

# ERIN GRUNDY

(Cell) 201-478-9146  
Email: [eringrundy8@gmail.com](mailto:eringrundy8@gmail.com)  
LinkedIn: [www.linkedin.com/in/eringrundy1020](http://www.linkedin.com/in/eringrundy1020)

**Home Address**  
36 Julia Court  
Township of Washington, NJ 07676

## Current Position

### George Washington University

- Graduate Student, Institute for Biomedical Sciences

August 2019-Present  
Washington, DC

## Work Experience

### Research Technician, Chodera Lab, Memorial Sloan Kettering Cancer Center

- Conducted high-throughput, automated wetlab experiments to investigate the interactions and determine the binding affinities of small molecules and their macromolecular targets

June 2018 – July 2019

## Education

### Bucknell University

B.S. Cell Biology & Biochemistry, overall GPA 3.61

*Phi Sigma Honors Society • cum laude • Alpha Sigma Chi Fraternity Prize*, awarded to the most deserving chemistry graduate chosen at the discretion of the chemistry department

Course List: Analytical Chemistry, Biophysical Chemistry, Biochemistry, Molecular Biology, Cell Biology

May 2018  
Lewisburg, PA

### University CEU San Pablo (study abroad)

Course List: Immunology

Fall 2016  
Madrid, Spain

### University Carlos III (study abroad)

Course List: Anatomy & Physiology, Biotechnology

Fall 2016  
Madrid, Spain

## Research Experience

### "Generation of repetitive element-specific T cells to target ovarian cancer"

- Laboratory rotation and dissertation research with Katherine Chiappinelli, PhD
- Research Summary: Assessing the ability of ERV-K-specific primary T cells to lyse several ovarian cancer cell lines and bioinformatic identification of novel tumor-associated antigens upregulated by epigenetic therapy.

May 2020-Present

### "Investigating the role of the p53 signaling pathway in HIV-1 infectivity and latency"

- Laboratory rotation with Alberto Bosque-Pardos, PhD
- Research Summary: Analyzed changes in HIV-1 infectivity and latency in response to the therapeutic perturbation of the p53 signaling pathway in cell lines and primary cells.

January-April 2020

### "Ex vivo expanded antigen-specific T cells targeting brain tumors"

- Laboratory rotation with C. Russell Y. Cruz, MD, PhD
- Research Summary: Generated *ex vivo* antigen-specific T cells for the WT1 antigen and identified additional candidate target peptides for glioblastoma multiforme via utilization of the GEO and SYFPEITHI databases.

September-December 2019

### "Structural determination of the RFTS domain of DNMT1 using 2D and 3D Nuclear Magnetic Resonance"

- Bucknell Undergraduate Summer Research Fellowship, Advisor: Rebecca Fagan Switzer, PhD
- Research Summary: Optimized a unique protocol for growing and labeling the RFTS domain of DNMT1 for use in NMR structural determination experiments.
- Funding: Merck Chemistry Undergraduate Research Fund

Summer 2017

### "Probing regulatory protein-protein interactions using Isothermal Titration Calorimetry"

- Bucknell Program for Undergraduate Research Summer Fellowship, Advisor: Rebecca Fagan Switzer, PhD
- Research Summary: Developed a distinctive protocol for isothermal titration calorimetry of the maintenance methylation proteins DNMT1 and UHRF1.
- Presented poster at Sigma Xi Symposium (Bucknell University, July 2016)

Summer 2016

## Technical Skills

### Automation Instruments:

- Tecan EVO- automated liquid handler
- LabChip- automated capillary electrophoresis
- Opentrons- open-source programming pipetting robot

### Laboratory Techniques & Instruments:

- Mammalian cell culture
- Bacterial cell culture
- Lentiviral transduction
- DNA transfection
- Plasmid transformation
- Flow cytometry
- ELISpot assays
- Western blots
- SDS-PAGE gels
- EMSAs
- Visible & UV light spectrophotometer
- Column chromatography
- Differential scanning fluorimetry
- Differential light scattering
- RT-qPCR
- Protein purification
- FPLC
- HPLC
- ITC
- Gas chromatography
- CE

### Programs:

- Python
- Prism
- FCS Express 7
- FlowJo
- R
- ImageJ