

Methods

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1 Background on study system

Starting in 2006, Monica Geber and collaborators have collected 12+ years of annual estimates for demographic data on the winter annual plant *Clarkia xantiana* ssp. *xantiana*. The data include annual estimates for survival of seedlings to fruiting adults, fruits per adult plant, and seeds per fruit. In addition the data on above-ground vital rates, experiments have been used to infer germination and seed survival in the seed bank. The demographic study has been used to test hypotheses about the geography of demography (Eckhart et al. (2011)) and species distributions (Pironon et al. (2018)).

2 Data

2.1 Seed survival and germination

2.1.1 Seed bag burial experiments

To assess germination and seed survival throughout the year in *C. xantiana*, we use data collected from a series of seed burial experiments. In June-July 2005, we collected seeds at each of the 20 populations included in this study. In October 2005, we buried 30 5×5-cm nylon mesh bags at each population. Each nylon mesh bag contained 100 seeds collected at that population. In January 2006, we removed 10 of these bags and counted the number of germinated seedlings and the number of ungerminated, intact seeds in each bag. We then returned the ungerminated, intact seeds to the resealed bag and returned the bag to the field. In October 2006, we removed these bags and counted the number of ungerminated, intact seeds. We collected the following data:

- n_{ijt} = observed count of seeds in the seed bags at the start of the experiment in October in the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly

- y_{ijt}^{intact} = observed count of ungerminated, intact seeds in the seed bags in January in the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly
- y_{ijt}^{germ} = observed count of germinated seedlings in the seed bags in January in the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly
- y_{ijt}^{total} = observed count of ungerminated, intact seeds plus germinated seedlings in the seed bags in January in the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly
- y_{ijt}^{surv} = observed count of ungerminated, intact seeds in the seed bags in October in the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly

We started these seed burial experiments in three subsequent years (2005, 2006, 2007) to obtain multiple estimates for seed survival and germination.

2.1.2 Viability trials

To assess what proportion of intact seeds are viable, we use data collected from viability assays conducted on seeds when they are unearthed in October. Only some proportion of the seeds that are unearthed intact in the seed burial experiments are likely to be viable—put another way, seeds that are intact may not be viable. Because seeds unearthed in January are reburied, we do not have direct estimates of viability in January.

Each year that we conducted seed burial experiments, we also conducted seed viability trials. After bags were removed from the field in October, we returned the bags to the lab. In the lab, we conducted germination trials and viability assays on subsets of the seeds from each bag to estimate the viability of the ungerminated, intact seeds. First, we placed up to 15 seeds from each bag on to moist filter paper in a disposable cup and observed germination over 10 days; we counted and removed germinants every 2 days.

After 10 days, all remaining ungerminated seeds (up to a total of 10 seeds) were sliced in half and individually placed into the wells of 96-well plates filled with a solution of tetrazolium chloride, which stains viable tissue red. [Eckhart et al. (2011): not all ungerminated seeds were tested; most were] We covered the plates with foil. Each 96-well plate contained seed from at least one bag per population of a given seed-age class. Two or three tests of up to 15 seeds each were conducted for each bag. We checked and counted for viable seeds every 2 days for 10 days.

We collected the following data:

- n_{ijt}^{germ} = observed count of seeds at the start of the X^{th} germination trial for the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly
- y_{ijt}^{germ} = observed count of germinated seedlings in the X^{th} germination trial for the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly
- n_{ijt}^{viab} = observed count of seeds at the start of the X^{th} viability trial for the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly
- y_{ijt}^{viab} = observed count of viable seedlings in the X^{th} viability trial for the i^{th} bag, from the j^{th} population, in the t^{th} year, assumed to be measured perfectly

2.2 Seedling survival to fruiting

To assess the survival of germinants to fruiting plants for *C. xantiana*, we use data of counts of seedlings and fruiting plants in 30 0.5 m² plots at 20 populations from 2006–present (Eckhart et al. (2011)). Each population was visited in February and June to count the number of seedlings and fruiting plants, respectively. Seedlings and fruiting plants in each plot are counted by a single person at each visit.

While we assume that the data on seedlings is measured perfectly (i.e. we did not under- or over-count seedlings), there are at least two possible sources of error: (1) measurement error that arises because we failed to count seedlings that were present and (2) error that arises because seedlings germinated after we visited the population. Germination phenology varies from year to year and with the position of the population in the range. For now, I ignore data that involved undercounting by filtering out those rows in the dataset. We may want to develop a model that relates our estimate of seedlings to the true number of seedlings in a plot because we sometimes observe more fruiting plants than seedlings but I had trouble developing such a model for the time being.

We assume that the data on fruiting plants is measured perfectly (i.e. we did not under- or over-count) because plants stand out from the background vegetation in June. Our model estimates the proportion of seedlings that survive to become fruiting plants. Define:

- n_{ijk} = observed counts of seedlings in the i^{th} plot, from the j^{th} population, from the k^{th} year
- y_{ijk} = observed counts of fruiting plants in the i^{th} plot, from the j^{th} population, from the k^{th} year, assumed to be measured perfectly

2.3 Fruits per plant

To assess the number of fruits per plant for *C. xantiana*, we use data on counts of the number of fruits per plant at 20 populations (Eckhart et al. (2011)). At each population, we made two sets of counts. First, we counted the number of fruits per plant on all plants in the 0.5m² permanent plots. Second, we counted the number of fruits per plant on plants that we sampled haphazardly across the site using throws of a 0.5m² grid.

From 2006–2012, we counted the number of undamaged fruits on a plant. We then took the damaged fruits on a plant and visually stacked them end to end to estimate how many

additional undamaged fruits that was equivalent to (e.g. two half fruits corresponded to one undamaged fruit). We used these counts to estimate the number fruits produced per plant.

From 2013–present, we counted the number of undamaged and damaged fruits on a plant. We used these counts to estimate the number of fruits produced per plant.

We seek to estimate the number of fruits produced per plant. Define:

- y_{ijk}^{TFE} = observed counts of total fruit equivalents per plant on the i^{th} plant, from the j^{th} population, from the k^{th} year, assumed to be measured perfectly
- n_{ijk} = observed counts of total fruits per plant (sum of y_{ijk}) on the i^{th} plant, from the j^{th} population, from the k^{th} year, assumed to be measured perfectly

2.4 Seeds per fruit

To assess the number of seeds per fruit for *C. xantiana*, we use data on counts of the seeds per fruit of fruits that were haphazardly collected in 20 populations (Eckhart et al. (2011)). In the field, we collected fruits that were undamaged. In the lab, we broke open the fruits to count the number of seeds per fruit. For each population in each year, we attempted to obtain 20-30 counts of seeds produced per undamaged fruit.

From 2006–2012, we collected one undamaged fruit from each of 20-30 plants that were haphazardly collected across each population.

From 2013–present, we collected one undamaged and one damaged fruit from each of 20-30 haphazardly selected plants distributed across each population. The plants were outside permanent plots to avoid affecting seed input. We used these fruits to estimate the mean number of seeds produced by undamaged and damaged fruits. In the lab, we broke open the fruits to count the number of seeds per fruit. For each population in each year, we attempted to obtain 20-30 counts of seeds produced per undamaged fruit and 20-30 counts of seeds produced per damaged fruit.

131 We seek to estimate the number of seeds per undamaged fruit. Define:

- 132 • y_{ijk}^{und} = observed counts of seeds in the i^{th} undamaged fruit, from the j^{th} population,
133 from the k^{th} year, assumed to be measured perfectly
- 134 • λ_{jk} = true, unobserved mean number of seeds per undamaged fruit from the j^{th} pop-
135 ulation, from the k^{th} year

3 Models

136 Across all datasets, we have data from multiple years and populations. The goal is to
137 obtain model-based estimates of vital rates. We use the models to separate variability in
138 parameters between temporal variability and variability due to sampling (e.g. [Gould and](#)
139 [Nichols \(1998\)](#)). This is particularly important for our estimates of per capita reproductive
140 success because we calculate annual estimates to quantify interannual variation. Failing to
141 account for sampling variation can inflate estimates of temporal variation. Sample sizes
142 for parameters vary both within years across populations, as well as across populations.
143 Accounting for sampling variability is important for making conclusions about differences
144 among populations.

3.1 Seed survival and germination

All data from seed bags and viability trials is in the form of binomial trials: we have counts of seeds at the start and end of an experimental window of time. All models have the same structure for seeds in bag i in year j in population k . If the number of seeds starting the trial (trials) is n_{ijk} and the number of seeds at the end of the trial (successes) is y_{ijk} , we write a model that has a population-level mean and year-level means drawn from the population-level distribution. Broadly, this is two-level hierarchical model with a population-level mean, and

year-level means drawn from the population-level distribution. The probability of success for each bag is drawn from this year- and population-level distribution. The model uses a binomial likelihood. The joint posterior for viability trials, seed bag experiments in the first year, and for germination of seeds in the second year thus has the following general form:

$$\begin{aligned}
[\boldsymbol{\alpha}, \boldsymbol{\mu}, \boldsymbol{\sigma}, \boldsymbol{\mu}^{\text{pop}}, \boldsymbol{\sigma}^{\text{pop}} | \mathbf{y}^{\text{tot}}] &\propto \prod_{i=1}^I \prod_{j=1}^J \prod_{k=1}^K \text{binomial}(y_{ijk} | n_{ijk}, \text{logit}^{-1}(\alpha_{ijk})) \\
&\times \text{normal}(\alpha_{ijk} | \mu_{jk}, \sigma_{jk}) \\
&\times \text{normal}(\mu_{jk} | \mu_j^{\text{pop}}, \sigma_j^{\text{pop}}) \\
&\times \text{half-normal}(\sigma_{jk} | 0, 1) \\
&\times \text{normal}(\mu_j^{\text{pop}} | 0, 1000) \text{half-normal}(\sigma_j^{\text{pop}} | 0, 1).
\end{aligned} \tag{1}$$

145 The number of trials (n_{ijk}) and successes (y_{ijk}) is different depending on the parameter
146 being modeled. Here, I use a centered parameterization.

The model for seed survival from October of year 1 to January of year 2 is a bit more complicated. We did not unearth these bags in year 1 of the experiment, so we do not have data on survival in the bags until we dig them up in January of year 2. We use the bags from the first year of the experiment to estimate survival up until October of year 1 and then infer survival between October of year 1 to January of year 2. I constructed a model to combine the experimental data from years one and two, jointly estimating survival rates. The model retains the two-level hierarchical structure with a population-level mean, and year-level means drawn from the population-level distribution. The probability of success for each bag is drawn from this year- and population-level distribution. The model uses a

binomial likelihood.

$$\begin{aligned}
& [\alpha_D, \alpha_4, \theta, \mu_D, \sigma_D, \mu_4, \sigma_4, \mu_D^{\text{pop}}, \sigma_D^{\text{pop}}, \mu_4^{\text{pop}}, \sigma_4^{\text{pop}} | \mathbf{y}^{\text{oct}}, \mathbf{y}^{\text{tot2}}] \propto \\
& \prod_{j=1}^J \left\{ \prod_{i=1}^{N_1} \prod_{k=1}^{K_1} \text{binomial}(y_{ijk}^{\text{oct}} | n_{ijk}^{\text{age1}}, \text{logit}^{-1}(\alpha_{D,ijk})) \right. \\
& \times \text{normal}(\alpha_{D,ijk} | \mu_{D,jk}, \sigma_{D,jk}) \text{normal}(\mu_{D,jk} | \mu_{D,j}^{\text{pop}}, \sigma_{D,j}^{\text{pop}}) \text{half-normal}(\sigma_{D,jk} | 0, 1) \\
& \times \text{normal}(\mu_{D,j}^{\text{pop}} | 0, 1000) \text{half-normal}(\sigma_{D,j}^{\text{pop}} | 0, 1) \left. \right\} \\
& \times \left\{ \prod_{i=1}^{N_2} \prod_{k=1}^{K_2} \text{binomial}(y_{ijk}^{\text{tot2}} | n_{ijk}^{\text{age2}}, \text{logit}^{-1}(\alpha_{4,ijk}) \times \theta_{jk}) \right. \\
& \times \text{normal}(\theta_{jk} | \mu_{D,jk}, \sigma_{D,jk}) \\
& \times \text{normal}(\alpha_{4,ijk} | \mu_{4,jk}, \sigma_{4,jk}) \text{normal}(\mu_{4,jk} | \mu_{4,j}^{\text{pop}}, \sigma_{4,j}^{\text{pop}}) \text{half-normal}(\sigma_{4,jk} | 0, 1) \\
& \times \text{normal}(\mu_{4,j}^{\text{pop}} | 0, 1000) \text{half-normal}(\sigma_{4,j}^{\text{pop}} | 0, 1) \left. \right\}. \tag{2}
\end{aligned}$$

3.1.1 Model for seed burial experiment data

147 We obtain the population-level posterior distribution of seed survival from October to Jan-
 148 uary (θ_1), germination in January (θ_2), and seed survival from January to October (θ_3) by
 149 marginalizing across years. We transform these posteriors to $[0, 1]$ by taking the inverse logit;
 150 this transforms the parameters into the probability of success.

151 We also obtain the population-level posterior distribution seed survival from October of
 152 year 1 to January of year 2 (θ_4) and germination in January of year 2 (θ_5) by marginal-
 153 izing across years. We transform these posteriors to $[0, 1]$ by taking the inverse logit; this
 154 transforms the parameters into the probability of success.

3.1.2 Model for viability trial data

155 We obtain the population- and population-by-year posterior distributions of success in ger-
 156 mination (ν^g) and viability (ν^v) trials by marginalization. We transform these posteriors

157 to $[0, 1]$ by taking the inverse logit; this transforms the parameters into the probability of
 158 SUCCESS.

3.2 Seedling survival to fruiting

We estimated survival across all populations taking into account both temporal and between-population variability with the following model. We write a model that has a population-level mean and year-level means drawn from the population-level distribution. The probability of success (seedling survival to fruiting) for each plot is drawn from this year- and population-level distribution. The model has a binomial likelihood, and thus thus has a similar structure as the model for data on seed survival:

$$\begin{aligned}
 [\alpha_S, \mu_S, \sigma_S, \mu_S^{\text{pop}}, \sigma_S^{\text{pop}} | \mathbf{y}^{\text{fruiting}}] &\propto \prod_{i=1}^I \prod_{j=1}^J \prod_{k=1}^K \text{binomial}(y_{ijk}^{\text{fruiting}} | n_{ijk}^{\text{seedling}}, \text{logit}^{-1}(\alpha_{S,ijk})) \\
 &\times \text{normal}(\alpha_{S,ijk} | \mu_{S,jk}, \sigma_{S,jk}) \\
 &\times \text{normal}(\mu_{S,jk} | \mu_{S,j}^{\text{pop}}, \sigma_{S,j}^{\text{pop}}) \\
 &\times \text{half-normal}(\sigma_{S,jk} | 0, 1) \\
 &\times \text{normal}(\mu_{S,j}^{\text{pop}} | 0, 1000) \text{half-normal}(\sigma_{S,j}^{\text{pop}} | 0, 1).
 \end{aligned} \tag{3}$$

159 We obtain the population- and population-by-year posterior distributions of seedling survival
 160 to fruiting (σ) by marginalization. We transform these posteriors to $[0, 1]$ by taking the
 161 inverse logit; this transforms the parameters into the probability of success.

3.3 Fruits per plant

We estimated fruits per plant across all populations taking into account both temporal and between-population variability with the following model. We constructed models with population-level mean and year-level means drawn from the population-level distribution.

We used a Poisson likelihood to model the count data of fruits per plant. We modeled sampling uncertainty with a lognormal, which draws the true number of fruits (the zs) from a distribution with the median (μ). Each combination of year and population is assigned its own sampling variance (σ^2). The general form of the model is the following:

$$\begin{aligned}
& [z, \mu_{\text{TF}}, \sigma_{\text{TF}}^2, (\sigma_{\text{TF}}^{\text{pop}})^2, \nu_{\text{TF}} | \mathbf{y}^{\text{TF}}] \propto \\
& \prod_{j=1}^J \prod_{i=1}^{N_1} \prod_{k=1}^{K_1} \text{Poisson}(y_{ijk}^{\text{TF}} | z_{\text{TF},ijk}) \text{lognormal}(z_{\text{TF},ijk} | \log(\mu_{\text{TF},jk}), \sigma_{\text{TF},jk}^2) \\
& \times \text{lognormal}(\mu_{\text{TF},jk} | \log(g(\nu_{\text{TF},j}), (\sigma_{\text{TF},j}^{\text{pop}})^2)) \\
& \times \text{inverse gamma}(\sigma_{\text{TF},jk}^2 | 0.001, 0.001) \\
& \times \text{gamma}(\nu_{\text{TF},j} | 1, 1) \text{inverse gamma}((\sigma_{\text{TF},j}^{\text{pop}})^2 | 0.001, 0.001).
\end{aligned} \tag{4}$$

162 We obtain the population- and population-by-year posterior distributions of fruits per plant
163 (F) by marginalization.

164 Estimating total fruit equivalents per plant required combining two types of data. First,
165 I used counts of total fruit equivalents per plant from (2006-2012). I modeled these counts
166 as observations (zs). Second, I used counts of undamaged and damaged fruits per plant from
167 (2013-present). I separately modeled counts of each as observations to obtain estimates of
168 undamaged and damaged fruits per plant.

3.4 Seeds per fruit

We estimated seeds per fruit across all populations taking into account both temporal and between-population variability with the following model. We constructed models with population-level mean and year-level means drawn from the population-level distribution. We used a Poisson likelihood to model the count data of seeds per fruit. We modeled sampling uncertainty with a lognormal, which draws the true number of seeds (the zs) from a

distribution with the median (μ). Each combination of year and population is assigned its own sampling variance (σ^2). The general form of the model is the following:

$$\begin{aligned}
& [\mathbf{z}, \boldsymbol{\mu}_{\text{TF}}, \boldsymbol{\sigma}_{\text{TF}}^2, (\boldsymbol{\sigma}_{\text{TF}}^{\text{pop}})^2, \boldsymbol{\nu}_{\text{TF}} | \mathbf{y}^{\text{TF}}] \propto \\
& \prod_{j=1}^J \prod_{i=1}^{N_1} \prod_{k=1}^{K_1} \text{Poisson}(y_{ijk}^{\text{TF}} | z_{\text{TF},ijk}) \text{lognormal}(z_{\text{TF},ijk} | \log(\mu_{\text{TF},jk}), \sigma_{\text{TF},jk}^2) \\
& \times \text{lognormal}(\mu_{\text{TF},jk} | \log(g(\nu_{\text{TF},j}), (\sigma_{\text{TF},j}^{\text{pop}})^2)) \\
& \times \text{inverse gamma}(\sigma_{\text{TF},jk}^2 | 0.001, 0.001) \\
& \times \text{gamma}(\nu_{\text{TF},j} | 1, 1) \text{inverse gamma}((\sigma_{\text{TF},j}^{\text{pop}})^2 | 0.001, 0.001).
\end{aligned} \tag{5}$$

169 We obtain the population- and population-by-year posterior distributions of seeds per fruit
170 (ϕ) by marginalization.

171 First, I used counts of seeds per undamaged fruit (2006-present). I modeled these counts
172 as observations (zs). Second, I used counts of seeds from damaged fruits per plant from
173 (2013-present). I separately modeled counts of each as observations to obtain estimates of
174 seeds per damaged fruit.

4 Demographic modeling framework

175 We use observational and experimental data from 20 populations of *Clarkia xantiana* to esti-
176 mate transition probabilities across the life cycle. We model population-specific estimates for
177 belowground vital rates, and model year- and population-specific estimates for aboveground
178 vital rates. We use these parameter estimates to analyze correlations between germination
179 probability and variance in per-capita reproductive success, correlation between germina-
180 tion probability and seed survival, and to compare the observed germination fraction to the
181 optimal germination fraction from a density-independent model.

4.1 Parameters for belowground transitions

To estimate population-specific estimates for belowground vital rates, I use data from seed burial experiments in the field and seed viability trials in the lab. I combine these data to infer germination and seed survival. Briefly, I estimate probabilities of success using data from seed burial experiments and viability trials $(\theta_1, \theta_2, \theta_3, \theta_4, \theta_5, \theta_g, \theta_v)$. I use these probabilities to compose transition probabilities that correspond to the life history of *Clarkia xantiana* (s_1, s_2, s_3, g_1) . I describe this approach in detail in the **Appendix on Conditional Probability**. Figure 1 illustrates the relationship between the data and the estimated probability of success.

The probability that seeds from the start of the experiment remain intact in January is represented as θ_1 . In January, all seeds are intact (this includes viable and non-viable seeds). I estimate the probability of a seedling emerging, conditional on being intact as θ_2 . I assume that there is no decay or loss of viability during germination. The number of intact seeds before germination is equal to the sum of the number of seeds and seedlings after germination. At this point, seeds transition into one of four possible states. Intact and viable seeds may have (1) germinated or (2) not germinated and remained dormant. All (3) other intact seeds are non-viable because (4) seeds that were not viable could not have germinated. Finally, we represent the probability of a seed being intact in October, conditional on being intact in January as θ_3 .

We use the viability trials to estimate the probability of viability (ν_1) for a seed that is intact in October one year after the seed bags were buried. The viability trials are a two-stage process: seeds are subject to germination trials before a fraction of the remaining, ungerminated seeds are tested for viability. The probability that a seed germinates in the germination trial is θ_g and the probability that it is viable, conditional on not germinating in the germination trial, is θ_v . I estimate the overall probability of viability, ν , as $\theta_g + \theta_v(1 -$

θ_g). This weights the estimates relative to the probability of germination (eg. if no seeds germinate the estimate of viability will mostly come from the viability test).

I make some assumptions in order to incorporate the loss of viability into the model. I assume that viability is lost at a constant rate, and that germination removes some number of seeds from the pool of viable seeds but does not change the rate of decay. Some fraction of the intact seeds in January are viable before germination ($\nu_1^{1/3}$), and some of those viable seeds germinate.

The seed burial experiments and viability trials provide information about the fate of seeds in the seed bank. We define s_1 as the probability of a seed being intact and viable from October in year t to January in year $t+1$. We define g_1 as the probability of germination for a seed that is intact and viable. We define s_2 as the probability of survival from January to October in year t for a seed that was intact and viable in January. Mathematically, we write each of the transition probabilities as follows:

$$\begin{aligned} s_1 &= \theta_1 \times (\theta_2 + (1 - \theta_2) \times \nu_1^{1/3}) \\ g_1 &= \frac{\theta_2}{1 - (1 - \nu_1^{1/3}) \times (1 - \theta_2)} \\ s_2 &= \theta_3 \times \nu_1^{2/3} \end{aligned} \tag{6}$$

To estimate the survival of two-year old seeds from October to January (s_3) requires some additional assumptions. First, we used a different set of bags for these estimates than for the estimates of first-year rates (s_1, s_2, g_1). We use the second set of bags to estimate the probability of viability (ν_2) as described above, combining θ_g and θ_v . We calculate the viability in the second January as the product of viability at the end of the first year (ν_1) and the fraction of first year viability remaining in January ($(\nu_2 \div \nu_1)^{1/3}$). For cases where $\nu_2 \geq \nu_1$, we assumed that all seeds in the bag were viable in October $t+1$ and calculated the fraction of first year viability remaining in January as $(\nu_2)^{1/3}$.

When I estimate the probability of seedling survival from the second October to second January (s_3), I incorporate the expected proportion of seeds that are expected to be intact at the end of the first year ($\theta_1(1 - \theta_2)\theta_3$). I define s_3 as the probability of survival from October in year t to January in year $t + 1$ for a seed that was intact and viable in October. Mathematically, we write the transition probability as follows:

$$s_3 = \theta_4 \times (\theta_5 + (1 - \theta_5) \times \nu_2^{1/3}) \quad (7)$$

4.2 Parameters for aboveground transitions

221 To obtain year- and population-specific estimates for aboveground vital rates, we use data
 222 from annual observational surveys of 20 *Clarkia xantiana* populations. We use data from
 223 surveys of plots to estimate seedling survival to fruiting (ϕ). We also estimate total fruit
 224 equivalents per plant (F) and seeds per fruit (ϕ). We combine these data to obtain annual
 225 estimates of per-capita reproductive reproductive success.

From 2006–2012, we counted total fruit equivalents in the field, and estimated the number of seeds in undamaged fruits in the lab. Starting in 2013, we began to count undamaged and damaged fruits in the field, and estimate the number of seeds in both undamaged and damaged fruits in the lab. To compare these components of fitness across all years of the study, we converted the number of undamaged and damaged fruits to total fruit equivalents with the following relationship:

$$\text{TFE} = \text{undamaged fruits} + \frac{\text{seeds per damaged fruit}}{\text{seeds per undamaged fruit}} \times \text{damaged fruits} \quad (8)$$

226 In these calculations, we use the marginalized, posterior distributions from the models for
 227 fruits per plant and seeds per fruit. We use the number of seeds per undamaged fruit to
 228 calculate per-fruit seed production. While using a single measure for counts of fruits per

plant might be ideal, such a metric is not available to us. We instead combine several sources of data to constrain our estimate while propagating uncertainty from those estimates.

5 Analysis

5.1 Correlation between germination probability and seed survival

Increased seed survivorship is predicted to decrease the optimal germination probability (Cohen (1966); Ellner (1985)). I assessed whether the observed germination probability was negatively correlated with seed survival (Gremer and Venable (2014)). I calculated seed survival, s_2s_3 , the probability that seeds which do not germinate in January remain in the seed bank until the following January. I obtained the posterior distribution for the correlation between germination and seed survival by calculating the correlation of g_1 and s_2s_3 across sites at each iteration of the MCMC output (Hobbs and Hooten (2015), pp. 194-5). Bet hedging models predict that germination probability should be negatively correlated with seed survival; 95% credible intervals that do not overlap zero provide support for this prediction.

5.2 Correlation between germination probability and variance in per-capita reproductive success

Increased variance in per-capita reproductive success is predicted to decrease the optimal germination probability (Cohen (1966); Ellner (1985)). I assessed whether the observed germination probability was negatively correlated with variance in per-capita reproductive success (Venable (2007)). Per-capita reproductive success F_{jk} at population j in year k was

calculated as:

$$F_{jk} = \phi_{jk} \times \lambda_{jk}^F \times \lambda_{jk}^P \quad (9)$$

where

$$\begin{aligned} \phi_{jk} &= \text{logit}^{-1}(\alpha_{0,j}^S + \beta_{jk}^S) \\ \lambda_{jk}^F &= \exp(\alpha_{0,j}^F + \beta_{jk}^F) \\ \lambda_{jk}^P &= \exp(\alpha_{0,j}^P + \beta_{jk}^P) \end{aligned} \quad (10)$$

241 To calculate the temporal variation in per-capita reproductive success for each population,
 242 I sampled the posterior distribution of reproductive success for each year and calculated the
 243 geometric SD of per capita reproductive success. I obtained the posterior distribution for
 244 the correlation between germination and geometric SD of per capita reproductive success at
 245 each iteration of the MCMC output (Hobbs and Hooten (2015), pp. 194-5). Bet hedging
 246 models predict that germination probability should be negatively correlated with temporal
 247 variance in fitness; 95% credible intervals that do not overlap zero provide support for this
 248 prediction.

5.3 Density-independent model for germination probability

We use estimates of seed survival and reproductive success to investigate the adaptive value of delayed germination (Gremer and Venable (2014)). We parameterize a model of population growth rate and calculate the optimal germination strategy for different combinations of seed survival and reproductive success. We describe *Clarkia xantiana*'s life cycle and calculate

population growth rate with the equation:

$$\lambda = g_1 Y(t) s_0 s_1 + (1 - g_1) s_2 s_3 \quad (11)$$

The parameters in this equation are described in [Table XX](#). Seed survival rates (s_0, s_1, s_2, s_3) are population-level estimates. Per capita reproductive success ($Y(t)$) is calculated as the product of seedling survival to fruiting, fruits per plant, and seeds per fruit (equation (9)). Temporal variation is incorporated into the model by varying the per-capita reproductive success, $Y(t)$, between years.

For each population, I numerically calculate the optimal germination probability for the observed variation in reproductive success and seed survival. In each case, I sampled parameters in the equation for density-independent growth (equation (11)) from the fitted vital rate distributions. I resampled the fitted values of per-capita reproductive success ($Y(t)$) to obtain a sequence of 1000 years. I used this same sequence of $Y(t)$ and the fitted seed survival probabilities to calculate long-term stochastic population growth rates (λ_s) at each germination probability along an evenly spaced grid of possible germination probabilities (G) between 0 and 1. The optimal germination probability is estimated as the value of G that maximizes the geometric mean of the population growth rate. I repeat the simulations 50 times for each population, resampling the sequence of per-capita reproductive success, $Y(t)$, each time. I then calculated the mean of the optimal germination fractions.

Models in which per-capita reproductive success is density-independent predict that germination probability should respond to variance in fitness (Cohen (1966)). To evaluate the density-independent model, I compared modeled germination probabilities to predicted germination optima.

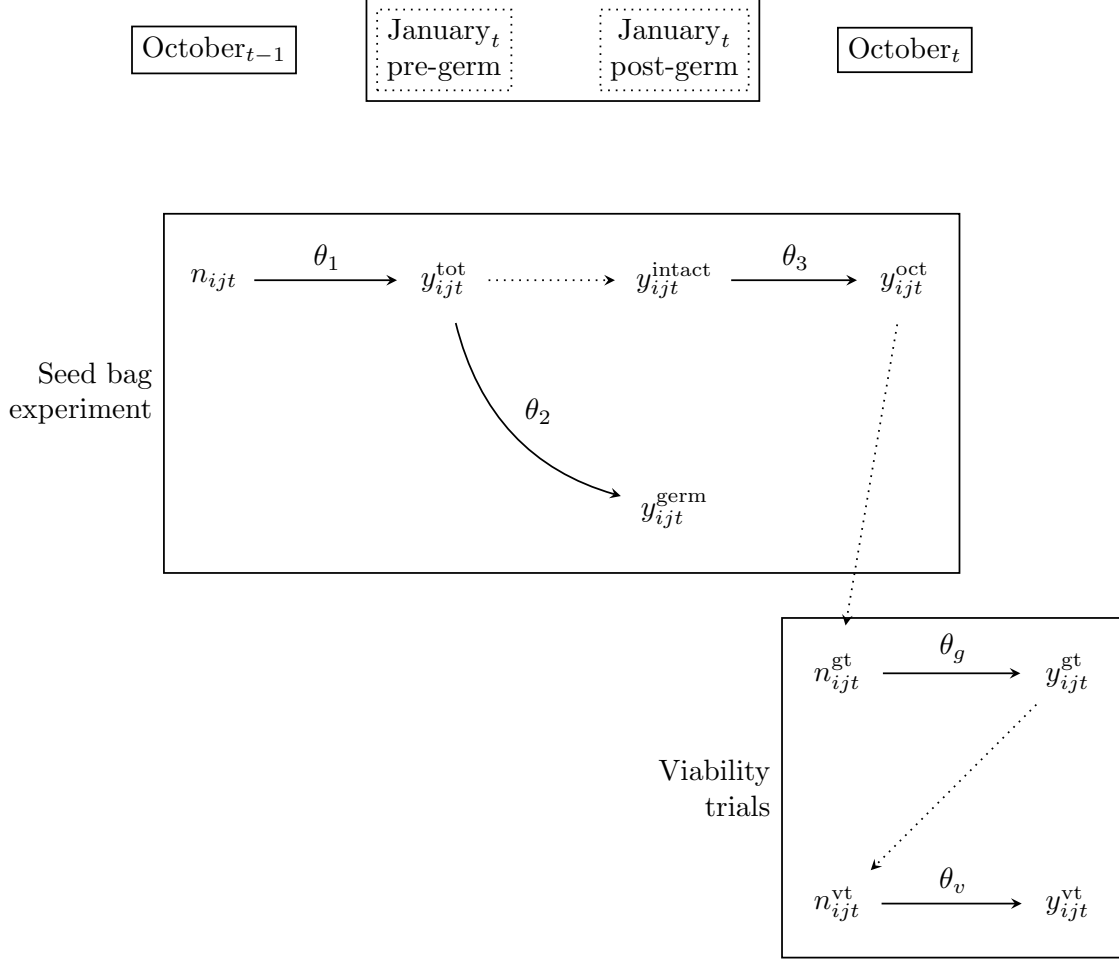


Figure 1: Diagram of data from the seed bag experiments and viability trials. There are two boxes: one for the seed bag experiment and one for the viability trials. In the seed bag experiment, I split January into two steps, one for just before germination and one for just after. Solid arrows represent probabilities estimated with a binomial experiment and are labeled with corresponding parameters. Dotted arrows represent cases where the seeds at the head of the arrow include some, possibly all, seeds at the tail of the arrow.

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