
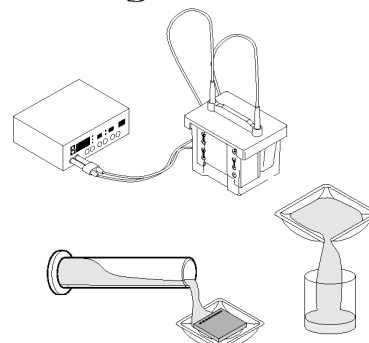
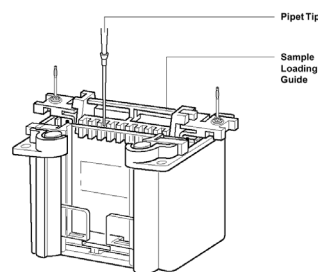
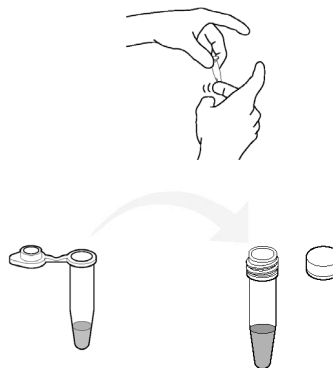
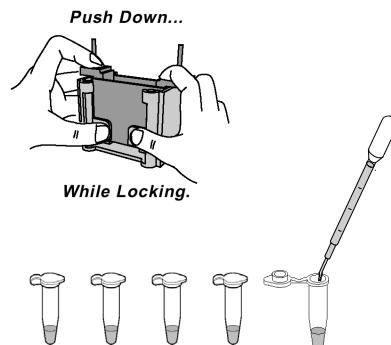


Protein Fingerprinting Kit – Quick Guide

1. Setup Mini-PROTEAN 3 gel box and add 1x TGS electrophoresis buffer to the chamber.
2. Label one 1.5 ml **flip-top** micro tube for each of five fish samples. Also label one **screwcap** micro tube for each fish sample.
3. Add 250 μ l of Bio-Rad Laemmli sample buffer to each labeled **flip-top** micro tube.
4. Cut a piece of each fish muscle about 0.25 x 0.25 x 0.25 cm³ () and transfer each piece into a labeled **flip-top** micro test tube. Close the lids.
5. Flick the microtubes 15 times to agitate the tissue in the sample buffer.
6. Incubate for 5 minutes at room temperature.
7. Carefully transfer the buffer by pouring from **each flip-top** microtube into a labeled **screwcap** microtube. Do not transfer the fish!
8. Obtain the Kaleidoscope prestained standards (KS) and the actin and myosin standard from your teacher.
9. Heat the fish samples and the actin and myosin standard (AM) in screwcap microtubes for 5 minutes at 95°C.
10. Load your gel:

<u>Lane</u>	<u>Volume</u>	<u>Sample</u>
1 & 2	empty	empty
3	10 μ l	Kaleidoscope standards (KS)
4	10 μ l	fish sample 1
5	10 μ l	fish sample 2
6	10 μ l	fish sample 3
7	10 μ l	fish sample 4
8	10 μ l	fish sample 5
9	10 μ l	Actin and myosin standard (AM)
10	empty	empty

11. Electrophorese for 30 minutes at 200 V in 1x TGS electrophoresis buffer.
12. After electrophoresis, remove gel from cassette and transfer gel to a container with 40 ml Bio-Safe Coomassie Blue stain and stain gel for 1 hour, with gentle shaking for best results.
13. Discard stain and destain gels in a large volume of water for at least 30 minutes overnight, changing the water at least once. Blue-stained bands will be visible on a clear gel after destaining.
14. Dry gels using GelAir cellophane.





**Bio-Rad
Laboratories, Inc.**

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