

Class 18: Pertussis Mini-project

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```
library(ggplot2)
library(jsonlite)
library(lubridate)
library(dplyr)
```

Pertussis (more commonly known as whooping cough) is a highly contagious respiratory disease caused by the bacterium *Bordetella pertussis*. People of all ages can be infected leading to violent coughing fits followed by a characteristic high-pitched “whoop” like intake of breath. Children have the highest risk for severe complications and death. Recent estimates from the WHO indicate that ~16 million cases and 200,000 infant deaths are due to pertussis annually.

Investigating pertussis cases by year

```
#install.packages("datapasta")

cdc <- data.frame(Year = c(1922L,
                           1923L, 1924L, 1925L, 1926L, 1927L, 1928L,
                           1929L, 1930L, 1931L, 1932L, 1933L, 1934L, 1935L,
                           1936L, 1937L, 1938L, 1939L, 1940L, 1941L,
                           1942L, 1943L, 1944L, 1945L, 1946L, 1947L, 1948L,
                           1949L, 1950L, 1951L, 1952L, 1953L, 1954L,
                           1955L, 1956L, 1957L, 1958L, 1959L, 1960L,
```

```

1961L,1962L,1963L,1964L,1965L,1966L,1967L,
1968L,1969L,1970L,1971L,1972L,1973L,
1974L,1975L,1976L,1977L,1978L,1979L,1980L,
1981L,1982L,1983L,1984L,1985L,1986L,
1987L,1988L,1989L,1990L,1991L,1992L,1993L,
1994L,1995L,1996L,1997L,1998L,1999L,
2000L,2001L,2002L,2003L,2004L,2005L,
2006L,2007L,2008L,2009L,2010L,2011L,2012L,
2013L,2014L,2015L,2016L,2017L,2018L,
2019L,2020L,2021L,2022L,2023L),
No..Reported.Pertussis.Cases = c(107473,
164191,165418,152003,202210,181411,
161799,197371,166914,172559,215343,179135,
265269,180518,147237,214652,227319,103188,
183866,222202,191383,191890,109873,
133792,109860,156517,74715,69479,120718,
68687,45030,37129,60886,62786,31732,28295,
32148,40005,14809,11468,17749,17135,
13005,6799,7717,9718,4810,3285,4249,
3036,3287,1759,2402,1738,1010,2177,2063,
1623,1730,1248,1895,2463,2276,3589,
4195,2823,3450,4157,4570,2719,4083,6586,
4617,5137,7796,6564,7405,7298,7867,
7580,9771,11647,25827,25616,15632,10454,
13278,16858,27550,18719,48277,28639,
32971,20762,17972,18975,15609,18617,6124,
2116,3044,7063)
)

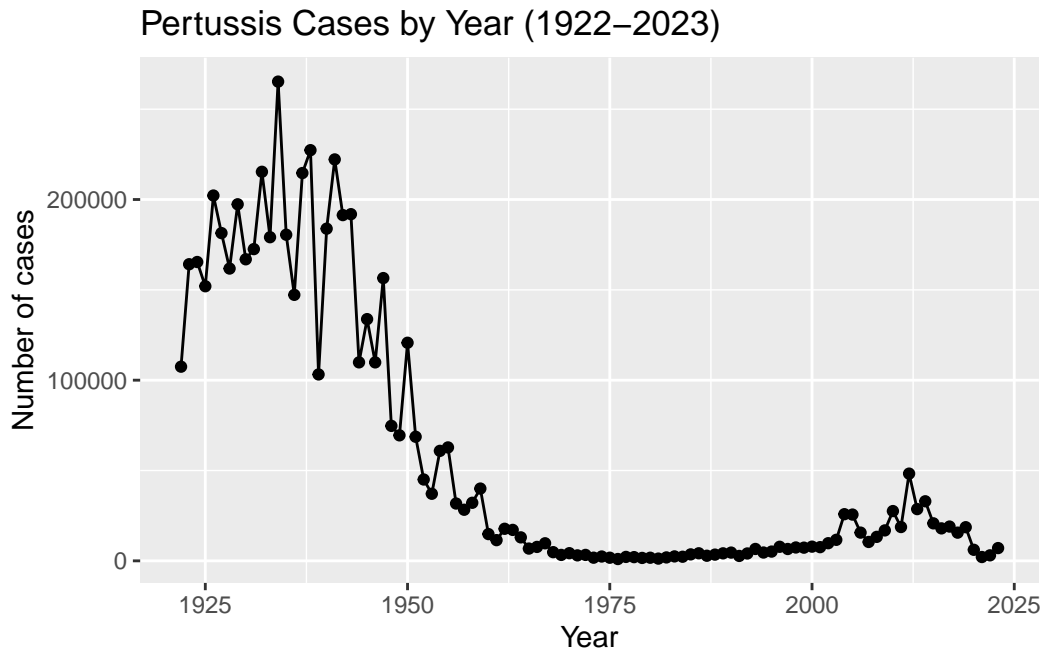
```

Q1: With the help of the R “addin” package **datapasta** assign the CDC pertussis case number data to a data frame called **cdc** and use **ggplot** to make a plot of cases numbers over time.

```

options(scipen = 999)
plot <- ggplot(cdc) +
  aes(x = Year, y = No..Reported.Pertussis.Cases) +
  geom_point() +
  geom_line() +
  labs(x = "Year", y = "Number of cases", title = "Pertussis Cases by Year (1922-2023)")
plot

```

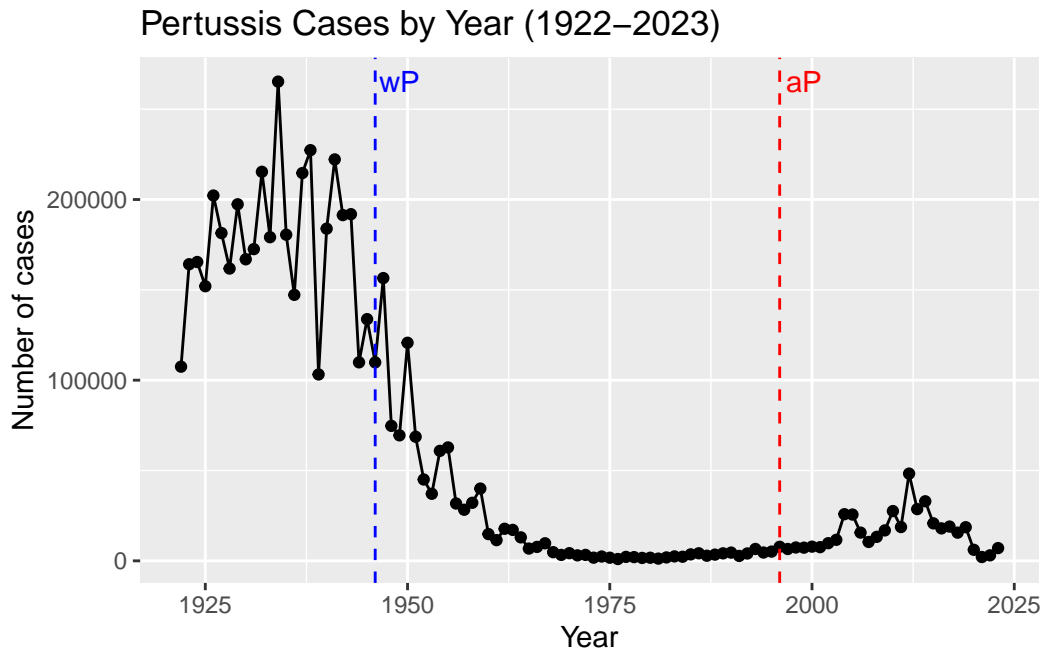


A tale of two vaccines

Two types of pertussis vaccines have been developed: **whole-cell pertussis (wP)** and **acellular pertussis (aP)**. The first vaccines were composed of ‘whole cell’ (wP) inactivated bacteria. The latter aP vaccines use purified antigens of the bacteria (the most important pertussis components for our immune system). These aP vaccines were developed to have less side effects than the older wP vaccines and are now the only form administered in the United States.

Q2: Using the ggplot `geom_vline()` function add lines to your previous plot for the 1946 introduction of the wP vaccine and the 1996 switch to aP vaccine (see example in the hint below). What do you notice?

```
plot +
  geom_vline(xintercept = 1946, col = "blue", linetype = 2) +
  annotate("text", label = "wP",
          x = 1946 + 3, y = max(cdc$No..Reported.Pertussis.Cases), col = "blue") +
  geom_vline(xintercept = 1996, col = "red", linetype = 2) +
  annotate("text", label = "aP",
          x = 1996 + 3, y = max(cdc$No..Reported.Pertussis.Cases), col = "red")
```



Q3: Describe what happened after the introduction of the aP vaccine? Do you have a possible explanation for the observed trend?

After the introduction of the aP vaccine, the number of cases rose again despite declining after the introduction of the wP vaccine. This could be because people were skeptical of the new vaccine and chose not to use it. This could also be because some strains of the bacterium evolved to acquire immunity from the vaccine, resulting in more cases.

Exploring CMI-PB data

The new and ongoing CMI-PB project aims to provide the scientific community with this very information. In particular, CMI-PB tracks and makes freely available long-term humoral and cellular immune response data for a large number of individuals who received either DTwP or DTaP combination vaccines in infancy followed by Tdap booster vaccinations. This includes complete API access to longitudinal RNA-Seq, AB Titer, Olink, and live cell assay results directly from their website: <https://www.cmi-pb.org/>.

```
subject <- read_json("https://www.cmi-pb.org/api/subject", simplifyVector = TRUE)
head(subject)
```

```
subject_id  infancy_vac  biological_sex      ethnicity  race
```

1	1	wP	Female Not Hispanic or Latino White
2	2	wP	Female Not Hispanic or Latino White
3	3	wP	Female Unknown White
4	4	wP	Male Not Hispanic or Latino Asian
5	5	wP	Male Not Hispanic or Latino Asian
6	6	wP	Female Not Hispanic or Latino White

	year_of_birth	date_of_boost	dataset
1	1986-01-01	2016-09-12	2020_dataset
2	1968-01-01	2019-01-28	2020_dataset
3	1983-01-01	2016-10-10	2020_dataset
4	1988-01-01	2016-08-29	2020_dataset
5	1991-01-01	2016-08-29	2020_dataset
6	1988-01-01	2016-10-10	2020_dataset

Q4: How many aP and wP infancy vaccinated subjects are in the dataset?

```
table(subject$infancy_vac)
```

```
aP wP
87 85
```

Q5: How many Male and Female subjects/patients are in the dataset?

```
table(subject$biological_sex)
```

```
Female    Male
   112     60
```

Q6: What is the breakdown of race and biological sex (e.g. number of Asian females, White males etc...)?

```
table(subject$biological_sex, subject$race)
```

	American Indian/Alaska Native	Asian	Black or African American
Female	0	32	2
Male	1	12	3

More Than One Race Native Hawaiian or Other Pacific Islander

Female	15	1
Male	4	1

Unknown or Not Reported White		
Female	14	48
Male	7	32

Q7: Using this approach determine (i) the average age of wP individuals, (ii) the average age of aP individuals; and (iii) are they significantly different?

```
subject$age <- today() - ymd(subject$year_of_birth)
```

```
wp <- subject %>% filter(infancy_vac == "wP")
ap <- subject %>% filter(infancy_vac == "aP")

summary(time_length(wp$age, "years"))
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
22.41	32.41	34.41	36.05	39.41	57.41

```
summary(time_length(ap$age, "years"))
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
22.41	26.41	27.41	27.30	28.41	34.41

```
t.test(time_length(wp$age, "years"), time_length(ap$age, "years"))
```

Welch Two Sample t-test

```
data: time_length(wp$age, "years") and time_length(ap$age, "years")
t = 12.918, df = 104.03, p-value < 0.000000000000000022
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
 7.407351 10.094058
sample estimates:
mean of x mean of y
36.05331 27.30260
```

The p-value for the t-test is very small, so the average ages of wP and aP individuals are significantly different from each other.

Q8: Determine the age of all individuals at time of boost?

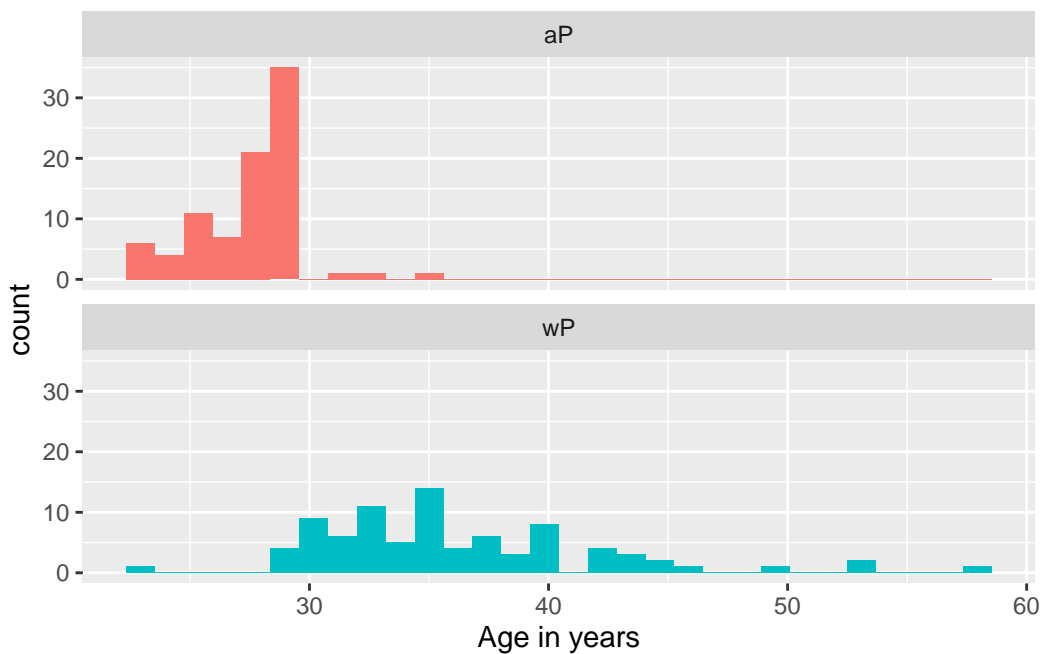
```
subject$age_at_boost <- time_length(ymd(subject$date_of_boost)- ymd(subject$year_of_birth), "year")
head(subject$age_at_boost)
```

```
[1] 30.69678 51.07461 33.77413 28.65982 25.65914 28.77481
```

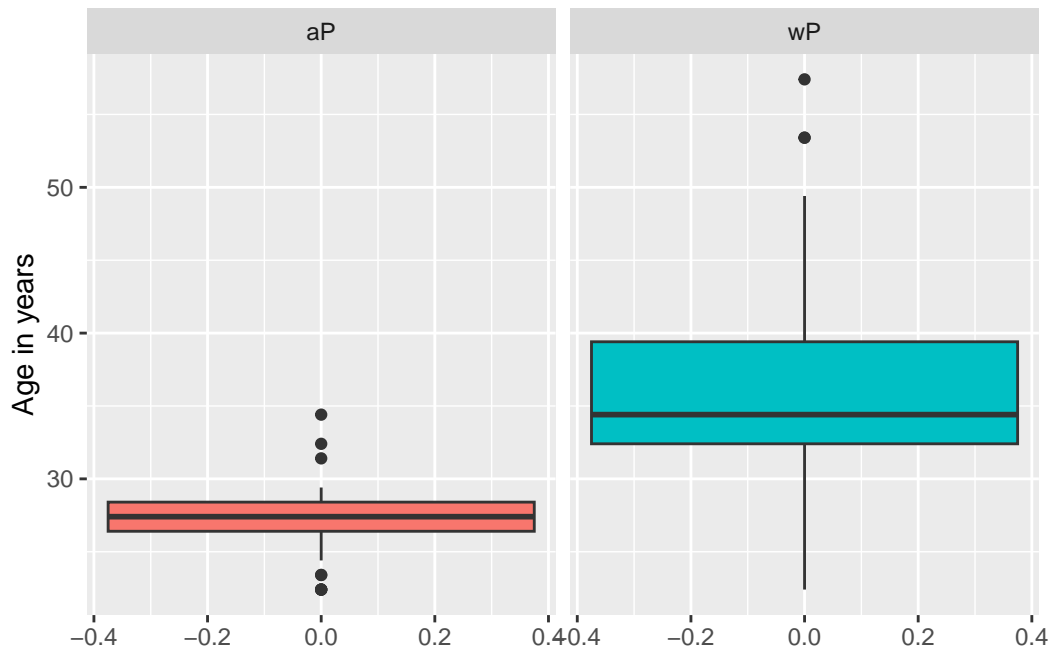
Q9: With the help of a faceted boxplot or histogram, do you think these two groups are significantly different?

```
ggplot(subject) +
  aes(time_length(age, "year"),
      fill=as.factor(infancy_vac)) +
  geom_histogram(show.legend=FALSE) +
  facet_wrap(vars(infancy_vac), nrow=2) +
  xlab("Age in years")
```

`stat_bin()` using `bins = 30`. Pick better value with `binwidth`.



```
ggplot(subject) +
  aes(y = time_length(age, "year"),
      fill=as.factor(infancy_vac)) +
  geom_boxplot(show.legend = FALSE) +
  facet_wrap(vars(infancy_vac), ncol=2) +
  ylab("Age in years")
```



```
specimen <- read_json("https://www.cmi-pb.org/api/specimen", simplifyVector = TRUE)
titer <- read_json("https://www.cmi-pb.org/api/plasma_ab_titer", simplifyVector = TRUE)
```

Q10: Complete the code to join `specimen` and `subject` tables to make a new merged data frame containing all specimen records along with their associated subject details:

```
meta <- inner_join(specimen, subject)
```

Joining with ``by = join_by(subject_id)``

```
dim(meta)
```

```
[1] 1503  15
```



```
head(meta)
```

```

specimen_id subject_id actual_day_relative_to_boost
1           1           1                      -3
2           2           1                       1
3           3           1                       3
4           4           1                       7
5           5           1                      11
6           6           1                      32
planned_day_relative_to_boost specimen_type visit infancy_vac biological_sex
1                           0         Blood      1          wP        Female
2                           1         Blood      2          wP        Female
3                           3         Blood      3          wP        Female
4                           7         Blood      4          wP        Female
5                          14         Blood      5          wP        Female
6                          30         Blood      6          wP        Female
ethnicity race year_of_birth date_of_boost dataset
1 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
2 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
3 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
4 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
5 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
6 Not Hispanic or Latino White 1986-01-01 2016-09-12 2020_dataset
age age_at_boost
1 14393 days 30.69678
2 14393 days 30.69678
3 14393 days 30.69678
4 14393 days 30.69678
5 14393 days 30.69678
6 14393 days 30.69678

```

Q11: Now using the same procedure join `meta` with `titer` data so we can further analyze this data in terms of time of visit aP/wP, male/female etc.

```
abdata <- inner_join(titer, meta)
```

Joining with ``by = join_by(specimen_id)``

```
dim(abdata)
```

```
[1] 52576    22
```

Q12: How many specimens (i.e. entries in `abdata`) do we have for each `isotype`?

```
table(abdata$isotype)
```

```

IgE   IgG  IgG1  IgG2  IgG3  IgG4
6698 5389 10117 10124 10124 10124

```

Q13: What are the different `$dataset` values in `abdata` and what do you notice about the number of rows for the most “recent” dataset?

```
table(abdata$dataset)
```

```

2020_dataset 2021_dataset 2022_dataset 2023_dataset
          31520           8085           7301           5670

```

The `$dataset` values show what year the data were collected, with the 2020 dataset presumably including previous years as well. The number of rows for the most recent dataset, 2023, is the smallest, likely because more data is being collected or entered. Alternatively, this could be because there were just fewer people in the most recent dataset as the number of subjects has been decreasing over time. (`abdata$subjectid`, `abdata$dataset`)

Examine IgG Ab titer levels

```

igg <- abdata %>% filter(isotype == "IgG")
head(igg)

```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgG	TRUE	PT	68.56614	3.736992
2	1	IgG	TRUE	PRN	332.12718	2.602350
3	1	IgG	TRUE	FHA	1887.12263	34.050956
4	19	IgG	TRUE	PT	20.11607	1.096366
5	19	IgG	TRUE	PRN	976.67419	7.652635
6	19	IgG	TRUE	FHA	60.76626	1.096457

	unit	lower_limit_of_detection	subject_id	actual_day_relative_to_boost
1	IU/ML	0.530000	1	-3
2	IU/ML	6.205949	1	-3
3	IU/ML	4.679535	1	-3

4	IU/ML	0.530000	3	-3
5	IU/ML	6.205949	3	-3
6	IU/ML	4.679535	3	-3

	planned_day_relative_to_boost	specimen_type	visit	infancy_vac	biological_sex
1	0	Blood	1	wP	Female
2	0	Blood	1	wP	Female
3	0	Blood	1	wP	Female
4	0	Blood	1	wP	Female
5	0	Blood	1	wP	Female
6	0	Blood	1	wP	Female

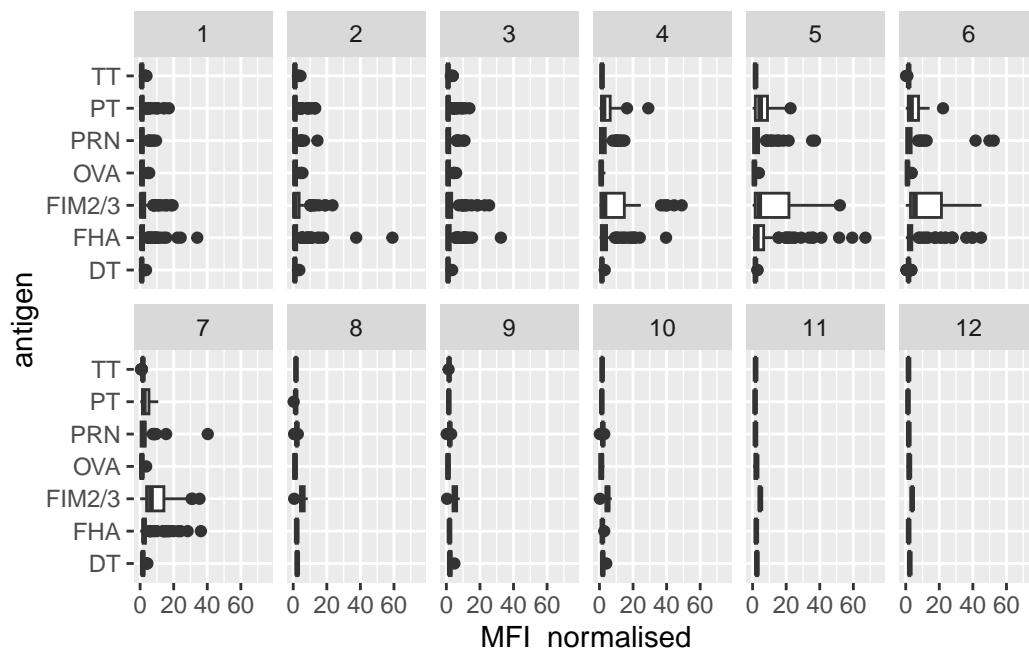
	ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
4	Unknown	White	1983-01-01	2016-10-10	2020_dataset
5	Unknown	White	1983-01-01	2016-10-10	2020_dataset
6	Unknown	White	1983-01-01	2016-10-10	2020_dataset

	age	age_at_boost
1	14393 days	30.69678
2	14393 days	30.69678
3	14393 days	30.69678
4	15489 days	33.77413
5	15489 days	33.77413
6	15489 days	33.77413

Q14: Complete the following code to make a summary boxplot of Ab titer levels (MFI) for all antigens:

```
ggplot(igg) +
  aes(MFI_normalised, antigen) +
  geom_boxplot() +
  xlim(0,75) +
  facet_wrap(vars(visit), nrow=2)
```

Warning: Removed 5 rows containing non-finite outside the scale range (`stat_boxplot()`).

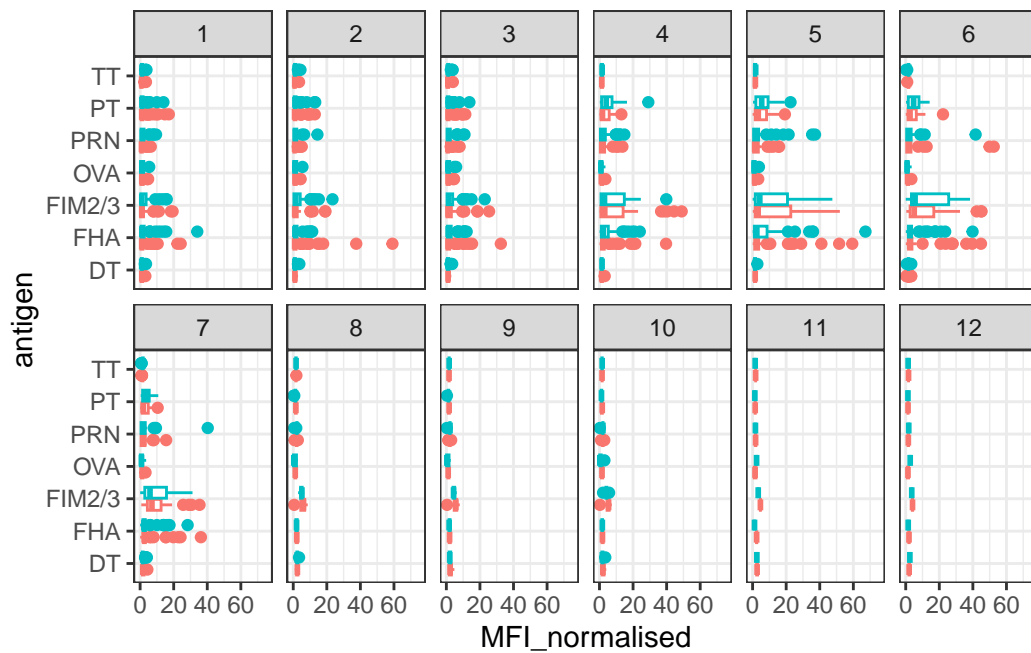


Q15: What antigens show differences in the level of IgG antibody titers recognizing them over time? Why these and not others?

The antigens PT and FIM2/3 show the most difference in IgG antibody recognition over time. A few others have some variation, but only in outliers. PT is the pertussis toxin complex, so it makes sense that it would have varying recognition over time. FIM2/3 is a mixture of fimbrial proteins 2 and 3, which are serotypes of a *B. pertussis* protein that seems to be important in attaching to host cells.

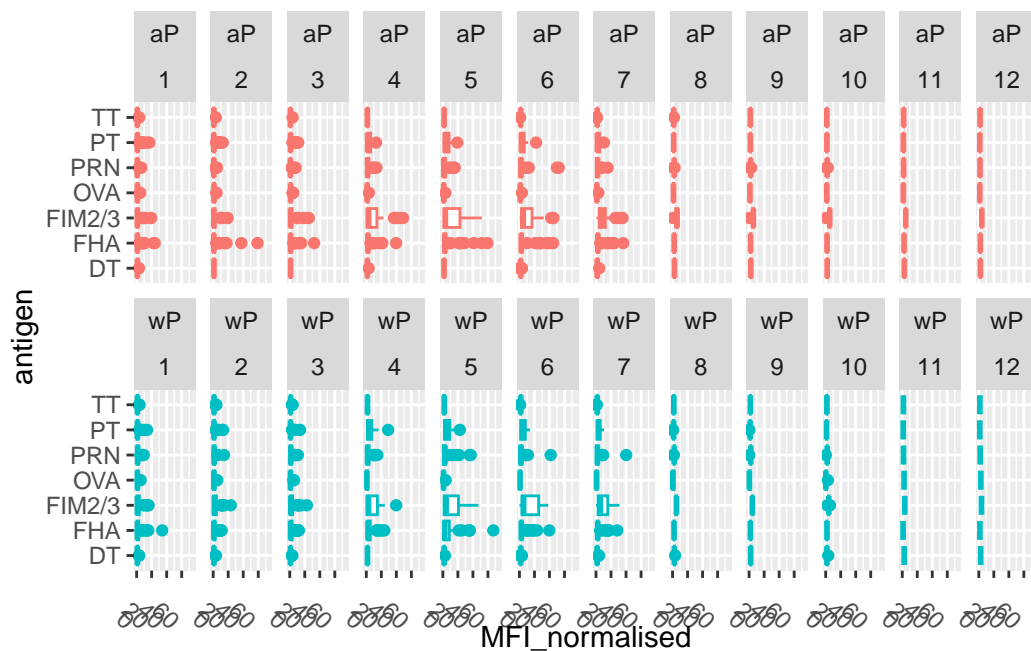
```
ggplot(igg) +
  aes(MFI_normalised, antigen, col=infancy_vac ) +
  geom_boxplot(show.legend = FALSE) +
  facet_wrap(vars(visit), nrow=2) +
  xlim(0,75) +
  theme_bw()
```

Warning: Removed 5 rows containing non-finite outside the scale range (``stat_boxplot()``).



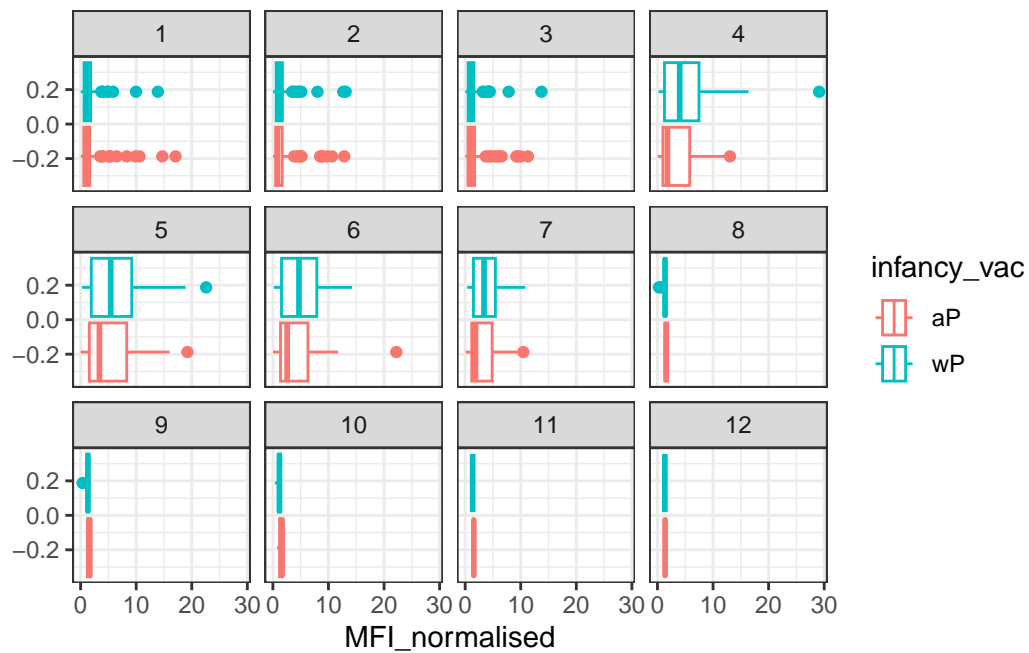
```
igg %>%
  ggplot() +
    aes(MFI_normalised, antigen, col=infancy_vac ) +
    geom_boxplot(show.legend = FALSE) +
    xlim(0,75) +
    facet_wrap(vars(infancy_vac, visit), nrow=2) +
    theme(axis.text.x = element_text(angle = -45, hjust=1))
```

Warning: Removed 5 rows containing non-finite outside the scale range (``stat_boxplot()``).

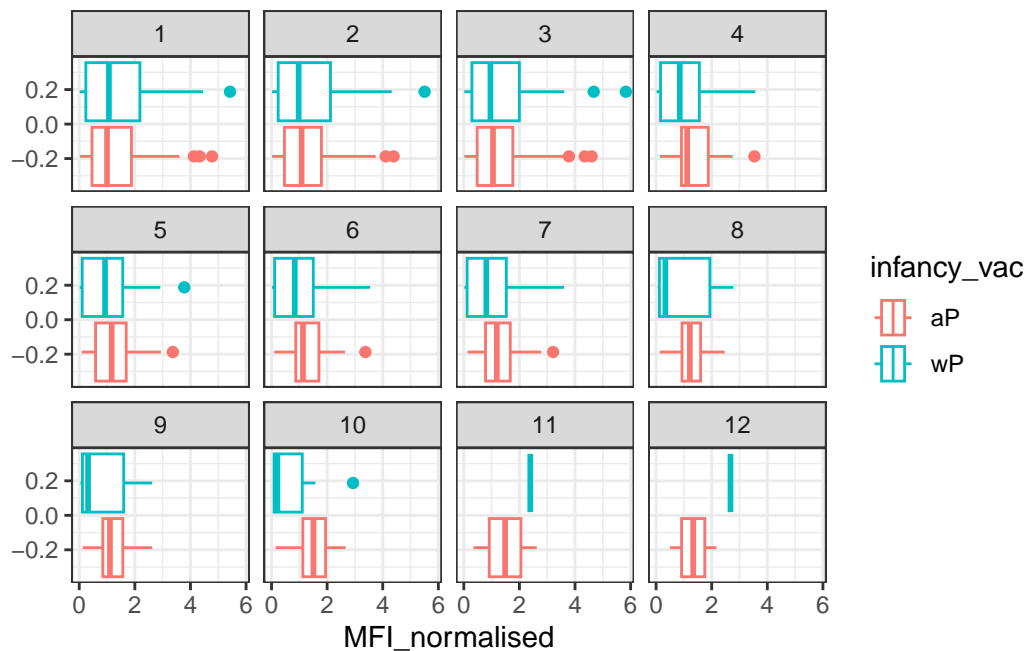


Q16: Filter to pull out only two specific antigens for analysis and create a boxplot for each. You can choose any you like. Below I picked a “control” antigen (“**OVA**”, that is not in our vaccines) and a clear antigen of interest (“**PT**”, Pertussis Toxin, one of the key virulence factors produced by the bacterium *B. pertussis*).

```
filter(igg, antigen == "PT") %>%
  ggplot() +
  aes(MFI_normalised, col=infancy_vac) +
  geom_boxplot(show.legend = T) +
  facet_wrap(vars(visit)) +
  theme_bw()
```



```
filter(igg, antigen == "OVA") %>%
  ggplot() +
  aes(MFI_normalised, col=infancy_vac) +
  geom_boxplot(show.legend = T) +
  facet_wrap(vars(visit)) +
  theme_bw()
```



Q17: What do you notice about these two antigens time courses and the PT data in particular?

OVA changes are on a much smaller scale than are PT changes and look roughly the same over time. PT starts with low MFI but rises to peak around the 5th or 6th visit, then falls back down.

Q18: Do you see any clear difference in aP vs. wP responses?

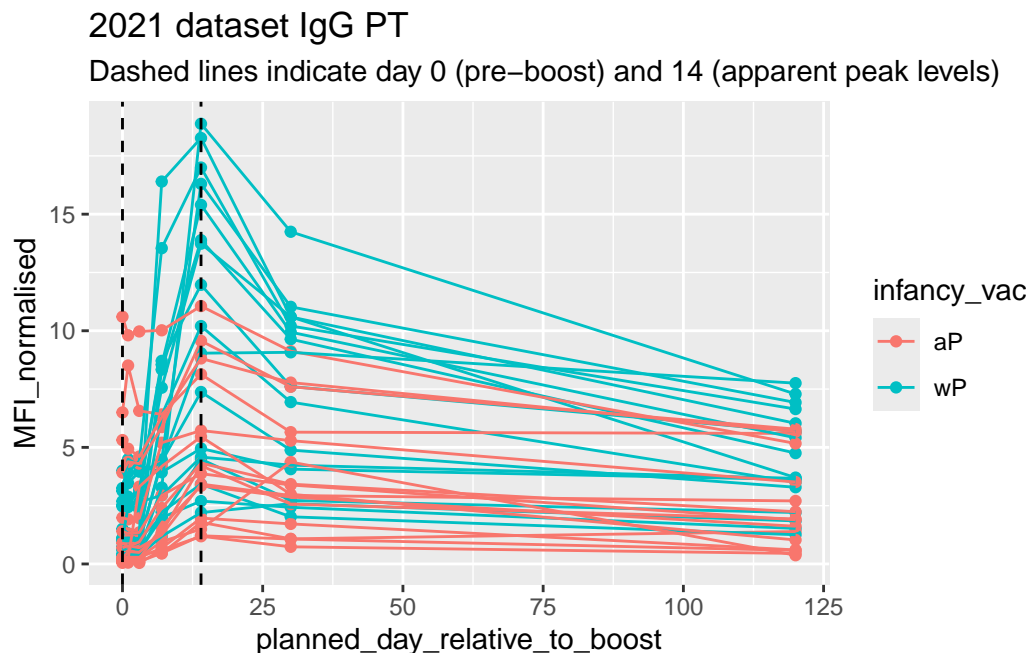
PT levels in wP individuals might be slightly higher when PT levels peak, but the difference does not look significant.

```
abdata.21 <- abdata %>% filter(dataset == "2021_dataset")

abdata.21 %>%
  filter(isotype == "IgG", antigen == "PT") %>%
  ggplot() +
    aes(x=planned_day_relative_to_boost,
         y=MFI_normalised,
         col=infancy_vac,
         group=subject_id) +
    geom_point() +
    geom_line() +
    geom_vline(xintercept=0, linetype="dashed") +
```



```
geom_vline(xintercept=14, linetype="dashed") +
labs(title="2021 dataset IgG PT",
      subtitle = "Dashed lines indicate day 0 (pre-boost) and 14 (apparent peak levels)")
```



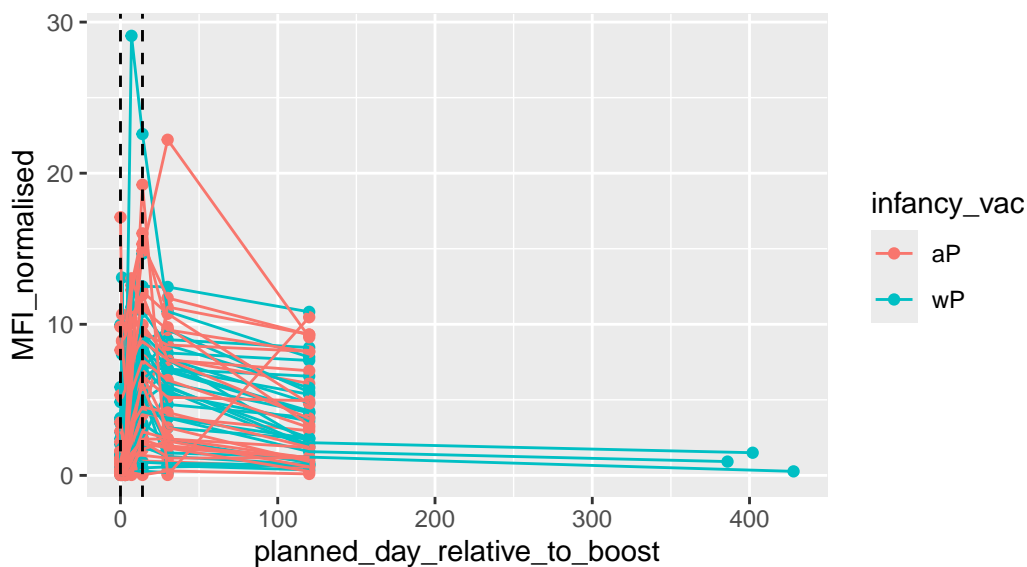
Q19: Does this trend look similar for the 2020 dataset?

```
abdata.20 <- abdata %>% filter(dataset == "2020_dataset")

abdata.20 %>%
  filter(isotype == "IgG", antigen == "PT") %>%
  ggplot() +
    aes(x=planned_day_relative_to_boost,
         y=MFI_normalised,
         col=infancy_vac,
         group=subject_id) +
    geom_point() +
    geom_line() +
    geom_vline(xintercept=0, linetype="dashed") +
    geom_vline(xintercept=14, linetype="dashed") +
    labs(title="2020 dataset IgG PT",
          subtitle = "Dashed lines indicate day 0 (pre-boost) and 14 (apparent peak levels)")
```

2020 dataset IgG PT

Dashed lines indicate day 0 (pre-boost) and 14 (apparent peak levels)



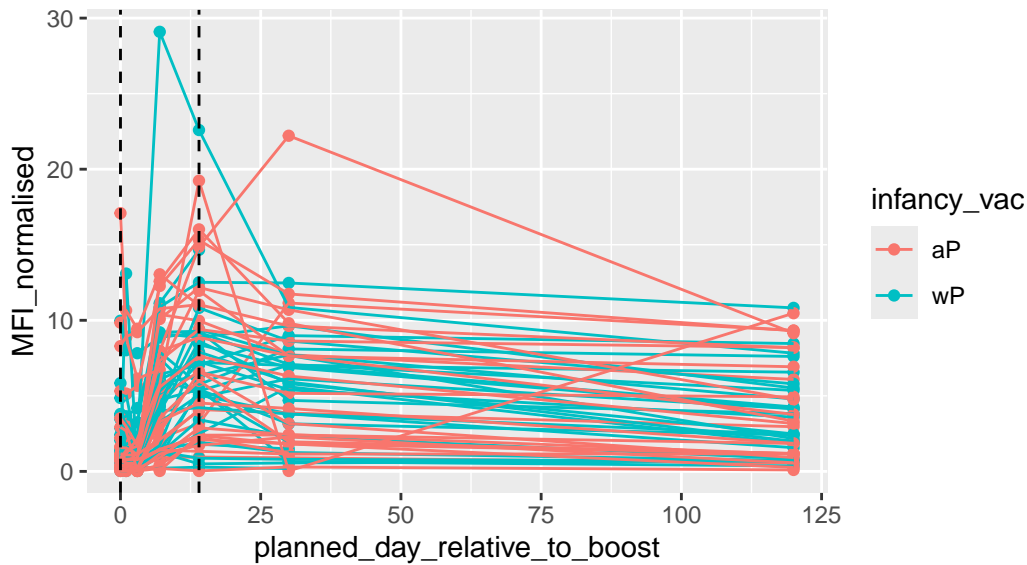
```
abdata.20 %>%
  filter(isotype == "IgG", antigen == "PT") %>%
  ggplot() +
    aes(x=planned_day_relative_to_boost,
         y=MFI_normalised,
         col=infancy_vac,
         group=subject_id) +
    geom_point() +
    geom_line() +
    geom_vline(xintercept=0, linetype="dashed") +
    geom_vline(xintercept=14, linetype="dashed") +
    labs(title="2020 dataset IgG PT",
         subtitle = "Dashed lines indicate day 0 (pre-boost) and 14 (apparent peak levels)") +
    xlim(0, 120)
```

Warning: Removed 3 rows containing missing values or values outside the scale range (`geom_point()`).

Warning: Removed 3 rows containing missing values or values outside the scale range (`geom_line()`).

2020 dataset IgG PT

Dashed lines indicate day 0 (pre-boost) and 14 (apparent peak levels)



The 2020 dataset is much messier. It looks like most of the individuals have PT antigen peaks before 14 days, although it's hard to tell because of how many datapoints there are. There are also individuals with peaks at or after 14 days, resulting in crossing in the lines. There is also an individual with a peak at 14 days but who also has increasing PT antigen levels after around 28 days, which might have a data entry error. Overall, the trend looks different from the 2021 dataset, which could be partly because I think the 2020 dataset also includes previous years.