

# Critical Reviews in Food Science and Nutrition



ISSN: 1040-8398 (Print) 1549-7852 (Online) Journal homepage: http://www.tandfonline.com/loi/bfsn20

# Regulation of glucose metabolism by bioactive phytochemicals for the management of type 2 diabetes mellitus

Chao Zhao, Chengfeng Yang, Sydney Tang Chi Wai, Yanbo Zhang, Maria P. Portillo, Paolo Paoli, Yijing Wu, Wai San Cheang, Bin Liu, Christian Carpéné, Jianbo Xiao & Hui Cao

To cite this article: Chao Zhao, Chengfeng Yang, Sydney Tang Chi Wai, Yanbo Zhang, Maria P. Portillo, Paolo Paoli, Yijing Wu, Wai San Cheang, Bin Liu, Christian Carpéné, Jianbo Xiao & Hui Cao (2018): Regulation of glucose metabolism by bioactive phytochemicals for the management of type 2 diabetes mellitus, Critical Reviews in Food Science and Nutrition, DOI: 10.1080/10408398.2018.1501658

To link to this article: <a href="https://doi.org/10.1080/10408398.2018.1501658">https://doi.org/10.1080/10408398.2018.1501658</a>

	Published online: 03 Dec 2018.
	Submit your article to this journal $oldsymbol{\mathcal{C}}$
CrossMark	View Crossmark data 🗗



#### **REVIEW**



# Regulation of glucose metabolism by bioactive phytochemicals for the management of type 2 diabetes mellitus

Chao Zhao<sup>a,b</sup>, Chengfeng Yang<sup>a,c</sup>, Sydney Tang Chi Wai<sup>d</sup>, Yanbo Zhang<sup>e</sup>, Maria P. Portillo<sup>f,g</sup>, Paolo Paoli<sup>h</sup>, Yijing Wu<sup>c,i</sup>, Wai San Cheang<sup>j</sup>, Bin Liu<sup>a</sup>, Christian Carpéné<sup>k</sup>, Jianbo Xiao<sup>j</sup>, and Hui Cao<sup>a,j</sup>

<sup>a</sup>College of Food Science, Fujian Agriculture and Forestry University, Fuzhou, China; <sup>b</sup>Department of Chemistry, University of California, Davis, CA, USA; <sup>c</sup>Institute of Oceanography, Minjiang University, Fuzhou, China; <sup>d</sup>Department of Medicine, LKS Faculty of Medicine, The University of Hong Kong, Queen Mary Hospital, Hong Kong; <sup>e</sup>School Chinese Medicine, University of Hong Kong, Hong Kong, China; <sup>f</sup>Department of Nutrition and Food Science, Faculty of Pharmacy, University of Basque Country (UPV/EHU) and Lucio Lascaray Research Center, Vitoria, Spain; <sup>g</sup>CIBEROBN Physiopathology of Obesity and Nutrition, Institute of Health Carlos III (ISCIII), Spain; <sup>h</sup>Department of Biomedical, Experimental and Clinical Sciences, University of Florence, Florence, Italy; <sup>i</sup>College of Food Science and Nutritional Engineering, China Agricultural University, China; <sup>j</sup>Institute of Chinese Medical Sciences, State Key Laboratory of Quality Control in Chinese Medicine, University of Macau, Macau SAR, China; <sup>k</sup>Institut des Maladies Métaboliques et Cardiovasculaires (I2MC), Institut National de la Santé et de la Recherche Médicale (INSERM U1048)/Université Paul Sabatier, Bât. L4, CHU Ranqueil, Toulouse cedex 4, France

#### **ABSTRACT**

Type 2 diabetes mellitus (T2DM) is the most prevalent disease and becoming a serious public health threat worldwide. It is a severe endocrine metabolic disorder that has the ability to induce serious complications in all kinds of organs. Although mechanisms of anti-diabetics have been described before, we focus here on the cellular and physiological mechanisms involved in the modulation of insulin and glucose blood levels. As obesity and inflammation are intimately associated with the development of T2DM, their possible relationships are also described. The effects of gut microbiota on insulin resistance have been recently investigated in clinical trials, and we discuss the potential mechanisms by which gut microbiota may improve glucose handling, especially via the metabolism of ingested phytochemicals. Among the historically supported effects of phytochemicals, their therapeutic potential for T2DM leads to consider these natural products as an important pool for the identification of novel anti-diabetic drug leads. This current research extends the descriptions of anti-diabetic effects of plants that are used in traditional medicines or as nutraceuticals. The objective of the present review is to make a systematic report on glucose metabolism in T2DM as well as to explore the relationships between natural phytochemicals and glucose handling.

#### **KEYWORDS**

Type 2 diabetes; metabolic pathways; gut microbiota; phytochemicals; new therapies

#### Introduction

Diabetes mellitus is a well-known public health issue, affecting 415 million people, causing around 5 million deaths and accounting for 14.5% of all-cause mortality worldwide in 2015 (International Diabetes Federation 2015). The World Health Organization reports that the worldwide prevalence of diabetes is expected to increase up to 642 million by the year 2040, with many new cases of diabetes occurring in developing countries, especially in Asia. This increase in diabetes prevalence will inevitably lead to the intensification of diabetes-related complications such as retinopathy, neuropathy, and cardiovascular diseases. Bommer et al. (2017) reported that the cost of diabetes worldwide was 1.31 trillion US dollars, or 1.8% of the global gross domestic product in 2015. The global estimation of diabetes expenditures is predicted to increase by 490 billions in the next 20 years

(Zhang et al. 2010a). Type 2 diabetes mellitus (T2DM), also known as "non-insulin-dependent diabetes," accounts for 90% of all cases of diabetes. It is the most prevalent disease in many modern societies and is becoming a serious public health threat worldwide. It is a complex metabolic disorder characterized by insulin resistance (IR) and impaired islet  $\beta$ -cell function, which together result in an inability to supply sufficient insulin to meet the body's demands and eventual  $\beta$  cell loss. Individuals with T2DM experience difficulty in controlling their blood sugar level, which leads to high blood glucose level, glucose in the urine, and high blood insulin level.

The prevalence of T2DM has dramatically increased in Europe with 60 million people living with diabetes and 32 million more at risk (Schwarz et al. 2008). The Centers for Disease Control and Prevention reported diabetes mellitus

CONTACT Christian Carpéné christian.carpene@inserm.fr linstitut des Maladies Métaboliques et Cardiovasculaires (I2MC), Institut National de la Santé et de la Recherche Médicale (INSERM U1048)/Université Paul Sabatier, Bât. L4, CHU Rangueil, Toulouse cedex 4, France; Jianbo Xiao Jianboxiao@yahoo.com Institute of Chinese Medical Sciences, State Key Laboratory of Quality Control in Chinese Medicine, University of Macau, Macau SAR, China. Color versions of one or more of the figures in the article can be found online at www.tandfonline.com/bfsn.

Supplemental data for this article can be accessed at https://doi.org/10.1080/10408398.2018.1501658.

affected 29.1 million Americans, or 9.3% of the population (Crawford 2017). Even more, the prevalence of T2DM in western countries reached an epidemic level, but which is worse in Asia (Kong et al. 2013). In Asia, rapid economic development in many countries has driven a great increase in diabetes prevalence in the recent decade. The prevalence of T2DM is rising, with many Asian countries featured in the top 10 countries with the highest numbers of persons with diabetes. Urbanization is linked to reduced physical activity, increased obesity rates, and a dietary shift toward more refined carbohydrates and increased fat intake (Ning et al. 2009; Ramachandran et al. 2008). Worldwide, China, and India are the top two countries with the most number of individuals suffering from diabetes, with Indonesia and Japan in seventh and ninth place, respectively (International Diabetes Federation 2015). The prevalence of diabetes in South East Asia is also expected to increase by 70% in the next 20 years. Risk scores derived in Caucasian populations might not perform well in Asian populations as there are different biological factors involved in the development of diabetes. Compared to Caucasians, the onset of diabetes in Asians occurs at lower body mass index (BMI) levels and at younger ages (Hu 2011). South Asians also experience early declines in  $\beta$ -cell function, as well as with more insulin resistant and a younger age of diabetes onset compared to other ethnic groups (DECODE Study Group 2003; Gujral et al. 2013). Studies have shown that Asian ethnicities have a 2- to 4-fold risk of developing T2DM compared to Caucasian ethnicity (Sacks et al. 2012; Urquia et al. 2011). In addition, Asians with T2DM are especially vulnerable to renal injury when compared to Caucasians (Kong et al. 2013). The waist-to-stature ratio is more strongly associated with T2DM than BMI in most Asian populations (He et al. 2009).

The mechanisms for this increased risk are likely a combination of both genetic and environmental factors (Tutino et al. 2014), including the fact that obesity may have a greater effect on IR in these populations compared to Caucasians (Retnakaran et al. 2006). More important, genetic factors play a crucial role in the pathogenesis of T2DM in the Asian population. A study reported that Asian Indians are excessively IR compared with Caucasians (Abate and Chandalia 2001). Recently, an excessive maternal transmission of T2DM was identified among Asian Indians (Chaithri et al. 2012). On the other hand, many environmental factors such as diet, lifestyle, are associated with the risk of T2DM in Indians (Ramachandran et al. 2001), as well as in other Asian populations such as Chinese. The high consumption of white rice, especially in East Asian, is significantly associated with a higher risk of T2DM (Hu et al. 2012).

The gut microbiota is essential for the development and regulation of host metabolism. The intestinal mucosal surface protects the host from pathogenic invasion; while it is tightly regulated with regard to its permeability and can influence the systemic energy balance. Consumption of diets high in sugar influences the microbiota composition and leads to an imbalanced microbial population in the gut. It has been hypothesized that the gut microbiota could be part

of a mechanistic link between the consumption of unbalanced diets and T2DM (Qin et al. 2012).

While the causes of T2DM are still not completely understood, it is generally believed that T2DM results from both genetic and environmental factors. In recent years, great attention has being paid by the scientific community to the beneficial effects of phytochemicals on this disease (Cao et al. 2018; Carpéné et al. 2015; Loizzo et al. 2016; Vinayagam, Xiao, and Xu 2017; Xiao et al. 2015; Xiao and Högger 2015; Xiao 2017). The current review is to summarize the advance on the glucose metabolism by natural phytochemicals in T2DM, as well as to explore the relationship among metabolic pathways, gut microbiota, obesity, and inflammation. How microbial metabolites of the dietary phytochemicals modulate host glucose metabolism is also discussed.

# What should reproduce the anti-diabetics: insulin mechanisms of action?

There are two main pathways for body to adjust the blood glucose: one by increasing the circulating levels of insulin and another by boosting the non-insulin-dependent glucose metabolism. Insulin secretion by  $\beta$ -cells of pancreatic islets plays one of the most important roles in our body for adjusting blood glucose and is essential for insulin-regulated glucose metabolism. One of the most rapid actions of insulin is to increase glucose uptake via the recruitment of the insulin-regulated glucose transporters (GLUT) in cells having functional insulin receptors, insulin signaling cascade and GLUT type 4 (GLUT4) carriers. By the way, insulin reduces blood glucose levels by removing it from the circulation. Insulin also triggers glycogen synthesis, glycolysis, and inhibition of hepatic glucose production, and at last regulates cell proliferation, apoptosis, and autophagy (Aikawa et al. 2000; Kane et al. 2002; Xing et al. 2015; Yamaguchi and Otsu 2012).

Insulin is not the sole glucoregulatory hormone, since also contributing to the glucose homeostasis are glucagon and amylin produced by the endocrine pancreas, and the incretins produced in the intestine: glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic peptide. Other hormones also participate, such as growth hormone, epinephrine, cortisol, and the atrial natriuretic peptides, which have recently shown to increase glucose uptake in GLUT4-containing tissues (Coué et al. 2018). However, this review will examine only the major metabolic pathways that have been researched and reported for explaining the mechanisms of anti-diabetics by insulin-like actions (Fig. 1).

#### IRS/PI3K/AKT-GLUT4 pathway activators

Insulin receptor substrate 1 (IRS-1) and IRS-2, the two major substrate proteins generated by phosphorylation of Ser/Tyr in insulin receptor, bind to and activate the phosphatidylinositol 3-kinase (PI3K). The activation of PI3K is a key step of insulin-induced glucose transport (Tanti et al. 1994). In some extent, the protein p85 sub-unit of PI3K can

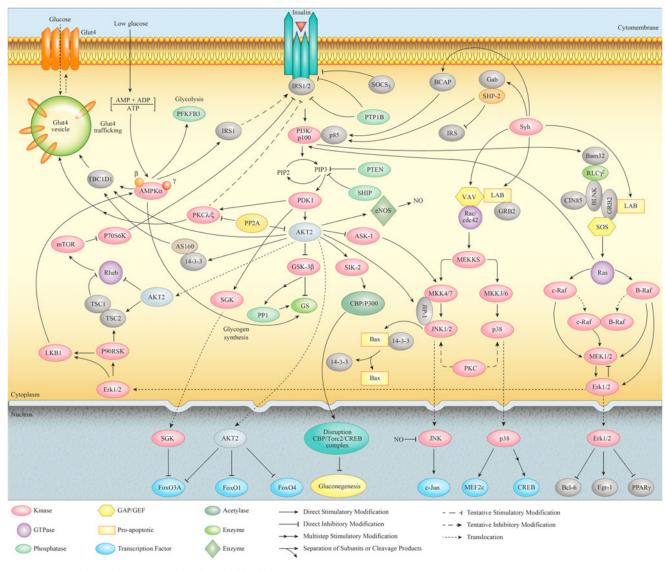


Figure 1. Main signaling pathways recruited to adjust the blood glucose.

improve the IR. The generation of phosphatidylinositol 3,4,5-trisphosphate, which can lead to activation of the three known AKT (protein kinase B) isoforms by pyruvate dehydrogenase kinase, isozyme 1 (PDK1), is accompanied by the activation of PI3K. AKT is by phosphorylation of Thr308 by PDK1 and is a key pleiotropic kinase that affects insulin function on glucose metabolism. It can deactivate glycogen synthase kinase (GSK) and inhibit some pro-apoptotic factors, such as FoxO3A, FoxO1, and FoxO4 in cell nucleus (Paradis and Ruvkun 1998), and cytoplasmic GSK- $3\beta$  with the development of cell (Mora et al. 2005). Meanwhile, it can also activate the translocation of GLUT4 by phosphofructokinase (Kadowaki et al. 2012; Kahn and Saltiel 2011; Manning and Cantley 2007). At the cell surface, GLUT4 permits the facilitated diffusion of circulating glucose down its concentration gradient into muscle and fat cells. Within cells, glucose is rapidly phosphorylated by glucokinase in the liver and hexokinase in other tissues to form glucose-6-phosphate, which then enters glycolysis or is polymerized into glycogen. Glucose-6-phosphate cannot diffuse back out of cells, which also serves to maintain the concentration gradient for glucose to passively enter cells (Watson et al. 2004). Moreover, GLUT4 finalizes the IRS1/2/PI3K/AKT signaling pathway, as it is the insulin-regulated glucose transporter.

# JNK/MAPK/ERK pathway

The mitogen-activated protein kinase (MAPK) signaling pathways, including c-Jun N-terminal kinase (JNK) and extracellular signal-regulated kinases (ERK) pathway, are among the more altered pathways in models of diabetes and obesity (Chen et al. 2018). The JNK signaling has been shown to contribute to a variety of pathological processes associated with diabetes, obesity, heart disease, and cancer (Kane et al. 2002). The molecules JNKs, key members of MAPK family, are named after their capacities to phosphorylate and activate the Jun family of AP-1 transcription factors (Chang and Karin 2001; Hibi et al. 1993). Compared with classical MAPK/ERK signaling, JNK pathway is more closely related to glycometabolism than lipid metabolism. The JNK/MAPK pathway comprises a sequential three-tiered

kinase cascade. An upstream MAP3K (MEKKs/ASK1) phosphorylates and activates the MAP2K (MKK4, MKK7, MKK3, or MKK6), which then regulates the downstream MAPKs (such as JNKs/p38 MAP kinase) (Chang and Karin 2001; Meloche and Pouysségur 2007). The ASK1 could lead to negative regulation of IRS1 and be deactivated by AKT2. MEKKs are activated by Vav proteins, being members of the guanine nucleotide-exchange factors which are GDP/GTP exchange factors for Rho/Rac GTPases. Vav is activated by tyrosine phosphorylation of Syh and SHP-2, which can directly inhibit phosphorylation of IRS. Small G-protein can be distributed into Rho, Rac, and Cdc42 for its function. In other words, the activation of MEKKs is accompanied with activation of regulators of cell cycle or cell reorganization such as Rac and Cdc42.

Advances in genomics and molecular genetics have revealed that the extracellular signal-regulated kinase (ERK) signaling pathway is known as MAPK pathway and a key signaling cascade for modulating multiple cellular functions by phosphorylating and inducing its downstream targets (Chen et al. 2001). ERK/MAPK pathway plays an important role on diabetes and malignancies by the regulation of cell differentiation, proliferation, growth, apoptosis, gene expression, and others (Degen et al. 2012; Mandal, Becker, and Strebhardt 2016; Mebratu and Tesfaigzi 2009; Reddy et al. 2003; Zhang and Liu 2002). The SHP-2 and Syh of Erk/ MAPK pathway as initial signaling is the same with JNK/ MAPK pathway, which initiates the formation of a "signalosome" composed of the tyrosine kinases, GRB2/ BLNK-related adaptor proteins, signaling enzymes such as PLCγ2, PI3K, and Vav, and small GTPases such as SOS and Ras (Goodnow et al. 2010; Harwood and Batista 2010). SOS is involved in Ras signaling activation and also acts as a guanine nucleotide exchange factor for Rac to transduce signals from Ras to Rac. In addition, the Ras GTPase subfamily plays a key role in this pathway and activates the MEKK1/2 by phosphorylation of B-Raf or c-Raf. Furthermore, MEK1/2 can activate ERK kinases (ERK1/2), which inhibits Bcl-6 and PPARy or induces Egr-1 DNA-binding activity in the nucleus.

#### IRS1/AKT/mTOR-AMPK signaling pathway

IRS1/AKT/mTOR signaling pathway represents a key pathway for genetic variation, diabetes, and obesity by cell growth control and autophagy inhibition in cytoplasm (Ganley et al. 2009; Magnuson et al. 2012). PI3K first converts phosphatidylinositol-4,5-bisphosphate (PIP2) into phosphatidylinositol-3,4,5-trisphosphate (PIP3), which activates AKT2 via PDK1 kinase. After that, the activated AKT2 inactivates the conserved serine/threonine protein kinases mTOR (mammalian target of rapamycin) by Rheb-GTPase, which is controlled directly or indirectly by AKT2, or indirectly through the control of tuberous sclerosis complexes (TSC1/TSC2). Finally, mTOR inhibits ribosomal protein S6 kinase (p70S6k), which intensely inhibits phosphorylation of IRS1 by the insulin receptor (Yin et al. 2017). However, AMP-dependent protein kinase (AMPK) signaling is

opposite to IRS1/AKT/mTOR pathway and gets in touch with each other by Erk1/2 in cytoplasm. AMPK has been identified as a critical positive regulator of autophagy, especially as an emerging drug target for T2DM and the metabolic syndrome (Hardie 2011; Zhang et al. 2009). AMPK not only directly activates IRS1 and participates in blood glucose metabolism, but also induces autophagy by inhibition of AKT/mTOR pathway. Since under low-glucose conditions, the increase in AMP/ATP ratio activates AMPK, this tends to stimulate hexose uptake.

# The major tissues involved in the regulation of glucose homeostasis

T2DM is characterized by the increase in blood glucose levels, and such excess of glucose alter in a non-enzymatic manner the circulating proteins by glycation. Moreover, T2DM is a non-contagious and severe endocrine metabolic disorder that induces serious complications in various organs (Su et al. 2017), either as a consequence of excessive glucose or in response to be altered levels of hormones and signaling molecules that impact all these organs toward an integrated function. Indeed insulin is increased in T2DM, leading to a state called hyperinsulinism. This aims to stimulate glucose uptake by the so-called insulin-responsive tissues (mainly skeletal and cardiac muscles as well as adipose tissues). Unfortunately, hyperinsulinism is often accompanied by IR, a state of weak responsiveness of the target tissues, thereby the increased hormone levels do not achieve to elevate sufficiently the glucose uptake by peripheral tissues and cannot efficiently elevate the rate of glucose removal from the circulation. Instead, chronically elevated insulin results in increased hepatic glucose output and de novo

In insulin-sensitive organs, such as skeletal muscle and adipose tissue, epigenetic modifications might be important in the pathogenesis of T2DM, as the changes alter the function profile of genes influencing the control of glucose metabolism (Fig. 2). T2DM also develops an inability of the cells to utilize the secreted insulin for helping in glucose management: then corresponds to glucose intolerance and IR (Catalogna et al. 2016; Gugliucci 2016; Sharabi et al. 2015). It can cause severe secondary complications such as liver dysfunction, kidney failure, heart attack, and nerve damage (Hoshino, Hoshino, and Nishino 2016; Jin et al. 2015; Lastra et al. 2010; Manna et al. 2010). The life-threatening T2DM associated complications include long-term damage, dysfunction, and failure of the vital organs such as eyes (retinopathy), kidneys (nephropathy), peripheral nerves (neuropathy), and heart vessels (cardiovascular diseases).

Insulin normally inhibits gluconeogenesis and initiates glucose uptake in the muscle and adipose tissues for the maintenance of normal blood glucose levels. Adipose tissue acts as a critical metabolic organ and produces a number of hormones and cytokines such as adiponectin, leptin, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin (IL)-6, and MCP1, that in turn modify the functions of other cells in the organism (Kristensen et al. 2015; Zhang et al. 2016). Adiponectin

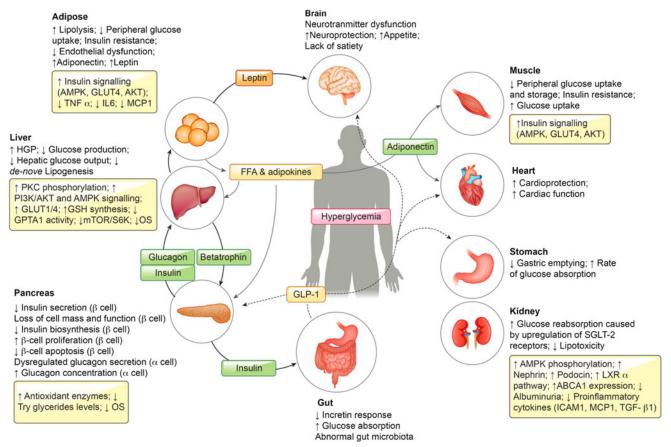


Figure 2. Tissues involved in the regulation of glucose homeostasis.

secreted from adipose tissue exerts insulin-sensitizing and anti-atherogenic activities. However, in adipose tissue, hyperglycemia enhances lipolysis and endothelial dysfunction. Lastly, in the digestive tract, high blood glucose levels inhibit incretin response and lead to glucose absorption and abnormal gut microbiota (Zappas et al. 2017).

Pancreas plays a critical role in glucose homeostasis since it is its endocrine part that secretes the glucose-lowering hormone insulin and its opponent glucagon (Grgurevic et al. 2016). Insulin decreases blood glucose levels. However, long-term high blood glucose levels impair insulin secretion by pancreatic  $\beta$ -cells and a loss of cell mass and insulin biosynthesis capacity can occur (Keane and Newsholme 2014; Mohan et al. 2015; Rutter et al. 2015). Similarly, sustained hyperglycemia can lead to dysregulated glucagon secretion by pancreatic α-cells and elevated glucagon concentration (Song et al. 2014).

Liver also plays an important role in ameliorating T2DM via participating in PKC phosphorylation, P13K/AKT and AMPK signaling regulation (Steinbrenner 2013), GLUT1/4 and glutathione (GSH) synthesis, as well as decreasing glutamic-pyruvic transaminase activity (GPTA), mTOR/S6K, and oxidative stress (Cordero-Herrera et al. 2015). While most tissues are unable to utilize the excess of secreted insulin to affect glucose metabolism, the resistance in liver produces increase in hepatic glucose production, high hepatic glucose output, and de novo lipogenesis (Catalogna et al. 2016; Gugliucci 2016; Sharabi et al. 2015).

In addition, hyperglycemia promotes lipid accumulation and glucose reabsorption caused by upregulation of sodiumdependent glucose transporters 2 (SGLT2) receptors in the diabetic kidney (Vallon and Thomson 2017; National Kidney Foundation 2012). Furthermore, it has been proven that proinflammatory cytokines such as MCP-1 and TGF- $\beta$ 1 play a key role in the development of diabetic nephropathy (Du et al. 2015). Persistent hyperglycemia not only promotes the rate of glucose absorption and slows gastric emptying in stomach, but also alters cardiac function (Chen et al. 2017; Maji and Samanta 2017). Similarly, IR induces neurotransmitter dysfunction (Hiriart et al. 2014), declines neuroprotection, increases appetite, and reduces satiety (João et al. 2016).

## Type 2 diabetes, obesity, and inflammation

Obesity is strongly associated with an increased risk of T2DM and cardiovascular disease. All these conditions are also now recognized as having an inflammatory component (Fig. 3). A number of cytokines and inflammatory signaling pathways have been shown to be involved in the development of T2DM, thus, increased serum levels of several inflammatory biomarkers, including tumor necrosis factor-α (TNF- $\alpha$ ), C reactive protein (CRP), high molecular weight adiponectin, and interleukin (IL)-6, produced at least in part by white adipose tissue in obese subjects, have been reported. The potential role of inflammation in obesity

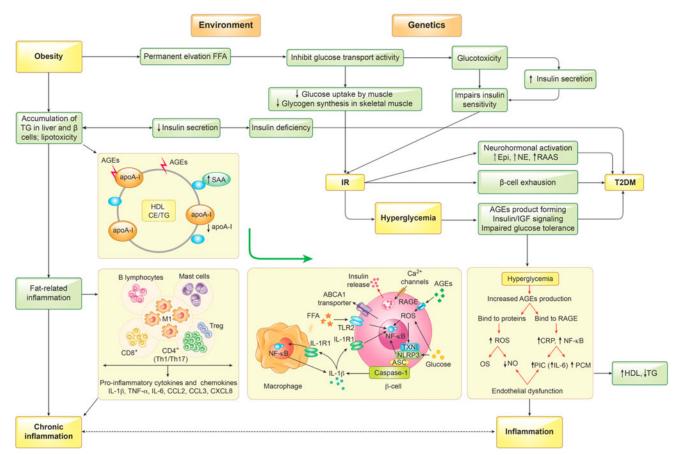


Figure 3. The relationships among type 2 diabetes, obesity, and inflammation.

complications is offering further insight into the relationship between T2DM and cardiovascular disease, and it has led to a greater interest on specific therapeutic targeting. Accordingly, poor glycemic control is positively correlated with levels of inflammatory cytokines such as IL-6 and IL- $1\beta$  in the circulating blood stream (Calle and Fernandez 2012). However, only scarce studies have also shown that the benefit of anti-inflammatory medication for the treatment of T2DM (Weisberg et al. 2008). In this context, there is evidence that a hyperglycemic state increases blood-brain barrier permeability (Hawkins et al. 2007), allowing these inflammatory cytokines to reach neuronal environment (Perry et al. 2017).

# Effects of gut microbiota and metabolic endotoxemia/bacteremia on diabetes

#### Altered gut microbiota composition in diabetes

The endogenous gut microbiota is considered to be a "forgotten organ" participating in the whole-body metabolism (O'Hara and Shanahan 2006). There are approximately 10<sup>14</sup> bacteria belonging to more than 1,000 phylotypes in human gut (Whitman et al. 1998). Although the composition of human gut microbiota shows great variation among individuals, most bacteria belong to six well-known bacterial divisions/phyla: Firmicutes, Bacteroidetes, Proteobacteria, Actinobacteria, Fusobacteria, and Verrucomicrobia (Eckburg et al. 2005; Lozupone et al. 2012). The Bacteroidetes and

Firmicutes account for 60-90% of the consortium (Neish 2009). Healthy human gut mainly harbors anaerobic bacteria, the number of which is far more than the aerobic and facultative anaerobic bacteria (Sommer and Bäckhed 2013). They comprised predominant obligatory anaerobes belonging to the genera Bacteroides, Eubacterium, Clostridium, Ruminococcus, Peptococcus, Peptostreptococcus, Bifidobacterium, and Fusobacterium. The following subdominant facultative anaerobes are Escherichia, Enterobacter, Enterococcus, Klebsiella, Lactobacillus, and Proteus (Guarner and Malagelada 2003). However, human gut harbors limited numbers of archaea (mainly Methanobrevibacter smithii), eukaryotes (mainly yeasts), and viruses (mainly phages) (Reyes et al. 2010). Several experimental reports have suggested that diet plays a dominant role in forming and changing the bacterial community composition of human gut (Hildebrandt et al. 2009). Moreover, the compositional changes in gut microbiota represent an etiological factor in the development of both IR and T2DM. Recent studies have shown that the gut microbial inhabitants have an influence on the onset of metabolic diseases such as obesity and diabetes. Turnbaugh et al. (2006) confirmed that obesity is associated with the shift in relative abundance of the two dominant bacterial phyla, the Firmicutes and the Bacteroidetes. Specifically, a larger proportion of Firmicutes and relatively lower abundance of the phylum Bacteroidetes were observed in obese individuals, suggesting that both of them were correlated with energy intake and adiposity (Murphy et al. 2010; Ravussin et al. 2012).

In contrast to obesity, T2DM-associated microbial dysbiosis is comparatively modest. The gut microbiome in T2DM is characterized by lower levels of short-chain fatty acids (SCFA)-producing bacteria (Eubacterium rectal, Faecalbacterium prausnitzii, Roseburia intestinalis, etc.) and higher levels of known or potential opportunistic pathogens (Clostridium hathewayi, C. ramosum, and Eggerthella lenta) (Qin et al. 2012). Pioneering studies have shown that functional changes in gut microbiota contribute to the increases in plasma glucose concentrations (Clemente et al. 2012). Influence of gut microbiota on the development of diabetes is summarized in the present review (Fig. 4).

# Gut microbiota and host energy balance and storage

The gut microbiota has been reported to participate in the host metabolic functions and energy balance by fermenting undigested carbohydrates, which activate host satiety and decrease food intake (Chambers et al. 2015). Undigested dietary carbohydrates are important sources of energy for human colon microbiota, which can utilize nearly all of the major plant and complex glycans, such as two members of the Bacteroides fragilis group: Bacteroides thetaiotaomicron and B. ovatus (Martens et al. 2011). Interestingly, Bifidobacteria are dominant and prevalent members of the (early) microbiota having access to glycans in the gut through mutualistic cross-feeding or resource-sharing activities, indicative of a "social behavior". The SCFAs

(principally acetate, propionate, and butyrate) modulate the intestinal barrier functions and can escape from the gut to influence systemic health as well. Being bacterial fermentation products, these SCFAs can act in colonic epithelium (butyrate) or be absorbed to act further in peripheral tissues (acetate and propionate) (Lin et al. 2012). SCFAs activate the gut hormones GLP-1 and peptide YY (PYY) through Gprotein coupled receptor 43/41 (GPR 43/41), also known as free fatty acid receptor 2/3 (FFAR 2/3) (Kaji et al. 2014). Several seminal research studies showed that GLP-1 and PYY suppress appetite and energy intake (Lin et al. 2012; Nøhr et al. 2013; Tolhurst et al. 2012). GLP-1 is a gut hormone, secreted from the intestine in response to meal ingestion, which stimulates insulin secretion and inhibits glucagon release in a dose-dependent fashion (Mazidi et al. 2017). Kjems et al. (2003) reported that GLP-1 led to increased insulin secretion in T2DM patients. The activated GPR 43 inhibits fat accumulation in adipose tissue by suppressing insulin signaling in adipocytes (Kimura et al. 2013). The same GPR 43 receptor also participates in regulation of inflammatory responses in immune cells (Maslowski et al. 2009). The impacts of microbiota on IR and metabolic syndrome were reported in males who received an autologous fecal microbiota transplant, or an allogenic transplant from healthy donors. Subjects showed a significant improvement in peripheral sensitivity to insulin corresponding to an increase in microbial diversity in the bowel and butyrateproducing bacteria (Vrieze et al. 2012).

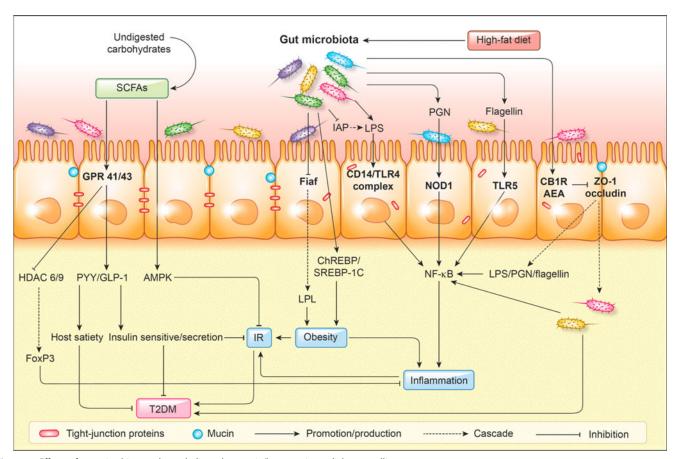


Figure 4. Effects of gut microbiota and metabolic endotoxemia/bacteremia on diabetes mellitus.

Butyrate and acetate were reported to elevate (AMPK) activity (Gao et al. 2009; Sakakibara et al. 2006), which then stimulates glucose uptake and fat oxidation as well as inhibits fatty acid and glycogen synthesis (Inoki et al. 2012). Moreover, acting as a histone deacetylase 6/9 (HDAC 6/9) inhibitor, butyrate was proved to promote FoxP3 expression and both number and function of FoxP3 (+) Treg cells (Beier et al. 2012; Tao et al. 2007). On the other hand, the gut microbiota promotes fat storage by suppressing the fasting-induced adipose factor (Fiaf), which is a circulating lipoprotein lipase inhibitor in the gut epithelium (Bäckhed et al. 2007). The resulting increased lipoprotein lipase activity was associated with the microbiota-induced enlargement of adipose tissue (Bäckhed et al. 2004). The gut microbiota also markedly enhances the hepatic triglyceride synthesis by activating the gene expression of the two key transcriptional factors: carbohydrate response element binding protein (ChREBP) and sterol response element binding protein 1c (SREBP-1c) (Shen et al. 2013). ChREBP and SREBP-1c are both critical for hepatocyte lipogenesis due to their independent effects in mediating glucose signaling and insulin action in liver, respectively, thus improving glucose absorption and insulin levels (Dentin et al. 2004). Hence, the influences of gut microbiota on host energy balance and storage represent a possible pathway linking gut microbiota and obesity and diabetes. The proper modification of diet resulting in functional rearrangement of microbiome could be considered to be treatment of first stages of T2DM (Zhao et al. 2018b).

## Metabolic endotoxemia/bacteremia, gut barrier function and diabetes

Gut microbiota-derived metabolic endotoxemia is reported to participate in the onset and progression of inflammation and metabolic complications. Several studies have demonstrated that dietary fat facilitates the development of metabolic endotoxemia, such as increased plasma levels of bacterial lipopolysaccharide (LPS) that is a component released from the cell wall of Gram-negative bacteria (Amar et al. 2008). LPS is one of the pathogen-associated molecular patterns (PAMPs) that also include peptidoglycan (PGN), flagellin, lipoproteins, and even more factors. The PAMPs are recognized by the pattern-recognition receptors (PRRs), including the Toll-like receptors (TLRs) and the Nod-like receptors (NLRs). The interaction between the PRRs and the PAMPs induces cytokine and interferon production, which triggers the proinflammatory signaling cascades in body peripheral tissues of the body (Icaza-Chávez 2013).

LPS and PGN molecules bind to TLR4 and NOD1 receptors, respectively, thus participating in diet-induced inflammation and IR (Cani et al. 2007; Schertzer et al. 2011). TLR5 is expressed on the apical and basolateral surface of intestinal epithelium, which detects bacterial flagellin from both Gram-positive (e.g., Listeria monocytogenes) and Gramnegative bacteria (e.g., Salmonella typhimurium) (Hayashi et al. 2001). The activation of TLR5 strongly promotes nuclear factor NF-κB production and driven inflammatory

responses (Letran et al. 2011). The gut mucosal surface is the key site of pathogenic bacteria and metabolic endotoxemia entry into the body and the initiation of metabolic endotoxemia. The endocannabinoid lipid anandamide (AEA) and its receptors cannabinoid receptors 1 (CBR1) in endocannabinoid (eCB) system are involved in the regulation of gut barrier function during obesity (Alhouayek and Muccioli 2012). Muccioli et al. (2010) found that gut microbiota regulates the CB1R expression and AEA content in the intestine. Furthermore, such endocannabinoid (eCB) system regulates epithelial permeability through the distribution and localization of tight-junction proteins (Muccioli et al. 2010). Additionally, mucus layer has long been recognized as an important ingredient providing protective gut barrier for the host. Muc2 is the major glycosylated mucin produced in the small and large intestine (Johansson et al. 2008). Normal gut microbiota can stimulate the secretion of Muc2 mucin in the goblet cells to ensure the integrity of the mucous layer structure and thus maintain gut barrier function (Johansson and Gordon 2011). Altogether, improvement of gut barrier integrity reduces metabolic endotoxemia and bacteremia, as well as lowers inflammation and glucose intolerance. Intestinal alkaline phosphatase (IAP) has been recognized to play a crucial role in LPS detoxification by dephosphorylating and detoxifying the phosphate residues of LPS (Bates et al. 2007). More interestingly, expression of IAP has been shown to be regulated by the gut microbiota and its activity could be increased by the diet (Lallès 2010). Enhanced IAP activity may, therefore, contribute to the reduction of metabolic endotoxaemia and gut permeability in T2DM and obesity.

Future physiological studies are needed to elucidate how the intestinal branched chain amino acids and other amino acids enter the bloodstream and from which intestinal location they are absorbed. Furthermore, investigations of how dietary changes alone or in combination with microbial or pharmacological interventions may impact the microbiome. Gut microbiota are also responsible for the extensive metabolism of phytochemicals such as polyphenols and hence contribute largely to the oral bioavailability of phytochemicals and thereby influences their potential antidiabetic activities (Eid et al. 2017).

Probiotics and/or prebiotics could be a promising approach to improve insulin sensitivity by favorably modifying the composition of the gut microbial community, reducing intestinal endotoxin concentrations, and decreasing energy harvest. The anti-diabetic effects of probiotics include reduction of proinflammatory cytokines via a NF-κB pathway, reduced intestinal permeability, and lowered oxidative stress. The ability to produce SCFAs is a feature shared by many different probiotic taxa. SCFAs play a key role in glucose homeostasis through multiple potential mechanisms of action in tissues: (1) insulinotropic and satiety effects mediated by gut hormones; (2) GLP-1 and peptide YY production; (3)  $\beta$ -cell-protective effect due to reduced oxidative stress; (4) decreased pro-inflammatory cytokines; (5) antilipolytic activity; and (6) enhanced insulin sensitivity via GLUT4 through the up-regulation of AMPK signaling

(Favaretto et al. 2014). Moreover, the antidiabetic effect has been shown after administration of probiotics containing certain strains of Lactobacillus, with a concomitant reduction in endotoxemia (Li et al. 2017).

The activation or suppression of the TLRs by microbial signals can dictate the tone of the immune response and thereby contribute to the regulation of the energy homeostasis (Spiljar et al. 2017). Since the microbiota-derived signals influence functions of distant organs and can change susceptibility to metabolic diseases, they can be considered as instrumental for the treatment of metabolic chronic diseases.

### Bioactive natural products for diabetes mellitus

Several plant and mushroom species, including a number of those have been used in traditional Chinese medicine, have now been shown to exert anti-diabetic effects. The future potential of the bioactive natural products used in diabetes treatment will be based on the modification of structures of biologically active compounds (leads), in order to obtain safer, more selective, or more bioavailable molecules, which is a primary requirement for drug development. New approaches for the identification and characterization of natural products are being developed that may address some of challenges related to the development of plant-based therapeutics. Resupplying from the original plant species is very unfeasible to meet the huge market demands upon commercialization of a natural product, and alternative resupply approaches are being developed that rely on biotechnological production or chemical synthesis. Over 9,000 herbs have known medicinal applications among various cultures and countries. Many plants have been investigated for their beneficial use in different types of diabetes and reported in numerous scientific journals. However, understanding the scientific material basis of traditional Chinese medicine herbal formulas at the molecular level remains a considerable challenge. Examples of these medicinal plants are described below since they play a key role in managing diabetes.

Scientists have discovered that plants have great efficacy to produce numerous bioactive molecules dealing with the problem of diabetes mellitus in recent years (Supplementary Table 1). These natural products consist of large quantities of bioactive compounds including phenolic compounds, oligo-/polysaccharides, terpenoids, curcumin, xanthones, thiosugar derivatives, tannins, chalcones, phenolic acids, alkaloids, and amino acids (Goto et al. 2010). And it is crucial to understand the mechanism behind the biological effects of these compounds for the prevention and treatment of T2DM.

Flavonoids are a type of polyphenols that includes a great number of compounds showing important differences in their structure. They can limit the hyperglycemic response following food intake. For example, apigenin regulates hyperglycemia by increasing the serum insulin levels while rutin improves IR and increases glucose uptake (Kappel et al. 2013); and quercetin mitigates the diabetes by stimulating insulin secretion and inhibiting aldolase reductase in

diabetic patients (Bahmani et al. 2014). In addition, resveratrol as a natural biologically active polyphenol compound present in different plant species has beneficial effects in relation to diabetes (Szkudelski and Szkudelska 2015). Thus, it appears that, alongside their widely recognized antioxidant capacity, other properties not totally characterized confer to flavonoids and anthocyanins their capacity to mitigate diabetes.

Polysaccharides are a class of important compounds preventing T2DM. Ganoderma lucidum polysaccharides (GLP) decreased plasma insulin concentration and reversed HFDinduced systemic IR (Xu et al. 2017). GLP ameliorated lowgrade chronic inflammation, induced lipolysis in adipose tissues, and decreased plasma triglyceride. GLP also regulated composition of gut microbiota implicated in obesity and T2DM development (Chang et al. 2015; Xu et al. 2017). So far, there are six possible mechanisms by which polysaccharides mitigate T2DM: (1) reducing carbohydrates decomposition and absorption; (2) restricting α-glycosidase enzymes in bowel; (3) scavenging free radicals and lipid peroxidation reduction; (4) elevating plasma insulin and declining pancreatic glucagon; (5) increasing hepatic glycogen and regulating inflammatory cytokines and gut microbiota composition; and thereby (6) restoring an adequate insulin sensitivity (Wang et al. 2017).

Plant terpenoids mitigate diabetes with almost the same efficacy as polysaccharides. The mechanisms of actions of terpenoids include reducing blood glucose level, increasing glycogenesis and decreasing glycogenolysis, as well as inhibiting aldose reductase. By these mechanisms, they can also achieve the same efficacy as the previously described compounds. As an example, curcumin is representative of the curcuminoids that can be extracted from the root of Curcumalonga. Curcumin has anti-carcinogenic and antioxidant effects (Duvoix et al. 2005; Shishodia et al. 2005). Although curcumin has been reported to inhibit glucose transporter activity (Green et al. 2014; Nabavi et al. 2015), such an action expected to impair glucose handling by peripheral tissues can instead reduce absorption of dietary glucose by direct inhibition of GLUT proteins in intestinal epithelial cells and contribute to a hypoglycemic effect (Gunnink et al. 2016).

Xanthones possess antidiabetic activity. The most known xanthonoid is mangiferin, which is a glucoside of norathyriol. It acts at least via two mechanisms such as decreasing resistance and/or increasing insulin sensitivity, and its glycoside moiety can impair several carbohydrate-metabolizing enzymes (Raut et al. 2016).

Thiosugar derivatives also show strong inhibitory activity against α-glucosidases. Tannins can produce denaturation of proteins and therefore, can trigger nonspecific inhibition of α-glucosidase, too. Indeed, condensed tannins can inhibit α-amylase and a higher inhibition can be achieved as the degree of polymerization increases. The antidiabetic effect of chalcones might be related to the anti-atherosclerotic activity of 2-hydroxy-4'-methoxychalcone, which has been reported to stimulate PPAR-y expression in human aortic smooth muscle cells. The group of chalcones could, therefore, be

considered as natural PPAR-y activators enhancing the cell sensitivity to insulin, somewhat reproducing the stimulation by the thiazolidinediones, the pharmacological agonists of reference for this nuclear factor.

The antidiabetic mechanism of alkaloids can be summarized by two axes: (1) repairing or stimulating proliferation of pancreatic  $\beta$ -cells, stimulating the secretion of insulin, increasing the sensitivity to insulin, decreasing resistance, increasing glycogenesis and inhibiting gluconeogenesis; (2) decreasing the level of glucogenic enzymes. Besides, S-allyl cysteine is a sulfur-containing amino acid found in garlic that has been found to promote glucose uptake and metabolism. Furthermore, it can also facilitate insulin secretion, thereby decreasing blood glucose levels.

The potential antidiabetic application of phenolic acids has been proposed when observing that they elevate glucokinase activity and accumulate glycogen in the liver. In addition, the antidiabetic properties of gallic acid rely on its ability to induce glucose uptake by stimulating GLUT4 translocation. Recent literature supports that polyphenols can inhibit pancreatic  $\alpha$ -amylase or  $\alpha$ -glucosidases, and this adds an additional mechanism to their hypoglycemic effect. Like gallic acid, another natural phenolic acid, ellagic acid, has been shown to inhibit glycogen phosphorylase and therefore to prevent hepatic glucose output when the liver has its glycogen stores intact (Kyriakis et al. 2015). Hence, the beneficial effects of dietary polyphenols rely on their modification to host digestive enzymes. It the case of the above-mentioned phenolic acids, their antidiabetic properties can be either increased (if active metabolites are generated) or diminished (if metabolites are inactive) after the digestion process. In fact, it has been demonstrated that ellagic acid undergoes a series of bacteria-mediated transformations leading to the appearance of urolithins (Cerda et al. 2005; Espin et al. 2013). Such metabolism probably limits the available amount of phytochemical able to act in the organism, but more importantly, once ingested, ellagic acid and ellagitannins generate among members of urolithins, urolithin A, recently shown to increase muscle function in rodents and to potentially delay age-related dysfunctions (Ryu et al. 2016). Since, in the polyphenol-related studies, a large amount of interindividual variation can be observed in the microbial metabolism and absorption of certain polyphenols (Shortt et al. 2018), the task to characterize the major agents involved in the promising antidiabetic actions of phytochemicals will likely conduct to a personalized nutritional approach, consisting in giving a combination of agents working in synergism, as in the case of polyphenols (Carpéné et al. 2015), or aiming at providing both the phytochemical of interest together with prebiotic or probiotic supplements to facilitate the generation of the active metabolites.

In addition to the multiple actions of dietary polyphenols in the gut, phytochemicals such as fibers, and polysaccharides play a vital role in modulating gut microbiota (phylum Bacteroidetes, Firmicutes and Firmicutes/Bacteroidetes ratio, and genus Akkermansia, Bifidobacteria, Lactobacillus, Bacteroides and Prevotella) through SCFAs, bile acids (BAs),

and LPS (Lyu et al. 2017). The administrations of natural phytochemicals lead to increase the abundance of phylum Bacteroidetes, and genus Akkermansia, Bifidobacteria, Lactobacillus, Bacteroides, and Prevotella, while reduced phylum Firmicutes and Firmicutes/Bacteroidetes ratio in gut. Additionally, the consumption of functional oligosaccharides can reduce the risk of chronic diseases, such as cardiovascular disease, obesity, and diabetes mellitus (Zhao et al. 2018a). Oligofructose was reported to promote the growth of beneficial bacteria such as Bifidobacterium, Lactobacillus, and Bacteroides, and to benefit diabetic individuals (Dehghan et al. 2014). Xylooligosaccharide highly promotes the population of Bifidobacteria population in vitro and decreases the abundances of the genera Enterorhabdus, Howardella, and Slackia in pre-diabetic subjects (Yang et al. 2015). There are also evidence that chitosan oligosaccharides (COS) improve the disturbance in glucose metabolism and reverse the dysbiosis of gut microbiota in diabetic mice. In vitro studies suggest that COS might reduce the glucose transport in Caco-2 cells and promote glucose uptake of adipocyte cells (Meng et al. 2016). COS could reduce fasting blood glucose and increase insulin secretion in db/db mice (Kim et al. 2014). COS treatment improves the disturbance in glucose metabolism and reverses the dysbiosis of gut microbiota in diabetic mice by promoting Akkermansia and suppressing Helicobacter (Zheng et al. 2018). This was confirmed by decreased blood glucose level, reversed IR, enhancement of intestinal integrity, suppression of inflammation and inhibition of lipogenesis in epididymal adipose tissue of diabetic mice after treatment of COS. Natural phytochemicals interact with gut microbiome and alter the microbial metabolites including SCFAs, BAs, and LPS, which are correlated with T2DM.

#### Challenges and perspectives

Why does T2DM get progressively worse over time? What is the most effective way to slow down or to prevent this progress? The growing prevalence of obesity and T2DM results in an overload of patients at very high short-term cardiovascular risk. There is an increasing need for new options to treat diabetes at its early stage, especially T2DM, due to an ineffective control of its long-term evolution in patients.

#### The modification of natural products for medical use

Although the natural products are normally valuable lead compounds, they rarely can be directly used in clinical applications. From the standpoint of drug innovation, it is necessary to modify natural product structures, because the aim in generation of secondary metabolites by organisms is to protect themselves from natural enemies as well as the environment. The final aim of modifying natural products is to develop active compounds into medicines. Structural modifications are necessary to achieve this goal. The strategy is therefore to increase potency and selectivity, to improve physicochemical, biochemical, and pharmacokinetic

properties, to eliminate or reduce adverse effects, to simplify the structural complexity, including removal of redundant atoms and chirality while retaining activities, and to generate patentable compounds. Lastly, another interest of biotechnology for synthesizing phytochemical-like derivatives is justified when the amount of active substance required per patient during prevention or chronic treatment, requires the equivalent of large quantities of plants, at the risk of putting the concerned species in danger of disappearing from the current biodiversity. As an example, the natural product Oglucoside phlorizin is a well-documented, potent glucosuric agent that was subsequently shown to be a nonselective SGLT inhibitor. Because of its inhibition of SGLT1 with poor metabolic stability due to its susceptibility to  $\beta$ -glucosidase-mediated cleavage, as a lead compound, O-glucoside phlorizin was modified. Canagliflozin, simplifying structures from phlorizin, entered into clinical trials and was approved by the FDA in 2013 for the treatment of T2DM (Nomura et al. 2010).

#### MicroRNAs as pharmacological targets in diabetes

Recently, small noncoding RNAs with approximately 22nucleotides, of a novel class termed microRNAs (miRNAs), were found to play a key role as important transcriptional and post-transcriptional inhibitors of gene expression in fine-tuning the target messenger RNAs. It is predicted that over 30% of human genes are regulated by miRNAs. Several of them are implicated in the pathogenesis of T2DM and have become an intriguing target for therapeutic intervention. Recent data suggest that miRNAs play a direct role in insulin secretion, pancreatic islet development,  $\beta$ -cell differentiation, and indirectly control glucose and lipid metabolbeing involved in diabetes-associated thereby complications. The ubiquity of miRNAs in body fluids and their association with the disease pathogenesis have made them important players for prognosis, diagnosis and management of T2DM. MiRNAs are regarded to regulate insulin biosynthesis and secretion in pancreatic  $\beta$ -cells, insulin sensitivity in skeletal muscle and adipose tissue, as well as glucose and lipid metabolism in liver. The first evidence of miRNAs controlling the  $\beta$  cell activities was demonstrated by Poy and his colleagues (Poy et al. 2004). They demonstrated that miR-375 regulates secretory activities of  $\beta$ -cells. Its silencing increases glucose-stimulated insulin secretion in murine pancreatic  $\beta$ -cell lines and in isolated primary  $\beta$ -cells. MiR-375 knockout mice exhibited increased pancreatic α-cells, elevated plasma glucagon levels, gluconeogenesis and hepatic glucose output. Moreover, MiR-7a was recognized to be a negative regulator of adult  $\beta$ -cell proliferation by targeting various components in mTOR signaling pathway (Wang et al. 2013). In mouse models, the specific overexpression of miR-200 in  $\beta$ -cell induced their apoptosis and promoted severe T2DM under stress conditions by negatively regulating  $\beta$ -cell chaperone Dnajc3 or p58IPK and caspase inhibitor Xiap (Belgardt et al. 2015). Other findings have reported various miRNA signatures associated with T2DM, newly diagnosed cases and vascular complications.

Currently, it is supposed that specific miRNAs play a critical role in the development of diabetic vascular complications. Although there are numerous studies implicating the role of specific miRNAs in  $\beta$ -cell biology, they were mainly performed in vitro on cell lines only. Thus, it is of great necessity and urgency to validate those studies by in vivo approaches. There is also a need to obtain more data on miRNA expression in human samples for highlighting their potential roles in T2DM progression. However, pharmacological over-inhibition or overexpression by administration of miRNA mimics or miRNA inhibitors may potentially have off-target effects. More substantial research and standardization of techniques are required to determine the efficacy and feasibility of miRNAs as diagnostic tools to be routinely used as prognostic markers of T2DM and complications.

#### Novel therapies for type 2 diabetes mellitus

Numerous recent studies on novel antidiabetic drugs aimed at improving insulinopenia with fewer side-effects than the current insulinotropes (glucokinase activators, G proteincoupled receptor ligands, ultra-long-acting insulins). Novel approaches for reducing hyperglycemia have focused interest on gluconeogenesis inhibitors and even on new drugs with unusual therapeutic pathways, such as the GLP-1 analogues or agents that prevent the catabolism of incretins, or even inhibitors of the urinary glucose reabsorption (sodium-glucose co-transporter inhibitors). The recent tests of the blockade of other metabolic pathways seem to indicate that it is possible to affect T2DM by influencing energy metabolism, as in the cases for diacylglycerol acyl-transferase inhibitors and  $11\beta$ -HSD1 inhibitors. In addition, herbal drugs have previously been reported to be beneficial in hyperglycemia control worldwide, and about 800-1,200 herbs or their products have largely been used as anti-diabetic medicines to regulate the diabetes (Kavishankar et al. 2011; Zhao et al. 2018b). In contrast to conventional synthetic drugs, they are generally perceived as safe, harmless and without deleterious side effects such as gastrointestinal or cardiovascular complications. Therefore, herbal drugs can provide an alternative therapy for this medical challenge.

#### **Conclusions**

The growing prevalence of T2DM results in an overload of patients with very high short-term cardiovascular risk. Recent surveys have shown that most of the diabetic patients using chronic prescription medications are also associating herbal supplements or vegetal rich diets without understanding the risks of such combination. The co-administration of natural products along with conventional medicines is believed to induce a modified bioavailability of the prescribed drugs and, to a lesser extent, to change the fluxes of various metabolic pathways. Systematic studies are still necessary in order to unravel the roles of phytochemicals since even the physicians are not always aware of the risk of such interactions. Unless validated, natural phytochemicals



should be avoided as supplements for patients undergoing chemotherapy in order to avoid the risk of decreased availability. To pour more complexity, natural products may lead to an increase of the drug concentration when administered in short-term regimen, but may induce an increased metabolism and decreased effect after prolonged intake. Alongside the recommended prevention involving diet modifications, optimal treatment with the good combination of drugs and natural phytochemicals remains to be established. Since dysbiosis of the human gut microbiota impacts the serum metabolome and contributes to IR, and since there are the inter-individual variations of the microbiota, a personalized approach should be considered. Nevertheless, these emergent microbial targets have the potential to diminish IR and to reduce the incidence of common metabolic and cardiovascular disorders. They, therefore, deserve to be considered to contribute as an essential step in the modification of the ingested phytochemicals capable of either generating active metabolites or hampering the beneficial effect of the ingested active principles. The objective of the present study was to make a systematic review on glucose metabolism in T2DM as well as to explore, among metabolic pathways, the multiple relationships between gut microbiota, inflammation, diabetes and obesity.

#### **Abbreviations**

AEA Endocannabinoid lipids anandamide **AMPK** AMP-dependent protein kinase

Bas Bile acids BMI Body mass index

CBR1 Receptors cannabinoid receptors 1

ChREBP Carbohydrate response element binding protein

COS Chitosan oligosaccharides C reactive protein CRP eCB endocannabinoid

Erk Extracellular signal-regulated kinases

FFAR 2/3 Free fatty acid receptor 2/3 GLP-1 Glucagon-like peptide-1 GLUT4 Glucose transporter type 4 GPR 43/41 G-protein coupled receptor 43/41 **GPTA** Glutamic-pyruvic transaminase activity

Glutathione synthesis **GSH** GSK Glycogen synthase kinase HDAC 6/9 Histone deacetylase 6/9 Intestinal alkaline phosphatase IAP

II.-6 Iinterleukin-6 IR Insulin resistance

Insulin receptor substrate 1 IRS-1 IRS-2 Insulin receptor substrate 2 INK c-Jun N-terminal kinase LPL lipoprotein lipase

MAPK Mitogen-activated protein kinase MIN6 Murine pancreatic  $\beta$ -cell lines

NLRs Nod-like receptors P70S6k protein S6 kinase

**PAMPs** Pathogen-associated molecular patterns PDK1 Pyruvate dehydrogenase kinase isozyme 1

**PGN** Peptidoglycan

PIP2 Phosphatidylinositol-4,5-bisphosphate PIP3 Phosphatidylinositol-3,4,5-trisphosphate

PI3K Phosphatidylinositol 3-kinase **PRRs** The pattern-recognition receptors PYY Peptide YY; SAC: S-allylcysteine **SCFA** short-chain fatty acids

SGLT2 Sodium-dependent glucose transporters 2 SREBP-1c Sterol response element binding protein 1c

T2DM Type 2 diabetes mellitus mTOR mammalian target of rapamycin TCM Traditional Chinese medicine  $TNF\text{-}\alpha$ Tumor necrosis factor-α TLRs The Toll-like receptors

TSC1/TSC2 Tuberous sclerosis complexes 1/2

7.0 - 1Zonula Occludens-1

#### **Acknowledgments**

The funders had no role in the planning, analysis, or writing of this article.

#### **Funding**

This work was financially supported by National Natural Science Foundation of China (81741163), Natural Science Foundation (2016J06009) of Fujian Province (China), Multi-Year Research Grant of University of Macau (MYRG2018-00169-ICMS), Key Project of Fuzhou Municipal Bureau of Science and Technology (2018-G-87), and grants from Innovation and Technology Support Programme, Government of Hong Kong (Project code: UIM/321), The University of Hong Kong. The project was also supported by FAFU International Science and Technology Cooperation Project (KXb16011A) and partially by HEPATIC/Refbio2 Project.

#### References

Abate, N., and M. Chandalia. 2001. Ethnicity and type 2 diabetes: focus on Asian Indians. Journal of Diabetes and Its Complications

Aikawa, R., M. Nawano, Y. Gu, H. Katagiri, T. Asano, W. Zhu, R. Nagai, and I. Komuro. 2000. Insulin prevents cardiomyocytes from oxidative stress-induced apoptosis through activation of PI3 kinase/ Akt. Circulation 102:2873-9.

Alhouayek, M., and G. G. Muccioli. 2012. The endocannabinoid system in inflammatory bowel diseases: from pathophysiology to therapeutic opportunity. Trends in Molecular Medicine 18:615-25.

Amar, J., R. Burcelin, J. B. Ruidavets, P. D. Cani, J. Fauvel, M. C. Alessi, B. Chamontin, and J. Ferriéres. 2008. Energy intake is associated with endotoxemia in apparently healthy men. American Journal of Clinical Nutrition 87:1219-23.

Bäckhed, F., H. Ding, T. Wang, L. V. Hooper, G. Y. Koh, A. Nagy, C. F. Semenkovich, and J. I. Gordon. 2004. The gut microbiota as an environmental factor that regulates fat storage. Proceedings of the National Academy of Sciences of the United States of America

Bäckhed, F., J. K. Manchester, C. F. Semenkovich, and J. I. Gordon. 2007. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. Proceedings of the National Academy of Sciences of the United States of America 104:979-84.

Bahmani, M., H. Golshahi, K. Saki, M. Rafieian-Kopaei, B. Delfan, and T. Mohammadi. 2014. Medicinal plants and secondary metabolites for diabetes mellitus control. Asian Pacific Journal of Tropical Disease/APJTD 4:S687-S92.

Bates, J. M., J. Akerlund, E. Mittge, and K. Guillemin. 2007. Intestinal alkaline phosphatase detoxifies lipopolysaccharide and prevents inflammation in response to the gut microbiota. Cell Host Microbe

Beier, U. H., L. Wang, R. Han, T. Akimova, Y. Liu, and W. W. Hancock. 2012. Histone deacetylases 6 and 9 and Sirtuin-1 control FoxP3+ regulatory T cell function through shared and isotypespecific mechanisms. Science Signaling 5:ra45.

- Belgardt, B. F., K. Ahmed, M. Spranger, M. Latreille, R. Denzler, N. Kondratiuk, F. von Meyenn, F. N. Villena, K. Herrmanns, D. Bosco, et al. 2015. The microRNA-200 family regulates pancreatic beta cell survival in type 2 diabetes. Nature Medicine 21:619-27.
- Bommer, C., E. Heesemann, V. Sagalova, J. Manne-Goehler, R. Atun, T. Bärnighausen, and S. Vollmer. 2017. The global economic burden of diabetes in adults aged 20-79: a cost-of-illness study. Lancet Diabetes Endocrinology 5:423-30.
- Calle, M. C. and M. L. Fernandez. 2012. Inflammation and type 2 diabetes. Diabetes Metabolism 38:183-91.
- Cani, P. D., J. Amar, M. A. Iglesias, M. Poggi, C. Knauf, D. Bastelica, A. M. Neyrinck, F. Fava, K. M. Tuohy, C. Chabo, et al. 2007. Metabolic endotoxemia initiates obesity and insulin resistance. Diabetes 56:1761-72.
- Cao, H., J. Y. Qu, L. Chen, Y. B. Zhang, T. Szkudelski, D. Delmas, M. Daglia, and J. B. Xiao. 2018. Dietary polyphenols and type 2 diabetes: human study and clinical trial. Critical Reviews in Food Science and Nutrition. DOI: 10.1080/10408398.2018.1492900.
- Carpéné, C., S. Gomez-Zorita, S. Deleruyelle, and M. A. Carpéné. 2015. Novel strategies for preventing diabetes and obesity complications with natural polyphenols. Current Medicinal Chemistry 22:150-64.
- Catalogna, M., S. Fishman, Z. Halpern, S. Ben-Shlomo, U. Nevo, and E. Ben-Jacob. 2016. Regulation of glucose dynamics by noninvasive peripheral electrical stimulation in normal and insulin-resistant rats. Metabolism 65:863-73.
- Cerda, B., P. Periago, J. C. Espin, and F. A. Tomas-Barberan. 2005. Identification of urolithin A as a metabolite produced by human colon microflora from ellagic acid and related compounds. Journal of Agricultural and Food Chemistry 53:5571-6
- Chaithri, P. K., P. Narne, M. Siraj, and M. Ishaq. 2012. Excess maternal trans-mission of type 2 diabetes mellitus in South India: indication from sibling recurrence risk ratio analysis. Asian Journal of Epidemiology 5:87-94.
- Chambers, E. S., D. J. Morrison, and G. Frost. 2015. Control of appetite and energy intake by SCFA: what are the potential underlying mechanisms? Proceedings of the Nutrition Society 74:328-36.
- Chang, C. J., C. S. Lin, C. C. Lu, J. Martel, Y. F. Ko, D. M. Ojcius, S. F. Tseng, T. R. Wu, Y. M. Chen, J. D. Young, et al. 2015. Ganoderma lucidum reduces obesity in mice by modulating the composition of the gut microbiota. Nature Communications 6:7489.
- Chang, L., and M. Karin. 2001. Mammalian MAP kinase signalling cascades. Nature 410:37-40.
- Chen, W. J. Y., M. Diamant, K. de Boer, H. J. Harms, L. F. H. J. Robbers, A. C. van Rossum, M. H. H. Kramer, A. A. Lammertsma, and P. Knaapen, 2017. Effects of exenatide on cardiac function, perfusion, and energetics in type 2 diabetic patients with cardiomyopathy: a randomized controlled trial against insulin glargine. Cardiovascular Diabetology 16:67.
- Chen, Y. Q., Y. Y. Liu, M. M. Sarker, X. Yan, C. F. Yang, L. N. Zhao, X. C. Lv, B. Liu, and C. Zhao. 2018. Structural characterization and antidiabetic potential of a novel heteropolysaccharide from Grifola frondosa via IRS1/PI3K-JNK signaling pathways. Carbohydrate Polymers 198:452-61.
- Chen, Z., T. B. Gibson, F. Robinson, L. Silvestro, G. Pearson, B. Xu, A. Wright, C. Vanderbilt, and M. H. Cobb. 2001. MAP kinases. Chemical Reviews 101:2449-76.
- Clemente, J. C., L. K. Ursell, L. W. Parfrey, and R. Knight. 2012. The impact of the gut microbiota on human health: an integrative view. Cell 148:1258-70.
- Cordero-Herrera, I., M. A. Martín, L. Goya, and S. Ramos. 2015. Cocoa intake ameliorates hepatic oxidative stress in young Zucker diabetic fatty rats. Food Research International 69:194-201.
- Coué, M., V. Barquissau, P. Morigny, K. Louche, C. Lefort, A. Mairal, C. Carpéné, N. Viguerie, P. Arner, D. Langin, et al. 2018. Natriuretic peptides promote glucose uptake in a cGMP-dependent manner in human adipocytes. Science Reports 8:1097.
- Crawford, K. 2017. Review of 2017 diabetes standards of care. Nursing Clinics of North America 52:621-63.

- DECODE Study Group, 2003. Age- and sex-specific prevalence of diabetes and impaired glucose regulation in 11 Asian cohorts. Diabetes
- Degen, M., E. Natarajan, P. Barron, H. R. Widlund, and J. G. Rheinwald, 2012. MAPK/ERK-dependent translation factor hyperactivation and dysregulated laminin γ2 expression in oral dysplasia and squamous cell carcinoma. American Journal of Pathology 180:2462-78.
- Dehghan, P., G. B. Pourghassem, and J. M. Asghari. 2014. Oligofructose-enriched inulin improves some inflammatory markers and metabolic endotoxemia in women with type 2 diabetes mellitus: a randomized controlled clinical trial. Nutrition 30:418-23.
- Dentin, R., J. P. Pégorier, F. Benhamed, F. Foufelle, P. Ferré, V. Fauveau, M. A. Magnuson, J. Girard, and C. Postic. 2004. Hepatic glucokinase is required for the synergistic action of ChREBP and SREBP-1c on glycolytic and lipogenic gene expression. Journal of Biological Chemistry 279:20314-26.
- Du, C., Y. Shi, Y. Ren, H. Wu, F. Yao, J. Wei, M. Wu, Y. Hou, and H. Duan. 2015. Anthocyanins inhibit high-glucose-induced cholesterol accumulation and inflammation by activating LXRα pathway in HK-2 cells. Drug Design Development and Therapy 9:5099-113.
- Duvoix, A., R. Blasius, S. Delhalle, M. Schnekenburger, F. Morceau, E. Henry, M. Dicato, and M. Diederich. 2005. Chemopreventive and therapeutic effects of curcumin. Cancer Letters 223:181-90.
- Eckburg, P. B., E. M. Bik, C. N. Bernstein, E. Purdom, L. Dethlefsen, M. Sargent, S. R. Gill, K. E. Nelson, and D. A. Relman. 2005. Diversity of the human intestinal microbial flora. Science 308:1635-8.
- Eid, H. M., M. L. Wright, N. V. Anil Kumar, A. Qawasmeh, S. T. S. Hassan, A. Mocan, S. M. Nabavi, L. Rastrelli, A. G. Atanasov, and P. S. Haddad. 2017. Significance of microbiota in obesity and metabolic diseases and the modulatory potential by medicinal plant and food ingredients. Frontiers of Pharmacology 8:387.
- Espin, J. C., M. Larrosa, M. T. Garcia-Conesa, and F. Tomas-Barberan. 2013. Biological significance of urolithins, the gut microbial ellagic acid-derived metabolites: the evidence so far. Evidence-Based Complementary and Alternative Medicine. 2013:270418.
- Favaretto, F., G. Milan, G. B. Collin, J. D. Marshall, F. Stasi, P. Maffei, R. Vettor, and J. K. Naggert. 2014. GLUT4 defects in adipose tissue are early signs of metabolic alterations in Alms1GT/GT, a mouse model for obesity and insulin resistance. PLoS One 9:e109540.
- Ganley, I. G., du. H. Lam, J. Wang, X. Ding, S. Chen, and X. Jiang. 2009. ULK1.ATG13.FIP200 complex mediates mTOR signaling and is essential for autophagy. Journal of Biological Chemistry 284:12297-305.
- Gao, Z., J. Yin, J. Zhang, R. E. Ward, R. J. Martin, M. Lefevre, W. T. Cefalu, and J. Ye. 2009. Butyrate improves insulin sensitivity and increases energy expenditure in mice. Diabetes 58:1509-17.
- García López, P. M., P. G. De La Mora, W. Wysocka, B. Maiztegui, M. E. Alzugaray, H. Del Zotto, and M. I. Borelli. 2004. Quinolizidine alkaloids isolated from Lupinus species enhance insulin secretion. European Journal of Pharmacology 504:139-42.
- Genet, C., A. Strehle, C. Schmidt, G. Boudjelal, A. Lobstein, K. Schoonjans, M. Souchet, J. Auwerx, R. Saladin, and A. Wagner. 2010. Structure-activity relationship study of betulinic acid, a novel and selective TGR5 agonist, and its synthetic derivatives: potential impact in diabetes. Journal of Medicinal Chemistry 53:178-90.
- Goodnow, C. C., C. G. Vinuesa, K. L. Randall, F. Mackay, and R. Brink. 2010. Control systems and decision making for antibody production. Nature Immunology 11:681-8.
- Goto, T., N. Takahashi, S. Hirai, and T. Kawada. 2010. Various terpenoids derived from herbal and dietary plants function as PPAR modulators and regulate carbohydrate and lipid metabolism. PPAR Research 2010:483958.
- Green, A., J. Krause, and J. M. Rumberger. 2014. Curcumin is a direct inhibitor of glucose transport in adipocytes. Phytomedicine *21*:118-122.
- Grgurevic, L., G. L. Christensen, T. J. Schulz, and S. Vukicevic. 2016. Bone morphogenetic proteins in inflammation, glucose homeostasis



- and adipose tissue energy metabolism. Cytokine Growth Factor Reviews 27:105-18.
- Guarner, F., and J. R. Malagelada. 2003. Gut flora in health and disease. Lancet 361:512-19.
- Gugliucci, A. 2016. Fructose surges damage hepatic adenosyl-monophosphate-dependent kinase and lead to increased lipogenesis and hepatic insulin resistance. Medical Hypotheses 93:87-92.
- Gujral, U. P., R. Pradeepa, M. B. Weber, K. M. V. Narayan, and V. Mohan. 2013. Type 2 diabetes in South Asians: similarities and differences with white Caucasian and other populations. Annals of the New York Academy of Sciences 1281:51-63.
- Gunnink, L. K., O. D. Alabi, B. D. Kuiper, S. M. Gunnink, S. J. Schuiteman, L. E. Strohbehn, K. E. Hamilton, K. E. Wrobel, and L. L. Louters. 2016. Curcumin directly inhibits the transport activity of GLUT1. Biochimie 125:179-85
- Guo, H., M. Xia, T. Zou, W. Ling, R. Zhong, and W. Zhang. 2012. Cyanidin 3-glucoside attenuates obesity-associated insulin resistance and hepatic steatosis in high-fat diet-fed and db/db mice via the transcription factor FoxO1. Journal of Nutritional Biochemistry
- Hafizur, R. M., A. Hameed, M. Shukrana, S. A. Raza, S. Chishti, N. Kabir, and R. A. Siddiqui. 2015. Cinnamic acid exerts anti-diabetic activity by improving glucose tolerance in vivo and by stimulating insulin secretion in vitro. Phytomedicine 22:297-300.
- Han, Z., S. Cai, X. Zhang, Q. Qian, Y. Huang, F. Dai, and G. Zhang. 2017. Development of predictive models for total phenolics and free p-coumaric acid contents in barley grain by near-infrared spectroscopy. Food Chemistry 227:342-8.
- Hardie, D. G. 2011. AMP-activated protein kinase: an energy sensor that regulates all aspects of cell function. Genes and Development 25:1895-908.
- Harwood, N. E., and F. D. Batista. 2010. Early events in B cell activation. Annual Review of Immunology 28:185-210.
- Hawkins, B. T., T. F. Lundeen, K. M. Norwood, H. L. Brooks, and R. D. Egleton. 2007. Increased blood-brain barrier permeability and altered tight junctions in experimental diabetes in the rat: contribution of hyperglycaemia and matrix metalloproteinases. Diabetologia 50:202-11.
- Hayashi, F., K. D. Smith, A. Ozinsky, T. R. Hawn, E. C. Yi, D. R. Goodlett, J. K. Eng, S. Akira, D. M. Underhill, and A. Aderem. 2001. The innate immune response to bacterial flagellin is mediated by Toll-like receptor 5. Nature 410:1099-103.
- He, Y., F. Zhai, G. Ma, E. J. Feskens, J. Zhang, and P. Fu. 2009. Abdominal obesity and the prevalence of diabetes and intermediate hyperglycaemia in Chinese adults. Public Health Nutrition 12:1078-84.
- Heleno, S. A., A. Martins, M. J. Queiroz, and I. C. Ferreira. 2015. Bioactivity of phenolic acids: Metabolites versus parent compounds: a review. Food Chemistry 173:501-13.
- Hibi, M., A. Lin, T. Smeal, A. Minden, and M. Karin. 1993. Identification of an oncoprotein- and UV-responsive protein kinase that binds and potentiates the c-Jun activation domain. Gene and Development 7:2135-48.
- Hildebrandt, M. A., C. Hoffmann, S. A. Sherrill-Mix, S. A. Keilbaugh, M. Hamady, Y. Y. Chen, R. Knight, R. S. Ahima, F. Bushman, and G. D. Wu. 2009. High-fat diet determines the composition of the murine gut microbiome independently of obesity. Gastroenterology 137:1716-24.
- Hiriart, M., M. Velasco, C. M. Diaz-Garcia, C. Larqué, C. Sánchez-Soto, A. Albarado-Ibañez, J. P. Chávez-Maldonado, A. Toledo, and N. García-Delgado. 2014. Pancreatic  $\beta$ -cells in metabolic syndrome. Islets Langerhans 817-44.
- Hoshinom, T., A. Hoshino, and J. Nishino. 2016. Assessment of associations between ischaemic attacks in patients with type 2 diabetes mellitus and air concentrations of particulate matter  $< 2.5 \,\mu\text{m}$ . Journal of International Medical Research 44:639-55.
- Hsu, C. Y., H. Y. Shih, Y. C. Chia, C. H. Lee, H. Ashida, Y. K. Lai, and C. F. Weng. 2014. Rutin potentiates insulin receptor kinase to enhance insulin-dependent glucose transporter 4 translocation. Molecular Nutrition & Food Research 58:1168-76.

- Hu, E. A., A. Pan, V. Malik, and Q. Sun. 2012. White rice consumption and risk of type 2 diabetes: meta-analysis and systematic review. British Medical Journal 44:e1454.
- Hu, F. B. 2011. Globalization of diabetes: the role of diet, lifestyle, and genes. Diabetes Care 34:1249-57.
- Huang, D. W., and S. C. Shen. 2012. Caffeic acid and cinnamic acid ameliorate glucose metabolism via modulating glycogenesis and gluconeogenesis in insulin-resistant mouse hepatocytes. Journal of Functional Foods 4:358-66.
- Icaza-Chávez, M. E. 2013. Gut microbiota in health and disease. Revista de gastroenterologia de Mexico 78:240-8.
- Inoki, K., J. Kim, and K. L. Guan. 2012. AMPK and mTOR in cellular energy homeostasis and drug targets. Annual Review of Pharmacology and Toxicology 52:381-400.
- International Diabetes Federation. 2015. IDF diabetes atlas. 7th ed. Brussels: International Diabetes Federation.
- Jahan, H., M. I. Choudhary, M. Manzoor, K. M. Khan, S. Perveen, and Atta-Ur-Rahman. 2017. Insulinotropic action of 2,4-dinitroanilinobenzoic acid through the attenuation of pancreatic beta-cell lesions in diabetic rats. Chemico-Biological Interactions 273:237-44.
- Jin, H. Y., S. B. Hong, and T. S. Park. 2015. Morphologic changes in autonomic nerves in diabetic autonomic neuropathy. Diabetes & Metabolism Journal 39:461-7.
- João A. L., F. Reis, and R. Fernandes. 2016. The incretin system ABCs in obesity and diabetes-novel therapeutic strategies for weight loss and beyond. Obesity Review 17:553-72.
- Johansson, M. E. V. and J. I. Gordon. 2011. The two mucus layers of colon are organized by the MUC2 mucin, whereas the outer layer is a legislator of host-microbial interactions. Proceedings of the National Academy of Sciences of the United States of America 108:4659-65.
- Johansson, M. E. V., M. Phillipson, J. Petersson, A. Velcich, L. Holm, and G. C. Hansson. 2008. The inner of the two Muc2 mucindependent mucus layers in colon is devoid of bacteria. Proceedings of the National Academy of Sciences of the United States of America 105:15064-9.
- Jung, M., M. Park, H. Lee, Y. H. Kang, E. Kang, and S. Kim. 2006. Antidiabetic agents from medicinal plants. Current Medicinal Chemistry 13:1203-18.
- Jung, U. J., M. K. Lee, Y. B. Park, M. A. Kang, and M. S. Choi. 2006. Effect of citrus flavonoids on lipid metabolism and glucose-regulating enzyme mRNA levels in type-2 diabetic mice. International Journal of Biochemistry & Cell Biology 38:1134-45.
- Kadowaki, T., K. Ueki, T. Yamauchi, and N. Kubota. 2012. SnapShot: insulin signaling pathways. Cell 148:624.
- Kaefer, C. M. and J. A. Milner. 2008. The role of herbs and spices in cancer prevention. Journal of Nutritional Biochemistry 19:347-61.
- Kahn, C. R. and A. R. Saltiel. 2011. Insulin signalling and the regulation of glucose and lipid metabolism. Nature 414:799-806.
- Kaji, I., S. Karaki, and A. Kuwahara. 2014. Short-chain fatty acid receptor and its contribution to glucagon-like peptide-1 release. Digestion 89:31-6.
- Kandasamy, N. and N. Ashokkumar. 2012. Myricetin, a natural flavonoid, normalizes hyperglycemia in streptozotocin-cadmium-induced experimental diabetic nephrotoxic rats. Biomedicine and Preventive Nutrition 2:246-51.
- Kane, S., H. Sano, S. C. Liu, J. M. Asara, W. S. Lane, C. C. Garner, and G. E. Lienhard. 2002. A method to identify serine kinase substrates. Akt phosphorylates a novel adipocyte protein with a Rab GTPaseactivating protein (GAP) domain. Journal of Biological Chemistry 277:22115-18.
- Kappel, V. D., L. H. Cazarolli, D. F. Pereira, B. G. Postal, A. Zamoner, F. H. Reginatto, and F. R. Silva. 2013. Involvement of GLUT-4 in the stimulatory effect of rutin on glucose uptake in rat soleus muscle. Journal of Pharmacy and Pharmacology 65:1179-86
- Kavishankar, G. B., N. Lakshmidevi, S. M. Murthy, and H. S. Prakash. 2011. Diabetes and medicinal plants-a review. International Journal of Pharmocology Biomedical 3:65-80.
- Keane, K. and P. Newsholme. 2014. Metabolic regulation of insulin secretion. Vitamins & Hormones 95:1-33.

- Kim, J. G., S. H. Jo, K. S. Ha, S. C. Kim, Y. C. Kim, E. Apostolidis, and Y. I. Kwon. 2014. Effect of long-term supplementation of low molecular weight chitosan oligosaccharide (go2ka1) on fasting blood glucose and hba1c in db/db mice model and elucidation of mechanism of action. BMC Complementary and Alternative Medicine 14:272.
- Kim, S. J., M. C. Kim, J. Y. Um, and S. H. Hong. 2010. The beneficial effect of vanillic acid on ulcerative colitis. Molecules 15:7208-17.
- Kimura, I., K. Ozawa, D. Inoue, T. Imamura, K. Kimura, T. Maeda, K. Terasawa, D. Kashihara, K. Hirano, T. Tani, et al. 2013. The gut microbiota suppresses insulin-mediated fat accumulation via the short-chain fatty acid receptor GPR43. Nature Communications
- Kjems, L. L., J. J. Holst, A. Vølund, and S. Madsbad. 2003. The influence of GLP-1 on glucose-stimulated insulin secretion: effects on beta-cell sensitivity in type 2 and nondiabetic subjects. Diabetes 52:380-6.
- Knowler, W. C., E. Barrett-Connor, S. E. Fowler, R. F. Hamman, J. M. Lachin, E. A. Walker, D. M. Nathan; Diabetes Prevention Program Research Group. 2002. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. New England Journal of Medicine 346:393-403.
- Kong, A. P., G. Xu, N. Brown, W. Y. So, R. C., Ma, and J. C. Chan. 2013. Diabetes and its comorbidities—where East meets West. Nature Reviews Endocrinology 9:537-47.
- Kristensen, T., M. Fredholm, and S. Cirera. 2015. Expression study of GLUT4 translocation-related genes in a porcine pre-diabetic model. Mammalian Genome 26:650-7.
- Kyriakis, E., G. A. Stravodimos, A. L. Kantsadi, D. S. Chatzileontiadou, V. T. Skamnaki, and D. D. Leonidas. 2015. Natural flavonoids as antidiabetic agents. The binding of gallic and ellagic acids to glycogen phosphorylase b. FEBS Letters 589:1787-94.
- Lallès, J. P. 2010. Intestinal alkaline phosphatase: multiple biological roles in maintenance of intestinal homeostasis and modulation by diet. Nutrition Reviews 68:323-32.
- Lastra, G., S. Dhuper, M. S. Johnson, and J. R. Sowerset. 2010. Salt, aldosterone, and insulin resistance: impact on the cardiovascular system. Nature Reviews Cardiology 7:577-84.
- Lei, H., Q. Wang, S. Z. Guo, J. C. Han, H. J. Sun, X. X. Zhang, and W. T. Wu. 2013. The protective effect of MT-alpha-glucan against streptozotocin (STZ)-inducedNIT-1 pancreatic beta-cell damage. Carbohydrate Polymers 92:1211-17.
- Lei, H., M. Zhang, Q. Wang, S. Guo, J. Han, H. Sun, and W. Wu, W. 2013. MT-α-Glucan from the fruit body of the maitake medicinal mushroom Grifola frondosa (higher basidiomyetes) shows protective effects for hypoglycemic pancreatic  $\beta$ -Cells. International Journal of Medicinal Mushrooms 15:373-81.
- Letran, S. E., S. J. Lee, A. Shaikh, S. Uematsu, S. Akira, and S. J. McSorley, 2011. TLR5 functions as an endocytic receptor to enhance flagellin-specific adaptive immunity. European Journal Immunology 41:29-38.
- Li, Y., T. H. Huang, and J. Yamahara. 2008. Salacia root, a unique ayurvedic medicine, meets multiple targets in diabetes and obesity. Life Science 82:1045-9.
- Li, K. K., P. J. Tian, S. D. Wang, P. Lei, L. Qu, J. P. Huang, Y. J. Shan, and L. B. Li. 2017. Targeting gut microbiota: Lactobacillus, alleviated type 2 diabetes via inhibiting LPS secretion and activating GPR43 pathway. Journal of Functional Foods 38:561-70.
- Lin, H. V., A. E. J. K. Frassetto, A. R. Nawrocki, M. M. Lu, J. R. Kosinski, J. A. Hubert, D. Szeto, X. Yao, G. Forrest, and D. J. Marsh. 2012. Butyrate and propionate protect against diet-induced obesity and regulate gut hormones via free fatty acid receptor 3independent mechanisms. Plos One 7:e35240.
- Loizzo M. R., A. Pugliese, M. Bonesi, M. C. Tenuta, F. Menichini, J. B. Xiao, and R. Tundis. 2016. Edible flowers: A rich source of phytochemicals with antioxidant and hypoglycaemic activity. Journal of Agricultural and Food Chemistry 64:2467-74.
- Lozupone, C. A., J. I. Stombaugh, J. I. Gordon, J. K. Jansson, and R. Knight. 2012. Diversity, stability and resilience of the human gut microbiota. Nature 489:220-30.

- Lovorka, G., L. C. Gitte, J. S. Tim, and V. Slobodan. 2016. Bone morphogenetic proteins in inflammation, glucose homeostasis and adipose tissue energy metabolism. Cytokine & Growth Factor Reviews 27:105-18.
- Lyu, M., Y. F. Wang, G. W. Fan, X. Y. Wang, S.Y. Xu, and Y. Zhu. 2017. Balancing herbal medicine and functional food for prevention and treatment of cardiometabolic diseases through modulating gut microbiota. Frontiers in Microbiology 8:2146.
- Magnuson, B., B. Ekim, and D. C. Fingar. 2012. Regulation and function of ribosomal protein S6 kinase (S6K) within mTOR signalling networks. Biochemical Journal 441:1-21.
- Maji, D. and S. Samanta, 2017. Novel thiazolidinedione-5-acetic-acidpeptide hybrid derivatives as potent antidiabetic and cardioprotective agents. Biomedicine & Pharmacotherapy 88:1163-72.
- Mandal, R., S. Becker, and K. Strebhardt. 2016. Stamping out RAF and MEK1/2 to inhibit the ERK1/2 pathway: an emerging threat to anticancer therapy. Oncogene 35:2547-61.
- Manna, P., J. Das, J. Ghosh, J., and P. C. Sil. 2010. Contribution of type 1 diabetes to rat liver dysfunction and cellular damage via activation of NOS, PARP, IκBα/NF-κB, MAPKs, and mitochondriadependent pathways: prophylactic role of arjunolic acid. Free Radical Biology and Medicine 48:1465-84.
- Manning, B. D. and L. C. Cantley, 2007. AKT/PKB signaling: navigating downstream. Cell 129:1261-74.
- Marles, R. J. and N. R. Farnsworth. 1995. Antidiabetic plants and their active constituents. Phytomedicine 2:137-89.
- Martens, E. C., E. C. Lowe, H. Chiang, N. A. Pudlo, M. Wu, N. P. McNulty, D. W. Abbott, B. Henrissat, H. J. Gilbert, D. N. Bolam, et al. 2011. Recognition and degradation of plant cell wall polysaccharides by two human gut symbionts. PLoS Biology 9:1-16.
- Maslowski, K. M., A. T. Vieira, A. Ng, J. Kranich, F. Sierro, D. Yu, H. C. Schilter, M. S. Rolph, F. Mackay, D. Artis, et al. 2009. Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. Nature 461:1282-6.
- Mazidi, M., E. Karimi, P. Rezaie, and G. A. Ferns. 2017. Treatment with GLP1 receptor agonists reduce serum CRP concentrations in patients with type 2 diabetes mellitus: a systematic review and metaanalysis of randomized controlled trials. Journal of Diabetes and Its Complications 31:1237-42.
- Mccranie, E. K., and B. O. Bachmann. 2014. Bioactive oligosaccharide natural products. Natural Product Reports 31:1026-42.
- Mebratu, Y., and Y. Tesfaigzi. 2009. How ERK1/2 activation controls cell proliferation and cell death: is subcellular localization the answer? Cell Cycle 8:1168-75.
- Meloche, S., and J. Pouysségur. 2007. The ERK1/2 mitogen-activated protein kinase pathway as a master regulator of the G1- to S-phase transition. Oncogene 26:3227-99.
- Meng, T., J. Wu, H. Yi, J. Liu, B. Lu, M. Yuan, X. Huang, H. Yuan, and F. Hu. 2016. A spermine conjugated stearic acid-g-chitosan oligosaccharide polymer with different types of amino groups for efficient p53 gene therapy. Colloids and Surfaces B 145:695-705.
- Mirmiranpour, H., S. Khaghani, R. Z. Bathaie, M. Nakhjavani, A. Kebriaeezadeh, M. Ebadi, and M. Zangooei. 2016. The preventive effect of L-lysine on lysozyme glycation in type 2 diabetes. Acta Medica Iranica 54:24-31.
- Mohan, R., Y. Mao, S. Zhang, Y. W. Zhang, C. R. Xu, G. Gradwohl, and X. Q. Tang. 2015. Differentially expressed microRNA-483 confers distinct functions in pancreatic  $\beta$ - and  $\alpha$ -cells. *Journal of* Biological Chemistry 290:19955-66.
- Mohan, S., and B.M. Pinto. 2007. Zwitterionic glycosidase inhibitors: salacinol and related analogues. Carbohydrate Research 242:1551-80.
- Mora, A., K. Sakamoto, E. J. McManus, and D. R. Alessi, 2005. Role of the PDK1-PKB-GSK3 pathway in regulating glycogen synthase and glucose uptake in the heart. FEBS Letters 579:3632-8.
- Muccioli, G. G., D. Naslain, F. Bäckhed, C. S. Reigstad, D. M. Lambert, N.M. Delzenne, and P. D. Cani. 2010. The endocannabinoid system links gut microbiota to adipogenesis. Molecular Systems Biology 6:392-406.
- Murphy, E. F., P. D. Cotter, S. Healy, S. Healy, T. M. Marques, O. O'Sullivan, F. Fouhy, S. F. Clarke, P. W. O'Toole, E. M. Quigley,



- et al. 2010. Composition and energy harvesting capacity of the gut microbiota: relationship to diet, obesity and time in mouse models.
- National Kidney Foundation. 2012. KDOQI clinical practice guideline for diabetes and CKD: 2012 update. American Journal of Kidney Diseases 60:850-86.
- Nabavi, S. F., R. Thiagarajan, L. Rastrelli, M. Daglia, E. Sobarzo-Sánchez, H. Alinezhad, and S. M. Nabavi. 2015. Curcumin: a natural product for diabetes and its complications. Current Topics in Medicinal Chemistry 15:2445-55.
- Neish, A. S. 2009. Microbes in gastrointestinal health and disease. Gastroenterology 136:65-80.
- Ning, F., Z. C. Pang, Y. H. Dong, W. G. Gao, H. R. Nan, S. J. Wang, L. Zhang, J. Ren, J. Tuomilehto, N. Hammar, et al; Qingdao Diabetes Survey Group. 2009. Risk factors associated with the dramatic increase in the prevalence of diabetes in the adult Chinese population in Qingdao, China. Diabetic Medicine 26:855-63.
- Nomura, S., S. Sakamaki, M. Hongu, E. Kawanishi, Y. Koga, T. Sakamoto, Y. Yamamoto, K. Ueta, H. Kimata, K. Nakayama, et al. 2010. Discovery of canagliflozin, a novel C-glucoside with thiophene ring, assodium-dependent glucose cotransporter 2 inhibitor for the treatment of type 2 diabetes mellitus. Journal of Medicinal Chemistry 53:6355-60.
- Nøhr, M. K., M. H. Pedersen, A. Gille, K. L. Egerod, M. S. Engelstoft, A. S. Husted, R. M. Sichlau, K. V. Grunddal, S. S. Poulsen, S. Han, et al. 2013. GPR41/FFAR3 and GPR43/FFAR2 as cosensors for short-chain fatty acids in enteroendocrine cells vs FFAR3 in enteric neurons and FFAR2 in enteric leukocytes. Endocrinology 154:3552-64.
- O'Hara, A. M., and Shanahan, F. 2006. The gut flora as a forgotten organ. EMBO Reports 7:688-93.
- Ohno, M., C. Shibata, T. Kishikawa, T. Yoshikawa, A. Takata, K. Kojima, M. Akanuma, Y. J. Kang, H. Yoshida, and M. Otsuka. 2013. The flavonoid apigenin improves glucose tolerance through inhibition of microRNA maturation in miRNA103 transgenic mice. Scientific Reports 3:2553.
- Panda, S. and A. Kar. 2007. Apigenin (4',5,7-trihydroxyflavone) regulates hyperglycaemia, thyroid dysfunction and lipid peroxidation in alloxan-induced diabetic mice. Journal of Pharmacy and Pharmacology 59:1543-8.
- Paradis, S. and G. Ruvkun. 1998. Caenorhabditis elegans Akt/PKB transduces insulin receptor-like signals from AGE-1 PI3 kinase to the DAF-16 transcription factor. Genes and Development 12:2488-98.
- Perry, B. I., D. Salimkumar, D. Green, A. Meakin, A. Gibson, D. Mahajan, T. Tahir, and S. P. Singh. 2017. Associated illness severity in schizophrenia and diabetes mellitus: A systematic review. Psychiatry Research 256:102-10.
- Poy, M. N., L. Eliasson, J. Krutzfeldt, S. Kuwajima, X. Ma, P. E. Macdonald, S. Pfeffer, T. Tuschl, N. Rajewsky, P. Rorsman, and M. Stoffel. 2004. A pancreatic islet-specific microRNA regulates insulin secretion. Nature 432:226-30.
- Qin, J., Y. Li, Z. Cai, S. Li, J. Zhu, F. Zhang, et al. 2012. A metagenome-wide association study of gut microbiota in type 2 diabetes. Nature 490:55-60.
- Ramachandran, A., N. Murugesan, S. Mary, C. Snehalatha, and A. Yamuna. 2008. High prevalence of diabetes and cardiovascular risk factors associated with urbanization in India. Diabetes Care 31:893-8.
- Ramachandran, A., C. Snehalatha, A. Kapur, V. Vijay, V. Mohan, A. K. Das, P. V. Rao, C. S. Yajnik, K. M. Prasanna Kumar, and J. D. Nair. 2001. High prevalence of diabetes and impaired glucose tolerance in India: National Urban Diabetes Survey. Diabetologia 44:1094-101.
- Raut, N. A., P. W. Dhore, S. D. Saoji, and D. M. Kokare. 2016. Selected bioactive natural products for diabetes mellitus. Studies in Natural Products Chemistry 48:287-322.
- Ravussin, Y., O. Koren, A. Spor, C. LeDuc, R. Gutman, J. Stombaugh, R. Knight, R. E. Ley, and R. L. Leibel. 2012. Responses of gut microbiota to diet composition and weight loss in lean and obese mice. Obesity 20:738-47.

- Reddy, K. B., S. M. Nabha, and N. Atanaskova. 2003. Role of MAP kinase in tumor progression and invasion. Cancer Metastasis Reviews 22:395-403.
- Ren, H., Q. L. Xu, M. Zhang, L. M. Dong, Q. Zhang, B. Luo, Q. W. Luo and J. W. Tan. 2017. Bioactive caffeic acid derivatives from wedelia trilobata. Phytochemistry Letters 19:18-22.
- Retnakaran, R., A. J. G. Hanley, P. W. Connelly, M. Sermer, and B. Zinman. 2006. Ethnicity modifies the effect of obesity on insulin resistance in pregnancy: a comparison of Asian, South Asian, and Caucasian women. Journal of Clinical Endocrinology and Metabolism 91:93-7.
- Reyes, A., M. Haynes, N. Hanson, F. E. Angly, A. C. Heath, F. Rohwer, and J. I. Gordon. 2010. Viruses in the fecal microbiota of monozygotic twins and their mothers. Nature 466:334-8.
- Rutter, G. A., T. J. Pullen, D. J. Hodson, and A. Martinez-Sanchez. 2015. Pancreatic  $\beta$ -cell identity, glucose sensing and the control of insulin secretion. Biochemical Journal 466:203-18.
- Ryu D, L. Mouchiroud, P. A. Andreux, E. Katsyuba, N. Moullan, A. A. Nicolet-Dit-Felix, E. G. Williams, P. Jha, G. Lo Sasso, D. Huzard, et al. 2016. Urolithin A induces mitophagy and prolongs lifespan in C. elegans and increases muscle function in rodents. Nature Medicine 22:879-88.
- Sacks, D. A., D. R. Hadden, M. Maresh, C. Deerochanawong, A. R. Dyer, B. E. Metzger, and E. R. Trimble. 2012. Frequency of gestational diabetes mellitus at collaborating centers based on IADPSG consensus panel-recommended criteria: the Hyperglycemia and Adverse Pregnancy Outcome (HAPO) Study. Diabetes Care 35:526-8.
- Sakakibara, S., T. Yamauchi, Y. Oshima, Y. Tsukamoto, and T. Kadowaki. 2006. Acetic acid activates hepatic AMPK and reduces hyperglycemia in diabetic KK-A(y) mice. Biochemical and Biophysical Research Communications 344:597-604.
- Schwarz, P.E., Lindström, J., Kissimovascarbeck, K., Szybinski, Z., Barengo, N.C., Peltonen, M. and Tuomilehto, J. 2008. The European perspective of type 2 diabetes prevention: diabetes in Europe-prevention using lifestyle, physical activity and nutritional intervention (de-plan) project. Experimental and Clinical Endocrinology & *Diabetes* 116:167–72.
- Schertzer, J. D., A. K. Tamrakar, J. G. Magalhães, S. Pereira, P. J. Bilan, M. D. Fullerton, Z. Liu, G. R. Steinberg, A. Giacca, D. J. Philpott, and A. Klip. 2011. NOD1 activators link innate immunity to insulin resistance. Diabetes 60:2206-15.
- Spiljar, M., D. Merkler, and M. Trajkovski. 2017. The immune system bridges the gut microbiota with systemic energy homeostasis-focus on TLRs, mucosal barrier and SCFAs. Frontiers in Immunology 8:1353.
- Sharabi K., C. D. J. Tavares, A. K. Rines and P. Puigserver. 2015. Molecular pathophysiology of hepatic glucose production. Molecular Aspects of Medicine 46:21-33.
- Shen, J., M. S. Obin, and L. Zhao. 2013. The gut microbiota, obesity and insulin resistance. Molecular Aspects of Medicine 34:39-58.
- Shi, X., T. Miyakawa, A. Nakamura, F. Hou, M. Hibi, J. Ogawa, and M. Tanokura. 2017. Engineering a short-chain dehydrogenase/reductase for the stereoselective production of (2S,3R,4S)-4-hydroxyisoleucine with three asymmetric centers. Scientific Reports 7:13703.
- Shibano, M., D. Tsukamoto, R. Fujimoto, Y. Masui, H. Sugimoto, and G. Kusano. 2000. Studies on the constituents of Broussonetia species. VII. Four new pyrrolidine alkaloids, broussonetines M, O, P, and Q, as inhibitors of glycosidase, from Broussonetia kazinoki SIEB. Chemical and Pharmaceutical Bulletin 48:1281-5.
- Shibano, M., D. Tsukamoto, A. Masuda, Y. Tanaka, and G. Kusano, 2001. Two new pyrrolidine alkaloids, radicamines A and B, as inhibitors of alpha-glucosidase from Lobelia chinensis Lour. Chemical and Pharmaceutical Bulletin 49:1362-5.
- Shimoda, H., N. Nishida, K. Ninomiya, H. Matsuda, and M. Yoshikawa. 2002. ChemInform Abstract: Javaberine A, new TNF-α and nitric oxide production inhibitor, from the roots of Talinum paniculatum. ChemInform 33:2002.
- Shishodia, S., G. Sethi, and B. B. Aggarwal. 2005. Curcumin: getting back to the roots. Annals of the New York Academy of Sciences 1056:206-17

- Shortt, C., O. Hasselwander, A. Meynier, A. Nauta, E. N. Fernández, P. Putz, I. Rowland, J. Swann, J. Türk, J. Vermeiren, and J. M. Antoine. 2018. Systematic review of the effects of the intestinal microbiota on selected nutrients and non-nutrients. European Journal of Nutrition 57:25-49.
- Si, H., Z. Fu, P. V. Babu, W. Zhen, T. Leroith, M. P. Meaney, K. A. Voelker, Z. Jia, R. W. Grange, and D. Liu. 2011. Dietary epicatechin promotes survival of obese diabetic mice and Drosophila melanogaster. Journal of Nutrition 141:1095-100.
- Sommer, F. and F. Bäckhed. 2013. The gut microbiota-masters of host development and physiology. Nature Reviews Microbiology 11:227-38.
- Song, W. J., P. Mondal, A. Wolfe, L. C. Alonso, R. Stamateris, B. W. Ong, O. C. Lim, K. S. Yang, S. Radovick, H. J. Novaira, et al. 2014. Glucagon regulates hepatic kisspeptin to impair insulin secretion. Cell Metabolism 19:667-81.
- Song, Y., T. Wu, Q. Yang, X. Chen, M. Wang, Y. Wang, X. C. Peng, and S. Y. Ou. 2014. Ferulic acid alleviates the symptoms of diabetes in obese rats. Journal of Functional Foods 9:141-7.
- Steinbrenner, H. 2013. Interference of selenium and selenoproteins with the insulin-regulated carbohydrate and lipid metabolism. Free Radical Biology and Medicine 65:1538-47.
- Su, Y., J. Wu, J. He, X. Liu, X. Chen, Y. Ding, C. Zhang, W. Chen, Y. Wang, and R. Gao. 2017. High insulin impaired ovarian function in early pregnant mice and the role of autophagy in this process. Endocrine Journal 64:613-21.
- Szkudelski, T., and K. Szkudelska. 2015. Resveratrol and diabetes: from animal to human studies. Biochimica et Biophysica Acta 1852:1145-54.
- Takada, K., T. Uehara, Y. Nakao, S. Matsunaga, R. W. M. van Soest, and N. Fusetani. 2004. Schulzeines A-C, new alpha-glucosidase inhibitors from the marine sponge Penares schulzei. Journal of the American Chemical Society 126:187-93.
- Tanti, J. F., T. Gremeaux, E. van Obberghen, and Y. Le Marchand-Brustel. 1994. Serine/threonine phosphorylation of insulin receptor substrate 1 modulates insulin receptor signaling. Journal of Biological Chemistry 269:6051-7.
- Tao, R., E. F. de Zoeten, E. Ozkaynak, C. Chen, L. Wang, P. M. Porrett, B. Li, L. A. Turka, E. N. Olson, M. I. Greene, A. D. Wells, and W.W. Hancock. 2007. Deacetylase inhibition promotes the generation and function of regulatory T cells. Nature Medicine 13:1299-307.
- Thea, K., F. Merete, Y. S. Lam, and C. Susanna. 2015. Expression study of GLUT4 translocation-related genes in a porcine pre-diabetic model. Mammalian Genome 26:650-7.
- Tolhurst, G., H. Heffron, Y. S. Lam, H. E. Parker, A. M. Habib, E. Diakogiannaki, J. Cameron, J. Grosse, F. Reimann, and F. M. Gribble. 2012. Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein-coupled receptor FFAR2. Diabetes 61:364-71.
- Tomas-Barberan, F. A. and M. N. Clifford. 2000. Dietary hydroxybenzoic acid derivatives-nature, occurrence and dietary burden. Journal of the Science of Food and Agriculture 80:1024-32.
- Tong, H., Z. Liang, and G. Wang. 2008. Structural characterization and hypoglycemic activity of a polysaccharide isolated from the fruit of Physalis alkekengi L. Carbohydrate Polymers 71:316-23.
- Turnbaugh, P. J., R. E. Ley, M. A. Mahowald, V. Magrini, E. R. Mardis, and J. I. Gordon. 2006. An obesity-associated gut microbiome with increased capacity for energy harvest. Nature 444:1027-31.
- Tutino, G. E., W. H. Tam, X. Yang, J. C. N. Chan, T. T. H. Lao, and R. C. W. Ma. 2014. Diabetes and pregnancy: perspectives from Asia. Diabetic Medicine 31:302-18.
- Urquia, M., R. H. Glazier, H. Berger, I. Ying, L. De Souza, and J. G. Ray. 2011. Gestational diabetes among immigrant women. Epidemiology 22:879-80.
- Vallon, V., and S. C. Thomson. 2017. Targeting renal glucose reabsorption to treat hyperglycaemia: the pleiotropic effects of SGLT2 inhibition. Diabetologia 60:215-25.
- Vinayagam, R., J. B. Xiao, and B. J. Xu. 2017. An insight into anti-diabetic properties of dietary phytochemicals. Phytochemistry Reviews 16:535-53.

- Vinothiya, K. and N. Ashokkumar. 2017. Modulatory effect of vanillic acid on antioxidant status in high fat diet-induced changes in diabetic hypertensive rats. Biomedicine & Pharmacotherapy 87:640-52.
- Vrieze, A., E. Van Nood, F. Holleman, J. Salojärvi, R. S. Kootte, J. F. Bartelsman, G. M. Dallinga-Thie, M. T. Ackermans, M. J. Serlie, R. Oozeer, et al. 2012. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. Gastroenterology 143:913-16.
- Wang, L., P. Gao, M. Zhang, Z. Huang, D. Zhang, Q. Deng, Y. Li, Z. Zhao, X. Qin, D. Jin, et al. 2017. Prevalence and ethnic pattern of diabetes and prediabetes in China in 2013. JAMA 317:2515-23.
- Wang, J., J. Zhang, B. Zhao, Y. Wu, C. Wang, and Y. Wang. 2010. Structural features and hypoglycaemic effects of Cynomorium songaricum polysaccharides on STZ-induced rats. Food Chemistry 120:443-51.
- Wang, L. Y., Y. Wang, D. S. Xu, K. F. Ruan, Y. Feng, and S. Wang. 2012. MDG-1, apolysaccharide from Ophiopogon japonicus exerts hypoglycemic effects through the PI3K/Akt pathway in a diabetic KKAy mouse model. Journal of Ethnopharmacology 143:347-54.
- Wang, Y., J. Liu, C. Liu, A. Naji, and D. A. Stoffers, 2013. MicroRNA-7 regulates the mTOR pathway and proliferation in adult pancreatic β-cells. *Diabetes* 62:887–95.
- Watson, R. T., M. Kanzaki, and J. E. Pessin. 2004. Regulated membrane trafficking of the insulin-responsive glucose transporter 4 in adipocytes. Endocrine Reviews 25:177-204.
- Weisberg, S. P., R. Leibel, and D. V. Tortoriello, 2008. Dietary curcumin significantly improves obesity-associated inflammation and diabetes in mouse models of diabesity. Endocrinology 149:3549-58.
- Whitman, W. B., D. C. Coleman, and W. J. Wiebe. 1998. Prokaryotes: the unseen majority. Proceedings of the National Academy of Sciences of the United States of America 95:6578-83.
- Xiao, J. B., X. L. Ni, G. Y. Kai, and X. Q. Chen. 2015. Advance in dietary polyphenols as aldose reductases inhibitors: structure-activity relationship aspect. Critical Reviews in Food Science and Nutrition
- Xiao, J. B., and P. Högger. 2015. Dietary polyphenols and type 2 diabetes: current insights and future perspectives. Current Medicinal Chemistry 22:23-38.
- Xiao, J. B. 2017. Dietary flavonoid aglycones and their glycosides: What show better biological benefits? Critical Reviews in Food Science and Nutrition 57:1874-905.
- Xiao, Z. Q., Y. L.Wang, S. R. Gan, and J. C. Chen. 2014. Polysaccharides from Liriopes Radix ameliorates hyperglycemia via various potential mechanisms in diabetic rats. Journal of the Science of Food and Agriculture 94:975-82.
- Xie, W., G. Tanabe, J. Akaki, T. Morikawa, K. Ninomiya, T. Minematsu, M. Yoshikawa, X. Wu, and O. Muraoka. 2011a. Isolation, structure identification and SAR studies on thiosugar sulfonium salts, neosalaprinol and neoponkoranol, as potent α-glucosidase inhibitors. Bioorganic & Medicinal Chemistry 19:2015-22.
- Xie, W., G. Tanabe, K. Matsuoka, M. F. Amer, T. Minematsu, X. Wu, M. Yoshikawa, and O. Muraoka. 2011b. Role of the side chain stereochemistry in the  $\alpha$ -glucosidase inhibitory activity of kotalanol, a potent natural α-glucosidase inhibitor. Bioorganic & Medicinal Chemistry Letters 19:2252-62.
- Xing, Y., D. Yang, J. Lu, and D. L. Dong. 2015. Insulin prevents bone morphogenetic protein-4 induced cardiomyocyte apoptosis through Biophysical Akt. Biochemical and Research activating Communications 456:605-9.
- Xu, J., Y. Wang, D. S. Xu, K. F. Ruan, Y. Feng, and S. Wang. 2011. Hypoglycemiceffects of MDG-1, a polysaccharide derived from Ophiopogon japonicas, in the ob/ob mouse model of type 2 diabetes mellitus. International Journal of Biological Macromolecules
- Xu, S., Y. Dou, B. Ye, Q. Wu, Y. Wang, M. Hu, F. Ma, X. Rong, and J. Guo. 2017. Ganoderma lucidum polysaccharides improve insulin sensitivity by regulating inflammatory cytokines and gut microbiota composition in mice. Journal of Functional Foods 38:545-55.
- Yamaguchi, O., and K. Otsu. 2012. Role of autophagy in aging. Journal of Cardiovascular Pharmacology 60:242-7.

- Yang, J., P. H. Summanen, S. M. Henning, M. Hsu, H. Lam, J. Huang, C. H. Tseng, S. E. Dowd, S. M. Finegold, D. Heber, et al. 2015. Xylooligosaccharide supplementation alters gut bacteria in both healthy and prediabetic adults: a pilot study. Frontiers in Physiology
- Yin, J., H. Lin, J. Li, Y. Wang, S. W. Cui, S. Nie, and M. Y. Xie. 2012. Structural characterization of a highly branched polysaccharide from the seeds of *Plantago asiatica* L. *Carbohydrate Polymers* 87:2416–24.
- Yin, X., Z. Xu, Z. Zhang, L. Li, Q. Pan, F. Zheng, and H. Li. 2017. Association of Pi3K/AKT/mTOR pathway genetic variants with type 2 diabetes mellitus in Chinese. Diabetes Research and Clinical Practice 128:127-35.
- Yoshikawa, M., T. Murakami, H. Shimada, H. Matsuda, J. Yamahara, G. Tanabe, and O. Muraoka, O. 1997. Salacinol, potent antidiabetic principle with unique thiosugar sulfonium sulfate structure from the Ayurvedic traditional medicine Salacia reticulata in Sri Lanka and India. Tetrahedron Letters 38:8367-70.
- Yousefi, H., P. Karimi, A. Alihemmati, M. R. Alipour, P. Habibi, and N. Ahmadiasl. 2017. Therapeutic potential of genistein in ovariectomy-induced pancreatic injury in diabetic rats: The regulation of MAPK pathway and apoptosis. Iranian Journal of Basic Medical Sciences 20:1009-15.
- Zappas, M. P., M. Gentes, and B. Walton-Moss. 2017. Use of incretin therapy in the treatment of type 2 diabetes mellitus. Journal for Nurse Practitioners 13:418-24.
- Zhang, B. B., Zhou, G, and Li, C. 2009. AMPK: an emerging drug target for diabetes and the metabolic syndrome. Cell Metabolism 9:407-16.
- Zhang, J., C. Wang, X. Ha, W. Li, P. Xu, Y. Gu, T. Wang, Y. Wang, and J. Xie. 2016. The DNA methylation of TNF-alpha, MCP-1, and adiponectin in visceral adipose tissue is related to type 2 diabetes in Xinjiang Uygur population. Journal of Diabetes 9:699-706.

- Zhang, P., X. Zhang, J. Brown, D. Vistisen, R. Sicree, J. Shaw, and G. Nichols. 2010a. Global healthcare expenditure on diabetes for 2010 and 2030. Diabetes Research and Clinical Practice 87:293-301.
- Zhang, W. and H. T. Liu 2002. MAPK signal pathways in the regulation of cell proliferation in mammalian cells. Cell Research 12:9-18.
- Zhang, Z. F., Q. Li, J., Liang, X. Q. Dai, Y. Ding, J. B. Wang, and Y. Li. 2010b. Epigallocatechin-3-O-gallate (EGCG) protects the insulin sensitivity in rat L6 muscle cells exposed to dexamethasone condition. Phytomedicine 17:14-18.
- Zhao, C., Y. J. Wu, X. Y. Liu, B. Liu, H. Cao, H. Yu, S. D. Sarker, L. Nahar, and J. B. Xiao. 2017. Functional properties, structural studies and chemo-enzymatic synthesis of oligosaccharides. Trends in Food Science and Technology 66:135-45.
- Zhao, C., C. F. Yang, M. J. Chen, X. C. Lv, B. Liu, L. Z. Yi, L. Cornara, M. C. Wei, Y. C. Yang, R. Tundis, et al. 2018a. Regulatory efficacy of brown seaweed Lessonia nigrescens extract on the gene expression profile and intestinal microflora in type 2 diabetic mice. Molecular Nutrition & Food Research 62:1700730.
- Zhao, C., C. F. Yang, B. Liu, L. Lin, S. D. Sarker, L. Nahar, H. Yu, H. Cao, and J. B. Xiao. 2018b. Bioactive compounds from marine macroalgae and their hypoglycemic benefits. Trends in Food Science and *Technology 72*:1–12.
- Zheng, J., X. Yuan, G. Cheng, S. Jiao, C. Feng, X. Zhao, H. Yin, Y. G. Du, and H. T. Liu. 2018. Chitosan oligosaccharides improve the disturbance in glucose metabolism and reverse the dysbiosis of gut microbiota in diabetic mice. Carbohydrate Polymers 190:77-86.
- Zou, S., X. Zhang, W. Yao, Y. Niu, and X. Gao. 2010. Structure characterization and hypoglycemic activity of a polysaccharide isolated from the fruit of Lycium barbarum L. Carbohydrate Polymers 80: 1161-7.
- Zygmunt, K., B. Faubert, J. Macneil, and E. Tsiani. 2010. Naringenin, a citrus flavonoid, increases muscle cell glucose uptake via AMPK. Biochemical and Biophysical Research Communications 398:178-83.