

# Class 10: Structural Bioinformatics (pt 1)

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## Table of contents

The PDB database . . . . .	1
Question 1 . . . . .	2
Question 2 . . . . .	2
Question 3 . . . . .	2
Visualizing with Mol-star . . . . .	3
Using the <code>bio3d</code> package in R . . . . .	5
Molecular visualization in R . . . . .	7
Predicting functional motions of a structure . . . . .	8

## The PDB database

The main repository of biomolecular structure data is called the [Protein Data Bank](#) (PDB). It is the second oldest database (after GenBank).

What is currently in the PDB?

```
stats <- read.csv("Data Export Summary.csv", row.names = 1)
head(stats)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	171,959	18,083	12,622	210	84	32
Protein/Oligosaccharide	10,018	2,968	34	10	2	0
Protein/NA	8,847	5,376	286	7	0	0
Nucleic acid (only)	2,947	185	1,535	14	3	1
Other	170	10	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
	Total					
Protein (only)	202,990					

Protein/Oligosaccharide	13,032
Protein/NA	14,516
Nucleic acid (only)	4,685
Other	213
Oligosaccharide (only)	22

### Question 1

What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy?

```
del_comma <- function(x){
  as.numeric(gsub(",", "", x))
}

stats <- data.frame(apply(stats, 2, del_comma), row.names = rownames(stats))
```

```
sum(stats$X.ray) / sum(stats$Total) * 100
```

```
[1] 82.37223
```

```
sum(stats$EM) / sum(stats$Total) * 100
```

```
[1] 11.30648
```

### Question 2

What proportion of structures in the PDB are protein?

```
sum(stats["Protein (only)", -ncol(stats)]) / sum(stats$Total) * 100
```

```
[1] 86.2107
```

### Question 3

Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

Skipped

## Visualizing with Mol-star

Explore the HIV-1 protease structure with PDB code 1HSG.

We will use [Mol-star](#).



Figure 1: Figure 1. A first view of HIV-Pr.

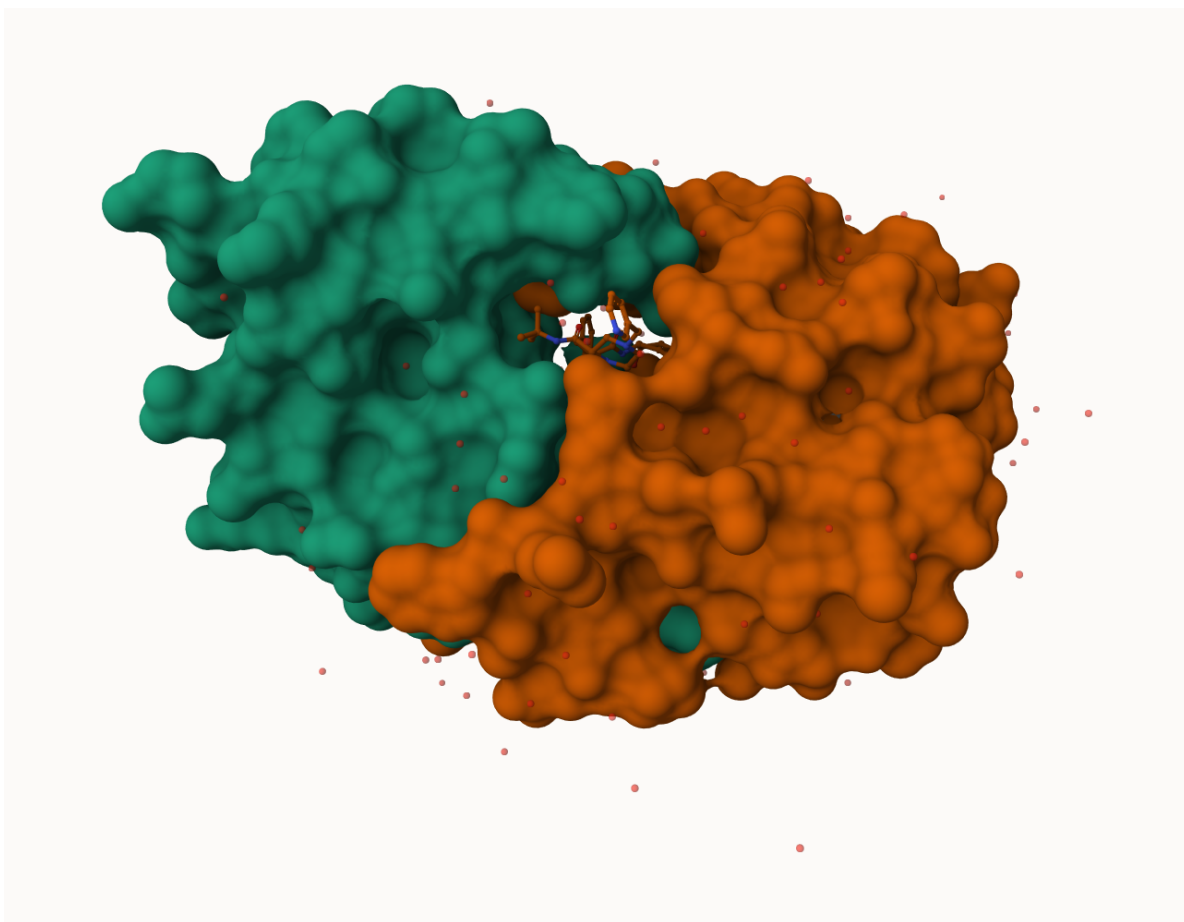


Figure 2: Figure 2. A view of the ligand-binding region of HIV-Pr.



Figure 3: Figure 3. The relevant Asp residues and water molecule (HOH 308) in MK1 binding.

### Using the bio3d package in R

The Bio3D package is focused on structural bioinformatics analysis and allows us to read and analyze PDB (and related) datasets.

```
library(bio3d)
```

Warning: package 'bio3d' was built under R version 4.4.3

```
pdb <- read.pdb("1HSG")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1HSG")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
```

```
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
```

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
attributes(pdb)
```

```
$names
```

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
```

```
$class
```

```
[1] "pdb" "sse"
```

We can see atom data with `pdb$atom`.

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

```
segid elesy charge
```

```

1  <NA>      N  <NA>
2  <NA>      C  <NA>
3  <NA>      C  <NA>
4  <NA>      O  <NA>
5  <NA>      C  <NA>
6  <NA>      C  <NA>

```

```
head(pdbseq(pdb))
```

```

      1      2      3      4      5      6
"P" "Q" "I" "T" "L" "W"

```

## Molecular visualization in R

We can make a quick 3d visualization with the `view.pdb()` function.

```

#install.packages("pak")
#pak::pak("bioboot/bio3dview")
library(bio3dview)

#install.packages("NGLViewer")
library(NGLViewer)

```

Warning: package 'NGLViewer' was built under R version 4.4.3

```

# view.pdb(pdb, backgroundColor = "cyan", colorScheme = "sse") |>
#   setSpin()

```

```

sel <- atom.select(pdb, resno = 25)

# view.pdb(pdb,
#           highlight = sel,
#           highlight.style = "spacefill",
#           cols = c("green", "orange")) |>
#   setRock()

```

## Predicting functional motions of a structure

We can finish off with a bioinformatics prediction of the functional motions of a protein.

We will run a Normal Mode Analysis (NMA).

```
adk <- read.pdb("6s36")
```

```
Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE
```

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
  Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

```
Protein sequence:
```

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
TDELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKFNPVKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

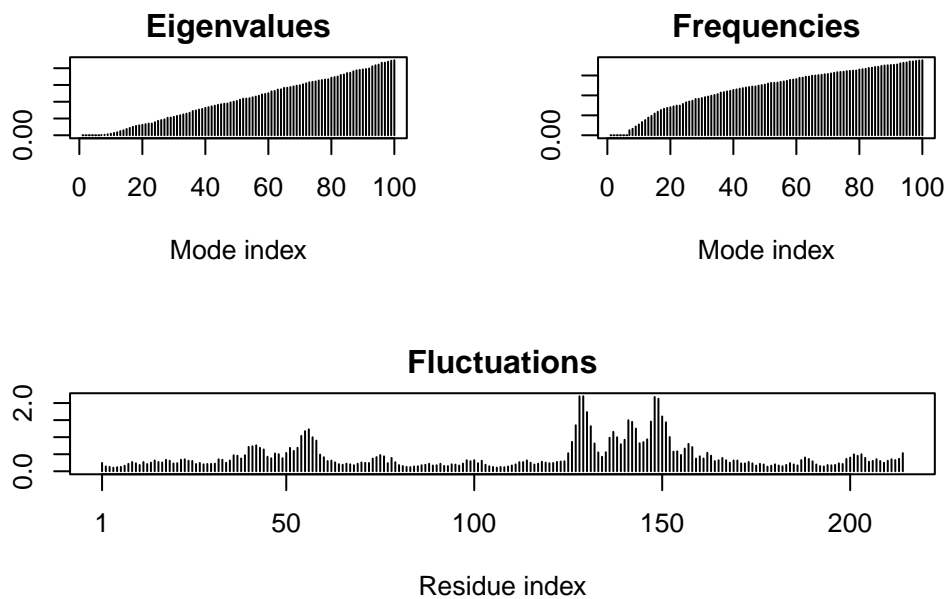
```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
m <- nma(adk)
```

```
Building Hessian...      Done in 0.03 seconds.
Diagonalizing Hessian... Done in 0.47 seconds.
```



```
plot(m)
```



```
# view.nma(m)
```

We can write out a trajectory of the predicted dynamics and view this in Mol-star.

```
mktrj(m, file = "nma.pdb")
```