

# Transcription Factors

ChIP-seq measures TF binding to DNA.

ChIP-seq also measures histone modification, cofactor, and RNA Polymerase genomic locations—however, their occupancy are a consequence of TF binding.

Mike Guertin

# Broad lecture goals:

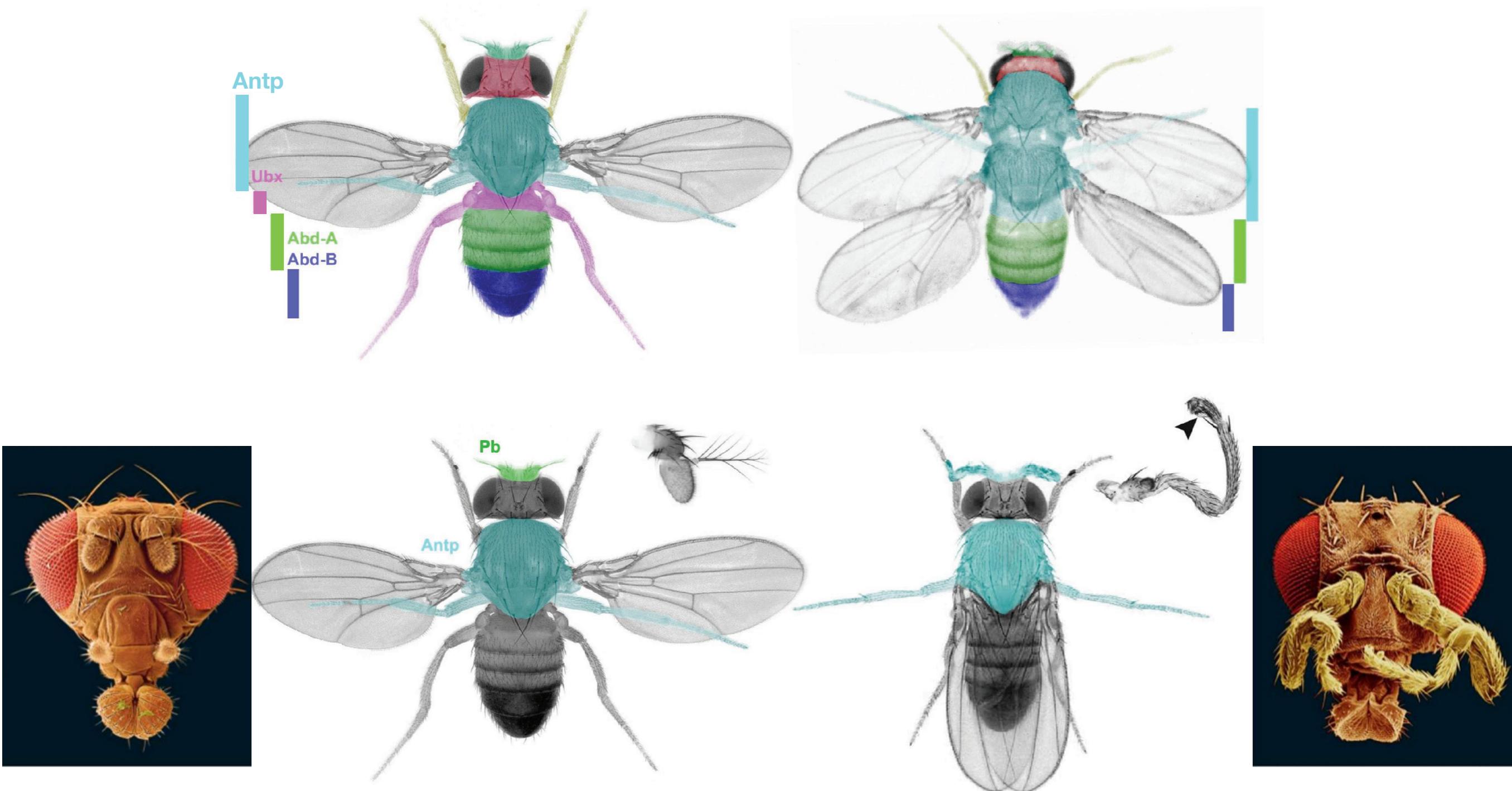
- Convince you of the importance of transcription factors in providing specificity in chromatin biology.
- Introduce classic experiments that defined principles of TF biology and provide references so one can follow up. Note that most molecular biology was interpreted through looking at bands on gels. **As a graduate student, you should aim to be able to take a well-written figure legend and figure and interpret the results.**
- Illustrate the point that biology is continuous, not discrete; relative quantification and controls are important.
- Emphasize the role of question-driven exploratory experiments (screens, molecular genomics, unbiased proximity label transfers, solving structures, etcetera) in defining principles of transcription factor biology.

# Transcription dysregulation alters developmental patterning



pseudocolored flies: Justin Crocker, Ed Lewis, Nicolas Gompel, and Welcome Bender

# Transcription dysregulation alters developmental patterning



pseudocolored flies: Justin Crocker, Ed Lewis, Nicolas Gompel, and Welcome Bender

pseudocolored SEM heads: Jürgen Berger

Classic genetics (perturb, observe, map) found that  
Transcription Factors control developmental patterning

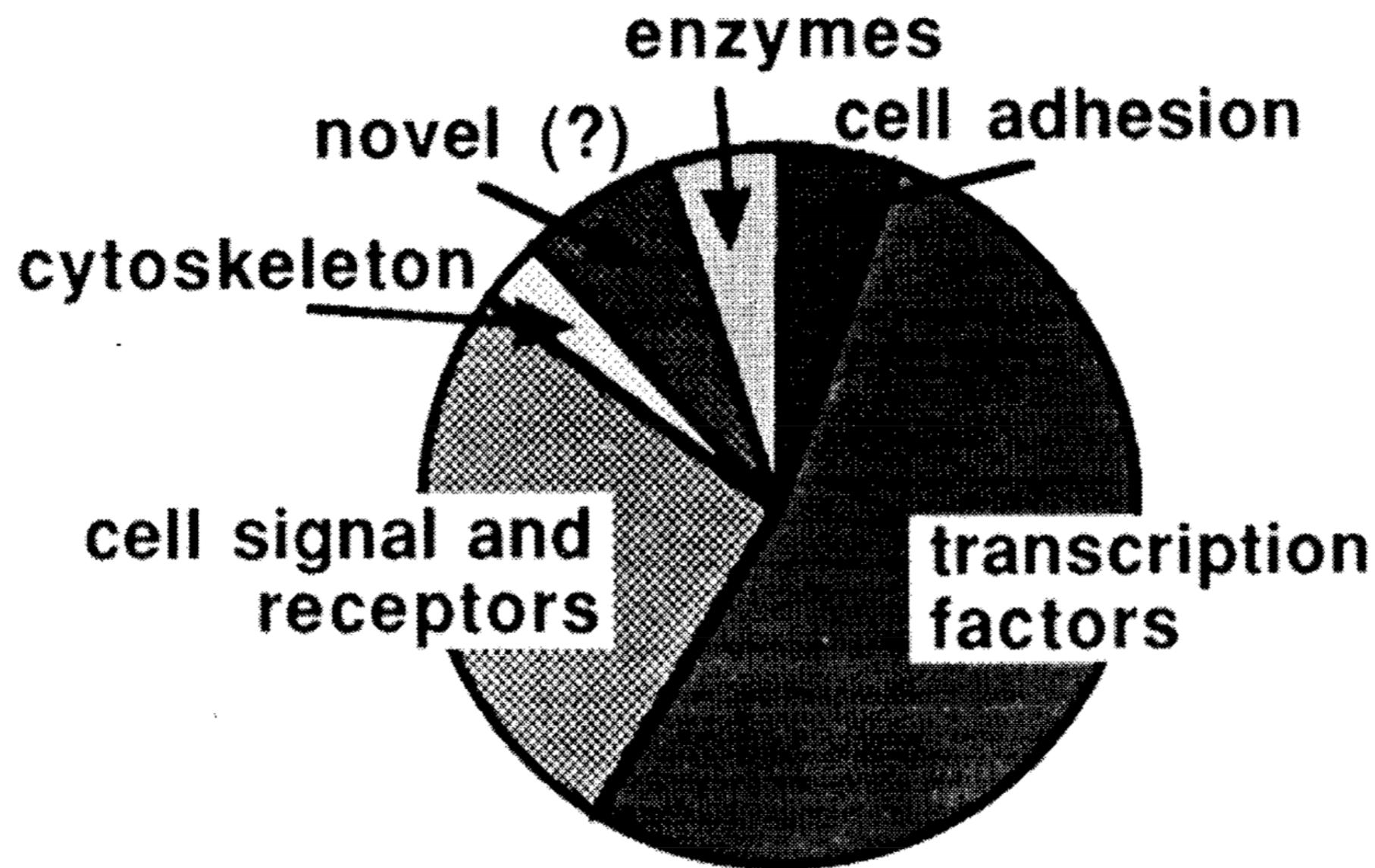
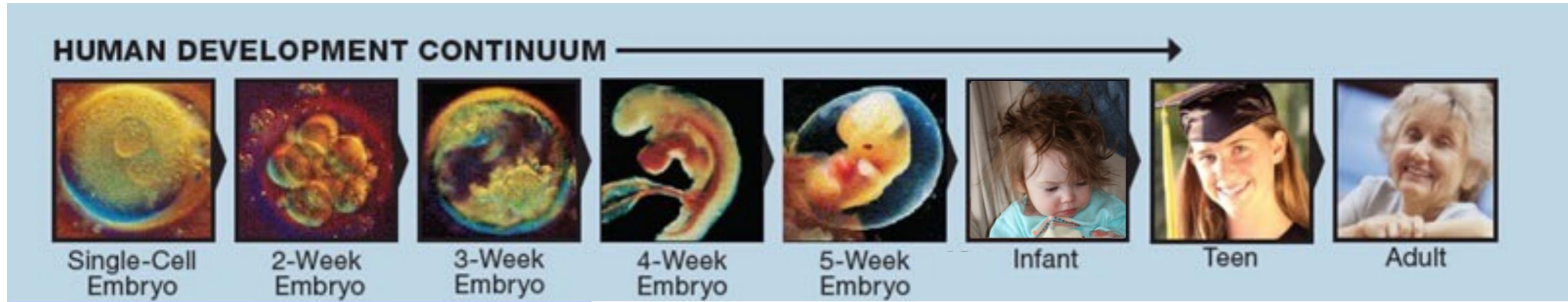
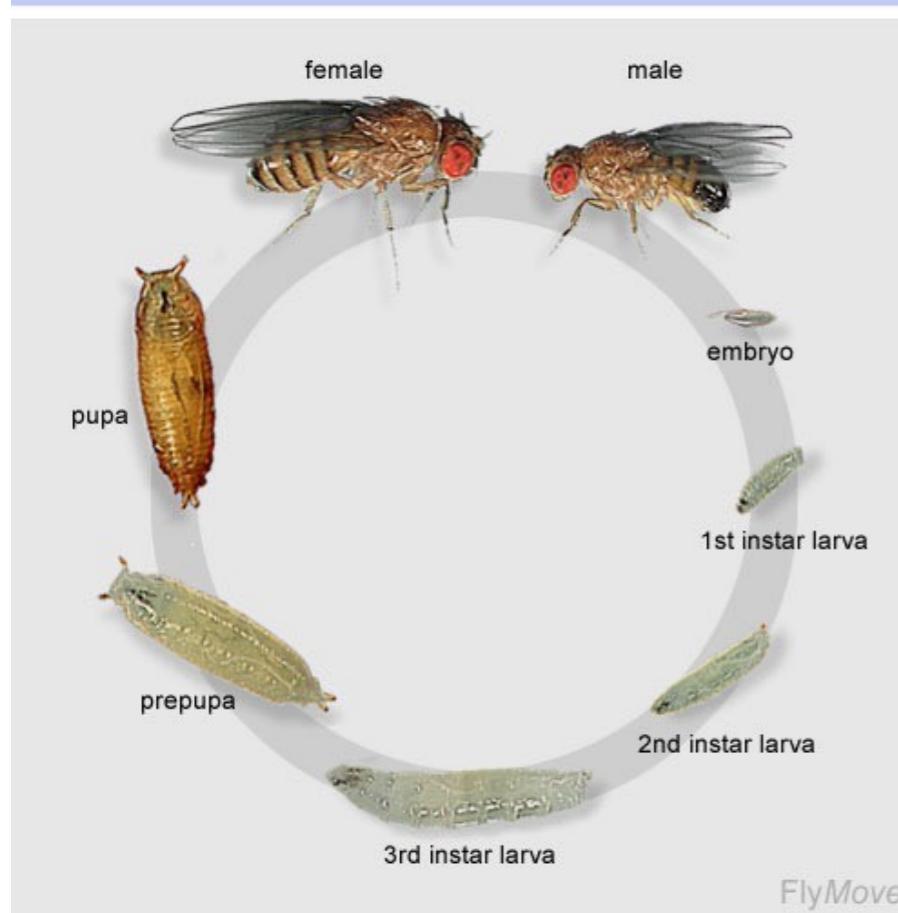


Figure 3. *Cellular Function of Heidelberg Mutations*. Based on the sequence of 75 cloned genes, most of the loci identified in Heidelberg encode transcription factors, or cell signals and receptors.

# Transcription control is key in development and homeostasis



The life cycle of *Drosophila melanogaster*



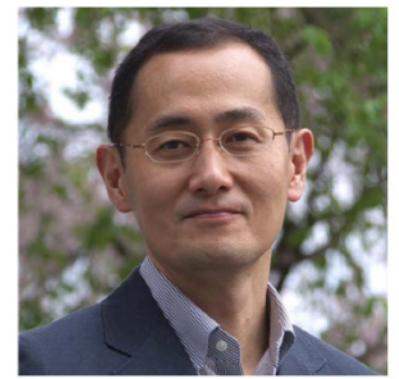
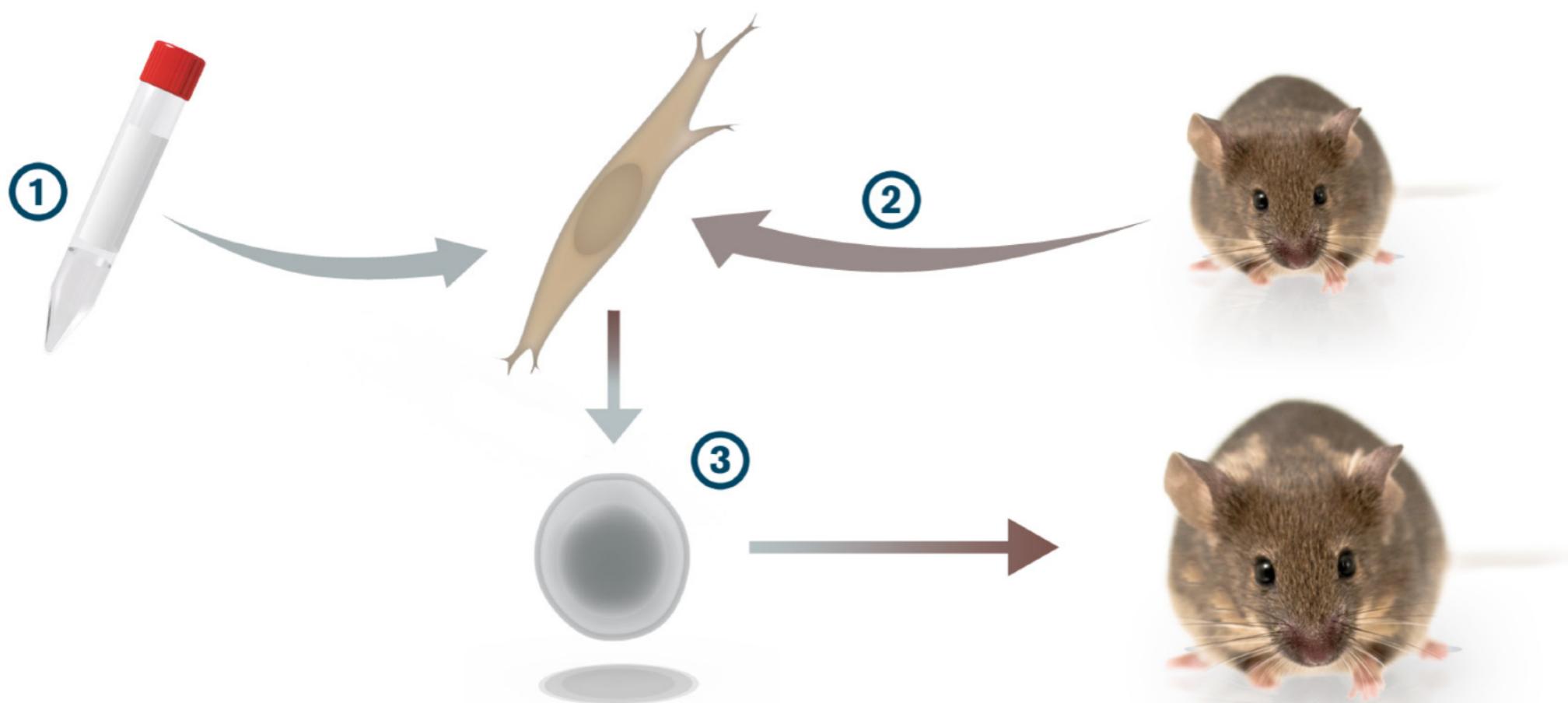
**Embryonic cells progress from totipotent to a spectrum of more specialized states.**

**Much of this developmental regulation starts at transcription.**

**Cells need to respond to changing nutrients and environments.**

**Organisms have sophisticated programs of transcription regulation.**

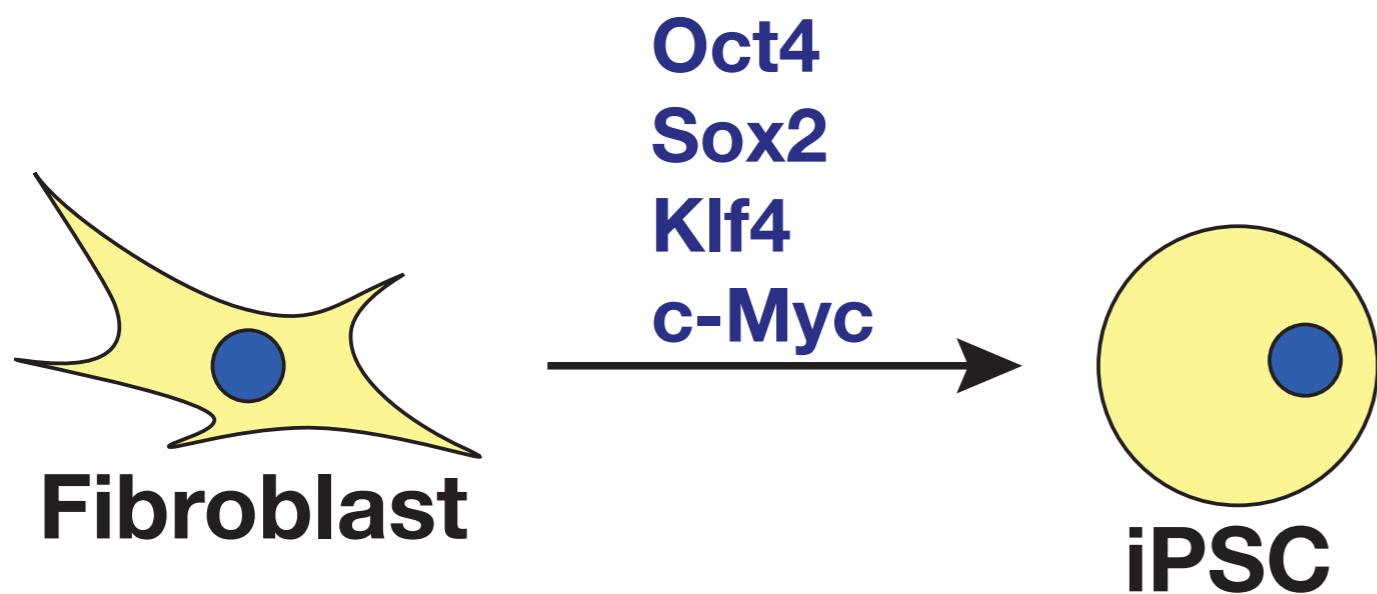
# 2012 Nobel in Physiology or Medicine: “for the discovery that mature cells can be reprogrammed to become pluripotent”

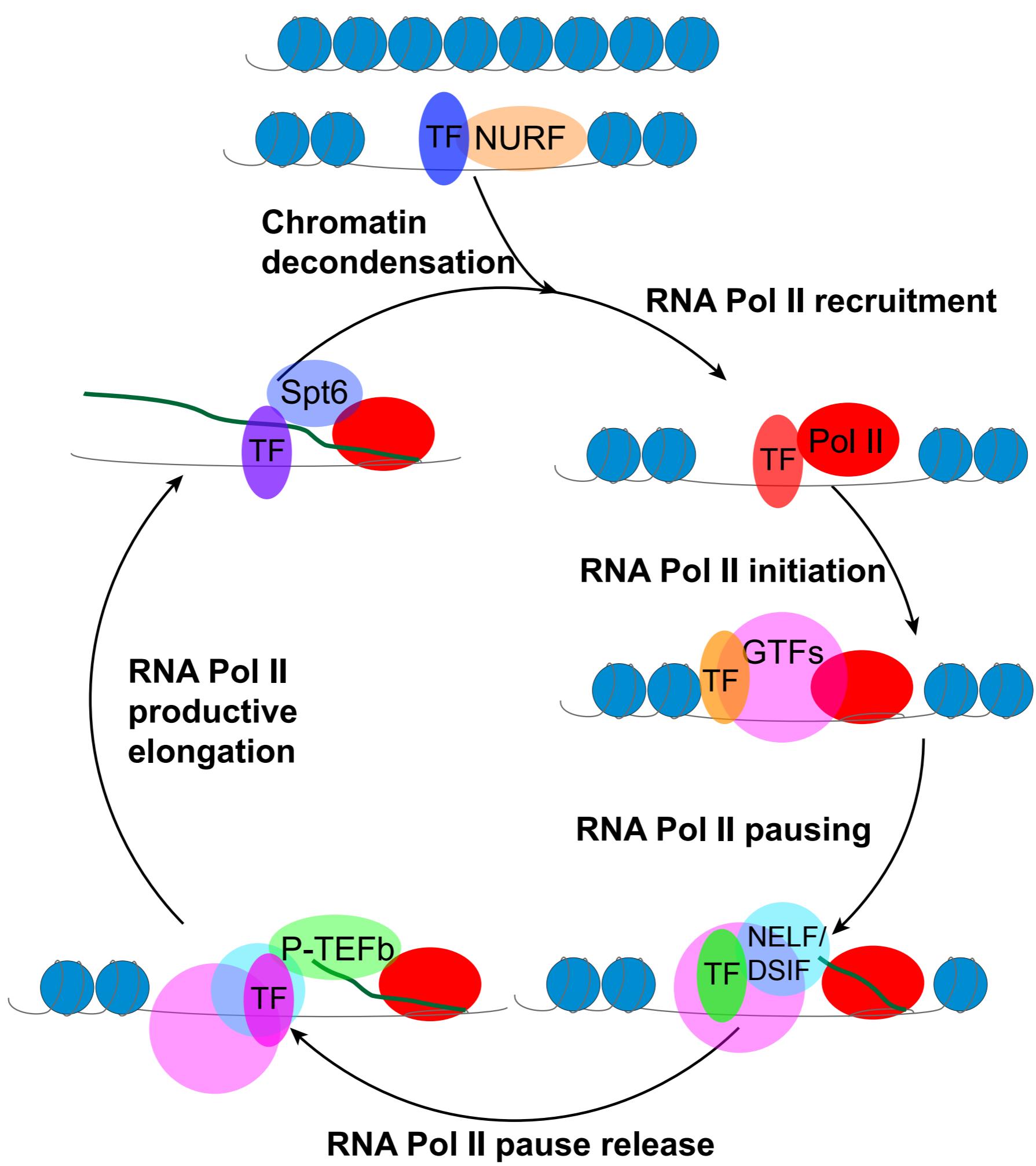


**Shinya Yamanaka**

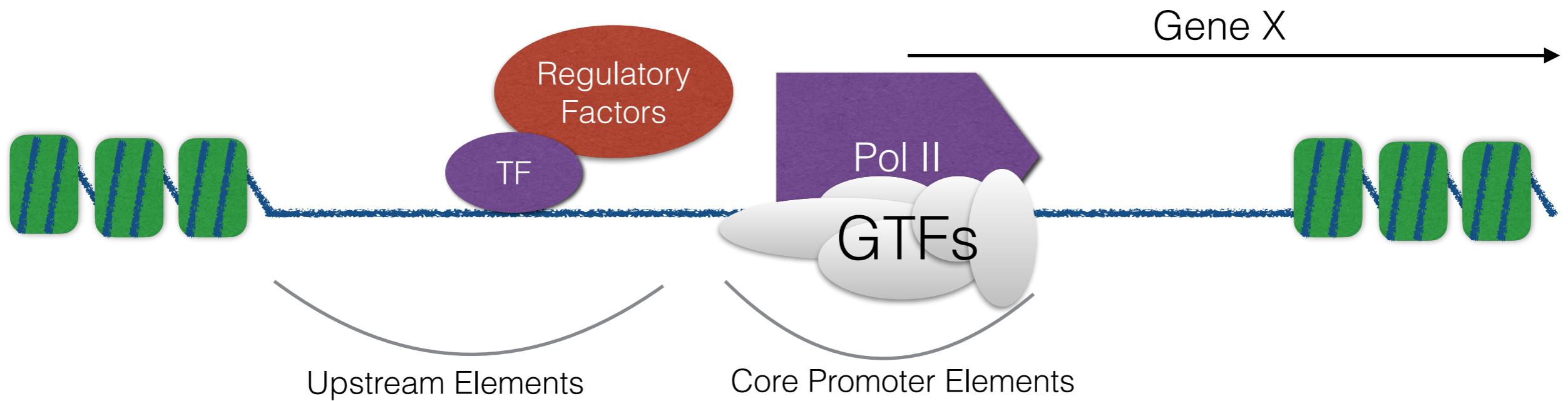
Shinya Yamanaka studied genes that are important for stem cell function. When he transferred four such genes (1) into cells taken from the skin (2), they were reprogrammed into pluripotent stem cells (3) that could develop into all cell types of an adult mouse. He named these cells induced pluripotent stem (iPS) cells.

# Activating transcription factors changes cell identity





# Transcription Regulation by Transcription Factors (TFs) is determined by DNA sequence



# Linker Scanning Mutations of the thymidine kinase gene of HSV

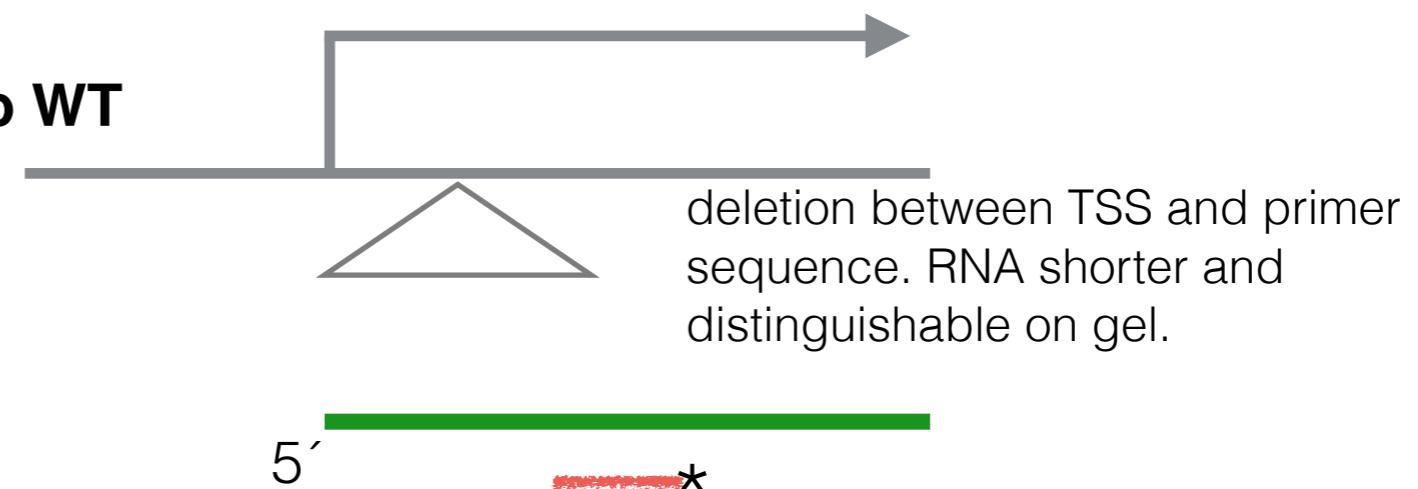
-120            -100            -80            -60            -40            -20            Cap            20  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      WILD TYPE  
**CCCGATCCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -119 -109  
 CTATGCCGGA **T****C****cc** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -115 -105  
 CTATGATGAC **CCG****A****T****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -111 -101  
 CTATGATGAC ACAAAAC**CGG****A** T**CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -105 -95  
 CTATGATGAC ACAAAACCCG CCCAGC**CGG** A**TC****cc** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -95 -85  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC **CCG****G****AT****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -84 -74  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC **CCCG****AT****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -80 -70  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC **G****CCCG****AT****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -79 -69  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC **CCG****C****A****TCC** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -70 -61  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC **A****CCG****G****ATC** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -59 -49  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTC**CCG****G****AT****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -56 -46  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG **CCG****C****G****ATC****C** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -47 -37  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG **CCG****A****TCC****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -42 -32  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT **C****CCG****C****A****TCC** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -29 -18  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA **C****GG****A****TCC****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -21 -12  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTG**CCG****AT****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -18 -6  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTG**CCG****AT****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS -7 +3  
 CTATGATGAC ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACAC**CCG****A****TCC****CCG** ACAAAACCCG CCCAGCGTCT TGTCAATTGGC GAATTGAAAC ACGCAGATGC AGTCGGGGCG GCGCGGTCCG AGGTCCACTT CGCATATTAA GGTGACGCGT GTGGCCTCGA ACACCGAGCG ACCCTGCAGC      LS +5 +15

Clusters of point mutants are generated at the point of joining 5' and 3' deletions, where linker sequence substitutes for tk sequence.

# Assay expression of tk promoter mutants

Plasmid DNAs

Inject mutant DNA into frog oocyte nuclei, include **pseudo WT standard** as internal control



Isolate RNA

Primer Extension Assay

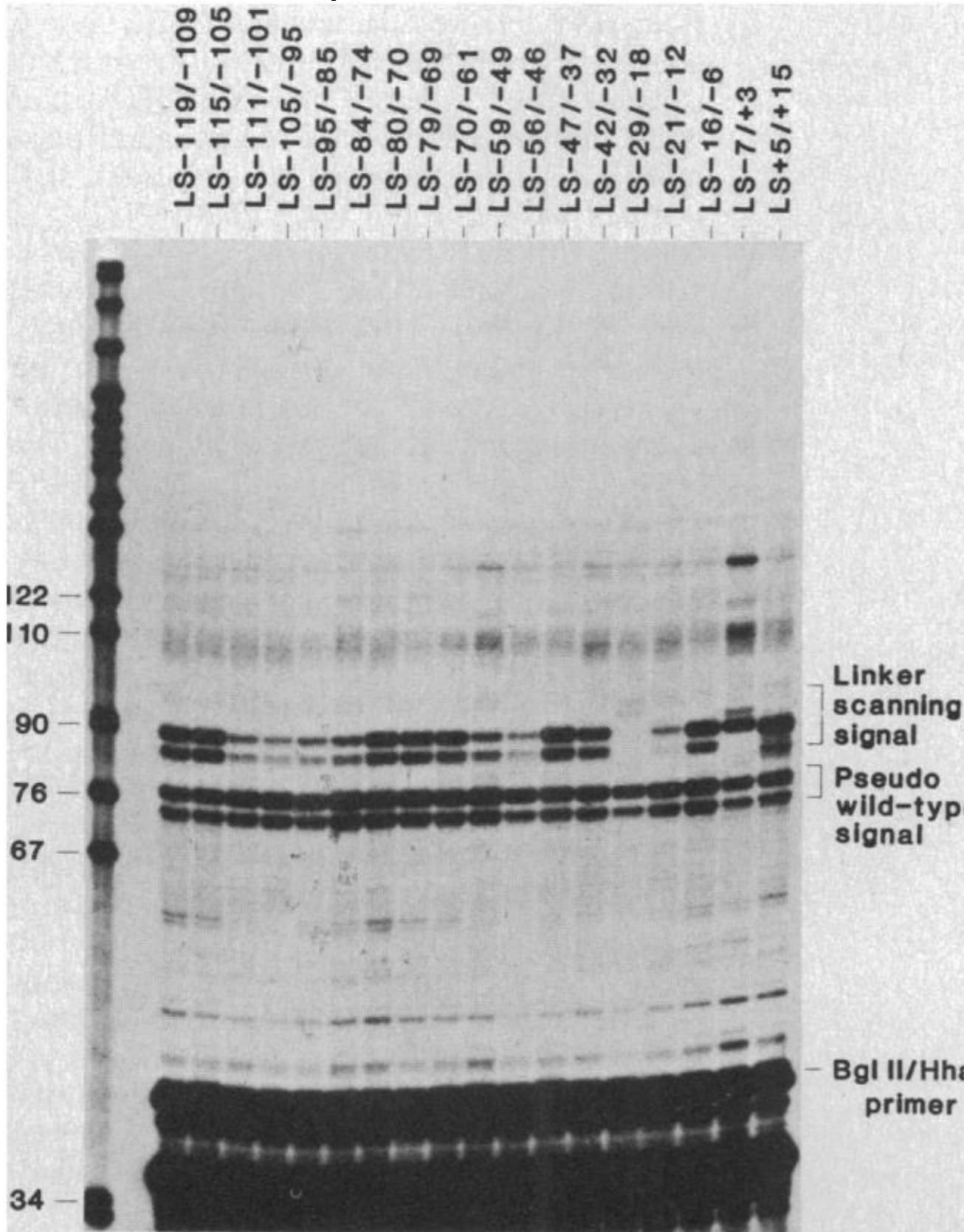
Gel Electrophoresis



tk promoter mutant

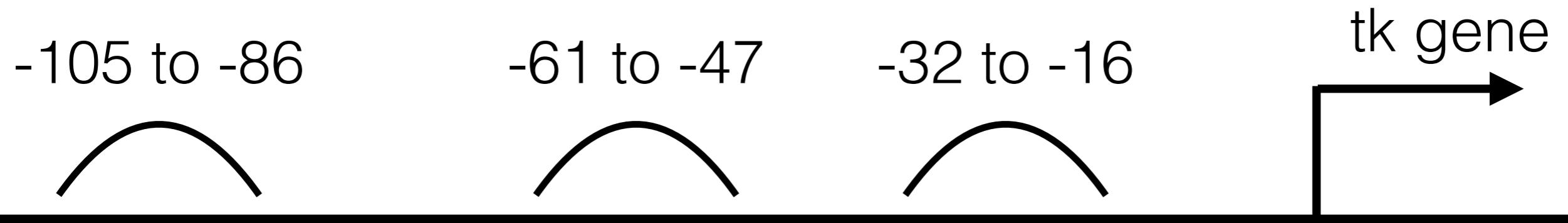
WT tk promoter; deletion of gene body

# Expression data from Linker Scanning Mutants



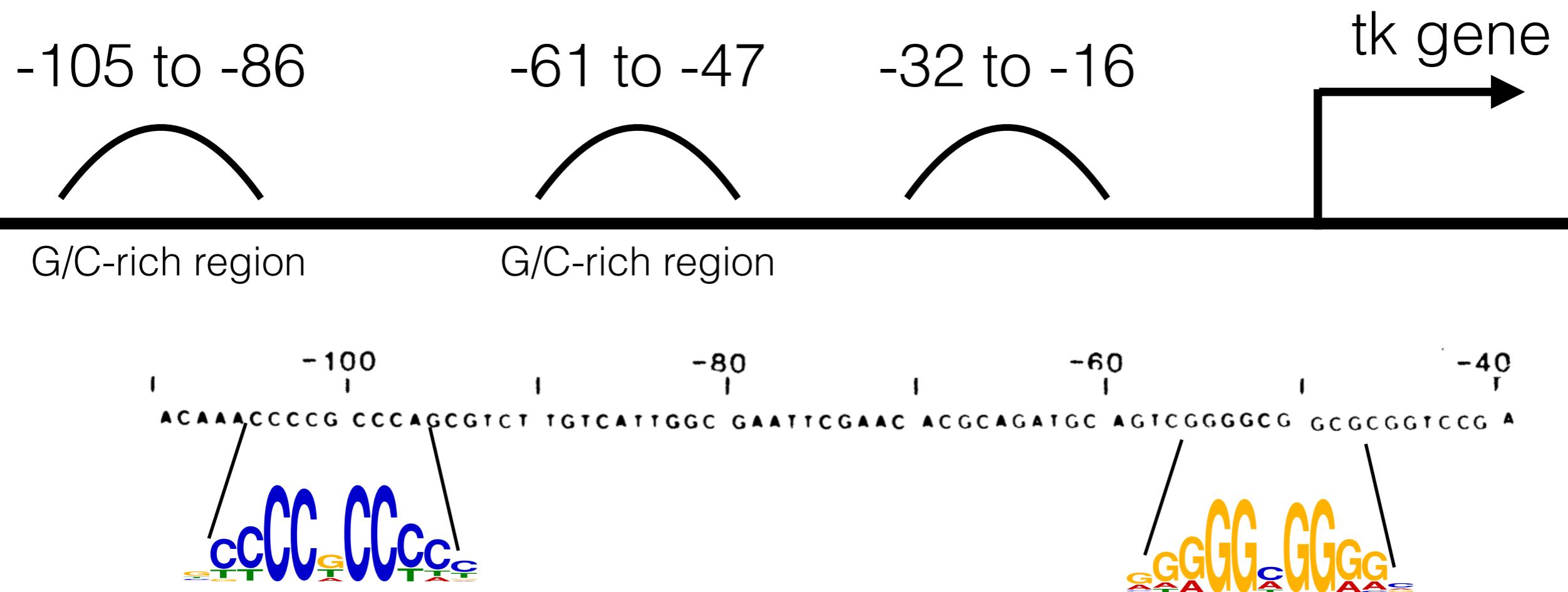
Short discontinuous regions of sequence are critical for basal expression.

Three promoter regions are critical for basal expression

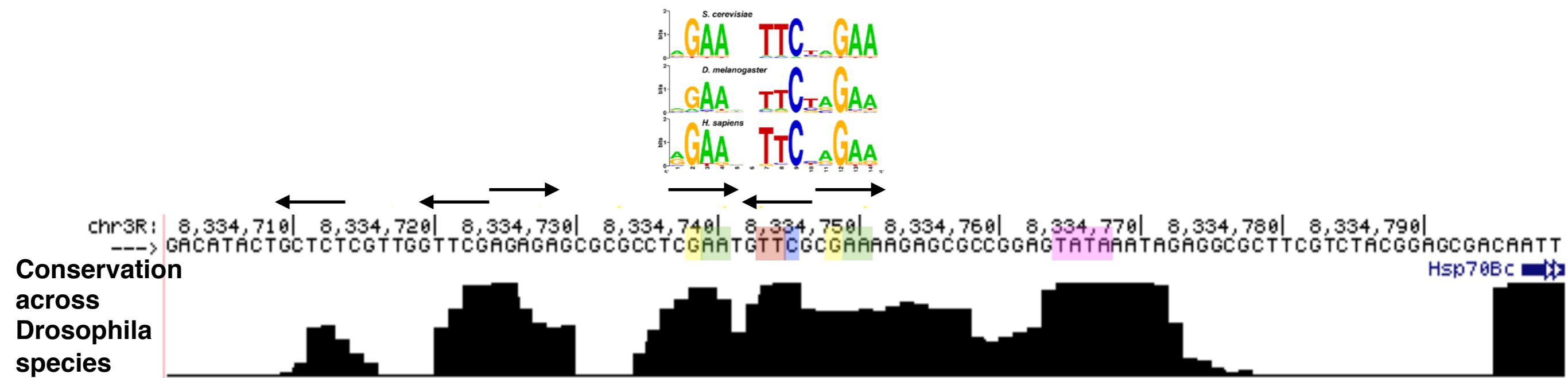


TATA element located  
approximately 30bp  
upstream of the TSS

Three promoter regions are critical for basal expression



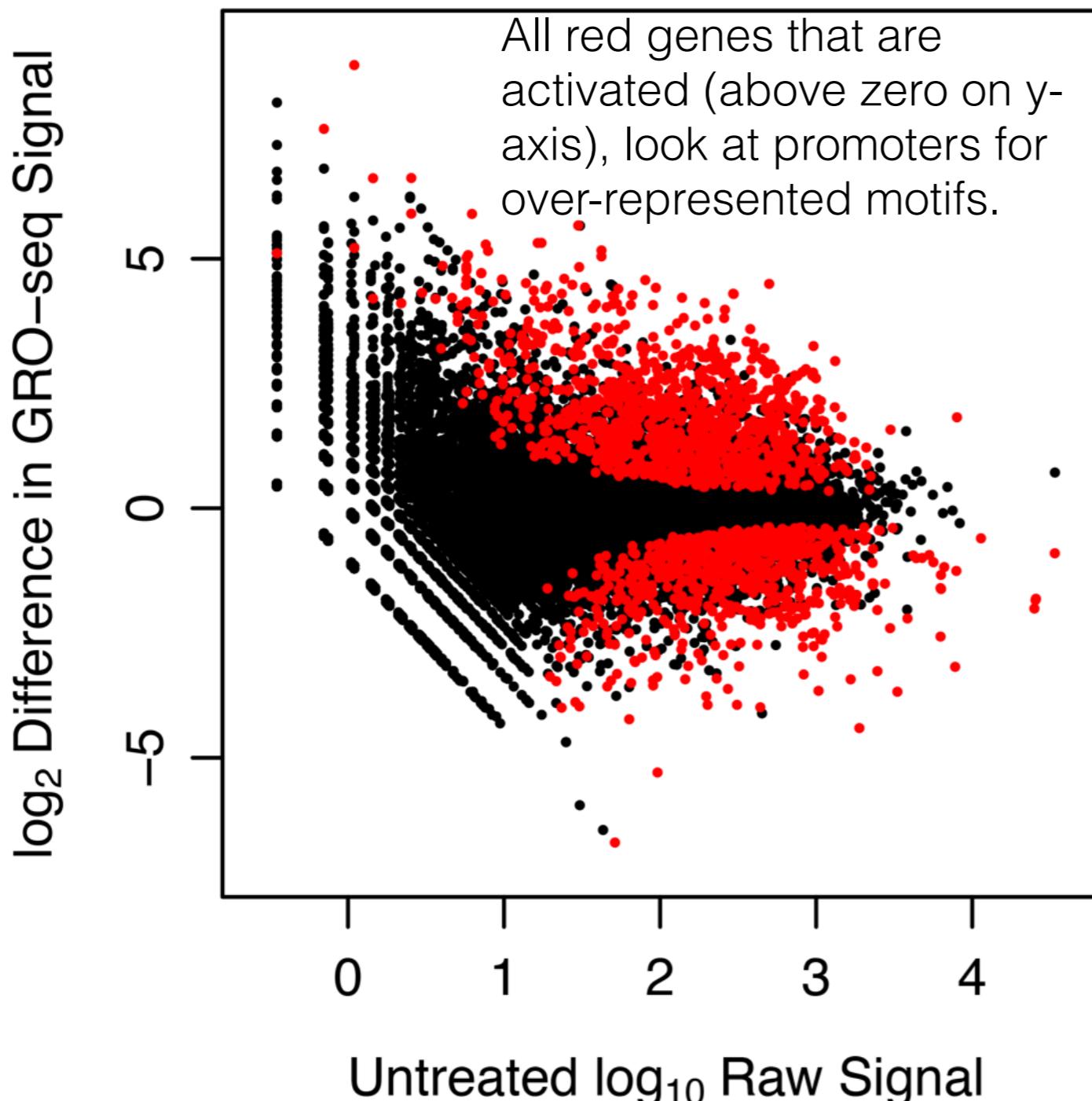
# Evolutionary conservation and comparative genomics can identify crucial elements



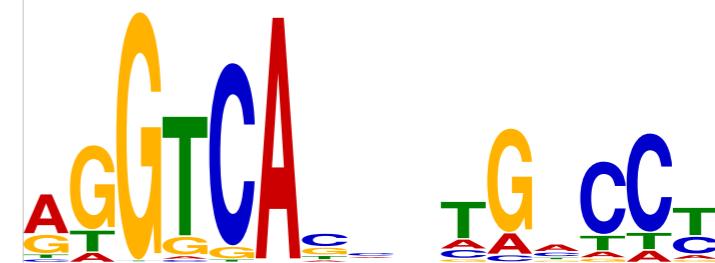
# Significant changes in nascent transcription upon estrogen treatment in breast cancer cells

Collect the sequences of multiple (co-regulated) promoters within a species, search for common sequence motifs

## Sustained Changes at both 10min and 40min



*de novo* motif analysis using MEME (or the alike) identifies the Estrogen Response Element, the known target of the Estrogen Receptor.

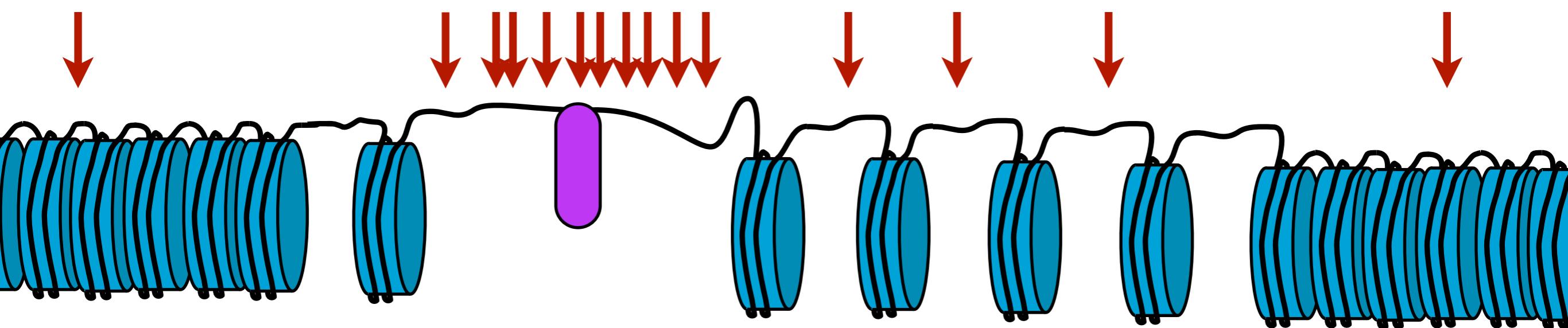


Note that not all regulatory elements bound by TFs are within promoters.

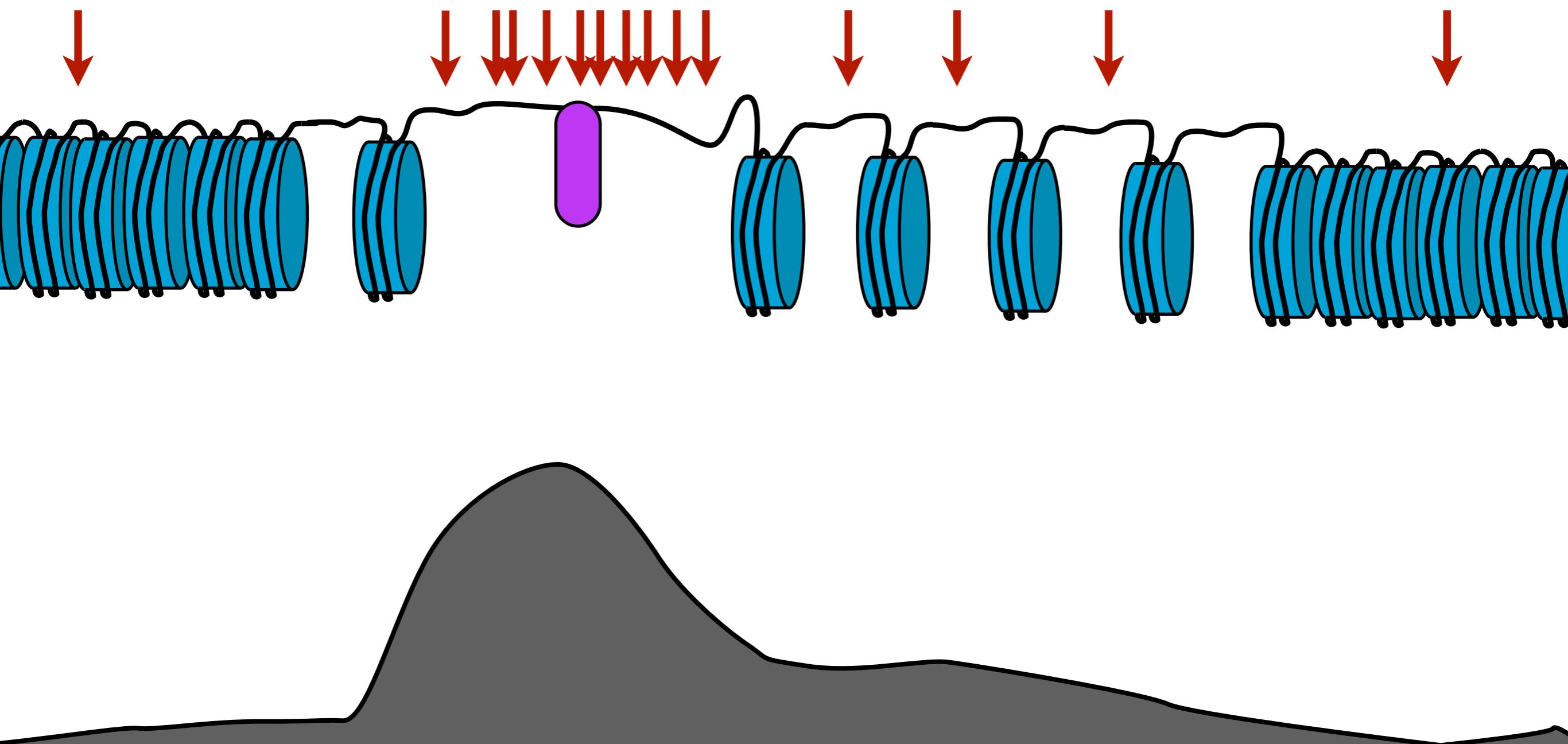
# Identify All Active Regulatory Elements in a Cell Type: Enzyme Hypersensitivity (DNase-seq & ATAC-seq)

- A general measure of chromatin structure.
  - Factor/species-general
  - Changes in enzyme hypersensitivity landscape after drug treatment or throughout development can be used to identify novel regulatory elements and factors
  - Generally unbiased, but challenging to deconvolve
  - **TFs controlling chromatin landscape can be inferred from the data**

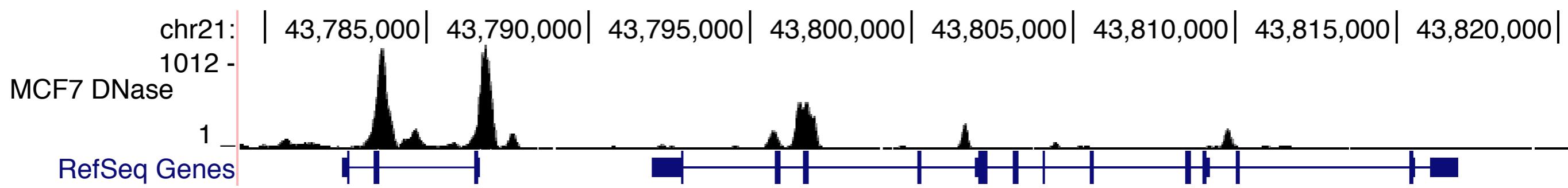
# Enzyme Hypersensitivity



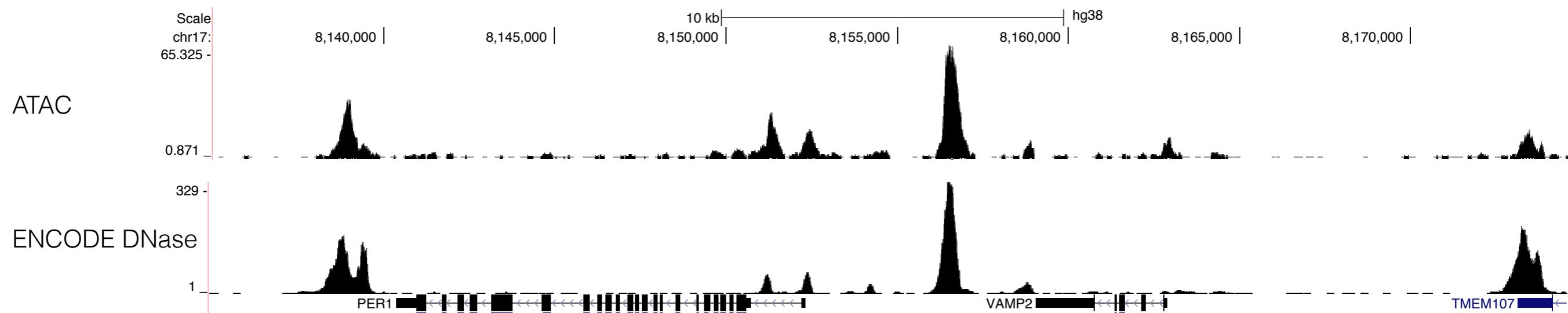
# Enzyme Hypersensitivity



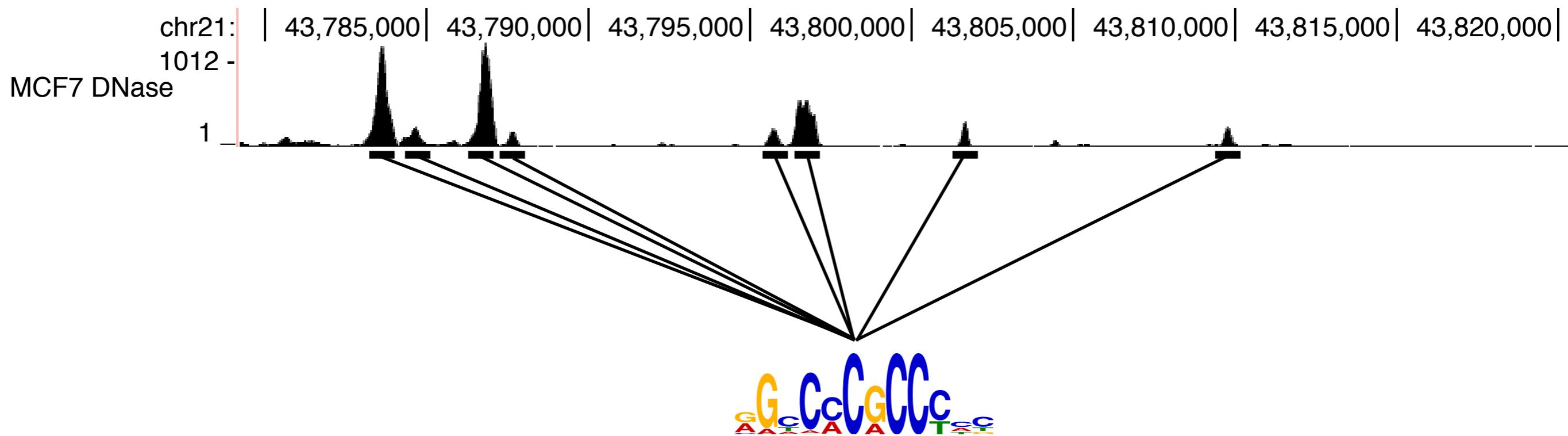
# DNase-seq Data



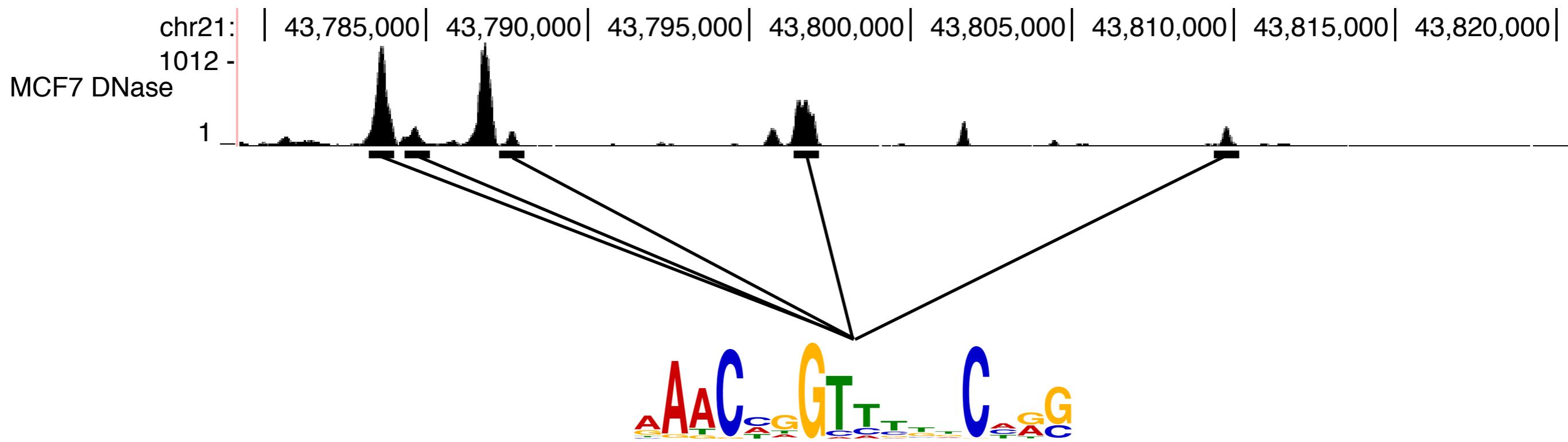
# ATAC-seq vs. DNase-seq



# DNase/ATAC identifies a repertoire of TF motifs

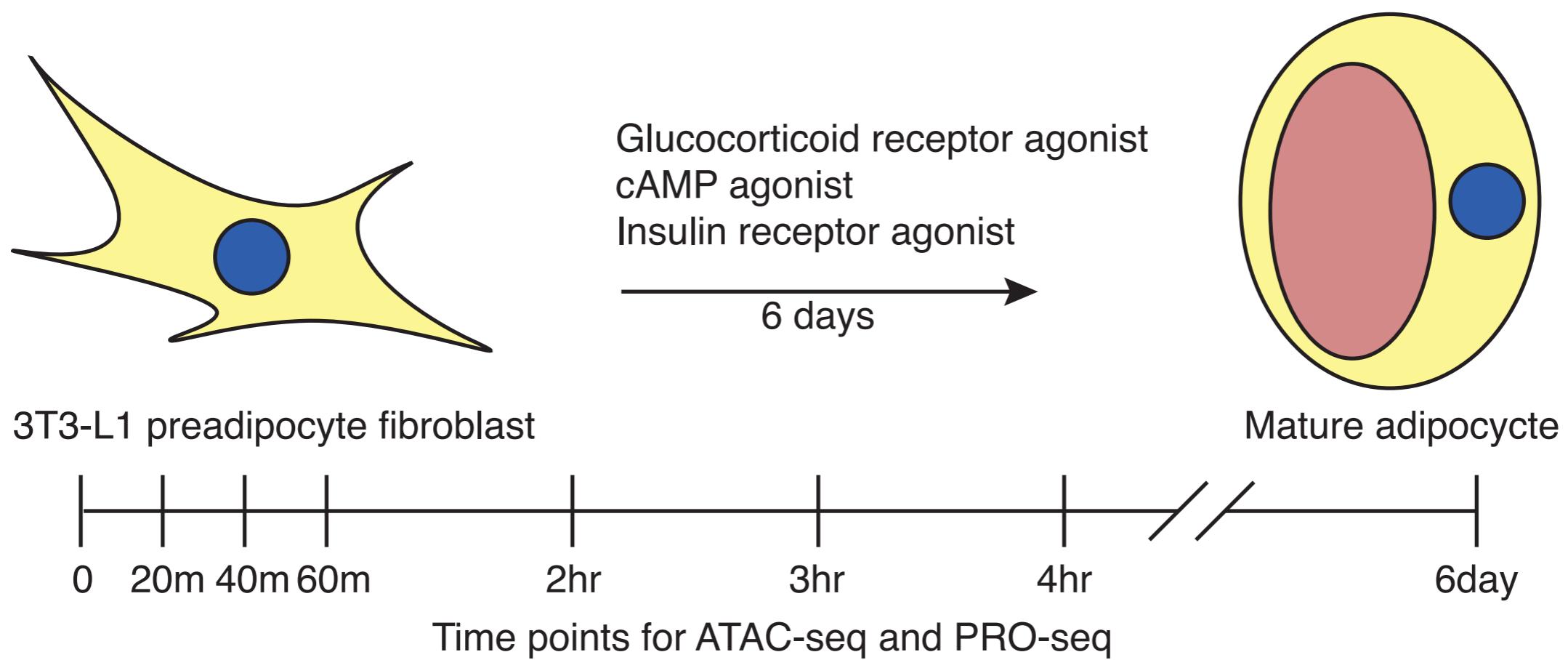


# DNase/ATAC identifies a repertoire of TF motifs

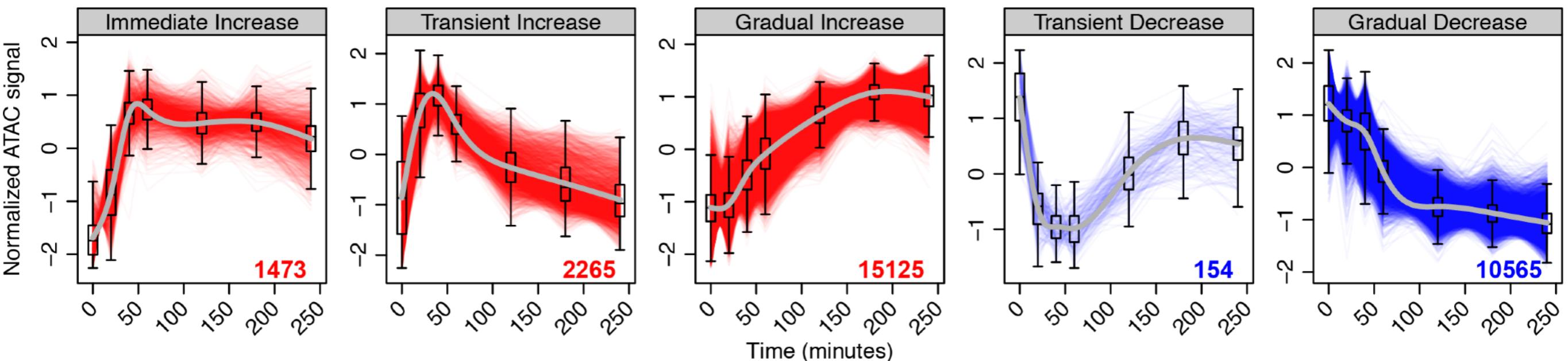


Identify sequence elements at hypersensitive site using iterative de novo motif analysis

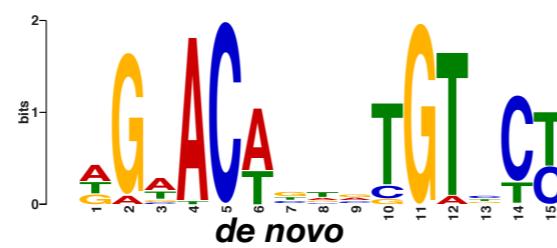
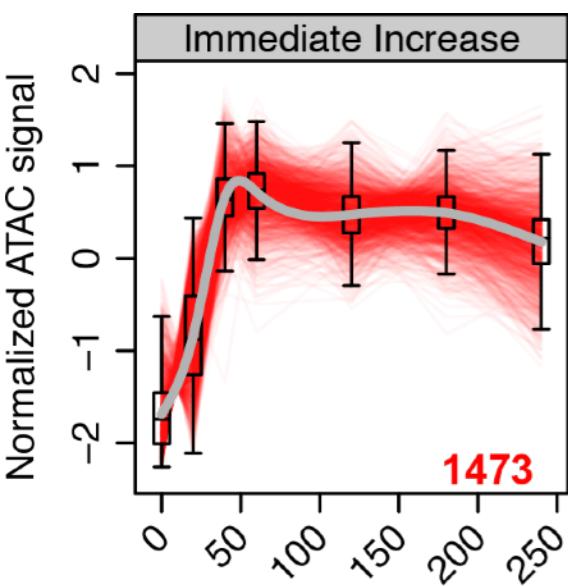
# Experimental Design



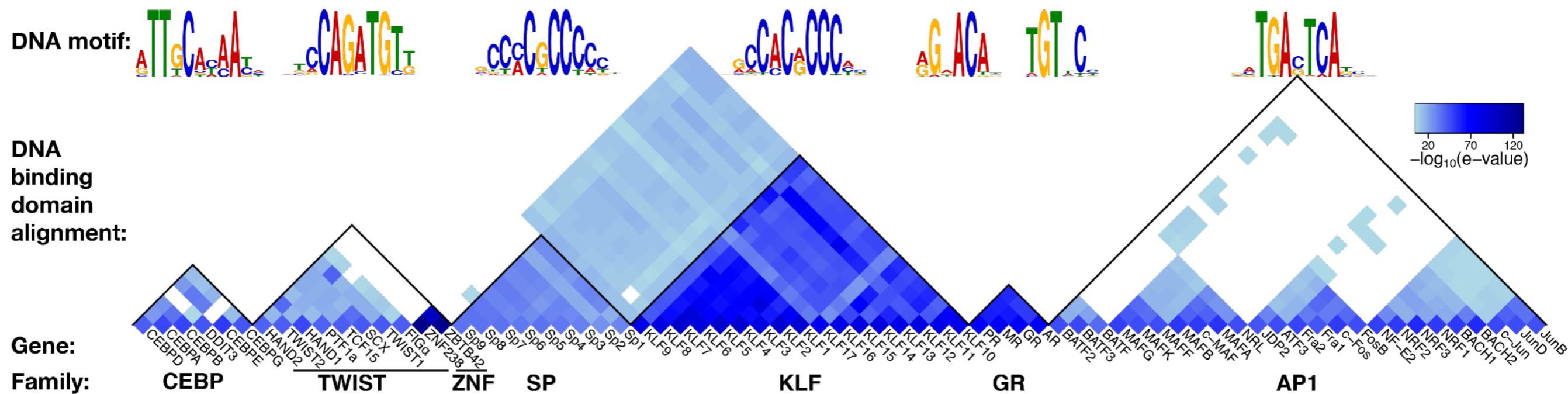
# ATAC-seq peaks have distinct accessibility kinetics



# *de novo* motif analysis identifies enriched sequence elements within dynamic ATAC peaks



# 14 TF-family motifs (top 6 shown) drive early adipogenesis changes in accessibility



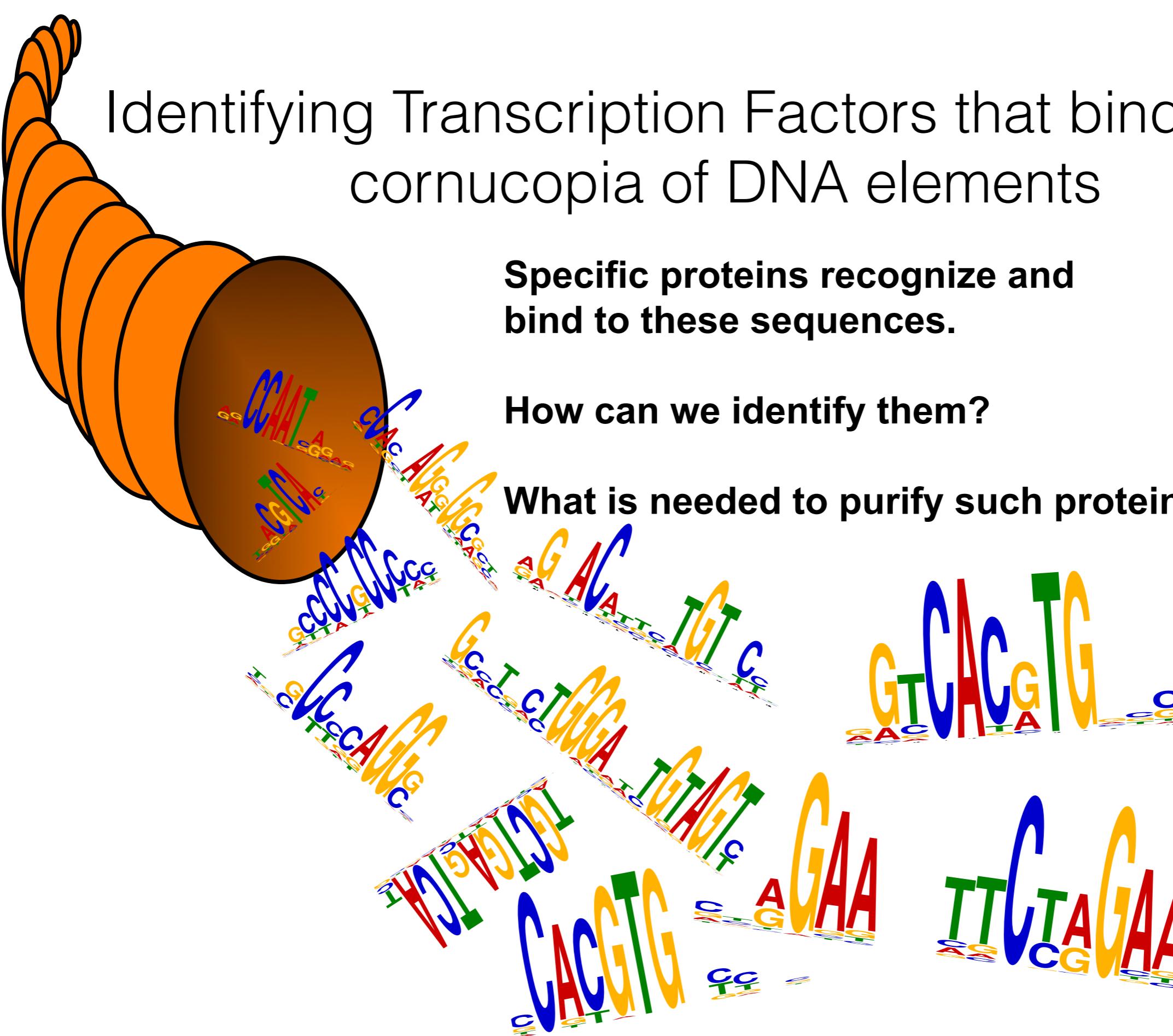
## Paralogous TF DBD families that recognize each motif

# Identifying Transcription Factors that bind to the cornucopia of DNA elements

Specific proteins recognize and bind to these sequences.

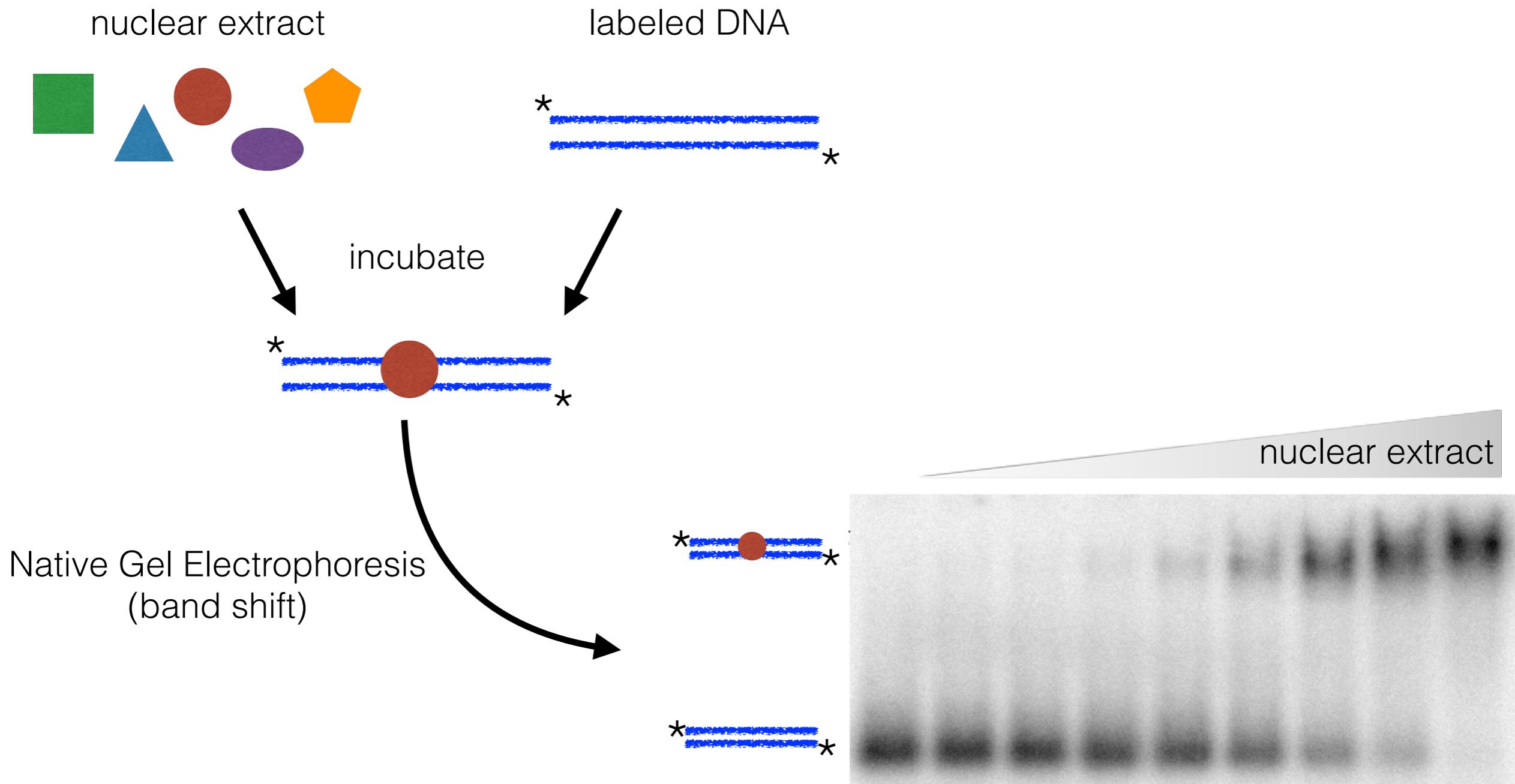
How can we identify them?

What is needed to purify such proteins?

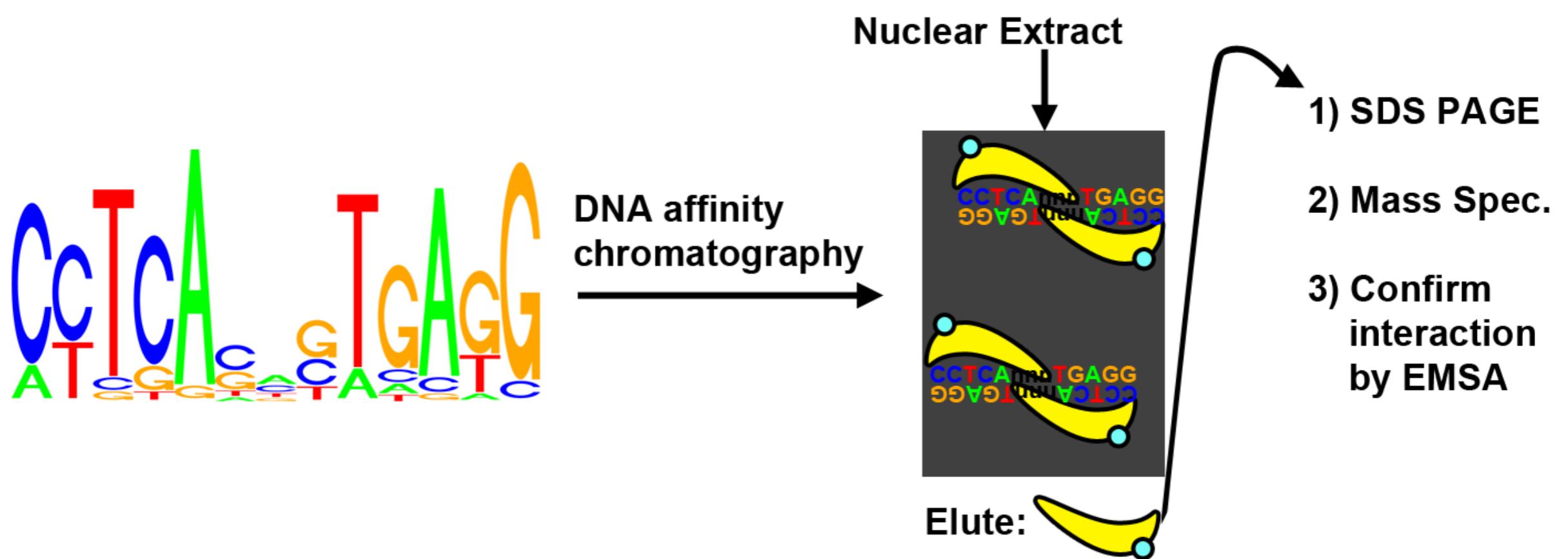


Knowing specific sequences helps create both an assay and tool for purifying.

# Electrophoretic Mobility Shift Assay (EMSA) detect DNA binding factors



# Purification of sequence-specific DNA-binding proteins



Order oligos with modest variants of your consensus sequence (include random flanking DNA). Biotinylate the ends of the duplexed DNA, bind to streptavidin beads/column, elute, compare eluate to nuclear extract by PAGE, and mass spec.

# Summary: Part I

- Transcription and its regulation is specified by short DNA sequence elements.
- These elements interact with particular transcription factors.
- See Lambert et. al., The Human Transcription Factors, Cell 2018 for a review of TF/DNA binding

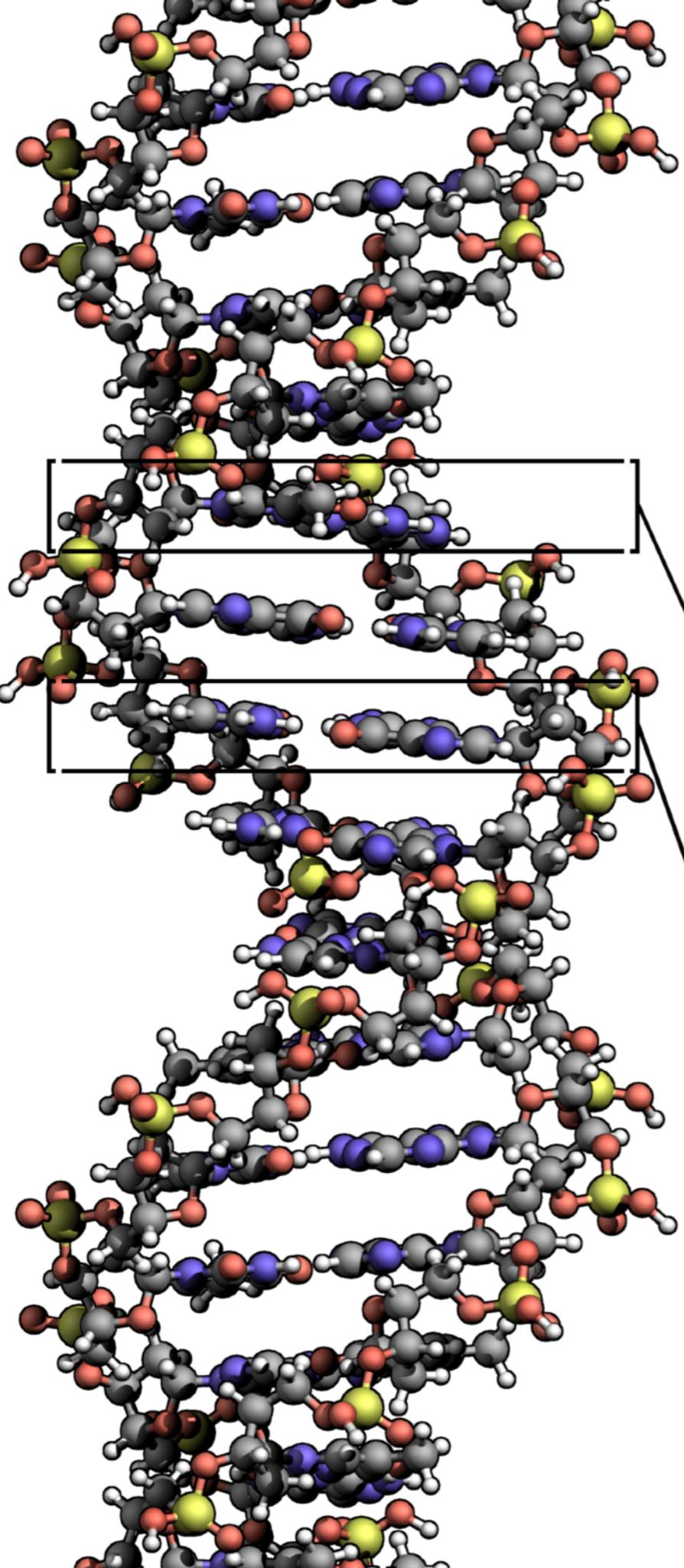
“Next thing is how a cell’s picking which GATs (stretches of nucleotides) get chosen, like Yogi in a picnic basket. Proteins and DNA? Some interesting chemistry. Cuz they getting jiggy with some different affinities.”

–Tom McFadden

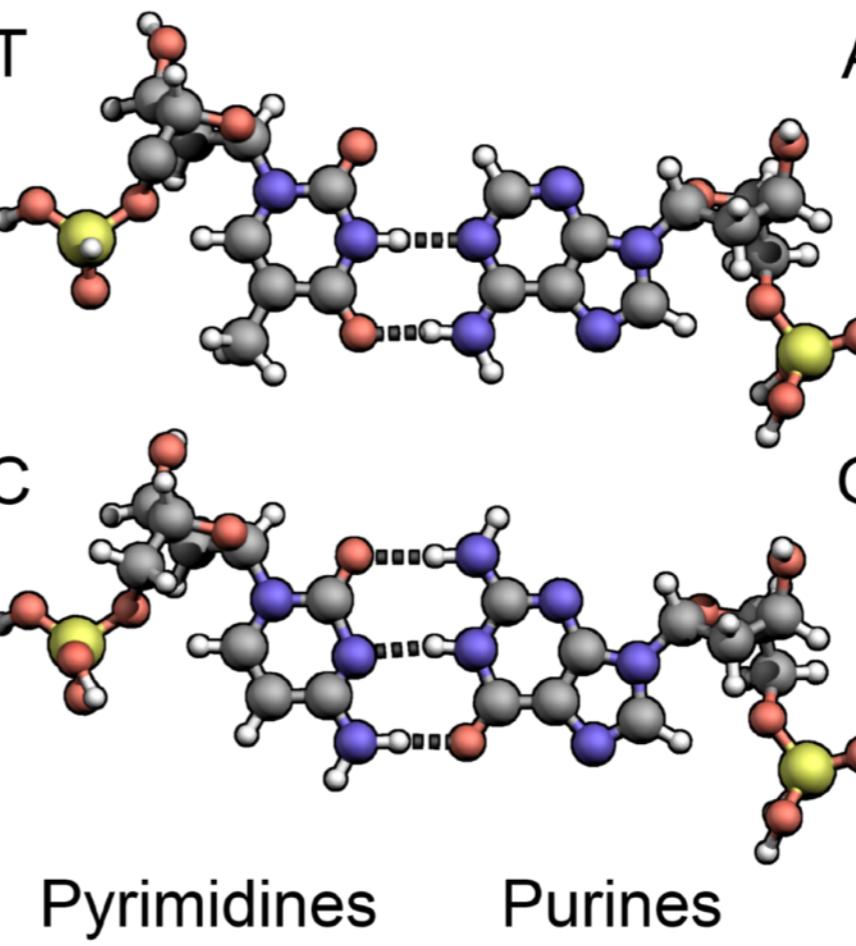
[https://www.youtube.com/watch?v=9k\\_oKK4Teco&list=RD9k\\_oKK4Teco](https://www.youtube.com/watch?v=9k_oKK4Teco&list=RD9k_oKK4Teco)

# How do proteins interact with specific DNA sequences?

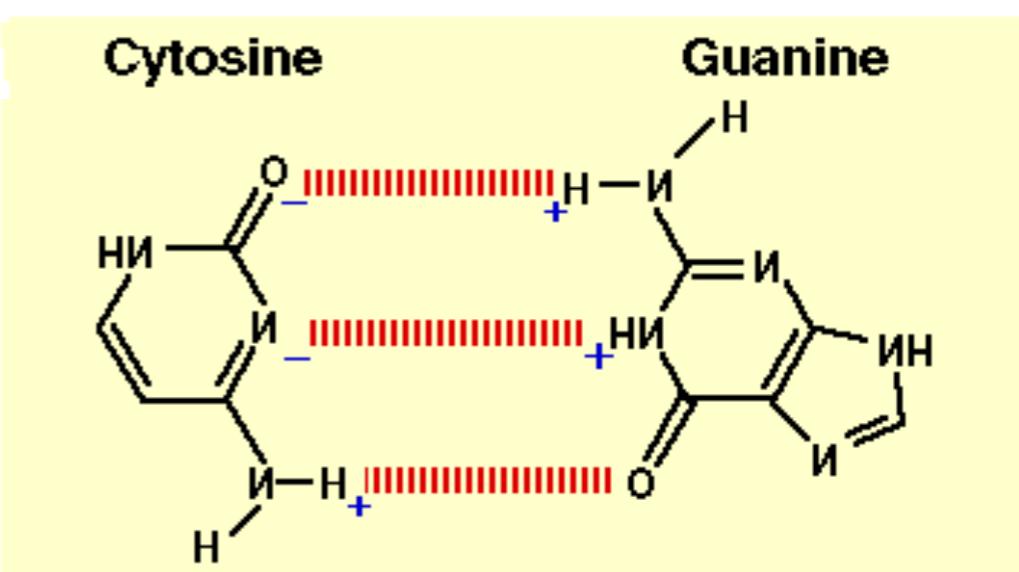
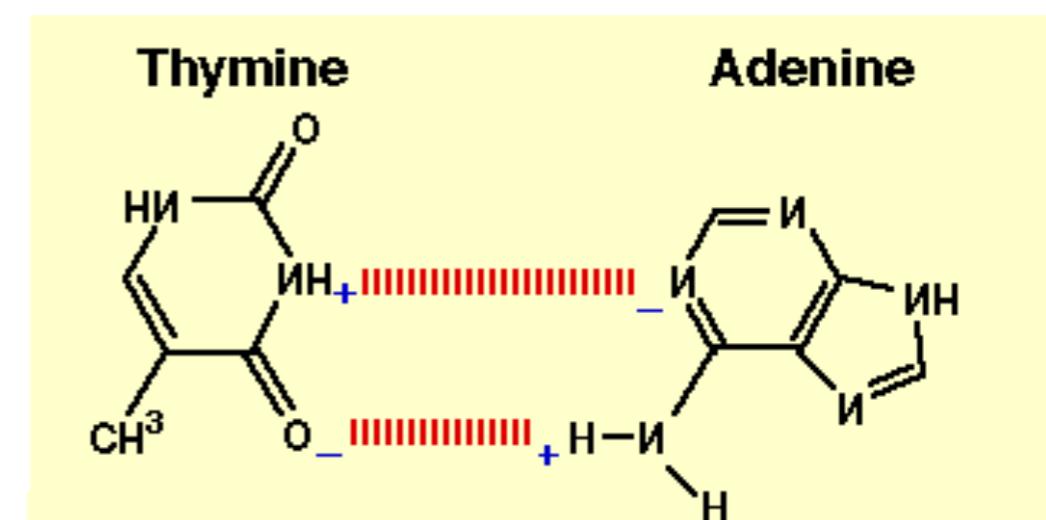
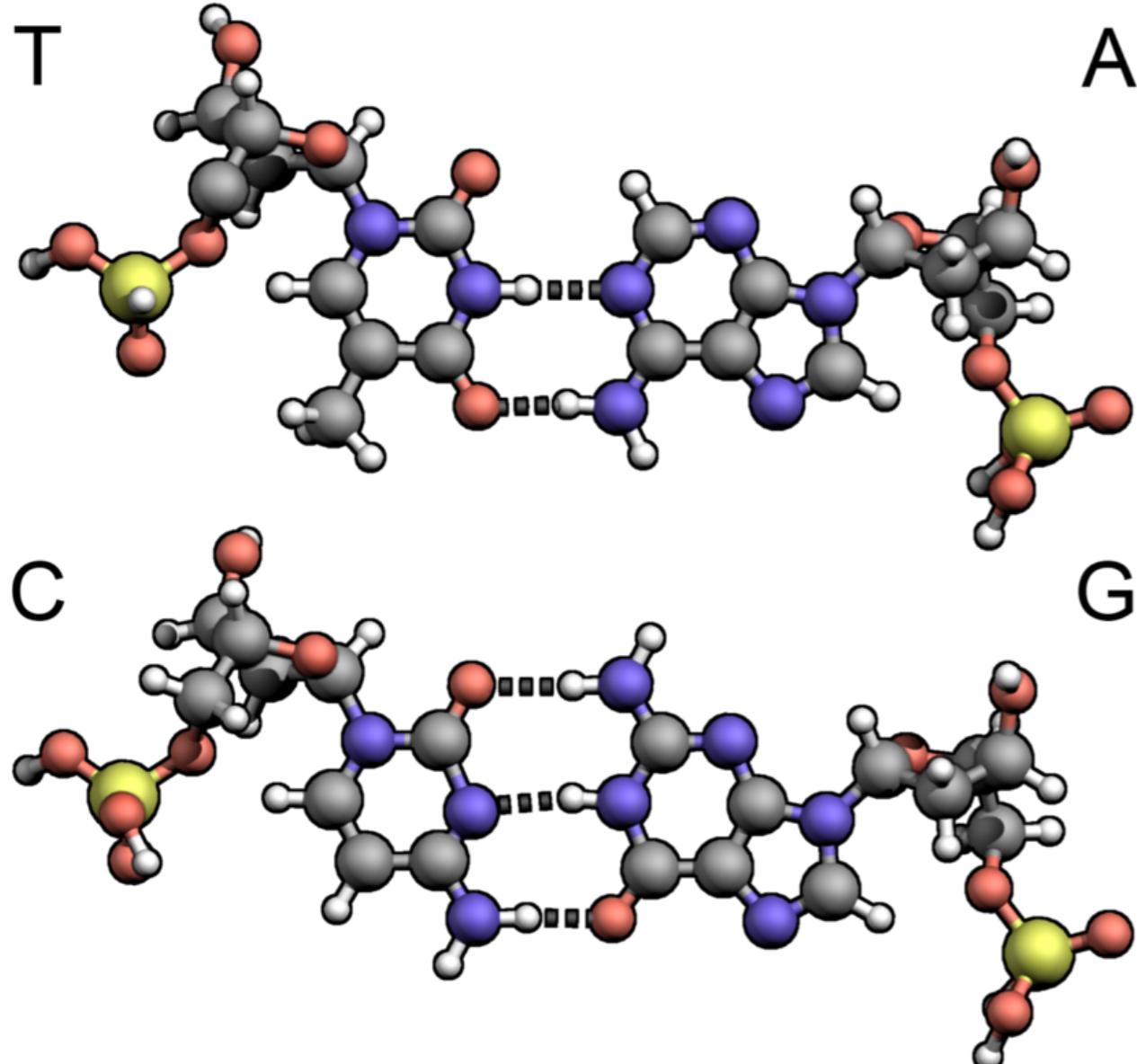
Major groove  
Minor groove



- Hydrogen
- Oxygen
- Nitrogen
- Carbon
- Phosphorus

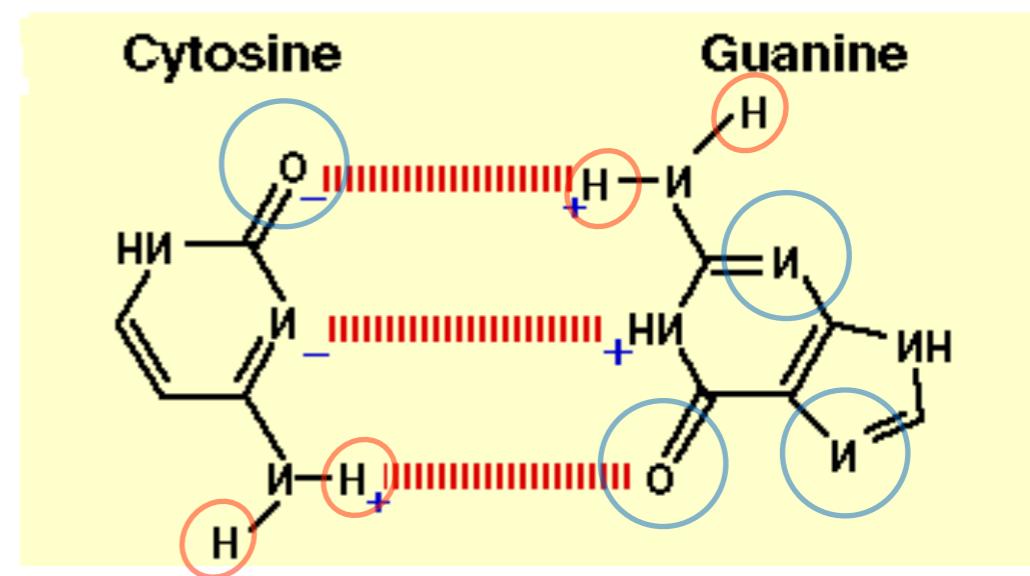
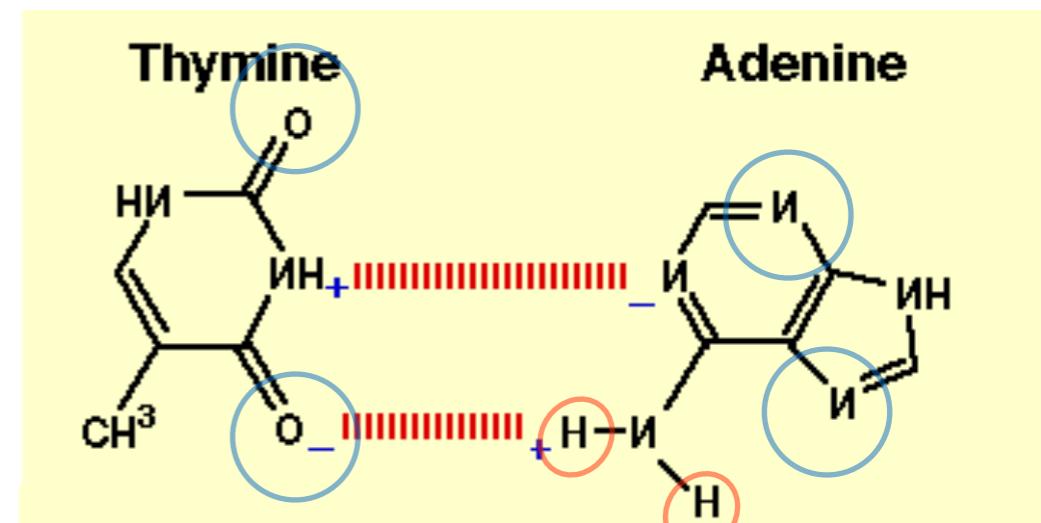
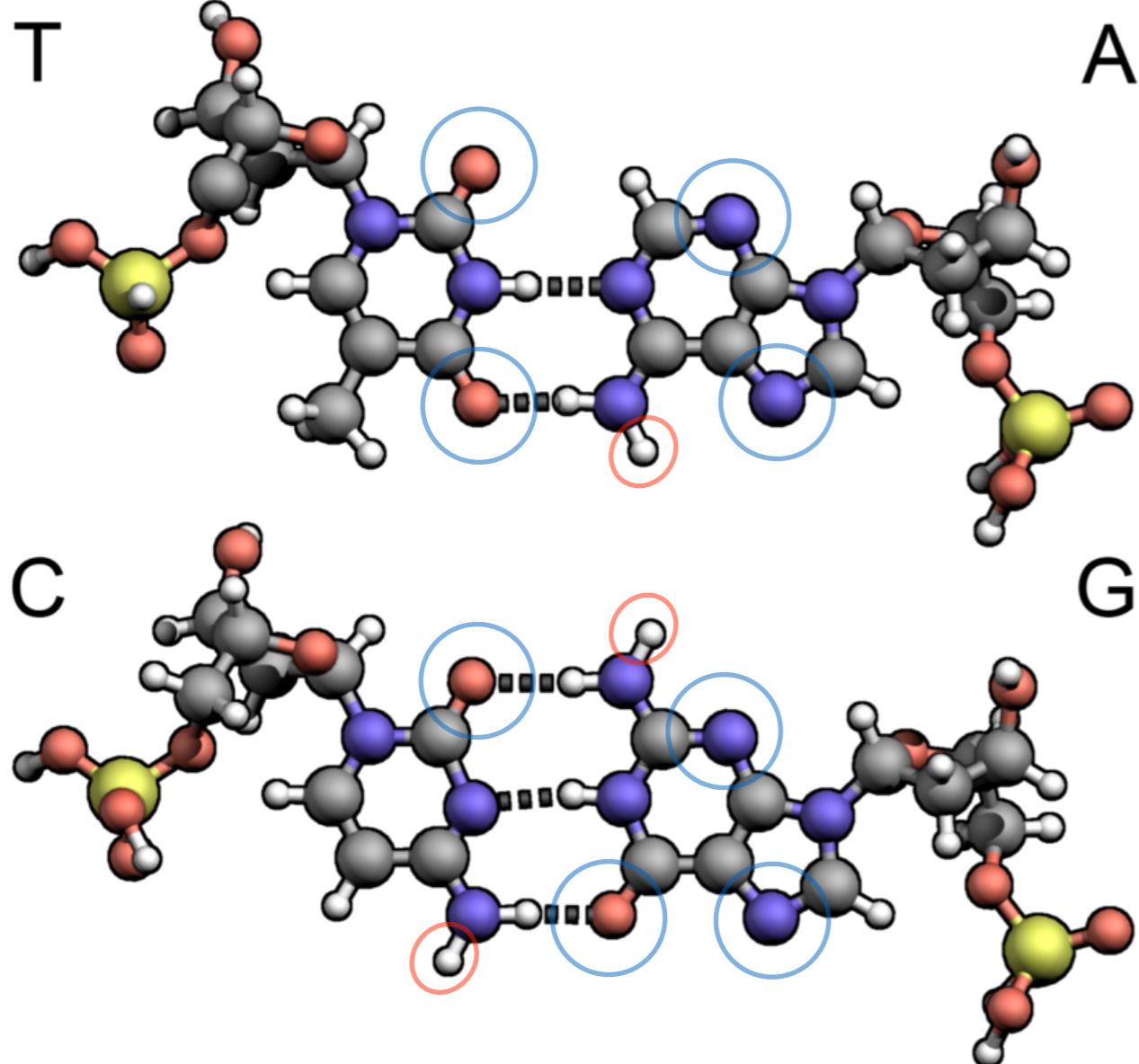


# Hydrogen bond is the electrostatic attraction between polar groups

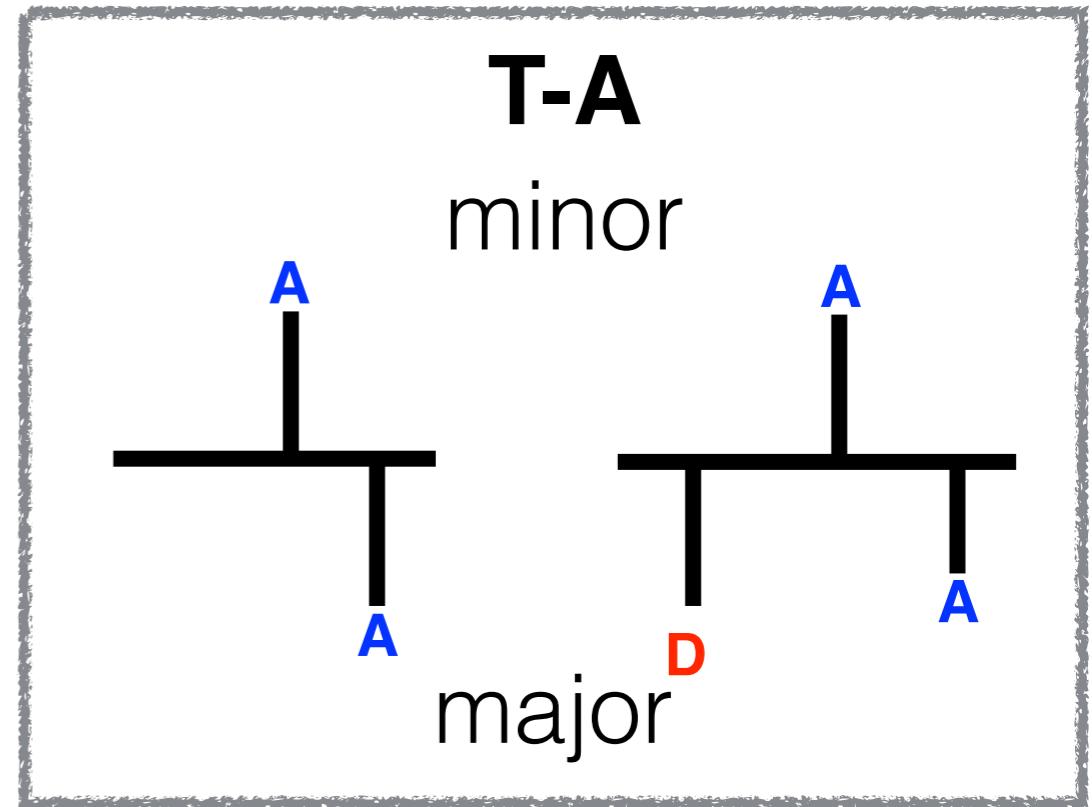
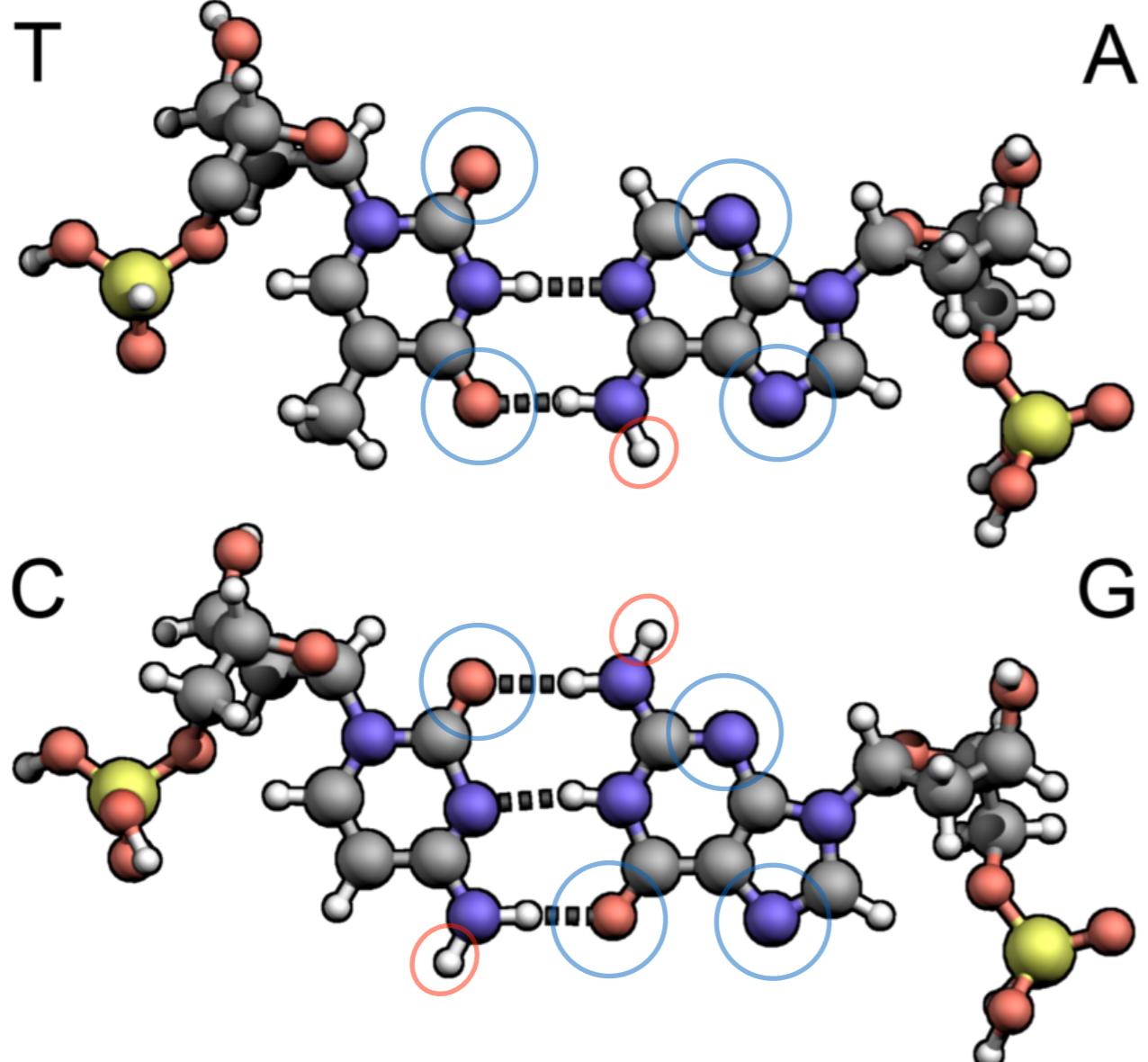


H-bond: a Hydrogen atom bound to a highly electronegative atom such as Nitrogen or Oxygen experiences attraction to another nearby highly electronegative atom.

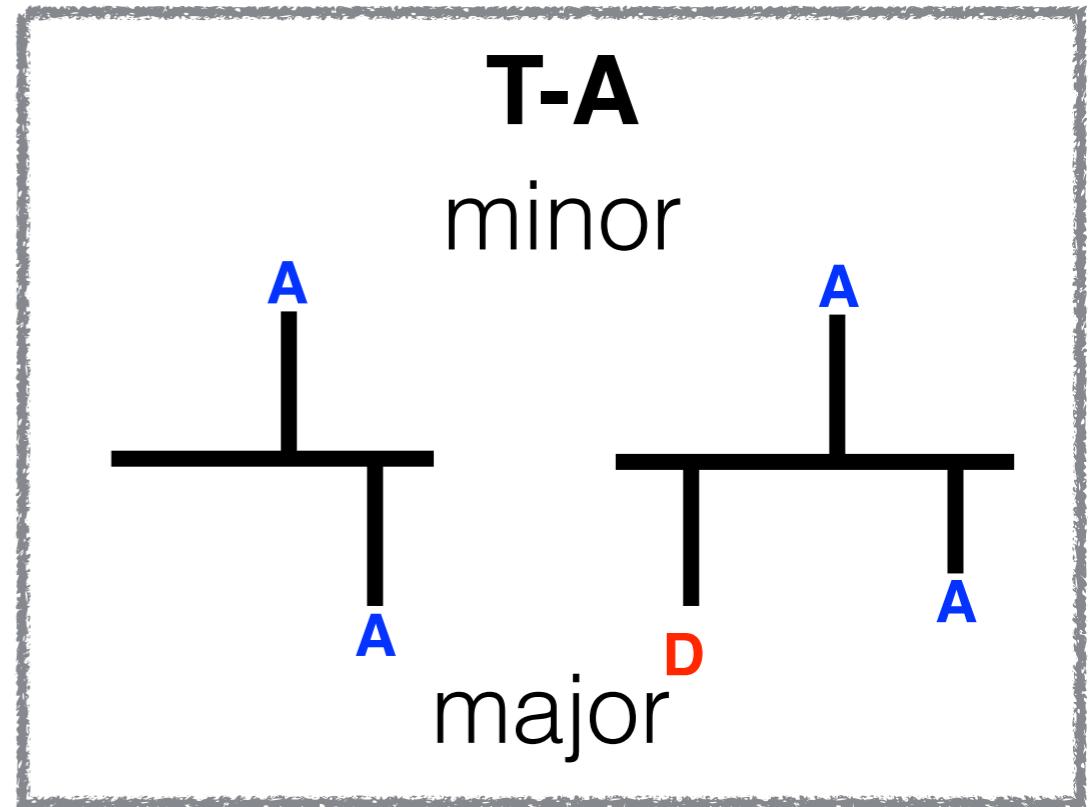
The atoms shown below are available to mediate protein/DNA interactions via H-bonds



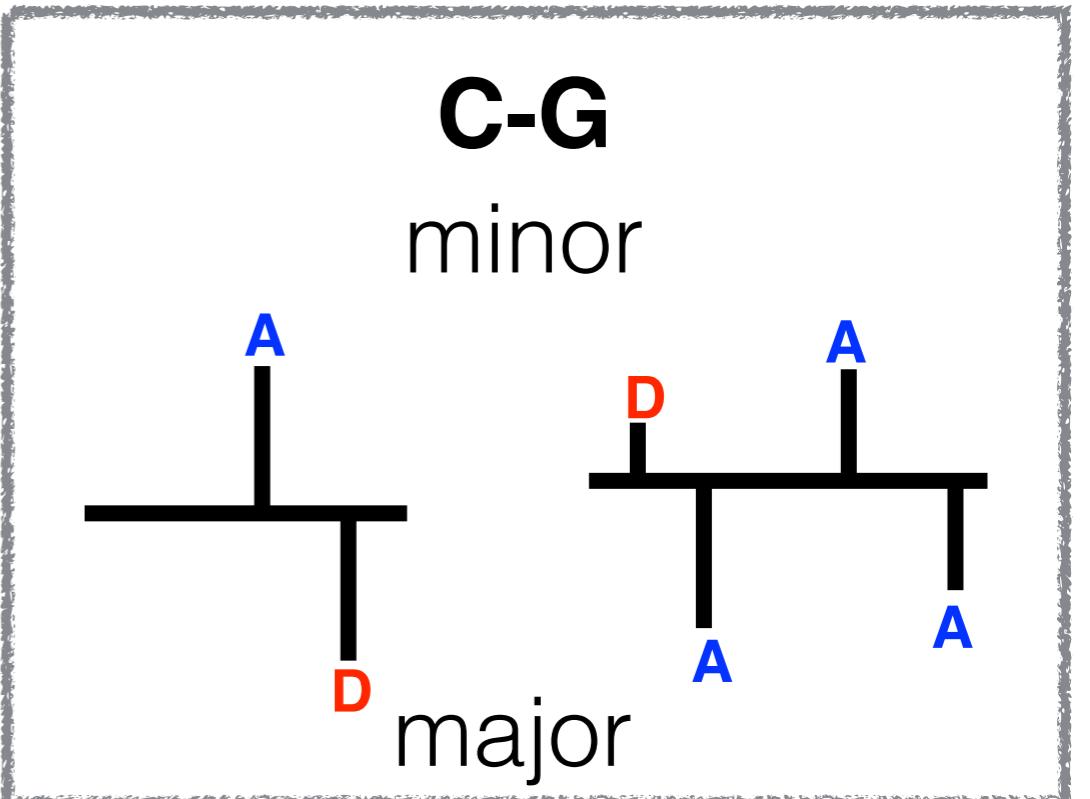
The atoms shown below are available to mediate protein/DNA interactions via H-bonds



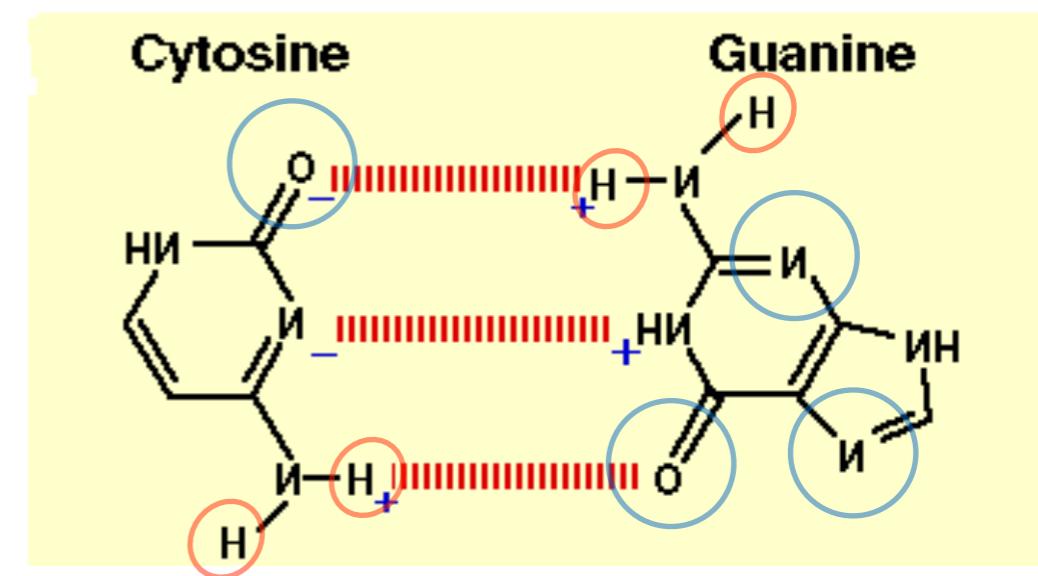
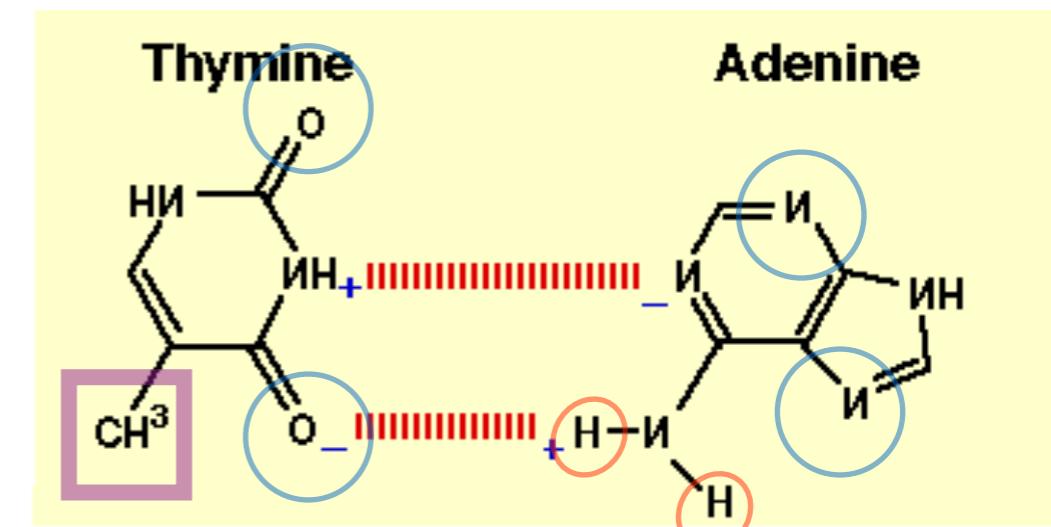
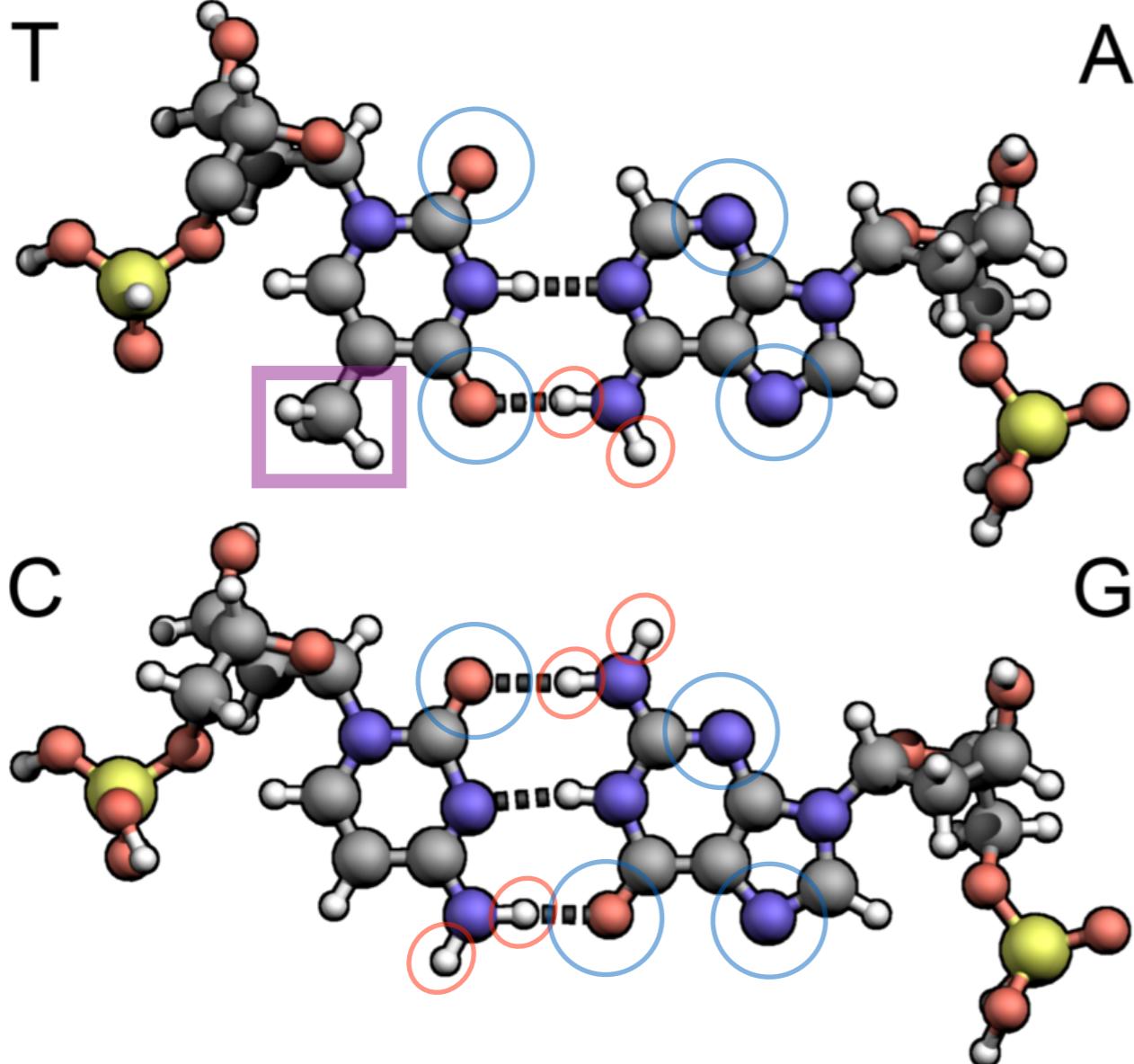
# Hydrogen Bond Donors and Acceptors are exposed in the Major and Minor Grooves



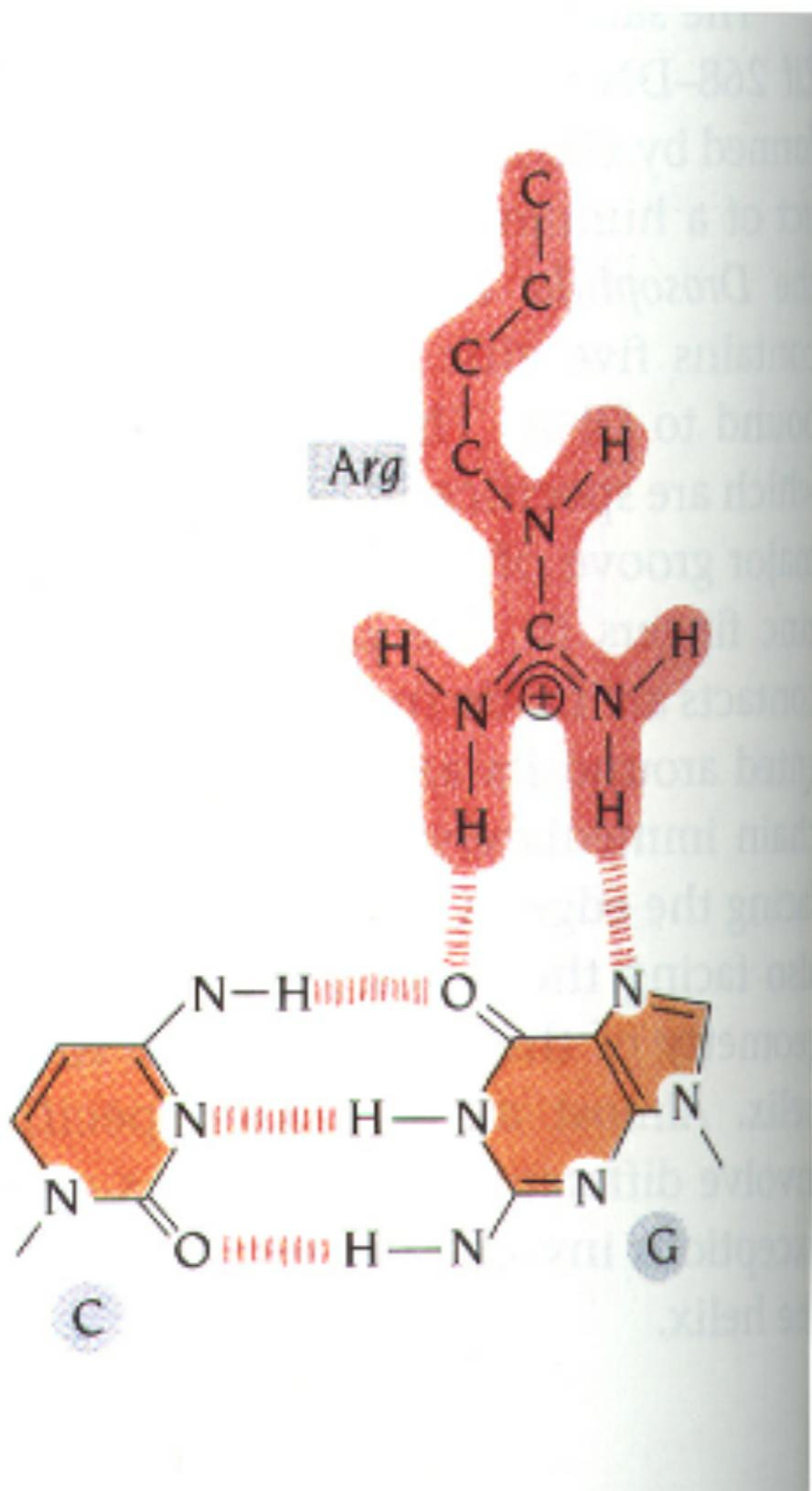
Base sequence is read  
and recognized by a  
protein probing the H-  
bonding possibilities in  
the Major and Minor  
Grooves



# Thymine's methyl group provides an additional source of recognition/specification

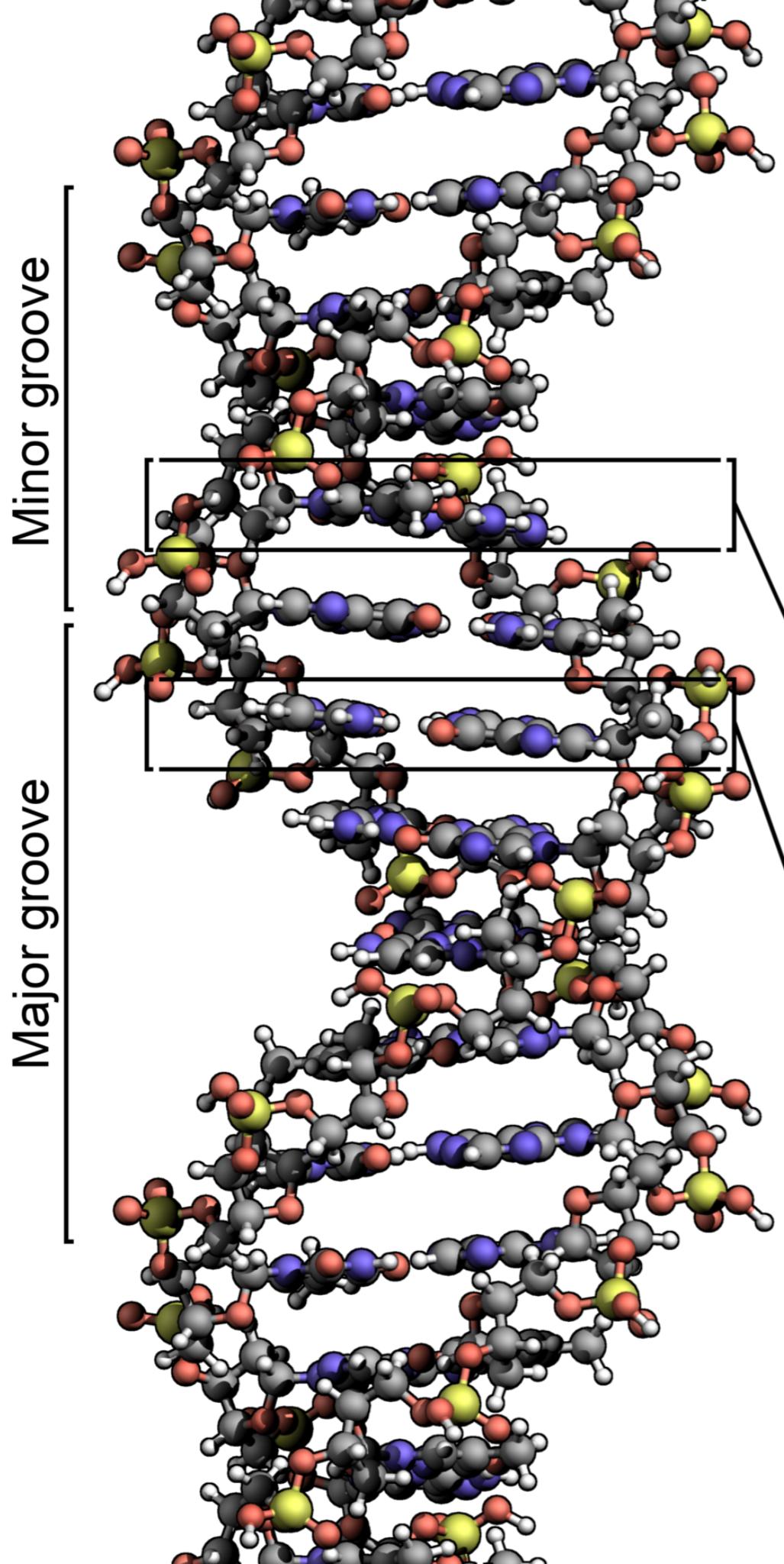


# An example of an Amino acid/DNA base interaction

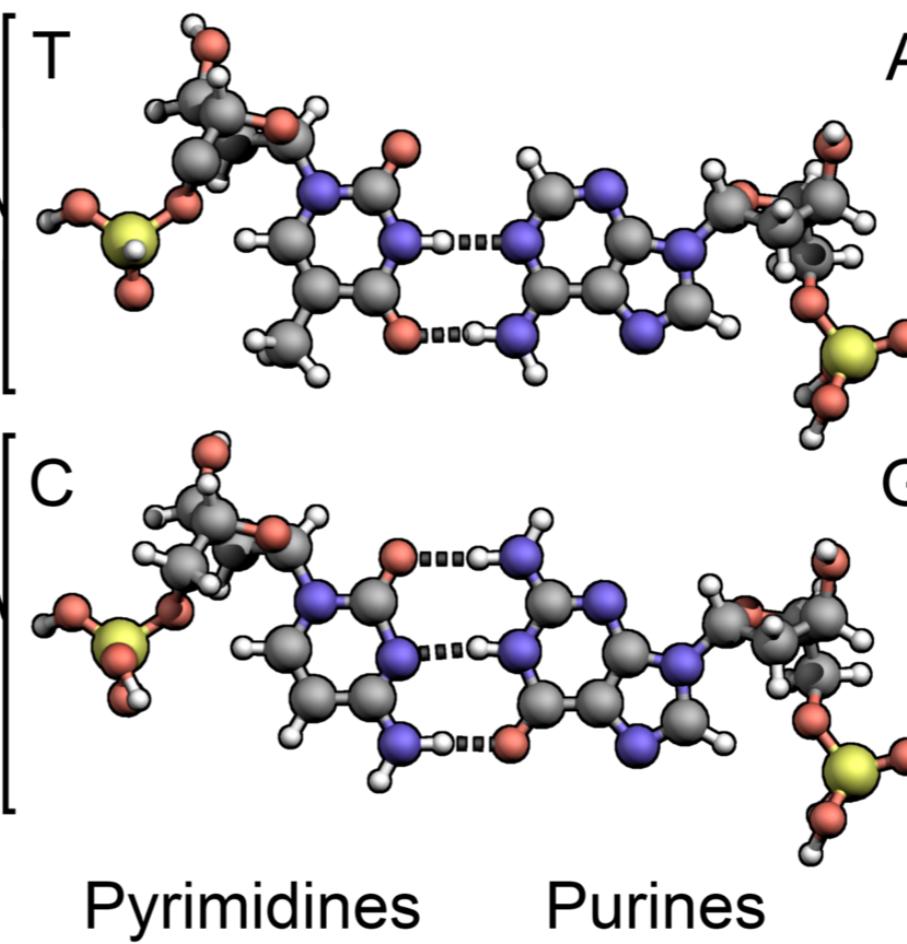


The interaction between arginine with its two hydrogen bond donors and a guanine base with its two acceptors in the major groove is an important component of many protein/DNA interactions.

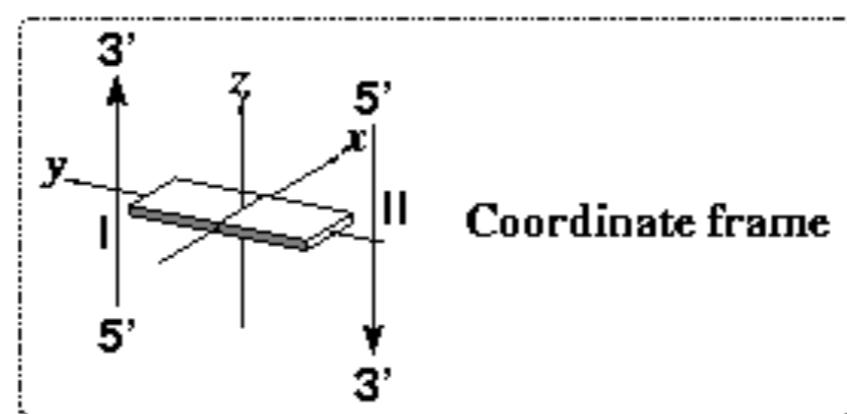
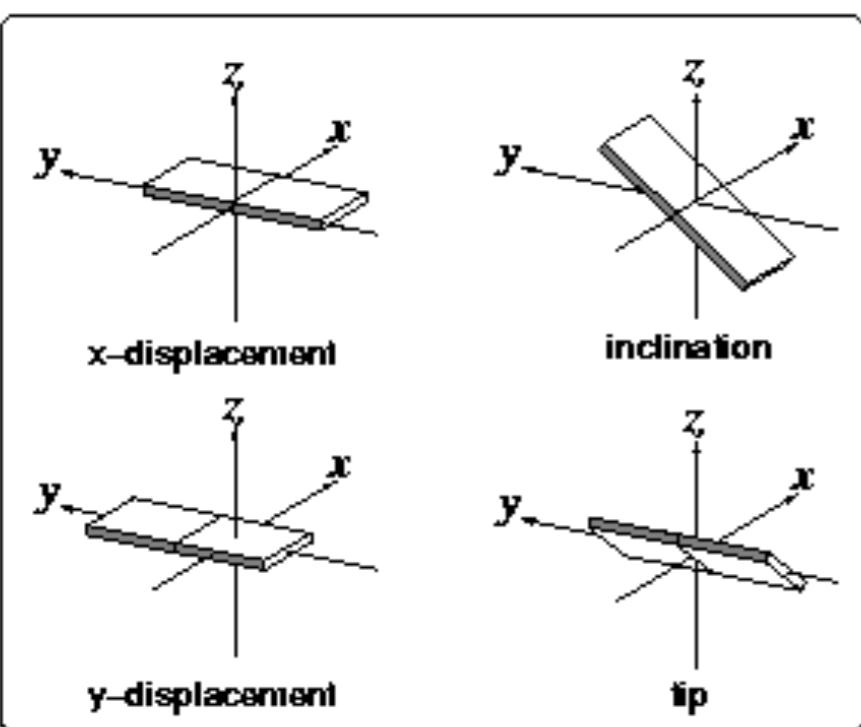
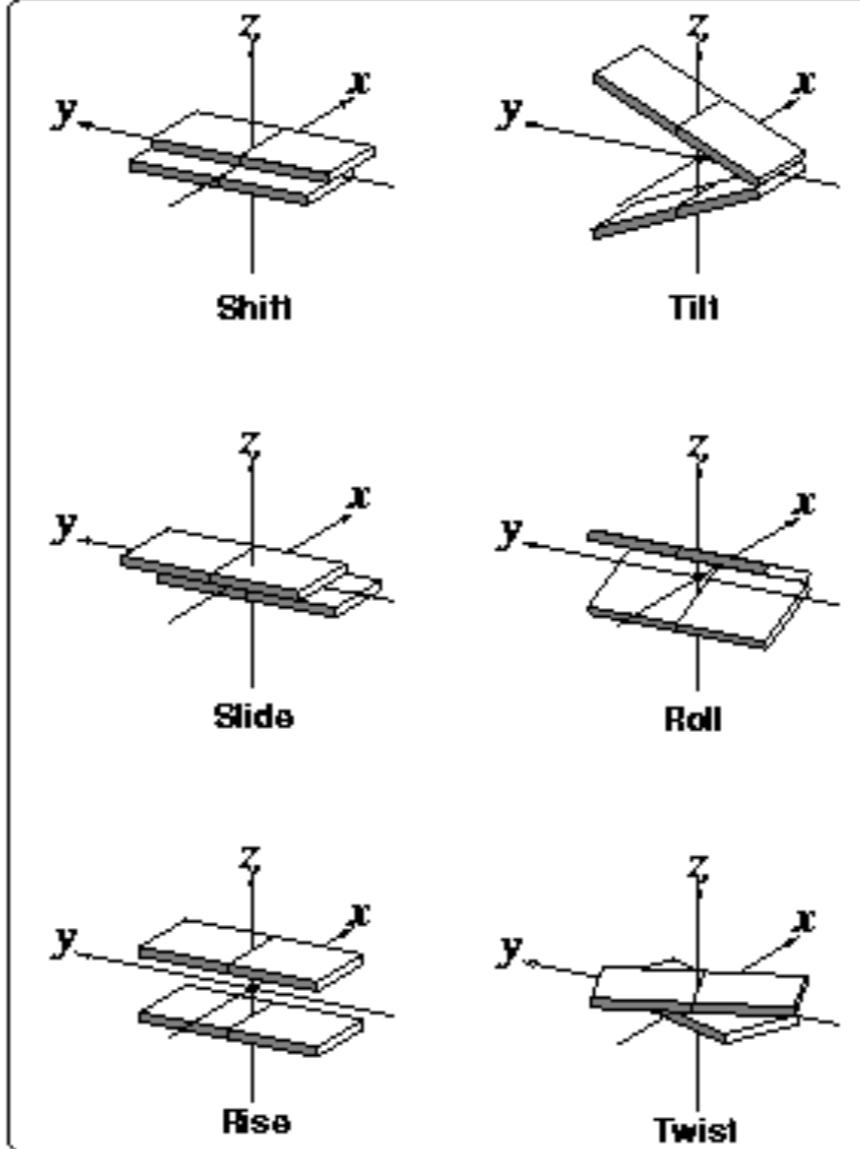
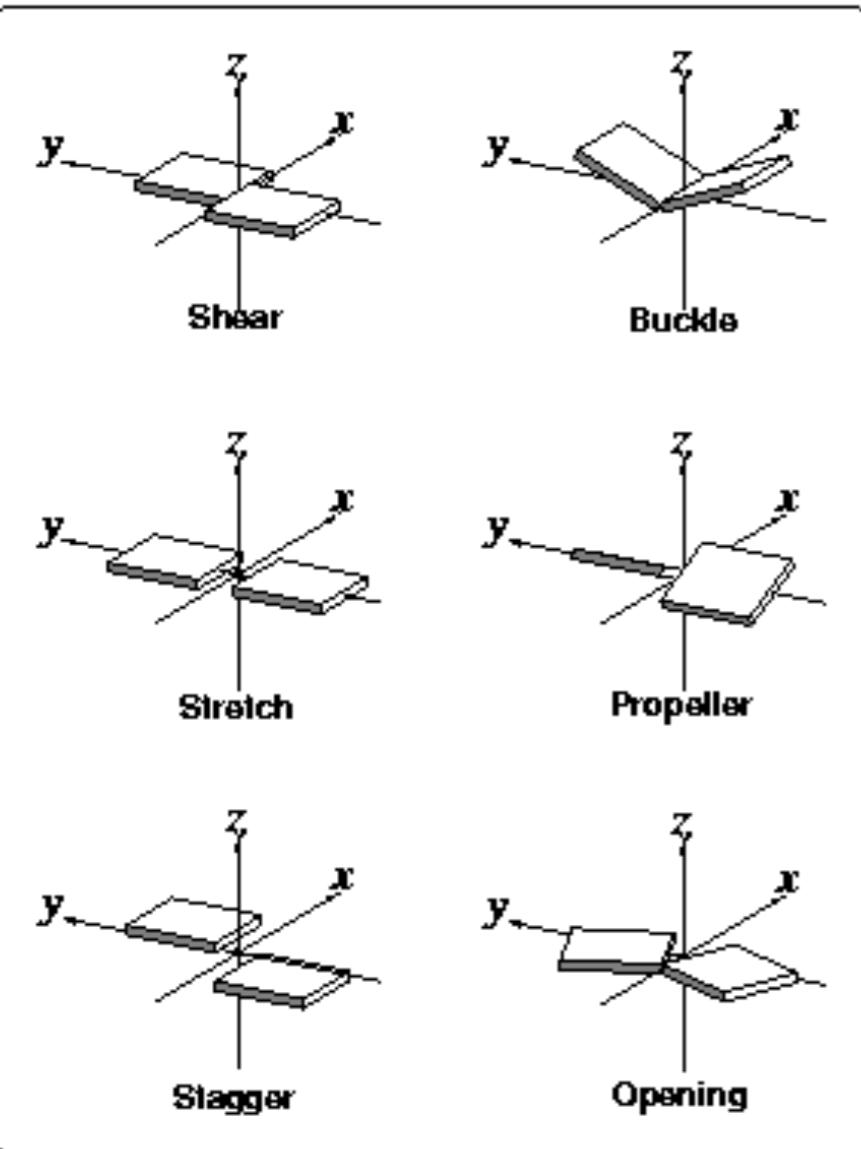
How do proteins interact with specific DNA sequences?



- Hydrogen
- Oxygen
- Nitrogen
- Carbon
- Phosphorus



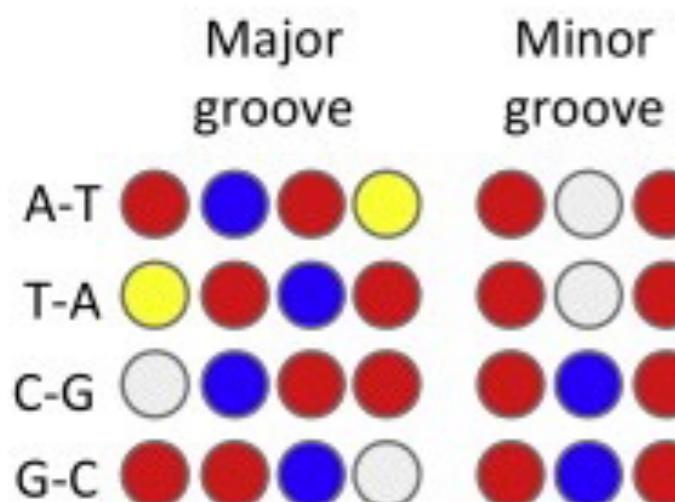
# Base Pair Geometry



Backbone phosphates are differentially positioned based on the degree of tilt, buckle, twist, roll, etc. relative to the preceding base pair.

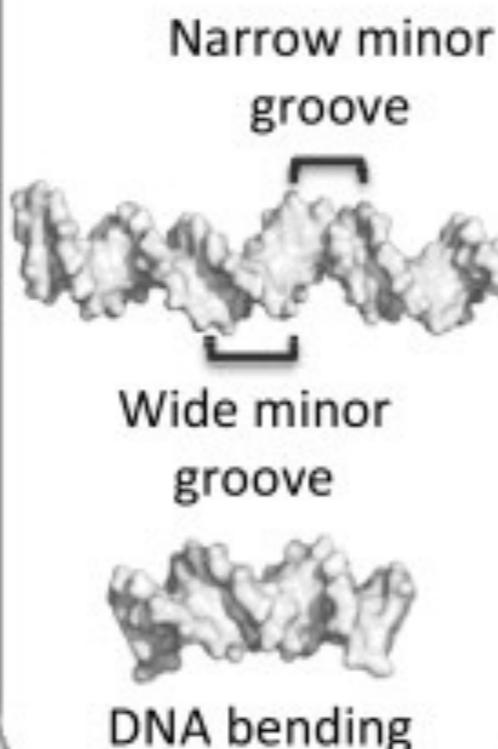
# Base Composition and Shape Contribute to TF-DNA Specificity

(A) Base readout:



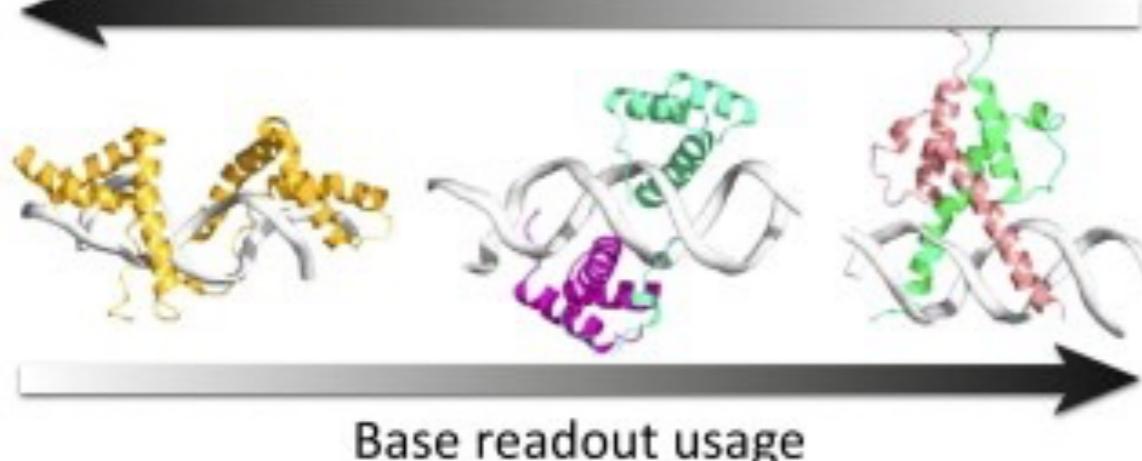
Key:  
● H-bond acceptor  
● Nonpolar hydrogen  
● H-bond donor  
● Methyl group

(B) Shape readout:



(C)

Shape readout usage



- Base readouts are specific for bp in major groove but degenerate for minor groove.
- Shape dominates for a minor groove-binding high motility group (HMG) box protein
- Base readout is a major contribution in DNA recognition by the bHLH protein Pho4
- Both readout modes are ~equally present in the DNA binding of a Hox–Exd heterodimer