

Assignment 3: Data Exploration

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Spring 2023

OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse, lubridate), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
setwd("C:/Users/17038/Documents/EDA Spring 2023/")
getwd()
```

```
## [1] "C:/Users/17038/Documents/EDA Spring 2023"
```

```
library(tidyverse)
library(lubridate)
```

```
Neonics <- read.csv("../Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv", stringsAsFactors = TRUE)
Litter <- read.csv("../Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv", stringsAsFactors = TRUE)
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Decreasing populations of pollinators is a pressing problem, and it's been established that insecticides contribute to excess pollinator death. Makes sense that we would want to look at the one of the most widely used classes of insecticides and their toxicological impact on insects.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Forest litter influences the temperature, duration, and veracity of wildfires, so studying forest litter would give us a better idea of how a wildfire will act in a given area. Seeing how wildfires have plagued western states recently, I can imagine folks in CO are interested in what will fuel the next fire.

4. How is litter and woody debris sampled as part of the NEON network? Read the `NEON_Litterfall_UserGuide.pdf` document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. The locations of tower plots, where woody debris sampling takes place, is randomly selected. 2. Trap placement within plots is either targeted or randomized, depending on vegetation. In sites with more than 50% aerial cover of woody vegetation greater than 2m in height, trap placement is randomized. In sites with less than 50% cover of woody vegetation or heterogeneously distributed, patchy vegetation, trap placement is targeted. 3. Ground traps are sampled once per year, and elevated traps are sampled at different frequencies depending on what vegetation is present.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Neonics)
```

```
## [1] 4623 30
```

6. Using the `summary` function on the "Effect" column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
summary(Neonics$Effect)
```

##	Accumulation	Avoidance	Behavior	Biochemistry
##	12	102	360	11
##	Cell(s)	Development	Enzyme(s)	Feeding behavior
##	9	136	62	255
##	Genetics	Growth	Histology	Hormone(s)
##	82	38	5	1
##	Immunological	Intoxication	Morphology	Mortality
##	16	12	22	1493
##	Physiology	Population	Reproduction	
##	7	1803	197	

Answer: The two most commonly studied effects are Population and Mortality. These effects would be of particular interest because pesticide is known to harm insect populations, particularly pollinator populations. I imagine scientists are trying to figure out exactly how pollinators are affected by this class of pesticides.

7. Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: The `sort()` command can sort the output of the summary command...]

```
summary(Neonics$Species.Common.Name) %>%
  sort(decreasing = TRUE)
```

##	(Other)	Honey Bee
##	670	667
##	Parasitic Wasp	Buff Tailed Bumblebee
##	285	183
##	Carniolan Honey Bee	Bumble Bee
##	152	140
##	Italian Honeybee	Japanese Beetle
##	113	94
##	Asian Lady Beetle	Euonymus Scale
##	76	75
##	Wireworm	European Dark Bee
##	69	66
##	Minute Pirate Bug	Asian Citrus Psyllid
##	62	60
##	Parastic Wasp	Colorado Potato Beetle
##	58	57
##	Parasitoid Wasp	Erythrina Gall Wasp
##	51	49
##	Beetle Order	Snout Beetle Family, Weevil
##	47	47
##	Sevenspotted Lady Beetle	True Bug Order
##	46	45
##	Buff-tailed Bumblebee	Aphid Family
##	39	38
##	Cabbage Looper	Sweetpotato Whitefly
##	38	37
##	Braconid Wasp	Cotton Aphid
##	33	33
##	Predatory Mite	Ladybird Beetle Family

##	33	30
##	Parasitoid	Scarab Beetle
##	30	29
##	Spring Tiphia	Thrip Order
##	29	29
##	Ground Beetle Family	Rove Beetle Family
##	27	27
##	Tobacco Aphid	Chalcid Wasp
##	27	25
##	Convergent Lady Beetle	Stingless Bee
##	25	25
##	Spider/Mite Class	Tobacco Flea Beetle
##	24	24
##	Citrus Leafminer	Ladybird Beetle
##	23	23
##	Mason Bee	Mosquito
##	22	22
##	Argentine Ant	Beetle
##	21	21
##	Flatheaded Appletree Borer	Horned Oak Gall Wasp
##	20	20
##	Leaf Beetle Family	Potato Leafhopper
##	20	20
##	Tooth-necked Fungus Beetle	Codling Moth
##	20	19
##	Black-spotted Lady Beetle	Calico Scale
##	18	18
##	Fairyfly Parasitoid	Lady Beetle
##	18	18
##	Minute Parasitic Wasps	Mirid Bug
##	18	18
##	Mulberry Pyralid	Silkworm
##	18	18
##	Vedalia Beetle	Araneoid Spider Order
##	18	17
##	Bee Order	Egg Parasitoid
##	17	17
##	Insect Class	Moth And Butterfly Order
##	17	17
##	Oystershell Scale Parasitoid	Hemlock Woolly Adelgid Lady Beetle
##	17	16
##	Hemlock Wooly Adelgid	Mite
##	16	16
##	Onion Thrip	Western Flower Thrips
##	16	15
##	Corn Earworm	Green Peach Aphid
##	14	14
##	House Fly	Ox Beetle
##	14	14
##	Red Scale Parasite	Spined Soldier Bug
##	14	14
##	Armoured Scale Family	Diamondback Moth
##	13	13
##	Eulophid Wasp	Monarch Butterfly

##		13		13
##		Predatory Bug		Yellow Fever Mosquito
##		13		13
##		Braconid Parasitoid		Common Thrip
##		12		12
##		Eastern Subterranean Termite		Jassid
##		12		12
##		Mite Order		Pea Aphid
##		12		12
##		Pond Wolf Spider		Spotless Ladybird Beetle
##		12		11
##		Glasshouse Potato Wasp		Lacewing
##		10		10
##		Southern House Mosquito		Two Spotted Lady Beetle
##		10		10
##		Ant Family		Apple Maggot
##		9		9

Answer: The six most commonly studied species, by common name, are: Honey Bee, Parasitic Wasp, Buff Tailed Bumblebe, Carniolan Honey Bee, Bumble Bee, Italian Honeybee. As it would happen, all of these species are pollinators! These are probably the most frequently studied for that exact reason, their populations are declining and we want to know what role insecticides play in that.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

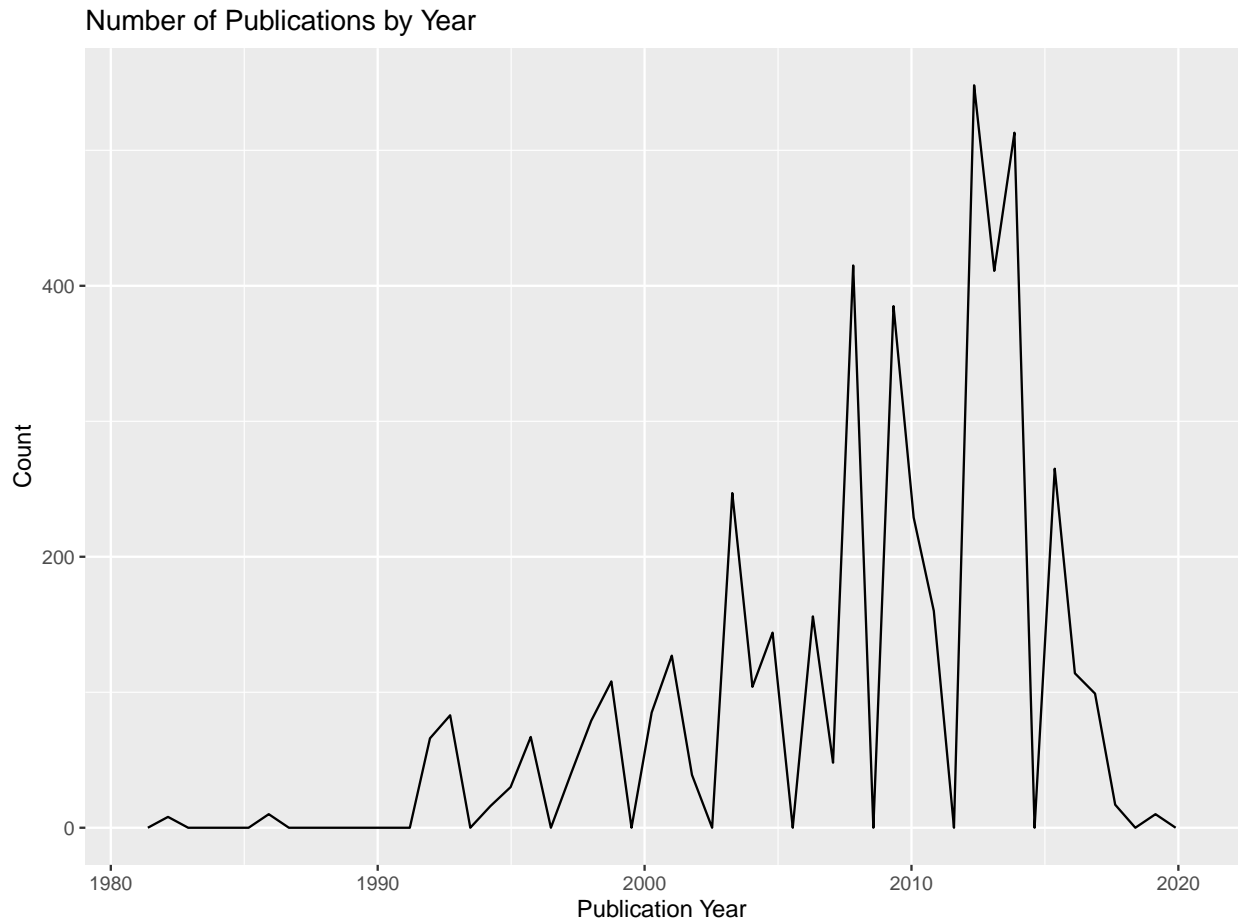
```
## [1] "factor"
```

Answer: `Conc.1..Author.` is factor data. R imported this column as factors because the column contains some non-numeric values, a lot of “/”s. R is kicking it all into factors because of those non-numeric characters.

Explore your data graphically (Neonics)

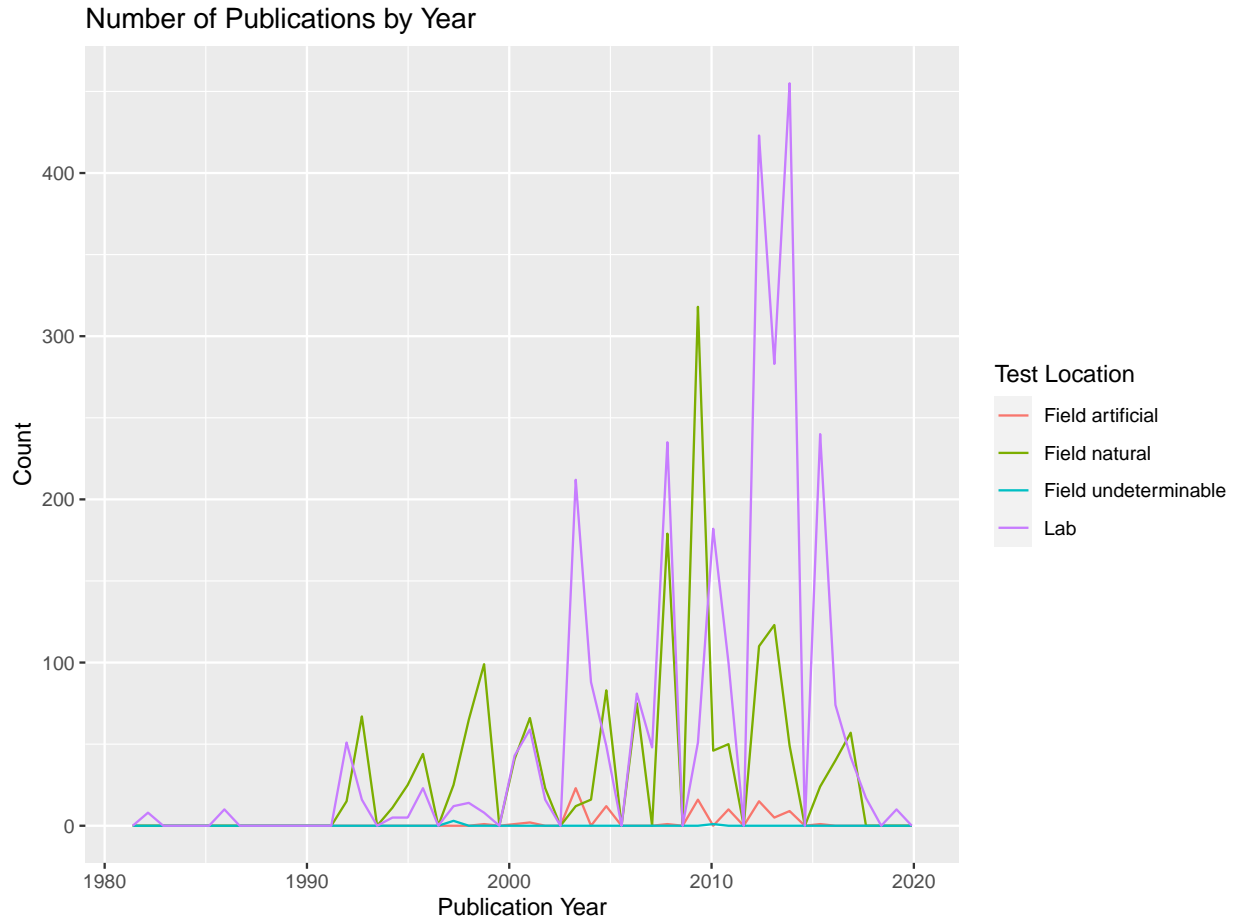
9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) +
  geom_freqpoly(aes(x = Publication.Year), bins = 50) + ggtitle("Number of Publications by Year") + xlab
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 50) + ggtitle("Number of Publications by Year and Test Location")
```



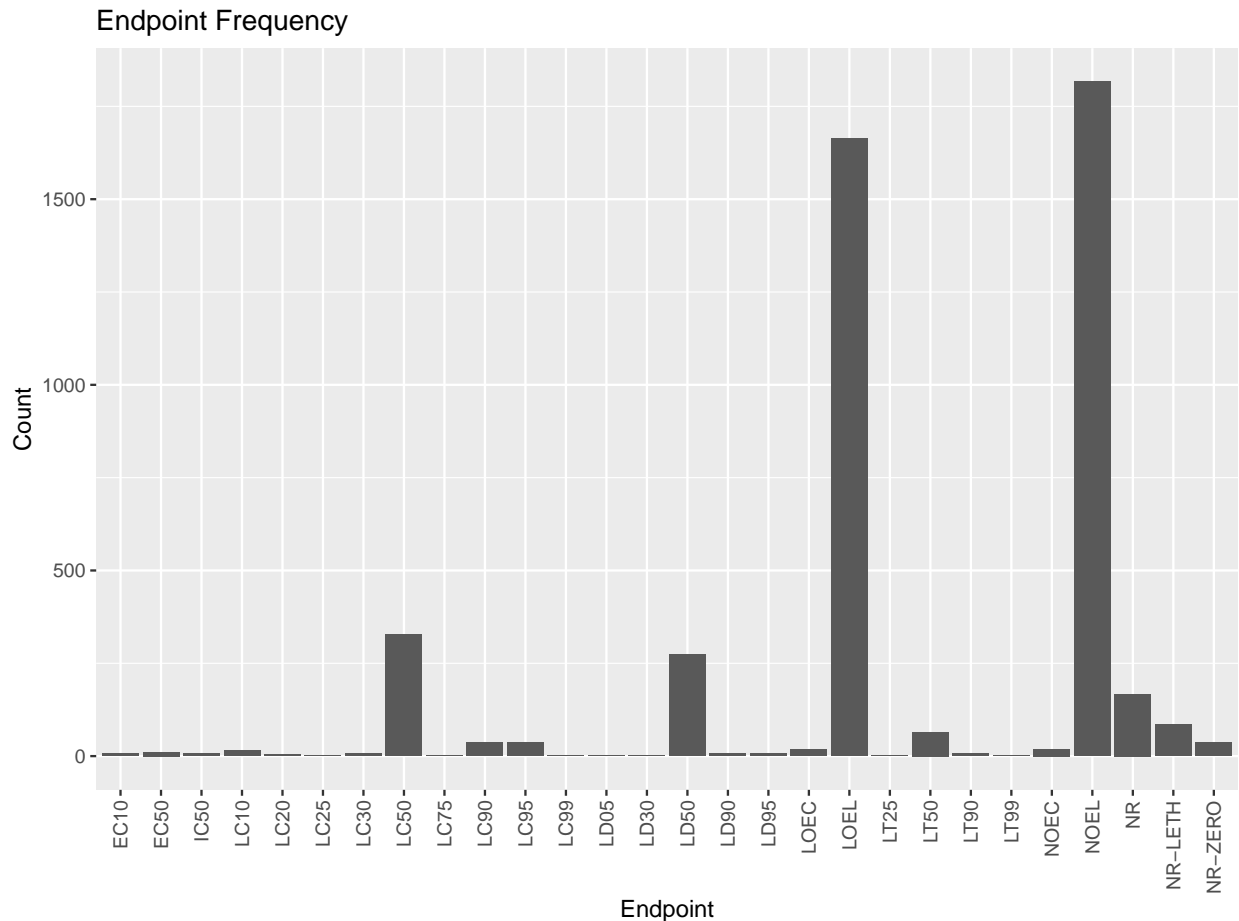
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are the lab and field natural. Field natural and lab experiments frequency are close through 2008 or so, but come 2015 lab is much more frequent than field natural.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[**TIP:** Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics) +
  geom_bar(aes(Endpoint)) + theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1)) + ylab(
```



Answer: The two most common endpoints are LOEL and NOEL. Loel is the lowest observable effect level at which a pesticide produced effects significantly different than the response to controls. Noel is the no-observable-effect-level which is the highest dose producing effects not significantly different from responses of controls.

Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate)
```

```
## [1] "factor"
```

```
Litter$collectDate <- ymd(Litter$collectDate)
class(Litter$collectDate)
```

```
## [1] "Date"
```



```
unique(Litter$collectDate)
```

```
## [1] "2018-08-02" "2018-08-30"
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
length(unique(Litter$plotID))
```

```
## [1] 12
```

```
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051  
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057  
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

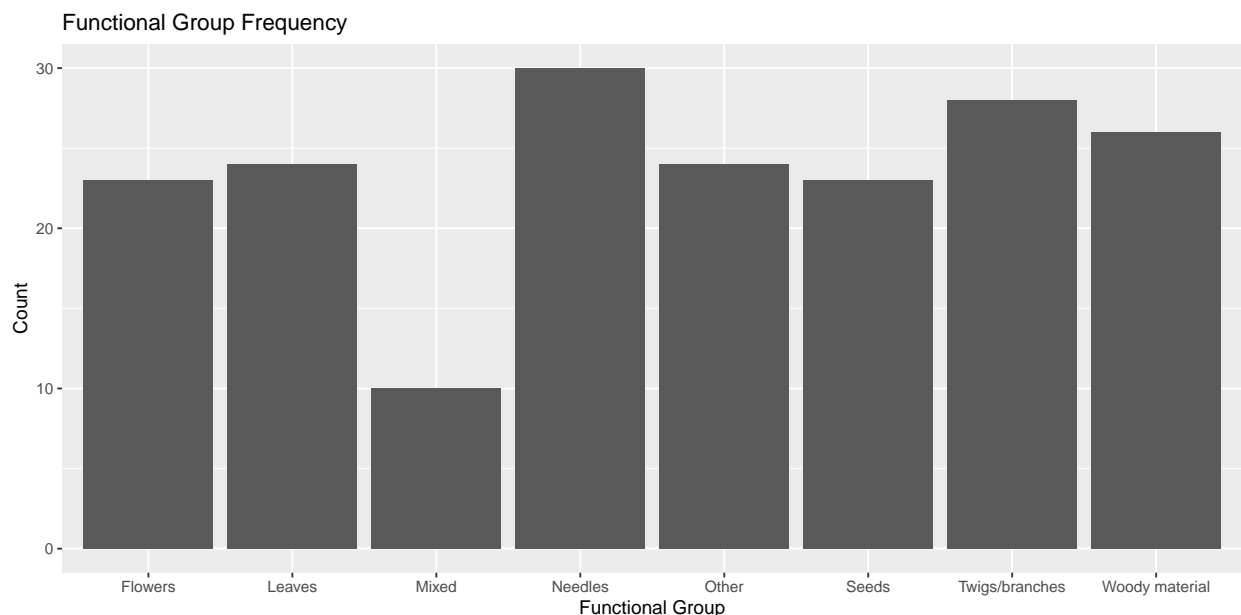
```
summary(Litter$plotID)
```

```
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061  
##      20      19      18      15      14       8      16      17  
## NIWO_062 NIWO_063 NIWO_064 NIWO_067  
##      14      14      16      17
```

Answer: 12 plots were sampled at Niwot Ridge. “Unique” generates a string of all unique values in a vector, “Summary” will do that and show a frequency count for each unique value.

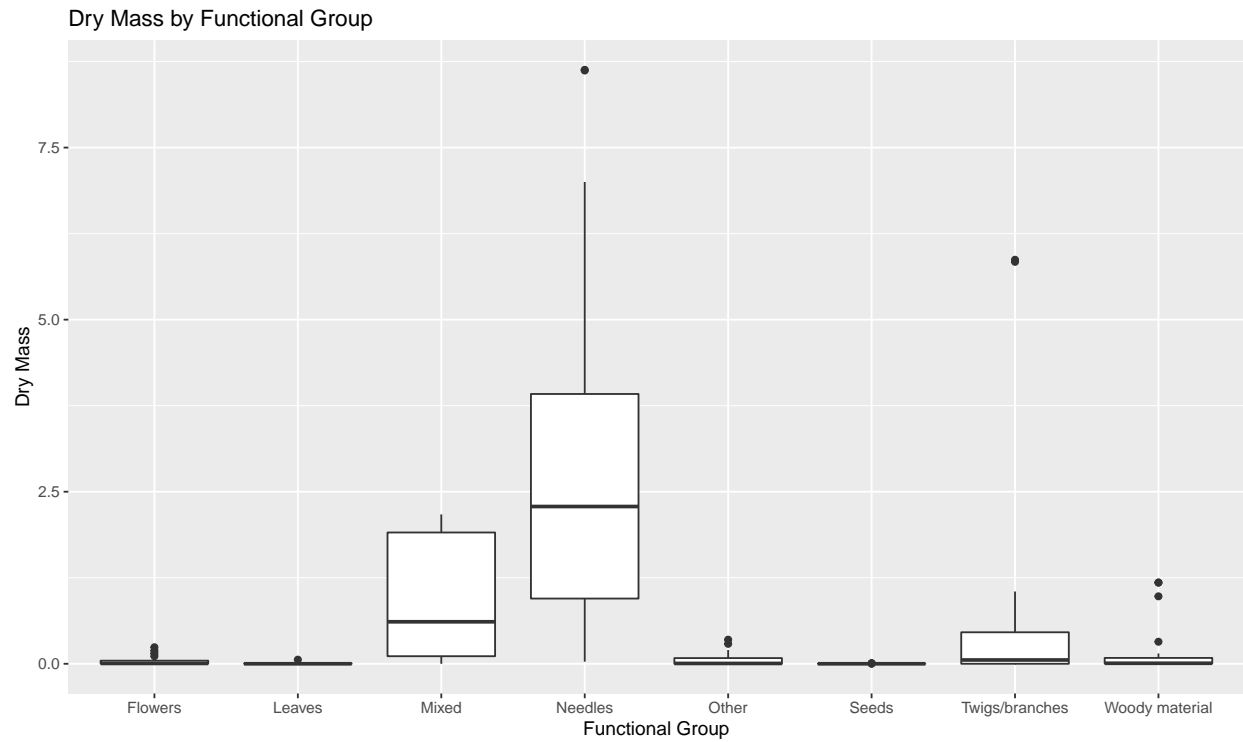
14. Create a bar graph of functional Group counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter) +  
  geom_bar(aes(functionalGroup)) + xlab("Functional Group") + ylab("Count") + ggtitle("Functional Group Frequency")
```

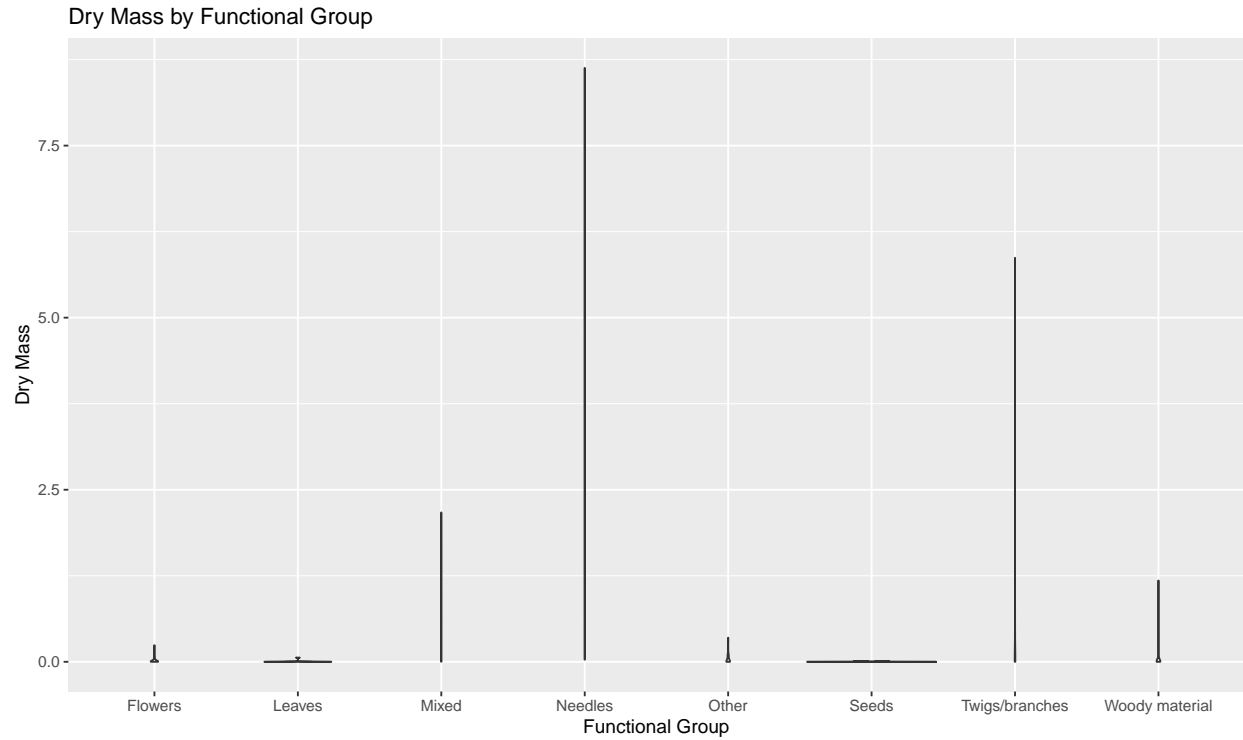


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
ggplot(Litter)+  
  geom_boxplot(aes(functionalGroup, dryMass)) + xlab("Functional Group") + ylab("Dry Mass") + ggtitle("D
```



```
ggplot(Litter)+  
  geom_violin(aes(functionalGroup, dryMass)) + xlab("Functional Group") + ylab("Dry Mass") + ggtitle("D
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: So many functional groups are being compared that it's difficult to see the definition of the "violin" for any one group, the box and whisker plot is much easier to read.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles, twigs/branches, and mixed litter have the highest biomass.