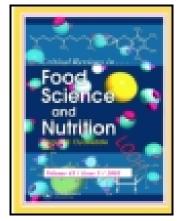
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# Health Benefits of Anthocyanins and Molecular Mechanisms: Update from Recent Decade

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Health benefits of anthocyanins and molecular mechanisms: Update from recent decade

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**Abbreviations:** 

**ABCA1**, ATP-binding cassette transporter A1; **ABCG1**, ATP-binding cassette transporter G1;

ACC, acetyl-CoA carboxylase; ACO, acyl-CoA oxidase; AD, Alzheimer's disease; AIF,

apoptosis-inducing factor; AMPK, AMP-activated protein kinase; AP-1, activator protein-1; aP2,

adipocyte fatty acid binding protein; apo E<sup>-/-</sup>, apolipoprotein E-deficient; APP, amyloid precursor

protein; Atg5, autophagy-related gene 5; ATGL, adipose triglyceride lipase; Aβ, amyloid-beta

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peptide; C/EBP6, CCAAT/enhancer-binding protein; COX-2, cyclooxygenase-2; CPT1A, carnitine palmitoyltransferase-1A; CRP, C-reactive protein; eIF2α, eukaryotic initiation factor 2α-subunit; Endo G, endonuclease G; FFAs, free fatty acids; FoxO1, forkhead box O1; G6Pase, glucose-6-phosphatase; GFAT, glutamine: fructose 6-phosphate aminotransferase; Glut2, glucose transporter 2; Glut4, glucose transporter 4; GSK3β, glycogen synthase kinase 3β; HBP, hexosamine biosynthetic pathway; IL, interleukin; iNOS, inducible nitric oxide synthase; JNK, c-Jun N-terminal kinase; LPL, lipoprotein lipaseand; LPS, anthocyanins in lipopolysaccharide; MAPK, mitogen-activated protein kinase; MMP, matrix metalloproteinase; mtGPAT1, mitochondrial acyl-CoA:glycerol-sn-3-phosphate acyltransferase 1; mTOR, mammalian target of rapamycin; NF- $\kappa$ B, nuclear factor  $\kappa$ B; NO, nitric oxide; OGD, oxygen-glucose deprivation; **oxLDL**, oxidative modification of low-density lipoprotein; **PARP**, poly ADP-ribose polymerase; PCA, protocatechuic acid; PD, Parkinson's disease; PEPCK, phosphoenol pyruvate carboxykinase; PGE<sub>2</sub>, prostaglandin E2; PhIP, 2-amino-1-methyl-6-phenylimidazo [4,5-b] pyridine; PKC $\zeta$ , protein kinase C  $\zeta$ ; PPAR $\gamma$ , peroxisome proliferator-activated receptor  $\gamma$ ; Pt, petunidin; RCT, reverse cholesterol transport; RR-ARFs, anthocyanin-rich fractions from red raspberries; SCI, spinal cord injury; SGLT1, sodium-dependent glucose transporter 1; STAT3, signal transducers and activators of transcription 3; TG, triglycerides; TNF, tumor necrosis factor; **TRAFs**, tumor necrosis factor receptor-associated factors; **UCP2**, uncoupling protein 2; **UDP-GlcNAc**, UDP-*N*-acetylglucosamine production

## <sup>2</sup> ACCEPTED MANUSCRIPT

**Abstract** 

Anthocyanins are one of the most widespread families of natural pigments in the plant kingdom.

Their health beneficial effects have been documented in many in vivo and in vitro studies. This

review summarizes the most recent literatures regarding the health benefits of anthocyanins and

molecular mechanisms. It appears that several signaling pathways including

mitogen-activated protein kinase, nuclear factor kB, AMP-activated protein kinase and

Wnt/β-catenin, as well as some crucial cellular processes, such as cell cycle, apoptosis,

autophagy and biochemical metabolism are involved in these beneficial effects and may provide

potential therapeutic targets and strategies for the improvement of a wide range of diseases in

future. In addition, specific anthocyanin metabolites contributing to the observed in vivo

biological activities, structure-activity relationships as well as additive and synergistic efficacy of

anthocyanins are also discussed.

Keywords: Anthocyanins; Benefits; Mechanism; Anti-cancer activity; Anti-inflammation activity;

Neuroprotective activity.

#### 1. Introduction

Anthocyanins are one of the most widespread families of natural pigments in the plant kingdom. They are responsible for the blue, purple, red and orange color of many fruits and vegetables. Anthocyanins belong to a large group of compounds collectively known as flavonoids, which are a subgroup of an even larger group of compounds known as polyphenolics (Andersen, 2001; Andersen and Markham, 2006; Mazza and Miniati, 1993; McGhie et al., 2007). Anthocyanins are present in nature mainly in the form of heterosides. The aglycon forms of anthocyanins, also called anthocyanidin, are structurally based on the flavilium ion or 2-phenylbenzopyrilium, and consist of hydroxyl and methoxyl groups in different positions. According to the number and position of the hydroxyl and methoxyl moieties, more than 635 anthocyanins have been identified. Among them, six most mentioned anthocyanidins in plants are pelargonidin, cyanidin, peonidin, delphinidin, petunidin and malvidin (Figure 1) (de Pascual-Teresa et al., 2008). Moreover, they are widely present in many fruits and vegetables. **Table 1** showed the daily consumption of anthocyanins from fruits, vegetables and beverages obtained from the National Health and Nutrition Examination Survey (Wu et al., 2006; NHANES, 2001).

During the last decade, several excellent reviews have illustrated these natural dietary phytochemicals in terms of their absorption (McGhie et al., 2007), metabolism (He and Giusti, 2010), bioavailability (Fernandes et al., 2014) and pharmacokinetics (Kay, 2006), as well as their

### <sup>4</sup> ACCEPTED MANUSCRIPT

various analytical techniques (Welch et al., 2008). The interests of knowing the health benefits of anthocyanins have also strongly increased in recent years. Ghosh and coworkers (2007) summarized the anti-diabetic and eye function properties of anthocyanins (Ghosh et al., 2007). de Pascual-Teresa discussed the beneficial effects of anthocyanins on the prevention of cardiovascular diseases and neurological conditions (de Pascual-Teresa et al., 2014). This review summarizes the most recent literatures regarding the biological benefits of dietary anthocyanins including anti-cancer activity, anti-inflammation activity, neuroprotective activity, prevention of cardiovascular disease, anti-obesity and anti-diabetes activity, especially focusing on the molecular mechanisms of action. We believe this present review will be helpful to better understand these dietary phytochemicals and apply them for the benefits of human health.

#### 2. Anti-cancer activity

Epidemiological studies revealing the anti-cancer activity of anthocyanins on gastrointestinal tract cancer risk in humans have been well summarized (Kocic et al., 2011). However, convincing epidemiological evidence indicating a positive association between the intake of anthocyanins-rich food and other cancers is needed to be further determined (Wang et al., 2008). We here discuss experimental findings related to the anti-cancer activity of anthocyanins (**Table 2**).

The anti-cancer activities of anthocyanins from many fruits and vegetables have been demonstrated to inhibit the initiation, promotion and progression of several cancers, such as breast

cancer (Singletary et al., 2007; Devi et al., 2011; Hui et al., 2010), prostate cancer (Reddivari et al., 2007), liver cancer (Bishayee et al., 2010; Bishayee et al., 2011), colorectal and intestinal cancers (Srivastava et al., 2007; Lim et al., 2013; Lala et al., 2006; Bobe et al., 2006; Koide et al., 1996; Koide et al., 1997; Hagiwara et al., 2001), blood cancer (Tsai et al., 2014), cervical cancer (Rugină et al., 2012), lung cancer (Aqil et al., 2012), fibrosarcoma (Filipiak et al., 2014) and metastatic melanoma (Bunea et al., 2013). In addition, some studies also demonstrated that anthocyanins from one fruit or vegetable may have the chemoprotective activity against various cancers. For example, anti-proliferative effect of anthocyanins-rich extract from blackberry on colon cancer HT-29, breast cancer MCF-7, lung cancer A549 and leukemia HL-60 cells have been determined (Dai et al., 2009; Agil et al., 2012). Barrios et al. demonstrated that the anthocyanins-rich extract from Pourouma cecropiifolia fruits, a tropical plant, significantly reduced the viability of laryngeal cancer HEp-2, gastric cancer MKN-45 and breast cancer MCF-7 cells (Barrios et al., 2010). Anthocyanin extract from blueberry significantly induced apoptosis in mouse melanoma B16-F10 and human colon cancer HT-29 cells (Srivastava et al., 2007; Bunea et al., 2013). However, there is little knowledge about synergistic or antagonastic effects of various anthocaynins on inhibiting the initiation, promotion, and progression of carcinogenesis, which should be a highlight in future. In addition, a recent study by Peiffer et al. (2014) has demonstrated that protocatechuic acid (PCA), a major anthocyanin metabolite, effectively inhibited the development of esophageal cancer in rats. Similarly, Forester et al. (2014) also found that three

anthocyanin metabolites, gallic acid, 3-*O*-methylgallic acid and 2,4,6-trihydroxybenzaldehyde, could decrease cell viability, cause cell cycle arrest and apoptosis in colon cancer Caco-2 cells. Therefore, it is indispensable to investigate the anti-cancer activity of the individual metabolite of anthocyanins in future.

Several studies have shown that the chemical structures of anthocyanins do have a significant impact on their biological activities. Konczak-Islam et al. demonstrated the anti-cancer property of the anthocyanins-rich extract from the sweet potato in HL-60 cells. Composition analysis showed that the dominated nonacylated cyanidin 3-sophoroside-5-glucoside might be related to the observed anti-cancer activity (Konczak-Islam et al., 2003). Further, Zhao et al. demonstrated that the chokeberry extracts, containing high levels of monoglycosylated cyanidin derivatives, showed a stronger chemoprotective activity than grape and bilberry extracts (Zhao et al., 2004). Moreover, Jing et al. used human colon cancer HT29 cell line to compare the anti-cancer properties of anthocyanins-rich extract from purple corn, chokeberry, bilberry, purple carrot, grape, radish and elderberry. The degrees of growth inhibitory activity were as follows: purple corn > chokeberry and bilberry > purple carrot and grape > radish and elderberry. Statistical analyses suggested that nonacylated monoglycosylated anthocyanins had greater anti-cancer property than anthocyanins with pelargonidin, triglycoside, and/or acylation with cinnamic acid (Jing et al., 2008). Therefore, it appears that the type of aglycones, sugars, and acylated acids, and the position and degree of glycosylation and acylation are the main factors influencing the

anti-cancer property. Nevertheless, the structure-activity relationship of anthocyanins as chemoprotective agents remains to be further elucidated.

Extensive investigations have been performed to determine the molecular mechanisms of the anti-cancer activity of anthocyanins, with results indicating that anthocyanins can inhibit several signaling pathways involved in tumor growth and apoptosis. In an early study, the anthocyanins-rich extract from Aronia meloncarpa induced G1/G0 and G2/M cell cycle arrest in HT-29 colon cancer cells by increasing the expression of the p21WAF1 and p27KIP1 and decreasing expression of cyclin A and cyclin B (Malik et al., 2003). Similarly, the anthocyanins-rich extract from potato also leaded to G1/G0 cell cycle arrest in prostate cancer LNCaP and PC-3 cells with the higher p27 protein levels. Moreover, the extract caused caspase-dependent apoptosis in LNCaP cells with the induction of poly ADP-ribose polymerase (PARP) cleavage and activation of caspase 3 (cleavage), as well as caspase-independent cell death in PC-3 cells with mitochondrial release and nuclear uptake of the proapoptotic endonuclease G (Endo G) and apoptosis-inducing factor (AIF) proteins. Notably, both apoptotic pathways depended on upstream activation of mitogen-activated protein kinase (MAPK) and c-Jun N-terminal kinase (JNK) pathways, indicating that MAPK signaling pathway might participate in the molecular mechanisms of the anti-cancer activity (Reddivari et al., 2007). In addition, the activation of nuclear factor κB (NF-κB) pathways might also been shown to be involved in tumor growth and development. Afaq et al. reported that the animals pretreated with anthocyanins-rich

extract from pomegranate fruit resulted in substantially reduced tumor incidence and lower tumor body burden in 7,12-dimethylbenz(a)anthracene-initiated CD-1 mouse through the inhibition of phosphorylation of extracellular signal-regulated kinase (ERK)1/2, p38 and JNK1/2, as well as activation of NF- $\kappa$ B and I $\kappa$ K $\alpha$  and phosphorylation and degradation of I $\kappa$ B $\alpha$  (Afaq et al., 2005).

The Wnt pathway is crucial to cell proliferation, differentiation and survival. β-catenin is a key component of the Wnt signaling pathway and overactivation of β-catenin in the cytosol is related to cancer metastasis. Under normal conditions, β-catenin is controlled by dephosphorylated glycogen synthase kinase 3\beta (GSK3\beta). When GSK3\beta is phosphorylated, GSK3\beta loses its activity and no longer controls β-catenin. Park et al. demonstrated that anthocyanin extract from Korea wild berry Meoru effectively inhibited liver cancer Hep3B cell migration and invasion by decreasing the expression of phospho-GSK3β and β-catenin. Moreover, AMP-activated protein kinase (AMPK) activation induced by anthocyanins might be an upstream regulator of the GSK3B/B-catenin pathway (Park et al., 2014). Additionally, autophagic pathway has also been reported to be a novel therapeutic target for cancer treatment. Longo et al. found that anthocyanin-induced autophagy was characterized by the upregulation of eukaryotic initiation factor 2α-subunit (eIF2α) and downregulation of mammalian target of rapamycin (mTOR) and Bcl-2. Inhibition of autophagy by either 3-methyladenine or autophagy-related gene 5 (Atg5) small interfering RNA enhanced anthocyanin-triggered apoptosis in liver cancer PLC/PRF/5 cells (Longo et al., 2008).

#### 3. Anti-inflammation activity

Many epidemiological and experimental studies have showed the anti-inflammation activity of anthocyanins in foods for the amelioration of inflammation-associated diseases or disorders, such as colitis (Akiyama et al., 2012), peyronie disease (Sohn et al., 2014), periodontal disease, laryngopharyngeal reflux (Samuels et al., 2013), postprandial inflammatory response (Edirisinghe et al. 2011) and pain behavior (Tall et al., 2004).

Inflammatory responses are a series of well-coordinated events that controlled by several factors including cytokines, enzymes, lipid mediators and vasoactive mediators (Hassimotto et al., 2013). Cyclooxygenases (COXs) are the key pro-inflammatory enzymes that are involved in arachidonic acid metabolism. COX-2 is the main enzyme for the synthesis of lipid mediators such as prostaglandin E2 (PGE2), which is a potent vasodilator enhancing oedema formation (Graf et al., 2013; Hassimotto et al., 2013). Hassimotto et al. showed that dietary supplements of the anthocyanins-rich extract from wild mulberry and the cyanidin 3-glucoside are effective in suppressing carrageenan-induced oedema and peritonitis through the downregulation of COX-2 expression and inhibition of PGE2 production (Hassimotto et al., 2013). The inducible nitric oxide synthase (iNOS) is another key pro-inflammatory enzyme for the production of nitric oxide (NO). Excessive production of NO appears to be associated with the progression of many inflammatory diseases (Li et al., 2014). An early study by Tsuda et al. (2002) demonstrated that cyanidin 3-O-β-glucoside suppressed the zymosan-induced inflammatory response in rats by reducing the

level of iNOS, tumor necrosis factor (TNF)-α, interleukin (IL)-1β and IL-6 in the peritoneal exudate cells. Poulose et al. also showed that pretreatment of BV-2 microglial cells with the anthocyanins-rich extract from acai pulp was protective against LPS-induced NO release, iNOS production and COX-2 expression (Poulose et al., 2012).

The NF-κB pathway plays an important role in triggering and regulating inflammatory processes. The NF-κB transcription factor exists in an inactive state in the cytosol by binding to IkB, upon its activation and translocation into the nucleus, genes involved in pro-inflammatory responses is induced, leading to the expression of cytokines and inflammatory enzymes including iNOS and COX-2 (Taverniti et al., 2014). Taverniti and coworkers (2014) showed that the anthocyanins-rich extract from wild blueberry displayed anti-inflammatory property by decreasing the activation of NF-κB in the presence of the pro-inflammatory stimulus IL-1β in human Caco-2 intestinal cells. In a retinal inflammatory mouse model, an anthocyanins-rich extract from bilberry showed anti-inflammatory activity by suppressing the expression of signal transducers and activators of transcription 3 (STAT3) and IL-6 through the reduction of NF-κB activation (Miyake et al., 2012). Moreover, delphinidin inhibited COX-2 expression in LPS-activated murine macrophage RAW264 cells by targeting the transcription factors NF-kB, CCAAT/enhancer-binding protein (C/EBPδ) and activator protein-1 (AP-1) (Hou et al., 2005). Xia et al. demonstrated that intracellular tumor necrosis factor receptor-associated factors (TRAFs) translocation to lipid rafts played pivotal role in CD40-mediated NF-κB activation, which induced

inflammatory response in human endothelial cells. However, cyanidin 3-O-β-glucoside and peonidin 3-O-β-glucoside significantly inhibited CD-40-induced inflammatory signaling through the reduction of cholesterol distribution in lipid rafts, indicating that anthocyanins could act as lipid-dependent regulators of inflammatory response (Xia et al., 2007). In addition, MAPK signaling pathway also played a critical role in the regulation of inflammatory response. Hou et al. reported that inhibition of COX-2 expression by blocking the activation of MAPK signaling pathway including JNK, ERK and p38 kinase contributed to the anti-inflammatory property of delphinidin (Hou et al., 2005). Xia et al. demonstrated that cyanidin 3-O-β-glucoside and peonidin 3-O-β-glucoside also significantly prevented CD40-induced endothelial activation and apoptosis by inhibiting production of pro-inflammatory cytokines and matrix metalloproteinases (MMP-1, MMP-9). Notably, the anti-inflammatory activity might be due to the downregulation of JNK and p38 activation (Xia et al., 2009). The above findings suggested that the NF-κB and MAPK signaling pathways may provide key targets for the application of anthocyanins to improve the inflammation-associated diseases. A recent study by Li and coworkers (2014) further confirmed this notion by determining the anti-inflammatory activity of anthocyanins-rich fraction from red raspberries (RR-ARFs) in RAW264.7 cells and an acute mouse colitis model. As shown in Figure 2, RR-ARFs significantly inhibited p65 phosphorylation and its nuclear translocation as well as the activation of IKK,  $I\kappa B\alpha$  and JNK, thereby suppressing the expression of pro-inflammatory genes such as iNOS, COX-2, IL-1\beta and IL-6.

## <sup>2</sup> ACCEPTED MANUSCRIPT

Interestingly, Graf et al. found that the anthocyanins-rich juice from grape-bilberry at physiological did not showed any anti-inflammatory effects on the systemic immune system, gut-associated lymphoid tissue and mesenteric adipose tissue in healthy rats, indicating that anti-inflammatory effects of anthocyanins might depend on various factors such as sources, doses and categories of anthoycanins used in the experiments (Graf et al., 2013). Hou and colleagues (2005) investigated the anti-inflammatory effects of five anthocyanins in lipopolysaccharide (LPS)-activated murine macrophage RAW264 cells. It seemed that only delphinidin and cyanidin inhibited LPS-induced COX-2 expression, but pelargonidin, peonidin and malvidin did not. Further analysis showed that the *ortho*-dihydroxyphenyl structure of anthocyanidins on the B-ring might be crucial for the inhibitory activity. Furthermore, long-term supplementation with purified anthocyanins (cyanidin 3-O-glucoside or delphenidin 3-O-glucosides) derived from bilberries and blackcurrants inhibited the inflammatory response in adults with hypercholesterolemia. Notably, this study showed that the mixture of the two purified anthocyanins caused a greater anti-inflammatory effect than single anthocyanin, which suggested that different anthocyanins might have additive or synergistic effects on mediating anti-inflammatory response (Zhu et al., 2013). Additionally, the synergistic effect of anthocyanins and antimicrobials has been confirmed. Yoon et al. reported that combination of anthocyanin extracts from black soybean and ciprofloxacin has more obvious anti-inflammatory and antimicrobial effects on treating chronic bacterial prostatitis than ciprofloxacin treatment alone (Yoon et al., 2013). Therefore, the

combined use of anthocyanins with other anti-inflammatory medicines may provide novel therapeutic strategies for the treatment of inflammation-associated diseases.

#### 4. Neuroprotective activity

Significant evidence from epidemiological studies has suggested the neuroprotective activity of anthocyanins for their improvement on cognitive performance, memory performance and motor performance, indicating their potential application for the prevention of many neurodegenerative diseases, such as Parkinson's disease (PD) and Alzheimer's disease (AD) (Youdim et al., 2004).

PD is a neurodegenerative disorder that involves a loss of dopaminergic neurons in a midbrain region. Dopaminergic cell death is suggested to be involved in the progression of PD. Strathearn et al. demonstrated that anthocyanins-rich extract from blueberries, grape seed, hibiscus, blackcurrant, and Chinese mulberry significantly suppressed rotenone-induced dopaminergic cell death *via* interference with microglial activation and amelioration of mitochondrial dysfunction (Strathearn et al., 2014). Mitochondrial dysfunction caused by oxidative stress also leads to neuronal damage after ischemic stroke. Cyanidin 3-*O*-glucoside has been shown to exert its neuroprotective effect against ischemic stroke in mice by blocking AIF release from mitochondria (Min et al., 2011). One of the physiological roles of cell autophagy is to remove damaged organelles including damaged mitochondria, thereby rescuing cells under stressed conditions. Kim and coworkers (2012) demonstrated that anthocyanin extract from black soybean increased survival of U87 glioma cells exposed to oxidative stress induced by oxygen-glucose deprivation

(OGD). Silencing Atg5 expression, an essential regulator of autophagy induction, reversed the cytoprotective effect of anthocyanin extract against OGD stress. However, treatment of U87 cells with rapamycin, an autophagy inducer, increased cell survival upon OGD stress, indicating autophagy might be a neuroprotective mechanism for anthocyanins against oxidative stress-induced cytotoxicity in glial cells. Whether induction of autophagy is the common neuroprotective mechanism of various anthocyanins with different chemical structures still need to be further determined. Anyway, anthocyanins are still potential candidates to be the effective neuroprotective dietary complementation for the prevention or reduction of neuronal cell death. In addition, anthocyanin extract from bog bilberry exerted its neuroprotective effects by reducing glial scar formation, axonal loss and inflammation and promoting remyelination and neuron survival in a rat model of spinal cord injury (SCI) (Wang et al., 2012). Similarly, treatment with cyanidin 3-O-β-glucoside could also reduce superoxide production, neuron cell damage, lesion volume and neurological dysfunction in a traumatic SCI rat model (Kim et al., 2011). Therefore, an anthocyanins-rich dietary therapy may be used to improve the condition of the SCI patients.

AD is an irreversible degenerative brain disease caused by the hyperphosphorylation of Tau protein aggregation. Two kinds of anthocyanins, cyanidin 3-*O*-glucoside and malvidin 3-*O*-glucoside have been shown to induce FK506 binding protein 52 (FKBP52) activation, leading to the reduction of hyper-phosphorylated Tau protein aggregation, and thereby improving the treatment of AD (Hung et al., 2014). It has also been reported that the altered amyloid precursor

protein (APP) processing leading to increased amyloid-beta peptide (A $\beta$ ) accumulation is a key pathogenic feature of AD. Vepsäläinen et al. observed that long-lasting supplementation with anthocyanins-rich extract from bilberry and blackcurrant exerted beneficial effects on APP and A $\beta$  metabolism and alleviated behavioral abnormalities in a mouse model of AD (Vepsäläinen et al., 2013). A $\beta$  is known to induce the redox imbalance, mitochondrial dysfunction and caspase activation, resulting in neuronal cell death. Pretreatment with anthocyanins-rich extract from purple sweet potato reduced A $\beta$ -induced apoptosis by inhibiting the ROS generation, lipid peroxidation, caspase-3 activity and decreasing levels of intracellular Ca<sup>2+</sup> and membrane potential loss in PC12 cells (Ye et al., 2010). In another study, anthocyanins-rich extract from mulberry significantly inhibited the accumulation of A $\beta$  and improved learning and memory ability in a senescence-accelerated mice model (Shih et al., 2010).

#### 5. Prevention of cardiovascular disease

The relationship between anthocyanins intake and the reduced risk of developing cardiovascular disease has been indicated in several epidemiological studies (Wallace, 2011; PrRimm et al., 1991). A recent study of young and middle-aged women showed a high intake of anthocyanins was associated with a 32% reduction in risk of myocardial infarction (Cassidy et al., 2013). In addition, an Iowa Women's Health Study with 34,489 women showed a significant reduction in cardiovascular disease mortality associated with anthocyanins-rich strawberry intake (Mink et al., 2007).

Atherosclerosis is characterized by vascular obstruction from the deposits of lipids, resulting in reduced blood flow (Mauray et al., 2012). The liver is the main organ regulating plasma lipids levels and lipoprotein metabolism. It plays a central role in atherosclerosis. Mauray et al. showed that 2-week supplementation with anthocyanins-rich extract from bilberry significantly reduced plasmatic total cholesterol and hepatic triglyceride levels in apolipoprotein E-deficient (apo E<sup>-/-</sup>) mice. Microarray analysis showed that numerous over-expressed genes (CYP7A1, HMGCR, LPL, NR1H4, INSIG2) were involved in bile acid synthesis and excretion, which enhances cholesterol elimination in the plasma and the reduction of hepatic lipogenesis (**Figure 3**). In addition, downregulation of many pro-inflammatory genes (*ALOX5AP*, *CX3CL1*, TNFRSF14) was also observed, indicating anti-inflammatory response might also participate in the protection against atherosclerosis (Mauray et al., 2010) and its mechanism of action was possibly inhibiting atherosclerotic plaque progression and increasing the stability of the vulnerable plaque (Xia et al., 2006). Further study showed that anthocyanin-rich extract from bilberry also markedly modulated the expression of genes in the aortas of apo E<sup>-/-</sup> mice. Bioinformatic analysis revealed that the identified numerous aortic genes seemed to be related to increased inter-cellular adhesion (CDH4, CTNNB1, JAM-A, VCAM1), decreased monocyte recruitment (RDX, ARPC5), cellular contractility (MYLC2B, PAK1) and vascular permeability (KDR, FAK, VEGFR2), thereby decreasing the risk of endothelial dysfunction, which is an early marker of the development of atherosclerosis (Mauray et al., 2012). In addition, endothelial

function has also been demonstrated to be improved by delphinidin 3-*O*-glucoside and cyanidin 3-*O*-glucoside through the activation of the NO-cGMP signaling pathway in hypercholesterolemic individuals (Zhu et al., 2011), inhibition of mitochondria-mediated apoptotic signaling pathway in human umbilical vein endothelial cells and bovine aortic endothelial cells (Zapolska-Downar et al., 2008; Paixão et al., 2011), as well as the activation of cAMP-PKA-eNOS signaling pathways in human aortic endothelial cells (Liu et al., 2014).

Oxidative modification of low-density lipoprotein (oxLDL) is involved in the pathogenesis of atherosclerosis through the formation of macrophage-derived foam cells. Therefore, the inhibition of oxLDL formation might effectively lower the risk of atherosclerosis. Anthocyanin extract from *Hibiscus* was demonstrated to inhibit oxLDL and foam cell formation by downregulating the expression of CD36 and increasing nuclear peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ) protein levels in mouse macrophage J774A.1 cells (Kao et al., 2009). Similarly, anthocyanin extract from mulberry also decreased macrophage death and foam cell formation in Cu<sup>2+</sup>-mediated oxLDL (Liu et al. 2008).

Reverse cholesterol transport (RCT) is a process that entails the efflux of excess cholesterol from macrophages into the liver. The promotion of macrophage RCT by upregulating the expression of ATP-binding cassette transporter A1 (ABCA1) and G1 (ABCG1) in macrophages may be a potential novel approach for prevention and treatment of atherosclerosis. PCA is determined as the main gut microbiota metabolite of cyanidin 3-*O*-β-glucoside. Wang et al.

demonstrated that PCA, but not its precursor, induced ABCA1 and ABCG1 expression in macrophages by decreasing the expression of miR-10b, which contributed to the accelerated macrophage RCT. Importantly, the intestinal microbiota ecosystem might be a potential target for the prevention and treatment of chronic diseases such as atherosclerosis because of its strong metabolic ability (Wang et al., 2012).

In summary, anthocyanins-rich extract from fruits and vegetables could effectively reduced risk of atherosclerosis by improving endothelial dysfunction, inhibiting oxLDL formation and promoting macrophage RCT.

#### 6. Anti-obesity and anti-diabetes activity

Many epidemiological studies have confirmed the inverse association between anthocyanin intake and the risk of obesity and diabetes. In a study of 1,997 females from the United Kingdom, higher intake of anthocyanins was associated with significantly lower concentrations of high-sensitivity C-reactive protein (CRP), a marker of obesity and diabetes (Wang et al., 2013). Similarly, a large cohort study of 200,994 health professionals from the United States revealed that consumption of anthocyanins-rich foods was inversely associated with the risk of diabetes (Muraki et al., 2013). In line with these findings, a cross-sectional study suggested consumption of high amounts of anthocyanins might have beneficial effect on improving lipid profile in Chinese women, indicating the efficacy of anthocynins for the treatment of obesity (Li et al., 2013).

Obesity is the result of accumulated excessive adipose tissue caused by the imbalance of

energy intake and expenditure. It is usually associated with various metabolic disorders. Jurgoński et al. reported that the anthocyanins-rich extract from Kamchatka honeysuckle berry was able to ameliorate the disturbances in lipid and glucose metabolism in rats (Jurgoński et al., 2013). In line with this result, Wu and coworkers (2013) observed that anthocyanin extract from honeysuckle suppressed body weight gain, reduced serum and liver lipid profiles, ameliorated impaired hepatic function and significantly increased serum adiponectin concentration while decreasing serum insulin and leptin levels in a high fat diet-induced mouse model. Continuous hyperglycemia could enhance lipolysis of triglycerides (TG) in the adipocytes resulting in elevated levels of plasma free fatty acids (FFAs), which have been demonstrated to potentially link obesity (Guo et al., 2012). Therefore, decreasing glucose uptake of the adipocytes may be a potential approach for the prevention of obesity. Alzaid et al. found that acute exposure to the anthocyanins-rich extract from berry significantly decreased both Na<sup>+</sup>-dependent and Na<sup>+</sup>-independent glucose uptake in Caco-2 cells. Further, longer-term exposure with berry extract markedly reduced the expression levels of glucose transporter 2 (Glut2) and sodium-dependent glucose transporter 1 (SGLT1) and resulted in significant inhibition of glucose uptake (Alzaid et al., 2013).

In addition, the adipose tissue secretes several adipocytokines (e.g. adiponectin, leptin, and resistin), which have important regulatory functions in the development of metabolic diseases such as obesity. Therefore, the regulation of adipocytokine secretion or the adipocyte-specific gene

expression is one of the most important strategies for the prevention of obesity. Graf et al. observed that rats fed anthocyanins-rich grape-bilberry juice showed reduced levels of serum cholesterol, triglycerides, leptin and resistin. In addition, increased proportion of polyunsaturated fatty acids and decreased amount of saturated fatty acids in plasma were also observed (Graf et al., 2013). Similarly, Tsuda et al. demonstrated that anthocyanins enhanced adipocytokine (adiponectin and leptin) secretion and the expression of PPARy, lipoprotein lipase (LPL), adipocyte fatty acid binding protein (aP2) and uncoupling protein 2 (UCP2) in isolated rat adipocytes. Notably, AMPK activation might be one of the possible mechanisms for the regulation of adipocyte-specific gene expression (Tsuda et al., 2004). Guo et al. revealed a novel mechanism by which anthocyanin cyanidin 3-O-β-glucoside effectively eliminated the impacts of high-glucose on the induction of adipocyte lipolysis (Figure 4). In that study, cyanidin 3-O-β-glucoside increased the activity of AMPK, decreased the activity of glutamine: fructose 6-phosphate aminotransferase (GFAT), reduced cellular UDP-N-acetylglucosamine production (UDP-GlcNAc), thereby suppressing the hexosamine biosynthetic pathway (HBP). In addition, it also attenuated high-glucose-promoted forkhead box O1 (FoxO1), resulting in decreased expression of adipose triglyceride lipase (ATGL), thus inhibited the lipolysis of TG and decreased the levels of FFAs in the plasma (Guo et al., 2012).

Insulin secreted from the  $\beta$ -cells of the pancreas is responsible for stimulation of blood glucose transport into skeletal muscle and adipose tissue as well as suppression of hepatic glucose

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production. Type 2 diabetes is a metabolic disorder associated in part with insulin resistance (Ghosh et al., 2007). Guo et al. showed that dietary anthocyanins-rich extract from black rice was capable of preventing the development of insulin resistance in fructose-fed rats and the underlying mechanism might be related mainly to inhibiting oxidative stress and improving the plasma lipid profile (Guo et al., 2007). Further, takikawa et al. (2010) showed that the dietary anthocyanins-rich extract from bilberry reduced the blood glucose level and improved insulin sensitivity via the activation of AMPK in type 2 diabetic mice. As shown in Figure 5, dietary bilberry extract activated AMPK in the white adipose tissue, skeletal muscle and liver. In the white adipose tissue and skeletal muscle, activation of AMPK increased the expression of glucose transporter 4 (Glut4), which resulted in the enhancing glucose uptake and utilization in these tissues. In the liver, activated AMPK decreased the expression of phosphoenol pyruvate carboxykinase (PEPCK) and glucose-6-phosphatase (G6Pase), leading to a decrease in the glucose output into the blood. In addition, AMPK activation in the liver also resulted in significantly decreased liver and serum lipid content via the phosphorylation of acetyl-CoA carboxylase (ACC) and upregulation of PPARα, acyl-CoA oxidase (ACO), and carnitine palmitoyltransferase-1A (CPT1A) gene expression (Takikawa et 2010). Mitochondrial acyl-CoA:glycerol-sn-3-phosphate al., acvltransferase 1 (mtGPAT1) controls the rate-limiting step of TG synthesis, which plays important roles in the development of metabolic diseases such as obesity and type 2 diabetes. Guo and coworkers (2011) demonstrated that protein kinase C  $\zeta$  (PKC $\zeta$ ) might be a potential target for

the prevention and treatment of steatotic liver associated with obesity and type 2 diabetes. In that study, anthocyanin cyanidin 3-O- $\beta$ -glucoside stimulated PKC $\zeta$  activation in HepG2 cells by increasing PKC $\zeta$  phosphorylation and membrane translocation from the endoplasmic reticulum to the outer mitochondrial membrane in order to phosphorylate the mtF0F1-ATPase  $\beta$ -subunit. As mentioned above, AMPK is an important energy metabolic regulator, whether AMPK activation was associated with the regulatory process of PKC $\zeta$  activation remain to be clarified.

In summary, anthocyanins are able to ameliorate disturbances in lipid and glucose metabolism, which are fundamental risk factors for obesity and diabetes. AMPK signaling pathway is one of the crucial factors for cellular energy homeostasis, which can be recognized as the key target in the prevention and treatment of obesity and diabetes.

#### 7. Other health benefits

Gout is a clinical syndrome in which tissue damage is induced by a chronic metabolic disorder associated with increased concentrations of uric acid in the blood. In a sodium oxonate-induced hyperuricemia mouse model, Hwa et al. reported the hypouricemic effects of anthocyanins-rich extract from the purple-fleshed sweet potato (Hwa et al., 2011). In addition, the ameliorative effect of anthocyanin extract from black rice against d-galactose-induced senescence has been demonstrated in a mice model (Lu et al., 2014). Jang and coworkers (2010) proposed that anthocyanins might be effective in treating prostatic hyperplasia. On the other hand, the protective roles of anthocyanin extracts from *Justicia secunda* Vahl against sickle cell disease

was indicated by both stabilizing the red blood cell membrane and inhibiting polymerisation of haemoglobin (Mpiana et al., 2010). Moreover, anthocyanin could even improve the developmental competence by stimulating nuclear reprogramming through the increased transcription factor expression (You et al., 2010). However, detailed mechanisms of action of anthocyanins for these beneficial effects remain to be clearly studied.

#### 8. Conclusion and future perspectives

Interests in anthocyanins have increased substantially during the past two decades. In this review, diverse health benefits of anthocyanins and molecular mechanisms have been described in detail. Several signaling pathways including MAPK, NF-κB, AMPK and Wnt/β-catenin, as well as some crucial cellular processes, such as cell cycle, apoptosis, autophagy and biochemical metabolism, are involved in these beneficial effects and may provide potential therapeutic targets and strategies for the improvement of a wide range of diseases in future. Nevertheless, much remains to be elucidated before better applying them for the health benefits of humans.

First, more studies on the biological activity of the certain anthocyanin metabolites are desired, since it has been suggested that anthocyanins are absorbed and transported in human serum and urine primarily as metabolites and these metabolites may totally or partially contribute to the above mentioned biological activities of anthocyanins.

Additionally, health benefits of dietary anthocyanins have been demonstrated in many *in vivo* and *in vitro* studies, as well as epidemiological and clinic research of human volunteers.

However, the low bioavailability of anthocyanins appears to be an obvious obstacle in achieving the desired beneficial effects (Xiao and Högger, 2015). Novel approaches for enhancing the bioavailability of these beneficial molecules in human bodies are desired to be developed. Fortunately, some new technologies such as nanotechnology may provide promising tools to solve this problem.

Moreover, purified anthocyanin individuals might exert different biological activities due to their specific chemical structures. Future studies should focus on careful and accurate characterization of the different anthocyanins, in order to better elucidating the molecular mechanism of their health benefits.

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Table 1 Estimation of daily consumption of anthocyanins from fruits, vegetables, and beveragesa.

Reproduced from the original source (Wu et al., 2006).

Food	Daily consumption (mg)
Fruits	
Apple, raw	0.70
Blackberry, raw	0.03
Blueberry, raw	3.39
Cherry, sweet, raw	0.56
Cranberry, raw	0.17
Grape, raw	1.77
Nectarine, raw	0.02
Peach, raw	0.12
Plum, raw	0.64
Raspberry, raw	0.93
Strawberry, raw	0.41
Vegetables	
Eggplant, raw	0.13
Cabbage, red, raw	0.82
Lettuce, red leaf, raw	0.01
Red radish, raw	0.14
Onion, raw	0.96
Bean, black, raw	0.13
Nuts	
Pistachio nut	0.004
Beverages	
Grape juice	0.93
Wine	0.66
Total	12.53

Table 2 Summary of the experimental findings on anti-cancer activity of anthocyanin-rich fruits and vegetables.

Sources	Major composition	Tumor tape	Experimental	Reference
			materials	
Blueberry	Malvidin 3- <i>O</i> -galactoside,	Metastatic	Mouse	Bunea et al.,
	petunidin 3- <i>O</i> -galactoside,	melanoma	melanoma	2013
	delphinidin 3-O-galactoside		B16-F10 cells	
Grape rinds,		Colon	Human colon	Koide et al.,
red rice, red		cancer	cancer HCT-15	1996; 1997
soybeans			cells; Balb/C	
and red			mice bearing	
beans			syngeneic	
			tumor-Meth/A	
			cell xenografts	
Seeds of	Cyanidin 3- <i>O</i> -β-glucoside	Colon	F344 rats	Hagiwara et
corn		cancer	treated with	al., 2001
			PhIP	
Sweetpotato	Nonacylated cyanidin	Leukaemia	Human	Konczak-Isla
	3-sophoroside-5-glucoside		leukemia	m et al.,
			HL-60 cells	2003
Grape,	Glucosylated derivatives for	Colon	Human colon	Zhao et al.,
bilberry and	grape and bilberry; cyanidin	cancer	cancer HT-29	2004
chokeberry	derivatives for chokeberry		cells	
Grape	Delphinidin	Breast	Human breast	Singletary et
		cancer	epithelial	al., 2007
			MCF-10F cells	
			treated with	
			benzo[a]pyrene	
Blackberry	Cyanidin 3-glucoside	Colon	Human colon	Dai et al.,
		cancer,	cancer HT-29,	2009
		breast	breast cancer	
		cancer and	MCF-7 and	
		leukemia	leukemia	
			HL-60 cells	

Black currant	Cyanidin 3- <i>O</i> -rutinoside	Liver cancer	Rats treated with diethylnitrosam ine and human liver cancer HepG2 cells	Bishayee et al., 2010; 2011
Black rice		Breast cancer	Human breast cancer MDA-MB-453 cells and mice bearing MDA-MB-453 cell xenografts	Hui et al., 2010
Purple, chokeberry, bilberry, purple carrot, grape, radish and elderberry	Nonacylated monoglycosylated anthocyanins	Colon cancer	Human colon cancer HT-29 cells	Jing et al., 2008
Uva caimarona fruits	Delphinidin 3- <i>O</i> -β-glucopyranoside and cyanidin 3- <i>O</i> -β-glucopyranoside	Larynx cancer, colon cancer, gastric cancer, breast cancer and cervical cancer	Human larynx cancer HEp-2, colon cancer HT-29, gastric cancer MKN-45, breast cancer MCF-7 and cervical cancer HeLa cells	Barrios et al., 2010
Chokeberry	Cyanidin glycosides	Cervical cancer	Human cervical tumor HeLa cells	Rugină et al., 2012
Blueberry	Malvidin glycoside	Colon cancer	Human colon cancer HT-29 cells	Srivastava et al., 2007
	Delphinidin 3-glucoside	Fibrosarco	Human	Filipiak et al.,

		ma	fibrosarcoma HT1080 cells	2014
Red sorghum bran		Breast Cancer	Human breast cancer MCF-7 cells	Devi et al., 2011
Purple-flesh ed sweet potato	Peonidin- and cyanidin-glucoside	Colon cancer	Human colon cancer SW480 cells and mice treated with azoxymethane	Lim et al., 2013
Bilberry, chokeberry and grape		Colon cancer	Rats treated with azoxymethane	Lala et al., 2006
Tart cherry	3-cyanidin 2"- <i>O</i> -β-glucopyranosyl-6"- <i>O</i> -α -rhamnopyranosyl-β-glucopyran oside	Colon cancer	APC <sup>Min</sup> mice	Bobe et al., 2006
Blackberry	Malvidin, petunidin and delphinidin	Lung cancer	Human lung cancer A549 cells	Aqil et al., 2012
Roselle	Delphinidin and cyanidin	Leukemia	Rats treated with nitrosomethylur ea	Tsai et al., 2014
Aronia meloncarpa E		Colon cancer	Human colon cancer HT-29 cells	Malik et al., 2003
Pomegranate fruit	Pelargonidin 3-glucoside, cyanidin 3-glucoside, delphinidin 3-glucoside, pelargonidin 3,5-diglucoside, cyanidin 3,5-diglucoside and delphinidin 3,5-diglucoside	Skin tumor	CD-1 mice treated with 12- <i>O</i> -tetradeca noylphorbol-13 -acetate	Afaq et al., 2005
Potato		Prostate cancer	Human prostate cancer PC-3 and LNCaP cells	Reddivari et al., 2007

Wild berry (Meoru)		Liver cancer	Human liver cancer Hep3B cells and mouse bearing Hep3B cell xenografts	Park et al., 2014
Anthocyanin metabolite	Protocatechuic acid	Esophageal cancer	Rats treated with N-nitrosomethy lbenzylamine	Peiffer et al., 2014
Anthocyanin metabolite	Gallic acid, 3-O-methylgallic acid and 2,4,6-trihydroxybenzaldehyde	Colon cancer	Human colon cancer cells (HCT-116, Caco-2, SW-480, HT-29 and HCT-15)	Forester et al 2014
Wild-grown berries	Cyanidin 3- <i>O</i> -rutinoside and cyanidin 3- <i>O</i> -glucoside	Liver cancer	Human liver cancer PLC/PRF/5 and HepG2 cells	Longo et al., 2008

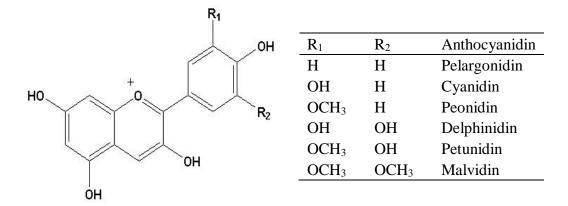
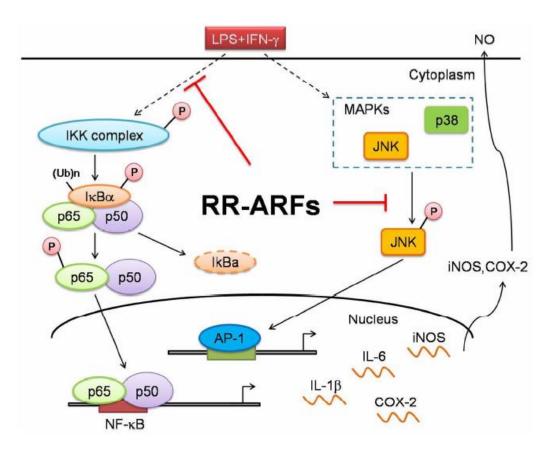
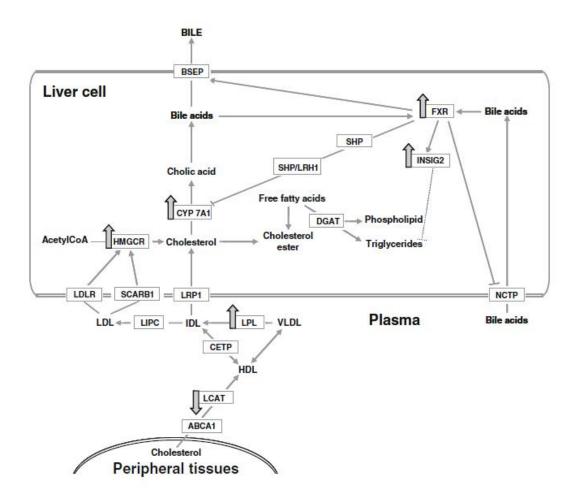


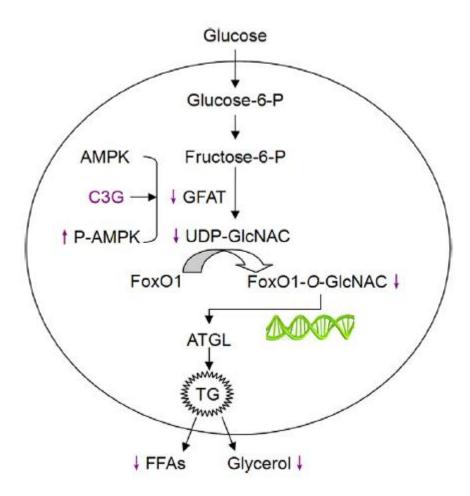
Figure 1 Structure of the anthocyanidins most commonly found in foods.



**Figure 2** Schematic model showing role of anthocyanin-rich fractions from red raspberries (RR-ARFs) in inflammatory signaling pathways. RR-ARFs attenuated LPS/IFN-γ-induced inflammatory responses through inhibition of NF-κB, MAPK/JNK activities, respectively, in LPS/IFN-γ-stimulated RAW264.7 cells. Reproduced from the original source (Li et al., 2014).



**Figure 3** Schematic model showing changes in the expression of genes involved in cholesterol metabolism by an anthocyanin-rich bilberry extract in apoE<sup>-/-</sup> mice. Large arrows indicate up- or down-regulation of genes involved in bile acid synthesis and excretion, which enhances cholesterol elimination in the plasma and the reduction of hepatic lipogenesis. Reproduced from the original source (Mauray et al., 2010).



**Figure 4** Schematic model showing the antilipolytic role of anthocyanin in highglucose-incubated adipocytes through regulating FoxO1-mediated transcription of ATGL. Cyanidin 3-*O*-β-glucoside treatment decreases the cellular GFAT activity resulting in diminished formation UDP-N-acetylglucosamine. This disrupts high-glucose-induced O-GlcNAc modification of FoxO1, inhibits ATGL transcription, and limits lipolysis. Reproduced from the original source (Guo et al., 2012).

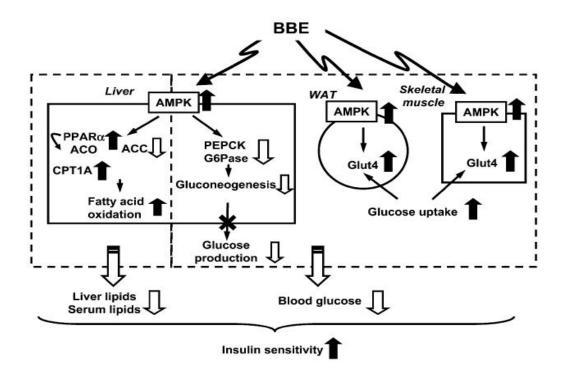


Figure 5 Proposed mechanisms for amelioration of hyperglycemia and insulin sensitivity by dietary bilberry extract (BBE). BBE activates AMPK in the white adipose tissue and skeletal muscle. This activation induces upregulation of Glut4 and enhancement of glucose uptake and utilization in these tissues. In the liver, dietary BBE reduces glucose production *via* AMPK activation, which ameliorates hyperglycemia in type 2 diabetic mice. The AMPK activation induces phosphorylation of acetylCoA carboxylase (ACC) and upregulation of PPARα, acylCoA oxidase (ACO), and carnitine palmitoyltransferase-1A (CPT1A) gene expression in the liver. These changes lead to reductions in lipid content and increase in insulin sensitivity *via* reduction of lipotoxicity. Reproduced from the original source (Takikawa et al., 2010).