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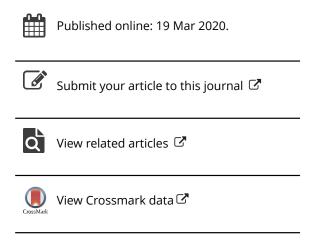
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REVIEW



Toxicological effects of marine seaweeds: a cautious insight for human consumption

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ABSTRACT

Marine environment is a rich and diverse source for many biologically active substances including functional foods and nutraceuticals. It is well exploited for useful compounds, natural products and aquaculture industry; and seaweeds is one of the major contributors in terms of both food security and healthy nutrition. They are well-known due to their enormous benefits and is consumed globally in many countries. However, there is lack of attention toward their toxicity reports which might be due toxic chemical compounds from seaweed, epiphytic bacteria or harmful algal bloom and absorbed heavy metals from seawater. The excess of these components might lead to harmful interactions with drugs and hormone levels in the human body. Due to their global consumption and to meet increasing demands, it is necessary to address their hazardous and toxic aspects. In this review, we have done extensive literature for healthy seaweeds, their nutritional composition while summarizing the toxic effects of selected seaweeds from red, brown and green group which includes- Gracilaria, Acanthophora, Caulerpa, Cladosiphon, and Laminaria sp. Spirulina, a microalgae (cyanobacteria) biomass is also included in toxicity discussion as it an important food supplement and many times shows adverse reactions and drug interactions. The identified compounds from seaweeds were concluded to be toxic to humans, though they exhibited certain beneficial effects too. They have an easy access in food chain and thus invade the higher trophic level organisms. This review will create an awareness among scientific and nonscientific community, as well as government organization to regulate edible seaweed consumption and keep them under surveillance for their beneficial and safe consumption.

KEYWORDS

Acanthophora; Caulerpa sp.; Cladosiphon; Gracilaria sp.; Laminaria japonica

Introduction

Most life forms on earth have evolved approximately 450 million years ago as marine habitats (Dring 1982). Considering volumetric scale, sea provides almost 90 percent of the living space on the planet. Among all the living forms of life in water, fish is the earliest and exclusive vertebrate habitat in water. Out of them, many evolved into land mammals such as seals, dolphins or whales later. There are total ~226,000 eukaryotic marine species, which includes 20,000 fish species, though there are about two million marine species which still haven't been documented (Appeltans et al. 2012). These species measure from microscopic (plankton and phytoplankton- 0.02 micrometres) to huge mammals (whales and dolphins-33 metres). Phytoplankton are primary producers and play an integral role in the ocean food chain. Water plants such as algae are the basis of some underwater ecosystems and are a part of well-established aquaculture industry, and accounts for approximately 10% growth rate (Ginneken and de Vries 2016). The Food and Agriculture Organization (FAO) states aquaculture as farming of aquatic organisms such as fish, mollusks, crustaceans and aquatic plants. It can be classified into fish, shrimp, oyster and ornamental fish farming; mariculture and alga culture (seaweed farming).

Source of bioactive compounds and nutrients

Bioactive compounds can be either natural or synthetic in origin, and are biologically assayed for activities in a number of key therapeutic areas (MacArtain et al. 2007). Seaweeds are rich source of biologically active compounds including minerals, lipids, proteins and polyphenols (Kumar et al. 2008). They synthesize a variety of these compounds such as amino acids, terpenoids, acetogenins, alkaloids, chlorophyll, carotenoids, xanthophylls, saturated fatty acids as well as unsaturated fatty acids, halogenated compounds, vitamins K, B, A; along with alginate, proteoglycans, laminarin, fucoidan, carrageenan and galactosyl glycerol (Renn 1997; Masuda et al. 1997; Ito and Hori 1989; Skrovankova 2011; Kadam et al. 2015; Manzelat et al. 2018). These compounds exhibit antimicrobial, anti-viral, anti-coagulant, herbivore deterrent, allelopathic, and antifouling activities (Richards et al. 1978; Del Val et al. 2001; Marechal et al. 2004; Athukorala et al. 2006). They can contain up to 47% protein of the dry

weight depending on the species and their cultivation and harvesting conditions.

Porphyra is one of the most valuable alga culture crop, belonging to group Rhodophyta. It is popularly known as nori and provides excellent nutrition. It has high vitamin C, protein, minerals and trace elements (Huisman 2006). It also contains bioactive polysaccharides (porphyrin), polyphenols, proteins, chlorophyll and carotenoids. It is helpful in reducing breast cancer risks in postmenopausal (35%) and premenopausal (65%) women (Brown et al. 2014). Porphyra umbilicalis contain 34.2 mg/100 g wet weight of calcium, 301.2 mg/100 g wet weight of potassium, 108.3 mg/100 g wet weight of magnesium, 119.7 mg/100 g wet weight of sodium, 0.1 mg/100 g wet weight of copper, 5.2 mg/100 g wet weight of iron, 1.3 mg/ 100 g wet weight of iodine, and 0.7 mg/100 g wet weight of zinc. 8 g of this seaweed can provide a carbohydrate content of almost 3.8 units of carbohydrates and 2.7 units of total fiber, whereas Institute of Grocery Distribution has a guideline of daily amount (GDA) of 11.25% (MacArtain et al. 2007).

Ulva is a blade-like species belonging to Chlorophyta group, commonly known as sea lettuce. They are thin and membranous leaves, with merely two cells thickness width. It is consumed directly in the raw form (Huisman 2006). They contain approximately 24.4% of proteins on dry weight basis (Kumar et al. 2008). 8 g of Ulva provides 3 units of total fiber which is more than what a banana contains, while the GDA limit for carbohydrate content is 12.5 units. It is also a novel source for minerals, consisting of 325 mg/100 g wet weight of calcium, 245 mg/100 g wet weight of potassium, 465 mg/100 g wet weight of magnesium, 340 mg/100 g wet weight of sodium, 0.3 mg/100 g wet weight of copper, 15.3 mg/100 g wet weight of iron, 1.6 iodine and 0.9 mg/ 100 g wet weight of zinc.

Ascophyllum nodosum regulates insulin level positively (Catarino, Silva, and Cardoso 2018). It is commonly known as Egg wrack contains a total fiber content of about 8.8 units per 8 g of seaweed. It has 575 mg of calcium, 765 mg of potassium, 225 mg of magnesium, 1173.8 mg of sodium, 0.8 mg of copper, 14.9 mg of iron, 18.2 mg of iodine, where all the minerals are in the unit mg/100 g wet weight (MacArtain et al. 2007). Undaria pinnatifida belong to the class of phaeophyte and contains approximately 9.14% of carbohydrates and dietary fibers, about 3.03% of proteins and approximately 0.64% of lipids (Wang et al. 2018). It reduces postprandial and fasting glucose levels, and also alters lipid profile by reducing the triglyceride levels and increasing HDL levels (Grasa-Lopez et al. 2016).

Hematococcus pluvialis contain astaxanthin, a natural antioxidant which is three times more potent than β carotene. It exhibits anti-inflammatory, immunomodulatory and anti-ageing activity (Chu 2011; Davinelli, Nielsen, and Scapagnini 2018).

Seaweeds also contain hypotensive peptides, which play an important role in combating cardiovascular disease risk factors. Thus, incorporation of hypotensive peptides containing seaweeds as functional food in form of beverages and soups is beneficial (Fitzgerald et al. 2011). Red, brown and green algae have been detected with cytostatic, anthelmintic, antibacterial,

antiviral and antifungal properties. Pharmaceutically, they are employed in drug development to treat AIDS, arthritis, cancer, inflammation and pain (Lee et al. 2013). They are claimed to produce purer form of fatty acids found in human milk, considered as the building blocks for visual and mental development (De Almeida et al. 2011). They also contain unique polysaccharides and phlorotannins with many health promoting benefits (Brown et al. 2014). Brown algae contain antioxidant and anticancer compounds like carotenoid- fucoxanthin and polyphenolic compound- phloroglucinol (Padua et al. 2015). In an attempt to determine the anticancer activities of fucoxanthin and phloroglucinol alone or in combination, studies were performed in two colorectal cancer cell lines (HCT116 and HT29), and was compared to a normal colon cell line (CCD-18Co). Fucoxanthin and phloroglucinol alone reduced cancer cell line viability without an effect on normal cells, and when used in combination it enhanced the cytotoxic effect of 5-fluorouracil in colon cancer cells (Costa et al. 2017). Brown algae is rich in biologically active compounds belonging to the class of carotenoids, $\omega - 3$ fatty acids, polysaccharides, lipids, proteins and also secondary metabolites, such as terpenes and polyphenols (Reboleira et al. 2019). Pectin, guar gum etc. are soluble polysaccharides from algae and possess hypocholesterolemic and hypoglycemic activity, whereas cellulose a water insoluble polysaccharaide is associated to cause a decrease indigestive tract transit time (Kolanjinath, Ganesh, and Sanraj 2014). Algae are also known as super foods due to their enormous benefits as a source of dietary protein, B-vitamins and iron. They help in weight loss, attention deficithyperactivity disorder (ADHD), maintain sugar level, relieve stress, fatigue, anxiety, help in premenstrual syndrome (PMS) and in other health issues in women. The enormous beneficial effects of seaweeds also offer its use in coronary heart disease, cancer and hyperlipidemia in Japan and Western countries (Kim et al. 2009; Iso 2011; Brown et al. 2014). Seaweeds belonging to the class Phaeophyta such as Laminaria and Undaria contain 83-97% of fucosterol in total sterol content (662-2320 μg/g dry weight) whereas Palmaria and Porphyra which belong to Rhodophyta contain 87-93% desmosterol of total sterol content (87–337 μ g/g dry weight). Few red seaweeds for example *C. crispus* has cholesterol as major sterol. It is reported that plant sterol like β -sitosterol and fucosterol leads to the decrease in the concentration of cholesterol in the serum experimentation in animals and human. Fucoidan (5) obtained from brown seaweeds is said to have reported in vivo and in vitro heparin like activity that demonstrate anti- thrombic and anticoagulant activity. A strong anticancer activity is demonstrated by Brown seaweeds extract obtained from Laminaria japonica (Kombu). Since it decreases the expansion of cancer. Regular use of Laminaria japonica inhibit the risk of the breast cancer considerably (Pal, Kamthania, Kumar 2014)

Toxic effects of seaweed

Seaweeds contain heavy metals and is a matter of concern due to the associated potential health risks. Inspite offering such extensive array of healthy effects, there are some

worrying incidents where avoiding excessive amounts of intake may prevent adverse effects on health of human individuals.

In Japanese population, iodine-induced goiter and transient hypothyroidism are common and gets altered by limiting the seaweed intake and thus balance the high iodine intake due to the seaweeds as a regular diet (Zava and Zava 2011). Seaweeds contain heavy metal arsenic in the form of arsenosugars, which can be further metabolized to various arsenic compounds in humans. Taylor et al. (2017) tested urinary arsenosugars and their metabolites in samples prior and post seaweed consumption in 24-hour urine samples while consuming seaweeds; tested compounds were dimethyl arsenate (DMA), thio-dimethylarsinoylethanol (thio-DMAE), thio-dimethylarsinovlacetate (thio-DMAA), dimethyl arsenate (thio-DMA) in the urine samples. Unbroken arsenosugars along with DMA, thio-DMAA, and thio-DMAE increased in urine, after ingesting nori, wakame, and kombu seaweeds. Trace amount of thio-DMA, a known toxic metabolite was present across the individuals. As commercial products from whole seaweed contain considerable concentrations of arsenic (12-84 µg/g), an assessment of these toxic compounds can help to monitor dietary intake of arsenic.

In earlier study carried out for the risk assessment of the inorganic arsenic in five edible seaweeds, they did not show any human health hazards. Laminaria japonica, Porphyra yezoensis, Undaria pinnatifida, Hizikia fusiformis and Enteromorpha prolifera were analyzed for arsenite, arsenate, methylarsonate, and dimethylarsinate (Zhao et al. 2014). Desideri et al. (2016) analyzed all the essential, nonessential and toxic elements present in seaweeds commonly consumed in Italy. Iodine level was highest in Laminaria digitata at 7316 mg/kg dry. Ulva lactuca contained the highest levels of Cu, Ni, Mn, and Pb. Lithothamnium calcareum had highest levels of Ca, Al, Si, Fe, and Ti; Palmaria palmata had the highest concentrations of K, Rb, and Cl; Chondrus crispus had the highest level of S. Laminaria digitata also exhibited highest concentrations of total As, Cd, Sn, Br, and I; Chlorella pyrenoidosa showed highest concentration of Zn. With a growing concern and awareness of heavy metals in seaweeds, Chen et al. (2018) recently investigated the distribution of 10 metals and metalloids in 295 dried brown and red seaweeds and calculated the hazard index for them. Mean value of elements in seaweeds were sequenced in descending order as: Al > Mn > As > Cu > Cr > Ni > Cd > Se > Pb > Hg. The levels of Cd, Cu, Mn and Ni were significantly higher in red seaweeds than brown seaweeds (P < 0.01). The results suggest a continuous surveillance for Cd, Pb and Hg in edible seaweeds. The high nutritional composition of essential elements found in algae might serve as an important source to fulfill the health requirements. However, the non-essential elements, which are toxic in nature is estimated with associated risks and its estimators.

There are no reports of poisonings by the most popular and commercially viable seaweeds such as Porphyra (nori), Laminaria (kombu), and Undaria (wakame), however there are few seaweeds reported to cause illness and death for e.g.

Gracilaria, Acanthophora, and Caulerpa. Till date, there are 14 deaths reported due to consuming seaweed, but the reports claim it to be due to the bacteria grown on the seaweed responsible for these deaths (Cheney 2016). U. fasciate, C. taxifolia, E. linza and S. johnstonii acute toxicity was tested by direct feeding of powdered samples to wistar-CFT strain (Rams norvegicus, albicus) rats of each sex (body weight 200-250 g) at 0.5, 2.0 and 5.0 g/kg body weight. The doses were equivalent to consumption of 10, 50 and 100 g of seaweed per 60 kg of human body weight. Hemagglutinating mechanism was understood to be reason for toxicity, which is generally given by lectins. This may also lead to growth retardation in animals, probably due to their ability to bind to specific receptor sites on the surface of the intestinal epithelial cells resulting in impairment of nutrient absorption (Naidu et al. 1992).

In this review, we have discussed seaweeds reported for illnesses and toxicity with their chemical compounds which might affect humans. We have selected seaweeds which have been reported for number of poisonings from Rhodophyta (red) - Gracilaria, Acanthophora; Chlorophyta (green) -Caulerpa and Ochrophyta (brown) - Cladosiphon, Laminaria species and have done extensive literature search for their chemical constituents. Figure 1 shows the selected seaweeds which we discussed in our study. Since Spirulina, a cyanobacteria is widely consumed globally and have been mostly reported for their significant beneficial properties, we have also added its toxic reaction, as it is imperative to review human population of their toxicity due to its high consumption mostly in coastal region and for their cautious use.

Toxic seaweeds

Gracilaria genus

Gracilaria Greville, 1830 is a Rhodophyta and belong to the Gracilariaceae family. There are total 160 species accepted taxonomically, among more than 300 species. They are mostly found in the tropical and subtropical region and widely cultivated in Asia, South America and Africa. It shows presence of important chemical compounds such as phycocolloid, acrylic acid and other bioactive metabolites (De Almeida et al. 2011). They generally grow as large beds either in the eulittoral or sublittoral zone, on muddy or sandy sediments which are protected from waves and are employed for nutraceutical, pharmaceutical and biotechnological applications (Francavilla et al. 2013).

Gracilaria is an important source of agar, which is a nontoxic sulfated carbohydrate gelatinous in nature and widely used in the preparation of various food products, ice-creams, cosmetics and bacteriological samples. It is employed pharmaceutically as a bulking agent, anticoagulant agent, and laxatives in capsules and tablets. Agarose, highly purified agar is used in molecular biology for separation techniques like electrophoresis, immunodiffusion and gel chromatography (Pal, Kamthania, and Kumar 2014). It is also employed as a producer of carrageenan and alginate in western countries like Venezuela, USA and Canada. Alginate is used to prolong the activity of drugs (Konecny 1969). Carrageenan can also be used as a potential pharmaceutical

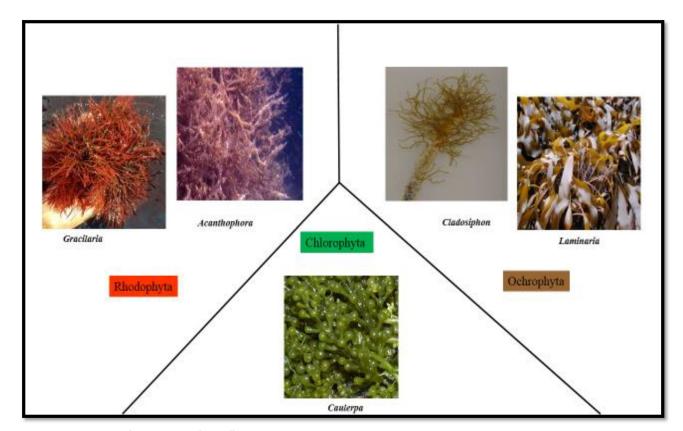


Figure 1. Seaweeds reported for their toxicity from different groups.

drug in fields such as anti-coagulant therapy, anti-tumour therapy, anti-viral therapy and immunomodulation activities (Pal, Kamthania, and Kumar 2014).

It is consumed widely in Japan, Hawaii, Guam, Malaysia and the Philippines. The human intoxications from Gracilaria ingestion have occurred in Japan, Sakata, Yamagata Prefecture, Toyo and Ehime (Konecny 1969). Till date, it has been found responsible for the maximum number of illnesses and deaths in the Pacific Rim countries. Majority of the incidents of death and illness were caused by the species G. verrucosa and G. edulis which is also known as Polycarvernosa tsudai (Cheney 2016). One of the major cases of human intoxication due to ingestion of red algae G. edulis reported 3 deaths and 13 people falling ill. The worst outbreaks with maximum number of deaths occurred in Japan in the year 1980, 1982 and 1993 along with one in Guam in the year 1991 and two in Philippines in the year 2002 and 2003. The other poisoning was reported in California in 1992, and in Hawaii in the year 1994 (Cheney 2016). In another incident, one of two patients died due to mere ingestion of G. verrucosa (Japanese name, ogonori) in Kanagawa, Japan in the year 1993 (Noguchi et al. 1994; Yotsu-Yamashita et al. 2004). The poisoning caused by G. verrucosa and G. chorda (tsurushiramo) occurred in 1980, 1982 and 1993 in Japan and were characterized by symptoms such as nausea, vomiting, stomach pain and diarrhea. G. chorda diseased four individuals and resulted into one death among them. G. verrucosa infected six and caused one death in 1982; later infected two and caused one death in 1993 (Fusetani and Hashimoto 1984). A similar incident had occurred due to consumption

of Gracillaria vermiculphylla, where symptoms appeared within one hour of consumption followed by the other gastric disturbances such as nausea, vomiting and diarrhea (Hammann et al. 2016). In the investigations carried out by Fusetani and Hashimoto (1984), water soluble fractions of G. verrucosa and G. chorda led to death of mice within 9 mins to 20 hours at a dose equivalent to 1-2 g of wet material, whereas diarrhea was reported in mice within 5-10 mins at a dose equivalent to 0.5 g of the extract or less. This study resulted into isolation of two prostaglandins i.e. prostaglandin A₂ (PGA₂) (1) and prostaglandin E₂ (PGE₂) (2), which were suspected for the ogonori poisoning. These prostaglandins were isolated from G. vermiculophylla and G. verrucosa, wherein PGE₂ (2) was found to be more toxic than PGA₂ (1) indicating it as a probable agent for causing poisoning. PGE₂ (2) is also reported to be present in G. verrucosa and G. lichenoides. This prostaglandin is known to cause a wide range of physiological activities in humans, such as diarrhea, nausea, bleeding and hypertension. It is administered for inducing child birth through artificial means. About 0.5 mg of PGE₂ (2) is administered orally to pregnant women (Osathanondh et al. 1980; Hammann et al. 2016). Symptoms such as temporary hypertension backed by hypotension with bleeding, headache, vomiting, diarrhea and nausea is observed in these cases. With an increase in the dose by almost twelve folds, the casualty can suffer from nausea, abdominal pain and vomiting instantaneously. Hypotension can also result in a very short span of time which would eventually lead to death due to hypotensive shock. The hypotension caused can be attributed to PGE₂ (2) due to its activity of shrinking the womb. It also leads to an excessive

Figure 2. Compounds obtained from different seaweeds found responsible for their toxicity.

excretion of urine (Konecny 1969). PGE₂ (2) can also increase cardiovascular risk by contributing to atherogenesis and influence inflammatory, immune, and oxidative stress responses, which are reported to alter growth, fibrosis, and apoptosis in renal cells (Nasrallah, Hassouneh, and Hebert 2016). The possible reason for the toxicity due to PGE₂ (2) can be attributed to the fact that, the cyclooxygenase (COX) in raw seaweeds use the highly unsaturated fatty acids to produce large amounts of PGE2 (2) in the stomach of victims in a short lapse of time. As per the paper recently a COX gene producing PGF_{2α} was cloned from this alga and heterologous expressed (Hsu et al. 2007; Di Costanzo et al. 2019).

In an unpleasant incident, a couple had fallen ill within 3 hours of consumption of 20-30 g of raw and freshly collected G. verrucosa. The wife developed low blood pressure and eventually leading to unconsciousness and death after 14 hours because of hypotensive shock, whereas the husband gradually recovered and survived. The sample consumed was investigated to have PGE2. Though the concentration of PGE2 (2) was not high, yet the toxic levels were produced due to the presence of an unsaturated fatty acid -arachidonic acid in combination with PGE₂ (Cheney 2016). However, it is also reported to show immunopotentiating activity when administered orally and intraperitoneally to mice at a dose of 4.0 mg, along with promoting phagocytosis (De Almeida et al. 2011).

When Hawaii was affected by G. coronopifolia poisoning, it affected seven people in 1994. The symptoms included vomiting and diarrhea, along with an additional symptom of burning sensation mouth throat.

Debromoaplysiatoxin (3) and Aplysiatoxin (4) were found to be the causative constituents for the additional symptoms (Figure 2). Debromoaplysiatoxin (3) was found to be less toxic than aplysiatoxin (4) from the toxicity studies performed in mice; the symptoms included immediate diarrhea within 30 minutes of consumption. Apart from this symptom, lethargy, muscular contractions and paralysis of hind legs were also observed. They also caused irritation on physical contact (Nagai, Yasumoto, and Hokama 1996). Aplysiatoxin (4) causes bleeding, followed by hemorrhagic shock in mice. When a morphological evaluation of diarrhea caused by aphysiatoxin (4) in mice was carried out, it showed that the diarrheic components apparently originated from capillaries in large intestine and were secreted directly into the lumen. The number of goblet cells increased exceptionally while showing fine cracks in the epithelium, the effect was observable within 4.5 hrs and then for 4 hrs intermittently even when the mice was administered a sub lethal dose of aplysiatoxin intraperitoneally (Ito and Nagai 1998). It was also found that different routes of administration had different site of actions but bleeding took place in both the cases. When the toxin was administered through an i.p. injection at about 250 mg/kg, initially the lymphatic vessels were dilated and later there was congestion in the lamina propria after 10 mins of the administration, where as an IV administration of about 100-200 μg/kg resulted in the death of the mice within 15 mins and bleeding was observed only in the lungs around the distended pulmonary artery (Ito and Nagai 2000).

The poisoning due to G. edulis (Polycavernosa tsudai) in Guam in the year 1991 and in the Philippines in the year 2002 is considered to be one of the deadliest poisonings (Yotsu-Yamashita, Haddock, and Yasumoto 1993). Both outbreaks collectively caused five deaths and sickness to twentytwo people. This outbreak symptom included vomiting, diarrhea and numbness of the extremities including muscle spasms. The chemical compounds responsible for the toxicity were found to be polycarvenoside A (5) and polycarvenoside B and were suspected to be the causative agent of the poisoning. The structure of polycarvenoside A (5) is very similar to that of the aplysiatoxin (4) which is found in G. cornopifolia and G. confervoides (Nagai, Yasumoto, and Hokama 1996). Polycarvenoside A (5) is a glycosidic toxin and constitutes of a lactone macrocyclic core. A trienyl side chain and a methylated disaccharide component is coupled in polycarvenoside A (5). Several toxicity studies were carried out for polycarvenoside A (5), which showed that it is responsible for neurological alterations like muscle spasms, scratching and paralysis. It also induces gastrointestinal disturbances like vomiting, diarrhea, depolarization of the neuroblastoma cells and increase the calcium level in the cytosol (Cagide et al. 2007). Estimated LD₉₉ for polycarvenoside A (5) and polycarvenoside B was determined to be 200-400 μg/kg in mice (Yotsu-Yamashita et al. 2004). G. edulis showed 100% sperm motility inhibition, due to the disruption of the plasma caused by the presence of spermicidal compounds (De Almeida et al. 2011). Apart from the above-mentioned compounds, Gracilaria poisoning could also be attributed to manauealides A, manauealide B, manauealide C, malyngamide M and malyngamides N. Nagai, Yasumoto, and Hokama (1997) reported that aplysiatoxinlike compounds, the manauealides, were present in Gracilaria coronopifolia. Weak cytotoxicity was shown by malyngamide compound in mice neuroblastoma cell line. It caused diarrhea in mice in the similar manner to aplysiatoxin (4) and debromoaplysiatoxin (3) (Higa and Kuniyoshi 2000).

On the basis of above reported studies, Gracilaria genus can be concluded to be taken with caution, and a detailed investigation of its or associated epiphytic compounds can be helpful to keep the species from this genus under surveillance.

Acanthophora genus

Acanthophora is a Rhodophyta, belonging to the family Rhodomelaceae (Preskitt 2001). It is widely distributed in Guam, Houtman Abrolhos and Hawaii. In Hawaii, it is considered a non-indigenous species, since it got involuntarily introduced from Guam to Oahu Pearl Harbor, Hawaii in the year 1952. Eventually, it turned out to be the most common non-indigenous algal species in the main Hawaiian Islands, supplanting many native prevalent species (Smith, Hunter, and Smith 2002). It successfully invaded the benthic habitats, due to its highly adaptable nature in diverse range of hydrological conditions, efficacious epiphytism and reproduction by both sexual and vegetative means as A. specifera undergoes fragmentation too. It is a highly favored and

palatable food for herbivorous fishes and is consumed by green sea turtles as well. It is a good source of β -carotene, antheraxanthin and carrageenan. It also contains large amount of other nutrients, polysaccharides, sterols, lipids, dipeptides and diketosterol. Recently two bioactive flavonoids having preventive procoagulant activity in adherent human monocytes, antiplatelet and antioxidant properties have been isolated (Zeng et al. 2010). Polycarvenoside already characterized in Gracilaria was also found in Acanthophora species, wherein it contained side chains of conjugated diene and triene along with an o-methylated Lfucosyl-D-xylose. The toxicity studies performed for the polycarvenoside analog in mice revealed that the aglycone part along with the conjugated polyene side is responsible for the fatal toxicity by A. specifera and G. edulis. The amount of polycarvenoside was found to be 72 nmol/kg by LC/ESI-MS; whereas other polycavernoside congeners, debromoaplysiatoxin (3) and aplysiatoxin (4) were beyond the detection limit (Yotsu-Yamashita et al. 2004).

A. specifera caused two major outbreaks in the year 2002 and 2003 in the Philippines. These outbreaks caused thirtythree deaths cumulatively. The symptoms were very similar to G. edulis poisoning, viz. neurological disorders accompanied by gastrointestinal disorders such as nausea, vomiting etc. A brine shrimp lethality test performed with the methanolic extract of the seaweed showed a dose dependent relationship, where the mortality was found to increase with the concentration in the range of 2-10 µg/ml. The same extract obtained from the seaweed has also reported in antioxidant effects which was found to be correlated to the total phenolic content (Ailli Zakaria et al. 2011). A. specifera showed presence of polycarvenoside A. G. edulis poisoning in the year 1991 in Guam and in the year 2002 in Philippines showed presence of polycarvenoside A (Cheney 2016). The synthesis of this lethal toxin from both edible G. edulis and A. specifera alga gained momentum since 2012, where many researchers started its synthesis in lab for its potential use in antibiotic activity model (Pereira, Latino, and Gaudencio 2014). Firstly, Kasai, Ito, and Sasaki (2012) achieved total synthesis of (-)-polycavernoside A (5), via a convergent approach; later total synthesis by Brewitz et al. (2013) and stereo selective synthesis of γ -butyrolactones subunit of polyeavernoside A (5) is reported by Kadari et al. (2018). It has also been reported for the accumulation of copper and cadmium from the Gulf of Mannar with 7.83 mg/g of copper that was accumulated in the seaweeds and the maximum concentration of cadmium that was found was 1.30 mg/g. Babu et al. (2014) revealed that copper was comparatively more toxic than cadmium and led to glutathione reduction, fluctuations of superoxide dismutase, catalase and glutathione peroxidase activity corresponding to the time of exposure. It also led to a decrease in the antioxidant properties due to the metal accumulation, which can be attributed to increased oxidative stress and induced antioxidant Defense system against ROS and DNA damage was also reported.

Further investigations revealed that it contains a flavone compound later to be identified as apigenin and on further studies it was found out that apigenin had promising analgesic and anti-inflammatory activities. To confirm the bioactivity hot plate and writhing test in mice, as well as tail-immersion, carrageenan-induced paw edema and cotton pellet-induced granuloma formation in rats were performed. The possible mechanism for the above responses was the ability of apigenin to inhibit PGE2 (2) as well as proinflammatory cytokines such as interleukin-1 β , interleukin-6, and TNF- α (El Shoubaky et al. 2016).

Caulerpa genus

Caulerpa genus belongs to the chlorophyta group in the family Caulerpaceae. The green algae Caulerpa genus consists of a thallus containing one cell with many nuclei and is coenocytic in nature. Three species, particularly C. racemosa, C. racemosa var. laetevirens and C. lentillifera are widely consumed (Nagappan and Vairappan 2014). It is a nonnative, invasive species in the Mediterranean, and an important stressor for several native organisms (Gorbi et al. 2014). Caulerpa racemosa var. cylindracea was introduced from south-western Australia (Klein and Verlaque 2008). Since 1990, it has been invading the Mediterranean Sea and the Canary Islands, raising ecological problems. It is widely distributed in the tropical seas. In the Southeast Asian waters, there are at least ten known species. It has become serious biological pollutant due to its invasive property and grows on substrates like sand, rocks and mud. It grows in the Mediterranean Sea, where it was accidently introduced in the year 1984. Due to its fast spreading property, it proliferated rapidly from Italy to Spain. They produce repulsive secondary metabolites which is used against grazers and epiphytes. Caulerpenyne (6) has been identified from total 11 species belonging to Caulerpa. Out of these 11 species, two species C. prolifera and C. taxifolia are found in the Mediterranean Sea, among which C. taxifolia garnered the nickname of "killer alga". Chemically, caulerpenyne (7) is a sesquiterpene having a diacetoxy-butadiene function (Figure 2) and exhibit antitumoral effect which has been hardly explored (Sfecci et al. 2017). There are many other minor metabolites which are also studied for their toxic activity, as they are transferred to the marine food chain. There are many poisoning cases reported due to consumption of fish Sarpa salpa, also known as the Mediterranean Sea bream.

Ciguatera poisoning a foodborne disease, is caused by consumption of S. salpa fish contaminated by ciguatera toxin. Chemically, it is a lipid soluble compound generally produced by dinoflagellates. It generally causes nausea and pain, and can lead to neurological disorders such as hallucinations, vertigo and amnesia along with cardiac symptoms in humans (Massin 1996; Kurt et al. 2009). However, later it was found that the causative fish had been feeding on C. prolifera. S. salpa from the Tunisian coast was also found to be toxic. As reported by Kurt et al. 2009 caulerpenyne (7) alters ATP-dependent calcium storage in the intracellular organelles, protein phosphorylation and in DNA synthesis. The antiproliferative activity of this toxin was similar to the anticancer drug -fotemustine and cisplatin. Cisplatin shows its action by binding with DNA and tubulin constituent of the cytoskeleton. Additionally, a few mollusks feeding on Caulerpa also exhibited a significant increase in their toxic metabolites, resulting into toxicity to its predators.

Its toxicity to humans in very high doses was also reported. It proved its toxicity to human melanocytes, keratinocytes and fibroblasts. It revealed cytotoxic activity at the concentration of 60-90 µM leading to the destruction of human hematopoietic progenitors. It didn't prove to be toxic to normal melanocytes in concentrations $< 10.5 \,\mu\text{M}$ and normal keratinocyte <12.6 µM (Massin 1996). Caulerpenyne (7) is found to be neurotoxic since it inhibits the onset of neurite which acted as a trigger for apoptosis leading to cell degeneration and death. Apoptosis was identified by chromatin condensation within the nucleus and intact nuclear boundaries whereas the late apoptotic cells were identified by nuclear fragmentation (Kurt et al. 2009). There are also reports of a toxic constituent named caulerpicin (7) (Figure 2), obtained from C. racemosa which has anesthetizing sensation and causes numbness of the tongue, lips and cold sensation in the fingers and the feet. Felline et al. (2012) showed an indirect mechanism which contributed to change in fish stocks by accumulation of caulerpin (8) in fish tissues. Some enzymatic pathways were activated while few got inactivated. Catalase, glutathione peroxidases, glutathione S-transferases, total glutathione were enzymes which were activated; acetylcholinesterase and acylCoA oxidase were inactivated which indicates their neurotoxic potential. Sudden difficulty in breathing is also reported. Caulerpin (8) isolated from C. racemosa, C. serrulata and C. sertularioides is an orangish red pigment. It was chemically characterized as a dimer of indole-3-acrylic acid (Nagappan and Vairappan 2014). It also demonstrated plant growth activities (Higa and Kuniyoshi 2000). Apart from the Caulerpa toxins, caulerpin and caulerpinic acid possess insecticidal activity against Culex pipiens, which is a common house mosquito (Cheney 2016). Caulerpin (8) produces mild anesthetic action, difficulty in breathing, sedation and loss of balance (Figure 2). These toxins can be explored for their use for diseases caused by mosquitoes for effective and efficient treatment.

Cladosiphon genus

These brown seaweeds belong to the family Chordariaceae. Marine brown algae belonging to families Nemacystaceae and Chordariaceae are extensively consumed in Japan. These have caused poisoning in people occasionally. One such poisoning was named as mozuku poisoning. There have been two mozuku poisonings. In the first incidence, Nemacystus decipiens commonly known as mozuku in Japanese, occurred in Ogashi, Akita in the year 1967. The second incident was caused by C. okamuranus commonly known as Okinawa-mozuku. This had taken place on the Yoron Island in the year 1974. Toxicity studies were performed in mice with water and fat-soluble extracts and were administered intraperitoneally and orally. The fat-soluble extracts killed the mice overnight. The symptoms of lacrimation, paralysis of hind legs, fatigue, piloerection, and difficulty in respiration, convulsions and death were recorded. Structural

elucidation of the chemical compound characterized it as a diethyl peroxide (9) namely α , α '-dihydroxy diethyl peroxide. The poisoning was also confirmed by a synthetic peroxide, which demonstrated the same toxicity when administered orally and intraperitoneally. Similar peroxide compounds are also found in brown alga Analipus japonicus and in the red alga Gracilariopsis chorda (Fusetani and Hashimoto 1981; Fusetani and Hashimoto 1984).

Fucoidan (10) (Figure 2) is an important polysaccharide found in C. okamuranus, being consumed since early times in countries like China, Korea and Japan (Kusaykin et al. 2008). They belong to the class of L-fucose enriched sulfated polysaccharides, mainly to be seen in the extracellular matrix of brown algae (phaeophyceae) (Cumashi et al. 2007; Li et al. 2008; Lim et al. 2019; Tako, Yoza, and Tohma 2000). It generally contains high amount of L-fucose and sulfate, along with few other sugars such as xylulose, glucuronic acid, galactose and mannose. Fucoidan (10) from Cladosiphon okamuranus contained fucose, glucuronic acid and sulfate in a molar ratio of about 6.1:1.0:2.9 (Nagaoka et al. 1999). Fucoidan (5) extracted from C. okamuranus contained 30.9% w/w of fucose, 0.7% w/w xylulose, 2.2% w/ w glucose, 23.4% w/w of uronic acid and 15.1% w/w of sulfonic acid with no mannose (Cumashi et al. 2007). It's antiulcer activity was investigated in gastric mucosa, which exhibited anti-peptic activity of fucoidan (10), dextran sulfate, carrageenan without any inflammatory activity Fucoidan (10) also prevents H. pylori infection thus reducing the risk of gastric cancer associated with it (Shibata et al. 2000). It also improved murine chronic colitis by down-regulating interleukin-6 production from colonic epithelial cells suggesting its usefulness in inflammatory bowel disease and thus suggested to be taken as dietary substance in humans. Fucoidan was accumulated in the sinusoidal non-parenchymal and mononuclear cells present in the jejunal lamina propria in female Balb/c mice and also blocked the adhesion of Helicobacter pylori to human gastric cell line (Matsumoto et al. 2004). They possess antithrombic, antiproliferative, antitumoral and anticoagulant activities. It is also found that fucoidan (5) from Cladosiphon seaweed could alter the ratio of CD_4^+/CD_8^+ cells, thus increasing the number of cytotoxic T cells. It is also reported to be an effective cardio protective agent by Thomes et al. (2010); it could protect the cardiac cells from isoproterenol induced myocardial infarction which could be attributed to detoxification of isoproterenol by antioxidant Defense system and alteration in the HDL and LDL values. Fucoidan (10) cannot be degraded by human digestive enzymes and is utilized in very low to negligible quantities by intestinal bacteria. Toxicity study of fucoidan (10) in wistar rats did not show any toxicity at low dose, but it significantly prolonged the clotting time at higher dose (Gideon and Rengasamy 2008) due to its binding to various adhesion proteins, growth factors, cytokines and enzymes such as coagulation proteases (Thomes et al. 2010); incidences of diarrhea occurred with a dose of about 6 g/day up to 13 months (Gideon and Rengasamy 2008; Fitton 2011). They are also known for their potent anti-inflammatory activities (Cumashi et al.

2007). It was successful in reducing total and low-lipoprotein cholesterol (Abe et al. 2013). Also fucoidan (10) obtained from C. okamuranus showed anti H. pylori, antiulcer and anti-functional dyspepsia effects (Bellmann et al. 2014). These benefits can have a promising future which needs to be explored.

Fucoidan is also proven to have an antagonistic effect to most of the pathogens, and to have antitumor activity (Kusaykin et al. 2008). Fucoidan (10) obtained from C. okamuranus causes enlargement of natural killer cells (Azuma et al. 2012). Fucoidan (10) from C. okamuranus exhibited a low-toxic antiviral compound to be used in the poultry industry (Elizondo-Gonzalez et al. 2012). Another study also evaluated its anti-viral activity against Newcastle disease virus (NDV) which is dreadful to the poultry (Trejo-Avila et al. 2016). Results showed that fucoidan (10) is also active against this virus when applied in ovo, while it was nontoxic at therapeutic doses (Aguilar-Briseño et al. 2015). Thus, it can be a prospective lead to serve as an anti-viral and antitumor drug, for which further studies are required.

C. okamuranus contains another compound structurally similar to toxin A, obtained from Sphaerotrichia divertica. Toxin A was found to be lethal to mice at a dose of around 250 μg/g (Anthoni et al. 1987). The fat soluble fractions from S. divaricata and C. okamuranus have been found toxic in the study carried out by Fusetani and Hashimoto (1981). The toxins were generated only when the algal body was heated with surrounding water.

Laminaria genus

Laminaria is a kelp that belongs to family Phaeophyceae. They are widely distributed in the North Atlantic Ocean, Pacific Ocean and in the warm waters of Mediterranean Sea. It is an economically important seaweed cultured in China and distributed widely in Korea and Japan (Li, Zhang and Song 2005). Laminaria consists of 50.7% dietary fibers, reported as the maximum content among all other seaweeds as well as plants. It contains 32.8% water-soluble and about 17.9% water insoluble dietary fibers. The water-soluble dietary fibers are bioactive to a larger extent in humans. It contains a high molecular weight polysaccharide namely algin, and good amount of iodine, potassium, magnesium, calcium and iron (Kim and Bhatnagar 2011). Apart from these, they are also reported to have cadmium and copper. These metal ions are considered to be highly toxic as they may cause damage to the liver, nerves and bones since they can interfere with membrane ion transport. Functional groups of various vital enzymes can be blocked by this seaweed (Ye, Wang, and Tseng 2005). However, it can be used for the treatment for goiter since it contains highest amount of iodine (Mason 1959). L. digitata arsenic is found in the form of arsenosugars is natural ingredient of L. japonica (Kuehnelt, Irgolic, and Goessler 2001; Hwang et al. 2016). L. japonica is reported to have high amounts of total arsenic, inorganic arsenic, total and chemical forms of cadmium (Zhao et al. 2012). Dibutyl phthalate (DBP) and di-(2-ethylhexyl) phthalate (DEHP) also occurs naturally in L. japonica (Namikoshi

et al. 2006). L. japonica, L. saccharina, and L. cloustoni are approved as a food additive as a source of iodine for direct human consumption. L. hyperborean is commonly used for thyroid regulation in herbal medicine (PDR for herbal medicines 2000). L. japonica contains 54% bio-accessible iodine (Alves et al. 2018). It has been a part of traditional Chinese medicine since a long time now. Studies show that the polysaccharides that are isolated from this seaweed has endogenous enzymes with antioxidant properties (Peng et al. 2012). It is also reported to have anticoagulant and antioxidant properties (Peng et al. 2013). Aqueous extract of L. japonica exhibited the hypotensive activity in rats with normal blood pressure. In the studies performed for their effects on the cardiac functions, the extracts from L. japonica decreased atrial beats in a dose dependent manner (Chiu and Fung 1997).

Laminaria used for cervical dilation is also predicted to cause toxic shock syndrome. Women experience dyspnea, tachycardia, and hypotension after application, and the sample showed presence of heavy growth of Staphylococcus aureus and expression of Staphylococcal enterotoxin C when amniotic membrane culture was tested (Sutkin et al. 2001). L. japonica also contains fucoidan (10) which demonstrated protective effect against chronic renal failure. Fucoidan increased the clotting time significantly in wistar rats during the toxicologic studies. The increase in clotting time was dose dependent (Li, Zhang and Song 2005). It was shown that at a dose of more than 300 mg/kg of body wt./day of fucoidan from L. japonica significantly increased clotting time and reduced aspartate transaminase (AST); significant reduction in glucose levels was also observed at the dose of 2,500 mg/kg of body wt./day (Gideon and Rengasamy 2008). Fucoidan (10) obtained from L. japonica has the potential to stop free radical chain reactions and act as a potent antioxidant (Zhao et al. 2004). L. japonica extract (up to 400 mg/ kg) caused decrease in body weight, fat-pad weights, and serum and hepatic lipid levels in rats, when administered by gavage for 6 weeks (Jang and Choung 2013). In patch test on human skin, cream containing a 50/50 aqueous/propylene glycol extract of L. japonica (10%; 20 mg) it exhibited nonirritating effect (Choi et al. 2013). A low-molecularweight fucoidan of approximately 7.6 kDa obtained from L. japonica had better absorption profile and antithrombotic activity in male wistar rats when compared to mediummolecular-weight fucoidan of approximately 35 kDa. Oral administration of the low-molecular-weight fucoidan at 400-800 mg/kg prolonged the activated partial thromboplastin time (APTT) and thrombin time (TT) in male wistar rats (Sanjeewa and Jeon 2018). Cao et al. (2016) stated that polysaccharides obtained from L. japonica can be used as a new therapy against respiratory syncytial virus (RSV) which belongs to the family Paramyxoviridae and responsible for respiratory viral infection.

Table 1 is an extensive compiled list of edible seaweeds belonging to different class and their nutritional composition with reference to the daily average requirements recommended by USFDA guidelines for adults and children above 4 years of age (CFR-USFDA 2018).

Impact of toxic seaweeds, microalgae and algal blooms in the food chain

Seaweeds are highly nutritious and have multiple benefits like it helps thyroid function, diabetes, gut health, weight loss and cardio protective action. Along with high nutrition value and therapeutic properties, they may suffer high risks due to the presence of excess iodine and heavy metals. Hizikia fusiforme has arsenic concentrations > 50 mg/kg (Ichikawa et al. 2006). Edible seaweeds if boiled, their arsenic content reduces by 43-50% (Laparra et al. 2003; 2004) and by soaking up to 60% reduction in arsenic content occurs (Hanaoka et al. 2001). In a study in Korea, 426 seaweed samples were tested for arsenic, lead, mercury, and cadmium levels. Arsenic was highest at 17.4 mg/kg dry weight, and assuming 8.5 g per day seaweed consumption, the intake was falling in the WHO recommended weekly limit (0.2-6.7%). The study confirmed minimum chances of health risks from the metals consumed via seaweed (Hwang et al. 2010).

In marine ecosystem, food webs constitute of food chain, which connect at different trophic levels. They support long, intricate, complicated, as well as short food chains. All the food webs have the same basic trophic levels throughout the world, with only variation in numbers and types of species that constitutes each level with the ecosystem. The consumers of seaweeds can be herbivores, omnivores or carnivores. People are mostly exposed to algal toxins through contaminated seafood products consumption. They are amnesic shellfish, ciguatera fish, diarrhetic shellfish, neurotoxic shellfish and paralytic shellfish poisoning. The most significant problem to human health have been caused by harmful algae, diatoms, dinoflagellates poisoning. Amnesic Shellfish Poisoning (ASP) is caused by diatom Pseudo-nitzschia sp. due to produced toxin named domoic acid. Fish and crab viscera can also contain domoic acid, causing high risk to human consumers and animals in food chain (Subba Rao, Quilliam, and Pocklington 1988; Lawrence et al. 1989; Novelli et al. 1992). Ciguatera fish poisoning (CFP) is caused by ciguatoxin/maitotoxin produced by Gambierdiscus toxicus, Prorocentrum spp., Ostreopsis sp., Coolia monotis, Thecadinium sp. and Amphidinium carterae. Diarrhetic shellfish poisoning (DSP) is caused by okadaic acid produced by Dinophysis sp. Karenia brevis, a marine dinoflagellate causes neurotoxic shellfish poisoning (NSP) by produced brevetoxins. Saxitoxins produced by Alexandrium sp., Gymnodinium catenatum, Pyrodinium bahamense causes paralytic shellfish poisoning (PSP). It is prevented by active monitoring programs to check toxin level in mussels, oysters, scallops, clams etc. (Baden, Fleming, and Bean 1995). Many fatal outbreaks have taken place due to shellfish poisoning leading to the death of birds, mammals from the marine environment. In 1998, sea lions suffered from an unknown disease with seizures symptoms at the California coast. While earlier it was thought of mercury poisoning, later it was co-related to a similar incidence with a few people in Canada; wherein they were diagnosed by amnesic shellfish poisoning caused by the consumption of blue mussels infected by domoic acid, which is generally produced by

Table 1. Edible seaweeds and their nutritional content with specific reference to the daily intake of minerals per day as per FAO guidelines.

USFDA avg. daily limits in adults and children above 4 years	Carbohydrate 1275 g	Dietary fiber 128 g	Lipid 178 g	Protein 150 g	Na 2300 mg	K 4700 mg	P 1250 mg	Ca 1300 mg	Mg 420 mg	Fe 18 mg	l 150 μg	Ref. (CFR-USFDA 2018)
Chlorophyta Caulerpa lentillifera	38–59	33	0.86–1.11	10–13	8917	700-1142	1030	780–1874	630–1650	9.3–21.4	I	(Matanjun et al. 2009; Pattam and
Caulerpa racemosa	33-41	64.9	8.6	17.8–18.4	2574	318	29.71	1852	384–1610	30–81	1	Chirapart 2008; Yuan 2008) (El-Sarraf and El-Shaarawy 1994; Akhtar and Sultana 2002; Santoso, Yoshie-Stark, and Suzuki 2006;
Codium fragile	39–67	5.1	0.5–1.5	08–11								Kumar, Gupta, et al. 2010) (Ortiz et al. 2009; Guerra-Rivas
Ulva compressa	48.2	29–45	0.3-4.2	21–32							ı	(Burtin 2003; Mamatha et al. 2007;
Ulva lactuca	36–43	29–55	0.6–1.6	01–25		I	140	840	ı	99	I	(Fleurence et al. 2010), Idail 2000 (Fleurence et al. 1995; Manivannan et al. 2008; Morrissey, Kraan, and
Ulva pertusa	47	1	I	20–26								Guiry 2001; Yuan 2008) (Fujiwara-Arasaki, Mino and Kuroda
Ulva rigida	43–56	38–41	0.9–2.0	18–19	1595	1561	210	524	2094	283		(García, Castroviejo, and Neira 1993; Kumar, Gupta, et al. 2010;
												Santoso, Youshie-Stark and Suzuki 2006; Taboada, Millán and Míguez 2010)
Ulva reticulata	50–58	65.7	1.7–2.3	17–20							I	(Kumar, Gupta, et al. 2010; Shanmugam and Palpandi 2008)
Phaeophyceae Alaria esculenta	46–51	42.86	1 to 2	9–20								(Abbott 1988; Morrissey, Kraan, and
Durvillaea antarctica	70.9 ± 2.7	71.4 ± 0.5	08 ± 0.2	10.4 ± 0.3	52.9 ± 1.0	15.6±2.7	I	I	I	13.7 ± 1.8	29.1 ± 2.7	(Ortiz et al. 2006; Smith, Summers,
Eisenia bicyclis Fucus spiralis	60.6	10–75	0.1	7.5	(6y /6)	(6y/6)				(gy kg)	(ga/giii)	and Worlg 2010) (Misurcová et al. 2010; Yuan 2008) (Patarra et al. 2010)
Fucus vesiculosus	46.8	45-59	1.9	3–14	2450–5469	2500–4322	315	725–938	670–994	4–11	14.5	(Applegate and Gray 1995; Díaz- Rubio, Pérez-Jiménez, and Saura-
												Calixto, 2009; Fleurence et al. 1995; Rupérez 2002; Saá 2002; Truns Vahor and Taure 2001)
Himanthalia elongata	44–61	33–37	0.5–1.1	5–15	4100	8250	240	720	435	59	14.7	(Brutin 2001) Jiménez-Escrig, and Rupérez 2010; López-López et al. 2009; Plaza, Cifuentes, and Bañez 2008: Saá
Laminaria digitata	48	36–37	-	8–15	3818	11,5 79	1	1005	629	3.29	1	2002; Yuan 2008) (Brutin 2003; Fleurence et al. 1995; Morrissey, Kraan, and Guiry 2001;
Laminaria ochroleuca Nereocystis luetkeana	38.82	1.30	0.92	7.49 15.28	5.08 ± 0.025	1.30 ± 0.099	ı	51	ı	43.3	I	nupelez 2002; Tudil 2008) (Yuan 2008) (Barta, Branen, and Leung 1981)
Saccharina japonica	51.9	10–41	1.0–1.9	7–8	(% dry wt.) 2532–3260	(% ary wt.) 4350–5951	150–300	(% dry wt.) 225–910	550-757	(% ary wt.) 1.19–43	130–690	(Funaki et al. 2001; Mabeau and Fleurence 1993; Mišurcová et al. 2010; Yuan 2008)
												(continued)

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USFDA avg. daily limits in adults and children	Carbohydrate	Dietary fiber	- - -	Protein	ν. V	×	۵	٣	Ş	ā	-	Rof
above 4 years	1275 g	128 g	178 g	150 g	2300 mg	4700 mg	1250 mg	1300 mg	420 mg	18 mg	150 µg	(CFR-USFDA 2018)
Saccharina latissima	52–61	30	0.5–1.1	6- 26	2620	4330	165	810	715	1	15.9	(Gómez-Ordóñez, Jiménez-Escrig and
j	Ċ	1	7	-				0	100	0		Ruperez 2010; Morrissey, Kraan, and Guiry 2001; Saá 2002)
sargassum tusirorme	30.6	1/-69	4:	9.[ı	I	I	1860	/89	988.0	I	(Funaki et al. 2001; Mabeau and Fleurence 1993; Mišurcová et al.
Care of the care o	00 13	67.7	0.45	16 76								2010; Yuan 2008; Sugawa- Katayama and Katayama 2009)
Surgussum vuigure Undaria pinnatifida	07.00 45–51	7.7. 16–51	1.05–4.5	12-23	1600-7000	5500-6810	235–450	680–1380	405–680	1.54–30	10–100	(Mailling-Solially et al. 2003) (Brutin 2003; Holdt and Kraan 2011;
												Kolb et al. 2004; López-López et al. 2009; Mišurcová et al. 2010; Rupérez 2002; Saá 2002; Vian 2008)
Rhodophyta	;	;	•	;			,			;		(dall 2000)
Chondrus crispus	25-68	10–34	1.0–3.0	11–21	1200-42/0	1350-3184	135	420-1120	600-/32	4-1/	17.3	(Holdt and Kraan 2011; Morrissey, Kraan, and Guiry 2001; Saá 2002;
:				;	;	!				,		Rupérez 2002; Yuan 2008)
Gracilaria changii		24.7	3.3	6.9	5465	3417	ı	402	265	3.6–5	I	(Krishnaiah et al. 2008; Yuan 2008)
Gracilaria chilensis	66.1	ı	1.3	13.7	5465	3417	1	402	265	3.6–5	I	(Krishnaiah et al. 2008; Ortiz
Palmaria palmata	46–56	29–46	0.7–3	8–35	1600-2500	7000-9000	235	560-1200	170-610	20	10-100	(Holdt and Kraan 2011; Morrissey,
												Kraan, and Guiry 2001; Saá 2002; Rupérez 2002; Yuan 2008)
Porphyra tenera	44.3	12–35	0.7-1.3	28-47	3627	3500	ı	390	565	10–11	1.7	(Brutin 2003; Fleurence et al. 1995;
												Mišurcová et al. 2010; Rupérez
Porphyra umbilicalis	43	29–35	0.3	29–39	940	2030	235	330	370	23	17.3	(I ópez-l ópez et al. 2009: Saá 2002)
Porphyra yezoensis	44.4	30–59	2.1	31–44	570	2400	ı	440	650	13	!	(Indergaard and Minsaas 1991; Noda
•												1993; Yuan 2008)

The composition of carbohydrate, dietary fibers, lipids and proteins are in % dry weight; minerals sodium (Na), potassium (K), phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe) are in mg and lodine (I) is in µg.



a certain type of algae belonging to Pseudo-nitzschia genus, and is essentially a neurotoxin. Further investigations also reported domoic acid being found in the gastrointestinal contents and body fluids of thirteen different mammal species from the Alaskan waters (Husson et al. 2016).

Over the last two decades, due to the increase in pollution and eutrophication there has been a growing concern about the impact of microalgal and cyanobacterial blooms. Harmful effects, including the development of high biomass and scums is decreasing the water quality and adversely affecting the aquatic ecosystems, simultaneously affecting the aquaculture industry and human health (Zerrifi et al. 2018). Hundreds of fish and shellfish are often found dead and following many people getting ill after consuming certain shellfish, only to later discover deadly toxins in the sick or dead species. Tiny phytoplankton cells produce potent toxins, which is eaten by zooplankton. Zooplanktons are microscopic animals, which remain unaffected by these toxins. Zooplankton and phytoplankton are eaten by many animals from tiny shrimp, krill, mussels and fish to the largest of mammals. The minuscule toxin produced by phytoplankton cell gets bio-accumulated through the food web as each organism consumes more and more of the toxin, ending in a bigger dose of toxin in higher mammals causing their death. It may be also transported to humans through the food web, mostly through contaminated shellfish. These toxins affect the nerve and muscle cells, causes respiratory irritation, distress, gastrointestinal disturbances, paralysis and death (Pokapū Akoranga Pūtaiao 2012)

Spirulina is used as a vegan source of vitamin B12 and protein, and is also known to improve lipid and glucose metabolism and reduce liver fat and exhibit cardio protective property. It has few active compounds such as phycocyanobilin, which mimics the body's bilirubin and inhibit the NADPH oxidase enzyme (McCarty 2007). It also provides potent anti-oxidative and anti-inflammatory activity. A daily dose of about 1-8 g daily is effective for cholesterol, 2-7.5 g daily is found to be impactful in muscle performance. Mild effects are observed with a daily consumption of 2 g per day for blood glucose control. Fatty liver results are effective at dose of about 4.5 g daily and blood pressure at a daily dose of 3.5-4.5 g daily. Polyphenols rich seaweed has an important additional role in cardiovascular-associated disorders (Gómez-Guzmán et al. 2018). Seaweeds act as preventive agents for cardiovascular diseases (Cardoso et al. 2015). Though consumption of these seaweeds seem to be very nutritious and advantageous but unchecked consumption of these may cause severe issues and autoimmune diseases like systemic lupus erythematosus (SLE), rheumatoid arthritis, pemphigus vulgaris etc. Spirulina is a widely consumed cyanobacteria biomass which is often contaminated with blue-green algae, such as

M. aeruginosa, that produces toxic microcystins. Spirulina plantensis contains good amount of phycocyanin and carotenoid, which are good antioxidant compounds (Liu et al. 2016). Its antiatherogenic properties were observed due to reduction of total cholesterol in rat, hamster and rabbit. It

depicts antiatherosclerotic properties among people suffering from atherogenic dyslipidemia and ischemic heart disease. It is also found to have an apoptosis preventing property in cardiomyocytes and plays a role in preventing oxidative stress. The chronic intake of Se-rich Spirulina phycocyanin averts the development of atherosclerosis, mainly due to reduction of pro-oxidant factors and by improving the serum lipid profile (Riss et al. 2007). One study reported an allergic reaction toward c-phycocyanin component present in spirulina (Petrus et al. 2009). It is also reported in case of allergic reaction within six hours of consuming 2.5 g of seaweed in a 14 years old volunteer. The Dietary Supplements Information Expert Committee (DSI-EC) of United States Pharmacopeial Convention (USP) reviewed 31 reported adverse events of spirulina to evaluate related health concerns. Spirulina maxima and S. platensis were admitted in USP-NF and class A safety grade dietary supplements (Marles et al. 2011).

Microcystins are cyclic heptapeptides produced by cyanobacteria during algal blooms; they are hepatotoxic and carcinogenic due to their property of inhibiting protein phosphatases and possible contamination with heavy metals such as cadmium, lead, mercury and arsenic (Marles et al. 2011; Marsan et al. 2018). The safe consumption of microcystin is set at 0.04 µg/kg/day (Dietrich and Hoeger 2005). An early analysis of fifteen commercial Spirulina supplements reported an average microcystin concentration of $0.15 \,\mu\text{g/g}$ (range: $0.06-0.32 \,\mu\text{g/g}$) and $0.52 \,\mu\text{g/g}$ (range: 0.0-2.12 µg/g) in the year 1998 and 1999 respectively. This suggests that an 80 kg adult can safely consume up to 6-21 g of Spirulina (3.2 μg of microcystin) based on the reported average contamination levels (Gilroy et al. 2000). Though, it is suggested that Spirulina does not show a health risk, however it may due to contaminants from other species such as toxic microcystins produced by other bacteria growing in close vicinity of Spirulina (Marles et al. 2011).

Recently, researchers explored the functional characterization of marine macroalgae, in order to understand its genome protecting potential. The antigenotoxic potential of Ulva rigida, Fucus vesiculosus, and Gracilaria sp. was assessed in Drosophila melanogaster following macroalgae diet exposure and somatic mutation and recombination test approach (SMART). All macroalgae species exhibited genoprotection activity, when challenged against streptonigrin as an exogenous material. Gracilaria species provided indecisive indications, as 10% of G. vermiculophylla inhibited the egg and larvae development and 10% G. gracilis increased genotoxicity. U. rigida demonstrated higher genoprotection in an aquaculture-controlled system. F. vesiculosus did not produce significant changes in the growing condition. These findings can help the role of seaweed as functional food and their advantages (Marques et al. 2018). Another study by Desideri et al. (2018) investigated the bio accessibility of toxic elements, including Al, As, Ni, Cd, and Pb in five commercially available and commonly consumed algae in Italy. The bio accessibility of the toxic elements showed some interesting results with Cd having



high bio accessibility; while As and Ni exhibited up to 40-55% bio accessibility, Al and Pb exhibited poor bio accessibility (5-15%).

Even though these seaweeds act as a source of essential antioxidants, minerals and vitamins; they may lead to toxic accumulation to a higher extent due to their high absorption capacity. An overview of toxicity of edible seaweeds is presented in Table 2. These organisms are even capable of absorbing and accumulating metals without suffering any damage (Paz et al. 2019). This aspect requires continuous monitoring of edible seaweeds as a functional food and its heavy metals limit.

Side effects and adverse interactions

Seaweeds are widely consumed and is one of the most nutritious food in nature, due to their rich nutrient content of iodine, calcium, iron, vitamins B6, and B12. The most widely consumed seaweed is Kelp viz. kombu, arame, and wakame belonging to the family Phaeophyceae. Kombu is a highly nutritious seaweed containing calcium, carotene, phytohormones, mannitol, and vitamins. Arame is an excellent source of iron and iodine. Wakame is the richest source of calcium. Nori has highest consumption and is a good source of protein. Dulse another seaweed, commonly known as sea lettuce is known for its nutritional value. It has high content of vitamins B12, B6, A, C, E, calcium, protein, iodine and magnesium. Hijiki is another widely consumed seaweed, and is rich source of algin, calcium and dietary fibers (Murray, Pizzorno, and Pizzorno 2005).

Nori is rich in various nutrients and minerals but also impose hazardous risk like congestive heart failure, cirrhosis and chronic kidney disease. These effects are seen due to the presence of large amount of sodium. It may also reduce blood pressure thus is also contraindicated for patients undergoing treatment for hypertension this effect is attributed to the presence of docosahexaenoic acid (DHA). They also impose risks such as contact dermatitis, goiter, cholera and inflammation in large consumption of these seaweeds (Seaweed 2011).

Kelp which is previously mentioned has been reported to have various interactions and side effects. Side effects such as allergic reactions, itching, rashes, trouble in breathing, tightness in the chest, arrhythmia, nervousness, trouble in sleeping, unusual bleeding and cramps are observed. They have a tendency to interact with drugs like digoxin, potassium supplements, spironolactone, amiloride, thyroid supplements, thus patients on these medications should be careful and avoid Kelp (Kobriger 2011).

Laminaria a widely consumed seaweed for its high iodine and iron content, and benefits in weight loss, blood pressure maintenance, heartburn and cancer prevention, may also cause issue in the thyroid functioning, since an ideal intake of 150 µg of iodine is required for the normal functioning of thyroid, but Laminaria can provide iodine content as high as 1000 µg which eventually can lead to deranged thyroid levels (Zava and Zava 2011). They also contain significant levels of arsenic, which is poisonous and rendered unsafe; if

administered directly to the cervix during childbirth or pregnancy, it may lead to rupture of cervix and infant death. High potassium levels in Laminaria may also lead to kidney damage and thus is contraindicated with anti-hypertensive agents. Laminaria contains large amount of potassium, which can increase the effects and side effects of digoxin (Atlas et al. 1998).

A study was performed in the UK, to identify all the seaweed containing products sold in the grocery market. Out of 224 products, only 22 products stated iodine content information on their labels; 40 products provided information which was sufficient to estimate the iodine content. There was a lack of information about the seaweed, its type, source and iodine content in the seaweed product, which makes the safety of these products questionable especially in people sensitive for high iodine exposure i.e. pregnant women (Bouga and Combet 2015). Thus it's very important to have the information of seaweed which is being consumed by people to predict its safety in individuals on respective diet and conditions.

Seaweeds may also decrease the pharmacological effect of anticoagulants (aspirin, warfarin) (Holbrook et al. 2005). A patient on high-iodine diet will require higher doses of antithyroid drugs. Antithyroid drugs (propylthiouracil or thyroxine) prevent iodine absorption in the stomach and with increase in the dose of antithyroid drugs, the chances of side effects such as rashes, hives, and liver disease increases (Bareuther 2008). Fucus vesiculosus is a slimming supplement which may compromise on the therapeutic efficacy of the amiodarone drug (Rodrigues et al. 2013). Major adverse reactions and drug interactions of seaweeds are compiled in Table 3. The herb-drug interaction between the consumable seaweeds and prescription or over the counter drugs should be studied further to accept its complete safety in patients. The amiodarone bioavailability decreased in rats, therefore the therapeutic efficacy of drugs might be being compromised by the concurrent administration of seaweed in diet or supplements. These aspects should be mostly looked into where the content and pharmaceutical composition are opposite in nature.

A toxicological study conducted by Banu and Umamageswari (2011) suggested that seaweeds are safe for human consumption and a daily consumption in the range of 5-10 g is considered safe.

Future prospects and conclusion

Seaweeds are a boon to the mankind if consumed appropriately, otherwise they may impose a major risk to humans. They are reported of many bioactive compounds which aren't found in other natural food source, and are beneficial for the human well-being by providing better health and preventing various diseases. There is need of an awareness for consuming seaweeds among people from countries where it still remains away from daily food list. As of now seaweeds are utilized mostly to extract the phycocolloid. It has great potential as a dietary source with almost all the nutrients and as reported earlier they can be looked upon as

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Name	Chemical compounds/extracts	Toxicity reported	Dose	Study data	Reference
Gracilaria sp. a) G. verrucosa	Prostaglandin E ₂ (PGE ₂) Prostaglandin A ₂ (PGA ₂)	Hypotension eventually leading to death	0.5 mg was found to be beneficial to control gastrointestinal disorders.	Toxic levels produced from arachidonic acid, in the combination of the seaweed and the raw fish consumption. Henorrhaged blood reported from	(Noguchi et al. 1994; Cheney 2016; Se-Kwon, 2017)
		Excessive phagocytosis	2 g/kg as supplement found to have a positive immune response 4.0 mg/animal (mice) showed immunopotentiating activity stimulating phagocytosis when administered orally and by ip route.	one stornach: positive effect with higher immune response in shrimps	(Jasmanindar et al. 2018; De Almeida et al. 2011)
b) G. cornopifolia	Aplysiatoxin Debromoaplysia-toxin	Gastrointestinal disturbances		Toxic effect studied in human adult by oral route	(Nagai, Yasumoto, and Hokama 1996)
c) G. edulis	Polycarvenoside A Polycarvenoside B	Gastrointestinal toxicity	Toxicity studied in mouse and LD-50 was reported as 0.825 mg/kg-ip	Semi purified toxic fraction studied on mouse bioassay contained	(Yotsu-Yamashita et al. 2004; De Almeida et al., 2011)
d) G. chorda	Prostaglandin E ₂ (PGE ₂) Prostaglandin A ₂ (PGA ₂)	Gastrointestinal toxicity	Toxic at concentrations of about 1.2 mg/rat administered ip; 0.5 mg was found beneficial to control	polycal verioside 'A	(Fusetani and Hashimoto 1984; Cheney 2016; Se-Kwon, 2017)
e) G. corticata	50% EtOH-H ₂ O extract	Autonomic effects in dogs, CNS effects, analgesic activity in mouse	gastrointestinal disorders 500 or 1000 mg/kg/day to female rats produced significant post-coital contraceptive activity; toxicity was assessed in mouse and LD50 reported at 1000 mg/kg-ip		(Kamat et al. 1992; De Almeida et al. 2011)
f) G.verrucosa		Mouse- 0.5 mg/	Toxicity assessment-mouse-1.2 mg/		(Fusetani and Hashimoto 1984)
g) G. chorda 2.Cladosiphon okamuranus	H_2O extract Diethyl peroxide $(\alpha_i\alpha'$ -Dihydroxy diethyl peroxide)	gasure mudaaton 1.2 mg/animal i.p. Psychotic disturbances, Paralysis	allillari.p. was positive	Toxicity was tested in mice, fat soluble fraction was given after emulsified with 1% Tween	(Fusetani and Hashimoto 1984) (Fusetani and Hashimoto 1981)
	Fucoidan	Significant increase in clotting	Increased clotting time $>1,200\mathrm{mg/}$ kg of body weight in wistar rats	ou saime solution	(Gideon and Rengasamy 2008)
3. Acanthophora specifera	Polycarvenoside A	Neurological disorders	A. spicifera 100 and 200 mg /kg body weight suggested the anti-tumour properties; at doses above 2000 mg/kg body weight reported mortality and clinical signs	No sufficient amount in initial studies by authors later claimed to be polycarvenoside A	(Yotsu-Yamashita et al. 2004; Lavakumar, Ahamed and Ravichandiran 2012)

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4 Complement		Gastrointestinal disorders		Semi purified toxic fraction studied on mouse bioassay contained polycarvenoside A	(Preskitt 2001)
4. Carierpu sp. a) C. taxifolia	Caulerpenyne	Risks of cutaneous and food intoxication was estimated minimal to humans		Skin cells, melanocytes, keratinocytes, immortalized keratinocytes, and bone marrow cells	(Massin 1996; Agardh 1822)
b) C. <i>prolifera</i>	Caulerpenyne	Neurotoxic activity		Nuclear fragmentation of appointed cells	(Kurt et al. 2009)
c) C. racemosa	Caulerpin	Cellular and Physiological Responses in fish		Inhibition of acetylcholinesterase and acylCoA oxidase	(Felline et al. 2012)
5. Laminaria japonica	Fucoidan	Chronotropic effect	Significant increase in clotting time at 900 and 2500 mg/kg body weight per day.	Protective effect against Diabetic cardiomyopathy	(Yu et al. 2014; Citkowska, Szekalska, and Winnicka 2019)
	Heavy metals	Hypotension		Damage to the liver, nerves	(Kim and Bhatnagar 2011)

Dose of G. verucosa is in mg and in g/kg for excessive phagocytosis, whereas for G. eduli, G. Chorda, A. specifera and L. japonica are in my/kg body weight

treatments for various acute and chronic ailments and diseases. They contain maximum amount of nutrients and vitamins compared to any other food product. Antioxidant compounds like sulfated polysaccharides from them can help in treating diseases such as AIDs and cancer.

There is a need of guidelines over the limit and quality of consumption of the seaweeds by the regulatory bodies including specific drug seaweed interactions. This is mostly understood in the respect of high level of iodine in seaweeds, for e.g. Laminaria spp. (kombu) are brown seaweeds especially high in iodine, thus daily intake of 3-6 g of dried seaweeds is sufficient for adequate dietary iodine. Individuals who are extremely sensitive to iodine, should be cautious about the amount of seaweed in their diet to avoid symptoms of hyperthyroidism such as nervousness, insomnia, increased heart rate etc. Too much of iodine consumption can develop goiter, by increasing thyroid-stimulating hormone and skin problems. High sodium content can lead to high blood pressure and heart disease. Seaweed supplementation causes gastrointestinal disturbances with abdominal pain which decreases over a period of time due to getting used to the supplementation. Any step toward regulation of heavy metal and arsenic contamination in seaweeds from water, and public awareness over the food security from consumption of seaweeds will be a good move to enjoy highly beneficial sea plants as vegetables.

There are other equally important aspects in applications such as soil fertilizers, animal feed, fish feed, cosmetics, in integrated aquaculture, and in wastewater treatment. Integrated aquaculture is developing fast as one of the solutions to environmental sustainability problems and for the management of disposal of effluents in coastal areas from large-scale aquaculture activities. They can also be used as efficient biofuels. Their abundance in marine environment make it an easy alternative source for biofuels and thus reducing the pollution as well. Their anti-clotting properties should be widely explored and used for the treatment of cardiac diseases. Their ability to increase the T-cell count can also be employed for the treatment of cancer. Studies should be performed in the area obesity, diabetes and thyroid related disorders that have affected a large population to make use of these seaweeds effectively. These properties and its constituents can prove to have a promising future but might be affected by ocean acidification and increased temperatures due to climate change. As seaweeds remain available, edible and sustainable source to mankind, a careful regime of consumption can prevent the toxic effects and can allow people to enjoy its beneficial properties, thus its efficacious use should be promoted. While buying seaweed based packed products, people should look for the relevant information such as the source of seaweed, water quality and processing etc. which can be helpful to assess the safety of the product, and will also help to understand the potential toxic compounds. These steps will strengthen the seaweed market and its commercialization and will also attain consumer's satisfaction.



Table 3. Side effects and drug interactions of seaweeds

Seaweeds	Side effects	Contraindications	Drug interactions	Reference
Porphyra sp.	Congestive heart failure, cirrhosis, chronic kidney disease, contact dermatitis, goiter, cholera and inflammation	Patients undergoing treatment for hypertension	No interactions are reported. Peptides from <i>Porphyra</i> show antihypertensive effects	(Lee 2019)
Kelp	Allergic reactions arrhythmia, nervousness, unusual bleeding and cramps	Patients on thyroid medication	It could potentially interact with: digoxin, potassium supplements, spironolactone, amiloride, thyroid medications blood pressure medication	(Kelp. Accessed November 24, 2018 from http:www. naturalstandard.com)
Laminaria	Hyper-thyroidism	Childbirth or pregnancy and to patients undergoing for hypertension	Digoxin	(Accessed September 28, 2019 from https://www.rxlist.com/laminaria/ supplements.htm)
Spirulina Blue green algae	might decrease the effectiveness of immunosuppressants	Patients taking immunosuppressants	Azathioprine (Imuran), basiliximab (Simulect), cyclosporine (Neoral, Sandimmune), daclizumab (Zenapax), muromonab-CD3 (OKT3, Orthoclone OKT3), mycophenolate (CellCept), tacrolimus (FK506, Prograf), sirolimus (Rapamune), prednisone (Deltasone, Orasone), corticosteroids (glucocorticoids), and others.	(Accessed September 27, 2019 from https://www.webmd.com/vitamins/ai/ingredientmono-923/blue-green-algae)
Fucoidon		Additive effects with anticoagulants. Because fucoidan can slow down the blood-clotting process, and may increase bleeding	Warfarin and heparin- due to its anti-thrombotic effects, there are chances of increased bleeding.	(Accessed September 28, 2019 from https://www.mskcc.org/cancer- care/integrative-medicine/ herbs/fucoidan)

Disclosure statement

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References

Abbott, I. A. 1988. Food and food products from seaweeds. In: Algae and human affairs, eds. C. A. Lembi, and J. R. Waaland, 135-147. New York: Cambridge University Press.

Abe, S., K. Hiramatsu, O. Ichikawa, H. Kawamoto, T. Kasagi, Y. Miki, T. Kimura, and T. Ikeda. 2013. Safety evaluation of excessive ingestion of Mozuku Fucoidan in Human. Journal of Toxicology and Chemical Food Safety 78:648-51. doi: 10.1111/j.1750-3841.2012. 02966.

Agardh, C. 1822. Algae: native, invasive elsewhere Caulerpa taxifolia. Botany, University of Hawai'i at Manoa, 43-4.

Agardh, J. 1852. Aplysiatoxin and debromoaplysiatoxin as the causative agents of a red alga Gracilaria Coronopifolia poisoning. Botany, University of Hawai'i at Manoa 37 (7):11-2.

Aguilar-Briseño, J., L. Cruz-Suarez, J. F. Sassi, D. Ricque-Marie, P. Zapata-Benavides, E. Mendoza-Gamboa, C. Rodríguez-Padilla, and L. Trejo-Avila. 2015. Sulphated polysaccharides from Ulva clathrata and Cladosiphon okamuranus seaweeds both inhibit viral attachment/entry and cell-cell fusion, in NDV infection. Marine Drugs 13 (2):697-712. doi: doi: 10.3390/md13020697.

Ailli Zakaria, N., D. Ibrahim, S. Fariza Sulaiman, and N. Afifah Supardy. 2011. Assessment of antioxidant activity, total phenolic content and in-vitro toxicity of Malaysian red seaweed, Acanthophora spicifera. Journal of Chemical and Pharmaceutical Research 3 (3):182-91.

Akhtar, P., and V. Sultana. 2002. Biochemical studies of some seaweed species from Karachi coast. Records Zoological Survey of Pakistan 14:

Alves, R. N., A. L. Maulvault, V. L. Barbosa, M. F. Tejedor, A. Tediosi, M. Kotterman, F. H. M. van den Heuvel, J. Robbens, J. O. Fernandes, R. R. Rasmussen, et al. 2018. Oral bioaccessibility of toxic and essential elements in raw and cooked commercial seafood



- species available in European markets. Food Chemistry 267:15-27. doi: 10.1016/j.foodchem.2017.11.045.
- Annette, K. M. 2011. Drug use in long-term care (effects on nutrition & health status), 2nd ed. Nutrition Dimension.
- Anthoni, U., C. Larsen, P. H. Nielsen, C. Christophersen, and G. W. Fischer. 1987. Haphazard isolation of diethyl ether and structures of mozuku toxin A. Acta Chemica Scandinavica 41b:216-8. doi: 10. 3891/acta.chem.scand0216.
- Appeltans, W., S. T. Ahyong, G. Anderson, M. V. Angel, T. Artois, N. Bailly, R. Bamber, A. Barber, I. Bartsch, A. Berta, et al. 2012. The magnitude of global marine species diversity. Current Biology 22 (23):2189-202. doi: 10.1016/j.cub.2012.09.036.
- Applegate, R. D., and P. B. Gray. 1995. Nutritional value of seaweed to ruminants. Rangifer 15 (1):15-8. doi: 10.7557/2.15.1.1152.
- Athukorala, Y., K. W. Lee, S. K. Kim, and Y. J. Jeon. 2006. Anticoagulant activity of marine green and brown algae collected from Jeju Island in Korea. Bioresource Technology (98):1711-6. doi: 10.1016/j.biortech.2006.07.034.
- Atlas, R., J. Lemus, J. Reed, D. Atkins, and L. Alger. 1998. Second trimester abortion using prostaglandin E2 suppositories with or without intracervical Laminaria japonica: A randomized study. Obstetrics & Gynecology 92 (3):398-402. doi: 10.1016/S0029-7844(98)00194-X.
- Azuma, K., T. Ishihara, H. Nakamoto, T. Amaha, T. Osaki, T. Tsuka, T. Imagawa, S. Minami, O. Takashima, S. Ifuku, et al. 2012. Effects of oral administration of fucoidan extracted from Cladosiphon okamuranus on tumor growth and survival time in a tumor-bearing mouse model. Marine Drugs 10 (12):2337-48.): doi: 10.3390/ md10102337.
- Babu, M., L. Palanikumar, N. Nagarani, V. J. Devi, S. R. Kumar, C. M. Ramakritinan, and A. K. Kumaraguru. 2014. Cadmium and copper toxicity in three marine macroalgae: Evaluation of the biochemical responses and DNA damage. Environmental Science and Pollution Research 21 (16):9604-16. doi: 10.1007/s11356-014-2999-0.
- Baden, D., L. E. Fleming, and J. A. Bean. 1995. Chapter: Marine toxins. In: Handbook of clinical neurology: Intoxications of the nervous system part ii. Natural toxins and drugs, ed. F. A. de Wolff, 141-175. Amsterdam: Elsevier Press.
- Banu, A., and S. Umamageswari. 2011. Toxicity study of seaweeds in rat. Continental Journal of Food Science and Technology 5 (2):23-
- Bareuther, C. M. 2008. Dangerous food-drug interactions: Aging well. Today's Geriatric Medicine 1:4.
- Barta, E. S., A. L. Branen, and H. K. Leung. 1981. Nutritional analysis of puget sound bull kelp (Nereocystis luetkeana). Journal of Food Science 46 (2):494-7. doi: 10.1111/j.1365-2621.1981.tb04894.x.
- Bellmann, S., K. Miyazaki, O. Chonan, F. Ishikawa, and R. Havenaar. 2014. Fucoidan from Cladosiphon okamuranus tokida added to food has no adverse effect on availability for absorption of divalent minerals in the dynamic multicompartmental model of the upper gastrointestinal tract. Food Digestion 5 (1-3):19-25. doi: 10.1007/ s13228-014-0036-x.
- Bouga, M., and E. Combet. 2015. Emergence of seaweed and seaweedcontaining foods in the UK: Focus on labelling, iodine content. Foods 15 (4):240-53. doi: 10.3390/foods4020240.
- Brewitz, L., J. Llaveria, A. Yada, and A. Furstner. 2013. Formal total synthesis of the algal toxin (-)-polycavernoside A. Chemistry - A European Journal 19 (14):4532-7. doi: 10.1002/chem.201204551.
- Brown, E. M., P. J. Allsopp, P. J. Magee, C. I. Gill, S. Nitecki, C. R. Strain, and E. M. McSorley. 2014. Seaweed and human health. Nutrition Reviews 72 (3):205-16. doi: 10.1111/nure.12091.
- Burtin, P. 2003. Nutritional value of seaweeds. Electronic Journal of Environmental, Agricultural and Food Chemistry (2):498-503.
- Buschmann, A. H., C. Camus, J. Infante, A. Neori, A. Israel, M. C. Hernández-González, S. V. Pereda, J. L. Gomez-Pinchetti, A. Golberg, N. Tadmor-Shalev, et al. 2017. Seaweed production: Overview of the global state of exploitation, farming and emerging research activity. European Journal of Phycology 52 (4):391-406. doi: 10.1080/09670262.2017.1365175.
- Cagide, E., M. C. Louzao, I. R. Ares, M. R. Vieytes, M. Yotsu-Yamashita, L. A. Paquette, T. Yasumoto, and L. M. Botana. 2007.

- Effects of a synthetic analog of polycavernoside A on human neuroblastoma cells. Cellular Physiology and Biochemistry 19 (1-4):185-94. doi: 10.1159/000099206.
- Cao, Y., Y. Hao, Z. H. Li, S. T. Liu, and L. X. Wang. 2016. Antiviral activity of polysaccharide extract from Laminaria japonica against Respiratory Syncytial Virus. Biomedicine and Pharmacotherapy doi: 10.1016/j.biopha.2016.10.082.
- Cardoso, S. M., O. R. Pereira, A. M. Seca, D. C. Pinto, and A. M. Silva. 2015. Seaweeds as preventive agents for cardiovascular diseases: From nutrients to functional foods. Marine Drugs 13 (11):6838-65. doi: 10.3390/md13116838.
- Catarino, M. D., M. S. Silva, and S. M. Cardoso. 2018. Phycochemical constituents and biological activities of Fucus Spp. Marine Drugs 16 (8):248. doi: 10.3390/md16080:249.
- Chen, Q., X.-D. Pan, B.-F. Huang, and J.-L. Han. 2018. Distribution of metals and metalloids in dried seaweeds and health risk to population in south-eastern China. Scientific Reports 8 Article number: 3578. doi: 10.1038/s41598-018-21732-z.
- Cheney, D. 2016. Toxic and harmful seaweeds, seaweed in health and disease prevention. Elsevier. doi: 10.1016/B978-0-12-802772-1.00013-0.
- Chiu, K. W., and A. Y. L. Fung. 1997. The cardiovascular effects of green beans (Phaseolus aureus), common rue (Ruta graveolens), and kelp (Laminaria japonica) in rats. General Pharmacology: The Vascular System 29 (5):859-62. doi: 10.1016/S0306-3623(97)00001-3.
- Choi, J.-S., W. S. Moon, J. N. Choi, D. K. H, S. H. Moon, K. K. Cho, C. J. Han, and I. S. Choi. 2013. Effects of seaweed Laminaria japonica extracts on skin moisturizing activity in vivo. Journal of Cosmetic Science 64 (3):193-205.
- Chu, W.-L. 2011. Potential applications of antioxidant compounds derived from algae. Current Topics in Nutraceutical Research 9 (3):
- Ciaran, F., G. Eimear, T. Deniz, and H. Maria. 2011. Heart Health peptides from macroalgae and their potential use in functional foods. Journal of Agricultural Food Chemistry 13 (59):6829-36. doi: 10. 1021/jf201114d.
- Citkowska, A., M. Szekalska, and K. Winnicka. 2019. Possibilities of fucoidan utilization in the development of pharmaceutical dosage forms. Marine Drugs 17 (8):458. doi: 10.3390/md17080458.
- Costa, E. L., M. Abreu, D. Gargiulo, E. Rocha, and A. A. Ramos. 2017. Anticancer effects of seaweed compounds fucoxanthin and phloroglucinol, alone and in combination with 5-fluorouracil in colon cells. Journal of Toxicology and Environmental Health, Part A 80:776-87. doi: 10.1080/15287394.2017.1357297.
- Cumashi, A., N. A. Ushakova, M. E. Preobrazhenskaya, A. D'Incecco, A. Piccoli, L. Totani, N. Tinari, G. E. Morozevich, A. E. Berman, M. I. Bilan, et al. 2007. A comparative study of the anti-inflammatory, anticoagulant, antiangiogenic, and antiadhesive activities of nine different fucoidans from brown seaweeds. Glycobiology 17 (5): 541-52. doi: 10.1093/glycob/cwm014.
- Davinelli, S., M. E. Nielsen, and G. Scapagnini. 2018. Astaxanthin in Skin health, repair, and disease: A comprehensive review. Nutrients 10 (4):522. doi: 10.3390/nu10040522.
- De Almeida, C. L. F., H. Falcão, S. de, G. R. d M. Lima, C. Montenegro, A. de, N. S. Lira, P. F. de Athayde-Filho, L. C. Rodrigues, M. F. V. de Souza, et al. 2011. Bioactivities from marine algae of the genus Gracilaria. International Journal of Molecular Sciences 12 (7):4550-73. doi: 10.3390/ijms12074550.
- Desideri, D., Cantaluppi, C. F. Ceccotto, M. A. Meli, C. Roselli, C., and L. Feduzi. 2016. Essential and toxic elements in seaweeds for human consumption. Journal of Toxicology and Environmental Health Part A 79 (3):112-22. doi: 10.1080/15287394.2015.1113598.
- Desideri, D., C. Roselli, L. Feduzi, L. Ugolini, and M. A. Meli. 2018. Applicability of an in vitro gastrointestinal digestion method to evaluation of toxic elements bioaccessibility from algae for human consumption. Journal of Toxicology and Environmental Health Part A 81 (8):212-7. doi: 10.1080/15287394.2018.1436480.
- Di Costanzo, F., V. D. Dato, A. Ianora, and G. Romano. 2019. Prostaglandins in marine organisms: A review. Marine Drugs 17 (7): 428. doi: 10.3390/md17070428.

- Díaz-Rubio, M. E., J. Pérez-Jiménez, and F. Saura-Calixto. 2009. Dietary fiber and antioxidant capacity in Fucus vesiculosus products. International Journal of Food Sciences and Nutrition 60 (Suppl. 2): 23-34. doi: 10.1080/09637480802189643.
- Dietrich, D., and S. Hoeger. 2005. Guidance values for microcystins in water and cyanobacterial supplement products (blue-green algal supplements): A reasonable or misguided approach?. Toxicology and Applied Pharmacology 203 (3):273-89. doi: 10.1016/j.taap.2004.09.
- Dring, M. J. 1982. Marine plants: Taxonomic, morphological and ecological categories. In The biology of marine plants. London: Edward Arnold British Pharmacological Society.
- El Shoubaky, A. G., M. M. Abdel-Daim, H. M. Mohamed, and E. A. Salem. 2016. Isolation and identification of a flavone apigenin from marine red alga Acanthophora spicifera with antinociceptive and anti-inflammatory activities. Journal of Experimental Neuroscience 10:21-9. doi: 10.4137/JEN.S25096.
- Elizondo-Gonzalez, R., L. E. Cruz-Suarez, D. Ricque-Marie, E. Mendoza-Gamboa, C. Rodriguez-Padilla, and L. M. Trejo-Avila. 2012. In vitro characterization of the antiviral activity of fucoidan from Cladosiphon okamuranus against Newcastle Disease Virus. Virology Journal 9 (1):307. doi: 10.1186/1743-422X-9-307.
- El-Sarraf, W., and G. El-Shaarawy. 1994. Chemical composition of some marine macroalgae from the Mediterranean Sea of Alexandria, Egypt. The Bulletin of the High Institute of Public Health 24:523-34.
- Felline, S., R. Caricato, A. Cutignano, S. Gorbi, M. G. Lionetto, E. Mollo, F. Regoli, and A. Terlizzi. 2012. Subtle effects of biological invasions: Cellular and physiological responses of fish eating the exotic pest Caulerpa racemosa. PloS One 7 (6):e38763. doi: 10.1371/ journal.pone.0038763.
- Ferdouse, F., S. L. Holdt, R. Smith, P. Murúa, and Z. Yang. 2018. The global status of seaweed production, trade and utilization. FAO Globefish Research Program 124:1-114.
- Fitton, J. H. 2011. Therapies from fucoidan; multifunctional marine polymers. Marine Drugs 9 (10):1731-60. doi: 10.3390/md9101731.
- Fitzgerald, C., E. Gallagher, D. Tasdemir, and M. Hayes. 2011. Heart health peptides from macroalgae and their potential use in functional foods. Journal of Agricultural and Food Chemistry 59 (13): 6829-36. doi: 10.1021/jf201114d.
- Fleurence, J., C. L. Coeur, S. Mabeau, M. Maurice, and A. Land-Rein. 1995. Comparison of different extractive procedures from the edible seaweeds Ulva rigida and Ulva rotundata. Journal of Applied Phycology 7 (6):577-82. doi: 10.1007/BF00003945.
- Food for Human Consumption, Code of Federal Regulations, Food and Drug Administration. 2011. https://www.accessdata.fda.gov/ scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=101.9&SearchTerm= nutrition%20label
- Francavilla, M., M. Franchi, M. Monteleone, and C. Caroppo. 2013. The red seaweed Gracilaria gracilis as a multi products source. Marine Drugs 11 (10):3754-76. doi: 10.3390/md11103754.
- Fujiwara-Arasaki, T., N. Mino, and M. Kuroda. 1984. The protein value in human nutrition of edible marine algae in Japan. Hydrobiologia 116-117 (1):513-6. doi: 10.1007/BF00027735.
- Funaki, M., M. Nishizawa, T. Sawaya, S. Inoue, and T. Yamagishi. 2001. Mineral composition in the holdfast of three brown algae of the genus Laminaria. Fisheries Science 67 (2):295-300. doi: 10.1046/j. 1444-2906.2001.00236.x.
- Fusetani, N., and K. Hashimoto. 1981. Diethyl peroxides probably responsible for Mozuku poisoning. Nippon Suisan Gakkaishi 47 (8): 1059-63. doi: 10.2331/suisan.47.1059.
- Fusetani, N., and K. Hashimoto. 1984. Prostaglandin E2: A candidate for causative agent of "ogoniri" poisoning. Nippon Suisan Gakkaishi 50 (3):465-9. doi: 10.2331/suisan.50.465.
- García, I., R. Castroviejo, and C. Neira. 1993. Las algas en Galicia: Alimentación y Otros Usos. La Coruña: Consellería de Pesca, Marisqueo e Acuicultura - Xunta de Galícia
- Gideon, T. P., and R. Rengasamy. 2008. Toxicological evaluation of fucoidan from Cladosiphon okamuranus. Journal of Medicinal Food 11 (4):638-42. doi: 10.1089/jmf.2007.0127.

- Gilroy, D. J., K. W. Kauffman, R. A. Hall, X. Huang, and F. S. Chu. 2000. Assessing potential health risks from microcystin toxins in blue-green algae dietary supplements. Environmental Health Perspectives 108 (5):435-9. doi: 10.1289/ehp.00108435.
- Ginneken, V. V., and E. de Vries. 2016. Towards a seaweed based economy: The global ten billion people issue at the midst of the 21st century. Journal of Fisheries Sciences 10 (2):001-11.
- Gómez-Guzmán, M., A. Rodríguez-Nogales, F. Algieri, and J. Gálvez. 2018. Potential role of seaweed polyphenols in cardiovascular-associated disorders. Marine Drugs 16 (8):250. doi: 10.3390/md16080250.
- Gómez-Ordóñez, E., A. Jiménez-Escrig, and P. Rupérez. 2010. Dietary fibre and physicochemical properties of several edible seaweeds from the northwestern Spanish coast. Food Research International 43 (9): 2289-94. doi: 10.1016/j.foodres.2010.08.005.
- Gorbi, S., M. E. Giuliani, L. Pittura, G. d'Errico, A. Terlizzi, S. Felline, L. Grauso, E. Mollo, A. Cutignano, and F. Regoli. 2014. Could molecular effects of Caulerpa racemosa metabolites modulate the impact on fish populations of Diplodus sargus? Marine Environmental Research 96:2-11. doi: 10.1016/j.marenvres.2014.01.
- Grasa-Lopez, A., A. Milliar-Gracia, L. Quevedo-Corona, N. Paniagua-Catro, G. Escalona-Cardoso, E. Reyes-Maldono, and M. E. Jaramillo-Flores. 2016. Undaria pinnatifida and Fucoxanthin ameliorate lipogenesis and markers of both inflammation and cardiovascular dysfunction in an animal model of diet-induced obesity. Marine Drugs 14 (8):148. doi: 10.3390/md14080148.
- Guerra-Rivas, G., C. M. Gómez-Gutiérrez, G. Alarcón-Arteaga, I. E. Soria-Mercado, and N. E. Ayala Sánchez. 2010. Screening for anticoagulant activity in marine algae from the Northwest Mexican Pacific coast. Journal of Applied Phycology 23:495-503. doi: 10.1007/ s10811-010-9618-3.
- Hammann, M., M. Rempt, G. Pohnert, G. Wang, S. M. Boo, and F. Weinberger. 2016. Increased potential for wound activated production of prostaglandin E2 and related toxic compounds in non-native populations of Gracilaria vermiculophylla. Harmful Algae 51:81-8. doi: 10.1016/j.hal.2015.11.009.
- Hanaoka, K., K. Yosida, M. Tamano, T. Kuroiwa, T. Kaise, and S. Maeda. 2001. Arsenic in the prepared edible brown alga hijiki, Hizikia fusiforme. Applied Organometallic Chemistry 15 (6):561-5. doi: 10.1002/aoc.195.
- Hassan, A. M., S. H. Abdel-Aziem, and M. A. Abdel-Wahhab. 2012. Modulation of DNA damage and alteration of gene expression during aflatoxicosis via dietary supplementation of Spirulina (Arthrospira) and Whey protein concentrate. Ecotoxicology and Environmental Safety (79):294-300. Epub 2012 Feb 9. doi: 10.1016/j. ecoenv.2012.01.017.
- Higa, T., and M. Kuniyoshi. 2000. Toxins associated with medicinal and edible seaweeds. Journal of Toxicology: Toxin Reviews 19 (2): 119-37. doi: 10.1081/TXR-100100317.
- Holbrook, A. M., J. A. Pereira, R. Labiris, H. McDonald, J. D. Douketis, M. Crowther, and P. S. Wells. 2005. Systematic overview of warfarin and its drug and food interactions. Archives of Internal Medicine 165:1095-106. doi: 10.1001/archinte.165.10.1095.
- Holdt, S. L., and S. Kraan. 2011. Bioactive compounds in seaweed: Functional food applications and legislation. Journal of Applied Phycology 23 (3):543-97. doi: 10.1007/s10811-010-9632-5.
- Hsu, B. Y., C.-Y. Tsao, T. K. Chiou, P. A. Hwang, and D. F. Hwang. 2007. HPLC determination for prostaglandins from seaweed. Food Control 18 (6):639-45. doi: 10.1016/j.foodcont.2006.02.013.
- Huisman, J. 2006. Vegetables from the sea. Landscope, summer edition 2006. Published by the Department of Environment and Conservation, Western Australia, 20-5.
- Husson, B., T. Hernández-Fariñas, R. L. Gendre, M. Schapira, and A. Chapell. 2016. Two decades of Pseudo-nitzschia spp. blooms and king scallop (Pecten maximus) contamination by domoic acid along the French Atlantic and English Channel coasts: Seasonal dynamics, spatial heterogeneity and interannual variability. Harmful Algae 51: 26-39. doi: 10.1016/j.hal.2015.10.017.
- Hwang, Y. O., S. G. Park, G. Y. Park, S. M. Choi, and M. Y. Kim. 2010. Total arsenic, mercury, lead, and cadmium contents in edible

- dried seaweed in Korea. Food Additives and Contaminants: Part B 3 (1):7-13. doi: 10.1080/19440040903532079.
- Hwang, P.-A., M.-D. Yan, H.-T V. Lin, K.-L. Li, and Y.-C. Lin. 2016. Toxicological evaluation of low molecular weight fucoidan in vitro and in vivo. Marine Drugs 14 (7):121. doi: 10.3390/md14070121.
- Ichikawa, S., M. Kamoshida, K. Hanaoka, M. Hamano, T. Maitani, and T. Kaise. 2006. Decrease of arsenic in edible brown algae Hijikia fusiforme by the cooking process. Applied Organometallic Chemistry 20 (9):585-90. doi: 10.1002/aoc.1102.
- Indergaard, M., and J. Minsaas. 1991. Animal and human nutrition. In Seaweed resources in Europe, eds. M. D. Guiry and G. Blunden, 21-64. Chichester: Wiley.
- Iso, H. 2011. Lifestyle and cardiovascular disease in Japan. Journal of Atherosclerosis and Thrombosis 18 (2):83-8. doi: 10.5551/jat.6866.
- Ito, K., and K. Hori. 1989. Seaweed: Chemical composition and potential food uses. Food Reviews International 5 (1):101-44. doi: 10.1080/ 87559128909540845.
- Ito, K., and H. Nagai. 1998. Morphological observations of diarrhoea in mice caused by aplysiatoxin, the causative agent of the red alga Gracilaria coronopifolia poisoning in Hawaii. Toxicon 36 (12): 1913-20. doi: 10.1016/S0041-0101(98)00113-5.
- Ito, K., and H. Nagai. 2000. Bleeding from the small intestine caused by aplysiatoxin, the causative agent of the red alga Gracilaria coronopifolia poisoning. Toxicon 38 (1):123-32. doi: 10.1016/S0041-0101(98)00113-5.
- Jang, W. S., and S. Y. Choung. 2013. Antiobesity effects of the ethanol extract of Laminaria japonica Areshoung in high fat diet induced obese rat. Evidence-Based Complementary and Alternative Medicine 2013:1-17. doi: 10.1155/2013/492807.
- Jasmanindar, Y., S. Sukenda, M. Jr. Zairin, A. Alimuddin, and N. B. P. Utomo. 2018. Dietary administration of Gracilaria verrucosa extract on Litopenaeus vannamei immune response, growth, and resistance to Vibrio harveyi. AACL Bioflux 11 (4):1069-80.
- Kadam, S., C. O'Donnell, D. Rai, M. Hossain, C. Burgess, D. Walsh, and B. Tiwari. 2015. Laminarin from Irish Brown Seaweeds Ascophyllum nodosum and Laminaria hyperborea: Ultrasound Assisted Extraction, Characterization and Bioactivity. Marine Drugs 13 (7):4270-80. doi: 10.3390/md13074270.
- Kadari, S., H. Yerrabelly, T. Gogula, Y. G. Erukala, J. R. Yerrabelly, and P. K. Begari. 2018. Stereoselective synthesis of γ -Butyrolactones subunit of polycavernoside A. Journal of Heterocyclic Chemistry 55 (8):1986-90. doi: 10.1002/jhet.3240.
- Kamat, S., Y. Wahidulla, S. L. D'Souza, C. G. Naik, V. Ambiye, D. S. Bhakuni, A. K. Goel, H. S. Garg, and R. C. Srimal. 1992. Bioactivity of marine organisms. VI. Antiviral evaluation of marine algal extracts from the Indian Coast. Botanica Marina 35 (2):161-4. doi: 10.1515/botm.1992.35.2.161.
- Kasai, Y., T. Ito, and M. Sasaki. 2012. Total synthesis of (-)-polycavernoside A: Suzuki- Miyaura coupling approach. Organic Letters 14 (12):3186-9. doi: 10.1021/ol301278e.
- Khan, M., S. Varadharaj, J. C. Shobha, M. U. Naidu, N. L. Parinandi, V. K. Kutala, and P. Kuppusamy. 2006. C-phycocyanin ameliorates doxorubicin-induced oxidative stress and apoptosis in adult rat cardiomyocytes. Journal of Cardiovascular Pharmacology 47 (1):9-20. doi: 10.1097/01.fjc.0000191520.48404.27.
- Kim, S.-K. 2017. Marine glycobiology: Principles and applications. Busan, South Korea: Taylor and Francis Group.
- Kim, S.-K., and I. Bhatnagar. 2011. Physical, chemical, and biological properties of wonder kelp-Laminaria. Advances in Food and Nutrition Research 64:85-96. doi: 10.1016/B978-0-12-387669-0.
- Kim, J., A. Shin, J. S. Lee, S. Youn, and K. Y. Yoo. 2009. Dietary factors and breast cancer in Korea: An ecological study. The Breast Journal 15 (6):683-6. 00817.x doi: 10.1111/j.1524-4741.2009.
- Klein, J., and M. Verlaque. 2008. The Caulerpa racemosa invasion: A critical review. Marine Pollution Bulletin 56 (2):205-25. doi: 10. 1016/j.marpolbul.2007.09.043.
- Kolanjinath, K., P. Ganesh, and P. Sanraj. 2014. Pharmacological importance of seaweeds: A review. World Journal of Fish and

- Marine Sciences 6 (1):1-15. doi: 10.5829/idosi.wjfms.2014.06.01.
- Kolb, N., L. Vallorani, N. Milanovi, and V. Stocchi. 2004. Evaluation of marine algae Wakame (Undaria pinnatifida) and Kombu (Laminaria digitata japonica) as food supplements. Food Technology and Biotechnology 42:57-61.
- Konecny, M. 1969. Doubling-time. Souborný referát Vnitr Lek 15: 969-76. doi: 10.1016/0041-0101(94)90311-5.
- Krishnaiah, D., S. Rosalam, D. M. R. Prasad, and A. Bono. 2008. Mineral content of some seaweeds from Sabah's South China sea. Asian Journal of Scientific Research 1 (2):166-70. doi: 10.3923/ajsr. 2008.166.170.
- Kuehnelt, D., K. J. Irgolic, and W. Goessler. 2001. Comparison of three methods for the extraction of arsenic compounds from the NRCC standard reference material DORM and the brown alga Hijiki fuziforme. Applied Organometallic Chemistry 15 (6):445-56. doi: 10. 1002/aoc.189.
- Kumar, M., V. Gupta, P. Kumari, and B. Jha. 2010. Assessment of nutrient composition and antioxidant potential of Caulerpaceae seaweeds. Journal of Food Composition and Analysis 24:270-278. doi: 10.1016/j.jfca.2010.07.007.
- Kumar, I. J. N., R. N. Kumar, K. Manmeet, A. Bora, and S. Chakraborty. 2010. Variation of biochemical composition of eighteen marine macroalgae collected from Okha coast, Gulf of Kutch. India. Electronic Journal of Environmental Agricultural and Food Chemistry 9:404-10.
- Kumar, C., G. Ponesakki, V. Suresh, and N. Bhaskar. 2008. Seaweeds as a source of nutritionally beneficial compounds - A review. Journal Food Science Technology 45 (1):1-13.
- Kurt, O., F. Ozdal Kurt, I. Tuglu, S. I. Deliloglu-Gurhan, and M. Ozturk. 2009. Neurotoxic effect of Caulerpa racemosa var. cylindracea by neurite inhibition on the neuroblastoma cell line. Russian Journal of Marine Biology 35 (4):342-50. doi: 10.1134/ S1063074009040105.
- Kusaykin, M., I. Bakunina, V. Sova, S. Ermakova, T. Kuznetsova, N. Besednova, T. Zaporozhets, and T. Zvyagintseva. 2008. Structure, biological activity, and enzymatic transformation of fucoidans from the brown seaweeds. Biotechnology Journal 3 (7):904-15. doi: 10. 1002/biot.200700054.
- Laparra, J. M., D. Velez, R. Montoro, R. Barbera, and R. Farre. 2003. Estimation of arsenic bioaccessibility in edible seaweed by an in vitro digestion method. Journal of Agricultural and Food Chemistry 51 (20):6080-5. doi: 10.1021/jf034537i.
- Laparra, J. M., D. Velez, R. Montoro, R. Barbera, and R. Farre. 2004. Bioaccessibility of inorganic arsenic species in raw and cooked Hizikia fusiforme seaweed. Applied Organometallic Chemistry 18 (12):662-9. doi: 10.1002/aoc.732.
- Lavakumar, V., K. F. H. Ahamed, and V. Ravichandiran. 2012. Anticancer and antioxidant effect of Acanthophora spicifera against EAC induced carcinoma in mice. Journal of Pharmacy Research 5 (3): 1503 - 7.
- Lawrence, J. F., C. F. Charbonneau, C. Menard, M. A. Quilliam, and P. G. Sim. 1989. Liquid chromatographic determination of domoic acid in shellfish products using the paralytic shellfish poison extraction procedure of the Association of Official Analytical Chemists. Journal of Chromatography A 462:349-56. doi: 10.1016/S0021-9673(00)91361-X.
- Lee, J. C., F. Hou, H. W. Huang, F. R. Chang, C. C. Yeh, J. Y. Tang, and H. W. Chang. 2013. Marine algal natural products with antioxidative, anti-inflammatory, and anti-cancer properties. Cancer Cell International 13 (1):55. doi: 10.1186/1475-2867-13-55.
- Lee, S. 2019. Hazards of eating nori seaweed. Last modified August 3, 2019. Accessed September 20, 2019. https://www.livestrong.com/article/496996-hazards-of-eating-nori-seaweed/
- Li, B., F. Lu, X. Wei, and R. Zhao. 2008. Fucoidan: Structure and bioactivity. Molecules 13 (8):1671-95. doi:. doi: molecules13081671.
- Lim, S. J., W. M. W. Aida, S. Schiehser, T. Rosenau, and S. Böhmdorfer. 2019. Structural elucidation of fucoidan from



- Cladosiphon okamuranus (Okinawa mozuku). Food Chemistry 272: 222-6. doi: 10.1016/j.foodchem.2018.08.034.
- Liu, Q., Y. Huang, R. Zhang, T. Cai, and Y. Cai. 2016. Medical application of Spirulina platensis derived C-phycocyanin. Evidence-Based Complementary and Alternative Medicine (2016):1-14. doi: 10.1155/ 2016/7803846.
- Li, N., Q. Zhang, and J. Song. 2005. Toxicological evaluation of fucoidan extracted from Laminaria japonica in Wistar rats. Food and Chemical Toxicology 43 (3):421-6. doi: 10.1016/j.fct.2004.12.001.
- López-López, I., S. Bastida, C. Ruiz-Capillas, L. Bravo, M. T. Larrea, F. Sánchez-Muniz, S. Cofrades, and F. Jiménez-Colmenero. 2009. Composition and antioxidant capacity of low-salt meat emulsion model systems containing edible seaweeds. Meat Science 83 (3): 492-8. doi: 10.1016/j.meatsci.2009.06.031.
- Mabeau, S., and J. Fleurence. 1993. Seaweed in food products: Biochemical and nutritional aspects. Trends in Food Science & Technology 4 (4):103-7. doi: 10.1016/0924-2244(93)90091-N.
- MacArtain, P., C. Gill, M. Brooks, R. Campbell, and I. Rowland. 2007. Nutritional value of edible sea weeds. Nutrition Reviews 65(1):535-543. doi: 10.1301/nr.2007.dec.535-543.
- Mamatha, B. S., K. K. Namitha, A. Senthil, J. Smitha, and G. A. Ravishankar. 2007. Studies on use of Enteromorpha in snack food. Food Chemistry 101 (4):1707-13. doi: 10.1016/j.foodchem.2006.04.
- Manivannan, K., G. Thirumaran, G. K. Devi, A. Hemalatha, and P. Anantharaman. 2008. Biochemical composition of seaweeds from Mandapam coastal regions along Southeast Coast of India. American-Eurasian Journal of Botany 1:32-7.
- Manzelat, S. F., A. M. Mufarrah, B. A. Hasan, and N. A. Hussain. 2018. Macroalgae of the Red Sea from Jizan, Saudi Arabia. Journal of Phycological Society 48 (1):88-108.
- Marechal, J. P., G. Culioli, C. Hellio, H. T-Guyon, M. E. Callow, A. S. Clare, and A. O-Magné. 2004. Seasonal variation in antifouling activity of crude extracts of the brown alga Bifurcaria bifurcate (Cystoseiraceae) against cyprids of Balanus amphitrite and the marine bacteria Cobetia marina and Pseudoalteromonas haloplanktis. Journal of Experimental Marine Biology and Ecology 313(1):47-62. doi: 10.1016/j.jembe.2004.07.016.
- Marinho-Soriano, E., P. C. Fonseca, M. A. A. Carneiro, and W. S. C. Moreira. 2005. Seasonal variation in the chemical composition of two tropical seaweeds. Bioresource Technology 97 (18):2402-6. doi: 10.1016/j.biortech.2005.10.014.
- Marles, R. J., M. L. Barrett, J. Barnes, M. L. Chavez, P. Gardiner, R. Ko, G. B. Mahady, T. L. Dog, N. D. Sarma, G. I. Giancaspro, et al. 2011. United States pharmacopeia safety evaluation of Spirulina. Critical Reviews in Food Science and Nutrition 51 (7):593-604. doi: 10.1080/10408391003721719.
- Marques, A., J. Ferreira, H. Abreu, R. Pereira, A. Rego, J. Serôdio, G. Christa, I. Gaivão, and M. Pacheco. 2018. Searching for antigenotoxic properties of marine macroalgae dietary supplementation against endogenous and exogenous challenges. Journal of Toxicology and Environmental Health Part A 81 (18):939-56. doi: 10.1080/ 15287394.2018.1507856.
- Marsan, D. W., S. M. Conrad, W. L. Stutts, C. H. Parker, and J. R. Deeds. 2018. Evaluation of microcystin contamination in blue-green algal dietary supplements using a protein phosphatase inhibitionbased test kit. Heliyon 4 (3):e00573. doi: 10.1016/j.heliyon.2018.
- Mason, B. 1959. Chemical composition of tektites. Nature 183 (4656): 254-5. doi: 10.1038/183254b0.
- Massin, D. 1996. Evaluation of the toxicological risk to humans of caulerpenyne using human hematopoietic progenitors, melanocytes, and keratinocytes in culture. Journal of Toxicology and Environmental Health 47(1):47-59. doi: 10.1080/009841096161924.
- Masuda, M., T. Abe, S. Sato, T. Suzuki, and M. Suzuki. 1997. Diversity of halogenated secondary metabolites in the red alga Laurencia nipponica (Rhodomelaceae, Ceramiales). Journal of Phycology 33 (2):
- Matanjun, P., S. Mohamed, N. M. Mustapha, and K. Muhammad. 2009. Nutrient content of tropical edible seaweeds, Eucheuma

- cottonii, Caulerpa lentillifera and Sargassum polycystum. Journal of Applied Phycology 21 (1):75-80. doi: 10.1007/s10811-008-9326-4.
- Matsumoto, S., M. Nagaoka, T. Hara, I. K-Takagi, K. Mistuyama, and S. Ueyama. 2004. Fucoidan derived from Cladosiphon okamuranus Tokida ameliorates murine chronic colitis through the down-regulation of interleukin-6 production on colonic epithelial cells. Clinical and Experimental Immunology 136:432-9. doi: 10.1111/j.1365-2249.
- McCarty, M. F. 2007. Clinical potential of Spirulina as a source of phycocyanobilin. Journal of Medicinal Food 10 (4):566-70. doi: 10.1089/ imf.2007.621.
- Mišurcová, L., S. Kráčmar, B. Klejdus, and J. Vacek. 2010. Nitrogen content, dietary fibre, and digestibility in algal food products. Czech Journal of Food Sciences 28 (No. 1):27-35. doi: 10.17221/111/2009-CIFS.
- Morrissey, J., S. Kraan, and M. D. Guiry. 2001. A guide to commercially important seaweeds on the Irish coast. Dun Laoghaire: Bord Iascaigh
- Murray, M., L. Pizzorno, and L. Pizzorno. 2005. The world's healthiest foods: Sea vegetables 7 ways to eat more seaweed (and why you should) the encyclopedia of healing foods. New York, NY: Atria
- Nagai, H., T. Yasumoto, and Y. Hokama. 1996. Aplysiatoxin and debromoaplysiatoxinas the causative agents of a red alga Gracilaria cornopifolia poisoning in Hawaii. Toxicon 34 (7):753-61. doi: 10. 1016/0041-0101(96)00014-1.
- Nagai, H., T. Yasumoto, and Y. Hokama. 1997. Manauealides, some of the causative agents of a red alga Gracilaria coronopifolia poisoning in Hawaii. Journal of Natural Products 60 (9):925-8. doi: 10.1021/ np970193c.
- Nagaoka, M., H. Shibata, I. Kimura-Takagi, S. Hashimoto, K. Kimura, T. Makino, R. Aiyama, S. Ueyama, and T. Yokokura. 1999. Structural study of fucoidan from Cladosiphon okamuranus TOKIDA. Glycoconjugate Journal 16 (1):19-26. doi: 10.1023/ A:1006945618657.
- Nagappan, T., and C. S. Vairappan. 2014. Nutritional and bioactive properties of three edible species of green algae, genus Caulerpa (Caulerpaceae). Journal of Applied Phycology 26 (2):1019-27. doi: 10. 1007/s10811-013-0147-8.
- Naidu, K., A. Tiwari, H. Joshi, S. Vishwanath, H. Ramesh, and S. Rao. 1992. Evaluation of nutritional quality of food and safety of seaweeds in India. Journal of Food Safety 13 (2):77-90. doi: 10.1111/j. 1745-4565.1993.tb00096.x.
- Namikoshi, M., T. Fujiwara, T. Nishikawa, and K. Ukai. 2006. Natural abundance 14C content of dibutyl phthalate (DBP) from three marine algae. Marine Drugs 4 (4):290-7. doi: 10.3390/md404290.
- Nasrallah, R., R. Hassouneh, and R. L. Hebert. 2016. PGE2, kidney disease, and cardiovascular risk: Beyond hypertension and diabetes. Journal of the American Society of Nephrology 27 (3):666-76. doi: 10. 1681/ASN.2015050528.
- Noda, H. 1993. Health benefits and nutritional properties of Nori. Journal of Applied Phycology 5 (2):255-8. doi: 10.1007/BF00004027.
- Noguchi, T., T. Matsui, K. Miyazawa, M. Asakawa, N. Iijima, Y. Shida, M. Fuse, Y. Hosaka, C. Kirigaya, K. Watabe, et al. 1994. Poisoning by the red alga "ogonori" (Gracilaria verrucosa) on the Nojima Coast, Yokohama, and Kanagawa Prefecture, Japan. Toxicon 32 (12): 1533-8. doi: 10.1016/0041-0101(94)90311-5.
- Novelli, A., J. Kispert, M. T. FernáNdez-SáNchez, A. Torreblanca, and V. Zitko. 1992. Domoic acid-containing toxic mussels produce neurotoxicity in neuronal cultures through a synergism between excitatory amino acids. Brain Research 577 (1):41-8. doi: 10.1016/ 0006-8993(92)90535-H.
- Ortiz, J., N. Romero, P. Robert, J. Araya, J. Lopez-HernáNdez, C. Bozzo, E. Navarrete, A. Osorio, and A. Rios. 2006. Dietary fiber, amino acid, fatty acid and tocopherol contents of the edible seaweeds Ulva lactuca and Durvillaea antarctica. Food Chemistry 99 (1):98-104. doi: 10.1016/j.foodchem.2005.07.027.
- Ortiz, J., E. P. Uquiche, N. Robert, V. Romero, C. Quitral, and C. Llantén. 2009. Functional and nutritional value of the Chilean seaweeds Codium fragile, Gracilaria chilensis and Macrocystis pyrifera.

- European Journal of Lipid Science and Technology 111 (4):320-7. doi: 10.1002/ejlt.200800140.
- Osathanondh, R., A. E. Donnenfeld, F. D. Frigoletto, S. G. Driscoll, and K. J. Ryan. 1980. Induction of labour with anencephalic foetus. Obstetrics and Gynecology 56 (5):655-7.
- Padua, D., E. Rocha, D. Gargiulo, and A. A. Ramos. 2015. Bioactive compounds from brown seaweeds: Phloroglucinol, fucoxanthin and fucoidan as promising therapeutic agents against breast cancer. Phytochemistry Letters 14:91-8. doi: 10.1016/j.phytol.2015.09.007.
- Pal, A., M. C. Kamthania, and A. Kumar. 2014. Bioactive compounds and properties of seaweeds— A review. Oalib 1:e752. doi: 10.4236/ oalib.1100752.
- Patarra, R. F., L. Paiva, A. I. Neto, E. Lima, and J. Baptista. 2010. Nutritional value of selected macroalgae. Journal of Applied Phycology 15:9-14. doi: 10.1007/s10811-010-9556-0.
- Pattam, R. A., and A. Chirapart. 2006. Nutritional evaluation of tropical green seaweeds Caulerpa lentillifera and Ulva reticulata. Kasetsart Journal: Natural Science 40:75-83.
- Paz, S., C. Rubio, I. Frías, A. J. Gutiérrez, D. G-Weller, V. Martín, C. Revert, and A. Hardisson. 2019. Toxic metals (Al, Cd, Pb and Hg) in the most consumed edible seaweeds in Europe. Chemosphere 218: 879-84. doi: doi: 10.1016/j.chemosphere.2018.11.165.
- PDR for herbal medicines. 2000. American Botanical Garden. 2nd ed. Montvale, NJ: Medical Economics Company, Inc.
- Peng, Z., M. Liu, Z. Fang, L. Chen, J. Wu, and Q. Zhang. 2013. In vitro antiproliferative effect of a water-soluble Laminaria japonica polysaccharide on human melanoma cell line A375. Food and Chemical Toxicology 58:56-60. doi: 10.1016/j.fct.2013.04.026.
- Peng, Z., M. Liu, Z. Fang, J. Wu, and Q. Zhang. 2012. Composition and cytotoxicity of a novel polysaccharide from brown alga (Laminaria japonica). Carbohydrate Polymers 89 (4):1022-6. doi: 10. 1016/j.carbpol.2012.03.043.
- Pereira, F., A. R. D. Latino, S., and S. P. Gaudencio. 2014. A chemoinformatics approach to the discovery of lead-like molecules from marine and microbial sources en route to antitumor and antibiotic drugs. Marine Drugs 12 (2):757-78. doi: 10.3390/md12020757.
- Petrus, M., R. Culerrier, M. Campistron, A. Barre, and P. Rouge. 2009. First case report of anaphylaxis to spirulin: Identification of phycocyanin as responsible allergen. Allergy 65 (7):924-5. doi: 10.1111/j. 1398-9995.2009.02257.x.
- Plaza, M., A. Cifuentes, and E. Ibáñez. 2008. In the search of new functional food ingredients from algae. Trends in Food Science & Technology 19 (1):31-9. doi: 10.1016/j.tifs.2007.07.012.
- Pokapū Akoranga Pūtaiao. 2012. Toxins and food webs. Accessed September 24, 2019. https://www.sciencelearn.org.nz/resources/367toxins-and-food-webs.
- Preskitt, L. 2001. Acanthophora spicifera. University of Hawaii at Manoa Botany, 2. Accessed September 24, 2019. https://www.hawaii. $edu/reefalgae/invasive_algae/rhodo/acanthophora_spicifera.htm$
- Reboleira, J., R. Freitas, S. Pinteus, J. Silva, C. Alves, R. Pedrosa, and S. Bernardino. 2019. Brown seaweeds, nonvitamin and nonmineral nutritional supplements. Plant and Algae Extracts :171-6. doi: 10. 1016/B978-0-12-812491-8.00024-2.
- Renn, D. 1997. Biotechnology and the red seaweed polysaccharide industry: Status, needs and prospects. Trends in Biotechnology 15 (1):9-14. doi: 10.1016/S0167-7799(96)10069-X.
- Richards, J. T., E. R. Kern, L. A. Glasgow, J. C. Overall, Jr, E. F. Deign, and M. T. Hatch. 1978. Antiviral activity of extracts from marine algae. Antimicrobial Agents and Chemotherapy 14 (1):24-30. doi: 10. 1128/AAC.14.1.24.
- Riss, J., K. Decorde, T. Sutra, M. Delage, J. C. Baccou, N. Jouy, J. P. Brune, H. Oreal, J. P. Cristol, and J. M. Rouanet. 2007. Phycobiliprotein C-phycocyanin from Spirulina platensis is powerfully responsible for reducing oxidative stress and NADPH oxidase expression induced by an atherogenic diet in hamsters. Journal of Agricultural and Food Chemistry 55 (19):7962-7. doi: 10.1021/ jf070529g.
- Rodrigues, M., G. Alves, J. Abrantes, and A. Falcao. 2013. Herb-drug interaction of Fucus vesiculosus extract and amiodarone in rats: A potential risk for reduced bioavailability of amiodarone in clinical

- practice. Food and Chemical Toxicology 52:121-8. doi: 10.1016/j.fct. 2012.11.012.
- Rupérez, P. 2002. Mineral content of edible marine seaweeds. Food Chemistry 79:23-6.
- Saá, C. F. 2002. Atlantic sea vegetables nutrition and health: Properties, recipes, description. Redondela - Pontevedra: Algamar.
- Sanjeewa, K. K. A., and Y. J. Jeon. 2018. Edible brown seaweeds: A review. Journal of Food Bioactives 2:37-50. doi: 10.31665/JFB.2018. 2139.
- Santoso, J., Y. Yoshie-Stark, and T. Suzuki. 2006. Comparative contents of minerals and dietary fibres in several tropical seaweeds. Bulletin Teknologi Hasil Perikanan 9:1-11.
- Seaweed. 2011, June. Review of natural products. Facts & comparisons. St. Louis, MO: Wolters Kluwer Health Inc.
- Sfecci, E., C. L. Quemener, T. Lacour, L. Massi, P. Amade, G. Audo, and M. Mehiri. 2017. Caulerpenyne from Caulerpa taxifolia: A comparative study between CPC and classical chromatographic techniques. Phytochemistry Letters 20:406-9. doi: 10.1016/j.phytol.2017.01.
- Shanmugam, A., and C. Palpandi. 2008. Biochemical composition and fatty acid profile of the green alga Ulva reticulata. Asian Journal of Biochemistry 3:26-31. doi: 10.3923/ajb.2008.26.31.
- Shibata, H., I. Kimura-Takagi, M. Nagaoka, S. Hashimoto, R. Aiyama, M. Iha, S. Ueyama, and T. Yokokura. 2000. Properties of fucoidan from Cladosiphon okamuranus tokida in gastric mucosal protection. Biofactors 11 (4):235-45. doi: 10.1002/biof.5520110402.
- Skrovankova, S. 2011. Seaweed vitamins as nutraceuticals. Advances in Food and Nutrition Research 64:357-69. doi: 10.1016/B978-0-12-387669-0.00028-4.
- Smith, J. E., C. L. Hunter, and C. M. Smith. 2002. Distribution and reproductive characteristics of nonindigenous and invasive marine algae in the Hawaiian Islands. Pacific Science 56 (3):299-315. doi: 10.1353/psc.2002.0030.
- Smith, J. L., G. Summers, and R. Wong. 2010. Nutrient and heavy metal content of edible seaweeds in New Zealand. New Zealand Journal of Crop and Horticultural Science 38 (1):19-28. doi: 10.1080/ 01140671003619290.
- Subba Rao, D. V., M. A. Quilliam, and R. Pocklington. 1988. Domoic acid - a neurotoxic amino acid produced by the marine diatom Nitzschia pungens in culture. Canadian Journal of Fisheries and Aquatic Sciences 45:2076-9. doi: 10.1139/f88-241.
- Sugawa-Katayama, Y., and M. Katayama. 2009. Release of minerals from dried Hijiki, Sargassum fusiforme (Harvey) Setchell, during water-soaking. Trace Nutrients Research (24):106-9.
- Sutkin, G., Capelle, S. D. Susan, P. M. Schlievert, and M. D. Creinin. 2001. Toxic shock syndrome after Laminaria insertion. Obstetrics & Gynecology 98 (5):959-61. doi: 10.1016/S0029-7844(01)01502-2.
- Taboada, C., R. Millán, and I. Míguez. 2010. Composition, nutritional aspects and effect on serum parameters of marine algae Ulva rigida. Journal of the Science of Food and Agriculture 93 (90):445-9. doi: 10. 1002/jsfa.5981.
- Tako, M., E. Yoza, and S. Tohma. 2000. Chemical characterization of acetyl Fucoidan and alginate from commercially cultured Cladosiphon okamuranus. Botanica Marina 43 (4):393-8. doi: 10. 1515/BOT.2000.040.
- Taylor, V. F., Z. Li, V. Sayarath, T. J. Palys, K. R. Morse, R. A. Scholz-Bright, and M. R. Karagas. 2017. Distinct arsenic metabolites following seaweed consumption in humans. Scientific Reports 7 (1):1-9. doi: 10.1038/s41598-017-03883-7.
- Thomes, P., M. Rajendran, B. Pasanban, and R. Rengasamy. 2010. Cardioprotective activity of Cladosiphon okamuranus fucoidan against isoproterenol induced myocardial infarction in rats. Phytomedicine 18 (1):52-7. doi: 10.1016/j.phymed.2010.06.006.
- Trejo-Avila, L. T., R. E-Gonzalez, P. R-Santillan, J. A. A. Briseño, D. R -Marie, C. R. Padilla, and L. E. Cruz-Suarez. 2016. Innocuity and anti-Newcastle-virus-activity of Cladosiphon okamuranus fucoidan in chicken embryos. Poultry Science 95 (12):2795-802. doi: 10.3382/ps/
- Truus, K., M. Vaher, and I. Taure. 2001. Algal biomass from Fucus vesiculosus (Phaeophyta): investigation of the mineral and alginate



- components. Proceedings of the Estonian Academy of Sciences 50: 95-103.
- Val, A., G. Platas, A. Basilio, A. Cabello, J. Gorrochategui, I. Suay, F. Vicente, E. Portillo, M. Río, G. Reina, et al. 2001. Screening of antimicrobial activities in red, green and brown macroalgae from Gran Canaria (Canary Islands, Spain). *International Microbiology* 4 (1): 35–40. doi: 10.1007/s101230100006.
- Wang, L., Y.-J. Park, Y.-J. Jeon, and B. M. Ryu. 2018. Bioactivities of the edible brown seaweed, *Undaria pinnatifida*: A review. *Aquaculture* (495):873–80. doi: 10.1016/j.aquaculture.2018.06.079.
- World Health Organization. 2000. Evaluation and use of epidemiological evidence for environmental health risk assessment: WHO guideline document. *Environmental Health Perspective* 108:99.
- Ye, N. H., G. C. Wang, and C. K. Tseng. 2005. Effect of heavy metals (Cd, Cu) on the gametophytes of *Laminaria japonica* Aresch. *Journal of Integrative Plant Biology* 47 (8):942–51. doi: 10.1111/j. 1744-7909.2005.00116.x.
- Yotsu-Yamashita, M., R. L. Haddock, and T. Yasumoto. 1993. Polycavernoside A: A novel glycosidic macrolide from the red alga *Polycavernosa tsudai* (*Gracilaria edulis*). *Journal of the American Chemical Society* 115 (3):1147–8. doi: 10.1021/ja00056a048.
- Yotsu-Yamashita, M., T. Yasumoto, S. Yamada, F. F. A. Bajarias, M. A. Formeloza, M. L. Romero, and Y. Fukuyo. 2004. Identification of polycavernoside A as the causative agent of the fatal food poisoning resulting from ingestion of the red alga *Gracilaria edulis* in the Philippines. *Chemical Research in Toxicology* 17 (9):1265–71. doi: 10. 1021/tx0498556.
- Yu, X., Q. Zhang, W. Cui, Z. Zeng, W. Yang, C. Zhang, H. Zhao, W. Gao, X. Wang, and D. Luo. 2014. Low molecular weight fucoidan alleviates cardiac dysfunction in diabetic Goto-Kakizaki rats by

- reducing oxidative stress and cardiomyocyte apoptosis. *Journal of Diabetes Research* 2014:1–13. doi: 10.1155/2014/420929.
- Yuan, Y. V. 2008. Marine algal constituents. In Marine nutraceuticals and functional foods, eds. C. Barrow and F. Shahidi, 259–296. New York: CRC Press, Taylor and Francis Group.
- Zava, T. T., and D. T. Zava. 2011. Assessment of Japanese iodine intake based on seaweed consumption in Japan: A literature-based analysis. *Thyroid Research* 4 (1):14. doi: 10.1186/1756-6614-4-14.
- Zeng, L.-M., C.-J. Wang, J.-Y. Su, D. Li, N. L. Owen, Y. Lu, N. Lu, and Q.-T. Zheng. 2010. Flavonoids from the Red Alga Acanthophora spicifera. Chinese Journal of Chemistry 19 (11):1097–100. doi: 10. 1002/cjoc.20010191116.
- Zerrifi, S. E. A., F. E. Khalloufi, B. Oudra, and V. Vasconcelos. 2018. Seaweed bioactive compounds against pathogens and microalgae: Potential uses on pharmacology and harmful algae bloom control. *Marine Drugs* 16 (2):55. doi: 10.3390/md16020055.
- Zhao, Y., D. Shang, J. Ning, and Y. Zhai. 2012. Arsenic and cadmium in the marine macroalgae (*Porphyra yezoensis* and *Laminaria Japonica*) forms and concentrations. *Chemical Speciation & Bioavailability* 24 (3):197–203. doi: 10.3184/095422912X13404690516133.
- Zhao, Y.-F., J.-F. Wu, D.-R. Shang, J.-S. Ning, H.-Y. Ding, and Y.-X. Zhai. 2014. Arsenic species in edible seaweeds using *In vitro* biomimetic digestion determined by high-performance liquid chromatography inductively coupled plasma mass spectrometry. *International Journal of Food Science* 2014:1–12. doi: 10.1155/2014/436347.
- Zhao, X., C. H. Xue, Z. J. Li, Y. P. Cai, H. Y. Liu, and H. T. Qi. 2004. Antioxidant and hepatoprotective activities of low molecular weight sulfated polysaccharide from *Laminaria japonica*. *Journal of Applied Phycology* 16 (2):111. doi: 10.1023/B:JAPH.0000044822.10744.59.