- [Working title]: Intraspecific variation in range-wide seed bank dynamics is not consistent with density-independent model of bet hedging
  - Gregor-Fausto Siegmund and Monica Geber
- Last updated: March 20, 2021

# Writing list

- 1. Read Cohen & Ellner to identify the role of complete reproductive failure in the original
- 6 models for the evolution of bet hedging
- <sup>7</sup> 2. Write description of modeling for the seed bag experiment
- 3. Write about strong and weaker test of the hypothesis (partial pooling, no partial pool-
- 9 ing)
- 4. Write paragraph about history of studies of bet hedging via seed bank, with emphasis
- on how this study addresses this question at an intraspecific level. Lower level of
- variation in intraspecific germination fraction. (link)
- 5. Write models for belowground vital rates
- 6. Revise description for density-independent model for germination probability
- 7. Write and implement model checking process

# Introduction

```
General introduction to seed banks/dormancy – Clarkia as a case where seed banks have
   been posited to be important – bet hedging theory and predictions – review of tests of bet
17
   hedging in plants – what we did
18
      Seed banks can buffer plant populations against environmental change and stochasticity
19
   (Eager et al. (2014); Paniw et al. (2017)), increase effective population size (Nunney (2002);
20
   Waples (2006)), and maintain genetic diversity (Mccue and Holtsford (1998)). Dormancy
21
   can affect the outcome of evolution (Heinrich et al. (2018); Ritland (1983)). Theory thus
   suggests that seed banks have ecological and evolutionary consequences (Evans and Dennehy
   (2005)).
      What drives the evolution of delayed germination? The theory developed by Cohen
25
   (1966) frames the problem in the following terms. What is the optimal germination fraction
26
   for a given level of interannual variation in fitness and seed survivorship? These models
27
   make it clear that the germination fraction that maximizes long-term population growth
   rate is a function of the distribution of fitness (characterized by the variation in fitness),
   the fitness values, and the rate of seed survivorship. For a given mean fitness, increasing
30
   the variance in fitness decreases the optimal germination fraction (see: Supplementary Ma-
31
   terial: Theoretical background for hypotheses). Increasing seed survivorship decreases the
32
   optimal germination fraction, and the degree to which it does so depends on the probability
33
   of a 'good year'. Specifically, as the probability of a high-fitness year decreases, the opti-
   mal germination fraction decreases. Bet hedging should evolve to maximize the long-term
35
   geometric population growth rate (as compared to the arithmetic population growth rate)
36
   (Cohen (1966, 1968); Ellner (1985a,b)).
37
      Cohen (1966) emphasizes the role of particularly bad years. This is highlighted by the
38
   inequality in equation (12), which states that for the optimal germination strategy to be
```

bet hedging, it is sufficient that the harmonic mean is less than the survival probability of seeds that do not germinate. The minimum fitness has a strong impact on the harmonic 41 mean of fitness (see definition of harmonic mean on Wikipedia), and a single year of very low fitness would tend to make the harmonic mean small. In particular, the harmonic mean is 0 when any of its values is 0. The probability of complete reproductive failure is thus particularly important. Years with no seedling survival in the plots or no germination are 45 those in which fitness would be zero; there are only 4/20 populations in which this does not happen. We fit two types of models to the aboveground data; one with partial pooling and 47 one with no pooling. The first was an attempt to correct for sampling variation; the latter an attempt to estimate per-capita reproductive success as-is, providing a more extreme estimate of interannual variation but one that reflects the combined effect of sampling variation and true variation. 51

Revisit theory to see particular role of complete reproductive failure versus low fitness years more generally.

In paragraph on theory, discuss density-independent models Cohen (1966, 1968), densitydependent models Ellner (1985a,b), predictive germination Cohen (1967), and unstructured
versus structured models Easterling and Ellner (2000) (Tuljaparkur 1993, Valleriani and
Tielborger 2006).

#### Paragraph about the history of empirical studies of bet hedging.

58

Population vital rates are known to vary across *Clarkia xantiana*'s geographic range.

Population growth rates determine species abundance and distribution, and are ultimately
what limit persistence beyond range edges. Geographic patterns to vital rates have so far
been studied to help understand the demography of geography. Seed banks are a strategy
that annual plants may use to buffer against environmental variation and may be part of
population persistence. I will begin by characterizing geographic variation in belowground
vital rates. [What is the geographic pattern to variation in germination or seed survival?]

[I think this question could be expanded to make clear predictions and/or address another aspect such as variation in time.]

A previous study with Clarkia xantiana suggests that the soil seed bank is important for population dynamics in Clarkia xantiana (Eckhart et al. (2011)). A separate set of seed burial experiments suggests that seeds of C. xantiana can remain viable in the soil for at least 10 years (Moeller personal communication). In the study of C. xantiana population dynamics that showed a decline of long-term stochastic population growth rate from west to east across the range, Eckhart et al. (2011) inferred a decrease in survival through winter (s1) and an increase in germination rate (g1) of first-year seeds from west to east.

See also the comments in Lewis 1953 on lifespan of seeds, absence of dormancy, and 75 cued germination. Lewis 1963 Presidential address at SSE on catastrophic selection is also 76 relevant. Discusses extinction and possible absence of a seed bank in populations of Clarkia 77 species. Specifically mentions a population in the upper Kern Valley that had both whites 78 and pinks; the pinks went extinct during a drought year while the whites maintained their population size. This work was followed up by others on different species in Clarkia genus. I think its important to consider the possibility that C. xantiana being studied in 1963 was a 81 xantiana/parviflora ssp. which may have different dynamics than the outcrossing xantiana/xantiana. Furthermore: the upper parts of the range have the highest rates of germination which may also trade off with low survival. It would be interesting to put that observation in the range-wide context! 85

Knies et al. 2004 studied 2 C. xantiana populations; seeds of outcrosser are heavier than selfers. Identify heavier seeds ( .1 mg heavier at SC than WH population). Seeds from both populations were bullet-shaped but differ in shape of exotesta cells; WH cells are gourd shaped, SC shapes are even width and spherical. Seed size/number trade-off: smaller size for dispersal; greater mass increases germination probability. Smaller seed size also facilitates seed integration into soil seed bank. Small seed size is also correlated with a persistent

- seed bank (Thompson et al. 1993). Larger seeds may germinate under more conditions and perhaps have smaller seed bank.
- Moeller et al. 2011 population genetic study.
- Eckhart and Geber 1999 identify more populations than those of Lewis, populations that they have since monitored intensively.
- Price et al. 1985 followed a population for 2 years, did an elimination experiment. determined a dormant seed bank. Snow 1960 reports eliminating plants and not observing plants the following year.

Geber and Eckhart 2005 observed a 30-fold difference in lifetime fitness calculated as 100 number of germinants times survival, flower number, fruit set and seed set between two 101 transplant years (one wet, one dry). Fitness was very low in the dry year of the study 102 (0.02-0.03 seeds per seed planted). They observed a 2-4 fold difference between years in 103 germination, seedling survival, and flower number. They did not observe a difference in 104 fruit or seed set. Western populations showed evidence of regional adaptation: survival and 105 flower number. Germination declined from the western to eastern transplant sites in contrast 106 to what we observe here? Germination is consistently higher in subspecies xantiana than 107 parviflora. From west to east, rainfall declines and becomes more variable. Low survivorship 108 is the product of end-of-season drought, small mammal herbivory (Geber and Eckhart 2005, Benning et al. 2019), competition by higher vegetation cover of grasses (survivorship increases 110 with plant size), fungal rust infection. Plants from western populations are larger. Note that 111 the transplant experiments took place on igneous soils, and later work by Vince has shown 112 that soil type is important. 113

Eckhart et al. 2010 IJPS: geographic patterns in precipitation, decline from west to east, mirrored geographic pattern in midday water potential (reduced performance). Metasedimentary soils were more resistant and finer. Soil characteristic curves indicated that metasedimentary soils can hold more water but that it becomes unavailable to plants at higher water contents. Higher water potential on steeper, eroding slopes; this soil was softer and coarser.

Decline in soil water potential (more negative) decreases plant size and fruit number. However, soil water potential is unrelated to seedling survival in the study years. Metasedimentary rocks may compound effect of decline in precipitation. Metasedimentary rocks may
hold more water but it may be more available to plants in igneous soils.

Bet hedging should evolve to maximize the long-term geometric population growth rate 123 (as compared to the arithmetic population growth rate) (Cohen (1966, 1968); Ellner (1985 a, b)). 124 Seed banks are more likely to be selected in populations which experience higher levels of in-125 terannual variation in per-capita reproductive success. To investigate this empirical relation-126 ship, I will estimate the correlation between interannual variation in per-capita reproductive 127 success and the proportion of seeds that germinate in the winter immediately following seed 128 production. I predict that germination is negatively correlated with interannual variation in 129 per-capita reproductive success. [What is the relationship between interannual variation in 130 fitness and dormancy? This is a question about whether fitness variation and dormancy are 131 positively correlated as expected? this is what selects for bet hedging.] 132

Seed dormancy and persistence in the soil seed bank may be a bet-hedging strategy
that is favored by environmental uncertainty. If this is the case in *C. xantiana*, I think we
might expect increased seed survival and decreased germination at the eastern range edge
(precipitation is more variable at eastern populations in winter and spring). This seems to be
opposite of what was observed in the previous study? though I could also be misinterpreting
this. This is could be one reason to revisit this question with a new analysis.

Why using the Clarkia system? We expect to see differences among populations in either seed survivorship or variance in fitness. In the study of *C. xantiana* population dynamics that showed a decline of long-term stochastic population growth rate from west to east across the range, Eckhart et al. 2011 inferred a decrease in survival through winter (s1) and an increase in germination rate (g1) of first-year seeds from west to east. [need to be clear

about this] also know that the environment changes across the range; that will affect the mean fitness

I expect bet hedging stages to be sensitive to variation in an environmental cue. For 146 something to be an adaptive strategy, it should respond to variation in the environment to 147 capitalize on good years such as ones with high precipitation. To investigate the sensitivity of 148 vital rates to the environment, I estimate the slope of the relationship between environmental 149 cues and vital rates at individual sites using a random coefficients model that estimates 150 covariation between intercepts and slopes. I predict that I will estimate variation both 151 in the intercept and slope for populations, and that variation in the modeled cue [to be 152 determined will be positively related to the estimated slope. What is the relationship 153 between dormancy and environmental cues? This is a question about whether we can make 154 any inferences about the mechanism responsible for bet hedging in this system. 155

Table 1: Models for germination delays: references and predictions

	Density-independent	Density-dependent fit-	Predictive germina-	Structured model
	fitness	ness	tion	
Key theory	Cohen (1966, 1968)	Ellner $(1985a,b)$	Cohen (1967)	Tuljapurkar and Is-
references				tock 1997 Easterling
				and Ellner (2000) Val-
				leriani and Tielborger
				2006
Key empirical	Venable (2007)	Gremer and Venable	Gremer et al. (2016)	
tests		(2014)		
Mean of	increase in $\bar{Y}$ increases	increase in $\bar{K}$ can in-		
seed yield	$G^*$	crease or decrease $\hat{G}$		
CV of	increasing $\rho_Y$ de-	increasing $\rho_K$ or $\rho_C$		
seed yield	creases $G^*$	decreases $\hat{G}$		
Seed	increasing $s$ decreases	increasing $s$ decreases		
survivorship	$G^*$ ; limit near $s=1$ is	$\hat{G}$ ; limit near $s=1$ is		
	p	0		

156

# 1 Methods

# 1.1 Clarkia life history

Clarkia xantiana is a winter annual that germinates with late fall and winter rains. In our 157 study region, the Kern Valley in the southern Sierra Nevada Mountains, historically happens 158 between October and late February or early March. Seedlings grow throughout the winter 159 and spring, and surviving plants flower in late spring and early summer, April into early 160 July. Pollinated fruits set seed in the early summer, June to July. Seeds of C. xantiana are produced in early summer, with fruits that dry out and gradually split open. Most seeds 162 appear to be shed from fruits within 3-4 months after production, but can remain on the 163 plant for up to a year. Seeds are small (< 1 mm in width) and have no structures to aid in 164 aerial dispersal. 165

We represent the Clarkia xantiana as a 166 life cycle graph (Figure 1A) that describes 167 transitions from October of year t to Octo-168 ber of year t+1 in terms of underlying vital 169 rates. The census period occurs when the 170 entire population is seeds, and corresponds 171 to the time at which seed bags are placed 172 into the field (see below). Seeds are grouped 173 into three stages: age 0 seeds, which were 174 produced in the current year; age 1 seeds, 175 which were produced in the previous year; 176 age 2+ seeds, which were produced two or more years ago. Persistence of seeds in the 178 seed bank is represented by transitions from

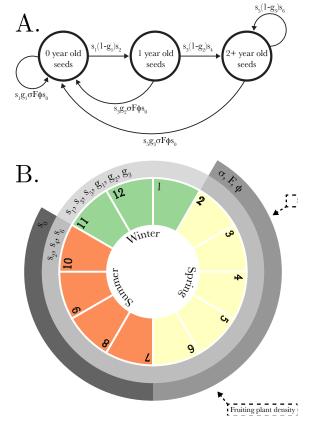


Figure 1: Diagram representations of Clarkia life cycle.

younger to older seeds. Production of new seeds is captured by transition to the age 0 seed state.

Transitions in the life cycle graph are the product of age-specific seed survival and germi-183 nation, and aboveground seedling survival to fruiting, fruit production, and seeds per fruit. 184 Seed-related rates are represented separately for age 0, 1, and 2+ seeds. Germination for 185 each age class is given as  $g_1$ ,  $g_2$ , and  $g_3$ , respectively. Seed survival from seed production to 186 the first October is given as  $s_0$ , and survival from October to February is given as  $s_1$ ,  $s_3$ , 187 and  $s_5$  for age 0, 1, and 2+ seeds, respectively. Survival from February to October is given 188 as  $s_2$ ,  $s_4$ , and  $s_6$  for age 0, 1, and 2+ seeds, respectively. We assume that vital rates remain 189 unchanged after age 2. We also we assume that all plants experience the same vital rates 190 upon germination seed age at germination does not affect seedling survival to fruiting  $(\sigma)$ , 191 fruits per plant (F), or seeds per fruit  $(\phi)$ . 192

The life cycle graph (Figure 1A) corresponds to the annual projection matrix

$$\mathbf{A} = \begin{bmatrix} s_1 g_1 \sigma F \phi s_0 & s_3 g_2 \sigma F \phi s_0 & s_5 g_3 \sigma F \phi s_0 \\ s_1 (1 - g_1) s_2 & 0 & 0 \\ 0 & s_3 (1 - g_2) s_4 & s_5 (1 - g_3) s_6 \end{bmatrix}$$
(1)

that summarizes transitions between stages.

Eckhart et al. 2004 conducted a transplant experiment in two years in which seed set varied ten-fold. There are additional studies that did elimination experiments (with other species of Clarkia) and suggested limited role/role for seed bank because plants did or did not come up with following year.

Lewis 1962 suggested that catastrophic selection played a major role in the dynamics of *Clarkia* populations. In 13 years of data on 20 populations across the species range,

we observed high seedling mortality and/or low germination in at least one year in all but four populations. Although most populations had at least some flowering plants in most years, there were also four populations in which no flowering plants were observed in the entire population. None of these populations, however, went extinct during the period of the study. Seeds can remain viable in the field and laboratory for at least 5 years, contrary to previous suggestions that seeds are unlikely to persist past a few years (Lewis 1962).

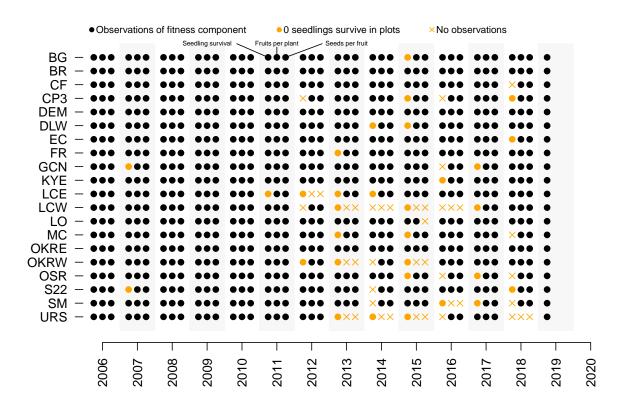


Figure 2: Summary of the aboveground observations, low fitness, and no observations.

## 1.2 Creating the dataset

We used field experiments and surveys to assemble observations of below- and above-ground demography for 20 populations of *Clarkia xantiana* (Table 2). Specifically, we used experiments to estimate transitions in the seed bank and surveys to estimate components of per-capita reproductive success. These demographic data have previously been used to test hypotheses about the geography of demography (Eckhart et al. 2011) and species distributions (Pironon et al. 2018). Here, we sought to obtain population-level estimates of germination and seed survival, and yearly estimates of per-capita reproductive success.

To estimate transitions in the seed bank, we used observations from a seed bag burial 213 experiment conducted in all populations from 2006-2009 (Figure ??). The experiment has 214 been previously described in Eckhart et al. 2011 and we reanalyze the data here. Geber and 215 collaborators buried seeds in bags and unearthed 1, 2, or 3 years after being buried to count 216 seedlings and intact, viable seeds. The experiment was repeated in 3 consecutive years and 217 ended in 2009. We thus have 3 sets of observations associated with 1 year old seeds, 2 sets 218 of observations associated with 2 year old seeds, and 1 set of observations associated with 219 3 year old seeds. We use data from the experiment to estimate age-specific germination 220 and seed survival (see Joint model for seed vital rates) but note that we test predictions of 221 bet-hedging theory that are based on an unstructured seed bank and use only the relevant 222 subset of transitions in our analysis (see Computing vital rates). 223

To estimate per-capita reproductive success, we combine censuses of seedlings and fruiting plants, surveys of fruits per plant, and lab counts of seeds per fruit. To assess the survival of seedlings to fruiting plants, we counted seedlings and fruiting plants in 30 0.5 m<sup>2</sup> permanent plots from 2006–2018 (Eckhart et al. 2011). To assess seed production by plants that survive to reproduction, we counted fruits per plant on individual plants in permanent plots, and on additional haphazardly chosen plants throughout the population. We then attempted to

obtain 20-30 fruits per population, which we used to count seeds per fruit (Eckhart2011).

Table 2: Summary of data sets used to estimate parameters.

Parameter data	Description	Data set	Time span
SEED VITAL RATES	_		
Seed survival and germination	Seed bag burial	$\mathbf{Y}_1$	2006-2009
Seed viability	Viability trials	$\mathbf{Y}_2$	2006-2009
Seed survival and germination	Seed pots	$\mathbf{Y}_3$	2013-2019
SEEDLING SURVIVAL			
Seedling survival to fruiting	Field surveys	$\mathbf{Y}_4$	2006-2019
FRUITS PER PLANT			
Total fruit equivalents per plant	Field surveys	$\mathbf{Y}_5$	2006-2012
Undamaged and damaged fruits per plant	Field surveys	$\mathbf{Y}_{6}$	2013-2019
Total fruit equivalents per plant	Extra plots	$\mathbf{Y}_7$	2006-2012
Undamaged and damaged fruits per plant	Extra plots	$\mathbf{Y}_{8}$	2013-2019
SEEDS PER FRUIT			
Seeds per undamaged fruit	Lab counts	$\mathbf{Y}_{9}$	2006-2019
Seeds per damaged fruit	Lab counts	$\mathbf{Y}_{10}$	2013-2019

# 1.3 Model

232

We use observational and experimental data from 20 populations of *Clarkia xantiana* to estimate transition probabilities across the life cycle. We fit multilevel models to obtain population-specific estimates for belowground vital rates, and year- and population-specific estimates for aboveground vital rates. Because we were interested in describing the life histories of individual populations, we built separate models for each population. Our general approach applies a common model structure to partially pool observations from each population.

We first explicitly describe our formulation in terms linear mixed models before defining the joint posterior (Evans et al. 2010, Ogle and Barber 2020). We assume that the latent mean of observations in year j at a population k,  $\theta_{jk}$ , is drawn from a normal distribution with mean  $\theta_{0,k}$  and variance  $\sigma_j^2$ .

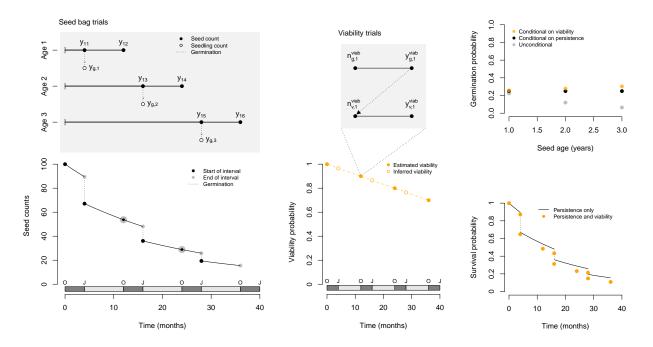


Figure 3: Summary of the seed bag burial experiments and viability trials. Figure will be labeled as (A-B: seed bag trials, C-D: viability trials, E-F: germination probability and survival probability. Add month markers to the y-axis in panels B, D, F. (A) The gray panel contains a graphical representation of the seed bag trials. Seeds were buried at the start of each experiment (100 seeds in month 0). Seed bags were unearthed and intact seeds  $(y_{\cdot\cdot\cdot})$  and germinants  $(y_{g,\cdot})$  counted. The graph below the panel shows a hypothetical survival function associated with persistence of seeds in the soil seed bank. (B) The gray panel contains a graphical representation of the viability trials. Seeds were tested in two rounds; germination trials were performed and then some or all of the ungerminated seeds were tested for viability. The graph below the panel shows hypothetical data from a series of viability trials and the interpolated, inferred viabilities at times when viability was unobserved. (C) Age-specific germination probably is summarized in three ways. (D) The graph shows the survival function for persistence of seeds in the soil seed bank (black line) and the estimated discrete survival probabilities for persistence and viability of seeds (orange points).

$$\theta_{jk} = \theta_{0,k} + \epsilon_{(jk)}. \tag{2}$$

Our model includes a population-level intercept  $\theta_{0,k}$  and random effects  $\epsilon_{(jk)}$ . The random

244

effects can be written as  $\epsilon_{(jk)} \sim N(0, \varsigma^2)$ . For the moment, we focus on describing the hierarchical structure of the model but note that we use link functions for transformation to parameters that are appropriate for the likelihoods we use to model different sets of observations (e.g. binomial for seed bag experiments; Poisson for counts of seed per fruit). We note that such a linear mixed effects model with random intercepts for years is one method commonly used to model interannual variation in demographic rates (e.g. Metcalf et al. 2015). Using hierarchical centering, the same model is rewritten as

$$\theta_{jk} = \alpha_{(jk)}. (3)$$

The mean  $\theta_{jk}$ , is now drawn from a normal distribution with mean  $\alpha_{(jk)}$  and variance  $\sigma_j^2$ . We place a prior on  $\alpha_{(jk)}$  such that  $\alpha_{(jk)} \sim N(\theta_{0,k}, \varsigma^2)$ . The expressions are related by  $\alpha_{(jk)} = \theta_{0,k} + \epsilon_{(jk)}$ . We thus draw year-level means from the population-level means.

For a single population (ie. suppressing subscript k), we write the posterior proportional to the joint distribution as

$$[\theta_i, \theta_0, \sigma_i^2, \varsigma^2 | y_{ii}] \propto [y_{ii}|\theta_i, \sigma_i^2] [\theta_i|\theta_0, \varsigma^2] [\theta_0] [\sigma_i^2] [\varsigma^2]. \tag{4}$$

The distribution of the observations  $y_{ij}$  is conditional on the year-specific parameters  $\theta_j$  and  $\sigma_j^2$ . In turn, the year-specific parameter  $\theta_j$  is conditional on the populationspecific parameters  $\theta_0$  and  $\varsigma^2$ . We placed priors on all parameters found only on the

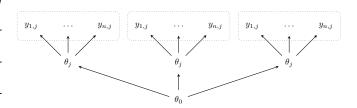


Figure 4: Graph depicting the general structure for the hierarchical models, for one population. Observations from each year,  $y_{ij}$ , are shown grouped and outlined by dotted lines. The observations are drawn from year-level palameters,  $\theta_j$ , which in turn are drawn from a population-level parameter,  $\theta_0$ . The graph omits variance terms.

right hand side of conditional statements  $(\theta_0, \sigma_i^2, \varsigma^2)$ . In practice, we implemented 264 this model by specifying the population- and 265 year-levels of the model with normal distri-266 butions; for example,  $[\theta_j|\theta_0,\varsigma^2]$  is  $\theta_j \sim N(\theta_0,\varsigma^2)$ . The model thus describes a structure in 267 which years are nested within populations. 268

#### 1.4 Model statements, implementation, and fitting

We include the expression for the posterior proportional to the joint distribution, and corresponding directed acyclic graphs, in Appendix: Joint Posterior. Priors for all parameters are defined in Table Priors. We applied the following principles for specifying priors: (1) we used weakly informative priors that avoided placing probability mass on biologically implausible values (Gelman; Lemoine; Wesner and Pomeranz), (2) we placed positive, unbounded priors 273 on variance components (REF), (3) we conducted prior predictive checks to assess the scale 274 of priors after parameter transformation (Hobbs and Hooten; Gabry; Wesner and Pomer-275 anz), and (4) we simulated prior predictive distributions to confirm that the joint likelihood 276 generated data within the observed range (Gabry; Conn; Hobbs and Hooten). We provide 277 additional detail regarding our choice of priors in Appendix: Priors. 278 We prepared data for analysis using the tidyverse and tidybayes packages (CITE) in R 279 VERSION; CITE. We wrote, fit all models, and estimated posterior distributions using 280 JAGS VERSION with rjags (Plummer 2016). We randomly generated initial conditions for 281 all parameters with a prior by drawing from the corresponding probability distribution in R 282 before passing the initial values to right we ran three chains for XX,000 iterations. The first 283 XX,000 samples were discarded as burn-in and we sampled the following XX,000 iterations. 284 We did not thin the chains (Elderd and Miller 2016).

285

We assessed convergence of the MCMC samples with visual inspection of trace plots, by calculating the Brooks-Gelman-Rubin diagnostic (R-hat), and by calculating the Heidelberg-Welch diagnostic (Elderd and Miller 2016. The Gelman-Rubin diagnostic is used to assess convergence between chains and the Heidelberg-Welch for stationarity within chains. Trace plots for all chains, histograms of R-hat, and the percentage of chains that passed the HW test are shown in the appendix.

To evaluate our model's fit to the data, we performed model checks that are described in 292 full in Appendix: Model Checking. We used our posterior distribution to simulate replicate 293 datasets based on the parameters of our model. We compared samples from the simulated 294 datasets to the real, observed datasets using both graphical, visual checks and by calculating 295 Bayesian p-values for test statistics calculated for the observed and simulated data. In the 296 following section, we describe how we used the models we fit to obtain the parameters that 297 describe the Clarkia life history. While we do not perform model checks for these derived 298 quantities (e.g. winter seed survival accounting for the combined effect of seed decay and 299 loss of viability) because we combine the output of multiple models, the model checks are 300 still essential to determine whether our inferences are reasonable. 301

# 1.5 Computing vital rates

#### 1.5.1 Belowground vital rates - seed bank paper

We used the age-specific germination probabilities, survival function, and viability estimates to account for viability in estimates for the probability of germination and survival. We first discretized the survival function to times at which we observed germination and counted seeds (January and October). Estimates of survival over these intervals are the probability that a seed remains intact, but does not account for loss of viability. Next, we used viability estimates from October to calculate viability for January by interpolation (Figure 3D). We

tested the viability of seeds in October, and were thus able to estimate the proportion of viable seeds (Figure 3B; filled points). We inferred the viability of intact seeds in January 309 by assuming that seeds lost viability at a constant rate (exponential decay). Further, we 310 interpolated between estimates by assuming that viability changed at a constant rate between 311 years, and that all seeds were viable at the start of the experiment (Figure 3B; open points). 312 We combined the discretized survival function and viability estimates to construct a 313 survival function for the probability that a seed remains intact and viable (Table 3, col-314 umn X). Specifically, we multiplied the posteriors of the discretized survival and viability 315 estimates. Because we combined estimates, some portions of the posterior for seed survival 316 probability was than 1, especially for later seed ages. We restricted the posterior to be less 317 than 1 by truncating the distribution and resampling to redistribute the probability mass. 318 We take this step to retain parameter uncertainty about survival probability in cases where 319 combining the estimates implies a high probability of survival. The survival function for 320 viable seeds  $(\phi)$  is composed of estimates of persistence over time  $(\theta)$ , estimates of viability 321  $(\nu)$ , and estimates of germination conditional on persistence  $(\gamma)$ . 322

We used the discretized survival function and age-specific germination probability to 323 obtain the estimates of germination and seed survival required to test predictions from bethedging theory. Table ?? defines the age-specific germination probabilities and survival probabilities for the structured model in Eckhart et al. 2011 in terms of the survival function 326 and age-specific germination probabilities. Figure 3E-F illustrate the relationship among 327 the various probabilities of germination and seed survival. Estimates from the seed bag 328 experiment correspond to the probability of germination or survival conditional on persis-329 tence (e.g.  $\gamma_1$ ). Multiplying these estimates by the probability of persistence up to a certain 330 time gives the unconditional probability (e.g.  $\theta_1 \times \gamma_1$ ). Finally, the probability conditional 331 on persistence and viability is estimated by incorporating loss of viability into the survival 332 function (e.g.  $\gamma_1/\phi_1$ ), and defines the parameters in the structured population model.

Table 3: Seed persistence and viability in the soil seed bank

	Persistence	Persistence & viability
Time $(x_i)$	$S(x_i)$	$S(x_i)$
$\operatorname{Oct}_0$	$\theta_0$	$\phi_0 = \theta_0$
$\mathrm{Jan}_{1,\mathrm{total}}$	$ heta_1$	$\phi_1 = \theta_1(\gamma_1 + (1 - \gamma_1)\nu_1^{1/3})$
$Jan_{1,intact}$	$\theta_2$	$\phi_2 = \theta_2 \nu_1^{1/3}$
$\operatorname{Oct}_1$	$\theta_3$	$\phi_3 = \theta_3 \nu_1$
$Jan_{2,total}$	$ heta_4$	$\phi_4 = \theta_4(\gamma_2 + (1 - \gamma_2)\nu_1(\nu_2/\nu_1)^{1/3})$
Description	Parameter	Probability
July-October	$s_0$	
October-January	$s_1$	$\mid \phi_1 \mid$
1-year old germination	$g_1$	$\gamma_1/\phi_1$
January-October	$s_2$	$\phi_3/\phi_2$
October-January	$s_3$	$\phi_4/\phi_3$

#### 1.5.2 Per-capita reproductive success

334

In order to make our analysis comparable to previous empirical studies of bet hedging,
we calculated per-capita reproductive success as the product of the probability of seedling
survival to fruiting, fruits per plant, and seeds per fruit. We thus calculate per-capita
reproductive success as the number of seeds produced per seedling, on average (e.g. Venable
2007, Gremer et al. 2014).

We used a consistent method to estimate seedling survival to fruiting throughout the experiment, and use the population- and year-level means  $(\mu_{S,jk})$  in our calculation. Because we estimated fruit production in 2 different ways during the study, we chose to use total fruit equivalents (TFE) per plant as our common estimate of fruit production. From 2006–2012, we used  $\mu_{TFE,jk}$ ) as estimated in the statistical model. From 2013–2018, we used the ratio of seeds per damaged to undamaged fruit to calculate a proportion of damaged fruits to add to undamaged fruit counts, as in

TFE = undamaged fruits + 
$$\frac{\text{seeds per damaged fruit}}{\text{seeds per undamaged fruit}} \times \text{damaged fruits}.$$
 (5)

We used posterior distributions for population- and year-level parameters (e.g.  $\mu_{{\rm US},jk}$ ) for these calculations and obtained estimates of  $\mu_{{\rm TFE},jk}$ ) for 2013–2018. Finally, we used estimates of seeds per undamaged fruit ( $\mu_{{\rm US},jk}$ ) as our estimate of seeds per fruit.

In terms of parameters from our statistical models, per-capita reproductive success  $F_{jk}$  at population j in year k is calculated as

$$F_{ik} = \phi_{ik} \times \lambda_{\text{TFE},ik} \times \lambda_{\text{US},ik}, \tag{6}$$

where

$$\phi_{jk} = \text{logit}^{-1}(\mu_{S,jk})$$

$$\lambda_{\text{TFE},jk} = \exp(\mu_{\text{TFE},jk})$$

$$\lambda_{\text{US},jk} = \exp(\mu_{\text{US},jk}).$$
(7)

Our multilevel models for aboveground vital rates pooled data more strongly in years
with relatively little data. A benefit of this approach is that it implicitly corrects for variation in sample size (e.g. an observation of 0/37 seeds surviving is given more weight than an
observation of 0/1 seeds surviving). While this is beneficial for distinguishing between spurious estimates and true temporal variation in reproductive success, it may also underestimate
variation in reproductive success. At the extreme, estimates in years without any data are
pooled to the population-level means. Years with zero seedling survivorship would thus have

estimates for fruits per plant that are pooled towards the population-mean (because there were no fruiting plants on which to count fruits).

We calculated the posterior mode of annual estimates for each parameter in Equation

6 before multiplying to obtain the per-capita reproductive success in that year. Using the

posterior mode is equivalent to taking the BLUP of a linear model, and allowed us to estimate

vital rates in years with small sample sizes.

# **Analysis**

Strict and less strict tests of the bet-hedging model. We consider two models for per-capita reproductive success. In the first, we use partial pooling to correct for sampling bias in estimates of seedling survival, fruits per plant, and seeds per fruit. However, our model with partial pooling pools years with few plants towards the overall population mean, which will reduce the variance in per-capita reproductive success. We thus also considered a second 370 model in which we did not pool to the population-level. In this model, we instead estimated 371 seedling survival, fruits per plant, and seeds per fruit each year separately and did not include 372 a population-level effect (in other words, we did not nest year in population). This would 373 have the effect of letting the prior have a stronger effect each year. We conducted model 374 checks for both of these. Years without data would be missing or true NAs. Finally, we could 375 also consider a model without pooling and in which the observed estimate is uncorrected. 376

#### Correlation between germination probability and seed survival

Increased seed survivorship is predicted to decrease the optimal germination probability
Cohen (1966); Ellner (1985a). I assessed whether the observed germination probability was
negatively correlated with seed survival (Gremer and Venable (2014)). I calculated the
probability that seeds which do not germinate in January remain in the seed bank until the

following January  $(s_2s_3)$ . I obtained the posterior distribution for the correlation between germination and seed survival by calculating the correlation of  $g_1$  and  $s_2s_3$  at each iteration 382 of the MCMC output Hobbs and Hooten 2015, p 194-5. Calculating the sample correla-383 tion in draws from the posterior. (quote Stan list-serv). Results of this analysis are shown 384 in Figure 8. Bet hedging models predict that germination probability should be negatively 385 correlated with seed survival; 95% credible intervals that do not overlap zero provide support 386 for this prediction. The bottom panel shows the posterior distribution of correlation between 387 the probability of germination and seed survival. The median correlation is negative (-0.07) 388 and the 95% credible interval overlaps 0. 389

Ref: https://discourse.mc-stan.org/t/computing-correlations-from-the-posterior/2633

# Correlation between germination probability and variance in per-capita reproductive success

Increased variance in per-capita reproductive success is predicted to decrease the optimal germination probability (Cohen (1966); Ellner (1985a)). I assessed whether the observed germination probability was negatively correlated with variance in per-capita reproductive success (Venable (2007)). Per-capita reproductive success  $F_{jk}$  at population j in year k was calculated as:

$$F_{jk} = \phi_{jk} \times \lambda_{jk}^F \times \lambda_{jk}^P \tag{8}$$

where

390

$$\phi_{jk} = \operatorname{logit}^{-1}(\alpha_{0,j}^{S} + \beta_{jk}^{S})$$

$$\lambda_{jk}^{F} = \exp(\alpha_{0,j}^{F} + \beta_{jk}^{F})$$

$$\lambda_{jk}^{P} = \exp(\alpha_{0,j}^{P} + \beta_{jk}^{P})$$
(9)

To calculate the temporal variation in per-capita reproductive success for each population, I sampled the posterior distribution of reproductive success for each year and calculated the 392 geometric SD of per capita reproductive success. I obtained the posterior distribution for 393 the correlation between germination and geometric SD of per capita reproductive success 394 at each iteration of the MCMC output Hobbs and Hooten 2015, p 194-5. Calculating the 395 sample correlation in draws from the posterior. (quote Stan list-serv). Using this approach, I 396 obtained a distribution of correlation estimates. Bet hedging models predict that germination 397 probability should be negatively correlated with temporal variance in fitness; 95% credible 398 intervals that do not overlap zero provide support for this prediction. Results of this analysis 399 are shown in Figure 9. 400

#### Density-independent model for germination probability

406

We use estimates of seed survival and reproductive success to investigate the adaptive value of delayed germination (Gremer and Venable (2014)). We parameterize a model of population growth rate and calculate the optimal germination strategy for different combinations of seed survival and reproductive success. We describe *C. xantiana*'s life cycle and calculate population growth rate with the equation:

$$\lambda = g_1 Y(t) s_0 s_1 + (1 - g_1) s_2 s_3 \tag{10}$$

The parameters in this equation were fit in models corresponding to equations (??), (??), and (??). Seed survival rates  $(s_0, s_1, s_2, s_3)$  are population-level estimates. Per capita reproductive success (Y(t)) is calculated as the product of seedling survival to fruiting, fruits per plant, and seeds per fruit (equation (8)). Temporal variation is incorporated into the model by varying the per-capita reproductive success, Y(t), between years.

For each population, I numerically calculate the optimal germination probability for

the observed variation in reproductive success and seed survival. In each case, I sampled parameters in the equation for density-independent growth (equation (10)) from the fitted 408 vital rate distributions. I resampled the fitted values of per-capita reproductive success (Y(t))409 to obtain a sequence of 1000 years. I used this same sequence of Y(t) and the fitted seed 410 survival probabilities to calculate long-term stochastic population growth rates  $(\lambda_s)$  at each 411 germination probability along an evenly spaced grid of possible germination probabilities 412 (G) between 0 and 1. The optimal germination probability is estimated as the value of G413 that maximizes the geometric mean of the population growth rate. I repeat the simulations 414 50 times for each population, resampling the sequence of per-capita reproductive success, 415 Y(t), each time. I then calculated the mean of the optimal germination fractions. 416

Models in which per-capita reproductive success is density-independent predict that germination probability should respond to variance in fitness (Cohen (1966)). To evaluate
the density-independent model, I compared modeled germination probabilities to predicted
germination optima. I plot this comparison in Figure 10. The dotted line indicates a 1:1
relationship between observations and predictions. Values below the line indicate that the
model predicts higher germination probabilities than observed; values above the line would
indicate that the model predicts lower germination probabilities than observed.

#### Density-dependent model for germination fraction

consequences of a germination strategy for an individual?s fitness depend on the strategies
being used by other individuals in the population (Gremer and Venable 2014).

Here is what I think the general strategy would be if I wanted to test this model. Because
we don't have data on all the species in the plot, we are solely focusing on the strength of
intraspecific competition, which may vary with regards to how good of a proxy for overall competition the seedling experiences. In years with high grass germination there may
be strong competition from other species (for example). Our data on seeds per fruit comes

from haphazard collections of fruits; there is no information in density-dependence in this
estimate. Our data on fruits per plant could be informed by the number of seedlings or
adult plants in the plot. However, then we are getting one step removed from where the
competition happens (among seedlings). We thus start by estimating seedling survival to
fruiting as a function of density, assuming that this is the stage at which density-dependence
is strongest.

-We use counts of seedlings in the plot to incorporate the number of seedlings into our model for seedling survival to fruiting. We assume 'low density' is a single plant, and so obtain an estimate for the probability of survival at low densities as the marginal posterior probability of survival in a plot with a single plant. In the end, we obtain an estimate of seedling survival per fruit at low density in a given year (*K* in Gremer and Venable) and a competition coefficient (from our logistic regression).

#### Age-structured model for germination fraction

Valleriani and Tielborger build on Tuljaparkur and Easterling/Ellner to show that an agestructured seed bank can modify the expectations for how dormancy should evolve. Here,
we would want to show that there is (1) age-structure in the seed bank and (2) that the
importance of age structure varies across the species range.

# Results

### Correlation between germination probability and seed survival

I examined the correlation between germination probability and seed survival in the seed bank. Results of this analysis are shown in Figure 8. The bottom panel shows the posterior distribution of correlation between modeled germination probability and the probability of seed survival; the median correlation is negative (-0.07) and the 95% credible interval overlaps 0. There is no correlation between germination and seed survival.

Correlation between germination probability and variance in per-capita repro-

ductive success

<sup>452</sup> I examined the correlation between germination probability and variance in per-capita repro-

ductive success. Results of this analysis are shown in Figure 9. The bottom panel shows the

posterior distribution of correlation between modeled germination probability and geometric

SD in per-capita reproductive success; the median correlation is positive (0.28) and the 95%

credible interval overlaps 0.

Optimal germination probability predicted by a density-independent model

157 ...

# Discussion

458 ...

# Figures

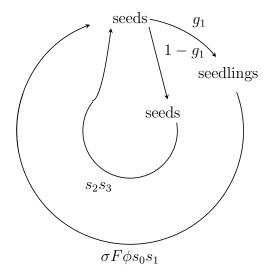


Figure 5: Life cycle diagram for *Clarkia xantiana*. I would like to draw a better life cycle diagram.

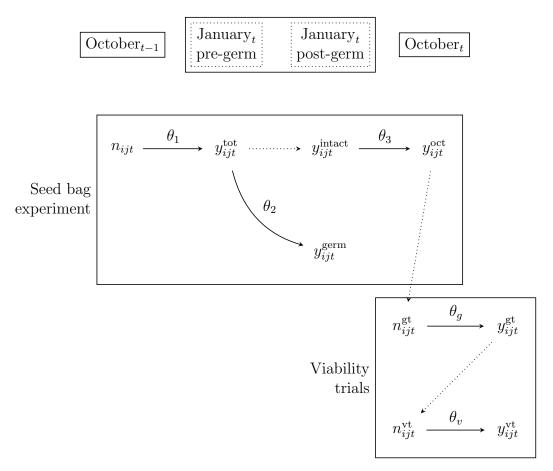


Figure 6: Diagram of data from the seed bag experiments and viability trials. There are two boxes: one for the seed bag experiment and one for the viability trials. In the seed bag experiment, I split January into two steps, one for just before germination and one for just after. Solid arrows represent probabilities estimated with a binomial experiment and are labeled with corresponding parameters. Dotted arrows represent cases where the seeds at the head of the arrow include some, possibly all, seeds at the tail of the arrow.

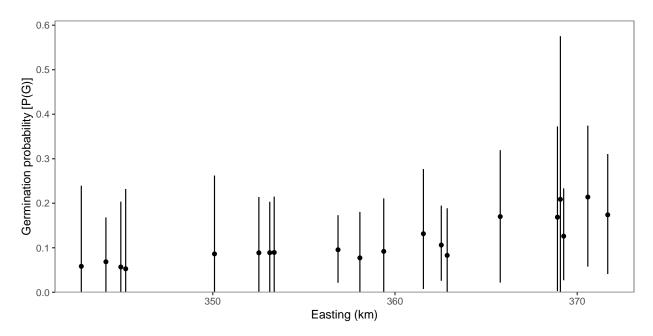


Figure 7: Germination probability plotted against easting (km). The plot shows the marginal posterior distribution for germination probability at each site. The points are the median of the posterior. The thinner line represents the 95% credible interval and the thicker line represents the 50% credible interval.

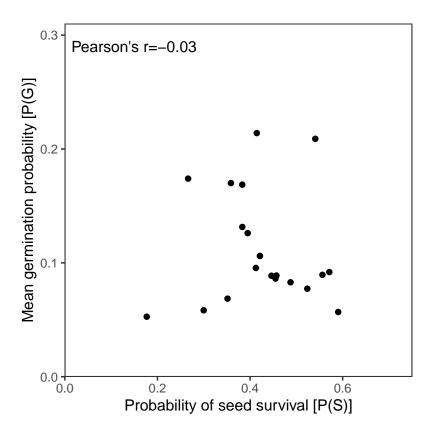


Figure 8: The top panel shows the observed germination probability plotted against probability of seed survival. The bottom panel shows the posterior distribution of correlation between observed germination probability and the probability of seed survival; the correlation is negative (-0.03).

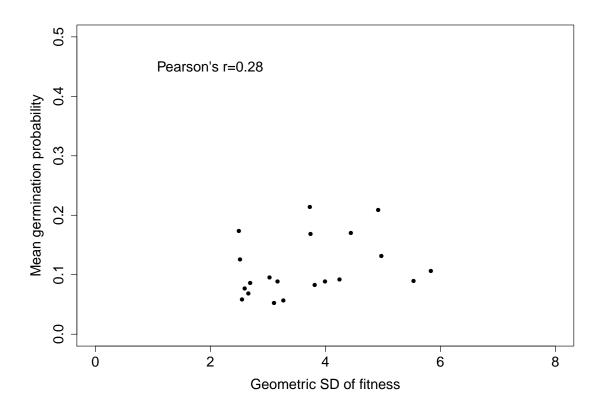


Figure 9: The top panel shows the observed germination probability plotted against the temporal variation in per capita reproductive success. The bottom panel shows the posterior distribution of correlation between observed germination probability and geometric SD of per capita reproductive success; the median correlation is negative (-0.16) but the 95% credible interval overlaps 0.

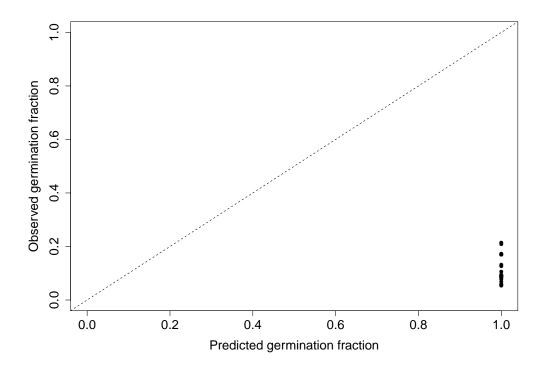


Figure 10: Observed germination probability plotted against the optimal germination probability predicted by a density-independent model. For each population, the observed germination probability is the obtained from the model for seed bank vital rates. Each point is the population-specific median of the posterior of  $g_1$  for a model fit to data from seed bag experiments from 2006–2009. Data was pooled across years. The dotted line indicates a 1:1 relationship between observations and predictions. Values below the line indicate that the model predicts higher germination probabilities than observed; values above the line would indicate that the model predicts lower germination probabilities than observed.

# References

- Cohen, D. 1966. Optimizing reproduction in a randomly varying environment. Journal of
  Theoretical Biology, **12**:119–129.
- 461 Cohen, D. 1967. Optimizing Reproduction in a Randoruly Varying Environment when a
- 462 Correlation May Exist between the Conditions at the Time a Choice has to be Made and
- the Subsequent Outcome. Journal of Theoretical Biology, **16**:1–14.
- Cohen, D. 1968. A General Model of Optimal Reproduction in a Randomly Varying Envi-
- ronment. The Journal of Ecology, **56**:219.
- Eager, E. A., R. Rebarber, and B. Tenhumberg. 2014. Modeling and Analysis of a Density-
- Dependent Stochastic Integral Projection Model for a Disturbance Specialist Plant and
- Its Seed Bank. Bulletin of Mathematical Biology, **76**:1809–1834.
- Easterling, M. R. and S. P. Ellner. 2000. Dormancy strategies in a random environment:
- 470 Comparing structured and unstructured models. Evolutionary Ecology Research, 2:387–
- 407.
- Eckhart, V. M., M. A. Geber, W. F. Morris, E. S. Fabio, P. Tiffin, and D. A. Moeller.
- 2011. The Geography of Demography: Long-Term Demographic Studies and Species
- Distribution Models Reveal a Species Border Limited by Adaptation. The American
- Naturalist, **178**:S26–S43.
- Ellner, S. 1985a. ESS germination strategies in randomly varying environments. I. Logistic-
- type models. Theoretical Population Biology, **28**:50–79.
- Ellner, S. 1985b. ESS germination strategies in randomly varying environments. II. Recip-
- rocal Yield-Law models. Theoretical Population Biology, **28**:80–116.

- Evans, M. E. K. and J. J. Dennehy. 2005. Germ Banking: Bet-Hedging and Variable Release
  From Egg and Seed Dormancy. The Quarterly Review of Biology, 80:431–451.
- Gremer, J. R., S. Kimball, and D. L. Venable. 2016. Within-and among-year germination in Sonoran Desert winter annuals: bet hedging and predictive germination in a variable
- environment. Ecology Letters, **19**:1209–1218.
- Gremer, J. R. and D. L. Venable. 2014. Bet hedging in desert winter annual plants: optimal germination strategies in a variable environment. Ecology Letters, 17:380–387.
- Heinrich, L., J. Müller, A. Tellier, and D. Živković. 2018. Effects of population- and seed bank size fluctuations on neutral evolution and efficacy of natural selection. Theoretical Population Biology, **123**:45–69.
- Mccue, K. A. and T. P. Holtsford. 1998. Seed bank influences on genetic diversity in the rare annual Clarkia springvillensis (Onagraceae). American Journal of Botany, 85:30–36.
- Nunney, L. 2002. The Effective Size of Annual Plant Populations: The Interaction of a Seed
  Bank with Fluctuating Population Size in Maintaining Genetic Variation. The American
  Naturalist, **160**:195.
- Paniw, M., P. F. Quintana-Ascencio, F. Ojeda, and R. Salguero-Gómez. 2017. Accounting for uncertainty in dormant life stages in stochastic demographic models. Oikos, **126**:900–909.
- Ritland, K. 1983. The joint evolution of seed dormancy and flowering time in annual plants living in variable environments. Theoretical Population Biology, **24**:213–243.
- Venable, D. L. 2007. Bet hedging in a guild of desert annuals. Ecology, 88:1086–1090.
- Waples, R. S. 2006. Seed Banks, Salmon, and Sleeping Genes: Effective Population Size in Semelparous, Age-Structured Species with Fluctuating Abundance. The American Naturalist, **167**:118.

# Supplementary material

## Theoretical background for hypotheses.

```
Explanation of key papers that develop theoretical results about seed banks. The docu-
ment describes results from these papers that are relevant to understanding and interpreting
the data in this manuscript. Link to document: https://github.com/gregor-fausto/
clarkiaSeedBanks/blob/master/products/appendices/appendix-cohen-results/appendix-x-cohen-
pdf
```

# Data summary.

Summary tables for all datasets used in the manuscript. The document summarizes the types
of data collected. The document provides a table summarizing each dataset (e.g. sample
size per each site and year). Link to document: https://github.com/gregor-fausto/
clarkiaSeedBanks/blob/master/products/tables/data-summary.pdf

# Data processing workflow.

Description of workflow for processing the data used in the analysis. The document describes how comma-separated value (.csv) and Excel (.xls and .xlsx) files were read and processed in R. Link to document: https://github.com/gregor-fausto/clarkiaSeedBanks/blob/
master/library/dataProcessingWorkflow.md

# Method for estimating seed bank parameters using conditional probabilities.

The document explains how we compose conditional probabilities to calculate probabilities of survival and germination of seeds in the seed burial experiment. Link to document: https://

github.com/gregor-fausto/clarkiaSeedBanks/blob/master/products/appendices/appendix-condi

appendix-x-conditional-probability.pdf