

BRAIN TISSUE TEMPERATURE DYNAMICS DURING FUNCTIONAL ACTIVITY AND  
POSSIBILITIES FOR OPTICAL MEASUREMENT TECHNIQUES

by

GREGGORY H. ROTHMEIER

Under the Direction of A. G. Unil Perera, Mukesh Dhamala

ABSTRACT

Regional tissue temperature dynamics in the brain is determined by the balance of the metabolic heat production rate and heat exchange with blood flowing through capillaries embedded in the tissue, the surrounding tissues and the environment. Local changes in blood flow and metabolism during functional activity can upset this balance and induce transient temperature changes. Invasive experimental studies in animal models have established that the brain temperature changes during functional activity are observable and a definitive relationship exists between temperature and brain activity. We present a theoretical framework that links tissue temperature dynamics with hemodynamic activity allowing us to non-invasively estimate brain temperature changes from experimentally measured blood-oxygen level dependent (BOLD) signals. With this unified approach, we are able to pinpoint the mechanisms for hemodynamic activity-related temperature increases and decreases. In addition to this, the potential uses and limitations of optical measurements are discussed.

INDEX WORDS: Functional magnetic resonance imaging, Blood oxygen level dependent, Brain temperature, Functional near-infrared spectroscopy

BRAIN TISSUE TEMPERATURE DYNAMICS DURING FUNCTIONAL ACTIVITY AND  
POSSIBILITIES FOR OPTICAL MEASUREMENT TECHNIQUES

by

GREGGORY H. ROTHMEIER

A Thesis Submitted in Partial Fulfillment of the Requirements for the Degree of

Master of Science

in the College of Arts and Sciences

Georgia State University

2012

Copyright by  
Greggory H. Rothmeier  
2012

BRAIN TISSUE TEMPERATURE DYNAMICS DURING FUNCTIONAL ACTIVITY AND  
POSSIBILITIES FOR OPTICAL MEASUREMENT TECHNIQUES

by

GREGGORY H. ROTHMEIER

Committee Chair:

A. G. Unil Perera

Mukesh Dhamala

Committee:

Brian Thoms

D. Michael Crenshaw

Electronic Version Approved:

Office of Graduate Studies

College of Arts and Sciences

Georgia State University

May 2012

## **Dedication**

This is dedicated to my parents who made me go to college and to Brooke who inspired me to go to graduate school. If I wasn't lucky enough to have all of you I would probably be working for Geek Squad.

## Acknowledgments

I want to thank my advisors A. G. Unil Perera and Mukesh Dhamala for their guidance and leadership through my graduate school career. Likewise, I must thank everyone in Dr. Perera's and Dr. Dhamala's labs for always being helpful over the past couple of years. Thank you. The work presented here was done while under the support of the GSU Brains and Behavior fellowship and a faculty seed grant awarded to Mukesh Dhamala and A. G. Unil Perera.

## Table of Contents

Dedication . . . . .	iv
Acknowledgments . . . . .	v
List of Tables . . . . .	viii
List of Figures . . . . .	ix
List of Abbreviations . . . . .	x
<b>1 Introduction . . . . .</b>	<b>1</b>
<b>2 Calculating Temperature Changes using the fMRI BOLD Response . . .</b>	<b>4</b>
2.1 Introduction . . . . .	4
2.1.1 Single-Voxel Methods . . . . .	4
2.1.2 Multi-Voxel Methods . . . . .	6
2.2 Our Approach . . . . .	7
2.2.1 Preparing the model of the head . . . . .	8
2.2.2 Calculating the equilibrium temperature . . . . .	10
2.2.3 Calculating Metabolism and Blood Flow Changes from the BOLD response . . . . .	10
2.2.4 Calculating the change in temperature in the active brain . . . . .	13
2.3 Results . . . . .	15
2.3.1 Using Synthetic BOLD Data . . . . .	17
2.3.2 Using Experimental BOLD Data . . . . .	18
<b>3 Optical techniques for brain activity measurements . . . . .</b>	<b>21</b>

3.1	Functional Near-Infrared fNIR Spectroscopy . . . . .	21
3.2	Thermal Imaging . . . . .	25
<b>4</b>	<b>Conclusion . . . . .</b>	<b>31</b>
	<b>References . . . . .</b>	<b>33</b>
	<b>Appendices . . . . .</b>	<b>41</b>
<b>A</b>	<b>Code . . . . .</b>	<b>41</b>
A.1	Creating the Head Matrix . . . . .	46
A.1.1	ImportSegmentedT1() . . . . .	46
A.1.2	fillAir() . . . . .	50
A.1.3	fillHoles() . . . . .	51
A.1.4	build_skin() . . . . .	53
A.1.5	repair_headdata() . . . . .	54
A.2	Loading the fMRI Data . . . . .	57
A.2.1	sample_bold_import() . . . . .	57
A.2.2	avg_NII_rest() . . . . .	62
A.2.3	avg_NII_normalize() . . . . .	65
A.2.4	BOLDtoMF() . . . . .	69
A.2.5	lambw() and lambw_mex() . . . . .	73
A.3	Calculating the Equilibrium Temperature . . . . .	75
A.3.1	tempCalcEquilibrium() . . . . .	75
A.4	Calculating the Temperature Change . . . . .	82
A.4.1	tempCalcDynMF . . . . .	82
<b>B</b>	<b>Visualization Tools . . . . .</b>	<b>89</b>
B.1.1	TempPlot() . . . . .	89
B.1.2	tsliceplot . . . . .	92



## List of Tables

2.1	Tissue-specific parameters . . . . .	9
2.2	Parameters used in the single-voxel approximation . . . . .	11
3.1	Comparison of fMRI and fNIR . . . . .	21

## List of Figures

1.1	Generation of the fMRI BOLD response and a corresponding temperature change . . . . .	2
2.1	Procedure used to calculate temperature change . . . . .	8
2.2	Slice of the segmented head . . . . .	9
2.3	Temperature changes: simulated BOLD data . . . . .	16
2.4	Temperature changes: experimental BOLD data . . . . .	19
3.1	Absorption spectra of water, deoxyhemoglobin and oxyhemoglobin . . . . .	22
3.2	Sample arrangement of detectors and emitters in a typical fNIRS setup . . .	23
3.3	Penetration by fNIR . . . . .	24
3.4	Black-body Spectrum at 250, 310 and 350 K . . . . .	27
3.5	Wide-band absorption spectra of water . . . . .	28
A.1	temptools: main window . . . . .	42
A.2	temptools: calculate the equilibrium temperature window . . . . .	43
A.3	temptools: calculate temperature during activity . . . . .	44
A.4	temptools: visualize the data . . . . .	45
B.1	Visualization using tsliceplot . . . . .	93
B.2	Visualization using tsliceplot (z v. t plane) . . . . .	94

## List of Abbreviations

BOLD	Blood Oxygen Level Dependent
CBF	Cerebral Blood Flow
CMRO <sub>2</sub>	Cerebral metabolic rate of O <sub>2</sub>
CSF	Cerebral Spinal Fluid
deoxyHb	Deoxyhemoglobin
fMRI	Functional Magnetic Resonance Imaging
fNIRS	Functional Near-Infrared Spectroscopy
OLM	Oxygen Limitation Model
oxyHb	Oxyhemoglobin
ROI	Region of Interest
TRS	Time Resolved Spectroscopy

## 1 Introduction

Since its invention in the 1950's [4] and later development in the 1970's [5], Magnetic Resonance Imaging (MRI) has allowed physicians and scientists a detailed view within the human body. Beginning in the 1990's, it was realized that changes in the local metabolic state affect the local magnetic resonance and provide an indication of brain activity [6, 7]. This is possible because hemoglobin is diamagnetic in an oxygenated state and paramagnetic when deoxygenated. Thus as the local concentration of deoxyhemoglobin changes, the local magnetic susceptibility is also altered.

By combining this effect with a discovery made earlier in 1986 [8], MRI becomes a powerful tool for measuring brain activity. Fox and Raichle [8] found that with an increase in neuronal activity came an increase in local cerebral blood flow (CBF) that exceeded the increase in cerebral metabolic rate of oxygen ( $CMRO_2$ ). The change in local tissue oxygenation created by uncoupled changes in CBF and  $CMRO_2$  is referred to as the blood oxygen dependent (BOLD) response [7]. A schematic of the model for generation of the fMRI BOLD response is provided in Fig. 1.1.

A stimulus within a region of the brain induces either an excitatory or inhibitory (or a combination) neuronal response. An increase in excitatory neuronal activity triggers an increase in CBF which overcompensates for the increase in  $CMRO_2$  [8]. Conversely, an increase in inhibitory neuronal activity does not induce a change in CBF. Increasing  $CMRO_2$  increases the concentration of deoxyhemoglobin (deoxyHb) while increasing CBF delivers more blood to the tissue thereby increasing the concentration of oxyhemoglobin (oxyHb) and increasing local tissue oxygenation. The change in blood oxygenation is detected by the fMRI as the BOLD response [7].

Along with changes in the BOLD response, changes in CBF and  $CMRO_2$  also affect the local tissue temperature. As glucose is metabolized, heat is released that is primarily

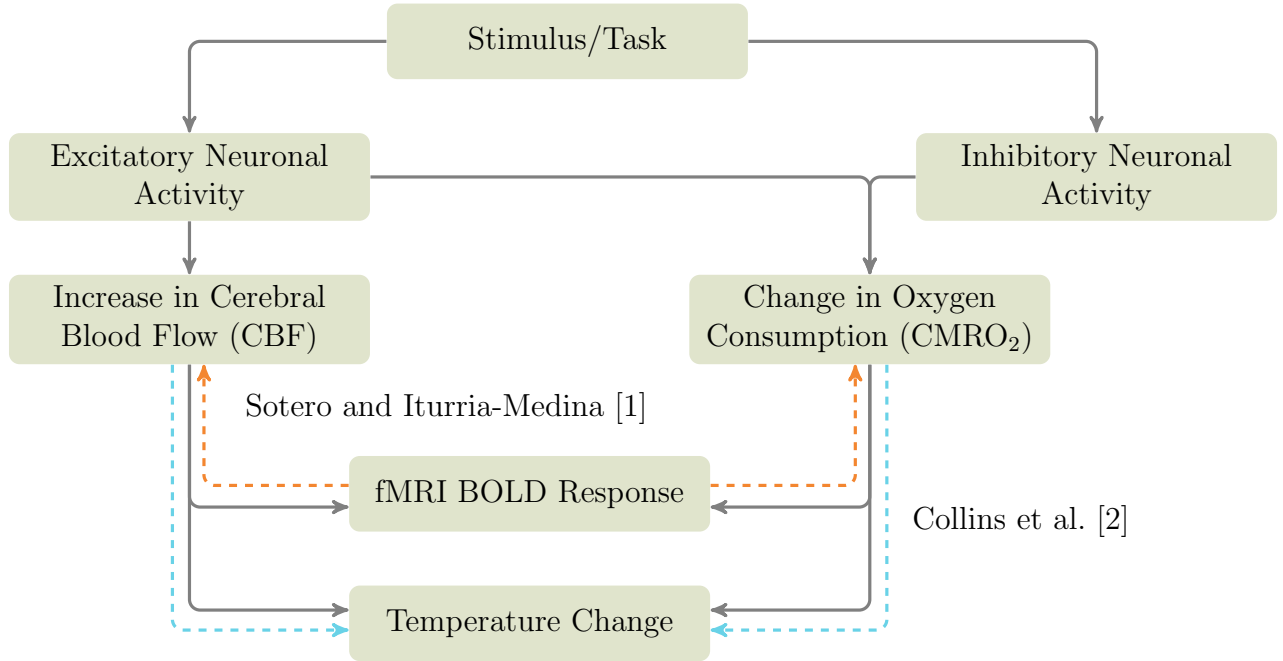


Figure 1.1 Generation of the fMRI BOLD response from changes in neuronal activity. Black arrows indicate a causal relationship while colored dashed-arrows indicate existing models for the relationship. The orange line (●) shows the model proposed by Sotero and Iturria-Medina [1] to calculate cerebral blood flow and metabolism and the blue line (●) shows how the model proposed by Collins et al. [2] is used to calculate temperature. Modified after Sotero and Trujillo-Barreto [3].

dissipated by convection to blood. The human brain relies on CBF to remove excess heat from the brain [9]; thus, a change in rate of blood flow will affect how quickly heat can be dissipated (or delivered as we'll explore in section 2.3). Since both BOLD and temperature are dependent on CBF and CMRO<sub>2</sub>, it is possible to use the BOLD data collected to calculate the temperature change. As will be further discussed in section 2.3.1, the resulting temperature change cannot be easily characterized for the entire brain because it's behavior is spatially dependent.

Experimental measurements of activity-induced brain temperature changes are mixed in whether an increase in brain activity increases or decreases local tissue temperature [10, 11, 12, 13, 14]. Current temperature models predict that an increase in activity will result in a decrease in temperature [1, 15, 16]. This is generally the prediction because these models

generalize the resting-state conditions of the voxel (ignoring spatial dependence) which puts the blood temperature below the resting state tissue temperature. An increase in blood flow then acts as a coolant for the tissue and lowers the temperature (more on this in section 2.1).

An accurate model for brain temperature could be valuable for many clinical applications [17]. A seizure has been shown to induce a large change in brain temperature [18], so if an fMRI measurement is made of a patient during a seizure, the temperature change could be calculated. It has been shown that treating severe head injuries with induced mild hypothermia greatly improves the clinical outcome [19]. A similar result has been shown in patients being treated for brain ischemia [20, 21] and stroke [22]. Additionally, it has been shown that mild hypothermia can be neuroprotective in infants undergoing heart surgery [23]. During treatment, the brain temperature is currently inferred by indirect means. This can be achieved by measuring the temperature of the temporalis muscle [24], the tympanic canal [25, 26], nasopharynx [27] or by the use of a venous bulb in the jugular [28, 29]. While commonly used, their reliability has not been validated [26, 29]. A model of brain temperature could let physicians better understand the relationship between brain temperature and the measurements they are able to make. This could make hypothermia-based treatments more effective since the brain temperature could be better controlled.

I will explore a model which calculates temperature using the fMRI BOLD response for the entire brain, thereby accounting for spatial dependencies. Additionally, in chapter 3 I'll explore optical measurement techniques including functional near-infrared spectroscopy (fNIRS) and the use of thermal imaging cameras to measure activity-induced brain temperature changes.

## 2 Calculating Temperature Changes using the fMRI BOLD Response

### 2.1 Introduction

Using fMRI to find brain temperature is enticing because it is noninvasive. Existing efforts to model temperature changes be can categorized into two classes. The first class approaches the problem by considering a single region of interest (ROI) deep within the brain (single-voxel approach) while the second approach considers the brain and head as an entire system (multi-voxel approach). The single-voxel approach is unable to explain the experimental observation of an increase in temperature during a task; however a multi-voxel approach is able to account for this observation.

#### 2.1.1 Single-Voxel Methods

Numerous single-voxel approaches have been created [1, 15, 16] which differ largely only in their formulation of Penne's Bioheat Equation [30]. Although different approaches consider different contributions to the temperature change, they all narrow the problem down to a single region of interest. By simplifying the model, the heat equation can be simplified and the calculation is much easier to undertake. However, since the brain is not homogenous, the values used for parameters such as heat production and thermal conductivity are taken from an average of the tissues. As a result, this reduces the level of accuracy these models can achieve.

The most recently published iteration of a single-voxel model was published by Sotero and Iturria-Medina [1] in 2011. Their formulation of Penne's Bioheat Equation is as follows [30, 1].

$$C_{tissue} \frac{dT(t)}{dt} = (\Delta H^\circ - \Delta H_b) CMRO_2 |_0 m(t) - \rho_b C_b CBF |_0 f(t)(T(t) - T_a) - \frac{C_t}{\tau}(T(t) - T_0) \quad (2.1)$$

where  $C_{tissue}$  is the specific heat of the tissue,  $\Delta H^\circ$  is the enthalpy released in the oxidation

of glucose,  $\Delta H_b$  is the enthalpy used to release oxygen from hemoglobin,  $CMRO_2|_0$  is the metabolic rate of oxygen at rest,  $\rho_b$  is the blood density,  $C_b$  is the specific heat of blood,  $CBF|_0$  is the cerebral blood flow at rest,  $T_a$  is the arterial blood temperature,  $C_T$  is the specific heat for the tissue, and  $\tau$  is a time constant for conductive heat loss. The values used are provided in Table 2.2.

One advantage of using Eq. (2.1) is that the resting state temperature can be analytically determined by substituting  $\frac{dT(t)}{dt} = 0$  [1].

$$T_0 = T_a + \frac{(\Delta H|^\circ - \Delta H_b)CMRO_2|_0}{\rho_b C_b CBF|_0} \quad (2.2)$$

If the values provided in Table 2.2 are substituted into Eq. (2.2), a resting temperature of 37.3057°C is found (using  $T_a = 37^\circ\text{C}$ ). Regardless of the arterial blood temperature, since the second term of Eq. (2.2) is always positive the resting state temperature will always be greater than the arterial blood temperature. Thus, an increase in blood flow will remove heat from the ROI, thereby lowering the temperature. As will be further discussed in section 2.3.1, this is an acceptable result for the majority of the brain; however a multi-voxel approach reveals that a region of the brain exists where a single-voxel model is unable to predict temperature changes. Since it is a simpler model than a multi-voxel model, it is easier to first understand the principles of Penne's Bioheat Equation using this model before discussing how it can be applied to the entire head.

While Eq. (2.1) appears complicated, conceptually Penne's Bioheat Equation can be easily understood:

$$\begin{aligned} \text{heat capacity} * \text{change in temperature} &= \text{heat generated by metabolism} \\ &- \text{heat lost due to convection} - \text{heat lost due to conduction} \\ &- \text{heat lost due to radiation} \end{aligned} \quad (2.3)$$

The system is a balance between heat generation (metabolism) and heat transfer (conduc-



tion, convection and radiation). The direction of heat transfer by convection is determined by the difference between the voxel temperature and the arterial blood temperature ( $T(t) - T_a$ ). Similarly, the direction of heat transfer by conduction is determined by the difference between the voxel temperature and the temperature of the surrounding tissue ( $T(t) - T_0$ ). Since  $T_a$  is less than  $T_0$ , an increase in blood flow ( $f(t)$ ) will remove heat from the voxel thereby decreasing the temperature. Conversely, an increase in metabolism ( $m(t)$ ) without a corresponding change in blood flow, will result in tissue warming. Equation (2.1) omits the effect of radiation because the contribution is small compared to the other terms. As we will see in section 3.2, mid-infrared photons are reabsorbed by tissue within  $\sim 10\text{--}100\text{ }\mu\text{m}$  (Fig. 3.5) so any energy lost due to radiation does not travel very far compared to the ROI size.

### 2.1.2 Multi-Voxel Methods

The multi-voxel approach to calculating brain tissue temperature alleviates many of the issues that a single-voxel approach has. The most prominent advantage a multi-voxel approach has is a result of it accounting for a voxel's location relative to the surface of the head and other voxels. By accounting for a voxel's location, the same BOLD response in two different locations can have vastly different effects on the local tissue temperature (more on this in section 2.3.1).

Multi-voxel methods use a three-dimensional implementation of Penne's Bioheat Equation [2].

$$\rho c \frac{dT}{dt} = k \nabla^2 T - \rho_{blood} f(t) w c_{blood} (T - T_{blood}) + m(t) Q_m \quad (2.4)$$

where  $\rho$  is the tissue density,  $c$  is the specific heat of the voxel,  $k$  is the thermal conductivity,  $\rho_{blood}$  is the blood density,  $w$  is perfusion by blood,  $c_{blood}$  is the specific heat of blood,  $T_{blood}$  is the arterial blood temperature, and  $Q_m$  is the baseline metabolic heat production.  $f(t)$  and  $m(t)$  are the time-dependent changes in blood flow and metabolism. These two factors determine the short-term change in temperature and are calculated from the fMRI BOLD response; however what makes this approach more complete than a single-voxel approach is

that the relatively slow conductive heat loss makes for a different equilibrium temperature at each voxel. This effect can only be captured by considering the entire head. The approach we use is a multi-voxel approach, so more details about this model are discussed in section 2.2.

## 2.2 Our Approach

Our approach [31, 32] combines a multi-voxel model (section 2.1.2) with a model for calculating the change in metabolism and blood flow from the BOLD response. Penne’s Bioheat Equation (Eq. (2.4)) [30, 1] includes three terms. The first and second terms describe heat exchange by conduction to surrounding tissues (or air) and convection to blood, respectively. The third term describes heat generated by metabolism. On shorter time scales, the first two terms dominate and are sufficient for determining activity-induced temperature changes; however, the third term becomes important on longer time scales because it plays a role in determining the resting-state temperature.

Conductive heat transfer to surrounding tissues is a comparatively slow process, but on larger time scales, conduction plays an important role in determining the resting state temperature. When calculating the temperature change, it is important to first have an accurate resting state temperature since it can either be greater than or less than the arterial blood temperature. By considering the entire head, the model we use is able to accurately determine a resting state temperature for each voxel, enabling far more accurate temperature calculations than what is capable with single-voxel approaches.

Figure 2.1 gives a schematic of the temperature calculation procedure. The orange blocks represent the required data. The first thing to be done is establish the resting-state temperature for each voxel within the head. The details of this procedure are given in section 2.2.2, but in summary a T1 contrast image is segmented using SPM8 (Fig. 2.2) and is combined with tissue-specific parameters (Table 2.1). The resulting dataset is then used to determine the resting-state temperature by repeatedly applying Eq. (2.4) until the temperature stabilizes for all voxels.

The resting-state time slices from the fMRI BOLD dataset are averaged to create a resting

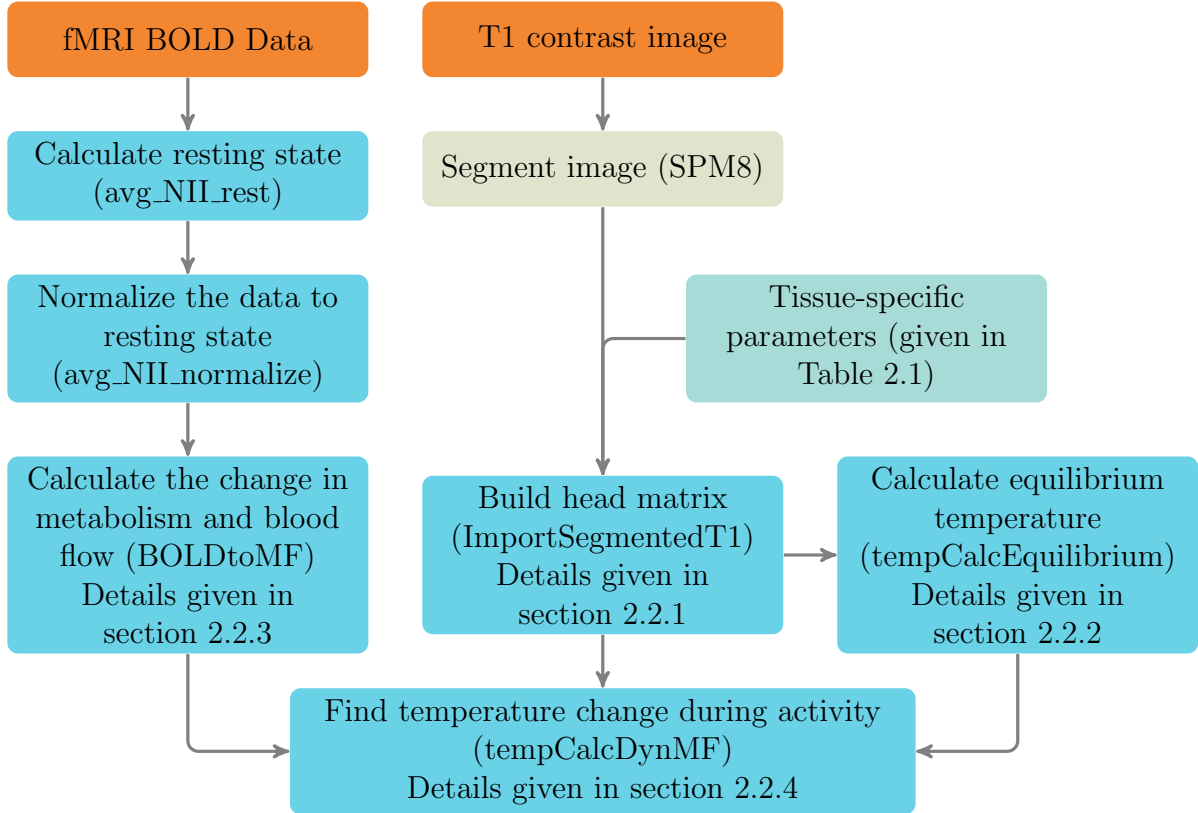


Figure 2.1 The procedure used to calculate temperature from BOLD data. Orange blocks (●) represent data, the sandy-colored block (●) is a step done using SPM8 and the teal blocks (●) are steps done using a function provided within temptools (appendix A). The name of the function used is in parentheses.

state representative slice. The remaining BOLD data is then normalized to this in order to have the normalized change in BOLD response ( $\frac{\Delta S}{S_0}$  in Eq. (2.16)). This can then be used with Eqs. (2.9), (2.15) and (2.16) to create a time series for the change in blood flow and metabolism. More details about this procedure are given in section 2.2.3.

The following sections provide a detailed explanation of the theory behind our modeling approach. The code used to implement this procedure is provided and documented in appendix A.

### 2.2.1 Preparing the model of the head

In order to begin the temperature calculating procedure, a model of the head must first be created. Using SPM8 (<http://www.fil.ion.ucl.ac.uk/spm/>), we segmented a T1 contrast

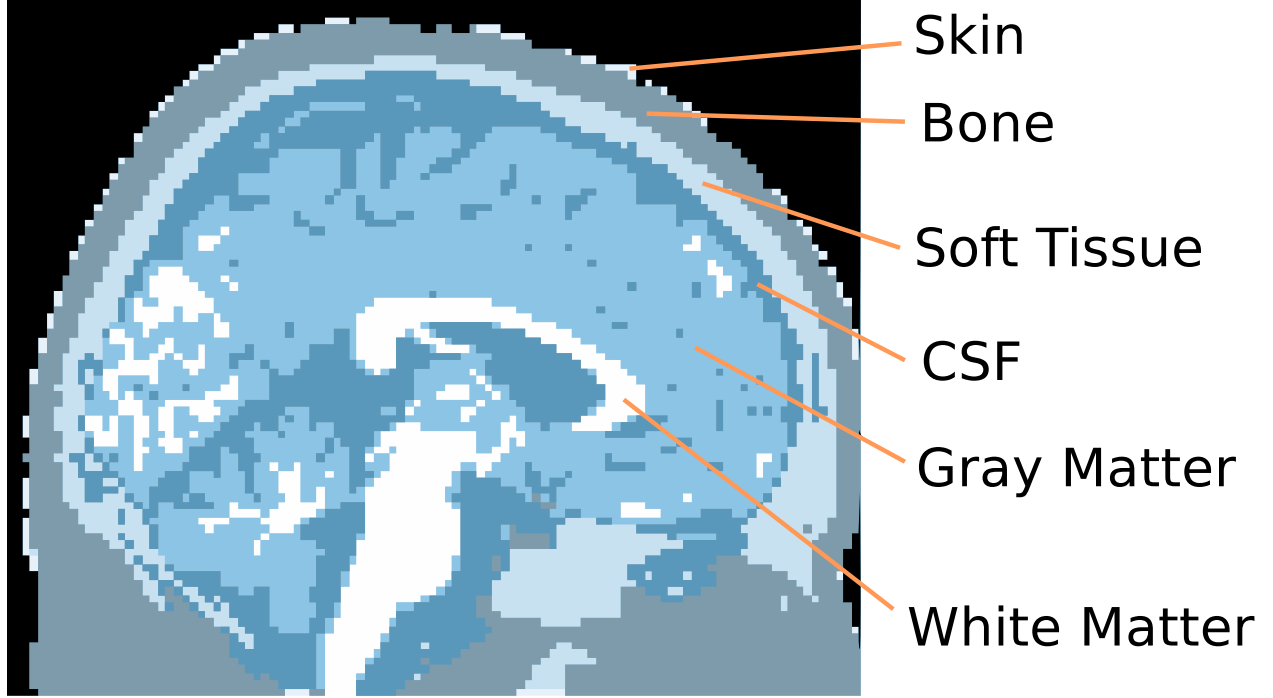


Figure 2.2 Slice of the segmented head. Each color represents a different tissue type.

image of the head into five different tissue types: bone, cerebral spinal fluid, gray matter, white matter and soft tissue. It was assumed that soft tissue voxels that are in contact with air are more appropriately labeled as skin, so in total we are left with a model of the head separated in to six tissue types (Fig. 2.2). The advantage this has is that we are able to use tissue specific parameters when doing the calculations, thereby improving the accuracy of the results. The parameters used are available in Table 2.1. The code used to create the

Tissue	$f_0$ 100 ml/(g min)	$\rho$ kg/m <sup>3</sup>	$c$ J kg <sup>-1</sup> °C <sup>-1</sup>	$k$ W m <sup>-1</sup> °C <sup>-1</sup>	$Q_m$ W/m <sup>3</sup>
Bone	3	1,080	2,110	0.65	26.1
Cerebrospinal Fluid	0	1,007	3,800	0.50	0
Gray Matter	67.1	1,035.5	3,680	0.565	15,575
White Matter	23.7	1,027.4	3,600	0.503	5,192
Muscle	3.8	1,041	3,720	0.4975	687
Skin	12	1,100	3,150	0.342	1,100

Table 2.1 Tissue-specific parameters used to calculate the temperature change (values from Collins et al. [2]).

head matrix is discussed in appendix A.1.

### 2.2.2 Calculating the equilibrium temperature

The first step in calculating the temperature change is to know the resting state temperature for each voxel within the head. Our approach was to have the initial temperature for all tissue voxels set to 37°C and air voxels are kept at 24°C. The starting temperature of the tissue does not affect the final resting state temperature; however, starting off at drastically different values could greatly increase the calculating time required before the temperature stabilizes. The finite difference implementation of Penne’s Bioheat Equation (Eq. (2.4)) is used to update the temperature (derivation provided in section 2.2.4). The temperature is updated until the temperature for every voxel has stabilized ( $\frac{dT}{dt} < 10^{-6} \text{ }^\circ\text{C/s}$ ). Since temperature changes due to changes in neuronal activity are typically greater than  $10^{-2} \text{ }^\circ\text{C}$ , a change in temperature less than  $10^{-6} \text{ }^\circ\text{C/s}$  is sufficiently small that transient temperature changes are negligible and temperature can be considered stabilized. The code used to calculate the equilibrium temperature is detailed in appendix A.3.

### 2.2.3 Calculating Metabolism and Blood Flow Changes from the BOLD response

This is the critical step where we use fMRI BOLD data to calculate the normalized change in metabolism and blood flow. The method used [1] is an assemblage of a couple other works [33, 34, 35, 36, 37, 38]. It starts by using the relation between metabolism and blood flow proposed by Buxton et al. [33]:

$$m(t) = f(t) \frac{E(t)}{E_0} \quad (2.5)$$

where  $E_0$  is the oxygen extraction at rest and  $E(f)$  is

$$E(f) = 1 - (1 - E_0)^{\frac{1}{f(t)}} \quad (2.6)$$

Table 2.2 Parameters used to solve the single-voxel implementation of Penne’s Bioheat Equation. (modified from Sotero and Iturria-Medina [1])

Parameter	Meaning	Value
$T_a$	Arterial blood temperature	37°C
$C_{tissue}$	Tissue Heat Capacity	3.664 J/(gK)
$\Delta H^\circ$	Enthalpy released by oxidation of glucose	4.710 <sup>5</sup> J
$\Delta H_b$	Enthalpy used to release O <sub>2</sub> from hemoglobin	2.810 <sup>4</sup> J
$CMRO_2 _0$	Cerebral metabolic rate of O <sub>2</sub> consumption at rest	0.026310 <sup>-6</sup> mol/(gs)
$CBF _0$	Cerebral blood flow at rest	0.0093 cm <sup>3</sup> /(gs)
$\rho_b$	Blood density	1.05 g/cm <sup>3</sup>
$C_B$	Blood heat capacity	3.894 J/(gK)
$\tau$	Time constant for conductive heat loss from the ROI to the surrounding tissue	190.52 s
a, b, c	Parameters of the gamma function fitted from E(f) vs. f	0.4492, 0.2216, -0.9872
A	Maximum BOLD signal change	0.22
$\alpha$	Steady state flow-volume relation	0.4
$\beta$	Field-strength dependent parameter	1.5
Variable	Meaning	
m(t)	CMRO <sub>2</sub> normalized to baseline	
f(t)	CBF normalized to baseline	
T(t)	Temperature	
W(t)	Lambert W Function	
$\frac{\Delta S(t)}{S_0}$	Change in BOLD signal normalized to rest	

in accordance with the oxygen limitation model [34]. Combining Eq. (2.5) with Eq. (2.6) yields

$$m(t) = \frac{f(t)}{E_0} \left[ 1 - (1 - E_0)^{\frac{1}{f(t)}} \right] \quad (2.7)$$

Sotero and Iturria-Medina [1] goes about solving Eq. (2.7) by adjusting  $E(t)$  data generated by Eq. (2.6) and fitting it to the gamma function for the  $f$  range (0.7–2.0) that is within experimentally reported values [35, 36, 37]:

$$\frac{E(f)}{E_0} = a f^c(t) e^{-b f(t)} \quad (2.8)$$

where  $a$ ,  $b$  and  $c$  are fitting parameters (values provided in Table 2.2). From this approximation we have the final form of metabolism:

$$m(t) = a f^{c+1}(t) e^{-b f(t)}. \quad (2.9)$$

As proposed by Davis et al. [38], the BOLD signal changes ( $\frac{\Delta S(t)}{S_0}$ ) can be described in terms of  $m(t)$  and  $f(t)$ :

$$\frac{\Delta S(t)}{S_0} = \frac{S(t) - S_0}{S_0} = A(1 - f^{\alpha-\beta}(t) m^\beta(t)) \quad (2.10)$$

Substituting Eq. (2.9) into Eq. (2.10) yields

$$f(t) e^{-\frac{b\beta}{\alpha+\beta c} f(t)} = \left( \frac{\left( A - \frac{\Delta S(t)}{S_0} \right)}{A a^\beta} \right)^{\frac{1}{\alpha+\beta c}} \quad (2.11)$$

where  $A$  is the maximum change in BOLD signal. Multiplying each side by  $-\frac{b\beta}{\alpha+\beta c}$  gives

$$-\frac{b\beta}{\alpha+\beta c} f(t) e^{-\frac{b\beta}{\alpha+\beta c} f(t)} = -\frac{b\beta}{\alpha+\beta c} \left( \frac{\left( A - \frac{\Delta S(t)}{S_0} \right)}{A a^\beta} \right)^{\frac{1}{\alpha+\beta c}} \quad (2.12)$$

which can be solved by using the Lambert W function

$$z = W(x) \quad (2.13)$$

where  $W(x)$  is a solution for  $z$  in the equation

$$ze^z = x \quad (2.14)$$

Finally,  $f(t)$  is obtained from Eq. (2.12)

$$f(t) = \frac{\alpha + \beta c}{b\beta} W(y(t)) \quad (2.15)$$

where

$$y(t) = -\frac{b\beta}{\alpha + \beta c} \left[ \frac{(A - \frac{S(t)}{S_0} - 1)}{Aa^\beta} \right]^{\left(\frac{1}{\alpha + \beta c}\right)} \quad (2.16)$$

is a function of the BOLD signal. Using Eqs. (2.9), (2.15) and (2.16) allows for the metabolism and blood flow to be calculated from the BOLD signal (values used are provided in Table 2.2).

In order to process the files, the BOLD dataset is stored as a separate NIFTI (\*.nii) file for each time step. The first step in processing the data for temperature calculations is to determine a resting state BOLD signal ( $S_0$ ). The resting state is calculated by taking the voxel-wise mean of the data when the subject is at rest (i.e. the first and last 20 seconds). This results in one data set where each voxel is a mean of all of the voxels at the location over time ( $S_0$ ). In order to calculate the metabolism and blood flow, the BOLD dataset needs to be normalized to this resting state ( $\frac{\Delta S(t)}{S_0}$ ).

Once  $\frac{\Delta S(t)}{S_0}$  is known for each time step, Eqs. (2.9), (2.15) and (2.16) can be used to calculate the metabolism and blood flow. The implementation of this procedure is discussed in appendix A.2.



#### 2.2.4 Calculating the change in temperature in the active brain

As discussed in section 2.1.2, the foundation of the model is Penne's Bioheat Equation, Eq. (2.4). The metabolism and blood flow time-courses calculated in section 2.2.3 are used to scale the baseline heat production and blood perfusion. This, in turn, induces a change in the temperature.

Penne's Bioheat Equation (Eq. (2.4)) is implemented using the first-order forward finite difference method:

$$\dot{T}(t) \approx \frac{T(t+l) - T(t)}{l} \quad (2.17)$$

where  $T(t)$  is the temperature at time  $t$  and  $l$  is the time step size. Next, we can approximate  $\nabla^2 T$  by using the second order central finite difference method applied for each coordinate:

$$\begin{aligned} \frac{\partial^2 T(t; x, y, z)}{\partial x^2} &\approx \frac{T(t; x+h, y, z) - 2T(t; x, y, z) + T(t; x-h, y, z)}{h^2} \\ \frac{\partial^2 T(t; x, y, z)}{\partial y^2} &\approx \frac{T(t; x, y+h, z) - 2T(t; x, y, z) + T(t; x, y-h, z)}{h^2} \\ \frac{\partial^2 T(t; x, y, z)}{\partial z^2} &\approx \frac{T(t; x, y, z+h) - 2T(t; x, y, z) + T(t; x, y, z-h)}{h^2} \end{aligned} \quad (2.18)$$

where

$$\nabla^2 T = \frac{\partial^2 T(t; x, y, z)}{\partial x^2} + \frac{\partial^2 T(t; x, y, z)}{\partial y^2} + \frac{\partial^2 T(t; x, y, z)}{\partial z^2} \quad (2.19)$$

and the step size in each coordinate direction is given by  $h$ . In the case of processing BOLD data,  $h$  is the voxel size. Equation (2.19) can be substituted into Penne's Bioheat Equation (Eq. (2.4))

$$\rho c \dot{T} = k_x \frac{\partial^2 T}{\partial x^2} + k_y \frac{\partial^2 T}{\partial y^2} + k_z \frac{\partial^2 T}{\partial z^2} - \rho_{blood} f(t) w_{C_{blood}} (T - T_{blood}) + m(t) Q_m \quad (2.20)$$

where  $\dot{T}$  is the first-derivative of temperature with respect to time,  $k_x$  is the thermal conductivity in the x-direction,  $k_y$  is the thermal conductivity in the y-direction and  $k_z$  is the thermal conductivity in the z-direction. Substituting Eqs. (2.17) and (2.18) into Eq. (2.20)

yields

$$\begin{aligned}
\rho c \frac{T(t+l; x, y, z) - T(t; x, y, z)}{l} \approx & \frac{1}{h^2} [k_x(T(t; x+h, y, z) - 2T(t; x, y, z) + T(t; x-h, y, z)) \\
& + k_y(T(t; x, y+h, z) - 2T(t; x, y, z) + T(t; x, y-h, z)) \\
& + k_z(T(t; x, y, z+h) - 2T(t; x, y, z) + T(t; x, y, z-h))] \\
& - \rho_{blood} f(t) w_{blood} (T - T_{blood}) + m(t) Q_m
\end{aligned} \tag{2.21}$$

which is then rearranged to solve for  $T(t+l; x, y, z)$

$$\begin{aligned}
T(t+l; x, y, z) \approx & T(t; x, y, z) + \frac{l}{\rho c h^2} [k_x(T(t; x+h, y, z) - 2T(t; x, y, z) + T(t; x-h, y, z)) \\
& + k_y(T(t; x, y+h, z) - 2T(t; x, y, z) + T(t; x, y-h, z)) \\
& + k_z(T(t; x, y, z+h) - 2T(t; x, y, z) + T(t; x, y, z-h))] \\
& - \rho_{blood} f(t) w_{blood} (T - T_{blood}) + m(t) Q_m
\end{aligned} \tag{2.22}$$

The time step size ( $l$ ) can be picked arbitrarily, however the spatial step size ( $h$ ) is limited to the voxel spacing.

The final equation (Eq. (2.22)) gives a method for calculating the next time step ( $T(t+l)$ ) from the current time step ( $T(t)$ ) for each voxel. By using the central difference to solve  $\nabla^2 T$ , the voxels on all six sides of the current voxel are considered in the heat conduction. The implementation of this equation is discussed in appendix A.4.1.

## 2.3 Results

In order to understand the behavior of the model, we first applied it simulated BOLD data that was generated by convolving a boxcar function with the hemodynamic response function provided by SPM8. After we understood the behavior of the model, we then applied it to experimental BOLD data that was collected by Dhamala et al. [39].

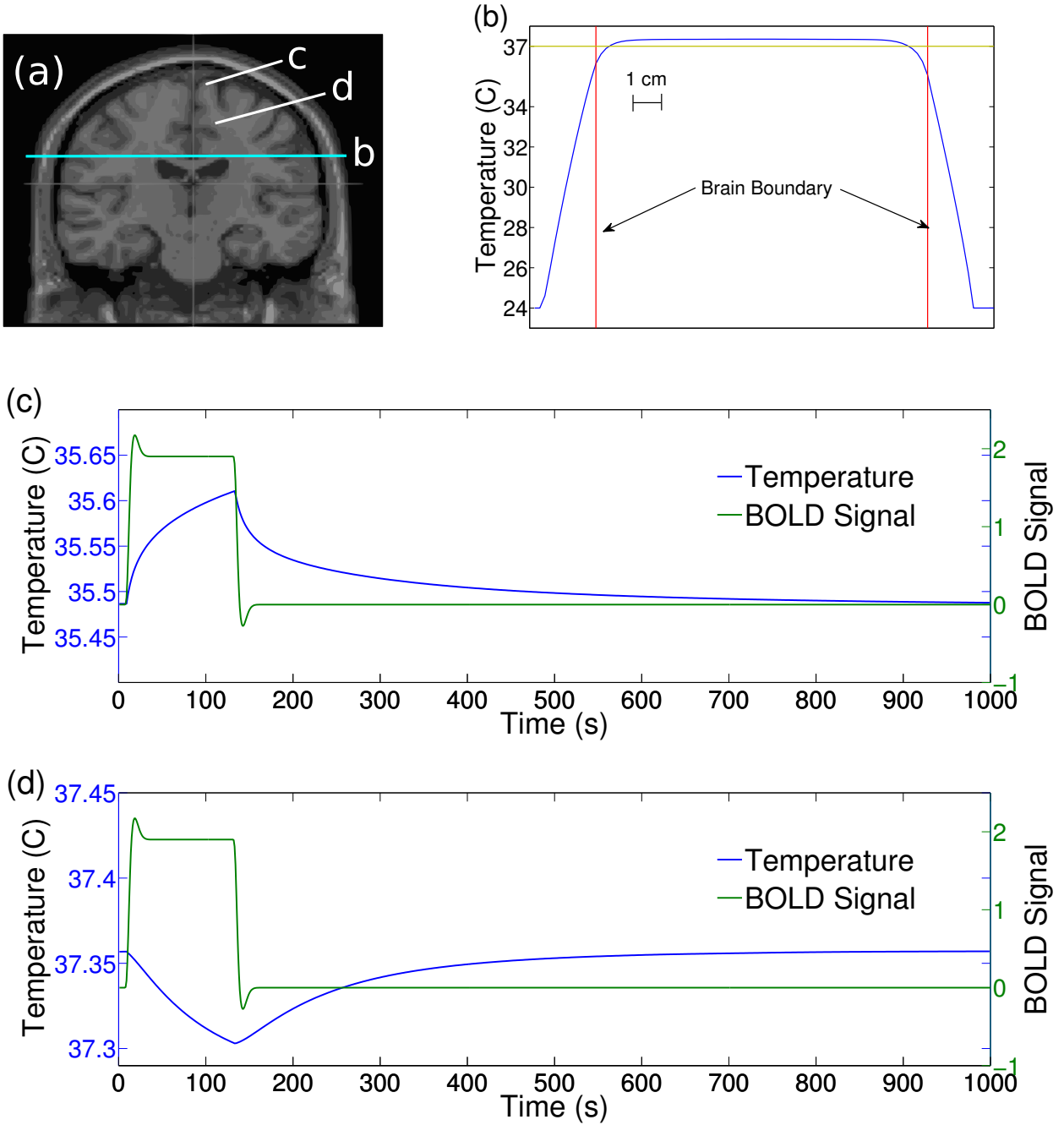


Figure 2.3 Temperature changes using simulated BOLD signals. (a) Slice of the head ( $y = -12$ ) with indicators of the locations for parts (b)-(d). (b) Equilibrium temperature along a line through the head. Red lines indicate the brain boundary and the gold line indicates the blood temperature ( $37^{\circ}\text{C}$ ) used for calculations. Inside the brain, a 4-6 mm thick shell is created where the equilibrium temperature is less than the blood temperature. Within this shell, (c) the temperature rises with increased brain activity while (d) tissue deeper in the brain experiences the opposite effect.

### 2.3.1 Using Synthetic BOLD Data

To better understand the behavior of the tissue temperature model and the characteristics of temperature changes, BOLD activity was simulated in two ways: (i) from a nonlinear hemodynamic model [40] using stimulus or response function, or (ii) by convolving a boxcar function with the canonical hemodynamic response function provided by SPM 8 and corresponding temperature changes were calculated. Figure 2.3(c,d) shows a typical simulated BOLD response used (green curve) along with the change in temperature (blue) for two voxels at different locations in the brain (locations indicated by Fig. 2.3(a)).

Although both voxels have the same BOLD data, they demonstrate contrasting changes in temperature. This can be best understood by considering the equilibrium temperature of each voxel. Figure 2.3(b) is a plot of the equilibrium temperature (blue line) along a line passing through the head (path indicated by the teal line in part (a) of the same figure). The vertical red lines indicate the boundary between the brain and surrounding tissues and the horizontal yellow line is an indication of the blood temperature ( $37^{\circ}\text{C}$ ). Two regions exist within the brain that lead to contrasting temperature behaviors.

The majority of the brain tissue is at a resting-state temperature that is less than the blood temperature (region 1). For voxels within this region, a behavior like that shown in Fig. 2.3(d) is to be expected. The primary contribution to an increase in the BOLD response is an increase in local blood flow. Since the blood temperature is cooler than the tissue temperature, blood flow removes heat from the tissue thereby lowering the temperature. Single-voxel models are able to account for this result because their assumptions about the location of a voxel are consistent with being located within this region.

The second region is comprised of a thin (4–6 mm) layer of brain tissue that is closest to the surface of the head. As a result of its proximity to the surface of the head, conductive heat lost to the air puts the resting-state temperature of voxels in this region below the arterial blood temperature. As a result, when there is an increase in blood flow (increase in BOLD), the warmer blood will increase the voxel temperature (Fig. 2.3(c)). Since single-

voxel models approximate voxel conditions, they are unable to account for this region of tissue.

Conduction is a slow process, so over shorter time scales (less than  $\sim 10$  minutes), conduction will contribute very little to the temperature change from a change in brain activity. However, conduction plays an important role in determining the resting-state temperature.

The primary advantage with this model is that it accounts for the contribution of all of the voxels when determining the temperature, thus the direction of the temperature change depends on how far away from the surface of the head the voxel is. For voxels within a 4–6 mm shell near the surface of the brain, the temperature increases with increased activity (Fig. 2.3(c)) while voxels deeper within the brain experience the opposite change (Fig. 2.3(d)).

### **2.3.2 Using Experimental BOLD Data**

Data from a previous fMRI study [39] was used to study the characteristics of temperature changes in a typical experiment. All participants in this experiment were right handed and between the ages of 23 and 27 years old. Signed informed consent was collected from each one prior to participating in the study. Institutional Review Boards of Emory University and Georgia State University approved this experiment. Twelve participants were asked to tap their right index fingers with rhythms of varying complexity for 320s.

This task resulted in a strong BOLD response in the motor cortex (Fig. 2.4). The experiment included 20s of rest at the beginning and end of the tapping periods. Here, the resting state response level is calculated for each voxel by averaging across 40s of resting-state fMRI data. Using equations Eqs. (2.9), (2.15) and (2.16), the time-dependent change in blood flow and metabolism can be determined for each voxel. Finally, these values are used in conjunction with Eq. (2.4) to find the change in temperature throughout the brain. In this task, a temperature increase of approximately  $0.02^{\circ}\text{C}$  was observed in the motor cortex (Fig. 2.4). This value is well within the range of temperature changes observed in experimental measurements [10, 11, 12, 13, 14].

The increase rather than decrease in temperature in the motor cortex during a functional

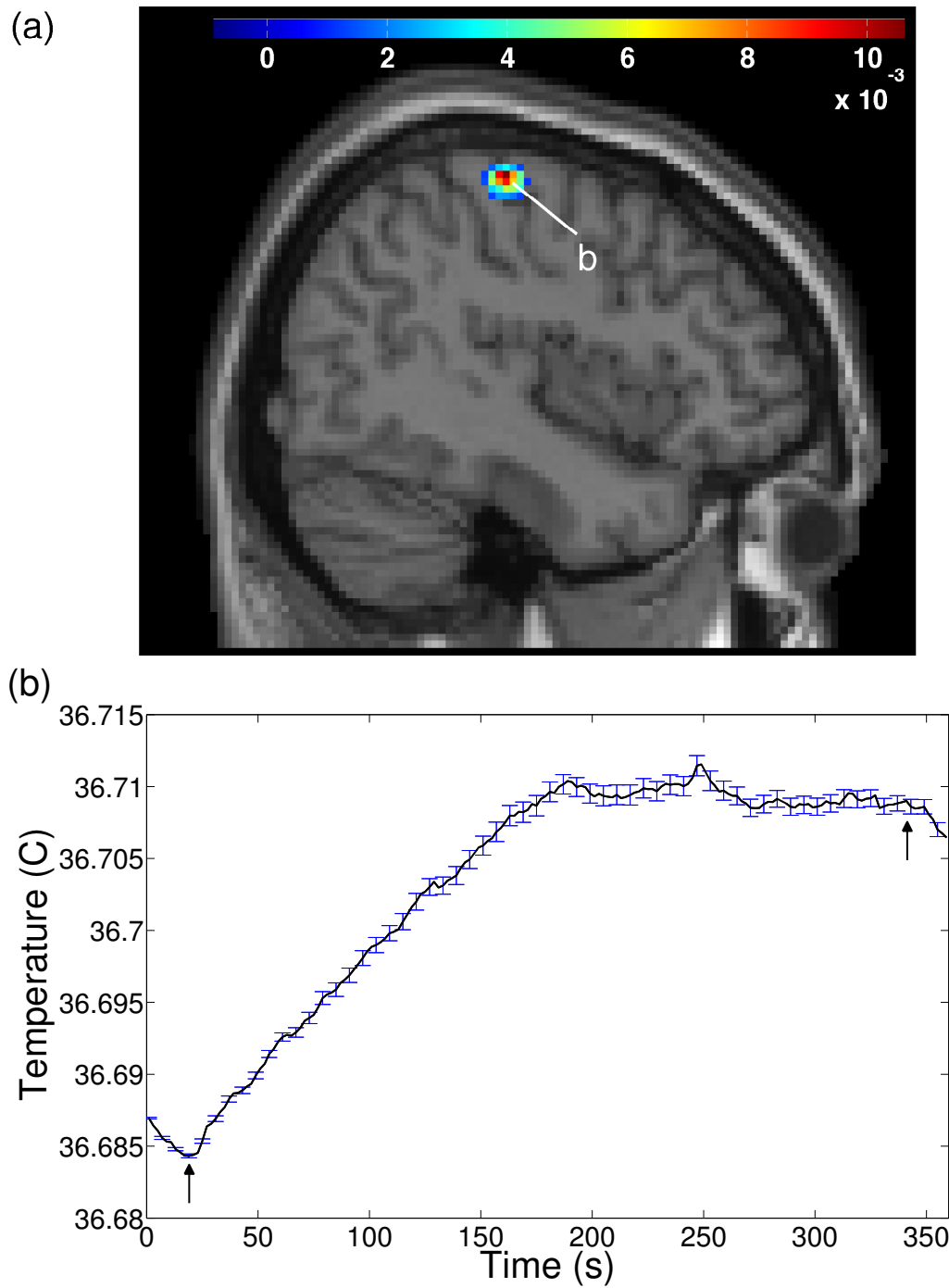


Figure 2.4 Temperature calculated from a voxel within the motor cortex. (a) A slice ( $x = -44$ ) showing the motor cortex warming during a finger-tapping task. (b) Temperature at a voxel within the motor cortex ( $-44, -24, 60$ ) with standard error indicated by blue error bars (Arrows indicate task onset and conclusion,  $N=24$ ).

activity is consistent with the idea that the temperature of the blood in the capillaries is slightly greater than the baseline tissue temperature in superficial cortical regions; however, single-voxel models would predict the opposite effect.

### 3 Optical techniques for brain activity measurements

The best method for measuring brain temperature is to use a thermocouple probe placed in direct contact with the tissue. Experimental measurements of brain temperature have achieved a precision as small as  $0.0003^{\circ}\text{C}$  using this method [10]. However, this method can not be used in humans without damaging the tissue. An optical method would be ideal for non-invasive measurements. Presently, there does not exist a method for accurately measuring the temperature of brain tissues optically. However, other optical measurements methods could be used in conjunction with a temperature model (such as the one proposed here) to calculate the temperature. The possible application of functional Near-Infrared Spectroscopy (fNIRS) and its possible use in brain temperature calculations is discussed along with the possibilities and limitations of a direct measurement technique such as thermal imaging.

#### 3.1 Functional Near-Infrared fNIR Spectroscopy

As discussed in chapter 1, changes in tissue activity can be detected by measuring the change in blood oxygenation levels. Functional Magnetic Resonance Imaging (fMRI) is one technique among several for measuring tissue oxygenation (the BOLD signal).

Blood oxygenation can be determined by measuring the relative concentrations of oxyhe-

	fMRI	fNIR
Spatial Resolution	8–27 mm <sup>3</sup>	$\sim 1\text{--}10\text{ cm}^3$
Temporal Resolution	1–2 s	$\sim 10^{-3}\text{ s}$
Measurement Parameter	blood volume, flow, and O <sub>2</sub> metabolism	oxyHb and deoxyHb concentrations
Motion	Must Remain Stationary	Small movements OK
Penetration	Whole-head	outer 2–4 mm of brain tissue

Table 3.1 Comparison of the capabilities and limitations of fMRI and fNIR techniques. Compiled from Bunce et al. [43], Elliott [44].



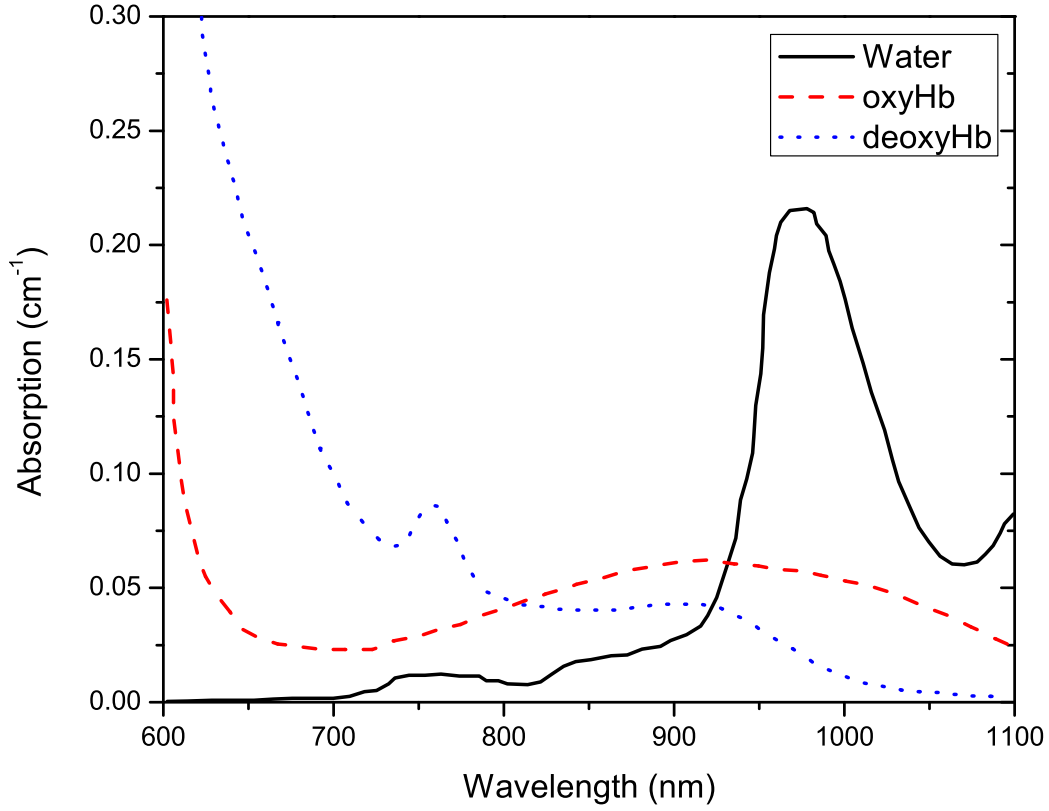


Figure 3.1 Absorption spectra of water Cope [41], oxyHb and deoxyHb Horecker [42].

myoglobin (oxyHb) and deoxyhemoglobin (deoxyHb) [6, 7, 8]. Since oxyHb and deoxyHb have different absorption spectra as shown in Fig. 3.1. These differences are possible to detect through optical techniques. Functional Near-Infrared Spectroscopy is a technique which utilizes two or more spectral bands in order to determine blood oxygenation. fNIRS has a high (millisecond) temporal resolution and a low ( $\sim 1 \text{ cm}^3$ ) spatial resolution compared to fMRI (as low as  $1 \text{ mm}^3$ ). Also, fNIRS is limited to only imaging the outer cortex (2–4 mm) [43]. A comparison of fMRI and fNIR is presented in Table 3.1.

fNIRS works by utilizing an array of near-infrared detectors and emitters (typically spaced 2–3 cm apart) placed in contact with the skin [45, 46]. A schematic of a typical fNIRS array setup is shown in Fig. 3.2. Each dashed line is a detection path. By illuminating the emitters

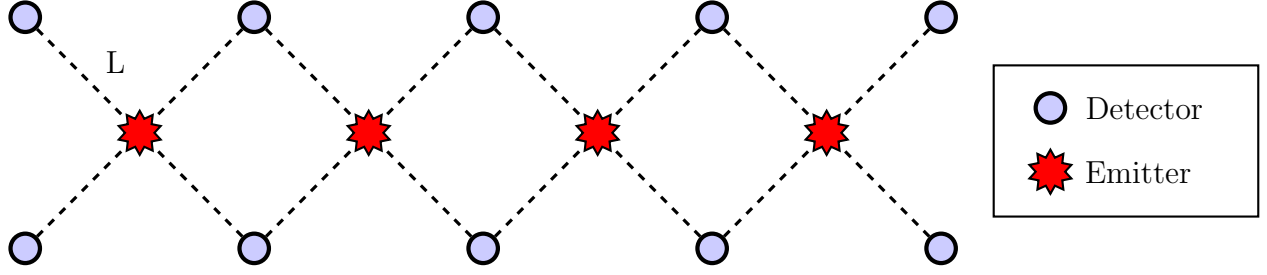


Figure 3.2 A Sample arrangement of detectors and emitters in a typical 16-channel fNIRS setup (based on Izzetoglu et al. [46]). Light coming from emitters (stars) is detected by the detectors (circles) set at a distance  $L$  away.

sequentially, it is possible to have 16 detection channels using the setup shown. The exact spacing between the emitters and detectors determines the depth the light is detected from. As shown in Fig. 3.3, the closer the spacing, the higher the resolution but at the expense of lower penetration. Conversely, in order to detect light passing through deeper tissue, a wider spacing is used which reduces the resolution. Systems use either two [45, 47, 48] or three [49] wavelengths selected based on differences in the absorption of oxyHb and deoxyHb. The exact wavelengths used vary, but all lie within an optical window between 700–1000 nm [45] where the near-infrared photon absorption in the tissue is low (Fig. 3.1).

Three techniques are used to illuminate the tissue: (i) time domain (or time resolved spectroscopy, TRS), (ii) frequency domain and, (iii) continuous wave illumination [46]. In TRS, short pulses of light are incident on the tissue and the temporal distribution of photons is measured. In frequency domain spectroscopy, the amplitude of the incident light is modulated at a high frequency (10–100 MHz) and the phase shift and amplitude decay of the detected light is compared to the incident light [51]. In continuous wave illumination, the incident light is not modulated so the detected light can only be compared for amplitude attenuation [46].

All of the techniques use the Beer-Lambert Law [52]

$$I = I_0 e^{-\alpha(\lambda)x} \quad (3.1)$$

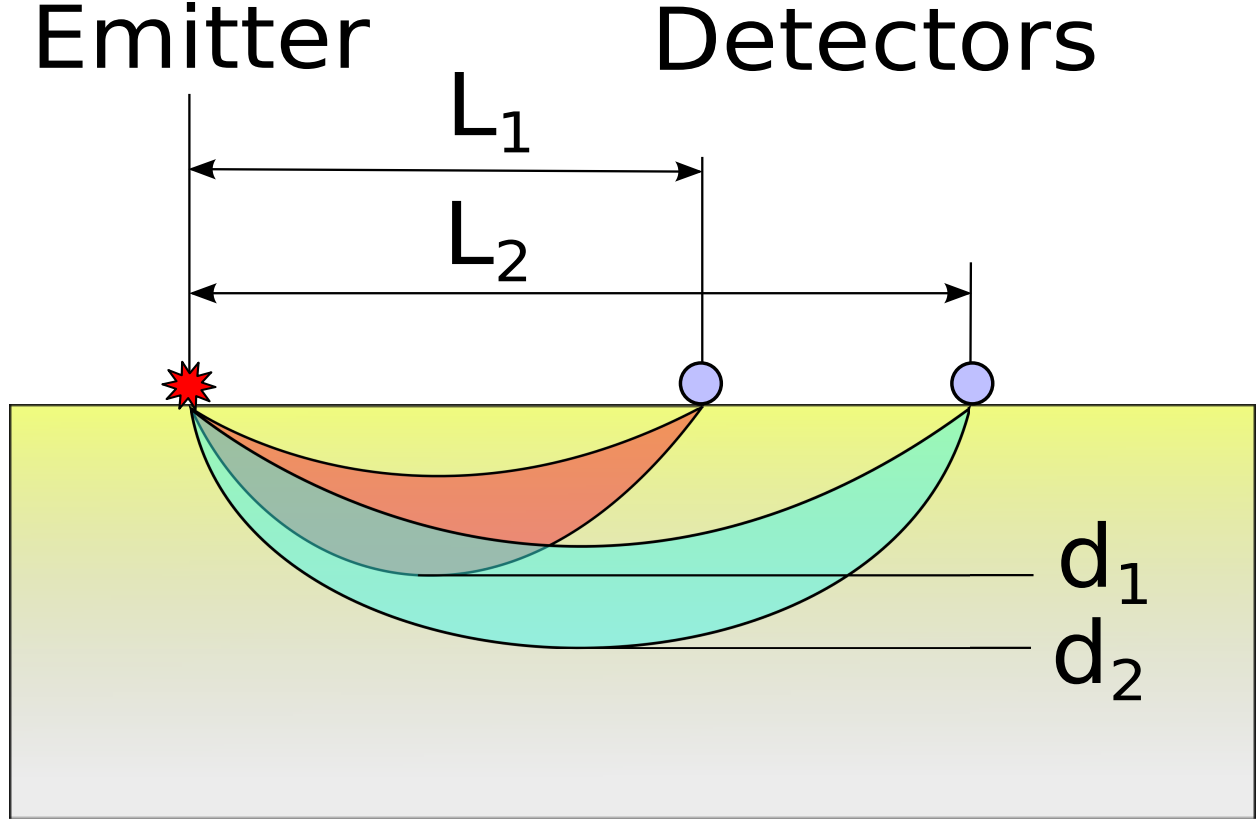


Figure 3.3 Penetration depth of a fNIR detector as a function of the distance between the NIR emitter and detector. Light emitted by either an LED or fiber optic (red star) from the left passes through the tissue before it is scattered back to be detected (blue circle). The path it takes through the tissue is determined by the distance between the emitter and detector. A larger separation between emitter and detector ( $L_2$ ) allows the light to penetrate deeper into the tissue ( $d_2$ ). Modified after [50]

modified to isolate the individual contributions from oxyHb and deoxyHb [41]:

$$I = GI_0 e^{-(\alpha_{\text{deoxyHb}} C_{\text{deoxyHb}} + \alpha_{\text{oxyHb}} C_{\text{oxyHb}})l} \quad (3.2)$$

where  $G$  is a factor to adjust for the measurement geometry,  $I_0$  is the incident light intensity,  $\alpha_{\text{oxyHb}}$  and  $\alpha_{\text{deoxyHb}}$  are the molar extinction coefficients for oxyHb and deoxyHb,  $C_{\text{oxyHb}}$  and  $C_{\text{deoxyHb}}$  are the chromophore concentrations for oxyHb and deoxyHb, and  $l$  is the path length [46]. By comparing a baseline measurement ( $I_b$ ) with a new measurement ( $I$ ),

the optical density can be determined [46]

$$\Delta OD = \log \frac{I_b}{I} = \alpha_{deoxyHb} \Delta C_{deoxyHb} + \alpha_{oxyHb} \Delta C_{oxyHb} \quad (3.3)$$

As discussed in Izzetoglu et al. [46], at least two wavelengths are utilized in the spectral window (700–1000 nm) in order to determine the change in concentration of chromophores  $\Delta C_{deoxyHb}$  and  $\Delta C_{oxyHb}$ . With these values, the oxygenation and total blood volume can be determined:

$$\begin{aligned} \text{Oxygenation} &= \Delta C_{HBO2} - \Delta C_{HB} \\ \text{Blood Volume} &= \Delta C_{HBO2} + \Delta C_{HB} \end{aligned} \quad (3.4)$$

Using this method to experimentally measure the blood oxygenation while measuring the fMRI BOLD response could be used to refine the present model for calculating the metabolism and blood flow from the BOLD response.

While fNIRS does not provide spatially-precise measurements as fMRI, it should be possible to modify the existing model for calculating temperature from the BOLD response to use fNIR data. This would be advantageous because fNIRS systems are cheaper and less disruptive than fMRI systems, meaning they can be used with a wider range of patients (children and the elderly). For this reason, developing a model which uses fNIRS data should be considered in future research.

### 3.2 Thermal Imaging

The primary challenge in brain temperature research is performing brain temperature experimental measurements. Since it is non-invasive, thermal imaging is appealing as a possible replacement for damaging thermocouple probes. Unfortunately, this technique is limited by the high absorption of mid-infrared photons by water.

Light absorption by a material is modeled using the Beer-Lambert law

$$I = I_0 e^{-\alpha(\lambda)x} \quad (3.5)$$

where  $I$  is the intensity at a depth  $x$  remaining from light with an incident intensity  $I_0$  in a material with absorption coefficient  $\alpha$ . The point at which the intensity has decayed to  $1/e$  (about 37%) of the incident intensity is called the penetration depth,  $\delta_p$

$$\delta_p = \frac{1}{\alpha(\lambda)} \quad (3.6)$$

This equation can be used along with the black-body spectrum at tissue temperatures (Fig. 3.4) we can estimate the penetration depth of mid-infrared photons passing through water.

Wien's Displacement Law is a solution to Planck's law for the peak light emission wavelength:

$$\lambda_{max} = \frac{b}{T} \quad (3.7)$$

$$b = 2.897\,7721 \times 10^{-3} \text{ K m}$$

where  $b$  is Wien's displacement constant and  $T$  is the temperature in kelvin. For  $T = 310 \text{ K}$  ( $T = 37^\circ \text{ C}$ ), Wien's law yields a peak black-body emission wavelength of  $9.347\,652 \text{ }\mu\text{m}$ . A physiologically reasonable temperature change to expect from stimulation is on the order of  $0.01^\circ\text{C}$  which corresponds to a new peak wavelength of  $9.347\,350 \text{ }\mu\text{m}$  ( $T=310.01 \text{ K}$ ) or a shift of  $0.302 \text{ nm}$ .

The values of the absorption coefficient and the penetration depth of photons in water is shown in Fig. 3.5. Looking at around  $9.3 \text{ }\mu\text{m}$ , the absorption coefficient is approximately  $700 \text{ cm}^{-1}$  which corresponds to a penetration depth of approximately  $0.14 \text{ }\mu\text{m}$ . This depth is roughly five orders of magnitude smaller than the distance from the surface of the brain to

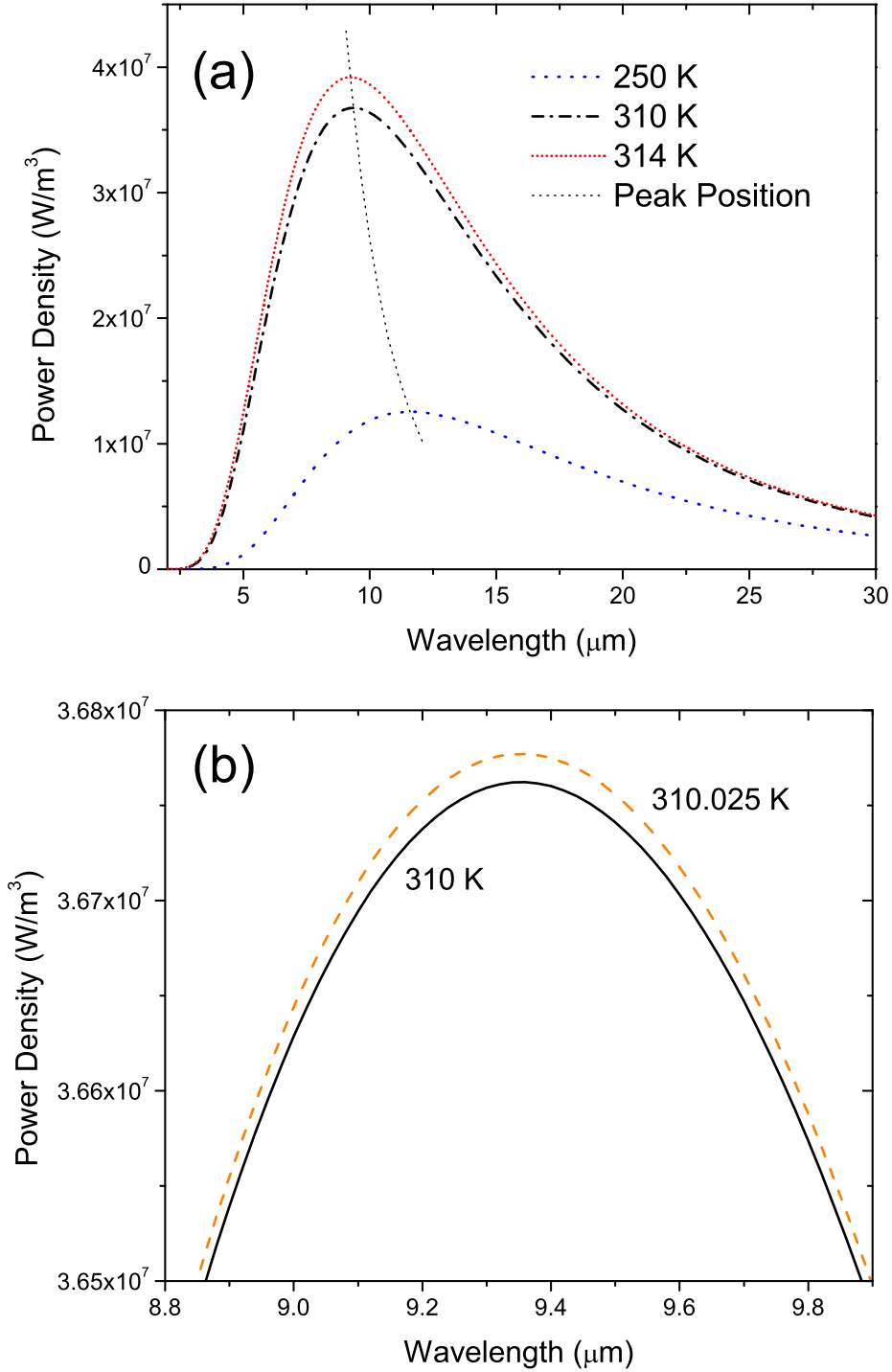


Figure 3.4 (a) Black-body spectrum at 250, 310 and 350 K calculated using Planck's Law. The black dashed line traces the peak in the spectrum as temperature changes. As the temperature increases, differences in temperature translate to smaller shifts in peak wavelength compared to cooler temperatures. (b) A comparison of the black-body spectrum at 310 K and 310.025 K. This is the shift in local temperature that would be expected under strong stimulation.

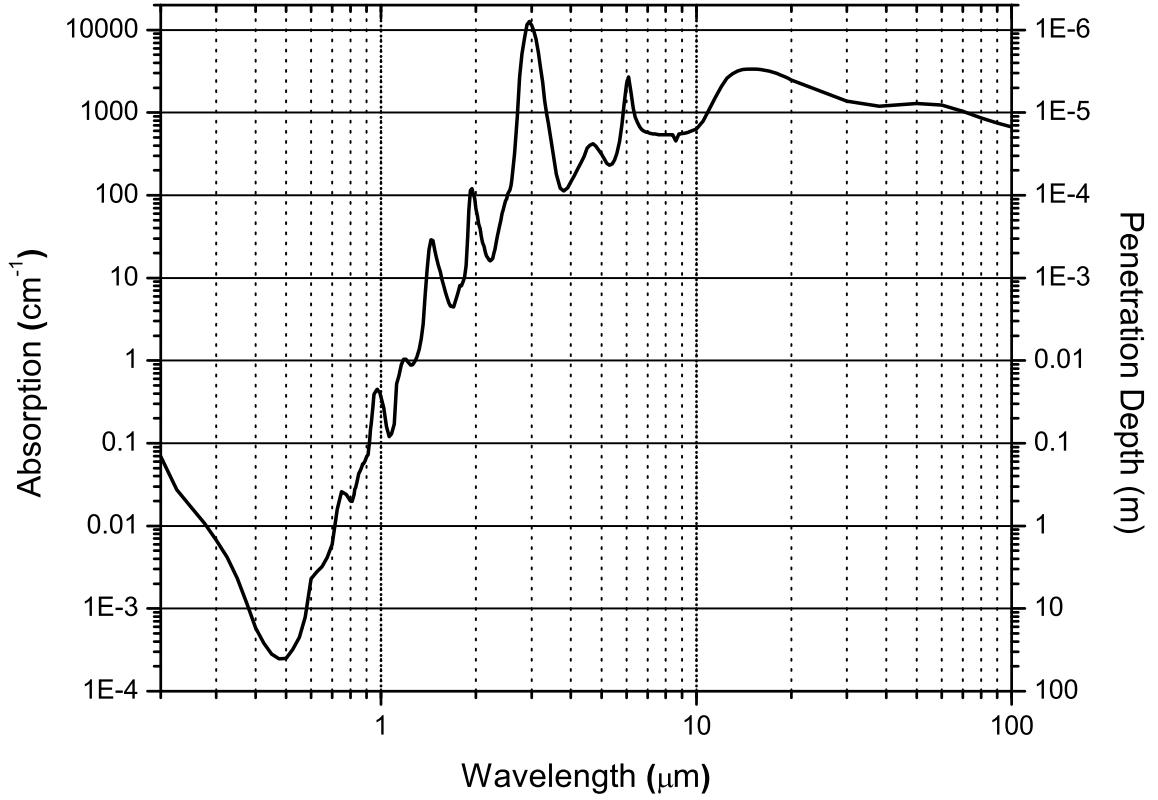


Figure 3.5 The absorptions spectra of water from UV to far-infrared. Modified from Hale and Querry [53].

the surface of the head. Further, all photons emitted as blackbody radiation (ranging from 3  $\mu\text{m}$  to over 30  $\mu\text{m}$ ) have a penetration depth of less than 100 microns. Thus, a thermal imaging camera is unable to image photons coming from the brain.

When thermal imaging is used, the photons collected come from the skin of the head rather than from any deeper tissues, thus it is not a viable form of brain activity detection unless direct line of sight to the brain is available (such as in an open skull surgery). The noise-equivalent temperature difference (NETD) of currently available cameras is greater than 14 mK [54, 55] so it would be limited to only the most extreme of excitations even if line of site to the brain is available. As a comparison, the finger-tapping task discussed in the results section (section 2.3.2) only induced a peak temperature change of 25 mK after

tapping for about 170 seconds. Detection of this activity would be at the limits of a thermal imaging camera.

While its applications to detecting brain activity are limited, thermal cameras could be useful in the operating room. It has been found that inducing mild hypothermia in patients being treated for cerebral ischemia improves the clinical outcome [20]. The same treatment has been shown to improve the outcome of patients who have experienced a stroke [22] and even in patients with severe head injuries [19]. The temperature of the brain is currently inferred from the core body temperature (which is monitored via an invasive thermistor catheter). If it is possible to directly image the brain (i.e. during surgery) then the hypothermia treatment can be better monitored through a thermal imaging camera. This would be especially useful since conductive and radiative heat loss to the air from an exposed brain could reduce how tightly the brain temperature is regulated by the arterial blood temperature. Since the tissue will be directly exposed to the surrounding air, Eq. (2.1) would need to include a term for the radiative heat loss:

$$C_{tissue} \frac{dT(t)}{dt} = \dots + \frac{A\sigma T^4}{\rho_{gm}V} \quad (3.8)$$

After rearranging to solve for  $\frac{dT}{dt}$  and substituting values in, it is found that the change in temperature is approximately 0.07 K/s:

$$\begin{aligned} \frac{dT}{dt} &= \frac{A\sigma T^4}{\rho_{gm}V C_{tissue}} \\ &= \frac{1}{6} \frac{(6 * 4 * 10^{-6} \text{ m}^2) (5.6704 * 10^{-8} \frac{\text{J}}{\text{s m}^2 \text{ K}^4}) (310 \text{ K})^4}{(1035 * 10^3 \frac{\text{g}}{\text{m}^3}) (8 * 10^{-9} \text{ m}^3) (3.664 \frac{\text{J}}{\text{g K}})} \\ &\approx 0.0690454 \frac{\text{K}}{\text{s}} \end{aligned} \quad (3.9)$$

where A and V are the surface area and volume of the voxel,  $\sigma$  is the StefanBoltzmann constant and  $\rho_{gm}$  is the density of gray matter. The factor 1/6 is there because only one face of the voxel is exposed to air. Radiation passing through the other faces will be reabsorbed



by the tissue (Fig. 3.5).

Optical detectors face many challenges working with biological tissue, the worst being infrared light absorption by water. fNIRS works within an optical window in the water absorption in order to measure changes in blood oxygenation, while thermal imaging is limited to measuring the temperature of tissue it has direct line of sight with because of the high absorption of water in the operating window. Despite their limitations, both of these techniques could be used in future studies to improve our understanding of brain temperature dynamics.

## 4 Conclusion

It has been shown that by considering the entire head within the model, brain temperature can be reliably calculated from non-invasive fMRI measurements. Experimental measurements of activity-induced brain temperature changes have shown that a simple relationship does not exist [10, 11, 12, 13, 14]. Single-voxel brain temperature modeling efforts predict that an increase in brain activity will induce a decrease in temperature. This one-dimensional perspective does not account for the spatial distribution of heat throughout the head like a multi-voxel approach.

Our model of brain temperature changes is able to account for the variability found in experimental brain temperature measurements. This is accomplished by modeling heat dynamics throughout the entire head rather than reducing the model to one ROI. It was found that the variability in experiment measurements is most likely due to differences in resting state temperatures throughout the brain. Since each voxel is at a slightly different temperature, the same change in the BOLD response may result in different changes in temperature. Additionally, it was found through the model that a thin (4–6 mm) region of outer cortical tissue is at a resting temperature below the blood temperature. In this region, an increase in brain activity (inducing an increase in CBF) will warm the tissue. Thus, with the same BOLD response, tissue may either be warmed or cooled depending on its proximity to the surface of the head.

The biggest shortcoming of our model is that we are unable to independently compare our calculations with experimental measurements of temperature and BOLD response. It was not possible for us to do this because there is currently not a method for non-invasively measuring temperature independent of an fMRI. An improvement to the model could also be gained by a more accurate method of CMRO<sub>2</sub> and CBF calculations from the BOLD response. The current method uses empirically fit formulas, so the accuracy is limited by

the data used for the fitting. A model that does not rely on experimental data would be ideal. The calculations could also be improved by segmenting the head into more tissue types. We used six tissue types, but the use of more would further improve the calculations since each tissue type has different physiological parameters (thermal conduction, baseline heat production, etc.). A separate line of research that could be pursued would be a model for calculating brain temperature changes from fNIRS data. Both fNIRS and fMRI BOLD response detect changes in local tissue oxygenation, so it should be possible to adapt our model to use fNIRS data. If such a model existed, calculations from it and our model could be compared to refine both models.

Although it is expected that the contribution would be negligible [56], our model does not take into account the effects of perspiration. It would likely not affect the change in temperature greatly because it takes place a couple of centimeters away from brain tissue. Another physiological affect not account for is temperature regulation by the pre-optic nucleus of the anterior hypothalamus [17]. It is responsible for balancing heat production and dissipation [57] and if the model were applied to cases where extreme brain temperatures are created then it would be important to account for how this would react.

How human brain temperature is affected by changes in local brain activity is not well understood because the changes are small and current experimental measurement techniques may require invasive procedures. Models such as the one proposed here allow for brain temperature to be understood through non-invasive measurements such as the fMRI BOLD response.

## References

- [1] Roberto C Sotero and Yasser Iturria-Medina. From Blood Oxygen Level Dependent (BOLD) signals to brain temperature maps. *Bulletin of Mathematical Biology*, 73(11): 2731–2747, 2011.
- [2] Christopher M. Collins, Michael B. Smith, and Robert Turner. Model of local temperature changes in brain upon functional activation. *Journal of Applied Physics*, 97: 2051–2055, 2004.
- [3] Roberto C Sotero and Nelson J Trujillo-Barreto. Modeling the role of excitatory and inhibitory neuronal activity in the generation of the BOLD signal. *NeuroImage*, 35: 149–165, 2007.
- [4] Herman Y Carr and E M Purcell. Effects of diffusion on free precession in nuclear magnetic resonance experiments. *Physical Review*, 94:630–638, 1954.
- [5] Paul Lauterbur. Image formation by induced local interactions: Examples employing nuclear magnetic resonance. *Nature*, 242:190–191, 1973.
- [6] Seiji Ogawa, David W. Tank, Ravi Menon, Jutta M . Ellermann, Seong-Gi Kim, Hellmut Merkle, and Kamil Ugurbil. Intrinsic signal changes accompanying sensory stimulation: Functional brain mapping with magnetic resonance imaging. *Proceedings of the National Academy of Science*, 89:5951–5955, 1992.
- [7] Kenneth K. Kwong, John W. Belliveau, David A. Chesler, Inna E. Goldberg, Robert M. Weisskoff, Brigitte P. Poncelet, David N. Kennedy, Bernice E. Hoppel, Mark S. Cohen, Robert Turner, Hong-Mind Cheng, Thomas J. Brady, and Bruce R. Rosen. Dynamic magnetic resonance imaging of human brain activity during primary sensory stimulation. *Proceedings of the National Academy of Science*, 89:5675–5679, 1992.

- [8] Peter T. Fox and Marcus E. Raichle. Focal physiological uncoupling of cerebral blood flow and oxidative metabolism during somatosensory stimulation in human subjects. *Proceedings of the National Academy of Science*, 83:1140–1144, 1986.
- [9] Jan Karbowski. Thermodynamic constraints on neural dimensions, firing rates, brain temperature and size. *Journal of Computational Neuroscience*, 27:415–436, 2009.
- [10] J. G. McElligott and R Melzack. Localized thermal changes evoked in the brain by visual and auditory stimulation. *Experimental Neurology*, 17:293–312, 1967.
- [11] Eugene A. Kiyatkin, P. Leon Brown, and Roy A. Wise. Brain temperature fluctuation: a reflection of functional neural activity. *European Journal of Neuroscience*, 16:164–168, 2002.
- [12] G. Zeschke and V. G. Krasilnikov. Decreases of local brain temperature due to convection (local brain blood flow) and increases of local brain temperature due to activity. *Acta Biologica et Medica Germanica*, 35:935–941, 1976.
- [13] J. S. George, J. D. Lewine, A. S. Goggin, R. B. Dyer, and E. R. Flynn. IR thermal imaging of a monkey’s head: local temperature changes in response to somatosensory stimulation. *Optical Imaging of Brain Function and Metabolism*, 333:125–136, 1993.
- [14] Shunro Tachibana. Local temperature, blood flow, and electrical activity correlations in the posterior hypothalamus of the cat. *Experimental Neurology*, 16:148–161, 1966.
- [15] Dmitriy A. Yablonskiy, Joseph J. H. Ackerman, and Marcus E. Raichle. Coupling between changes in human brain temperature and oxidative metabolism during prolonged visual stimulation. *Proceedings of the National Academy of Science*, 97(13):7603–7608, 2000.
- [16] Hubert K. F. Trübel, Laura I. Sacolick, and Fahmeed Hyder. Regional temperature

- changes in the brain during somatosensory stimulation. *Journal of Cerebral Blood Flow & Metabolism*, 26:68–78, 2006.
- [17] Gianluca Bertolizio, Linda Mason, and Bruno Bissonnette. Brain temperature: heat production, elimination and clinical relevance. *Pediatric Anesthesia*, 21(4):347–358, 2011.
- [18] Hubert Trübel, Peter Herman, Christoph Kampmann, Ralf Huth, Paul K Maciejewski, Edward Novotny, and Fahmeed Hyder. A novel approach for selective brain cooling: implications for hypercapnia and seizure activity. *Intensive care medicine*, 30(9):1829–33, 2004.
- [19] Jens Soukup, Alois Zauner, Egon M.R. Doppenberg, Matthias Menzel, Charlotte Gilman, Harold F. Young, and Ross Bullock. The importance of brain temperature in patients after severe head injury: Relationship to intracranial pressure, cerebral perfusion pressure, cerebral blood flow, and outcome. *Journal of Neurotrauma*, 19(5):559–571, 2002.
- [20] J Maher and V Hachinski. Hypothermia as a potential treatment for cerebral ischemia. *Cerebrovascular and brain metabolism reviews*, 5(4):277–300, 1993.
- [21] M D Ginsberg, L L Sternau, M Y Globus, W D Dietrich, and R Busto. Therapeutic modulation of brain temperature: relevance to ischemic brain injury. *Cerebrovascular and brain metabolism reviews*, 4(3):189–225, 1992.
- [22] Derk W. Krieger, Michael A. De Georgia, Alex Abou-Chebl, John C. Andrefsky, Cathy A. Sila, Irene L. Katzan, Marc R. Mayberg, and Anthony J. Furlan. Cooling for acute ischemic brain damage (cool aid). *Stroke*, 32(8):1847–1854, 2001.
- [23] Troy E. Dominguez, Gil Wernovsky, and J. William Gaynor. Cause and prevention of central nervous system injury in neonates undergoing cardiac surgery. *Seminars in Thoracic and Cardiovascular Surgery*, 19(3):269 – 277, 2007.

- [24] Eiichi Suehiro, Yuji Ueda, Enoch P. Wei, Hermes A. Kontos, and John T. Povlishock. Posttraumatic hypothermia followed by slow rewarming protects the cerebral microcirculation. *Journal of Neurotrauma*, 20(4):381–390, 2003.
- [25] M A Baker, R A Stocking, and J P Meehan. Thermal relationship between tympanic membrane and hypothalamus in conscious cat and monkey. *Journal of applied physiology*, 32(6):739–742, 1972.
- [26] K. Shiraki, S. Sagawa, F. Tajima, A. Yokota, M. Hashimoto, and G. L. Brengelmann. Independence of brain and tympanic temperatures in an unanesthetized human. *Journal of Applied Physiology*, 65(1):482–486, 1988.
- [27] Mark M Stecker, Albert T Cheung, Alberto Pochettino, Glenn P Kent, Terry Patterson, Stuart J Weiss, and Joseph E Bavaria. Deep hypothermic circulatory arrest: I. effects of cooling on electroencephalogram and evoked potentials. *The Annals of Thoracic Surgery*, 71(1):14 – 21, 2001.
- [28] Christopher S Rumana, Shankar P Gopinath, Masahiko Uzura, Alex B Valadka, and Claudia S Robertson. Brain temperature exceeds systemic temperature in head-injured patients. *Critical Care Medicine*, 26(3), 1998.
- [29] Bruno Bissonnette, Helen M Holtby, Annette J Davis, Hweeleng Pua, Fay J Gilder, and Michael Black. Cerebral Hyperthermia in Children after Cardiopulmonary Bypass. *Anesthesiology*, 93(3), 2000.
- [30] H H Pennes. Analysis of tissue and arterial blood temperatures in the resting human forearm. *Journal of Applied Physiology*, 1(2):93–122, 1948.
- [31] Gregory H. Rothmeier, A. G. Unil Perera, and Mukeshwar Dhamala. Brain tissue temperature dynamics during functional activity. Society for Neuroscience Abstract, Prog. No. 823.09, 2011.

- [32] Gregory H. Rothmeier, A. G. Unil Perera, and Mukeshwar Dhamala. Determining brain tissue temperature from fmri blood-oxygen level dependent response. With Reviewers, 2012.
- [33] Richard B. Buxton, Kâmil Uludağ, David J. Dubowitz, and Thomas T. Liu. Modeling the hemodynamic response to brain activation. *NeuroImage*, 23, Supplement 1(0):S220–S233, 2004.
- [34] Richard B. Buxton, Eric C. Wong, and Lawrence R. Frank. Dynamics of blood flow and oxygen changes during brain activation: The Balloon Model. *Magnetic Resonance in Medicine*, 39:855–864, 1998.
- [35] PT Fox, ME Raichle, MA Mintun, and C Dence. Nonoxidative glucose consumption during focal physiologic neural activity. *Science*, 241(4864):462–464, 1988.
- [36] Christoph Leithner, Georg Rojl, Nikolas Offenhauser, Martina Fuchtemeier, Matthias Kohl-Bareis, Arno Villringer, Ulrich Dirnagl, and Ute Lindauer. Pharmacological uncoupling of activation induced increases in CBF and CMRO<sub>2</sub>. *Journal of Cerebral Blood Flow & Metabolism*, 30(2):311–322, Sep 2009.
- [37] Ai-Ling Lin, Peter T. Fox, Jean Hardies, Timothy Q. Duong, and Jia-Hong Gao. Non-linear coupling between cerebral blood flow, oxygen consumption, and ATP production in human visual cortex. *Proceedings of the National Academy of Sciences*, 107(18):8446–8451, 2010.
- [38] Timothy L. Davis, Kenneth K. Kwong, Robert M. Weisskoff, and Bruce R. Rosen. Calibrated functional MRI: Mapping the dynamics of oxidative metabolism. *Proceedings of the National Academy of Sciences*, 95(4):1834–1839, 1998.
- [39] Mukeshwar Dhamala, Giuseppe Pagnoni, Kurt Wiesenfeld, Caroline Zink, Megan Martin, and Gregory Berns. Neural correlates of the complexity of rhythmic finger tapping. *NeuroImage*, 20:918–926, 2003.



- [40] K. J. Friston, A. Mechelli, R. Turner, and C. J. Price. Nonlinear response in fMRI: The Balloon Model, Volterra Kernels, and other hemodynamics. *NeuroImage*, 12:466–477, 2000.
- [41] M Cope. *The development of a near infrared spectroscopy system and its application for non invasive monitoring of cerebral blood and tissue oxygenation in the newborn infants*. PhD thesis, London University, 1991.
- [42] B L Horecker. The absorption spectra of hemoglobin and its derivatives in the visible and near infra-red regions. *The Journal of Biological Chemistry*, 1942.
- [43] S.C. Bunce, M. Izzetoglu, K. Izzetoglu, B. Onaral, and K. Pourrezaei. Functional near-infrared spectroscopy. *Engineering in Medicine and Biology Magazine, IEEE*, 25(4):54–62, July–August 2006. ISSN 0739-5175. doi: 10.1109/MEMB.2006.1657788.
- [44] Mark Elliott. Functional imaging with diffuse optical tomography. URL <http://www.mmrrcc.upenn.edu/mediawiki/images/3/30/FNIR.ppt>.
- [45] Arno Villringer and Britton Chance. Non-invasive optical spectroscopy and imaging of human brain function. *Trends in Neurosciences*, 20(10):435–442, 1997. ISSN 01662236. doi: 10.1016/S0166-2236(97)01132-6.
- [46] Kurtulus Izzetoglu, Scott Bunce, Banu Onaral, and Kambiz Pourrezaei. Functional optical brain imaging using near-infrared during cognitive tasks. *International Journal of Human-Computer Interaction*, 17(2):211–227, 2004.
- [47] Meltem Izzetoglu, Scott Bunce, and Banu Onaral. Single trial hemodynamic response estimation in event related fnir spectroscopy. In *Biomedical Topical Meeting*, page WF9. Optical Society of America, 2004.
- [48] Hiroki Sato, Masashi Kiguchi, Fumio Kawaguchi, and Atsushi Maki. Practicality of

- wavelength selection to improve signal-to-noise ratio in near-infrared spectroscopy. *NeuroImage*, 21(4):1554 – 1562, 2004.
- [49] Yoko Hoshi. Functional near-infrared optical imaging: utility and limitations in human brain mapping. *Psychophysiology*, 40(4):511–20, July 2003.
- [50] L.M. Head and R.M. Pierson. Functional near infrared spectroscopy for Hb and HbO<sub>2</sub> detection using remote sensing. In *Sensors Applications Symposium (SAS), 2010 IEEE*, pages 166 –169, Feb. 2010. doi: 10.1109/SAS.2010.5439393.
- [51] David A. Boas, Maria Angela Franceschini, Andy K. Dunn, and Gary Strangman. Noninvasive imaging of cerebral activation with diffuse optical tomography. In Ron D. Frostig, editor, *In-vivo optical imaging of brain function*, pages 193–221. 2002.
- [52] Beer. Bestimmung der absorption des rothen lichts in farbigen flssigkeiten (determination of the absorption of red light in colored liquids). In Ludwig Wilhelm Gilbert, editor, *Annalen der Physik und Chemie*, volume 86, pages 78–88. 1852.
- [53] George M. Hale and Marvin R. Querry. Optical constants of water in the 200-nm to 200- $\mu$ m wavelength region. *Appl. Opt.*, 12(3):555–563, Mar 1973. doi: 10.1364/AO.12.000555.
- [54] FLIR Advanced Thermal Solutions: FLIR GF335 Broadband Infrared Cameras for Research & Science, February 2012. URL <http://www.flir.com/thermography/americas/us/view/?id=46792&collectionid=519&col=4586>.
- [55] Mirage Infrared Camera - Infrared Cameras Inc, February 2012. URL <http://www.infraredcamerasinc.com/infrared-camera-Mirage.html>.
- [56] E. R. Nadel, J. W. Mitchell, B. Saltin, and J. A. Stolwijk. Peripheral modifications to the central drive for sweating. *Journal of Applied Physiology*, 31(6):828–833, 1971. URL <http://jap.physiology.org/content/31/6/828.short>.

- [57] H B Simon. Hyperthermia. *The New England journal of medicine*, 329(7):483–487, 1993.

## Appendix A Code

The following sections include the code used. It was written for Matlab R2011b and requires SPM8 to run. Additionally, it is recommended that you have at least 4 GB of RAM in order to work with the large datasets that are produced. For information about how to visualize the data produced, see appendix B. All of the code is available through the temptools github page (<https://github.com/greggroth/temptools>). Additionally, many of the tasks can be completed using the temptools gui (Figs. (A.1) - (A.4)) which can be invoked by running

`temptools`

at the Matlab command prompt (make sure the temptools directory and subdirectories have been added to the Matlab path). The procedure used is explained in section 2.2 and a graphical representation is available in Fig. 2.1.

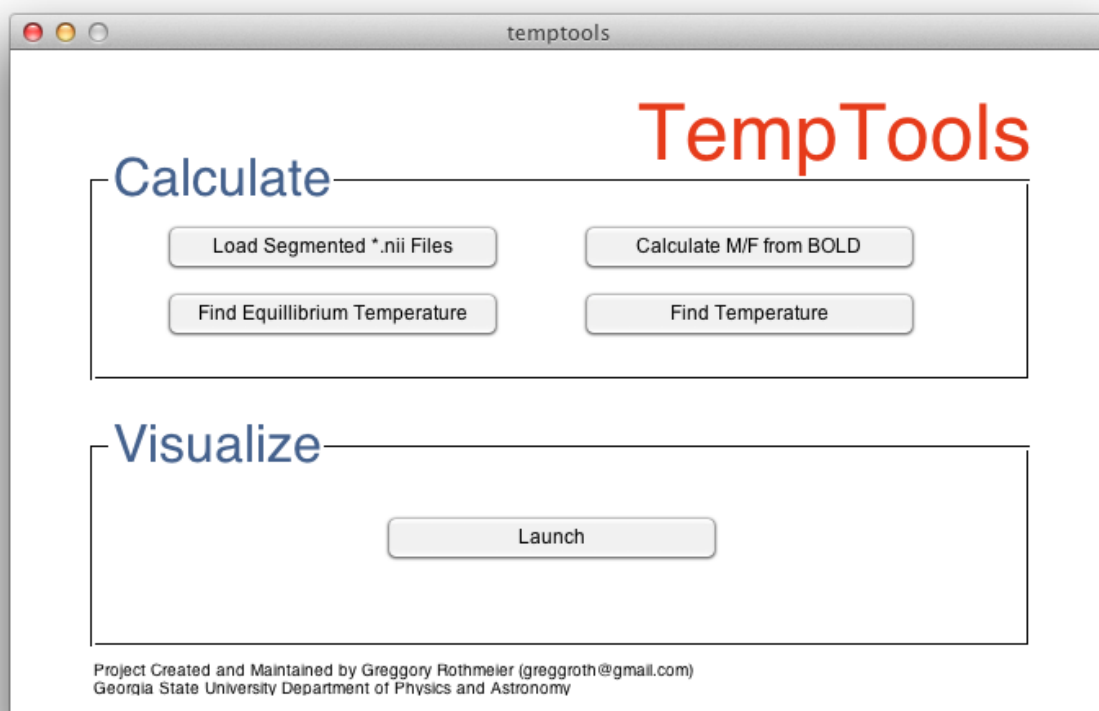


Figure A.1 The main window of temptools. From here, you can go through the calculation steps and launch the visualization tool.

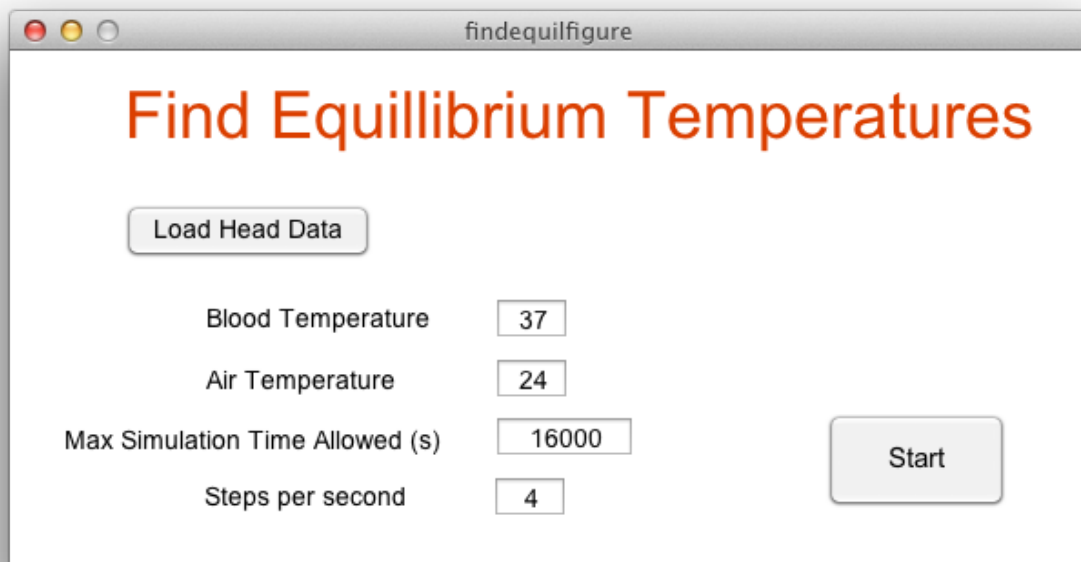


Figure A.2 This is the interface for calculating the equilibrium temperature (method explained in appendix A.3) under certain conditions.

findtempactivation

## Temperature with Activity

Load Head Data

Load Equilibrium Temp Data

Load Region of Interest

Load Metabolism Timecourse

Load Blood Flow Timecourse

Blood Temperature

Air temperature

Max Time

Step Frequency

Save Every  Steps 15 steps saved

Total Steps: 90

Use Generated Activity

Normalized Change in Metabolism

Normalized Change in Blood Flow

Start (Steps)  1.67 s

Stop (Steps)  3.33 s

Activation Duration 1.67 s

Start

Figure A.3 The interface for calculating temperature changes when blood flow and metabolism are time dependent. This can be achieved by either loading metabolism and blood flow datasets or by using generated activity.

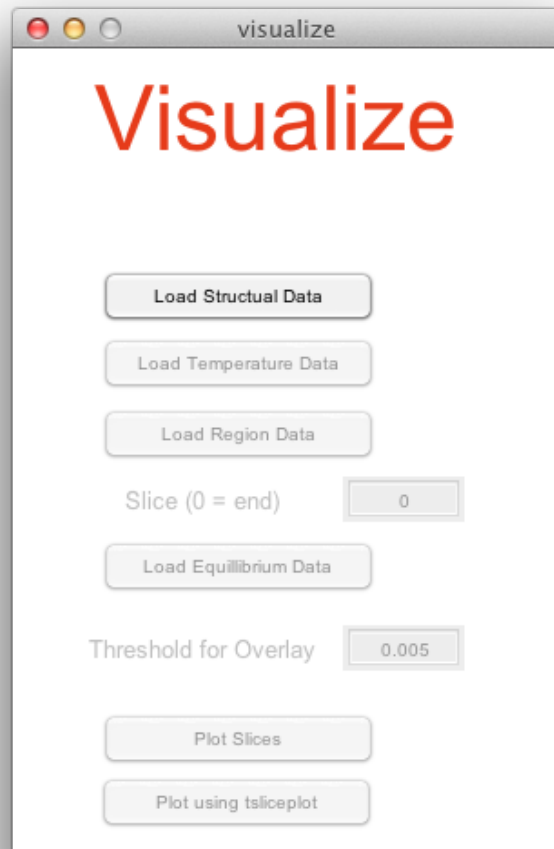


Figure A.4 Visualize your data using the temptools visualization window. This loads all of the required data and launches a slice browser or tsliceplot (see appendix B for more details).



## A.1 Creating the Head Matrix

Before any calculations can be done, a matrix containing tissue-specific parameters must be created. First, a T1 contrast image should be segmented using SPM8 (<http://www.fil.ion.ucl.ac.uk/spm/software/spm8/>). For ease of consistency, the one provided by SPM8 in `./canonical/` is best to use. Using SPM's "New Segmentation" algorithm will segment the image into five different tissue types (gray matter, white matter, cerebral spinal fluid, soft tissue and bone). Once this is complete, run `ImportSegmentedT1()` within this directory and it will return a matrix that has been populated with the tissue-specific parameters required for accurate temperature calculations. The functions `fillAir()` (A.1.2), `fillHoles()` (A.1.3), `build_skin()` (A.1.4) and `repair_headdata()` (A.1.5) are functions required by `BulkImportNII()`. More information about this procedure is in section 2.2.1.

### A.1.1 ImportSegmentedT1()

```
1  function [ total ] = ImportSegmentedT1(varargin)
2  %   ImportSegmentedT1 Import NII files from a directory
3  %   Must be run within the directory containing the files
4  %
5  %   Output: head data as single with variables stored in the 4th
6  %   dimension.
7  %
8  %   Author:   Gregory Rothmeier (greggroth@gmail.com)
9  %   Georgia State University
10 %   Created:  5/31/11
11
12  statusbar = waitbar(0, 'Initializing');
13
14  if size(varargin) == 1
15      oldFolder = cd(varargin{1});
```

```

16  end
17
18
19  % =====
20  % = Tissue Parameters =
21  % =====
22  % Each tissue type is assigned an integer index (i.e. gray matter
    -> 11)
23  % such that tissue-specific parameters can be found by looking at
24  % that element within the corresponding storage matrix
25  % (i.e. QmSTORE(11) -> gray matter Qm)
26
27  % Parameters taken from Colins, 2004
28
29  tisorder = [11 15 5 13 3]; % Using: [GM WM CSF Muscle Bone]
30
31  QmSTORE = [0 0 26.1 11600 0 26.1 697 0 0 302 15575 0 697 1100
    5192];
32  cSTORE = [1006 4600 2110 3640 3800 1300 3720 3000 4200 2300 3680
    3500 3720 3150 3600];
33  rhoSTORE = [1.3 1057 1080 1035.5 1007 1850 1126 1076 1009 916
    1035.5 1151 1041 1100 1027.4];
34  kSTORE = [0.026 0.51 0.65 0.534 0.5 0.65 0.527 0.4 0.594 0.25 0.565
    0.4975 0.4975 .342 .503];
35  wSTORE = [0 1000 3 45.2 0 1.35 40 0 0 2.8 67.1 3.8 3.8 12 23.7];
36
37  % =====
38  % = Import the pre-segmented T1 files =
39  % =====

```

```

40 % The T1 contrast image should be segmented using SPM8.
41 % This loop needs to complete before the next one can begin
42 % Import all of the datat and store as 'cdat1','cdat2', etc.
43 for i = 1:5
44     eval(strcat('dat',num2str(i),' = loadNII(''rc', num2str(i), '
        single_subj_T1.nii'');'))
45     % Preallocate
46     eval(strcat('out', num2str(i),' = zeros(cat(2,size(dat',
        num2str(i),'),7));'))
47 end
48
49 % =====
50 % = Populate the head matrix =
51 % =====
52 % For each data file, it fills in the data from the data storage
53 % arrays for that particular type of tissue. It picks which
54 % ever tissue is the most likely candidate for that voxel based
55 % on the segmented data
56
57 % PROBLEM: It returns 0 (later filled with air) if there is
58 % equal probability of a voxel being two or more different types
59 % of tissue.
60 % SOLVED BY fillHoles()
61
62
63 for i = 1:5
64     % Preallocate
65     holder = zeros(cat(2,size(dat1),7),'single');
66     mask = zeros(size(dat1));

```

```

67     final = zeros(size(holder),'single');
68
69     % Create a mask that indicates whether it is the mostly likely
        tissue type
70     guide = [1 2 3 4 5 1 2 3 4 5]; % This guides it through the
        data correctly
71     eval(strcat('mask = (dat',num2str(i),'>dat',num2str(guide(i+1))
        ,') & (dat',num2str(i),'>dat',num2str(guide(i+2)),') & (dat',
        num2str(i),'>dat',num2str(guide(i+3)),') & (dat',num2str(i),'
        >dat',num2str(guide(i+4)),') & (dat',num2str(i),'~=0);'))
72     % move structure data to new matrix
73     holder(:,:,:,1) = mask;
74     % get indicies of tissues
75     a = find(holder(:,:,:,1) == 1);
76     % gets coordinates from index
77     [x y z t] = ind2sub(size(holder),a);
78
79     % go to each tissue point and store the info
80     for j = 1:length(a)
81         final(x(j),y(j),z(j),:) = [tisorder(i) 0 QmSTORE(tisorder(i)
            )) cSTORE(tisorder(i)) rhoSTORE(tisorder(i)) kSTORE(
            tisorder(i)) wSTORE(tisorder(i))];
82     end
83
84     % Saves the result to a unique output variable (out1, out2,
        etc)
85     eval(strcat('out',num2str(i),'= final;'))
86
87     clearvars a x y z t holder final;

```

```

88     waitbar(i/6,statusbar,sprintf(['File ',num2str(i),' Import
      Compete']));
89 end
90
91 % The filleAir() function checks for any voxels which were not
92 % assigned a tissue type and fills them in with air
93 almostthere = fillAir(out1+out2+out3+out4+out5);
94 % The fillHoles() function corrects for a voxel having two
95 % equally-probable tissue types
96 total = single(buildskin(fillHoles(dat1,dat2,dat3,dat4,dat5,
      almostthere))));
97 waitbar(1,statusbar,'Saving Data')
98
99 cd(oldFolder);
100 close(statusbar);
101
102 end

```

### A.1.2 fillAir()

```

1  function [ output ] = fillAir( tissue )
2  % fillAir() fills gaps in data with air
3  % Once you import all of the data using loadNII(), run it though
4  % this to fill in the remaining spaces with air.
5
6  airdata = [1 0 0 1006 1.3 0.026 0];
7
8  % Picks out air spots
9  a = find(tissue(:,:,,1) == 0);
10 [x y z t] = ind2sub(size(tissue),a);

```

```

11
12 for i = 1:length(a)
13     tissue(x(i),y(i),z(i),:) = airdata;
14 end
15
16 output = tissue;
17 end

```

### A.1.3 fillHoles()

```

1 function [ out_head ] = fillHoles( in1,in2,in3,in4,in5,headin)
2 % fillHoles() checks for misassigned voxels
3 %
4 % Solves an issue where a voxel with two equally probable tissue
5 % types resulted in being assigned as air. This checks for air
6 % voxels that are surrounded by tissue and decides a tissue it
7 % it would be best suited as
8
9 % I only need the tissue indices so this makes things easier down
   the line
10 head = squeeze(headin(:,:,:,:),1));
11
12 %% Data Storage
13 QmSTORE = [0 0 26.1 11600 0 26.1 697 0 0 302 15575 0 697 1100
   5192];
14 cSTORE = [1006 4600 2110 3640 3800 1300 3720 3000 4200 2300 3680
   3500 3720 3150 3600];
15 rhoSTORE = [1.3 1057 1080 1035.5 1007 1850 1126 1076 1009 916
   1035.5 1151 1041 1100 1027.4];

```

```

16 kSTORE = [0.026 0.51 0.65 0.534 0.5 0.65 0.527 0.4 0.594 0.25 0.565
            0.4975 0.4975 .342 .503];
17 wSTORE = [0 1000 3 45.2 0 1.35 40 0 0 2.8 67.1 3.8 3.8 12 23.7];
18
19 %% Get locations of holes
20 % Where two tissue types have the same probability
21
22 idx1 = (in1==in2 | in1 == in3 | in1==in4 | in1==in5) & logical(in1)
        ;
23 idx2 = (in1==in2 | in2 == in3 | in2==in4 | in2==in5) & logical(in2)
        ;
24 idx3 = (in1==in3 | in2 == in3 | in3==in4 | in3==in5) & logical(in3)
        ;
25 idx4 = (in1==in4 | in2 == in4 | in3==in4 | in4==in5) & logical(in4)
        ;
26 idx5 = (in1==in5 | in2 == in5 | in3==in5 | in4==in5) & logical(in5)
        ;
27 % This array will have a zero anywhere there were two or more
28 % common elements between any of the five arrays.
29 idx = idx1|idx2|idx3|idx4|idx5;
30
31 [xmax ymax zmax] = size(in1)
32 [x y z] = ind2sub(size(in1),find(idx)); % get x, y and z
        coordinates of the holes
33
34 for i = 1:length(x) % go to each hole and do work
35     if (x(i)~=1)&&(y(i)~=1)&&(z(i)~=1)&&(x(i)~=xmax)&&(y(i)~=ymax)
        &&(z(i)~=zmax)&&(headin(x(i),y(i),z(i),1)==1) % keeps away
        from the edge and only looks at voxels that were assigned air

```

```

36     [commonesttissue nouse secondbest] = mode([head(x(i)+1,y(i)
        ,z(i)) head(x(i)-1,y(i),z(i)) head(x(i),y(i)+1,z(i)) head
        (x(i),y(i)-1,z(i)) head(x(i),y(i),z(i)+1) head(x(i),y(i),
        z(i)-1)]);
37     % if air and something else are equally common, it'll
        choose air. This forces it to pick the tissue if
        possible.
38     if commonesttissue == 1 && length(secondbest{1})>=2
39         commonesttissue = secondbest{1}(2);
40     end
41     headin(x(i),y(i),z(i),:) = [commonesttissue 0 QmSTORE(
        commonesttissue) cSTORE(commonesttissue) rhoSTORE(
        commonesttissue) kSTORE(commonesttissue) wSTORE(
        commonesttissue)];
42     end
43 end
44
45 out_head = headin;
46
47 end

```

#### A.1.4 build\_skin()

```

1  function [ head_out ] = build_skin( head_in )
2  % build_skin() Creates a layer of skin around the head
3  %
4  % This will check all voxels that were previously labeled
5  % as soft tissue and checks if it has a neighbor which is air.
6  % If so, then it is reassigned as skin.
7

```



```

8  if ndims(head_in)==4
9      head_in = head_in(:,:,:,1);
10 end
11
12 % Git a list of all voxels labeled as muscle
13 muscle_voxels = find(head_in==13);
14
15 % Go through each of them and check for neighboring air voxels
16 for i=1:length(muscle_voxels)
17     [x y z] = ind2sub(size(head_in), muscle_voxels(i));
18     % makes sure we're not at a voxel at the boundry of the dataset
19     if (x~=1) && (x~=size(head_in,1)) && (y~=1) && (y~=size(head_in
        ,2)) && (z~=1) && (z~=size(head_in,3))
20         % Looks for neighboring voxels that are air
21         if ((head_in(x+1,y,z)==1) || (head_in(x-1,y,z)==1) || (head_in
            (x,y+1,z)==1) || (head_in(x,y-1,z)==1) || (head_in(x,y,z+1)
                ==1) || (head_in(x,y,z-1)==1))
22             head_in(x,y,z) = 14;
23         end
24     end
25 end
26
27 head_out = repair_headdata(head_in);
28
29 end

```

### A.1.5 repair\_headdata()

This function will go through the dataset and make sure the tissue-specific parameters are correct for the tissue type assigned for that voxel. fillAir(), fillHoles() and build\_skin() all

correct mislabeled voxels, but they only correct the tissue assignment. After using any of these functions, the data must be passed through `repair_headdata` to update the stored parameters.

```
1  function [ head_out ] = repair_headdata( head_in )
2  % repaid_headdata repopulates the headdata matrix
3  %   If any changes are made to the index column in the headdata
4  % matrix, use this function to repopulate and correct the
5  % parameter values before running any other functions.
6  %   head_in can be either 3 or 4 dimenisions
7
8
9  % =====
10 % = Parameter Storage =
11 % =====
12
13 QmSTORE = [0 0 26.1 11600 0 26.1 697 0 0 302 15575 0 500 1100
14           5192];
15 cSTORE = [1006 4600 2110 3640 3800 1300 3720 3000 4200 2300 3680
16           3500 3010 3150 3600];
17 rhoSTORE = [1.3 1057 1080 1035.5 1007 1850 1126 1076 1009 916
18             1035.5 1151 978.5 1100 1027.4];
19 kSTORE = [0.026 0.51 0.65 0.534 0.5 0.65 0.527 0.4 0.594 0.25 0.565
20           0.4975 0.3738 .342 .503];
21 wSTORE = [0 1000 3 45.2 0 1.35 40 0 0 2.8 67.1 3.8 3.3 12 23.7];
22
23 if ndims(head_in)==4
24     head_in = head_in(:,:,:,1);
25 end
```

```

23 % Reassign the parameter values
24 head_out = cat(4, head_in, zeros(size(head_in)), QmSTORE(head_in),
    cSTORE(head_in), rhoSTORE(head_in), kSTORE(head_in), wSTORE(
    head_in));
25
26 end

```

## A.2 Loading the fMRI Data

The following sections details the processing required to convert the BOLD data (in NIFTI format) to metabolism and blood flow time-courses that can then be used to calculate temperature.

### A.2.1 `sample_bold_import()`

The following code automates the procedure of processing and doing all the calculations on the dataset reported in Dhamala et al. [39]. It's is written for my data on my machine, but it can be used to gain a better understanding of the procedure. For a conceptual explanation, see section 2.2.3.

```
1  %%=====
2  %% How to process preprocessed BOLD data to calculate temperature
3  %%=====
4
5  % This Matlab script was used to automate the the process of using
6  % BOLD data stored in NIFTI (*.nii) format to calculate temperature
7  % changes. The particulars of the code may be specific to this
8  % case, but the procedure should be the same when doing these
9  % calculations on other datasets. All required functions are
10 % included as an attachment to my thesis and are available on my
11 % github (https://github.com/greggroth/tempcalc)
12
13 cd('/Users/Greggory/Documents/Data/fmri_rhythmic_tapping01/NIFTI')
14
15 directories = dir('*01');
16
17 %% Move coregistered files to new Directory
18 for i = 1:length(directories)
```

```

19     dir_name = directories(i).name;
20     main_path = cd( [dir_name filesep dir_name '_NIFTI_1'] );
21     mkdir 'Coregistered'
22     movefile('r*.nii','Coregistered')
23     main_path = cd( [dir_name filesep dir_name '_NIFTI_2'] );
24     mkdir 'Coregistered'
25     movefile('r*.nii','Coregistered')
26     cd(main_path)
27 end
28
29 %% Calculate Rest State
30 disp('Calculating Rest State')
31 for i = 1:length(directories)
32     dir_name = directories(i).name;
33     avg_NII_rest([dir_name filesep dir_name '_NIFTI_1' filesep '
        Coregistered']);
34     avg_NII_rest([dir_name filesep dir_name '_NIFTI_2' filesep '
        Coregistered']);
35 end
36
37
38 %% Normalize to Rest and Mask
39 disp('Normalize to Rest and Mask')
40 for i = 1:length(directories)
41     dir_name = directories(i).name;
42     avg_NII_normalize([dir_name filesep dir_name '_NIFTI_1' filesep
        'Coregistered'], fullfile(dir_name, [dir_name '_NIFTI_1'], '
        Coregistered', 'RestState', 'RestStateAvg.nii'), '
        fullBrainMask.nii');

```

```

43     avg_NII_normalize([dir_name filesep dir_name '_NIFTI_2' filesep
        'Coregistered'], fullfile(dir_name, [dir_name '_NIFTI_2'], '
        Coregistered', 'RestState', 'RestStateAvg.nii'), '
        fullBrainMask.nii');
44 end
45
46
47 %% Calculate metabolism and blood flow change
48 disp('Calculate metabolism and blood flow change')
49 for i = 1:length(directories)
50     dir_1 = [ directories(i).name filesep directories(i).name '
        _NIFTI_1' filesep 'Coregistered' filesep 'Normalized_to_rest'
        ];
51     dir_2 = [ directories(i).name filesep directories(i).name '
        _NIFTI_2' filesep 'Coregistered' filesep 'Normalized_to_rest'
        ];
52     BOLDtoMF(dir_1);
53     BOLDtoMF(dir_2);
54 end
55
56
57 %% Calculate the change in temperature based on metabolism and
58 % blood flow
59
60 % load('equil.mat'); % equilibriumT
61 % load('tt_headdata.mat'); % headdata
62 mask = loadNII('fullBrainMask.nii');
63
64 for i = 1:length(directories)

```

```

65     disp([int2str(i) ' -1 started'])
66     tic
67     % Part I
68     actResult.dat = tempCalcDynMF(headdata, 37, 24, 720, 360,
        equilibriumT, ...
69         fullfile(directories(i).name,[directories(i).name '_NIFTI_1
            '], 'Coregistered', 'Normalized_to_rest', 'Output_18-Sep
            -2011', 'm.mat'), ...
70         fullfile(directories(i).name,[directories(i).name '_NIFTI_1
            '], 'Coregistered', 'Normalized_to_rest', 'Output_18-Sep
            -2011', 'f.mat'), ...
71         4, mask);
72     % Store the parameters used for the calculations for reference
        in the future
73     [c lmax] = max(actResult.dat(:));
74     [likelymax x y z] = ind2sub(size(actResult.dat),lmax);
75     actResult.likelymaxslice = round(likelymax/2);
76     actResult.bloodT = 37;
77     actResult.airT = 24;
78     actResult.tmax = 360;
79     actResult.stepf = 2;
80     actResult.savestepf = 4;
81     actResult.metabandflowdata = 'From Dataset';
82     save(fullfile(directories(i).name,[directories(i).name '
        _NIFTI_1'], 'Coregistered', 'Normalized_to_rest', 'Output_18-
        Sep-2011', 'tt_act_res.mat'), 'actResult');
83     old = cd([directories(i).name,filesep,[directories(i).name '
        _NIFTI_1'],filesep,'Coregistered', filesep,'
        Normalized_to_rest', filesep,'Output_18-Sep-2011']));

```

```

84     writeT_to_nii(actResult, equilibriumT, exp_nii);
85     cd(old)
86     clear actResult
87     % Part II
88     disp([int2str(i) '-2 started'])
89     actResult.dat = tempCalcDynMF(headdata, 37, 24, 720, 360,
        equilibriumT, ...
90         fullfile(directories(i).name,[directories(i).name '_NIFTI_2
            '], 'Coregistered', 'Normalized_to_rest', 'Output_18-Sep
            -2011', 'm.mat'), ...
91         fullfile(directories(i).name,[directories(i).name '_NIFTI_2
            '], 'Coregistered', 'Normalized_to_rest', 'Output_18-Sep
            -2011', 'f.mat'), ...
92         4, mask);
93     [c lmax] = max(actResult.dat(:));
94     [likelymax x y z] = ind2sub(size(actResult.dat),lmax);
95     actResult.likelymaxslice = round(likelymax/2);
96     actResult.bloodT = 37;
97     actResult.airT = 24;
98     actResult.tmax = 360;
99     actResult.stepf = 2;
100    actResult.savestepf = 4;
101    actResult.metabandflowdata = 'From Dataset';
102    save(fullfile(directories(i).name,[directories(i).name '
        '_NIFTI_2'], 'Coregistered', 'Normalized_to_rest', 'Output_18-
        Sep-2011', 'tt_act_res.mat'), 'actResult');
103    old = cd([directories(i).name,filesep,[directories(i).name '
        '_NIFTI_2'],filesep, 'Coregistered', filesep, '
        Normalized_to_rest', filesep, 'Output_18-Sep-2011']));

```



```

104     writeT_to_nii(actResult, equilibriumT, exp_nii);
105     cd(old)
106     clear actResult
107     disp([int2str(i) ' finished in ' num2str(toc)])
108 end

```

### A.2.2 avg\_NII\_rest()

```

1  function [ ] = avg_NII_rest( varargin )
2  % Collects datasets which are part of the
3  % resting state and averages them together to
4  % give a resting-state image
5  %
6  % THIS MUST BE EDITED TO WORK
7  % This is written for my data and you should read
8  % and understand what it is doing before you use it.
9  % It will almost certainly require some editing
10 % to select the right range of data.
11
12 %% Setup
13 switch length(varargin)
14     case 0
15         fold_name = uigetdir;
16         if ~fold_name % Cancel Button
17             return
18         end
19     case 1
20         fold_name = varargin{1};
21     otherwise
22 end

```

```

23
24 % Go to the folder containing the files
25 oldfold = cd(fold_name);
26 file_list = dir('*.nii');
27
28 % Select resting state images
29 % (first and last 10 steps in my case).
30 % EDIT THIS TO FIT YOUR CASE
31 file_list = file_list([1:10 170:180]);
32 file_count = length(file_list);
33
34 % Cell array to store all of the datasets in.
35 datHolder = cell(file_count,1);
36
37 statusbar = waitbar(0,'Initializing');
38
39 for j=1:file_count
40     try
41         waitbar(j/file_count,statusbar,sprintf('%d%%',round((j/
42             file_count)*100)));
43     catch err
44         return
45     end
46     fi = load_nii(file_list(j).name);
47     datHolder{j} = fi.img;
48
49 %% Calculate the mean
50 ymax = size(datHolder{1},2);

```

```

51  zmax = size(datHolder{1},3);
52  output = zeros(size(datHolder{1}));
53
54  for i=1:ymax
55      try
56          waitbar(i/ymax,statusbar,sprintf('%d%%',round((i/ymax)*100)
57              ));
58      catch err
59          return
60      end
61      for k=1:zmax
62          excStr = cell(length(datHolder),1);
63          for l=1:length(datHolder)
64              excStr{l} = [',datHolder{' int2str(l) '}'(:,' int2str(i)
65                  ',' int2str(k) ')'''];
66          end
67          comb = eval(['cat(1' cell2mat(excStr') ')']);
68          output(:,i,k) = mean(comb);
69      end
70  end
71
72  close(statusbar)
73
74  fi.img = output;
75  mkdir('RestState')
76  save_nii(fi,fullfile('RestState','RestStateAvg.nii'));
77
78  cd(olderd)
79  end

```

### A.2.3 avg\_NII\_normalize()

```
1  function [ ] = avg_NII_normalize( varargin )
2  % Uses the resting-state image calculated using
3  % avg_NII_rest() to normalize the rest of the data
4
5  % If no inputs are given, the "open file..." UI will
6  % prompt for the required information.
7
8  %% Setup
9  switch length(varargin)
10     case 0
11         fold_name = uigetdir('Directory Containing Data');
12         if ~fold_name % Cancel Button
13             return
14         end
15
16         [rest_file rest_path rest_index]= uigetfile('*.nii','
17             Resting State NIFTI File');
18         switch rest_index
19             case 0
20                 return
21             case 1
22                 rest_dat = load_nii(fullfile(rest_path,rest_file));
23                 rest_dat = double(rest_dat.img);
24             otherwise
25                 error('An error has occurred loading the resting
26                     state data')
27         end
28     end
```

```

26
27     [mask_file mask_path mask_index] = uigetfile('*.nii','Mask'
28         );
29     switch mask_index
30         case 0
31             return
32         case 1
33             mask_dat = load_nii(fullfile(mask_path, mask_file))
34             ;
35             mask_dat = logical(mask_dat.img);
36             if max(size(mask_dat) ~= size(rest_dat))
37                 error('The Mask and Resting State files must
38                     have the same size')
39             end
40         otherwise
41             error('An error has occurred loading the resting
42                 state data')
43     end
44 case 1
45     fold_name = varargin{1};
46     [rest_file rest_path rest_index]= uigetfile('*.nii','
47         Resting State NIFTI File');
48     switch rest_index
49         case 0
50             return
51         case 1
52             rest_dat = load_nii(fullfile(rest_path,rest_file));
53             rest_dat = double(rest_dat.img);
54         otherwise

```

```

50         error('An error has ocured loading the resting
51             state data')
52     end
53 case 2
54     fold_name = varargin{1};
55     rest_dat = loadNII(varargin{2});
56     [mask_file mask_path mask_index] = uigetfile('*.nii','Mask'
57         );
58     switch mask_index
59         case 0
60             return
61         case 1
62             mask_dat = load_nii(fullfile(mask_path, mask_file))
63             ;
64             mask_dat = logical(mask_dat.img);
65             if max(size(mask_dat) ~= size(rest_dat))
66                 error('The Mask and Resting State files must
67                     have the same size')
68             end
69         otherwise
70             error('An error has ocured loading the resting
71                 state data')
72     end
73 case 3
74     fold_name = varargin{1};
75     rest_dat = loadNII(varargin{2});
76     mask_dat = loadNII(varargin{3});
77 otherwise
78     return

```

```

74 end
75
76 % Go to the folder containing the files
77 oldfold = cd(fold_name);
78 file_list = dir('*.nii');
79 file_count = length(file_list);
80
81 % Make a directoy to save the normalized data to
82 saveDir = 'Normalized_to_rest';
83 if ~.isdir(saveDir)
84     mkdir(saveDir);
85 end
86
87 statusbar = waitbar(0,'Initializing');
88
89 % for each file: load it, devide by the rest state and save it
90 for i=1:file_count
91     try
92         waitbar(i/file_count,statusbar,[fold_name sprintf('%d%',
93             round((i/file_count)*100))]);
94     catch err
95         return
96     end
97     [file_path file_name file_ext] = fileparts(file_list(i).name);
98     file_hold = load_nii(file_list(i).name);
99     file_hold.img = double(file_hold.img)./rest_dat - 1;
100    file_hold.img(~mask_dat) = 0; % set everything
    outside the mask to 0
100    file_hold.img(isnan(file_hold.img)) = 0; % set all NaN's to 0

```

```

101     file_hold.img(isinf(file_hold.img)) = 0; % set all inf's to 0
102     file_hold.img(file_hold.img == -1) = 0; % correct these for
        voxels that are giving me problems
103     file_hold.hdr.dime.datatype = 16; % set the datatype to single
104     file_hold.hdr.dime.bitpix = 16;
105     save_nii(file_hold,fullfile(saveDir,[file_name '_rn' file_ext])
        )
106 end
107
108 close(statusbar)
109 cd(oldfold)
110
111 end

```

#### A.2.4 BOLDtoMF()

```

1  function [ ] = BOLDtoMF( varargin)
2  %BOLDtoMF Calculate metabolism and blood flow from BOLD response
3  %
4  %   Input: Directory containing a series of *.nii files of the BOLD
5  %   response.
6  %
7  %   Output: Two new files will be created in a new subdirectory
8  %   with a variable for each time step.
9  %
10 %   Usage:
11 %       BOLDtoMF
12 %       BOLDtoMF(directory)
13 %
14 %   If a directory is not provided, one will be requested.

```



```

15 %
16 %   Method from Sotero, et. al. 2011
17
18 % =====
19 % = Setup =
20 % =====
21 % if a folder isn't an argument, it'll prompt for one
22 switch length(varargin)
23     case 0
24         fold_name = uigetdir;
25         if ~fold_name % Cancel Button pressed
26             return
27         end
28     case 1
29         fold_name = varargin{1};
30     otherwise
31         error('Input is not understood')
32 end
33
34 % Go to the folder containing the files
35 oldfold = cd(fold_name);
36 file_list = dir('*.nii');
37 file_count = length(file_list);
38
39 % Set up a directory for the outputs
40 newFolder = ['Output_',datestr(clock,1)];
41 mkdir(newFolder)
42
43 % Make *.mat files to append the data to

```

```

44 m0001 = 0; f0001 = 0;
45 save(['./' newFolder '/m.mat'], 'm0001');
46 save(['./' newFolder '/f.mat'], 'f0001');
47
48 s = loadNII(file_list(1).name);
49 norm = ones(size(s));
50
51 % =====
52 % = Do Work =
53 % =====
54 % This will calculate the metabolism and blood flow. The output is
55 % appended to 'm.mat' and 'f.mat' within a new folder created
56 % within the directory containing the data.
57
58 statusbar = waitbar(0, 'Initializing');
59
60 maxBOLD = 0.22;
61
62 % Required Parameters
63 % [alpha beta a      b      c      A      ]
64 p = [0.4 1.5 0.1870 0.1572 -0.6041 maxBOLD];
65
66 % Calc flow and metabolism for when BOLD = 1
67 s = 0;
68 y = -((p(4)*p(2))/(p(1)+p(2)*p(5)))*((p(6)-s)/(p(6)*p(3)^p(2)))
    ^ (1/(p(1)+p(2)*p(5)));
69 fNOACT = -((p(1)+p(2)*p(5))/(p(4)*p(2)))*lambertw(y);
70 mNOACT = p(3)*fNOACT^(p(5)+1)*exp(-p(4)*fNOACT);
71

```

```

72
73 %% Calc flow and metabolism
74 disp(fold_name)
75 for j=1:file_count
76     try
77         waitbar(j/file_count, statusbar, sprintf('%d%%', round((j/
            file_count)*100)));
78     catch err
79         return
80     end
81     s = loadNII(file_list(j).name); % Load up the file
82     s(isnan(s)) = 1;
83     s(isinf(s)) = 1;
84     y = -((p(4)*p(2))/(p(1)+p(2)*p(5))) .* ((p(6)-s)./(p(6)*p(3)^p(2)
        )) .^(1/(p(1)+p(2)*p(5)));
85     if (size(y,1)==91)&&(size(y,2)==109)&&(size(y,3)==91)
86         f = -((p(1)+p(2)*p(5))/(p(4)*p(2))) .* lambw_mex(real(y));
87     else
88         f = -((p(1)+p(2)*p(5))/(p(4)*p(2))) .* lambw(y);
89     end
90     m = p(3)*f.^(p(5)+1).*exp(-p(4)*f);
91     % Clean up NaNs that may have popped up
92     m(isnan(m))=1;
93     f(isnan(f))=1;
94     % Normalize to resting m and f
95     m = m./mNOACT;
96     f = f./fNOACT;
97
98     % Rename and save the data

```

```

99     eval(['m' sprintf('%04d',j) ' = m;']);
100    eval(['f' sprintf('%04d',j) ' = f;']);
101    eval(['save(''./' newFolder '/m.mat'', 'm' sprintf('%04d',j) '
        ''-append'');']);
102    eval(['save(''./' newFolder '/f.mat'', 'f' sprintf('%04d',j) '
        ''-append'');']);
103    clear m0* f0*
104 end
105
106 close(statusbar)
107 cd(oldfold)
108 end

```

### A.2.5 lambw() and lambw\_mex()

The `lambw()` function is a wrapper for the `wapr()` function available on Matlab FileExchange (<http://www.mathworks.com/matlabcentral/fileexchange/3644-real-values-of-the-lambert-w-function/content/Lambert/wapr.m>). A compiled version of this function (`lambw_mex()`) runs much faster and is recommended. This function is used over Matlab's built-in Lambert-W function for the sake of performance.

```

1 function [ array_out ] = lambw( array_in )
2 % lambw Wrapper for wapr()
3 % Available:  http://www.mathworks.com/matlabcentral/fileexchange
    /3644-real-values-of-the-lambert-w-function/content/Lambert/wapr.
    m
4 % Dwapr() doesn't work any arrays over Nx1, so this steps through
5 % the full matrix and gives the rows to wapr. Works pretty fast.
6 %#codegen
7
8 if ndims(array_in) ~= 3

```

```

9      error('This only works (for now) with a three dimensional array
      .')
10 end
11
12 xmax = size(array_in,1);
13 ymax = size(array_in,2);
14
15 array_out = zeros(size(array_in));
16 for ix=1:xmax
17     for iy=1:ymax
18         array_out(ix,iy,:) = wapr(array_in(ix,iy,:));
19     end
20 end
21 end

```

### A.3 Calculating the Equilibrium Temperature

In order to determine the temperature fluctuations due to changes in activity, the baseline temperature must first be established for each voxel. The function `tempCalcEquilibrium()` will update the temperature using the Penne's bioheat equation (Eq. (2.4)) until the change in temperature for each voxel falls below a certain threshold. Details about this procedure are available in section 2.2.2.

#### A.3.1 `tempCalcEquilibrium()`

```
1  function temperature_Out = tempCalcEquilibrium(tissue,bloodT,airT,
      nt,tmax,pastCalc,printprogress)
2  % tempCalcEquilibrium Find the equilibrium values
3  %   tissue: holds all of the structural information
4  %   bloodT: Temperature of the blood
5  %   airT:   Temperature of the surrounding air
6  %   nt:     Max number of time steps
7  %   tmax:   Total amount of time the simulation should run over
8  %
9  %   This is based off of tempCalc() but loops until the rate of
10 % change of a each voxel is sufficiently small then outputs
11 % what's calculated. If it takes too long to do all at once,
12 % split it up into smaller time chunks and use the last step
13 % from the previous dataset as pastCalc in order to resume.
14 %
15 %   Note: This does not save the time course because it can take
16 % a lot of steps to find the equilibrium. It outputs the last
17 % time step.
18 %
19 %   Written by Gregory Rothmeier (greggroth@gmail.com)
```

```

20 %   Georgia State University Dept. Physics and Astronomy
21 %   May, 2011
22 tic
23 %%   Default Values
24 if nargin<2, bloodT = 37;           end
25 if nargin<3, airT = 24;           end
26 if nargin<4, nt = 100;           end
27 if nargin<5, tmax = 50;           end
28 if nargin<6, pastCalc = 0;        end
29 if nargin<7, printprogress = 1; end
30
31 % These rescue the data if the calculation is interrupted.
32 global temperature
33 global dirty
34
35 c = onCleanup(@InterCatch);
36 dirty = 1;
37
38 dx = 2*10^-3;           % Voxel size (m)
39
40 if nt<(2*tmax),
41     warning('Time step size is not large enough. Results will be
42             unreliable. Consider increasing the number of steps or
43             reducing tmax.')
44 end
45
46 % Constants used that aren't already stored in tissue
47 [xmax ymax zmax t] = size(tissue);

```

```

47  clear t;
48  dt = tmax/(nt-1);
49  % rhoBlood = 1057;
50  % wBlood = 1000;
51  % cBlood = 3600;
52
53  % =====
54  % = Setup =
55  % =====
56  %   Starts all tissue voxels at bloodT (default 37) and maintains
57  %   air at airT (default 24)
58  %   The condition squeeze(tissue(:,:,:),~airIndex picks out the
59  %   elements that are tissue
60
61  temperature = ones(3,xmax,ymax,zmax,'single')*airT;
62  if pastCalc == 0
63      temperature(1,squeeze(tissue(:,:,:,1))~=1) = bloodT;
64  else
65      temperature(1,:,:,:) = pastCalc;
66  end
67  numElements = numel(temperature(1,:,:,:));
68
69  % =====
70  % = Do Work =
71  % =====
72  %   This is a vectorized version of the next section. For the love
73  %   of god don't make any changes to this without first looking below
74  %   to make sure you know what you're changing. This is [nearly]
75  %   impossible to understand, so take your time and don't break it.

```



```

76 % data is stored in 'tissue' as such :
77 % [tissuetype 0 Qm c rho k w]; <-- second element is blank for
    all.
78 % [ 1 2 3 4 5 6 7
79
80 averagedk = (circshift(tissue(:,:,:,6),[1 0 0])+circshift(tissue
    (:,:,:,6),[-1 0 0])+circshift(tissue(:,:,:,6),[0 1 0])+circshift(
    tissue(:,:,:,6),[0 -1 0])+circshift(tissue(:,:,:,6),[0 0 1])+
    circshift(tissue(:,:,:,6),[0 0 -1])+tissue(:,:,:,6))/7;
81 rhoblood = 1057;
82 cblood = 3600;
83
84 %% Specify Percision Goal
85 tolerance = 1; % fraction of voxels have a slope less than '
    zeropoint'
86 zeropoint = 2.5e-7; % point at which the slope between two *steps*
    is considered essentially zero
87
88
89 goal = numElements - tolerance*numElements;
90 goon = numElements; % Forces the while loop to run the first time
91 format shortG;
92 % temperature(1,:,:,) = Current Temperature
93 % temperature(2,:,:,) = Next Temperature
94 % Resets after each update
95 if printprogress
96     disp(['Goal: ', num2str(goal), ' remaining voxels'])
97 end
98 t2 = 1;

```

```

99 while goon(1)>goal && t2<=nt % runs until either 'goal' elements
    have a slope greater than 'zeropoint' or it exceeds nt
100 if printprogress
101     disp([t2 goon(1) ((numElements-goan(1))/numElements)*100]) %
        progress
102 end
103 temperature(2,:,:,:) = squeeze(temperature(1,:,:,:)) + ...
104     dt/(tissue(:,:,:5).*tissue(:,:,:4)).* ...
105     ((averagedk/dx^2).*.
106     (circshift(squeeze(temperature(1,:,:,:)),[1 0 0])-2*squeeze
        (temperature(1,:,:,:))+circshift(squeeze(temperature
        (1,:,:,:)),[-1 0 0])+... % shift along x
107     circshift(squeeze(temperature(1,:,:,:)),[0 1 0])-2*squeeze
        (temperature(1,:,:,:))+circshift(squeeze(temperature
        (1,:,:,:)),[0 -1 0])+... % shift along y
108     circshift(squeeze(temperature(1,:,:,:)),[0 0 1])-2*squeeze
        (temperature(1,:,:,:))+circshift(squeeze(temperature
        (1,:,:,:)),[0 0 -1]))... % shift along z
109     -(1/6000)*rhoblood*tissue(:,:,:7)*cblood.*(squeeze(
        temperature(1,:,:,:))-bloodT)+tissue(:,:,:3));
110 % resets the air temperature back since it's also modified
111 % above, but it needs to be kept constant throughout the
112 % calculations
113 temperature(2,squeeze(tissue(:,:,:1))==1) = airT;
114 % checks how quickly the temperature is changing and if it is
115 % close enough to zero to be considered stopped ('zeropoint')
116 goon = size(temperature(abs(squeeze(temperature(2,:,:,:)-
        temperature(1,:,:,:))>zeropoint)));
117 temperature(1,:,:,:) = temperature(2,:,:,:);

```

```

118         t2 = t2 + 1;
119     end
120
121     temperature_Out = temperature(2,:,:,:);
122     dirty = 0;
123
124     % equilTemperature = temperature_Out;
125     % save('equil.mat','equilTemperature');
126
127     %% To Combine Datasets
128     % use this technique if there are seperate datasets that need
129     % combining
130     %     vertcat(squeeze(res1(:,:,:,:)),squeeze(res2(2:end,:,:,:)))
131     % Where for all by the first dataset, you need to do the time from
132     % 2:end so that there are no repeats (remember that the last
133     % timestep from the previous dataset serves as the first for the
134     % new one)
135
136
137     time = toc;
138     end
139
140     % Recovers the data if calculation was interrupted
141     function InterCatch
142     global dirty
143     if dirty
144         disp('Interupt Intercepted.  Inprepretating Interworkspace Data
145             Interfaces.')
```

```
146     equilibriumT = temperature;
147     save('equiltempAbortDump.mat','equilibriumT');
148     % setappdata(0,'InterpOut',temperature);
149 end
150 end
```

## A.4 Calculating the Temperature Change

The following function takes as inputs the head data matrix (appendix A.1), the metabolism and blood flow time courses (appendix A.2) and the equilibrium temperatures (appendix A.3) and calculates the temperature time-course. More details about this algorithm can be found in section 2.2.4.

### A.4.1 tempCalcDynMF

```
1  function temperatureOut = tempCalcDynMF(tissue,bloodT,airT,nt,tmax,
    pastCalc,metab,flow,savesteps,region)
2  % tempCalcDynMF  How does changing metabolism and blood flow
3  % affect things?
4  %
5  %  tissue: holds all of the structural information
6  %  bloodT: Temperature of the blood
7  %  airT:   Temperature of the surrounding air
8  %  nt:     Number of time steps
9  %  tmax:   Amount of model time the simulation should span
10 %
11 %  region: logical matrix same size as head that is used
12 %           as a mask
13 %
14 %  Written by Gregory Rothmeier (greggroth@gmail.com)
15 %  Georgia State University Dept. Physics and Astronomy
16 %  May, 2011
17
18  statusbar = waitbar(0,'Initializing');
19
20 %%  Default Values
```

```

21  if nargin<2,  bloodT = 37;          end
22  if nargin<3,  airT = 24;           end
23  if nargin<4,  nt = 3;              end
24  if nargin<5,  tmax = 1;           end
25  if nargin<6,  pastCalc = 0;       end
26
27
28  % Length of one side of a voxel (m)
29  dx = 2*10^-3;
30
31  if nt<(2*tmax),
32      warning('Time step size is not large enough. Results will be
              unreliable. Consider increasing the number of steps or
              reducing tmax.')
33  end
34
35  [xmax ymax zmax t] = size(tissue);
36  clear t;
37  dt = ones([xmax ymax zmax])*(tmax/(nt-1));
38
39  %% Determine Metabolism/Blood Flow Data Storage System
40  if ischar(metab)&&ischar(flow)
41      % if file locations are given rather than data
42      option = 1;
43  else
44      % Preallocate matrices for holding metabolism and blood flow data
45      metabMulti = ones([xmax ymax zmax],'single');
46      flowMulti = ones([xmax ymax zmax],'single');
47      option = 0;

```

```

48 end
49
50 %% Maps
51 % Creates a map that identifies where there is tissue
52 % the condition squeeze(tissue(:,:,:),~airIndex picks out the
53 % elements that are tissue
54
55 tmax = ceil((nt-1)/savesteps);
56 temperatureOut = ones(tmax,xmax,ymax,zmax,'single');
57 temperature = ones(2,xmax,ymax,zmax,'single')*airT;
58 if pastCalc == 0
59     temperature(1,squeeze(tissue(:,:,:,1))~=1) = bloodT;
60 else
61     % Starts everything off at the pre-determined equilibrium
        temperatures
62     temperature(1,:,:,:) = pastCalc(end,:,:,:);
63 end
64 temperatureOut(1,:,:,:) = temperature(1,:,:,:);
65
66
67 % =====
68 % = Do Work =
69 % =====
70 % This is a vectorized version of the next section. For the love
71 % of god don't make any changes to this without first looking below
72 % to make sure you know what you're changing. This is [nearly]
73 % impossible to understand because it's been vectorized, so take
74 % your time and don't break it. Data is stored in 'tissue' as such
    :

```

```

75 % [tissuetype 0 Qm c rho k w] <-- second element is blank for all
    .
76 % [      1      2 3 4 5 6 7]
77
78 averagedk = (circshift(tissue(:,:,,6),[1 0 0])+circshift(tissue
    (:,:,6),[-1 0 0])+circshift(tissue(:,:,,6),[0 1 0])+circshift(
    tissue(:,:,,6),[0 -1 0])+circshift(tissue(:,:,,6),[0 0 1])+
    circshift(tissue(:,:,,6),[0 0 -1])+tissue(:,:,,6))/7;
79 rhoblood = 1057;
80 cblood = 3600;
81
82 %% Only saves every 4 steps to reduce the final matrix size
83 for t2 = 1:nt-1
84     waitbar(t2/(nt-1),statusbar,sprintf('%d%%',round(t2/(nt-1)*100))
        );
85
86 % if a variable needs to be used multiple times for the same time
    step.
87     t3 = floor((t2-1)/4)+1; % 1 1 1 1 2 2 2 2 3 3 . . .
88
89     % if a file is specified, pulls the data from the file for each
        step
90     if option
91         eval(strcat('load(fullfile(metab),''-mat'', ''m'',sprintf('%04
            d',t3),'');'));
92         eval(strcat('load(fullfile(flow),''-mat'', ''f'',sprintf('%04d
            ',t3),'');'));
93         eval(strcat('metabMulti = m',sprintf('%04d',t3),''));
94         eval(strcat('flowMulti = f',sprintf('%04d',t3),''));

```



```

95         eval(strcat('clear f', sprintf('%04d',t3),' m',sprintf('%04d',t3)))
96     else
97         metabMulti(region) = metab(t2);
98         flowMulti(region) = flow(t2);
99     end
100
101     temperature(2,:,:,:) = squeeze(temperature(1,:,:,:)) + ...
102         dt./(tissue(:,:,:5).*tissue(:,:,:4)).* ...
103         ((averagedk/dx^2).*...
104         (circshift(squeeze(temperature(1,:,:,:)),[1 0 0])-2*squeeze
105             (temperature(1,:,:,:))+circshift(squeeze(temperature
106             (1,:,:,:)),[-1 0 0])+... % shift along x
107         circshift(squeeze(temperature(1,:,:,:)),[0 1 0])-2*squeeze
108             (temperature(1,:,:,:))+circshift(squeeze(temperature
109             (1,:,:,:)),[0 -1 0])+... % shift along y
110         circshift(squeeze(temperature(1,:,:,:)),[0 0 1])-2*squeeze
111             (temperature(1,:,:,:))+circshift(squeeze(temperature
112             (1,:,:,:)),[0 0 -1]))... % shift along z
113         -(1/6000)*rhoblood*flowMulti.*tissue(:,:,:7)*cblood.*(
114             squeeze(temperature(1,:,:,:))-bloodT)+metabMulti.*
115             tissue(:,:,:3));
116
117     % resets the air temperature back since it's also modified
118     % above,
119     % but it needs to be kept constant throughout the calculations
120     temperature(2,squeeze(tissue(:,:,:1))==1) = airT;
121     temperatureOut(ceil(t2/savesteps),,:,:,:) = temperature(2,:,:,:)
122     ;
123     temperature(1,:,:,:) = temperature(2,:,:,:);

```

```

113         clear metabMulti flowMulti
114     end
115     close(statusbar);
116
117     % =====
118     % = Old Code =
119     % =====
120     % This is what used to be used. It's much slower (~60 times
121     % slower), but it's much easier to understand compared to the
122     % above code. If any changes need to be made above, first look
123     % through this code to ensure you understand it before making
124     % changes. It's really easy to mess up the code above and nearly
125     % impossible to figure out where.
126     %
127     % good luck.
128
129     % for t2 = 1:nt-1
130     %     for x2 = 2:xmax-1
131     %         for y2 = 2:ymax-1
132     %             for z2 = 2:zmax-1
133     %                 if tissue(x2,y2,z2,1) ~= 1,
134     %                     temperature(t2+1,x2,y2,z2) = temperature(t2,
135     %                         x2,y2,z2) + (dt/(tissue(x2,y2,z2,5)*tissue(x2,y2,z2,4)))*((tissue
136     %                         (x2,y2,z2,6)/dx^2)*...
137     %                         (temperature(t2,x2+1,y2,z2)-2*temperature(
138     %                         t2,x2,y2,z2)+temperature(t2,x2-1,y2,z2)+...
139     %                         temperature(t2,x2,y2+1,z2)-2*temperature(t2
140     %                         ,x2,y2,z2)+temperature(t2,x2,y2-1,z2)+...

```

```

137 %             temperature(t2,x2,y2,z2+1)-2*temperature(t2
      ,x2,y2,z2)+temperature(t2,x2,y2,z2-1))...
138 %             -(1/6000)*rhoBlood*wBlood*cBlood*(
      temperature(t2,x2,y2,z2)-bloodT)+tissue(x2,y2,z2,3));
139 %             end
140 %         end
141 %     end
142 % end
143 % end
144
145 end

```

## Appendix B Visualization Tools

The temperature data is a four dimensional dataset (time, x, y and z), so good visualizations tools are necessary to analyzing the results. The primary tool I use is a modification of SliceBrowser (<http://www.mathworks.com/matlabcentral/fileexchange/20604>) and is provided as part of temptools (<https://github.com/greggroth/temptools/tree/master/lib/SliceBrowser>). In working with this, I also created a function (TempPlot()) to act as a wrapper and handle possible plotting situations depending on the number of inputs.

### B.1.1 TempPlot()

```
1 function [ ] = TempPlot( head, tempdata, highlightRegion, slice,
    equil, threshold, point)
2 %TempPlot Plot data from tempCalc() or BulkImportNII()
3 % INPUT TempPlot(structuredata)
4 % TempPlot(structuredata, temperaturedata)
5 % TempPlot(structuredata, temperaturedata, highlightRegion)
6 % TempPlot(structuredata, temperaturedata, highlightRegion,
    slice)
7 % TempPlot(structuredata, temperaturedata, highlightRegion,
    slice, EquillibriumData)
8 %
9 % This function with determine which type of data it is and then
10 % plot it appropriately.
11 %
12 % equil - Equillibrium state data
13 % threshold - threshold value for being displayed as an overlay
14 % REQUIRES: SliceBrowser (http://www.mathworks.com/matlabcentral
    /fileexchange/20604)
```

```

15 %% Error checking and data restructuring where necessary
16 if ndims(head) == 4
17     head = head(:,:,:,1);
18 elseif ndims(head) ~= 3
19     error('Input ''head'' must have either 3 or 4 dimensions');
20 end
21
22 if nargin > 1
23     if ndims(tempdata) == 3 % should only happen when comparing
24         two equilibrium datasets
25         temp = tempdata;
26         tempdata = zeros([1 size(temp)]);
27         tempdata(1,:,:,:) = temp;
28     elseif ndims(tempdata) ~= 4
29         error('Input ''tempdata'' must have either 3 or 4 dimensions'
30             ');
31     end
32     tempdataShort = squeeze(tempdata(end,:,:,:));
33 end
34
35 if nargin > 2
36     if ndims(highlightRegion) ~= 3
37         error('Input ''highlightRegion'' must have 3 dimensions');
38     end
39     if size(highlightRegion) ~= size(head)
40         error('Input ''highlightRegion'' must be of the same size as
41             ''head''');
42     end
43     tempdataShort = squeeze(tempdata(end,:,:,:));

```

```

41 end
42
43 if nargin > 3
44     if slice > size(tempdata,1)
45         error('Input ''slice'' must be less or equal to the length of
46             the first dimension of ''tempdata''');
47     end
48     tempdataShort = squeeze(tempdata(slice,:,:,:));
49
50 if nargin > 4
51     if ndims(equil) == 3
52         eq = equil;
53     elseif ndims(equil) == 4
54         eq = squeeze(equil(1,:,:,:));
55     else
56         error('Input ''equil'' must have either 3 or 4 dimensions')
57         ;
58     end
59     clear 'equil';
60 end
61 %% Pick how to format the call of SliceBrowser()
62 switch nargin
63     case 1
64         SliceBrowser(head,1,head);
65         colormap(gray);
66     case 2
67         SliceBrowser(tempdataShort,tempdata,head);

```

```

68     case 3
69     SliceBrowser(tempdataShort,tempdata,head,highlightRegion);
70     case 4
71     SliceBrowser(tempdataShort,tempdata,head,highlightRegion);
72     case 5
73     SliceBrowser(tempdataShort-eq,tempdata,head,highlightRegion);
74     case 6
75     SliceBrowserOverlay(tempdataShort-eq,tempdata,head,
76         highlightRegion,threshold);
77     case 7
78     imgoverlay(head,tempdataShort-eq,point,threshold)
79 end
80 end

```

### B.1.2 tsliceplot

This is a visualization tool I wrote that allows you to view the change in temperature versus time for a line passing through the head. Screenshots of the tool can be seen in Figs. B.1 and B.2.

Usage:

```
tsliceplot(temperature_data, equilibrium_temperature_data)
```

or

```
tsliceplot(change_in_temperature_data)
```

The inputs `temperature_data` and `change_in_temperature_data` should be four dimensional matrices (time, x,y,z) and `equilibrium_temperature_data` is also a four dimensional matrix (1,x,y,z).

The script is available as part of temptools (<https://github.com/greggroth/temptools/tree/master/lib/tsliceplot>).

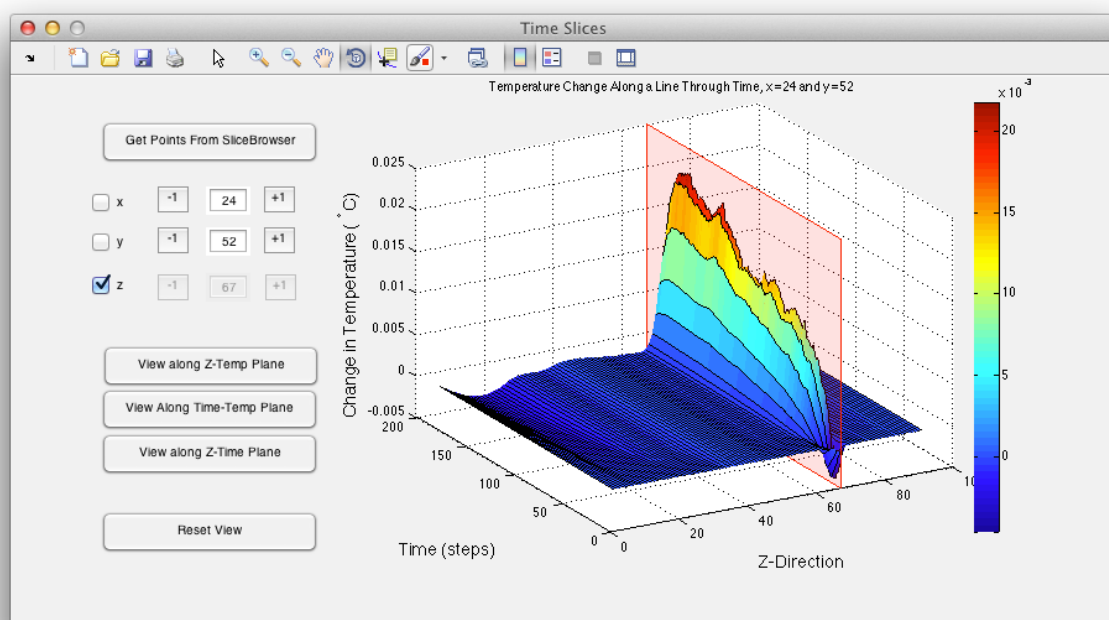


Figure B.1 Experimental data for activity in the motor cortex visualized with tsliceplot.



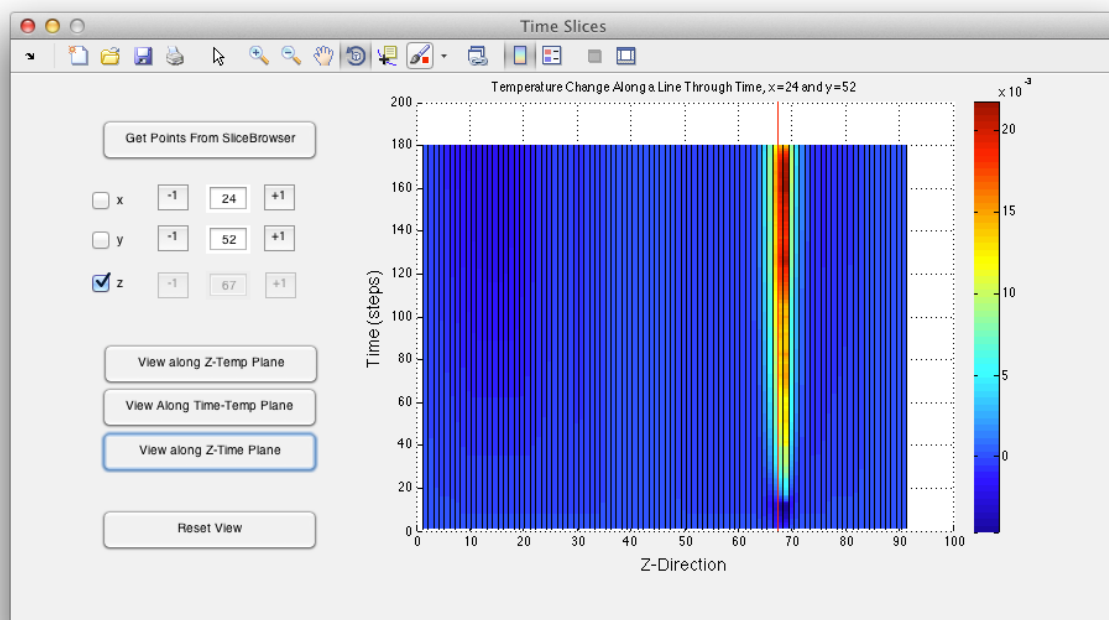


Figure B.2 The same data as is presented in Fig. B.1, but viewed flat-on along the z vs. time plane.