapidly isolated potent neutralizing antibody, provides protection in a non-human primate model of SARS-CoV-2 infection**

Bryan E Jones, Patricia L Brown-Augsburger, Kizzmekia S Corbett, Kathryn Westendorf, Julian Davies, Thomas P Cujec, Christopher M Wiethoff, Jamie L Blackbourne, Beverly A Heinz, Denisa Foster, ... Ester Falconer

Cold Spring Harbor Laboratory (2020-10-01) <https://doi.org/gh4sjm>

DOI: [10.1101/2020.09.30.318972](https://doi.org/10.1101/2020.09.30.318972) · PMID: [33024963](#) · PMCID: [PMC7536866](#)

897. **A Randomized, Placebo-Controlled, Double-Blind, Sponsor Unblinded, Single Ascending Dose, Phase 1 First in Human Study to Evaluate the Safety, Tolerability, Pharmacokinetics and Pharmacodynamics of Intravenous LY3819253 in Participants Hospitalized for COVID-19**

Eli Lilly and Company

clinicaltrials.gov (2021-11-10)

<https://clinicaltrials.gov/ct2/show/NCT04411628>

898. **A Phase 1, Randomized, Placebo-Controlled Study to Evaluate the Tolerability, Safety, Pharmacokinetics, and Immunogenicity of LY3832479 Given as a Single Intravenous Dose in Healthy Participants**

Eli Lilly and Company

clinicaltrials.gov (2021-11-01)

<https://clinicaltrials.gov/ct2/show/NCT04441931>

899. **Effect of Bamlanivimab as Monotherapy or in Combination With Etesevimab on Viral Load in Patients With Mild to Moderate COVID-19**

Robert L Gottlieb, Ajay Nirula, Peter Chen, Joseph Boscia, Barry Heller, Jason Morris, Gregory Huhn, Jose Cardona, Bharat Mocherla, Valentina Stosor, ... Daniel M Skovronsky

JAMA (2021-02-16) <https://doi.org/ghvnrr>

DOI: [10.1001/jama.2021.0202](https://doi.org/10.1001/jama.2021.0202) · PMID: [33475701](#) · PMCID: [PMC7821080](#)

900. **A Randomized, Double-blind, Placebo-Controlled, Phase 2/3 Study to Evaluate the Efficacy and Safety of LY3819253 and LY3832479 in Participants With Mild to Moderate COVID-19 Illness**

Eli Lilly and Company

clinicaltrials.gov (2022-02-14)

<https://clinicaltrials.gov/ct2/show/NCT04427501>

901. **SARS-CoV-2 Neutralizing Antibody LY-CoV555 in Outpatients with Covid-19**

Peter Chen, Ajay Nirula, Barry Heller, Robert L Gottlieb, Joseph Boscia, Jason Morris, Gregory Huhn, Jose Cardona, Bharat Mocherla, Valentina

Stosor, ... Daniel M Skovronsky
New England Journal of Medicine (2021-01-21) <https://doi.org/fgtm>
DOI: [10.1056/nejmoa2029849](https://doi.org/10.1056/nejmoa2029849) · PMID: [33113295](#) · PMCID: [PMC7646625](#)

902. **Bamlanivimab and Etesevimab EUA Letter of Authorization**
U.S. Food and Drug Administration
(2021-02-25) <https://www.fda.gov/media/145801/download>
903. **Studies in humanized mice and convalescent humans yield a SARS-CoV-2 antibody cocktail**
Johanna Hansen, Alina Baum, Kristen E Pascal, Vincenzo Russo, Stephanie Giordano, Elzbieta Wloga, Benjamin O Fulton, Ying Yan, Katrina Koon, Krunal Patel, ... Christos A Kyratsous
Science (2020-08-21) <https://doi.org/fcqh>
DOI: [10.1126/science.abd0827](https://doi.org/10.1126/science.abd0827) · PMID: [32540901](#) · PMCID: [PMC7299284](#)
904. **A Master Protocol Assessing the Safety, Tolerability, and Efficacy of Anti-Spike (S) SARS-CoV-2 Monoclonal Antibodies for the Treatment of Hospitalized Patients With COVID-19**
Regeneron Pharmaceuticals
clinicaltrials.gov (2021-11-29)
<https://clinicaltrials.gov/ct2/show/NCT04426695>
905. **A Master Protocol Assessing the Safety, Tolerability, and Efficacy of Anti-Spike (S) SARS-CoV-2 Monoclonal Antibodies for the Treatment of Ambulatory Patients With COVID-19**
Regeneron Pharmaceuticals
clinicaltrials.gov (2022-02-10)
<https://clinicaltrials.gov/ct2/show/NCT04425629>
906. **REGN-COV2, a Neutralizing Antibody Cocktail, in Outpatients with Covid-19**
David M Weinreich, Sumathi Sivapalasingam, Thomas Norton, Shazia Ali, Haitao Gao, Rafia Bhore, Bret J Musser, Yuhwen Soo, Diana Rofail, Joseph Im, ... George D Yancopoulos
New England Journal of Medicine (2021-01-21) <https://doi.org/gh4sjh>
DOI: [10.1056/nejmoa2035002](https://doi.org/10.1056/nejmoa2035002) · PMID: [33332778](#) · PMCID: [PMC7781102](#)
907. **Coronavirus (COVID-19) Update: FDA Authorizes Monoclonal Antibodies for Treatment of COVID-19**
Office of the Commissioner
FDA (2020-11-23) <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-monoclonal-antibodies-treatment-covid-19>
908. **An efficient method to make human monoclonal antibodies from memory B cells: potent neutralization of SARS coronavirus**
Elisabetta Traggiai, Stephan Becker, Kanta Subbarao, Larissa Kolesnikova, Yasushi Uematsu, Maria Rita Gismondo, Brian R Murphy, Rino Rappuoli, Antonio Lanzavecchia
Nature Medicine (2004-07-11) <https://doi.org/b9867c>
DOI: [10.1038/nm1080](https://doi.org/10.1038/nm1080) · PMID: [15247913](#) · PMCID: [PMC7095806](#)

909. **'Super-antibodies' could curb COVID-19 and help avert future pandemics**
Elie Dolgin
Nature Biotechnology (2021-06-22) <https://doi.org/gmg2fx>
DOI: [10.1038/s41587-021-00980-x](https://doi.org/10.1038/s41587-021-00980-x) · PMID: [34158667](#) · PMCID: [PMC8218965](#)
910. **Passive immunotherapy of viral infections: 'super-antibodies' enter the fray**
Laura M Walker, Dennis R Burton
Nature Reviews Immunology (2018-01-30) <https://doi.org/gcwgp>
DOI: [10.1038/nri.2017.148](https://doi.org/10.1038/nri.2017.148) · PMID: [29379211](#) · PMCID: [PMC5918154](#)
911. **A Randomized, Multi-center, Double-blind, Placebo-controlled Study to Assess the Safety and Efficacy of Monoclonal Antibody VIR-7831 for the Early Treatment of Coronavirus Disease 2019 (COVID-19) in Non-hospitalized Patients**
Vir Biotechnology, Inc.
clinicaltrials.gov (2021-09-24)
<https://clinicaltrials.gov/ct2/show/NCT04545060>
912. **Early Covid-19 Treatment With SARS-CoV-2 Neutralizing Antibody Sotrovimab**
Anil Gupta, Yaneicy Gonzalez-Rojas, Erick Juarez, Manuel Crespo Casal, Jaynier Moya, Diego Rodrigues Falci, Elias Sarkis, Joel Solis, Hanzhe Zheng, Nicola Scott, ...
Cold Spring Harbor Laboratory (2021-05-28) <https://doi.org/gmg2fz>
DOI: [10.1101/2021.05.27.21257096](https://doi.org/10.1101/2021.05.27.21257096)
913. **Identification of SARS-CoV-2 spike mutations that attenuate monoclonal and serum antibody neutralization**
Zhuoming Liu, Laura A VanBlargan, Louis-Marie Bloyet, Paul W Rothlauf, Rita E Chen, Spencer Stumpf, Haiyan Zhao, John M Errico, Elitza S Theel, Mariel J Liebeskind, ... Sean PJ Whelan
Cell Host & Microbe (2021-03) <https://doi.org/gh4m7j>
DOI: [10.1016/j.chom.2021.01.014](https://doi.org/10.1016/j.chom.2021.01.014) · PMID: [33535027](#) · PMCID: [PMC7839837](#)
914. **SARS-CoV-2 variants show resistance to neutralization by many monoclonal and serum-derived polyclonal antibodies**
Michael Diamond, Rita Chen, Xuping Xie, James Case, Xianwen Zhang, Laura VanBlargan, Yang Liu, Jianying Liu, John Errico, Emma Winkler, ... Pavlo Gilchuk
Research Square Platform LLC (2021-02-10) <https://doi.org/gh4sjz>
DOI: [10.21203/rs.3.rs-228079/v1](https://doi.org/10.21203/rs.3.rs-228079/v1) · PMID: [33594356](#) · PMCID: [PMC7885928](#)
915. **Impact of the B.1.1.7 variant on neutralizing monoclonal antibodies recognizing diverse epitopes on SARS-CoV-2 Spike**
Carl Graham, Jeffrey Seow, Isabella Huettner, Hataf Khan, Neophytos Kouphou, Sam Acors, Helena Winstone, Suzanne Pickering, Rui Pedro Galao, Maria Jose Lista, ... Katie J Doores
Cold Spring Harbor Laboratory (2021-02-03) <https://doi.org/gh4sjq>
DOI: [10.1101/2021.02.03.429355](https://doi.org/10.1101/2021.02.03.429355) · PMID: [33564766](#) · PMCID: [PMC7872354](#)

916. **Antibody Resistance of SARS-CoV-2 Variants B.1.351 and B.1.1.7**
Pengfei Wang, Manoj S Nair, Lihong Liu, Sho Iketani, Yang Luo, Yicheng Guo, Maple Wang, Jian Yu, Baoshan Zhang, Peter D Kwong, ... David D Ho
Cold Spring Harbor Laboratory (2021-01-26) <https://doi.org/gh4sjp>
DOI: [10.1101/2021.01.25.428137](https://doi.org/10.1101/2021.01.25.428137) · PMID: [33532778](https://pubmed.ncbi.nlm.nih.gov/33532778/) · PMCID: [PMC7852271](https://pubmed.ncbi.nlm.nih.gov/PMC7852271/)
917. **Pause in the Distribution of bamlanivimab/etesevimab**
U.S. Department of Health and Human Services
(2021-06-25)
<https://www.phe.gov/emergency/events/COVID19/investigation-MCM/Bamlanivimab-etesevimab/Pages/bamlanivimab-etesevimab-distribution-pause.aspx>
918. **HIV-1 Broadly Neutralizing Antibody Extracts Its Epitope from a Kinked gp41 Ectodomain Region on the Viral Membrane**
Zhen-Yu J Sun, Kyoung Joon Oh, Mikyung Kim, Jessica Yu, Vladimir Brusic, Likai Song, Zhisong Qiao, Jia-huai Wang, Gerhard Wagner, Ellis L Reinherz
Immunity (2008-01) <https://doi.org/ftw7t3>
DOI: [10.1016/j.immuni.2007.11.018](https://doi.org/10.1016/j.immuni.2007.11.018) · PMID: [18191596](https://pubmed.ncbi.nlm.nih.gov/18191596/)
919. **Antibody Recognition of a Highly Conserved Influenza Virus Epitope**
Damian C Ekiert, Gira Bhabha, Marc-André Elsliger, Robert HE Friesen, Mandy Jongeneelen, Mark Throsby, Jaap Goudsmit, Ian A Wilson
Science (2009-04-10) <https://doi.org/ffsb4r>
DOI: [10.1126/science.1171491](https://doi.org/10.1126/science.1171491) · PMID: [19251591](https://pubmed.ncbi.nlm.nih.gov/19251591/) · PMCID: [PMC2758658](https://pubmed.ncbi.nlm.nih.gov/PMC2758658/)
920. **Identification and characterization of novel neutralizing epitopes in the receptor-binding domain of SARS-CoV spike protein: Revealing the critical antigenic determinants in inactivated SARS-CoV vaccine**
Yuxian He, Jingjing Li, Lanying Du, Xuxia Yan, Guangan Hu, Yusen Zhou, Shibo Jiang
Vaccine (2006-06) <https://doi.org/b99b68>
DOI: [10.1016/j.vaccine.2006.04.054](https://doi.org/10.1016/j.vaccine.2006.04.054) · PMID: [16725238](https://pubmed.ncbi.nlm.nih.gov/16725238/) · PMCID: [PMC7115380](https://pubmed.ncbi.nlm.nih.gov/PMC7115380/)
921. **Escape from Human Monoclonal Antibody Neutralization Affects In Vitro and In Vivo Fitness of Severe Acute Respiratory Syndrome Coronavirus**
Barry Rockx, Eric Donaldson, Matthew Frieman, Timothy Sheahan, Davide Corti, Antonio Lanzavecchia, Ralph S Baric
The Journal of Infectious Diseases (2010-03-15) <https://doi.org/cdgqjd>
DOI: [10.1086/651022](https://doi.org/10.1086/651022) · PMID: [20144042](https://pubmed.ncbi.nlm.nih.gov/20144042/) · PMCID: [PMC2826557](https://pubmed.ncbi.nlm.nih.gov/PMC2826557/)
922. **Stanford Coronavirus Antiviral & Resistance Database (CoVDB)**
<https://covdb.stanford.edu/page/susceptibility-data>
923. **Broad and potent activity against SARS-like viruses by an engineered human monoclonal antibody**
CGarrett Rappazzo, Longping V Tse, Chengzi I Kaku, Daniel Wrapp, Mrunal Sakharkar, Deli Huang, Laura M Deveau, Thomas J

Yockachonis, Andrew S Herbert, Michael B Battles, ... Laura M Walker
Science (2021-02-19) <https://doi.org/fsbc>
DOI: [10.1126/science.abf4830](https://doi.org/10.1126/science.abf4830) · PMID: [33495307](https://pubmed.ncbi.nlm.nih.gov/33495307/) · PMCID: [PMC7963221](https://pubmed.ncbi.nlm.nih.gov/PMC7963221/)

924. **Drug repurposing: a promising tool to accelerate the drug discovery process**

Vineela Parvathaneni, Nishant S Kulkarni, Aaron Muth, Vivek Gupta
Drug Discovery Today (2019-10) <https://doi.org/gj3v46>
DOI: [10.1016/j.drudis.2019.06.014](https://doi.org/10.1016/j.drudis.2019.06.014) · PMID: [31238113](https://pubmed.ncbi.nlm.nih.gov/31238113/)

925. **Drug repurposing screens and synergistic drug-combinations for infectious diseases**

Wei Zheng, Wei Sun, Anton Simeonov
British Journal of Pharmacology (2017-07-09) <https://doi.org/gj3v6j>
DOI: [10.1111/bph.13895](https://doi.org/10.1111/bph.13895) · PMID: [28685814](https://pubmed.ncbi.nlm.nih.gov/28685814/) · PMCID: [PMC5758396](https://pubmed.ncbi.nlm.nih.gov/PMC5758396/)

926. **A critical overview of computational approaches employed for COVID-19 drug discovery**

Eugene N Muratov, Rommie Amaro, Carolina H Andrade, Nathan Brown, Sean Ekins, Denis Fourches, Olexandr Isayev, Dima Kozakov, José L Medina-Franco, Kenneth M Merz, ... Alexander Tropsha
Chemical Society Reviews (2021) <https://doi.org/gmg9nm>
DOI: [10.1039/d0cs01065k](https://doi.org/10.1039/d0cs01065k) · PMID: [34212944](https://pubmed.ncbi.nlm.nih.gov/34212944/) · PMCID: [PMC8371861](https://pubmed.ncbi.nlm.nih.gov/PMC8371861/)

927. **Drug repurposing: a better approach for infectious disease drug discovery?**

GLynn Law, Jennifer Tisoncik-Go, Marcus J Korth, Michael G Katze
Current Opinion in Immunology (2013-10) <https://doi.org/f5jvrt>
DOI: [10.1016/j.coи.2013.08.004](https://doi.org/10.1016/j.coи.2013.08.004) · PMID: [24011665](https://pubmed.ncbi.nlm.nih.gov/24011665/) · PMCID: [PMC4015799](https://pubmed.ncbi.nlm.nih.gov/PMC4015799/)

928. **Origin and evolution of high throughput screening**

DA Pereira, JA Williams
British Journal of Pharmacology (2007-09) <https://doi.org/brs35w>
DOI: [10.1038/sj.bjp.0707373](https://doi.org/10.1038/sj.bjp.0707373) · PMID: [17603542](https://pubmed.ncbi.nlm.nih.gov/17603542/) · PMCID: [PMC1978279](https://pubmed.ncbi.nlm.nih.gov/PMC1978279/)

929. **Developing predictive assays: The phenotypic screening “rule of 3”**

Fabien Vincent, Paula Loria, Marko Pregel, Robert Stanton, Linda Kitching, Karl Nocka, Regis Doyonnas, Claire Steppan, Adam Gilbert, Thomas Schroeter, Marie-Claire Peakman
Science Translational Medicine (2015-06-24) <https://doi.org/ggp3tk>
DOI: [10.1126/scitranslmed.aab1201](https://doi.org/10.1126/scitranslmed.aab1201) · PMID: [26109101](https://pubmed.ncbi.nlm.nih.gov/26109101/)

930. **Phenotypic vs. Target-Based Drug Discovery for First-in-Class Medicines**

DC Swinney
Clinical Pharmacology & Therapeutics (2013-04) <https://doi.org/f4q6gz>
DOI: [10.1038/clpt.2012.236](https://doi.org/10.1038/clpt.2012.236) · PMID: [23511784](https://pubmed.ncbi.nlm.nih.gov/23511784/)

931. **Phenotypic screening in cancer drug discovery — past, present and future**

John G Moffat, Joachim Rudolph, David Bailey
Nature Reviews Drug Discovery (2014-07-18) <https://doi.org/f6cnfw>
DOI: [10.1038/nrd4366](https://doi.org/10.1038/nrd4366) · PMID: [25033736](https://pubmed.ncbi.nlm.nih.gov/25033736/)

932. **ChemBridge | Screening Libraries | Diversity Libraries | DIVERSet**
https://www.chembridge.com/screening_libraries/diversity_libraries/
933. **The Power of Sophisticated Phenotypic Screening and Modern Mechanism-of-Action Methods**
Bridget K Wagner, Stuart L Schreiber
Cell Chemical Biology (2016-01) <https://doi.org/gfsdbh>
DOI: [10.1016/j.chembiol.2015.11.008](https://doi.org/10.1016/j.chembiol.2015.11.008) · PMID: [26933731](https://pubmed.ncbi.nlm.nih.gov/26933731/) · PMCID: [PMC4779180](https://pubmed.ncbi.nlm.nih.gov/PMC4779180/)
934. **Drug Repurposing for Viral Infectious Diseases: How Far Are We?**
Beatrice Mercorelli, Giorgio Palù, Arianna Loregian
Trends in Microbiology (2018-10) <https://doi.org/gfbp3h>
DOI: [10.1016/j.tim.2018.04.004](https://doi.org/10.1016/j.tim.2018.04.004) · PMID: [29759926](https://pubmed.ncbi.nlm.nih.gov/29759926/) · PMCID: [PMC7126639](https://pubmed.ncbi.nlm.nih.gov/PMC7126639/)
935. **Systematically Prioritizing Candidates in Genome-Based Drug Repurposing**
Anup P Challa, Robert R Lavieri, Judith T Lewis, Nicole M Zaleski, Jana K Shirey-Rice, Paul A Harris, David M Aronoff, Jill M Pulley
ASSAY and Drug Development Technologies (2019-12-01)
<https://doi.org/gj3v6d>
DOI: [10.1089/adt.2019.950](https://doi.org/10.1089/adt.2019.950) · PMID: [31769998](https://pubmed.ncbi.nlm.nih.gov/31769998/) · PMCID: [PMC6921094](https://pubmed.ncbi.nlm.nih.gov/PMC6921094/)
936. **What Are the Odds of Finding a COVID-19 Drug from a Lab Repurposing Screen?**
Aled Edwards
Journal of Chemical Information and Modeling (2020-09-11)
<https://doi.org/gjkv79>
DOI: [10.1021/acs.jcim.0c00861](https://doi.org/10.1021/acs.jcim.0c00861) · PMID: [32914973](https://pubmed.ncbi.nlm.nih.gov/32914973/)
937. **Proteases Essential for Human Influenza Virus Entry into Cells and Their Inhibitors as Potential Therapeutic Agents**
Hiroshi Kido, Yuushi Okumura, Hiroshi Yamada, Trong Quang Le, Mihiro Yano
Current Pharmaceutical Design (2007-02-01) <https://doi.org/bts3xp>
DOI: [10.2174/138161207780162971](https://doi.org/10.2174/138161207780162971) · PMID: [17311557](https://pubmed.ncbi.nlm.nih.gov/17311557/)
938. **Protease inhibitors targeting coronavirus and filovirus entry**
Yanchen Zhou, Punitha Vedantham, Kai Lu, Juliet Agudelo, Ricardo Carrion, Jerritt W Nunneley, Dale Barnard, Stefan Pöhlmann, James H McKerrow, Adam R Renslo, Graham Simmons
Antiviral Research (2015-04) <https://doi.org/ggr984>
DOI: [10.1016/j.antiviral.2015.01.011](https://doi.org/10.1016/j.antiviral.2015.01.011) · PMID: [25666761](https://pubmed.ncbi.nlm.nih.gov/25666761/) · PMCID: [PMC4774534](https://pubmed.ncbi.nlm.nih.gov/PMC4774534/)
939. **Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors**
Zhenming Jin, Xiaoyu Du, Yechun Xu, Yongqiang Deng, Meiqin Liu, Yao Zhao, Bing Zhang, Xiaofeng Li, Leike Zhang, Chao Peng, ... Haitao Yang
Nature (2020-04-09) <https://doi.org/ggrp42>
DOI: [10.1038/s41586-020-2223-y](https://doi.org/10.1038/s41586-020-2223-y) · PMID: [32272481](https://pubmed.ncbi.nlm.nih.gov/32272481/)
940. **Design of Wide-Spectrum Inhibitors Targeting Coronavirus Main Proteases**

Haitao Yang, Weiqing Xie, Xiaoyu Xue, Kailin Yang, Jing Ma, Wenzhong Liang, Qi Zhao, Zhe Zhou, Duanqing Pei, John Ziebuhr, ... Zihe Rao
PLoS Biology (2005-09-06) <https://doi.org/bcm9k7>
DOI: [10.1371/journal.pbio.0030324](https://doi.org/10.1371/journal.pbio.0030324) · PMID: [16128623](#) · PMCID: [PMC1197287](#)

941. **The newly emerged SARS-Like coronavirus HCoV-EMC also has an “Achilles’ heel”: current effective inhibitor targeting a 3C-like protease**
Zhilin Ren, Liming Yan, Ning Zhang, Yu Guo, Cheng Yang, Zhiyong Lou, Zihe Rao
Protein & Cell (2013-04-03) <https://doi.org/ggr7vh>
DOI: [10.1007/s13238-013-2841-3](https://doi.org/10.1007/s13238-013-2841-3) · PMID: [23549610](#) · PMCID: [PMC4875521](#)
942. **Structure of Main Protease from Human Coronavirus NL63: Insights for Wide Spectrum Anti-Coronavirus Drug Design**
Fenghua Wang, Cheng Chen, Wenjie Tan, Kailin Yang, Haitao Yang
Scientific Reports (2016-03-07) <https://doi.org/f8cfx9>
DOI: [10.1038/srep22677](https://doi.org/10.1038/srep22677) · PMID: [26948040](#) · PMCID: [PMC4780191](#)
943. **Structures of Two Coronavirus Main Proteases: Implications for Substrate Binding and Antiviral Drug Design**
Xiaoyu Xue, Hongwei Yu, Haitao Yang, Fei Xue, Zhixin Wu, Wei Shen, Jun Li, Zhe Zhou, Yi Ding, Qi Zhao, ... Zihe Rao
Journal of Virology (2008-03) <https://doi.org/b2zbhv>
DOI: [10.1128/jvi.02114-07](https://doi.org/10.1128/jvi.02114-07) · PMID: [18094151](#) · PMCID: [PMC2258912](#)
944. **Ebselen, a promising antioxidant drug: mechanisms of action and targets of biological pathways**
Gajendra Kumar Azad, Raghuvir S Tomar
Molecular Biology Reports (2014-05-28) <https://doi.org/f6cnq3>
DOI: [10.1007/s11033-014-3417-x](https://doi.org/10.1007/s11033-014-3417-x) · PMID: [24867080](#)
945. **Molecular characterization of ebselen binding activity to SARS-CoV-2 main protease**
Cintia A Menéndez, Fabian Byléhn, Gustavo R Perez-Lemus, Walter Alvarado, Juan J de Pablo
Science Advances (2020-09-11) <https://doi.org/gmhshj>
DOI: [10.1126/sciadv.abd0345](https://doi.org/10.1126/sciadv.abd0345) · PMID: [32917717](#) · PMCID: [PMC7486088](#)
946. **Target discovery of ebselen with a biotinylated probe**
Zhenzhen Chen, Zhongyao Jiang, Nan Chen, Qian Shi, Lili Tong, Fanpeng Kong, Xiufen Cheng, Hao Chen, Chu Wang, Bo Tang
Chemical Communications (2018) <https://doi.org/ggrrtcm>
DOI: [10.1039/c8cc04258f](https://doi.org/10.1039/c8cc04258f) · PMID: [30091742](#)
947. **FDA Clears SPI’s Ebselen for Phase II COVID-19 Trials**
Contract Pharma
https://www.contractpharma.com/contents/view_breaking-news/2020-08-31/fda-clears-spis-ebselen-for-phase-ii-covid-19-trials/
948. **A Phase 2, Randomized, Double-Blind, Placebo-Controlled, Dose Escalation Study to Evaluate the Safety and Efficacy of SPI-1005 in Moderate COVID-19 Patients**

Sound Pharmaceuticals, Incorporated
clinicaltrials.gov (2022-04-05)
<https://clinicaltrials.gov/ct2/show/NCT04484025>

949. **A Phase 2, Randomized, Double-Blind, Placebo-Controlled, Dose Escalation Study to Evaluate the Safety and Efficacy of SPI-1005 in Severe COVID-19 Patients**

Sound Pharmaceuticals, Incorporated
clinicaltrials.gov (2022-04-05)
<https://clinicaltrials.gov/ct2/show/NCT04483973>

950. **A PHASE 1B, 2-PART, DOUBLE-BLIND, PLACEBO-CONTROLLED, SPONSOR-OPEN STUDY, TO EVALUATE THE SAFETY, TOLERABILITY AND PHARMACOKINETICS OF SINGLE ASCENDING (24-HOUR, PART 1) AND MULTIPLE ASCENDING (120-HOUR, PART 2) INTRAVENOUS INFUSIONS OF PF-07304814 IN HOSPITALIZED PARTICIPANTS WITH COVID-19**

Pfizer
clinicaltrials.gov (2021-06-23)
<https://clinicaltrials.gov/ct2/show/NCT04535167>

951. **AN INTERVENTIONAL EFFICACY AND SAFETY, PHASE 2/3, DOUBLE-BLIND, 2-ARM STUDY TO INVESTIGATE ORALLY ADMINISTERED PF-07321332/RITONAVIR COMPARED WITH PLACEBO IN NONHOSPITALIZED SYMPTOMATIC ADULT PARTICIPANTS WITH COVID-19 WHO ARE AT INCREASED RISK OF PROGRESSING TO SEVERE ILLNESS**

Pfizer
clinicaltrials.gov (2022-04-28)
<https://clinicaltrials.gov/ct2/show/NCT04960202>

952. **Use of big data in drug development for precision medicine: an update**

Tongqi Qian, Shijia Zhu, Yujin Hoshida
Expert Review of Precision Medicine and Drug Development (2019-05-04) <https://doi.org/gmpgx2>
DOI: [10.1080/23808993.2019.1617632](https://doi.org/23808993.2019.1617632) · PMID: [31286058](#) · PMCID: [PMC6613936](#)

953. **The COVID-19 Drug and Gene Set Library**

Maxim V Kuleshov, Daniel J Stein, Daniel JB Clarke, Eryk Kropiwnicki, Kathleen M Jagodnik, Alon Bartal, John E Evangelista, Jason Hom, Minxuan Cheng, Allison Bailey, ... Avi Ma'ayan
Patterns (2020-09) <https://doi.org/gg56f3>
DOI: [10.1016/j.patter.2020.100090](https://doi.org/10.1016/j.patter.2020.100090) · PMID: [32838343](#) · PMCID: [PMC7381899](#)

954. **Exploring the SARS-CoV-2 virus-host-drug interactome for drug repurposing**

Sepideh Sadegh, Julian Matschinske, David B Blumenthal, Ghanna Galindez, Tim Kacprowski, Markus List, Reza Nasirigerdeh, Mhaned Oubounyt, Andreas Pichlmair, Tim Daniel Rose, ... Jan Baumbach
Nature Communications (2020-07-14) <https://doi.org/gg477d>
DOI: [10.1038/s41467-020-17189-2](https://doi.org/10.1038/s41467-020-17189-2) · PMID: [32665542](#) · PMCID: [PMC7360763](#)

955. **Artificial intelligence in COVID-19 drug repurposing**
Yadi Zhou, Fei Wang, Jian Tang, Ruth Nussinov, Feixiong Cheng
The Lancet Digital Health (2020-12) <https://doi.org/gr9>
DOI: [10.1016/s2589-7500\(20\)30192-8](https://doi.org/10.1016/s2589-7500(20)30192-8) · PMID: [32984792](https://pubmed.ncbi.nlm.nih.gov/32984792/) · PMCID: [PMC7500917](https://pubmed.ncbi.nlm.nih.gov/PMC7500917/)
956. **The pharmacology of sigma-1 receptors**
Tangui Maurice, Tsung-Ping Su
Pharmacology & Therapeutics (2009-11) <https://doi.org/fhm455>
DOI: [10.1016/j.pharmthera.2009.07.001](https://doi.org/10.1016/j.pharmthera.2009.07.001) · PMID: [19619582](https://pubmed.ncbi.nlm.nih.gov/19619582/) · PMCID: [PMC2785038](https://pubmed.ncbi.nlm.nih.gov/PMC2785038/)
957. **Comparative host-coronavirus protein interaction networks reveal pan-viral disease mechanisms.**
David E Gordon, Joseph Hiatt, Mehdi Bouhaddou, Veronica V Rezelj, Svenja Ulferts, Hannes Braberg, Alexander S Jureka, Kirsten Obernier, Jeffrey Z Guo, Jyoti Batra, ... Nevan J Krogan
Science (New York, N.Y.) (2020-10-15)
<https://www.ncbi.nlm.nih.gov/pubmed/33060197>
DOI: [10.1126/science.abe9403](https://doi.org/10.1126/science.abe9403) · PMID: [33060197](https://pubmed.ncbi.nlm.nih.gov/33060197/) · PMCID: [PMC7808408](https://pubmed.ncbi.nlm.nih.gov/PMC7808408/)
958. **Repurposing Sigma-1 Receptor Ligands for COVID-19 Therapy?**
José Miguel Vela
Frontiers in Pharmacology (2020-11-09) <https://doi.org/gmh3mh>
DOI: [10.3389/fphar.2020.582310](https://doi.org/10.3389/fphar.2020.582310) · PMID: [33364957](https://pubmed.ncbi.nlm.nih.gov/33364957/) · PMCID: [PMC7751758](https://pubmed.ncbi.nlm.nih.gov/PMC7751758/)
959. **The Sigma Receptor: Evolution of the Concept in Neuropsychopharmacology**
T Hayashi, T-Su
Current Neuropharmacology (2005-10-01) <https://doi.org/fwpwcs>
DOI: [10.2174/157015905774322516](https://doi.org/10.2174/157015905774322516) · PMID: [18369400](https://pubmed.ncbi.nlm.nih.gov/18369400/) · PMCID: [PMC2268997](https://pubmed.ncbi.nlm.nih.gov/PMC2268997/)
960. **Drug-induced phospholipidosis confounds drug repurposing for SARS-CoV-2**
Tia A Tummino, Veronica V Rezelj, Benoit Fischer, Audrey Fischer, Matthew J O'Meara, Blandine Monel, Thomas Vallet, Kris M White, Ziyang Zhang, Assaf Alon, ... Brian K Shoichet
Science (2021-07-30) <https://doi.org/gmgx79>
DOI: [10.1126/science.abi4708](https://doi.org/10.1126/science.abi4708) · PMID: [34326236](https://pubmed.ncbi.nlm.nih.gov/34326236/) · PMCID: [PMC8501941](https://pubmed.ncbi.nlm.nih.gov/PMC8501941/)
961. **Emerging mechanisms of drug-induced phospholipidosis**
Bernadette Breiden, Konrad Sandhoff
Biological Chemistry (2019-08-13) <https://doi.org/gjkv8x>
DOI: [10.1515/hsz-2019-0270](https://doi.org/10.1515/hsz-2019-0270) · PMID: [31408430](https://pubmed.ncbi.nlm.nih.gov/31408430/)
962. **Zebra-like bodies in COVID-19: is phospholipidosis evidence of hydroxychloroquine induced acute kidney injury?**
Mohammad Obeidat, Alexandra L Isaacson, Stephanie J Chen, Marina Ivanovic, Danniele Holanda
Ultrastructural Pathology (2020-11-20) <https://doi.org/gj3v6c>
DOI: [10.1080/01913123.2020.1850966](https://doi.org/10.1080/01913123.2020.1850966) · PMID: [33274661](https://pubmed.ncbi.nlm.nih.gov/33274661/)

963. **Drug repurposing screens reveal cell-type-specific entry pathways and FDA-approved drugs active against SARS-CoV-2**

Mark Dittmar, Jae Seung Lee, Kanupriya Whig, Elisha Segrist, Minghua Li, Brinda Kamalia, Lauren Castellana, Kasirajan Ayyanathan, Fabian L Cardenas-Diaz, Edward E Morrisey, ... Sara Cherry

Cell Reports (2021-04) <https://doi.org/gj3v44>

DOI: [10.1016/j.celrep.2021.108959](https://doi.org/10.1016/j.celrep.2021.108959) · PMID: [33811811](#) · PMCID: [PMC7985926](#)

964. **No shortcuts to SARS-CoV-2 antivirals**

Aled Edwards, Ingo V Hartung

Science (2021-07-30) <https://doi.org/gmh3mg>

DOI: [10.1126/science.abj9488](https://doi.org/10.1126/science.abj9488) · PMID: [34326222](#)

965. **Repurposing of CNS drugs to treat COVID-19 infection: targeting the sigma-1 receptor**

Kenji Hashimoto

European Archives of Psychiatry and Clinical Neuroscience (2021-01-05) <https://doi.org/ghth6q>

DOI: [10.1007/s00406-020-01231-x](https://doi.org/10.1007/s00406-020-01231-x) · PMID: [33403480](#) · PMCID: [PMC7785036](#)

966. **Fluvoxamine vs Placebo and Clinical Deterioration in Outpatients With Symptomatic COVID-19**

Eric J Lenze, Caline Mattar, Charles F Zorumski, Angela Stevens, Julie Schweiger, Ginger E Nicol, JPhilip Miller, Lei Yang, Michael Yingling, Michael S Avidan, Angela M Reiersen

JAMA (2020-12-08) <https://doi.org/ghjtd5>

DOI: [10.1001/jama.2020.22760](https://doi.org/10.1001/jama.2020.22760) · PMID: [33180097](#) · PMCID: [PMC7662481](#)

967. **Prospective Cohort of Fluvoxamine for Early Treatment of Coronavirus Disease 19**

David Seftel, David R Boulware

Open Forum Infectious Diseases (2021-02-01) <https://doi.org/gmj9sz>

DOI: [10.1093/ofid/ofab050](https://doi.org/10.1093/ofid/ofab050) · PMID: [33623808](#) · PMCID: [PMC7888564](#)

968. **Modulation of the sigma-1 receptor-IRE1 pathway is beneficial in preclinical models of inflammation and sepsis**

Dorian A Rosen, Scott M Seki, Anthony Fernández-Castañeda, Rebecca M Beiter, Jacob D Eccles, Judith A Woodfolk, Alban Gaultier

Science Translational Medicine (2019-02-06) <https://doi.org/gmj9s2>

DOI: [10.1126/scitranslmed.aau5266](https://doi.org/10.1126/scitranslmed.aau5266) · PMID: [30728287](#) · PMCID: [PMC6936250](#)

969. **Fluvoxamine: A Review of Its Mechanism of Action and Its Role in COVID-19**

Vikas P Sukhatme, Angela M Reiersen, Sharat J Vayttaden, Vidula V Sukhatme

Frontiers in Pharmacology (2021-04-20) <https://doi.org/gmj9tg>

DOI: [10.3389/fphar.2021.652688](https://doi.org/10.3389/fphar.2021.652688) · PMID: [33959018](#) · PMCID: [PMC8094534](#)

970. **Too Many Papers**

Derek Lowe

In the Pipeline (2021-07-19)

<https://blogs.sciencemag.org/pipeline/archives/2021/07/19/too-many-papers>

971. **Believe it or not: how much can we rely on published data on potential drug targets?**

Florian Prinz, Thomas Schlange, Khusru Asadullah

Nature Reviews Drug Discovery (2011-08-31) <https://doi.org/dfsxxb>

DOI: [10.1038/nrd3439-c1](https://doi.org/10.1038/nrd3439-c1) · PMID: [21892149](https://pubmed.ncbi.nlm.nih.gov/21892149/)

972. **How were new medicines discovered?**

David C Swinney, Jason Anthony

Nature Reviews Drug Discovery (2011-06-24) <https://doi.org/bbg5wh>

DOI: [10.1038/nrd3480](https://doi.org/10.1038/nrd3480) · PMID: [21701501](https://pubmed.ncbi.nlm.nih.gov/21701501/)

973. **Big studies dim hopes for hydroxychloroquine**

Kai Kupferschmidt

Science (2020-06-12) <https://doi.org/gh7d7p>

DOI: [10.1126/science.368.6496.1166](https://doi.org/10.1126/science.368.6496.1166) · PMID: [32527806](https://pubmed.ncbi.nlm.nih.gov/32527806/)

974. **Trends in COVID-19 therapeutic clinical trials**

Kevin Bugin, Janet Woodcock

Nature Reviews Drug Discovery (2021-02-25) <https://doi.org/gmj9sj>

DOI: [10.1038/d41573-021-00037-3](https://doi.org/10.1038/d41573-021-00037-3) · PMID: [33633370](https://pubmed.ncbi.nlm.nih.gov/33633370/)

975. **Clinical Trial Data Sharing for COVID-19-Related Research**

Louis Dron, Alison Dillman, Michael J Zoratti, Jonas Haggstrom, Edward J Mills, Jay JH Park

Journal of Medical Internet Research (2021-03-12) <https://doi.org/gk6zfj>

DOI: [10.2196/26718](https://doi.org/10.2196/26718) · PMID: [33684053](https://pubmed.ncbi.nlm.nih.gov/33684053/) · PMCID: [PMC7958972](https://pubmed.ncbi.nlm.nih.gov/PMC7958972/)

976. **The Rise and Fall of Hydroxychloroquine for the Treatment and Prevention of COVID-19**

Zelyn Lee, Craig R Rayner, Jamie I Forrest, Jean B Nachega, Esha Senchaudhuri, Edward J Mills

The American Journal of Tropical Medicine and Hygiene (2021-01-06)

<https://doi.org/gmj9s3>

DOI: [10.4269/ajtmh.20-1320](https://doi.org/10.4269/ajtmh.20-1320) · PMID: [33236703](https://pubmed.ncbi.nlm.nih.gov/33236703/) · PMCID: [PMC7790108](https://pubmed.ncbi.nlm.nih.gov/PMC7790108/)

977. **Moving forward in clinical research with master protocols**

Jay JH Park, Louis Dron, Edward J Mills

Contemporary Clinical Trials (2021-07) <https://doi.org/gmj9sd>

DOI: [10.1016/j.cct.2021.106438](https://doi.org/10.1016/j.cct.2021.106438) · PMID: [34000408](https://pubmed.ncbi.nlm.nih.gov/34000408/) · PMCID: [PMC8120789](https://pubmed.ncbi.nlm.nih.gov/PMC8120789/)

978. **Main protease structure and XChem fragment screen**

Diamond

(2020-05-05) <https://www.diamond.ac.uk/covid-19/for-scientists/Main-protease-structure-and-XChem.html>

979. **Non-traditional cytokines: How catecholamines and adipokines influence macrophages in immunity, metabolism and the central nervous system**

Mark A Barnes, Monica J Carson, Meera G Nair

Cytokine (2015-04) <https://doi.org/f65c59>

980. **Stress Hormones, Proinflammatory and Antiinflammatory Cytokines, and Autoimmunity**

ILIA J ELENKOV, GEORGE P CHROUSOS

Annals of the New York Academy of Sciences (2002-06)

<https://doi.org/fmwpx2>

DOI: [10.1111/j.1749-6632.2002.tb04229.x](https://doi.org/10.1111/j.1749-6632.2002.tb04229.x) · PMID: [12114286](https://pubmed.ncbi.nlm.nih.gov/12114286/)

981. **Recovery of the Hypothalamic-Pituitary-Adrenal Response to Stress**

Arantxa García, Octavi Martí, Astrid Vallès, Silvina Dal-Zotto, Antonio Armario

Neuroendocrinology (2000) <https://doi.org/b2cq8n>

DOI: [10.1159/000054578](https://doi.org/10.1159/000054578) · PMID: [10971146](https://pubmed.ncbi.nlm.nih.gov/10971146/)

982. **Modulatory effects of glucocorticoids and catecholamines on human interleukin-12 and interleukin-10 production: clinical implications.**

IJ Elenkov, DA Papanicolaou, RL Wilder, GP Chrousos

Proceedings of the Association of American Physicians (1996-09)

<https://www.ncbi.nlm.nih.gov/pubmed/8902882>

PMID: [8902882](https://pubmed.ncbi.nlm.nih.gov/8902882/)

983. **Dexamethasone for COVID-19? Not so fast.**

TC Theoharides

JOURNAL OF BIOLOGICAL REGULATORS AND HOMEOSTATIC AGENTS

(2020-08-31) <https://doi.org/ghfkjx>

DOI: [10.23812/20-editorial_1-5](https://doi.org/10.23812/20-editorial_1-5) · PMID: [32551464](https://pubmed.ncbi.nlm.nih.gov/32551464/)

984. **Dexamethasone in hospitalised patients with COVID-19: addressing uncertainties**

Michael A Matthay, BTaylor Thompson

The Lancet Respiratory Medicine (2020-12) <https://doi.org/ftk4>

DOI: [10.1016/s2213-2600\(20\)30503-8](https://doi.org/10.1016/s2213-2600(20)30503-8) · PMID: [33129421](https://pubmed.ncbi.nlm.nih.gov/33129421/) · PMCID: [PMC7598750](https://pubmed.ncbi.nlm.nih.gov/PMC7598750/)

985. **Dexamethasone for COVID-19: data needed from randomised clinical trials in Africa**

Helen Brotherton, Effua Usuf, Behzad Nadjm, Karen Forrest, Kalifa Bojang, Ahmadou Lamin Samateh, Mustapha Bittaye, Charles AP Roberts, Umberto d'Alessandro, Anna Roca

The Lancet Global Health (2020-09) <https://doi.org/gg42kx>

DOI: [10.1016/s2214-109x\(20\)30318-1](https://doi.org/10.1016/s2214-109x(20)30318-1) · PMID: [32679038](https://pubmed.ncbi.nlm.nih.gov/32679038/) · PMCID: [PMC7833918](https://pubmed.ncbi.nlm.nih.gov/PMC7833918/)

986. **Experimental Treatment with Favipiravir for COVID-19: An Open-Label Control Study**

Qingxian Cai, Minghui Yang, Dongjing Liu, Jun Chen, Dan Shu, Junxia Xia, Xuejiao Liao, Yuanbo Gu, Qie Cai, Yang Yang, ... Lei Liu

Engineering (2020-10) <https://doi.org/ggpprd>

DOI: [10.1016/j.eng.2020.03.007](https://doi.org/10.1016/j.eng.2020.03.007) · PMID: [32346491](https://pubmed.ncbi.nlm.nih.gov/32346491/)

987. **Lopinavir-ritonavir in patients admitted to hospital with COVID-19 (RECOVERY): a randomised, controlled, open-label, platform trial**

Peter W Horby, Marion Mafham, Jennifer L Bell, Louise Linsell, Natalie Staplin, Jonathan Emberson, Adrian Palfreeman, Jason Raw, Einas Elmahi, Benjamin Prudon, ... Martin J Landray

The Lancet (2020-10) <https://doi.org/fnx2>

DOI: [10.1016/s0140-6736\(20\)32013-4](https://doi.org/s0140-6736(20)32013-4) · PMID: [33031764](https://pubmed.ncbi.nlm.nih.gov/33031764/) · PMCID: [PMC7535623](https://pubmed.ncbi.nlm.nih.gov/PMC7535623/)

988. **A Large, Simple Trial Leading to Complex Questions**

David P Harrington, Lindsey R Baden, Joseph W Hogan

New England Journal of Medicine (2020-12-02) <https://doi.org/ghnhnx>

DOI: [10.1056/nejme2034294](https://doi.org/10.1056/nejme2034294) · PMID: [33264557](https://pubmed.ncbi.nlm.nih.gov/33264557/) · PMCID: [PMC7727323](https://pubmed.ncbi.nlm.nih.gov/PMC7727323/)

989. **Retracted coronavirus (COVID-19) papers**

Retraction Watch

(2020-04-29) <https://retractionwatch.com/retracted-coronavirus-covid-19-papers/>

990. **Clinical Outcomes and Plasma Concentrations of Baloxavir Marboxil and Favipiravir in COVID-19 Patients: An Exploratory Randomized, Controlled Trial**

Yan Lou, Lin Liu, Hangping Yao, Xingjiang Hu, Junwei Su, Kaijin Xu, Rui Luo, Xi Yang, Lingjuan He, Xiaoyang Lu, ... Yunqing Qiu

European Journal of Pharmaceutical Sciences (2021-02)

<https://doi.org/ghx88n>

DOI: [10.1016/j.ejps.2020.105631](https://doi.org/10.1016/j.ejps.2020.105631) · PMID: [33115675](https://pubmed.ncbi.nlm.nih.gov/33115675/) · PMCID: [PMC7585719](https://pubmed.ncbi.nlm.nih.gov/PMC7585719/)

991. **Efficacy of favipiravir in COVID-19 treatment: a multi-center randomized study**

Hany M Dabbous, Sherief Abd-Elsalam, Manal H El-Sayed, Ahmed F Sherief, Fatma FS Ebeid, Mohamed Samir Abd El Ghafar, Shaimaa Soliman, Mohamed Elbahnaawy, Rehab Badawi, Mohamed Awad Tageldin

Archives of Virology (2021-01-25) <https://doi.org/ghx874>

DOI: [10.1007/s00705-021-04956-9](https://doi.org/s00705-021-04956-9) · PMID: [33492523](https://pubmed.ncbi.nlm.nih.gov/33492523/) · PMCID: [PMC7829645](https://pubmed.ncbi.nlm.nih.gov/PMC7829645/)

992. **AVIFAVIR for Treatment of Patients With Moderate Coronavirus Disease 2019 (COVID-19): Interim Results of a Phase II/III Multicenter Randomized Clinical Trial**

Andrey A Ivashchenko, Kirill A Dmitriev, Natalia V Vostokova, Valeria N Azarova, Andrew A Blinow, Alina N Egorova, Ivan G Gordeev, Alexey P Ilin, Ruben N Karapetian, Dmitry V Kravchenko, ... Alexandre V Ivachtchenko

Clinical Infectious Diseases (2020-08-09) <https://doi.org/ghx9c2>

DOI: [10.1093/cid/ciaa1176](https://doi.org/10.1093/cid/ciaa1176) · PMID: [32770240](https://pubmed.ncbi.nlm.nih.gov/32770240/) · PMCID: [PMC7454388](https://pubmed.ncbi.nlm.nih.gov/PMC7454388/)

993. **A review of the safety of favipiravir – a potential treatment in the COVID-19 pandemic?**

Victoria Pilkington, Toby Pepperrell, Andrew Hill

Journal of Virus Eradication (2020-04) <https://doi.org/ftgm>

994. **A Randomized, Controlled Trial of Ebola Virus Disease Therapeutics**

Sabue Mulangu, Lori E Dodd, Richard T Davey, Olivier Tshiani Mbaya, Michael Proschan, Daniel Mukadi, Mariano Lusakibanza Manzo, Didier Nzolo, Antoine Tshomba Oloma, Augustin Ibanda, ... the PALM Writing Group

New England Journal of Medicine (2019-12-12) <https://doi.org/gggmx4>
DOI: [10.1056/nejmoa1910993](https://doi.org/10.1056/nejmoa1910993) · PMID: [31774950](#)

995. **Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses**

Timothy P Sheahan, Amy C Sims, Rachel L Graham, Vineet D Menachery, Lisa E Gralinski, James B Case, Sarah R Leist, Krzysztof Pyrc, Joy Y Feng, Iva Trantcheva, ... Ralph S Baric

Science Translational Medicine (2017-06-28) <https://doi.org/gc3grb>
DOI: [10.1126/scitranslmed.aal3653](https://doi.org/10.1126/scitranslmed.aal3653) · PMID: [28659436](#) · PMCID: [PMC5567817](#)

996. **Did an experimental drug help a U.S. coronavirus patient?**

Jon Cohen

Science (2020-03-13) <https://doi.org/ggqm62>
DOI: [10.1126/science.abb7243](https://doi.org/10.1126/science.abb7243)

997. **First 12 patients with coronavirus disease 2019 (COVID-19) in the United States**

Stephanie A Kujawski, Karen K Wong, Jennifer P Collins, Lauren Epstein, Marie E Killerby, Claire M Midgley, Glen R Abedi, NSeema Ahmed, Olivia Almendares, Francisco N Alvarez, ... The COVID-19 Investigation Team

Cold Spring Harbor Laboratory (2020-03-12) <https://doi.org/ggqm6z>
DOI: [10.1101/2020.03.09.20032896](https://doi.org/10.1101/2020.03.09.20032896)

998. **First Case of 2019 Novel Coronavirus in the United States**

Michelle L Holshue, Chas DeBolt, Scott Lindquist, Kathy H Lofy, John Wiesman, Holianne Bruce, Christopher Spitters, Keith Ericson, Sara Wilkerson, Ahmet Tural, ... Satish K Pillai

New England Journal of Medicine (2020-03-05) <https://doi.org/ggjvr6>
DOI: [10.1056/nejmoa2001191](https://doi.org/10.1056/nejmoa2001191) · PMID: [32004427](#) · PMCID: [PMC7092802](#)

999. **Remdesivir for 5 or 10 Days in Patients with Severe Covid-19**

Jason D Goldman, David CB Lye, David S Hui, Kristen M Marks, Raffaele Bruno, Rocio Montejano, Christoph D Spinner, Massimo Galli, Mi-Young Ahn, Ronald G Nahass, ... Aruna Subramanian

New England Journal of Medicine (2020-11-05) <https://doi.org/ggz7qy>
DOI: [10.1056/nejmoa2015301](https://doi.org/10.1056/nejmoa2015301) · PMID: [32459919](#) · PMCID: [PMC7377062](#)

1000. **Remdesivir EUA Letter of Authorization**

Denise M Hinton

(2020-05-01) <https://www.fda.gov/media/137564/download>

1001. **A Phase 3 Randomized Study to Evaluate the Safety and Antiviral Activity of Remdesivir (GS-5734™) in Participants With Moderate**

COVID-19 Compared to Standard of Care Treatment

Gilead Sciences

clinicaltrials.gov (2021-01-21)

<https://clinicaltrials.gov/ct2/show/NCT04292730>

1002. **Multi-centre, adaptive, randomized trial of the safety and efficacy of treatments of COVID-19 in hospitalized adults**

EU Clinical Trials Register

(2020-03-09) <https://www.clinicaltrialsregister.eu/ctr-search/trial/2020-000936-23/FR>

1003. **A Trial of Remdesivir in Adults With Mild and Moderate COVID-19 - Full Text View - ClinicalTrials.gov**

<https://clinicaltrials.gov/ct2/show/NCT04252664>

1004. **A Phase 3 Randomized, Double-blind, Placebo-controlled, Multicenter Study to Evaluate the Efficacy and Safety of Remdesivir in Hospitalized Adult Patients With Severe COVID-19.**

Bin Cao

clinicaltrials.gov (2020-04-13)

<https://clinicaltrials.gov/ct2/show/NCT04257656>

1005. **FDA Approves First Treatment for COVID-19**

Office of the Commissioner

FDA (2020-10-22) <https://www.fda.gov/news-events/press-announcements/fda-approves-first-treatment-covid-19>

1006. **Gilead Sciences Statement on the Solidarity Trial**

<https://www.gilead.com/news-and-press/company-statements/gilead-sciences-statement-on-the-solidarity-trial>

1007. **Conflicting results on the efficacy of remdesivir in hospitalized Covid-19 patients: comment on the Adaptive Covid-19 Treatment Trial**

Leonarda Galiuto, Carlo Patrono

European Heart Journal (2020-12-07) <https://doi.org/ghp4kw>

DOI: [10.1093/eurheartj/ehaa934](https://doi.org/10.1093/eurheartj/ehaa934) · PMID: [33306101](https://pubmed.ncbi.nlm.nih.gov/33306101/) · PMCID: [PMC7799042](https://pubmed.ncbi.nlm.nih.gov/PMC7799042/)

1008. **The 'very, very bad look' of remdesivir, the first FDA-approved COVID-19 drug**

Jon Cohen, Kai Kupferschmidt

Science / AAAS (2020-10-28)

<https://www.sciencemag.org/news/2020/10/very-very-bad-look-remdesivir-first-fda-approved-covid-19-drug>

1009. **Effect of Remdesivir vs Standard Care on Clinical Status at 11 Days in Patients With Moderate COVID-19**

Christoph D Spinner, Robert L Gottlieb, Gerard J Criner, José Ramón Arribas López, Anna Maria Cattelan, Alex Soriano Viladomiu, Onyema Ogbuagu, Prashant Malhotra, Kathleen M Mullane, Antonella Castagna, ... for the GS-US-540-5774 Investigators

JAMA (2020-09-15) <https://doi.org/ghhz6g>

DOI: [10.1001/jama.2020.16349](https://doi.org/10.1001/jama.2020.16349) · PMID: [32821939](https://pubmed.ncbi.nlm.nih.gov/32821939/) · PMCID: [PMC7442954](https://pubmed.ncbi.nlm.nih.gov/PMC7442954/)

1010. **Baricitinib plus Remdesivir for Hospitalized Adults with Covid-19**
Andre C Kalil, Thomas F Patterson, Aneesh K Mehta, Kay M Tomashek, Cameron R Wolfe, Varduhi Ghazaryan, Vincent C Marconi, Guillermo M Ruiz-Palacios, Lanny Hsieh, Susan Kline, ... John H Beigel
New England Journal of Medicine (2020-12-11) <https://doi.org/ghpbd2>
DOI: [10.1056/nejmoa2031994](https://doi.org/10.1056/nejmoa2031994) · PMID: [33306283](#) · PMCID: [PMC7745180](#)
1011. **Letter of Authorization: EUA for baricitinib (Olumiant), in combination with remdesivir (Veklury), for the treatment of suspected or laboratory confirmed coronavirus disease 2019 (COVID-19)**
Denise M Hinton
Food and Drug Administration (2020-01-19)
<https://www.fda.gov/media/143822/download>
1012. **Lysosomotropic agents as HCV entry inhibitors**
Usman A Ashfaq, Tariq Javed, Sidra Rehman, Zafar Nawaz, Sheikh Riazuddin
Virology Journal (2011-04-12) <https://doi.org/dr5g4m>
DOI: [10.1186/1743-422x-8-163](https://doi.org/10.1186/1743-422x-8-163) · PMID: [21481279](#) · PMCID: [PMC3090357](#)
1013. **Mechanism of Action of Hydroxychloroquine in the Antiphospholipid Syndrome**
Nadine Müller-Calleja, Davit Manukyan, Wolfram Ruf, Karl Lackner
Blood (2016-12-02) <https://doi.org/ggrm82>
DOI: [10.1182/blood.v128.22.5023.5023](https://doi.org/10.1182/blood.v128.22.5023.5023)
1014. **14th International Congress on Antiphospholipid Antibodies Task Force Report on Antiphospholipid Syndrome Treatment Trends**
Doruk Erkan, Cassyanne L Aguiar, Danieli Andrade, Hannah Cohen, Maria J Cuadrado, Adriana Danowski, Roger A Levy, Thomas L Ortel, Anisur Rahman, Jane E Salmon, ... Michael D Lockshin
Autoimmunity Reviews (2014-06) <https://doi.org/ggp8r8>
DOI: [10.1016/j.autrev.2014.01.053](https://doi.org/10.1016/j.autrev.2014.01.053) · PMID: [24468415](#)
1015. **What is the role of hydroxychloroquine in reducing thrombotic risk in patients with antiphospholipid antibodies?**
Tzu-Fei Wang, Wendy Lim
Hematology (2016-12-02) <https://doi.org/ggrn3k>
DOI: [10.1182/asheducation-2016.1.714](https://doi.org/10.1182/asheducation-2016.1.714) · PMID: [27913551](#) · PMCID: [PMC6142483](#)
1016. **COVID-19: a recommendation to examine the effect of hydroxychloroquine in preventing infection and progression**
Dan Zhou, Sheng-Ming Dai, Qiang Tong
Journal of Antimicrobial Chemotherapy (2020-07)
<https://doi.org/ggq84c>
DOI: [10.1093/jac/dkaa114](https://doi.org/10.1093/jac/dkaa114) · PMID: [32196083](#) · PMCID: [PMC7184499](#)
1017. **Hydroxychloroquine treatment of patients with human immunodeficiency virus type 1**
Kirk Sperber, Michael Louie, Thomas Kraus, Jacqueline Proner, Erica Sapira, Su Lin, Vera Stecher, Lloyd Mayer
Clinical Therapeutics (1995-07) <https://doi.org/cq2hx9>

DOI: [10.1016/0149-2918\(95\)80039-5](https://doi.org/10.1016/0149-2918(95)80039-5)

1018. **Hydroxychloroquine augments early virological response to pegylated interferon plus ribavirin in genotype-4 chronic hepatitis C patients**

Gouda Kamel Helal, Magdy Abdelmawgoud Gad, Mohamed Fahmy Abd-Ellah, Mahmoud Saied Eid

Journal of Medical Virology (2016-12) <https://doi.org/f889nt>

DOI: [10.1002/jmv.24575](https://doi.org/10.1002/jmv.24575) · PMID: [27183377](https://pubmed.ncbi.nlm.nih.gov/27183377/) · PMCID: [PMC7167065](https://pubmed.ncbi.nlm.nih.gov/PMC7167065/)

1019. **Making the Best Match: Selecting Outcome Measures for Clinical Trials and Outcome Studies**

Wendy J Coster

The American Journal of Occupational Therapy (2013-03-01)

<https://doi.org/f4rf5s>

DOI: [10.5014/ajot.2013.006015](https://doi.org/10.5014/ajot.2013.006015) · PMID: [23433270](https://pubmed.ncbi.nlm.nih.gov/23433270/) · PMCID: [PMC3628620](https://pubmed.ncbi.nlm.nih.gov/PMC3628620/)

1020. **No evidence of rapid antiviral clearance or clinical benefit with the combination of hydroxychloroquine and azithromycin in patients with severe COVID-19 infection**

JM Molina, C Delaugerre, J Le Goff, B Mela-Lima, D Ponscarme, L Goldwirt, N de Castro

Médecine et Maladies Infectieuses (2020-06) <https://doi.org/ggqzrb>

DOI: [10.1016/j.medmal.2020.03.006](https://doi.org/10.1016/j.medmal.2020.03.006) · PMID: [32240719](https://pubmed.ncbi.nlm.nih.gov/32240719/)

1021. **Efficacy of hydroxychloroquine in patients with COVID-19: results of a randomized clinical trial**

Zhaowei Chen, Jijia Hu, Zongwei Zhang, Shan Jiang, Shoumeng Han, Dandan Yan, Ruhong Zhuang, Ben Hu, Zhan Zhang

Cold Spring Harbor Laboratory (2020-04-10) <https://doi.org/ggqm4v>

DOI: [10.1101/2020.03.22.20040758](https://doi.org/10.1101/2020.03.22.20040758)

1022. **Therapeutic effect of hydroxychloroquine on novel coronavirus pneumonia (COVID-19)**

Chinese Clinical Trial Registry

(2020-02-12) <http://www.chictr.org.cn/showprojen.aspx?proj=48880>

1023. **A pilot study of hydroxychloroquine in treatment of patients with common coronavirus disease-19 (COVID-19)**

CHEN Jun, LIU Danping, LIU Li, LIU Ping, XU Qingnian, XIA Lu, LING Yun, HUANG Dan, SONG Shuli, ZHANG Dandan, ... LU Hongzhou

Journal of Zhejiang University (Medical Sciences) (2020-03)

<https://doi.org/10.3785/j.issn.1008-9292.2020.03.03>

DOI: [10.3785/j.issn.1008-9292.2020.03.03](https://doi.org/10.3785/j.issn.1008-9292.2020.03.03)

1024. **Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies**

Jianjun Gao, Zhenxue Tian, Xu Yang

BioScience Trends (2020-02-29) <https://doi.org/ggm3mv>

DOI: [10.5582/bst.2020.01047](https://doi.org/10.5582/bst.2020.01047) · PMID: [32074550](https://pubmed.ncbi.nlm.nih.gov/32074550/)

1025. **Targeting the Endocytic Pathway and Autophagy Process as a Novel Therapeutic Strategy in COVID-19**

Naidi Yang, Han-Ming Shen
International Journal of Biological Sciences (2020)
<https://doi.org/ggqspm>
DOI: [10.7150/ijbs.45498](https://doi.org/10.7150/ijbs.45498) · PMID: [32226290](https://pubmed.ncbi.nlm.nih.gov/32226290/) · PMCID: [PMC7098027](https://pubmed.ncbi.nlm.nih.gov/PMC7098027/)

1026. **SARS-CoV-2: an Emerging Coronavirus that Causes a Global Threat**

Jun Zheng
International Journal of Biological Sciences (2020)
<https://doi.org/ggqspr>
DOI: [10.7150/ijbs.45053](https://doi.org/10.7150/ijbs.45053) · PMID: [32226285](https://pubmed.ncbi.nlm.nih.gov/32226285/) · PMCID: [PMC7098030](https://pubmed.ncbi.nlm.nih.gov/PMC7098030/)

1027. **RETRACTED: Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis**

Mandeep R Mehra, Sapan S Desai, Frank Ruschitzka, Amit N Patel
The Lancet (2020-05) <https://doi.org/ggwzsb>
DOI: [10.1016/s0140-6736\(20\)31180-6](https://doi.org/10.1016/s0140-6736(20)31180-6) · PMID: [32450107](https://pubmed.ncbi.nlm.nih.gov/32450107/) · PMCID: [PMC7255293](https://pubmed.ncbi.nlm.nih.gov/PMC7255293/)

1028. **Retraction—Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis**

Mandeep R Mehra, Frank Ruschitzka, Amit N Patel
The Lancet (2020-06) <https://doi.org/ggzqng>
DOI: [10.1016/s0140-6736\(20\)31324-6](https://doi.org/10.1016/s0140-6736(20)31324-6) · PMID: [32511943](https://pubmed.ncbi.nlm.nih.gov/32511943/) · PMCID: [PMC7274621](https://pubmed.ncbi.nlm.nih.gov/PMC7274621/)

1029. **Life Threatening Severe QTc Prolongation in Patient with Systemic Lupus Erythematosus due to Hydroxychloroquine**

John P O'Laughlin, Parag H Mehta, Brian C Wong
Case Reports in Cardiology (2016) <https://doi.org/ggqzrc>
DOI: [10.1155/2016/4626279](https://doi.org/10.1155/2016/4626279) · PMID: [27478650](https://pubmed.ncbi.nlm.nih.gov/27478650/) · PMCID: [PMC4960328](https://pubmed.ncbi.nlm.nih.gov/PMC4960328/)

1030. **Keep the QT interval: It is a reliable predictor of ventricular arrhythmias**

Dan M Roden
Heart Rhythm (2008-08) <https://doi.org/d5rchx>
DOI: [10.1016/j.hrthm.2008.05.008](https://doi.org/10.1016/j.hrthm.2008.05.008) · PMID: [18675237](https://pubmed.ncbi.nlm.nih.gov/18675237/) · PMCID: [PMC3212752](https://pubmed.ncbi.nlm.nih.gov/PMC3212752/)

1031. **Safety of hydroxychloroquine, alone and in combination with azithromycin, in light of rapid wide-spread use for COVID-19: a multinational, network cohort and self-controlled case series study**

Jennifer C.E.Lane, James Weaver, Kristin Kostka, Talita Duarte-Salles, Maria Tereza F Abrahao, Heba Alghoul, Osaid Alser, Thamir M Alshammari, Patricia Biedermann, Edward Burn, ... Daniel Prieto-Alhambra
Cold Spring Harbor Laboratory (2020-04-10) <https://doi.org/ggrn7s>
DOI: [10.1101/2020.04.08.20054551](https://doi.org/10.1101/2020.04.08.20054551)

1032. **Chloroquine diphosphate in two different dosages as adjunctive therapy of hospitalized patients with severe respiratory syndrome in the context of coronavirus (SARS-CoV-2) infection:**

Preliminary safety results of a randomized, double-blinded, phase IIb clinical trial (CloroCovid-19 Study)

Mayla Gabriela Silva Borba, Fernando Fonseca Almeida Val, Vanderson Souza Sampaio, Marcia Almeida Araújo Alexandre, Gisely Cardoso Melo, Marcelo Brito, Maria Paula Gomes Mourão, José Diego Brito-Sousa, Djane Baía-da-Silva, Marcus Vinitius Farias Guerra, ... CloroCovid-19 Team

Cold Spring Harbor Laboratory (2020-04-16) <https://doi.org/ggr3nj>

DOI: [10.1101/2020.04.07.20056424](https://doi.org/10.1101/2020.04.07.20056424)

1033. Heart risk concerns mount around use of chloroquine and hydroxychloroquine for Covid-19 treatment

Jacqueline Howard, Elizabeth Cohen, Nadia Kounang, Per Nyberg
CNN (2020-04-14)

<https://www.cnn.com/2020/04/13/health/chloroquine-risks-coronavirus-treatment-trials-study/index.html>

1034. WHO Director-General's opening remarks at the media briefing on COVID-19

World Health Organization

(2020-05-25) <https://www.who.int/dg/speeches/detail/who-director-general-s-opening-remarks-at-the-media-briefing-on-covid-19--25-may-2020>

1035. Hydroxychloroquine in patients mainly with mild to moderate COVID-19: an open-label, randomized, controlled trial

Wei Tang, Zhujun Cao, Mingfeng Han, Zhengyan Wang, Junwen Chen, Wenjin Sun, Yaojie Wu, Wei Xiao, Shengyong Liu, Erzhen Chen, ... Qing Xie

Cold Spring Harbor Laboratory (2020-05-07) <https://doi.org/ggr68m>

DOI: [10.1101/2020.04.10.20060558](https://doi.org/10.1101/2020.04.10.20060558)

1036. Outcomes of hydroxychloroquine usage in United States veterans hospitalized with Covid-19

Joseph Magagnoli, Siddharth Narendran, Felipe Pereira, Tammy Cummings, James W Hardin, Scott Sutton, Jayakrishna Ambati
Cold Spring Harbor Laboratory (2020-04-21) <https://doi.org/ggspt6>
DOI: [10.1101/2020.04.16.20065920](https://doi.org/10.1101/2020.04.16.20065920) · PMID: [32511622](#) · PMCID: [PMC7276049](#)

1037. Hydroxychloroquine for Early Treatment of Adults With Mild Coronavirus Disease 2019: A Randomized, Controlled Trial

Oriol Mitjà, Marc Corbacho-Monné, Maria Ubals, Cristian Tebé, Judith Peñafiel, Aurelio Tobias, Ester Ballana, Andrea Alemany, Núria Riera-Martí, Carla A Pérez, ... Martí Vall-Mayans

Clinical Infectious Diseases (2020-07-16) <https://doi.org/gg5f9x>

DOI: [10.1093/cid/ciaa1009](https://doi.org/10.1093/cid/ciaa1009) · PMID: [32674126](#) · PMCID: [PMC7454406](#)

1038. A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19

David R Boulware, Matthew F Pullen, Ananta S Bangdiwala, Katelyn A Pastick, Sarah M Lofgren, Elizabeth C Okafor, Caleb P Skipper, Alanna A Nascene, Melanie R Nicol, Mahsa Abassi, ... Kathy H Hullsieck

New England Journal of Medicine (2020-08-06) <https://doi.org/dxkv>

DOI: [10.1056/nejmoa2016638](https://doi.org/10.1056/nejmoa2016638) · PMID: [32492293](#) · PMCID: [PMC7289276](#)

1039. **Efficacy and Safety of Hydroxychloroquine vs Placebo for Pre-exposure SARS-CoV-2 Prophylaxis Among Health Care Workers**
Benjamin S Abella, Eliana L Jolkovsky, Barbara T Biney, Julie E Uspal, Matthew C Hyman, Ian Frank, Scott E Hensley, Saar Gill, Dan T Vogl, Ivan Maillard, ... Prevention and Treatment of COVID-19 With Hydroxychloroquine (PATCH) Investigators
JAMA Internal Medicine (2021-02-01) <https://doi.org/ghd6nj>
DOI: [10.1001/jamainternmed.2020.6319](https://doi.org/10.1001/jamainternmed.2020.6319) · PMID: [33001138](#) · PMCID: [PMC7527945](#)
1040. **Mechanisms of action of hydroxychloroquine and chloroquine: implications for rheumatology**
Eva Schrezenmeier, Thomas Dörner
Nature Reviews Rheumatology (2020-02-07) <https://doi.org/ggzjnh>
DOI: [10.1038/s41584-020-0372-x](https://doi.org/10.1038/s41584-020-0372-x) · PMID: [32034323](#)
1041. **Elimination or Prolongation of ACE Inhibitors and ARB in Coronavirus Disease 2019 - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04338009>
1042. **Stopping ACE-inhibitors in COVID-19 - Full Text View - ClinicalTrials.gov** <https://clinicaltrials.gov/ct2/show/NCT04353596>
1043. **Losartan for Patients With COVID-19 Not Requiring Hospitalization - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04311177>
1044. **Losartan for Patients With COVID-19 Requiring Hospitalization - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04312009>
1045. **The CORONAVirus Disease 2019 Angiotensin Converting Enzyme Inhibitor/Angiotensin Receptor Blocker InvestigatiON (CORONACION) Randomized Clinical Trial**
Prof John William McEvoy
clinicaltrials.gov (2020-06-26)
<https://clinicaltrials.gov/ct2/show/NCT04330300>
1046. **Ramipril for the Treatment of COVID-19 - Full Text View - ClinicalTrials.gov** <https://clinicaltrials.gov/ct2/show/NCT04366050>
1047. **Suspension of Angiotensin Receptor Blockers and Angiotensin-converting Enzyme Inhibitors and Adverse Outcomes in Hospitalized Patients With Coronavirus Infection (COVID-19). A Randomized Trial**
D'Or Institute for Research and Education
clinicaltrials.gov (2020-07-01)
<https://clinicaltrials.gov/ct2/show/NCT04364893>
1048. **Response by Cohen et al to Letter Regarding Article, "Association of Inpatient Use of Angiotensin-Converting Enzyme Inhibitors and Angiotensin II Receptor Blockers With Mortality Among Patients With Hypertension Hospitalized With COVID-19"**
Jordana B Cohen, Thomas C Hanff, Andrew M South, Matthew A Sparks, Swapnil Hiremath, Adam P Bress, JBrian Byrd, Julio A Chirinos

1049. **Sound Science before Quick Judgement Regarding RAS Blockade in COVID-19**

Matthew A Sparks, Andrew South, Paul Welling, JMatt Luther, Jordana Cohen, James Brian Byrd, Louise M Burrell, Daniel Batlle, Laurie Tomlinson, Vivek Bhalla, ... Swapnil Hiremath
Clinical Journal of the American Society of Nephrology (2020-05-07)
<https://doi.org/ggg8gn>
DOI: [10.2215/cjn.03530320](https://doi.org/10.2215/cjn.03530320) · PMID: [32220930](https://pubmed.ncbi.nlm.nih.gov/32220930/) · PMCID: [PMC7269218](https://pubmed.ncbi.nlm.nih.gov/PMC7269218/)

1050. **The Coronavirus Conundrum: ACE2 and Hypertension Edition**

Matthew Sparks, Swapnil Hiremath
NephJC <http://www.nephjc.com/news/covidace2>

1051. **Hall of Fame among Pro-inflammatory Cytokines: Interleukin-6 Gene and Its Transcriptional Regulation Mechanisms**

Yang Luo, Song Guo Zheng
Frontiers in Immunology (2016-12-19) <https://doi.org/ggqmgy>
DOI: [10.3389/fimmu.2016.00604](https://doi.org/10.3389/fimmu.2016.00604) · PMID: [28066415](https://pubmed.ncbi.nlm.nih.gov/28066415/) · PMCID:
[PMC5165036](https://pubmed.ncbi.nlm.nih.gov/PMC5165036/)

1052. **IL-6 Trans-Signaling via the Soluble IL-6 Receptor: Importance for the Pro-Inflammatory Activities of IL-6**

Stefan Rose-John
International Journal of Biological Sciences (2012)
<https://doi.org/f4c4hf>
DOI: [10.7150/ijbs.4989](https://doi.org/10.7150/ijbs.4989) · PMID: [23136552](https://pubmed.ncbi.nlm.nih.gov/23136552/) · PMCID: [PMC3491447](https://pubmed.ncbi.nlm.nih.gov/PMC3491447/)

1053. **Interleukin-6 and its receptor: from bench to bedside**

Jürgen Scheller, Stefan Rose-John
Medical Microbiology and Immunology (2006-05-31)
<https://doi.org/ck8xch>
DOI: [10.1007/s00430-006-0019-9](https://doi.org/10.1007/s00430-006-0019-9) · PMID: [16741736](https://pubmed.ncbi.nlm.nih.gov/16741736/)

1054. **Plasticity and cross-talk of Interleukin 6-type cytokines**

Christoph Garbers, Heike M Hermanns, Fred Schaper, Gerhard Müller-Newen, Joachim Grötzingler, Stefan Rose-John, Jürgen Scheller
Cytokine & Growth Factor Reviews (2012-06) <https://doi.org/f3z743>
DOI: [10.1016/j.cytogfr.2012.04.001](https://doi.org/10.1016/j.cytogfr.2012.04.001) · PMID: [22595692](https://pubmed.ncbi.nlm.nih.gov/22595692/)

1055. **Soluble receptors for cytokines and growth factors: generation and biological function**

S Rose-John, PC Heinrich
Biochemical Journal (1994-06-01) <https://doi.org/ggqmgd>
DOI: [10.1042/bj3000281](https://doi.org/10.1042/bj3000281) · PMID: [8002928](https://pubmed.ncbi.nlm.nih.gov/8002928/) · PMCID: [PMC1138158](https://pubmed.ncbi.nlm.nih.gov/PMC1138158/)

1056. **Interleukin-6; pathogenesis and treatment of autoimmune inflammatory diseases**

Toshio Tanaka, Masashi Narazaki, Kazuya Masuda, Tadamitsu Kishimoto
Inflammation and Regeneration (2013) <https://doi.org/ggqmgt>
DOI: [10.2492/inflammregen.33.054](https://doi.org/10.2492/inflammregen.33.054)

1057. **Systematic Review and Meta-Analysis of Case-Control Studies from 7,000 COVID-19 Pneumonia Patients Suggests a Beneficial Impact of Tocilizumab with Benefit Most Evident in Non-Corticosteroid Exposed Subjects.**
Abdulla Watad, Nicola Luigi Bragazzi, Charlie Bridgewood, Muhammad Mansour, Naim Mahroum, Matteo Riccò, Ahmed Nasr, Amr Hussein, Omer Gendelman, Yehuda Shoenfeld, ... Dennis McGonagle
SSRN Electronic Journal (2020) <https://doi.org/gg62hz>
DOI: [10.2139/ssrn.3642653](https://doi.org/10.2139/ssrn.3642653)
1058. **The efficacy of IL-6 inhibitor Tocilizumab in reducing severe COVID-19 mortality: a systematic review**
Avi Gurion Kaye, Robert Siegel
PeerJ (2020-11-02) <https://doi.org/ghx8r4>
DOI: [10.7717/peerj.10322](https://doi.org/10.7717/peerj.10322) · PMID: [33194450](#) · PMCID: [PMC7643559](#)
1059. **Rationale and evidence on the use of tocilizumab in COVID-19: a systematic review**
A Cortegiani, M Ippolito, M Greco, V Granone, A Protti, C Gregoretti, A Giarratano, S Einav, M Cecconi
Pulmonology (2021-01) <https://doi.org/gg5xv3>
DOI: [10.1016/j.pulmoe.2020.07.003](https://doi.org/10.1016/j.pulmoe.2020.07.003) · PMID: [32713784](#) · PMCID: [PMC7369580](#)
1060. **New insights and long-term safety of tocilizumab in rheumatoid arthritis**
Graeme Jones, Elena Panova
Therapeutic Advances in Musculoskeletal Disease (2018-10-07)
<https://doi.org/gffsd>
DOI: [10.1177/1759720x18798462](https://doi.org/10.1177/1759720x18798462) · PMID: [30327685](#) · PMCID: [PMC6178374](#)
1061. **Tocilizumab during pregnancy and lactation: drug levels in maternal serum, cord blood, breast milk and infant serum**
Jumpei Saito, Naho Yakuwa, Kayoko Kaneko, Chinatsu Takai, Mikako Goto, Ken Nakajima, Akimasa Yamatani, Atsuko Murashima
Rheumatology (2019-08) <https://doi.org/ggzhks>
DOI: [10.1093/rheumatology/kez100](https://doi.org/10.1093/rheumatology/kez100) · PMID: [30945743](#)
1062. **Short-course tocilizumab increases risk of hepatitis B virus reactivation in patients with rheumatoid arthritis: a prospective clinical observation**
Le-Feng Chen, Ying-Qian Mo, Jun Jing, Jian-Da Ma, Dong-Hui Zheng, Lie Dai
International Journal of Rheumatic Diseases (2017-07)
<https://doi.org/f9pbc5>
DOI: [10.1111/1756-185x.13010](https://doi.org/10.1111/1756-185x.13010) · PMID: [28160426](#)
1063. **Why tocilizumab could be an effective treatment for severe COVID-19?**
Binqing Fu, Xiaoling Xu, Haiming Wei
Journal of Translational Medicine (2020-04-14) <https://doi.org/ggv5c8>
DOI: [10.1186/s12967-020-02339-3](https://doi.org/10.1186/s12967-020-02339-3) · PMID: [32290839](#) · PMCID: [PMC7154566](#)

1064. Risk of adverse events including serious infections in rheumatoid arthritis patients treated with tocilizumab: a systematic literature review and meta-analysis of randomized controlled trials

L Campbell, C Chen, SS Bhagat, RA Parker, AJK Ostor

Rheumatology (2010-11-14) <https://doi.org/crqn7c>

DOI: [10.1093/rheumatology/keq343](https://doi.org/10.1093/rheumatology/keq343) · PMID: [21078627](https://pubmed.ncbi.nlm.nih.gov/21078627/)

1065. Risk of serious infections in tocilizumab versus other biologic drugs in patients with rheumatoid arthritis: a multidatabase cohort study

Ajinkya Pawar, Rishi J Desai, Daniel H Solomon, Adrian J Santiago Ortiz, Sara Gale, Min Bao, Khaled Sarsour, Sebastian Schneeweiss, Seoyoung C Kim

Annals of the Rheumatic Diseases (2019-04) <https://doi.org/gg62hx>

DOI: [10.1136/annrheumdis-2018-214367](https://doi.org/10.1136/annrheumdis-2018-214367) · PMID: [30679153](https://pubmed.ncbi.nlm.nih.gov/30679153/)

1066. Risk of infections in rheumatoid arthritis patients treated with tocilizumab

Veronika R Lang, Matthias Englbrecht, Jürgen Rech, Hubert Nüsslein, Karin Manger, Florian Schuch, Hans-Peter Tony, Martin Fleck, Bernhard Manger, Georg Schett, Jochen Zwerina

Rheumatology (2012-05) <https://doi.org/d3b3rh>

DOI: [10.1093/rheumatology/ker223](https://doi.org/10.1093/rheumatology/ker223) · PMID: [21865281](https://pubmed.ncbi.nlm.nih.gov/21865281/)

1067. Use of Tocilizumab for COVID-19-Induced Cytokine Release Syndrome

Jared Radbel, Navaneeth Narayanan, Pinki J Bhatt

Chest (2020-07) <https://doi.org/ggtxvs>

DOI: [10.1016/j.chest.2020.04.024](https://doi.org/10.1016/j.chest.2020.04.024) · PMID: [32343968](https://pubmed.ncbi.nlm.nih.gov/32343968/) · PMCID: [PMC7195070](https://pubmed.ncbi.nlm.nih.gov/PMC7195070/)

1068. Incidence of ARDS and outcomes in hospitalized patients with COVID-19: a global literature survey

Susan J Tzotzos, Bernhard Fischer, Hendrik Fischer, Markus Zeitlinger

Critical Care (2020-08-21) <https://doi.org/gh294r>

DOI: [10.1186/s13054-020-03240-7](https://doi.org/10.1186/s13054-020-03240-7) · PMID: [32825837](https://pubmed.ncbi.nlm.nih.gov/32825837/) · PMCID: [PMC7441837](https://pubmed.ncbi.nlm.nih.gov/PMC7441837/)

1069. The Efficacy of IL-6 Inhibitor Tocilizumab in Reducing Severe COVID-19 Mortality: A Systematic Review

Avi Kaye, Robert Siegel

Cold Spring Harbor Laboratory (2020-07-14) <https://doi.org/gg62hv>

DOI: [10.1101/2020.07.10.20150938](https://doi.org/10.1101/2020.07.10.20150938)

1070. Utilizing tocilizumab for the treatment of cytokine release syndrome in COVID-19

Ali Hassoun, Elizabeth Dilip Thottacherry, Justin Muklewiecz, Qurrat-ul-Aziz, Jonathan Edwards

Journal of Clinical Virology (2020-07) <https://doi.org/ggx359>

DOI: [10.1016/j.jcv.2020.104443](https://doi.org/10.1016/j.jcv.2020.104443) · PMID: [32425661](https://pubmed.ncbi.nlm.nih.gov/32425661/) · PMCID: [PMC7229471](https://pubmed.ncbi.nlm.nih.gov/PMC7229471/)

1071. The antiviral effect of interferon-beta against SARS-CoV-2 is not mediated by MxA protein

Martin Spiegel, Andreas Pichlmair, Elke Mühlberger, Otto Haller,

Friedemann Weber

Journal of Clinical Virology (2004-07) <https://doi.org/cmc3ds>

DOI: [10.1016/j.jcv.2003.11.013](https://doi.org/10.1016/j.jcv.2003.11.013) · PMID: [15135736](#)

1072. Coronavirus virulence genes with main focus on SARS-CoV envelope gene

Marta L DeDiego, Jose L Nieto-Torres, Jose M Jimenez-Guardeño, Jose A Regla-Nava, Carlos Castaño-Rodriguez, Raul Fernandez-Delgado, Fernando Usera, Luis Enjuanes

Virus Research (2014-12) <https://doi.org/f6wm24>

DOI: [10.1016/j.virusres.2014.07.024](https://doi.org/10.1016/j.virusres.2014.07.024) · PMID: [25093995](#) · PMCID: [PMC4261026](#)

1073. Synairgen to start trial of SNG001 in COVID-19 imminently

Synairgen plc press release

(2020-03-18)

<http://synairgen.web01.hosting.bdci.co.uk/umbraco/Surface/Download/GetFile?cid=23c9b12c-508b-48c3-9081-36605c5a9ccd>

1074. Nebulised interferon beta-1a for patients with COVID-19

Nathan Peiffer-Smadja, Yazdan Yazdanpanah

The Lancet Respiratory Medicine (2021-02) <https://doi.org/ftmj>

DOI: [10.1016/s2213-2600\(20\)30523-3](https://doi.org/s2213-2600(20)30523-3) · PMID: [33189160](#) · PMCID: [PMC7833737](#)

1075. Effect of Intravenous Interferon β-1a on Death and Days Free From Mechanical Ventilation Among Patients With Moderate to Severe Acute Respiratory Distress Syndrome

VMarco Ranieri, Ville Pettilä, Matti K Karvonen, Juho Jalkanen, Peter Nightingale, David Brealey, Jordi Mancebo, Ricard Ferrer, Alain Mercat, Nicolò Patroniti, ...

JAMA (2020-02-25) <https://doi.org/ghzkww>

DOI: [10.1001/jama.2019.22525](https://doi.org/10.1001/jama.2019.22525) · PMID: [32065831](#)

1076. A Randomized Clinical Trial of the Efficacy and Safety of Interferon β-1a in Treatment of Severe COVID-19

Effat Davoudi-Monfared, Hamid Rahmani, Hossein Khalili, Mahboubeh Hajiabdolbaghi, Mohamadreza Salehi, Ladan Abbasian, Hossein Kazemzadeh, Mir Saeed Yekaninejad

Antimicrobial Agents and Chemotherapy (2020-08-20)

<https://doi.org/gg5xvm>

DOI: [10.1128/aac.01061-20](https://doi.org/10.1128/aac.01061-20) · PMID: [32661006](#) · PMCID: [PMC7449227](#)

1077. A Multicenter, Adaptive, Randomized Blinded Controlled Trial of the Safety and Efficacy of Investigational Therapeutics for the Treatment of COVID-19 in Hospitalized Adults (ACTT-3)

National Institute of Allergy and Infectious Diseases (NIAID)

clinicaltrials.gov (2022-03-09)

<https://clinicaltrials.gov/ct2/show/NCT04492475>

1078. Tocilizumab (Actemra): Adult Patients with Moderately to Severely Active Rheumatoid Arthritis

Canadian Agency for Drugs and Technologies in Health

CADTH Common Drug Reviews (2015-08)
<https://www.ncbi.nlm.nih.gov/books/NBK349513/table/T43/>

1079. **A Cost Comparison of Treatments of Moderate to Severe Psoriasis**

Cheryl Hankin, Steven Feldman, Andy Szczotka, Randolph Stinger,
Leslie Fish, David Hankin
Drug Benefit Trends (2005-05)
https://escholarship.umassmed.edu/meyers_pp/385

1080. **TNF- α inhibition for potential therapeutic modulation of SARS coronavirus infection**

Edward Tobinick
Current Medical Research and Opinion (2008-09-22)
<https://doi.org/bq4cx2>
DOI: [10.1185/030079903125002757](https://doi.org/10.1185/030079903125002757) · PMID: [14741070](#)

1081. **Sanofi and Regeneron begin global Kevzara® (sarilumab) clinical trial program in patients with severe COVID-19**

Sanofi
(2020-03-16) <http://www.news.sanofi.us/2020-03-16-Sanofi-and-Regeneron-begin-global-Kevzara-R-sarilumab-clinical-trial-program-in-patients-with-severe-COVID-19>

1082. **Sarilumab COVID-19 - Full Text View - ClinicalTrials.gov**

<https://clinicaltrials.gov/ct2/show/NCT04327388>

1083. **COVID-19: combining antiviral and anti-inflammatory treatments**

Justin Stebbing, Anne Phelan, Ivan Griffin, Catherine Tucker, Olly Oechsle, Dan Smith, Peter Richardson
The Lancet Infectious Diseases (2020-04) <https://doi.org/dph5>
DOI: [10.1016/s1473-3099\(20\)30132-8](https://doi.org/10.1016/s1473-3099(20)30132-8) · PMID: [32113509](#) · PMCID: [PMC7158903](#)

1084. **Baricitinib as potential treatment for 2019-nCoV acute respiratory disease**

Peter Richardson, Ivan Griffin, Catherine Tucker, Dan Smith, Olly Oechsle, Anne Phelan, Michael Rawling, Edward Savory, Justin Stebbing
The Lancet (2020-02) <https://doi.org/ggnrsx>
DOI: [10.1016/s0140-6736\(20\)30304-4](https://doi.org/10.1016/s0140-6736(20)30304-4) · PMID: [32032529](#) · PMCID: [PMC7137985](#)

1085. **Lilly Begins Clinical Testing of Therapies for COVID-19 | Eli Lilly and Company** <https://investor.lilly.com/news-releases/news-release-details/lilly-begins-clinical-testing-therapies-covid-19>

1086. **Baricitinib Combined With Antiviral Therapy in Symptomatic Patients Infected by COVID-19: an Open-label, Pilot Study**

Fabrizio Cantini
clinicaltrials.gov (2020-04-19)
<https://clinicaltrials.gov/ct2/show/NCT04320277>

1087. **Design and Synthesis of Hydroxyferroquine Derivatives with Antimalarial and Antiviral Activities**

Christophe Biot, Wassim Daher, Natascha Chavain, Thierry Fandeur, Jamal Khalife, Daniel Dive, Erik De Clercq

1088. An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice

Timothy P Sheahan, Amy C Sims, Shuntai Zhou, Rachel L Graham, Andrea J Pruijssers, Maria L Agostini, Sarah R Leist, Alexandra Schäfer, Kenneth H Dinnon, Laura J Stevens, ... Ralph S Baric
Science Translational Medicine (2020-04-29) <https://doi.org/ggrqd2>
DOI: [10.1126/scitranslmed.abb5883](https://doi.org/10.1126/scitranslmed.abb5883) · PMID: [32253226](https://pubmed.ncbi.nlm.nih.gov/32253226/) · PMCID: [PMC7164393](https://pubmed.ncbi.nlm.nih.gov/PMC7164393/)

1089. Antiviral Monoclonal Antibodies: Can They Be More Than Simple Neutralizing Agents?

Mireia Pelegrin, Mar Naranjo-Gomez, Marc Piechaczyk
Trends in Microbiology (2015-10) <https://doi.org/f7vzrf>
DOI: [10.1016/j.tim.2015.07.005](https://doi.org/10.1016/j.tim.2015.07.005) · PMID: [26433697](https://pubmed.ncbi.nlm.nih.gov/26433697/) · PMCID: [PMC7127033](https://pubmed.ncbi.nlm.nih.gov/PMC7127033/)

1090. Intranasal Treatment with Poly(I{middle dot}C) Protects Aged Mice from Lethal Respiratory Virus Infections

J Zhao, C Wohlford-Lenane, J Zhao, E Fleming, TE Lane, PB McCray, S Perlman
Journal of Virology (2012-08-22) <https://doi.org/f4bzfp>
DOI: [10.1128/jvi.01410-12](https://doi.org/10.1128/jvi.01410-12) · PMID: [22915814](https://pubmed.ncbi.nlm.nih.gov/22915814/) · PMCID: [PMC3486278](https://pubmed.ncbi.nlm.nih.gov/PMC3486278/)

1091. History of vaccination

Stanley Plotkin
Proceedings of the National Academy of Sciences (2014-08-18) <https://doi.org/f6fcwk>
DOI: [10.1073/pnas.1400472111](https://doi.org/10.1073/pnas.1400472111) · PMID: [25136134](https://pubmed.ncbi.nlm.nih.gov/25136134/) · PMCID: [PMC4151719](https://pubmed.ncbi.nlm.nih.gov/PMC4151719/)

1092. A strategic approach to COVID-19 vaccine R&D

Lawrence Corey, John R Mascola, Anthony S Fauci, Francis S Collins
Science (2020-05-29) <https://doi.org/ggwfck>
DOI: [10.1126/science.abc5312](https://doi.org/10.1126/science.abc5312) · PMID: [32393526](https://pubmed.ncbi.nlm.nih.gov/32393526/)

1093. Neutralizing Monoclonal Antibodies as Promising Therapeutics against Middle East Respiratory Syndrome Coronavirus Infection

Hui-Ju Han, Jian-Wei Liu, Hao Yu, Xue-Jie Yu
Viruses (2018-11-30) <https://doi.org/ggp87v>
DOI: [10.3390/v10120680](https://doi.org/10.3390/v10120680) · PMID: [30513619](https://pubmed.ncbi.nlm.nih.gov/30513619/) · PMCID: [PMC6315345](https://pubmed.ncbi.nlm.nih.gov/PMC6315345/)

1094. Developing Covid-19 Vaccines at Pandemic Speed

Nicole Lurie, Melanie Saville, Richard Hatchett, Jane Halton
New England Journal of Medicine (2020-05-21) <https://doi.org/ggg8bc>
DOI: [10.1056/nejmp2005630](https://doi.org/10.1056/nejmp2005630) · PMID: [32227757](https://pubmed.ncbi.nlm.nih.gov/32227757/)

1095. Editorial: Reverse Vaccinology

Richard Moxon, Pedro A Reche, Rino Rappuoli
Frontiers in Immunology (2019-12-03) <https://doi.org/gjjtwg>
DOI: [10.3389/fimmu.2019.02776](https://doi.org/10.3389/fimmu.2019.02776) · PMID: [31849959](https://pubmed.ncbi.nlm.nih.gov/31849959/) · PMCID: [PMC6901788](https://pubmed.ncbi.nlm.nih.gov/PMC6901788/)

1096. **Rapid response to an emerging infectious disease – Lessons learned from development of a synthetic DNA vaccine targeting Zika virus**

Sagar B Kudchodkar, Hyeree Choi, Emma L Reuschel, Rianne Esquivel, Jackie Jin-Ah Kwon, Moonsup Jeong, Joel N Maslow, Charles C Reed, Scott White, Jjoseph Kim, ... Kar Muthumani

Microbes and Infection (2018-12) <https://doi.org/gfrn5h>

DOI: [10.1016/j.micinf.2018.03.001](https://doi.org/10.1016/j.micinf.2018.03.001) · PMID: [29555345](https://pubmed.ncbi.nlm.nih.gov/29555345/) · PMCID: [PMC6593156](https://pubmed.ncbi.nlm.nih.gov/PMC6593156/)

1097. **Newer Vaccine Technologies Deployed to Develop COVID-19 Shot**

Abby Olena

The Scientist Magazine (2020-02-21) <https://www.the-scientist.com/news-opinion/newer-vaccine-technologies-deployed-to-develop-covid-19-shot-67152>

1098. **DNA vaccines: ready for prime time?**

Michele A Kutzler, David B Weiner

Nature Reviews Genetics (2008-10) <https://doi.org/fvzbws>

DOI: [10.1038/nrg2432](https://doi.org/10.1038/nrg2432) · PMID: [18781156](https://pubmed.ncbi.nlm.nih.gov/18781156/) · PMCID: [PMC4317294](https://pubmed.ncbi.nlm.nih.gov/PMC4317294/)

1099. **Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK**

Merryn Voysey, Sue Ann Costa Clemens, Shabir A Madhi, Lily Y Weckx, Pedro M Folegatti, Parvinder K Aley, Brian Angus, Vicky L Baillie, Shaun L Barnabas, Qasim E Bhorat, ... Peter Zuidewind

The Lancet (2021-01) <https://doi.org/fmq2>

DOI: [10.1016/s0140-6736\(20\)32661-1](https://doi.org/10.1016/s0140-6736(20)32661-1) · PMID: [33306989](https://pubmed.ncbi.nlm.nih.gov/33306989/) · PMCID: [PMC7723445](https://pubmed.ncbi.nlm.nih.gov/PMC7723445/)

1100. **The SARS-CoV-2 Spike Glycoprotein Biosynthesis, Structure, Function, and Antigenicity: Implications for the Design of Spike-Based Vaccine Immunogens**

Liangwei Duan, Qianqian Zheng, Hongxia Zhang, Yuna Niu, Yunwei Lou, Hui Wang

Frontiers in Immunology (2020-10-07) <https://doi.org/gjkthw>

DOI: [10.3389/fimmu.2020.576622](https://doi.org/10.3389/fimmu.2020.576622) · PMID: [33117378](https://pubmed.ncbi.nlm.nih.gov/33117378/) · PMCID: [PMC7575906](https://pubmed.ncbi.nlm.nih.gov/PMC7575906/)

1101. **SARS-CoV-2 vaccines in development**

Florian Krammer

Nature (2020-09-23) <https://doi.org/ghdprn>

DOI: [10.1038/s41586-020-2798-3](https://doi.org/10.1038/s41586-020-2798-3) · PMID: [32967006](https://pubmed.ncbi.nlm.nih.gov/32967006/)

1102. **An mRNA Vaccine against SARS-CoV-2 — Preliminary Report**

Lisa A Jackson, Evan J Anderson, Nadine G Roushanel, Paul C Roberts, Mamodikoe Makhene, Rhea N Coler, Michele P McCullough, James D Chappell, Mark R Denison, Laura J Stevens, ... John H Beigel

New England Journal of Medicine (2020-11-12) <https://doi.org/d3tt>

DOI: [10.1056/nejmoa2022483](https://doi.org/10.1056/nejmoa2022483) · PMID: [32663912](https://pubmed.ncbi.nlm.nih.gov/32663912/) · PMCID: [PMC7377258](https://pubmed.ncbi.nlm.nih.gov/PMC7377258/)

1103. **BioRender**

BioRender

<https://biorender.com/>

1104. **New Images of Novel Coronavirus SARS-CoV-2 Now Available | NIH: National Institute of Allergy and Infectious Diseases**
<https://www.niaid.nih.gov/news-events/novel-coronavirus-sarscov2-images>

1105. **Structural features of coronavirus SARS-CoV-2 spike protein: Targets for vaccination**
Ariane Sternberg, Cord Naujokat
Life Sciences (2020-09) <https://doi.org/gg4cmp>
DOI: [10.1016/j.lfs.2020.118056](https://doi.org/10.1016/j.lfs.2020.118056) · PMID: [32645344](https://pubmed.ncbi.nlm.nih.gov/32645344/) · PMCID: [PMC7336130](https://pubmed.ncbi.nlm.nih.gov/PMC7336130/)

1106. **Pre-fusion structure of a human coronavirus spike protein**
Robert N Kirchdoerfer, Christopher A Cottrell, Nianshuang Wang, Jesper Pallesen, Hadi M Yassine, Hannah L Turner, Kizzmekia S Corbett, Barney S Graham, Jason S McLellan, Andrew B Ward
Nature (2016-03) <https://doi.org/f8b8zb>
DOI: [10.1038/nature17200](https://doi.org/10.1038/nature17200) · PMID: [26935699](https://pubmed.ncbi.nlm.nih.gov/26935699/) · PMCID: [PMC4860016](https://pubmed.ncbi.nlm.nih.gov/PMC4860016/)

1107. **The Architecture of Inactivated SARS-CoV-2 with Postfusion Spikes Revealed by Cryo-EM and Cryo-ET**
Chuang Liu, Luiza Mendonça, Yang Yang, Yuanzhu Gao, Chenguang Shen, Jiwei Liu, Tao Ni, Bin Ju, Congcong Liu, Xian Tang, ... Peijun Zhang
Structure (2020-11) <https://doi.org/ghhwtg>
DOI: [10.1016/j.str.2020.10.001](https://doi.org/10.1016/j.str.2020.10.001) · PMID: [33058760](https://pubmed.ncbi.nlm.nih.gov/33058760/) · PMCID: [PMC7557167](https://pubmed.ncbi.nlm.nih.gov/PMC7557167/)

1108. **Structures and distributions of SARS-CoV-2 spike proteins on intact virions**
Zunlong Ke, Joaquin Oton, Kun Qu, Mirko Cortese, Vojtech Zila, Lesley McKeane, Takanori Nakane, Jasenko Zivanov, Christopher J Neufeldt, Berati Cerikan, ... John AG Briggs
Nature (2020-08-17) <https://doi.org/d6sf>
DOI: [10.1038/s41586-020-2665-2](https://doi.org/10.1038/s41586-020-2665-2) · PMID: [32805734](https://pubmed.ncbi.nlm.nih.gov/32805734/) · PMCID: [PMC7116492](https://pubmed.ncbi.nlm.nih.gov/PMC7116492/)

1109. **Immunogenicity and structures of a rationally designed prefusion MERS-CoV spike antigen**
Jesper Pallesen, Nianshuang Wang, Kizzmekia S Corbett, Daniel Wrapp, Robert N Kirchdoerfer, Hannah L Turner, Christopher A Cottrell, Michelle M Becker, Lingshu Wang, Wei Shi, ... Jason S McLellan
Proceedings of the National Academy of Sciences (2017-08-14) <https://doi.org/gbwk7p>
DOI: [10.1073/pnas.1707304114](https://doi.org/10.1073/pnas.1707304114) · PMID: [28807998](https://pubmed.ncbi.nlm.nih.gov/28807998/) · PMCID: [PMC5584442](https://pubmed.ncbi.nlm.nih.gov/PMC5584442/)

1110. **Structure-based design of prefusion-stabilized SARS-CoV-2 spikes**
Ching-Lin Hsieh, Jory A Goldsmith, Jeffrey M Schaub, Andrea M DiVenere, Hung-Che Kuo, Kamyab Javanmardi, Kevin C Le, Daniel Wrapp, Alison G Lee, Yutong Liu, ... Jason S McLellan
Science (2020-09-18) <https://doi.org/gg8k5r>
DOI: [10.1126/science.abd0826](https://doi.org/10.1126/science.abd0826) · PMID: [32703906](https://pubmed.ncbi.nlm.nih.gov/32703906/) · PMCID: [PMC7402631](https://pubmed.ncbi.nlm.nih.gov/PMC7402631/)

1111. **Ad26 vector-based COVID-19 vaccine encoding a prefusion-stabilized SARS-CoV-2 Spike immunogen induces potent humoral**

and cellular immune responses

Rinke Bos, Lucy Rutten, Joan EM van der Lubbe, Mark JG Bakkers, Gijs Hardenberg, Frank Wegmann, David Zuidgeest, Adriaan H de Wilde, Annemart Koornneef, Annemieke Verwilligen, ... Hanneke Schuitemaker
npj Vaccines (2020-09-28) <https://doi.org/ghjkr8>
DOI: [10.1038/s41541-020-00243-x](https://doi.org/10.1038/s41541-020-00243-x) · PMID: [33083026](https://pubmed.ncbi.nlm.nih.gov/33083026/) · PMCID: [PMC7522255](https://pubmed.ncbi.nlm.nih.gov/PMC7522255/)

1112. Towards an understanding of the adjuvant action of aluminium

Philippa Marrack, Amy S McKee, Michael W Munk
Nature Reviews Immunology (2009-04) <https://doi.org/drcwvf>
DOI: [10.1038/nri2510](https://doi.org/10.1038/nri2510) · PMID: [19247370](https://pubmed.ncbi.nlm.nih.gov/19247370/) · PMCID: [PMC3147301](https://pubmed.ncbi.nlm.nih.gov/PMC3147301/)

1113. DAMP-Inducing Adjuvant and PAMP Adjuvants Parallelly Enhance Protective Type-2 and Type-1 Immune Responses to Influenza Split Vaccination

Tomoya Hayashi, Masatoshi Momota, Etsushi Kuroda, Takato Kusakabe, Shingo Kobari, Kotaro Makisaka, Yoshitaka Ohno, Yusuke Suzuki, Fumika Nakagawa, Michelle SJ Lee, ... Hidetoshi Arima
Frontiers in Immunology (2018-11-20) <https://doi.org/gfqq89>
DOI: [10.3389/fimmu.2018.02619](https://doi.org/10.3389/fimmu.2018.02619) · PMID: [30515151](https://pubmed.ncbi.nlm.nih.gov/30515151/) · PMCID: [PMC6255964](https://pubmed.ncbi.nlm.nih.gov/PMC6255964/)

1114. Better Adjuvants for Better Vaccines: Progress in Adjuvant Delivery Systems, Modifications, and Adjuvant-Antigen Codelivery

Zhi-Biao Wang, Jing Xu
Vaccines (2020-03-13) <https://doi.org/gg35vj>
DOI: [10.3390/vaccines8010128](https://doi.org/10.3390/vaccines8010128) · PMID: [32183209](https://pubmed.ncbi.nlm.nih.gov/32183209/) · PMCID: [PMC7157724](https://pubmed.ncbi.nlm.nih.gov/PMC7157724/)

1115. Our Story

Moderna
<https://www.modernatx.com/en-US/about-us/our-story?slug=about-us%2Four-story>

1116. The COVID-19 vaccine development landscape

Tung Thanh Le, Zacharias Andreadakis, Arun Kumar, Raúl Gómez Román, Stig Tollefsen, Melanie Saville, Stephen Mayhew
Nature Reviews Drug Discovery (2020-04-09) <https://doi.org/ggrnbr>
DOI: [10.1038/d41573-020-00073-5](https://doi.org/10.1038/d41573-020-00073-5) · PMID: [32273591](https://pubmed.ncbi.nlm.nih.gov/32273591/)

1117. The history of the smallpox vaccine

Alexandra J Stewart, Phillip M Devlin
Journal of Infection (2006-05) <https://doi.org/d455hw>
DOI: [10.1016/j.jinf.2005.07.021](https://doi.org/10.1016/j.jinf.2005.07.021) · PMID: [16176833](https://pubmed.ncbi.nlm.nih.gov/16176833/)

1118. "Variolation" and Vaccination in Late Imperial China, Ca 1570–1911

Angela Ki Che Leung
History of Vaccine Development (2011) <https://doi.org/fftx2m>
DOI: [10.1007/978-1-4419-1339-5_2](https://doi.org/10.1007/978-1-4419-1339-5_2)

1119. The History Of Vaccines And Immunization: Familiar Patterns, New Challenges

Alexandra Minna Stern, Howard Markel
Health Affairs (2005-05) <https://doi.org/dzcgw5>
DOI: [10.1377/hlthaff.24.3.611](https://doi.org/10.1377/hlthaff.24.3.611) · PMID: [15886151](https://pubmed.ncbi.nlm.nih.gov/15886151/)

1120. **Equination (inoculation of horsepox): An early alternative to vaccination (inoculation of cowpox) and the potential role of horsepox virus in the origin of the smallpox vaccine**

José Esparza, Livia Schrick, Clarissa R Damaso, Andreas Nitsche
Vaccine (2017-12) <https://doi.org/gcsnbp>
DOI: [10.1016/j.vaccine.2017.11.003](https://doi.org/10.1016/j.vaccine.2017.11.003) · PMID: [29137821](https://pubmed.ncbi.nlm.nih.gov/29137821/)

1121. **Live attenuated vaccines: Historical successes and current challenges**

Philip D Minor
Virology (2015-05) <https://doi.org/f7cnmj>
DOI: [10.1016/j.virol.2015.03.032](https://doi.org/10.1016/j.virol.2015.03.032) · PMID: [25864107](https://pubmed.ncbi.nlm.nih.gov/25864107/)

1122. **Advances in mRNA Vaccines for Infectious Diseases**

Cuiling Zhang, Giulietta Maruggi, Hu Shan, Junwei Li
Frontiers in Immunology (2019-03-27) <https://doi.org/ggsnm7>
DOI: [10.3389/fimmu.2019.00594](https://doi.org/10.3389/fimmu.2019.00594) · PMID: [30972078](https://pubmed.ncbi.nlm.nih.gov/30972078/) · PMCID: [PMC6446947](https://pubmed.ncbi.nlm.nih.gov/PMC6446947/)

1123. **DNA Vaccine**

Zhengrong Cui
Non-Viral Vectors for Gene Therapy, Second Edition: Part 2 (2005)
<https://doi.org/dn299p>
DOI: [10.1016/s0065-2660\(05\)54011-2](https://doi.org/10.1016/s0065-2660(05)54011-2) · PMID: [16096015](https://pubmed.ncbi.nlm.nih.gov/16096015/) · PMCID: [PMC7119308](https://pubmed.ncbi.nlm.nih.gov/PMC7119308/)

1124. **Types of Vaccines – COVID19 Vaccine Tracker**

<https://covid19.trackvaccines.org/types-of-vaccines/>

1125. **A single-dose live-attenuated YF17D-vectored SARS-CoV-2 vaccine candidate**

Lorena Sanchez-Felipe, Thomas Vercruyse, Sapna Sharma, Ji Ma, Viktor Lemmens, Dominique Van Looveren, Mahadesh Prasad Arkalagud Javarappa, Robbert Boudewijns, Bert Malengier-Devlies, Laurens Liesenborghs, ... Kai Dallmeier
Nature (2020-12-01) <https://doi.org/ghn8jk>
DOI: [10.1038/s41586-020-3035-9](https://doi.org/10.1038/s41586-020-3035-9) · PMID: [33260195](https://pubmed.ncbi.nlm.nih.gov/33260195/)

1126. **Principles of Virology, Volume I: Molecular Biology**

Jane Flint, Anna Marie Skalka, Glenn F Rall, Vincent R Racaniello
American Society of Microbiology (2015-01-01) <https://doi.org/gmqjck>
DOI: [10.1128/9781555818951](https://doi.org/10.1128/9781555818951)

1127. **Vaccine Immunology**

Claire-Anne Siegrist
Plotkin's Vaccines (2018) <https://doi.org/gmqjcc>
DOI: [10.1016/b978-0-323-35761-6.00002-x](https://doi.org/10.1016/b978-0-323-35761-6.00002-x)

1128. **COVID-19: Coronavirus Vaccine Development Updates**

Jing Zhao, Shan Zhao, Junxian Ou, Jing Zhang, Wendong Lan, Wenyi Guan, Xiaowei Wu, Yuqian Yan, Wei Zhao, Jianguo Wu, ... Qiwei Zhang

1129. **Griffith University researchers on the road to COVID-19 vaccine**
Deborah Marshall
<https://news.griffith.edu.au/2020/04/23/griffith-university-researchers-on-the-road-to-covid-19-vaccine/>
1130. **Milken Institute's COVID-19 Treatment and Vaccine Tracker tracks the development of treatments and vaccines for COVID-19 at covid-19tracker.milkeninstitute.org #COVID19 #coronavirus #COVID19treatment #COVID19vaccine @MilkenInstitute @FirstPersonSF** <https://covid-19tracker.milkeninstitute.org/>
1131. **Scalable live-attenuated SARS-CoV-2 vaccine candidate demonstrates preclinical safety and efficacy**
Ying Wang, Chen Yang, Yutong Song, JRobert Coleman, Marcin Stawowczyk, Juliana Tafrova, Sybil Tasker, David Boltz, Robert Baker, Liliana Garcia, ... Steffen Mueller
Proceedings of the National Academy of Sciences (2021-06-30)
<https://doi.org/gmc76v>
DOI: [10.1073/pnas.2102775118](https://doi.org/10.1073/pnas.2102775118) · PMID: [34193524](#) · PMCID:
[PMC8307828](#)
1132. **A Lassa Fever Live-Attenuated Vaccine Based on Codon Deoptimization of the Viral Glycoprotein Gene**
Yingyun Cai, Chengjin Ye, Benson Cheng, Aitor Nogales, Masaharu Iwasaki, Shuiqing Yu, Kurt Cooper, David X Liu, Randy Hart, Ricky Adams, ... Luis Martínez-Sobrido
mBio (2020-02-25) <https://doi.org/gnvf9q>
DOI: [10.1128/mbio.00039-20](https://doi.org/10.1128/mbio.00039-20) · PMID: [32098811](#) · PMCID: [PMC7042690](#)
1133. **Recent Progress in Vaccine Development Against Chikungunya Virus**
Shan Gao, Siqi Song, Leiliang Zhang
Frontiers in Microbiology (2019-12-19) <https://doi.org/gh7rn6>
DOI: [10.3389/fmicb.2019.02881](https://doi.org/10.3389/fmicb.2019.02881) · PMID: [31921059](#) · PMCID:
[PMC6930866](#)
1134. **BCG-induced trained immunity: can it offer protection against COVID-19?**
Luke AJ O'Neill, Mihai G Netea
Nature Reviews Immunology (2020-05-11) <https://doi.org/ggvzp3>
DOI: [10.1038/s41577-020-0337-y](https://doi.org/10.1038/s41577-020-0337-y) · PMID: [32393823](#) · PMCID:
[PMC7212510](#)
1135. **Non-specific effects of BCG vaccine on viral infections**
SJC FM Moorlag, RJW Arts, R van Crevel, MG Netea
Clinical Microbiology and Infection (2019-12) <https://doi.org/ggq62z>
DOI: [10.1016/j.cmi.2019.04.020](https://doi.org/10.1016/j.cmi.2019.04.020) · PMID: [31055165](#)
1136. **Efficacy of BCG Vaccination Against Respiratory Tract Infections in Older Adults During the Coronavirus Disease 2019 Pandemic**

Simone JCFM Moorlag, Esther Taks, Thijs ten Doesschate, Thomas W van der Vaart, Axel B Janssen, Lisa Müller, Philipp Ostermann, Helga Dijkstra, Heidi Lemmers, Elles Simonetti, ... Mihai G Netea
Clinical Infectious Diseases (2022-03-05) <https://doi.org/gptcjh>
DOI: [10.1093/cid/ciac182](https://doi.org/10.1093/cid/ciac182) · PMID: [35247264](https://pubmed.ncbi.nlm.nih.gov/35247264/) · PMCID: [PMC8903481](https://pubmed.ncbi.nlm.nih.gov/PMC8903481/)

1137. Replicating and non-replicating viral vectors for vaccine development

Marjorie Robert-Guroff
Current Opinion in Biotechnology (2007-12) <https://doi.org/dgfz6w>
DOI: [10.1016/j.copbio.2007.10.010](https://doi.org/10.1016/j.copbio.2007.10.010) · PMID: [18063357](https://pubmed.ncbi.nlm.nih.gov/18063357/) · PMCID: [PMC2245896](https://pubmed.ncbi.nlm.nih.gov/PMC2245896/)

1138. Vaccine Types

Office of Infectious Disease and HIV/AIDS Policy (OIDP)
HHS.gov (2021-04-26)
<https://www.hhs.gov/immunization/basics/types/index.html>

1139. Vero cell technology for rapid development of inactivated whole virus vaccines for emerging viral diseases

PNoel Barrett, Sara J Terpening, Doris Snow, Ronald R Cobb, Otfried Kistner
Expert Review of Vaccines (2017-07-27) <https://doi.org/ggt7vf>
DOI: [10.1080/14760584.2017.1357471](https://doi.org/10.1080/14760584.2017.1357471) · PMID: [28724343](https://pubmed.ncbi.nlm.nih.gov/28724343/)

1140. Vaccine delivery: a matter of size, geometry, kinetics and molecular patterns

Martin F Bachmann, Gary T Jennings
Nature Reviews Immunology (2010-10-15) <https://doi.org/fg5dx9>
DOI: [10.1038/nri2868](https://doi.org/10.1038/nri2868) · PMID: [20948547](https://pubmed.ncbi.nlm.nih.gov/20948547/)

1141. Animal models and vaccines for SARS-CoV infection

Anjeanette Roberts, Elaine W Lamirande, Leatrice Vogel, Jadon P Jackson, Christopher D Paddock, Jeannette Guarner, Sherif R Zaki, Timothy Sheahan, Ralph Baric, Kanta Subbarao
Virus Research (2008-04) <https://doi.org/brrg6k>
DOI: [10.1016/j.virusres.2007.03.025](https://doi.org/10.1016/j.virusres.2007.03.025) · PMID: [17499378](https://pubmed.ncbi.nlm.nih.gov/17499378/) · PMCID: [PMC2323511](https://pubmed.ncbi.nlm.nih.gov/PMC2323511/)

1142. Functional analysis of influenza-specific helper T cell clones in vivo. T cells specific for internal viral proteins provide cognate help for B cell responses to hemagglutinin.

PA Scherle, W Gerhard
Journal of Experimental Medicine (1986-10-01) <https://doi.org/bp47qh>
DOI: [10.1084/jem.164.4.1114](https://doi.org/10.1084/jem.164.4.1114) · PMID: [2944982](https://pubmed.ncbi.nlm.nih.gov/2944982/) · PMCID: [PMC2188433](https://pubmed.ncbi.nlm.nih.gov/PMC2188433/)

1143. Severe acute respiratory syndrome (SARS) coronavirus: application of monoclonal antibodies and development of an effective vaccine

Yasuko Tsunetsugu-Yokota, Kazuo Ohnishi, Toshitada Takemori
Reviews in Medical Virology (2006) <https://doi.org/dskzwh>
DOI: [10.1002/rmv.492](https://doi.org/10.1002/rmv.492) · PMID: [16518829](https://pubmed.ncbi.nlm.nih.gov/16518829/) · PMCID: [PMC7169118](https://pubmed.ncbi.nlm.nih.gov/PMC7169118/)

1144. A subcutaneously injected UV-inactivated SARS coronavirus vaccine elicits systemic humoral immunity in mice

N Takasuka

International Immunology (2004-08-31) <https://doi.org/bkd6xq>

DOI: [10.1093/intimm/dxh143](https://doi.org/10.1093/intimm/dxh143) · PMID: [15314040](#) · PMCID: [PMC7108621](#)

1145. **Learning from the past: development of safe and effective COVID-19 vaccines**

Shan Su, Lanying Du, Shibo Jiang

Nature Reviews Microbiology (2020-10-16) <https://doi.org/ghmtgp>

DOI: [10.1038/s41579-020-00462-y](https://doi.org/10.1038/s41579-020-00462-y) · PMID: [33067570](#) · PMCID: [PMC7566580](#)

1146. **Immunodominant SARS Coronavirus Epitopes in Humans Elicited both Enhancing and Neutralizing Effects on Infection in Non-human Primates**

Qidi Wang, Lianfeng Zhang, Kazuhiko Kuwahara, Li Li, Zijie Liu, Taisheng Li, Hua Zhu, Jiangning Liu, Yanfeng Xu, Jing Xie, ... Gang Liu

ACS Infectious Diseases (2016-03-30) <https://doi.org/ggrcdk>

DOI: [10.1021/acsinfecdis.6b00006](https://doi.org/10.1021/acsinfecdis.6b00006) · PMID: [27627203](#) · PMCID: [PMC7075522](#)

1147. **Immunization with inactivated Middle East Respiratory Syndrome coronavirus vaccine leads to lung immunopathology on challenge with live virus**

Anurodh Shankar Agrawal, Xinrong Tao, Abdullah Algaissi, Tania Garron, Krishna Narayanan, Bi-Hung Peng, Robert B Couch, Chien-Te K Tseng

Human Vaccines & Immunotherapeutics (2016-06-07)

<https://doi.org/gmkb76>

DOI: [10.1080/21645515.2016.1177688](https://doi.org/10.1080/21645515.2016.1177688) · PMID: [27269431](#) · PMCID: [PMC5027702](#)

1148. **Single-Dose, Intranasal Immunization with Recombinant Parainfluenza Virus 5 Expressing Middle East Respiratory Syndrome Coronavirus (MERS-CoV) Spike Protein Protects Mice from Fatal MERS-CoV Infection**

Kun Li, Zhuo Li, Christine Wohlford-Lenane, David K Meyerholz, Rudragouda Channappanavar, Dong An, Stanley Perlman, Paul B McCray Jr., Biao He

mBio (2020-04-28) <https://doi.org/ggrzk2>

DOI: [10.1128/mbio.00554-20](https://doi.org/10.1128/mbio.00554-20) · PMID: [32265331](#) · PMCID: [PMC7157776](#)

1149. **Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine (CoronaVac) in healthy adults aged 60 years and older: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial**

Zhiwei Wu, Yaling Hu, Miao Xu, Zhen Chen, Wanqi Yang, Zhiwei Jiang, Minjie Li, Hui Jin, Guoliang Cui, Panpan Chen, ... Weidong Yin

The Lancet Infectious Diseases (2021-06) <https://doi.org/fx8z>

DOI: [10.1016/s1473-3099\(20\)30987-7](https://doi.org/10.1016/s1473-3099(20)30987-7) · PMID: [33548194](#) · PMCID: [PMC7906628](#)

1150. **Development of an inactivated vaccine candidate for SARS-CoV-2**

Qiang Gao, Linlin Bao, Haiyan Mao, Lin Wang, Kangwei Xu, Minnan Yang, Yajing Li, Ling Zhu, Nan Wang, Zhe Lv, ... Chuan Qin

Science (2020-07-03) <https://doi.org/ggvckc>

1151. **Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine in healthy adults aged 18–59 years: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial**

Yanjun Zhang, Gang Zeng, Hongxing Pan, Changgui Li, Yaling Hu, Kai Chu, Weixiao Han, Zhen Chen, Rong Tang, Weidong Yin, ... Fengcai Zhu
The Lancet Infectious Diseases (2021-02) <https://doi.org/fpcx>

DOI: [10.1016/s1473-3099\(20\)30843-4](https://doi.org/s1473-3099(20)30843-4) · PMID: [33217362](https://pubmed.ncbi.nlm.nih.gov/33217362/) · PMCID: [PMC7832443](https://pubmed.ncbi.nlm.nih.gov/PMC7832443/)

1152. **Background document on the inactivated vaccine Sinovac-CoronaVac against COVID-19** <https://www.who.int/publications-detail-redirect/WHO-2019-nCoV-vaccines-SAGE-recommendation-Sinovac-CoronaVac-background-2021.1>

1153. **Efficacy and safety of an inactivated whole-virion SARS-CoV-2 vaccine (CoronaVac): interim results of a double-blind, randomised, placebo-controlled, phase 3 trial in Turkey**

Mine Durusu Tanrıover, Hamdi Levent Doğanay, Murat Akova, Hatice Rahmet Güner, Alpay Azap, Sila Akhan, Şükran Köse, Fatma Şebnem Erdinç, Emin Halis Akalın, Ömer Fehmi Tabak, ... Kurtuluş Aksu
The Lancet (2021-07) <https://doi.org/gk898z>

DOI: [10.1016/s0140-6736\(21\)01429-x](https://doi.org/s0140-6736(21)01429-x) · PMID: [34246358](https://pubmed.ncbi.nlm.nih.gov/34246358/) · PMCID: [PMC8266301](https://pubmed.ncbi.nlm.nih.gov/PMC8266301/)

1154. **Interim report: Safety and immunogenicity of an inactivated vaccine against SARS-CoV-2 in healthy Chilean adults in a phase 3 clinical trial**

Susan M Bueno, Katia Abarca, Pablo A González, Nicolás MS Gálvez, Jorge A Soto, Luisa F Duarte, Bárbara M Schultz, Gaspar A Pacheco, Liliana A González, Yaneisi Vázquez, ... Alexis M Kalergis
Cold Spring Harbor Laboratory (2021-04-01) <https://doi.org/gmwn42>

DOI: [10.1101/2021.03.31.21254494](https://doi.org/10.1101/2021.03.31.21254494)

1155. **Coronavirus Vaccine Tracker**

Carl Zimmer, Jonathan Corum, Sui-Lee Wee, Matthew Kristoffersen
The New York Times (2020-06-10)

<https://www.nytimes.com/interactive/2020/science/coronavirus-vaccine-tracker.html>

1156. **SARS-CoV-2 vaccines strategies: a comprehensive review of phase 3 candidates**

Nikolaos C Kyriakidis, Andrés López-Cortés, Eduardo Vásconez González, Alejandra Barreto Grimaldos, Esteban Ortiz Prado
npj Vaccines (2021-02-22) <https://doi.org/gjsgc4>

DOI: [10.1038/s41541-021-00292-w](https://doi.org/s41541-021-00292-w) · PMID: [33619260](https://pubmed.ncbi.nlm.nih.gov/33619260/) · PMCID: [PMC7900244](https://pubmed.ncbi.nlm.nih.gov/PMC7900244/)

1157. **Development of an Inactivated Vaccine Candidate, BBIBP-CorV, with Potent Protection against SARS-CoV-2**

Hui Wang, Yuntao Zhang, Baoying Huang, Wei Deng, Yaru Quan, Wenling Wang, Wenbo Xu, Yuxiu Zhao, Na Li, Jin Zhang, ... Xiaoming Yang
Cell (2020-08) <https://doi.org/ghms9s>

1158. **Effect of an Inactivated Vaccine Against SARS-CoV-2 on Safety and Immunogenicity Outcomes**

Shengli Xia, Kai Duan, Yuntao Zhang, Dongyang Zhao, Huajun Zhang, Zhiqiang Xie, Xinguo Li, Cheng Peng, Yanbo Zhang, Wei Zhang, ...
Xiaoming Yang

JAMA (2020-09-08) <https://doi.org/gg72mg>

DOI: [10.1001/jama.2020.15543](https://doi.org/10.1001/jama.2020.15543) · PMID: [32789505](https://pubmed.ncbi.nlm.nih.gov/32789505/) · PMCID: [PMC7426884](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7426884/)

1159. **Immunogenicity and protective efficacy of BBV152: a whole virion inactivated SARS CoV-2 vaccine in the Syrian hamster model**

Sreelekshmy Mohandas, Pragya D Yadav, Anita Shete, Priya Abraham, Krishna Mohan, Gajanan Sapkal, Chandrashekhar Mote, Dimpal Nyayanit, Nivedita Gupta, VK Srini, ... Balram Bhargava

Research Square Platform LLC (2020-09-16) <https://doi.org/gmwn5d>

DOI: [10.21203/rs.3.rs-76768/v1](https://doi.org/10.21203/rs.3.rs-76768/v1)

1160. **Immunogenicity and protective efficacy of inactivated SARS-CoV-2 vaccine candidate, BBV152 in rhesus macaques**

Pragya D Yadav, Raches Ella, Sanjay Kumar, Dilip R Patil, Sreelekshmy Mohandas, Anita M Shete, Krishna M Vadrevu, Gaurav Bhati, Gajanan Sapkal, Himanshu Kaushal, ... Balram Bhargava

Nature Communications (2021-03-02) <https://doi.org/gmwn4c>

DOI: [10.1038/s41467-021-21639-w](https://doi.org/10.1038/s41467-021-21639-w) · PMID: [33654090](https://pubmed.ncbi.nlm.nih.gov/33654090/) · PMCID: [PMC7925524](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7925524/)

1161. **Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBV152: a double-blind, randomised, phase 1 trial**

Raches Ella, Krishna Mohan Vadrevu, Harsh Jogdand, Sai Prasad, Siddharth Reddy, Vamshi Sarangi, Brunda Ganneru, Gajanan Sapkal, Pragya Yadav, Priya Abraham, ... Balram Bhargava

The Lancet Infectious Diseases (2021-05) <https://doi.org/gkrthh>

DOI: [10.1016/s1473-3099\(20\)30942-7](https://doi.org/10.1016/s1473-3099(20)30942-7) · PMID: [33485468](https://pubmed.ncbi.nlm.nih.gov/33485468/) · PMCID: [PMC7825810](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7825810/)

1162. **Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBV152: interim results from a double-blind, randomised, multicentre, phase 2 trial, and 3-month follow-up of a double-blind, randomised phase 1 trial**

Raches Ella, Siddharth Reddy, Harsh Jogdand, Vamshi Sarangi, Brunda Ganneru, Sai Prasad, Dipankar Das, Dugyala Raju, Usha Praturi, Gajanan Sapkal, ... Krishna Mohan Vadrevu

The Lancet Infectious Diseases (2021-07) <https://doi.org/gh7597>

DOI: [10.1016/s1473-3099\(21\)00070-0](https://doi.org/10.1016/s1473-3099(21)00070-0) · PMID: [33705727](https://pubmed.ncbi.nlm.nih.gov/33705727/) · PMCID: [PMC8221739](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8221739/)

1163. **Bharat Biotech: Covaxin – COVID19 Vaccine Tracker**

<https://covid19.trackvaccines.org/vaccines/9/>

1164. **Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBIBP-CorV: a randomised, double-blind, placebo-controlled, phase 1/2 trial**

Shengli Xia, Yuntao Zhang, Yanxia Wang, Hui Wang, Yunkai Yang,
George Fu Gao, Wenjie Tan, Guizhen Wu, Miao Xu, Zhiyong Lou, ...
Xiaoming Yang
The Lancet Infectious Diseases (2021-01) <https://doi.org/ghjkrf>
DOI: [10.1016/s1473-3099\(20\)30831-8](https://doi.org/10.1016/s1473-3099(20)30831-8) · PMID: [33069281](https://pubmed.ncbi.nlm.nih.gov/33069281/) · PMCID: [PMC7561304](https://pubmed.ncbi.nlm.nih.gov/PMC7561304/)

1165. **How the Sinovac Vaccine Works**

Jonathan Corum, Carl Zimmer
The New York Times (2020-12-24)
<https://www.nytimes.com/interactive/2020/health/sinovac-covid-19-vaccine.html>

1166. **Officials Stress That the Pandemic 'Is Not Over Yet' as U.S.**

Vaccinations Begin
Karen Zraick
The New York Times (2020-12-16)
<https://www.nytimes.com/live/2020/12/16/world/covid-19-coronavirus>

1167. **Ensayo Clínico de Fase III, Aleatorio, Doble Ciego y Controlado Con Placebo Paralelo, Para Evaluar la Seguridad y la Eficacia Protectora de la Vacuna Inactivada Contra el SARS-CoV-2 en la Población Sana de 18 años o más, en Perú**

Universidad Peruana Cayetano Heredia
[clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/NCT04612972) (2021-04-27)
<https://clinicaltrials.gov/ct2/show/NCT04612972>

1168. **Effect of 2 Inactivated SARS-CoV-2 Vaccines on Symptomatic COVID-19 Infection in Adults**

Nawal Al Kaabi, Yuntao Zhang, Shengli Xia, Yunkai Yang, Manaf M Al Qahtani, Najiba Abdulrazzaq, Majed Al Nusair, Mohamed Hassany, Jaleela S Jawad, Jihad Abdalla, ... Xiaoming Yang
JAMA (2021-05-26) <https://doi.org/gj7khd>
DOI: [10.1001/jama.2021.8565](https://doi.org/10.1001/jama.2021.8565) · PMID: [34037666](https://pubmed.ncbi.nlm.nih.gov/34037666/) · PMCID: [PMC8156175](https://pubmed.ncbi.nlm.nih.gov/PMC8156175/)

1169. **Efficacy, safety, and lot to lot immunogenicity of an inactivated SARS-CoV-2 vaccine (BBV152): a, double-blind, randomised, controlled phase 3 trial**

Raches Ella, Siddarth Reddy, William Blackwelder, Varsha Potdar, Pragya Yadav, Vamshi Sarangi, Vinay Kumar Aileni, Suman Kanungo, Sanjay Rai, Prabhakar Reddy, ...
Cold Spring Harbor Laboratory (2021-07-02) <https://doi.org/gmns9m>
DOI: [10.1101/2021.06.30.21259439](https://doi.org/10.1101/2021.06.30.21259439)

1170. **Sinopharm's COVID-19 vaccine 79% effective, seeks approval in China**

Reuters
(2020-12-30) <https://www.reuters.com/article/health-coronavirus-china-vaccine-int-idUSKBN2940CA>

1171. **Brazil institute says CoronaVac efficacy above 50%, but delays full results**

Reuters
(2020-12-23) <https://www.reuters.com/article/us-health-coronavirus-sinovac-brazil-idUSKBN28X2CR>

1172. Turkey and Brazil Say Chinese Vaccine Effective, With Sparse Supporting Data

Carl Zimmer, Ernesto Londoño

The New York Times (2020-12-25)

<https://www.nytimes.com/2020/12/25/health/turkey-brazil-sinovac-coronavirus-vaccine.html>

1173. Effectiveness of an Inactivated SARS-CoV-2 Vaccine in Chile

Alejandro Jara, Eduardo A Undurraga, Cecilia González, Fabio Paredes, Tomás Fontecilla, Gonzalo Jara, Alejandra Pizarro, Johanna Acevedo, Katherinne Leo, Francisco Leon, ... Rafael Araos

New England Journal of Medicine (2021-09-02) <https://doi.org/gk475w>

DOI: [10.1056/nejmoa2107715](https://doi.org/10.1056/nejmoa2107715) · PMID: [34233097](https://pubmed.ncbi.nlm.nih.gov/34233097/) · PMCID: [PMC8279092](https://pubmed.ncbi.nlm.nih.gov/PMC8279092/)

1174. Susceptibility of Circulating SARS-CoV-2 Variants to Neutralization

Guo-Lin Wang, Zhuang-Ye Wang, Li-Jun Duan, Qing-Chuan Meng, Ming-Dong Jiang, Jing Cao, Lin Yao, Ka-Li Zhu, Wu-Chun Cao, Mai-Juan Ma

New England Journal of Medicine (2021-06-17) <https://doi.org/gjnrhz>

DOI: [10.1056/nejmc2103022](https://doi.org/10.1056/nejmc2103022) · PMID: [33822491](https://pubmed.ncbi.nlm.nih.gov/33822491/) · PMCID: [PMC8063885](https://pubmed.ncbi.nlm.nih.gov/PMC8063885/)

1175. Comparable neutralization of SARS-CoV-2 Delta AY.1 and Delta in individuals sera vaccinated with BBV152

Pragya D Yadav, Rima R Sahay, Gajanan Sapkal, Dimpal Nyayanit, Anita M Shete, Gururaj Deshpande, Deepak Y Patil, Nivedita Gupta, Sanjay Kumar, Priya Abraham, ... Balram Bhargava

Cold Spring Harbor Laboratory (2021-08-01) <https://doi.org/gmx72g>

DOI: [10.1101/2021.07.30.454511](https://doi.org/10.1101/2021.07.30.454511)

1176. Neutralization of Beta and Delta variant with sera of COVID-19 recovered cases and vaccinees of inactivated COVID-19 vaccine BBV152/Covaxin

Pragya D Yadav, Gajanan N Sapkal, Raches Ella, Rima R Sahay, Dimpal A Nyayanit, Deepak Y Patil, Gururaj Deshpande, Anita M Shete, Nivedita Gupta, VKrishna Mohan, ... Balram Bhargava

Journal of Travel Medicine (2021-07-06) <https://doi.org/gmwn4x>

DOI: [10.1093/jtm/taab104](https://doi.org/10.1093/jtm/taab104) · PMID: [34230972](https://pubmed.ncbi.nlm.nih.gov/34230972/) · PMCID: [PMC8344909](https://pubmed.ncbi.nlm.nih.gov/PMC8344909/)

1177. Inactivated COVID-19 vaccine BBV152/COVAXIN effectively neutralizes recently emerged B.1.1.7 variant of SARS-CoV-2

Gajanan N Sapkal, Pragya D Yadav, Raches Ella, Gururaj R Deshpande, Rima R Sahay, Nivedita Gupta, Krishna Mohan Vadrevu, Priya Abraham, Samiran Panda, Balram Bhargava

Journal of Travel Medicine (2021-03-27) <https://doi.org/gjs7m8>

DOI: [10.1093/jtm/taab051](https://doi.org/10.1093/jtm/taab051) · PMID: [33772577](https://pubmed.ncbi.nlm.nih.gov/33772577/) · PMCID: [PMC8083765](https://pubmed.ncbi.nlm.nih.gov/PMC8083765/)

1178. Neutralization of SARS-CoV-2 VOC 501Y.V2 by human antisera elicited by both inactivated BBIBP-CorV and recombinant dimeric RBD ZF2001 vaccines

Baoying Huang, Lianpan Dai, Hui Wang, Zhongyu Hu, Xiaoming Yang, Wenjie Tan, George F Gao

Cold Spring Harbor Laboratory (2021-02-02) <https://doi.org/gh2px7>

DOI: [10.1101/2021.02.01.429069](https://doi.org/10.1101/2021.02.01.429069)

1179. Antibody and T cell responses to Sinopharm/BBIBP-CorV in naïve and previously infected individuals in Sri Lanka

Chandima Jeewandara, Inoka Sepali Aberathna, Pradeep Darshana Pushpakumara, Achala Kamaladasa, Dinuka Guruge, Deshni Jayathilaka, Banuri Gunasekara, Shyrar Tanussiya, Heshan Kuruppu, Thushali Ranasinghe, ... Gathsaurie Neelika Malavige
Cold Spring Harbor Laboratory (2021-07-19) <https://doi.org/gk86qx>
DOI: [10.1101/2021.07.15.21260621](https://doi.org/10.1101/2021.07.15.21260621)

1180. **They Relied on Chinese Vaccines. Now They're Battling Outbreaks.**

Sui-Lee Wee
The New York Times (2021-06-22)
<https://www.nytimes.com/2021/06/22/business/economy/china-vaccines-covid-outbreak.html>

1181. **An Open-label, Phase IV Clinical Trial to Evaluate the Immunogenicity and Safety of the Inactivated SARS-CoV-2 Vaccine (Vero Cell) in Healthy Population Aged From 18 to 59 Years.**

Sinovac Research and Development Co., Ltd.
clinicaltrials.gov (2021-09-22)
<https://clinicaltrials.gov/ct2/show/NCT04962308>

1182. **Covaxin booster dose: What is it? What does govt say about this?**

Hindustan Times
(2021-07-07) <https://www.hindustantimes.com/india-news/covaxin-booster-dose-what-is-it-what-does-govt-say-about-this-101625644184446.html>

1183. **A booster dose is immunogenic and will be needed for older adults who have completed two doses vaccination with CoronaVac: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial**

Minjie Li, Juan Yang, Lin Wang, Qianhui Wu, Zhiwei Wu, Wen Zheng, Lei Wang, Wanying Lu, Xiaowei Deng, Cheng Peng, ... Weidong Yin
Cold Spring Harbor Laboratory (2021-08-08) <https://doi.org/grsh>
DOI: [10.1101/2021.08.03.21261544](https://doi.org/10.1101/2021.08.03.21261544)

1184. **Immunogenicity and safety of a third dose, and immune persistence of CoronaVac vaccine in healthy adults aged 18-59 years: interim results from a double-blind, randomized, placebo-controlled phase 2 clinical trial**

Hongxing Pan, Qianhui Wu, Gang Zeng, Juan Yang, Deyu Jiang, Xiaowei Deng, Kai Chu, Wen Zheng, Fengcai Zhu, Hongjie Yu, Weidong Yin
Cold Spring Harbor Laboratory (2021-07-25) <https://doi.org/gm2g4h>
DOI: [10.1101/2021.07.23.21261026](https://doi.org/10.1101/2021.07.23.21261026)

1185. **Robust induction of B cell and T cell responses by a third dose of inactivated SARS-CoV-2 vaccine**

Yihao Liu, Qin Zeng, Caiguanxi Deng, Mengyuan Li, Liubing Li, Dayue Liu, Ming Liu, Xinyuan Ruan, Jie Mei, Ruohui Mo, ... Haipeng Xiao
Cold Spring Harbor Laboratory (2021-09-15) <https://doi.org/gmx72k>
DOI: [10.1101/2021.09.12.21263373](https://doi.org/10.1101/2021.09.12.21263373)

1186. **China approves first mixed-vaccine trial as Delta spreads**

<https://medicalxpress.com/news/2021-08-china-mixed-vaccine-trial-delta.html>

1187. **Reactogenicidad, Seguridad e Inmunogenicidad de Dosis de Refuerzo de Vacunas Contra SARS-CoV-2 en Chile (Estudio REFUERZO)**

Rafael Araos

clinicaltrials.gov (2021-08-04)

<https://clinicaltrials.gov/ct2/show/NCT04992182>

1188. **Heterologous prime-boost immunization with CoronaVac and Convidecia**

Jingxin Li, Lihua Hou, Xiling Guo, Pengfei Jin, Shipo Wu, Jiahong Zhu, Hongxing Pan, Xue Wang, Zhizhou Song, Jingxuan Wan, ... Fengcai Zhu
Cold Spring Harbor Laboratory (2021-09-06) <https://doi.org/gmx72j>

DOI: [10.1101/2021.09.03.21263062](https://doi.org/2021.09.03.21263062)

1189. **Vaccines, new opportunities for a new society**

Rino Rappuoli, Mariagrazia Pizza, Giuseppe Del Giudice, Ennio De Gregorio

Proceedings of the National Academy of Sciences (2014-08-18)

<https://doi.org/f6fdps>

DOI: [10.1073/pnas.1402981111](https://doi.org/10.1073/pnas.1402981111) · PMID: [25136130](https://pubmed.ncbi.nlm.nih.gov/25136130/) · PMCID: [PMC4151714](https://pubmed.ncbi.nlm.nih.gov/PMC4151714/)

1190. **Virus-Like Particles, a Versatile Subunit Vaccine Platform**

Braeden Donaldson, Farah Al-Barwani, Vivienne Young, Sarah Scullion, Vernon Ward, Sarah Young

Advances in Delivery Science and Technology (2014-11-01)

<https://doi.org/gptgmx>

DOI: [10.1007/978-1-4939-1417-3_9](https://doi.org/10.1007/978-1-4939-1417-3_9) · PMCID: [PMC7121566](https://pubmed.ncbi.nlm.nih.gov/PMC7121566/)

1191. **Recombinant protein vaccines, a proven approach against coronavirus pandemics**

Jeroen Pollet, Wen-Hsiang Chen, Ulrich Strych

Advanced Drug Delivery Reviews (2021-03) <https://doi.org/gh7wss>

DOI: [10.1016/j.addr.2021.01.001](https://doi.org/10.1016/j.addr.2021.01.001) · PMID: [33421475](https://pubmed.ncbi.nlm.nih.gov/33421475/) · PMCID: [PMC7788321](https://pubmed.ncbi.nlm.nih.gov/PMC7788321/)

1192. **Virus-like particles: preparation, immunogenicity and their roles as nanovaccines and drug nanocarriers**

Saghi Nooraei, Howra Bahrulolum, Zakeh Sadat Hoseini, Camellia Katalani, Abbas Hajizade, Andrew J Easton, Gholamreza Ahmadian
Journal of Nanobiotechnology (2021-02-25) <https://doi.org/gk635z>

DOI: [10.1186/s12951-021-00806-7](https://doi.org/10.1186/s12951-021-00806-7) · PMID: [33632278](https://pubmed.ncbi.nlm.nih.gov/33632278/) · PMCID: [PMC7905985](https://pubmed.ncbi.nlm.nih.gov/PMC7905985/)

1193. **Vaccine Design**

Michael F Powell, Mark J Newman (editors)

Pharmaceutical Biotechnology (1995) <https://doi.org/gh3zp9>

DOI: [10.1007/978-1-4615-1823-5](https://doi.org/10.1007/978-1-4615-1823-5)

1194. **Virus-like particles as immunogens**

Rob Noad, Polly Roy

Trends in Microbiology (2003-09) <https://doi.org/dg35hw>

DOI: [10.1016/s0966-842x\(03\)00208-7](https://doi.org/10.1016/s0966-842x(03)00208-7)

1195. **Peptide Vaccine: Progress and Challenges**

Weidang Li, Medha Joshi, Smita Singhania, Kyle Ramsey, Ashlesh

Murthy

Vaccines (2014-07-02) <https://doi.org/gcfszb>

DOI: [10.3390/vaccines2030515](https://doi.org/10.3390/vaccines2030515) · PMID: [26344743](https://pubmed.ncbi.nlm.nih.gov/26344743/) · PMCID:

[PMC4494216](https://pubmed.ncbi.nlm.nih.gov/PMC4494216/)

1196. Role of AS04 in human papillomavirus vaccine: mode of action and clinical profile

Nathalie Garçon, Martine Wettendorff, Marcelle Van Mechelen

Expert Opinion on Biological Therapy (2011-04-04)

<https://doi.org/bvtmpk>

DOI: [10.1517/14712598.2011.573624](https://doi.org/10.1517/14712598.2011.573624) · PMID: [21457083](https://pubmed.ncbi.nlm.nih.gov/21457083/)

1197. Vaccine adjuvants: Understanding the structure and mechanism of adjuvanticity

Shuting Shi, Haoru Zhu, Xinyu Xia, Zhihui Liang, Xuehu Ma, Bingbing Sun

Vaccine (2019-05) <https://doi.org/gk6vqb>

DOI: [10.1016/j.vaccine.2019.04.055](https://doi.org/10.1016/j.vaccine.2019.04.055) · PMID: [31047671](https://pubmed.ncbi.nlm.nih.gov/31047671/)

1198. Immunogenicity of a receptor-binding domain of SARS coronavirus spike protein in mice: Implications for a subunit vaccine

Alexander N Zakhartchouk, Chetna Sharon, Malathy Satkunarajah, Thierry Auperin, Sathiyanarayanan Viswanathan, George Mutwiri, Martin Petric, Raymond H See, Robert C Brunham, BBrett Finlay, ... Lorne A Babiuk

Vaccine (2007-01) <https://doi.org/b92cpk>

DOI: [10.1016/j.vaccine.2006.06.084](https://doi.org/10.1016/j.vaccine.2006.06.084) · PMID: [16919855](https://pubmed.ncbi.nlm.nih.gov/16919855/) · PMCID: [PMC7115608](https://pubmed.ncbi.nlm.nih.gov/PMC7115608/)

1199. Receptor-binding domain of SARS-CoV spike protein induces long-term protective immunity in an animal model

Lanying Du, Guangyu Zhao, Yuxian He, Yan Guo, Bo-Jian Zheng, Shibo Jiang, Yusen Zhou

Vaccine (2007-04) <https://doi.org/drpspr>

DOI: [10.1016/j.vaccine.2006.10.031](https://doi.org/10.1016/j.vaccine.2006.10.031) · PMID: [17092615](https://pubmed.ncbi.nlm.nih.gov/17092615/) · PMCID: [PMC7115660](https://pubmed.ncbi.nlm.nih.gov/PMC7115660/)

1200. A 219-mer CHO-Expressing Receptor-Binding Domain of SARS-CoV S Protein Induces Potent Immune Responses and Protective Immunity

Lanying Du, Guangyu Zhao, Chris CS Chan, Lin Li, Yuxian He, Yusen Zhou, Bo-Jian Zheng, Shibo Jiang

Viral Immunology (2010-04) <https://doi.org/b5ghkz>

DOI: [10.1089/vim.2009.0090](https://doi.org/10.1089/vim.2009.0090) · PMID: [20374001](https://pubmed.ncbi.nlm.nih.gov/20374001/) · PMCID: [PMC2883479](https://pubmed.ncbi.nlm.nih.gov/PMC2883479/)

1201. Antigenic and Immunogenic Characterization of Recombinant Baculovirus-Expressed Severe Acute Respiratory Syndrome Coronavirus Spike Protein: Implication for Vaccine Design

Yuxian He, Jingjing Li, Susanne Heck, Sara Lustigman, Shibo Jiang

Journal of Virology (2006-06-15) <https://doi.org/bkcf55>

DOI: [10.1128/jvi.00083-06](https://doi.org/10.1128/jvi.00083-06) · PMID: [16731915](https://pubmed.ncbi.nlm.nih.gov/16731915/) · PMCID: [PMC1472569](https://pubmed.ncbi.nlm.nih.gov/PMC1472569/)

1202. **Immunogenicity and Protection Efficacy of Monomeric and Trimeric Recombinant SARS Coronavirus Spike Protein Subunit Vaccine Candidates**
Jie Li, Laura Ulitzky, Erica Silberstein, Deborah R Taylor, Raphael Viscidi
Viral Immunology (2013-04) <https://doi.org/f4tdd4>
DOI: [10.1089/vim.2012.0076](https://doi.org/10.1089/vim.2012.0076) · PMID: [23573979](https://pubmed.ncbi.nlm.nih.gov/23573979/) · PMCID: [PMC3624630](https://pubmed.ncbi.nlm.nih.gov/PMC3624630/)
1203. **Antigenicity and immunogenicity of SARS-CoV S protein receptor-binding domain stably expressed in CHO cells**
Lanying Du, Guangyu Zhao, Lin Li, Yuxian He, Yusen Zhou, Bo-Jian Zheng, Shibo Jiang
Biochemical and Biophysical Research Communications (2009-07) <https://doi.org/brx5bg>
DOI: [10.1016/j.bbrc.2009.05.003](https://doi.org/10.1016/j.bbrc.2009.05.003) · PMID: [19422787](https://pubmed.ncbi.nlm.nih.gov/19422787/) · PMCID: [PMC2750803](https://pubmed.ncbi.nlm.nih.gov/PMC2750803/)
1204. **Elicitation of Immunity in Mice After Immunization with the S2 Subunit of the Severe Acute Respiratory Syndrome Coronavirus**
Yingjun Guo, Shuhan Sun, Kaiyu Wang, Shu Zhang, Weijia Zhu, Ze Chen
DNA and Cell Biology (2005-08) <https://doi.org/bqrpd7>
DOI: [10.1089/dna.2005.24.510](https://doi.org/10.1089/dna.2005.24.510) · PMID: [16101349](https://pubmed.ncbi.nlm.nih.gov/16101349/)
1205. **Identification of Immunodominant Epitopes on the Membrane Protein of the Severe Acute Respiratory Syndrome-Associated Coronavirus**
Yuxian He, Yusen Zhou, Pamela Siddiqui, Jinkui Niu, Shibo Jiang
Journal of Clinical Microbiology (2005-08) <https://doi.org/bn4tfg>
DOI: [10.1128/jcm.43.8.3718-3726.2005](https://doi.org/10.1128/jcm.43.8.3718-3726.2005) · PMID: [16081901](https://pubmed.ncbi.nlm.nih.gov/16081901/) · PMCID: [PMC1234014](https://pubmed.ncbi.nlm.nih.gov/PMC1234014/)
1206. **Boosted expression of the SARS-CoV nucleocapsid protein in tobacco and its immunogenicity in mice**
Nuoyan Zheng, Ran Xia, Cuiping Yang, Bojiao Yin, Yin Li, Chengguo Duan, Liming Liang, Huishan Guo, Qi Xie
Vaccine (2009-08) <https://doi.org/cmwbzj>
DOI: [10.1016/j.vaccine.2009.05.073](https://doi.org/10.1016/j.vaccine.2009.05.073) · PMID: [19523911](https://pubmed.ncbi.nlm.nih.gov/19523911/) · PMCID: [PMC7115566](https://pubmed.ncbi.nlm.nih.gov/PMC7115566/)
1207. **Immunological characterizations of the nucleocapsid protein based SARS vaccine candidates**
S LIU, C LENG, S LIEN, H CHI, C HUANG, C LIN, W LIAN, C CHEN, S HSIEH, P CHONG
Vaccine (2006-04-12) <https://doi.org/crmzqd>
DOI: [10.1016/j.vaccine.2006.01.058](https://doi.org/10.1016/j.vaccine.2006.01.058) · PMID: [16494977](https://pubmed.ncbi.nlm.nih.gov/16494977/) · PMCID: [PMC7115648](https://pubmed.ncbi.nlm.nih.gov/PMC7115648/)
1208. **Subunit Vaccines Against Emerging Pathogenic Human Coronaviruses**
Ning Wang, Jian Shang, Shibo Jiang, Lanying Du
Frontiers in Microbiology (2020-02-28) <https://doi.org/ggpqxng>
DOI: [10.3389/fmicb.2020.00298](https://doi.org/10.3389/fmicb.2020.00298) · PMID: [32265848](https://pubmed.ncbi.nlm.nih.gov/32265848/) · PMCID: [PMC7105881](https://pubmed.ncbi.nlm.nih.gov/PMC7105881/)
1209. **Yeast-expressed recombinant protein of the receptor-binding domain in SARS-CoV spike protein with deglycosylated forms as a**

SARS vaccine candidate

Wen-Hsiang Chen, Lanying Du, Shivali M Chag, Cuiqing Ma, Nancy Tricoche, Xinrong Tao, Christopher A Seid, Elissa M Hudspeth, Sara Lustigman, Chien-Te K Tseng, ... Shibo Jiang

Human Vaccines & Immunotherapeutics (2013-12-30)

<https://doi.org/ghms54>

DOI: [10.4161/hv.27464](https://doi.org/10.4161/hv.27464) · PMID: [24355931](https://pubmed.ncbi.nlm.nih.gov/24355931/) · PMCID: [PMC4130269](https://pubmed.ncbi.nlm.nih.gov/PMC4130269/)

1210. Evaluation of candidate vaccine approaches for MERS-CoV

Lingshu Wang, Wei Shi, MGordon Joyce, Kayvon Modjarrad, Yi Zhang, Kwanyee Leung, Christopher R Lees, Tongqing Zhou, Hadi M Yassine, Masaru Kanekiyo, ... Barney S Graham

Nature Communications (2015-07-28) <https://doi.org/f7mqhd>

DOI: [10.1038/ncomms8712](https://doi.org/10.1038/ncomms8712) · PMID: [26218507](https://pubmed.ncbi.nlm.nih.gov/26218507/) · PMCID: [PMC4525294](https://pubmed.ncbi.nlm.nih.gov/PMC4525294/)

1211. MERS-CoV spike protein: Targets for vaccines and therapeutics

Qihui Wang, Gary Wong, Guangwen Lu, Jinghua Yan, George F Gao

Antiviral Research (2016-09) <https://doi.org/f86fvj>

DOI: [10.1016/j.antiviral.2016.07.015](https://doi.org/10.1016/j.antiviral.2016.07.015) · PMID: [27468951](https://pubmed.ncbi.nlm.nih.gov/27468951/) · PMCID: [PMC7113765](https://pubmed.ncbi.nlm.nih.gov/PMC7113765/)

1212. Recombinant Receptor Binding Domain Protein Induces Partial Protective Immunity in Rhesus Macaques Against Middle East Respiratory Syndrome Coronavirus Challenge

Jiaming Lan, Yanfeng Yao, Yao Deng, Hong Chen, Guangwen Lu, Wen Wang, Linlin Bao, Wei Deng, Qiang Wei, George F Gao, ... Wenjie Tan

EBioMedicine (2015-10) <https://doi.org/gfn5b7>

DOI: [10.1016/j.ebiom.2015.08.031](https://doi.org/10.1016/j.ebiom.2015.08.031) · PMID: [26629538](https://pubmed.ncbi.nlm.nih.gov/26629538/) · PMCID: [PMC4634622](https://pubmed.ncbi.nlm.nih.gov/PMC4634622/)

1213. Tailoring Subunit Vaccine Immunity with Adjuvant Combinations and Delivery Routes Using the Middle East Respiratory

Coronavirus (MERS-CoV) Receptor-Binding Domain as an Antigen

Jiaming Lan, Yao Deng, Hong Chen, Guangwen Lu, Wen Wang, Xiaojuan Guo, Zhuozhuang Lu, George F Gao, Wenjie Tan

PLoS ONE (2014-11-18) <https://doi.org/gh4zb6>

DOI: [10.1371/journal.pone.0112602](https://doi.org/10.1371/journal.pone.0112602) · PMID: [25405618](https://pubmed.ncbi.nlm.nih.gov/25405618/) · PMCID: [PMC4236105](https://pubmed.ncbi.nlm.nih.gov/PMC4236105/)

1214. Engineering a stable CHO cell line for the expression of a MERS-coronavirus vaccine antigen

Mun Peak Nyon, Lanying Du, Chien-Te Kent Tseng, Christopher A Seid, Jeroen Pollet, Kevin S Naceanceno, Anurodh Agrawal, Abdullah Algaissi, Bi-Hung Peng, Wanbo Tai, ... Peter J Hotez

Vaccine (2018-03) <https://doi.org/gdd62m>

DOI: [10.1016/j.vaccine.2018.02.065](https://doi.org/10.1016/j.vaccine.2018.02.065) · PMID: [29496347](https://pubmed.ncbi.nlm.nih.gov/29496347/) · PMCID: [PMC5860679](https://pubmed.ncbi.nlm.nih.gov/PMC5860679/)

1215. A recombinant receptor-binding domain of MERS-CoV in trimeric form protects human dipeptidyl peptidase 4 (hDPP4) transgenic mice from MERS-CoV infection

Wanbo Tai, Guangyu Zhao, Shihun Sun, Yan Guo, Yufei Wang, Xinrong Tao, Chien-Te K Tseng, Fang Li, Shibo Jiang, Lanying Du, Yusen Zhou

Virology (2016-12) <https://doi.org/f9c5sn>

1216. **The Amino Acids 736–761 of the MERS-CoV Spike Protein Induce Neutralizing Antibodies: Implications for the Development of Vaccines and Antiviral Agents**

Yang Yang, Yao Deng, Bo Wen, Huijuan Wang, Xin Meng, Jiaming Lan, George F Gao, Wenjie Tan

Viral Immunology (2014-12) <https://doi.org/f6rjbb>

DOI: [10.1089/vim.2014.0080](https://doi.org/10.1089/vim.2014.0080) · PMID: [25387086](#) · PMCID: [PMC4259179](#)

1217. **The recombinant N-terminal domain of spike proteins is a potential vaccine against Middle East respiratory syndrome coronavirus (MERS-CoV) infection**

Lan Jiaming, Yao Yanfeng, Deng Yao, Hu Yawei, Bao Linlin, Huang Baoying, Yan Jinghua, George F Gao, Qin Chuan, Tan Wenjie

Vaccine (2017-01) <https://doi.org/f9htwb>

DOI: [10.1016/j.vaccine.2016.11.064](https://doi.org/10.1016/j.vaccine.2016.11.064) · PMID: [27899228](#) · PMCID: [PMC7115548](#)

1218. **Vaccines based on virus-like nano-particles for use against Middle East Respiratory Syndrome (MERS) coronavirus**

Alireza Hashemzadeh, Amir Avan, Gordon A Ferns, Majid Khazaei

Vaccine (2020-08) <https://doi.org/gg4rkj>

DOI: [10.1016/j.vaccine.2020.07.003](https://doi.org/10.1016/j.vaccine.2020.07.003) · PMID: [32684497](#) · PMCID: [PMC7837099](#)

1219. **Purified coronavirus spike protein nanoparticles induce coronavirus neutralizing antibodies in mice**

Christopher M Coleman, Ye V Liu, Haiyan Mu, Justin K Taylor, Michael Massare, David C Flyer, Gregory M Glenn, Gale E Smith, Matthew B Frieman

Vaccine (2014-05) <https://doi.org/f2rn4w>

DOI: [10.1016/j.vaccine.2014.04.016](https://doi.org/10.1016/j.vaccine.2014.04.016) · PMID: [24736006](#) · PMCID: [PMC4058772](#)

1220. **MERS-CoV virus-like particles produced in insect cells induce specific humoral and cellular immunity in rhesus macaques**

Chong Wang, Xuexing Zheng, Weiwei Gai, Yongkun Zhao, Hualei Wang, Haijun Wang, Na Feng, Hang Chi, Boning Qiu, Nan Li, ... Xianzhu Xia

Oncotarget (2016-03-30) <https://doi.org/f92z2j>

DOI: [10.18632/oncotarget.8475](https://doi.org/10.18632/oncotarget.8475) · PMID: [27050368](#) · PMCID: [PMC5355045](#)

1221. **Significant Spike-Specific IgG and Neutralizing Antibodies in Mice Induced by a Novel Chimeric Virus-Like Particle Vaccine Candidate for Middle East Respiratory Syndrome Coronavirus**

Jiaming Lan, Yao Deng, Jingdong Song, Baoying Huang, Wenling Wang, Wenjie Tan

Virologica Sinica (2018-10) <https://doi.org/gpmnq7>

DOI: [10.1007/s12250-018-0064-8](https://doi.org/10.1007/s12250-018-0064-8) · PMID: [30374826](#) · PMCID: [PMC6235757](#)

1222. **Coronavirus vaccine development: from SARS and MERS to COVID-19**

Yen-Der Li, Wei-Yu Chi, Jun-Han Su, Louise Ferrall, Chien-Fu Hung, T-C

Wu

Journal of Biomedical Science (2020-12) <https://doi.org/gmf6bk>

DOI: [10.1186/s12929-020-00695-2](https://doi.org/s12929-020-00695-2) · PMID: [33341119](https://pubmed.ncbi.nlm.nih.gov/33341119/) · PMCID:

[PMC7749790](https://pubmed.ncbi.nlm.nih.gov/PMC7749790/)

1223. Production of recombinant subunit vaccines: protein immunogens, live delivery systems and nucleic acid vaccines

Sissela Liljeqvist, Stefan Ståhl

Journal of Biotechnology (1999-07) <https://doi.org/d4g86c>

DOI: [10.1016/s0168-1656\(99\)00107-8](https://doi.org/10.1016/s0168-1656(99)00107-8)

1224. Acellular Pertussis Vaccines and Pertussis Resurgence: Revise or Replace?

Clara Maria Ausiello, Antonio Cassone

mBio (2014-07) <https://doi.org/ggi6mm>

DOI: [10.1128/mbio.01339-14](https://doi.org/10.1128/mbio.01339-14) · PMID: [24917600](https://pubmed.ncbi.nlm.nih.gov/24917600/) · PMCID: [PMC4056554](https://pubmed.ncbi.nlm.nih.gov/PMC4056554/)

1225. Pertussis: Challenges Today and for the Future

James D Cherry

PLoS Pathogens (2013-07-25) <https://doi.org/gg74fv>

DOI: [10.1371/journal.ppat.1003418](https://doi.org/10.1371/journal.ppat.1003418) · PMID: [23935481](https://pubmed.ncbi.nlm.nih.gov/23935481/) · PMCID:

[PMC3723573](https://pubmed.ncbi.nlm.nih.gov/PMC3723573/)

1226. Advancements in the development of subunit influenza vaccines

Naru Zhang, Bo-Jian Zheng, Lu Lu, Yusen Zhou, Shibo Jiang, Lanying Du

Microbes and Infection (2015-02) <https://doi.org/gngp52>

DOI: [10.1016/j.micinf.2014.12.006](https://doi.org/10.1016/j.micinf.2014.12.006) · PMID: [25529753](https://pubmed.ncbi.nlm.nih.gov/25529753/) · PMCID:

[PMC4336774](https://pubmed.ncbi.nlm.nih.gov/PMC4336774/)

1227. Vaccines – COVID19 Vaccine Tracker

<https://covid19.trackvaccines.org/vaccines/>

1228. Recent advances in the production of recombinant subunit vaccines in *Pichia pastoris*

Man Wang, Shuai Jiang, Yefu Wang

Bioengineered (2016-04-08) <https://doi.org/ghqkt8>

DOI: [10.1080/21655979.2016.1191707](https://doi.org/10.1080/21655979.2016.1191707) · PMID: [27246656](https://pubmed.ncbi.nlm.nih.gov/27246656/) · PMCID:

[PMC4927204](https://pubmed.ncbi.nlm.nih.gov/PMC4927204/)

1229. Coronavirus Pandemic (COVID-19)

Hannah Ritchie, Edouard Mathieu, Lucas Rodés-Guirao, Cameron Appel, Charlie Giattino, Esteban Ortiz-Ospina, Joe Hasell, Bobbie Macdonald, Diana Beltekian, Max Roser

Our World in Data (2020-03-05) <https://ourworldindata.org/covid-vaccinations>

1230. SARS-CoV-2 spike glycoprotein vaccine candidate NVX-CoV2373 immunogenicity in baboons and protection in mice

Jing-Hui Tian, Nita Patel, Robert Haupt, Haixia Zhou, Stuart Weston, Holly Hammond, James Logue, Alyse D Portnoff, James Norton, Mimi Guebre-Xabier, ... Gale Smith

Nature Communications (2021-01-14) <https://doi.org/gjh782>

DOI: [10.1038/s41467-020-20653-8](https://doi.org/10.1038/s41467-020-20653-8) · PMID: [33446655](https://pubmed.ncbi.nlm.nih.gov/33446655/) · PMCID:

[PMC7809486](https://pubmed.ncbi.nlm.nih.gov/PMC7809486/)

1231. The Coming Age of Insect Cells for Manufacturing and Development of Protein Therapeutics

Christine M Yee, Andrew J Zak, Brett D Hill, Fei Wen

Industrial & Engineering Chemistry Research (2018-07-09)

<https://doi.org/gd332h>

DOI: [10.1021/acs.iecr.8b00985](https://doi.org/10.1021/acs.iecr.8b00985) · PMID: [30886455](https://pubmed.ncbi.nlm.nih.gov/30886455/) · PMCID:

[PMC6420222](https://pubmed.ncbi.nlm.nih.gov/PMC6420222/)

1232. Phase 1–2 Trial of a SARS-CoV-2 Recombinant Spike Protein Nanoparticle Vaccine

Cheryl Keech, Gary Albert, Iksung Cho, Andreana Robertson, Patricia Reed, Susan Neal, Joyce S Plested, Mingzhu Zhu, Shane Cloney-Clark, Haixia Zhou, ... Gregory M Glenn

New England Journal of Medicine (2020-12-10) <https://doi.org/gg9q7d>

DOI: [10.1056/nejmoa2026920](https://doi.org/10.1056/nejmoa2026920) · PMID: [32877576](https://pubmed.ncbi.nlm.nih.gov/32877576/) · PMCID: [PMC7494251](https://pubmed.ncbi.nlm.nih.gov/PMC7494251/)

1233. Safety and Efficacy of NVX-CoV2373 Covid-19 Vaccine

Paul T Heath, Eva P Galiza, David N Baxter, Marta Boffito, Duncan Browne, Fiona Burns, David R Chadwick, Rebecca Clark, Catherine Cosgrove, James Galloway, ... Seth Toback

New England Journal of Medicine (2021-09-23) <https://doi.org/gk3zvz>

DOI: [10.1056/nejmoa2107659](https://doi.org/10.1056/nejmoa2107659) · PMID: [34192426](https://pubmed.ncbi.nlm.nih.gov/34192426/) · PMCID: [PMC8262625](https://pubmed.ncbi.nlm.nih.gov/PMC8262625/)

1234. Novavax COVID-19 vaccine Nuvaxovid approved by MHRA

GOV.UK

<https://www.gov.uk/government/news/novavax-covid-19-vaccine-nuvaxovid-approved-by-mhra>

1235. EMA recommends Nuvaxovid for authorisation in the EU

EMA

European Medicines Agency (2021-12-20)

<https://www.ema.europa.eu/en/news/ema-recommends-nuvaxovid-authorisation-eu>

1236. Novavax Submits Request to the U.S. FDA for Emergency Use Authorization of COVID-19 Vaccine

Novavax Investor Relations

<https://ir.novavax.com/2022-01-31-Novavax-Submits-Request-to-the-U-S-FDA-for-Emergency-Use-Authorization-of-COVID-19-Vaccine>

1237. 'They rushed the process': Vaccine maker's woes hamper global inoculation campaign

POLITICO

<https://www.politico.com/news/2021/10/19/novavax-vaccine-rush-process-global-campaign-516298>

1238. Why is WHO pushing back on a Health Canada-approved Medicago SARS-CoV-2 vaccine?

Diana Duong, Lauren Vogel

Canadian Medical Association Journal (2022-04-03)

<https://doi.org/gp37s9>

DOI: [10.1503/cmaj.1095992](https://doi.org/10.1503/cmaj.1095992) · PMID: [35379666](https://pubmed.ncbi.nlm.nih.gov/35379666/) · PMCID: [PMC8985905](https://pubmed.ncbi.nlm.nih.gov/PMC8985905/)

1239. Covifenz

Medicago

<https://medicago.com/en/our-products/our-vaccines/covifenz-covid-19-vlp-vaccine/>

1240. **Efficacy, immunogenicity, and safety of a plant-derived, quadrivalent, virus-like particle influenza vaccine in adults (18–64 years) and older adults (≥65 years): two multicentre, randomised phase 3 trials**

Brian J Ward, Alexander Makarkov, Annie Séguin, Stéphane Pillet, Sonia Trépanier, Jiwanjeet Dhaliwall, Michael D Libman, Timo Vesikari, Nathalie Landry

The Lancet (2020-11) <https://doi.org/gjn28x>

DOI: [10.1016/s0140-6736\(20\)32014-6](https://doi.org/10.1016/s0140-6736(20)32014-6)

1241. **Phase 1 randomized trial of a plant-derived virus-like particle vaccine for COVID-19**

Brian J Ward, Philipe Gobeil, Annie Séguin, Judith Atkins, Iohann Boulay, Pierre-Yves Charbonneau, Manon Couture, Marc-André D'Aoust, Jiwanjeet Dhaliwall, Carolyn Finkle, ... Nathalie Landry

Nature Medicine (2021-05-18) <https://doi.org/gm7br3>

DOI: [10.1038/s41591-021-01370-1](https://doi.org/s41591-021-01370-1) · PMID: [34007070](#) · PMCID:

[PMC8205852](#)

1242. [10.1056/NEJMoa2201300](https://doi.org/10.1056/NEJMoa2201300)

1243. **Medicago and GSK announce the approval by Health Canada of COVIFENZ®, an adjuvanted plant-based COVID-19 vaccine | GSK**

<https://www.gsk.com/en-gb/media/press-releases/medicago-and-gsk-announce-the-approval-by-health-canada-of-covifenz/>

1244. **Influenza virus-like particles produced by transient expression in *Nicotiana benthamiana* induce a protective immune response against a lethal viral challenge in mice**

Marc-André D'Aoust, Pierre-Olivier Lavoie, Manon M-J Couture, Sonia Trépanier, Jean-Martin Guay, Michèle Dargis, Sébastien Mongrand, Nathalie Landry, Brian J Ward, Louis-P Vézina

Plant Biotechnology Journal (2008-12) <https://doi.org/b8xqkk>

DOI: [10.1111/j.1467-7652.2008.00384.x](https://doi.org/10.1111/j.1467-7652.2008.00384.x) · PMID: [19076615](#)

1245. **Covid-19: WHO set to reject Canadian plant based vaccine because of links with tobacco industry**

Owen Dyer

BMJ (2022-03-28) <https://doi.org/gp37s8>

DOI: [10.1136/bmj.o811](https://doi.org/10.1136/bmj.o811) · PMID: [35346968](#)

1246. **Plant-derived virus-like particles as vaccines**

Qiang Chen, Huafang Lai

Human Vaccines & Immunotherapeutics (2013-01)

<https://doi.org/f4rrfj>

DOI: [10.4161/hv.22218](https://doi.org/10.4161/hv.22218) · PMID: [22995837](#) · PMCID: [PMC3667944](#)

1247. **Efficacy and Safety of NVX-CoV2373 in Adults in the United States and Mexico**

Lisa M Dunkle, Karen L Kotloff, Cynthia L Gay, Germán Áñez, Jeffrey M Adelglass, Alejandro Q Barrat Hernández, Wayne L Harper, Daniel M Duncanson, Monica A McArthur, Diana F Florescu, ...

1248. **Novavax Statement on UK and Mexico Phase 3 Clinical Trial Participants Considered Fully Vaccinated in the US**

Novavax Investor Relations

<https://ir.novavax.com/Novavax-Statement-on-UK-and-Mexico-Phase-3-Clinical-Trial-Participants-Considered-Fully-Vaccinated-in-the-US>

1249. **Efficacy and Safety of a Recombinant Plant-Based Adjuvanted Covid-19 Vaccine**

Karen J Hager, Gonzalo Pérez Marc, Philipe Gobeil, Ricardo S Diaz, Gretchen Heizer, Conrado Llapur, Alexander I Makarkov, Eduardo Vasconcellos, Stéphane Pillet, Fernando Riera, ... Brian J Ward

New England Journal of Medicine (2022-05-04) <https://doi.org/gp3zrx>

DOI: [10.1056/nejmoa2201300](https://doi.org/10.1056/nejmoa2201300) · PMID: [35507508](#)

1250. **Efficacy of NVX-CoV2373 Covid-19 Vaccine against the B.1.351 Variant**

Vivek Shinde, Sutika Bhikha, Zaheer Hoosain, Moherndran Archary, Qasim Bhorat, Lee Fairlie, Umesh Laloo, Mduduzi SL Masilela, Dhayendre Moodley, Sherika Hanley, ... Shabir A Madhi

New England Journal of Medicine (2021-05-20) <https://doi.org/gjzcxc>

DOI: [10.1056/nejmoa2103055](https://doi.org/10.1056/nejmoa2103055) · PMID: [33951374](#) · PMCID: [PMC8091623](#)

1251. **Towards Vaccine 3.0: new era opened in vaccine research and industry**

Joon Haeng Rhee

Clinical and Experimental Vaccine Research (2014)

<https://doi.org/gmqm4>

DOI: [10.7774/cevr.2014.3.1.1](https://doi.org/10.7774/cevr.2014.3.1.1) · PMID: [24427757](#) · PMCID: [PMC3890443](#)

1252. **Developing vaccines in the era of genomics: a decade of reverse vaccinology**

KL Seib, X Zhao, R Rappuoli

Clinical Microbiology and Infection (2012-10) <https://doi.org/gkbn9x>

DOI: [10.1111/j.1469-0691.2012.03939.x](https://doi.org/10.1111/j.1469-0691.2012.03939.x) · PMID: [22882709](#)

1253. **A Comparison of Plasmid DNA and mRNA as Vaccine Technologies**

Liu

Vaccines (2019-04-24) <https://doi.org/ggwd7r>

DOI: [10.3390/vaccines7020037](https://doi.org/10.3390/vaccines7020037) · PMID: [31022829](#) · PMCID: [PMC6631684](#)

1254. **New Vaccine Technologies**

Ronald W Ellis

JAMA: The Journal of the American Medical Association (1994-03-23)

<https://doi.org/b8gn86>

DOI: [10.1001/jama.1994.03510360055036](https://doi.org/10.1001/jama.1994.03510360055036)

1255. **DNA vaccines: a review**

MA Liu

Journal of Internal Medicine (2003-04) <https://doi.org/c9z766>

DOI: [10.1046/j.1365-2796.2003.01140.x](https://doi.org/10.1046/j.1365-2796.2003.01140.x) · PMID: [12653868](#)

1256. **Plasmid DNA vaccines: where are we now?**

F Ghaffarifar

Drugs of Today (2018) <https://doi.org/gdsqgg>

DOI: [10.1358/dot.2018.54.5.2807864](https://doi.org/10.1358/dot.2018.54.5.2807864) · PMID: [29911696](#)

1257. **Recent innovations in mRNA vaccines**

Jeffrey B Ulmer, Andrew J Geall

Current Opinion in Immunology (2016-08) <https://doi.org/f82bgg>

DOI: [10.1016/j.coি.2016.05.008](https://doi.org/10.1016/j.coि.2016.05.008) · PMID: [27240054](#)

1258. **DNA Vaccines—How Far From Clinical Use?**

Dominika Hobernik, Matthias Bros

International Journal of Molecular Sciences (2018-11-15)

<https://doi.org/gmqmg3>

DOI: [10.3390/ijms19113605](https://doi.org/10.3390/ijms19113605) · PMID: [30445702](#) · PMCID: [PMC6274812](#)

1259. **Engineered Nanodelivery Systems to Improve DNA Vaccine Technologies**

Michael Lim, Abu Zayed Md Badruddoza, Jannatul Firdous, Mohammad Azad, Adnan Mannan, Taslim Ahmed Al-Hilal, Chong-Su Cho, Mohammad Ariful Islam

Pharmaceutics (2020-01-01) <https://doi.org/ghwmkd>

DOI: [10.3390/pharmaceutics12010030](https://doi.org/10.3390/pharmaceutics12010030) · PMID: [31906277](#) · PMCID: [PMC7022884](#)

1260. **DNA** <https://www.who.int/teams/health-product-and-policy-standards/standards-and-specifications/vaccines-quality/dna>

1261. **Safety, Tolerability and Immunogenicity of INO-4800 for COVID-19 in Healthy Volunteers - Full Text View - ClinicalTrials.gov**

<https://clinicaltrials.gov/ct2/show/NCT04336410>

1262. **Electroporation delivery of DNA vaccines: prospects for success**

Niranjan Y Sardesai, David B Weiner

Current Opinion in Immunology (2011-06) <https://doi.org/cq8b4p>

DOI: [10.1016/j.coি.2011.03.008](https://doi.org/10.1016/j.coি.2011.03.008) · PMID: [21530212](#) · PMCID: [PMC3109217](#)

1263. **Tolerability of intramuscular and intradermal delivery by CELLECTRA® adaptive constant current electroporation device in healthy volunteers**

Malissa C Diehl, Jessica C Lee, Stephen E Daniels, Pablo Tebas, Amir S Khan, Mary Giffear, Niranjan Y Sardesai, Mark L Bagarazzi

Human Vaccines & Immunotherapeutics (2014-10-27)

<https://doi.org/ggrj7h>

DOI: [10.4161/hv.24702](https://doi.org/10.4161/hv.24702) · PMID: [24051434](#) · PMCID: [PMC3906411](#)

1264. **Multivalent and Multipathogen Viral Vector Vaccines**

Katharina B Lauer, Ray Borrow, Thomas J Blanchard

Clinical and Vaccine Immunology (2017-01) <https://doi.org/f9tsw2>

DOI: [10.1128/cvi.00298-16](https://doi.org/10.1128/cvi.00298-16) · PMID: [27535837](#) · PMCID: [PMC5216423](#)

1265. **Viral vectors as vaccine platforms: from immunogenicity to impact**

Katie J Ewer, Teresa Lambe, Christine S Rollier, Alexandra J Spencer, Adrian VS Hill, Lucy Dorrell
Current Opinion in Immunology (2016-08) <https://doi.org/f82tb6>
DOI: [10.1016/j.coi.2016.05.014](https://doi.org/10.1016/j.coi.2016.05.014) · PMID: [27286566](#)

1266. Clinical Assessment of a Novel Recombinant Simian Adenovirus ChAdOx1 as a Vectored Vaccine Expressing Conserved Influenza A Antigens

Richard D Antrobus, Lynda Coughlan, Tamara K Berthoud, Matthew D Dicks, Adrian VS Hill, Teresa Lambe, Sarah C Gilbert
Molecular Therapy (2014-03) <https://doi.org/f5vhv3>
DOI: [10.1038/mt.2013.284](https://doi.org/10.1038/mt.2013.284) · PMID: [24374965](#) · PMCID: [PMC3944330](#)

1267. Vaccine Candidates against Coronavirus Infections. Where Does COVID-19 Stand?

Jawad Al-Kassmy, Jannie Pedersen, Gary Kobinger
Viruses (2020-08-07) <https://doi.org/ghsfmc>
DOI: [10.3390/v12080861](https://doi.org/10.3390/v12080861) · PMID: [32784685](#) · PMCID: [PMC7472384](#)

1268. Poxviruses as vaccine vectors

P-P Pastoret, A Vanderplasschen
Comparative Immunology, Microbiology and Infectious Diseases (2003-10) <https://doi.org/cnw6vw>
DOI: [10.1016/s0147-9571\(03\)00019-5](https://doi.org/10.1016/s0147-9571(03)00019-5)

1269. Enhancing poxvirus vectors vaccine immunogenicity

Juan García-Arriaza, Mariano Esteban
Human Vaccines & Immunotherapeutics (2014-05-05)
<https://doi.org/ghz9tw>
DOI: [10.4161/hv.28974](https://doi.org/10.4161/hv.28974) · PMID: [25424927](#) · PMCID: [PMC4896794](#)

1270. New Insights on Adenovirus as Vaccine Vectors

Marcio O Lasaro, Hildegund CJ Ertl
Molecular Therapy (2009-08) <https://doi.org/dcz549>
DOI: [10.1038/mt.2009.130](https://doi.org/10.1038/mt.2009.130) · PMID: [19513019](#) · PMCID: [PMC2835230](#)

1271. Attenuated vesicular stomatitis viruses as vaccine vectors.

A Roberts, L Buonocore, R Price, J Forman, JK Rose
Journal of virology (1999-05)
<https://www.ncbi.nlm.nih.gov/pubmed/10196265>
DOI: [10.1128/jvi.73.5.3723-3732.1999](https://doi.org/10.1128/jvi.73.5.3723-3732.1999) · PMID: [10196265](#) · PMCID:
[PMC104148](#)

1272. Vesicular stomatitis virus: re-inventing the bullet

Brian D Lichtry, Anthony T Power, David F Stojdl, John C Bell
Trends in Molecular Medicine (2004-05) <https://doi.org/fg6wv5>
DOI: [10.1016/j.molmed.2004.03.003](https://doi.org/10.1016/j.molmed.2004.03.003) · PMID: [15121047](#)

1273. Viral vectors as vaccine platforms: deployment in sight

Christine S Rollier, Arturo Reyes-Sandoval, Matthew G Cottingham, Katie Ewer, Adrian VS Hill
Current Opinion in Immunology (2011-06) <https://doi.org/d8p72q>
DOI: [10.1016/j.coi.2011.03.006](https://doi.org/10.1016/j.coi.2011.03.006) · PMID: [21514130](#)

1274. Progress and prospects: immune responses to viral vectors

S Nayak, RW Herzog
Gene Therapy (2009-11-12) <https://doi.org/ctbtwq>
DOI: [10.1038/gt.2009.148](https://doi.org/10.1038/gt.2009.148) · PMID: [19907498](#) · PMCID: [PMC3044498](#)

1275. Developments in Viral Vector-Based Vaccines

Takehiro Ura, Kenji Okuda, Masaru Shimada
Vaccines (2014-07-29) <https://doi.org/gcfnx9>
DOI: [10.3390/vaccines2030624](https://doi.org/10.3390/vaccines2030624) · PMID: [26344749](#) · PMCID: [PMC4494222](#)

1276. Development and Applications of Viral Vectored Vaccines to Combat Zoonotic and Emerging Public Health Threats

Sophia M Vrba, Natalie M Kirk, Morgan E Brisse, Yuying Liang, Hinh Ly
Vaccines (2020-11-13) <https://doi.org/gh23ww>
DOI: [10.3390/vaccines8040680](https://doi.org/10.3390/vaccines8040680) · PMID: [33202961](#) · PMCID: [PMC7712223](#)

1277. A Vaccine against Ebola Virus

Erica Ollmann Saphire
Cell (2020-04) <https://doi.org/gprj4w>
DOI: [10.1016/j.cell.2020.03.011](https://doi.org/10.1016/j.cell.2020.03.011) · PMID: [32243796](#)

1278. Viral Vector Malaria Vaccines Induce High-Level T Cell and Antibody Responses in West African Children and Infants

Carly M Bliss, Abdoulie Drammeh, Georgina Bowyer, Guillaume S Sanou, Ya Jankey Jagne, Oumarou Ouedraogo, Nick J Edwards, Casimir Tarama, Nicolas Ouedraogo, Mireille Ouedraogo, ... Katie J Ewer
Molecular Therapy (2017-02) <https://doi.org/f9xwv3>
DOI: [10.1016/j.ymthe.2016.11.003](https://doi.org/10.1016/j.ymthe.2016.11.003) · PMID: [28153101](#) · PMCID: [PMC5368405](#)

1279. Viral vectors for malaria vaccine development

Shengqiang Li, Emily Locke, Joseph Bruder, David Clarke, Denise L Doolan, Menzo JE Havenga, Adrian VS Hill, Peter Liljestrom, Thomas P Monath, Hussein Y Naim, ... Filip Dubovsky
Vaccine (2007-03) <https://doi.org/fh9fn6>
DOI: [10.1016/j.vaccine.2006.07.035](https://doi.org/10.1016/j.vaccine.2006.07.035) · PMID: [16914237](#) · PMCID: [PMC7131149](#)

1280. Chimpanzee Adenovirus Vector Ebola Vaccine

Julie E Ledgerwood, Adam D DeZure, Daphne A Stanley, Emily E Coates, Laura Novik, Mary E Enama, Nina M Berkowitz, Zonghui Hu, Gyan Joshi, Aurélie Ploquin, ... Barney S Graham
New England Journal of Medicine (2017-03-09) <https://doi.org/xdr>
DOI: [10.1056/nejmoa1410863](https://doi.org/10.1056/nejmoa1410863) · PMID: [25426834](#)

1281. Recombinant Vesicular Stomatitis Virus-Based Vaccines Against Ebola and Marburg Virus Infections

Thomas W Geisbert, Heinz Feldmann
The Journal of Infectious Diseases (2011-11) <https://doi.org/fcvgxq>
DOI: [10.1093/infdis/jir349](https://doi.org/10.1093/infdis/jir349) · PMID: [21987744](#) · PMCID: [PMC3218670](#)

1282. Ebola virus vaccines: an overview of current approaches

Andrea Marzi, Heinz Feldmann
Expert Review of Vaccines (2014-02-27) <https://doi.org/f52bn6>

1283. **Development of replication-competent viral vectors for HIV vaccine delivery**

Christopher L Parks, Louis J Picker, CRichter King

Current Opinion in HIV and AIDS (2013-09) <https://doi.org/f5b5qm>

DOI: [10.1097/coh.0b013e328363d389](https://doi.org/10.1097/coh.0b013e328363d389) · PMID: [23925000](https://pubmed.ncbi.nlm.nih.gov/23925000/) · PMCID: [PMC4040527](https://pubmed.ncbi.nlm.nih.gov/PMC4040527/)

1284. **Different HIV pox viral vector-based vaccines and adjuvants can induce unique antigen presenting cells that modulate CD8 T cell avidity**

Shubhanshi Trivedi, Ronald J Jackson, Charani Ranasinghe

Virology (2014-11) <https://doi.org/f6ngrk>

DOI: [10.1016/j.virol.2014.09.004](https://doi.org/10.1016/j.virol.2014.09.004) · PMID: [25261870](https://pubmed.ncbi.nlm.nih.gov/25261870/)

1285. **Comparative evaluation of two severe acute respiratory syndrome (SARS) vaccine candidates in mice challenged with SARS coronavirus**

Raymond H See, Alexander N Zakhartchouk, Martin Petric, David J Lawrence, Catherine PY Mok, Robert J Hogan, Thomas Rowe, Lois A Zitzow, Karuna P Karunakaran, Mary M Hitt, ... BBrett Finlay

Journal of General Virology (2006-03-01) <https://doi.org/fm9v5c>

DOI: [10.1099/vir.0.81579-0](https://doi.org/10.1099/vir.0.81579-0) · PMID: [16476986](https://pubmed.ncbi.nlm.nih.gov/16476986/)

1286. **Severe acute respiratory syndrome vaccine efficacy in ferrets: whole killed virus and adenovirus-vectored vaccines**

Raymond H See, Martin Petric, David J Lawrence, Catherine PY Mok, Thomas Rowe, Lois A Zitzow, Karuna P Karunakaran, Thomas G Voss, Robert C Brunham, Jack Gauldie, ... Rachel L Roper

Journal of General Virology (2008-09-01) <https://doi.org/c5wc6w>

DOI: [10.1099/vir.0.2008/001891-0](https://doi.org/10.1099/vir.0.2008/001891-0) · PMID: [18753223](https://pubmed.ncbi.nlm.nih.gov/18753223/)

1287. **Update with the development of Ebola vaccines and implications of emerging evidence to inform future policy recommendations**

Hoi Ting Yeung

World Health Organization SAGE meeting background (2018-09-19)

https://www.who.int/immunization/sage/meetings/2018/october/2_Ebola_SAGE2018Oct_BgDoc_20180919.pdf

1288. **ChAdOx1 and MVA based vaccine candidates against MERS-CoV elicit neutralising antibodies and cellular immune responses in mice**

Naif Khalaf Alharbi, Eriko Padron-Regalado, Craig P Thompson, Alexandra Kupke, Daniel Wells, Megan A Sloan, Keith Grehan, Nigel Temperton, Teresa Lambe, George Warimwe, ... Sarah C Gilbert

Vaccine (2017-06) <https://doi.org/gbms8z>

DOI: [10.1016/j.vaccine.2017.05.032](https://doi.org/10.1016/j.vaccine.2017.05.032) · PMID: [28579232](https://pubmed.ncbi.nlm.nih.gov/28579232/) · PMCID: [PMC5516308](https://pubmed.ncbi.nlm.nih.gov/PMC5516308/)

1289. **A single dose of ChAdOx1 MERS provides protective immunity in rhesus macaques**

Neeltje van Doremale, Elaine Haddock, Friederike Feldmann, Kimberly Meade-White, Trenton Bushmaker, Robert J Fischer, Atsushi

Okumura, Patrick W Hanley, Greg Saturday, Nick J Edwards, ... Vincent J

Munster

Science Advances (2020-06-12) <https://doi.org/gjkthy>

DOI: [10.1126/sciadv.aba8399](https://doi.org/10.1126/sciadv.aba8399) · PMID: [32577525](https://pubmed.ncbi.nlm.nih.gov/32577525/) · PMCID: [PMC7286676](https://pubmed.ncbi.nlm.nih.gov/PMC7286676/)

1290. **Safety and immunogenicity of a candidate Middle East respiratory syndrome coronavirus viral-vectored vaccine: a dose-escalation, open-label, non-randomised, uncontrolled, phase 1 trial**

Pedro M Folegatti, Mustapha Bittaye, Amy Flaxman, Fernando Ramos Lopez, Duncan Bellamy, Alexandra Kupke, Catherine Mair, Rebecca Makinson, Jonathan Sheridan, Cornelius Rohde, ... Sarah Gilbert

The Lancet Infectious Diseases (2020-07) <https://doi.org/ggtxgp>

DOI: [10.1016/s1473-3099\(20\)30160-2](https://doi.org/10.1016/s1473-3099(20)30160-2) · PMID: [32325038](https://pubmed.ncbi.nlm.nih.gov/32325038/) · PMCID: [PMC7172901](https://pubmed.ncbi.nlm.nih.gov/PMC7172901/)

1291. **ChAdOx1 nCoV-19 vaccine prevents SARS-CoV-2 pneumonia in rhesus macaques**

Neeltje van Doremalen, Teresa Lambe, Alexandra Spencer, Sandra Belij-Rammerstorfer, Jyothi N Purushotham, Julia R Port, Victoria A Avanzato, Trenton Bushmaker, Amy Flaxman, Marta Ulaszewska, ... Vincent J Munster

Nature (2020-07-30) <https://doi.org/gg67jr>

DOI: [10.1038/s41586-020-2608-y](https://doi.org/10.1038/s41586-020-2608-y) · PMID: [32731258](https://pubmed.ncbi.nlm.nih.gov/32731258/) · PMCID: [PMC8436420](https://pubmed.ncbi.nlm.nih.gov/PMC8436420/)

1292. **Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial**

Pedro M Folegatti, Katie J Ewer, Parvinder K Aley, Brian Angus, Stephan Becker, Sandra Belij-Rammerstorfer, Duncan Bellamy, Sagida Bibi, Mustapha Bittaye, Elizabeth A Clutterbuck, ... Yasmine Yau

The Lancet (2020-08) <https://doi.org/gg5gwk>

DOI: [10.1016/s0140-6736\(20\)31604-4](https://doi.org/10.1016/s0140-6736(20)31604-4) · PMID: [32702298](https://pubmed.ncbi.nlm.nih.gov/32702298/) · PMCID: [PMC7445431](https://pubmed.ncbi.nlm.nih.gov/PMC7445431/)

1293. **The Russian vaccine for COVID-19**

Talha Khan Burki

The Lancet Respiratory Medicine (2020-11) <https://doi.org/ft7j>

DOI: [10.1016/s2213-2600\(20\)30402-1](https://doi.org/10.1016/s2213-2600(20)30402-1) · PMID: [32896274](https://pubmed.ncbi.nlm.nih.gov/32896274/) · PMCID: [PMC7837053](https://pubmed.ncbi.nlm.nih.gov/PMC7837053/)

1294. **Operation Warp Speed: Accelerated COVID-19 Vaccine Development Status and Efforts to Address Manufacturing Challenges**

USGovernment Accountability Office

<https://www.gao.gov/products/gao-21-319>

1295. **Johnson & Johnson Announces a Lead Vaccine Candidate for COVID-19; Landmark New Partnership with U.S. Department of Health & Human Services; and Commitment to Supply One Billion Vaccines Worldwide for Emergency Pandemic Use | Johnson & Johnson**

Content Lab U.S.

<https://www.jnj.com/johnson-johnson-announces-a-lead-vaccine-candidate-for-covid-19-landmark-new-partnership-with-u-s->

[department-of-health-human-services-and-commitment-to-supply-one-billion-vaccines-worldwide-for-emergency-pandemic-use](#)

1296. **Interim Results of a Phase 1–2a Trial of Ad26.COV2.S Covid-19 Vaccine**

Jerald Sadoff, Mathieu Le Gars, Georgi Shukarev, Dirk Heerwagh, Carla Truyers, Anne M de Groot, Jeroen Stoop, Sarah Tete, Wim Van Damme, Isabel Leroux-Roels, ... Hanneke Schuitemaker

New England Journal of Medicine (2021-05-13) <https://doi.org/fqnt>

DOI: [10.1056/nejmoa2034201](https://doi.org/10.1056/nejmoa2034201) · PMID: [33440088](https://pubmed.ncbi.nlm.nih.gov/33440088/) · PMCID: [PMC7821985](https://pubmed.ncbi.nlm.nih.gov/PMC7821985/)

1297. **Single-shot Ad26 vaccine protects against SARS-CoV-2 in rhesus macaques**

Noe B Mercado, Roland Zahn, Frank Wegmann, Carolin Loos, Abishek Chandrashekar, Jingyou Yu, Jinyan Liu, Lauren Peter, Katherine McMahan, Lisa H Tostanoski, ... Dan H Barouch

Nature (2020-07-30) <https://doi.org/d5d4>

DOI: [10.1038/s41586-020-2607-z](https://doi.org/10.1038/s41586-020-2607-z) · PMID: [32731257](https://pubmed.ncbi.nlm.nih.gov/32731257/) · PMCID: [PMC7581548](https://pubmed.ncbi.nlm.nih.gov/PMC7581548/)

1298. **Ad26 vaccine protects against SARS-CoV-2 severe clinical disease in hamsters**

Lisa H Tostanoski, Frank Wegmann, Amanda J Martinot, Carolin Loos, Katherine McMahan, Noe B Mercado, Jingyou Yu, Chi N Chan, Stephen Bondoc, Carly E Starke, ... Dan H Barouch

Nature Medicine (2020-09-03) <https://doi.org/gjhd2>

DOI: [10.1038/s41591-020-1070-6](https://doi.org/10.1038/s41591-020-1070-6) · PMID: [32884153](https://pubmed.ncbi.nlm.nih.gov/32884153/) · PMCID: [PMC7671939](https://pubmed.ncbi.nlm.nih.gov/PMC7671939/)

1299. **Immunogenicity and protective efficacy of one- and two-dose regimens of the Ad26.COV2.S COVID-19 vaccine candidate in adult and aged rhesus macaques**

Laura Solforosi, Harmjan Kuipers, Sietske K Rosendahl Huber, Joan EM van der Lubbe, Liesbeth Dekking, Dominika N Czapska-Casey, Ana Izquierdo Gil, Miranda RM Baert, Joke Drijver, Joost Vaneman, ... Roland C Zahn

Cold Spring Harbor Laboratory (2020-11-17) <https://doi.org/ghwzk9>

DOI: [10.1101/2020.11.17.368258](https://doi.org/10.1101/2020.11.17.368258)

1300. **SARS-CoV-2 binding and neutralizing antibody levels after vaccination with Ad26.COV2.S predict durable protection in rhesus macaques**

Ramon Rozendaal, Laura Solforosi, Daniel Stieh, Jan Serroyen, Roel Straetemans, Frank Wegmann, Sietske K Rosendahl Huber, Joan EM van der Lubbe, Jenny Hendriks, Mathieu le Gars, ... Roland Zahn

Cold Spring Harbor Laboratory (2021-01-30) <https://doi.org/gjhd4>

DOI: [10.1101/2021.01.30.428921](https://doi.org/10.1101/2021.01.30.428921)

1301. **Safety and efficacy of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine: an interim analysis of a randomised controlled phase 3 trial in Russia**

Denis Y Logunov, Inna V Dolzhikova, Dmitry V Shcheglyakov, Amir I Tukhvatulin, Olga V Zubkova, Alina S Dzharullaeva, Anna V Kovyrshina, Nadezhda L Lubenets, Daria M Groussova, Alina S Erokhova, ... Alexander L Gintsburg

1302. **Janssen Investigational COVID-19 Vaccine: Interim Analysis of Phase 3 Clinical Data Released**
National Institutes of Health (NIH)
(2021-01-29) <https://www.nih.gov/news-events/news-releases/janssen-investigational-covid-19-vaccine-interim-analysis-phase-3-clinical-data-released>
1303. **Johnson & Johnson Announces Single-Shot Janssen COVID-19 Vaccine Candidate Met Primary Endpoints in Interim Analysis of its Phase 3 ENSEMBLE Trial**
Janssen
(2021-01-29)
https://www.janssen.com/emea/sites/www_janssen_com_emea/files/johnson_johnson_announces_single-shot_janssen_covid-19_vaccine_candidate_met_primary_endpoints_in_interim_analysis_of_its_phase_3_ensemble_trial.pdf
1304. **AstraZeneca's COVID-19 vaccine authorised for emergency supply in the UK** <https://www.astrazeneca.com/media-centre/press-releases/2020/astrazenecas-covid-19-vaccine-authorised-in-uk.html>
1305. **The Brussels Times** <https://www.brusselstimes.com/news-contents/world/149039/1-5-million-people-have-received-sputnik-v-vaccine-russia-says-russian-direct-investment-fund-mikhail-murashko>
1306. **Hungary becomes first EU country to deploy Russia's COVID-19 vaccine**
Michael Daventry
euronews (2021-02-12)
<https://www.euronews.com/2021/02/12/hungary-to-begin-using-russia-s-sputnik-v-vaccine-today>
1307. **San Marino buys Russia's Sputnik V after EU vaccine delivery delays**
euronews
(2021-02-24) <https://www.euronews.com/2021/02/24/san-marino-buys-russia-s-sputnik-v-after-eu-vaccine-delivery-delays>
1308. **Belarus Starts Coronavirus Vaccination With Sputnik V**
AFP
The Moscow Times (2020-12-29)
<https://www.themoscowtimes.com/2020/12/29/belarus-starts-coronavirus-vaccination-with-sputnik-v-a72512>
1309. **Russia's coronavirus vaccine is alluring for Eastern Europe, creating a headache for the EU**
Holly Ellyatt
CNBC (2021-03-02) <https://www.cnbc.com/2021/03/02/russias-sputnik-vaccine-is-luring-eastern-europe-worrying-the-eu.html>
1310. **Three decades of messenger RNA vaccine development**

Rein Verbeke, Ine Lentacker, Stefaan C De Smedt, Heleen Dewitte
Nano Today (2019-10) <https://doi.org/ghm43s>
DOI: [10.1016/j.nantod.2019.100766](https://doi.org/10.1016/j.nantod.2019.100766)

1311. **Developing mRNA-vaccine technologies**

Thomas Schlake, Andreas Thess, Mariola Fotin-Mleczek, Karl-Josef Kallen
RNA Biology (2012-11) <https://doi.org/f4qzdb>
DOI: [10.4161/rna.22269](https://doi.org/10.4161/rna.22269) · PMID: [23064118](https://pubmed.ncbi.nlm.nih.gov/23064118/) · PMCID: [PMC3597572](https://pubmed.ncbi.nlm.nih.gov/PMC3597572/)

1312. **mRNA vaccines — a new era in vaccinology**

Norbert Pardi, Michael J Hogan, Frederick W Porter, Drew Weissman
Nature Reviews Drug Discovery (2018-01-12) <https://doi.org/gcsmgr>
DOI: [10.1038/nrd.2017.243](https://doi.org/10.1038/nrd.2017.243) · PMID: [29326426](https://pubmed.ncbi.nlm.nih.gov/29326426/) · PMCID: [PMC5906799](https://pubmed.ncbi.nlm.nih.gov/PMC5906799/)

1313. **Induction of virus-specific cytotoxic T lymphocytes in vivo by liposome-entrapped mRNA**

Frédéric Martinon, Sivadasan Krishnan, Gerlinde Lenzen, Rémy Magné, Elisabeth Gomard, Jean-Gérard Guillet, Jean-Paul Lévy, Pierre Meulien
European Journal of Immunology (1993-07) <https://doi.org/b6jb3z>
DOI: [10.1002/eji.1830230749](https://doi.org/10.1002/eji.1830230749) · PMID: [8325342](https://pubmed.ncbi.nlm.nih.gov/8325342/)

1314. **mRNA vaccine delivery using lipid nanoparticles**

Andreas M Reichmuth, Matthias A Oberli, Ana Jaklenec, Robert Langer, Daniel Blankschtein
Therapeutic Delivery (2016-05) <https://doi.org/f8xfzc>
DOI: [10.4155/tde-2016-0006](https://doi.org/10.4155/tde-2016-0006) · PMID: [27075952](https://pubmed.ncbi.nlm.nih.gov/27075952/) · PMCID: [PMC5439223](https://pubmed.ncbi.nlm.nih.gov/PMC5439223/)

1315. **Mechanism of action of mRNA-based vaccines**

Carlo Iavarone, Derek T O'hagan, Dong Yu, Nicolas F Delahaye, Jeffrey B Ulmer
Expert Review of Vaccines (2017-07-28) <https://doi.org/ggsnm6>
DOI: [10.1080/14760584.2017.1355245](https://doi.org/10.1080/14760584.2017.1355245) · PMID: [28701102](https://pubmed.ncbi.nlm.nih.gov/28701102/)

1316. **RNA vaccines: an introduction**

PHG Foundation
<https://www.phgfoundation.org/briefing/rna-vaccines>

1317. **T Follicular Helper Cell Differentiation, Function, and Roles in Disease**

Shane Crotty
Immunity (2014-10) <https://doi.org/ggsp64>
DOI: [10.1016/j.immuni.2014.10.004](https://doi.org/10.1016/j.immuni.2014.10.004) · PMID: [25367570](https://pubmed.ncbi.nlm.nih.gov/25367570/) · PMCID: [PMC4223692](https://pubmed.ncbi.nlm.nih.gov/PMC4223692/)

1318. **SARS-CoV-2 Vaccines: Status Report**

Fatima Amanat, Florian Krammer
Immunity (2020-04) <https://doi.org/ggrdj4>
DOI: [10.1016/j.immuni.2020.03.007](https://doi.org/10.1016/j.immuni.2020.03.007) · PMID: [32259480](https://pubmed.ncbi.nlm.nih.gov/32259480/) · PMCID: [PMC7136867](https://pubmed.ncbi.nlm.nih.gov/PMC7136867/)

1319. **Study in Healthy Adults to Evaluate Gene Activation After Vaccination With GlaxoSmithKline (GSK) Biologicals' Candidate Tuberculosis (TB) Vaccine GSK 692342 - Full Text View - ClinicalTrials.gov**

<https://clinicaltrials.gov/ct2/show/NCT01669096>

1320. **Nucleoside-modified mRNA immunization elicits influenza virus hemagglutinin stalk-specific antibodies**

Norbert Pardi, Kaela Parkhouse, Ericka Kirkpatrick, Meagan McMahon, Seth J Zost, Barbara L Mui, Ying K Tam, Katalin Karikó, Christopher J Barbosa, Thomas D Madden, ... Drew Weissman

Nature Communications (2018-08-22) <https://doi.org/gd49qt>

DOI: [10.1038/s41467-018-05482-0](https://doi.org/s41467-018-05482-0) · PMID: [30135514](#) · PMCID: [PMC6105651](#)

1321. **Cell specific delivery of modified mRNA expressing therapeutic proteins to leukocytes**

Nuphar Veiga, Meir Goldsmith, Yasmin Granot, Daniel Rosenblum, Niels Dammes, Ranit Kedmi, Srinivas Ramishetti, Dan Peer

Nature Communications (2018-10-29) <https://doi.org/gfmcrt>

DOI: [10.1038/s41467-018-06936-1](https://doi.org/s41467-018-06936-1) · PMID: [30374059](#) · PMCID: [PMC6206083](#)

1322. **Managing intellectual property to develop medicines for the world's poorest**

Sylvie Fonteilles-Drabek, David Reddy, Timothy NC Wells

Nature Reviews Drug Discovery (2017-02-24) <https://doi.org/gmqmgy>

DOI: [10.1038/nrd.2017.24](https://doi.org/10.1038/nrd.2017.24) · PMID: [28232725](#)

1323. **Immunology of COVID-19: Current State of the Science**

Nicolas Vabret, Graham J Britton, Conor Gruber, Samarth Hegde, Joel Kim, Maria Kuksin, Rachel Levantovsky, Louise Malle, Alvaro Moreira, Matthew D Park, ... Uri Laserson

Immunity (2020-06) <https://doi.org/ggt54g>

DOI: [10.1016/j.immuni.2020.05.002](https://doi.org/10.1016/j.immuni.2020.05.002) · PMID: [32505227](#) · PMCID: [PMC7200337](#)

1324. **Synthetic Chemically Modified mRNA (modRNA): Toward a New Technology Platform for Cardiovascular Biology and Medicine**

KR Chien, L Zangi, KO Lui

Cold Spring Harbor Perspectives in Medicine (2014-10-09)

<https://doi.org/f3pvsr>

DOI: [10.1101/cshperspect.a014035](https://doi.org/10.1101/cshperspect.a014035) · PMID: [25301935](#) · PMCID: [PMC4292072](#)

1325. **Pfizer and BioNTech Announce Early Positive Data from an Ongoing Phase 1/2 study of mRNA-based Vaccine Candidate Against SARS-CoV-2 | Pfizer** [https://www\(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-announce-early-positive-data-ongoing-0](https://www(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-announce-early-positive-data-ongoing-0)

1326. **Phase I, Open-Label, Dose-Ranging Study of the Safety and Immunogenicity of 2019-nCoV Vaccine (mRNA-1273) in Healthy Adults**

National Institute of Allergy and Infectious Diseases (NIAID)

clinicaltrials.gov (2020-12-17)

<https://clinicaltrials.gov/ct2/show/NCT04283461>

1327. **Efficacy and Safety of the mRNA-1273 SARS-CoV-2 Vaccine**

Lindsey R Baden, Hana M El Sahly, Brandon Essink, Karen Kotloff, Sharon Frey, Rick Novak, David Diemert, Stephen A Spector, Nadine

1328. **FDA Takes Key Action in Fight Against COVID-19 By Issuing Emergency Use Authorization for First COVID-19 Vaccine**

Office of the Commissioner

FDA (2020-12-14) <https://www.fda.gov/news-events/press-announcements/fda-takes-key-action-fight-against-covid-19-issuing-emergency-use-authorization-first-covid-19>

1329. **The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Moderna COVID-19 Vaccine — United States, December 2020**

Sara E Oliver

MMWR. Morbidity and Mortality Weekly Report (2021)
<https://www.cdc.gov/mmwr/volumes/69/wr/mm695152e1.htm>
DOI: [10.15585/mmwr.mm695152e1](https://doi.org/10.15585/mmwr.mm695152e1)

1330. **Prevention and Attenuation of Covid-19 with the BNT162b2 and mRNA-1273 Vaccines**

Mark G Thompson, Jefferey L Burgess, Allison L Naleway, Harmony Tyner, Sarang K Yoon, Jennifer Meece, Lauren EW Olsho, Alberto J Caban-Martinez, Ashley L Fowlkes, Karen Lutrick, ... Manjusha Gaglani
New England Journal of Medicine (2021-07-22) <https://doi.org/gk3bzr>
DOI: [10.1056/nejmoa2107058](https://doi.org/10.1056/nejmoa2107058) · PMID: [34192428](#) · PMCID: [PMC8262622](#)

1331. **Increased risk of SARS-CoV-2 reinfection associated with emergence of Omicron in South Africa**

Juliet RC Pulliam, Cari van Schalkwyk, Nevan Shan Govender, Anne von Gottberg, Cheryl Cohen, Michelle J Groome, Jonathan Dushoff, Koleka Mlisana, Harry Moultrie

Cold Spring Harbor Laboratory (2021-11-11) <https://doi.org/g8gj>
DOI: [10.1101/2021.11.11.21266068](https://doi.org/10.1101/2021.11.11.21266068)

1332. **Coronavirus Disease 2019 (COVID-19)**

CDC

Centers for Disease Control and Prevention (2020-02-11)
<https://www.cdc.gov/coronavirus/2019-ncov/science/science-briefs/scientific-brief-omicron-variant.html>

1333. **Sensitivity of infectious SARS-CoV-2 B.1.1.7 and B.1.351 variants to neutralizing antibodies**

Delphine Planas, Timothée Bruel, Ludivine Grzelak, Florence Guivel-Benhassine, Isabelle Staropoli, Françoise Porrot, Cyril Planchais, Julian Buchrieser, Maaran Michael Rajah, Elodie Bishop, ... Olivier Schwartz
Nature Medicine (2021-03-26) <https://doi.org/gjmwwr>

DOI: [10.1038/s41591-021-01318-5](https://doi.org/10.1038/s41591-021-01318-5) · PMID: [33772244](#)

1334. **Antibody resistance of SARS-CoV-2 variants B.1.351 and B.1.1.7**

Pengfei Wang, Manoj S Nair, Lihong Liu, Sho Iketani, Yang Luo, Yicheng Guo, Maple Wang, Jian Yu, Baoshan Zhang, Peter D Kwong, ... David D Ho

Nature (2021-03-08) <https://doi.org/gjhdxm>
DOI: [10.1038/s41586-021-03398-2](https://doi.org/10.1038/s41586-021-03398-2) · PMID: [33684923](#)

1335. Neutralizing Antibodies Against SARS-CoV-2 Variants After Infection and Vaccination

Venkata Viswanadh Edara, William H Hudson, Xuping Xie, Rafi Ahmed, Mehul S Suthar

JAMA (2021-05-11) <https://doi.org/gj29vq>

DOI: [10.1001/jama.2021.4388](https://doi.org/10.1001/jama.2021.4388) · PMID: [33739374](https://pubmed.ncbi.nlm.nih.gov/33739374/) · PMCID: [PMC7980146](https://pubmed.ncbi.nlm.nih.gov/PMC7980146/)

1336. Classification of Omicron (B.1.1.529): SARS-CoV-2 Variant of Concern [https://www.who.int/news-room/detail/26-11-2021-classification-of-omicron-\(b.1.1.529\)-sars-cov-2-variant-of-concern](https://www.who.int/news-room/detail/26-11-2021-classification-of-omicron-(b.1.1.529)-sars-cov-2-variant-of-concern)

1337. Breakthrough Infections with SARS-CoV-2 Omicron Variant Despite Booster Dose of mRNA Vaccine

Constanze Kuhlmann, Carla Konstanze Mayer, Mathilda Claassen, Tongai G Maponga, Andrew D Sutherland, Tasnim Suliman, Megan Shaw, Wolfgang Preiser

SSRN Electronic Journal (2021) <https://doi.org/gpnbfz>

DOI: [10.2139/ssrn.3981711](https://doi.org/10.2139/ssrn.3981711)

1338. Reduced neutralization of SARS-CoV-2 B.1.617 by vaccine and convalescent serum

Chang Liu, Helen M Ginn, Wanwisa Dejnirattisai, Piyada Supasa, Beibei Wang, Aekkachai Tuekprakhon, Rungtiwa Nutalai, Daming Zhou, Alexander J Mentzer, Yuguang Zhao, ... Gavin R Screamton

Cell (2021-08) <https://doi.org/gmdhv>

DOI: [10.1016/j.cell.2021.06.020](https://doi.org/10.1016/j.cell.2021.06.020) · PMID: [34242578](https://pubmed.ncbi.nlm.nih.gov/34242578/) · PMCID: [PMC8218332](https://pubmed.ncbi.nlm.nih.gov/PMC8218332/)

1339. Broadly neutralizing antibodies overcome SARS-CoV-2 Omicron antigenic shift

Elisabetta Cameroni, Christian Saliba, John E Bowen, Laura E Rosen, Katja Culap, Dora Pinto, Laura A VanBlargan, Anna De Marco, Samantha K Zepeda, Julia di Iulio, ... Davide Corti

Cold Spring Harbor Laboratory (2021-12-14) <https://doi.org/hb7x>

DOI: [10.1101/2021.12.12.472269](https://doi.org/10.1101/2021.12.12.472269) · PMID: [34931194](https://pubmed.ncbi.nlm.nih.gov/34931194/) · PMCID: [PMC8687478](https://pubmed.ncbi.nlm.nih.gov/PMC8687478/)

1340. SARS-CoV-2 Omicron: evasion of potent humoral responses and resistance to clinical immunotherapeutics relative to viral variants of concern

Anupriya Aggarwal, Alberto Ospina Stella, Gregory Walker, Anouschka Akerman, Vanessa Milogiannakis, Fabienne Brilot, Supavadee Amatayakul-Chantler, Nathan Roth, Germano Coppola, Peter Schofield, ... Stuart Turville

Cold Spring Harbor Laboratory (2021-12-15) <https://doi.org/hb73>

DOI: [10.1101/2021.12.14.2126777](https://doi.org/10.1101/2021.12.14.2126777)

1341. The SARS-CoV-2 B.1.1.529 Omicron virus causes attenuated infection and disease in mice and hamsters

Michael Diamond, Peter Halfmann, Tadashi Maemura, Kiyoko Iwatsuki-Horimoto, Shun Iida, Maki Kiso, Suzanne Scheaffer, Tamarand Darling, Astha Joshi, Samantha Loeber, ... Viviana Simon

Research Square Platform LLC (2021-12-29) <https://doi.org/gpnbf>

DOI: [10.21203/rs.3.rs-1211792/v1](https://doi.org/10.21203/rs.3.rs-1211792/v1) · PMID: [34981044](https://pubmed.ncbi.nlm.nih.gov/34981044/) · PMCID: [PMC8722607](https://pubmed.ncbi.nlm.nih.gov/PMC8722607/)

1342. **SARS-CoV-2 Omicron-B.1.1.529 Variant leads to less severe disease than Pango B and Delta variants strains in a mouse model of severe COVID-19**

Eleanor G Bentley, Adam Kirby, Parul Sharma, Anja Kipar, Daniele F Mega, Chloe Bramwell, Rebekah Penrice-Randal, Tessa Prince, Jonathan C Brown, Jie Zhou, ... James P Stewart
Cold Spring Harbor Laboratory (2021-12-28) <https://doi.org/gpnfb>
DOI: [10.1101/2021.12.26.474085](https://doi.org/10.1101/2021.12.26.474085)

1343. **The omicron (B.1.1.529) SARS-CoV-2 variant of concern does not readily infect Syrian hamsters**

Rana Abdelnabi, Caroline S Foo, Xin Zhang, Viktor Lemmens, Piet Maes, Bram Slechten, Joren Raymenants, Emmanuel André, Birgit Weynand, Kai Dallemeier, Johan Neyts
Cold Spring Harbor Laboratory (2021-12-26) <https://doi.org/gpnbd9>
DOI: [10.1101/2021.12.24.474086](https://doi.org/10.1101/2021.12.24.474086)

1344. **Reduced Pathogenicity of the SARS-CoV-2 Omicron Variant in Hamsters**

Katherine McMahan, Victoria Giffin, Lisa H Tostanoski, Benjamin Chung, Mazuba Siamatu, Mehul S Suthar, Peter Halfmann, Yoshihiro Kawaoka, Cesar Piedra-Mora, Amanda J Martinot, ... Dan H Barouch
Cold Spring Harbor Laboratory (2022-01-03) <https://doi.org/gpnbf>
DOI: [10.1101/2022.01.02.474743](https://doi.org/10.1101/2022.01.02.474743)

1345. **SARS-CoV-2 Omicron spike mediated immune escape and tropism shift**

Bo Meng, Isabella ATM Ferreira, Adam Abdullahi, Niluka Goonawardane, Akatsuki Saito, Izumi Kimura, Daichi Yamasoba, Pehuén Perera Gerba, Saman Fatihi, Surabhi Rathore, ...
Cold Spring Harbor Laboratory (2021-12-21) <https://doi.org/gpnbd7>
DOI: [10.1101/2021.12.17.473248](https://doi.org/10.1101/2021.12.17.473248)

1346. **Coronavirus in the U.S.: Latest Map and Case Count**

The New York Times
<https://www.nytimes.com/interactive/2021/us/covid-cases.html>

1347. **Analysis | Four charts that analyze how omicron's wave compares to previous coronavirus peaks**

Shelly Tan
Washington Post
<https://www.washingtonpost.com/health/interactive/2022/omicron-comparison-cases-deaths-hospitalizations/>

1348. **Effectiveness of BNT162b2 Vaccine against Omicron Variant in South Africa**

Shirley Collie, Jared Champion, Harry Moultrie, Linda-Gail Bekker, Glenda Gray
New England Journal of Medicine (2022-02-03) <https://doi.org/gnxj2w>
DOI: [10.1056/nejmc2119270](https://doi.org/10.1056/nejmc2119270) · PMID: [34965358](#) · PMCID: [PMC8757569](#)

1349. **A meta-analysis of Early Results to predict Vaccine efficacy against Omicron**

David S Khoury, Megan Steain, James A Triccas, Alex Sigal, Miles P Davenport, Deborah Cromer

1350. **Waning 2-Dose and 3-Dose Effectiveness of mRNA Vaccines Against COVID-19-Associated Emergency Department and Urgent Care Encounters and Hospitalizations Among Adults During Periods of Delta and Omicron Variant Predominance — VISION Network, 10 States, August 2021–January 2022**
Jill M Ferdinands, Suchitra Rao, Brian E Dixon, Patrick K Mitchell, Malini B DeSilva, Stephanie A Irving, Ned Lewis, Karthik Natarajan, Edward Stenehjem, Shaun J Grannis, ... Bruce Fireman
MMWR. Morbidity and Mortality Weekly Report (2022-02-18)
<https://doi.org/gpfrkh>
DOI: [10.15585/mmwr.mm7107e2](https://doi.org/10.15585/mmwr.mm7107e2) · PMID: [35176007](https://pubmed.ncbi.nlm.nih.gov/35176007/) · PMCID: [PMC8853475](https://pubmed.ncbi.nlm.nih.gov/PMC8853475/)
1351. **COVID-19 mRNA booster vaccines elicit strong protection against SARS-CoV-2 Omicron variant in patients with cancer**
Cong Zeng, John P Evans, Karthik Chakravarthy, Panke Qu, Sarah Reisinger, No-Joon Song, Mark P Rubinstein, Peter G Shields, Zihai Li, Shan-Lu Liu
Cancer Cell (2022-02) <https://doi.org/gn2gcf>
DOI: [10.1016/j.ccr.2021.12.014](https://doi.org/10.1016/j.ccr.2021.12.014) · PMID: [34986328](https://pubmed.ncbi.nlm.nih.gov/34986328/) · PMCID: [PMC8716174](https://pubmed.ncbi.nlm.nih.gov/PMC8716174/)
1352. **Immune correlates analysis of the mRNA-1273 COVID-19 vaccine efficacy clinical trial**
Peter B Gilbert, David C Montefiori, Adrian B McDermott, Youyi Fong, David Benkeser, Weiping Deng, Honghong Zhou, Christopher R Houchens, Karen Martins, Lakshmi Jayashankar, ...
Science (2022-01-07) <https://doi.org/gpnbff>
DOI: [10.1126/science.abm3425](https://doi.org/10.1126/science.abm3425) · PMID: [34812653](https://pubmed.ncbi.nlm.nih.gov/34812653/)
1353. **Towards a population-based threshold of protection for COVID-19 vaccines**
David Goldblatt, Andrew Fiore-Gartland, Marina Johnson, Adam Hunt, Christopher Bengt, Dace Zavadska, Hilda Darta Snipe, Jeremy S Brown, Lesley Workman, Heather J Zar, ... Donna Ambrosino
Vaccine (2022-01) <https://doi.org/gpnbdz>
DOI: [10.1016/j.vaccine.2021.12.006](https://doi.org/10.1016/j.vaccine.2021.12.006) · PMID: [34933765](https://pubmed.ncbi.nlm.nih.gov/34933765/) · PMCID: [PMC8673730](https://pubmed.ncbi.nlm.nih.gov/PMC8673730/)
1354. **Evidence for antibody as a protective correlate for COVID-19 vaccines**
Kristen A Earle, Donna M Ambrosino, Andrew Fiore-Gartland, David Goldblatt, Peter B Gilbert, George R Siber, Peter Dull, Stanley A Plotkin
Vaccine (2021-07) <https://doi.org/gnr5w7>
DOI: [10.1016/j.vaccine.2021.05.063](https://doi.org/10.1016/j.vaccine.2021.05.063) · PMID: [34210573](https://pubmed.ncbi.nlm.nih.gov/34210573/) · PMCID: [PMC8142841](https://pubmed.ncbi.nlm.nih.gov/PMC8142841/)
1355. **Towards Internationally standardised humoral Immune Correlates of Protection from SARS-CoV-2 infection and COVID-19 disease**
Javier Castillo-Olivares, David A Wells, Matteo Ferrari, Andrew Chan, Peter Smith, Angalee Nadesalingam, Minna Paloniemi, George Carnell,

1356. **A longitudinal study of SARS-CoV-2-infected patients reveals a high correlation between neutralizing antibodies and COVID-19 severity**

Vincent Legros, Solène Denolly, Manon Vogrig, Bertrand Boson, Eglantine Siret, Josselin Rigaill, Sylvie Pillet, Florence Grattard, Sylvie Gonzalo, Paul Verhoeven, ... Bruno Pozzetto
Cellular & Molecular Immunology (2021-01-06)
<https://doi.org/gh5qjr>
DOI: [10.1038/s41423-020-00588-2](https://doi.org/s41423-020-00588-2) · PMID: [33408342](https://pubmed.ncbi.nlm.nih.gov/33408342/) · PMCID: [PMC7786875](https://pubmed.ncbi.nlm.nih.gov/PMC7786875/)

1357. **4th Dose COVID mRNA Vaccines' Immunogenicity & Efficacy Against Omicron VOC**

Gili Regev-Yochay, Tal Gonen, Mayan Gilboa, Michal Mandelboim, Victoria Indenbaum, Sharon Amit, Lilac Meltzer, Keren Asraf, Carmit Cohen, Ronen Fluss, ... Yaniv Lustig
Cold Spring Harbor Laboratory (2022-02-15) <https://doi.org/gpnbfd>
DOI: [10.1101/2022.02.15.22270948](https://doi.org/10.1101/2022.02.15.22270948)

1358. **Safety and immunogenicity of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia**

Denis Y Logunov, Inna V Dolzhikova, Olga V Zubkova, Amir I Tukhvatulin, Dmitry V Shchegolyakov, Alina S Dzharullaeva, Daria M Grousova, Alina S Erokhova, Anna V Kovyrshina, Andrei G Botikov, ... Alexander L Gintzburg
The Lancet (2020-09) <https://doi.org/gg96hq>
DOI: [10.1016/s0140-6736\(20\)31866-3](https://doi.org/s0140-6736(20)31866-3) · PMID: [32896291](https://pubmed.ncbi.nlm.nih.gov/32896291/) · PMCID: [PMC7471804](https://pubmed.ncbi.nlm.nih.gov/PMC7471804/)

1359. **WHO lists additional COVID-19 vaccine for emergency use and issues interim policy recommendations**

<https://www.who.int/news-room/detail/07-05-2021-who-lists-additional-covid-19-vaccine-for-emergency-use-and-issues-interim-policy-recommendations>

1360. **Immunogenicity and safety of the CoronaVac inactivated vaccine in patients with autoimmune rheumatic diseases: a phase 4 trial**

Ana C Medeiros-Ribeiro, Nadia E Aikawa, Carla GS Saad, Emily FN Yuki, Tatiana Pedrosa, Solange RG Fusco, Priscila T Rojo, Rosa MR Pereira, Samuel K Shinjo, Danieli CO Andrade, ... Eloisa Bonfa
Nature Medicine (2021-07-30) <https://doi.org/gmwn4j>
DOI: [10.1038/s41591-021-01469-5](https://doi.org/10.1038/s41591-021-01469-5) · PMID: [34331051](https://pubmed.ncbi.nlm.nih.gov/34331051/)

1361. **Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine (CoronaVac) in healthy children and adolescents: a double-blind, randomised, controlled, phase 1/2 clinical trial**

Bihua Han, Yufei Song, Changgui Li, Wanqi Yang, Qingxia Ma, Zhiwei Jiang, Minjie Li, Xiaojuan Lian, Wenbin Jiao, Lei Wang, ... Qiang Gao
The Lancet Infectious Diseases (2021-12) <https://doi.org/gn93>

1362. **Viral dynamics in asymptomatic patients with COVID-19**

Rui Zhou, Furong Li, Fengjuan Chen, Huamin Liu, Jiazhen Zheng, Chunliang Lei, Xianbo Wu

International Journal of Infectious Diseases (2020-07)

<https://doi.org/ggxs96>

DOI: [10.1016/j.ijid.2020.05.030](https://doi.org/j.ijid.2020.05.030) · PMID: [32437933](#) · PMCID:

[PMC7211726](#)

1363. **Higher viral loads in asymptomatic COVID-19 patients might be the invisible part of the iceberg**

Imran Hasanoglu, Gulay Korukluoglu, Dilek Asilturk, Yasemin Cosgun, Ayse Kaya Kalem, Ayse Basak Altas, Bircan Kayaaslan, Fatma Eser, Esra Akkan Kuzucu, Rahmet Guner

Infection (2020-11-24) <https://doi.org/ghxsp4>

DOI: [10.1007/s15010-020-01548-8](https://doi.org/s15010-020-01548-8) · PMID: [32231841](#) · PMCID:

[PMC7685188](#)

1364. **Relationships between Viral Load and the Clinical Course of COVID-19**

Hiroyuki Tsukagoshi, Daisuke Shinoda, Mariko Saito, Kaori Okayama, Mitsuru Sada, Hirokazu Kimura, Nobuhiro Saruki

Viruses (2021-02-15) <https://doi.org/gndc5c>

DOI: [10.3390/v13020304](https://doi.org/v13020304) · PMID: [33672005](#) · PMCID: [PMC7919281](#)

1365. **Estimating the extent of asymptomatic COVID-19 and its potential for community transmission: Systematic review and meta-analysis**

Oyungerel Byambasuren, Magnolia Cardona, Katy Bell, Justin Clark, Mary-Louise McLaws, Paul Glasziou

Official Journal of the Association of Medical Microbiology and Infectious Disease Canada (2020-12) <https://doi.org/gh7qmm>

DOI: [10.3138/jammi-2020-0030](https://doi.org/10.3138/jammi-2020-0030)

1366. **SARS-CoV-2 Infection after Vaccination in Health Care Workers in California**

Jocelyn Keehner, Lucy E Horton, Michael A Pfeffer, Christopher A Longhurst, Robert T Schooley, Judith S Currier, Shira R Abeles, Francesca J Torriani

New England Journal of Medicine (2021-05-06) <https://doi.org/gjjr6h>

DOI: [10.1056/nejmc2101927](https://doi.org/10.1056/nejmc2101927) · PMID: [33755376](#) · PMCID: [PMC8008750](#)

1367. **Impact of the Coronavirus Disease 2019 (COVID-19) Vaccine on Asymptomatic Infection Among Patients Undergoing Preprocedural COVID-19 Molecular Screening**

Aaron J Tande, Benjamin D Pollock, Nilay D Shah, Gianrico Farrugia, Abinash Virk, Melanie Swift, Laura Breeher, Matthew Binnicker, Elie F Berbari

Clinical Infectious Diseases (2021-03-10) <https://doi.org/gjg8qf>

DOI: [10.1093/cid/ciab229](https://doi.org/10.1093/cid/ciab229) · PMID: [33704435](#) · PMCID: [PMC7989519](#)

1368. **Association Between Vaccination With BNT162b2 and Incidence of Symptomatic and Asymptomatic SARS-CoV-2 Infections Among**

Health Care Workers

Yoel Angel, Avishay Spitzer, Oryan Henig, Esther Saiag, Eli Sprecher,

Hagit Padova, Ronen Ben-Ami

JAMA (2021-06-22) <https://doi.org/gjwp6b>

DOI: [10.1001/jama.2021.7152](https://doi.org/10.1001/jama.2021.7152) · PMID: [33956048](https://pubmed.ncbi.nlm.nih.gov/33956048/) · PMCID: [PMC8220476](https://pubmed.ncbi.nlm.nih.gov/PMC8220476/)

1369. Initial report of decreased SARS-CoV-2 viral load after inoculation with the BNT162b2 vaccine

Matan Levine-Tiefenbrun, Idan Yelin, Rachel Katz, Esma Herz, Ziv Golan, Licia Schreiber, Tamar Wolf, Varda Nadler, Amir Ben-Tov, Jacob Kuint, ... Roy Kishony

Nature Medicine (2021-03-29) <https://doi.org/gjmx9h>

DOI: [10.1038/s41591-021-01316-7](https://doi.org/10.1038/s41591-021-01316-7) · PMID: [33782619](https://pubmed.ncbi.nlm.nih.gov/33782619/)

1370. Virological and serological kinetics of SARS-CoV-2 Delta variant vaccine-breakthrough infections: a multi-center cohort study

Po Ying Chia, Sean Wei Xiang Ong, Calvin J Chiew, Li Wei Ang, Jean-Marc Chavatte, Tze-Minn Mak, Lin Cui, Shirin Kalimuddin, Wan Ni Chia, Chee Wah Tan, ... Barnaby Edward Young

Cold Spring Harbor Laboratory (2021-07-31) <https://doi.org/gmd72x>

DOI: [10.1101/2021.07.28.21261295](https://doi.org/10.1101/2021.07.28.21261295)

1371. Shedding of Infectious SARS-CoV-2 Despite Vaccination

Kasen K Riemersma, Brittany E Grogan, Amanda Kita-Yarbro, Peter J Halfmann, Hannah E Segaloff, Anna Kocharian, Kelsey R Florek, Ryan Westergaard, Allen Bateman, Gunnar E Jeppson, ... Katarina M Grande

Cold Spring Harbor Laboratory (2021-07-31) <https://doi.org/gmfh6j>

DOI: [10.1101/2021.07.31.21261387](https://doi.org/10.1101/2021.07.31.21261387)

1372. No Significant Difference in Viral Load Between Vaccinated and Unvaccinated, Asymptomatic and Symptomatic Groups When Infected with SARS-CoV-2 Delta Variant

Charlotte B Acharya, John Schrom, Anthea M Mitchell, David A Coil, Carina Marquez, Susana Rojas, Chung Yu Wang, Jamin Liu, Genay Pilarowski, Leslie Solis, ... Diane Havlir

Cold Spring Harbor Laboratory (2021-09-29) <https://doi.org/gndc47>

DOI: [10.1101/2021.09.28.21264262](https://doi.org/10.1101/2021.09.28.21264262)

1373. Safety, Immunogenicity, and Efficacy of a COVID-19 Vaccine (NVX-CoV2373) Co-administered With Seasonal Influenza Vaccines

Seth Toback, Eva Galiza, Catherine Cosgrove, James Galloway, Anna L Goodman, Pauline A Swift, Sankarasubramanian Rajaram, Alison Graves-Jones, Jonathan Edelman, Fiona Burns, ... Paul T Heath

Cold Spring Harbor Laboratory (2021-06-13) <https://doi.org/gngp57>

DOI: [10.1101/2021.06.09.21258556](https://doi.org/10.1101/2021.06.09.21258556)

1374. Novavax Initiates Phase 1/2 Clinical Trial of Combination Vaccine for COVID-19 and Seasonal Influenza

Novavax Investor Relations

<https://ir.novavax.com/2021-09-08-Novavax-Initiates-Phase-1-2-Clinical-Trial-of-Combination-Vaccine-for-COVID-19-and-Seasonal-Influenza>

1375. Combination Respiratory Vaccine Containing Recombinant SARS-CoV-2 Spike and Quadrivalent Seasonal Influenza Hemagglutinin

Nanoparticles with Matrix-M Adjuvant

Michael J Massare, Nita Patel, Bin Zhou, Sonia Maciejewski, Rhonda Flores, Mimi Guebre-Xabier, Jing-Hui Tian, Alyse D Portnoff, Louis Fries, Vivek Shinde, ... Gale Smith

Cold Spring Harbor Laboratory (2021-05-05) <https://doi.org/gngp56>

DOI: [10.1101/2021.05.05.442782](https://doi.org/10.1101/2021.05.05.442782)

1376. **Reactogenicidad, Seguridad e Inmunogenicidad de Dosis de Refuerzo de Vacunas Contra SARS-CoV-2 en Chile (Estudio REFUERZO)**

Rafael Araos

clinicaltrials.gov (2021-08-04)

<https://clinicaltrials.gov/ct2/show/NCT04992182>

1377. **SARS Vaccine Development**

Shibo Jiang, Yuxian He, Shuwen Liu

Emerging Infectious Diseases (2005-07) <https://doi.org/gm2qkj>

DOI: [10.3201/eid1107.050219](https://doi.org/eid1107.050219) · PMID: [16022774](https://pubmed.ncbi.nlm.nih.gov/16022774/) · PMCID: [PMC3371787](https://pubmed.ncbi.nlm.nih.gov/PMC3371787/)

1378. **A decade after SARS: strategies for controlling emerging coronaviruses**

Rachel L Graham, Eric F Donaldson, Ralph S Baric

Nature Reviews Microbiology (2013-11-11) <https://doi.org/ggwrzg>

DOI: [10.1038/nrmicro3143](https://doi.org/nrmicro3143) · PMID: [24217413](https://pubmed.ncbi.nlm.nih.gov/24217413/) · PMCID: [PMC5147543](https://pubmed.ncbi.nlm.nih.gov/PMC5147543/)

1379. **Vaccines for emerging infectious diseases: Lessons from MERS coronavirus and Zika virus**

Joel N Maslow

Human Vaccines & Immunotherapeutics (2017-08-28)

<https://doi.org/gk7gb4>

DOI: [10.1080/21645515.2017.1358325](https://doi.org/21645515.2017.1358325) · PMID: [28846484](https://pubmed.ncbi.nlm.nih.gov/28846484/) · PMCID: [PMC5718785](https://pubmed.ncbi.nlm.nih.gov/PMC5718785/)

1380. **Ebola vaccine: Little and late**

Jon Cohen

Science (2014-09-19) <https://doi.org/gm2qkd>

DOI: [10.1126/science.345.6203.1441](https://doi.org/10.1126/science.345.6203.1441) · PMID: [25237082](https://pubmed.ncbi.nlm.nih.gov/25237082/)

1381. **Clinical development of a recombinant Ebola vaccine in the midst of an unprecedented epidemic**

Beth-Ann G Coller, Jeffrey Blue, Rituparna Das, Sheri Dubey, Lynn Finelli, Swati Gupta, Frans Helmond, Rebecca J Grant-Klein, Kenneth Liu, Jakub Simon, ... Thomas P Monath

Vaccine (2017-08) <https://doi.org/gbw3rt>

DOI: [10.1016/j.vaccine.2017.05.097](https://doi.org/10.1016/j.vaccine.2017.05.097) · PMID: [28647166](https://pubmed.ncbi.nlm.nih.gov/28647166/)

1382. **First-in-human, Randomised, Double-blind, Placebo-controlled, Dose-escalation Study in Healthy Young Adults Evaluating the Safety and Immunogenicity of COVI-VAC, a Live Attenuated Vaccine Candidate for Prevention of COVID-19**

Codagenix, Inc

clinicaltrials.gov (2021-07-26)

<https://clinicaltrials.gov/ct2/show/NCT04619628>

1383. **Phase 1, Open-Label, Dose-Escalation Study to Evaluate Tolerability, Safety, and Immunogenicity of an Intranasal Live Attenuated Respiratory Syncytial Virus Vaccine Expressing Spike Protein of SARS-CoV-2 in Healthy Adults Ages 18 - 69 Years**
Meissa Vaccines, Inc.
clinicaltrials.gov (2021-07-01)
<https://clinicaltrials.gov/ct2/show/NCT04798001>
1384. **BCG Vaccination to Protect Healthcare Workers Against COVID-19**
- Full Text View - ClinicalTrials.gov
<https://clinicaltrials.gov/ct2/show/NCT04327206>
1385. **BCG Vaccine for Health Care Workers as Defense Against COVID 19**
- Full Text View - ClinicalTrials.gov
<https://clinicaltrials.gov/ct2/show/NCT04348370>
1386. **Safety and immunogenicity of a novel human Enterovirus 71 (EV71) vaccine: A randomized, placebo-controlled, double-blind, Phase I clinical trial**
Yan-Ping Li, Zheng-Lun Liang, Qiang Gao, Li-Rong Huang, Qun-Ying Mao, Shu-Qun Wen, Yan Liu, Wei-Dong Yin, Rong-Cheng Li, Jun-Zhi Wang
Vaccine (2012-05) <https://doi.org/gh7tjn>
DOI: [10.1016/j.vaccine.2012.03.010](https://doi.org/10.1016/j.vaccine.2012.03.010) · PMID: [22426327](#)
1387. **Double-Blind, Randomized, Placebo-Controlled Phase III Clinical Trial to Evaluate the Efficacy and Safety of treating Healthcare Professionals with the Adsorbed COVID-19 (Inactivated) Vaccine Manufactured by Sinovac – PROFISCOV: A structured summary of a study protocol for a randomised controlled trial**
Ricardo Palacios, Elizabeth González Patiño, Roberta de Oliveira Piorelli, Monica Tilli Reis Pessoa Conde, Ana Paula Batista, Gang Zeng, Qianqian Xin, Esper G Kallas, Jorge Flores, Christian F Ockenhouse, Christopher Gast
Trials (2020-10-15) <https://doi.org/ghjkrh>
DOI: [10.1186/s13063-020-04775-4](https://doi.org/10.1186/s13063-020-04775-4) · PMID: [33059771](#) · PMCID: [PMC7558252](#)
1388. **Sinovac: CoronaVac – COVID19 Vaccine Tracker**
<https://covid19.trackvaccines.org/vaccines/7/>
1389. **Bharat Biotech Announces Phase 3 Results of COVAXIN**
Bharat Biotech
(2021-03-03) <https://www.bharatbiotech.com/images/press/covaxin-phase3-efficacy-results.pdf>
1390. **Ocugen's COVID-19 Vaccine Co-Development Partner, Bharat Biotech shares Phase 3 Interim Results of COVAXIN, Demonstrates Efficacy of 81%**
Biogenetech
(2021-03-05) <https://www.biogenetech.co.th/wp-content/uploads/2021/03/5-Ocugen.pdf>
1391. **Zimbabwe authorizes use of India's first indigenous COVID-19 vaccine - Xinhua | English.news.cn**

http://www.xinhuanet.com/english/2021-03/04/c_139783893.htm

1392. **Booster dose: Bharat Biotech's nasal vaccine may be used with Covaxin**

Sohini Das

Business Standard India (2021-09-25) https://www.business-standard.com/article/current-affairs/booster-dose-bharat-biotech-s-nasal-vaccine-may-be-used-with-covaxin-121092500034_1.html

1393. **Covaxin kids trial over, Bharat Biotech to submit data to DCGI next week**

The Times of India

(2021-09-21) <https://timesofindia.indiatimes.com/india/bharat-biotech-to-submit-covaxin-kids-trials-data-to-dcgi-in-weeks-time/articleshow/86392325.cms>

1394. **WHO emergency approval for Covaxin delayed till October 5**

India Today

<https://www.indiatoday.in/coronavirus-outbreak/video/who-emergency-approval-for-covaxin-delayed-till-october-5-1856178-2021-09-23>

1395. **A Snapshot of the Global Race for Vaccines Targeting SARS-CoV-2 and the COVID-19 Pandemic**

Colin D Funk, Craig Laferrière, Ali Ardkani

Frontiers in Pharmacology (2020-06-19) <https://doi.org/gg4hxd>

DOI: [10.3389/fphar.2020.00937](https://doi.org/10.3389/fphar.2020.00937) · PMID: [32636754](#) · PMCID:

[PMC7317023](#)

1396. **Moderna's COVID-19 Vaccine Candidate Meets its Primary Efficacy Endpoint in the First Interim Analysis of the Phase 3 COVE Study | Moderna, Inc.** <https://investors.modernatx.com/news-releases/news-release-details/modernas-covid-19-vaccine-candidate-meets-its-primary-efficacy/>

1397. **Vaccines and Related Biological Products Advisory Committee December 17, 2020 Meeting Announcement - 12/17/2020 - 12/17/2020**

FDA

(2021-01-27) <https://www.fda.gov/advisory-committees/advisory-committee-calendar/vaccines-and-related-biological-products-advisory-committee-december-17-2020-meeting-announcement>

1398. **Vaccines and Related Biological Products Advisory Committee December 17, 2020 Meeting Briefing Document - FDA**

FDA/CBER

(2020-12-15) <https://www.fda.gov/media/144434/download>

1399. **Moderna Has Completed Case Accrual for First Planned Interim Analysis of its mRNA Vaccine Against COVID-19 (mRNA-1273) | Moderna, Inc.** <https://investors.modernatx.com/news-releases/news-release-details/moderna-has-completed-case-accrual-first-planned-interim/>

1400. **Guidance for Industry: Toxicity Grading Scale for Healthy Adult and Adolescent Volunteers Enrolled in Preventive Vaccine Clinical Trials**
CBER
(2018-10-08) <https://www.fda.gov/media/73679/download>
1401. **Health Canada Authorizes Moderna COVID-19 Vaccine in Canada | Moderna, Inc.** <https://investors.modernatx.com/news-releases/news-release-details/health-canada-authorizes-moderna-covid-19-vaccine-canada/>
1402. **EMA recommends COVID-19 Vaccine Moderna for authorisation in the EU**
Daniel GLANVILLE
European Medicines Agency (2021-01-06)
<https://www.ema.europa.eu/en/news/ema-recommends-covid-19-vaccine-moderna-authorisation-eu>
1403. **Pfizer and BioNTech Announce Vaccine Candidate Against COVID-19 Achieved Success in First Interim Analysis from Phase 3 Study | Pfizer** [https://www\(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-announce-vaccine-candidate-against](https://www(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-announce-vaccine-candidate-against)
1404. **Phase I/II study of COVID-19 RNA vaccine BNT162b1 in adults**
Mark J Mulligan, Kirsten E Lyke, Nicholas Kitchin, Judith Absalon, Alejandra Gurtman, Stephen Lockhart, Kathleen Neuzil, Vanessa Raabe, Ruth Bailey, Kena A Swanson, ... Kathrin U Jansen
Nature (2020-08-12) <https://doi.org/gg7ww9>
DOI: [10.1038/s41586-020-2639-4](https://doi.org/10.1038/s41586-020-2639-4) · PMID: [32785213](#)
1405. **COVID-19 vaccine BNT162b1 elicits human antibody and TH1 T cell responses**
Ugur Sahin, Alexander Muik, Evelyn Derhovanessian, Isabel Vogler, Lena M Kranz, Mathias Vormehr, Alina Baum, Kristen Pascal, Jasmin Quandt, Daniel Maurus, ... Özlem Türeci
Nature (2020-09-30) <https://doi.org/ghfmb2>
DOI: [10.1038/s41586-020-2814-7](https://doi.org/10.1038/s41586-020-2814-7) · PMID: [32998157](#)
1406. **Coronavirus COVID-19 Vaccine Update: Latest Developments | Pfizer** [https://www\(pfizer.com/science/coronavirus/vaccine](https://www(pfizer.com/science/coronavirus/vaccine)
1407. **Pfizer and BioNTech Conclude Phase 3 Study of COVID-19 Vaccine Candidate, Meeting All Primary Efficacy Endpoints | Pfizer**
[https://www\(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-conclude-phase-3-study-covid-19-vaccine](https://www(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-conclude-phase-3-study-covid-19-vaccine)
1408. **Covid-19: UK approves Pfizer and BioNTech vaccine with rollout due to start next week**
Elisabeth Mahase
BMJ (2020-12-02) <https://doi.org/ghpnhg>
DOI: [10.1136/bmj.m4714](https://doi.org/10.1136/bmj.m4714) · PMID: [33268330](#)
1409. **Covid-19 vaccine: First person receives Pfizer jab in UK**
BBC News
(2020-12-08) <https://www.bbc.com/news/uk-55227325>

1410. **Early effectiveness of COVID-19 vaccination with BNT162b2 mRNA vaccine and ChAdOx1 adenovirus vector vaccine on symptomatic disease, hospitalisations and mortality in older adults in England**
Jamie Lopez Bernal, Nick Andrews, Charlotte Gower, Julia Stowe, Chris Robertson, Elise Tessier, Ruth Simmons, Simon Cottrell, Richard Roberts, Mark O'Doherty, ... Mary Ramsay
Cold Spring Harbor Laboratory (2021-03-02) <https://doi.org/gh63t4>
DOI: [10.1101/2021.03.01.21252652](https://doi.org/10.1101/2021.03.01.21252652)
1411. **The arrival of Sputnik V**
Vijay Shankar Balakrishnan
The Lancet Infectious Diseases (2020-10) <https://doi.org/ghs3sn>
DOI: [10.1016/s1473-3099\(20\)30709-x](https://doi.org/10.1016/s1473-3099(20)30709-x) · PMID: [32979327](#) · PMCID: [PMC7511201](#)
1412. **Sputnik V COVID-19 vaccine candidate appears safe and effective**
Ian Jones, Polly Roy
The Lancet (2021-02) <https://doi.org/ghx7xz>
DOI: [10.1016/s0140-6736\(21\)00191-4](https://doi.org/10.1016/s0140-6736(21)00191-4) · PMID: [33545098](#) · PMCID: [PMC7906719](#)
1413. **International seroepidemiology of adenovirus serotypes 5, 26, 35, and 48 in pediatric and adult populations**
Dan H Barouch, Sandra V Kik, Gerrit J Weverling, Rebecca Dilan, Sharon L King, Lori F Maxfield, Sarah Clark, David Ng'ang'a, Kara L Brandariz, Peter Abbink, ... Jaap Goudsmit
Vaccine (2011-07) <https://doi.org/bmzpdx>
DOI: [10.1016/j.vaccine.2011.05.025](https://doi.org/10.1016/j.vaccine.2011.05.025) · PMID: [21619905](#) · PMCID: [PMC3138857](#)
1414. **Oxford-AstraZeneca COVID-19 vaccine efficacy**
Maria Deloria Knoll, Chizoba Wonodi
The Lancet (2021-01) <https://doi.org/ghpghz>
DOI: [10.1016/s0140-6736\(20\)32623-4](https://doi.org/10.1016/s0140-6736(20)32623-4) · PMID: [33306990](#) · PMCID: [PMC7832220](#)
1415. **Lead SARS-CoV-2 Candidate Vaccines: Expectations from Phase III Trials and Recommendations Post-Vaccine Approval**
Ebenezer Tumban
Viruses (2020-12-31) <https://doi.org/gh2z7h>
DOI: [10.3390/v13010054](https://doi.org/10.3390/v13010054) · PMID: [33396343](#) · PMCID: [PMC7824305](#)
1416. **A dangerous rush for vaccines**
HHolden Thorp
Science (2020-08-21) <https://doi.org/gh2pwb>
DOI: [10.1126/science.abe3147](https://doi.org/10.1126/science.abe3147) · PMID: [32792466](#)
1417. **Scientists worry whether Russia's 'Sputnik V' coronavirus vaccine is safe and effective**
Berkeley Lovelace Jr
CNBC (2020-08-11) <https://www.cnbc.com/2020/08/11/scientists-worry-whether-russias-sputnik-v-coronavirus-vaccine-is-safe-and-effective.html>

1418. Russia's claim of a successful COVID-19 vaccine doesn't pass the 'smell test,' critics say

Jon Cohen

Science (2020-11-11) <https://doi.org/gh2pwc>

DOI: [10.1126/science.abf6791](https://doi.org/10.1126/science.abf6791)

1419. Russia announces positive COVID-vaccine results from controversial trial

Ewen Callaway

Nature (2020-11-11) <https://doi.org/gh2pv9>

DOI: [10.1038/d41586-020-03209-0](https://doi.org/10.1038/d41586-020-03209-0) · PMID: [33177689](#)

1420. Covid-19: Russian vaccine efficacy is 91.6%, show phase III trial results

Elisabeth Mahase

BMJ (2021-02-02) <https://doi.org/gh2pwd>

DOI: [10.1136/bmj.n309](https://doi.org/10.1136/bmj.n309) · PMID: [33531342](#)

1421. Russia cuts size of COVID-19 vaccine study, stops enrollment

ABC News

ABC News <https://abcnews.go.com/Health/wireStory/russia-cuts-size-covid-19-vaccine-study-stops-74885458>

1422. About Sputnik V <https://sputnikvaccine.com/about-vaccine/>

1423. ACTIVE SURVEILLANCE OF THE SPUTNIK V VACCINE IN HEALTH WORKERS

Vanina Pagotto, Analía Ferloni, María Mercedes Soriano, Morena Díaz, Manuel Braguisnky Golde, María Isabel González, Valeria Asprea, Inés Staneloni, Gustavo Vidal, Martín Silveira, ... Silvana Figar

Cold Spring Harbor Laboratory (2021-02-05) <https://doi.org/gprj3m>

DOI: [10.1101/2021.02.03.21251071](https://doi.org/10.1101/2021.02.03.21251071)

1424. UPDATE 1-Russia's Sputnik V vaccine found safe in India mid-stage trial -Dr.Reddy's

Reuters

(2021-01-11) <https://www.reuters.com/article/health-coronavirus-india-vaccine-idUSL4N2JM2XA>

1425. RDIF, The Gamaleya National Center, AstraZeneca and R-Pharm sign an agreement to cooperate on COVID-19 vaccine development

The Russian Direct Investment Fund (RDIF)

<https://www.prnewswire.com/ae/news-releases/rdif-the-gamaleya-national-center-astrazeneca-and-r-pharm-sign-an-agreement-to-cooperate-on-covid-19-vaccine-development-301196874.html>

1426. Open-label, Non-randomized, Non-comparative, Phase II Study in Adult Subjects to Assess Safety and Immunogenicity of Combination of AZD1222, a Non-replicating ChAdOx1 Vector Vaccine, and rAd26-S, a Recombinant Adenovirus Type 26 Component of Gam-COVID-Vac Vaccine, for COVID-19 Prevention

R-Pharm

clinicaltrials.gov (2021-12-20)

<https://clinicaltrials.gov/ct2/show/NCT04686773>

1427. **The first registered COVID-19 vaccine** <https://sputnikvaccine.com/>
1428. **Johnson & Johnson Initiates Pivotal Global Phase 3 Clinical Trial of Janssen's COVID-19 Vaccine Candidate | Johnson & Johnson**
Content Lab U.S.
<https://www.jnj.com/johnson-johnson-initiates-pivotal-global-phase-3-clinical-trial-of-janssens-covid-19-vaccine-candidate>
1429. **Low-Dose Ad26.COV2.S Protection Against SARS-CoV-2 Challenge in Rhesus Macaques**
Xuan He, Abishek Chandrashekhar, Roland Zahn, Frank Wegmann, Jingyou Yu, Noe B Mercado, Katherine McMahan, Amanda J Martinot, Cesar Piedra-Mora, Sidney Beecy, ... Dan H Barouch
Cold Spring Harbor Laboratory (2021-01-27) <https://doi.org/gjhd3>
DOI: [10.1101/2021.01.27.428380](https://doi.org/10.1101/2021.01.27.428380) · PMID: [33532782](https://pubmed.ncbi.nlm.nih.gov/33532782/) · PMCID: [PMC7852276](https://pubmed.ncbi.nlm.nih.gov/PMC7852276/)
1430. **Safety and immunogenicity of the Ad26.COV2.S COVID-19 vaccine candidate: interim results of a phase 1/2a, double-blind, randomized, placebo-controlled trial**
Jerald Sadoff, Mathieu Le Gars, Georgi Shukarev, Dirk Heerwagh, Carla Truyers, Anne Marit de Groot, Jeroen Stoop, Sarah Tete, Wim Van Damme, Isabel Leroux-Roels, ... Hanneke Schuitemaker
Cold Spring Harbor Laboratory (2020-09-25) <https://doi.org/ghjk2q>
DOI: [10.1101/2020.09.23.20199604](https://doi.org/10.1101/2020.09.23.20199604)
1431. **Immunogenicity of the Ad26.COV2.S Vaccine for COVID-19**
Kathryn E Stephenson, Mathieu Le Gars, Jerald Sadoff, Anne Marit de Groot, Dirk Heerwagh, Carla Truyers, Caroline Atyeo, Carolin Loos, Abishek Chandrashekhar, Katherine McMahan, ... Dan H Barouch
JAMA (2021-04-20) <https://doi.org/gjhdz>
DOI: [10.1001/jama.2021.3645](https://doi.org/10.1001/jama.2021.3645) · PMID: [33704352](https://pubmed.ncbi.nlm.nih.gov/33704352/) · PMCID: [PMC7953339](https://pubmed.ncbi.nlm.nih.gov/PMC7953339/)
1432. **Correlates of protection against SARS-CoV-2 in rhesus macaques**
Katherine McMahan, Jingyou Yu, Noe B Mercado, Carolin Loos, Lisa H Tostanoski, Abishek Chandrashekhar, Jinyan Liu, Lauren Peter, Caroline Atyeo, Alex Zhu, ... Dan H Barouch
Nature (2020-12-04) <https://doi.org/fmjk>
DOI: [10.1038/s41586-020-03041-6](https://doi.org/10.1038/s41586-020-03041-6) · PMID: [33276369](https://pubmed.ncbi.nlm.nih.gov/33276369/) · PMCID: [PMC7906955](https://pubmed.ncbi.nlm.nih.gov/PMC7906955/)
1433. **A Randomized, Double-blind, Placebo-controlled Phase 3 Study to Assess the Efficacy and Safety of Ad26.COV2.S for the Prevention of SARS-CoV-2-mediated COVID-19 in Adults Aged 18 Years and Older**
Janssen Vaccines & Prevention B.V.
clinicaltrials.gov (2022-04-11)
<https://clinicaltrials.gov/ct2/show/NCT04505722>
1434. **Johnson & Johnson Prepares to Resume Phase 3 ENSEMBLE Trial of its Janssen COVID-19 Vaccine Candidate in the U.S. | Johnson & Johnson**
Content Lab U.S.
<https://www.jnj.com/our-company/johnson-johnson-prepares-to-resume-phase-3-ensemble-trial-of-its-janssen-covid-19-vaccine->

[candidate-in-the-us](#)

1435. **Johnson & Johnson Initiates Second Global Phase 3 Clinical Trial of its Janssen COVID-19 Vaccine Candidate | Johnson & Johnson**
Content Lab U.S.
<https://www.jnj.com/johnson-johnson-initiates-second-global-phase-3-clinical-trial-of-its-janssen-covid-19-vaccine-candidate>
1436. **A Review of the Progress and Challenges of Developing a Vaccine for COVID-19**
Omna Sharma, Ali A Sultan, Hong Ding, Chris R Triggle
Frontiers in Immunology (2020-10-14) <https://doi.org/gh65wd>
DOI: [10.3389/fimmu.2020.585354](https://doi.org/10.3389/fimmu.2020.585354) · PMID: [33163000](#) · PMCID: [PMC7591699](#)
1437. **Recombinant protein vaccines produced in insect cells**
Manon MJ Cox
Vaccine (2012-02) <https://doi.org/fx3mh3>
DOI: [10.1016/j.vaccine.2012.01.016](https://doi.org/10.1016/j.vaccine.2012.01.016) · PMID: [22265860](#) · PMCID: [PMC7115678](#)
1438. **Applications of nanotechnology for immunology**
Douglas M Smith, Jakub K Simon, James R Baker Jr
Nature Reviews Immunology (2013-07-25) <https://doi.org/gfzq8x>
DOI: [10.1038/nri3488](https://doi.org/10.1038/nri3488) · PMID: [23883969](#) · PMCID: [PMC7097370](#)
1439. **Matrix-M™ adjuvant enhances immunogenicity of both protein- and modified vaccinia virus Ankara-based influenza vaccines in mice**
Sofia E Magnusson, Arwen F Altenburg, Karin Lövgren Bengtsson, Fons Bosman, Rory D de Vries, Guus F Rimmelzwaan, Linda Stertman
Immunologic Research (2018-03-28) <https://doi.org/gdd2fw>
DOI: [10.1007/s12026-018-8991-x](https://doi.org/10.1007/s12026-018-8991-x) · PMID: [29594879](#) · PMCID: [PMC5899102](#)
1440. **Immune enhancing properties of the novel Matrix-M™ adjuvant leads to potentiated immune responses to an influenza vaccine in mice**
Sofia E Magnusson, Jenny M Reimer, Karin H Karlsson, Lena Lilja, Karin Lövgren Bengtsson, Linda Stertman
Vaccine (2013-03) <https://doi.org/f2ntg8>
DOI: [10.1016/j.vaccine.2013.01.039](https://doi.org/10.1016/j.vaccine.2013.01.039) · PMID: [23384754](#)
1441. **Intramuscular Matrix-M-adjuvanted virosomal H5N1 vaccine induces high frequencies of multifunctional Th1 CD4+ cells and strong antibody responses in mice**
Abdullah S Madhun, Lars R Haaheim, Mona V Nilsen, Rebecca J Cox
Vaccine (2009-12) <https://doi.org/d6cthn>
DOI: [10.1016/j.vaccine.2009.09.044](https://doi.org/10.1016/j.vaccine.2009.09.044) · PMID: [19781678](#)
1442. **Matrix-M adjuvanted virosomal H5N1 vaccine confers protection against lethal viral challenge in a murine model**
Gabriel Pedersen, Diane Major, Sarah Roseby, John Wood, Abdullah S Madhun, Rebecca J Cox

1443. **Evaluation of a virosomal H5N1 vaccine formulated with Matrix M™ adjuvant in a phase I clinical trial**

Rebecca J Cox, Gabriel Pedersen, Abdullah S Madhun, Signe Svindland, Marianne Sævik, Lucy Breakwell, Katja Hoschler, Marieke Willemsen, Laura Campitelli, Jane Kristin Nøstbakken, ... Haakon Sjursen

Vaccine (2011-10) <https://doi.org/dq7db6>

DOI: [10.1016/j.vaccine.2011.08.042](https://doi.org/10.1016/j.vaccine.2011.08.042) · PMID: [21864624](#)

1444. **A 2-Part, Phase 1/2, Randomized, Observer-Blinded Study To Evaluate The Safety And Immunogenicity Of A SARS-CoV-2 Recombinant Spike Protein Nanoparticle Vaccine (SARS-CoV-2 rS) With Or Without MATRIX-M™ Adjuvant In Healthy Subjects**

Novavax

clinicaltrials.gov (2021-12-02)

<https://clinicaltrials.gov/ct2/show/NCT04368988>

1445. **Evaluation of a SARS-CoV-2 Vaccine NVX-CoV2373 in Younger and Older Adults**

Neil Formica, Raburn Mallory, Gary Albert, Michelle Robinson, Joyce S Plested, Iksung Cho, Andreana Robertson, Filip Dubovsky, Gregory M Glenn

Cold Spring Harbor Laboratory (2021-03-01) <https://doi.org/gjh94p>

DOI: [10.1101/2021.02.26.21252482](https://doi.org/10.1101/2021.02.26.21252482)

1446. **Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC**

VStalin Raj, Huihui Mou, Saskia L Smits, Dick HW Dekkers, Marcel A Müller, Ronald Dijkman, Doreen Muth, Jeroen AA Demmers, Ali Zaki, Ron AM Fouchier, ... Bart L Haagmans

Nature (2013-03) <https://doi.org/f4qf89>

DOI: [10.1038/nature12005](https://doi.org/10.1038/nature12005) · PMID: [23486063](#) · PMCID: [PMC7095326](#)

1447. **Molecular basis of binding between novel human coronavirus MERS-CoV and its receptor CD26**

Guangwen Lu, Yawei Hu, Qihui Wang, Jianxun Qi, Feng Gao, Yan Li, Yanfang Zhang, Wei Zhang, Yuan Yuan, Jinku Bao, ... George F Gao

Nature (2013-07-07) <https://doi.org/m8z>

DOI: [10.1038/nature12328](https://doi.org/10.1038/nature12328) · PMID: [23831647](#) · PMCID: [PMC7095341](#)

1448. **The Receptor Binding Domain of the New Middle East Respiratory Syndrome Coronavirus Maps to a 231-Residue Region in the Spike Protein That Efficiently Elicits Neutralizing Antibodies**

Huihui Mou, VStalin Raj, Frank JM van Kuppeveld, Peter JM Rottier, Bart L Haagmans, Berend Jan Bosch

Journal of Virology (2013-08-15) <https://doi.org/ggg9rg>

DOI: [10.1128/jvi.01277-13](https://doi.org/10.1128/jvi.01277-13) · PMID: [23785207](#) · PMCID: [PMC3754068](#)

1449. **A Truncated Receptor-Binding Domain of MERS-CoV Spike Protein Potently Inhibits MERS-CoV Infection and Induces Strong**

Neutralizing Antibody Responses: Implication for Developing Therapeutics and Vaccines

Lanying Du, Zhihua Kou, Cuiqing Ma, Xinrong Tao, Lili Wang, Guangyu Zhao, Yaoqing Chen, Fei Yu, Chien-Te K Tseng, Yusen Zhou, Shibo Jiang

PLoS ONE (2013-12-04) <https://doi.org/gk657p>

DOI: [10.1371/journal.pone.0081587](https://doi.org/10.1371/journal.pone.0081587) · PMID: [24324708](#) · PMCID: [PMC3852489](#)

1450. Identification of an ideal adjuvant for receptor-binding domain-based subunit vaccines against Middle East respiratory syndrome coronavirus

Naru Zhang, Rudragouda Channappanavar, Cuiqing Ma, Lili Wang, Jian Tang, Tania Garron, Xinrong Tao, Sumaiya Tasneem, Lu Lu, Chien-Te K Tseng, ... Lanying Du

Cellular & Molecular Immunology (2015-02-02) <https://doi.org/f8vdhn>

DOI: [10.1038/cmi.2015.03](https://doi.org/10.1038/cmi.2015.03) · PMID: [25640653](#) · PMCID: [PMC4786625](#)

1451. Defining trained immunity and its role in health and disease

Mihai G Netea, Jorge Domínguez-Andrés, Luis B Barreiro, Triantafyllos Chavakis, Maziar Divangahi, Elaine Fuchs, Leo AB Joosten, Jos WM van der Meer, Musa M Mhlanga, Willem JM Mulder, ... Eicke Latz

Nature Reviews Immunology (2020-03-04) <https://doi.org/gg28pr>

DOI: [10.1038/s41577-020-0285-6](https://doi.org/s41577-020-0285-6) · PMID: [32132681](#) · PMCID: [PMC7186935](#)

1452. Trained Immunity: a Tool for Reducing Susceptibility to and the Severity of SARS-CoV-2 Infection

Mihai G Netea, Evangelos J Giamarellos-Bourboulis, Jorge Domínguez-Andrés, Nigel Curtis, Reinout van Crevel, Frank L van de Veerdonk, Marc Bonten

Cell (2020-05) <https://doi.org/gg2584>

DOI: [10.1016/j.cell.2020.04.042](https://doi.org/10.1016/j.cell.2020.04.042) · PMID: [32437659](#) · PMCID: [PMC7196902](#)

1453. Reducing Health Care Workers Absenteeism in Covid-19 Pandemic Through BCG Vaccine - Full Text View - ClinicalTrials.gov

<https://clinicaltrials.gov/ct2/show/NCT04328441>

1454. Application of BCG Vaccine for Immune-prophylaxis Among Egyptian Healthcare Workers During the Pandemic of COVID-19

Adel Khattab

clinicaltrials.gov (2020-04-17)

<https://clinicaltrials.gov/ct2/show/NCT04350931>

1455. Performance Evaluation of BCG Vaccination in Healthcare Personnel to Reduce the Severity of SARS-COV-2 Infection in Medellín, Colombia, 2020

Universidad de Antioquia

clinicaltrials.gov (2020-11-24)

<https://clinicaltrials.gov/ct2/show/NCT04362124>

1456. COVID-19: BCG As Therapeutic Vaccine, Transmission Limitation, and Immunoglobulin Enhancement - Full Text View - ClinicalTrials.gov

<https://clinicaltrials.gov/ct2/show/NCT04369794>

1457. **Using BCG Vaccine to Protect Health Care Workers in the COVID-19 Pandemic - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04373291>

1458. **Reducing Morbidity and Mortality in Health Care Workers Exposed to SARS-CoV-2 by Enhancing Non-specific Immune Responses Through Bacillus Calmette-Guérin Vaccination, a Randomized Controlled Trial**
TASK Applied Science
clinicaltrials.gov (2020-05-06)
<https://clinicaltrials.gov/ct2/show/NCT04379336>

1459. **Randomized Controlled Trial Evaluating the Efficacy of Vaccination With Bacillus Calmette and Guérin (BCG) in the Prevention of COVID-19 Via the Strengthening of Innate Immunity in Health Care Workers**
Assistance Publique - Hôpitaux de Paris
clinicaltrials.gov (2020-08-17)
<https://clinicaltrials.gov/ct2/show/NCT04384549>

1460. **Study to Assess VPM1002 in Reducing Healthcare Professionals' Absenteeism in COVID-19 Pandemic - Full Text View - ClinicalTrials.gov** <https://clinicaltrials.gov/ct2/show/NCT04387409>

1461. **A Randomized Clinical Trial for Enhanced Trained Immune Responses Through Bacillus Calmette-Guérin Vaccination to Prevent Infections by COVID-19: The ACTIVATE II Trial**
Hellenic Institute for the Study of Sepsis
clinicaltrials.gov (2020-07-10)
<https://clinicaltrials.gov/ct2/show/NCT04414267>

1462. **Reducing Hospital Admission of Elderly in SARS-CoV-2 Pandemic Via the Induction of Trained Immunity by Bacillus Calmette-Guérin Vaccination, a Randomized Controlled Trial**
Radboud University
clinicaltrials.gov (2020-06-03)
<https://clinicaltrials.gov/ct2/show/NCT04417335>

1463. **Study to Assess VPM1002 in Reducing Hospital Admissions and/or Severe Respiratory Infectious Diseases in Elderly in COVID-19 Pandemic - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04435379>

1464. **Efficacy and Safety of VPM1002 in Reducing SARS-CoV-2 (COVID-19) Infection Rate and Severity - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04439045>

1465. **Complex Immune Dysregulation in COVID-19 Patients with Severe Respiratory Failure**
Evangelos J Giamarellos-Bourboulis, Mihai G Netea, Nikoletta Rovina, Karolina Akinosoglou, Anastasia Antoniadou, Nikolaos Antonakos, Georgia Damoraki, Theologia Gkavogianni, Maria-Evangelia Adami, Paraskevi Katsaounou, ... Antonia Koutsoukou
Cell Host & Microbe (2020-06) <https://doi.org/ggthxs>

1466. **Type I and Type III Interferons – Induction, Signaling, Evasion, and Application to Combat COVID-19**

Annsea Park, Akiko Iwasaki

Cell Host & Microbe (2020-06) <https://doi.org/gg2ccp>

DOI: [10.1016/j.chom.2020.05.008](https://doi.org/10.1016/j.chom.2020.05.008) · PMID: [32464097](https://pubmed.ncbi.nlm.nih.gov/32464097/) · PMCID: [PMC7255347](https://pubmed.ncbi.nlm.nih.gov/PMC7255347/)

1467. **Viral Mutation Rates**

Rafael Sanjuán, Miguel R Nebot, Nicola Chirico, Louis M Mansky, Robert Belshaw

Journal of Virology (2010-10) <https://doi.org/bc7c55>

DOI: [10.1128/jvi.00694-10](https://doi.org/10.1128/jvi.00694-10) · PMID: [20660197](https://pubmed.ncbi.nlm.nih.gov/20660197/) · PMCID: [PMC2937809](https://pubmed.ncbi.nlm.nih.gov/PMC2937809/)

1468. **SARS-CoV-2 and influenza: a comparative overview and treatment implications**

Laura D Manzanares-Meza, Oscar Medina-Contreras

Boletín Médico del Hospital Infantil de México (2020-10-23)

<https://doi.org/gjj2n7>

DOI: [10.24875/bmhim.20000183](https://doi.org/10.24875/bmhim.20000183) · PMID: [33064680](https://pubmed.ncbi.nlm.nih.gov/33064680/)

1469. **Influenza evolution and H3N2 vaccine effectiveness, with application to the 2014/2015 season**

Xi Li, Michael W Deem

Protein Engineering Design and Selection (2016-06-16)

<https://doi.org/f856v5>

DOI: [10.1093/protein/gzw017](https://doi.org/10.1093/protein/gzw017) · PMID: [27313229](https://pubmed.ncbi.nlm.nih.gov/27313229/) · PMCID: [PMC4955871](https://pubmed.ncbi.nlm.nih.gov/PMC4955871/)

1470. **Neutralizing Activity of BNT162b2-Elicited Serum**

Yang Liu, Jianying Liu, Hongjie Xia, Xianwen Zhang, Camila R Fontes-Garfias, Kena A Swanson, Hui Cai, Ritu Sarkar, Wei Chen, Mark Cutler, ... Pei-Yong Shi

New England Journal of Medicine (2021-04-15) <https://doi.org/fwsc>

DOI: [10.1056/nejmc2102017](https://doi.org/10.1056/nejmc2102017) · PMID: [33684280](https://pubmed.ncbi.nlm.nih.gov/33684280/) · PMCID: [PMC7944950](https://pubmed.ncbi.nlm.nih.gov/PMC7944950/)

1471. **Covid-19 vaccine effectiveness affected by variants**

Pharmaceutical Technology

(2021-03-03) <https://www.pharmaceutical-technology.com/marketdata/covid-19-vaccine-effectiveness-affected-by-variants/>

1472. **The effects of virus variants on COVID-19 vaccines**

<https://www.who.int/news-room/feature-stories/detail/the-effects-of-virus-variants-on-covid-19-vaccines>

1473. **Predicting Influenza H3N2 Vaccine Efficacy From Evolution of the Dominant Epitope**

Melia E Bonomo, Michael W Deem

Clinical Infectious Diseases (2018-04-17) <https://doi.org/gf33js>

DOI: [10.1093/cid/ciy323](https://doi.org/10.1093/cid/ciy323) · PMID: [29672670](https://pubmed.ncbi.nlm.nih.gov/29672670/)

1474. **Looking beyond COVID-19 vaccine phase 3 trials**

Jerome H Kim, Florian Marks, John D Clemens

1475. **The challenges of distributing COVID-19 vaccinations**

Melinda C Mills, David Salisbury
EClinicalMedicine (2021-01) <https://doi.org/gh77b5>
DOI: [10.1016/j.eclim.2020.100674](https://doi.org/10.1016/j.eclim.2020.100674) · PMID: [33319186](#) · PMCID: [PMC7725651](#)

1476. **An ethical framework for global vaccine allocation**

Ezekiel J Emanuel, Govind Persad, Adam Kern, Allen Buchanan, Cécile Fabre, Daniel Halliday, Joseph Heath, Lisa Herzog, RJ Leland, Ephrem T Lemango, ... Henry S Richardson
Science (2020-09-03) <https://doi.org/ghz7k6>
DOI: [10.1126/science.abe2803](https://doi.org/10.1126/science.abe2803) · PMID: [32883884](#) · PMCID: [PMC8691258](#)

1477. **Vaccine optimization for COVID-19: Who to vaccinate first?**

Laura Matrajt, Julia Eaton, Tiffany Leung, Elizabeth R Brown
Science Advances (2021-02-05) <https://doi.org/ghz7k7>
DOI: [10.1126/sciadv.abf1374](https://doi.org/10.1126/sciadv.abf1374) · PMID: [33536223](#) · PMCID: [PMC8128110](#)

1478. **Model-informed COVID-19 vaccine prioritization strategies by age and serostatus**

Kate M Bubar, Kyle Reinholt, Stephen M Kissler, Marc Lipsitch, Sarah Cobey, Yonatan H Grad, Daniel B Larremore
Science (2021-02-26) <https://doi.org/ght4xk>
DOI: [10.1126/science.abe6959](https://doi.org/10.1126/science.abe6959) · PMID: [33479118](#) · PMCID: [PMC7963218](#)

1479. **Strategic spatiotemporal vaccine distribution increases the survival rate in an infectious disease like Covid-19**

Jens Grauer, Hartmut Löwen, Benno Liebchen
Scientific Reports (2020-12) <https://doi.org/ghq7vp>
DOI: [10.1038/s41598-020-78447-3](https://doi.org/10.1038/s41598-020-78447-3) · PMID: [33299029](#) · PMCID: [PMC7726577](#)

1480. **How should we conduct pandemic vaccination?**

Jane Williams, Chris Degeling, Jodie McVernon, Angus Dawson
Vaccine (2021-02) <https://doi.org/gh77b7>
DOI: [10.1016/j.vaccine.2020.12.059](https://doi.org/10.1016/j.vaccine.2020.12.059) · PMID: [33423839](#) · PMCID: [PMC7792561](#)

1481. **Vaccine ethics: an ethical framework for global distribution of COVID-19 vaccines**

Nancy S Jecker, Aaron G Wightman, Douglas S Diekema
Journal of Medical Ethics (2021-02-16) <https://doi.org/gh77cg>
DOI: [10.1136/medethics-2020-107036](https://doi.org/10.1136/medethics-2020-107036) · PMID: [33593876](#) · PMCID: [PMC7887861](#)

1482. **Optimal SARS-CoV-2 vaccine allocation using real-time seroprevalence estimates in Rhode Island and Massachusetts**

Thu Nguyen-Anh Tran, Nathan Wikle, Joseph Albert, Haider Inam, Emily Strong, Karel Brinda, Scott M Leighow, Fuhan Yang, Sajid Hossain, Justin R Pritchard, ... Maciej F Boni

1483. **Coronavirus Pandemic (COVID-19)**

Hannah Ritchie, Edouard Mathieu, Lucas Rodés-Guirao, Cameron Appel, Charlie Giattino, Esteban Ortiz-Ospina, Joe Hasell, Bobbie Macdonald, Diana Beltekian, Max Roser

Our World in Data (2020-03-05) <https://ourworldindata.org/covid-vaccination-policy>

1484. **Tracking Coronavirus Vaccinations Around the World**

Josh Holder

The New York Times (2021-01-29)

<https://www.nytimes.com/interactive/2021/world/covid-vaccinations-tracker.html>

1485. **Bloomberg - Are you a robot?**

<https://www.bloomberg.com/tosv2.html?vid=&uuid=fbf8301a-cfa1-11ec-a314-77575062466d&url=L2dyYXBoaWNzL2NvdmlkLXZhY2NpbmUtdHJhY2tlci1nbG9iYWwtZGlzdHJpYnV0aW9u>

1486. **One Vaccine Side Effect: Global Economic Inequality**

Peter S Goodman

The New York Times (2020-12-25)

<https://www.nytimes.com/2020/12/25/business/coronavirus-vaccines-global-economy.html>

1487. **Vaccine development and approval in Canada**

Health Canada

(2020-12-08) <https://www.canada.ca/en/health-canada/services/drugs-health-products/covid19-industry/drugs-vaccines-treatments/vaccines/development-approval-infographic.html>

1488. **Vaccines and treatments for COVID-19: Safety after authorization**

Public Health Agency of Canada

(2020-12-03) <https://www.canada.ca/en/public-health/services/diseases/2019-novel-coronavirus-infection/prevention-risks/covid-19-vaccine-treatment/safety-after-authorization.html>

1489. **Drug and vaccine authorizations for COVID-19: Applications received**

Health Canada

(2020-09-17) <https://www.canada.ca/en/health-canada/services/drugs-health-products/covid19-industry/drugs-vaccines-treatments/authorization/applications.html>

1490. **An earlier end date for vaccination campaign is 'possible', Trudeau says | CBC News**

John Paul Tasker · CBC News ·

CBC <https://www.cbc.ca/news/politics/trudeau-possible-vaccination-campaign-ends-sooner-1.5934994>

1491. **The Advisory Committee on Immunization Practices' Interim Recommendation for Allocating Initial Supplies of COVID-19 Vaccine — United States, 2020**

Kathleen Dooling, Nancy McClung, Mary Chamberland, Mona Marin, Megan Wallace, Beth P Bell, Grace M Lee, HKeipp Talbot, José R Romero, Sara E Oliver

MMWR. Morbidity and Mortality Weekly Report (2020-12-11)

<https://doi.org/gjkxrm>

DOI: [10.15585/mmwr.mm6949e1](https://doi.org/10.15585/mmwr.mm6949e1) · PMID: [33301429](https://pubmed.ncbi.nlm.nih.gov/33301429/) · PMCID: [PMC7737687](https://pubmed.ncbi.nlm.nih.gov/PMC7737687/)

1492. **The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Pfizer-BioNTech COVID-19 Vaccine — United States, December 2020**

Sara E Oliver, Julia W Gargano, Mona Marin, Megan Wallace, Kathryn G Curran, Mary Chamberland, Nancy McClung, Doug Campos-Outcalt, Rebecca L Morgan, Sarah Mbaeyi, ... Kathleen Dooling

MMWR. Morbidity and Mortality Weekly Report (2020-12-18)

<https://doi.org/ghvnsf>

DOI: [10.15585/mmwr.mm6950e2](https://doi.org/10.15585/mmwr.mm6950e2) · PMID: [33332292](https://pubmed.ncbi.nlm.nih.gov/33332292/) · PMCID: [PMC7745957](https://pubmed.ncbi.nlm.nih.gov/PMC7745957/)

1493. **US administers 1st doses of Pfizer coronavirus vaccine**

ABC News

ABC News <https://abcnews.go.com/US/us-administer-1st-doses-pfizer-coronavirus-vaccine/story?id=74703018>

1494. **The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Moderna COVID-19 Vaccine — United States, December 2020**

Sara E Oliver, Julia W Gargano, Mona Marin, Megan Wallace, Kathryn G Curran, Mary Chamberland, Nancy McClung, Doug Campos-Outcalt, Rebecca L Morgan, Sarah Mbaeyi, ... Kathleen Dooling

MMWR. Morbidity and Mortality Weekly Report (2021-01-01)

<https://doi.org/gh77ch>

DOI: [10.15585/mmwr.mm695152e1](https://doi.org/10.15585/mmwr.mm695152e1) · PMID: [33382675](https://pubmed.ncbi.nlm.nih.gov/33382675/)

1495. **The Advisory Committee on Immunization Practices' Updated Interim Recommendation for Allocation of COVID-19 Vaccine — United States, December 2020**

Kathleen Dooling, Mona Marin, Megan Wallace, Nancy McClung, Mary Chamberland, Grace M Lee, HKeipp Talbot, José R Romero, Beth P Bell, Sara E Oliver

MMWR. Morbidity and Mortality Weekly Report (2021-01-01)

<https://doi.org/ghqfv>

DOI: [10.15585/mmwr.mm695152e2](https://doi.org/10.15585/mmwr.mm695152e2) · PMID: [33382671](https://pubmed.ncbi.nlm.nih.gov/33382671/)

1496. **The Moderna vaccine is now in some Americans' arms as Covid-19 cases in the US pass 18 million**

Madeline Holcombe CNN Holly Yan and Steve Almasy

CNN <https://www.cnn.com/2020/12/21/health/us-coronavirus-monday/index.html>

1497. **Janssen COVID-19 Vaccine**

Office of the Commissioner

FDA (2022-05-09) <https://www.fda.gov/emergency-preparedness-and-response/coronavirus-disease-2019-covid-19/janssen-covid-19-vaccine>

1498. **The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Janssen COVID-19 Vaccine — United States, February 2021**

Sara E Oliver, Julia W Gargano, Heather Scobie, Megan Wallace, Stephen C Hadler, Jessica Leung, Amy E Blain, Nancy McClung, Doug Campos-Outcalt, Rebecca L Morgan, ... Kathleen Dooling
MMWR. Morbidity and Mortality Weekly Report (2021-03-05)
<https://doi.org/gh77cj>
DOI: [10.15585/mmwr.mm7009e4](https://doi.org/10.15585/mmwr.mm7009e4) · PMID: [33661860](https://pubmed.ncbi.nlm.nih.gov/33661860/) · PMCID: [PMC7948932](https://pubmed.ncbi.nlm.nih.gov/PMC7948932/)

1499. **WHO adds Janssen vaccine to list of safe and effective emergency tools against COVID-19** <https://www.who.int/news-room/detail/12-03-2021-who-adds-janssen-vaccine-to-list-of-safe-and-effective-emergency-tools-against-covid-19>

1500. **COVID-19 Vaccines**

Assistant Secretary for Public Affairs (ASPA)
HHS.gov (2020-12-12) <https://www.hhs.gov/coronavirus/covid-19-vaccines/index.html>

1501. **Biden now says US will have enough vaccine for every adult by the end of May | CNN Politics**

Kevin Liptak Harwood Jeff Zeleny, John
CNN (2021-03-02) <https://www.cnn.com/2021/03/02/politics/biden-merck-johnson--johnson-vaccine/index.html>

1502. **Covid-19: Was US vaccine rollout a 'dismal failure' under Trump?**

BBC News
(2021-01-26) <https://www.bbc.com/news/world-us-canada-55721437>

1503. **Structured to Fail: Lessons from the Trump Administration's Faulty Pandemic Planning and Response**

Alejandro E Camacho, Robert L Glicksman
(2021-01-21) <https://papers.ssrn.com/abstract=3770368>

1504. **The US Regulatory System and COVID-19 Vaccines**

Joshua M Sharfstein, Jesse L Goodman, Luciana Borio
JAMA (2021-03-23) <https://doi.org/gh77b3>
DOI: [10.1001/jama.2021.1961](https://doi.org/10.1001/jama.2021.1961) · PMID: [33587124](https://pubmed.ncbi.nlm.nih.gov/33587124/)

1505. **South Africa starts administering Janssen COVID-19 vaccine to health workers**

Rachel Arthur
BioPharma-Reporter (2021-02-18) <https://www.biopharma-reporter.com/Article/2021/02/18/South-Africa-starts-administering-Janssen-COVID-19-vaccine-to-health-workers>

1506. **EMA receives application for conditional marketing authorisation of COVID-19 Vaccine Janssen**

EMA

European Medicines Agency (2021-02-16)
<https://www.ema.europa.eu/en/news/ema-receives-application-conditional-marketing-authorisation-covid-19-vaccine-janssen>

1507. **Merck will help make Johnson & Johnson coronavirus vaccine as rivals team up to help Biden accelerate shots**

Washington Post

<https://www.washingtonpost.com/health/2021/03/02/merck-johnson-and-johnson-covid-vaccine-partnership/>

1508. **The UK has approved a COVID vaccine — here's what scientists now want to know**

Heidi Ledford, David Cyranoski, Richard Van Noorden

Nature (2020-12-03) <https://doi.org/gh4xmm>

DOI: [10.1038/d41586-020-03441-8](https://doi.org/10.1038/d41586-020-03441-8) · PMID: [33288887](#)

1509. **EMA recommends first COVID-19 vaccine for authorisation in the EU**

EMA

European Medicines Agency (2020-12-21)

<https://www.ema.europa.eu/en/news/ema-recommends-first-covid-19-vaccine-authorisation-eu>

1510. **Regulatory approval of COVID-19 Vaccine AstraZeneca**

GOV.UK

<https://www.gov.uk/government/publications/regulatory-approval-of-covid-19-vaccine-astrazeneca>

1511. **EMA recommends COVID-19 Vaccine AstraZeneca for authorisation in the EU**

EMA

European Medicines Agency (2021-01-29)

<https://www.ema.europa.eu/en/news/ema-recommends-covid-19-vaccine-astrazeneca-authorisation-eu>

1512. **Covid: Brian Pinker, 82, first to get Oxford-AstraZeneca vaccine**

BBC News

(2021-01-04) <https://www.bbc.com/news/uk-55525542>

1513. **Spikevax (previously COVID-19 Vaccine Moderna)**

EMA

European Medicines Agency (2021-01-04)

<https://www.ema.europa.eu/en/medicines/human/EPAR/spikevax>

1514. **Regulatory approval of Spikevax (formerly COVID-19 Vaccine Moderna)**

GOV.UK

<https://www.gov.uk/government/publications/regulatory-approval-of-covid-19-vaccine-moderna>

1515. **Bloomberg - Are you a robot?**

<https://www.bloomberg.com/tosv2.html?vid=&uuid=fc44db96-cfa1-11ec-81b7-4f63454e516a&url=L25Id3MvYXJ0aWNsZXMvcMjAyMC0xMi0wMi93aXRo>

[aW4taG91cnMtb2YtdS1rLXB1dGluLW9yZGVycy1zdGFydC1vZi1tYXNzLWNvdmlkLTE5LXNob3Rz](#)

1516. **Facing Record Covid-19 Case Rise, Russia Rolls Out Sputnik V Vaccine**

James Rodgers

Forbes

<https://www.forbes.com/sites/jamesrodgerseurope/2020/12/05/facing-record-covid-19-case-rise-russia-rolls-out-sputnik-v-vaccine/>

1517. **Clarification on Sputnik V vaccine in the EU approval process**

EMA

European Medicines Agency (2021-02-10)

<https://www.ema.europa.eu/en/news/clarification-sputnik-v-vaccine-eu-approval-process>

1518. **Countries are lining up for Russia's once-scorned Sputnik vaccine after strong efficacy results**

Fortune

<https://fortune.com/2021/02/08/international-sputnik-russia-demand/>

1519. **RDIF announces delivery of the first batch of Sputnik V vaccine to Venezuela for clinical trials**

<https://sputnikvaccine.com/newsroom/pressreleases/rdif-announces-delivery-of-the-first-batch-of-sputnik-v-vaccine-to-venezuela-for-clinical-trials/>

1520. **Unable to get U.S. vaccines, world turns to Russia and China**

Ryan Heath

POLITICO <https://www.politico.com/news/2021/02/25/global-vaccine-public-relations-war-471665>

1521. **Germany moves to bring Russian vaccine into EU orbit**

France 24

(2021-02-03) <https://www.france24.com/en/live-news/20210203-germany-moves-to-bring-russian-vaccine-into-eu-orbit>

1522. **Russia approves its third COVID-19 vaccine, Covivac**

Reuters

(2021-02-20) <https://www.reuters.com/article/us-health-coronavirus-russia-vaccine-idUSKBN2AK07H>

1523. **Intranasal Vaccine For Covid-19 | Bharat Biotech**

<https://www.bharatbiotech.com/intranasal-vaccine.html>

1524. **Novavax aims for 2 billion COVID-19 vaccine doses with expanded India deal**

Reuters

(2020-09-15) <https://www.reuters.com/article/health-coronavirus-novavax-idUSKBN2661PI>

1525. <https://ir.novavax.com/news-releases/news-release-details/novavax-announces-expanded-collaboration-and-license-agreement>

1526. **Covaxin: India approves two Covid vaccines for children under 12**

BBC News

(2022-04-26) <https://www.bbc.com/news/world-asia-india-55748124>

1527. **Vaccine Supply** <https://www.meaindia.gov.in/vaccine-supply.htm>

1528. **Coronavirus Vaccine Tracker**

Carl Zimmer, Jonathan Corum, Sui-Lee Wee, Matthew Kristoffersen
The New York Times (2020-06-10)

<https://www.nytimes.com/interactive/2020/science/coronavirus-vaccine-tracker.html>

1529. **China Wanted to Show Off Its Vaccines. It's Backfiring.**

Sui-Lee Wee
The New York Times (2021-01-25)
<https://www.nytimes.com/2021/01/25/business/china-covid-19-vaccine-backlash.html>

1530. **Coronavirus Vaccine Tracker**

Carl Zimmer, Jonathan Corum, Sui-Lee Wee, Matthew Kristoffersen
The New York Times (2020-06-10)
<https://www.nytimes.com/interactive/2020/science/coronavirus-vaccine-tracker.html>

1531. **Coronavirus Vaccine Tracker**

Carl Zimmer, Jonathan Corum, Sui-Lee Wee, Matthew Kristoffersen
The New York Times (2020-06-10)
<https://www.nytimes.com/interactive/2020/science/coronavirus-vaccine-tracker.html>

1532. **Philippines receives COVID-19 vaccine after delays**

ABC News
ABC News <https://abcnews.go.com/Health/wireStory/philippines-receive-covid-19-vaccine-delays-76163594>

1533. **China's Covid-19 Vaccine Makers Struggle to Meet Demand**

Chao Deng in Taipei and Jared Malsin in Dubai
Wall Street Journal (2021-02-10) <https://www.wsj.com/articles/chinas-covid-19-vaccine-makers-struggle-to-meet-demand-11612958560>

1534. **With First Dibs on Vaccines, Rich Countries Have 'Cleared the Shelves'**

Megan Twohey, Keith Collins, Katie Thomas
The New York Times (2020-12-15)
<https://www.nytimes.com/2020/12/15/us/coronavirus-vaccine-doses-reserved.html>

1535. **Covid-19 vaccinations: African nations miss WHO target**

BBC News
(2021-12-31) <https://www.bbc.com/news/56100076>

1536. **International Collaboration to Ensure Equitable Access to Vaccines for COVID-19: The ACT-Accelerator and the COVAX Facility**

MARK ECCLESTON-TURNER, HARRY UPTON
The Milbank Quarterly (2021-03-02) <https://doi.org/10.1111/1468-0009.12503>
DOI: [10.1111/1468-0009.12503](https://doi.org/10.1111/1468-0009.12503) · PMID: [33650737](https://pubmed.ncbi.nlm.nih.gov/33650737/) · PMCID: [PMC8014072](https://pubmed.ncbi.nlm.nih.gov/PMC8014072/)

1537. **Fair allocation mechanism for COVID-19 vaccines through the COVAX Facility** <https://www.who.int/publications/m/item/fair-allocation-mechanism-for-covid-19-vaccines-through-the-covax-facility>
1538. **Global plan seeks to promote vaccine equity, spread risks**
Kai Kupferschmidt
Science (2020-07-31) <https://doi.org/gh77cd>
DOI: [10.1126/science.369.6503.489](https://doi.org/10.1126/science.369.6503.489) · PMID: [32732400](#)
1539. **Covax must go beyond proportional allocation of covid vaccines to ensure fair and equitable access**
Lisa M Herzog, Ole F Norheim, Ezekiel J Emanuel, Matthew S McCoy
BMJ (2021-01-05) <https://doi.org/gjggjv>
DOI: [10.1136/bmj.m4853](https://doi.org/10.1136/bmj.m4853) · PMID: [33402340](#)
1540. **COVAX** <https://www.who.int/initiatives/act-accelerator/covax>
1541. **Countries now scrambling for COVID-19 vaccines may soon have surpluses to donate**
AAAS Articles DO Group
American Association for the Advancement of Science (AAAS) (2021-08-16) <https://doi.org/gh77cf>
DOI: [10.1126/science.abh4476](https://doi.org/10.1126/science.abh4476)
1542. **First COVID-19 COVAX vaccine doses administered in Africa**
<https://www.who.int/news/item/01-03-2021-first-covid-19-covax-vaccine-doses-administered-in-africa>
1543. **Pfizer and BioNTech to Submit Emergency Use Authorization Request Today to the U.S. FDA for COVID-19 Vaccine | Pfizer**
[https://www\(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-submit-emergency-use-authorization](https://www(pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-submit-emergency-use-authorization)
1544. **Moderna Announces First Participants Dosed in Phase 2/3 Study of COVID-19 Vaccine Candidate in Adolescents | Moderna, Inc.**
<https://investors.modernatx.com/news-releases/news-release-details/moderna-announces-first-participants-dosed-phase-23-study-covid/>
1545. **COVID-19: The Inflammation Link and the Role of Nutrition in Potential Mitigation**
Ioannis Zabetakis, Ronan Lordan, Catherine Norton, Alexandros Tsoupras
Nutrients (2020-05-19) <https://doi.org/ggxdq3>
DOI: [10.3390/nu12051466](https://doi.org/10.3390/nu12051466) · PMID: [32438620](#) · PMCID: [PMC7284818](#)
1546. **Could nutrition modulate COVID-19 susceptibility and severity of disease? A systematic review**
Philip T James, Zakari Ali, Andrew E Armitage, Ana Bonell, Carla Cerami, Hal Drakesmith, Modou Jobe, Kerry S Jones, Zara Liew, Sophie E Moore, ... Andrew M Prentice
Cold Spring Harbor Laboratory (2020-10-21) <https://doi.org/ghr94g>
DOI: [10.1101/2020.10.19.20214395](https://doi.org/10.1101/2020.10.19.20214395)
1547. **Coronavirus Disease 2019 (COVID-19) and Nutritional Status: The Missing Link?**

Renata Silverio, Daniela Caetano Gonçalves, Márcia Fábia Andrade,

Marilia Seelaender

Advances in Nutrition (2020-09-25) <https://doi.org/ghhqd>

DOI: [10.1093/advances/nmaa125](https://doi.org/10.1093/advances/nmaa125) · PMID: [32975565](https://pubmed.ncbi.nlm.nih.gov/32975565/) · PMCID:

[PMC7543263](https://pubmed.ncbi.nlm.nih.gov/PMC7543263/)

1548. Nutritional status of patients with COVID-19

Jae Hyoung Im, Young Soo Je, Jihyeon Baek, Moon-Hyun Chung, Hea Yoon Kwon, Jin-Soo Lee

International Journal of Infectious Diseases (2020-11)

<https://doi.org/gg7t5t>

DOI: [10.1016/j.ijid.2020.08.018](https://doi.org/10.1016/j.ijid.2020.08.018) · PMID: [32795605](https://pubmed.ncbi.nlm.nih.gov/32795605/) · PMCID:

[PMC7418699](https://pubmed.ncbi.nlm.nih.gov/PMC7418699/)

1549. Optimal Nutritional Status for a Well-Functioning Immune System

Is an Important Factor to Protect against Viral Infections

Philip Calder, Anitra Carr, Adrian Gombart, Manfred Eggersdorfer

Nutrients (2020-04-23) <https://doi.org/gg29hh>

DOI: [10.3390/nu12041181](https://doi.org/10.3390/nu12041181) · PMID: [32340216](https://pubmed.ncbi.nlm.nih.gov/32340216/) · PMCID: [PMC7230749](https://pubmed.ncbi.nlm.nih.gov/PMC7230749/)

1550. Peak dietary supplement sales leveling off during COVID-19 pandemic, but growth still remains strong over last year, market researchers report during webcast

Nutritional Outlook

<https://www.nutritionaloutlook.com/view/peak-dietary-supplement-sales-leveling-during-covid-19-pandemic-growth-still-remains-strong>

1551. Dietary Diversity among Chinese Residents during the COVID-19 Outbreak and Its Associated Factors

Ai Zhao, Zhongyu Li, Yalei Ke, Shanshan Huo, Yidi Ma, Yumei Zhang, Jian Zhang, Zhongxia Ren

Nutrients (2020-06-06) <https://doi.org/ghc6d9>

DOI: [10.3390/nu12061699](https://doi.org/10.3390/nu12061699) · PMID: [32517210](https://pubmed.ncbi.nlm.nih.gov/32517210/) · PMCID: [PMC7352896](https://pubmed.ncbi.nlm.nih.gov/PMC7352896/)

1552. Lockdown impact: Grocery stores bolstered NZ supplements sales as pharmacies slumped

nutraingredients-asia.com

[nutraingredients-asia.com https://www.nutraingredients-asia.com/Article/2020/07/06/Lockdown-impact-Grocery-stores-bolstered-NZ-supplements-sales-as-pharmacies-slumped](https://www.nutraingredients-asia.com/Article/2020/07/06/Lockdown-impact-Grocery-stores-bolstered-NZ-supplements-sales-as-pharmacies-slumped)

1553. COVID-19 temporarily bolsters European interest in supplements

.nutritioninsight.com/

<https://ni.cnsmedia.com/a/EHHJsDOG2oc=>

1554. India's immune health surge: Nation leads APAC in number of new product launches – new data

nutraingredients.com

[nutraingredients.com https://www.nutraingredients.com/Article/2020/07/21/India-s-immune-health-surge-Nation-leads-APAC-in-number-of-new-product-launches-new-data](https://www.nutraingredients.com/Article/2020/07/21/India-s-immune-health-surge-Nation-leads-APAC-in-number-of-new-product-launches-new-data)

1555. Food policy, nutrition and nutraceuticals in the prevention and management of COVID-19: Advice for healthcare professionals

Yasemin Ipek Ayseli, Nazli Aytekin, Derya Buyukkayhan, Ismail Aslan,

Mehmet Turan Ayseli

Trends in Food Science & Technology (2020-11) <https://doi.org/ghjtcp>

DOI: [10.1016/j.tifs.2020.09.001](https://doi.org/10.1016/j.tifs.2020.09.001) · PMID: [33519086](https://pubmed.ncbi.nlm.nih.gov/33519086/) · PMCID:

[PMC7834257](https://pubmed.ncbi.nlm.nih.gov/PMC7834257/)

1556. **5 Food and Beverage Trends in Europe During COVID-19**

<https://kerry.com/insights/kerrydigest/2020/5-food-and-beverage-trends-in-europe-during-covid-19>

1557. **Structural Design Principles for Delivery of Bioactive Components in Nutraceuticals and Functional Foods**

David Julian McClements, Eric Andrew Decker, Yeonhwa Park, Jochen Weiss

Critical Reviews in Food Science and Nutrition (2009-06-16)

<https://doi.org/dt68m4>

DOI: [10.1080/10408390902841529](https://doi.org/10.1080/10408390902841529) · PMID: [19484636](https://pubmed.ncbi.nlm.nih.gov/19484636/)

1558. **Nutraceutical therapies for atherosclerosis**

Joe WE Moss, Dipak P Ramji

Nature Reviews Cardiology (2016-07-07) <https://doi.org/f9g389>

DOI: [10.1038/nrcardio.2016.103](https://doi.org/10.1038/nrcardio.2016.103) · PMID: [27383080](https://pubmed.ncbi.nlm.nih.gov/27383080/) · PMCID:

[PMC5228762](https://pubmed.ncbi.nlm.nih.gov/PMC5228762/)

1559. **Nutraceutical-definition and introduction**

Ekta K Kalra

AAPS PharmSci (2015-07-10) <https://doi.org/cg5wc9>

DOI: [10.1208/ps050325](https://doi.org/10.1208/ps050325) · PMID: [14621960](https://pubmed.ncbi.nlm.nih.gov/14621960/) · PMCID: [PMC2750935](https://pubmed.ncbi.nlm.nih.gov/PMC2750935/)

1560. **Dietary Supplement Health and Education Act of 1994**

National Institutes of Health Office of Dietary Supplements

https://ods.od.nih.gov/About/DSHEA_Wording.aspx

1561. **Food and Drug Administration Modernization Act (FDAMA) of 1997**

Office of the Commissioner

FDA (2018-11-03) <https://www.fda.gov/regulatory-information/selected-amendments-fdc-act/food-and-drug-administration-modernization-act-fdama-1997>

1562. **Nutraceuticals - shedding light on the grey area between pharmaceuticals and food**

Antonello Santini, Ettore Novellino

Expert Review of Clinical Pharmacology (2018-04-23)

<https://doi.org/ggwztk>

DOI: [10.1080/17512433.2018.1464911](https://doi.org/10.1080/17512433.2018.1464911) · PMID: [29667442](https://pubmed.ncbi.nlm.nih.gov/29667442/)

1563. <https://eur-lex.europa.eu/legal-content/EN/ALL>

1564. <https://eur-lex.europa.eu/legal-content/EN/TXT>

1565. **EU Register of nutrition and health claims made on foods (v.3.5)**

https://ec.europa.eu/food/safety/labelling_nutrition/claims/register/public/

1566. **Nutraceuticals: opening the debate for a regulatory framework**

Antonello Santini, Silvia Miriam Cammarata, Giacomo Capone, Angela Ianaro, Gian Carlo Tenore, Luca Pani, Ettore Novellino
British Journal of Clinical Pharmacology (2018-04)
<https://doi.org/ggwztm>
DOI: [10.1111/bcp.13496](https://doi.org/10.1111/bcp.13496) · PMID: [29433155](https://pubmed.ncbi.nlm.nih.gov/29433155/) · PMCID: [PMC5867125](https://pubmed.ncbi.nlm.nih.gov/PMC5867125/)

1567. **Reviewing the Nutrition and Health Claims Regulation (EC) No. 1924/2006: What do we know about its challenges and potential impact on innovation?**

Stefanie Bröring, Sukhada Khedkar, Stefano Ciliberti
International Journal of Food Sciences and Nutrition (2016-08-02)
<https://doi.org/ghr936>
DOI: [10.1080/09637486.2016.1212816](https://doi.org/10.1080/09637486.2016.1212816) · PMID: [27484163](https://pubmed.ncbi.nlm.nih.gov/27484163/)

1568. **Dietary Supplements: Regulatory Challenges and Research Resources**

Johanna Dwyer, Paul Coates, Michael Smith
Nutrients (2018-01-04) <https://doi.org/ghr949>
DOI: [10.3390/nu10010041](https://doi.org/10.3390/nu10010041) · PMID: [29300341](https://pubmed.ncbi.nlm.nih.gov/29300341/) · PMCID: [PMC5793269](https://pubmed.ncbi.nlm.nih.gov/PMC5793269/)

1569. **Noetic Nutraceuticals - 607572 - 05/15/2020**

Center for Drug Evaluation and Research
Center for Drug Evaluation and Research (2020-05-18)
<https://www.fda.gov/inspections-compliance-enforcement-and-criminal-investigations/warning-letters/noetic-nutraceuticals-607572-05152020>

1570. **Regulations.gov** <https://beta.regulations.gov/document/FDA-2020-S-0023-0068>

1571. **Spartan Enterprises Inc. dba Watershed Wellness Center - 610876 - 10/30/2020**

Center for Drug Evaluation and Research
Center for Drug Evaluation and Research (2020-11-02)
<https://www.fda.gov/inspections-compliance-enforcement-and-criminal-investigations/warning-letters/spartan-enterprises-inc-dba-watershed-wellness-center-610876-10302020>

1572. **FTC Sues California Marketer of \$23,000 COVID-19 "Treatment" Plan**

Federal Trade Commission
(2020-07-31) <https://www.ftc.gov/news-events/press-releases/2020/07/ftc-sues-california-marketer-23000-covid-19-treatment-plan>

1573. <https://cen.acs.org/biological-chemistry/natural-products/oleandrin-compound-touted-possible-COVID/98/web/2020/08>

1574. **Reducing mortality from 2019-nCoV: host-directed therapies should be an option**

Alimuddin Zumla, David S Hui, Esam I Azhar, Ziad A Memish, Markus Maeurer
The Lancet (2020-02) <https://doi.org/ggkd3b>
DOI: [10.1016/s0140-6736\(20\)30305-6](https://doi.org/10.1016/s0140-6736(20)30305-6)

1575. **Diet Supplementation, Probiotics, and Nutraceuticals in SARS-CoV-2 Infection: A Scoping Review**
Fabio Infusino, Massimiliano Marazzato, Massimo Mancone, Francesco Fedele, Claudio Maria Mastroianni, Paolo Severino, Giancarlo Ceccarelli, Letizia Santinelli, Elena Cavarretta, Antonino GM Marullo, ...
Gabriella d'Ettorre
Nutrients (2020-06-08) <https://doi.org/gg8k58>
DOI: [10.3390/nu12061718](https://doi.org/10.3390/nu12061718) · PMID: [32521760](https://pubmed.ncbi.nlm.nih.gov/32521760/) · PMCID: [PMC7352781](https://pubmed.ncbi.nlm.nih.gov/PMC7352781/)
1576. **Potential interventions for novel coronavirus in China: A systematic review**
Lei Zhang, Yunhui Liu
Journal of Medical Virology (2020-03-03) <https://doi.org/ggpx57>
DOI: [10.1002/jmv.25707](https://doi.org/10.1002/jmv.25707) · PMID: [32052466](https://pubmed.ncbi.nlm.nih.gov/32052466/)
1577. **Nutraceuticals have potential for boosting the type 1 interferon response to RNA viruses including influenza and coronavirus**
Mark F McCarty, James J DiNicolantonio
Progress in Cardiovascular Diseases (2020-05) <https://doi.org/ggpwx2>
DOI: [10.1016/j.pcad.2020.02.007](https://doi.org/10.1016/j.pcad.2020.02.007) · PMID: [32061635](https://pubmed.ncbi.nlm.nih.gov/32061635/)
1578. **Inflammation and cardiovascular disease: are marine phospholipids the answer?**
Ronan Lordan, Shane Redfern, Alexandros Tsoupras, Ioannis Zabetakis
Food & Function (2020) <https://doi.org/gg29hg>
DOI: [10.1039/c9fo01742a](https://doi.org/10.1039/c9fo01742a) · PMID: [32270798](https://pubmed.ncbi.nlm.nih.gov/32270798/)
1579. **The Potential Beneficial Effect of EPA and DHA Supplementation Managing Cytokine Storm in Coronavirus Disease**
Zoltán Szabó, Tamás Marosvölgyi, Éva Szabó, Péter Bai, Mária Figler, Zsófia Verzár
Frontiers in Physiology (2020-06-19) <https://doi.org/gg4hz4>
DOI: [10.3389/fphys.2020.00752](https://doi.org/10.3389/fphys.2020.00752) · PMID: [32636763](https://pubmed.ncbi.nlm.nih.gov/32636763/) · PMCID: [PMC7318894](https://pubmed.ncbi.nlm.nih.gov/PMC7318894/)
1580. **Exploitation of Microalgae Species for Nutraceutical Purposes: Cultivation Aspects**
Sushanta Saha, Patrick Murray
Fermentation (2018-06-14) <https://doi.org/ghv64j>
DOI: [10.3390/fermentation4020046](https://doi.org/10.3390/fermentation4020046)
1581. **Prospective options of algae-derived nutraceuticals as supplements to combat COVID-19 and human coronavirus diseases**
Sachitra K Ratha, Nirmal Renuka, Ismail Rawat, Faizal Bux
Nutrition (2021-03) <https://doi.org/ghr93z>
DOI: [10.1016/j.nut.2020.111089](https://doi.org/10.1016/j.nut.2020.111089) · PMID: [33412367](https://pubmed.ncbi.nlm.nih.gov/33412367/) · PMCID: [PMC7680017](https://pubmed.ncbi.nlm.nih.gov/PMC7680017/)
1582. **Safety Aspects of Fish Oils**
Erik Berg Schmidt, Jørn Munkhof Møller, Niels Svaneborg, Jørn Dyerberg
Drug Investigation (2012-10-14) <https://doi.org/ghvqm8>
DOI: [10.1007/bf03257413](https://doi.org/10.1007/bf03257413)

1583. **Update on Seafood Consumption During Pregnancy**
[https://www.acog.org/en/Clinical/Clinical Guidance/Practice Advisory/Articles/2017/01/Update on Seafood Consumption During Pregnancy](https://www.acog.org/en/Clinical/Clinical%20Guidance/Practice%20Advisory/Articles/2017/01/Update%20on%20Seafood%20Consumption%20During%20Pregnancy)
1584. **Omega-3 Fatty Acid supplementation during pregnancy**
James A Greenberg, Stacey J Bell, Wendy Van Ausdal
Reviews in obstetrics & gynecology (2008)
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2621042/>
PMID: [19173020](#) · PMCID: [PMC2621042](#)
1585. **Omega-3 polyunsaturated fatty acids and inflammatory processes: nutrition or pharmacology?**
Philip C Calder
British Journal of Clinical Pharmacology (2013-03)
<https://doi.org/gggmrg>
DOI: [10.1111/j.1365-2125.2012.04374.x](#) · PMID: [22765297](#) · PMCID: [PMC3575932](#)
1586. **Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance**
Philip C Calder
Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids (2015-04) <https://doi.org/gf8pc6>
DOI: [10.1016/j.bbalip.2014.08.010](#) · PMID: [25149823](#)
1587. **N-3 polyunsaturated fatty acids modulate B cell activity in pre-clinical models: Implications for the immune response to infections**
Jarrett Whelan, Kymberly M Gowdy, Saame Raza Shaikh
European Journal of Pharmacology (2016-08) <https://doi.org/f8xn7q>
DOI: [10.1016/j.ejphar.2015.03.100](#) · PMID: [26022530](#) · PMCID: [PMC4662641](#)
1588. **Blood omega-3 fatty acids and death from COVID-19: A pilot study**
Arash Asher, Nathan L Tintle, Michael Myers, Laura Lockshon, Heribert Bacareza, William S Harris
Prostaglandins, Leukotrienes and Essential Fatty Acids (2021-03)
<https://doi.org/ghv63m>
DOI: [10.1016/j.plefa.2021.102250](#) · PMID: [33516093](#) · PMCID: [PMC7816864](#)
1589. **n-3 Polyunsaturated Fatty Acids Improve Inflammation via Inhibiting Sphingosine Kinase 1 in a Rat Model of Parenteral Nutrition and CLP-Induced Sepsis**
Tao Tian, Yunzhao Zhao, Qian Huang, Jieshou Li
Lipids (2016-02-08) <https://doi.org/ghvqm9>
DOI: [10.1007/s11745-016-4129-x](#) · PMID: [26856322](#)
1590. **Polyunsaturated fatty acids and sepsis**
Undurti N Das
Nutrition (2019-09) <https://doi.org/ghvqn8>
DOI: [10.1016/j.nut.2019.02.016](#) · PMID: [31029920](#)

1591. **Effects of an omega-3 fatty acid-enriched lipid emulsion on eicosanoid synthesis in acute respiratory distress syndrome (ARDS): A prospective, randomized, double-blind, parallel group study**

Joan Sabater, Joan Masclans, Judit Sacanell, Pilar Chacon, Pilar Sabin, Mercè Planas

Nutrition & Metabolism (2011) <https://doi.org/bg2kp>

DOI: [10.1186/1743-7075-8-22](https://doi.org/10.1186/1743-7075-8-22) · PMID: [21477318](https://pubmed.ncbi.nlm.nih.gov/21477318/) · PMCID: [PMC3080285](https://pubmed.ncbi.nlm.nih.gov/PMC3080285/)

1592. **Immunonutrition for Adults With ARDS: Results From a Cochrane Systematic Review and Meta-Analysis**

Ahilanandan Dushianthan, Rebecca Cusack, Victoria A Burgess, Michael PW Grocott, Philip Calder

Respiratory Care (2020-01) <https://doi.org/fj2p>

DOI: [10.4187/respcares.06965](https://doi.org/10.4187/respcares.06965) · PMID: [31506339](https://pubmed.ncbi.nlm.nih.gov/31506339/)

1593. **Correlation analysis of omega-3 fatty acids and mortality of sepsis and sepsis-induced ARDS in adults: data from previous randomized controlled trials**

HuaiSheng Chen, Su Wang, Ying Zhao, YuTian Luo, HuaSheng Tong, Lei Su

Nutrition Journal (2018-05-31) <https://doi.org/gdpjtq>

DOI: [10.1186/s12937-018-0356-8](https://doi.org/10.1186/s12937-018-0356-8) · PMID: [29859104](https://pubmed.ncbi.nlm.nih.gov/29859104/) · PMCID: [PMC5984323](https://pubmed.ncbi.nlm.nih.gov/PMC5984323/)

1594. **Proresolving Lipid Mediators and Mechanisms in the Resolution of Acute Inflammation**

Christopher D Buckley, Derek W Gilroy, Charles N Serhan

Immunity (2014-03) <https://doi.org/f5wntr>

DOI: [10.1016/j.immuni.2014.02.009](https://doi.org/10.1016/j.immuni.2014.02.009) · PMID: [24656045](https://pubmed.ncbi.nlm.nih.gov/24656045/) · PMCID: [PMC4004957](https://pubmed.ncbi.nlm.nih.gov/PMC4004957/)

1595. **Specialized pro-resolving mediators: endogenous regulators of infection and inflammation**

Maria C Basil, Bruce D Levy

Nature Reviews Immunology (2015-12-21) <https://doi.org/f9fgtd>

DOI: [10.1038/nri.2015.4](https://doi.org/10.1038/nri.2015.4) · PMID: [26688348](https://pubmed.ncbi.nlm.nih.gov/26688348/) · PMCID: [PMC5242505](https://pubmed.ncbi.nlm.nih.gov/PMC5242505/)

1596. **Specialized mediators in infection and lung injury**

Shayna Sandhaus, Andrew G Swick

BioFactors (2020-11-28) <https://doi.org/ghr93m>

DOI: [10.1002/biof.1691](https://doi.org/10.1002/biof.1691) · PMID: [33249673](https://pubmed.ncbi.nlm.nih.gov/33249673/) · PMCID: [PMC7744833](https://pubmed.ncbi.nlm.nih.gov/PMC7744833/)

1597. **Pro-resolving lipid mediators are leads for resolution physiology**

Charles N Serhan

Nature (2014-06-04) <https://doi.org/ggv53d>

DOI: [10.1038/nature13479](https://doi.org/10.1038/nature13479) · PMID: [24899309](https://pubmed.ncbi.nlm.nih.gov/24899309/) · PMCID: [PMC4263681](https://pubmed.ncbi.nlm.nih.gov/PMC4263681/)

1598. **The Specialized Proresolving Mediator 17-HDHA Enhances the Antibody-Mediated Immune Response against Influenza Virus: A New Class of Adjuvant?**

Sesquile Ramon, Steven F Baker, Julie M Sahler, Nina Kim, Eric A Feldsott, Charles N Serhan, Luis Martínez-Sobrido, David J Topham, Richard P Phipps

The Journal of Immunology (2014-12-15) <https://doi.org/f6spr8>

1599. **The Lipid Mediator Protectin D1 Inhibits Influenza Virus Replication and Improves Severe Influenza**

Masayuki Morita, Keiji Kuba, Akihiko Ichikawa, Mizuho Nakayama, Jun Katahira, Ryo Iwamoto, Tokiko Watanebe, Saori Sakabe, Tomo Daidoji, Shota Nakamura, ... Yumiko Imai

Cell (2013-03) <https://doi.org/f4rbgb>

DOI: [10.1016/j.cell.2013.02.027](https://doi.org/10.1016/j.cell.2013.02.027) · PMID: [23477864](https://pubmed.ncbi.nlm.nih.gov/23477864/)

1600. **Inflammation resolution: a dual-pronged approach to averting cytokine storms in COVID-19?**

Dipak Panigrahy, Molly M Gilligan, Sui Huang, Allison Gartung, Irene Cortés-Puch, Patricia J Sime, Richard P Phipps, Charles N Serhan, Bruce D Hammock

Cancer and Metastasis Reviews (2020-05-08) <https://doi.org/ggvv7w>

DOI: [10.1007/s1055-020-09889-4](https://doi.org/10.1007/s1055-020-09889-4) · PMID: [32385712](https://pubmed.ncbi.nlm.nih.gov/32385712/) · PMCID: [PMC7207990](https://pubmed.ncbi.nlm.nih.gov/PMC7207990/)

1601. **Pro resolving inflammatory effects of the lipid mediators of omega 3 fatty acids and its implication in SARS COVID-19**

Pedro-Antonio Regidor, Fernando Gonzalez Santos, Jose Miguel Rizo, Fernando Moreno Egea

Medical Hypotheses (2020-12) <https://doi.org/ghr93x>

DOI: [10.1016/j.mehy.2020.110340](https://doi.org/10.1016/j.mehy.2020.110340) · PMID: [33069094](https://pubmed.ncbi.nlm.nih.gov/33069094/) · PMCID: [PMC7543931](https://pubmed.ncbi.nlm.nih.gov/PMC7543931/)

1602. **Obesity-Driven Deficiencies of Specialized Pro-resolving Mediators May Drive Adverse Outcomes During SARS-CoV-2 Infection**

Anandita Pal, Kymberly M Gowdy, Kenneth J Oestreich, Melinda Beck, Saame Raza Shaikh

Frontiers in Immunology (2020-08-11) <https://doi.org/ght38j>

DOI: [10.3389/fimmu.2020.01997](https://doi.org/10.3389/fimmu.2020.01997) · PMID: [32983141](https://pubmed.ncbi.nlm.nih.gov/32983141/) · PMCID: [PMC7438933](https://pubmed.ncbi.nlm.nih.gov/PMC7438933/)

1603. **Fish Oil-Fed Mice Have Impaired Resistance to Influenza Infection**

Nicole MJ Schwerbrock, Erik A Karlsson, Qing Shi, Patricia A Sheridan, Melinda A Beck

The Journal of Nutrition (2009-08) <https://doi.org/dv45f4>

DOI: [10.3945/jn.109.108027](https://doi.org/10.3945/jn.109.108027) · PMID: [19549756](https://pubmed.ncbi.nlm.nih.gov/19549756/) · PMCID: [PMC2709305](https://pubmed.ncbi.nlm.nih.gov/PMC2709305/)

1604. **Modulation of host defence against bacterial and viral infections by omega-3 polyunsaturated fatty acids**

Marie-Odile Husson, Delphine Ley, Céline Portal, Madeleine Gottrand, Thomas Hueso, Jean-Luc Desseyn, Frédéric Gottrand

Journal of Infection (2016-12) <https://doi.org/f9pp2h>

DOI: [10.1016/j.jinf.2016.10.001](https://doi.org/10.1016/j.jinf.2016.10.001) · PMID: [27746159](https://pubmed.ncbi.nlm.nih.gov/27746159/)

1605. **Bioactive products formed in humans from fish oils**

Carsten Skarke, Naji Alamuddin, John A Lawson, Xuanwen Li, Jane F Ferguson, Muredach P Reilly, Garret A FitzGerald

Journal of Lipid Research (2015-09) <https://doi.org/f7pm5g>

DOI: [10.1194/jlr.m060392](https://doi.org/10.1194/jlr.m060392) · PMID: [26180051](https://pubmed.ncbi.nlm.nih.gov/26180051/) · PMCID: [PMC4548785](https://pubmed.ncbi.nlm.nih.gov/PMC4548785/)

1606. **Resolving Inflammatory Storm in COVID-19 Patients by Omega-3 Polyunsaturated Fatty Acids - A Single-blind, Randomized, Placebo-controlled Feasibility Study**

Magnus Bäck

clinicaltrials.gov (2020-11-27)

<https://clinicaltrials.gov/ct2/show/NCT04647604>

1607. **Stimulating the Resolution of Inflammation Through Omega-3 Polyunsaturated Fatty Acids in COVID-19: Rationale for the COVID-Omega-F Trial**

Hildur Arnardottir, Sven-Christian Pawelzik, Ulf Öhlund Wistbacka, Gonzalo Artiach, Robin Hofmann, Ingallil Reinholdsson, Frieder Braunschweig, Per Tornvall, Dorota Religa, Magnus Bäck

Frontiers in Physiology (2021-01-11) <https://doi.org/ghv64h>

DOI: [10.3389/fphys.2020.624657](https://doi.org/fphys.2020.624657) · PMID: [33505321](https://pubmed.ncbi.nlm.nih.gov/33505321/) · PMCID: [PMC7830247](https://pubmed.ncbi.nlm.nih.gov/PMC7830247/)

1608. **COVID-19 and its implications for thrombosis and anticoagulation**

Jean M Connors, Jerrold H Levy

Blood (2020-06-04) <https://doi.org/ggy35b>

DOI: [10.1182/blood.2020006000](https://doi.org/blood.2020006000) · PMID: [32339221](https://pubmed.ncbi.nlm.nih.gov/32339221/) · PMCID: [PMC7273827](https://pubmed.ncbi.nlm.nih.gov/PMC7273827/)

1609. **COVID-19 and Thrombotic or Thromboembolic Disease: Implications for Prevention, Antithrombotic Therapy, and Follow-Up**

Behnoor Bikdeli, Mahesh V Madhavan, David Jimenez, Taylor Chuich, Isaac Dreyfus, Elissa Driggin, Caroline Der Nigoghossian, Walter Ageno, Mohammad Madjid, Yutao Guo, ... Gregory YH Lip

Journal of the American College of Cardiology (2020-06)

<https://doi.org/ggsppk>

DOI: [10.1016/j.jacc.2020.04.031](https://doi.org/10.1016/j.jacc.2020.04.031) · PMID: [32311448](https://pubmed.ncbi.nlm.nih.gov/32311448/) · PMCID: [PMC7164881](https://pubmed.ncbi.nlm.nih.gov/PMC7164881/)

1610. **Thrombosis and COVID-19: The Potential Role of Nutrition**

Alexandros Tsoupras, Ronan Lordan, Ioannis Zabetakis

Frontiers in Nutrition (2020-09-25) <https://doi.org/ghr945>

DOI: [10.3389/fnut.2020.583080](https://doi.org/fnut.2020.583080) · PMID: [33102511](https://pubmed.ncbi.nlm.nih.gov/33102511/) · PMCID: [PMC7545367](https://pubmed.ncbi.nlm.nih.gov/PMC7545367/)

1611. **Regulation of platelet function and thrombosis by omega-3 and omega-6 polyunsaturated fatty acids**

Reheman Adili, Megan Hawley, Michael Holinstat

Prostaglandins & Other Lipid Mediators (2018-11)

<https://doi.org/ggv73>

DOI: [10.1016/j.prostaglandins.2018.09.005](https://doi.org/10.1016/j.prostaglandins.2018.09.005) · PMID: [30266534](https://pubmed.ncbi.nlm.nih.gov/30266534/) · PMCID: [PMC6242736](https://pubmed.ncbi.nlm.nih.gov/PMC6242736/)

1612. **Platelet activation and prothrombotic mediators at the nexus of inflammation and atherosclerosis: Potential role of antiplatelet agents**

Ronan Lordan, Alexandros Tsoupras, Ioannis Zabetakis

Blood Reviews (2020-04) <https://doi.org/ggv7x>

DOI: [10.1016/j.blre.2020.100694](https://doi.org/10.1016/j.blre.2020.100694) · PMID: [32340775](https://pubmed.ncbi.nlm.nih.gov/32340775/)

1613. **An Investigation on the Effects of Icosapent Ethyl (VascepaTM) on Inflammatory Biomarkers in Individuals With COVID-19 - Full Text View - ClinicalTrials.gov**

<https://clinicaltrials.gov/ct2/show/NCT04412018>

1614. **Cardiovascular Risk Reduction with Icosapent Ethyl for Hypertriglyceridemia**

Deepak L Bhatt, PGabriel Steg, Michael Miller, Eliot A Brinton, Terry A Jacobson, Steven B Ketchum, Ralph T Doyle, Rebecca A Juliano, Lixia Jiao, Craig Granowitz, ... Christie M Ballantyne

New England Journal of Medicine (2019-01-03) <https://doi.org/gfj3w9>

DOI: [10.1056/nejmoa1812792](https://doi.org/10.1056/nejmoa1812792) · PMID: [30415628](#)

1615. **A Randomised, Double-blind, Placebo Controlled Study of Eicosapentaenoic Acid (EPA-FFA) Gastro-resistant Capsules to Treat Hospitalised Subjects With Confirmed SARS-CoV-2**

S.L.A. Pharma AG

clinicaltrials.gov (2020-10-29)

<https://clinicaltrials.gov/ct2/show/NCT04335032>

1616. **Anti-inflammatory/Antioxidant Oral Nutrition Supplementation on the Cytokine Storm and Progression of COVID-19: A Randomized Controlled Trial**

Mahmoud Abulmeaty FACN M. D.

clinicaltrials.gov (2020-09-18)

<https://clinicaltrials.gov/ct2/show/NCT04323228>

1617. **Functional Role of Dietary Intervention to Improve the Outcome of COVID-19: A Hypothesis of Work**

Giovanni Messina, Rita Polito, Vincenzo Monda, Luigi Cipolloni, Nunzio Di Nunno, Giulio Di Mizio, Paolo Murabito, Marco Carotenuto, Antonietta Messina, Daniela Pisanelli, ... Francesco Sessa

International Journal of Molecular Sciences (2020-04-28)

<https://doi.org/ggvb88>

DOI: [10.3390/ijms21093104](https://doi.org/10.3390/ijms21093104) · PMID: [32354030](#) · PMCID: [PMC7247152](#)

1618. **Zinc and immunity: An essential interrelation**

Maria Maares, Hajo Haase

Archives of Biochemistry and Biophysics (2016-12)

<https://doi.org/f9c9b5>

DOI: [10.1016/j.abb.2016.03.022](https://doi.org/10.1016/j.abb.2016.03.022) · PMID: [27021581](#)

1619. **Zinc-Dependent Suppression of TNF- α Production Is Mediated by Protein Kinase A-Induced Inhibition of Raf-1, I κ B Kinase β , and NF- κ B**

Verena von Bülow, Svenja Dubben, Gabriela Engelhardt, Silke Hebel, Birgit Plümäkers, Holger Heine, Lothar Rink, Hajo Haase

The Journal of Immunology (2007-09-15) <https://doi.org/f3vs45>

DOI: [10.4049/jimmunol.179.6.4180](https://doi.org/10.4049/jimmunol.179.6.4180) · PMID: [17785857](#)

1620. **Zinc activates NF- κ B in HUT-78 cells**

Ananda S Prasad, Bin Bao, Frances WJ Beck, Fazlul H Sarkar

Journal of Laboratory and Clinical Medicine (2001-10)

<https://doi.org/cnc6fr>

DOI: [10.1067/mlc.2001.118108](https://doi.org/10.1067/mlc.2001.118108) · PMID: [11574819](#)

1621. Innate or Adaptive Immunity? The Example of Natural Killer Cells

E Vivier, DH Raulet, A Moretta, MA Caligiuri, L Zitvogel, LL Lanier, WM Yokoyama, S Ugolini

Science (2011-01-06) <https://doi.org/ckzg9g>

DOI: [10.1126/science.1198687](https://doi.org/10.1126/science.1198687) · PMID: [21212348](https://pubmed.ncbi.nlm.nih.gov/21212348/) · PMCID: [PMC3089969](https://pubmed.ncbi.nlm.nih.gov/PMC3089969/)

1622. Zinc supplementation decreases incidence of infections in the elderly: effect of zinc on generation of cytokines and oxidative stress

Ananda S Prasad, Frances WJ Beck, Bin Bao, James T Fitzgerald, Diane C Snell, Joel D Steinberg, Lavoisier J Cardozo

The American Journal of Clinical Nutrition (2007-03)

<https://doi.org/ggqmgs>

DOI: [10.1093/ajcn/85.3.837](https://doi.org/10.1093/ajcn/85.3.837) · PMID: [17344507](https://pubmed.ncbi.nlm.nih.gov/17344507/)

1623. The Role of Zinc in Antiviral Immunity

Scott A Read, Stephanie Obeid, Chantelle Ahlenstiel, Golo Ahlenstiel

Advances in Nutrition (2019-07) <https://doi.org/ggqmgr>

DOI: [10.1093/advances/nmz013](https://doi.org/10.1093/advances/nmz013) · PMID: [31305906](https://pubmed.ncbi.nlm.nih.gov/31305906/) · PMCID:

[PMC6628855](https://pubmed.ncbi.nlm.nih.gov/PMC6628855/)

1624. Efficacy of Zinc Against Common Cold Viruses: An Overview

Darrell Hulisz

Journal of the American Pharmacists Association (2004-09)

<https://doi.org/cf6pmt>

DOI: [10.1331/1544-3191.44.5.594.hulisz](https://doi.org/10.1331/1544-3191.44.5.594.hulisz) · PMID: [15496046](https://pubmed.ncbi.nlm.nih.gov/15496046/)

1625. Zinc Lozenges May Shorten the Duration of Colds: A Systematic Review

Harri Hemilä

The Open Respiratory Medicine Journal (2011-06-23)

<https://doi.org/bndmfq>

DOI: [10.2174/1874306401105010051](https://doi.org/10.2174/1874306401105010051) · PMID: [21769305](https://pubmed.ncbi.nlm.nih.gov/21769305/) · PMCID:

[PMC3136969](https://pubmed.ncbi.nlm.nih.gov/PMC3136969/)

1626. COVID-19: Poor outcomes in patients with zinc deficiency

Dinesh Jothimani, Ezhilarasan Kailasam, Silas Danielraj, Balaji Nallathambi, Hemalatha Ramachandran, Padmini Sekar, Shruthi Manoharan, Vidyalakshmi Ramani, Gomathy Narasimhan, Ilankumaran Kaliamoorthy, Mohamed Rela

International Journal of Infectious Diseases (2020-11)

<https://doi.org/ghr93t>

DOI: [10.1016/j.ijid.2020.09.014](https://doi.org/10.1016/j.ijid.2020.09.014) · PMID: [32920234](https://pubmed.ncbi.nlm.nih.gov/32920234/) · PMCID:

[PMC7482607](https://pubmed.ncbi.nlm.nih.gov/PMC7482607/)

1627. Zn²⁺ Inhibits Coronavirus and Arterivirus RNA Polymerase Activity In Vitro and Zinc Ionophores Block the Replication of These Viruses in Cell Culture

Aartjan JW te Velthuis, Sjoerd HE van den Worm, Amy C Sims, Ralph S Baric, Eric J Snijder, Martijn J van Hemert

PLoS Pathogens (2010-11-04) <https://doi.org/d95x4g>

DOI: [10.1371/journal.ppat.1001176](https://doi.org/10.1371/journal.ppat.1001176) · PMID: [21079686](https://pubmed.ncbi.nlm.nih.gov/21079686/) · PMCID:

[PMC2973827](https://pubmed.ncbi.nlm.nih.gov/PMC2973827/)

1628. **The SARS-coronavirus papain-like protease: Structure, function and inhibition by designed antiviral compounds**
Yahira M Báez-Santos, Sarah E St. John, Andrew D Mesecar
Antiviral Research (2015-03) <https://doi.org/f63hjp>
DOI: [10.1016/j.antiviral.2014.12.015](https://doi.org/10.1016/j.antiviral.2014.12.015) · PMID: [25554382](#) · PMCID: [PMC5896749](#)
1629. **A Randomized Study Evaluating the Safety and Efficacy of Hydroxychloroquine and Zinc in Combination With Either Azithromycin or Doxycycline for the Treatment of COVID-19 in the Outpatient Setting**
Avni Thakore MD
clinicaltrials.gov (2020-12-08)
<https://clinicaltrials.gov/ct2/show/NCT04370782>
1630. **A Study of Hydroxychloroquine and Zinc in the Prevention of COVID-19 Infection in Military Healthcare Workers - Full Text View - ClinicalTrials.gov** <https://clinicaltrials.gov/ct2/show/NCT04377646>
1631. **Therapies to Prevent Progression of COVID-19, Including Hydroxychloroquine, Azithromycin, Zinc, Vitamin D, Vitamin B12 With or Without Vitamin C, a Multi-centre, International, Randomized Trial: The International ALLIANCE Study**
National Institute of Integrative Medicine, Australia
clinicaltrials.gov (2020-09-09)
<https://clinicaltrials.gov/ct2/show/NCT04395768>
1632. **Early Intervention in COVID-19: Favipiravir Versus Standard Care - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04373733>
1633. **Hydroxychloroquine with or without Azithromycin in Mild-to-Moderate Covid-19**
Alexandre B Cavalcanti, Fernando G Zampieri, Regis G Rosa, Luciano CP Azevedo, Viviane C Veiga, Alvaro Avezum, Lucas P Damiani, Aline Marcadenti, Letícia Kawano-Dourado, Thiago Lisboa, ... Otávio Berwanger
New England Journal of Medicine (2020-11-19) <https://doi.org/gg5343>
DOI: [10.1056/nejmoa2019014](https://doi.org/10.1056/nejmoa2019014) · PMID: [32706953](#) · PMCID: [PMC7397242](#)
1634. **The efficacy and safety of hydroxychloroquine for COVID-19 prophylaxis: A systematic review and meta-analysis of randomized trials**
Kimberley Lewis, Dipayan Chaudhuri, Fayed Alshamsi, Laiya Carayannopoulos, Karin Dearness, Zain Chagla, Waleed Alhazzani, for the GUIDE Group
PLOS ONE (2021-01-06) <https://doi.org/ghsv36>
DOI: [10.1371/journal.pone.0244778](https://doi.org/10.1371/journal.pone.0244778) · PMID: [33406138](#) · PMCID: [PMC7787432](#)
1635. **Effect of hydroxychloroquine with or without azithromycin on the mortality of coronavirus disease 2019 (COVID-19) patients: a systematic review and meta-analysis**
Thibault Fiolet, Anthony Guihur, Mathieu Edouard Rebeaud, Matthieu Mulot, Nathan Peiffer-Smadja, Yahya Mahamat-Saleh

1636. Zinc sulfate in combination with a zinc ionophore may improve outcomes in hospitalized COVID-19 patients

Philip M Carlucci, Tania Ahuja, Christopher Petrilli, Harish Rajagopalan, Simon Jones, Joseph Rahimian

Journal of Medical Microbiology (2020-10-01) <https://doi.org/ghnws7>

DOI: [10.1099/jmm.0.001250](https://doi.org/10.1099/jmm.0.001250) · PMID: [32930657](https://pubmed.ncbi.nlm.nih.gov/32930657/) · PMCID: [PMC7660893](https://pubmed.ncbi.nlm.nih.gov/PMC7660893/)

1637. The Minimal Effect of Zinc on the Survival of Hospitalized Patients With COVID-19

Jasper Seth Yao, Joseph Alexander Paguio, Edward Christopher Dee, Hanna Clementine Tan, Achintya Moulick, Carmelo Milazzo, Jerry Jurado, Nicolás Della Penna, Leo Anthony Celi

Chest (2021-01) <https://doi.org/gg5w36>

DOI: [10.1016/j.chest.2020.06.082](https://doi.org/10.1016/j.chest.2020.06.082) · PMID: [32710890](https://pubmed.ncbi.nlm.nih.gov/32710890/) · PMCID: [PMC7375307](https://pubmed.ncbi.nlm.nih.gov/PMC7375307/)

1638. Coronavirus Disease 2019- Using Ascorbic Acid and Zinc Supplementation (COVIDAtoZ) Research Study A Randomized, Open Label Single Center Study

Milind Desai

clinicaltrials.gov (2021-01-28)

<https://clinicaltrials.gov/ct2/show/NCT04342728>

1639. Vitamin B12 May Inhibit RNA-Dependent-RNA Polymerase Activity of nsp12 from the COVID-19 Virus

Naveen Narayanan, Deepak T Nair

Preprints (2020-03-22) <https://doi.org/ggqmjc>

DOI: [10.20944/preprints202003.0347.v1](https://doi.org/10.20944/preprints202003.0347.v1)

1640. The Long History of Vitamin C: From Prevention of the Common Cold to Potential Aid in the Treatment of COVID-19

Giuseppe Cerullo, Massimo Negro, Mauro Parimbelli, Michela Pecoraro, Simone Perna, Giorgio Liguori, Mariangela Rondanelli, Hellas Cena, Giuseppe D'Antona

Frontiers in Immunology (2020-10-28) <https://doi.org/ghr943>

DOI: [10.3389/fimmu.2020.574029](https://doi.org/10.3389/fimmu.2020.574029) · PMID: [33193359](https://pubmed.ncbi.nlm.nih.gov/33193359/) · PMCID: [PMC7655735](https://pubmed.ncbi.nlm.nih.gov/PMC7655735/)

1641. The Emerging Role of Vitamin C in the Prevention and Treatment of COVID-19

Anitra C Carr, Sam Rowe

Nutrients (2020-10-27) <https://doi.org/ghr95c>

DOI: [10.3390/nu12113286](https://doi.org/10.3390/nu12113286) · PMID: [33121019](https://pubmed.ncbi.nlm.nih.gov/33121019/) · PMCID: [PMC7693980](https://pubmed.ncbi.nlm.nih.gov/PMC7693980/)

1642. Vitamin C Mitigates Oxidative Stress and Tumor Necrosis Factor-Alpha in Severe Community-Acquired Pneumonia and LPS-Induced Macrophages

Yuanyuan Chen, Guangyan Luo, Jiao Yuan, Yuanyuan Wang, Xiaoqiong Yang, Xiaoyun Wang, Guoping Li, Zhiguang Liu, Nanshan Zhong

Mediators of Inflammation (2014) <https://doi.org/f6nb5f>

DOI: [10.1155/2014/426740](https://doi.org/10.1155/2014/426740) · PMID: [25253919](https://pubmed.ncbi.nlm.nih.gov/25253919/) · PMCID: [PMC4165740](https://pubmed.ncbi.nlm.nih.gov/PMC4165740/)

1643. **Intravenous infusion of ascorbic acid decreases serum histamine concentrations in patients with allergic and non-allergic diseases**
Alexander F Hagel, Christian M Layritz, Wolfgang H Hagel, Hans-Jürgen Hagel, Edith Hagel, Wolfgang Dauth, Jürgen Kressel, Tanja Regnet, Andreas Rosenberg, Markus F Neurath, ... Martin Raithel
Naunyn-Schmiedeberg's Archives of Pharmacology (2013-05-11)
<https://doi.org/f48jsb>
DOI: [10.1007/s00210-013-0880-1](https://doi.org/10.1007/s00210-013-0880-1) · PMID: [23666445](https://pubmed.ncbi.nlm.nih.gov/23666445/)
1644. **Vitamin C and Immune Function**
Anitra Carr, Silvia Maggini
Nutrients (2017-11-03) <https://doi.org/gfzrjs>
DOI: [10.3390/nu9111211](https://doi.org/10.3390/nu9111211) · PMID: [29099763](https://pubmed.ncbi.nlm.nih.gov/29099763/) · PMCID: [PMC5707683](https://pubmed.ncbi.nlm.nih.gov/PMC5707683/)
1645. **Changes in Leucocyte Ascorbic Acid during the Common Cold**
R Hume, Elspeth Weyers
Scottish Medical Journal (2016-06-25) <https://doi.org/ggqrjf>
DOI: [10.1177/003693307301800102](https://doi.org/10.1177/003693307301800102) · PMID: [4717661](https://pubmed.ncbi.nlm.nih.gov/4717661/)
1646. **ASCORBIC ACID FUNCTION AND METABOLISM DURING COLDS**
CWM Wilson
Annals of the New York Academy of Sciences (1975-09)
<https://doi.org/bjfdtb>
DOI: [10.1111/j.1749-6632.1975.tb29312.x](https://doi.org/10.1111/j.1749-6632.1975.tb29312.x) · PMID: [1106304](https://pubmed.ncbi.nlm.nih.gov/1106304/)
1647. **Metabolism of ascorbic acid (vitamin C) in subjects infected with common cold viruses**
JEW Davies, RE Hughes, Eleri Jones, Sylvia E Reed, JW Craig, DAJ Tyrrell
Biochemical Medicine (1979-02) <https://doi.org/fd22sv>
DOI: [10.1016/0006-2944\(79\)90058-9](https://doi.org/10.1016/0006-2944(79)90058-9)
1648. **Vitamin C and Infections**
Harri Hemilä
Nutrients (2017-03-29) <https://doi.org/gfkbn>
DOI: [10.3390/nu9040339](https://doi.org/10.3390/nu9040339) · PMID: [28353648](https://pubmed.ncbi.nlm.nih.gov/28353648/) · PMCID: [PMC5409678](https://pubmed.ncbi.nlm.nih.gov/PMC5409678/)
1649. **Vitamin C and the common cold**
Harri Hemilä
British Journal of Nutrition (2007-03-09) <https://doi.org/fszhc6>
DOI: [10.1079/bjn19920004](https://doi.org/10.1079/bjn19920004) · PMID: [1547201](https://pubmed.ncbi.nlm.nih.gov/1547201/)
1650. **Vitamin C Can Shorten the Length of Stay in the ICU: A Meta-Analysis**
Harri Hemilä, Elizabeth Chalker
Nutrients (2019-03-27) <https://doi.org/gfzscg>
DOI: [10.3390/nu11040708](https://doi.org/10.3390/nu11040708) · PMID: [30934660](https://pubmed.ncbi.nlm.nih.gov/30934660/) · PMCID: [PMC6521194](https://pubmed.ncbi.nlm.nih.gov/PMC6521194/)
1651. **Serum Levels of Vitamin C and Vitamin D in a Cohort of Critically Ill COVID-19 Patients of a North American Community Hospital Intensive Care Unit in May 2020: A Pilot Study**
Cristian Arvinte, Maharaj Singh, Paul E Marik
Medicine in Drug Discovery (2020-12) <https://doi.org/ghnwqt>
DOI: [10.1016/j.medidd.2020.100064](https://doi.org/10.1016/j.medidd.2020.100064) · PMID: [32964205](https://pubmed.ncbi.nlm.nih.gov/32964205/) · PMCID: [PMC7499070](https://pubmed.ncbi.nlm.nih.gov/PMC7499070/)

1652. **Vitamin C levels in patients with SARS-CoV-2-associated acute respiratory distress syndrome**

Luis Chiscano-Camón, Juan Carlos Ruiz-Rodriguez, Adolf Ruiz-Sanmartin, Oriol Roca, Ricard Ferrer

Critical Care (2020-08-26) <https://doi.org/ghbr97>

DOI: [10.1186/s13054-020-03249-y](https://doi.org/10.1186/s13054-020-03249-y) · PMID: [32847620](#) · PMCID: [PMC7447967](#)

1653. **Targeting coagulation activation in severe COVID-19 pneumonia: lessons from bacterial pneumonia and sepsis**

Ricardo J José, Andrew Williams, Ari Manuel, Jeremy S Brown, Rachel C Chambers

European Respiratory Review (2020-10-01) <https://doi.org/ghr94s>

DOI: [10.1183/16000617.0240-2020](https://doi.org/10.1183/16000617.0240-2020) · PMID: [33004529](#) · PMCID: [PMC7537941](#)

1654. **Vitamin C and Microvascular Dysfunction in Systemic Inflammation**

Karel Tyml

Antioxidants (2017-06-29) <https://doi.org/ghr947>

DOI: [10.3390/antiox6030049](https://doi.org/10.3390/antiox6030049) · PMID: [28661424](#) · PMCID: [PMC5618077](#)

1655. **The use of IV vitamin C for patients with COVID-19: a case series**

Raul Hiedra, Kevin Bryan Lo, Mohammad Elbashabsheh, Fahad Gul, Robert Matthew Wright, Jeri Albano, Zurab Azmaiparashvili, Gabriel Patarroyo Aponte

Expert Review of Anti-infective Therapy (2020-08-01)

<https://doi.org/ghr938>

DOI: [10.1080/14787210.2020.1794819](https://doi.org/10.1080/14787210.2020.1794819) · PMID: [32662690](#) · PMCID: [PMC7441798](#)

1656. **Vitamin C for preventing and treating the common cold**

Harri Hemilä, Elizabeth Chalker

Cochrane Database of Systematic Reviews (2013-01-31)

<https://doi.org/xz5>

DOI: [10.1002/14651858.cd000980.pub4](https://doi.org/10.1002/14651858.cd000980.pub4) · PMID: [23440782](#)

1657. **Vitamin C intake and susceptibility to pneumonia**

HARRI HEMILÄ

The Pediatric Infectious Disease Journal (1997-09)

<https://doi.org/fkvs9d>

DOI: [10.1097/00006454-199709000-00003](https://doi.org/10.1097/00006454-199709000-00003) · PMID: [9306475](#)

1658. **Effect of Vitamin C Infusion on Organ Failure and Biomarkers of Inflammation and Vascular Injury in Patients With Sepsis and Severe Acute Respiratory Failure**

Alpha A Fowler, Jonathon D Truwit, RDuncan Hite, Peter E Morris, Christine DeWilde, Anna Priday, Bernard Fisher, Leroy R Thacker, Ramesh Natarajan, Donald F Brophy, ... Matthew Halquist

JAMA (2019-10-01) <https://doi.org/ggqmh8>

DOI: [10.1001/jama.2019.11825](https://doi.org/10.1001/jama.2019.11825) · PMID: [31573637](#) · PMCID: [PMC6777268](#)

1659. **Intravenous high-dose vitamin C for the treatment of severe COVID-19: study protocol for a multicentre randomised controlled**

trial

Fang Liu, Yuan Zhu, Jing Zhang, Yiming Li, Zhiyong Peng
BMJ Open (2020-07-08) <https://doi.org/gg4sgj>
DOI: [10.1136/bmjopen-2020-039519](https://doi.org/bmjopen-2020-039519) · PMID: [32641343](#) · PMCID: [PMC7348463](#)

1660. Pilot Trial of High-dose vitamin C in critically ill COVID-19 patients

Jing Zhang, Xin Rao, Yiming Li, Yuan Zhu, Fang Liu, Guangling Guo, Guoshi Luo, Zhongji Meng, Daniel De Backer, Hui Xiang, Zhi-Yong Peng
Research Square (2020-08-03) <https://doi.org/ghr94x>
DOI: [10.21203/rs.3.rs-52778/v2](https://doi.org/10.21203/rs.3.rs-52778/v2)

1661. High-dose vitamin C infusion for the treatment of critically ill COVID-19

Jing Zhang, Xin Rao, Yiming Li, Yuan Zhu, Fang Liu, Guangling Guo, Guoshi Luo, Zhongji Meng, Daniel De Backer, Hui Xiang, Zhi-Yong Peng
Research Square (2020-08-03) <https://doi.org/ghr94v>
DOI: [10.21203/rs.3.rs-52778/v1](https://doi.org/10.21203/rs.3.rs-52778/v1)

1662. Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids

Panel on Dietary Antioxidants and Related Compounds, Subcommittee on Upper Reference Levels of Nutrients, Subcommittee on Interpretation and Uses of Dietary Reference Intakes, Standing Committee on the Scientific Evaluation of Dietary Reference Intakes, Food and Nutrition Board, Institute of Medicine
The National Academies Press (2000-07-27) <https://doi.org/ghtvqx>
DOI: [10.17226/9810](https://doi.org/10.17226/9810) · PMID: [25077263](#)

1663. Vitamin D and Infectious Diseases: Simple Bystander or Contributing Factor?

Pedro Gois, Daniela Ferreira, Simon Olenski, Antonio Seguro
Nutrients (2017-06-24) <https://doi.org/ggpcwr>
DOI: [10.3390/nu9070651](https://doi.org/10.3390/nu9070651) · PMID: [28672783](#) · PMCID: [PMC5537771](#)

1664. Vitamin D and Influenza—Prevention or Therapy?

Beata M Gruber-Bzura
International Journal of Molecular Sciences (2018-08-16)
<https://doi.org/ggnndrj>
DOI: [10.3390/ijms19082419](https://doi.org/10.3390/ijms19082419) · PMID: [30115864](#) · PMCID: [PMC6121423](#)

1665. Immunologic Effects of Vitamin D on Human Health and Disease

Nipith Charoenngam, Michael F Holick
Nutrients (2020-07-15) <https://doi.org/gg45fp>
DOI: [10.3390/nu12072097](https://doi.org/10.3390/nu12072097) · PMID: [32679784](#) · PMCID: [PMC7400911](#)

1666. Vitamin D and respiratory health

DA Hughes, R Norton
Clinical & Experimental Immunology (2009-10) <https://doi.org/b3n6wc>
DOI: [10.1111/j.1365-2249.2009.04001.x](https://doi.org/10.1111/j.1365-2249.2009.04001.x) · PMID: [19737226](#) · PMCID: [PMC2759054](#)

1667. Regulation of Immune Function by Vitamin D and Its Use in Diseases of Immunity

An-Sofie Vanherwegen, Conny Gysemans, Chantal Mathieu

Endocrinology and Metabolism Clinics of North America (2017-12)

<https://doi.org/gcm7h9>

DOI: [10.1016/j.ecl.2017.07.010](https://doi.org/10.1016/j.ecl.2017.07.010) · PMID: [29080635](https://pubmed.ncbi.nlm.nih.gov/29080635/)

1668. Vitamin D and the Immune System

Cynthia Aranow

Journal of Investigative Medicine (2015-12-15) <https://doi.org/f3wh87>

DOI: [10.2310/jim.0b013e31821b8755](https://doi.org/10.2310/jim.0b013e31821b8755) · PMID: [21527855](https://pubmed.ncbi.nlm.nih.gov/21527855/)

1669. Vitamin D in the prevention of acute respiratory infection:

Systematic review of clinical studies

David A Jolliffe, Christopher J Griffiths, Adrian R Martineau

The Journal of Steroid Biochemistry and Molecular Biology (2013-07)

<https://doi.org/ggqmh9>

DOI: [10.1016/j.jsbmb.2012.11.017](https://doi.org/10.1016/j.jsbmb.2012.11.017) · PMID: [23220552](https://pubmed.ncbi.nlm.nih.gov/23220552/)

1670. Vitamin D: modulator of the immune system

Femke Baeke, Tatiana Takiishi, Hannelie Korf, Conny Gysemans, Chantal Mathieu

Current Opinion in Pharmacology (2010-08) <https://doi.org/d43qtf>

DOI: [10.1016/j.coph.2010.04.001](https://doi.org/10.1016/j.coph.2010.04.001) · PMID: [20427238](https://pubmed.ncbi.nlm.nih.gov/20427238/)

1671. Evidence that Vitamin D Supplementation Could Reduce Risk of Influenza and COVID-19 Infections and Deaths

William B Grant, Henry Lahore, Sharon L McDonnell, Carole A Baggerly, Christine B French, Jennifer L Aliano, Harjit P Bhattoa

Nutrients (2020-04-02) <https://doi.org/ggr2v5>

DOI: [10.3390/nu12040988](https://doi.org/10.3390/nu12040988) · PMID: [32252338](https://pubmed.ncbi.nlm.nih.gov/32252338/) · PMCID: [PMC7231123](https://pubmed.ncbi.nlm.nih.gov/PMC7231123/)

1672. Perspective: Vitamin D deficiency and COVID-19 severity – plausibly linked by latitude, ethnicity, impacts on cytokines, ACE2 and thrombosis

JM Rhodes, S Subramanian, E Laird, G Griffin, RA Kenny

Journal of Internal Medicine (2020-07-22) <https://doi.org/ghc7dh>

DOI: [10.1111/joim.13149](https://doi.org/10.1111/joim.13149) · PMID: [32613681](https://pubmed.ncbi.nlm.nih.gov/32613681/) · PMCID: [PMC7361294](https://pubmed.ncbi.nlm.nih.gov/PMC7361294/)

1673. COVID-19 fatalities, latitude, sunlight, and vitamin D

Paul B Whittemore

American Journal of Infection Control (2020-09) <https://doi.org/ghr93r>

DOI: [10.1016/j.ajic.2020.06.193](https://doi.org/10.1016/j.ajic.2020.06.193) · PMID: [32599103](https://pubmed.ncbi.nlm.nih.gov/32599103/) · PMCID: [PMC7319635](https://pubmed.ncbi.nlm.nih.gov/PMC7319635/)

1674. Editorial: low population mortality from COVID-19 in countries south of latitude 35 degrees North supports vitamin D as a factor determining severity

Jonathan M Rhodes, Sreedhar Subramanian, Eamon Laird, Rose A Kenny

Alimentary Pharmacology & Therapeutics (2020-06)

<https://doi.org/ggtw4b>

DOI: [10.1111/apt.15777](https://doi.org/10.1111/apt.15777) · PMID: [32311755](https://pubmed.ncbi.nlm.nih.gov/32311755/) · PMCID: [PMC7264531](https://pubmed.ncbi.nlm.nih.gov/PMC7264531/)

1675. 25-Hydroxyvitamin D Concentrations Are Lower in Patients with Positive PCR for SARS-CoV-2

Antonio D'Avolio, Valeria Avataneo, Alessandra Manca, Jessica Cusato, Amedeo De Nicolò, Renzo Lucchini, Franco Keller, Marco Cantù

1676. Vitamin D deficiency as risk factor for severe COVID-19: a convergence of two pandemics

D De Smet, K De Smet, P Herroelen, S Gryspeerdt, GA Martens
Cold Spring Harbor Laboratory (2020-05-05) <https://doi.org/ggv75>
DOI: [10.1101/2020.05.01.20079376](https://doi.org/10.1101/2020.05.01.20079376)

1677. Vitamin D sufficiency, a serum 25-hydroxyvitamin D at least 30 ng/mL reduced risk for adverse clinical outcomes in patients with COVID-19 infection

Zhila Maghbooli, Mohammad Ali Sahraian, Mehdi Ebrahimi, Marzieh Pazoki, Samira Kafan, Hedieh Moradi Tabriz, Azar Hadadi, Mahnaz Montazeri, Mehrad Nasiri, Arash Shirvani, Michael F Holick
PLOS ONE (2020-09-25) <https://doi.org/ghdzx8>
DOI: [10.1371/journal.pone.0239799](https://doi.org/10.1371/journal.pone.0239799) · PMID: [32976513](#) · PMCID: [PMC7518605](#)

1678. Role of vitamin D in preventing of COVID-19 infection, progression and severity

Nurshad Ali
Journal of Infection and Public Health (2020-10) <https://doi.org/ghdw9>
DOI: [10.1016/j.jiph.2020.06.021](https://doi.org/10.1016/j.jiph.2020.06.021) · PMID: [32605780](#) · PMCID: [PMC7305922](#)

1679. Low plasma 25(OH) vitamin D level is associated with increased risk of COVID-19 infection: an Israeli population-based study

Eugene Merzon, Dmitry Tworowski, Alessandro Gorohovski, Shlomo Vinker, Avivit Golan Cohen, Ilan Green, Milana Frenkel-Morgenstern
The FEBS Journal (2020-08-28) <https://doi.org/gg7b5c>
DOI: [10.1111/febs.15495](https://doi.org/10.1111/febs.15495) · PMID: [32700398](#) · PMCID: [PMC7404739](#)

1680. Association of Vitamin D Status and Other Clinical Characteristics With COVID-19 Test Results

David O Meltzer, Thomas J Best, Hui Zhang, Tamara Vokes, Vineet Arora, Julian Solway
JAMA Network Open (2020-09-03) <https://doi.org/ghdzw6>
DOI: [10.1001/jamanetworkopen.2020.19722](https://doi.org/10.1001/jamanetworkopen.2020.19722) · PMID: [32880651](#) · PMCID: [PMC7489852](#)

1681. Vitamin D Status in Hospitalized Patients with SARS-CoV-2 Infection

José L Hernández, Daniel Nan, Marta Fernandez-Ayala, Mayte García-Unzueta, Miguel A Hernández-Hernández, Marcos López-Hoyos, Pedro Muñoz-Cacho, José M Olmos, Manuel Gutiérrez-Cuadra, Juan J Ruiz-Cubillán, ... Víctor M Martínez-Taboada
The Journal of Clinical Endocrinology & Metabolism (2020-10-27) <https://doi.org/ghh737>
DOI: [10.1210/clinem/dgaa733](https://doi.org/10.1210/clinem/dgaa733) · PMID: [33159440](#) · PMCID: [PMC7797757](#)

1682. Analysis of vitamin D level among asymptomatic and critically ill COVID-19 patients and its correlation with inflammatory markers

Anshul Jain, Rachna Chaurasia, Narendra Singh Sengar, Mayank Singh, Sachin Mahor, Sumit Narain

1683. **Low 25-Hydroxyvitamin D Levels on Admission to the Intensive Care Unit May Predispose COVID-19 Pneumonia Patients to a Higher 28-Day Mortality Risk: A Pilot Study on a Greek ICU Cohort**

Alice G Vassiliou, Edison Jahaj, Maria Pratikaki, Stylianos E Orfanos, Ioanna Dimopoulou, Anastasia Kotanidou

Nutrients (2020-12-09) <https://doi.org/ghr95d>

DOI: [10.3390/nu12123773](https://doi.org/10.3390/nu12123773) · PMID: [33316914](https://pubmed.ncbi.nlm.nih.gov/33316914/) · PMCID: [PMC7764169](https://pubmed.ncbi.nlm.nih.gov/PMC7764169/)

1684. **Vitamin D deficiency as a predictor of poor prognosis in patients with acute respiratory failure due to COVID-19**

GE Carpagnano, V Di Lecce, VN Quaranta, A Zito, E Buonamico, E Capozza, A Palumbo, G Di Gioia, VN Valerio, O Resta

Journal of Endocrinological Investigation (2020-08-09)

<https://doi.org/gg7kqp>

DOI: [10.1007/s40618-020-01370-x](https://doi.org/10.1007/s40618-020-01370-x) · PMID: [32772324](https://pubmed.ncbi.nlm.nih.gov/32772324/) · PMCID:
[PMC7415009](https://pubmed.ncbi.nlm.nih.gov/PMC7415009/)

1685. **Vitamin D Deficiency and Outcome of COVID-19 Patients**

Aleksandar Radujkovic, Theresa Hippchen, Shilpa Tiwari-Heckler, Saida Dreher, Monica Boxberger, Uta Merle

Nutrients (2020-09-10) <https://doi.org/ghgfmp>

DOI: [10.3390/nu12092757](https://doi.org/10.3390/nu12092757) · PMID: [32927735](https://pubmed.ncbi.nlm.nih.gov/32927735/) · PMCID: [PMC7551780](https://pubmed.ncbi.nlm.nih.gov/PMC7551780/)

1686. **Impact of Vitamin D Deficiency on COVID-19—A Prospective Analysis from the CovILD Registry**

Alex Pizzini, Magdalena Aichner, Sabina Sahanic, Anna Böhm, Alexander Egger, Gregor Hoermann, Katharina Kurz, Gerlig Widmann, Rosa Bellmann-Weiler, Günter Weiss, ... Judith Löffler-Ragg

Nutrients (2020-09-11) <https://doi.org/ghr95b>

DOI: [10.3390/nu12092775](https://doi.org/10.3390/nu12092775) · PMID: [32932831](https://pubmed.ncbi.nlm.nih.gov/32932831/) · PMCID: [PMC7551662](https://pubmed.ncbi.nlm.nih.gov/PMC7551662/)

1687. **Does Serum Vitamin D Level Affect COVID-19 Infection and Its Severity?-A Case-Control Study**

Kun Ye, Fen Tang, Xin Liao, Benjamin A Shaw, Meiqiu Deng, Guangyi Huang, Zhiqiang Qin, Xiaomei Peng, Hewei Xiao, Chunxia Chen, ...

Jianrong Yang

Journal of the American College of Nutrition (2020-10-13)

<https://doi.org/ghr935>

DOI: [10.1080/07315724.2020.1826005](https://doi.org/10.1080/07315724.2020.1826005) · PMID: [33048028](https://pubmed.ncbi.nlm.nih.gov/33048028/)

1688. **Lower levels of vitamin D are associated with SARS-CoV-2 infection and mortality in the Indian population: An observational study**

Sunali Padhi, Subham Suvankar, Venketesh K Panda, Abhijit Pati, Aditya K Panda

International Immunopharmacology (2020-11) <https://doi.org/ghr93w>

DOI: [10.1016/j.intimp.2020.107001](https://doi.org/10.1016/j.intimp.2020.107001) · PMID: [33182040](https://pubmed.ncbi.nlm.nih.gov/33182040/) · PMCID:
[PMC7489890](https://pubmed.ncbi.nlm.nih.gov/PMC7489890/)

1689. **Vitamin D Deficiency Is Associated with COVID-19 Incidence and Disease Severity in Chinese People**

Xia Luo, Qing Liao, Ying Shen, Huijun Li, Liming Cheng
The Journal of Nutrition (2021-01) <https://doi.org/ghr939>
DOI: [10.1093/jn/nxaa332](https://doi.org/10.1093/jn/nxaa332) · PMID: [33188401](#)

1690. **Vitamin D concentrations and COVID-19 infection in UK Biobank**
Claire E Hastie, Daniel F Mackay, Frederick Ho, Carlos A Celis-Morales, Srinivasa Vittal Katikireddi, Claire L Niedzwiedz, Bhautesh D Jani, Paul Welsh, Frances S Mair, Stuart R Gray, ... Jill P Pell
Diabetes & Metabolic Syndrome: Clinical Research & Reviews (2020-07) <https://doi.org/ggvv72>
DOI: [10.1016/j.dsx.2020.04.050](https://doi.org/10.1016/j.dsx.2020.04.050) · PMID: [32413819](#) · PMCID: [PMC7204679](#)
1691. **Vitamin D and COVID-19 infection and mortality in UK Biobank**
Claire E Hastie, Jill P Pell, Naveed Sattar
European Journal of Nutrition (2020-08-26) <https://doi.org/ghr93p>
DOI: [10.1007/s00394-020-02372-4](https://doi.org/10.1007/s00394-020-02372-4) · PMID: [32851419](#) · PMCID: [PMC7449523](#)
1692. **Low serum 25-hydroxyvitamin D (25[OH]D) levels in patients hospitalized with COVID-19 are associated with greater disease severity**
Grigoris Panagiotou, Su Ann Tee, Yasir Ihsan, Waseem Athar, Gabriella Marchitelli, Donna Kelly, Christopher S Boot, Nadia Stock, James Macfarlane, Adrian R Martineau, ... Richard Quinton
Clinical Endocrinology (2020-08-06) <https://doi.org/gg5gbj>
DOI: [10.1111/cen.14276](https://doi.org/10.1111/cen.14276) · PMID: [32621392](#) · PMCID: [PMC7361912](#)
1693. **Letter in response to the article: Vitamin D concentrations and COVID-19 infection in UK biobank (Hastie et al.)**
WB Grant, SL McDonnell
Diabetes & Metabolic Syndrome: Clinical Research & Reviews (2020-09) <https://doi.org/ghc7p4>
DOI: [10.1016/j.dsx.2020.05.046](https://doi.org/10.1016/j.dsx.2020.05.046) · PMID: [32563941](#) · PMCID: [PMC7293469](#)
1694. **Vitamin D deficiency in African Americans is associated with a high risk of severe disease and mortality by SARS-CoV-2**
Virna Margarita Martín Giménez, Felipe Inserra, León Ferder, Joxel García, Walter Manucha
Journal of Human Hypertension (2020-08-13) <https://doi.org/ghr933>
DOI: [10.1038/s41371-020-00398-z](https://doi.org/10.1038/s41371-020-00398-z) · PMID: [32792611](#) · PMCID: [PMC7425793](#)
1695. **Evidence for possible association of vitamin D status with cytokine storm and unregulated inflammation in COVID-19 patients**
Ali Daneshkhah, Vasundhara Agrawal, Adam Eshein, Hariharan Subramanian, Hemant Kumar Roy, Vadim Backman
Aging Clinical and Experimental Research (2020-09-02) <https://doi.org/ghr93q>
DOI: [10.1007/s40520-020-01677-y](https://doi.org/10.1007/s40520-020-01677-y) · PMID: [32876941](#) · PMCID: [PMC7465887](#)

1696. **Short term, high-dose vitamin D supplementation for COVID-19 disease: a randomised, placebo-controlled, study (SHADE study)**
Ashu Rastogi, Anil Bhansali, Niranjan Khare, Vikas Suri, Narayana Yaddanapudi, Naresh Sachdeva, GD Puri, Pankaj Malhotra
Postgraduate Medical Journal (2020-11-12) <https://doi.org/ghnpg>
DOI: [10.1136/postgradmedj-2020-139065](https://doi.org/10.1136/postgradmedj-2020-139065) · PMID: [33184146](#)
1697. **"Effect of calcifediol treatment and best available therapy versus best available therapy on intensive care unit admission and mortality among patients hospitalized for COVID-19: A pilot randomized clinical study"**
Marta Entrenas Castillo, Luis Manuel Entrenas Costa, José Manuel Vaquero Barrios, Juan Francisco Alcalá Díaz, José López Miranda, Roger Bouillon, José Manuel Quesada Gomez
The Journal of Steroid Biochemistry and Molecular Biology (2020-10) <https://doi.org/ghd79r>
DOI: [10.1016/j.jsbmb.2020.105751](https://doi.org/10.1016/j.jsbmb.2020.105751) · PMID: [32871238](#) · PMCID: [PMC7456194](#)
1698. **COVID-19 rapid evidence summary: vitamin D for COVID-19 | Advice | NICE** <https://www.nice.org.uk/advice/es28>
1699. **Mathematical analysis of Córdoba calcifediol trial suggests strong role for Vitamin D in reducing ICU admissions of hospitalized COVID-19 patients**
Irwin Jungreis, Manolis Kellis
Cold Spring Harbor Laboratory (2020-12-21) <https://doi.org/ghr94h>
DOI: [10.1101/2020.11.08.20222638](https://doi.org/10.1101/2020.11.08.20222638)
1700. **High-Dose Cholecalciferol Booster Therapy is Associated with a Reduced Risk of Mortality in Patients with COVID-19: A Cross-Sectional Multi-Centre Observational Study**
Stephanie F Ling, Eleanor Broad, Rebecca Murphy, Joseph M Pappachan, Satveer Pardesi-Newton, Marie-France Kong, Edward B Jude
Nutrients (2020-12-11) <https://doi.org/ghr95f>
DOI: [10.3390/nu12123799](https://doi.org/10.3390/nu12123799) · PMID: [33322317](#) · PMCID: [PMC7763301](#)
1701. **Effect of Vitamin D ₃ Supplementation vs Placebo on Hospital Length of Stay in Patients with Severe COVID-19: A Multicenter, Double-blind, Randomized Controlled Trial**
Igor H Murai, Alan L Fernandes, Lucas P Sales, Ana J Pinto, Karla F Goessler, Camila SC Duran, Carla BR Silva, André S Franco, Marina B Macedo, Henrique HH Dalmolin, ... Rosa MR Pereira
Cold Spring Harbor Laboratory (2020-11-17) <https://doi.org/ghr94j>
DOI: [10.1101/2020.11.16.20232397](https://doi.org/10.1101/2020.11.16.20232397)
1702. **Effect of Vitamin D Administration on Prevention and Treatment of Mild Forms of Suspected Covid-19**
Manuel Castillo Garzón
clinicaltrials.gov (2020-04-03)
<https://clinicaltrials.gov/ct2/show/NCT04334005>
1703. **Improving Vitamin D Status in the Management of COVID-19**
Aldo Montano-Loza

clinicaltrials.gov (2020-06-03)
<https://clinicaltrials.gov/ct2/show/NCT04385940>

1704. **Cholecalciferol to Improve the Outcomes of COVID-19 Patients - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04411446>
1705. **Covid-19 and Vitamin D Supplementation: a Multicenter Randomized Controlled Trial of High Dose Versus Standard Dose Vitamin D3 in High-risk COVID-19 Patients (CoVitTrial) - Full Text View - ClinicalTrials.gov**
<https://clinicaltrials.gov/ct2/show/NCT04344041>
1706. **The LEAD COVID-19 Trial: Low-risk, Early Aspirin and Vitamin D to Reduce COVID-19 Hospitalizations**
Louisiana State University Health Sciences Center in New Orleans
clinicaltrials.gov (2020-04-24)
<https://clinicaltrials.gov/ct2/show/NCT04363840>
1707. **Randomized Double-Blind Placebo-Controlled Proof-of-Concept Trial of a Plant Polyphenol for the Outpatient Treatment of Mild Coronavirus Disease (COVID-19)**
Marvin McCreary MD
clinicaltrials.gov (2020-09-22)
<https://clinicaltrials.gov/ct2/show/NCT04400890>
1708. **Current vitamin D status in European and Middle East countries and strategies to prevent vitamin D deficiency: a position statement of the European Calcified Tissue Society**
Paul Lips, Kevin D Cashman, Christel Lamberg-Allardt, Heike Annette Bischoff-Ferrari, Barbara Obermayer-Pietsch, Maria Luisa Bianchi, Jan Stepan, Ghada El-Hajj Fuleihan, Roger Bouillon
European Journal of Endocrinology (2019-04) <https://doi.org/ggr42p>
DOI: [10.1530/eje-18-0736](https://doi.org/10.1530/eje-18-0736) · PMID: [30721133](#)
1709. **Communiqué de l'Académie nationale de Médecine : Vitamine D et Covid-19 – Académie nationale de médecine | Une institution dans son temps** <https://www.academie-medecine.fr/communique-de-lacademie-nationale-de-medecine-vitamine-d-et-covid-19/>
1710. **Covid-19: NHS bosses told to assess risk to ethnic minority staff who may be at greater risk**
Gareth Iacobucci
BMJ (2020-05-04) <https://doi.org/ggy2zq>
DOI: [10.1136/bmj.m1820](https://doi.org/10.1136/bmj.m1820) · PMID: [32366503](#)
1711. **Covid-19: Public health agencies review whether vitamin D supplements could reduce risk**
Ingrid Torjesen
BMJ (2020-06-19) <https://doi.org/ghr94p>
DOI: [10.1136/bmj.m2475](https://doi.org/10.1136/bmj.m2475) · PMID: [32561509](#)
1712. **Avoidance of vitamin D deficiency to slow the COVID-19 pandemic**
Martin Kohlmeier
BMJ Nutrition, Prevention & Health (2020-06) <https://doi.org/ghr94q>

1713. **COVID-19 rapid guideline: vitamin D**

National Institute for Health and Care Excellence (NICE)

<https://www.nice.org.uk/guidance/ng187/resources/covid19-rapid-guideline-vitamin-d-pdf-66142026720709>

1714. **Prevalence of Vitamin D Deficiency and Associated Risk Factors in the US Population (2011-2012)**

Naveen R Parva, Satish Tadepalli, Pratiksha Singh, Andrew Qian, Rajat Joshi, Hyndavi Kandala, Vinod K Nookala, Pramil Cheriyath

Cureus (2018-06-05) <https://doi.org/gg7kqq>

DOI: [10.7759/cureus.2741](https://doi.org/10.7759/cureus.2741) · PMID: [30087817](https://pubmed.ncbi.nlm.nih.gov/30087817/) · PMCID: [PMC6075634](https://pubmed.ncbi.nlm.nih.gov/PMC6075634/)

1715. **Vitamin D**

COVID-19 Treatment Guidelines

<https://www.covid19treatmentguidelines.nih.gov/adjunctive-therapy/vitamin-d/>

1716. **Dietary Supplements during COVID-19 Outbreak. Results of Google Trends Analysis Supported by PLifeCOVID-19 Online Studies**

Jadwiga Hamulka, Marta Jeruszka-Bielak, Magdalena Górnicka, Małgorzata E Drywień, Monika A Zielinska-Pukos

Nutrients (2020-12-27) <https://doi.org/ghtvq3>

DOI: [10.3390/nu13010054](https://doi.org/10.3390/nu13010054) · PMID: [33375422](https://pubmed.ncbi.nlm.nih.gov/33375422/) · PMCID: [PMC7823317](https://pubmed.ncbi.nlm.nih.gov/PMC7823317/)

1717. **Court Orders Georgia Defendants to Stop Selling Vitamin D Products as Treatments for Covid-19 and Other Diseases (2021-01-08)**

<https://www.justice.gov/opa/pr/court-orders-georgia-defendants-stop-selling-vitamin-d-products-treatments-covid-19-and-other>

1718. **The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic**

Colin Hill, Francisco Guarner, Gregor Reid, Glenn R Gibson, Daniel J Merenstein, Bruno Pot, Lorenzo Morelli, Roberto Berni Canani, Harry J Flint, Seppo Salminen, ... Mary Ellen Sanders

Nature Reviews Gastroenterology & Hepatology (2014-06-10)

<https://doi.org/f6ndv7>

DOI: [10.1038/nrgastro.2014.66](https://doi.org/10.1038/nrgastro.2014.66) · PMID: [24912386](https://pubmed.ncbi.nlm.nih.gov/24912386/)

1719. **The Effect of Probiotics on Prevention of Common Cold: A Meta-Analysis of Randomized Controlled Trial Studies**

En-Jin Kang, Soo Young Kim, In-Hong Hwang, Yun-Jeong Ji

Korean Journal of Family Medicine (2013) <https://doi.org/gg3knf>

DOI: [10.4082/kjfm.2013.34.1.2](https://doi.org/10.4082/kjfm.2013.34.1.2) · PMID: [23372900](https://pubmed.ncbi.nlm.nih.gov/23372900/) · PMCID: [PMC3560336](https://pubmed.ncbi.nlm.nih.gov/PMC3560336/)

1720. **Probiotics and Paraprobiotics in Viral Infection: Clinical Application and Effects on the Innate and Acquired Immune Systems**

Osamu Kanauchi, Akira Andoh, Sazaly AbuBakar, Naoki Yamamoto

Current Pharmaceutical Design (2018-05-10) <https://doi.org/gdjnpk>

1721. **Using Probiotics to Flatten the Curve of Coronavirus Disease COVID-2019 Pandemic**

David Baud, Varvara Dimopoulou Agri, Glenn R Gibson, Gregor Reid, Eric Giannoni

Frontiers in Public Health (2020-05-08) <https://doi.org/gg3knd>

DOI: [10.3389/fpubh.2020.00186](https://doi.org/10.3389/fpubh.2020.00186) · PMID: [32574290](#) · PMCID: [PMC7227397](#)

1722. **Next-generation probiotics: the spectrum from probiotics to live biotherapeutics**

Paul W O'Toole, Julian R Marchesi, Colin Hill

Nature Microbiology (2017-04-25) <https://doi.org/ggzggv>

DOI: [10.1038/nmicrobiol.2017.57](https://doi.org/10.1038/nmicrobiol.2017.57) · PMID: [28440276](#)

1723. **Mechanisms of Action of Probiotics**

Julio Plaza-Díaz, Francisco Javier Ruiz-Ojeda, Mercedes Gil-Campos, Angel Gil

Advances in Nutrition (2019-01) <https://doi.org/gft8sh>

DOI: [10.1093/advances/nmy063](https://doi.org/10.1093/advances/nmy063) · PMID: [30721959](#) · PMCID: [PMC6363529](#)

1724. **Probiotic mechanisms of action**

Katrina Halloran, Mark A Underwood

Early Human Development (2019-08) <https://doi.org/gg3jc4>

DOI: [10.1016/j.earlhumdev.2019.05.010](https://doi.org/10.1016/j.earlhumdev.2019.05.010) · PMID: [31174927](#)

1725. **Probiotic Mechanisms of Action**

Miriam Bermudez-Brito, Julio Plaza-Díaz, Sergio Muñoz-Quezada, Carolina Gómez-Llorente, Angel Gil

Annals of Nutrition and Metabolism (2012) <https://doi.org/gg3knb>

DOI: [10.1159/000342079](https://doi.org/10.1159/000342079) · PMID: [23037511](#)

1726. **A novel eukaryotic cell culture model to study antiviral activity of potential probiotic bacteria**

T BOTIC, T KLINGBERG, H WEINGARTL, A CENCIC

International Journal of Food Microbiology (2007-04-30)

<https://doi.org/fks5cz>

DOI: [10.1016/j.ijfoodmicro.2006.10.044](https://doi.org/10.1016/j.ijfoodmicro.2006.10.044) · PMID: [17261339](#)

1727. **Oral administration of < i>Lactobacillus brevis</i> KB290 to mice alleviates clinical symptoms following influenza virus infection**

N Waki, N Yajima, H Suganuma, BM Buddle, D Luo, A Heiser, T Zheng

Letters in Applied Microbiology (2014-01) <https://doi.org/f5j37w>

DOI: [10.1111/lam.12160](https://doi.org/10.1111/lam.12160) · PMID: [24329975](#)

1728. **Antiviral activity of Lactobacillus brevis towards herpes simplex virus type 2: Role of cell wall associated components**

Paola Mastromarino, Fatima Cacciotti, Alessandra Masci, Luciana Mosca

Anaerobe (2011-12) <https://doi.org/bcpvm5>

DOI: [10.1016/j.anaerobe.2011.04.022](https://doi.org/10.1016/j.anaerobe.2011.04.022) · PMID: [21621625](#)

1729. Critical Adverse Impact of IL-6 in Acute Pneumovirus Infection

Caroline M Percopo, Michelle Ma, Todd A Brenner, Julia O Krumholz, Timothy J Break, Karen Laky, Helene F Rosenberg
The Journal of Immunology (2019-02-01) <https://doi.org/ghr95h>
DOI: [10.4049/jimmunol.1800927](https://doi.org/10.4049/jimmunol.1800927) · PMID: [30578308](#) · PMCID: [PMC6365009](#)

1730. Antiviral Activity of Exopolysaccharides Produced by Lactic Acid Bacteria of the Genera *Pediococcus*, *Leuconostoc* and *Lactobacillus* against Human Adenovirus Type 5

Biliavska, Pankivska, Povnitsa, Zagorodnya
Medicina (2019-08-22) <https://doi.org/ghr948>
DOI: [10.3390/medicina55090519](https://doi.org/10.3390/medicina55090519) · PMID: [31443536](#) · PMCID: [PMC6780409](#)

1731. Prevention of respiratory syncytial virus infection with probiotic lactic acid bacterium *Lactobacillus gasseri* SBT2055

Kei Eguchi, Naoki Fujitani, Hisako Nakagawa, Tadaaki Miyazaki
Scientific Reports (2019-03-18) <https://doi.org/ghr934>
DOI: [10.1038/s41598-019-39602-7](https://doi.org/10.1038/s41598-019-39602-7) · PMID: [30886158](#) · PMCID: [PMC6423325](#)

1732. Effect of probiotic on innate inflammatory response and viral shedding in experimental rhinovirus infection – a randomised controlled trial

RB Turner, JA Woodfolk, L Borish, JW Steinke, JT Patrie, LM Muehling, S Lahtinen, MJ Lehtinen
Beneficial Microbes (2017-04-26) <https://doi.org/f955fh>
DOI: [10.3920/bm2016.0160](https://doi.org/10.3920/bm2016.0160) · PMID: [28343401](#) · PMCID: [PMC5797652](#)

1733. Immunobiotic lactobacilli reduce viral-associated pulmonary damage through the modulation of inflammation-coagulation interactions

Hortensia Zelaya, Kohichiro Tsukida, Eriko Chiba, Gabriela Marranzino, Susana Alvarez, Haruki Kitazawa, Graciela Agüero, Julio Villena
International Immunopharmacology (2014-03) <https://doi.org/f5wd93>
DOI: [10.1016/j.intimp.2013.12.020](https://doi.org/10.1016/j.intimp.2013.12.020) · PMID: [24394565](#)

1734. Nasal priming with immunobiotic lactobacilli improves the adaptive immune response against influenza virus

Fernanda Raya Tonetti, MdAminul Islam, Maria Guadalupe Vizoso-Pinto, Hideki Takahashi, Haruki Kitazawa, Julio Villena
International Immunopharmacology (2020-01) <https://doi.org/ghr93v>
DOI: [10.1016/j.intimp.2019.106115](https://doi.org/10.1016/j.intimp.2019.106115) · PMID: [31841753](#)

1735. The potential application of probiotics and prebiotics for the prevention and treatment of COVID-19

Amin N Olaimat, Iman Aolymat, Murad Al-Holy, Mutamed Ayyash, Mahmoud Abu Ghoush, Anas A Al-Nabulsi, Tareq Osaili, Vasso Apostolopoulos, Shao-Quan Liu, Nagendra P Shah
npj Science of Food (2020-10-05) <https://doi.org/ghggq4>
DOI: [10.1038/s41538-020-00078-9](https://doi.org/10.1038/s41538-020-00078-9) · PMID: [33083549](#) · PMCID: [PMC7536434](#)

1736. Pulmonary-intestinal cross-talk in mucosal inflammatory disease

1737. The role of the lung microbiota and the gut-lung axis in respiratory infectious diseases

Alexia Dumas, Lucie Bernard, Yannick Poquet, Geanncarlo Lugo-Villarino, Olivier Neyrolles
Cellular Microbiology (2018-12) <https://doi.org/gfjds9>
DOI: [10.1111/cmi.12966](https://doi.org/10.1111/cmi.12966) · PMID: [30329198](https://pubmed.ncbi.nlm.nih.gov/30329198/)

1738. Gut microbiota and Covid-19- possible link and implications

Debojoyoti Dhar, Abhishek Mohanty
Virus Research (2020-08) <https://doi.org/gg3jc5>
DOI: [10.1016/j.virusres.2020.198018](https://doi.org/10.1016/j.virusres.2020.198018) · PMID: [32430279](https://pubmed.ncbi.nlm.nih.gov/32430279/) · PMCID: [PMC7217790](https://pubmed.ncbi.nlm.nih.gov/PMC7217790/)

1739. Oral Microbiome and SARS-CoV-2: Beware of Lung Co-infection

Lirong Bao, Cheng Zhang, Jiajia Dong, Lei Zhao, Yan Li, Jianxun Sun
Frontiers in Microbiology (2020-07-31) <https://doi.org/ghr944>
DOI: [10.3389/fmicb.2020.01840](https://doi.org/10.3389/fmicb.2020.01840) · PMID: [32849438](https://pubmed.ncbi.nlm.nih.gov/32849438/) · PMCID: [PMC7411080](https://pubmed.ncbi.nlm.nih.gov/PMC7411080/)

1740. Lung microbiome and coronavirus disease 2019 (COVID-19): Possible link and implications

Saroj Khatiwada, Astha Subedi
Human Microbiome Journal (2020-08) <https://doi.org/gg7m83>
DOI: [10.1016/j.humic.2020.100073](https://doi.org/10.1016/j.humic.2020.100073) · PMID: [32835135](https://pubmed.ncbi.nlm.nih.gov/32835135/) · PMCID: [PMC7405772](https://pubmed.ncbi.nlm.nih.gov/PMC7405772/)

1741. Probiotics in respiratory virus infections

L Lehtoranta, A Pitkäranta, R Korpela
European Journal of Clinical Microbiology & Infectious Diseases (2014-03-18) <https://doi.org/f583jr>
DOI: [10.1007/s10096-014-2086-y](https://doi.org/10.1007/s10096-014-2086-y) · PMID: [24638909](https://pubmed.ncbi.nlm.nih.gov/24638909/) · PMCID: [PMC7088122](https://pubmed.ncbi.nlm.nih.gov/PMC7088122/)

1742. Probiotics for preventing acute upper respiratory tract infections

Qiukui Hao, Bi Rong Dong, Taixiang Wu
Cochrane Database of Systematic Reviews (2015-02-03)
<https://doi.org/gg3jc3>
DOI: [10.1002/14651858.cd006895.pub3](https://doi.org/10.1002/14651858.cd006895.pub3) · PMID: [25927096](https://pubmed.ncbi.nlm.nih.gov/25927096/)

1743. Probiotics for the prevention of respiratory tract infections: a systematic review

Evridiki K Vouloumanou, Gregory C Makris, Drosos E Karageorgopoulos, Matthew E Falagas
International Journal of Antimicrobial Agents (2009-09)
<https://doi.org/dn8kw8>
DOI: [10.1016/j.ijantimicag.2008.11.005](https://doi.org/10.1016/j.ijantimicag.2008.11.005) · PMID: [19179052](https://pubmed.ncbi.nlm.nih.gov/19179052/)

1744. Effectiveness of probiotics on the duration of illness in healthy children and adults who develop common acute respiratory infectious conditions: a systematic review and meta-analysis

Sarah King, Julie Glanville, Mary Ellen Sanders, Anita Fitzgerald, Danielle Varley
British Journal of Nutrition (2014-04-29) <https://doi.org/f57hq5>
DOI: [10.1017/s0007114514000075](https://doi.org/10.1017/s0007114514000075) · PMID: [24780623](https://pubmed.ncbi.nlm.nih.gov/24780623/) · PMCID: [PMC4054664](https://pubmed.ncbi.nlm.nih.gov/PMC4054664/)

1745. **Effect of probiotics on the incidence of ventilator-associated pneumonia in critically ill patients: a randomized controlled multicenter trial**

Juan Zeng, Chun-Ting Wang, Fu-Shen Zhang, Feng Qi, Shi-Fu Wang, Shuang Ma, Tie-Jun Wu, Hui Tian, Zhao-Tao Tian, Shu-Liu Zhang, ... Yu-Ping Wang
Intensive Care Medicine (2016-04-04) <https://doi.org/f8jnrt>
DOI: [10.1007/s00134-016-4303-x](https://doi.org/10.1007/s00134-016-4303-x) · PMID: [27043237](https://pubmed.ncbi.nlm.nih.gov/27043237/)

1746. **Probiotic Prophylaxis of Ventilator-associated Pneumonia**

Lee E Morrow, Marin H Kollef, Thomas B Casale
American Journal of Respiratory and Critical Care Medicine (2010-10-15) <https://doi.org/d5hh4t>
DOI: [10.1164/rccm.200912-1853oc](https://doi.org/10.1164/rccm.200912-1853oc) · PMID: [20522788](https://pubmed.ncbi.nlm.nih.gov/20522788/) · PMCID: [PMC2970846](https://pubmed.ncbi.nlm.nih.gov/PMC2970846/)

1747. **Synbiotics modulate gut microbiota and reduce enteritis and ventilator-associated pneumonia in patients with sepsis: a randomized controlled trial**

Kentaro Shimizu, Tomoki Yamada, Hiroshi Ogura, Tomoyoshi Mohri, Takeyuki Kiguchi, Satoshi Fujimi, Takashi Asahara, Tomomi Yamada, Masahiro Ojima, Mitsunori Ikeda, Takeshi Shimazu
Critical Care (2018-09-27) <https://doi.org/gfdggi>
DOI: [10.1186/s13054-018-2167-x](https://doi.org/10.1186/s13054-018-2167-x) · PMID: [30261905](https://pubmed.ncbi.nlm.nih.gov/30261905/) · PMCID: [PMC6161427](https://pubmed.ncbi.nlm.nih.gov/PMC6161427/)

1748. **Probiotics for the Prevention of Ventilator-Associated Pneumonia: A Meta-Analysis of Randomized Controlled Trials**

Minmin Su, Ying Jia, Yan Li, Dianyou Zhou, Jinsheng Jia
Respiratory Care (2020-05) <https://doi.org/gg3kng>
DOI: [10.4187/respcares.07097](https://doi.org/10.4187/respcares.07097) · PMID: [32127415](https://pubmed.ncbi.nlm.nih.gov/32127415/)

1749. **COVID-19: An Alert to Ventilator-Associated Bacterial Pneumonia**

Helvécio Cardoso Corrêa Póvoa, Gabriela Ceccon Chianca, Natalia Lopes Pontes Póvoa Iorio
Adis Journals (2020) <https://doi.org/gg3knh>
DOI: [10.6084/m9.figshare.12340496](https://doi.org/10.6084/m9.figshare.12340496)

1750. **The challenge of ventilator-associated pneumonia diagnosis in COVID-19 patients**

Bruno François, Pierre-François Laterre, Charles-Edouard Luyt, Jean Chastre
Critical Care (2020-06-05) <https://doi.org/gg3knc>
DOI: [10.1186/s13054-020-03013-2](https://doi.org/10.1186/s13054-020-03013-2) · PMID: [32503590](https://pubmed.ncbi.nlm.nih.gov/32503590/) · PMCID: [PMC7273812](https://pubmed.ncbi.nlm.nih.gov/PMC7273812/)

1751. **Prophylactic use of probiotics for gastrointestinal disorders in children**

Celine Perceval, Hania Szajewska, Flavia Indrio, Zvi Weizman, Yvan

Vandenplas

The Lancet Child & Adolescent Health (2019-09) <https://doi.org/d2qp>

DOI: [10.1016/s2352-4642\(19\)30182-8](https://doi.org/s2352-4642(19)30182-8)

1752. **Effect of Gastrointestinal Symptoms in Patients With COVID-19**

Zili Zhou, Ning Zhao, Yan Shu, Shengbo Han, Bin Chen, Xiaogang Shu

Gastroenterology (2020-06) <https://doi.org/ggq8x8>

DOI: [10.1053/j.gastro.2020.03.020](https://doi.org/j.gastro.2020.03.020) · PMID: [32199880](https://pubmed.ncbi.nlm.nih.gov/32199880/) · PMCID:

[PMC7270807](https://pubmed.ncbi.nlm.nih.gov/PMC7270807/)

1753. **The digestive system is a potential route of 2019-nCov infection: a bioinformatics analysis based on single-cell transcriptomes**

Hao Zhang, Zijian Kang, Haiyi Gong, Da Xu, Jing Wang, Zifu Li, Xingang

Cui, Jianru Xiao, Tong Meng, Wang Zhou, ... Huji Xu

Cold Spring Harbor Laboratory (2020-01-31) <https://doi.org/ggjvx2>

DOI: [10.1101/2020.01.30.927806](https://doi.org/10.1101/2020.01.30.927806)

1754. **Cryo-electron microscopy structures of the SARS-CoV spike glycoprotein reveal a prerequisite conformational state for receptor binding**

Miao Gui, Wenfei Song, Haixia Zhou, Jingwei Xu, Silian Chen, Ye Xiang, Xinquan Wang

Cell Research (2016-12-23) <https://doi.org/f9m247>

DOI: [10.1038/cr.2016.152](https://doi.org/10.1038/cr.2016.152) · PMID: [28008928](https://pubmed.ncbi.nlm.nih.gov/28008928/) · PMCID: [PMC5223232](https://pubmed.ncbi.nlm.nih.gov/PMC5223232/)

1755. **Enteric involvement of coronaviruses: is faecal-oral transmission of SARS-CoV-2 possible?**

Charleen Yeo, Sanghvi Kaushal, Danson Yeo

The Lancet Gastroenterology & Hepatology (2020-04)

<https://doi.org/ggpzx7s>

DOI: [10.1016/s2468-1253\(20\)30048-0](https://doi.org/s2468-1253(20)30048-0) · PMID: [32087098](https://pubmed.ncbi.nlm.nih.gov/32087098/) · PMCID:

[PMC7130008](https://pubmed.ncbi.nlm.nih.gov/PMC7130008/)

1756. **Characteristics of pediatric SARS-CoV-2 infection and potential evidence for persistent fecal viral shedding**

Yi Xu, Xufang Li, Bing Zhu, Huiying Liang, Chunxiao Fang, Yu Gong, Qiaozhi Guo, Xin Sun, Danyang Zhao, Jun Shen, ... Sitang Gong

Nature Medicine (2020-03-13) <https://doi.org/ggpwx5>

DOI: [10.1038/s41591-020-0817-4](https://doi.org/10.1038/s41591-020-0817-4) · PMID: [32284613](https://pubmed.ncbi.nlm.nih.gov/32284613/) · PMCID:

[PMC7095102](https://pubmed.ncbi.nlm.nih.gov/PMC7095102/)

1757. **Modulation of rotavirus severe gastroenteritis by the combination of probiotics and prebiotics**

Guadalupe Gonzalez-Ochoa, Lilian K Flores-Mendoza, Ramona Icedo-Garcia, Ricardo Gomez-Flores, Patricia Tamez-Guerra

Archives of Microbiology (2017-06-20) <https://doi.org/gbsb4d>

DOI: [10.1007/s00203-017-1400-3](https://doi.org/s00203-017-1400-3) · PMID: [28634691](https://pubmed.ncbi.nlm.nih.gov/28634691/) · PMCID:

[PMC5548957](https://pubmed.ncbi.nlm.nih.gov/PMC5548957/)

1758. **Multicenter Trial of a Combination Probiotic for Children with Gastroenteritis**

Stephen B Freedman, Sarah Williamson-Urquhart, Ken J Farion, Serge Gouin, Andrew R Willan, Naveen Poonai, Katrina Hurley, Philip M Sherman, Yaron Finkelstein, Bonita E Lee, ... Suzanne Schuh

1759. **Synbiotic Therapy of Gastrointestinal Symptoms During Covid-19 Infection: A Randomized, Double-blind, Placebo Controlled, Telemedicine Study (SynCov Study)**
Medical University of Graz
clinicaltrials.gov (2021-01-14)
<https://clinicaltrials.gov/ct2/show/NCT04420676>
1760. **Multicentric Study to Assess the Effect of Consumption of Lactobacillus Coryniformis K8 on Healthcare Personnel Exposed to COVID-19**
Biosearch S.A.
clinicaltrials.gov (2020-04-28)
<https://clinicaltrials.gov/ct2/show/NCT04366180>
1761. **The Intestinal Microbiota as a Therapeutic Target in Hospitalized Patients With COVID-19 Infection**
Bioithas SL
clinicaltrials.gov (2021-01-26)
<https://clinicaltrials.gov/ct2/show/NCT04390477>
1762. **Probiotics: definition, scope and mechanisms of action**
Gregor Reid
Best Practice & Research Clinical Gastroenterology (2016-02)
<https://doi.org/f8m79k>
DOI: [10.1016/j.bpg.2015.12.001](https://doi.org/10.1016/j.bpg.2015.12.001) · PMID: [27048893](https://pubmed.ncbi.nlm.nih.gov/27048893/)
1763. **Health benefits and health claims of probiotics: bridging science and marketing**
Ger T Rijkers, Willem M de Vos, Robert-Jan Brummer, Lorenzo Morelli, Gerard Corthier, Philippe Marteau
British Journal of Nutrition (2011-08-24) <https://doi.org/cb78rx>
DOI: [10.1017/s000711451100287x](https://doi.org/10.1017/s000711451100287x) · PMID: [21861940](https://pubmed.ncbi.nlm.nih.gov/21861940/)
1764. **Probiotics and COVID-19: one size does not fit all**
Joyce WY Mak, Francis KL Chan, Siew C Ng
The Lancet Gastroenterology & Hepatology (2020-07)
<https://doi.org/d2qq>
DOI: [10.1016/s2468-1253\(20\)30122-9](https://doi.org/10.1016/s2468-1253(20)30122-9) · PMID: [32339473](https://pubmed.ncbi.nlm.nih.gov/32339473/) · PMCID: [PMC7182525](https://pubmed.ncbi.nlm.nih.gov/PMC7182525/)
1765. **Bloomberg - Are you a robot?**
<https://www.bloomberg.com/tosv2.html?vid=&uuid=b91b9b90-6a34-11eb-a07d-15fd64b6d7f0&url=L3ByZXNzLXJlbGVhc2VzLzlwMjAtMDgtMDMvcHjvYmlvdGljcy1tYXJrZXQtd29ydGgtNzYtNy1iaWxsaW9uLWJ5LTlwlMjctZXhjbHVzaXZILXJlcG9ydC1jb3ZlcmluZy1wcmUtYW5kLXBvc3QtY292aWQtMTktbWFya2V0LWFuYWx5c2IzLWJ5LW1IdGljdWxvdXM=>
1766. **Vitamin D deficiency aggravates COVID-19: systematic review and meta-analysis**
Marcos Pereira, Alialdo Dantas Damascena, Laylla Mirella Galvão Azevedo, Tarcio de Almeida Oliveira, Jerusa da Mota Santana

1767. Diet and Inflammation

Leo Galland

Nutrition in Clinical Practice (2010-12-07) <https://doi.org/b7qgx7>

DOI: [10.1177/0884533610385703](https://doi.org/10884533610385703) · PMID: [21139128](#)

1768. Obesogenic diet in aging mice disrupts gut microbe composition and alters neutrophil:lymphocyte ratio, leading to inflamed milieu in acute heart failure

Vasundhara Kain, William Van Der Pol, Nithya Mariappan, Aftab Ahmad, Peter Eipers, Deanna L Gibson, Cecile Gladine, Claire Vigor, Thierry Durand, Casey Morrow, Ganesh V Halade

The FASEB Journal (2019-02-15) <https://doi.org/ghwfq8>

DOI: [10.1096/fj.201802477r](https://doi.org/fj.201802477r) · PMID: [30768364](#) · PMCID: [PMC6463911](#)

1769. Colloidal Silver

NCCIH

<https://www.nccih.nih.gov/health/colloidal-silver>

1770. Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2

Yadi Zhou, Yuan Hou, Jiayu Shen, Yin Huang, William Martin, Feixiong Cheng

Cell Discovery (2020-03-16) <https://doi.org/ggg84x>

DOI: [10.1038/s41421-020-0153-3](https://doi.org/s41421-020-0153-3) · PMID: [32194980](#) · PMCID: [PMC7073332](#)

1771. Role of Melatonin on Virus-Induced Neuropathogenesis—A Concomitant Therapeutic Strategy to Understand SARS-CoV-2 Infection

Prapimpun Wongchitrat, Mayuri Shukla, Ramaswamy Sharma, Piyarat Govitrapong, Russel J Reiter

Antioxidants (2021-01-02) <https://doi.org/ghr946>

DOI: [10.3390/antiox10010047](https://doi.org/10.3390/antiox10010047) · PMID: [33401749](#) · PMCID: [PMC7823793](#)

1772. Nutraceutical Strategies for Suppressing NLRP3 Inflammasome Activation: Pertinence to the Management of COVID-19 and Beyond

Mark F McCarty, Simon Bernard Iloki Assanga, Lidianys Lewis Luján, James H O'Keefe, James J DiNicolantonio

Nutrients (2020-12-25) <https://doi.org/ghr95g>

DOI: [10.3390/nu13010047](https://doi.org/nu13010047) · PMID: [33375692](#) · PMCID: [PMC7823562](#)

1773. Update: Here's what is known about Trump's COVID-19 treatment

Jon Cohen

Science (2020-10-05) <https://doi.org/ghr94n>

DOI: [10.1126/science.abf0974](https://doi.org/10.1126/science.abf0974)

1774. Dietary supplements during the COVID-19 pandemic: insights from 1.4M users of the COVID Symptom Study app - a longitudinal app-based community survey

Panayiotis Louca, Benjamin Murray, Kerstin Klaser, Mark S Graham, Mohsen Mazidi, Emily R Leeming, Ellen Thompson, Ruth Bowyer, David A Drew, Long H Nguyen, ... Cristina Menni
Cold Spring Harbor Laboratory (2020-11-30) <https://doi.org/ghr94k>
DOI: [10.1101/2020.11.27.20239087](https://doi.org/10.1101/2020.11.27.20239087)

1775. **ESPEN expert statements and practical guidance for nutritional management of individuals with SARS-CoV-2 infection**

Rocco Barazzoni, Stephan C Bischoff, Joao Breda, Kremlin Wickramasinghe, Zeljko Krznaric, Dorit Nitzan, Matthias Pirlich, Pierre Singer
Clinical Nutrition (2020-06) <https://doi.org/ggtzjq>
DOI: [10.1016/j.clnu.2020.03.022](https://doi.org/10.1016/j.clnu.2020.03.022) · PMID: [32305181](https://pubmed.ncbi.nlm.nih.gov/32305181/) · PMCID: [PMC7138149](https://pubmed.ncbi.nlm.nih.gov/PMC7138149/)

1776. **Nutritional status assessment in patients with Covid-19 after discharge from the intensive care unit**

Nassim Essabah Haraj, Siham El Aziz, Asma Chadli, Asma Dafir, Amal Mjabber, Ouissal Aissaoui, Lhoucine Barrou, Chafik El Kettani El Hamidi, Afak Nsiri, Rachid AL Harrar, ... Moulay Hicham Afif
Clinical Nutrition ESPEN (2021-02) <https://doi.org/ghjhdq>
DOI: [10.1016/j.clnesp.2020.09.214](https://doi.org/10.1016/j.clnesp.2020.09.214) · PMID: [33487301](https://pubmed.ncbi.nlm.nih.gov/33487301/) · PMCID: [PMC7552965](https://pubmed.ncbi.nlm.nih.gov/PMC7552965/)

1777. **Nutrition Status Affects COVID-19 Patient Outcomes**

Mette M Berger
Journal of Parenteral and Enteral Nutrition (2020-07-15)
<https://doi.org/gg5qv4>
DOI: [10.1002/jpen.1954](https://doi.org/10.1002/jpen.1954) · PMID: [32613691](https://pubmed.ncbi.nlm.nih.gov/32613691/) · PMCID: [PMC7361441](https://pubmed.ncbi.nlm.nih.gov/PMC7361441/)

1778. **Evaluation of Nutrition Risk and Its Association With Mortality Risk in Severely and Critically Ill COVID-19 Patients**

Xiaobo Zhao, Yan Li, Yanyan Ge, Yuxin Shi, Ping Lv, Jianchu Zhang, Gui Fu, Yanfen Zhou, Ke Jiang, Nengxing Lin, ... Xin Li
Journal of Parenteral and Enteral Nutrition (2020-07-20)
<https://doi.org/ghr93n>
DOI: [10.1002/jpen.1953](https://doi.org/10.1002/jpen.1953) · PMID: [32613660](https://pubmed.ncbi.nlm.nih.gov/32613660/) · PMCID: [PMC7361906](https://pubmed.ncbi.nlm.nih.gov/PMC7361906/)

1779. **Multisystem inflammatory syndrome in children: A systematic review**

Mubbasher Ahmed, Shailesh Advani, Axel Moreira, Sarah Zoretic, John Martinez, Kevin Chorath, Sebastian Acosta, Rija Naqvi, Finn Burmeister-Morton, Fiona Burmeister, ... Alvaro Moreira
EClinicalMedicine (2020-09) <https://doi.org/ghsv27>
DOI: [10.1016/j.eclinm.2020.100527](https://doi.org/10.1016/j.eclinm.2020.100527) · PMID: [32923992](https://pubmed.ncbi.nlm.nih.gov/32923992/) · PMCID: [PMC7473262](https://pubmed.ncbi.nlm.nih.gov/PMC7473262/)

1780. **Nutritional management of COVID-19 patients in a rehabilitation unit**

Luigia Brugliera, Alfio Spina, Paola Castellazzi, Paolo Cimino, Pietro Arcuri, Alessandra Negro, Elise Houdayer, Federica Alemanno, Alessandra Giordani, Pietro Mortini, Sandro Iannaccone
European Journal of Clinical Nutrition (2020-05-20)
<https://doi.org/gg29hf>

1781. **The frontier between nutrition and pharma: The international regulatory framework of functional foods, food supplements and nutraceuticals**
Laura Domínguez Díaz, Virginia Fernández-Ruiz, Montaña Cámara
Critical Reviews in Food Science and Nutrition (2019-03-29)
<https://doi.org/ggqs3w>
DOI: [10.1080/10408398.2019.1592107](https://doi.org/10408398.2019.1592107) · PMID: [30924346](https://pubmed.ncbi.nlm.nih.gov/30924346/)
1782. **Coronavirus Update: FDA and FTC Warn Seven Companies Selling Fraudulent Products that Claim to Treat or Prevent COVID-19**
Office of the Commissioner
FDA (2020-03-27) <https://www.fda.gov/news-events/press-announcements/coronavirus-update-fda-and-ftc-warn-seven-companies-selling-fraudulent-products-claim-treat-or>
1783. **COVID-19 and Your Health**
CDC
Centers for Disease Control and Prevention (2021-02-04)
<https://www.cdc.gov/coronavirus/2019-ncov/prevent-getting-sick/prevention.html>
1784. **Potential roles of social distancing in mitigating the spread of coronavirus disease 2019 (COVID-19) in South Korea**
Sang Woo Park, Kaiyuan Sun, Cécile Viboud, Bryan T Grenfell, Jonathan Dushoff
Cold Spring Harbor Laboratory (2020-03-30) <https://doi.org/gg3mhg>
DOI: [10.1101/2020.03.27.20045815](https://doi.org/10.1101/2020.03.27.20045815) · PMID: [32511429](https://pubmed.ncbi.nlm.nih.gov/32511429/) · PMCID: [PMC7217070](https://pubmed.ncbi.nlm.nih.gov/PMC7217070/)
1785. **Evaluating the Effectiveness of Social Distancing Interventions to Delay or Flatten the Epidemic Curve of Coronavirus Disease**
Laura Matrajt, Tiffany Leung
Emerging Infectious Diseases (2020-08) <https://doi.org/ggtx3k>
DOI: [10.3201/eid2608.201093](https://doi.org/10.3201/eid2608.201093) · PMID: [32343222](https://pubmed.ncbi.nlm.nih.gov/32343222/) · PMCID: [PMC7392458](https://pubmed.ncbi.nlm.nih.gov/PMC7392458/)
1786. **Association of Race With Mortality Among Patients Hospitalized With Coronavirus Disease 2019 (COVID-19) at 92 US Hospitals**
Baligh R Yehia, Angela Winegar, Richard Fogel, Mohamad Fakih, Allison Ottenbacher, Christine Jesser, Angelo Bufalino, Ren-Huai Huang, Joseph Cacchione
JAMA Network Open (2020-08-18) <https://doi.org/ghcspt>
DOI: [10.1001/jamanetworkopen.2020.18039](https://doi.org/10.1001/jamanetworkopen.2020.18039) · PMID: [32809033](https://pubmed.ncbi.nlm.nih.gov/32809033/) · PMCID: [PMC7435340](https://pubmed.ncbi.nlm.nih.gov/PMC7435340/)
1787. **Characteristics of and Important Lessons From the Coronavirus Disease 2019 (COVID-19) Outbreak in China**
Zunyou Wu, Jennifer M McGoogan
JAMA (2020-04-07) <https://doi.org/ggmq43>
DOI: [10.1001/jama.2020.2648](https://doi.org/10.1001/jama.2020.2648) · PMID: [32091533](https://pubmed.ncbi.nlm.nih.gov/32091533/)
1788. **Critical Care Utilization for the COVID-19 Outbreak in Lombardy, Italy**

Giacomo Grasselli, Antonio Pesenti, Maurizio Cecconi

JAMA (2020-04-28) <https://doi.org/gqgf6g>

DOI: [10.1001/jama.2020.4031](https://doi.org/jama.2020.4031) · PMID: [32167538](#)

1789. Hospitalization Rates and Characteristics of Patients Hospitalized with Laboratory-Confirmed Coronavirus Disease 2019 — COVID-NET, 14 States, March 1–30, 2020

Shikha Garg, Lindsay Kim, Michael Whitaker, Alissa O'Halloran, Charisse Cummings, Rachel Holstein, Mila Prill, Shua J Chai, Pam D Kirley, Nisha B Alden, ... Alicia Fry

MMWR. Morbidity and Mortality Weekly Report (2020-04-17)

<https://doi.org/ggsppz>

DOI: [10.15585/mmwr.mm6915e3](https://doi.org/mmwr.mm6915e3) · PMID: [32298251](#) · PMCID: [PMC7755063](#)

1790. COVID-19 and Your Health

CDC

Centers for Disease Control and Prevention (2020-02-11)

<https://www.cdc.gov/coronavirus/2019-ncov/need-extra-precautions/older-adults.html>

1791. Disparities In Outcomes Among COVID-19 Patients In A Large Health Care System In California

Kristen MJ Azar, Zijun Shen, Robert J Romanelli, Stephen H Lockhart, Kelly Smits, Sarah Robinson, Stephanie Brown, Alice R Pressman

Health Affairs (2020-07-01) <https://doi.org/gx4mf>

DOI: [10.1377/hlthaff.2020.00598](https://doi.org/hlthaff.2020.00598) · PMID: [32437224](#)

1792. Characteristics Associated with Hospitalization Among Patients with COVID-19 — Metropolitan Atlanta, Georgia, March–April 2020

Marie E Killerby, Ruth Link-Gelles, Sarah C Haight, Caroline A Schrodt, Lucinda England, Danica J Gomes, Mays Shamout, Kristen Pettrone, Kevin O'Laughlin, Anne Kimball, ... CDC COVID-19 Response Clinical Team

MMWR. Morbidity and Mortality Weekly Report (2020-06-26)

<https://doi.org/gg3k6h>

DOI: [10.15585/mmwr.mm6925e1](https://doi.org/mmwr.mm6925e1) · PMID: [32584797](#) · PMCID: [PMC7316317](#)

1793. Demographic science aids in understanding the spread and fatality rates of COVID-19

Jennifer Beam Dowd, Liliana Andriano, David M Brazel, Valentina Rotondi, Per Block, Xuejie Ding, Yan Liu, Melinda C Mills

Proceedings of the National Academy of Sciences (2020-05-05)

<https://doi.org/ggsd5b>

DOI: [10.1073/pnas.2004911117](https://doi.org/pnas.2004911117) · PMID: [32300018](#) · PMCID: [PMC7211934](#)

1794. -19 and Older Adults: What We Know

Zainab Shahid, Ricci Kalayanamitra, Brendan McClafferty, Douglas Kepko, Devyani Ramgobin, Ravi Patel, Chander Shekher Aggarwal, Ramarao Vunnam, Nitasa Sahu, Dhirisha Bhatt, ... Rohit Jain

Journal of the American Geriatrics Society (2020-04-20)

<https://doi.org/ggxgsb>

DOI: [10.1111/jgs.16472](https://doi.org/10.1111/jgs.16472) · PMID: [32255507](#) · PMCID: [PMC7262251](#)

1795. **Features of 20 133 UK patients in hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol: prospective observational cohort study**

Annemarie B Docherty, Ewen M Harrison, Christopher A Green, Hayley E Hardwick, Riinu Pius, Lisa Norman, Karl A Holden, Jonathan M Read, Frank Dondelinger, Gail Carson, ... Malcolm G Semple

BMJ (2020-05-22) <https://doi.org/ggw4nh>

DOI: [10.1136/bmj.m1985](https://doi.org/10.1136/bmj.m1985) · PMID: [32444460](https://pubmed.ncbi.nlm.nih.gov/32444460/) · PMCID: [PMC7243036](https://pubmed.ncbi.nlm.nih.gov/PMC7243036/)

1796. **Impact of sex and gender on COVID-19 outcomes in Europe**

Catherine Gebhard, Vera Regitz-Zagrosek, Hannelore K Neuhauser, Rosemary Morgan, Sabra L Klein

Biology of Sex Differences (2020-05-25) <https://doi.org/ghbvck>

DOI: [10.1186/s13293-020-00304-9](https://doi.org/10.1186/s13293-020-00304-9) · PMID: [32450906](https://pubmed.ncbi.nlm.nih.gov/32450906/) · PMCID: [PMC7247289](https://pubmed.ncbi.nlm.nih.gov/PMC7247289/)

1797. **The Sex, Gender and COVID-19 Project | Global Health 50/50**

<https://globalhealth5050.org/the-sex-gender-and-covid-19-project/>

1798. **Biological sex impacts COVID-19 outcomes**

Sabra L Klein, Santosh Dhakal, Rebecca L Ursin, Sharvari Deshpande, Kathryn Sandberg, Franck Mauvais-Jarvis

PLOS Pathogens (2020-06-22) <https://doi.org/gg3hwv>

DOI: [10.1371/journal.ppat.1008570](https://doi.org/10.1371/journal.ppat.1008570) · PMID: [32569293](https://pubmed.ncbi.nlm.nih.gov/32569293/) · PMCID: [PMC7307725](https://pubmed.ncbi.nlm.nih.gov/PMC7307725/)

1799. **Sex-specific clinical characteristics and prognosis of coronavirus disease-19 infection in Wuhan, China: A retrospective study of 168 severe patients**

Yifan Meng, Ping Wu, Wanrong Lu, Kui Liu, Ke Ma, Liang Huang, Jiaoqiao Cai, Hong Zhang, Yu Qin, Haiying Sun, ... Peng Wu

PLOS Pathogens (2020-04-28) <https://doi.org/ggv3zn>

DOI: [10.1371/journal.ppat.1008520](https://doi.org/10.1371/journal.ppat.1008520) · PMID: [32343745](https://pubmed.ncbi.nlm.nih.gov/32343745/) · PMCID: [PMC7209966](https://pubmed.ncbi.nlm.nih.gov/PMC7209966/)

1800. **Sex differences in renal angiotensin converting enzyme 2 (ACE2) activity are 17 β -oestradiol-dependent and sex chromosome-independent**

Jun Liu, Hong Ji, Wei Zheng, Xie Wu, Janet J Zhu, Arthur P Arnold, Kathryn Sandberg

Biology of Sex Differences (2010) <https://doi.org/bbx3r6>

DOI: [10.1186/2042-6410-1-6](https://doi.org/10.1186/2042-6410-1-6) · PMID: [21208466](https://pubmed.ncbi.nlm.nih.gov/21208466/) · PMCID: [PMC3010099](https://pubmed.ncbi.nlm.nih.gov/PMC3010099/)

1801. **COVID-19 in nursing homes**

A Fallon, T Dukelow, SP Kennelly, D O'Neill

QJM: An International Journal of Medicine (2020-06)

<https://doi.org/ggy4xx>

DOI: [10.1093/qjmed/hcaa136](https://doi.org/10.1093/qjmed/hcaa136) · PMID: [32311049](https://pubmed.ncbi.nlm.nih.gov/32311049/) · PMCID: [PMC7188176](https://pubmed.ncbi.nlm.nih.gov/PMC7188176/)

1802. **Vulnerabilities to COVID-19 Among Transgender Adults in the U.S.**

Jody L Herman, Kathryn O'Neill

(2020-04-01) <https://escholarship.org/uc/item/55t297mc>

1803. **Association of Blood Glucose Control and Outcomes in Patients with COVID-19 and Pre-existing Type 2 Diabetes**

Lihua Zhu, Zhi-Gang She, Xu Cheng, Juan-Juan Qin, Xiao-Jing Zhang, Jingjing Cai, Fang Lei, Haitao Wang, Jing Xie, Wenxin Wang, ... Hongliang Li

Cell Metabolism (2020-06) <https://doi.org/ggvcc9>

DOI: [10.1016/j.cmet.2020.04.021](https://doi.org/10.1016/j.cmet.2020.04.021) · PMID: [32369736](https://pubmed.ncbi.nlm.nih.gov/32369736/) · PMCID: [PMC7252168](https://pubmed.ncbi.nlm.nih.gov/PMC7252168/)

1804. **Diabetes increases the mortality of patients with COVID-19: a meta-analysis**

Zeng-hong Wu, Yun Tang, Qing Cheng

Acta Diabetologica (2020-06-24) <https://doi.org/gg3k55>

DOI: [10.1007/s00592-020-01546-0](https://doi.org/10.1007/s00592-020-01546-0) · PMID: [32583078](https://pubmed.ncbi.nlm.nih.gov/32583078/) · PMCID: [PMC7311595](https://pubmed.ncbi.nlm.nih.gov/PMC7311595/)

1805. **COVID-19 infection may cause ketosis and ketoacidosis**

Juyi Li, Xiufang Wang, Jian Chen, Xiuran Zuo, Hongmei Zhang, Aiping Deng

Diabetes, Obesity and Metabolism (2020-05-18) <https://doi.org/ggv4tm>

DOI: [10.1111/dom.14057](https://doi.org/10.1111/dom.14057) · PMID: [32314455](https://pubmed.ncbi.nlm.nih.gov/32314455/) · PMCID: [PMC7264681](https://pubmed.ncbi.nlm.nih.gov/PMC7264681/)

1806. **COVID-19 pandemic, coronaviruses, and diabetes mellitus**

Ranganath Muniyappa, Sriram Gubbi

American Journal of Physiology-Endocrinology and Metabolism (2020-05-01) <https://doi.org/ggq79v>

DOI: [10.1152/ajpendo.00124.2020](https://doi.org/10.1152/ajpendo.00124.2020) · PMID: [32228322](https://pubmed.ncbi.nlm.nih.gov/32228322/) · PMCID: [PMC7191633](https://pubmed.ncbi.nlm.nih.gov/PMC7191633/)

1807. **Severe obesity, increasing age and male sex are independently associated with worse in-hospital outcomes, and higher in-hospital mortality, in a cohort of patients with COVID-19 in the Bronx, New York**

Leonidas Palaiodimos, Damianos G Kokkinidis, Weijia Li, Dimitrios Karamanis, Jennifer Ognibene, Shitij Arora, William N Southern, Christos S Mantzoros

Metabolism (2020-07) <https://doi.org/ggx229>

DOI: [10.1016/j.metabol.2020.154262](https://doi.org/10.1016/j.metabol.2020.154262) · PMID: [32422233](https://pubmed.ncbi.nlm.nih.gov/32422233/) · PMCID: [PMC7228874](https://pubmed.ncbi.nlm.nih.gov/PMC7228874/)

1808. **Features of 16,749 hospitalised UK patients with COVID-19 using the ISARIC WHO Clinical Characterisation Protocol**

AB Docherty, EM Harrison, CA Green, H Hardwick, R Pius, L Norman, KA Holden, JM Read, F Dondelinger, G Carson, ... ISARIC4C Investigators
Cold Spring Harbor Laboratory (2020-04-28) <https://doi.org/ggtedb>

DOI: [10.1101/2020.04.23.20076042](https://doi.org/10.1101/2020.04.23.20076042)

1809. **When Two Pandemics Meet: Why Is Obesity Associated with Increased COVID-19 Mortality?**

Sam M Lockhart, Stephen O'Rahilly

Med (2020-12) <https://doi.org/gg3k57>

DOI: [10.1016/j.medj.2020.06.005](https://doi.org/10.1016/j.medj.2020.06.005) · PMID: [32838359](https://pubmed.ncbi.nlm.nih.gov/32838359/) · PMCID: [PMC7323660](https://pubmed.ncbi.nlm.nih.gov/PMC7323660/)

1810. **Besides population age structure, health and other demographic factors can contribute to understanding the COVID-19 burden**

Marília R Nepomuceno, Enrique Acosta, Diego Alburez-Gutierrez, José Manuel Aburto, Alain Gagnon, Cássio M Turra
Proceedings of the National Academy of Sciences (2020-06-23)
<https://doi.org/gg33qx>
DOI: [10.1073/pnas.2008760117](https://doi.org/10.1073/pnas.2008760117) · PMID: [32576710](https://pubmed.ncbi.nlm.nih.gov/32576710/) · PMCID: [PMC7322063](https://pubmed.ncbi.nlm.nih.gov/PMC7322063/)

1811. <https://www.who.int/publications/i/item/report-of-the-who-china-joint-mission-on-coronavirus-disease-2019>

1812. **The SOFA (Sepsis-related Organ Failure Assessment) score to describe organ dysfunction/failure**

J-L Vincent, R Moreno, J Takala, S Willatts, A De Mendonça, H Bruining, CK Reinhart, PM Suter, LG Thijs
Intensive Care Medicine (1996-07) <https://doi.org/bpkxdw>
DOI: [10.1007/bf01709751](https://doi.org/10.1007/bf01709751) · PMID: [8844239](https://pubmed.ncbi.nlm.nih.gov/8844239/)

1813. **The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3)**

Mervyn Singer, Clifford S Deutschman, Christopher Warren Seymour, Manu Shankar-Hari, Djillali Annane, Michael Bauer, Rinaldo Bellomo, Gordon R Bernard, Jean-Daniel Chiche, Craig M Coopersmith, ... Derek C Angus
JAMA (2016-02-23) <https://doi.org/gdrcdh>
DOI: [10.1001/jama.2016.0287](https://doi.org/10.1001/jama.2016.0287) · PMID: [26903338](https://pubmed.ncbi.nlm.nih.gov/26903338/) · PMCID: [PMC4968574](https://pubmed.ncbi.nlm.nih.gov/PMC4968574/)

1814. **COVID-19 and African Americans**

Clyde W Yancy
JAMA (2020-05-19) <https://doi.org/ggv494>
DOI: [10.1001/jama.2020.6548](https://doi.org/10.1001/jama.2020.6548) · PMID: [32293639](https://pubmed.ncbi.nlm.nih.gov/32293639/)

1815. **COVID-19 and Racial/Ethnic Disparities**

Monica Webb Hooper, Anna María Nápoles, Eliseo J Pérez-Stable
JAMA (2020-06-23) <https://doi.org/ggvzqn>
DOI: [10.1001/jama.2020.8598](https://doi.org/10.1001/jama.2020.8598) · PMID: [32391864](https://pubmed.ncbi.nlm.nih.gov/32391864/)

1816. **Covid-19: Black people and other minorities are hardest hit in US**

Owen Dyer
BMJ (2020-04-14) <https://doi.org/ggy5br>
DOI: [10.1136/bmj.m1483](https://doi.org/10.1136/bmj.m1483) · PMID: [32291262](https://pubmed.ncbi.nlm.nih.gov/32291262/)

1817. **Susceptibility of Southwestern American Indian Tribes to Coronavirus Disease 2019 (COVID-19)**

Monika Kakol, Dona Upson, Akshay Sood
The Journal of Rural Health (2020-06) <https://doi.org/ggtzkq>
DOI: [10.1111/jrh.12451](https://doi.org/10.1111/jrh.12451) · PMID: [32304251](https://pubmed.ncbi.nlm.nih.gov/32304251/) · PMCID: [PMC7264672](https://pubmed.ncbi.nlm.nih.gov/PMC7264672/)

1818. **The Fullest Look Yet at the Racial Inequity of Coronavirus**

Richard AOppel Jr, Robert Gebeloff, KKRebecca Lai, Will Wright, Mitch Smith
The New York Times (2020-07-05)
<https://www.nytimes.com/interactive/2020/07/05/us/coronavirus-latino-african-americans-cdc-data.html>

1819. **Addressing inequities in COVID-19 morbidity and mortality: research and policy recommendations**

Monica L Wang, Pamela Behrman, Akilah Dulin, Monica L Baskin, Joanna Buscemi, Kassandra I Alcaraz, Carly M Goldstein, Tiffany L Carson, Megan Shen, Marian Fitzgibbon

Translational Behavioral Medicine (2020-06) <https://doi.org/gg3389>

DOI: [10.1093/tbm/iba055](https://doi.org/10.1093/tbm/iba055) · PMID: [32542349](https://pubmed.ncbi.nlm.nih.gov/32542349/) · PMCID: [PMC7337775](https://pubmed.ncbi.nlm.nih.gov/PMC7337775/)

1820. **Historical Environmental Racism, Structural Inequalities, and Dik'os Ntsaaígíí-19 (COVID-19) on Navajo Nation**

Nicholet A Deschine Parkhurst, Kimberly R Huyser, Aggie J Yellow Horse
Journal of Indigenous Social Development (2020-11-02)

<https://journalhosting.ucalgary.ca/index.php/jisd/article/view/70753>

1821. **Protect Indigenous peoples from COVID-19**

Lucas Ferrante, Philip M Fearnside

Science (2020-04-16) <https://doi.org/gg3k6f>

DOI: [10.1126/science.abc0073](https://doi.org/10.1126/science.abc0073) · PMID: [32299940](https://pubmed.ncbi.nlm.nih.gov/32299940/)

1822. **Factors associated with COVID-19-related death using OpenSAFELY**

Elizabeth J Williamson, Alex J Walker, Krishnan Bhaskaran, Seb Bacon, Chris Bates, Caroline E Morton, Helen J Curtis, Amir Mehrkar, David Evans, Peter Inglesby, ... Ben Goldacre

Nature (2020-07-08) <https://doi.org/gg39n7>

DOI: [10.1038/s41586-020-2521-4](https://doi.org/10.1038/s41586-020-2521-4) · PMID: [32640463](https://pubmed.ncbi.nlm.nih.gov/32640463/)

1823. **Implications of biogeography of human populations for 'race' and medicine**

Sarah A Tishkoff, Kenneth K Kidd

Nature Genetics (2004-10-26) <https://doi.org/d2xq92>

DOI: [10.1038/ng1438](https://doi.org/10.1038/ng1438) · PMID: [15507999](https://pubmed.ncbi.nlm.nih.gov/15507999/)

1824. **African Genetic Diversity: Implications for Human Demographic History, Modern Human Origins, and Complex Disease Mapping**

Michael C Campbell, Sarah A Tishkoff

Annual Review of Genomics and Human Genetics (2008-09)

<https://doi.org/cphggp>

DOI: [10.1146/annurev.genom.9.081307.164258](https://doi.org/10.1146/annurev.genom.9.081307.164258) · PMID: [18593304](https://pubmed.ncbi.nlm.nih.gov/18593304/) ·

PMCID: [PMC2953791](https://pubmed.ncbi.nlm.nih.gov/PMC2953791/)

1825. **NIH must confront the use of race in science**

Michael Yudell, Dorothy Roberts, Rob DeSalle, Sarah Tishkoff, 70 signatories

Science (2020-09-10) <https://doi.org/ghcm7s>

DOI: [10.1126/science.abd4842](https://doi.org/10.1126/science.abd4842) · PMID: [32913094](https://pubmed.ncbi.nlm.nih.gov/32913094/)

1826. **APOE e4 Genotype Predicts Severe COVID-19 in the UK Biobank Community Cohort**

Chia-Ling Kuo, Luke C Pilling, Janice L Atkins, Jane AH Masoli, João Delgado, George A Kuchel, David Melzer

The Journals of Gerontology: Series A (2020-11) <https://doi.org/ggx4ng>

DOI: [10.1093/gerona/glaa131](https://doi.org/10.1093/gerona/glaa131) · PMID: [32451547](https://pubmed.ncbi.nlm.nih.gov/32451547/) · PMCID: [PMC7314139](https://pubmed.ncbi.nlm.nih.gov/PMC7314139/)

1827. **Genome-wide CRISPR screen reveals host genes that regulate SARS-CoV-2 infection**

Jin Wei, Mia Madel Alfajaro, Ruth E Hanna, Peter C DeWeirdt, Madison S Strine, William J Lu-Culligan, Shang-Min Zhang, Vincent R Graziano, Cameron O Schmitz, Jennifer S Chen, ... Craig B Wilen
Cold Spring Harbor Laboratory (2020-06-17) <https://doi.org/dzz3>
DOI: [10.1101/2020.06.16.155101](https://doi.org/10.1101/2020.06.16.155101) · PMID: [32869025](https://pubmed.ncbi.nlm.nih.gov/32869025/) · PMCID: [PMC7457610](https://pubmed.ncbi.nlm.nih.gov/PMC7457610/)

1828. **New insights into genetic susceptibility of COVID-19: an ACE2 and TMPRSS2 polymorphism analysis**

Yuan Hou, Junfei Zhao, William Martin, Asha Kallianpur, Mina K Chung, Lara Jehi, Nima Sharifi, Serpil Erzurum, Charis Eng, Feixiong Cheng
BMC Medicine (2020-07-15) <https://doi.org/gg445n>
DOI: [10.1186/s12916-020-01673-z](https://doi.org/10.1186/s12916-020-01673-z) · PMID: [32664879](https://pubmed.ncbi.nlm.nih.gov/32664879/) · PMCID: [PMC7360473](https://pubmed.ncbi.nlm.nih.gov/PMC7360473/)

1829. **Seroprevalence of anti-SARS-CoV-2 IgG antibodies in Kenyan blood donors**

Sophie Uyoga, Ifedayo M.O. Adetifa, Henry K Karanja, James Nyagwange, James Tuju, Perpetual Wanjiku, Rashid Aman, Mercy Mwangangi, Patrick Amoth, Kadondi Kasera, ... George M Warimwe
Cold Spring Harbor Laboratory (2020-07-29) <https://doi.org/ghcm7p>
DOI: [10.1101/2020.07.27.20162693](https://doi.org/10.1101/2020.07.27.20162693)

1830. **High SARS-CoV-2 seroprevalence in Health Care Workers but relatively low numbers of deaths in urban Malawi**

Marah G Chibwana, Khuzwayo C Jere, Raphael Kamn'gona, Jonathan Mandolo, Vincent Katunga-Phiri, Dumizulu Tembo, Ndaona Mitole, Samantha Musasa, Simon Sichone, Agness Lakudzala, ... Kondwani C Jambo

Cold Spring Harbor Laboratory (2020-08-01) <https://doi.org/ghcm7q>
DOI: [10.1101/2020.07.30.20164970](https://doi.org/10.1101/2020.07.30.20164970) · PMID: [32766597](https://pubmed.ncbi.nlm.nih.gov/32766597/) · PMCID: [PMC7402052](https://pubmed.ncbi.nlm.nih.gov/PMC7402052/)

1831. **Africa's pandemic puzzle: why so few cases and deaths?**

Linda Nordling
Science (2020-08-13) <https://doi.org/ghcm7r>
DOI: [10.1126/science.369.6505.756](https://doi.org/10.1126/science.369.6505.756) · PMID: [32792376](https://pubmed.ncbi.nlm.nih.gov/32792376/)

1832. **Are some ethnic groups more vulnerable to COVID-19 than others?** <https://www.ifs.org.uk/inequality/chapter/are-some-ethnic-groups-more-vulnerable-to-covid-19-than-others/>

1833. **Quantifying the social distancing privilege gap: a longitudinal study of smartphone movement**

Nabarun Dasgupta, Michele Jonsson Funk, Allison Lazard, Benjamin Eugene White, Stephen W Marshall
Cold Spring Harbor Laboratory (2020-05-08) <https://doi.org/gg79qk>
DOI: [10.1101/2020.05.03.20084624](https://doi.org/10.1101/2020.05.03.20084624)

1834. **Uncovering socioeconomic gaps in mobility reduction during the COVID-19 pandemic using location data**

Samuel P Fraiberger, Pablo Astudillo, Lorenzo Candeago, Alex Chunet, Nicholas KW Jones, Maham Faisal Khan, Bruno Lepri, Nancy Lozano

1835. **Mobility network models of COVID-19 explain inequities and inform reopening**

Serina Chang, Emma Pierson, Pang Wei Koh, Jaline Gerardin, Beth Redbird, David Grusky, Jure Leskovec

Nature (2020-11-10) <https://doi.org/ghjmt2>

DOI: [10.1038/s41586-020-2923-3](https://doi.org/10.1038/s41586-020-2923-3) · PMID: [33171481](#)

1836. **A Basic Demographic Profile of Workers in Frontline Industries**

Hye Jin Rho;Shawn Fremstad;Hayley Brown

(2020-06) <https://mronline.org/wp-content/uploads/2020/06/2020-04-Frontline-Workers.pdf>

1837. **Differential occupational risk for COVID-19 and other infection exposure according to race and ethnicity**

Devan Hawkins

American Journal of Industrial Medicine (2020-06-15)

<https://doi.org/gg3rb2>

DOI: [10.1002/ajim.23145](https://doi.org/10.1002/ajim.23145) · PMID: [32539166](#) · PMCID: [PMC7323065](#)

1838. **Estimating the burden of United States workers exposed to infection or disease: A key factor in containing risk of COVID-19 infection**

Marissa G Baker, Trevor K Peckham, Noah S Seixas

PLOS ONE (2020-04-28) <https://doi.org/ggtx7c>

DOI: [10.1371/journal.pone.0232452](https://doi.org/10.1371/journal.pone.0232452) · PMID: [32343747](#) · PMCID: [PMC7188235](#)

1839. **Coronavirus (COVID-19) related deaths by occupation, England and Wales: deaths registered up to and including 20 April 2020**

Ben Windsor-Shellard, Jasveer Kaur

(2020-05-11)

<https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/causesofdeath/bulletins/coronaviruscovid19relateddeathsbyoccupationenglandandwales/deathsregistereduptoandincluding20april2020>

1840. **Which occupations have the highest potential exposure to the coronavirus (COVID-19)?**

Office for National Statistics

(2020-05-11)

<https://www.ons.gov.uk/employmentandlabourmarket/peopleinwork/employmentandemployeetypes/articles/whichoccupationshavethehighestpotentialexposuretothecoronaviruscovid19/2020-05-11>

1841. **Disparities in the risk and outcomes from COVID-19**

Public Health England

(2020-06-12)

https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/892085/disparities_review.pdf

1842. **Exclusive: deaths of NHS staff from covid-19 analysed**

Tim Cook, Emira Kursumovic, Simon Lennane 2020-04-22T12:42:00+01:00

Health Service Journal <https://www.hsj.co.uk/exclusive-deaths-of-nhs-staff-from-covid-19-analysed/7027471.article>

1843. <https://act.nationalnursesunited.org/page>

1844. Racial Disparity in COVID-19 Deaths: Seeking Economic Roots with Census data.

John McLaren

National Bureau of Economic Research (2020-06-22)

<https://www.nber.org/papers/w27407>

1845. Mortality, Admissions, and Patient Census at SNFs in 3 US Cities During the COVID-19 Pandemic

Michael L Barnett, Lissy Hu, Thomas Martin, David C Grabowski

JAMA (2020-08-04) <https://doi.org/gg3387>

DOI: [10.1001/jama.2020.11642](https://doi.org/10.1001/jama.2020.11642) · PMID: [32579161](https://pubmed.ncbi.nlm.nih.gov/32579161/) · PMCID: [PMC7315390](https://pubmed.ncbi.nlm.nih.gov/PMC7315390/)

1846. COVID-19 in Prisons and Jails in the United States

Laura Hawks, Steffie Woolhandler, Danny McCormick

JAMA Internal Medicine (2020-08-01) <https://doi.org/ggtxw6>

DOI: [10.1001/jamainternmed.2020.1856](https://doi.org/10.1001/jamainternmed.2020.1856) · PMID: [32343355](https://pubmed.ncbi.nlm.nih.gov/32343355/)

1847. COVID-19 Cases and Deaths in Federal and State Prisons

Brendan Saloner, Kalind Parish, Julie A Ward, Grace DiLaura, Sharon Dolovich

JAMA (2020-08-11) <https://doi.org/gg4dcv>

DOI: [10.1001/jama.2020.12528](https://doi.org/10.1001/jama.2020.12528) · PMID: [32639537](https://pubmed.ncbi.nlm.nih.gov/32639537/) · PMCID: [PMC7344796](https://pubmed.ncbi.nlm.nih.gov/PMC7344796/)

1848. State Rates of Incarceration Race & Ethnicity_updated2

nlizanna

(2018-03-24) <https://www.issuelab.org/resources/695/695.pdf>

1849. Under One Roof: A Review of Research on Intergenerational Coresidence and Multigenerational Households in the United States

Jennifer Reid Keene, Christie D Batson

Sociology Compass (2010-08) <https://doi.org/fsrcr7>

DOI: [10.1111/j.1751-9020.2010.00306.x](https://doi.org/10.1111/j.1751-9020.2010.00306.x)

1850. Chaos and the macrosetting: The role of poverty and socioeconomic status.

Gary W Evans, John Eckenrode, Lyscha A Marcynyszyn

American Psychological Association (APA) (2010-01-12)

<https://doi.org/c76n3g>

DOI: [10.1037/12057-014](https://doi.org/10.1037/12057-014)

1851. Housing insecurity among urban fathers

Marah A Curtis, Amanda B Geller

Columbia University (2010) <https://doi.org/ghdjn2>

DOI: [10.7916/d8wh2w9t](https://doi.org/10.7916/d8wh2w9t)

1852. Housing and Employment Insecurity among the Working Poor

Matthew Desmond, Carl Gershenson
Social Problems (2016-02) <https://doi.org/f8crm2>
DOI: [10.1093/socpro/spv025](https://doi.org/10.1093/socpro/spv025)

1853. **Obesity and its comorbid conditions**

Lalita Khaodhiar, Karen C McCowen, George L Blackburn
Clinical Cornerstone (1999-01) <https://doi.org/bpp37d>
DOI: [10.1016/s1098-3597\(99\)90002-9](https://doi.org/10.1016/s1098-3597(99)90002-9)

1854. **Aging, Male Sex, Obesity, and Metabolic Inflammation Create the Perfect Storm for COVID-19**

Franck Mauvais-Jarvis
Diabetes (2020-09) <https://doi.org/gg47zk>
DOI: [10.2337/dbi19-0023](https://doi.org/10.2337/dbi19-0023) · PMID: [32669390](https://pubmed.ncbi.nlm.nih.gov/32669390/) · PMCID: [PMC7458034](https://pubmed.ncbi.nlm.nih.gov/PMC7458034/)

1855. **Non-communicable disease syndemics: poverty, depression, and diabetes among low-income populations**

Emily Mendenhall, Brandon A Kohrt, Shane A Norris, David Ndetei, Dorairaj Prabhakaran
The Lancet (2017-03) <https://doi.org/gddg84>
DOI: [10.1016/s0140-6736\(17\)30402-6](https://doi.org/10.1016/s0140-6736(17)30402-6) · PMID: [28271846](https://pubmed.ncbi.nlm.nih.gov/28271846/) · PMCID: [PMC5491333](https://pubmed.ncbi.nlm.nih.gov/PMC5491333/)

1856. **Obesity and poverty paradox in developed countries**

Wioletta Żukiewicz-Sobczak, Paula Wróblewska, Jacek Zwoliński, Jolanta Chmielewska-Badora, Piotr Adamczuk, Ewelina Krasowska, Jerzy Zagórski, Anna Oniszczuk, Jacek Piątek, Wojciech Silny
Annals of Agricultural and Environmental Medicine (2014-09-04) <https://doi.org/f6jhzc>
DOI: [10.5604/12321966.1120608](https://doi.org/10.5604/12321966.1120608) · PMID: [25292135](https://pubmed.ncbi.nlm.nih.gov/25292135/)

1857. **Impact of the COVID-19 Pandemic on Unhealthy Eating in Populations with Obesity**

Nathaniel JS Ashby
Obesity (2020-08-20) <https://doi.org/ghd6qc>
DOI: [10.1002/oby.22940](https://doi.org/10.1002/oby.22940) · PMID: [32589788](https://pubmed.ncbi.nlm.nih.gov/32589788/) · PMCID: [PMC7361200](https://pubmed.ncbi.nlm.nih.gov/PMC7361200/)

1858. **Fast Food Patronage and Obesity Prevalence During the COVID-19 Pandemic: An Alternative Explanation**

Candice A Myers, Stephanie T Broyles
Obesity (2020-09-03) <https://doi.org/gg6v84>
DOI: [10.1002/oby.22993](https://doi.org/10.1002/oby.22993) · PMID: [32741130](https://pubmed.ncbi.nlm.nih.gov/32741130/) · PMCID: [PMC7435526](https://pubmed.ncbi.nlm.nih.gov/PMC7435526/)

1859. **The global food syndemic: The impact of food insecurity, Malnutrition and obesity on the healthspan amid the COVID-19 pandemic**

Martha I Huizar, Ross Arena, Deepika R Laddu
Progress in Cardiovascular Diseases (2020-07) <https://doi.org/gg4r3h>
DOI: [10.1016/j.pcad.2020.07.002](https://doi.org/10.1016/j.pcad.2020.07.002) · PMID: [32653438](https://pubmed.ncbi.nlm.nih.gov/32653438/) · PMCID: [PMC7347484](https://pubmed.ncbi.nlm.nih.gov/PMC7347484/)

1860. **Stress, chronic inflammation, and emotional and physical well-being: Concurrent effects and chronic sequelae**

George P Chrousos

Journal of Allergy and Clinical Immunology (2000-11)

<https://doi.org/bgx7hn>

DOI: [10.1067/mai.2000.110163](https://doi.org/ma.2000.110163) · PMID: [11080744](https://pubmed.ncbi.nlm.nih.gov/11080744/)

1861. **Chronic psychological stress and the regulation of pro-inflammatory cytokines: A glucocorticoid-resistance model.**

Gregory E Miller, Sheldon Cohen, AKim Ritchey

Health Psychology (2002) <https://doi.org/dj5r8b>

DOI: [10.1037/0278-6133.21.6.531](https://doi.org/10.1037/0278-6133.21.6.531)

1862. **Chronic stress, daily stressors, and circulating inflammatory markers.**

Jean-Philippe Gouin, Ronald Glaser, William B Malarkey, David Beversdorf, Janice Kiecolt-Glaser

Health Psychology (2012-03) <https://doi.org/dkz9tr>

DOI: [10.1037/a0025536](https://doi.org/10.1037/a0025536) · PMID: [21928900](https://pubmed.ncbi.nlm.nih.gov/21928900/) · PMCID: [PMC3253267](https://pubmed.ncbi.nlm.nih.gov/PMC3253267/)

1863. **Turning Up the Heat**

Gregory E Miller, Ekin Blackwell

Current Directions in Psychological Science (2016-06-24)

<https://doi.org/bft9mv>

DOI: [10.1111/j.1467-8721.2006.00450.x](https://doi.org/10.1111/j.1467-8721.2006.00450.x)

1864. **Sick of Poverty**

Robert Sapolsky

Scientific American (2005-12) <https://doi.org/fxf5kp>

DOI: [10.1038/scientificamerican1205-92](https://doi.org/10.1038/scientificamerican1205-92) · PMID: [16323696](https://pubmed.ncbi.nlm.nih.gov/16323696/)

1865. **Exposure to air pollution and COVID-19 mortality in the United States: A nationwide cross-sectional study**

Xiao Wu, Rachel C Nethery, MBenjamin Sabath, Danielle Braun, Francesca Dominici

Cold Spring Harbor Laboratory (2020-04-27) <https://doi.org/ggrpcj>

DOI: [10.1101/2020.04.05.20054502](https://doi.org/10.1101/2020.04.05.20054502) · PMID: [32511651](https://pubmed.ncbi.nlm.nih.gov/32511651/) · PMCID: [PMC7277007](https://pubmed.ncbi.nlm.nih.gov/PMC7277007/)

1866. **Global estimates of mortality associated with long-term exposure to outdoor fine particulate matter**

Richard Burnett, Hong Chen, Mieczysław Szyszkowicz, Neal Fann, Bryan Hubbell, CArden Pope, Joshua S Apte, Michael Brauer, Aaron Cohen, Scott Weichenthal, ... Joseph V Spadaro

Proceedings of the National Academy of Sciences (2018-09-18)

<https://doi.org/gfgbcx>

DOI: [10.1073/pnas.1803222115](https://doi.org/10.1073/pnas.1803222115) · PMID: [30181279](https://pubmed.ncbi.nlm.nih.gov/30181279/) · PMCID: [PMC6156628](https://pubmed.ncbi.nlm.nih.gov/PMC6156628/)

1867. **Early-Life Air Pollution Exposure, Neighborhood Poverty, and Childhood Asthma in the United States, 1990–2014**

Nicole Kravitz-Wirtz, Samantha Teixeira, Anjum Hajat, Bongki Woo, Kyle Crowder, David Takeuchi

International Journal of Environmental Research and Public Health

(2018-05-30) <https://doi.org/gdvwp9>

DOI: [10.3390/ijerph15061114](https://doi.org/10.3390/ijerph15061114) · PMID: [29848979](https://pubmed.ncbi.nlm.nih.gov/29848979/) · PMCID: [PMC6025399](https://pubmed.ncbi.nlm.nih.gov/PMC6025399/)

1868. **Early life stress, air pollution, inflammation, and disease: An integrative review and immunologic model of social-environmental adversity and lifespan health**

Hector A Olvera Alvarez, Laura D Kubzansky, Matthew J Campen, George M Slavich

Neuroscience & Biobehavioral Reviews (2018-09)

<https://doi.org/gd46bm>

DOI: [10.1016/j.neubiorev.2018.06.002](https://doi.org/10.1016/j.neubiorev.2018.06.002) · PMID: [29874545](https://pubmed.ncbi.nlm.nih.gov/29874545/) · PMCID: [PMC6082389](https://pubmed.ncbi.nlm.nih.gov/PMC6082389/)

1869. **Covid-19 and Disparities in Nutrition and Obesity**

Matthew J Belanger, Michael A Hill, Angeliki M Angelidi, Maria Dalamaga, James R Sowers, Christos S Mantzoros

New England Journal of Medicine (2020-09-10) <https://doi.org/gg475x>

DOI: [10.1056/nejmp2021264](https://doi.org/10.1056/nejmp2021264) · PMID: [32668105](https://pubmed.ncbi.nlm.nih.gov/32668105/)

1870. **Systemic racism, chronic health inequities, and -19: A syndemic in the making?**

Clarence C Gravlee

American Journal of Human Biology (2020-08-04)

<https://doi.org/ghcxwk>

DOI: [10.1002/ajhb.23482](https://doi.org/10.1002/ajhb.23482) · PMID: [32754945](https://pubmed.ncbi.nlm.nih.gov/32754945/) · PMCID: [PMC7441277](https://pubmed.ncbi.nlm.nih.gov/PMC7441277/)

1871. **Racial and Ethnic Health Disparities Related to COVID-19**

Leo Lopez III, Louis H Hart III, Mitchell H Katz

JAMA (2021-02-23) <https://doi.org/gh6bsm>

DOI: [10.1001/jama.2020.26443](https://doi.org/10.1001/jama.2020.26443) · PMID: [33480972](https://pubmed.ncbi.nlm.nih.gov/33480972/)

1872. **Coronavirus Disease (COVID-19): A primer for emergency physicians**

Summer Chavez, Brit Long, Alex Koyfman, Stephen Y Liang

The American Journal of Emergency Medicine (2020-03)

<https://doi.org/ggr22z>

DOI: [10.1016/j.ajem.2020.03.036](https://doi.org/10.1016/j.ajem.2020.03.036) · PMID: [32265065](https://pubmed.ncbi.nlm.nih.gov/32265065/) · PMCID: [PMC7102516](https://pubmed.ncbi.nlm.nih.gov/PMC7102516/)

1873. **Insurers May Only Pay For Coronavirus Tests When They're 'Medically Necessary'**

NPR.org

<https://www.npr.org/sections/health-shots/2020/06/19/880543755/insurers-may-only-pay-for-coronavirus-tests-when-theyre-medically-necessary>

1874. **Private Health Insurance Coverage in the COVID-19 Public Health Emergency | Commonwealth Fund**

<https://www.commonwealthfund.org/blog/2020/private-health-insurance-coverage-covid-19-public-health-emergency>

1875. **COVID-19 and racial disparities**

Monica Shah, Muskaan Sachdeva, Roni P Dodiuk-Gad

Journal of the American Academy of Dermatology (2020-07)

<https://doi.org/ggtwm7>

DOI: [10.1016/j.jaad.2020.04.046](https://doi.org/10.1016/j.jaad.2020.04.046) · PMID: [32305444](https://pubmed.ncbi.nlm.nih.gov/32305444/) · PMCID: [PMC7162783](https://pubmed.ncbi.nlm.nih.gov/PMC7162783/)

1876. **FAQs for COVID-19 Claims Reimbursement to Health Care Providers and Facilities for Testing, Treatment and Vaccine Administration**

Official web site of the U.S. Health Resources & Services Administration
<https://www.hrsa.gov/coviduninsuredclaim/frequently-asked-questions>

1877. **Potential association between COVID-19 mortality and health-care resource availability**

Yunpeng Ji, Zhongren Ma, Maikel P Peppelenbosch, Qiuwei Pan
The Lancet Global Health (2020-04) <https://doi.org/ggqscd>
DOI: [10.1016/s2214-109x\(20\)30068-1](https://doi.org/s2214-109x(20)30068-1) · PMID: [32109372](#) · PMCID: [PMC7128131](#)

1878. **Combating COVID-19: health equity matters**

Zhicheng Wang, Kun Tang
Nature Medicine (2020-03-26) <https://doi.org/ggs4p6>
DOI: [10.1038/s41591-020-0823-6](https://doi.org/s41591-020-0823-6) · PMID: [32284617](#)

1879. **lockup black**

chen0307
(2020-10-14)
<https://www.census.gov/content/dam/Census/library/publications/2019/demo/p60-267.pdf>

1880. **An Ethical Dilemma in SARS-Cov-2 Pandemic : Who Gets the Ventilator?**

Dumache Raluca, Ciocan Veronica, Muresan Camelia Oana, Enache Alexandra
European Scientific Journal ESJ (2020-07-31) <https://doi.org/ghfprk>
DOI: [10.19044/esj.2020.v16n21p24](https://doi.org/10.19044/esj.2020.v16n21p24)

1881. **Planning Hospital Needs for Ventilators and Respiratory Therapists in the COVID-19 Crisis**

John Raffensperger, Marygail Brauner, R Briggs
Rand Corporation (2020) <https://doi.org/ghfprp>
DOI: [10.7249/pea228-1](https://doi.org/10.7249/pea228-1)

1882. **Fair Allocation of Vaccines, Ventilators and Antiviral Treatments: Leaving No Ethical Value Behind in Health Care Rationing**

Parag A Pathak, Tayfun Sönmez, MUTKU ÜNVER, MBUMIN YENMEZ
arXiv (2021-01-21) <https://arxiv.org/abs/2008.00374>

1883. **Reallocating ventilators during the coronavirus disease 2019 pandemic: Is it ethical?**

Quyen Chu, Ricardo Correa, Tracey L Henry, Kyle A McGregor, Hanni Stoklosa, Loren Robinson, Sachin Jha, Alagappan Annamalai, Benson S Hsu, Rohit Gupta, ... SreyRam Kuy
Surgery (2020-09) <https://doi.org/ghfprb>
DOI: [10.1016/j.surg.2020.04.044](https://doi.org/10.1016/j.surg.2020.04.044) · PMID: [32616345](#) · PMCID: [PMC7205622](#)

1884. **Ethics Lessons From Seattle's Early Experience With COVID-19**

Denise M Dudzinski, Benjamin Y Hoisington, Crystal E Brown
The American Journal of Bioethics (2020-06-18) <https://doi.org/ghfprc>

1885. **Rationing Limited Healthcare Resources in the COVID-19 Era and Beyond: Ethical Considerations Regarding Older Adults**

Timothy W Farrell, Leslie Francis, Teneille Brown, Lauren E Ferrante, Eric Widera, Ramona Rhodes, Tony Rosen, Ula Hwang, Leah J Witt, Niranjan Thothala, ... Debra Saliba

Journal of the American Geriatrics Society (2020-06-14)

<https://doi.org/ggvt7z>

DOI: [10.1111/jgs.16539](https://doi.org/10.1111/jgs.16539) · PMID: [32374466](#) · PMCID: [PMC7267288](#)

1886. **Paediatric ethical issues during the COVID-19 pandemic are not just about ventilator triage**

Marlyse F Haward, Gregory P Moore, John Lantos, Annie Janvier

Acta Paediatrica (2020-05-20) <https://doi.org/ggv24n>

DOI: [10.1111/apa.15334](https://doi.org/10.1111/apa.15334) · PMID: [32364256](#) · PMCID: [PMC7267437](#)

1887. **Ethical Challenges Arising in the COVID-19 Pandemic: An Overview from the Association of Bioethics Program Directors (ABPD) Task Force**

Amy L McGuire, Mark P Aulisio, FDaniel Davis, Cheryl Erwin, Thomas D Harter, Reshma Jaggi, Robert Klitzman, Robert Macauley, Eric Racine, Susan M Wolf, ... The COVID-19 Task Force of the Association of Bioethics Program Directors (ABPD)

The American Journal of Bioethics (2020-06-08) <https://doi.org/gg6c5k>

DOI: [10.1080/15265161.2020.1764138](https://doi.org/10.1080/15265161.2020.1764138) · PMID: [32511078](#)

1888. **Disability, Ethics, and Health Care in the COVID-19 Pandemic**

Maya Sabatello, Teresa Blankmeyer Burke, Katherine E McDonald, Paul S Appelbaum

American Journal of Public Health (2020-10) <https://doi.org/ghfprm>

DOI: [10.2105/ajph.2020.305837](https://doi.org/10.2105/ajph.2020.305837) · PMID: [32816541](#) · PMCID: [PMC7483109](#)

1889. **Allocating Ventilators During the COVID-19 Pandemic and Conscientious Objection**

Mark Wicclair

The American Journal of Bioethics (2020-07-27) <https://doi.org/gg6nk4>

DOI: [10.1080/15265161.2020.1777347](https://doi.org/10.1080/15265161.2020.1777347) · PMID: [32716798](#)

1890. **Colorblind Algorithms: Racism in the Era of COVID-19**

JCorey Williams, Nientara Anderson, Myra Mathis, Ezelle Sanford, Jeffrey Eugene, Jessica Isom

Journal of the National Medical Association (2020-10)

<https://doi.org/ghfpq8>

DOI: [10.1016/j.jnma.2020.05.010](https://doi.org/10.1016/j.jnma.2020.05.010) · PMID: [32563687](#)

1891. **Structural Racism, Social Risk Factors, and Covid-19 — A Dangerous Convergence for Black Americans**

Leonard E Egede, Rebekah J Walker

New England Journal of Medicine (2020-09-17) <https://doi.org/gg56nc>

DOI: [10.1056/nejmp2023616](https://doi.org/10.1056/nejmp2023616) · PMID: [32706952](#) · PMCID: [PMC7747672](#)

1892. **Allocating Remdesivir Under Scarcity: Social Justice or More Systemic Racism**

Eli Weber, Mark J Bliton
The American Journal of Bioethics (2020-08-25) <https://doi.org/ghfprf>
DOI: [10.1080/15265161.2020.1795538](https://doi.org/10.1080/15265161.2020.1795538) · PMID: [32840451](#)

1893. **Revisiting the equity debate in COVID-19: ICU is no panacea**

Angela Ballantyne, Wendy A Rogers, Vikki Entwistle, Cindy Towns
Journal of Medical Ethics (2020-10) <https://doi.org/gg33nq>
DOI: [10.1136/medethics-2020-106460](https://doi.org/10.1136/medethics-2020-106460) · PMID: [32571847](#) · PMCID: [PMC7335695](#)

1894. **Ethical Dilemmas in Covid-19 Medical Care: Is a Problematic Triage Protocol Better or Worse than No Protocol at All?**

Sheri Fink
The American Journal of Bioethics (2020-07-27) <https://doi.org/gg6nqn>
DOI: [10.1080/15265161.2020.1788663](https://doi.org/10.1080/15265161.2020.1788663) · PMID: [32716771](#)

1895. **Developing an Ethics Framework for Allocating Remdesivir in the COVID-19 Pandemic**

Sarah Lim, Debra A DeBruin, Jonathon P Leider, Nneka Sederstrom, Ruth Lynfield, Jason V Baker, Susan Kline, Sarah Kesler, Stacey Rizza, Joel Wu, ... Susan M Wolf
Mayo Clinic Proceedings (2020-09) <https://doi.org/ghfpq9>
DOI: [10.1016/j.mayocp.2020.06.016](https://doi.org/10.1016/j.mayocp.2020.06.016) · PMID: [32861338](#) · PMCID: [PMC7305893](#)

1896. **Ethically Allocating COVID-19 Drugs Via Pre-approval Access and Emergency Use Authorization**

Jamie Webb, Lesha D Shah, Holly Fernandez Lynch
The American Journal of Bioethics (2020-08-25) <https://doi.org/ghfprd>
DOI: [10.1080/15265161.2020.1795529](https://doi.org/10.1080/15265161.2020.1795529)

1897. (2011-07-27)

https://www.cdc.gov/about/advisory/pdf/VentDocument_Release.pdf

1898. **Adopting an Anti-Racist Model of COVID-19 Drug Allocation and Prioritization**

Akilah A Jefferson
The American Journal of Bioethics (2020-08-25) <https://doi.org/ghggz5>
DOI: [10.1080/15265161.2020.1795541](https://doi.org/10.1080/15265161.2020.1795541)

1899. **Equitably Sharing the Benefits and Burdens of Research: Covid-19 Raises the Stakes**

Carl H Coleman
Ethics & Human Research (2020-05-14) <https://doi.org/ghgg2q>
DOI: [10.1002/eahr.500055](https://doi.org/10.1002/eahr.500055) · PMID: [32410347](#) · PMCID: [PMC7272984](#)

1900. **Ensuring global access to COVID-19 vaccines**

Gavin Yamey, Marco Schäferhoff, Richard Hatchett, Muhammad Pate, Feng Zhao, Kaci Kennedy McDade
The Lancet (2020-05) <https://doi.org/ggq7mf>
DOI: [10.1016/s0140-6736\(20\)30763-7](https://doi.org/10.1016/s0140-6736(20)30763-7) · PMID: [32243778](#) · PMCID: [PMC7271264](#)

1901. **The Equitable Distribution of COVID-19 Therapeutics and Vaccines**

Thomas J Bollyky, Lawrence O Gostin, Margaret A Hamburg

1902. **Recruitment and participation in clinical trials: Socio-demographic, rural/urban, and health care access predictors**
Claudia R Baquet, Patricia Commisskey, C Daniel Mullins, Shiraz I Mishra
Cancer Detection and Prevention (2006-01) <https://doi.org/bk2k4g>
DOI: [10.1016/j.cdp.2005.12.001](https://doi.org/10.1016/j.cdp.2005.12.001) · PMID: [16495020](https://pubmed.ncbi.nlm.nih.gov/16495020/) · PMCID: [PMC3276312](https://pubmed.ncbi.nlm.nih.gov/PMC3276312/)
1903. **COVID-19 vaccine trials in Africa**
Munyaradzi Makoni
The Lancet Respiratory Medicine (2020-11) <https://doi.org/fgzk>
DOI: [10.1016/s2213-2600\(20\)30401-x](https://doi.org/10.1016/s2213-2600(20)30401-x) · PMID: [32896275](https://pubmed.ncbi.nlm.nih.gov/32896275/) · PMCID: [PMC7831818](https://pubmed.ncbi.nlm.nih.gov/PMC7831818/)
1904. **Racial/ethnic differences in clinical trial enrollment, refusal rates, ineligibility, and reasons for decline among patients at sites in the National Cancer Institute's Community Cancer Centers Program**
Aisha T Langford, Ken Resnicow, Eileen P Dimond, Andrea M Denicoff, Diane St Germain, Worta McCaskill-Stevens, Rebecca A Enos, Angela Carrigan, Kathy Wilkinson, Ronald S Go
Cancer (2014-03-15) <https://doi.org/ghgg2p>
DOI: [10.1002/cncr.28483](https://doi.org/10.1002/cncr.28483) · PMID: [24327389](https://pubmed.ncbi.nlm.nih.gov/24327389/) · PMCID: [PMC3947654](https://pubmed.ncbi.nlm.nih.gov/PMC3947654/)
1905. **Participation in Cancer Clinical Trials**
Vivek H Murthy, Harlan M Krumholz, Cary P Gross
JAMA (2004-06-09) <https://doi.org/bbh8h7>
DOI: [10.1001/jama.291.22.2720](https://doi.org/10.1001/jama.291.22.2720) · PMID: [15187053](https://pubmed.ncbi.nlm.nih.gov/15187053/)
1906. **Participation in Surgical Oncology Clinical Trials: Gender-, Race/Ethnicity-, and Age-based Disparities**
John H Stewart, Alain G Bertoni, Jennifer L Staten, Edward A Levine, Cary P Gross
Annals of Surgical Oncology (2007-08-08) <https://doi.org/cq7wqs>
DOI: [10.1245/s10434-007-9500-y](https://doi.org/10.1245/s10434-007-9500-y) · PMID: [17682824](https://pubmed.ncbi.nlm.nih.gov/17682824/)
1907. **Inclusion, Analysis, and Reporting of Sex and Race/Ethnicity in Clinical Trials: Have We Made Progress?**
Stacie E Geller, Abby Koch, Beth Pellettieri, Molly Carnes
Journal of Women's Health (2011-03) <https://doi.org/dhzxk7>
DOI: [10.1089/jwh.2010.2469](https://doi.org/10.1089/jwh.2010.2469) · PMID: [21351877](https://pubmed.ncbi.nlm.nih.gov/21351877/) · PMCID: [PMC3058895](https://pubmed.ncbi.nlm.nih.gov/PMC3058895/)
1908. **The Representation of Gender and Race/Ethnic Groups in Randomized Clinical Trials of Individuals with Systemic Lupus Erythematosus**
Titilola Falasinnu, Yashaar Chaichian, Michelle B Bass, Julia F Simard
Current Rheumatology Reports (2018-03-17) <https://doi.org/ghjmpz>
DOI: [10.1007/s11926-018-0728-2](https://doi.org/10.1007/s11926-018-0728-2) · PMID: [29550947](https://pubmed.ncbi.nlm.nih.gov/29550947/) · PMCID: [PMC5857270](https://pubmed.ncbi.nlm.nih.gov/PMC5857270/)
1909. **Racial Disproportionality in Covid Clinical Trials**
Daniel B Chastain, Sharmon P Osae, Andrés F Henao-Martínez, Carlos Franco-Paredes, Joeanna S Chastain, Henry N Young
New England Journal of Medicine (2020-08-27) <https://doi.org/gg7vcf>

1910. **Othering and Being Othered in the Context of Health Care Services**

Joy L Johnson, Joan L Bottorff, Annette J Browne, Sukhdev Grewal, BAnn Hilton, Heather Clarke

Health Communication (2004-04) <https://doi.org/cvxqm4>

DOI: [10.1207/s15327027hc1602_7](https://doi.org/10.1207/s15327027hc1602_7) · PMID: [15090288](#)

1911. **A decade of studying implicit racial/ethnic bias in healthcare providers using the implicit association test**

Ivy W Maina, Tanisha D Belton, Sara Ginzberg, Ajit Singh, Tiffani J Johnson

Social Science & Medicine (2018-02) <https://doi.org/gdfwd9>

DOI: [10.1016/j.socscimed.2017.05.009](https://doi.org/10.1016/j.socscimed.2017.05.009) · PMID: [28532892](#)

1912. **A Systematic Review of the Impact of Physician Implicit Racial Bias on Clinical Decision Making**

Erin Dehon, Nicole Weiss, Jonathan Jones, Whitney Faulconer, Elizabeth Hinton, Sarah Sterling

Academic Emergency Medicine (2017-08) <https://doi.org/gbw5pk>

DOI: [10.1111/acem.13214](https://doi.org/10.1111/acem.13214) · PMID: [28472533](#)

1913. **Aversive racism and medical interactions with Black patients: A field study**

Louis A Penner, John F Dovidio, Tessa V West, Samuel L Gaertner, Terrance L Albrecht, Rhonda K Dailey, Tsveti Markova

Journal of Experimental Social Psychology (2010-03)

<https://doi.org/dc5342>

DOI: [10.1016/j.jesp.2009.11.004](https://doi.org/10.1016/j.jesp.2009.11.004) · PMID: [20228874](#) · PMCID: [PMC2835170](#)

1914. **Intersection of Bias, Structural Racism, and Social Determinants With Health Care Inequities**

Tiffani J Johnson

Pediatrics (2020-08) <https://doi.org/ghjmqz>

DOI: [10.1542/peds.2020-003657](https://doi.org/10.1542/peds.2020-003657) · PMID: [32690807](#)

1915. **A Systematic Review Of The Food And Drug Administration's 'Exception From Informed Consent' Pathway**

William B Feldman, Spencer Phillips Hey, Aaron S Kesselheim

Health Affairs (2018-10) <https://doi.org/ghjmqw>

DOI: [10.1377/hlthaff.2018.0501](https://doi.org/10.1377/hlthaff.2018.0501) · PMID: [30273035](#)

1916. **The legacy of the tuskegee syphilis experiments for emergency exception from informed consent**

Terri A Schmidt

Annals of Emergency Medicine (2003-01) <https://doi.org/fw3kvs>

DOI: [10.1067/mem.2003.17](https://doi.org/10.1067/mem.2003.17) · PMID: [12514686](#)

1917. **CDC officials are considering a plan to distribute COVID-19 vaccines to the most vulnerable first — including people of color**

Sarah Al-Arshani

Business Insider <https://www.businessinsider.com/cdc-official-considering-giving-covid-19-vaccine-most-vulnerable-first-2020-10>

1918. **Racial Differences in T-Lymphocyte Response to Glucocorticoids**
Monica J Federico, Ronina A Covar, Eleanor E Brown, Donald YM Leung, Joseph D Spahn
Chest (2005-02) <https://doi.org/bjfcf6>
DOI: [10.1378/chest.127.2.571](https://doi.org/10.1378/chest.127.2.571) · PMID: [15705998](#)
1919. **ENDOCRINOLOGY OF THE STRESS RESPONSE**
Evangelia Charmandari, Constantine Tsigos, George Chrousos
Annual Review of Physiology (2005-03-17) <https://doi.org/brcm9n>
DOI: [10.1146/annurev.physiol.67.040403.120816](https://doi.org/10.1146/annurev.physiol.67.040403.120816) · PMID: [15709959](#)
1920. **Chronic stress, glucocorticoid receptor resistance, inflammation, and disease risk**
S Cohen, D Janicki-Deverts, WJ Doyle, GE Miller, E Frank, BS Rabin, RB Turner
Proceedings of the National Academy of Sciences (2012-04-02)
<https://doi.org/f4n6r8>
DOI: [10.1073/pnas.1118355109](https://doi.org/10.1073/pnas.1118355109) · PMID: [22474371](#) · PMCID: [PMC3341031](#)
1921. **Proliferation of Papers and Preprints During the Coronavirus Disease 2019 Pandemic: Progress or Problems With Peer Review?**
Caitlyn Vlasschaert, Joel M Topf, Swapnil Hiremath
Advances in Chronic Kidney Disease (2020-09) <https://doi.org/gg7nk4>
DOI: [10.1053/j.ackd.2020.08.003](https://doi.org/10.1053/j.ackd.2020.08.003) · PMID: [33308508](#) · PMCID: [PMC7409832](#)
1922. **How to fight an infodemic**
John Zarocostas
The Lancet (2020-02) <https://doi.org/ggpx67>
DOI: [10.1016/s0140-6736\(20\)30461-x](https://doi.org/10.1016/s0140-6736(20)30461-x) · PMID: [32113495](#) · PMCID: [PMC7133615](#)
1923. **CORD-19: The COVID-19 Open Research Dataset**
Lucy Lu Wang, Kyle Lo, Yoganand Chandrasekhar, Russell Reas, Jiangjiang Yang, Doug Burdick, Darrin Eide, Kathryn Funk, Yannis Katsis, Rodney Kinney, ... Sebastian Kohlmeier
arXiv (2020-07-14) <https://arxiv.org/abs/2004.10706>
1924. **Analyzing the vast coronavirus literature with CoronaCentral**
Jake Lever, Russ B Altman
Proceedings of the National Academy of Sciences (2021-05-20)
<https://doi.org/gj9qfh>
DOI: [10.1073/pnas.2100766118](https://doi.org/10.1073/pnas.2100766118) · PMID: [34016708](#) · PMCID: [PMC8202008](#)
1925. **The impact of preprint servers and electronic publishing on biomedical research**
Gunther Eysenbach
Current Opinion in Immunology (2000-10) <https://doi.org/d3bmnv>
DOI: [10.1016/s0952-7915\(00\)00127-8](https://doi.org/10.1016/s0952-7915(00)00127-8)
1926. **An alarming retraction rate for scientific publications on Coronavirus Disease 2019 (COVID-19)**
Nicole Shu Ling Yeo-Teh, Bor Luen Tang

1927. **An “alarming” and “exceptionally high” rate of COVID-19 retractions?**

Alison Abritis, Adam Marcus, Ivan Oransky

Accountability in Research (2020-07-11) <https://doi.org/gg4f67>

DOI: [10.1080/08989621.2020.1793675](https://doi.org/10.1080/08989621.2020.1793675) · PMID: [32634321](#)

1928. **Queries on the COVID-19 quick publishing ethics**

Govindasamy Agoramoorthy, Minna J Hsu, Pochuen Shieh

Bioethics (2020-06) <https://doi.org/gjtm8x>

DOI: [10.1111/bioe.12772](https://doi.org/10.1111/bioe.12772) · PMID: [32433777](#) · PMCID: [PMC7276831](#)

1929. **Idle medical students review emerging COVID-19 research**

Carl Boodman, Santina Lee, Jared Bullard

Medical Education Online (2020-01-01) <https://doi.org/gjtm8v>

DOI: [10.1080/10872981.2020.1770562](https://doi.org/10.1080/10872981.2020.1770562) · PMID: [32441229](#) · PMCID: [PMC7448910](#)

1930. **Scientists are drowning in COVID-19 papers. Can new tools keep them afloat?**

Jeffrey Brainard

Science (2020-05-13) <https://doi.org/gg924n>

DOI: [10.1126/science.abc7839](https://doi.org/10.1126/science.abc7839)

1931. **Advancing scientific knowledge in times of pandemics**

Nicolas Vabret, Robert Samstein, Nicolas Fernandez, Miriam Merad,
The Sinai Immunology Review Project, Trainees, Faculty

Nature Reviews Immunology (2020-04-23) <https://doi.org/ghjs79>

DOI: [10.1038/s41577-020-0319-0](https://doi.org/10.1038/s41577-020-0319-0) · PMID: [32327718](#) · PMCID: [PMC7187143](#)

1932. **COVID-19: Epidemiology, Evolution, and Cross-Disciplinary Perspectives**

Jiumeng Sun, Wan-Ting He, Lifang Wang, Alexander Lai, Xiang Ji,
Xiaofeng Zhai, Gairu Li, Marc A Suchard, Jin Tian, Jiyong Zhou, ... Shuo
Su

Trends in Molecular Medicine (2020-05) <https://doi.org/ggqgwq>

DOI: [10.1016/j.molmed.2020.02.008](https://doi.org/10.1016/j.molmed.2020.02.008) · PMID: [32359479](#) · PMCID: [PMC7118693](#)

1933. **COVID-19 diagnostics in context**

Ralph Weissleder, Hakho Lee, Jina Ko, Mikael J Pittet

Science Translational Medicine (2020-06-03) <https://doi.org/gg339m>

DOI: [10.1126/scitranslmed.abc1931](https://doi.org/10.1126/scitranslmed.abc1931) · PMID: [32493791](#)

1934. **Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19)**

James M Sanders, Marguerite L Monogue, Tomasz Z Jodlowski, James B
Cutrell

JAMA (2020-04-13) <https://doi.org/ggr27x>

DOI: [10.1001/jama.2020.6019](https://doi.org/10.1001/jama.2020.6019) · PMID: [32282022](#)

1935. **COVID-19 Research in Brief: December, 2019 to June, 2020**

Thiago Carvalho

Nature Medicine (2020-06-26) <https://doi.org/gg3kd2>

DOI: [10.1038/d41591-020-00026-w](https://doi.org/10.1038/d41591-020-00026-w) · PMID: [32778824](#)

1936. **Pathophysiology, Transmission, Diagnosis, and Treatment of Coronavirus Disease 2019 (COVID-19)**

WJoost Wiersinga, Andrew Rhodes, Allen C Cheng, Sharon J Peacock, Hallie C Prescott

JAMA (2020-08-25) <https://doi.org/gg4ht4>

DOI: [10.1001/jama.2020.12839](https://doi.org/10.1001/jama.2020.12839) · PMID: [32648899](#)

1937. **Open collaborative writing with Manubot**

Daniel S Himmelstein, Vincent Rubinetti, David R Slochower, Dongbo Hu, Venkat S Malladi, Casey S Greene, Anthony Gitter

PLOS Computational Biology (2019-06-24) <https://doi.org/c7np>

DOI: [10.1371/journal.pcbi.1007128](https://doi.org/journal.pcbi.1007128) · PMID: [31233491](#) · PMCID: [PMC6611653](#)

1938. **Introducing Massively Open Online Papers (MOOPs)**

Jonathan P Tennant, Natalia Bielczyk, Bastian Greshake Tzovaras, Paola Masuzzo, Tobias Steiner

KULA: Knowledge Creation, Dissemination, and Preservation Studies (2020-04-20) <https://doi.org/gg89rt>

DOI: [10.5334/kula.63](https://doi.org/10.5334/kula.63)

1939. **How you can help with COVID-19 modelling**

Julia R Gog

Nature Reviews Physics (2020-04-08) <https://doi.org/ggrsq3>

DOI: [10.1038/s42254-020-0175-7](https://doi.org/10.1038/s42254-020-0175-7) · PMCID: [PMC7144181](#)

1940. **Advancing Open Science with Version Control and Blockchains**

Jonathan Bell, Thomas D LaToza, Foteini Baldmitzi, Angelos Stavrou

2017 IEEE/ACM 12th International Workshop on Software Engineering for Science (SE4Science) (2017-05) <https://doi.org/gjtt3z>

DOI: [10.1109/se4science.2017.11](https://doi.org/10.1109/se4science.2017.11)

1941. **Curating Research Assets: A Tutorial on the Git Version Control System**

Matti Vuorre, James P Curley

Advances in Methods and Practices in Psychological Science (2018-04-11) <https://doi.org/gdj7ch>

DOI: [10.1177/2515245918754826](https://doi.org/10.1177/2515245918754826)

1942. **Git can facilitate greater reproducibility and increased transparency in science**

Karthik Ram

Source Code for Biology and Medicine (2013-02-28) <https://doi.org/krv>

DOI: [10.1186/1751-0473-8-7](https://doi.org/10.1186/1751-0473-8-7) · PMID: [23448176](#) · PMCID: [PMC3639880](#)

1943. **Using the MAARIE Framework To Read the Research Literature**

M Corcoran

American Journal of Occupational Therapy (2006-07-01)

<https://doi.org/bqh97x>

DOI: [10.5014/ajot.60.4.367](https://doi.org/10.5014/ajot.60.4.367) · PMID: [16915865](#)

1944. **Studying a study & testing a test: reading evidence-based health research**

Richard K Riegelman

Wolters Kluwer/Lippincott Williams & Wilkins Heath (2013)

ISBN: 9780781774260

1945. **Matplotlib: A 2D Graphics Environment**

John D Hunter

Computing in Science & Engineering (2007) <https://doi.org/drjhg>

DOI: [10.1109/mcse.2007.55](https://doi.org/mcse.2007.55)

1946. **scite: a smart citation index that displays the context of citations and classifies their intent using deep learning**

JM Nicholson, M Mordaunt, P Lopez, A Uppala, D Rosati, NP Rodrigues, P Grabitz, SC Rife

Cold Spring Harbor Laboratory (2021-03-16) <https://doi.org/gjt36w>

DOI: [10.1101/2021.03.15.435418](https://doi.org/2021.03.15.435418)

1947. **Synchronized editing: the future of collaborative writing**

Jeffrey M Perkel

Nature (2020-03-31) <https://doi.org/ggqk8s>

DOI: [10.1038/d41586-020-00916-6](https://doi.org/d41586-020-00916-6) · PMID: [32235940](#)

1948. **Coronavirus pandemic (COVID-19)**

Max Roser, Hannah Ritchie, Esteban Ortiz-Ospina, Joe Hasell

Our World in Data (2020) <https://ourworldindata.org/coronavirus>

1949. **Linguistic Analysis of the bioRxiv Preprint Landscape**

David N Nicholson, Vincent Rubinetti, Dongbo Hu, Marvin Thielk, Lawrence E Hunter, Casey S Greene

Cold Spring Harbor Laboratory (2021-03-04) <https://doi.org/gh7rph>

DOI: [10.1101/2021.03.04.433874](https://doi.org/2021.03.04.433874)

1950. **Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody**

Xiaolong Tian, Cheng Li, Ailing Huang, Shuai Xia, Sicong Lu, Zhengli Shi, Lu Lu, Shibo Jiang, Zhenlin Yang, Yanling Wu, Tianlei Ying

Cold Spring Harbor Laboratory (2020-01-28) <https://doi.org/ggjqfd>

DOI: [10.1101/2020.01.28.923011](https://doi.org/2020.01.28.923011)

1951. **Integrative Bioinformatics Analysis Provides Insight into the Molecular Mechanisms of 2019-nCoV**

Xiang He, Lei Zhang, Qin Ran, Junyi Wang, Anying Xiong, Dehong Wu, Feng Chen, Guoping Li

Cold Spring Harbor Laboratory (2020-02-05) <https://doi.org/ggrbd8>

DOI: [10.1101/2020.02.03.20020206](https://doi.org/2020.02.03.20020206)

1952. **Diarrhea may be underestimated: a missing link in 2019 novel coronavirus**

Weicheng Liang, Zhijie Feng, Shitao Rao, Cuicui Xiao, Ze-Xiao Lin, Qi Zhang, Qi Wei

Cold Spring Harbor Laboratory (2020-02-17) <https://doi.org/ggrbdw>

DOI: [10.1101/2020.02.03.20020289](https://doi.org/2020.02.03.20020289)

1953. **Specific ACE2 Expression in Cholangiocytes May Cause Liver Damage After 2019-nCoV Infection**

Xiaoqiang Chai, Longfei Hu, Yan Zhang, Weiyu Han, Zhou Lu, Aiwu Ke, Jian Zhou, Guoming Shi, Nan Fang, Jia Fan, ... Fei Lan
Cold Spring Harbor Laboratory (2020-02-04) <https://doi.org/ggq626>
DOI: [10.1101/2020.02.03.931766](https://doi.org/10.1101/2020.02.03.931766)

1954. **Recapitulation of SARS-CoV-2 Infection and Cholangiocyte Damage with Human Liver Organoids**

Bing Zhao, Chao Ni, Ran Gao, Yuyan Wang, Li Yang, Jinsong Wei, Ting Lv, Jianqing Liang, Qisheng Zhang, Wei Xu, ... Xinhua Lin
Cold Spring Harbor Laboratory (2020-03-17) <https://doi.org/ggq648>
DOI: [10.1101/2020.03.16.990317](https://doi.org/10.1101/2020.03.16.990317)

1955. **ACE2 expression by colonic epithelial cells is associated with viral infection, immunity and energy metabolism**

Jun Wang, Shanmeizi Zhao, Ming Liu, Zhiyao Zhao, Yiping Xu, Ping Wang, Meng Lin, Yanhui Xu, Bing Huang, Xiaoyu Zuo, ... Yuxia Zhang
Cold Spring Harbor Laboratory (2020-02-07) <https://doi.org/ggrfbx>
DOI: [10.1101/2020.02.05.20020545](https://doi.org/10.1101/2020.02.05.20020545)

1956. **The Pathogenicity of SARS-CoV-2 in hACE2 Transgenic Mice**

Linlin Bao, Wei Deng, Baoying Huang, Hong Gao, Jiangning Liu, Lili Ren, Qiang Wei, Pin Yu, Yanfeng Xu, Feifei Qi, ... Chuan Qin
Cold Spring Harbor Laboratory (2020-02-28) <https://doi.org/dph2>
DOI: [10.1101/2020.02.07.939389](https://doi.org/10.1101/2020.02.07.939389)

1957. **Caution on Kidney Dysfunctions of COVID-19 Patients**

Zhen Li, Ming Wu, Jiwei Yao, Jie Guo, Xiang Liao, Siji Song, Jiali Li, Guangjie Duan, Yuanxiu Zhou, Xiaojun Wu, ... Junan Yan
Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggq627>
DOI: [10.1101/2020.02.08.20021212](https://doi.org/10.1101/2020.02.08.20021212)

1958. **Acute renal impairment in coronavirus-associated severe acute respiratory syndrome**

Kwok Hong Chu, Wai Kay Tsang, Colin S Tang, Man Fai Lam, Fernand M Lai, Ka Fai To, Ka Shun Fung, Hon Lok Tang, Wing Wa Yan, Hilda WH Chan, ... Kar Neng Lai
Kidney International (2005-02) <https://doi.org/b7tgtx>
DOI: [10.1111/j.1523-1755.2005.67130.x](https://doi.org/10.1111/j.1523-1755.2005.67130.x) · PMID: [15673319](#) · PMCID: [PMC7112337](#)

1959. **Single-cell Analysis of ACE2 Expression in Human Kidneys and Bladders Reveals a Potential Route of 2019-nCoV Infection**

Wei Lin, Longfei Hu, Yan Zhang, Joshua D Ooi, Ting Meng, Peng Jin, Xiang Ding, Longkai Peng, Lei Song, Zhou Xiao, ... Yong Zhong
Cold Spring Harbor Laboratory (2020-02-18) <https://doi.org/ggq629>
DOI: [10.1101/2020.02.08.939892](https://doi.org/10.1101/2020.02.08.939892)

1960. **The immune vulnerability landscape of the 2019 Novel Coronavirus, SARS-CoV-2**

James Zhu, Jiwoong Kim, Xue Xiao, Yunguan Wang, Danni Luo, Shuang Jiang, Ran Chen, Lin Xu, He Zhang, Lenny Moise, ... Yang Xie
Cold Spring Harbor Laboratory (2020-09-04) <https://doi.org/ggq628>
DOI: [10.1101/2020.02.08.939553](https://doi.org/10.1101/2020.02.08.939553) · PMID: [32908981](#)

1961. **Clinical Course and Outcomes of Critically Ill Patients With Middle East Respiratory Syndrome Coronavirus Infection**

Yaseen M Arabi, Ahmed A Arifi, Hanan H Balkhy, Hani Najm, Abdulaziz S Aldawood, Alaa Ghabashi, Hassan Hawa, Adel Alothman, Abdulaziz Khaldi, Basel Al Riy

Annals of Internal Medicine (2014-03-18) <https://doi.org/ggptxw>

DOI: [10.7326/m13-2486](https://doi.org/10.7326/m13-2486) · PMID: [24474051](#)

1962. **Neutrophil-to-Lymphocyte Ratio Predicts Severe Illness Patients with 2019 Novel Coronavirus in the Early Stage**

Jingyuan Liu, Yao Liu, Pan Xiang, Lin Pu, Haofeng Xiong, Chuansheng Li, Ming Zhang, Jianbo Tan, Yanli Xu, Rui Song, ... Xianbo Wang

Cold Spring Harbor Laboratory (2020-02-12) <https://doi.org/ggrbdx>

DOI: [10.1101/2020.02.10.20021584](https://doi.org/10.1101/2020.02.10.20021584)

1963. **Dysregulation of Immune Response in Patients With Coronavirus 2019 (COVID-19) in Wuhan, China**

Chuan Qin, Luoqi Zhou, Ziwei Hu, Shuoqi Zhang, Sheng Yang, Yu Tao, Cuihong Xie, Ke Ma, Ke Shang, Wei Wang, Dai-Shi Tian

Clinical Infectious Diseases (2020-08-01) <https://doi.org/ggpvcf>

DOI: [10.1093/cid/ciaa248](https://doi.org/10.1093/cid/ciaa248) · PMID: [32161940](#) · PMCID: [PMC7108125](#)

1964. **Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP)**

Suxin Wan, Qingjie Yi, Shibing Fan, Jinglong Lv, Xianxiang Zhang, Lian Guo, Chunhui Lang, Qing Xiao, Kaihu Xiao, Zhengjun Yi, ... Yongping Chen

Cold Spring Harbor Laboratory (2020-02-12) <https://doi.org/ggq63b>

DOI: [10.1101/2020.02.10.20021832](https://doi.org/10.1101/2020.02.10.20021832)

1965. **Longitudinal Characteristics of Lymphocyte Responses and Cytokine Profiles in the Peripheral Blood of SARS-CoV-2 Infected Patients**

Jing Liu, Sumeng Li, Jia Liu, Boyun Liang, Xiaobei Wang, Wei Li, Hua Wang, Qiaoxia Tong, Jianhua Yi, Lei Zhao, ... Xin Zheng

SSRN Electronic Journal (2020) <https://doi.org/ggq655>

DOI: [10.2139/ssrn.3539682](https://doi.org/10.2139/ssrn.3539682)

1966. **Epidemiological and Clinical Characteristics of 17 Hospitalized Patients with 2019 Novel Coronavirus Infections Outside Wuhan, China**

Jie Li, Shilin Li, Yurui Cai, Qin Liu, Xue Li, Zhaoping Zeng, Yanpeng Chu, Fangcheng Zhu, Fanxin Zeng

Cold Spring Harbor Laboratory (2020-02-12) <https://doi.org/ggq63c>

DOI: [10.1101/2020.02.11.20022053](https://doi.org/10.1101/2020.02.11.20022053)

1967. **ACE2 Expression in Kidney and Testis May Cause Kidney and Testis Damage After 2019-nCoV Infection**

Caibin Fan, Kai Li, Yanhong Ding, Wei Lu, Jianqing Wang

Cold Spring Harbor Laboratory (2020-02-13) <https://doi.org/ggq63d>

DOI: [10.1101/2020.02.12.20022418](https://doi.org/10.1101/2020.02.12.20022418)

1968. **Aberrant pathogenic GM-CSF ⁺ T cells and inflammatory CD14 ⁺ CD16 ⁺ monocytes**

in severe pulmonary syndrome patients of a new coronavirus

Yonggang Zhou, Binqing Fu, Xiaohu Zheng, Dongsheng Wang, Changcheng Zhao, Yingjie qi, Rui Sun, Zhigang Tian, Xiaoling Xu, Haiming Wei

Cold Spring Harbor Laboratory (2020-02-20) <https://doi.org/ggq63f>

DOI: [10.1101/2020.02.12.945576](https://doi.org/10.1101/2020.02.12.945576)

1969. Clinical Characteristics of 2019 Novel Infected Coronavirus Pneumonia: A Systemic Review and Meta-analysis

Kai Qian, Yi Deng, Yong-Hang Tai, Jun Peng, Hao Peng, Li-Hong Jiang

Cold Spring Harbor Laboratory (2020-02-17) <https://doi.org/ggrgbq>

DOI: [10.1101/2020.02.14.20021535](https://doi.org/10.1101/2020.02.14.20021535)

1970. Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of SARS-CoV-2 infected patients

Jing Liu, Sumeng Li, Jia Liu, Boyun Liang, Xiaobei Wang, Hua Wang, Wei Li, Qiaoxia Tong, Jianhua Yi, Lei Zhao, ... Xin Zheng

Cold Spring Harbor Laboratory (2020-02-22) <https://doi.org/ggq63g>

DOI: [10.1101/2020.02.16.20023671](https://doi.org/10.1101/2020.02.16.20023671)

1971. Clinical and immunologic features in severe and moderate forms of Coronavirus Disease 2019

Guang Chen, Di Wu, Wei Guo, Yong Cao, Da Huang, Hongwu Wang, Tao Wang, Xiaoyun Zhang, Huilong Chen, Haijing Yu, ... Qin Ning

Cold Spring Harbor Laboratory (2020-02-19) <https://doi.org/ggq63h>

DOI: [10.1101/2020.02.16.20023903](https://doi.org/10.1101/2020.02.16.20023903)

1972. Protection of Rhesus Macaque from SARS-Coronavirus challenge by recombinant adenovirus vaccine

Yiyou Chen, Qiang Wei, Ruobing Li, Hong Gao, Hua Zhu, Wei Deng, Linlin Bao, Wei Tong, Zhe Cong, Hong Jiang, Chuan Qin

Cold Spring Harbor Laboratory (2020-02-21) <https://doi.org/ggq63k>

DOI: [10.1101/2020.02.17.951939](https://doi.org/10.1101/2020.02.17.951939)

1973. Reduction and Functional Exhaustion of T Cells in Patients with Coronavirus Disease 2019 (COVID-19)

Bo Diao, Chenhui Wang, Yingjun Tan, Xiewan Chen, Ying Liu, Lifen Ning, Li Chen, Min Li, Yueping Liu, Gang Wang, ... Yongwen Chen

Cold Spring Harbor Laboratory (2020-02-20) <https://doi.org/ggq63m>

DOI: [10.1101/2020.02.18.20024364](https://doi.org/10.1101/2020.02.18.20024364)

1974. Clinical characteristics of 25 death cases with COVID-19: a retrospective review of medical records in a single medical center, Wuhan, China

Xun Li, Luwen Wang, Shaonan Yan, Fan Yang, Longkui Xiang, Jiling Zhu, Bo Shen, Zuojiong Gong

Cold Spring Harbor Laboratory (2020-02-25) <https://doi.org/ggq63n>

DOI: [10.1101/2020.02.19.20025239](https://doi.org/10.1101/2020.02.19.20025239)

1975. SARS-CoV-2 infection does not significantly cause acute renal injury: an analysis of 116 hospitalized patients with COVID-19 in a single hospital, Wuhan, China

Luwen Wang, Xun Li, Hui Chen, Shaonan Yan, Yan Li, Dong Li, Zuojiong Gong

1976. **Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China**
Dawei Wang, Bo Hu, Chang Hu, Fangfang Zhu, Xing Liu, Jing Zhang, Binbin Wang, Hui Xiang, Zhenshun Cheng, Yong Xiong, ... Zhiyong Peng
JAMA (2020-03-17) <https://doi.org/ggkh48>
DOI: [10.1001/jama.2020.1585](https://doi.org/10.1001/jama.2020.1585) · PMID: [32031570](https://pubmed.ncbi.nlm.nih.gov/32031570/) · PMCID: [PMC7042881](https://pubmed.ncbi.nlm.nih.gov/PMC7042881/)
1977. **Clinical characteristics of 2019 novel coronavirus infection in China**
Wei-jie Guan, Zheng-yi Ni, Yu Hu, Wen-hua Liang, Chun-quan Ou, Jian-xing He, Lei Liu, Hong Shan, Chun-liang Lei, David SC Hui, ... Nan-shan Zhong
Cold Spring Harbor Laboratory (2020-02-09) <https://doi.org/ggkj9s>
DOI: [10.1101/2020.02.06.20020974](https://doi.org/10.1101/2020.02.06.20020974)
1978. **Potential T-cell and B-cell Epitopes of 2019-nCoV**
Ethan Fast, Russ B Altman, Binbin Chen
Cold Spring Harbor Laboratory (2020-03-18) <https://doi.org/ggq63q>
DOI: [10.1101/2020.02.19.955484](https://doi.org/10.1101/2020.02.19.955484)
1979. **Structure, function and antigenicity of the SARS-CoV-2 spike glycoprotein**
Alexandra C Walls, Young-Jun Park, MAlexandra Tortorici, Abigail Wall, Andrew T McGuire, David Veesler
Cold Spring Harbor Laboratory (2020-02-20) <https://doi.org/ggrgb>
DOI: [10.1101/2020.02.19.956581](https://doi.org/10.1101/2020.02.19.956581)
1980. **Breadth of concomitant immune responses underpinning viral clearance and patient recovery in a non-severe case of COVID-19**
Irani Thevarajan, Thi HO Nguyen, Marios Koutsakos, Julian Druce, Leon Caly, Carolien E van de Sandt, Xiaoxiao Jia, Suellen Nicholson, Mike Catton, Benjamin Cowie, ... Katherine Kedzierska
Cold Spring Harbor Laboratory (2020-02-23) <https://doi.org/ggq63r>
DOI: [10.1101/2020.02.20.20025841](https://doi.org/10.1101/2020.02.20.20025841)
1981. **The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing**
Minfeng Liao, Yang Liu, Jin Yuan, Yanling Wen, Gang Xu, Juanjuan Zhao, Lin Chen, Jinxiu Li, Xin Wang, Fuxiang Wang, ... Zheng Zhang
Cold Spring Harbor Laboratory (2020-02-26) <https://doi.org/ggq63s>
DOI: [10.1101/2020.02.23.20026690](https://doi.org/10.1101/2020.02.23.20026690)
1982. **Influenza A Virus Infection Induces Hyperresponsiveness in Human Lung Tissue-Resident and Peripheral Blood NK Cells**
Marlena Scharenberg, Sindhu Vangeti, Eliisa Kekäläinen, Per Bergman, Mamdoh Al-Ameri, Niclas Johansson, Klara Sondén, Sara Falck-Jones, Anna Färnert, Hans-Gustaf Ljunggren, ... Nicole Marquardt
Frontiers in Immunology (2019-05-17) <https://doi.org/ggq656>
DOI: [10.3389/fimmu.2019.01116](https://doi.org/10.3389/fimmu.2019.01116) · PMID: [31156653](https://pubmed.ncbi.nlm.nih.gov/31156653/) · PMCID: [PMC6534051](https://pubmed.ncbi.nlm.nih.gov/PMC6534051/)

1983. **A serological assay to detect SARS-CoV-2 seroconversion in humans**

Fatima Amanat, Daniel Stadlbauer, Shirin Strohmeier, Thi HO Nguyen, Veronika Chromikova, Meagan McMahon, Kaijun Jiang, Guha Asthagiri Arunkumar, Denise Jurczyszak, Jose Polanco, ... Florian Krammer
Cold Spring Harbor Laboratory (2020-04-16) <https://doi.org/ggpn83>
DOI: [10.1101/2020.03.17.20037713](https://doi.org/10.1101/2020.03.17.20037713) · PMID: [32511441](#)

1984. **Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China**

Chaolin Huang, Yeming Wang, Xingwang Li, Lili Ren, Jianping Zhao, Yi Hu, Li Zhang, Guohui Fan, Jiuyang Xu, Xiaoying Gu, ... Bin Cao
The Lancet (2020-02) <https://doi.org/ggjfnn>
DOI: [10.1016/s0140-6736\(20\)30183-5](https://doi.org/10.1016/s0140-6736(20)30183-5)

1985. **Alveolar Macrophages in the Resolution of Inflammation, Tissue Repair, and Tolerance to Infection**

Benoit Allard, Alice Panariti, James G Martin
Frontiers in Immunology (2018-07-31) <https://doi.org/gd3bnz>
DOI: [10.3389/fimmu.2018.01777](https://doi.org/10.3389/fimmu.2018.01777) · PMID: [30108592](#) · PMCID: [PMC6079255](#)

1986. **PPAR-γ in Macrophages Limits Pulmonary Inflammation and Promotes Host Recovery following Respiratory Viral Infection**

Su Huang, Bibo Zhu, In Su Cheon, Nick P Goplen, Li Jiang, Ruixuan Zhang, R Stokes Peebles, Matthias Mack, Mark H Kaplan, Andrew H Limper, Jie Sun
Journal of Virology (2019-04-17) <https://doi.org/ggg652>
DOI: [10.1128/jvi.00030-19](https://doi.org/10.1128/jvi.00030-19) · PMID: [30787149](#) · PMCID: [PMC6475778](#)

1987. **Can routine laboratory tests discriminate 2019 novel coronavirus infected pneumonia from other community-acquired pneumonia?**

Yunbao Pan, Guangming Ye, Xiantao Zeng, Guohong Liu, Xiaoqiao Zeng, Xianghu Jiang, Jin Zhao, Liangjun Chen, Shuang Guo, Qiaoling Deng, ... Xinghuan Wang
Cold Spring Harbor Laboratory (2020-02-25) <https://doi.org/ggg63t>
DOI: [10.1101/2020.02.25.20024711](https://doi.org/10.1101/2020.02.25.20024711)

1988. **Correlation Analysis Between Disease Severity and Inflammation-related Parameters in Patients with COVID-19 Pneumonia**

Jing Gong, Hui Dong, Qingsong Xia, Zhaoyi Huang, Dingkun Wang, Yan Zhao, Wenhua Liu, Shenghao Tu, Mingmin Zhang, Qi Wang, Fuer Lu
Cold Spring Harbor Laboratory (2020-02-26) <https://doi.org/ggq63v>
DOI: [10.1101/2020.02.25.20025643](https://doi.org/10.1101/2020.02.25.20025643)

1989. **An Effective CTL Peptide Vaccine for Ebola Zaire Based on Survivors' CD8+ Targeting of a Particular Nucleocapsid Protein Epitope with Potential Implications for COVID-19 Vaccine Design**

CV Herst, S Burkholz, J Sidney, A Sette, PE Harris, S Massey, T Brasel, E Cunha-Neto, DS Rosa, WCH Chao, ... R Rubsamen
Cold Spring Harbor Laboratory (2020-04-06) <https://doi.org/ggq63x>
DOI: [10.1101/2020.02.25.963546](https://doi.org/10.1101/2020.02.25.963546)

1990. **Epitope-based peptide vaccine design and target site characterization against novel coronavirus disease caused by**

SARS-CoV-2

Lin Li, Ting Sun, Yufei He, Wendong Li, Yubo Fan, Jing Zhang
Cold Spring Harbor Laboratory (2020-02-27) <https://doi.org/ggnqwt>
DOI: [10.1101/2020.02.25.965434](https://doi.org/10.1101/2020.02.25.965434)

1991. **The definition and risks of Cytokine Release Syndrome-Like in 11 COVID-19-Infected Pneumonia critically ill patients: Disease Characteristics and Retrospective Analysis**

Wang Wenjun, Liu Xiaoqing, Wu Sipei, Lie Puyi, Huang Liyan, Li Yimin, Cheng Linling, Chen Sibei, Nong Lingbo, Lin Yongping, He Jianxing
Cold Spring Harbor Laboratory (2020-02-27) <https://doi.org/ggrgbs>
DOI: [10.1101/2020.02.26.20026989](https://doi.org/10.1101/2020.02.26.20026989)

1992. **Clinical characteristics of 36 non-survivors with COVID-19 in Wuhan, China**

Ying Huang, Rui Yang, Ying Xu, Ping Gong
Cold Spring Harbor Laboratory (2020-03-05) <https://doi.org/gqq63z>
DOI: [10.1101/2020.02.27.20029009](https://doi.org/10.1101/2020.02.27.20029009)

1993. **Risk factors related to hepatic injury in patients with corona virus disease 2019**

Lu Li, Shuang Li, Manman Xu, Pengfei Yu, Sujun Zheng, Zhongping Duan, Jing Liu, Yu Chen, Junfeng Li
Cold Spring Harbor Laboratory (2020-03-10) <https://doi.org/gqq632>
DOI: [10.1101/2020.02.28.20028514](https://doi.org/10.1101/2020.02.28.20028514)

1994. **Detectable serum SARS-CoV-2 viral load (RNAemia) is closely associated with drastically elevated interleukin 6 (IL-6) level in critically ill COVID-19 patients**

Xiaohua Chen, Binghong Zhao, Yueming Qu, Yurou Chen, Jie Xiong, Yong Feng, Dong Men, Qianchuan Huang, Ying Liu, Bo Yang, ... Feng Li
Cold Spring Harbor Laboratory (2020-03-03) <https://doi.org/gqq633>
DOI: [10.1101/2020.02.29.20029520](https://doi.org/10.1101/2020.02.29.20029520)

1995. **Prognostic factors in the acute respiratory distress syndrome**

Wei Chen, Lorraine B Ware
Clinical and Translational Medicine (2015-07-02) <https://doi.org/gqq653>
DOI: [10.1186/s40169-015-0065-2](https://doi.org/10.1186/s40169-015-0065-2) · PMID: [26162279](#) · PMCID: [PMC4534483](#)

1996. **Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study**

Li Tan, Qi Wang, Duanyang Zhang, Jinya Ding, Qianchuan Huang, Yi-Quan Tang, Qiongshu Wang, Hongming Miao
Cold Spring Harbor Laboratory (2020-03-03) <https://doi.org/gqq634>
DOI: [10.1101/2020.03.01.20029074](https://doi.org/10.1101/2020.03.01.20029074)

1997. **The potential role of IL-6 in monitoring severe case of coronavirus disease 2019**

Tao Liu, Jieying Zhang, Yuhui Yang, Hong Ma, Zhengyu Li, Jiaoyu Zhang, Ji Cheng, Xiaoyun Zhang, Yanxia Zhao, Zihan Xia, ... Jianhua Yi
Cold Spring Harbor Laboratory (2020-03-10) <https://doi.org/gqq635>
DOI: [10.1101/2020.03.01.20029769](https://doi.org/10.1101/2020.03.01.20029769)

1998. **Clinical and Laboratory Profiles of 75 Hospitalized Patients with Novel Coronavirus Disease 2019 in Hefei, China**
Zonghao Zhao, Jiajia Xie, Ming Yin, Yun Yang, Hongliang He, Tengchuan Jin, Wenting Li, Xiaowu Zhu, Jing Xu, Changcheng Zhao, ... Xiaoling Ma
Cold Spring Harbor Laboratory (2020-03-06) <https://doi.org/ggq636>
DOI: [10.1101/2020.03.01.20029785](https://doi.org/10.1101/2020.03.01.20029785)
1999. **Exuberant elevation of IP-10, MCP-3 and IL-1ra during SARS-CoV-2 infection is associated with disease severity and fatal outcome**
Yang Yang, Chenguang Shen, Jinxiu Li, Jing Yuan, Minghui Yang, Fuxiang Wang, Guobao Li, Yanjie Li, Li Xing, Ling Peng, ... Yingxia Liu
Cold Spring Harbor Laboratory (2020-03-06) <https://doi.org/ggq637>
DOI: [10.1101/2020.03.02.20029975](https://doi.org/10.1101/2020.03.02.20029975)
2000. **Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019**
Juanjuan Zhao, Quan Yuan, Haiyan Wang, Wei Liu, Xuejiao Liao, Yingying Su, Xin Wang, Jing Yuan, Tingdong Li, Jinxiu Li, ... Zheng Zhang
Cold Spring Harbor Laboratory (2020-03-02) <https://doi.org/ggrbj6>
DOI: [10.1101/2020.03.02.20030189](https://doi.org/10.1101/2020.03.02.20030189)
2001. **Restoration of leukomonocyte counts is associated with viral clearance in COVID-19 hospitalized patients**
Xiaoping Chen, Jiaxin Ling, Pingzheng Mo, Yongxi Zhang, Qunqun Jiang, Zhiyong Ma, Qian Cao, Wenjia Hu, Shi Zou, Liangjun Chen, ... Yong Xiong
Cold Spring Harbor Laboratory (2020-03-06) <https://doi.org/ggq639>
DOI: [10.1101/2020.03.03.20030437](https://doi.org/10.1101/2020.03.03.20030437)
2002. **Effects of Systemically Administered Hydrocortisone on the Human Immunome**
Matthew J Olnes, Yuri Kotliarov, Angélique Bianotto, Foo Cheung, Jinguo Chen, Rongye Shi, Huizhi Zhou, Ena Wang, John S Tsang, Robert Nussenblatt, The CHI Consortium
Scientific Reports (2016-03-14) <https://doi.org/f8dmvw>
DOI: [10.1038/srep23002](https://doi.org/10.1038/srep23002) · PMID: [26972611](https://pubmed.ncbi.nlm.nih.gov/26972611/) · PMCID: [PMC4789739](https://pmcid.ncbi.nlm.nih.gov/pmc/articles/PMC4789739/)
2003. **Procalcitonin in patients with severe coronavirus disease 2019 (COVID-19): A meta-analysis**
Giuseppe Lippi, Mario Plebani
Clinica Chimica Acta (2020-06) <https://doi.org/ggpwp7>
DOI: [10.1016/j.cca.2020.03.004](https://doi.org/10.1016/j.cca.2020.03.004) · PMID: [32145275](https://pubmed.ncbi.nlm.nih.gov/32145275/) · PMCID: [PMC7094472](https://pmcid.ncbi.nlm.nih.gov/pmc/articles/PMC7094472/)
2004. **Clinical findings in critically ill patients infected with SARS-CoV-2 in Guangdong Province, China: a multi-center, retrospective, observational study**
Yonghao Xu, Zhiheng Xu, Xuesong Liu, Lihua Cai, Haichong Zheng, Yongbo Huang, Lixin Zhou, Linxi Huang, Yun Lin, Liehua Deng, ... Yimin Li
Cold Spring Harbor Laboratory (2020-03-06) <https://doi.org/ggq64b>
DOI: [10.1101/2020.03.03.20030668](https://doi.org/10.1101/2020.03.03.20030668)
2005. **Multi-epitope vaccine design using an immunoinformatics approach for 2019 novel coronavirus (SARS-CoV-2)**

Ye Feng, Min Qiu, Liang Liu, Shengmei Zou, Yun Li, Kai Luo, Qianpeng Guo, Ning Han, Yingqiang Sun, Kui Wang, ... Fan Mo
Cold Spring Harbor Laboratory (2020-06-30) <https://doi.org/ggq64c>
DOI: [10.1101/2020.03.03.962332](https://doi.org/10.1101/2020.03.03.962332)

2006. **Clinical Features of Patients Infected with the 2019 Novel Coronavirus (COVID-19) in Shanghai, China**
Min Cao, Dandan Zhang, Youhua Wang, Yunfei Lu, Xiangdong Zhu, Ying Li, Honghao Xue, Yunxiao Lin, Min Zhang, Yiguo Sun, ... Hongzhou Lu
Cold Spring Harbor Laboratory (2020-03-06) <https://doi.org/ggq64d>
DOI: [10.1101/2020.03.04.20030395](https://doi.org/10.1101/2020.03.04.20030395) · PMID: [32511465](#)
2007. **Serological detection of 2019-nCoV respond to the epidemic: A useful complement to nucleic acid testing**
Jin Zhang, Jianhua Liu, Na Li, Yong Liu, Rui Ye, Xiaosong Qin, Rui Zheng
Cold Spring Harbor Laboratory (2020-03-06) <https://doi.org/ggq64f>
DOI: [10.1101/2020.03.04.20030916](https://doi.org/10.1101/2020.03.04.20030916)
2008. **Human Kidney is a Target for Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection**
Bo Diao, Chenhui Wang, Rongshuai Wang, Zeqing Feng, Yingjun Tan, Huiming Wang, Changsong Wang, Liang Liu, Ying Liu, Yueping Liu, ... Yongwen Chen
Cold Spring Harbor Laboratory (2020-04-10) <https://doi.org/ggq64g>
DOI: [10.1101/2020.03.04.20031120](https://doi.org/10.1101/2020.03.04.20031120)
2009. **COVID-19 early warning score: a multi-parameter screening tool to identify highly suspected patients**
Cong-Ying Song, Jia Xu, Jian-Qin He, Yuan-Qiang Lu
Cold Spring Harbor Laboratory (2020-03-08) <https://doi.org/ggq64h>
DOI: [10.1101/2020.03.05.20031906](https://doi.org/10.1101/2020.03.05.20031906)
2010. **LY6E impairs coronavirus fusion and confers immune control of viral disease**
Stephanie Pfaender, Katrina B Mar, Eleftherios Michailidis, Annika Kratzel, Dagny Hirt, Philip V'kovski, Wenchun Fan, Nadine Ebert, Hanspeter Stalder, Hannah Kleine-Weber, ... Volker Thiel
Cold Spring Harbor Laboratory (2020-03-07) <https://doi.org/dpvn>
DOI: [10.1101/2020.03.05.979260](https://doi.org/10.1101/2020.03.05.979260) · PMID: [32511345](#)
2011. **A preliminary study on serological assay for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in 238 admitted hospital patients**
Lei Liu, Wanbing Liu, Yaqiong Zheng, Xiaojing Jiang, Guomei Kou, Jinya Ding, Qiongshu Wang, Qianchuan Huang, Yinjuan Ding, Wenxu Ni, ... Shangen Zheng
Cold Spring Harbor Laboratory (2020-03-08) <https://doi.org/ggq64j>
DOI: [10.1101/2020.03.06.20031856](https://doi.org/10.1101/2020.03.06.20031856)
2012. **Monoclonal antibodies for the S2 subunit of spike of SARS-CoV cross-react with the newly-emerged SARS-CoV-2**
Zhiqiang Zheng, Vanessa M Monteil, Sebastian Maurer-Stroh, Chow Wenn Yew, Carol Leong, Nur Khairiah Mohd-Ismail, Suganya Cheyyatraivendran Arularasu, Vincent Tak Kwong Chow, Raymond Lin Tzer Pin, Ali Mirazimi, ... Yee-Joo Tan

2013. **Mortality of COVID-19 is Associated with Cellular Immune Function Compared to Immune Function in Chinese Han Population**

Qiang Zeng, Yong-zhe Li, Gang Huang, Wei Wu, Sheng-yong Dong, Yang Xu

Cold Spring Harbor Laboratory (2020-03-13) <https://doi.org/ggq64k>

DOI: [10.1101/2020.03.08.20031229](https://doi.org/10.1101/2020.03.08.20031229)

2014. **Retrospective Analysis of Clinical Features in 101 Death Cases with COVID-19**

Hua Fan, Lin Zhang, Bin Huang, Muxin Zhu, Yong Zhou, Huan Zhang, Xiaogen Tao, Shaohui Cheng, Wenhui Yu, Liping Zhu, Jian Chen

Cold Spring Harbor Laboratory (2020-03-17) <https://doi.org/ggq64n>

DOI: [10.1101/2020.03.09.20033068](https://doi.org/10.1101/2020.03.09.20033068)

2015. **The inhaled corticosteroid ciclesonide blocks coronavirus RNA replication by targeting viral NSP15**

Shutoku Matsuyama, Miyuki Kawase, Naganori Nao, Kazuya Shirato, Makoto Ujike, Wataru Kamitani, Masayuki Shimojima, Shuetsu Fukushi

Cold Spring Harbor Laboratory (2020-03-12) <https://doi.org/ggq64p>

DOI: [10.1101/2020.03.11.987016](https://doi.org/10.1101/2020.03.11.987016)

2016. **Immune phenotyping based on neutrophil-to-lymphocyte ratio and IgG predicts disease severity and outcome for patients with COVID-19**

Bicheng Zhang, Xiaoyang Zhou, Chengliang Zhu, Fan Feng, Yanru Qiu, Jia Feng, Qingzhu Jia, Qibin Song, Bo Zhu, Jun Wang

Cold Spring Harbor Laboratory (2020-03-16) <https://doi.org/ggq64q>

DOI: [10.1101/2020.03.12.20035048](https://doi.org/10.1101/2020.03.12.20035048)

2017. **Lack of Reinfection in Rhesus Macaques Infected with SARS-CoV-2**

Linlin Bao, Wei Deng, Hong Gao, Chong Xiao, Jiayi Liu, Jing Xue, Qi Lv, Jiangning Liu, Pin Yu, Yanfeng Xu, ... Chuan Qin

Cold Spring Harbor Laboratory (2020-05-01) <https://doi.org/ggn8r8>

DOI: [10.1101/2020.03.13.990226](https://doi.org/10.1101/2020.03.13.990226)

2018. **A highly conserved cryptic epitope in the receptor-binding domains of SARS-CoV-2 and SARS-CoV**

Meng Yuan, Nicholas C Wu, Xueyong Zhu, Chang-Chun D Lee, Ray TY So, Huibin Lv, Chris KP Mok, Ian A Wilson

Cold Spring Harbor Laboratory (2020-03-14) <https://doi.org/ggq64s>

DOI: [10.1101/2020.03.13.991570](https://doi.org/10.1101/2020.03.13.991570)

2019. **Highly accurate and sensitive diagnostic detection of SARS-CoV-2 by digital PCR**

Lianhua Dong, Junbo Zhou, Chunyan Niu, Quanyi Wang, Yang Pan, Sitong Sheng, Xia Wang, Yongzhuo Zhang, Jiayi Yang, Manqing Liu, ... Xiang Fang

Cold Spring Harbor Laboratory (2020-03-30) <https://doi.org/ggqnqh>

DOI: [10.1101/2020.03.14.20036129](https://doi.org/10.1101/2020.03.14.20036129)

2020. **SARS-CoV-2 invades host cells via a novel route: CD147-spike protein**

Ke Wang, Wei Chen, Yu-Sen Zhou, Jian-Qi Lian, Zheng Zhang, Peng Du, Li Gong, Yang Zhang, Hong-Yong Cui, Jie-Jie Geng, ... Zhi-Nan Chen
Cold Spring Harbor Laboratory (2020-03-14) <https://doi.org/ggq64t>
DOI: [10.1101/2020.03.14.988345](https://doi.org/10.1101/2020.03.14.988345)

2021. **CD147 (EMMPRIN/Basigin) in kidney diseases: from an inflammation and immune system viewpoint**

Tomoki Kosugi, Kayaho Maeda, Waichi Sato, Shoichi Maruyama, Kenji Kadomatsu
Nephrology Dialysis Transplantation (2015-07) <https://doi.org/ggq624>
DOI: [10.1093/ndt/gfu302](https://doi.org/10.1093/ndt/gfu302) · PMID: [25248362](#)

2022. **The roles of CyPA and CD147 in cardiac remodelling**

Hongyan Su, Yi Yang
Experimental and Molecular Pathology (2018-06)
<https://doi.org/ggq622>
DOI: [10.1016/j.yexmp.2018.05.001](https://doi.org/10.1016/j.yexmp.2018.05.001) · PMID: [29772453](#)

2023. **Cancer-related issues of CD147.**

Ulrich H Weidle, Werner Scheuer, Daniela Eggle, Stefan Klostermann, Hannes Stockinger
Cancer genomics & proteomics
<https://www.ncbi.nlm.nih.gov/pubmed/20551248>
PMID: [20551248](#)

2024. **Blood single cell immune profiling reveals the interferon-MAPK pathway mediated adaptive immune response for COVID-19**

Lulin Huang, Yi Shi, Bo Gong, Li Jiang, Xiaoqi Liu, Jialiang Yang, Juan Tang, Chunfang You, Qi Jiang, Bo Long, ... Zhenglin Yang
Cold Spring Harbor Laboratory (2020-03-17) <https://doi.org/ggq64v>
DOI: [10.1101/2020.03.15.20033472](https://doi.org/10.1101/2020.03.15.20033472)

2025. **Cross-reactive antibody response between SARS-CoV-2 and SARS-CoV infections**

Huabin Lv, Nicholas C Wu, Owen Tak-Yin Tsang, Meng Yuan, Ranawaka APM Perera, Wai Shing Leung, Ray TY So, Jacky Man Chun Chan, Garrick K Yip, Thomas Shiu Hong Chik, ... Chris KP Mok
Cold Spring Harbor Laboratory (2020-03-17) <https://doi.org/ggq64w>
DOI: [10.1101/2020.03.15.993097](https://doi.org/10.1101/2020.03.15.993097) · PMID: [32511317](#)

2026. **The feasibility of convalescent plasma therapy in severe COVID-19 patients: a pilot study**

Kai Duan, Bende Liu, Cesheng Li, Huajun Zhang, Ting Yu, Jieming Qu, Min Zhou, Li Chen, Shengli Meng, Yong Hu, ... Xiaoming Yang
Cold Spring Harbor Laboratory (2020-03-23) <https://doi.org/dqrs>
DOI: [10.1101/2020.03.16.20036145](https://doi.org/10.1101/2020.03.16.20036145)

2027. **Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial**

Philippe Gautret, Jean-Christophe Lagier, Philippe Parola, Van Thuan Hoang, Line Meddeb, Morgane Mailhe, Barbara Doudier, Johan Courjon, Valérie Giordanengo, Vera Esteves Vieira, ... Didier Raoult
Cold Spring Harbor Laboratory (2020-03-20) <https://doi.org/dqbv>

DOI: [10.1101/2020.03.16.20037135](https://doi.org/10.1101/2020.03.16.20037135)

2028. **Chloroquine: Modes of action of an undervalued drug**

Rodolfo Thomé, Stefanie Costa Pinto Lopes, Fabio Trindade Maranhão Costa, Liana Verinaud

Immunology Letters (2013-06) <https://doi.org/f5b5cr>

DOI: [10.1016/j.imlet.2013.07.004](https://doi.org/10.1016/j.imlet.2013.07.004) · PMID: [23891850](#)

2029. **Patients with LRBA deficiency show CTLA4 loss and immune dysregulation responsive to abatacept therapy**

B Lo, K Zhang, W Lu, L Zheng, Q Zhang, C Kanelloupolou, Y Zhang, Z Liu, JM Fritz, R Marsh, ... MB Jordan

Science (2015-07-23) <https://doi.org/f7kc8d>

DOI: [10.1126/science.aaa1663](https://doi.org/10.1126/science.aaa1663) · PMID: [26206937](#)

2030. **The sequence of human ACE2 is suboptimal for binding the S spike protein of SARS coronavirus 2**

Erik Procko

Cold Spring Harbor Laboratory (2020-05-11) <https://doi.org/ggrbj8>

DOI: [10.1101/2020.03.16.994236](https://doi.org/10.1101/2020.03.16.994236) · PMID: [32511321](#)

2031. **Comparative Pathogenesis Of COVID-19, MERS And SARS In A Non-Human Primate Model**

Barry Rockx, Thijs Kuiken, Sander Herfst, Theo Bestebroer, Mart M Lamers, Dennis de Meulder, Geert van Amerongen, Judith van den Brand, Nisreen MA Okba, Debby Schipper, ... Bart L Haagmans

Cold Spring Harbor Laboratory (2020-03-17) <https://doi.org/ggq649>

DOI: [10.1101/2020.03.17.995639](https://doi.org/10.1101/2020.03.17.995639)

2032. **Lethal Infection of K18-hACE2 Mice Infected with Severe Acute Respiratory Syndrome Coronavirus**

Paul B McCray, Lecia Pewe, Christine Wohlford-Lenane, Melissa Hickey, Lori Manzel, Lei Shi, Jason Netland, Hong Peng Jia, Carmen Halabi, Curt D Sigmund, ... Stanley Perlman

Journal of Virology (2007-01-15) <https://doi.org/b2dr3s>

DOI: [10.1128/jvi.02012-06](https://doi.org/10.1128/jvi.02012-06) · PMID: [PMC1797474](#)

2033. **Modeling the Impact of Asymptomatic Carriers on COVID-19 Transmission Dynamics During Lockdown**

Jacob B Aguilar, Jeremy Samuel Faust, Lauren M Westafer, Juan B Gutierrez

Cold Spring Harbor Laboratory (2020-08-11) <https://doi.org/ggqnvp>

DOI: [10.1101/2020.03.18.20037994](https://doi.org/10.1101/2020.03.18.20037994)

2034. **Antibody responses to SARS-CoV-2 in COVID-19 patients: the perspective application of serological tests in clinical practice**

Quan-xin Long, Hai-jun Deng, Juan Chen, Jie-li Hu, Bei-zhong Liu, Pu Liao, Yong Lin, Li-hua Yu, Zhan Mo, Yin-yin Xu, ... Ai-long Huang

Cold Spring Harbor Laboratory (2020-03-20) <https://doi.org/ggpvz3>

DOI: [10.1101/2020.03.18.20038018](https://doi.org/10.1101/2020.03.18.20038018)

2035. **Heat inactivation of serum interferes with the immunoanalysis of antibodies to SARS-CoV-2**

Xiumei Hu, Taixue An, Bo Situ, Yuhai Hu, Zihao Ou, Qiang Li, Xiaojing He, Ye Zhang, Peifu Tian, Dehua Sun, ... Lei Zheng

2036. **SARS-CoV-2 specific antibody responses in COVID-19 patients**

Nisreen MA Okba, Marcel A Müller, Wentao Li, Chunyan Wang, Corine H GeurtsvanKessel, Victor M Corman, Mart M Lamers, Reina S Sikkema, Erwin de Bruin, Felicity D Chandler, ... Bart L Haagmans
Cold Spring Harbor Laboratory (2020-03-20) <https://doi.org/ggpvz2>
DOI: [10.1101/2020.03.18.20038059](https://doi.org/10.1101/2020.03.18.20038059)

2037. **A brief review of antiviral drugs evaluated in registered clinical trials for COVID-19**

Drifa Belhadi, Nathan Peiffer-Smadja, François-Xavier Lescure, Yazdan Yazdanpanah, France Mentré, Cédric Laouénan
Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggq65b>
DOI: [10.1101/2020.03.18.20038190](https://doi.org/10.1101/2020.03.18.20038190)

2038. **ACE-2 Expression in the Small Airway Epithelia of Smokers and COPD Patients: Implications for COVID-19**

Janice M Leung, Chen X Yang, Anthony Tam, Tawimas Shaipanich, Tillie-Louise Hackett, Gurpreet K Singhera, Delbert R Dorscheid, Don D Sin
Cold Spring Harbor Laboratory (2020-03-23) <https://doi.org/dqx2>
DOI: [10.1101/2020.03.18.20038455](https://doi.org/10.1101/2020.03.18.20038455)

2039. **Dynamic profile of severe or critical COVID-19 cases**

Yang Xu
Cold Spring Harbor Laboratory (2020-03-20) <https://doi.org/ggrbj9>
DOI: [10.1101/2020.03.18.20038513](https://doi.org/10.1101/2020.03.18.20038513)

2040. **Association between Clinical, Laboratory and CT Characteristics and RT-PCR Results in the Follow-up of COVID-19 patients**

Hang Fu, Huayan Xu, Na Zhang, Hong Xu, Zhenlin Li, Huizhu Chen, Rong Xu, Ran Sun, Lingyi Wen, Linjun Xie, ... Yingkun Guo
Cold Spring Harbor Laboratory (2020-03-23) <https://doi.org/ggq65c>
DOI: [10.1101/2020.03.19.20038315](https://doi.org/10.1101/2020.03.19.20038315)

2041. **An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 and multiple endemic, epidemic and bat coronavirus**

Timothy P Sheahan, Amy C Sims, Shuntai Zhou, Rachel L Graham, Collin S Hill, Sarah R Leist, Alexandra Schäfer, Kenneth H Dinnon, Stephanie A Montgomery, Maria L Agostini, ... Ralph S Baric
Cold Spring Harbor Laboratory (2020-03-20) <https://doi.org/ggrbkb>
DOI: [10.1101/2020.03.19.997890](https://doi.org/10.1101/2020.03.19.997890)

2042. **Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs**

Sangeun Jeon, Meehyun Ko, Jihye Lee, Inhee Choi, Soo Young Byun, Soonju Park, David Shum, Seungtaek Kim
Cold Spring Harbor Laboratory (2020-03-28) <https://doi.org/ggq65h>
DOI: [10.1101/2020.03.20.999730](https://doi.org/10.1101/2020.03.20.999730)

2043. **Respiratory disease and virus shedding in rhesus macaques inoculated with SARS-CoV-2**

Vincent J Munster, Friederike Feldmann, Brandi N Williamson, Neeltje van Doremalen, Lizzette Pérez-Pérez, Jonathan Schulz, Kimberly

Meade-White, Atsushi Okumura, Julie Callison, Beniah Brumbaugh, ...

Emmie de Wit

Cold Spring Harbor Laboratory (2020-03-21) <https://doi.org/ggq65j>

DOI: [10.1101/2020.03.21.001628](https://doi.org/10.1101/2020.03.21.001628) · PMID: [32511299](#)

2044. **Ocular conjunctival inoculation of SARS-CoV-2 can cause mild COVID-19 in Rhesus macaques**

Wei Deng, Linlin Bao, Hong Gao, Zhiguang Xiang, Yajin Qu, Zhiqi Song, Shunran Gong, Jiayi Liu, Jiangning Liu, Pin Yu, ... Chuan Qin

Cold Spring Harbor Laboratory (2020-03-30) <https://doi.org/ggq64r>

DOI: [10.1101/2020.03.13.990036](https://doi.org/10.1101/2020.03.13.990036)

2045. **ACE2 Expression is Increased in the Lungs of Patients with Comorbidities Associated with Severe COVID-19**

Bruna GG Pinto, Antonio ER Oliveira, Youvika Singh, Leandro Jimenez, Andre NA Gonçalves, Rodrigo LT Ogava, Rachel Creighton, Jean Pierre Schatzmann Peron, Helder I Nakaya

Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggq65k>

DOI: [10.1101/2020.03.21.20040261](https://doi.org/10.1101/2020.03.21.20040261) · PMID: [32511627](#)

2046. **Meplazumab treats COVID-19 pneumonia: an open-labelled, concurrent controlled add-on clinical trial**

Huijie Bian, Zhao-Hui Zheng, Ding Wei, Zheng Zhang, Wen-Zhen Kang, Chun-Qiu Hao, Ke Dong, Wen Kang, Jie-Lai Xia, Jin-Lin Miao, ... Ping Zhu

Cold Spring Harbor Laboratory (2020-07-15) <https://doi.org/ggq65m>

DOI: [10.1101/2020.03.21.20040691](https://doi.org/10.1101/2020.03.21.20040691)

2047. **CD147 facilitates HIV-1 infection by interacting with virus-associated cyclophilin A**

T Pushkarsky, G Zybarth, L Dubrovsky, V Yurchenko, H Tang, H Guo, B Toole, B Sherry, M Bukrinsky

Proceedings of the National Academy of Sciences (2001-05-15)

<https://doi.org/cc4c7p>

DOI: [10.1073/pnas.111583198](https://doi.org/10.1073/pnas.111583198) · PMID: [11353871](#) · PMCID: [PMC33473](#)

2048. **CD147/EMMPRIN Acts as a Functional Entry Receptor for Measles Virus on Epithelial Cells**

Akira Watanabe, Misako Yoneda, Fusako Ikeda, Yuri Terao-Muto, Hiroki Sato, Chieko Kai

Journal of Virology (2010-05-01) <https://doi.org/dpcsqg>

DOI: [10.1128/jvi.02168-09](https://doi.org/10.1128/jvi.02168-09) · PMID: [20147391](#) · PMCID: [PMC2863760](#)

2049. **Basigin is a receptor essential for erythrocyte invasion by Plasmodium falciparum**

Cécile Crosnier, Leyla Y Bustamante, SJosefin Bartholdson, Amy K Bei, Michel Theron, Makoto Uchikawa, Souleymane Mboup, Omar Ndir, Dominic P Kwiatkowski, Manoj T Duraisingh, ... Gavin J Wright

Nature (2011-11-09) <https://doi.org/dm59hf>

DOI: [10.1038/nature10606](https://doi.org/10.1038/nature10606) · PMID: [22080952](#) · PMCID: [PMC3245779](#)

2050. **Function of HAb18G/CD147 in Invasion of Host Cells by Severe Acute Respiratory Syndrome Coronavirus**

Zhinan Chen, Li Mi, Jing Xu, Jiyun Yu, Xianhui Wang, Jianli Jiang, Jinliang Xing, Peng Shang, Airong Qian, Yu Li, ... Ping Zhu

The Journal of Infectious Diseases (2005-03) <https://doi.org/cd8snd>

2051. **CD147 mediates intrahepatic leukocyte aggregation and determines the extent of liver injury**

Christine Yee, Nathan M Main, Alexandra Terry, Igor Stevanovski, Annette Maczurek, Alison J Morgan, Sarah Calabro, Alison J Potter, Tina Lemma, David G Bowen, ... Nicholas A Shackel

PLOS ONE (2019-07-10) <https://doi.org/ggq654>

DOI: [10.1371/journal.pone.0215557](https://doi.org/10.1371/journal.pone.0215557) · PMID: [31291257](https://pubmed.ncbi.nlm.nih.gov/31291257/) · PMCID: [PMC6619953](https://pubmed.ncbi.nlm.nih.gov/PMC6619953/)

2052. **Characterisation of the transcriptome and proteome of SARS-CoV-2 using direct RNA sequencing and tandem mass spectrometry reveals evidence for a cell passage induced in-frame deletion in the spike glycoprotein that removes the furin-like cleavage site**

Andrew D Davidson, Maia Kavanagh Williamson, Sebastian Lewis, Deborah Shoemark, Miles W Carroll, Kate Heesom, Maria Zambon, Joanna Ellis, Phillip A Lewis, Julian A Hiscox, David A Matthews

Cold Spring Harbor Laboratory (2020-03-24) <https://doi.org/ggq65n>

DOI: [10.1101/2020.03.22.002204](https://doi.org/10.1101/2020.03.22.002204)

2053. **Modifications to the Hemagglutinin Cleavage Site Control the Virulence of a Neurotropic H1N1 Influenza Virus**

Xiangjie Sun, Longping V Tse, ADamon Ferguson, Gary R Whittaker
Journal of Virology (2010-09-01) <https://doi.org/drs2zt>

DOI: [10.1128/jvi.00797-10](https://doi.org/10.1128/jvi.00797-10) · PMID: [20554779](https://pubmed.ncbi.nlm.nih.gov/20554779/) · PMCID: [PMC2919019](https://pubmed.ncbi.nlm.nih.gov/PMC2919019/)

2054. **The architecture of SARS-CoV-2 transcriptome**

Dongwan Kim, Joo-Yeon Lee, Jeong-Sun Yang, Jun Won Kim, VNarry Kim, Hyeshik Chang

Cold Spring Harbor Laboratory (2020-03-14) <https://doi.org/ggpx9q>

DOI: [10.1101/2020.03.12.988865](https://doi.org/10.1101/2020.03.12.988865)

2055. **A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug-Repurposing**

David E Gordon, Gwendolyn M Jang, Mehdi Bouhaddou, Jiewei Xu, Kirsten Obernier, Matthew J O'Meara, Jeffrey Z Guo, Danielle L Swaney, Tia A Tummino, Ruth Hüttenhain, ... Nevan J Krogan

Cold Spring Harbor Laboratory (2020-03-22) <https://doi.org/ggpptg>

DOI: [10.1101/2020.03.22.002386](https://doi.org/10.1101/2020.03.22.002386) · PMID: [32511329](https://pubmed.ncbi.nlm.nih.gov/32511329/)

2056. **First Clinical Study Using HCV Protease Inhibitor Danoprevir to Treat Naïve and Experienced COVID-19 Patients**

Hongyi Chen, Zhicheng Zhang, Li Wang, Zhihua Huang, Fanghua Gong, Xiaodong Li, Yahong Chen, Jinzi J Wu

Cold Spring Harbor Laboratory (2020-03-24) <https://doi.org/ggrgbt>

DOI: [10.1101/2020.03.22.20034041](https://doi.org/10.1101/2020.03.22.20034041)

2057. **Preclinical Characteristics of the Hepatitis C Virus NS3/4A Protease Inhibitor ITMN-191 (R7227)**

Scott D Seiwert, Steven W Andrews, Yutong Jiang, Vladimir Serebryany, Hua Tan, Karl Kossen, PTRavi Rajagopalan, Shawn Misialek, Sarah K Stevens, Antitsa Stoycheva, ... Lawrence M Blatt

Antimicrobial Agents and Chemotherapy (2008-12)

<https://doi.org/btpg52>

2058. **Efficacy and Safety of All-oral, 12-week Ravidasvir Plus Ritonavir-boosted Danoprevir and Ribavirin in Treatment-naïve Noncirrhotic HCV Genotype 1 Patients: Results from a Phase 2/3 Clinical Trial in China**

Xiaoyuan Xu, Bo Feng, Yujian Guan, Sujun Zheng, Jifang Sheng, Xingxiang Yang, Yuanji Ma, Yan Huang, Yi Kang, Xiaofeng Wen, ... Lai Wei

Journal of Clinical and Translational Hepatology (2019-09-30)

<https://doi.org/ggrbkd>

DOI: [10.14218/jcth.2019.00033](https://doi.org/10.14218/jcth.2019.00033) · PMID: [31608212](https://pubmed.ncbi.nlm.nih.gov/31608212/) · PMCID: [PMC6783683](https://pubmed.ncbi.nlm.nih.gov/PMC6783683/)

2059. **Potentially highly potent drugs for 2019-nCoV**

Duc Duy Nguyen, Kaifu Gao, Jiahui Chen, Rui Wang, Guo-Wei Wei

Cold Spring Harbor Laboratory (2020-02-13) <https://doi.org/ggrbj5>

DOI: [10.1101/2020.02.05.936013](https://doi.org/10.1101/2020.02.05.936013) · PMID: [32511344](https://pubmed.ncbi.nlm.nih.gov/32511344/)

2060. **Serology characteristics of SARS-CoV-2 infection since the exposure and post symptoms onset**

Bin Lou, Ting-Dong Li, Shu-Fa Zheng, Ying-Ying Su, Zhi-Yong Li, Wei Liu, Fei Yu, Sheng-Xiang Ge, Qian-Da Zou, Quan Yuan, ... Yu Chen

Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggrbkc>

DOI: [10.1101/2020.03.23.20041707](https://doi.org/10.1101/2020.03.23.20041707)

2061. **SARS-CoV-2 launches a unique transcriptional signature from in vitro, ex vivo, and in vivo systems**

Daniel Blanco-Melo, Benjamin E Nilsson-Payant, Wen-Chun Liu, Rasmus Møller, Maryline Panis, David Sachs, Randy A Albrecht, Benjamin R tenOever

Cold Spring Harbor Laboratory (2020-03-24) <https://doi.org/ggq65q>

DOI: [10.1101/2020.03.24.2004655](https://doi.org/10.1101/2020.03.24.2004655)

2062. **A New Predictor of Disease Severity in Patients with COVID-19 in Wuhan, China**

Ying Zhou, Zhen Yang, Yanan Guo, Shuang Geng, Shan Gao, Shenglan Ye, Yi Hu, Yafei Wang

Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggq65r>

DOI: [10.1101/2020.03.24.20042119](https://doi.org/10.1101/2020.03.24.20042119)

2063. **Metabolic disturbances and inflammatory dysfunction predict severity of coronavirus disease 2019 (COVID-19): a retrospective study**

Shuke Nie, Xueqing Zhao, Kang Zhao, Zhaojun Zhang, Zhentao Zhang, Zhan Zhang

Cold Spring Harbor Laboratory (2020-03-26) <https://doi.org/ggq65s>

DOI: [10.1101/2020.03.24.20042283](https://doi.org/10.1101/2020.03.24.20042283)

2064. **Viral Kinetics and Antibody Responses in Patients with COVID-19**

Wenting Tan, Yanqiu Lu, Juan Zhang, Jing Wang, Yunjie Dan, Zhaoxia Tan, Xiaoqing He, Chunfang Qian, Qiangzhong Sun, Qingli Hu, ... Guohong Deng

Cold Spring Harbor Laboratory (2020-03-26) <https://doi.org/ggq65t>

DOI: [10.1101/2020.03.24.20042382](https://doi.org/10.1101/2020.03.24.20042382)

2065. **Global profiling of SARS-CoV-2 specific IgG/ IgM responses of convalescents using a proteome microarray**
He-wei Jiang, Yang Li, Hai-nan Zhang, Wei Wang, Dong Men, Xiao Yang, Huan Qi, Jie Zhou, Sheng-ce Tao
Cold Spring Harbor Laboratory (2020-03-27) <https://doi.org/ggq65g>
DOI: [10.1101/2020.03.20.20039495](https://doi.org/10.1101/2020.03.20.20039495)
2066. **Anti-spike IgG causes severe acute lung injury by skewing macrophage responses during acute SARS-CoV infection**
Li Liu, Qiang Wei, Qingqing Lin, Jun Fang, Haibo Wang, Hauyee Kwok, Hangying Tang, Kenji Nishiura, Jie Peng, Zhiwu Tan, ... Zhiwei Chen
JCI Insight (2019-02-21) <https://doi.org/ggqbwp>
DOI: [10.1172/jci.insight.123158](https://doi.org/10.1172/jci.insight.123158) · PMID: [30830861](#) · PMCID: [PMC6478436](#)
2067. **COVID-19 infection induces readily detectable morphological and inflammation-related phenotypic changes in peripheral blood monocytes, the severity of which correlate with patient outcome**
Dan Zhang, Rui Guo, Lei Lei, Hongjuan Liu, Yawen Wang, Yili Wang, Hongbo Qian, Tongxin Dai, Tianxiao Zhang, Yanjun Lai, ... Jinsong Hu
Cold Spring Harbor Laboratory (2020-03-26) <https://doi.org/ggq65v>
DOI: [10.1101/2020.03.24.20042655](https://doi.org/10.1101/2020.03.24.20042655)
2068. **Correlation between universal BCG vaccination policy and reduced morbidity and mortality for COVID-19: an epidemiological study**
Aaron Miller, Mac Josh Reandelar, Kimberly Fasciglione, Violeta Roumenova, Yan Li, Gonzalo H Otazu
Cold Spring Harbor Laboratory (2020-03-28) <https://doi.org/ggq65w>
DOI: [10.1101/2020.03.24.20042937](https://doi.org/10.1101/2020.03.24.20042937)
2069. **BCG vaccination to reduce the impact of COVID-19 in healthcare workers (The BRACE Trial)**
Murdoch Children's Research Institute
<https://www.mcri.edu.au/BRACE>
2070. **Non-neuronal expression of SARS-CoV-2 entry genes in the olfactory system suggests mechanisms underlying COVID-19-associated anosmia**
David H Brann, Tatsuya Tsukahara, Caleb Weinreb, Marcela Lipovsek, Koen Van den Berge, Boying Gong, Rebecca Chance, Iain C Macaulay, Hsin-jung Chou, Russell Fletcher, ... Sandeep Robert Datta
Cold Spring Harbor Laboratory (2020-05-18) <https://doi.org/ggqr4m>
DOI: [10.1101/2020.03.25.009084](https://doi.org/10.1101/2020.03.25.009084)
2071. **Cigarette smoke exposure and inflammatory signaling increase the expression of the SARS-CoV-2 receptor ACE2 in the respiratory tract**
Joan C Smith, Erin L Sausville, Vishruth Girish, Monet Lou Yuan, Kristen M John, Jason M Sheltzer
Cold Spring Harbor Laboratory (2020-04-26) <https://doi.org/ggq65x>
DOI: [10.1101/2020.03.28.013672](https://doi.org/10.1101/2020.03.28.013672)
2072. **The comparative superiority of IgM-IgG antibody test to real-time reverse transcriptase PCR detection for SARS-CoV-2 infection**

diagnosis

Rui Liu, Xinghui Liu, Huan Han, Muhammad Adnan Shereen, Zhili Niu,
Dong Li, Fang Liu, Kailang Wu, Zhen Luo, Chengliang Zhu
Cold Spring Harbor Laboratory (2020-03-30) <https://doi.org/ggqtp5>
DOI: [10.1101/2020.03.28.20045765](https://doi.org/10.1101/2020.03.28.20045765)

12 Appendix A

This appendix contains reviews produced by the Immunology Institute of the Icahn School of Medicine

12.1 Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody

Tian et al. *Emerg Microbes Infect* 2020 [[1950](#)]

12.1.1 Keywords

- Monoclonal antibody
- Cross-reactivity
- receptor binding domain

12.1.2 Summary

Considering the relatively high identity of the receptor binding domain (RBD) of the spike proteins from 2019-nCoV and SARS-CoV (73%), this study aims to assess the cross-reactivity of several anti-SARS-CoV monoclonal antibodies with 2019-nCoV. The results showed that the SARS-CoV-specific antibody CR3022 can potently bind 2019-nCoV RBD.

12.1.3 Main Findings

The structure of the 2019-nCoV spike RBD and its conformation in complex with the receptor angiotensin-converting enzyme (ACE2) was modeled *in silico* and compared with the SARS-CoV RBD structure. The models predicted very similar RBD-ACE2 interactions for both viruses. The binding capacity of representative SARS-CoV-RBD specific monoclonal antibodies (m396, CR3014, and CR3022) to recombinant 2019-nCoV RBD was then investigated by ELISA and their binding kinetics studied using biolayer interferometry. The analysis showed that only CR3022 was able to bind 2019-nCoV RBD with high affinity (KD of 6.3 nM), however it did not interfere with ACE2 binding. Antibodies m396 and CR3014, which target the ACE2 binding site of SARS-CoV failed to bind 2019-nCoV spike protein.

12.1.4 Limitations

The 2019-nCoV RBD largely differ from the SARS-CoV at the C-terminus residues, which drastically impact the cross-reactivity of antibodies described for other B beta-coronaviruses, including SARS-CoV. This study claims that CR3022 antibody could be a potential candidate for therapy. However, none of the antibodies assayed in this work showed cross-reactivity with the ACE2 binding site of 2019-nCoV, essential for the replication of this virus. Furthermore, neutralization assays with 2019-nCoV virus or pseudovirus

were not performed. Although the use of neutralizing antibodies is an interesting approach, these results suggest that it is critical the development of novel monoclonal antibodies able to specifically bind 2019-nCoV spike protein.

12.1.5 Credit

Review by D.L.O as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.2 Integrative Bioinformatics Analysis Provides Insight into the Molecular Mechanisms of 2019-nCoV

He et al. *medRxiv* [[1951](#)]

12.2.1 Keywords

- ACE2
- lungs
- smoking
- COPD
- asthma
- SARS-CoV
- IL-1
- IL-10
- IL-6
- IL-8

12.2.2 Main Findings

The authors used bioinformatics tools to identify features of ACE2 expression in the lungs of different patient groups: healthy, smokers, patients with chronic airway disease (i.e., COPD) or asthma. They used gene expression data publicly available from GEO that included lung tissues, bronchoalveolar lavage, bronchial epithelial cells, small airway epithelial cells, or SARS-CoV infected cells.

The authors describe no significant differences in ACE2 expression in lung tissues of Healthy, COPD, and Asthma groups ($p=0.85$); or in BAL of Healthy and COPD ($p=0.48$); or in epithelial brushings of Healthy and Mild/Moderate/Severe Asthma ($p=0.99$). ACE2 was higher in the small airway epithelium of long-term smokers vs non-smokers ($p<0.001$). Consistently, there was a trend of higher ACE2 expression in the bronchial airway epithelial cells 24h post-acute smoking exposure ($p=0.073$). Increasing ACE2 expression at 24h and 48h compared to 12h post SARS-CoV infection ($p=0.026$; $n=3$ at each time point) was also detected.

15 lung samples' data from healthy participants were separated into high and low ACE2 expression groups. "High" ACE2 expression was associated with the following GO pathways: innate and adaptive immune responses, B cell

mediated immunity, cytokine secretion, and IL-1, IL-10, IL-6, IL-8 cytokines. The authors speculate that a high basal ACE2 expression will increase susceptibility to SARS-CoV infection.

In 3 samples SARS-CoV infection was associated with IL-1, IL-10 and IL-6 cytokine production (GO pathways) at 24h. And later, at 48h, with T-cell activation and T-cell cytokine production. It is unclear whether those changes were statistically significant.

The authors describe a time course quantification of immune infiltrates in epithelial cells infected with SARS-CoV infection. They state that in healthy donors ACE2 expression did not correlate with the immune cell infiltration. However, in SARS-CoV samples, at 48h they found that ACE2 correlated with neutrophils, NK-, Th17-, Th2-, Th1- cells, and DCs. Again, while authors claim significance, the corresponding correlation coefficients and p-values are not presented in the text or figures. In addition, the source of the data for this analysis is not clear.

Using network analysis, proteins SRC, FN1, MAPK3, LYN, MBP, NLRC4, NLRP1 and PRKCD were found to be central (Hub proteins) in the regulating network of cytokine secretion after coronavirus infection. Authors conclude this indicates that these molecules were critically important in ACE2-induced inflammatory response. Additionally, authors speculate that the increased expression of ACE2 affected RPS3 and SRC, which were the two hub genes involved in viral replication and inflammatory response.

12.2.3 Limitations

The methods section is very limited and does not describe any of the statistical analyses; and description of the construction of the regulatory protein networks is also limited. For the findings in Figures 2 authors claim significance, which is not supported by p-values or coefficients. For the sample selection, would be useful if sample sizes and some of the patients' demographics (e.g. age) were described.

For the analysis of high vs low ACE2 expression in healthy subjects, it is not clear what was the cut off for 'high' expression and how it was determined. Additionally, further laboratory studies are warranted to confirm that high ACE2 gene expression would have high correlation with the amount of ACE2 protein on cell surface. For the GO pathway analysis significance was set at $p<0.05$, but not adjusted for multiple comparisons.

There were no samples with SARS-CoV-2 infection. While SARS-CoV and SARS-CoV-2 both use ACE2 to enter the host cells, the analysis only included data on SARS-CoV and any conclusions about SARS-CoV2 are limited.

Upon checking GSE accession numbers of the datasets references, two might not be cited correctly: GSE37758 ("A spurgillus niger: Control (fructose) vs. steam-exploded sugarcane induction (SEB)" was used in this paper as "lung tissue" data) and GSE14700 ("Steroid Pretreatment of Organ Donors to Prevent Postischemic Renal Allograft Failure: A Randomized, Controlled Trial" - was used as SARS-CoV infection data).

12.2.4 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.3 Diarrhea may be underestimated: a missing link in 2019 novel coronavirus

Liang et al. *medRxiv* [[1952](#)]

12.3.1 Keywords

- SARS-CoV-2
- diarrhea
- ACE2
- scRNA-seq

12.3.2 Main Findings

This study examined the incidence of diarrhea in patients infected with SARS-CoV-2 across three recently published cohorts and found that there are statistically significant differences by Fisher's exact test. They report that this could be due to subjective diagnosis criterion for diarrhea or from patients first seeking medical care from gastroenterologist. In order to minimize nosocomial infections arising from unsuspected patients with diarrhea and gain comprehensive understanding of transmission routes for this viral pathogen, they compared the transcriptional levels of ACE2 of various human tissues from NCBI public database as well as in small intestine tissue from CD57BL/6 mice using single cell sequencing. They show that ACE2 expression is not only increased in the human small intestine, but demonstrate a particular increase in mice enterocytes positioned on the surface of the intestinal lining exposed to viral pathogens. Given that ACE2 is the viral receptor for SARS-CoV-2 and also reported to regulate diarrhea, their data suggests the small intestine as a potential transmission route and diarrhea as a potentially underestimated symptom in COVID19 patients that must be carefully monitored. Interestingly, however, they show that ACE2 expression level is not elevated in human lung tissue.

12.3.3 Limitations

Although this study demonstrates a statistical difference in the incidence of diarrhea across three separate COVID19 patient cohorts, their conclusions are limited by a small sample size. Specifically, the p-value computed by Fisher's exact test is based on a single patient cohort of only six cases of which 33% are reported to have diarrhea, while the remaining two larger cohorts with 41 and 99 cases report 3% and 2% diarrhea incidence, respectively. Despite showing significance, they would need to acquire larger sample sizes and cohorts to minimize random variability and draw meaningful conclusions. Furthermore, they do not address why ACE2 expression level is not elevated in human lung tissue despite it being a major

established route of transmission for SARS-CoV-2. It could be helpful to validate this result by looking at ACE2 expression in mouse lung tissue. Finally, although this study is descriptive and shows elevated ACE2 expression in small intestinal epithelial cells, it does not establish a mechanistic link to SARS-CoV-2 infection of the host. Overall, their claim that infected patients exhibiting diarrhea pose an increased risk to hospital staff needs to be further substantiated.

12.3.4 Significance

This study provides a possible transmission route and a potentially underappreciated clinical symptom for SARS-CoV-2 for better clinical management and control of COVID19.

12.3.5 Credit

Summary generated as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.4 Specific ACE2 Expression in Cholangiocytes May Cause Liver Damage After 2019-nCoV Infection

Chai et al. *bioRxiv* [[1953](#)]

12.4.1 Keywords

- ACE2
- Cholangiocytes
- COVID-associated Liver Damage

12.4.2 Summary

Using both publicly available scRNA-seq dataset of liver samples from colorectal patients and scRNA-sequencing of four liver samples from healthy volunteers, the authors show that ACE2 is significantly enriched in the majority of cholangiocytes (59.7 %) but not in hepatocytes (2.6%).

12.4.3 Main Findings

Using bioinformatics approaches of RNASeq analysis, this study reveals that ACE2 dominates in cholangiocytes and is present at very low levels in hepatocytes.

12.4.4 Limitations

The study does not provide mechanistic insights into how SARS-CoV-2 can infect and replicate in cholangiocytes and the types of intrinsic anti-viral responses induced by cholangiocytes when infected. In addition, because the

study relies on the assumption that SARS-CoV-2 infects cells only through ACE2, it cannot discount the possibility that the virus can infect hepatocytes through mechanisms other than ACE2-mediated entry. Furthermore, because the scRNA-seq analysis were performed on healthy liver samples, one cannot draw any definitive conclusions about gene expression states (including ACE2 expression in liver cell types) in system-wide inflammatory contexts.

12.4.5 Significance

This article with other studies on liver damage in COVID patients suggests that liver damage observed in COVID patients is more due to inflammatory cytokines than direct infection of the liver. Even if cholangiocytes are infectable by SARS-CoV-2 (which was demonstrated by human liver ductal organoid study ([1954])), published clinical data show no significant increase in bile duct injury related indexes (i.e. alkaline phosphatase, gamma-glutamyl transpeptidase and total bilirubin). In sum, it underscores the importance of future studies characterizing cellular responses of extra-pulmonary organs in the context of COVID or at least in viral lung infections..

12.4.6 Credit

Summary generated by Chang Moon as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.5 ACE2 expression by colonic epithelial cells is associated with viral infection, immunity and energy metabolism

Wang et al. *medRxiv*. [1955]

12.5.1 Keywords

- single cell RNA seq
- ACE2 expression
- human colonic biopsy

12.5.2 Main Findings

Colonic enterocytes primarily express ACE2. Cellular pathways associated with ACE2 expression include innate immune signaling, HLA up regulation, energy metabolism and apoptotic signaling.

12.5.3 Limitations

This is a study of colonic biopsies taken from 17 children with and without IBD and analyzed using scRNAseq to look at ACE2 expression and identify gene families correlated with ACE2 expression. The authors find ACE2 expression to be primarily in colonocytes. It is not clear why both healthy and

IBD patients were combined for the analysis. Biopsies were all of children so extrapolation to adults is limited. The majority of genes found to be negatively correlated with ACE2 expression include immunoglobulin genes (IGs). IG expression will almost certainly be low in colonocytes irrespective of ACE2 expression.

12.5.4 Significance

This study performs a retrospective analysis of ACE2 expression using an RNAseq dataset from intestinal biopsies of children with and without IBD. The implications for the CoV-19 epidemic are modest, but do provide support that ACE2 expression is specific to colonocytes in the intestines. The ontological pathway analysis provides some limited insights into gene expression associated with ACE2.

12.5.5 Credit

Summary generated as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.6 The Pathogenicity of 2019 Novel Coronavirus in hACE2 Transgenic Mice

Bao et al. *bioRxiv* [1956]

12.6.1 Keywords

- Covid-19 mouse model
- hACE2 mice
- 2019-nCoV model
- ACE2
- 2019-nCoV

12.6.2 Main Findings

Using a transgenic human Angiotensin-converting enzyme 2 (hACE2) mouse that has previously been shown susceptible to infection by SARS-CoV, Bao et al. create a model of pandemic 2019-nCoV strain coronavirus. The model includes interstitial hyperplasia in lung tissue, moderate inflammation in bronchioles and blood vessels, and histology consistent with viral pneumonia at 3 days post infection. Wildtype did not experience these symptoms. In addition, viral antigen and hACE2 receptor were found to co-localize the lung by immunofluorescence 3-10 days post infection only in the hACE2 infected mice.

12.6.3 Limitations

The characterization of the infection remains incomplete, as well as lacking characterization of the immune response other than the presence of a single antiviral antibody. Though they claim to fulfill Koch's postulates, they only

isolate the virus and re-infect Vero cells, rather than naive mice.

12.6.4 Significance

This paper establishes a murine model for 2019-nCoV infection with symptoms consistent with viral pneumonia. Though not fully characterized, this model allows *in vivo* analysis of viral entry and pathology that is important for the development of vaccines and antiviral therapeutics.

12.6.5 Credit

Review by Dan Fu Ruan, Evan Cody and Venu Pothula as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.7 Caution on Kidney Dysfunctions of 2019-nCoV Patients

Li et al. *medRxiv*. [\[1957\]](#)

12.7.1 Keywords

CoVID-19, 2019-nCoV, SARS-CoV-2, kidney, clinical, creatinine, proteinuria, albuminuria, CT

12.7.2 Main Findings

- Retrospective study of 59 patients assayed key function indicators of the kidney—including urine protein, blood urea nitrogen (BUN), plasma creatinine (Cre), and renal CT scan data.
- Found that 34% of patients developed massive albuminuria on the first day of admission, and 63% developed proteinuria during their stay in hospital; and 19% of patients had high plasma creatinine, especially the terminal cases.
- CT analyses of 27 patients showed all patients to have abnormal kidney damage; indicate that inflammation and edema of the renal parenchyma very common.

12.7.3 Limitations

- No analysis of immunity-dependent damage and cytokines in blood/plasma/urine. Will be worth correlating disease progression with cytokine production, immune activity and kidney function.
- Extrapolating to earlier SARS-CoV studies provides the only rationale for viral-damage in kidney and resultant pathologic immune response (*understandable for this clinical study*).

12.7.4 Significance

- Multiple lines of evidence along this study's finding point to the idea that renal impairment/injury is a key risk factor in 2019-nCoV patients similar to what has been reported for SARS-CoV[[1958](#)]; this may be one of the major causes of virally-induced damage and contribute to multiorgan failure.
- ACE2 expression in kidney proximal tubule epithelia and bladder epithelia [[1959](#)] support these clinical findings.
- Study argues for closely monitoring kidney function, and applying potential interventions including continuous renal replacement therapies (CRRT) for protecting kidney functions as early as possible, particularly for those with rising plasma creatinine.

12.7.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.8 Profiling the immune vulnerability landscape of the 2019 Novel Coronavirus

Zhu et al. *bioRxiv* [[1960](#)]

12.8.1 Keywords

- epitope prediction
- vaccine development.

12.8.2 Main Findings

This study harnesses bioinformatic profiling to predict the potential of COV2 viral proteins to be presented on MHC I and II and to form linear B-cell epitopes. These estimates suggest a T-cell antigenic profile distinct from SARS-CoV or MERS-CoV, identify focused regions of the virus with a high density of predicted epitopes, and provide preliminary evidence for adaptive immune pressure in the genetic evolution of the virus.

12.8.3 Limitations

While the study performs a comprehensive analysis of potential epitopes within the virus genome, the analysis relies solely on bioinformatic prediction to examine MHC binding affinity and B-cell epitope potential and does not capture the immunogenicity or recognition of these epitopes. Future experimental validation in data from patients infected with SARS-CoV-2 will be important to validate and refine these findings. Thus some of

the potential conclusions stated, including viral evolution toward lower immunogenicity or a dominant role for CD4+ T-cells rather than CD8+ T-cells in viral clearance, require further validation.

12.8.4 Significance

These findings may help direct peptide vaccine design toward relevant epitopes and provide intriguing evidence of viral evolution in response to immune pressure.

12.8.5 Credit

Summary generated as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.9 Single-cell Analysis of ACE2 Expression in Human Kidneys and Bladders Reveals a Potential Route of 2019-nCoV Infection

Lin et al. *bioRxiv* [1959]

12.9.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- ACE2
- scRNAseq
- kidney
- bladder
- public dataset

12.9.2 Main Findings

- To investigate the possible cause of kidney damage in 2019-nCoV patients, authors used published kidney and bladder cell atlas data (GSE131685, GSE108097; 3 healthy donors each) as well as an unpublished kidney single-cell RNA-Seq data (in-house from 2 transplant donors) to evaluate ACE2 gene expressions in all cell types of healthy kidneys and bladders.
- They find enriched expression of ACE2 transcript in all subtypes of proximal tubule cells of kidney, with 5%-15% of both straight and convoluted proximal tubule cells expressing ACE2.
- They also find detectable levels of ACE2 in bladder epithelial cells, noting expression from around 1.5% of cells in the outer layer umbrella cells of the bladder epithelium and decreasing in the basal cells.
- Importantly endothelial or immune cells in kidney/bladder do not express ACE2.

12.9.3 Limitations

- This study primarily characterizes ACE2 expression (amongst other genes) from a small healthy-donor dataset, and will benefit from supporting data in (expired) patient samples to show functional viral damage. ACE2 transcript does not necessarily translate to viral permissiveness in kidney/bladder epithelia or cytokine release.
- This study focuses on only healthy tissue; it will be useful to analyze kidney/bladder epithelial ACE2 expression under inflammatory conditions or in patients with underlying kidney conditions.
- Given what is known about protease TMPRSS2 expression during SARS-CoV-2 infection, ACE2+TMPRSS2+ double-positive cell identification would be useful in these datasets.

12.9.4 Significance

- ACE2 protein is spatially restricted to brush border of proximal tubules and in bladder umbrella cells [65], such cells in direct contact with viral particles are likely to be highly sensitive to viral-induced damage.
- SARS-CoV and MERS-CoV have been shown to be detected in urine of patients and associate with higher mortality [1958,1961], thus worth understanding kidney damage and resultant immune response in SARS-CoV-2 as well.
- This study argues for a potential mode of viral infectivity and resultant inflammatory responses in these tissue in addition to reported infectivity in the lung and digestive system, which is supported by clinical data showing acute and early kidney complications in 2019-nCoV patients [1957].
- Clinically, thus very important to track urinary CoVID-19 shedding as well as study acute kidney injury-related co-morbidities.

12.9.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.10 Neutrophil-to-Lymphocyte Ratio Predicts Severe Illness Patients with 2019 Novel Coronavirus in the Early Stage

Liu et al. *medRxiv* [1962]

12.10.1 Keywords

- severe disease
- pneumonia
- lymphocytes
- neutrophils

12.10.2 Main Findings

This study aimed to find prognostic biomarkers of COVID-19 pneumonia severity. Sixty-one (61) patients with COVID-19 treated in January at a hospital in Beijing, China were included. On average, patients were seen within 5 days from illness onset. Samples were collected on admission; and then patients were monitored for the development of severe illness with a median follow-up of 10 days].

Patients were grouped as “mild” (N=44) or “moderate/severe” (N=17) according to symptoms on admission and compared for different clinical/laboratory features. “Moderate/severe” patients were significantly older (median of 56 years old, compared to 41 years old). Whereas comorbidities rates were largely similar between the groups, except for hypertension, which was more frequent in the severe group ($p= 0.056$). ‘Severe’ patients had higher counts of neutrophils, and serum glucose levels; but lower lymphocyte counts, sodium and serum chlorine levels. The ratio of neutrophils to lymphocytes (NLR) was also higher for the ‘severe’ group. ‘Severe’ patients had a higher rate of bacterial infections (and antibiotic treatment) and received more intensive respiratory support and treatment.

26 clinical/laboratory variables were used to select NLR and age as the best predictors of the severe disease. Predictive cutoffs for a severe illness as $\text{NLR} \geq 3.13$ or $\text{age} \geq 50$ years.

12.10.3 Limitations

Identification of early biomarkers is important for making clinical decisions, but large sample size and validation cohorts are necessary to confirm findings. It is worth noting that patients classified as “mild” showed pneumonia by imaging and fever, and in accordance with current classifications this would be consistent with “moderate” cases. Hence it would be more appropriate to refer to the groups as “moderate” vs “severe/critical”. Furthermore, there are several limitations that could impact the interpretation of the results: e.g. classification of patients was based on symptoms presented on admission and not based on disease progression, small sample size, especially the number of ‘severe’ cases (with no deaths among these patients). Given the small sample size, the proposed NLR and age cut offs might not hold for a slightly different set of patients. For example, in a study of >400 patients, ‘non-severe’ and ‘severe’ NLR were 3.2 and 5.5, respectively [1963].

12.10.4 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.11 Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP)

Wan et. al. *medRxiv* [[1964](#)]

12.11.1 Keywords

- Cytokines
- lymphocyte subsets
- CD8 + T
- B cells
- NK cells,
- PBMCs
- IL-6
- IL-10

12.11.2 Main Findings

The authors analyzed lymphocyte subsets and cytokines of 102 patients with mild disease and 21 with severe disease. CD8+T cells and CD4+T cells were significantly reduced in both cohort, particularly in severe patients. The cytokines IL6 and IL10 were significantly elevated in severe patients as compared to mild. No significant differences were observed in frequency of B cells and NK cells.

The authors argue that the measurement of T cell frequencies and cytokine levels of IL6 and IL10 can be used to predict progression of disease from Mild to severe Cov-2 infection.

12.11.3 Limitations

The study demonstrates in a limited cohort similar associations to several other reported studies. The authors didn't compare the changes in lymphocyte and cytokine with healthy individual (Covid-19 Negative) rather used an internal standard value. The recently preprint in LANCET shows The degree of lymphopenia and a pro-inflammatory cytokine storm is higher in severe COVID-19 patients than in mild cases, and is associated with the disease severity [[1965](#)].

12.11.4 Significance

This translational data identifies key cytokines and lymphopenia associated with disease severity although mechanism and key cellular players are still unknown. Higher level IL-6 production in severe patient suggests potential role of Tocilizumab (anti-IL6R) biologic although clinical trial will be necessary.

12.11.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.12 Epidemiological and Clinical Characteristics of 17 Hospitalized Patients with 2019 Novel Coronavirus Infections Outside Wuhan, China

Li et al. *medRxiv* [1966]

12.12.1 Keywords

- epidemiology
- clinical characteristics

12.12.2 Major Findings

These authors looked at 17 hospitalized patients with COVID-19 confirmed by RT-PCR in Dazhou, Sichuan. Patients were admitted between January 22 and February 10 and the final data were collected on February 11. Of the 17 patients, 12 remained hospitalized while 5 were discharged after meeting national standards. The authors observed no differences based on the sex of the patients but found that the discharged patients were younger in age ($p = 0.026$) and had higher lymphocyte counts ($p = 0.005$) and monocyte counts ($p = 0.019$) upon admission.

12.12.3 Limitations

This study is limited in the sample size of the study and the last data collection point was only one day after some of the patients were admitted.

12.12.4 Significance

These findings have been somewhat supported by subsequent studies that show that older age and an immunocompromised state are more likely to result in a more severe clinical course with COVID-19. However, other studies have been published that report on larger numbers of cases.

12.12.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.13 ACE2 Expression in Kidney and Testis May Cause Kidney and Testis Damage After 2019-nCoV Infection

[1967]

12.13.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- kidney
- testis
- ACE2
- scRNAseq

12.13.2 Main Findings

- Study used online datasets (scRNAseq GSE131685, scRNAseq GSE107585, Human Protein Atlas, GTEx portal, CCLE) to analyze ACE2 expression in different human organs.
- Study re-analyzed three clinical datasets (n=6, n=99, and n=41) to show 3~10% of 2019-nCoV patients present with abnormal renal function.
- results indicate ACE2 highly expressed in renal tubular cells, Leydig cells and seminiferous ductal cells of testis.

12.13.3 Limitations

- Very preliminary transcript/protein dataset analysis in healthy cohorts; does not necessarily translate to actual viral tropism and permissiveness.
- Clinically, would be important to determine with larger longitudinal dataset if SARS-CoV-2 infection changes sperm quality or testicular inflammation.
- Similarly, would be important to determine if simultaneous HBV or syphilis infection and orchitis impacts SARS-CoV-2 severity.
- Examination and follow-up of renal function and viral orchitis/sperm quality of CoVID-19 patients not done in this preliminary study.

12.13.4 Significance

- Kidney ACE2 result supports other concurrent sequencing studies [1959] and clinical reports of abnormal renal function or even kidney damage in patients infected with 2019-nCoV [1957].
- High ACE2 expression in testis suggests potential tropism of the virus to testicular tissues and indicates potential risks for male fertility. Viral orchitis reported for SARS-CoV previously [1], but no clear evidence so far of infertility in SARS, MERS or CoVID-19 patients.

12.13.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.14 Aberrant pathogenic GM-CSF+ T cells and inflammatory CD14+CD16+ monocytes in severe pulmonary syndrome patients of a new coronavirus

[[1968](#)]

12.14.1 Keywords

- immunopathology
- Th1
- inflammatory monocytes
- GM-CSF
- IFN-γ
- IL-6

12.14.2 Main Findings

The authors of this study sought to characterize the immune mechanism causing severe pulmonary disease and mortality in 2019-nCoV (COVID-19) patients. Peripheral blood was collected from hospitalized ICU (n=12) and non-ICU (n=21) patients with confirmed 2019-nCoV and from healthy controls (n=10) in The First Affiliated Hospital of University of Science and Technology China (Hefei, Anhui). Immune analysis was conducted by flow cytometry. 2019-nCoV patients had decreased lymphocyte, monocyte, and CD4 T cell counts compared to healthy controls. ICU patients had fewer lymphocytes than non-ICU patients. CD4 T cells of 2019-nCoV patients expressed higher levels of activation markers (OX40, CD69, CD38, CD44) and exhaustion markers (PD-1 and Tim3) than those of healthy controls. CD4 cells of ICU patients expressed significantly higher levels of OX40, PD-1, and Tim3 than those of non-ICU patients. 2019-nCoV patients had higher percentages of CD4 T cells co-expressing GM-CSF and IL-6 compared to healthy controls, while ICU patients had a markedly higher percentage of GM-CSF+ IFN-γ+ CD4 T cells than non-ICU patients. The CD4 T cells of nCoV patients and healthy controls showed no differences in TNF-α secretion.

The CD8 T cells of 2019-nCoV patients also showed higher expression of activation markers CD69, CD38, and CD44, as well as exhaustion markers PD-1 and Tim3, compared to healthy controls. CD8 T cells of ICU patients expressed higher levels of GM-CSF than those of non-ICU patients and healthy controls. No IL-6 or TNF-α was found in the CD8 T cells of any group. There were no differences in numbers of NK cells or B cells in 2019-nCoV patients and healthy controls, nor was there any GM-CSF or IL-6 secretion from these cells in either group.

Percentages of CD14+ CD16+ GM-CSF+ and CD14+ CD16+ IL-6+ inflammatory monocytes were significantly increased in nCoV patients compared to healthy controls; in particular, patients in the ICU had greater percentages of CD14+ CD16+ IL-6+ monocytes than non-ICU patients. The authors suggest that in 2019-nCoV patients, pathogenic Th1 cells produce GM-CSF, recruiting CD14+ CD16+ inflammatory monocytes that secrete high levels of IL-6. These may enter pulmonary circulation and damage lung tissue while initiating the cytokine storm that causes mortality in severe cases. This is consistent with the cytokine storm seen in similar coronaviruses, as IL-6, IFN- γ , and GM-CSF are key inflammatory mediators seen in patients with SARS-CoV-1 and MERS-CoV.

12.14.3 Limitations

Though the results of this study open questions for further investigation, this is an early study on a small cohort of patients, and as such there are a number of limitations. The study included only 12 ICU patients and 21 non-ICU patients, and ideally would be repeated with a much larger patient cohort. Though the authors make claims about differences in lymphocyte and monocyte counts between patients and healthy controls, they did not report baseline laboratory findings for the control group. Additionally, severity of disease was classified based on whether or not patients were in the ICU. It would be interesting to contextualize the authors' immunological findings with more specific metrics of disease severity or time course. Noting mortality, time from disease onset, pre-existing conditions, or severity of lung pathology in post-mortem tissue samples would paint a fuller picture of how to assess risk level and the relationship between severity of disease and immunopathology. Another limitation is the selection of cytokines and immune markers for analysis, as the selection criteria were based on the cell subsets and cytokine storm typically seen in SARS-CoV-1 and MERS-CoV patients. Unbiased cytokine screens and immune profiling may reveal novel therapeutic targets that were not included in this study.

12.14.4 Significance

This study identifies potential therapeutic targets that could prevent acute respiratory disease syndrome (ARDS) and mortality in patients most severely affected by COVID-19. The authors propose testing monoclonal antibodies against IL6-R or GM-CSF to block recruitment of inflammatory monocytes and the subsequent cytokine storm in these patients.

12.14.5 Credit

Review by Gabrielle Lubitz as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.15 Clinical Characteristics of 2019 Novel Infected Coronavirus Pneumonia : A Systemic Review and Meta-analysis

12.15.1 Keywords

- White Blood Cells
- Lymphocytes
- Neutrophils

12.15.2 Main Findings

The authors performed a meta analysis of literature on clinical, laboratory and radiologic characteristics of patients presenting with pneumonia related to SARSCoV2 infection, published up to Feb 6 2020. They found that symptoms that were mostly consistent among studies were sore throat, headache, diarrhea and rhinorrhea. Fever, cough, malaise and muscle pain were highly variable across studies. Leukopenia (mostly lymphocytopenia) and increased white blood cells were highly variable across studies. They identified three most common patterns seen on CT scan, but there was high variability across studies. Consistently across the studies examined, the authors found that about 75% of patients need supplemental oxygen therapy, about 23% mechanical ventilation and about 5% extracorporeal membrane oxygenation (ECMO). The authors calculated a staggering pooled mortality incidence of 78% for these patients.

12.15.3 Limitations

The authors mention that the total number of studies included in this meta analysis is nine, however they also mentioned that only three studies reported individual patient data. It is overall unclear how many patients in total were included in their analysis. This is mostly relevant as they reported an incredibly high mortality (78%) and mention an absolute number of deaths of 26 cases overall. It is not clear from their report how the mortality rate was calculated.

The data is based on reports from China and mostly from the Wuhan area, which somewhat limits the overall generalizability and applicability of these results.

12.15.4 Significance

This meta analysis offers some important data for clinicians to refer to when dealing with patients with COVID-19 and specifically with pneumonia. It is very helpful to set expectations about the course of the disease.

12.15.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.16 Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of SARS-CoV-2 infected patients

Liu et al. *medRxiv* [1970]

12.16.1 Keywords

- Lymphopenia
- Neutrophil to CD8 T cell ratio (N8R)
- inflammatory cytokines

12.16.2 Main Findings

Liu et al. enrolled a cohort of 40 patients from Wuhan including 27 mild cases and 13 severe cases of COVID-19. They performed a 16-day kinetic analysis of peripheral blood from time of disease onset. Patients in the severe group were older (medium age of 59.7, compared to 48.7 in mild group) and more likely to have hypertension as a co-morbidity. Lymphopenia was observed in 44.4% of the mild patients and 84.6% of the severe patients. Lymphopenia was due to low T cell count, specially CD8 T cells. Severe patients showed higher neutrophil counts and an increase of cytokines in the serum (IL2, IL6, IL10 and IFNy). The authors measured several other clinical laboratory parameters were also higher in severe cases compared to mild, but concluded that neutrophil to CD8 T cell ratio (N8R) as the best prognostic factor to identify the severe cases compared to other receiver operating characteristic (ROC).

12.16.3 Limitations

This was a small cohort (N=40), and two of the patients initially included in the severe group (N=13) passed away and were excluded from the analysis due to lack of longitudinal data. However, it would be most important to be able to identify patients with severe disease with higher odds of dying. It seems that the different time points analyzed relate to hospital admission, which the authors describe as disease onset. The time between first symptoms and first data points is not described. It would have been important to analyze how the different measured parameters change according to health condition, and not just time (but that would require a larger cohort). The predictive value of N8R compared to the more commonly used NLR needs to be assessed in other independent and larger cohorts. Lastly, it is important to note that pneumonia was detected in patients included in the "mild" group, but according to the Chinese Clinical Guidance for COVID-19 Pneumonia Diagnosis and Treatment (7th edition) this group should be considered "moderate".

12.16.4 Significance

Lymphopenia and cytokine storm have been described to be detrimental in many other infections including SARS-CoV1 and MERS-CoV. However, it was necessary to confirm that this dramatic immune response was also observed in the SARS-CoV2 infected patients. These results and further validation of the N8R ratio as a predictor of disease severity will contribute for the management of COVID19 patients and potential development of therapies.

12.16.5 Credit

Review by Pauline Hamon as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.17 Clinical and immunologic features in severe and moderate forms of Coronavirus Disease 2019

Chen et al. *medRxiv* [[1971](#)]

12.17.1 Keywords

- severe disease
- lymphocytes
- cytokines
- IFNy
- CD4 Tcells
- HLA-DR CD8
- Tcells

12.17.2 Main Findings

This study retrospectively evaluated clinical, laboratory, hematological, biochemical and immunologic data from 21 subjects admitted to the hospital in Wuhan, China (late December/January) with confirmed SARS-CoV-2 infection. The aim of the study was to compare 'severe' (n=11, ~64 years old) and 'moderate' (n=10, ~51 years old) COVID-19 cases. Disease severity was defined by patients' blood oxygen level and respiratory output. They were classified as 'severe' if SpO₂ 93% or respiratory rates 30 per min.

In terms of the clinical laboratory measures, 'severe' patients had higher CRP and ferritin, alanine and aspartate aminotransferases, and lactate dehydrogenase but lower albumin concentrations.

The authors then compared plasma cytokine levels (ELISA) and immune cell populations (PBMCs, Flow Cytometry). 'Severe' cases had higher levels of IL-2R, IL-10, TNFa, and IL-6 (marginally significant). For the immune cell counts, 'severe' group had higher neutrophils, HLA-DR+ CD8 T cells and total B cells; and lower total lymphocytes, CD4 and CD8 T cells (except for HLA-DR+), CD45RA Tregs, and IFNy-expressing CD4 T cells. No significant differences were observed for IL-8, counts of NK cells, CD45+RO Tregs, IFNy-expressing CD8 T and NK cells.

12.17.3 Limitations

Several potential limitations should be noted: 1) Blood samples were collected 2 days post hospital admission and no data on viral loads were available; 2) Most patients were administered medications (e.g. corticosteroids), which could have affected lymphocyte counts. Medications are briefly mentioned in the text of the manuscript; authors should include medications as part of Table 1. 3) 'Severe' cases were significantly older and 4/11 'severe' patients died within 20 days. Authors should consider a sensitivity analysis of biomarkers with the adjustment for patients' age.

12.17.4 Significance

Although the sample size was small, this paper presented a broad range of clinical, biochemical, and immunologic data on patients with COVID-19. One of the main findings is that SARS-CoV-2 may affect T lymphocytes, primarily CD4+ T cells, resulting in decreased IFNy production. Potentially, diminished T lymphocytes and elevated cytokines can serve as biomarkers of severity of COVID-19.

12.17.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.18 SARS-CoV-2 and SARS-CoV Spike-RBD Structure and Receptor Binding Comparison and Potential Implications on Neutralizing Antibody and Vaccine Development

Sun et al. *bioRxiv* [891]

12.18.1 Keywords

- SARS-CoV
- SARS-CoV-2
- ACE2
- Spike (S) protein
- receptor binding domain (RBD)
- receptor binding motif (RBM)
- neutralizing antibody

12.18.2 Main Findings

This study compared the structure of SARS-CoV and SARS-CoV-2 Spike (S) protein receptor binding domain (RBD) and interactions with ACE2 using computational modeling, and interrogated cross-reactivity and cross-neutralization of SARS-CoV-2 by antibodies against SARS-CoV. While SARS-

CoV and SARS-CoV-2 have over 70 % sequence homology and share the same human receptor ACE2, the receptor binding motif (RBM) is only 50% homologous.

Computational prediction of the SARS-CoV-2 and ACE2 interactions based on the previous crystal structure data of SARS-CoV, and measurement of binding affinities against human ACE2 using recombinant SARS-CoV and SARS-CoV-2 S1 peptides, demonstrated similar binding of the two S1 peptides to ACE2, explaining the similar transmissibility of SARS-CoV and SARS-CoV-2 and consistent with previous data (Wall et al Cell 2020).

The neutralization activity of SARS-CoV-specific rabbit polyclonal antibodies were about two-order of magnitude less efficient to neutralize SARS-CoV-2 than SARS-CoV, and four potently neutralizing monoclonal antibodies against SARS-CoV had poor binding and neutralizing activity against SARS-CoV-2. In contrast, 3 poor SARS-CoV-binding monoclonal antibodies show some efficiency to bind and neutralize SARS-CoV-2. The results suggest that that antibodies to more conserved regions outside the RBM motif might possess better cross-protective neutralizing activities between two strains.

12.18.3 Limitations

It would have been helpful to show the epitopes recognized by the monoclonal antibodies tested on both SARS-CoV, SARS-CoV-2 to be able to make predictions for induction of broadly neutralizing antibodies. The data on monoclonal antibody competition with ACE2 for binding to SARS-CoV RBD should have also included binding on SARS-CoV2, especially for the three monoclonal antibodies that showed neutralization activity for SARS-CoV2. Because of the less homology in RBM sequences between viruses, it still may be possible that these antibodies would recognize the ACE2 RBD in SARS-CoV-2.

12.18.4 Significance

It is noteworthy that immunization to mice and rabbit with SARS-CoV S1 or RBD protein could induce monoclonal antibodies to cross-bind and cross-neutralize SARS-CoV-2 even if they are not ACE2-blocking. If these types of antibodies could be found in human survivors or in the asymptomatic populations as well, it might suggest that exposure to previous Coronavirus strains could have induced cross-neutralizing antibodies and resulted in the protection from severe symptoms in some cases of SARS-CoV2.

12.18.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.19 Protection of Rhesus Macaque from SARS-CoV-1 challenge by recombinant adenovirus vaccine

Chen et al. *bioRxiv* [1972]

12.19.1 Keywords

- SARS-CoV-1
- rhesus macaque
- recombinant adenovirus vaccine

12.19.2 Main Findings

Rhesus macaques were immunized intramuscularly twice (week 0 and week 4) with SV8000 carrying the information to express a S1-orf8 fusion protein and the N protein from the BJ01 strain of SARS-CoV-1. By week 8, immunized animals had signs of immunological protection (IgG and neutralization titers) against SARS-CoV-1 and were protected against challenge with the PUMC-1 strain, with fewer detectable symptoms of respiratory distress, lower viral load, shorter periods of viral persistence, and less pathology in the lungs compared to non-immunized animals.

12.19.3 Limitations

The authors should write clearer descriptions of the methods used in this article. They do not describe how the IgG titers or neutralization titers were determined. There are some issues with the presentation of data, for example, in Figure 1a, y-axis should not be Vmax; forming cells and 1d would benefit from showing error bars. Furthermore, although I inferred that the animals were challenged at week 8, the authors did not explicitly detail when the animals were challenged. The authors should explain the design of their vaccine, including the choice of antigens and vector. The authors also do not include a description of the ethical use of animals in their study.

12.19.4 Significance

The authors describe a vaccine for SARS-CoV-1 with no discussion of possible implications for the current SARS-CoV-2 pandemic. Could a similar vaccine be designed to protect against SARS-CoV-2 and would the concerns regarding emerging viral mutations that the authors describe as a limitation for SARS-CoV-1 also be true in the context of SARS-CoV-2?

12.19.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.20 Reduction and Functional Exhaustion of T cells in Patients with Coronavirus Disease 2019 (COVID-19)

[1973]

12.20.1 Keywords

- T cell exhaustion
- T cell lymphopenia
- IL-6
- IL-10
- TNF- α

12.20.2 Main Findings

Based on a retrospective study of 522 COVID patients and 40 healthy controls from two hospitals in Wuhan, China, authors show both age-dependent and clinical severity-dependent decrease in T cell numbers with elderly patients and patients who are in ICU-care showing the most dramatic decrease in T cell counts. Cytokine profiling of COVID patients reveal that TNF- α , IL-6 and IL-10 are increased in infected patients with patients in the ICU showing the highest levels. Interestingly, these three cytokine levels were inversely correlated with T cell counts and such inverse relationship was preserved throughout the disease progression. Surface staining of exhaustion markers (PD-1 and Tim-3) and flow cytometry of stained peripheral blood of 14 patients and 3 healthy volunteers demonstrate that T cells of COVID patients have increased expression of PD-1 with patients in ICU having the highest number of CD8 $^{+}$ PD-1 $^{+}$ cells than their counterparts in non-ICU groups.

12.20.3 Limitations

Compared to the number of patients, number of control (n= 40) is small and is not controlled for age. Additional data linking inflammatory cytokines and the quality of the adaptive response including humoral and antigen specific T cell response is much needed. T cell exhaustion study relies on marker-dependent labeling of T cell functionality of a very limited sample size (n=17)—a functional/mechanistic study of these T cells from PBMCs would have bolstered their claims.

12.20.4 Significance

Limited but contains interesting implications. It is already known in literature that in the context of acute respiratory viral infections CD8 T cells exhibit exhaustion-like phenotypes which further underscores the importance of mechanistic studies that can elucidate how COVID infection leads to lymphopenia and T cell exhaustion-like phenotype.

However, as authors have noted, the data does point to an interesting question: How these inflammatory cytokines (TNF- α , IL-6 and IL-10) correlate with or affect effective viral immunity and what types of cells produce these cytokines? Answering that question will help us refine our targets for immune-modulatory therapies especially in patients suffering from cytokine storms.

12.20.5 Credit

This review by Chang Moon was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.21 Clinical Characteristics of 25 death cases infected with COVID-19 pneumonia: a retrospective review of medical records in a single medical center, Wuhan, China

[1974]

12.21.1 Keywords

- COVID-19
- pneumonia
- hypertension
- diabetes
- biomarker
- neutrophilia
- lymphopenia

12.21.2 Main Findings

Most common chronic conditions among 25 patients that died from COVID-19 related respiratory failure were hypertension (64%) and diabetes (40%). Disease progression was marked by progressive organ failure, starting first with lung dysfunction, then heart (e.g. increased cTnI and pro-BNP), followed by kidney (e.g. increased BUN, Cr), and liver (e.g. ALT, AST). 72% of patients had neutrophilia and 88% also had lymphopenia. General markers of inflammation were also increased (e.g. PCT, D-Dimer, CRP, LDH, and SAA).

12.21.3 Limitations

The limitations of this study include small sample size and lack of measurements for some tests for several patients. This study would also have been stronger with comparison of the same measurements to patients suffering from less severe disease to further validate and correlate proposed biomarkers with disease severity.

12.21.4 Significance

This study identifies chronic conditions (i.e. hypertension and diabetes) that strongly correlates with disease severity. In addition to general markers of inflammation, the authors also identify concomitant neutrophilia and lymphopenia among their cohort of patients. This is a potentially interesting immunological finding because we would typically expect increased lymphocytes during a viral infection. Neutrophilia may also be contributing to cytokine storm. In addition, PCT was elevated in 90.5% of patients, suggesting a role for sepsis or secondary bacterial infection in COVID-19 related respiratory failure.

12.21.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.22 SARS-CoV-2 infection does not significantly cause acute renal injury: an analysis of 116 hospitalized patients with COVID-19 in a single hospital, Wuhan, China

[1975]

12.22.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- kidney
- clinical
- longitudinal

12.22.2 Main Findings

- Clinical data from 116 hospitalized CoVID-19 patients analyzed over 4 weeks for correlation with renal injury. Comorbidities included chronic renal failure (CRF) in 5 patients (4.3%).
- 10.8% of patients with no prior kidney disease showed elevations in blood urea or creatinine, and 7.2% of patients with no prior kidney disease showed albuminuria.
- Patients with pre-existing CRF underwent continuous renal replacement therapy (CRRT) alongside CoVID-19 treatment. Renal functions remained stable in these patients.
- All 5 patients with CRF survived CoVID-19 therapy without progression to ARDS or worsening of CRF.

12.22.3 Limitations

- Renal injury biomarkers in patients with incipient kidney abnormalities not tabulated separately, making overall data hard to interpret. It will be critical to separately examine kidney function (BUN, urine creatinine and eGFR) in patients that developed any kidney abnormalities (7.2~10.8% of cohort).
- No information on type of CoVID-19 therapy used across cohort; will be useful to correlate how treatment modality influences kidney function (and other parameters).
- Invokes previous clinical-correlation studies that indicate low instances of kidney damage [[1976](#),[1977](#)], but those studies did not track longitudinal urine samples for acute renal injury markers and viral shedding.
- CRRT in patients with CRF is standard therapy irrespective of CoVID-19 status; it will be important to compare clinical parameters of these patients (n=5) with virus-naïve CRF patients (none in this study) to make any meaningful conclusions.

12.22.4 Significance

- This study argues that renal impairment is uncommon in CoVID-19 and not associated with high mortality, in stark contrast with a concurrent study [[1957](#)]. If supported by further studies, this argues kidney impairment is secondary to cytokine storm/inflammation-induced organ failure, and not due to direct viral replication.
- Will be important to comprehensively characterize large-datasets of CoVID-19 patients to conclude if kidney function actively disrupted due to viral infection.

12.22.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.23 Potential T-cell and B-Cell Epitopes of 2019-nCoV

[[1978](#)]

12.23.1 Keywords

- COVID-19
- vaccine
- epitopes
- spike protein
- MHC-I
- MHC-II
- neutralizing antibodies

- ACE2

12.23.2 Main Findings

The authors use 2 neural network algorithms, NetMHCpan4 and MARIA, to identify regions within the COVID-19 genome that are presentable by HLA. They identify 405 viral epitopes that are presentable on MHC-I and MHC-II and validate using known epitopes from SARS-CoV. To determine whether immune surveillance drives viral mutations to evade MHC presentation, the authors analyzed 68 viral genomes from 4 continents. They identified 93 point mutations that occurred preferentially in regions predicted to be presented by MHC-I ($p=0.02$) suggesting viral evolution to evade CD8 T-cell mediated killing. 2 nonsense mutations were also identified that resulted in loss of presentation of an associated antigen (FGDSVEEV) predicted to be good antigen for presentation across multiple HLA alleles.

To identify potential sites of neutralizing antibody binding, the authors used homology modeling to the SARS-CoV's spike protein (S protein) to determine the putative structure of the CoV2 spike protein. They used Discotope2 to identify antibody binding sites on the protein surface in both the down and up conformations of the S protein. The authors validate this approach by first identifying antibody binding site in SARS-CoV S protein. In both the down and up conformation of the CoV2 S protein, the authors identified a potential antibody binding site on the S protein receptor binding domain (RBD) of the ACE2 receptor (residues 440-460, 494-506). While RBDs in both SARS-CoV and CoV2 spike proteins may be important for antibody binding, the authors note that SARS-CoV has larger attack surfaces than CoV2. These results were later validated on published crystal structures of the CoV2 S protein RBD and human ACE2. Furthermore, analysis of 68 viral genomes did not identify any mutations in this potential antibody binding site in CoV2.

Finally, the authors compile a list of potential peptide vaccine candidates across the viral genome that can be presented by multiple HLA alleles. Several of the peptides showed homology to SARS-CoV T-cell and B-cell epitopes.

12.23.3 Limitations

While the authors used computational methods of validation, primarily through multiple comparisons to published SARS-CoV structures and epitopes, future work should include experimental validation of putative T-cell and B-cell epitopes.

12.23.4 Significance

The authors identified potential T-cell and B-cell epitopes that may be good candidates for peptide based vaccines against CoV2. They also made interesting observations in comparing SARS-CoV and CoV2 potential antibody binding sites, noting that SARS-CoV had larger attack surfaces for potential neutralizing antibody binding. One of the highlights of this paper was the authors' mutation analysis of 68 viral genomes from 4 continents. This analysis not only validated their computational method for identifying T-cell

epitopes, but showed that immune surveillance likely drives viral mutation in MHC-I binding peptides. The smaller attack surface may point to potential mechanisms of immune evasion by CoV2. However, absence of mutations in the RBD of CoV2 and the small number of mutations in peptides presentable to T cells suggests that vaccines against multiple epitopes could still elicit robust immunity against CoV2.

12.23.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.24 Structure, Function, and Antigenicity of the SARSCoV-2 Spike Glycoprotein

Walls et al. *bioRxiv*. [1979] now [23]

12.24.1 Keywords

- binding affinity
- antigenicity
- neutralizing antibody

12.24.2 Main Findings

The authors highlight a human angiotensin-converting enzyme 2 (hACE2), as a potential receptor used by the current Severe Acute respiratory syndrome coronavirus-2 (SARS-CoV-2) as a host factor that allows the virus target human cells. This virus-host interaction facilitates the infection of human cells with a high affinity comparable with SARS-CoV. The authors propose this mechanism as a probable explanation of the efficient transmission of SARS-CoV-2 between humans. Besides, Walls and colleagues described SARS-CoV-2 S glycoprotein S by Cryo-EM along with neutralizing polyclonal response against SAR-CoV-2 S from mice immunized with SAR-CoV and blocking SAR-CoV-2 S-mediated entry into VeroE6 infected cells.**

12.24.3 Limitations

The SARS-CoV-2 depends on the cell factors ACE2 and TMPRSS2, this last, according to a recent manuscript by Markus Hoffman et al., *Cell*, 2020. The authors used green monkey (VeroE6) and hamster (BHK) cell lines in the experiments to drive its conclusions to humans; however, it is well known the caucasian colon adenocarcinoma human cell line (CaCo-2), highly express the hACE2 receptor as the TMPRSS2 protease as well. In humans, ACE2 protein is highly expressed in the gastrointestinal tract, which again, makes the CaCo-2 cell line suitable for the following SARS-CoV-2 studies.

12.24.4 Significance

The results propose a functional receptor used by SARS-CoV-2 to infect humans worldwide and defining two distinct conformations of spike (S) glycoprotein by cryogenic electron microscopy (Cryo-EM). This study might help establish a precedent for initial drug design and treatment of the current global human coronavirus epidemic.

12.24.5 Credit

Review by postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.25 Breadth of concomitant immune responses underpinning viral clearance and patient recovery in a non-severe case of COVID-19

Thevarajan et al. *medRxiv* [1980]

12.25.1 Keywords

- IgG
- IgM
- TfH cells
- NK cells
- SNP

12.25.2 Main Findings

The authors characterized the immune response in peripheral blood of a 47-year old COVID-19 patient.

SARS-CoV2 was detected in nasopharyngeal swab, sputum and faeces samples, but not in urine, rectal swab, whole blood or throat swab. 7 days after symptom onset, the nasopharyngeal swab test turned negative, at day 10 the radiography infiltrates were cleared and at day 13 the patient became asymptomatic.

Immunofluorescence staining shows from day 7 the presence of **COVID-19-binding IgG and IgM** antibodies in plasma, that increase until day 20.

Flow cytometry on whole blood reveals a plasmablast peak at day 8, a gradual increase in T follicular helper cells, stable HLA-DR⁺ NK frequencies and decreased monocyte frequencies compared to healthy counterparts. The expression of CD38 and HLA-DR peaked on T cells at D9 and was associated with higher production of cytotoxic mediators by CD8⁺ T cells.

IL-6 and IL-8 were undetectable in plasma.

The authors further highlight the presence of the **IFITM3 SNP-rs12252-C/C variant** in this patient, which is associated with higher susceptibility to influenza virus.

12.25.3 Limitations

These results need to be confirmed in additional patients.

COVID-19 patients have increased infiltration of macrophages in their lungs [1981]. Monitoring monocyte proportions in blood earlier in the disease might help to evaluate their eventual migration to the lungs.

The stable concentration of HLA-DR⁺ NK cells in blood from day 7 is not sufficient to rule out NK cell activation upon SARS-CoV2 infection. In response to influenza A virus, NK cells express higher levels of activation markers CD69 and CD38, proliferate better and display higher cytotoxicity [1982]. Assessing these parameters in COVID-19 patients is required to better understand NK cell role in clearing this infection.

Neutralization potential of the COVID-19-binding IgG and IgM antibodies should be assessed in future studies.

This patient was able to clear the virus, while presenting a SNP associated with severe outcome following influenza infection. The association between this SNP and outcome upon SARS-CoV2 infection should be further investigated.

12.25.4 Significance

This study is among the first to describe the appearance of COVID-19-binding IgG and IgM antibodies upon infection. The emergence of new serological assays might contribute to monitor more precisely the seroconversion kinetics of COVID-19 patients [1983]. Further association studies between IFITM3 SNP-rs12252-C/C variant and clinical data might help to refine the COVID-19 outcome prediction tools.

12.25.5 Credit

Review by Bérengère Salomé as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.26 The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing

Liao et al. medRxiv [1981]

12.26.1 Keywords

- COVID-19
- SARS-CoV-2
- Broncho-alveolar lavage
- macrophages
- NK cells
- T cells
- cytokine storm
- scRNAseq

12.26.2 Main Findings

The authors performed single-cell RNA sequencing (scRNAseq) on bronchoalveolar lavage fluid (BAL) from 6 COVID-19 patients (n=3 mild cases, n=3 severe cases). Data was compared to previously generated scRNAseq data from healthy donor lung tissue (n=8).

Clustering analysis of the 6 patients revealed distinct immune cell organization between mild and severe disease. Specifically, they found that transcriptional clusters annotated as tissue resident alveolar macrophages were strongly reduced while monocytes-derived FCN1⁺SPP1⁺ inflammatory macrophages dominated the BAL of patients with severe COVID19 diseases. They show that inflammatory macrophages upregulated interferon-signaling genes, monocytes recruiting chemokines including CCL2, CCL3, CCL4 as well as IL-6, TNF, IL-8 and profibrotic cytokine TGF-β, while alveolar macrophages expressed lipid metabolism genes, such as PPARG.

The lymphoid compartment was overall enriched in lungs from patients. Clonally expanded CD8 T cells were enriched in mild cases suggesting that CD8 T cells contribute to viral clearance as in Flu infection, whereas proliferating T cells were enriched in severe cases.

SARS-CoV-2 viral transcripts were detected in severe patients, but considered here as ambient contaminations.

12.26.3 Limitations

These results are based on samples from 6 patients and should therefore be confirmed in the future in additional patients. Longitudinal monitoring of BAL during disease progression or resolution would have been most useful.

The mechanisms underlying the skewing of the macrophage compartment in patients towards inflammatory macrophages should be investigated in future studies.

Deeper characterization of the lymphoid subsets is required. The composition of the “proliferating” cluster and how these cells differ from conventional T cell clusters should be assessed. NK and CD8 T cell transcriptomic profile, in particular the expression of cytotoxic mediator and immune checkpoint transcripts, should be compared between healthy and diseased lesions.

12.26.4 Significance

COVID-19 induces a robust inflammatory cytokine storm in patients that contributes to severe lung tissue damage and ARDS [1984]. Accumulation of monocyte-derived inflammatory macrophages at the expense of Alveolar macrophages known to play an anti-inflammatory role following respiratory viral infection, in part through the PPAR γ pathway [1985,1986] are likely contributing to lung tissue injuries. These data suggest that reduction of monocyte accumulation in the lung tissues could help modulate COVID-19-induced inflammation. Further analysis of lymphoid subsets is required to understand the contribution of adaptive immunity to disease outcome.

12.26.5 Credit

Review by Bérengère Salomé and Assaf Magen as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.27 Can routine laboratory tests discriminate 2019 novel coronavirus infected pneumonia from other community-acquired pneumonia?

Pan et al. *medRxiv* [1987]

12.27.1 Keywords

- Routine laboratory testing

12.27.2 Main Findings

In an attempt to use standard laboratory testing for the discrimination between “Novel Coronavirus Infected Pneumonia” (NCIP) and a usual community acquired pneumonia (CAP), the authors compared laboratory testing results of 84 NCIP patients with those of a historical group of 316 CAP patients from 2018 naturally COVID-19 negative. The authors describe significantly lower white blood- as well as red blood- and platelet counts in NCIP patients. When analyzing differential blood counts, lower absolute counts were measured in all subsets of NCIP patients. With regard to clinical chemistry parameters, they found increased AST and bilirubin in NCIP patients as compared to CAP patients.

12.27.3 Limitations

The authors claim to describe a simple method to rapidly assess a pre-test probability for NCIP. However, the study has substantial weakpoints. The deviation in clinical laboratory values in NCIP patients described here can usually be observed in severely ill patients. The authors do not comment on how severely ill the patients tested here were in comparison to the historical control. Thus, the conclusion that the tests discriminate between CAP and NCIP lacks justification.

12.27.4 Significance

The article strives to compare initial laboratory testing results in patients with COVID-19 pneumonia as compared to patients with a usual community acquired pneumonia. The implications of this study for the current clinical situation seem restricted due to a lack in clinical information and the use of a control group that might not be appropriate.

12.27.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.28 Correlation Analysis Between Disease Severity and Inflammation-related Parameters in Patients with COVID-19 Pneumonia

[1988]

12.28.1 Keywords

- cytokine
- COVID-19 pneumonia
- severity
- disease progression

12.28.2 Main Findings

This study is a cross-sectional analysis of 100 patients with COVID-19 pneumonia, divided into mild ($n = 34$), severe ($n = 34$), and critical ($n = 32$) disease status based on clinical definitions.

The criteria used to define disease severity are as follows:

1. *Severe* – any of the following: respiratory distress or respiratory rate ≥ 30 respirations/minute; oxygen saturation $\leq 93\%$ at rest; oxygen partial pressure (PaO₂)/oxygen concentration (FiO₂) in arterial blood $\leq 300\text{mmHg}$, progression of disease on imaging to $>50\%$ lung involvement in the short term.
2. *Critical* – any of the following: respiratory failure that requires mechanical ventilation; shock; other organ failure that requires treatment in the ICU.
3. Patients with pneumonia who test positive for COVID-19 who do not have the symptoms delineated above are considered *mild*.

Peripheral blood inflammatory markers were correlated to disease status. Disease severity was significantly associated with levels of IL-2R, IL-6, IL-8, IL-10, TNF- α , CRP, ferroprotein, and procalcitonin. Total WBC count, lymphocyte count, neutrophil count, and eosinophil count were also significantly correlated with disease status. Since this is a retrospective, cross-sectional

study of clinical laboratory values, these data may be extrapolated for clinical decision making, but without studies of underlying cellular causes of these changes this study does not contribute to a deeper understanding of SARS-CoV-2 interactions with the immune system.

It is also notable that the mean age of patients in the mild group was significantly different from the mean ages of patients designated as severe or critical ($p < 0.001$). The mean patient age was not significantly different between the severe and critical groups. However, IL-6, IL-8, procalcitonin (Table 2), CRP, ferroprotein (Figure 3A, 3B), WBC count, and neutrophil count (Figure 4A, 4B) were all significantly elevated in the critical group compared to severe. These data suggest underlying differences in COVID-19 progression that is unrelated to age.

12.28.3 Significance

Given the inflammatory profile outlined in this study, patients who have mild or severe COVID-19 pneumonia, who *also* have any elevations in the inflammatory biomarkers listed above, should be closely monitored for potential progression to critical status.

12.28.4 Credit

This review by JJF was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.29 An Effective CTL Peptide Vaccine for Ebola Zaire Based on Survivors' CD8+ Targeting of a Particular Nucleocapsid Protein Epitope with Potential Implications for COVID-19 Vaccine Design

Herst et al. *bioRxiv* [1989]

12.29.1 Keywords

- Peptide vaccine
- Ebolavirus
- nucleocapsid
- epitope
- vaccine design
- microsphere

12.29.2 Main Findings

Vaccination of mice with a single dose of a 9-amino-acid peptide NP44-52 located in a conserved region of ebolavirus (EBOV) nucleocapsid protein (NP) confers CD8+ T-cell-mediated immunity against mouse adapted EBOV

(maEBOV). Bioinformatic analyses predict multiple conserved CD8+ T cell epitopes in the SARS-CoV-2 NP, suggesting that a similar approach may be feasible for vaccine design against SARS-CoV-2.

The authors focus on a site within a 20-peptide region of EBOV NP which was commonly targeted by CD8+ T cells in a group of EBOV survivors carrying the HLA-A*30:01:01 allele. To justify the testing of specific vaccine epitopes in a mouse challenge setting, the authors cite known examples of human pathogen-derived peptide antigens that are also recognized by C57BL/6 mice, as well as existing data surrounding known mouse immunogenicity of peptides related to this EBOV NP region. Testing 3 distinct 9mer peptides over an 11 amino-acid window and comparing to vaccination with the 11mer with a T-cell reactivity readout demonstrated that optimizing peptide length and position for immunogenicity may be crucial, likely due to suboptimal peptide processing and MHC-class-I loading.

Vaccines for maEBOV challenge studies were constructed by packaging NP44-52 in d,l poly(lactic-co-glycolic) acid microspheres. CpG was also packaged within the microspheres, while Monophosphoryl Lipid A (a TLR4 ligand) was added to the injectate solution. A second peptide consisting of a predicted MHC-II epitope from the EBOV VG19 protein was added using a separate population of microspheres, and the formulation was injected by intraperitoneal administration. The vaccine was protective against a range of maEBOV doses up to at least 10,000 PFU. Survival was anticorrelated with levels of IL6, MCP-1 (CCL2), IL9, and GM-CSF, which recapitulated trends seen in human EBOV infection.

While HLA-A*30:01:01 is only present in a minority of humans, the authors state that MHC binding algorithms predict NP44-52 to be a strong binder of a set of more common HLA-A*02 alleles. The authors predict that a peptide vaccine based on the proposed formulation could elicit responses in up to 50% of people in Sudan or 30% of people in North America.

SARS-CoV-2 NP, meanwhile, has conserved regions which may provide peptide-vaccine candidates. Scanning the SARS-CoV-2 NP sequence for HLA-binding 9mers identified 53 peptides with predicted binding affinity < 500nM, including peptides that are predicted to bind to HLA-class-I alleles of 97% of humans, 7 of which have previously been tested *in-vitro*.

The results support previously appreciated correlations between certain cytokines and disease severity, specifically IL6 which relates to multiple trial therapies. Prediction of HLA-class-I binding of SARS-CoV-2 NP peptides suggests the plausibility of a peptide vaccine targeting conserved regions of SARS-CoV-2 NP although further validation in previously infected patient samples will be essential.

12.29.3 Credit

Review by Andrew M. Leader as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.30 Epitope-based peptide vaccines predicted against novel coronavirus disease caused by SARS-CoV-2

Li et al. *bioRxiv*. [1990]

12.30.1 Keywords

- SARS-CoV-2
- immune-informatics
- vaccine design
- T cell epitope
- B cell epitope

12.30.2 Main Findings

This study employs a series of bioinformatic pipelines to identify T and B cell epitopes on spike (S) protein of SARS-CoV-2 and assess their properties for vaccine potential. To identify B cell epitopes, they assessed structural accessibility, hydrophilicity, and beta-turn and flexibility which are all factors that promote their targeting by antibodies. To identify T cell epitopes, they filtered for peptides with high antigenicity score and capacity to bind 3 or more MHC alleles. Using the protein digest server, they also demonstrated that their identified T and B cell epitopes are stable, having multiple non-digesting enzymes per epitope. Epitopes were also determined to be non-allergenic and non-toxin as assessed by Allergen FP 1.0 and ToxinPred, respectively. For T cell epitopes, they assessed the strength of epitope-HLA interaction via PepSite. Overall, they predict four B cell and eleven T cell epitopes (two MHC I and nine MHC II binding) to pass stringent computational thresholds as candidates for vaccine development. Furthermore, they performed sequence alignment between all identified SARS-CoV-2 S protein mutations and predicted epitopes, and showed that the epitopes are conserved across 134 isolates from 38 locations worldwide. However, they report that these conserved epitopes may soon become obsolete given the known mutation rate of related SARS-CoV is estimated to be 4×10^{-4} /site/year, underscoring the urgency of anti-viral vaccine development.

12.30.3 Limitations

While spike (S) protein may have a critical role in viral entry into host cells and their epitope prediction criterion were comprehensive, this study did not examine other candidate SARS-CoV-2 proteins. This point is particularly important given that a single epitope may not be sufficient to induce robust immune memory, and recent approaches involve multi-epitope vaccine design. Furthermore, their study only included a direct implementation of various published methods, but did not validate individual bioinformatic tools with controls to demonstrate robustness. Finally, it is critical that these predicted epitopes are experimentally validated before any conclusions can be drawn about their potential as vaccine candidates or their clinical efficacy.

12.30.4 Significance

This study provides a computational framework to rapidly identify epitopes that may serve as potential vaccine candidates for treating SARS-CoV-2.

12.30.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.31 The definition and risks of Cytokine Release Syndrome-Like in 11 COVID-19-Infected Pneumonia critically ill patients: Disease Characteristics and Retrospective Analysis

Wang Jr. et al. *medRxiv*. [[1991](#)]

12.31.1 Keywords

- Cytokine release syndrome (CRS)
- biomarkers
- ARDS
- IL-6
- lymphopenia

12.31.2 Main Findings

This study describes the occurrence of a cytokine release syndrome-like (CRSL) toxicity in ICU patients with COVID-19 pneumonia. The median time from first symptom to acute respiratory distress syndrome (ARDS) was 10 days. All patients had decreased CD3, CD4 and CD8 cells, and a significant increase of serum IL-6. Furthermore, 91% had decreased NK cells. The changes in IL-6 levels preceded those in CD4 and CD8 cell counts. All of these parameters correlated with the area of pulmonary inflammation in CT scan images. Mechanical ventilation increased the numbers of CD4 and CD8 cells, while decreasing the levels of IL-6, and improving the immunological parameters.

12.31.3 Limitations

The number of patients included in this retrospective single center study is small ($n=11$), and the follow-up period very short (25 days). Eight of the eleven patients were described as having CRSL, and were treated by intubation (7) or ECMO (2). Nine patients were still in the intensive care unit at the time of publication of this article, so their disease outcome is unknown.

12.31.4 Significance

The authors define a cytokine release syndrome-like toxicity in patients with COVID-19 with clinical radiological and immunological criteria: 1) decrease of circulating CD4, CD8 and NK cells; 2) substantial increase of IL-6 in peripheral blood; 3) continuous fever; 4) organ and tissue damage. This event seems to occur very often in critically ill patients with COVID-19 pneumonia.

Interestingly, the increase of IL-6 in the peripheral blood preceded other laboratory alterations, thus, IL-6 might be an early biomarker for the severity of COVID-19 pneumonia. The manuscript will require considerable editing for organization and clarity.

12.31.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.32 Clinical characteristics of 36 non-survivors with COVID-19 in Wuhan, China

Huang et al. *medRxiv*. [[1992](#)]

12.32.1 Keywords

- Clinical Characteristics
- Non Survivors
- retrospective study

12.32.2 Main Findings

This is a simple study reporting clinical characteristics of patients who did not survive COVID-19. All patients (mean age=69.22 years) had acute respiratory distress syndrome (ARDS) and their median time from onset to ARDS was 11 days. The median time from onset to death was 17 days. Most patients were older male (70% male) with co-morbidities and only 11 % were smokers. 75% patients showed bilateral pneumonia. Many patients had chronic diseases, including hypertension (58.33%), cardiovascular disease (22.22%) and diabetes (19.44%). Typical clinical feature measured in these patients includes lymphopenia and elevated markers of inflammation.

12.32.3 Limitations

As noted by the authors, the conclusions of this study are very limited because this is single-centered study focusing on a small cohort of patients who did not survive. Many clinical parameters observed by the authors (such* as increase levels of serum CRP, PCT, IL-6) have also been described in other COVID19 patients who survived the infection

12.32.4 Significance

This study is essentially descriptive and may be useful for clinical teams monitoring COVID19 patients.

12.32.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.33 Risk Factors Related to Hepatic Injury in Patients with Corona Virus Disease 2019

[1993]

12.33.1 Keywords

- COVID-related Hepatic Injury

12.33.2 Main Findings

Based on a retrospective study of 85 hospitalized COVID patients in a Beijing hospital, authors showed that patients with elevated ALT levels ($n = 33$) were characterized by significantly higher levels of lactic acid and CRP as well as lymphopenia and hypoalbuminemia compared to their counterparts with normal ALT levels. Proportion of severe and critical patients in the ALT elevation group was significantly higher than that of normal ALT group. Multivariate logistic regression performed on clinical factors related to ALT elevation showed that $CRP \geq 20\text{mg/L}$ and low lymphocyte count ($<1.1*10^9 \text{ cells/L}$) were independently related to ALT elevation—a finding that led the authors to suggest cytokine storm as a major mechanism of liver damage.

12.33.3 Limitations

The article's most attractive claim that liver damage seen in COVID patients is caused by cytokine storm (rather than direct infection of the liver) hinges solely on their multivariate regression analysis. Without further mechanistic studies a) demonstrating how high levels of inflammatory cytokines can induce liver damage and b) contrasting types of liver damage incurred by direct infection of the liver vs. system-wide elevation of inflammatory cytokines, their claim remains thin. It is also worth noting that six of their elevated ALT group ($n=33$) had a history of liver disease (i.e. HBV infection, alcoholic liver disease, fatty liver) which can confound their effort to pin down the cause of hepatic injury to COVID.

12.33.4 Significance

Limited. This article confirms a rich body of literature describing liver damage and lymphopenia in COVID patients.

12.33.5 Credit

12.34 Detectable serum SARS-CoV-2 viral load (RNAaemia) is closely associated with drastically elevated interleukin 6 (IL-6) level in critically ill COVID-19 patients

[[1994](#)]

12.34.1 Keywords

- ARDS
- interleukin-6 (IL-6)
- procalcitonin (PCT)
- pro-inflammatory cytokines
- SARS-CoV-2 RNAaemia

12.34.2 Main Findings

48 adult patients diagnosed with Covid19 according to Chinese guidelines for Covid19 diagnosis and treatment version 6 were included in this study. Patients were further sub-divided into three groups based on clinical symptoms and disease severity: (1) mild, positive Covid19 qPCR with no or mild clinical symptoms (fever; respiratory; radiological abnormalities); (2) severe, at least one of the following: shortness of breath/respiratory rate >30/min, oxygen saturation $\text{SaO}_2 < 93\%$, Horowitz index $\text{paO}_2/\text{FiO}_2 < 300 \text{ mmHg}$ (indicating moderate pulmonary damage); and (3) critically ill, at least one additional complicating factor: respiratory failure with need for mechanical ventilation; systemic shock; multi-organ failure and transfer to ICU. Serum samples and throat-swabs were collected from all 48 patients enrolled. SARS-CoV-2 RNA was assessed by qPCR with positive results being defined as Ct values < 40, and serum interleukin-6 (IL-6) was quantified using a commercially available detection kit. Briefly, patient characteristics in this study confirm previous reports suggesting that higher age and comorbidities are significant risk factors of clinical severity. Of note, 5 out of 48 of patients (10.41%), all in the critically ill category, were found to have detectable serum SARS-CoV-2 RNA levels, so-called RNAaemia. Moreover, serum IL-6 levels in these patients were found to be substantially higher and this correlated with the presence of detectable SARS-CoV-2 RNA levels. The authors hypothesize that viral RNA might be released from acutely damaged tissues in moribund patients during the course of Covid19 and that RNAaemia along with IL-6 could potentially be used as a prognostic marker.

12.34.3 Limitations

While this group's report generally confirms some of the major findings of a more extensive study, published in early February 2020, [[1984](#)], there are limitations that should be taken into account. First, the number of patients enrolled is relatively small; second, interpretation of these data would benefit

from inclusion of information about study specifics as well as providing relevant data on the clinical course of these patients other than the fact that some were admitted to ICU (i.e. demographics on how many patients needed respiratory support, dialysis, APACHE II/III or other standard ICU scores as robust prognostic markers for mortality etc). It also remains unclear at which time point the serum samples were taken, i.e. whether at admission, when the diagnosis was made or during the course of the hospital stay (and potentially after onset of therapy, which could have affected both IL-6 and RNA levels). The methods section lacks important information on the qPCR protocol employed, including primers and cycling conditions used. From a technical point of view, Ct values >35 seem somewhat non-specific (although Ct <40 was defined as the CDC cutoff as well) indicating that serum RNA levels are probably very low, therefore stressing the need for highly specific primers and high qPCR efficiency. In addition, the statistical tests used (t-tests, according to the methods section) do not seem appropriate as the organ-specific data such as BUN and troponin T values seem to be not normally distributed across groups (n= 5 RNAemia+ vs. n= 43 RNAemia-). Given the range of standard deviations and the differences in patient sample size, it is difficult to believe that these data are statistically significantly different.

12.34.4 Significance

This study is very rudimentary and lacks a lot of relevant clinical details. However, it corroborates some previously published observations regarding RNAemia and IL-6 by another group. Generally, regarding future studies, it would be important to address the question of IL-6 and other inflammatory cytokine dynamics in relation to Covid19 disease kinetics (high levels of IL-6, IL-8 and plasma leukotriene were shown to have prognostic value at the onset of ARDS ; serum IL-2 and IL-15 have been associated with mortality; reviewed by Chen W & Ware L, Clin Transl Med. 2015 [[1995](#)]).

12.34.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.35 Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study

[[1996](#)]

12.35.1 Keywords

- Lymphopenia

12.35.2 Main Findings

Based on a retrospective study of 162 COVID patients from a local hospital in Wuhan, China, the authors show an inverse correlation between lymphocyte % (LYM%) of patients and their disease severity. The authors have also tracked LYM% of 70 cases (15 deaths; 15 severe; 40 moderate) throughout the disease progression with fatal cases showing no recovery of lymphocytes (<5%) even after 17-19 days post-onset. The temporal data of LYM % in COVID patients was used to construct a Time-Lymphocyte% model which is used to categorize and predict patients' disease severity and progression. The model was validated using 92 hospitalized cases and kappa statistic test was used to assess agreement between predicted disease severity and the assigned clinical severity ($k = 0.49$).

12.35.3 Limitations

Time-Lymphocyte % Model (TLM) that authors have proposed as a predictive model for clinical severity is very simple in its construction and derives from correlative data of 162 patients. In order for the model to be of use, it needs validation using a far more robust data set and possibly a mechanistic study on how COVID leads to lymphopenia in the first place. In addition, it should be noted that no statistical test assessing significance of LYM % values between disease severities was performed.

12.35.4 Significance

This article is of limited significance as it simply reports similar descriptions of COVID patients made in previous literature that severe cases are characterized by lymphopenia.

12.35.5 Credit

Review by Chang Moon as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.36 The potential role of IL-6 in monitoring severe case of coronavirus disease 2019

Liu et al. *medRxiv*. [1997]

12.36.1 Keywords

- Cytokine Release Syndrome
- lymphocytopenia
- IL-6
- CRP
- COVID19
- pneumonia

12.36.2 Main Findings

Study on blood biomarkers on 80 COVID19 patients (69 severe and 11 non-severe). Patients with severe symptoms at admission (baseline) showed obvious lymphocytopenia and significantly increased interleukin-6 (IL-6) and CRP, which was positively correlated with symptoms severity. IL-6 at baseline positively correlates with CRP, LDH, ferritin and D-Dimer abundance in blood.

Longitudinal analysis of 30 patients (before and after treatment) showed significant reduction of IL-6 in remission cases.

12.36.3 Limitations

Limited sample size at baseline, especially for the non-severe leads to question on representativeness. The longitudinal study method is not described in detail and suffers from non-standardized treatment. Limited panel of pro-inflammatory cytokine was analyzed. Patients with severe disease show a wide range of altered blood composition and biomarkers of inflammation, as well as differences in disease course (53.6% were cured, about 10% developed acute respiratory distress syndrome). The authors comment on associations between IL-6 levels and outcomes, but these were not statistically significant (maybe due to the number of patients, non-standardized treatments, etc.) and data is not shown. Prognostic biomarkers could have been better explored. Study lacks multivariate analysis.

12.36.4 Significance

IL-6 could be used as a pharmacodynamic marker of disease severity. Cytokine Release Syndrome (CRS) is a well-known side effect for CAR-T cancer therapy and there are several effective drugs to manage CRS. Drugs used to manage CRS could be tested to treat the most severe cases of COVID19.

12.36.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.37 Clinical and Laboratory Profiles of 75 Hospitalized Patients with Novel Coronavirus Disease 2019 in Hefei, China

Zhao et al. *medRxiv*. [[1998](#)]

12.37.1 Keywords

- Routine laboratory testing

12.37.2 Main Findings

The authors of this study provide a comprehensive analysis of clinical laboratory assessments in 75 patients (median age 47 year old) hospitalized for Corona virus infection in China measuring differential blood counts including T-cell subsets (CD4, CD8), coagulation function, basic blood chemistry, of infection-related biomarkers including CRP, Procalcitonin (PCT) (Precursor of calcitonin that increases during bacterial infection or tissue injury), IL-6 and erythrocyte sedimentation rate as well as clinical parameters. Among the most common hematological changes they found increased neutrophils, reduced CD4 and CD8 lymphocytes, increased LDH, CRP and PCT

When looking at patients with elevated IL-6, the authors describe significantly reduced CD4 and CD8 lymphocyte counts and elevated CRP and PCT levels were significantly increased in infected patients suggesting that increased IL-6 may correlate well with disease severity in COVID-19 infections

12.37.3 Limitations

The authors performed an early assessment of clinical standard parameters in patients infected with COVID-19. Overall, the number of cases (75) is rather low and the snapshot approach does not inform about dynamics and thus potential relevance in the assessment of treatment options in this group of patients.

12.37.4 Significance

The article summarizes provides a good summary of some of the common changes in immune cells inflammatory cytokines in patients with a COVID-19 infection and. Understanding how these changes can help predict severity of disease and guide therapy including IL-6 cytokine receptor blockade using Tocilizumab or Sarilumab will be important to explore.

12.37.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.38 Exuberant elevation of IP-10, MCP-3 and IL-1ra during SARS-CoV-2 infection is associated with disease severity and fatal outcome

Yang et al. *medRxiv* [[1999](#)]

12.38.1 Keywords

- cytokine
- IP-10
- MCP-3
- IL-1Ra
- lymphocyte

- neutrophil
- stratification
- disease severity
- viral load
- lung function
- complications
- clinical data

12.38.2 Summary

Plasma cytokine analysis (48 cytokines) was performed on COVID-19 patient plasma samples, who were sub-stratified as severe (N=34), moderate (N=19), and compared to healthy controls (N=8). Patients were monitored for up to 24 days after illness onset: viral load (qRT-PCR), cytokine (multiplex on subset of patients), lab tests, and epidemiological/clinical characteristics of patients were reported.

12.38.3 Main Findings

- Many elevated cytokines with COVID-19 onset compared to healthy controls (IFNy, IL-1Ra, IL-2Ra, IL-6, IL-10, IL-18, HGF, MCP-3, MIG, M-CSF, G-CSF, MIG-1a, and IP-10).
- IP-10, IL-1Ra, and MCP-3 (esp. together) were associated with disease severity and fatal outcome.
- IP-10 was correlated to patient viral load ($r=0.3006$, $p=0.0075$).
- IP-10, IL-1Ra, and MCP-3 were correlated to loss of lung function ($\text{PaO}_2/\text{FaO}_2$ (arterial/atmospheric O₂) and Murray Score (lung injury) with MCP-3 being the most correlated ($r=0.4104$ $p<0.0001$ and $r=0.5107$ $p<0.0001$ respectively).
- Viral load (Lower Ct Value from qRT-PCR) was associated with upregulated IP-10 only (not IL-1Ra or MCP-3) and was mildly correlated with decreased lung function: $\text{PaO}_2/\text{FaO}_2$ (arterial/atmospheric O₂) and Murray Score (lung injury).
- Lymphopenia (decreased CD4 and CD8 T cells) and increased neutrophil correlated w/ severe patients.
- Complications were associated with COVID severity (ARDS, hepatic insufficiency, renal insufficiency).

12.38.4 Limitations

Collection time of clinical data and lab results not reported directly (likely 4 days (2,6) after illness onset), making it very difficult to determine if cytokines were predictive of patient outcome or reflective of patient compensatory immune response (likely the latter). Small N for cytokine analysis (N=2 fatal and N=5 severe/critical, and N=7 moderate or discharged). Viral treatment strategy not clearly outlined.

12.38.5 Expanded Results

NOTE: Moderate COVID-19 was classified by fever, respiratory manifestations, and radiological findings consistent with pneumonia while severe patients had one or more of the following: 1) respiratory distress, resting O₂ saturation, or 3) arterial PaO₂/FiO₂ < 300 mmHg.

Cytokine Results (Human Cytokine Screening Panel, Bio-Rad):

- **Significant elevation of cytokines observed in COVID patients compared to healthy controls: IFNy, IL-1Ra, IL-2Ra, IL-6, IL-10, IL-18, HGF, MCP-3, MIG, M-CSF, G-CSF, MIP-1a, and IP-10.**
- Severity was correlated **with increase in measured IP-10, MCP-3, and IL-Ra** as measured by area under the curve analysis during sample timecourse (2-24 days after illness onset).
- IL-1Ra incr. significant 0-7 days after onset, MCP-3 signif. upregulated throughout observation timecourse, and IP-10 increased and upregulated throughout (trending downwards over time).
- **The three cytokines together (IP-10, IL-1Ra, and MCP-3 AUC) served as the best predictors of disease deterioration and fatal outcome.**
- No significant differences between moderate/severe observed between groups in IL-2Ra, IL-6, IL-10, IL-18, CTACK, G-CSF, HGF, M-CSF, MIP-1a, MIG, and IFNy at any timepoints.
- **Viral load (Lower Ct Value from qRT-PCR) was associated with upregulated IP-10 only (not IL-1Ra or MCP-3) and was highly correlated with decreased lung function: PaO₂/FaO₂ (arterial/atmospheric O₂) and Murray Score (lung injury).**
- **Antibodies against these cytokines (esp. anti-IP-10) may serve as a potential treatment for amelioration of COVID-19 (and associated ARDS).**

Lab results:

- **Decreased lymphocytes (%) in all patients – lymphopenia corr. w/ severe patients**
 - **Decreased CD4 and CD8 T cells** – no monocyte or eosino/basophil % measured
- **Increased neutrophils (%)**
- Increased BUN (mmol/L) – other kidney markers, liver markers, and LDH were not significantly different between groups and were not compared to healthy controls.

Clinical features (between moderate vs. severe patient groups):

- Complications were associated with severity (ARDS, hepatic insufficiency, renal insufficiency).
- Coexisting conditions between groups were not significantly different (chronic heart/lung/renal/liver disease, diabetes, or cancer) and patient time courses (onset to admission and onset to viral tx) also not significantly different – 4 days (2, 6) on average for admission and 4 (3,7) for antiviral.
- Increased corticosteroids and mechanical/ invasive mechanical ventilation in severe patients.
- Increased median age in severe group (Median (Range = 63.5 (42-74) vs. 51 (22-78)) and patients >60 yrs had higher ratio of severe patients as compared patients 16-59 yrs.
- Higher incidence of fever in severe patients (91.2 vs. 68.4%), myalgia (57.7 vs. 48.1%), and chill (17.6% vs. 0%).
- No differences in cough, headache, nausea/vomiting, or diarrhea.

12.38.6 Significance

Outline of pathological time course (implicating innate immunity esp.) and identification key cytokines associated with disease severity and prognosis (+ comorbidities). Anti-IP-10 as a possible therapeutic intervention (ex: Eldelumab).

12.38.7 Credit

Review by Natalie Vaninov as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.39 Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019

Zhao Jr. et al. *medRxiv*. [[2000](#)]

12.39.1 Keywords

- SARS-CoV-2 IgG
- seroconversion rate
- total Ab
- Ig
- IgM

12.39.2 Main Findings

This study examined antibody responses in the blood of COVID-19 patients during the early SARS CoV2 outbreak in China. Total 535 plasma samples were collected from 173 patients (51.4% female) and were tested for seroconversion rate using ELISA. Authors also compared the sensitivity of RNA and antibody tests over the course of the disease . The key findings are:

- Among 173 patients, the seroconversion rates for total antibody (Ab), IgM and IgG were 93.1% (161/173), 82.7% (143/173) and 64.7% (112/173), respectively.
- The seroconversion sequentially appeared for Ab, IgM and then IgG, with a median time of 11, 12 and 14 days, respectively. Overall, the seroconversion of Ab was significantly quicker than that of IgM ($p = 0.012$) and IgG ($p < 0.001$). Comparisons of seroconversion rates between critical and non-critical patients did not reveal any significant differences.
- RNA tests had higher sensitivity in early phase and within 7 days of disease onset than antibody assays (66.7% Vs 38.3% respectively).
- The sensitivity of the Ab assays was higher 8 days after disease onset, reached 90% at day 13 and 100% at later time points (15-39 days). In contrast, RNA was only detectable in 45.5% of samples at days 15-39.
- In patients with undetectable RNA in nasal samples collected during day 1-3, day 4-7, day 8-14 and day 15-39 since disease onset, 28.6% (2/7), 53.6% (15/28), 98.2% (56/57) and 100% (30/30) had detectable total Ab titers respectively Combining RNA and antibody tests significantly raised the sensitivity for detecting COVID-19 patients in different stages of the disease ($p < 0.001$).
- There was a strong positive correlation between clinical severity and antibody titer 2-weeks after illness onset.
- Dynamic profiling of viral RNA and antibodies in representative COVID-19 patients ($n=9$) since onset of disease revealed that antibodies may not be sufficient to clear the virus. It should be noted that increases in of antibody titers were not always accompanied by RNA clearance.

12.39.3 Limitations

Because different types of ELISA assays were used for determining antibody concentrations at different time points after disease onset, sequential seroconversion of total Ab, IgM and IgG may not represent actual temporal differences but rather differences in the affinities of the assays used. Also, due to the lack of blood samples collected from patients in the later stage of illness, how long the antibodies could last remain unknown. For investigative dynamics of antibodies, more samples were required.

12.39.4 Significance

Total and IgG antibody titers could be used to understand the epidemiology of SARS CoV-2 infection and to assist in determining the level of humoral immune response in patients.

The findings provide strong clinical evidence for routine serological and RNA testing in the diagnosis and clinical management of COVID-19 patients. The understanding of antibody responses and their half-life during and after SARS CoV2 infection is important and warrants further investigations.

12.39.5 Credit

This review was undertaken by Zafar Mahmood and edited by K Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.40 Restoration of leukomonocyte counts is associated with viral clearance in COVID-19 hospitalized patients

Chen et al. *medRxiv* [[2001](#)]

12.40.1 Keywords

- COVID-19
- T cell
- B cell
- NK cell
- IL-6
- pro-calcitonin
- cytokine storm

12.40.2 Main Findings

The authors collected data on 25 COVID-19 patients (n=11 men, n=14 women) using standard laboratory tests and flow cytometry. All patients were treated with antibiotics. Twenty-four of the 25 patients were also treated with anti-viral Umefinovir and 14 of the patients were treated with corticosteroids. 14 patients became negative for the virus after 8-14 days of treatment. The same treatment course was extended to 15-23 days for patients who were still positive for the virus at day 14.

The authors found a negative association between age and resolution of infection. Patients with hypertension, diabetes, malignancy or chronic liver disease were all unable to clear the virus at day 14, though not statistically significant.

Elevated procalcitonin and a trend for increased IL-6 were also found in peripheral blood prior to the treatment.

A trend for lower NK cell, T cell and B cell counts in patients was also reported. B cell, CD4 and CD8 T cell counts were only increased upon treatment in patients who cleared the virus. NK cell frequencies remained unchanged after treatment in all the patients.

12.40.3 Limitations

73% of the patients who remained positive for SARS-CoV2 after the 1st treatment, and 43% of all patients who cleared the virus were treated with corticosteroids. Corticosteroids have strong effects on the immune compartment in blood [2002]. The authors should have accounted for corticosteroid treatment when considering changes in T, NK and B cell frequencies.

Assessing if IL-6 concentrations were back to baseline levels following treatment would have provided insights into the COVID-19 cytokine storm biology. Patients with higher baseline levels of IL-6 have been reported to have lower CD8 and CD4 T cell frequencies [1998]. Correlating IL-6 with cell counts before and after treatment would thus have also been of interest. The report of the laboratory measures in table 2 is incomplete and should include the frequencies of patients with increased/decreased levels for each parameter.

Correction is needed for the 1st paragraph of the discussion as data does not support NK cell restoration upon treatment in patients who cleared the virus. NK cells remain unchanged after the 1st treatment course and only seem to increase in 2 out of 6 donors after the 2nd treatment course in those patients.

12.40.4 Significance

Previous reports suggest an association between disease severity and elevated IL-6 or pro-calcitonin concentrations in COVID-19 patients [1994,2003]. IL-6 receptor blockade is also being administered to patients enrolled in clinical trials (NCT04317092). This report thus contributes to highlight elevated concentrations of these analytes in COVID-19 patients. Mechanisms underlying the association between viral clearance and restoration of the T cell and B cell frequencies suggests viral-driven immune dysregulation, which needs to be investigated in further studies.

12.40.5 Credit

Review by Bérengère Salomé as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.41 Clinical findings in critically ill patients infected with SARS-CoV-2 in Guangdong Province, China: a multi-center, retrospective, observational study

12.41.1 Keywords

- clinical outcomes
- prognosis
- critically ill patients
- ICU
- lymphopenia
- LDH

12.41.2 Main Findings:

This work analyses laboratory and clinical data from 45 patients treated in the in ICU in a single province in China. Overall, 44% of the patients were intubated within 3 days of ICU admission with only 1 death.

Lymphopenia was noted in 91% of patient with an inverse correlation with LDH.

Lymphocyte levels are negatively correlated with Sequential Organ Failure Assessment (SOFA) score (clinical score, the higher the more critical state), LDH levels are positively correlated to SOFA score. Overall, older patients (>60yo), with high SOFA score, high LDH levels and low lymphocytes levels at ICU admission are at higher risk of intubation.

Of note, convalescent plasma was administered to 6 patients but due to limited sample size no conclusion can be made.

12.41.3 Limitations

While the study offers important insights into disease course and clinical lab correlates of outcome, the cohort is relatively small and is likely skewed towards a less-severe population compared to other ICU reports given the outcomes observed. Analysis of laboratory values and predictors of outcomes in larger cohorts will be important to make triage and treatment decisions. As with many retrospective analyses, pre-infection data is limited and thus it is not possible to understand whether lymphopenia was secondary to underlying comorbidities or infection.

Well-designed studies are necessary to evaluate the effect of convalescent plasma administration.

12.41.4 Significance

This clinical data enables the identification of at-risk patients and gives guidance for research for treatment options. Indeed, further work is needed to better understand the causes of the lymphopenia and its correlation with outcome.

12.41.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.42 Immune Multi-epitope vaccine design using an immunoinformatics approach for 2019 novel coronavirus in China (SARS-CoV-2)

[2005]

12.42.1 Keywords

- Vaccine
- in silico
- B cell epitopes
- T cell epitopes

12.42.2 Main Findings

Using in silico bioinformatic tools, this study identified putative antigenic B-cell epitopes and HLA restricted T-cell epitopes from the spike, envelope and membrane proteins of SARS-CoV-2, based on the genome sequence available on the NCBI database. T cell epitopes were selected based on predicted affinity for the more common HLA-I alleles in the Chinese population.

Subsequently, the authors designed vaccine peptides by bridging selected B-cell epitopes and adjacent T-cell epitopes. Vaccine peptides containing only T-cell epitopes were also generated.

From 61 predicted B-cell epitopes, only 19 were exposed on the surface of the virion and had a high antigenicity score. A total of 499 T-cell epitopes were predicted. Based on the 19 B-cell epitopes and their 121 adjacent T-cell epitopes, 17 candidate vaccine peptides were designed. Additionally, another 102 vaccine peptides containing T-cell epitopes only were generated. Based on the epitope counts and HLA score, 13 of those were selected. Thus, a total of 30 peptide vaccine candidates were designed.

12.42.3 Limitations

While this study provides candidates for the development of vaccines against SARS-CoV-2, in vitro and in vivo trials are required to validate the immunogenicity of the selected B and T cell epitopes. This could be done using serum and cells from CoV-2-exposed individuals, and in preclinical studies. The implication of this study for the current epidemic are thus limited. Nevertheless, further research on this field is greatly needed.

12.42.4 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.43 Clinical Features of Patients Infected with the 2019 Novel Coronavirus (COVID-19) in Shanghai, China

Cao et al. *medRxiv* [2006]

12.43.1 Keywords

- Disease severity
- clinical features
- laboratory abnormalities

12.43.2 Main Findings

This single-center cohort study analyzes the clinical and laboratory features of 198 patients with confirmed COVID-19 infection in Shanghai, China and correlated these parameters with clinical disease severity, including subsequent intensive care unit (ICU) admission. 19 cases (9.5%) required ICU admission after developing respiratory failure or organ dysfunction. Age, male sex, underlying cardiovascular disease, and high symptom severity (high fever, dyspnea) were all significantly correlated with ICU admission. Additionally, ICU admission was more common in patients who presented with lymphopenia and elevated neutrophil counts, among other laboratory abnormalities. Flow cytometric analysis revealed that patients admitted to the ICU had significantly reduced circulating CD3+ T cell, CD4+ T cell, CD8+ T cell, and CD45+ leukocyte populations compared to the cohort of patients not requiring ICU admission.

12.43.3 Limitations

The limitations of this study include the relatively small sample size and lack of longitudinal testing. The authors also did not assess whether respiratory comorbidity – such as asthma or chronic obstructive lung disease – in addition to immunosuppression affected ICU admission likelihood.

12.43.4 Significance

COVID-19 has already sickened thousands across the globe, though the severity of these infections is markedly diverse, ranging from mild symptoms to respiratory failure requiring maximal intervention. Understanding what clinical, laboratory, and immunologic factors predict the clinical course of COVID-19 infection permits frontline providers to distribute limited medical resources more effectively.

12.43.5 Credit

Review by Andrew Charap as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine at Mount Sinai.

12.44 Serological detection of 2019-nCoV respond to the epidemic: A useful complement to nucleic acid testing

Zhang et al. *medRxiv*. [2007]

12.44.1 Keywords

- Immunoassay
- serum IgM and IgG
- specific antibodies

12.44.2 Main Finding

This study showed that both anti-2019-nCov IgM and IgG were detected by automated chemiluminescent immunoassay in the patients who had been already confirmed as positive by nucleic acid detection, while single positivity of IgM or IgG were detected in a very few cases in the other population including 225 non-COVID-19 cases. In addition to the increase of anti-2019-nCov IgM 7-12 days after morbidity, the increase of IgG was detected in three patients with COVID-19 within a very short of time (0-1 day).

12.44.3 Limitations

The limitation of this study is only 3 confirmed COVID-19 cases were included, so that the relationship between anti-2019-nCov antibodies and disease progression might not be clearly defined. Another limitation is that they did not show the course of 2019-nCov specific antibodies in the cases with positive for COVID-19 but without clinical symptoms.

12.44.4 Significance

The detection of anti-2019-nCov antibodies can be an alternative method to diagnose and treat COVID-19 more comprehensively by distinguish non COVID-19 patients. It may be helpful to understand the course of individual cases with COVID-19 to predict the prognosis if more cases will be evaluated.

12.44.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.45 Human Kidney is a Target for Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection

[2008]

12.45.1 Keywords

- Kidney/Renal Failure
- Macrophage Infiltration
- Complement Activation

12.45.2 Main Finding

Analyzing the eGFR (effective glomerular flow rate) of 85 Covid-19 patients and characterizing tissue damage and viral presence in post-mortem kidney samples from 6 Covid-19 patients, the authors conclude that significant damage occurs to the kidney, following Covid-19 infection. This is in contrast to the SARS infection from the 2003 outbreak. They determine this damage to be more prevalent in patients older than 60 years old, as determined by analysis of eGFR. H&E and IHC analysis in 6 Covid-19 patients revealed that damage was in the tubules, not the glomeruli of the kidneys and suggested that macrophage accumulation and C5b-9 deposition are key to this process.

12.45.3 Limitations

Severe limitations include that the H&E and IHC samples were performed on post-mortem samples of unknown age, thus we cannot assess how/if age correlates with kidney damage, upon Covid-19 infection. Additionally, eGFR was the only *in-vivo* measurement. Blood urea nitrogen and proteinuria are amongst other measurements that could have been obtained from patient records. An immune panel of the blood was not performed to assess immune system activation. Additionally, patients are only from one hospital.

12.45.4 Significance

This report makes clear that kidney damage is prevalent in Covid-19 patients and should be accounted for.

12.45.5 Credit

Review by Dan Fu Ruan, Evan Cody and Venu Pothula as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine at Mount Sinai.

12.46 COVID-19 early warning score: a multi-parameter screening tool to identify highly suspected patients

Song et al. *medRxiv*. [[2009](#)]

12.46.1 Keywords

- retrospective
- electronic health records
- blood counts

- diagnostic
- prognostic
- modeling

12.46.2 Main Findings

The aim of this study was to identify diagnostic or prognostic criteria which could identify patients with COVID-19 and predict patients who would go on to develop severe respiratory disease. The authors use EMR data from individuals taking a COVID-19 test at Zhejiang hospital, China in late January/Early February. A large number of clinical parameters were different between individuals with COVID-19 and also between 'severe' and 'non-severe' infections and the authors combine these into a multivariate linear model to derive a weighted score, presumably intended for clinical use.

12.46.3 Limitations

Unfortunately, the paper is lacking a lot of crucial information, making it impossible to determine the importance or relevance of the findings. Most importantly, the timings of the clinical measurements are not described relative to the disease course, so it is unclear if the differences between 'severe' and 'non-severe' infections are occurring before progression to severe disease (which would make them useful prognostic markers), or after (which would not).

12.46.4 Significance

This paper is one of many retrospective studies coming from hospitals in China studying individuals with COVID-19. Because of the sparse description of the study design, this paper offers little new information. However, studies like this could be very valuable and we would strongly encourage the authors to revise this manuscript to include more information about the timeline of clinical measurements in relation to disease onset and more details of patient outcomes.

12.46.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.47 LY6E impairs coronavirus fusion and confers immune control of viral disease

Pfaender et al. *bioRxiv*. [2010]

12.47.1 Keywords

- interferon-stimulated genes
- antiviral interferons

- human coronaviruses (CoV)
- murine hepatitis virus (MHV)

12.47.2 Main Findings

Screening a cDNA library of >350 human interferon-stimulated genes for antiviral activity against endemic human coronavirus HCoV-229E (associated with the common cold), Pfaender S & Mar K *et al.* identify lymphocyte antigen 6 complex, locus E (Ly6E) as an inhibitor of cellular infection of Huh7 cells, a human hepatoma cell line susceptible to HCoV-229E and other coronaviruses. In a series of consecutive *in vitro* experiments including both stable Ly6E overexpression and CRISPR-Cas9-mediated knockout the authors further demonstrate that Ly6E reduces cellular infection by various other coronaviruses including human SARS-CoV and SARS-CoV-2 as well as murine CoV mouse hepatitis virus (MHV). Their experiments suggest that this effect is dependent on Ly6E inhibition of CoV strain-specific spike protein-mediated membrane fusion required for viral cell entry.

To address the function of Ly6E *in vivo*, hematopoietic stem cell-specific Ly6E knock-out mice were generated by breeding Ly6E^{f/f} mice (referred to as functional wild-type mice) with transgenic *Vav-iCre* mice (offspring referred to as Ly6E HSC ko mice); wild-type and Ly6E HSC ko mice of both sexes were infected intraperitoneally with varying doses of the natural murine coronavirus MHV, generally causing a wide range of diseases in mice including hepatitis, enteritis and encephalomyelitis. Briefly, compared to wild-type controls, mice lacking hematopoietic cell-expressed Ly6E were found to present with a more severe disease phenotype as based on serum ALT levels (prognostic of liver damage), liver histopathology, and viral titers in the spleen. Moreover, bulk RNAseq analysis of infected liver and spleen tissues indicated changes in gene expression pathways related to tissue damage and antiviral immune responses as well as a reduction of genes associated with type I IFN response and inflammation. Finally, the authors report substantial differences in the numbers of hepatic and splenic APC subsets between wild-type and knockout mice following MHV infection and show that Ly6E-deficient B cells and to a lesser extent also DCs are particularly susceptible to MHV infection *in vitro*.

12.47.3 Limitations

Experiments and data in this study are presented in an overall logical and coherent fashion; however, some observations and the conclusions drawn are problematic and should be further addressed & discussed by the authors. Methodological & formal limitations include relatively low replicate numbers as well as missing technical replicates for some *in vitro* experiments (*cf.* Fig. legend 1; Fig. legend 2e); the omission of “outliers” in Fig. legend 2 without an apparent rationale as to why this approach was chosen; the lack of detection of actual Ly6E protein levels in Ly6E HSC ko or wild-type mice; and most importantly, missing information on RNAseq data collection & analysis in the method section and throughout the paper. A more relevant concern though is that the interpretation of the experimental data presented and the language used tend to overrate and at times overgeneralize findings: for example, while the authors demonstrate statistically significant, Ly6E-mediated reduction of coronavirus titers in stable cells lines *in vitro*, it

remains unclear whether a viral titer reduction by one log decade would be of actual biological relevance in face of high viral titers *in vivo*. After high-dose intraperitoneal MHV infection *in vivo*, early viral titers in Ly6E HSC knockout vs. wt mice only showed an elevation in the spleen (~1.5 log decades) but not liver of the ko mice (other tissue not evaluated), and while ko mice presented with only modestly increased liver pathology, both male and female ko mice exhibited significantly higher mortality. Thus, the manuscript tile statement that “Ly6E ... confers immune control of viral disease” is supported by only limited *in vivo* data, and gain-of-function experiments (eg. Ly6E overexpression) were not performed. Of additional note here, tissue tropism and virulence differ greatly among various MHV strains and isolates whereas dose, route of infection, age, genetic background and sex of the mice used may additionally affect disease outcome and phenotype (*cf.* Taguchi F & Hirai-Yuki A, <https://doi.org/10.3389/fmicb.2012.00068>; Kanolkhar A et al, <https://jvi.asm.org/content/ 83/18/9258>). Observations attributed to hematopoietic stem cell-specific Ly6E deletion could therefore be influenced by the different genetic backgrounds of floxed and cre mice used, and although it appears that littermates wt and ko littermates were used in the experiments, the potentially decisive impact of strain differences should at least have been discussed. Along these lines, it should also be taken into account that the majority of human coronaviruses cause respiratory symptoms, which follow a different clinical course engaging other primary cellular mediators than the hepatotropic murine MHV disease studied here. It therefore remains highly speculative how the findings reported in this study will translate to human disease and it would therefore be important to test other routes of MHV infection and doses that have been described to produce a more comparable phenotype to human coronavirus disease (*cf.* Kanolkhar A et al, <https://jvi.asm.org/content/ 83/18/9258>). Another important shortcoming of this study is the lack of any information on functional deficits or changes in Ly6E-deficient immune cells and how this might relate to the phenotype observed. Overall, the *in vitro* experiments are more convincing than the *in vivo* studies which appear somewhat limited.

12.47.4 Significance

Despite some shortcomings, the experiments performed in this study suggest a novel and somewhat unexpected role of Ly6E in the protection against coronaviruses across species. These findings are of relevance and should be further explored in ongoing research on potential coronavirus therapies. Yet an important caveat pertains to the authors' suggestion that “therapeutic mimicking of Ly6E action” may constitute a first line of defense against novel coronaviruses since their own prior work demonstrated that Ly6E can enhance rather than curtail infection with influenza A and other viruses.

12.47.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.48 A preliminary study on serological assay for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in 238 admitted hospital patients

Liu et al. *medRxiv*. [2011]

12.48.1 Keywords

- diagnosis
- serological assay
- ELISA
- RT-PCR

12.48.2 Main Findings

While RT-PCR is being used currently to routinely diagnose infection with SARS-CoV-2, there are significant limitations to the use of a nucleic acid test that lead to a high false-negative rate. This article describes ELISAs that can measure IgM and IgG antibodies against the N protein of SARS-CoV-2 to test samples from 238 patients (153 positive by RT-PCR and 85 negative by RT-PCR) at different times after symptom onset. The positivity rate of the IgM and/or IgG ELISAs was greater than that of the RT-PCR (81.5% compared to 64.3%) with similar positive rates in the confirmed and suspected cases (83% and 78.8%, respectively), suggesting that many of the suspected but RT-PCR-negative cases were also infected. The authors also found that the ELISAs have higher positive rates later after symptom onset while RT-PCR is more effective as a diagnostic test early during the infection.

12.48.3 Limitations

I cannot identify any limitations to this study.

12.48.4 Significance

The authors make a strong case for using a combination of ELISA and RT-PCR for diagnosis of infection with SARS-CoV-2, especially considering the dynamics of positivity rates of RT-PCR and ELISA. Fewer false-negative diagnoses would improve infection control and patient management.

12.48.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.49 Monoclonal antibodies for the S2 subunit of spike of SARS-CoV cross-react with the newly-emerged SARS-CoV-2

[2012]

12.49.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- Spike protein
- Cross- reactive antibodies

12.49.2 Main Findings

Whole genome sequencing-based comparisons of the 2003 Severe Acute Respiratory Syndrome Coronavirus (SARS-CoV) and the 2019 SARS-CoV-2 revealed conserved receptor binding domain (RBD) and host cell receptor, angiotensin-converting enzyme 2 (ACE2). In line with this, the authors tested cross-reactivity of murine monoclonal antibodies (mAbs) previously generated against the SARS-CoV spike (S) glycoprotein involved in viral entry. One of the screened mAb, 1A9, was able to bind and cross-neutralize multiple strains of SARS-CoV, as well as, detect the S protein in SARS-CoV-2-infected cells. mAb 1A9 was generated using an immunogenic fragment in the S2 subunit of SARS-CoV and binds through a novel epitope within the S2 subunit at amino acids 1111-1130. It is important to note that CD8+ T lymphocyte epitopes overlap with these residues, suggesting that S2 subunit could be involved in inducing both, humoral and cell-mediated immunity.

12.49.3 Limitations

The authors used previously generated mouse mAbs against the S protein in SARS-CoV expressed in mammalian cell line. Future experimental validation using COVID-19 patient samples is needed to validate these findings. In addition, the results of these studies are predominantly based on in vitro experiments and so, evaluating the effects of the mAb 1A9 in an animal model infected with this virus will help us better understand the host immune responses in COVID-19 and potential therapeutic vaccines.

12.49.4 Significance

This study identified mAbs that recognize the new coronavirus, SARS-CoV-2. These cross-reactive mAbs will help in developing diagnostic assays for COVID-19.

12.49.5 Credit

This review was undertaken by Tamar Plitt and Katherine Lindblad as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.50 Mortality of COVID-19 is Associated with Cellular Immune Function Compared to Immune Function in Chinese Han Population

Zeng et al. *medRxiv*. [2013]

12.50.1 Keywords

- WBC
- peripheral blood
- CD4
- CD8 T cells

12.50.2 Main Findings

Retrospective study of the clinical characteristics of 752 patients infected with COVID-19 at Chinese PLA General Hospital, Peking Union Medical College Hospital, and affiliated hospitals at Shanghai University of medicine & Health Sciences. This study is the first one that compares PB from healthy controls from the same regions in Shanghai and Beijing, and infected COVID-19 patients to standardize a reference range of WBCs of people at high risk.

12.50.3 Limitations

Lower levels of leukocyte counts -B cells, CD4 and CD8 T cells- correlated with mortality (WBCs are significantly lower in severe or critical UCI patients vs mild ones). Based on 14,117 normal controls in Chinese Han population (ranging in age from 18-86) it is recommended that reference ranges of people at high risk of COVID-19 infection are CD3+ lymphocytes below 900 cells/mm³, CD4+ lymphocytes below 500 cells/mm³, and CD8+ lymphocytes below 300 cells/mm³. Importantly, this study also reported that the levels of D-dimer, C-reactive protein and IL-6 were elevated in COVID-19 pts., indicating clot formation, severe inflammation and cytokine storm.

12.50.4 Significance

This study sets a threshold to identify patients at risk by analyzing their levels of leukocytes, which is an easy and fast approach to stratify individuals that require hospitalization. Although the study is limited (only counts of WBC are analyzed and not its profile) the data is solid and statistically robust to correlate levels of lymphopenia with mortality.

12.50.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.51 Retrospective Analysis of Clinical Features in 101 Death Cases with COVID-19

Chen et al. *medRxiv*. [2014]

12.51.1 Keywords

- death biomarkers
- cardiac damage
- Troponin
- Blood type
- respiratory failure
- hypertension

12.51.2 Main Findings

This is a retrospective study involving 101 death cases with COVID-19 in Wuhan Jinyintan Hospital. The aim was to describe clinical, epidemiological and laboratory features of fatal cases in order to identify the possible primary mortality causes related to COVID-19.

Among 101 death cases, 56.44% were confirmed by RT-PCR and 43.6% by clinical diagnostics. Males dominated the number of deaths and the average age was 65.46 years. All patients died of respiratory failure and multiple organs failure, except one (acute coronary syndrome). The predominant comorbidities were hypertension (42.57%) and diabetes (22.77%). 25.74% of the patients presented more than two underlying diseases. 82% of patients presented myocardial enzymes abnormalities at admission and further increase in myocardial damage indicators with disease progression: patients with elevated Troponin I progressed faster to death. Alterations in coagulation were also detected. Indicators of liver and kidney damage increased 48 hours before death. The authors studied the deceased patients' blood type and presented the following results: type A (44.44%), type B (29.29%), type AB (8.08%) and type O (18.19%), which is inconsistent with the distribution in Han population in Wuhan.

Clinical analysis showed that the most common symptom was fever (91.9%), followed by cough and dyspnea. The medium time from onset of symptoms to acute respiratory distress syndrome (ARDS) development was 12 days. Unlike SARS, only 2 patients with COVID-19 had diarrhea. 98% presented abnormal lung imaging at admission and most had double-lung abnormalities. Related to the laboratorial findings some inflammatory indicators gradually increased during the disease progression, such as IL-6 secretion in the circulation, procalcitonin (PCT) and C-reactive protein (CRP), while platelets numbers decreased. The authors also reported an initial lymphopenia that was followed by an increase in the lymphocytes numbers. Neutrophil count increased with disease progression.

The patients received different treatments such as antiviral drugs (60.40%), glucocorticoids, thymosin and immunoglobulins. All patients received antibiotic treatment and some received antifungal drugs. All patients received oxygen therapy (invasive or non-invasive ones).

12.51.3 Limitations

This study involves just fatal patients, lacking comparisons with other groups of patients e.g. patients that recovered from COVID-19. The authors didn't discuss the different approaches used for treatments and how these may affect the several parameters measured. The possible relationship between the increase of inflammatory indicators and morbidities of COVID-19 are not discussed.

12.51.4 Significance

This study has the largest cohort of fatal cases reported so far. The authors show that COVID-19 causes fatal respiratory distress syndrome and multiple organ failure. This study highlights prevalent myocardial damage and indicates that cardiac function of COVID-19 patients should be carefully monitored. The data suggest that Troponin I should be further investigated as an early indicator of patients with high risk of accelerated health deterioration. Secondary bacterial and fungal infections were frequent in critically ill patients and these need to be carefully monitored in severe COVID-19 patients. Differences in blood type distribution were observed, suggesting that type A is detrimental while type O is protective – but further studies are needed to confirm these findings and elucidate if blood type influences infection or disease severity. Several inflammatory indicators (neutrophils, PCT, CRP and IL-6, D-dimer) increased according to disease severity and should be assessed as biomarkers and to better understand the biology of progression to severe disease.

12.51.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.52 Relationship between the ABO Blood Group and the COVID-19 Susceptibility

Zhao et al. *medRxiv*. [439]

12.52.1 Keywords

- ABO blood group
- COVID-19 susceptibility

12.52.2 Main Findings

These authors compared the ABO blood group of 2,173 patients with RT-PCR-confirmed COVID-19 from hospitals in Wuhan and Shenzhen with the ABO blood group distribution in unaffected people in the same cities from previous studies (2015 and 2010 for Wuhan and Shenzhen, respectively). They found that people with blood group A are statistically over-represented

in the number of those infected and who succumb to death while those with blood group O are statistically underrepresented with no influence of age or sex.

12.52.3 Limitations

This study compares patients with COVID-19 to the general population but relies on data published 5 and 10 years ago for the control. The mechanisms that the authors propose may underlie the differences they observed require further study.

12.52.4 Significance

Risk stratification based on blood group may be beneficial for patients and also healthcare workers in infection control. Additionally, investigating the mechanism behind these findings could lead to better developing prophylactic and therapeutic targets for COVID-19.

12.52.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.53 The inhaled corticosteroid ciclesonide blocks coronavirus RNA replication by targeting viral NSP15

Matsuyama et al. *bioRxiv* [2015]

12.53.1 Keywords

- Corticosteroids
- ciclesonide
- mometasone
- NSP15
- MERS-CoV

12.53.2 Main Findings

This study reconsiders the use of inhaled corticosteroids in the treatment of pneumonia by coronavirus. Corticosteroids were associated with increased mortality for SARS in 2003 and for MERS in 2013, probably due to that fact that systemic corticosteroids suppress the innate immune system, resulting in increased viral replication. However, some steroid compounds might block coronavirus replication. The authors screened steroids from a chemical library and assessed the viral growth suppression and drug cytotoxicity. Ciclesonide demonstrated low cytotoxicity and potent suppression of MERS-CoV viral growth. The commonly used systemic steroids cortisone, prednisolone and dexamethasone did not suppress viral growth, nor did the

commonly used inhaled steroid fluticasone. To identify the drug target of virus replication, the authors conducted 11 consecutive MERS-CoV passages in the presence of ciclesonide or mometasone, and they could generate a mutant virus that developed resistance to ciclesonide, but not to mometasone. Afterwards, they performed next-generation sequencing and identified an amino acid substitution in nonstructural protein 15 (NSP15) as the predicted mechanism for viral resistance to ciclesonide. The authors were able to successfully generate a recombinant virus carrying that amino acid substitution, which overcome the antiviral effect of ciclesonide, suggesting that ciclosenide interacts with NSP15. The mutant virus was inhibited by mometasone, suggesting that the antiviral target of mometasone is different from that of ciclesonide. Lastly, the effects of ciclesonide and mometasone on suppressing the replication of SARS-CoV-2 were evaluated. Both compounds were found to suppress viral replication with a similar efficacy to lopinavir.

12.53.3 Limitations

Most of the experiments, including the identification of the mutation in NSP15 were conducted with MERS-CoV. This is not the closest related virus to SARS-CoV-2, as that would be SARS-CoV. Thus, to repeat the initial experiments with SARS-CoV, or preferably SARS-CoV-2, is essential. The manuscript should address this and, therefore, it will require considerable editing for organization and clarity. Also, in terms of cell immunogenic epitopes, while SARS-CoV-2 spike protein contains several predicted B and T cell immunogenic epitopes that are shared with other coronaviruses, some studies have shown critical differences between MERS-CoV, SARS-CoV and SARS-CoV-2. A main criticism is that the authors only used VeroE6/TMPRSS2 cells to gauge the direct cytotoxic effects of viral replication. To evaluate this in other cell lines, including human airway epithelial cells, is crucial, as the infectivity of coronavirus strains greatly varies in different cell lines,

12.53.4 Significance

Nevertheless, these findings encourage evaluating ciclesonide and mometasone as better options for patients with COVID-19 in need of inhaled steroids, especially as an alternative to other corticosteroids that have been shown to increase viral replication in vitro. This should be evaluated in future clinical studies.

12.53.5 Credit

This review was undertaken by Alvaro Moreira, MD as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.54 A human monoclonal antibody blocking SARS-CoV-2 infection **

12.54.1 Keywords

- Monoclonal antibodies
- SARS-CoV2
- cross-neutralization
- potential treatment
- spike receptor

12.54.2 Main Findings

The authors reported a human monoclonal antibody that neutralizes SARS-CoV-2 and SARS-CoV which belong to same family of corona viruses. For identifying mAbs, supernatants of a collection of 51 hybridomas raised against the spike protein of SARS-CoV (SARS-S) were screened by ELISA for cross-reactivity against the spike protein of SARS-CoV2 (SARS2-S).

Hybridomas were derived from immunized transgenic H2L2 mice (chimeric for fully human VH-VL and rat constant region). Four SARS-S hybridomas displayed cross-reactivity with SARS2-S, one of which (47D11) exhibited cross-neutralizing activity for SARS-S and SARS2-S pseudotyped VSV infection. A recombinant, fully human IgG1 isotype antibody was generated and used for further characterization.

The humanized 47D11 antibody inhibited infection of VeroE6 cells with SARS-CoV and SARS-CoV-2 with IC₅₀ values of 0.19 and 0.57 µg/ml respectively. 47D11 mAb bound a conserved epitope on the spike receptor binding domain (RBD) explaining its ability to cross-neutralize SARS-CoV and SARS-CoV-2. 47D11 was shown to target the S1B RBD of SARS-S and SARS2-S with similar affinities. Interestingly, binding of 47D11 to SARS-S1B and SARS2-S1B did not interfere with S1B binding to ACE2 receptor-expressing cells assayed by flow cytometry.

12.54.3 Limitations

These results show that the human 47D11 antibody neutralizes SARS-CoV and SARS-CoV2 infectivity via an as yet unknown mechanism that is different from receptor binding interference. Alternative mechanisms were proposed but these as yet remain to be tested in the context of SARS-CoV2. From a therapeutic standpoint and in the absence of in vivo data, it is unclear whether the 47D11 ab can alter the course of infection in an infected host through virus clearance or protect an uninfected host that is exposed to the virus. There is a precedent for the latter possibility as it relates to SARS-CoV that was cited by the authors and could turn out to be true for SARS-CoV2.

12.54.4 Significance

This study enabled the identification of novel neutralizing antibody against COV-that could potentially be used as first line of treatment in the near future to reduce the viral load and adverse effects in infected patients. In addition, neutralizing antibodies such as 47D11 represent promising reagents for developing antigen-antibody-based detection test kits and assays.

12.54.5 Credit

This review was edited by K. Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Heat inactivation of serum interferes with the immunoanalysis of antibodies to SARS-CoV-2

Heat inactivation, immunochromatography, diagnosis, serum antibodies, IgM, IgG

Summary

The use of heat inactivation to neutralize pathogens in serum samples collected from suspected COVID-19 patients reduces the sensitivity of a fluorescent immunochromatographic assay to detect anti-SARS-CoV-2 IgM and IgG.

Major findings

Coronaviruses can be killed by heat inactivation, and this is an important safety precaution in laboratory manipulation of clinical samples. However, the effect of this step on downstream SARS-CoV-2-specific serum antibody assays has not been examined. The authors tested the effect of heat inactivation (56 deg C for 30 minutes) versus no heat inactivation on a fluorescence immunochromatography assay. Heat inactivation reduced all IgM measurements by an average of 54% and most IgG measurements (22/36 samples, average reduction of 50%), consistent with the lower thermal stability of IgM than that of IgG. Heat inactivation caused a subset of IgM but not IgG readings to fall below a specified positivity threshold.

Limitations

Limitations included the use of only one type of assay for testing heat inactivated vs non-inactivated sera, and the use of the same baseline for heat inactivated and non-inactivated sera. The results indicate that heat inactivation affects the quantification of SARS-CoV-2-antibody response, specially IgM, but still allows to distinguish positive specific IgG. Therefore, the effect of heat inactivation should be studied when designing assays that quantitatively associate immunoglobulin levels (especially IgM) to immune state.

Review by Andrew M. Leader as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn school of medicine, Mount Sinai.

12.55 Immune phenotyping based on neutrophil-to-lymphocyte ratio and IgG predicts disease severity and outcome for patients with

COVID-19

Zhang et al. *medRxiv* [2016]

12.55.1 Keywords

- Biomarkers
- cytokines
- IgG
- immune cells

12.55.2 Main Findings

In a cohort of 222 patients, anti-SARS-CoV-2 IgM and IgG levels were analyzed during acute and convalescent phases (up to day 35) and correlated to the diseases' severity. The same was done with neutrophil-to-lymphocyte ratio. High IgG levels and high neutrophil-to-lymphocyte ratio in convalescence were both independently associated to the severity of the disease. The simultaneous occurrence of both of these laboratory findings correlated even stronger to the diseases' severity.

Severe cases with high neutrophil-to-lymphocyte ratios had clearly higher levels of IL-6. The authors propose that a robust IgG response leads to immune-mediated tissue damage, thus explaining the worse outcome in patients with overexuberant antibody response.

12.55.3 Limitations

A main criticism is that the criteria for stratifying patients in severe vs. non-severe are not described. The only reference related to this is the difference between the percentage of patients who needed mechanical ventilation, which was greater in patients with both high IgG levels and high neutrophil-to-lymphocyte ratio. No patient with both low IgG levels and low neutrophil-to-lymphocyte ratio was treated with mechanical ventilation.

The proposed correlation of severity with IL-2 and IL-10 levels is not very strong.

Furthermore, although mostly ignored in the paper's discussion, one of the most interesting findings is that an early increase in anti-SARS-CoV-2 IgM levels also seems to correlate with severe disease. However, as only median values are shown for antibody kinetics curves, the extent of variation in acute phase cannot be assessed.

12.55.4 Significance

Anti-SARS-CoV-2 IgG levels and with neutrophil-to-lymphocyte ratio predict severity of COVID-19 independently of each other. An additive predictive value of both variables is noticeable. Importantly, an early-on increase in anti-SARS-CoV-2 IgM levels also seem to predict outcome.

12.55.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.56 Reinfection could not occur in SARS-2 CoV-2 infected rhesus macaques

Bao et al. *bioRxiv* [2017]

12.56.1 Keywords

- SARS-CoV-2
- viral load
- reinfection
- relapse
- non-human primate model

12.56.2 Main Findings

This study addresses the issue of acquired immunity after a primary COVID-19 infection in rhesus monkeys. Four Chinese rhesus macaques were intratracheally infected with SARS-CoV-2 and two out of the four were re-infected at 28 days post initial infection (dpi) with the same viral dose after confirming the recovery by the absence of clinical symptoms, radiological abnormalities and viral detection (2 negative RT-PCR tests). While the initial infection led to viral loads in nasal and pharyngeal swabs that reach approximately $6.5 \log_{10}$ RNA copies/ml at 3 dpi in all four monkeys, viral loads in the swabs tested negative after re-infection in the two reinfected monkeys. In addition, the necropsies from a monkey (M1) at 7 days after primary infection, and another monkey (M3) at 5 days post re-infection, revealed histopathological damages and viral replication in the examined tissues from M1, while no viral replication as well as no histological damages were detected in the tissues from M3. Furthermore, sera from three monkeys at 21 and 28 dpi exhibited neutralizing activity against SARS-CoV-2 in vitro, suggesting the production of protective neutralizing antibodies in these monkeys. Overall, this study indicates that primary infection with SARS-CoV-2 may protect from subsequent exposure to the same virus.

12.56.3 Limitations

In humans, virus has been detected by nasopharyngeal swabs until 9 to 15 days after the onset of symptoms. In the infected monkeys in this study, virus were detected from day 1 after the infection, declining to undetectable level by day 15 post infection. It may suggest that there is a faster viral clearance mechanism in monkeys, therefore the conclusions of re-infection protection for humans need to be carefully considered. In addition, only two monkeys were re-infected in this study and the clinical signs of these monkeys were not similar: M3 did not show weight loss and M4 showed relatively higher fever on the day of infection and the day of re-challenge.

12.56.4 Significance

This study showed clear viral clearance and no indications of relapse or viremia after a secondary infection with SARS-CoV-2 in a Chinese rhesus macaque model. These results support the idea that patients with full recovery (two negative RT-PCR results) may also be protected from secondary SARS-CoV-2 infection. Recovered patients may be able to re-integrate to normal public life and provide protective serum perhaps even if having had a mild infection. The results are also encouraging for successful vaccine development against SARS-CoV-2.

12.56.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.57 A highly conserved cryptic epitope in the receptor-binding domains of SARS-CoV-2 and SARS-CoV

[2018]

12.57.1 Keywords

- neutralizing antibody
- cross-reactivity

12.57.2 Main Findings

Given the sequence similarity of the surface spike glycoprotein (S) of SARS-CoV-2 and SARS-CoV, Yuan et al. (2020) propose that neutralizing antibodies isolated from convalescent SARS-CoV patients may offer insight into cross-reactive antibodies targeting SARS-CoV-2. In particular, they find that the receptor-binding domain (RBD) of SARS-CoV-2 S protein shares 86% sequence similarity with the RBD of SARS-CoV S protein that binds to the CR3022 neutralizing antibody. CR3022 also displays increased affinity for the “up” conformation of the SARS-CoV-2 S protein compared to the “down” conformation as it does for the SARS-CoV S protein. Therefore, the authors propose that this cross-reactive antibody may confer some degree of protection *in vivo* even if it fails to neutralize *in vitro*.

12.57.3 Limitations

Although the authors offer a logical rationale for identifying cross-reactive neutralizing antibodies derived from SARS-CoV, their study using only CR3022 failed to demonstrate whether this approach will be successful. After all, CR3022 failed to neutralize *in vitro* despite the binding affinity to a similar

epitope on SARS-CoV-2. They would benefit from testing more candidates and using an *in vivo* model to demonstrate their claim that protection may be possible in the absence neutralization if combinations are used *in vivo*.

12.57.4 Significance

The ability to make use of previously characterized neutralizing antibodies for conserved epitopes can expedite drug design and treatment options.

12.57.5 Credit

This review was undertaken by Dan Fu Ruan, Evan Cody and Venu Pothula as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.58 Highly accurate and sensitive diagnostic detection of SARS-CoV-2 by digital PCR

Dong et al. *medRxiv* [[2019](#)]

12.58.1 Keywords

- Diagnosis
- digital PCR

12.58.2 Main Findings

The authors present a digital PCR (dPCR) diagnostic test for SARS-CoV-2 infection. In 103 individuals that were confirmed in a follow-up to be infected, the standard qPCR test had a positivity rate of 28.2% while the dPCR test detected 87.4% of the infections by detecting an additional 61 positive cases. The authors also tested samples from close contacts (early in infection stage) and convalescing individuals (late in infection stage) and were able to detect SARS-CoV-2 nucleic acid in many more samples using dPCR compared to qPCR.

12.58.3 Limitations

I did not detect limitations.

12.58.4 Significance

The authors make a strong case for the need for a highly sensitive and accurate confirmatory method for diagnosing COVID-19 during this outbreak and present a potential addition to the diagnostic arsenal. They propose a dPCR test that they present has a dramatically lower false negative rate than the standard RT-qPCR tests and can be especially beneficial in people with low viral load, whether they are in the earlier or later stages of infection.

12.58.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.59 SARS-CoV-2 invades host cells via a novel route: CD147-spike protein

Wang et al. *bioRxiv* [2020]

12.59.1 Keywords

- spike protein
- viral entry
- CD147
- SARS-CoV-2

12.59.2 Main Findings

The authors propose a novel mechanism of SARS-CoV-2 viral entry through the interaction of the viral spike protein (SP) and the immunoglobulin superfamily protein CD147 (also known as Basigin). Using an in-house developed humanized antibody against CD147 (maplazumab), they show that blocking CD147 decreases viral replication in Vero E6 cells. Using surface plasmon resonance (SPR), ELISA, and Co-IP assays, they show that the spike protein of SARS-CoV-2 directly interacts with CD147. Lastly, they utilize immune-electron microscopy to show spike protein and CD147 localize to viral inclusion bodies of Vero E6 cells.

12.59.3 Limitations

The authors claim that an anti-CD147 antibody (Meplazumab) inhibits SARS-CoV-2 replication by testing cell growth and viral load in cells infected with SARS-CoV-2, however there are key pieces of this experiment that are missing. First, the authors fail to use a non-specific antibody control. Second, the authors claim that viral replication is inhibited, and that they test this by qPCR, however this data is **not shown**. To further prove specificity, the authors should introduce CD147 to non-susceptible cells and show that they become permissive.

The authors claim that there is a direct interaction between CD147 and SP through SPR, ELISA, and Co-IP, and this data seems generally convincing. The electron microscopy provides further correlative evidence that SARS-CoV-2 may interact with CD147 as they are both found in the same viral inclusion body. A quantification of this data would make the findings more robust.

Finally, the data in this paper lacks replicates, error bars, and statistics to show that the data are reproducible and statistically significant.

12.59.4 Significance

It has been shown in various studies that SARS-CoV-2 binds to the cell surface protein ACE2 for cell entry, yet ACE2 is highly expressed in heart, kidney, and intestinal cells, raising the concern that blocking ACE2 would result in harmful side effects [2021] CD147 on the other hand is highly expressed in various tumor types, inflamed tissues, and pathogen infected cells, suggesting that the inhibition of CD147 would not result in major side effects [2022,2023] The research in this paper has resulted in an ongoing clinical trial in China to test the safety and efficacy of anti-CD147 Meplazumab to treat COVID-19. (ClinicalTrials.gov identifier NCT04275245).

12.59.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.60 Blood single cell immune profiling reveals that interferon-MAPK pathway mediated adaptive immune response for COVID-19

Huang et al. *medRxiv* [2024]

12.60.1 Keywords

- COVID-19
- SARS-CoV-2
- PBMC
- single cell
- MAPK

12.60.2 Main Findings

The authors performed single-cell RNA sequencing (scRNASeq) of peripheral blood mononuclear cells isolated from whole blood samples of COVID-19 patients (n=10). Data was compared to scRNASeq of samples collected from patients with influenza A (n=1), acute pharyngitis (n=1), and cerebral infarction (n=1), as well as, three healthy controls. COVID-19 patients were categorized into those with moderate (n=6), severe (n=1), critical (n=1), and cured (n=2) disease. Analysis across all COVID-19 disease levels revealed 56 different cellular subtypes, among 17 immune cell types; comparisons between each category to the normal controls revealed **increased proportions of CD1c⁺ dendritic cells, CD8⁺ CTLs, and plasmacytoid dendritic cells and a decrease in proportions of B cells and CD4⁺ T cells.**

TCR sequencing revealed that greater clonality is associated with milder COVID-19 disease; BCR sequencing revealed that COVID-19 patients have circulating antibodies against known viral antigens, including EBV, HIV,

influenza A, and other RNA viruses. This may suggest that the immune response to SARS-CoV-2 infection elicits production of antibodies against known RNA viruses.

Excluding enriched pathways shared by COVID-19 patients and patients with other conditions (influenza A, acute pharyngitis, and cerebral infarction), the authors identified the **interferon-MAPK signaling pathway as a major response to SARS-CoV-2 infection**. The authors performed quantitative real-time reverse transcriptase polymerase chain reaction (RT-PCR) for interferon-MAPK signaling genes: *IRF27*, *BST2*, and *FOS*. These samples were collected from a separate cohort of COVID-19 patients (critical, n=3; severe, n=3; moderate, n=19; mild, n=3; and cured, n=10; and healthy controls, n=5). Notably, consistent with the original scRNAseq data, *FOS* showed up-regulation in COVID-19 patients and down-regulation in cured patients. **The authors propose that *FOS* may be a candidate marker gene for curative COVID-19 disease.**

12.60.3 Limitations

The sample size of this study is limited. To further delineate differences in the immune profile of peripheral blood of COVID-19 patients, a greater sample size is needed, and longitudinal samples are needed, as well. A better understanding of the immunological interactions in cured patients, for example, would require a profile before and after improvement.

Moreover, the conclusions drawn from this scRNAseq study point to potential autoimmunity and immune deficiency to distinguish different severities of COVID-19 disease. However, this requires an expanded number of samples and a more robust organization of specific immune cell subtypes that can be compared across different patients. Importantly, this criterion is likely needed to ensure greater specificity in identifying markers for COVID-19 infection and subsequent immune response.

12.60.4 Significance

At the single-cell level, COVID-19 disease has been characterized in the lung, but a greater understanding of systemic immunological responses is furthered in this study. Type I interferon is an important signaling molecule for the anti-viral response. The identification of the interferon-MAPK signaling pathway and the differential expression of MAPK regulators between patients of differing COVID-19 severity and compared to cured patients may underscore the importance of either immune deficiency or autoimmunity in COVID-19 disease.

12.60.5 Credit

This review was undertaken by Matthew D. Park as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.61 Cross-reactive antibody response between SARS-CoV-2 and SARS-CoV infection.

Lv et al. *bioRxiv* [2025]

12.61.1 Keywords

SARS-CoV-2, SARS-CoV, spike protein, RBD, cross-reactivity, cross-neutralization, antibody, human patients, mouse

12.61.2 Main Findings

The authors explore the antigenic differences between SARS-CoV-2 and SARS-CoV by analyzing plasma samples from SARS-CoV-2 ($n = 15$) and SARS-CoV ($n = 7$) patients. Cross-reactivity in antibody binding to the spike protein between SARS-CoV-2 and SARS-CoV was found to be common, mostly targeting non-RBD regions in plasma from SARS-CoV-2 patients. Only one SARS-CoV-2 plasma sample was able to cross-neutralize SARS-CoV, with low neutralization activity. No cross-neutralization response was detected in plasma from SARS-CoV patients.

To further investigate the cross-reactivity of antibody responses to SARS-CoV-2 and SARS-CoV, the authors analyzed the antibody response of plasma collected from mice infected or immunized with SARS-CoV-2 or SARS-CoV ($n = 5$ or 6 per group). Plasma from mice immunized with SARS-CoV-2 displayed cross-reactive responses to SARS-CoV S ectodomain and, to a lesser extent, SARS-CoV RBD. Similarly, plasma from mice immunized with SARS-CoV displayed cross-reactive responses to SARS-CoV-2 S ectodomain. Cross-neutralization activity was not detected in any of the mouse plasma samples.

12.61.3 Limitations

The size of each patient cohort is insufficient to accurately determine the frequency of cross-reactivity and cross-neutralization in the current SARS-CoV-2 pandemic. Recruitment of additional patients from a larger range of geographical regions and time points would also enable exploration into the effect of the genetic diversity and evolution of the SARS-CoV-2 virus on cross-reactivity. This work would also benefit from the mapping of specific epitopes for each sample. Future studies may determine whether the non-neutralizing antibody responses can confer *in vitro* protection or lead to antibody-dependent disease enhancement.

12.61.4 Significance

The cross-reactive antibody responses to S protein in the majority of SARS-CoV-2 patients is an important consideration for development of serological assays and vaccine development during the current outbreak. The limited extent of cross-neutralization demonstrated in this study indicates that vaccinating to cross-reactive conserved epitopes may have limited efficacy,

presenting a key concern for the development of a more universal coronavirus vaccine to address the global health risk of novel coronavirus outbreaks.

12.61.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.62 The feasibility of convalescent plasma therapy in severe COVID-19 patients: a pilot study

Duan et al. *medRxiv* [[2026](#)]

12.62.1 Keywords

- COVID-19
- SARS-CoV-2
- convalescent plasma
- treatment outcome
- pilot
- therapy
- transfusion

12.62.2 Main Findings

This is the first report to date of convalescent plasma therapy as a therapeutic against COVID-19 disease. This is a feasibility pilot study. The authors report the administration and clinical benefit of 200 mL of convalescent plasma (CP) (1:640 titer) derived from recently cured donors (CP selected among 40 donors based on high neutralizing titer and ABO compatibility) to 10 severe COVID-19 patients with confirmed viremia. The primary endpoint was the safety of CP transfusion. The secondary endpoint were clinical signs of improvement based on symptoms and laboratory parameters.

The authors reported use of methylene blue photochemistry to inactivate any potential residual virus in the plasma samples, without compromising neutralizing antibodies, and no virus was detected before transfusion.

The authors report the following:

- No adverse events were observed in all patients, except 1 patient who exhibited transient facial red spotting.
- All patients showed significant improvement in or complete disappearance of clinical symptoms, including fever, cough, shortness of breath, and chest pain after 3 days of CP therapy.

- Reduction of pulmonary lesions revealed by chest CT.
- Elevation of lymphocyte counts in patients with lymphocytopenia.
- Increase in SaO₂ in all patients, indicative of recuperating lung function.
- Resolution of SARS-CoV-2 viremia in 7 patients and increase in neutralizing antibody titers in 5 patients. Persistence of neutralizing antibody levels in 4 patients.

12.62.3 Limitations

It is important to note that most recipients had high neutralization titers of antibodies before plasma transfusion and even without transfusion it would be expected to see an increase in neutralizing antibodies over time. In addition to the small sample set number (n=10), there are additional limitations to this pilot study:

1. All patients received concurrent therapy, in addition to the CP transfusion. Therefore, it is unclear whether a combinatorial or synergistic effect between these standards of care and CP transfusion contributed to the clearance of viremia and improvement of symptoms in these COVID-19 patients.
2. The kinetics of viral clearance was not investigated, with respect to the administration of CP transfusion. So, the definitive impact of CP transfusion on immune dynamics and subsequent viral load is not well defined.
3. Comparison with a small historical control group is not ideal.

12.62.4 Significance

For the first time, a pilot study provides promising results involving the use of convalescent plasma from cured COVID-19 patients to treat others with more severe disease. The authors report that the administration of a single, high-dose of neutralizing antibodies is safe. In addition, there were encouraging results with regards to the reduction of viral load and improvement of clinical outcomes. It is, therefore, necessary to expand this type of study with more participants, in order to determine optimal dose and treatment kinetics. It is important to note that CP has been studied to treat H1N1 influenza, SARS-CoV-1, and MERS-CoV, although it has not been proven to be effective in treating these infections.

12.62.5 Credit

Review by Matthew D. Park and revised by Alice O. Kamphorst and Maria A. Curotto de Lafaille as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.63 Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open label non-randomized clinical trial

[2027]

12.63.1 Keywords

- hydroxychloroquine
- clearance
- viral load
- clinical trial

12.63.2 Main Findings

This study was a single-arm, open label clinical trial with 600 mg hydroxychloroquine (HCQ) in the treatment arm ($n = 20$). Patients who refused participation or patients from another center not treated with HCQ were included as negative controls ($n = 16$). Among the patients in the treatment arm, 6 received concomitant azithromycin to prevent superimposed bacterial infection. The primary endpoint was respiratory viral loads on day 6 post enrollment, measured by nasopharyngeal swab followed by real-time reverse transcription-PCR.

HCQ alone was able to significantly reduce viral loads by day 6 ($n = 8/14$, 57.1% complete clearance, $p < 0.001$); azithromycin appears to be synergistic with HCQ, as 6/6 patients receiving combined treatment had complete viral clearance ($p < 0.001$).

12.63.3 Limitations

Despite what is outlined above, this study has a number of limitations that must be considered. First, there were originally $n = 26$ patients in the treatment arm, with 6 lost to follow up for the following reasons: 3 transferred to ICU, 1 discharge, 1 self-discontinued treatment d/t side effects, and 1 patient expired. Total length of clinical follow up was 14 days, but the data beyond day 6 post-inclusion are not shown.

Strikingly, in supplementary table 1, results of the real-time RT-PCR are listed for the control and treatment arms from D0 – D6. However, the data are not reported in a standard way, with a mix of broadly positive or negative result delineation with Ct (cycle threshold) values, the standard output of real time PCR. It is impossible to compare what is defined as a positive value between the patients in the control and treatment arms without a standardized threshold for a positive test. Further, the starting viral loads reported at D0 in the groups receiving HCQ or HCQ + azithromycin were significantly different (ct of 25.3 vs 26.8 respectively), which could explain in part the differences observed in the response to treatment between 2 groups. Finally, patients in the control arm from outside the primary medical center in this study

(Marseille) did not actually have samples tested by PCR daily. Instead, positive test results from every other day were extrapolated to mean positive results on the day before and after testing as well (Table 2, footnote ³).

Taken together, the results of this study suggest that HCQ represents a promising treatment avenue for COVID-19 patients. However, the limited size of the trial, and the way in which the results were reported does not allow for other medical centers to extrapolate a positive or negative result in the treatment of their own patients with HCQ +/- azithromycin. Further larger randomized clinical trials will be required to ascertain the efficacy of HCQ +/- azithromycin in the treatment of COVID-19.

12.63.4 Significance

Chloroquine is thought to inhibit viral infection, including SARS-CoV-2, by increasing pH within endosomes and lysosomes, altering the biochemical conditions required for viral fusion [[784,2028](#)]. However, chloroquine also has immuno-modulatory effects that I think may play a role. Chloroquine has been shown to increase CTLA-4 expression at the cell surface by decreasing its degradation in the endo-lysosome pathway; AP-1 traffics the cytoplasmic tail of CTLA-4 to lysosomes, but in conditions of increased pH, the protein machinery required for degradation is less functional [[2029](#)]. As such, more CTLA-4 remains in endosomes and is trafficked back to the cell surface. It is possible that this may also contribute to patient recovery via reduction of cytokine storm, in addition to the direct anti-viral effects of HCQ.

12.63.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.64 Recapitulation of SARS-CoV-2 Infection and Cholangiocyte Damage with Human Liver Organoids

[[1954](#)]

12.64.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- liver
- organoids
- Cholangiocyte

12.64.2 Main Findings

- Used human liver ductal organoids to determine ACE2+ cholangiocytes in healthy liver (2.92% of all cells) are infectable and support SARS-CoV-2 viral replication.
- Plaque-purified SARS-CoV-2 viral infection disrupted organoid barrier and bile transporting functions of cholangiocytes through dysregulation of genes involved in tight junction formation (CLDN1) and bile acid transportation (ASBT and CFTR).

12.64.3 Limitations:

- Unclear if liver damage observed in patients due to direct cholangiocyte infection or due to secondary immune/cytokine effects. This study argues for direct damage as it lacks immune contexture; but further studies needed with autopsy samples or organoid-immune cell co-culture to conclude strongly.
- Would be important to measure cholangiocyte-intrinsic anti-viral response and alarmins secreted upon infection, and furthermore study tropism of various immune cells to conditioned media from organoids infected with SARS-CoV-2.
- Does not address how cirrhotic liver or alcohol/smoking/obesity-associated liver organoids respond to SARS-CoV-2 infectivity and replication, worth pursuing to experimentally address clinical data indicating co-morbidities.

12.64.4 Significance

- Useful model to rapidly study drug activity against SARS-CoV-2 infection in liver, while monitoring baseline liver damage.
- Liver abnormality observed in >50% of CoVID-19 patients; the results from this study could explain the bile acid accumulation and consequent liver damage observed.

12.64.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.65 The sequence of human ACE2 is suboptimal for binding the S spike protein of SARS coronavirus 2

[2030]

12.65.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- Spike protein S
- ACE2

12.65.2 Main Findings

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) infects cells through S spike glycoprotein binding angiotensin-converting enzyme (ACE2) on host cells. S protein can bind both membrane-bound ACE2 and soluble ACE2 (sACE2), which can serve as a decoy that neutralizes infection.

Recombinant sACE2 is now being tested in clinical trials for COVID-19. To determine if a therapeutic sACE2 with higher affinity for S protein could be designed, authors generated a library containing every amino acid substitution possible at the 117 sites spanning the binding interface with S protein. The ACE2 library was expressed in human Expi293F cells and cells were incubated with medium containing the receptor binding domain (RBD) of SARS-CoV-2 fused to GFP. Cells with high or low affinity mutant ACE2 receptor compared to affinity of wild type ACE2 for the RBD were FACS sorted and transcripts from these sorted populations were deep sequenced. Deep mutagenesis identified numerous mutations in ACE2 that enhance RBD binding. This work serves to identify putative high affinity ACE2 therapeutics for the treatment of CoV-2.

12.65.3 Limitations

The authors generated a large library of mutated ACE2, expressed them in human Expi293F cells, and performed deep mutagenesis to identify enhanced binders for the RBD of SARS-CoV-2 S protein. While these data serve as a useful resource, the ability of the high affinity ACE2 mutants identified to serve as therapeutics needs further validation in terms of conformational stability when purified as well as efficacy/safety both *in vitro* and *in vivo*. Additionally, authors mentioned fusing the therapeutic ACE2 to Fc receptors to elicit beneficial host immune responses, which would need further design and validation.

12.65.4 Significance

This study identified structural ACE2 mutants that have potential to serve as therapeutics in the treatment of SARS-CoV-2 upon further testing and validation.

12.65.5 Credit

This review was undertaken by Katherine Lindblad and Tamar Plitt as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Title: A serological assay to detect SARS-Cov-2 seroconversion in humans

Immunology keywords: specific serological assay - ELISA - seroconversion - antibody titers

Note: the authors of this review work in the same institution as the authors of the study

Main findings:

Production of recombinant whole Spike (S) protein and the smaller Receptor Binding Domain (RBD) based on the sequence of Wuhan-Hu-1 SARS-CoV-2 isolate. The S protein was modified to allow trimerization and increase stability. The authors compared the antibody reactivity of 59 banked human serum samples (non-exposed) and 3 serum samples from confirmed SARS-CoV-2 infected patients. All Covid-19 patient sera reacted to the S protein and RBD domain compared to the control sera.

The authors also characterized the antibody isotypes from the Covid-19 patients, and observed stronger IgG3 response than IgG1. IgM and IgA responses were also prevalent.

Limitations of the study:

The authors analyzed a total of 59 control human serum samples, and samples from only three different patients to test for reactivity against the RBD domain and full-length spike protein. It will be important to follow up with a larger number of patient samples to confirm the data obtained. Furthermore, it would be interesting to assess people at different age groups and determine whether unexposed control kids have a higher "background".

Applications of the assay described in this study in diagnosis are limited, since antibody response should start to be detectable only one to two weeks after infection. Future studies will be required to assess how long after infection this assay allows to detect anti-CoV2 antibodies. Finally, while likely, the association of seroconversion with protective immunity against SARS-CoV-2 infection still needs to be fully established.

Relevance:

This study has strong implications in the research against SARS-CoV-2. First, it is now possible to perform serosurveys and determine who has been infected, allowing a more accurate estimate of infection prevalence and death rate. Second, if it is confirmed that re-infection does not happen (or is rare), this assay can be used as a tool to screen healthcare workers and prioritize immune ones to work with infected patients. Third, potential convalescent plasma donors can now be screened to help treat currently infected patients. Of note, this assay does not involve live virus handling. Experimentally, this is an advantage as the assay does not require the precautions required by manipulation of live virus. Finally, the recombinant proteins described in this study represent new tools that can be used for further applications, including vaccine development.

12.66 COMPARATIVE PATHOGENESIS OF COVID-19, MERS AND SARS IN A NON-HUMAN PRIMATE MODEL

12.66.1 Keywords

- SARS-CoV2
- cynomolgus macaque
- SARS-CoV

12.66.2 Main Findings

This work assesses SARS-CoV-2 infection in young adult and aged cynomolgus macaques. 4 macaques per age group were infected with low-passage clinical sample of SARS-CoV-2 by intranasal and intratracheal administration. Viral presence was assessed in nose, throat and rectum through RT-PCR and viral culture. SARS-CoV-2 replication was confirmed in the respiratory track (including nasal samples), and it was also detected in ileum. Viral nucleocapsid detection by IHC showed infection of type I and II pneumocytes and epithelia. Virus was found to peak between 2 and 4 days after administration and reached higher levels in aged vs. young animals. The early peak is consistent with data in patients and contrasts to SARS-CoV replication. SARS-CoV-2 reached levels below detection between 8 and 21 days after inoculation and macaques established antibody immunity against the virus by day 14. There were histopathological alteration in lung, but no overt clinical signs. At day 4 post inoculation of SARS-CoV-2, two of four animals presented foci of pulmonary consolidation, with limited areas of alveolar edema and pneumonia, as well as immune cell infiltration. In sum, cynomolgus macaques are permissive to SARS-CoV-2 and develop lung pathology (less severe than SARC-CoV, but more severe than MERS-CoV).

12.66.3 Limitations

Even though cynomolgus macaques were permissive to SARS-CoV-2 replication, it is unclear if the viral load reaches levels comparable to humans and there wasn't overt clinical pathology.

12.66.4 Significance

The development of platforms in which to carry out relevant experimentation on SARS-CoV-2 pathophysiology is of great urgency. Cynomolgus macaques offer an environment in which viral replication can happen, albeit in a limited way and without translating into clinically relevant symptoms. Other groups are contributing to SARS-CoV2 literature using this animal model [2017], potentially showing protection against reinfection in cured macaques. Therefore, this platform could be used to examine SARS-CoV2 pathophysiology while studies in other animal models are also underway [1956,2032].

12.66.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.67 Investigating the Impact of Asymptomatic Carriers on COVID-19 Transmission

[2033]

12.67.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- asymptomatic carriers
- mathematical model
- transmission

12.67.2 Main Findings

Multiple studies reported the same level of infectiousness between symptomatic and asymptomatic carriers of SARS-CoV-2. Given that asymptomatic and undocumented carriers escape public health surveillance systems, a better mathematical model of transmission is needed to determine a more accurate estimate of the basic reproductive number (R_0) of the virus to assess the contagiousness of virus. The authors developed a SEYAR dynamical model for transmission of the new coronavirus that takes into account asymptomatic and undocumented carriers. The model was validated using data reported from thirteen countries during the first three weeks of community transmission. While current studies estimate R_0 to be around 3, this model indicates that the value could range between 5.5 to 25.4.

12.67.3 Limitations

The SEYAR model realistically depicts transmission of the virus only during the initial stages of the disease. More data is necessary to better fit the model with current trends. In addition, multiple factors (e.g. behavioral patterns, surveillance capabilities, environmental and socioeconomic factors) affect transmission of the virus and so, these factors must be taken into consideration when estimating the R_0 .

12.67.4 Significance

Public health authorities use the basic reproductive number to determine the severity of disease. An accurate estimate of R_0 will inform intervention strategies. This model can be applied to different locations to assess the potential impact of COVID-19.

12.67.5 Credit

This review was undertaken by Tamar Plitt and Katherine Lindblad as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.68 Antibody responses to SARS-CoV-2 in COVID-19 patients: the perspective application of serological tests in clinical practice

Long et al. *medRxiv* [2034]

12.68.1 Keywords

- Serum antibodies
- IgM
- IgG
- immunoassay
- diagnosis
- seroconversion

12.68.2 Main Findings

This study investigated the profile of the acute antibody response against SARS-CoV-2 and provided proposals for serologic tests in clinical practice. Magnetic Chemiluminescence Enzyme Immunoassay was used to evaluate IgM and IgG seroconversion in 285 hospital admitted patients who tested positive for SARS-CoV-2 by RT-PCR and in 52 COVID-19 suspected patients that tested negative by RT-PCR. A follow up study with 63 patients was performed to investigate longitudinal effects. In addition, IgG and IgM titers were evaluated in a cohort of close contacts (164 persons) of an infected couple.

The median day of seroconversion for both IgG and IgM was 13 days after symptom onset. Patients varied in the order of IgM/ IgG seroconversion and there was no apparent correlation of order with age, severity, or hospitalization time. This led the authors to conclude that for diagnosis IgM and IgG should be detected simultaneously at the early phase of infection.

IgG titers, but not IgM titers were higher in severe patients compared to non-severe patients after controlling for days post-symptom onset. Importantly, 12% of COVID-19 patients (RT-PCR confirmed) did not meet the WHO serological diagnosis criterion of either seroconversion or >4-fold increase in IgG titer in sequential samples. This suggests the current serological criteria may be too stringent for COVID-19 diagnosis.

Of note, 4 patients from a group of 52 suspects (negative RT-PCR test) had anti-SARS-CoV-2 IgM and IgG. Similarly, 4.3% (7/162) of “close contacts” who had negative RT-PCR tests were positive for IgG and/or IgM. This highlights the usefulness of a serological assay to identify asymptomatic infections and/or infections that are missed by RT-PCR.

12.68.3 Limitations

This group's report generally confirms the findings of others that have evaluated the acute antibody response to SARS-CoV-2. However, these data would benefit from inclusion of data on whether the participants had a

documented history of viral infection. Moreover, serum samples that were collected prior to SARS-CoV-2 outbreak from patients with other viral infections would serve as a useful negative control for their assay. Methodological limitations include that only one serum sample per case was tested as well as the heat inactivation of serum samples prior to testing. It has previously been reported that heat inactivation interferes with the level of antibodies to SARS-CoV-2 and their protocol may have resulted in diminished quantification of IgM, specifically [2035].

12.68.4 Significance

Understanding the features of the antibody responses against SARS-CoV is useful in the development of a serological test for the diagnosis of COVID-19. This paper addresses the need for additional screening methods that can detect the presence of infection despite lower viral titers. Detecting the production of antibodies, especially IgM, which are produced rapidly after infection can be combined with PCR to enhance detection sensitivity and accuracy and map the full spread of infection in communities. Moreover, serologic assays would be useful to screen health care workers in order to identify those with immunity to care for patients with COVID19.

12.68.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.69 SARS-CoV-2 specific antibody responses in COVID-19 patients

[2036]

12.69.1 Keywords

- immunoassay
- antibody specificity
- serology
- cross-reactivity

12.69.2 Main findings

Antibodies specific to SARS-CoV-2 S protein, the S1 subunit and the RBD (receptor-binding domain) were detected in all SARS-CoV-2 patient sera by 13 to 21 days post onset of disease. Antibodies specific to SARS-CoV N protein (90% similarity to SARS-CoV-2) were able to neutralize SARS-CoV-2 by PRNT (plaque reduction neutralizing test). SARS-CoV-2 serum cross-reacted with SARS-CoV S and S1 proteins, and to a lower extent with MERS-CoV S protein, but not with the MERS-CoV S1 protein, consistent with an analysis of genetic similarity. No reactivity to SARS-CoV-2 antigens was observed in serum from patients with ubiquitous human CoV infections (common cold) or to non-CoV viral respiratory infections.

12.69.3 Limitations

Authors describe development of a serological ELISA based assay for the detection of neutralizing antibodies towards regions of the spike and nucleocapsid domains of the SARS-CoV-2 virus. Serum samples were obtained from PCR-confirmed COVID-19 patients. Negative control samples include a cohort of patients with confirmed recent exposure to non-CoV infections (i.e. adenovirus, bocavirus, enterovirus, influenza, RSV, CMV, EBV) as well as a cohort of patients with confirmed infections with ubiquitous human CoV infections known to cause the common cold. The study also included serum from patients with previous MERS-CoV and SARS-CoV zoonotic infections. This impressive patient cohort allowed the authors to determine the sensitivity and specificity of the development of their in-house ELISA assay. Of note, seroconversion was observed as early as 13 days following COVID-19 onset but the authors were not clear how disease onset was determined.

12.69.4 Significance

Validated serological tests are urgently needed to map the full spread of SARS-CoV-2 in the population and to determine the kinetics of the antibody response to SARS-CoV-2. Furthermore, clinical trials are ongoing using plasma from patients who have recovered from SARS-CoV-2 as a therapeutic option. An assay such as the one described in this study could be used to screen for strong antibody responses in recovered patients. Furthermore, the assay could be used to screen health care workers for antibody responses to SARS-CoV-2 as personal protective equipment continues to dwindle. The challenge going forward will be to standardize and scale-up the various in-house ELISA's being developed in independent laboratories across the world.

12.70 A brief review of antiviral drugs evaluated in registered clinical trials for COVID-19

Belhadi et al. [[2037](#)]

12.70.1 Keywords

- Clinical trials
- COVID-19
- SARS CoV-2
- 2019-nCoV
- SARS Cov-2
- Hcov-19
- novel coronal virus
- new corona virus
- antiviral drugs

12.70.2 Main Findings

Summary of clinical trials registered as of March 7, 2020 from U.S, Chinese, Korean, Iranian and European registries. Out of the 353 studies identified, 115 were selected for data extraction. 80% of the trials were randomized with parallel assignment and the median number of planned inclusions was 63 (IQR, 36-120). Most frequent therapies in the trials included; 1) antiviral drugs [lopinavir/ritonavir (n=15); umifenovir (n=9); favipiravir (n=7); remdesivir (n=5)]; 2) anti-malaria drugs [chloroquine (n=11); hydroxychloroquine (n=7)]; immunosuppressant drugs [methylprednisolone (n=5)]; and stem cell therapies (n=23). Medians of the total number of planned inclusions per trial for these therapies were also included. Stem cells and lopinavir/ritonavir were the most frequently evaluated candidate therapies (23 and 15 trials respectively), whereas remdesivir was only tested in 5 trials but these trials had the highest median number of planned inclusions per trial (400, IQR 394-453). Most of the agents used in the different trials were chosen based on preclinical assessments of antiviral activity against SARS CoV and MERS CoV corona viruses.

The primary outcomes of the studies were clinical (66%); virological (23%); radiological (8%); or immunological (3%). The trials were classified as those that included patients with severe disease only; trials that included patients with moderate disease; and trials that included patients with severe or moderate disease.

12.70.3 Limitations

The trials evaluated provided incomplete information: 23% of these were phase IV trials but the bulk of the trials (54%) did not describe the phase of the study. Only 52% of the trials (n=60) reported treatment dose and only 34% (n=39) reported the duration. A lot of the trials included a small number of patients and the trials are still ongoing, therefore no insight was provided on the outcome of the trials.

12.70.4 Significance

Nonetheless, this review serves as framework for identifying COVID-19 related trials, which can be expanded upon as new trials begin at an accelerated rate as the disease spreads around the world.

12.70.5 Credit

This review was undertaken by K Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.71 ACE-2 Expression in the Small Airway Epithelia of Smokers and COPD Patients: Implications for COVID-19

12.71.1 Keywords

- chronic obstructive pulmonary disease
- COPD
- smokingE-2
- risk factors

12.71.2 Main Findings

In bronchial epithelial samples from 3 different cohorts of individuals, ACE-2 gene expression was found to be significantly increased in both COPD patients and smokers relative to healthy controls. Across all test subjects, ACE-2 gene expression was also highly correlated with decreased forced expiratory volume in 1 second (FEV1), which may explain the increased COVID-19 disease severity in COPD patients. Former smokers were also found to show decreased ACE2 expression relative to current smokers and had no significant difference when compared to non-smokers.

12.71.3 Limitations

While the upregulation of ACE-2 is an interesting hypothesis for COVID-19 disease severity in COPD patients, this study leaves many more unanswered questions than it addresses. Further studies are required to show whether the specific cell type isolated in these studies is relevant to the pathophysiology of COVID-19. Furthermore, there is no attempt to show whether that increased ACE-2 expression contributes to greater disease severity. Does the increased ACE-2 expression lead to greater infectivity with SARS-CoV-2? There is no mechanistic explanation for why ACE-2 levels are increased in COPD patients. The authors could also have considered the impact of co-morbidities and interventions such as corticosteroids or bronchodilators on ACE-2 expression. Finally, given the extensive sequencing performed, the authors could have conducted significantly more in-depth analyses into gene signature differences.

12.71.4 Significance

This study attempts to address an important clinical finding that both smokers and COPD patients show increased mortality from COVID-19. The novel finding that ACE-2 expression is induced in smokers and COPD patients suggests not only a mechanism for the clinical observation, but also highlights the potential benefit of smoking cessation in reducing the risk of severe COVID-19 disease.

12.71.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.72 Dynamic profile of severe or critical COVID-19 cases

12.72.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- progressive lymphopenia (PLD)
- T-lymphocytes
- clinical data
- co-infection
- influenza A

12.72.2 Main Findings

Authors evaluate clinical correlates of 10 patients (6 male and 4 female) hospitalized for severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). All patients required oxygen support and received broad spectrum antibiotics and 6 patients received anti-viral drugs. Additionally, 40% of patients were co-infected with influenza A. All 10 patients developed lymphopenia, two of which developed progressive lymphopenia (PLD) and died. Peripheral blood (PB) lymphocytes were analyzed – low CD4 and CD8 counts were noted in most patients, though CD4:CD8 ratio remained normal.

12.72.3 Limitations

The authors evaluated a small cohort of severe SARS-CoV-2 cases and found an association between T cell lymphopenia and adverse outcomes. However, this is an extremely small and diverse cohort (40% of patients were co-infected with influenza A). These findings need to be validated in a larger cohort. Additionally, the value of this data would be greatly increased by adding individual data points for each patient as well as by adding error bars to each of the figures.

12.72.4 Significance

This study provides a collection of clinical data and tracks evolution of T lymphocyte in 10 patients hospitalized for SARS-CoV-2, of which 4 patients were co-infected with influenza A.

12.72.5 Credit

This review was undertaken by Katherine Lindblad and Tamar Plitt as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.73 Association between Clinical, Laboratory and CT Characteristics and RT-PCR Results in the Follow-up of COVID-19 patients

12.73.1 Keywords

- COVID-19
- clinical
- lymphocyte
- CRP
- LDH
- HSST TNT
- PCR test
- readmission
- CT
- GGO
- disease progression

12.73.2 Study description

Data analyzed from 52 COVID-19 patients admitted and then discharged with COVID-19. Clinical, laboratory, and radiological data were longitudinally recorded with illness timecourse (PCR + to PCR-) and 7 patients (13.5%) were readmitted with a follow up positive test (PCR+) within two weeks of discharge.

12.73.3 Main Findings

- At admission:
 - The majority of patients had increased CRP at admission (63.5%).
 - LDH, and HSST TNT were significantly increased at admission.
 - Radiographic signs via chest CT showed increased involvement in lower lobes: right lower lobe (47 cases, 90.4%), left lower lobe (37 cases, 71.2%).
 - GGO (90.4%), interlobular septal thickening (42.3%), vascular enlargement (42.3%), and reticulation (11.5%) were most commonly observed.
- After negative PCR test (discharge):
 - CRP levels decreased lymphocyte counts (#/L) increased significantly (CD3+, CD3+/8+ and CD3+/4+) after negative PCR.
 - Consolidation and mixed GGO observed in longitudinal CT imaging w different extents of inflammatory exudation in lungs, with overall tendency for improvement (except 2/7 patients that were readmitted after discharge with re-positive test) after negative PCR.
- Seven patients repeated positive RT-PCR test and were readmitted to the hospital (9 to 17 day after initial discharge).

- Follow up CT necessary to monitor improvement during recovery and patients with lesion progression should be given more attention.
- Dynamic CT in addition to negative test essential in clinical diagnosis due to nasal swab PCR sampling bias (false-negatives).
- Increase in CRP occurred in 2 readmitted patients (and decr. in lymphocytes in one patient), but was not correlated with new lesions or disease progression vs. improvement (very low N).
- Patients readmitted attributed to false-negative PCR vs. re-exposure.

12.73.4 Limitations

Patients sampled in this study were generally younger (65.4% < 50 yrs) and less critically ill/all discharged. Small number of recovered patients (N=18). Time of follow up was relatively short.. Limited clinical information available about patients with re-positive test (except CRP and lymph tracking).

12.73.5 Extended Results

NOTE: Patients sampled in this study were generally younger (65.4% < 50 yrs) and less critically ill/all discharged. After two consecutive negative PCR tests, patients were discharged.

Clinical Results at Admission

- Median interval disease onset to admission (5 days, IQR: 3-7)
- Most common symptoms included fever, fatigue, dry cough, and expectoration.
- Fifteen patients had reduced lymphocyte counts (28.8%).
- No change in WBC or Neutrophil counts.
- **The majority of patients had increased CRP at admission (63.5%).**
- **LDH, and HSST TNT were significantly increased at admission.**
- Fibrinogen was trending high though not significant.
- No major changes in liver function observed.
- **Radiographic signs via chest CT showed increased involvement in lower lobes: right lower lobe (47 cases, 90.4%), left lower lobe (37 cases, 71.2%).**
- **GGO (90.4%), interlobular septal thickening (42.3%), vascular enlargement (42.3%), and reticulation (11.5%)** were most commonly observed.

Change in Clinical Results following Negative Test

- **CRP levels decreased after negative PCR.**
- **Lymphocyte counts (#/L) increased significantly (CD3+, CD3+/8+ and CD3+/4+).**
- No significant change to CD4/8 ratio.
- LDH, HSST TNT, and Fibronectin remained high throughout, though range observed decreased over time.
- **Consolidation and mixed GGO observed in longitudinal CT imaging.**
- **Patients showed different extents of inflammatory exudation in lungs, with overall tendency for improvement (except 2/7 patients that were readmitted after discharge with re-positive test).**

Patients Readmitted with PCR+ test

- **Seven patients repeated positive RT-PCR test and were readmitted to the hospital (9 to 17 day after initial discharge).**
- Improvement during readmission in 4 patients and observation of segmental progression CT in 2 patients (2/18 or 11.1% - re-positive 9 and 10 days post-discharge).
- Two patients showed new GGO, while others improved greatly.
- **Follow up CT necessary to monitor improvement during recovery and patients with lesion progression should be given more attention.**
- **Dynamic CT in addition to negative test essential in clinical diagnosis due to nasal swab PCR sampling bias (false-negatives).**
- **Increase in CRP occurred in 2 readmitted patients (and decr. in lymphocytes in one patient), but was not correlated with new lesions or disease progression vs. improvement (very low N).**

12.73.6 Significance

Study tracked key clinical features associated with disease progression, recovery, and determinants of clinical diagnosis/management of COVID-19 patients.

12.73.7 Credit

This review was undertaken by Natalie Vaninov as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.74 An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 and multiple 2 endemic, epidemic and bat coronavirus

Sheahan et al. *bioRxiv*. [2041]

12.74.1 Keywords

- Treatment
- Antiviral
- Broad spectrum antiviral
- ribonucleoside analog β-D-N4 30 hydroxycytidine (NHC)
- Remdesivir

12.74.2 Main Findings

β-D-N4 30 -hydroxycytidine (NHC, EIDD-1931) is an orally bioavailable ribonucleoside with antiviral activity against various RNA viruses including Ebola, Influenza and CoV. NHC activity introduced mutations in the viral (but not cellular) RNA in a dose dependent *manner* that directly correlated with a decrease in viral titers. Authors show that NHC inhibited multiple genetically distinct Bat-CoV viruses in human primary epithelial cells *without affecting cell viability even at high concentrations (100 µM)*. Prophylactic oral administration of NHC in C57BL/6 mice reduce lung titers of SARS-CoV and prevented weight loss and hemorrhage. Therapeutic administration of NHC in C57BL/6 mice 12 hours post infected with SARS-CoV reduced acute lung injury, viral titer, and lung hemorrhage. The degree of clinical benefit was dependent on the time of treatment initiation post infection. The authors also demonstrate that NHC reduces MERS-CoV infection titers, pathogenesis, and viral RNA in prophylactic and therapeutic settings.

12.74.3 Limitations

Most of the experiments were conducted using MERS-CoV, and SARS-CoV and a few experiments were conducted using other strains of CoV as opposed to SARS-CoV-2. The authors note the core residues that make up the RNA interaction sites (which constitutes the NHC interaction sites) are highly conserved among CoV and because of this conservation their understanding is that NHC can inhibit a broad-spectrum of CoV including SARS-CoV-2.

The increased viral mutation rates associated with NHC activity may have adverse effects if mutations cause the virus to become drug resistant, more infectious or speed-up immune evasion. *In addition, the temporal diminishing effectiveness of NHC on clinical outcome when NHC was used therapeutically is concerning. However, the longer window (7-10 days) for clinical disease onset in human patients from the time of infection compared to that of mice (24-48 hours), may associate with increased NHC effectiveness in the clinic.*

12.74.4 Significance

Prophylactic or therapeutic oral administration of NHC reduces lung titers and prevents acute lung failure in C57BL/6 mice infected with CoV. Given its *broad-spectrum antiviral activity*, NHC could turn out to be a useful drug for treating current, emerging and future corona virus outbreaks. ##### Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.75 Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs

Sangeun Jeon et al. [[2042](#)]

12.75.1 Keywords

- COVID-19
- SARS CoV-2
- antiviral drugs
- niclosamide
- ciclesonide

12.75.2 Main Findings

A panel of ~3,000 FDA- and IND-approved antiviral drugs were previously screened for inhibitory efficacy against SARS CoV, a coronavirus related to the novel coronavirus SARS CoV-2 (79.5%) homology. 35 of these drugs along with another 15 (suggested by infectious disease specialists) were tested in vitro for their ability to inhibit SARS CoV-2 infectivity of Vero cells while preserving cell viability. The infected cells were scored by immunofluorescence analysis using an antibody against the N protein of SARS CoV-2. Chloroquine, lopinavir and remdesivir were used as reference drugs.

Twenty four out of 50 drugs exhibited antiviral activity with IC₅₀ values ranging from 0.1-10µM. Among these, two stood out: 1) the anti-helminthic drug niclosamide which exhibited potent antiviral activity against SARS CoV-2 (IC₅₀=0.28 µM). The broad-spectrum antiviral effect of niclosamide against SARS and MERS-CoV have been previously documented and recent evidence suggests that it may inhibit autophagy and reduce MERS CoV replication. 2) Ciclesonide, a corticosteroid used to treat asthma and allergic rhinitis, also exhibited antiviral efficacy but with a lower IC₅₀ (4.33µM) compared to niclosamide. The antiviral effects of ciclesonide were directed against NSP15, a viral ribonucleic acid (RNA) dependent RNA polymerase which is the molecular target of this drug.

12.75.3 Limitations

The drugs were tested against SARS CoV-2 infectivity in vitro only, therefore preclinical studies in animals and clinical trials in patients will be needed for validation of these drugs as therapeutic agents for COVID-19. In addition, niclosamide exhibits low adsorption pharmokinetically which could be alleviated with further development of drug formulation to increase effective delivery of this drug to target tissues. Nonetheless, niclosamide and ciclesonide represent promising therapeutic agents against SARS CoV-2 given that other compounds tested in the same study including favipiravir (currently used in clinical trials) and atazanavir (predicted as the most potent antiviral drug by AI-inference modeling) did not exhibit antiviral activity in the current study.

12.75.4 Credit

This review was undertaken by K Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.76 Respiratory disease and virus shedding in rhesus macaques inoculated with SARS-CoV-2

Munster et al. *bioRxiv*. [[2043](#)]

12.76.1 Keywords

animal model, pulmonary infiltrates, dynamic of antibody response, cytokine

12.76.2 Main Findings

Inoculation of 8 Rhesus macaques with SARS-CoV-2, which all showed clinical signs of infection (respiratory pattern, reduced appetite, weight loss, elevated body temperature) resulting in moderate, transient disease. Four animals were euthanized at 3 dpi, the 4 others at 21 dpi. Study of viral loads in different organs showed that nose swab and throat swabs were the most sensitive, with broncho-alveolar lavage. Interstitial pneumonia was visible in radiographies and at the histological scale too. Clinically, the macaques had similar symptoms as described in human patients with moderate disease.

Viral shedding was consistently detected in nose swabs and throat swabs immediately after infection but less consistent thereafter which could reflect virus administration route (intranasal, oral). Bronchoalveolar lavages performed as a measure of virus replication in the lower respiratory tract on animals maintained for 21 days, contained high viral loads in 1 and 3 dpi. The majority of the animals exhibited pulmonary edema and mild to moderate interstitial pneumonia on terminal bronchioles. In addition to the lung, viral RNA could also be detected throughout the respiratory track where viral replication mainly occurred.

Immunologic responses included leukocytosis, neutrophilia, moncytosis and lymphopenia in the majority of the animals at 1 dpi. Lymphocytes and monocytes re-normalized at 2 dpi. Neutrophils declined after 3 dpi and

through 10dpi after which they started to recover. After infection, serum analysis revealed significant increases in **IL1ra, IL6, IL10, IL15, MCP-1, MIP-1b, but quick normalization** (3dpi). **Antibody response started around 7dpi, and the antibody titers stayed elevated until 21dpi** (day of animal euthanasia).

12.76.3 Limitations

The macaques were inoculated via a combination of intratracheal, intranasal, ocular and oral routes, which might not reproduce how humans get infected. Maybe this can lead to different dynamics in the host immune response. Also, the authors noted that the seroconversion was not directly followed by a decline in viral loads, as observed in covid19 patients.

12.76.4 Significance

This work confirms that rhesus macaques can be a good model to study Covid-19, as it has been shown by other groups [[2017](#),[2031](#),[2044](#)]. While these experiments recapitulate moderate COVID-19 in humans, the mode of inoculation via a combination of intratracheal, intranasal, ocular and oral routes, might not reproduce how humans get infected and may lead to different dynamics in the host immune response. For example, the authors noted that the seroconversion was not directly followed by a decline in viral loads, as observed in COVID-19 patients. Therefore, it will be interesting to follow their antibody titers longer and further assess the possibility/effect of reinfection in these macaques. It is essential to be able to understand the dynamic of the disease and associated immune responses, and to work on vaccine development and antiviral drug testing.

12.76.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.77 ACE2 Expression is Increased in the Lungs of Patients with Comorbidities Associated with Severe COVID-19

[[2045](#)]

12.77.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- lung
- comorbidities
- histone
- epigenetics

12.77.2 Main Findings

- Transcriptomic analysis using systems-level meta-analysis and network analysis of existing literature to determine ACE2 regulation in patients who have frequent COVID-19 comorbidities [eg- cardiovascular diseases, familial pulmonary hypertension, cancer].
- Enrichment analyses indicated pathways associated with inflammation, metabolism, macrophage autophagy, and ER stress.
- ACE2 higher in adenocarcinoma compared to adjacent normal lung; ACE2 higher in COPD patients compared to normal.
- Co-expression analysis identified genes important to viral entry such as RAB1A, ADAM10, HMGBs, and TLR3 to be associated with ACE2 in diseased lungs.
- ACE2 expression could be potentially regulated by enzymes that modify histones, including HAT1, HDAC2, and KDM5B.

12.77.3 Limitations:

- Not actual CoVID-19 patients with co-morbidities, so interpretations in this study need to be confirmed by analyzing upcoming transcriptomics from CoVID-19 patients having co-morbidity metadata.
- As mentioned by authors, study does not look at diabetes and autoimmunity as risk factors in CoVID-19 patients due to lack of data; would be useful to extend such analyses to those datasets when available.
- Co-expression analysis is perfunctory and needs validation-experiments especially in CoVID-19 lung samples to mean anything.
- Epigenomic analyses are intriguing but incomplete, as existence of histone marks does not necessarily mean occupancy. Would be pertinent to check cell-line data (CCLE) or actual CoVID-19 patient samples to confirm ACE2 epigenetic control.

12.77.4 Significance

- Study implies vulnerable populations have ACE2 upregulation that could promote CoVID-19 severity. Shows important data-mining strategy to find gene-networks associated with ACE2 upregulation in co-morbid patients.
- Several of the genes co-upregulated with ACE2 in diseased lung might play an important role in CoVID-19 and can be preliminary targets for therapeutics
- If in silico findings hold true, epigenetic control of ACE2 expression could be a new target for CoVID-19 therapy with strategies such as KDM5 demethylases.

12.77.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.78 Meplazumab treats COVID-19 pneumonia: an open-labelled, concurrent controlled add-on clinical trial

Bian et al. *medRxiv*. [2046]

12.78.1 Keywords

- Meplazumab
- CD147
- humanized antibody
- clinical trial

12.78.2 Main Findings

This work is based on previous work by the same group that demonstrated that SARS-CoV-2 can also enter host cells via CD147 (also called Basigin, part of the immunoglobulin superfamily, is expressed by many cell types) consistent with their previous work with SARS-CoV-1.¹ A prospective clinical trial was conducted with 17 patients receiving Meplazumab, a humanized anti-CD147 antibody, in addition to all other treatments. 11 patients were included as a control group (non-randomized).

They observed a faster overall improvement rate in the Meplazumab group (e.g. at day 14 47% vs 17% improvement rate) compared to the control patients. Also, virological clearance was more rapid with median of 3 days in the Meplazumab group vs 13 days in control group. In laboratory values, a faster normalization of lymphocyte counts in the Meplazumab group was observed, but no clear difference was observed for CRP levels.

12.78.3 Limitations

While the results from the study are encouraging, this study was non-randomized, open-label and on a small number of patients, all from the same hospital. It offers evidence to perform a larger scale study. Selection bias as well as differences between treatment groups (e.g. age 51yo vs 64yo) may have contributed to results. The authors mention that there was no toxic effect to Meplazumab injection but more patient and longer-term studies are necessary to assess this.

12.78.4 Significance

These results seem promising as for now there are limited treatments for Covid-19 patients, but a larger cohort of patient is needed. CD147 has already been described to facilitate HIV [2047], measles virus [2048], and malaria [2049] entry into host cells. This group was the first to describe the CD147-spike route of SARS-CoV-2 entry in host cells [2020] p147. Indeed, they had previously shown in 2005 that SARS-CoV could enter host cells via this transmembrane protein [2050]. Further biological understanding of how SARS-CoV-2 can enter host cells and how this integrates with ACE2R route of entry is needed. Also, the specific cellular targets of the anti-CD147 antibody need to be assessed, as this protein can be expressed by many cell types and has been shown to be involved in leukocytes aggregation [2051]. Lastly, Meplazumab is not a commercially-available drug and requires significant health resources to generate and administer which might prevent rapid development and use.

12.78.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.79 Potent human neutralizing antibodies elicited 1 by SARS-CoV-2 infection

Ju et al. *bioRxiv*. [184]

12.79.1 Keywords

- monoclonal antibodies
- neutralization
- antibody cross-reactivity
- Receptor Binding Domain

12.79.2 Main Findings

In this study the authors report the affinity, cross reactivity (with SARS-CoV and MERS-CoV virus) and viral neutralization capacity of 206 monoclonal antibodies engineered from isolated IgG memory B cells of patients suffering from SARS-CoV-2 infection in Wuhan, China. All patients but one recovered from disease. Interestingly, the patient that did not recover had less SARS-CoV-2 specific B cells circulating compared to other patients.

Plasma from all patients reacted to trimeric Spike proteins from SARS-CoV-2, SARS-CoV and MERS-CoV but no HIV BG505 trimer. Furthermore, plasma from patients recognized the receptor binding domain (RBD) from SARS-CoV-2 but had little to no cross-reactivity against the RBD of related viruses SARS-CoV and MERS-CoV, suggesting significant differences between the RBDs of the different viruses. Negligible levels of cross-neutralization using pseudoviruses bearing Spike proteins of SARS-CoV-2, SARS-CoV or MERS-CoV, were observed, corroborating the ELISA cross-reactivity assays on the RBDs.

SARS-CoV-2 RBD specific B cells constituted 0.005-0.065% of the total B cell population and 0.023-0.329% of the memory subpopulation. SARS-CoV specific IgG memory B cells were single cell sorted to sequence the antibody genes that were subsequently expressed as recombinant IgG1 antibodies. From this library, 206 antibodies with different binding capacities were obtained. No discernible patterns of VH usage were found in the 206 antibodies suggesting immunologically distinct responses to the infection. Nevertheless, most high-binding antibodies were derived by clonal expansion. Further analyses in one of the patient derived clones, showed that the antibodies from three different timepoints did not group together in phylogenetic analysis, suggesting selection during early infection.

Using surface plasmon resonance (SPR) 13 antibodies were found to have 10^8 to 10^{-9} dissociation constants (Kd). Of the 13 antibodies, two showed 98-99% blocking of SARS-CoV-2 RBD-ACE2 receptor binding in competition assays. Thus, low Kd values alone did not predict ACE2 competing capacities. Consistent with competition assays the two antibodies that show high ACE2 blocking (P2C-2F6 and P2C-1F11) were the most capable of neutralizing pseudoviruses bearing SARS-CoV-2 spike protein (IC_{50} of 0.06 and 0.03 $\mu\text{g/mL}$, respectively). Finally, using SPR the neutralizing antibodies were found to recognize both overlapping and distinct epitopes of the RBD of SARS-CoV-2.

12.79.3 Limitations

1. Relatively low number of patients
 - a. No significant conclusion can be drawn about the possible > correlation between humoral response and disease severity
2. *In vitro* Cytopathic Effect Assay (CPE) for neutralization activity
 - a. Huh7 cells were used in neutralization assays with > pseudoviruses, and they may not entirely mimic what happens in > the upper respiratory tract
 - b. CPE assay is not quantitative
3. Duplicated panel in Figure 4C reported (has been fixed in version 2)

12.79.4 Significance

This paper offers an explanation as to why previously isolated antibodies against SARS-CoV do not effectively block SARS-CoV-2. Also, it offers important insight into the development of humoral responses at various time points during the first weeks of the disease in small but clinically diverse group of patients. Furthermore, it provides valuable information and well characterized antibody candidates for the development of a recombinant antibody treatment for SARS-CoV-2. Nevertheless, it also shows that plasmapheresis might have variability in its effectiveness, depending on the donor's antibody repertoire at the time of donation.

12.79.5 Credit

Review by Jovani Catalan-Dibene as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.80 Characterisation of the transcriptome and proteome of SARS-CoV-2 using direct RNA sequencing and tandem mass spectrometry reveals evidence for a cell passage induced in-frame deletion in the spike glycoprotein that removes the furin-like cleavage site.

Davidson et al. [2052]

12.80.1 Keywords

- Transcription
- RNA-seq
- proteomics
- mass spec
- furin cleavage site
- mutation
- pathogenicity

12.80.2 Main Findings

The authors performed long read RNA sequencing using an Oxford Nanopore MinION as well as tandem mass spec (MS) on Vero cells (a cell line derived from kidney cells of the African green monkey that is deficient in interferon) infected with SARS-CoV-2.

The authors found that passage of the virus in Vero cells gave rise to a spontaneous 9 amino acid deletion (679-NSPRRARSV-687 to I) in the spike (S) protein. The deleted sequence overlaps a predicted furin cleavage site at the S1 / S2 domain boundary that is present in SARS-CoV-2 but not SARS-CoV or the closely related bat coronavirus RaTG13, which are cleaved at S1 / S2 by other proteases [14]. Furin cleavage sites at similar positions in other viruses have been linked to increased pathogenicity and greater cell tropism [2053]. Loss of this site in SARS-CoV-2 has also already been shown to increase viral entry into Vero but not BHK cells (which are also interferon deficient) [23]. The authors therefore make an important contribution in demonstrating that passage in Vero cells may lead to spontaneous loss of a key pathogenicity-conferring element in SARS-CoV-2.

12.80.3 Limitations

As the authors note, a similar study posted earlier by Kim et al., which also passed SARS-CoV-2 in Vero cells, did not identify any loss in the S protein furin cleavage site [2054]. It therefore remains to be determined how likely it is that this mutation spontaneously arises. A quantitative investigation using multiple experimental replicas to understand the spontaneous viral mutation rate at this site and elsewhere would be informative. Also, the mechanistic basis for the higher viral fitness conferred by loss of the furin cleavage site in Vero cells – but, evidently, not *in vivo* in humans, as this site is maintained in all currently sequenced circulating isolates - remains to be understood.

Due to the high base-call error rate of MinION sequencing, the authors' bioinformatic pipeline required aligning transcripts to a reference to correct sequencing artifacts. This presumably made it difficult or impossible to identify other kinds of mutations, such as single nucleotide substitutions, which may occur even more frequently than the deletions identified in this work. Pairing long read sequencing with higher-accuracy short-read sequencing may be one approach to overcome this issue.

12.80.4 Significance

As the authors suggest, animal studies using live virus challenge may need to periodically verify the genomic integrity of the virus, or potentially risk unknowingly using a likely less-pathogenic variant of the virus.

More broadly, the results emphasize the complexity and plasticity of the SARS-CoV-2 viral transcriptome and proteome. For example, the authors found multiple versions of transcripts encoding the nucleocapsid (N) protein, each with different small internal deletions, some of which were verified for translation by MS. A number of peptides arising from translation of unexpected rearrangements of transcripts were also detected. Additionally, the authors identified phosphorylation of a number of viral proteins (N, M, ORF 3a, nsp3, nsp9, nsp12 and S). For any cases where these have functional consequences, targeting the kinases responsible could be an avenue for drug development. Understanding the functional consequences of the mutations, transcript variations, and post translational modifications identified in this study will be important future work.

12.80.5 Credit

This review was undertaken by Tim O'Donnell, Maria Kuksin as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.81 A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug- Repurposing

Gordon et al. *bioRxiv* [2055]

12.81.1 Keywords

- protein-protein interactions
- mass spectrometry
- drug targets

12.81.2 Main Findings

Gordon et al cloned, tagged and expressed 26 of the 29 SARS-CoV-2 proteins individually in HEK293T cells and used mass spectrometry to identify protein-protein interactions. They identified 332 viral-host protein-protein interactions. Furthermore, they used these interactions to identify 66 existing drugs known to target host proteins or host pathways (eg SARS-CoV-2 N and Orf8 proteins interact with proteins regulated by the mTOR pathway, so mTOR inhibitors Silmitasertib and Rapamycin are possible drug candidates).

12.81.3 Limitations

The main limitation of the study stems from the reductionist model: overexpression of plasmids encoding individual viral proteins in HEK293T cells. This precludes any interactions between the viral proteins, or the combined effects of multiple proteins on the host, as they are expressed individually. Moreover, HEK293T cells come from primary embryonic kidney and therefore might not reflect how SARS-CoV-2 interacts with its primary target, the lung. However, the authors found that the proteins found to interact with viral proteins in their experiments are enriched in lung tissue compared to HEK293Ts.

12.81.4 Significance

The authors provide a “SARS-CoV-2 interaction map,” which may provide potential hypotheses as to how the virus interacts with the host. Further, they identified existing drugs that could disrupt these host-viral interactions and curb SARS-CoV-2 infection. Although these interactions have not been validated, this paper acts as a valuable resource.

12.81.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.82 First Clinical Study Using HCV Protease Inhibitor Danoprevir to Treat Naïve and Experienced COVID-19 Patients

Chen et al. *medRxiv*. [2056]

12.82.1 Keywords

- Clinical study
- HCV protease inhibitor

- Danoprevir
- Ritonavir
- Covid19 treatment

12.82.2 Main Findings

The authors treated 11 Covid-19 patients with Danoprevir, a commercialized HCV protease inhibitor [2057]^(p4), boosted by ritonavir [2058], a CYP3A4 inhibitor (which enhances the plasma concentration and bioavailability of Danoprevir). Two patients had never received anti-viral therapy before (=naïve), whereas nine patients were on Lopinavir/Ritonavir treatment before switching to Danoprevir/Ritonavir (=experienced). The age ranged from 18 to 66yo.

Naïve patients that received Danoprevir/Ritonavir treatment had a decreased hospitalization time. Patients treated with Lopinavir/Ritonavir did not have a negative PCR test, while after switching to Danoprevir/Ritonavir treatment, the first negative PCR test occurred at a median of two days.

12.82.3 Limitations

The results of the study are very hard to interpret as there is no control group not receiving Danoprevir/Ritonavir treatment. This was especially true in naïve patients who seemed to have more mild symptoms before the start of the study and were younger (18 and 44yo) compared to the experienced patients (18 to 66yo). The possibility that the patients would have recovered without Danoprevir/Ritonavir treatment cannot be excluded.

12.82.4 Significance

The authors of the study treated patients with Danoprevir, with the rational to that this is an approved and well tolerated drug for HCV patients [2058], and that it could also target the protease from SARS-CoV-2 (essential for viral replication and transcription). Indeed, homology modelling data indicated that HCV protease inhibitors have the highest binding affinity to Sars-Cov2 protease among other approved drugs [2059].

While this study shows that the combination of Danoprevir and Ritonavir might be beneficial for Covid-19 patients, additional clinical trials with more patients and with better methodology (randomization and control group) are needed to make further conclusions.

12.82.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.83 Efficacy of hydroxychloroquine in patients with COVID-19: results of a randomized clinical trial

[1021]

12.83.1 Keywords

- hydroxychloroquine

12.83.2 Study Description

This is a randomized clinical trial of hydroxychloroquine (HCQ) efficacy in the treatment of COVID-19. From February 4 – February 28, 2020 142 COVID-19 positive patients were admitted to Renmin Hospital of Wuhan University. 62 patients met inclusion criteria and were enrolled in a double blind, randomized control trial, with 31 patients in each arm.

Inclusion criteria:

1. Age \geq 18 years
2. Positive diagnosis COVID-19 by detection of SARS-CoV-2 by RT-PCR
3. Diagnosis of pneumonia on chest CT
4. Mild respiratory illness, defined by $\text{SaO}_2/\text{SPO}_2$ ratio $> 93\%$ or $\text{PaO}_2/\text{FIO}_2$ ratio $> 300 \text{ mmHg}$ in hospital room conditions (Note: relevant clinical references described below.)
 - a. Hypoxia is defined as an SpO_2 of 85-94%; severe hypoxia $< 85\%$.
 - b. The $\text{PaO}_2/\text{FIO}_2$ (ratio of arterial oxygen tension to fraction of inspired oxygen) is used to classify the severity of acute respiratory distress syndrome (ARDS). Mild ARDS has a $\text{PaO}_2/\text{FIO}_2$ of 200-300 mmHg, moderate is 100-200, and severe < 100 .
5. Willing to receive a random assignment to any designated treatment group; not participating in another study at the same time

Exclusion criteria:

1. Severe or critical respiratory illness (not explicitly defined, presumed to be respiratory function worse than outlined in inclusion criteria); or participation in trial does not meet patient's maximum benefit or safe follow up criteria
2. Retinopathy or other retinal diseases
3. Conduction block or other arrhythmias

4. Severe liver disease, defined by Child-Pugh score \geq C or AST > twice the upper limit
5. Pregnant or breastfeeding
6. Severe renal failure, defined by eGFR \leq 30 mL/min/1.73m², or on dialysis
7. Potential transfer to another hospital within 72h of enrollment
8. Received any trial treatment for COVID-19 within 30 days before the current study

All patients received the standard of care: oxygen therapy, antiviral agents, antibacterial agents, and immunoglobulin, with or without corticosteroids. Patients in the HCQ treatment group received additional oral HCQ 400 mg/day, given as 200 mg 2x/day. HCQ was administered from days 1-5 of the trial. The primary endpoint was 5 days post enrollment or a severe adverse reaction to HCQ. The primary outcome evaluated was time to clinical recovery (TTCR), defined as return to normal body temperature and cough cessation for > 72h. Chest CT were imaged on days 0 and 6 of the trial for both groups; body temperature and patient reports of cough were collected 3x/day from day 0 – 6. The mean age and sex distribution between the HCQ and control arms were comparable.

12.83.3 Main Findings

There were 2 patients showing mild secondary effects of HCQ treatment. More importantly, while 4 patients in the control group progressed to severe disease, none progressed in the treatment group.

TTCR was significantly decreased in the HCQ treatment arm; recovery from fever was shortened by one day (3.2 days control vs. 2.2 days HCQ, p = 0.0008); time to cessation of cough was similarly reduced (3.1 days control vs. 2.0 days HCQ, p = 0.0016).

Overall, it appears that HCQ treatment of patients with mild COVID-19 has a modest effect on clinical recovery (symptom relief on average 1 day earlier) but may be more potent in reducing the progression from mild to severe disease.

12.83.4 Limitations

This study is limited in its inclusion of only patients with mild disease, and exclusion of those on any treatment other than the standard of care. It would also have been important to include the laboratory values of positive RT-PCR detection of SARS-CoV-2 to compare the baseline and evolution of the patients' viral load.

12.83.5 Limitations

Despite its limitations, the study design has good rigor as a double blind RCT and consistent symptom checks on each day of the trial. Now that the FDA has approved HCQ for treatment of COVID-19 in the USA, this study supports the efficacy of HCQ use early in treatment of patients showing mild symptoms, to improve time to clinical recovery, and possibly reduce disease progression. However, most of the current applications of HCQ have been in patients with severe disease and for compassionate use, which are out of the scope of the findings presented in this trial. Several additional clinical trials to examine [hydroxychloroquine](#) are now undergoing; their results will be critical to further validate these findings.

12.83.6 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Structure-based modeling of SARS-CoV-2 peptide/HLA-A02 antigens

<https://doi.org/10.1101/2020.03.23.004176>

Immunology keywords:

CoVID-19, 2019-nCoV, SARS-CoV-2, comparative, homology, peptide, modeling, simulation, HLA-A, antigen

Summary of Findings:

- The authors utilize homology modeling to identify peptides from the SARS-CoV-2 proteome that potentially bind HLA-A*02:01.
- They utilize high-resolution X-ray structures of peptide/MHC complexes on Protein Data Bank, substitute homologous peptides with SARS-CoV-2 peptides, and calculate MHC/SARS-CoV-2 peptide Rosetta binding energy.
- They select MHC/SARS-CoV-2 complex models with highest binding energy for further study and publish models in an online database (<https://rosettamhc.chemistry.ucsc.edu>).

Limitations:

- The authors only utilize computational methods and predicted SARS-CoV-2 peptides must be validated experimentally for immunogenicity and clinical response.
- Due to computational burden and limited availability of high resolution X-ray structures on PDB, authors only simulate 9-mer and 10-mer peptide binding to HLA-A*02:01.
- Since the authors compare select existing X-ray structures as a starting point, backbone conformations that deviate significantly between test and template peptides are not captured. Furthermore, Rosetta modeling

protocols do not capture all possible structures and binding energy scoring does not fully recapitulate fundamental forces.^{1,2}

Importance/Relevance:

- The authors identify and publish high-scoring SARS-CoV-2 peptides that may direct a targeted, experimental validation approach toward a COVID-19 vaccine.
- The authors utilize Rosetta simulation to further filter results from NetMHCpan 4.0, supporting machine learning prediction with structural analysis.
- The authors develop RosettaMHC, a computationally efficient method of leveraging existing X-ray structures for identification of immunogenic peptides.

References:

1. Bender, B. J., Cisneros, A., 3rd, Duran, A. M., Finn, J. A., Fu, D., Lokits, A. D., . . . Moretti, R. (2016). Protocols for Molecular Modeling with Rosetta3 and RosettaScripts. *Biochemistry*, 55(34), 4748-4763. doi:10.1021/acs.biochem.6b00444
2. Alford, R. F., Leaver-Fay, A., Jeliazkov, J. R., O'Meara, M. J., DiMaio, F. P., Park, H., . . . Gray, J. J. (2017). The Rosetta All-Atom Energy Function for Macromolecular Modeling and Design. *J Chem Theory Comput*, 13(6), 3031-3048. doi:10.1021/acs.jctc.7b00125

Review by Jonathan Chung as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn school of medicine, Mount Sinai.

12.84 Serology characteristics of SARS-CoV-2 infection since the exposure and post symptoms onset

Lou et al. *medRxiv*. [2060]

12.84.1 Keywords

- Seroconversion rate
- Total Antibody
- Ab
- IgG and IgM
- antibody

12.84.2 Main Findings

Currently, the diagnosis of SARS-CoV-2 infection entirely depends on the detection of viral RNA using polymerase chain reaction (PCR) assays. False negative results are common, particularly when the samples are collected from upper respiratory. Serological detection may be useful as an additional testing strategy. In this study the authors reported that a typical acute antibody response was induced during the SARS-CoV-2 infection, which was discussed earlier¹. The seroconversion rate for Ab, IgM and IgG in COVID-19 patients was 98.8% (79/80), 93.8% (75/80) and 93.8% (75/80), respectively. The first detectable serology marker was total antibody followed by IgM and IgG, with a median seroconversion time of 15, 18 and 20 days-post exposure (d.p.e) or 9, 10- and 12-days post-onset (d.p.o). Seroconversion was first detected at day 7d.p.e in 98.9% of the patients. Interestingly they found that viral load declined as antibody levels increased. This was in contrast to a previous study [2000], showing that increased antibody titers did not always correlate with RNA clearance (low number of patient sample).

12.84.3 Limitations

Current knowledge of the antibody response to SAR-CoV-2 infection and its mechanism is not yet well elucidated. Similar to the RNA test, the absence of antibody titers in the early stage of illness could not exclude the possibility of infection. A diagnostic test, which is the aim of the authors, would not be useful at the early time points of infection but it could be used to screen asymptomatic patients or patients with mild disease at later times after exposure.

12.84.4 Significance

Understanding the antibody responses against SARS-CoV2 is useful in the development of a serological test for the diagnosis of COVID-19. This manuscript discussed acute antibody responses which can be deducted in plasma for diagnostic as well as prognostic purposes. Thus, patient-derived plasma with known antibody titers may be used therapeutically for treating COVID-19 patients with severe illness.

12.84.5 Credit

This review was undertaken and edited by Konstantina A as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.85 SARS-CoV-2 launches a unique transcriptional signature from in vitro, ex vivo, and in vivo systems

Blanco-Melo et al. *bioRxiv*. [2061]

12.85.1 Keywords

- host cellular response

- host-pathogen interaction
- type I interferon
- type III interferon
- inflammation
- RNA-seq
- comparative analysis

12.85.2 Main Findings

Given the high mortality rate of SARS-CoV-2 relative to other respiratory viruses such seasonal IAV and RSV, there may be underlying host-pathogen interactions specific to SARS-CoV-2 that predispose to a worse clinical outcome. Using *in vivo*, *ex vivo*, and *in vitro* systems, the authors profiled host cell transcriptional responses to SARS-CoV-2 and to other common respiratory viruses (seasonal IAV and RSV). SARS-CoV-2 infection *in vitro* led to an induction of type I interferon response signaling and the upregulation of cytokine/chemokines transcripts. In comparison with IAV and RSV infection, SARS-CoV-2 *in vitro* appears to uniquely induce less type I and type III interferon expression and higher levels of two cytokines previously implicated in respiratory inflammation. Lastly, *in vivo* data from ferrets showed a reduced induction of cytokines and chemokines by SARS-CoV-2 infection relative to IAV infection.

12.85.3 Limitations

While these results are promising, there are several key weaknesses of this paper. 1) As the authors point out, there is an undetectable level of SARS-CoV-2 putative receptor (ACE2) and protease (TMPRSS2) expression in the lung epithelial cell line used for the *in vitro* studies. This raises the important question of whether viral replication actually occurs in any of the models used, which may explain the lack of interferon production observed *in vitro* in SARS-CoV-2 treated cells. Further studies characterizing viral titers across timepoints are needed. 2) Furthermore, these studies only characterize the host response at a single dose and timepoint per virus, and it is unclear why these doses/timepoints were chosen. This leaves open the possibility that the observed differences between viruses could be due to differences in dose, timing, host response, or a combination of all of these. 3) It is unclear whether ferrets are productively infected, which cell types are infected, and the extent/timing of the clinical course of infection. Moreover, the *in vitro* and *in vivo* data do not strongly correlate and the reasons for this are unclear.

12.85.4 Significance

This paper describes potentially unique transcriptional signatures of host cells exposed to SARS-CoV-2. If validated, these findings may help explain clinical outcomes and could be targeted in future therapeutic interventions.

12.85.5 Potential Conflicts of Interest Disclosure

The reviewers are also researchers at the Icahn School of Medicine at Mount Sinai.

12.85.6 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.86 A New Predictor of Disease Severity in Patients with COVID-19 in Wuhan, China

Zhou et al. *bioRxiv*. [2062]

12.86.1 Keywords

- disease severity
- clinical data
- Neutrophils/lymphocytes ratio
- CRP
- D-dimer

12.86.2 Main Findings

377 hospitalized patients were divided into two groups: severe and non-severe pneumonia. The laboratory results of their first day of admission were retrospectively analyzed to identify predictors of disease severity.

After adjusting for confounding factors from chronic comorbidities (such as high blood pressure, type 2 diabetes, coronary heart disease, and chronic obstructive pulmonary disease), the independent risk factors identified for severe pneumonia were **age**, the **ratio of neutrophil/lymphocytes counts**, **CRP** and **D-dimer** levels.

To further increase the specificity and sensibility of these markers, they showed that their multiplication **[(Neutrophil/lymphocyte count) * CRP * D-dimer]** was a better predictor of disease severity, with higher sensitivity (95.7%) and specificity (63.3%), with a cutoff value of 2.68.

12.86.3 Limitations

This study included 377 hospitalized patients. Among them, 45.6% patients tested positive for SARS-CoV-2 nucleic acid test results, and others were included in the study based on clinically diagnosis even if the molecular diagnosis was negative. Thus, additional studies are needed to verify this on a larger number of covid-19 certified patients and the cutoff value might be adjusted. Also, all the patients that did not have the clinical characteristics of severe pneumonia were included in the non-severe pneumonia group, but usually patients are also divided into moderate and mild disease.

Also, studying different subset of lymphocytes could lead to a more specific predictor. Another study showed that the neutrophils to CD8+ T cells ratio was a strong predictor of disease severity [1970]. Another more precise study showed that the percentage of helper T cells and regulatory T cells decrease

but the percentage of naïve helper T cells increases in severe cases [1963]. Taking these subpopulations into account might make the predictor more powerful.

Other studies also noted an inverse correlation between disease severity and LDH [2004] or IL6 [2013] levels, but the authors here do not discuss LDH nor IL6 levels, although this could help to strengthen the predictor.

The study is based on the results obtained on the first day of admission, studying the dynamic of the changes in patients might also be interesting to better predict disease severity.

12.86.4 Significance

This study confirms that the neutrophil to lymphocyte ratio can be a predictor of disease severity as shown by many others [1962,1963,1976]. The novelty here is that they show that a combination with other markers can enhance the specificity and sensibility of the predictor, although the study could be improved by taking into account sub-populations of lymphocytes and more biological factors from patients such as LDH and IL6.

12.86.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.87 Metabolic disturbances and inflammatory dysfunction predict severity of coronavirus disease 2019 (COVID-19): a retrospective study

Shuke Nie et al. *medRxiv*. [2063]

12.87.1 Keywords

- metabolism
- fasting blood glucose
- serum total protein
- albumin
- blood lipid
- HDL-C
- APOA1
- lymphocytopenia
- IL-6
- CRP
- severity prediction of COVID19

12.87.2 Main Findings

Retrospective Study on 97 COVID-19 hospitalized patients (25 severe and 72 non-severe) analyzing clinical and laboratory parameter to predict transition from mild to severe disease based on more accessible indicators (such as fasting blood glucose, serum protein or blood lipid) than inflammatory indicators. In accordance with other studies, age and hypertension were risk factors for disease severity, and lymphopenia and increased IL-6 was observed in severe patients. The authors show that fasting blood glucose (FBG) was altered and patients with severe disease were often hyperglycemic. Data presented support that hypoproteinaemia, hypoalbuminemia, and reduction in high-densitylipoprotein (HDL-C) and ApoA1 were associated with disease severity.

12.87.3 Limitations

In this study non-severe patients were divided in two groups based on average course of the disease: mild group 1 (14 days, n=28) and mild group 2 (30 days, n=44). However mild patients with a longer disease course did not show an intermediate phenotype (between mild patients with shorter disease course and severe patients), hence it is unclear whether this was a useful and how it impacted the analysis. Furthermore, the non-exclusion of co-morbidity factors in the analysis may bias the results (e.g. diabetic patients and glucose tests) It is not clear at what point in time the laboratory parameters are sampled. In table 3, it would have been interesting to explore a multivariate multiple regression. The correlation lacks of positive control to assess the specificity of the correlation to the disease vs. correlation in any inflammatory case. The dynamic study assessing the predictability of the laboratory parameter is limited to 2 patients. Hence there are several associations with disease severity, but larger studies are necessary to test the independent predictive value of these potential biomarkers.

12.87.4 Significance

As hospital are getting overwhelmed a set of easily accessible laboratory indicators (such as serum total protein) would potentially provide a triage methodology between potentially severe cases and mild ones. This paper also opens the question regarding metabolic deregulation and COVID-19 severity.

12.87.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.88 Viral Kinetics and Antibody Responses in Patients with COVID-19

[2064]

12.88.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- IgG
- IgM
- clinical
- kinetics
- antibodies

12.88.2 Main Findings

- Prospective cohort of 67 patients, clinical specimens taken and follow-up conducted.
- Viral shedding, serum IgM, IgG antibody against NP evaluated and correlated to disease severity and clinical outcome
- Viral RNA levels peaked at 1 week from febrile/cough symptom onset in sputum, nasal swabs, and stool samples. Shedding ranged from 12-19 days (median ranges) and was longer in severe patients.
- IgM and IgG titers stratified patients into three archetypes as 'strong vs weak vs non-responders'. Strong responders (with higher IgM/IgG titers) were significantly higher in severe patients.

12.88.3 Limitations

Specific for immune monitoring.

- Not clear if stool RNA captured from live infection in intestine/liver or from swallowed sputum. Transmission electron microscopy (TEM) carried out on sputum samples as proof of concept, but not stools. TEM unreasonable for actual clinical diagnosis.
- Several patients had co-morbidities (such as pulmonary and liver disease) that were not accounted for when tracking antibody responses. Viral kinetics and IgM/IgG titers in subsets of patients with underlying conditions/undergoing certain medication would be informative.

12.88.4 Significance

- Three archetypes of antibody response to SARS-CoV-2 with different disease progression and kinetics is useful to stratify patients, and for future serological tests.
- Strong spike-IgG levels often correlate with lymphopenia and CoVID-19 disease severity [2065], similar to macaque studies in SARS [2066]. It would be critical to see if anti-NP or anti-Spike IgG antibodies for SARS-CoV-2 also elicit similar detrimental effects before clinical use.

12.88.5 Credit

12.89 COVID-19 infection induces readily detectable morphological and inflammation-related phenotypic changes in peripheral blood monocytes, the severity of which correlate with patient outcome

[[2067](#)]

12.89.1 Keywords

- Monocytes
- FSC-high
- PBMC
- ACE2
- inflammatory cytokines

12.89.2 Main Findings

This study is based on flow cytometry immunophenotyping of PBMCs from 28 patients diagnosed positive for SARS-Cov2 (COVID19). The authors identify a population of abnormally large (FSC-hi) monocytes, present in COVID19 patients, but absent in PBMCs of healthy volunteers (n=16) or patients with different infections (AIDS, malaria, TB). This FSC-hi monocytic population contains classical, intermediate and non-classical (monocytes (based on CD14 and CD16 expression) that produce inflammatory cytokines (IL-6, TNF and IL-10). The authors suggest an association of FSC-hi monocytes with poor outcome and correlate a high percentage of FSC-low monocytes, or higher ratio of FSC-low/hi monocytes, with faster hospital discharge.

12.89.3 Limitations

While identification of the monocytic population based on FSC is rather robust, the characterization of these cells remains weak. A comprehensive comparison of FSC-hi monocytes with FSC-low monocytes from patients and healthy controls would be of high value. It is unclear if percentages in blood are among CD45+ cells. Furthermore, it would have been important to include absolute numbers of different monocytic populations (in table 1 there are not enough samples and it is unclear what the authors show).

The authors show expression of the ACE2 receptor on the surface of the monocytes, and highlight these cells as potential targets of SARS-Cov2. However, appropriate controls are needed. CD16 has high affinity to rabbit IgG and it is unclear whether the authors considered unspecific binding of rabbit anti-ACE2 to Fc receptors. Gene expression of ACE-2 on monocytes

needs to be assessed. Furthermore, it would be important to confirm infection of monocytes by presence of viral proteins or viral particles by microscopy.

Considering the predictive role of FSC-hi monocytes on the development of the disease and its severity, some data expected at this level are neither present nor addressed. Although the cohort is small, it does include 3 ICU patients. What about their ratio of FSC-low vs FSC-hi monocytes in comparison to other patients? Was this apparent early in the disease course? Does this population of FSC-hi monocytes differ between ICU patients and others in terms of frequency, phenotype or cytokine secretion?

In general, figures need to revised to make the data clear. For example, in Fig. 5, according to the legend it seems that patients with FSC-high monocytes are discharged faster from the hospital. However according to description in the text, patients were grouped in high or low levels of FSC-low monocytes.

12.89.4 Significance

Despite the limitations of this study, the discovery of a FSC-high monocyte population in COVID-19 patients is of great interest. With similar implication, a the recent study by Zhou et al. [1968] identified a connection between an inflammatory CD14+CD16+ monocyte population and pulmonary immunopathology leading to deleterious clinical manifestations and even acute mortality after SARS-CoV-2 infections. Although the presence of these monocytes in the lungs has yet to be demonstrated, such results support the importance of monocytes in the critical inflammation observed in some COVID19 patients.

12.89.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.90 Correlation between universal BCG vaccination policy and reduced morbidity and mortality for COVID-19: an epidemiological study

Miller et al. *medRxiv*. [2068]

12.90.1 Keywords

- BCG vaccine
- epidemiology
- vaccination policy

12.90.2 Main Findings

The authors compared middle and high income countries that never had a universal BCG vaccination policy (Italy, Lebanon, Nederland, Belgium) and countries with a current policy (low income countries were excluded from the analysis as their number of cases and deaths might be underreported for the moment). **Countries that never implement BCG vaccination have a higher mortality rate than countries which have a BCG vaccination policy** (16.38 deaths per million people vs 0.78). Next, **the authors show that an earlier start of vaccination correlates with a lower number of deaths per million inhabitants**. They interpret this as the vaccine protecting a larger fraction of elderly people, which are usually more affected by COVID-19. Moreover, higher number of COVID-19 **cases** were presented in countries that never implemented a universal BCG vaccination policy.

12.90.3 Limitations

While this study aims to test an intriguing hypothesis unfortunately, the data is not sufficient at this time to accurately make any determinations. Several caveats must be noted including: not all countries are in the same stage of the pandemic, the number of cases/deaths is still changing very rapidly in a lot of countries and thus the association may only reflect exposure to the virus. This analysis would need to be re-evaluated when all the countries are passed the pandemic and more accurate numbers are available. Additionally, very few middle and high-income countries ever implemented universal BCG vaccination, which can be a source of bias (5 countries, vs 55 that have a BCG vaccine policy). Effective screening and social isolation policies also varied considerable across the countries tested and may reflect another important confounder. The authors could consider analyzing the Case Fatality Rate (CFR, % of patients with COVID-19 that die), to more correct for exposure although testing availability will still bias this result. Variability in mortality within countries or cities with variable vaccination and similar exposure could also be appropriate although confounders will still be present.

12.90.4 Significance

BCG vaccine is a live attenuated strain derived from *Mycobacterium bovis* and used for a vaccine for tuberculosis (TB). This vaccine has been proven to be efficient in preventing childhood meningitis TB, but doesn't prevent adult TB as efficiently. For this reason, several countries are now only recommending this vaccine for at-risk population only.

This study shows that there is a correlation between BCG vaccination policy and reduced mortality for Covid-19. Indeed, BCG vaccine has been shown to protect against several viruses and enhance innate immunity [1135], which could explain why it could protect against SARS-CoV-2 infection, but the exact mechanism is still unknown. **Moreover, the efficiency of adult/older people vaccination and protection against Covid-19 still needs to be assessed.** Regarding this, Australian researchers are starting a clinical trial of BCG vaccine for healthcare workers [2069], to assess if it can protect them against Covid-19.

12.90.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.91 Non-neural expression of SARS-CoV-2 entry genes in the olfactory epithelium

[2070]

12.91.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- anosmia
- olfaction
- scRNaseq

12.91.2 Main Findings

- Study analyzed bulk and scRNaseq data of olfactory cell types from publicly-available mouse, nonhuman primate and human datasets.
- show that *ACE2* and *TMPRSS2* (genes involved in SARS-CoV-2 entry) are expressed in olfactory epithelial (OE) cells, basal stem cells and respiratory epithelium (RE), but not sensory neurons.
- Comparison of human RE and OE datasets (Deprez et al. 2019; Durante et al. 2020) revealed that *ACE2* and *TMPRSS2* expression in OE sustentacular cells was similar to expression in the remainder of the non-nasal respiratory tract.

12.91.3 Limitations

- Transcript data alone from healthy respiratory/olfactory cells is not sufficient to confirm infectivity of nasal passage, or to indicate damage to epithelia.
- No mechanism defined for anosmia; it is not clear if epithelial injury leads to reduced sensitivity or increased inflammation and altered immune contexture drives neural/epithelial dysfunction. Will be critical to test this in CoVID-19 patient samples or mouse models.

12.91.4 Significance

- Study provides possible rationale for anosmia observed in several CoVID-19 patients.
- Raises possibility that nasal respiratory goblet, ciliated cells, and olfactory epithelia may serve as a viral reservoir after initial SARS-CoV-2 infection.

- Human olfactory sensory neurons express several other molecules important to CoV (not CoV-19) entry such as *FURIN*, *ST6GAL1*, *ST3GAL4*; this suggests wider mechanism of neuronal infectivity in other coronaviruses.

12.91.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Title:

SARS-CoV-2 proteome microarray for mapping COVID-19 antibody interactions at amino acid resolution

Immunology keywords: SARS-CoV-2, COVID-19, high throughput, peptide microarray, antibody epitope screening

The main finding of the article:

This study screened the viral protein epitopes recognized by antibodies in the serum of 10 COVID-19 patients using a new SARS-CoV-2 proteome peptide microarray. The peptide library was constructed with 966 linear peptides, each 15 amino acids long with a 5 amino acid overlap, based on the protein sequences encoded by the genome of the Wuhan-Hu-1 strain.

To investigate crossreactivity between SARS-CoV-1 and SARS-CoV-2, they tested rabbit monoclonal and polyclonal antibodies against SARS-CoV-1 nucleocapsid (N) in the microarray. Antibodies against SARS-CoV-1 N displayed binding to the SARS-CoV-2 nucleocapsid (N) peptides. Polyclonal antibodies showed some crossreactivity to other epitopes from membrane (M), spike (S), ORF1ab and ORF8. This suggests that previous exposure to SARS-CoV-1 may induced antibodies recognizing both viruses.

Screening of IgM and IgG antibodies from 10 COVID-19 patients showed that many antibodies targeted peptides on M, N, S, Orf1ab, Orf3a, Orf7a, and Orf8 from SARS-CoV-2, while immunodominant epitopes with antibodies in more than 80 % COVID-19 patients were present in N, S and Orf3. It is shown that the receptor binding domain (RBD) resides on S protein and RBD is important for SARS-CoV-2 to enter the host cells via ACE2. Among six epitopes on S protein, structural analysis predicted that three epitopes were located at the surface and three epitopes were located inside of the protein. Furthermore, some IgM antibodies from 1 patient and IgG antibodies from 2 patients bound to the same epitope (residue 456-460, FRKSN) which resided within the RBD, and structural analysis determined that this epitope was located in the region of the RBD loop that engages with ACE2.

Critical analysis of the study:

In addition to the limitations mentioned in the manuscript, it would have been informative to do the analysis over the course of the disease. The pattern of antibody recognition, especially on S protein, and the course of

antibodies of different isotypes recognizing the same peptide might correlate to the clinical course in these patients. It would also have been informative to analyze the presence of cross-reactive antibodies from patients previously exposed to SARS-CoV-1.

The importance and implications for the current epidemics:

This study identified linear immunodominant epitopes on SARS-CoV-2, Wuhan-Hu-1 strain. This is a valuable information to design vaccines that will elicit desirable immune responses.

The Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Directly Decimates Human Spleens and Lymph Nodes

Review by Matthew D. Park

Revised by Miriam Merad

Keywords: COVID-19, SARS-CoV-2, spleen, lymph node, ACE2, macrophage

Main findings

It has been previously reported that COVID-19 patients exhibit severe lymphocytopenia, but the mechanism through which this depletion occurs has not been described. In order to characterize the cause and process of lymphocyte depletion in COVID-19 patients, the authors performed gross anatomical and *in situ* immune-histochemical analyses of spleens and lymph nodes (hilar and subscapular) obtained from post-mortem autopsies of 6 patients with confirmed positive viremia and 3 healthy controls (deceased due to vehicle accidents).

Primary gross observations noted significant splenic and LN atrophy, hemorrhaging, and necrosis with congestion of interstitial blood vessels and large accumulation of mononuclear cells and massive lymphocyte death. They found that CD68⁺ CD169⁺ cells in the spleens, hilar and subscapular LN, and capillaries of these secondary lymphoid organs expressed the ACE2 receptor and stain positive for the SARS-CoV-2 nucleoprotein (NP) antigen, while CD3⁺ T cells and B220⁺ B cells lacked both the ACE2 receptor and SARS-CoV-2 NP antigen. ACE2⁺ NP⁺ CD169⁺ macrophages were positioned in the splenic marginal zone (MZ) and in the marginal sinuses of LN, which suggests that these macrophages were positioned to encounter invading pathogens first and may contribute to virus dissemination.

Since SARS-CoV-2 does not directly infect lymphocytes, the authors hypothesized that the NP⁺ CD169⁺ macrophages are responsible for persistent activation of lymphocytes via Fas::FasL interactions that would mediate activation-induced cell death (AICD). Indeed, the expression of Fas was significantly higher in virus-infected tissue than that of healthy controls, and TUNEL staining showed significant lymphocytic apoptosis. Since pro-inflammatory cytokines like IL-6 and TNF- α can also engage cellular apoptosis and necrosis, the authors interrogated the cytokine expression of the secondary lymphoid organs from COVID-19 patients; IL-6, not TNF- α , was elevated in virus-infected splenic and lymph node tissues, compared to those

of healthy controls, and immunofluorescent staining showed that IL-6 is primarily produced by the infected macrophages. *In vitro* infection of THP1 cells with SARS-CoV-2 spike protein resulted in selectively increased *Il6* expression, as opposed to *Il1b* and *Tnfa* transcription. Collectively, the authors concluded that a combination of Fas up-regulation and IL-6 production by NP⁺ CD169⁺ macrophages induce AICD in lymphocytes in secondary lymphoid organs, resulting in lymphocytopenia.

In summary, this study reports that CD169⁺ macrophages in the splenic MZ, subscapular LN, and the lining capillaries of the secondary lymphoid tissues express ACE2 and are susceptible to SARS-CoV-2 infection. The findings point to the potential role of these macrophages in viral dissemination, immunopathology of these secondary lymphoid organs, hyperinflammation and lymphopenia.

Limitations

Technical

A notable technical limitation is the small number of samples (n=6); moreover, the analysis of these samples using multiplexed immunohistochemistry and immunofluorescence do not necessarily provide the depth of unbiased interrogation needed to better identify the cell types involved.

Biological

The available literature and ongoing unpublished studies, including single-cell experiments of spleen and LN from organ donors, do not indicate that ACE2 is expressed by macrophages; however, it remains possible that ACE2 expression may be triggered by type I IFN in COVID-19 patients. Importantly, the SARS-CoV-2 NP staining of the macrophages does not necessarily reflect direct infection of these macrophages; instead, positive staining only indicates that these macrophages carry SARS-CoV-2 NP as antigen cargo, which may have been phagocytosed. Direct viral culture of macrophages isolated from the secondary lymphoid organs with SARS-CoV-2 is required to confirm the potential for direct infection of macrophages by SARS-CoV-2. Additionally, it is important to note that the low to negligible viremia reported in COVID-19 patients to-date does not favor a dissemination route via the blood, as suggested by this study, which would be necessary to explain the presence of virally infected cells in the spleen.

Relevance

Excess inflammation in response to SARS-CoV-2 infection is characterized by cytokine storm in many COVID-19 patients. The contribution of this pathology to the overall fatality rate due to COVID-19, not even necessarily directly due to SARS-CoV-2 infection, is significant. A better understanding of the full effect and source of some of these major cytokines, like IL-6, as well as the deficient immune responses, like lymphocytopenia, is urgently needed. In this study, the authors report severe tissue damage in spleens and lymph nodes of COVID-19 patients and identify the role that CD169⁺ macrophages may play in the hyperinflammation and lymphocytopenia that are both

characteristic of the disease. It may, therefore, be important to note the effects that IL-6 inhibitors like Tocilizumab and Sarilumab may specifically have on splenic and LN function. It is important to note that similar observations of severe splenic and LN necrosis and inflammation in patients infected with SARS-CoV-1 further support the potential importance and relevance of this study.

12.92 Cigarette smoke triggers the expansion of a subpopulation of respiratory epithelial cells that express the SARS-CoV-2 receptor ACE2

[2071]

12.92.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- respiration
- cigarette
- ACE2
- lung

12.92.2 Main Findings

- Study uses scRNASeq, bulk seq data and air-liquid interface culture experiments to show that cigarette smoke causes a dose-dependent upregulation of ACE2 in mouse and human lungs (transplantation, tumor resection, or IPF datasets).
- ACE2 was not up-regulated in patients with asthma or lung-sarcoidosis or in mouse models of cystic fibrosis or carcinogen exposure.
- Cathepsin B (alternate protease involved in viral entry) is increased in smoke-exposed mouse or human lungs.
- Smoke triggers a protective expansion of mucus-secreting MUC5AC+ goblet and SCGB1A1+ club cells; ACE2 presence in these cells is increased upon smoke exposure.

12.92.3 Limitations:

- Long-term smokers usually have several co-morbidities including immune dysfunction, which can affect interpretation of CoV-2 susceptibility in these datasets. Ideally, analyses can control for major co-morbidities across smokers and non-smokers (immune suppression, cardiovascular disease and atherosclerosis).
- Hyperplasia of ACE2+ goblet cells upon smoking needs to be separated from ACE2 upregulation in existing goblet cells.

- ACE2 expression increase alone does not confirm increased viral entry into goblet cells; future studies with air-liquid interface cultures testing CoV-2 infectivity in *ex vivo* epithelial cells from human epithelial lines, *ex vivo* samples or hACE2 mice will be very informative.

12.92.4 Significance

- This study may partially explain why smokers are more likely to develop severe SARS-CoV-2 infections. Also, the reversibility of ACE2 expression upon smoking cessation suggests that quitting smoking could lessen CoV-2 susceptibility.
- Absence of ACE2 upregulation in other lung inflammation pathologies implies CoV-2 susceptibility might be smoking-specific, and not fibrosis-specific.
- Another preprint showed ACE2 expression increases in lung of patients with CoV-2 co-morbidities such as hypertension [[2045](#)]; these studies collectively paint a better picture of CoV-2 susceptibility before actual experiments can be carried out.

12.92.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

12.93 The comparative superiority of IgM-IgG antibody test to real-time reverse transcriptase PCR detection for SARS-CoV-2 infection diagnosis

Liu et al. *medRxiv*. [[2072](#)]

12.93.1 Keywords

- IgM/IgG antibody test
- nucleic acid test
- SARS-CoV-2 detection

12.93.2 Main Findings

The study compares IgM and IgG antibody testing to RT-PCR detection of SARS-CoV-2 infection. 133 patients diagnosed with SARS-CoV-2 in Renmin Hospital (Wuhan University, China) were analyzed. The positive ratio was 78.95% (105/133) in IgM antibody test (SARS-CoV-2 antibody detection kit from YHLO Biotech) and 68.42% (91/133) in RT-PCR (SARS-CoV-2 ORF1ab/N qPCR detection kit). There were no differences in the sensitivity of SARS-CoV-2 diagnosis in patients grouped according to disease severity. For example, IgG responses were detected in 93.18% of moderate cases, 100% of severe

cases and 97.3% of critical cases. In sum, positive ratios were higher in antibody testing compared to RT-PCR detection, demonstrating a higher detection sensitivity of IgM-IgG testing for patients hospitalized with COVID-19 symptoms.

12.93.3 Limitations

This analysis only included one-time point of 133 hospitalized patients, and the time from symptom onset was not described. There was no discussion about specificity of the tests and no healthy controls were included. It would be important to perform similar studies with more patients, including younger age groups and patients with mild symptoms as well as asymptomatic individuals. It is critical to determine how early after infection/symptom onset antibodies can be detected and the duration of this immune response.

12.93.4 Significance

The IgM-IgG combined testing is important to improve clinical sensitivity and diagnose COVID-19 patients. The combined antibody test shows higher sensitivity than individual IgM and IgG tests or nucleic acid-based methods, at least in patients hospitalized with symptoms.

12.93.5 Credit

Review by Erica Dalla as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Title: Lectin-like Intestinal Defensin Inhibits 2019-nCoV Spike binding to ACE2

Immunology keywords: defensins, spike protein, intestinal Paneth cells, ACE2 binding

Main Findings:

Human ACE2 was previously identified as the host receptor for SARS-CoV-2. Despite ACE2 being expressed in both lung alveolar epithelial cells and small intestine enterocytes, respiratory problems are the most common symptom after viral infection while intestinal symptoms are much less frequent. Thus, the authors here investigate the biology behind the observed protection of the intestinal epithelium from SARS-CoV-2. Human defensin 5 (HD5), produced by Paneth cells in the small intestine, was shown to interact with human ACE2, with a binding affinity of 39.3 nM by biolayer interferometry (BLI). A blocking experiment using different doses of HD5 coating ACE2 showed that HD5 lowered viral spike protein S1 binding to ACE2. Further, a molecular dynamic simulation demonstrated a strong intermolecular interaction between HD5 and the ACE2 ligand binding domain. To test HD5 inhibitory effect on S1 binding to ACE2, human intestinal epithelium Caco-2 cells were preincubated with HD5. Preincubation strongly reduced adherence of S1 to surface of cells. HD5 was effective at a concentration as low as 10 µg/mL, comparable to the concentration found in the intestinal fluid.

Limitations:

The study focuses exclusively on intestinal cells. However, HD5 could have been tested to block ACE2-S1 binding in human lung epithelial cells as a potential treatment strategy. It would be useful to know whether HD5 could also prevent viral entry in lung cells.

Relevance:

This work provides the first understanding of the different efficiency of viral entry and infection among ACE2-expressing cells and tissues. Specifically, the authors show that human defensin 5 produced in the small intestine is able to block binding between S1 and ACE2 necessary for viral entry into cells. The study provides a plausible explanation on why few patients show intestinal symptoms and suggests that patients with intestinal disease that decrease defensins' production may be more susceptible to SARS-CoV-2. It also indicates that HD5 could be used as a molecule to be exogenously administered to patients to prevent viral infection in lung epithelial cells.

Title:

Susceptibility of ferrets, cats, dogs and different domestic animals to SARS-coronavirus-2

Immunology keywords: SARS-CoV-2, ferret, cat, laboratory animal, domestic animals

The main finding of the article:

This study evaluated the susceptibility of different model laboratory animals (ferrets), as well as companion (cats and dogs), and domestic animals (pigs, chickens and ducks) to SARS-CoV-2. They tested infection with two SARS-CoV2 isolates, one from an environmental sample collected in the Huanan Seafood Market in Wuhan (F13-E) and the other from a human patient in Wuhan (CTan-H).

Ferrets were inoculated with either of the two viruses by intranasal route with 10^5 pfu, and the viral replication was evaluated. Two ferrets from each group were euthanized on day 4 post infection (p.i.). At day 4 p.i., viral RNA and infectious viruses were detected only in upper respiratory tract (nasal turbinate, upper palate, tonsils, but not in the trachea, lungs or other tissues. Viral RNA and virus titer in the remaining ferrets were monitored in nasal washes and rectal swabs on days 2, 4, 6, 8 and 10 p.i. Viral RNA and infectious viruses were detected in nasal washes until day 8 p.i. One ferret in each group developed fever and loss of appetite on days 10 and 12 p.i., however, viral RNA was practically undetectable. These two ferrets showed severe lymphoplasmacytic perivasculitis and vasculitis in the lungs and lower antibody titers compare to other 4 ferrets.

Cats. Five subadult 8-month-old domestic cats were inoculated with CTan-h virus and three uninfected cats were placed in a cage adjacent to each of the infected cats to monitor respiratory droplet transmission. Viral RNA was detected in the upper respiratory organs from all infected cats and in one out

of three exposed cats. All infected (inoculated and exposed) cats developed elevated antibodies against SARS-CoV2. Viral replication studies with juvenile cats (70-100 days) revealed massive lesions in the nasal and tracheal mucosa epithelium and lungs of two inoculated cats which died or were euthanized on day 3 p.i., and infection in one out of three exposed cats. These results indicated SARS-CoV2 could replicate in cats, that juvenile cats were more susceptible than adults, and that SARS-CoV2 could be transmitted via respiratory droplets between cats.

Dogs and others. Five 3-month-old beagle dogs were inoculated and housed with two uninoculated beagles in a room. Two virus inoculated dogs seroconverted, but others including two contact dogs were all seronegative for SARS-CoV2 and infectious virus was not detected in any swabs collected. Viral RNA was not detected in swabs from pigs, chickens, and ducks inoculated or contacted. These results indicated that dogs, pigs, chickens, and ducks might have low or no susceptibility to SARS-CoV2.

Critical analysis of the study:

This manuscript describes the viral replication and clinical symptoms of SARS-CoV2 infection in ferrets, and the SARS-CoV2 infection and transmission in cats. Clinical and pathological analysis was not performed in cats, therefore the correlation of virus titer with symptoms severity in the adult and juvenile cats could not be determined.

The importance and implications for the current epidemics:

SARS-CoV-2 transmission to tigers, cats and dogs has been previously reported. It should be noted that this manuscript did not evaluate the transmission from cats to human. Nevertheless, it clearly showed higher susceptibility of ferrets and domestic cats to SARS-CoV-2. This data strongly indicates the need for surveillance of possible infection and transmission of SARS-CoV-2 by domestic cats.

12.94 Virus-host interactome and proteomic survey of PBMCs from COVID-19 patients reveal potential virulence factors influencing SARS-CoV-2 pathogenesis

Li et al. *bioRxiv*. [193]

12.94.1 Keywords

- PBMC
- virulence factors – interaction network – nsp9
- nsp10 – NKRF

12.94.2 Main findings

The authors identified **intra-viral protein-protein interactions** (PPI) with two different approaches: genome wide yeast-two hybrid (Y2H) and co-immunoprecipitation (co-IP). A total of 58 distinct PPI were characterized. A screen of **viral-host PPI** was also established by overexpressing all the SARS-CoV-2 genes with a Flag epitope into HEK293 cells and purifying each protein complex. Interacting host proteins were then identified by liquid chromatography and tandem mass spectrometry. 251 cellular proteins were identified, such as subunits of ATPase, 40S ribosomal proteins, T complex proteins and proteasome related proteins, for a total of 631 viral-host PPI. Several interactions suggesting protein-mediated modulation of the immune response were identified, highlighting the multiple ways SARS-CoV-2 might reprogram infected cells.

Subsequently, the authors compared global proteome profiles of PBMCs from healthy donors ($n=6$) with PBMC from COVID-19 patients with mild ($n=22$) or severe ($n=13$) symptoms. 220 proteins were found to be differentially expressed between *healthy donors and mild COVID-19 patients*, and a pathway analysis showed **a general activation of the innate immune response**. 553 proteins were differentially expressed between the PBMC of *mild and severe COVID-19 patients*, most of them (95%) being downregulated in severe patients. Functional pathway analysis indicated a defect of T cell activation and function in severe COVID-19. There was also evidence suggesting reduced antibody secretion by B cells. Together, these results suggest a **functional decline of adaptive immunity**. A FACS analysis of PBMC from severe patients indicated higher levels of IL6 and IL8 but not IL17 compared to mild patients.

Finally, the authors focused on NKRF, an endogenous repressor of IL8/IL6 synthesis that was previously identified as interacting with SARS-CoV-2 nsp9,10,12,13 and 15. Individually expressed nsp9 and nsp10 (but not nsp12, nsp13, nsp15) induced both IL6 and IL8 in lung epithelial A459 cells, indicating that nsp9 and nsp10 may be directly involved in the induction of these pro-inflammatory cytokines. The authors finally argue that nsp9 and nsp10 represent potential drug targets to prevent over-production of IL6 and IL8 in infected cells, and reducing the over-activation of neutrophils.

12.94.3 Limitations

First, the authors seem to have forgotten to include the extended data in the manuscript, and their proteomic data does not seem to be publicly available for the moment, which limits greatly our analysis of their results.

While this work provides important data on host and viral PPI, only 19 interactions were identified by Y2H system but 52 with co-IP. The authors do not comment about what could lead to such differences between the two techniques and they don't specify whether they detected the same interactions using the two techniques.

Moreover, the PBMC protein quantification was performed comparing bulk PBMC. Consequently, protein differences likely reflect differences in cell populations rather than cell-intrinsic differences in protein expression. While

this analysis is still interesting, a similar experiment performed on pre-sorted specific cell populations would allow measuring proteome dynamics at a higher resolution.

Finally, the authors did not discuss their results in regards to another SARS-CoV-2 interactome of host-viral PPI that had been published previously¹. This study reported 332 host-virus PPI, but no interaction of viral proteins with NKRF was found. Some interactions were found in both studies (eg. N and G3BP1, Orf6 and RAE1). However, the time point used to lyse the cells were different (40h previously vs 72h here), which could explain some of the differences.

12.94.4 Relevance

The identification of many interactions between intra-viral and host-virus PPI provides an overview of host protein and pathways that are modulated by SARS-CoV-2, which can lead to the identification of potential targets for drug development.

In the model proposed by the authors, nsp9 and nsp10 from SARS-CoV-2 induce an over-expression of IL6 and IL8 by lung epithelial cells, which recruits neutrophils and could lead to an excess in lung infiltration. Nsp9 has been shown to be essential for viral replication for SARS-CoV-1², and shares a 97% homology with nsp9 from SARS-CoV-2³. Further, nsp9 crystal structure was recently solved³, which can help to develop drug inhibitors if this protein is further confirmed as being important for the virulence of SARS-CoV-2.

1. Gordon DE, Jang GM, Bouhaddou M, et al. A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug-Repurposing. *bioRxiv*. March 2020;2020.03.22.002386.
doi:10.1101/2020.03.22.002386

2. Miknis ZJ, Donaldson EF, Umland TC, Rimmer RA, Baric RS, Schultz LW. Severe acute respiratory syndrome coronavirus nsp9 dimerization is essential for efficient viral growth. *J Virol*. 2009;83(7):3007-3018.
doi:10.1128/JVI.01505-08

3. Littler DR, Gully BS, Colson RN, Rossjohn J. *Crystal Structure of the SARS-CoV-2 Non-Structural Protein 9, Nsp9*. Molecular Biology; 2020.
doi:10.1101/2020.03.28.013920

Title: Prediction and Evolution of B Cell Epitopes of Surface Protein in SARS-CoV-2

Keywords: SARS-CoV-2; Epitopes; Bioinformatics; Evolution

Summary/Main findings:

Lon et al. used a bioinformatic analysis of the published SARS-CoV-2 genomes in order to identify conserved linear and conformational B cell epitopes found on the spike (S), envelope (E), and membrane (M) proteins. The characterization of the surface proteins in this study began with an assessment of the peptide sequences in order to identify hydrophilicity

indices and protein instability indices using the Port-Param tool in ExPASy. All three surface proteins were calculated to have an instability score under 40 indicating that they were stable. Linear epitopes were identified on the basis of surface probability and antigenicity, excluding regions of glycosylation. Using BepiPred 2.0 (with a cutoff value of 0.35) and ABCpred (with a cutoff value of 0.51), 4 linear B cell epitopes were predicted for the S protein, 1 epitope for the E protein, and 1 epitope for the M protein. For structural analysis, SARS-CoV assemblies published in the Protein Data Bank (PDB) acting as scaffolds for the SARS-CoV-2 S and E amino acid sequences were used for input into the SWISS-MODEL server in order to generate three-dimensional structural models for the assessment of conformational epitopes. Using Ellipro (cutoff value of 0.063) and SEPPA (cutoff value of 0.5), 1 conformational epitope was identified for the S protein and 1 epitope was identified for the E protein, both of which are accessible on the surface of the virus. Finally, the Consurf Server was used to assess the conservation of these epitopes. All epitopes were conserved across the published SARS-CoV-2 genomes and one epitope of the spike protein was predicted to be the most stable across coronavirus phylogeny.

Critical Analysis/Limitations:

While this study provides a preliminary identification of potential linear and conformational B cell epitopes, the translational value of the epitopes described still needs extensive experimental validation to ascertain whether these elicit a humoral immune response. The conformational epitope analyses are also limited by the fact that they are based off of predicted 3D structure from homology comparisons and not direct crystal structures of the proteins themselves. Additionally, since there was not a published M protein with a high homology to SARS-CoV-2, no conformational epitopes were assessed for this protein. Finally, while evolutionary conservation is an important consideration in understanding the biology of the virus, conservation does not necessarily imply that these sites neutralize the virus or aid in non-neutralizing *in vivo* protection.

Relevance/Implications:

With further experimental validation that confirms that these epitopes induce effective antibody responses to the virus, the epitopes described can be used for the development of treatments and vaccines as well as better characterize the viral structure to more deeply understand pathogenesis.

13 Appendix B

Contributors were asked to complete this template to summarize and evaluate new papers related to diagnostics.

Title: Please edit the title to add the name of the paper after the colon

Please paste a link to the paper or a citation here:

Link:

What is the paper's [Manubot-style citation?](#)

Citation:

Please list some keywords (3-10) that help identify the relevance of this paper to COVID-19

- keyword 1 (replace me, copy and paste more than three if needed)
- keyword 2 (replace me, copy and paste more than three if needed)
- keyword 3 (replace me, copy and paste more than three if needed)

Please note the publication / review status

- Pre-print
- New Peer-Reviewed Paper
- Peer-Reviewed Paper Pre-2020

Which areas of expertise are particularly relevant to the paper?

- virology
- epidemiology
- biostatistics
- immunology
- pharmacology

Questions to answer about each paper:

Please provide 1-2 sentences introducing the study and its main findings

Study question(s) being investigated:

What type of testing scenario is being considered?

Is it a screening test (used for individuals with no symptoms), diagnostic test (used for individuals with symptoms), or definitive test (used for individuals who have had previous positive test results on diagnostic or screening tests)?

Study population:

What is the model system (e.g., human study, animal model, cell line study)?

What is the sample size?

What is the “pre-test” probability of disease in the study population (i.e., what is the anticipated prevalence of the disease?)

For human studies, the following are related to the pre-test probability:

What countries/regions are considered?

What is the age range, gender, other relevant characteristics?

What is the setting of the study (e.g., random sample of school children, retirement communities, etc.)?

What other specific inclusion-exclusion criteria are considered?

Reference test:

What reference test is considered as a “gold standard” comparator for the test under investigation?

Test assignment:

How are the new and reference tests assigned?

Examples of assignment could include: Recruited individuals have initially undergone neither the new nor the reference test; individuals tested as positive or negative by the reference test undergo the new test; individuals who have undertaken the new test are assessed by the standard test.

Are there any other relevant details about the study design?

Depending on how individuals are chosen, the test may be biasing towards more sick or less sick individuals or very clear-cut positive/negative cases. Any factors that would influence this bias should be included here.

Test conduct:

How were tests performed?

Describe technical details of assays used, when measurements were taken and by whom, etc. for both the new and standard tests.

Test Assessment

Describe how individuals are classified as positive or negative, e.g. if a threshold is used.

Is there evidence that the test is precise/reproducible when repeated more than once?

Are measurements complete?

For example: Do some participants undergo just one test (the new or the reference test)? Are there individuals with inconclusive results?

Results summary:

What are the estimated sensitivity, specificity, positive predictive value (PPV), and negative predicted value (NPV)?

Note that the PPV and NPV represent “post-test” probabilities of disease and are generally more meaningful than sensitivity and specificity. Sometimes the post-test odds will be given instead.

What are the confidence bounds around these intervals?

Interpretation of results for study population:

How good is the test at ruling in or ruling out a disease based on the post-test probabilities?

Are there identified side affects of the test?

Is patient adherence to the test likely to be an issue?

Extrapolation of conclusions to other groups of individuals

How well is the test likely to work in populations with different pretest odds?

For example, if the prevalence is lower, then the PPV will also be lower, but the NPV will be higher.

How costly is the test?

How difficult is it to perform the test in different settings?

Could the test be combined with other existing tests?

Summary of reliability

1-2 sentences on concluding remarks, including summary of strengths, weaknesses, limitations.

Progress

Check off the components as they are completed. If the component is not applicable, check the box as well.

- 1-2 sentences introducing the study and its main findings
- Describe testing scenario
- Describe model system
- Sample size

- Describe prevalence of disease
- Describe countries/regions are considered
- Describe age range, gender, other relevant characteristics
- Describe setting of the study
- Describe other specific inclusion-exclusion criteria
- Describe "gold standard"
- Describe how the new and reference tests assigned
- Describe other relevant details about the study design
- Describe how the tests were performed
- Describe how individuals are classified as positive or negative
- Describe if test is precise/reproducible
- Describe whether measurements are complete
- What are the estimated sensitivity, specificity, positive predictive value (PPV), and negative predicted value (NPV)?
- What are the confidence bounds around these intervals?
- Describe post-test probabilities
- Describe side affects of the test
- Describe patient adherence
- Describe how it will extrapolate
- How costly is the test?
- How difficult is it to perform the test in different settings?
- Could the test be combined with other existing tests?
- Summary of reliability

14 Appendix C

Contributors were asked to complete this template to summarize and evaluate new papers related to therapeutics.

Title: Please edit the title to add the name of the paper after the colon

Please paste a link to the paper or a citation here:

Link:

What is the paper's [Manubot-style citation?](#)

Citation:

Please list some keywords (3-10) that help identify the relevance of this paper to COVID-19

- keyword 1 (replace me, copy and paste more than three if needed)
- keyword 2 (replace me, copy and paste more than three if needed)
- keyword 3 (replace me, copy and paste more than three if needed)

Please note the publication / review status

- Pre-print
- New Peer-Reviewed Paper
- Peer-Reviewed Paper Pre-2020

Which areas of expertise are particularly relevant to the paper?

- virology
- epidemiology
- biostatistics
- immunology
- pharmacology

Questions to answer about each paper:

Please provide 1-2 sentences introducing the study and its main findings

Study question(s) being investigated:

How many/what drugs/combinations are being considered?

What are the main hypotheses being tested?

Study population:

What is the model system (e.g., human study, animal model, cell line study)?

What is the sample size? If multiple groups are considered, give sample size for each group (including controls).

- number treated with treatment A
- number treated with treatment B

For human studies:

What countries/regions are considered?

What is the age range, gender, other relevant characteristics?

What is the setting of the study (random sample of school children, inpatient, outpatient, etc)?

What other specific inclusion-exclusion criteria are considered?

For example, do the investigators exclude patients with diagnosed neoplasms or patients over/under a certain age?

Treatment assignment:

How are treatments assigned?

For example, is it an interventional or an observational study?

Is the study randomized?

A study can be interventional but not randomized (e.g., a phase I or II clinical trial is interventional but often not randomized).

Provide other relevant details about the design.

This includes possible treatment stratification (e.g., within litters for animal studies, within hospitals for human studies), possible confounding variables (e.g., having a large age range of individuals), possible risks of bias and how they are addressed (e.g., is there masking in a clinical trial? how are individuals chosen in an observational study?).

Outcome Assessment:

Describe the outcome that is assessed and whether it is appropriate.

For example: Is the outcome assessed by a clinician or is it self-reported? Is the outcome based on viral load or a functional measurement (e.g., respiratory function, discharge from hospital)? What method is used to measure the outcome? How long after a treatment is the outcome measured?

Are outcome measurements complete?

For example, are there individuals lost to follow up?

Are outcome measurements subject to various kinds of bias?

For example, a lack of masking in randomized clinical trials.

Statistical Methods Assessment:

What methods are used for inference?

For example, logistic regression, nonparametric methods.

Are the methods appropriate for the study?

For example, are clustered data treated independently or are clusters adjusted for, such as different hospitals or litters?

Are adjustments made for possible confounders?

For example, adjustment for age, sex, or comorbidities.

Results Summary:

What is the estimated association?

For example, is it an estimated odds ratio, a median difference in detected cases, etc?

What measures of confidence or statistical significance are provided?

For example, confidence intervals, p-values, and/or Bayes factors.

Interpretation of results for study population:

Can we make a causal interpretation for the individuals in the study of drug -> outcome, such as "taking drug A improves likelihood of survival twofold over taking drug B."

For example, with a well-performed animal study or randomized trial it is often possible to infer causality. If it is an observational study, does it match up with some of the Bradford Hill criteria?

<https://www.edwardtufte.com/tufte/hill>

https://en.wikipedia.org/wiki/Bradford_Hill_criteria

Are there identified side effects or interactions with other drugs?

For example, is the treatment known to cause liver damage or to not be prescribed for individuals with certain comorbidities?

Are there specific subgroups with different findings?

For example, do individuals with a specific baseline seem to do particularly well? Be particularly cautious with respect to multiple testing here.

Extrapolation of conclusions to other groups of individuals not specifically included in the study:

If the study is an animal study, which animal and how relevant is that model?

Is the model system appropriate? Is there evidence from past use that it's highly-relevant to therapeutic design in this context?

If it is a human study, what characteristics of the study population may support/limit extrapolation?

- Can results extrapolate easily to other similar groups? (e.g., same country, similar age groups)
- What would happen if conditions are extended in terms of dose or duration?
- Can results be extrapolated to other populations or in very different settings? (e.g., different age group, primary care setting vs emergency department etc)

Summary of reliability

1-2 sentences on concluding remarks, including summary of strengths, weaknesses, limitations.

Progress

Check off the components as they are completed. If the component is not applicable, check the box as well.

- 1-2 sentences introducing the study and its main findings
- Describe How many/what drugs/combinations are being considered
- Describe the model system
- What is the sample size?
- What countries/regions are considered
- What is the age range, gender, other relevant characteristics
- Describe study setting
- Describe other specific inclusion-exclusion criteria
- Describe how treatments are assigned
- Describe randomization (or not) and other relavent details about the design
- Describe the outcome that is assessed and whether it is appropriate.
- Describe whether the outcome measurements are complete
- Are outcome measurements subject to various kinds of bias?
- Describe methods used for inference
- Describe whether the methods are appropriate for the study
- Are adjustments made for possible confounders?
- Describe the estimated association
- What measures of confidence or statistical significance are provided?
- Describe whether a causal interpretation can be made
- Are there identified side effects or interactions with other drugs?
- Are there specific subgroups with different findings?

- If the study is an animal study, which animal and how relevant is that model?
- If it is a human study, what characteristics of the study population may support/limit extrapolation?
- Summary of reliability

15 Appendix D

Contributors were asked to complete this template to summarize and evaluate new papers related to topics besides therapeutics and diagnostics.

Title: Please edit the title to add the name of the paper after the colon.

General Information Please paste a link to the paper or a citation here:

Link:

What is the paper's [Manubot-style citation?](#)

Citation:

Is this paper primarily relevant to Background or Pathogenesis?

- Background
- Pathogenesis
- Methods

Please list some keywords (3-10) that help identify the relevance of this paper to COVID-19

- keyword 1 (replace me, copy and paste more than three if needed)
- keyword 2 (replace me, copy and paste more than three if needed)
- keyword 3 (replace me, copy and paste more than three if needed)

Please note the publication / review status

- Pre-print
- New Peer-Reviewed Paper
- Peer-Reviewed Paper Pre-2020

Which areas of expertise are particularly relevant to the paper?

- virology
- epidemiology
- biostatistics
- immunology
- pharmacology
- other:

Summary

Suggested questions to answer about each paper: - What did they analyze? - What methods did they use? - Does this paper study COVID-19, SARS-CoV-2, or a related disease and/or virus? - What is the main finding (or a few main takeaways)? - What does this paper tell us about the background and/or

diagnostics/therapeutics for COVID-19 / SARS-CoV-2? - Do you have any concerns about methodology or the interpretation of these results beyond this analysis?

Any comments or notes?

1. <https://asapbio.org/preprints-and-covid-19> as well as <https://retractionwatch.com/retracted-coronavirus-covid-19-papers> ↵
2. <https://depts.washington.edu/pandemicalliance/covid-19-literature-report/latest-reports> ↵
3. <https://outbreaksci.prereview.org> ↵
4. <https://asapbio.org/preprints-and-covid-19> ↵
5. <https://disqus.com/by/sinaiimmunologyreviewproject> ↵
6. <https://rapidreviews covid19.mitpress.mit.edu> ↵
7. <https://greenelab.github.io/covid19-review> ↵
8. <https://casrai.org/credit> ↵
9. <https://www.gitter.im> ↵
10. <https://greenelab.github.io/covid19-review> ↵
11. <https://greenelab.github.io/covid19-review/manuscript.pdf> ↵
12. Vaccines: <https://github.com/owid/covid-19-data>; Clinical Trials: https://github.com/ebmdata/covid_trials_tracker-covid; Cases and Deaths: <https://github.com/CSSEGISandData/COVID-19> ↵
13. <https://github.com/greenelab/covid19-review/blob/master/.github/workflows/update-external-resources.yaml> ↵
14. <https://github.com/greenelab/covid19-review/tree/external-resources> ↵
15. <https://github.com/greenelab/covid19-review/blob/external-resources/environment.yml> ↵
16. <https://forums.zotero.org/discussion/74933/import-from-clinical-trials-registry> and <https://forums.zotero.org/discussion/77721/add-reference-from-clinical-trials-org> ↵
17. <https://www.zotero.org> and <https://github.com/zotero/translation-server> ↵
18. <https://github.com/zotero/translators/pull/2153> ↵
19. <https://identifiers.org> ↵

20. <https://pandoc.org> ↵
21. <http://aspell.net> ↵
22. <https://github.com/pandoc/lua-filters/tree/master/spellcheck> ↵
23. https://twitter.com/j_perkel/status/1245454628235309057 ↵
24. CONTRIBUTING.md and INSTRUCTIONS.md within the repository ↵
25. https://github.com/ismms-himc/covid-19_sinai_reviews ↵
26. https://github.com/CSSEGISandData/COVID-19/tree/master/csse_covid_19_data/csse_covid_19_time_series ↵
27. <https://github.com/owid/covid-19-data> ↵