**BIO609**: Bash scripting. Bash is a unix shell and command language. It is mostly the default login shell (the shell you work with when you login to a Unix machine).

**Exercise 1: writing and executing a shell script**

Try to copy/paste the below simple bash script into a file and execute it. The hello world bash script:

#!/bin/bash  
echo "Hello World"

Copy the above 2 lines and save them to the file “hello.sh”. The first line tells the Unix shell to interpret the program. In order to run the program you still need to make the file "executable" (x flag).

Make your file executable by typing:

$ chmod +x hello.sh

And now you can simply type:

$ ./hello.sh

Note the “./” at the start of the command. This is because the directory where we stored hello.sh is not in the system variable $PATH.

 **Exercise 2: write a simple bash script with a for loop**

Often, you would like to run the same command with different parameters. As an exercise, write a simple bash script that will output numbers from 1 to 100. Use a for loop.

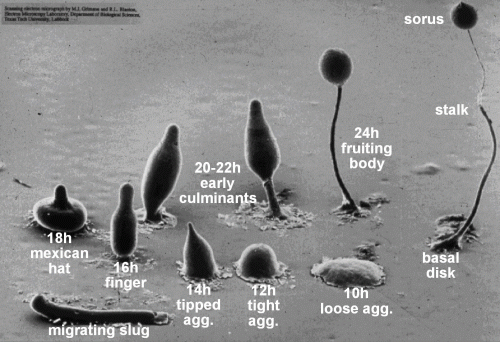
#!/bin/bash  
for i in {1..100}  
do  
 echo $i  
done

Save the above code to a file (e.g. script.sh), make the file executable (+x flag) and run it.

What is the output?

 **Exercise 3: download the Dictyostelium discoideum (dd) genome in FASTA format**

*Dictyostelium discoideum* ([www.dictybase.org](http://www.dictybase.org)) is an interesting social amoeba and a well studied model organism. The amoeba lives in the soil, and when nutrients run out, 100K amoebas aggregate into a fruiting body. The stalk amoebas "sacrifice" themselves and the sorus is blown away by the wind to a new location with more nutrients (hopefully).



You can download the **dd genome FASTA** file directly from our server:

wget https://bioinfo.evolution.uzh.ch/share/bio609/dicty/dd.fasta

The FASTA format is widely used in sequence distribution, see the description at <http://en.wikipedia.org/wiki/FASTA_format>.

Use **grep** to find out how many chromosomes are present in the FASTA file. Then use **grep** (-v) to only print out the genomic sequence (without chromosome identifiers). How large is the genome?

**Exercise 4: searching for short sequences in the *Dictyostelium discoideum* genome**

Is the sequence “AAAAAGAGATACAT” present in the DD genome (dd.fasta)?

You can simply use **grep** to find out.

What is the downside of this approach (using seach / grep to find short sequences) and why do we use short-read aligners like STAR, bowtie and others?

 **Exercise 5: handling RNA-seq data**

Next-generation (short-read) sequencing data is usually stored in FASTQ files:

<https://en.wikipedia.org/wiki/FASTQ_format>

FASTQ files are similar to FASTA files, but they also contain sequence quality.

We will work with 10 FASTQ files (10 samples) containing sequences from Dictyostelium discoideum RNA (RNA-seq).

Write a short script to download \*.fastq.gz files from:

https://bioinfo.evolution.uzh.ch/share/bio609/dicty

Open the link in a browser and you should see 10 FASTQ files. Instead of clicking and downloading each file separately, write a script to download the files.

for sample\_id in {1..10}

In the for loop, you would need to adjust the name of the file and use command **wget** to download the files.

 **Exercise 6: map the RNA-seq data to the reference genome**

We will now try to use STAR and map our 10 samples of RNA-seq data to the reference genome. First, you need to create an "index" for the dd.fasta reference genome. You can do this with:

mkdir **istar** # folder for genome index  
STAR --runMode genomeGenerate --genomeDir **istar** --genomeFastaFiles **dd.fasta**

The command should take around 1-2 minutes to finish. The index is now stored in the folder **istar**.

Then you can write a script to map the RNA-seq data to the reference genome using the newly created index above.

An example command to map sample1 would be:

STAR --genomeDir istar --readFilesIn sample1.fastq.gz --readFilesCommand zcat

After finishing, this creates a SAM file (alignments) named **Aligned.out.sam**. For each of the 10 samples, rename this file into sample**n**.sam (where n=1,2,3..10) and convert the SAM file to a BAM file.

The **"pipeline" for sample1** could look something like this:

mv Aligned.out.sam sample1.sam  
samtools view -S -b sample1.sam > sample1.bam # convert sam to bam  
samtools sort sample1.bam sample1 # sort bam file  
samtools index sample1.bam # index bam file  
rm sample1.sam # remove sample1.sam, since we don't need it anymore

Write a short script to repeat the above steps for all the 10 samples (sample1, sample2, ... sample10). Of course here comes the power of bash, simply use a for loop.

 **Exercise 7: download genome annotation in GFF format and count reads aligned to genes**

Download the GTF file (genome annotation) for *Dictyosrtelium discoideum* from:

wget https://bioinfo.evolution.uzh.ch/share/bio609/dicty/dd.gtf  
htseq-count -f bam sample1.bam dd.gtf > sample1.tab

Repeat this for all the samples, so in the end you should get 10 tab files with gene counts.

**\*** **Exercise 8: combine gene expression tables into one single table**

This one is tricky, but you can use awk:

awk 'NF > 1{ a[$1] = a[$1]"\t"$2} END {for( i in a ) print i a[i]}' \*.tab > samples.tab

Stores the big table into the file **samples.tab**.

**\*** **Exercise 9: try to download and compile bowtie2, make an index of the genome and map the reads to the reference genome, count the reads aligning to genes and create the samples.tab table. Compare the STAR to the bowtie2 mappings.**

**System information, processes and other useful commands**

|  |  |
| --- | --- |
| **uname -a** | display system information |
| **man** *command* | display manual page of command |
| **df -h** | list mounted disks with available space |
| **du -h** *path* | show space usage |
| **top** | display running processes |
| **kill -9** *pid* | kill process |

**File and folder manipulation, compression**

|  |  |
| --- | --- |
| **pwd** | display current folder |
| **ls -l** *path* | list files and folders |
| **cd** *path* | change folder to path |
| **cd** ~ | change folder to home folder |
| **mkdir** *name* | make folder |
| **rmdir** *name* | remove folder |
| **cp** *source* *dest* | copy file/folder and all its contents |
| **less** *filename* | display file content |
| **wc** *filename* | count number of lines in file |
| **head** *filename* | shows first few lines of file |
| **tail** *filename* | shows last few lines of file |
| **gzip** *filename* | compress file with gzip (adds .gz extension) |
| **gunzip** *filename* | decompress filuncompress and remove .gz extension |
| **tar xfz** *filename.tar.gz* | uncompress files from tar.gz archive |
| **tar** **zcvf** *archive.tar.gz folder\_to\_compress* | creates archive.tar.gz |
| **unzip** filename.zip | unzip archive |

**Network and file transfer**

|  |  |
| --- | --- |
| **wget** URL | download file (html page) and save to current folder |
| **ssh** *username@host* | remote login to host with username |
| **sftp** username@host | remote login to host with username and transfer files |

**“vi” editor**

|  |  |
| --- | --- |
| **$ vi** *filename* | start editing file with vi |
| **i** | switch to “insert” mode |
| **ESC** | switch to “command” mode |
| **:w** | save |
| **:q** | quit |
| **:x** | save and quit |
| **/<pattern>** | search for pattern, <n> gives you the next match |
| **:q!** | quit without saving changes |