Email: <a href="mailto:thomas.richards@zoo.ox.ac.uk">thomas.richards@zoo.ox.ac.uk</a>

Web: https://protists.co.uk

Qualifications

D. Phil. University of Oxford and the Natural History Museum, London

(BBSRC Studentship) Awarded: 2006

Title: Horizontal Gene Transfer and the Evolution of Eukaryotes

Supervisors: Prof. T. Martin Embley FRS (University of Newcastle)

Prof. Tom Cavalier-Smith FRS (University of Oxford)

M.Sc. University of Oxford, (BBSRC studentship) Awarded: 2001

B.Sc. University College, London Awarded: 1999

**Appointments** 

2019- Professor of Evolutionary Genomics (University of Oxford) & Fellow Merton

College Oxford

2015- Professor of Evolutionary Genomics (University of Exeter)

2014- Royal Society University Research Fellow (University of Exeter)

2013 Senior Research Fellow (University of Exeter)

2010 80% FTE, Group Leader (Natural History Museum, London)

20% FTE Senior Research Fellow (University of Exeter)

2009 Senior Research Fellow (University of Exeter)

2007 Leverhulme Early Career Fellow (University of Exeter)

2005 DEFRA Postdoctoral Fellow (University of Exeter)

**Personal Awards** 

2019 Royal Society University Research Fellowship Renewal (£483,000 personal salary

and research award for 3 years)

2018 Hutner Award (International Society of Protistology, \$1000)

2016/17 Miller Visiting Professorship, University of California Berkeley (\$31,000 salary and

travel award for semester sabbatical award)

2015 Elected as Fellow of the American Academy of Microbiology

2014 Philip Leverhulme Award (£100,000)

2014 Royal Society University Research Fellowship (£448,000 personal salary and

research award for 5 years)

2012 EMBO Young Investigator (~€70,000 in travel and research awards)

2012 Canadian Institute for Advanced research, Fellow of the Integrated Microbial

Biodiversity program (~\$80,000 CAD in travel and research awards)

2012 Berkeley Award, British Mycology Society (£500)

2009 President's Medal for Cell Biology, Society for Experimental Biology

2007 Leverhulme Early Career Fellowship (£60,000 salary and research award for 2

years)

2000 NASA Planetary Biology Internship, Rensselaer Polytechnic Institute, USA (\$3500)

1999 Royal Society Research Studentship, University College, London (£1600)

### Awards for People and Work under My Supervision

2020	Marie Curie Fellowship; Luis Javier Galindo (€213,000)
2019	Merton College, Oxford, Junior Research Fellowship; Nick Irwin (£90,000)
2017	Royal Society Newton Fellowship; Elisabet Alacid-Fernandez (£113,000)
2015	Marie Curie Fellowship; Estelle Kilas (€183,454)
2013	EMBO Long Term Fellowship; Jeremy Wideman (£70,000)
2012	Royal Society Newton Fellowship; Adam Monier (£82,875)
2011	EMBO Long Term Fellowship; Aurélie Chambouvet (£50,000)
2011	Marie Curie Fellowship; Aurélie Chambouvet (€209,033)
2009	Young Systematists' Forum, London, UK: Oral Presentation Prize, James Harrison
2009	Fungal Genetics Conference, Asilomar, USA: Eukaryotic Cell Outstanding Young

# **External Responsibilities and Commissions of Trust**

2021-	PARC PARC Disease-Task-Team (Dept. Interior, U.S. Geological-Survey), attending
	online-meetings and assisting with writing mitigation/monitoring wildlife-disease

factsheets

2020- Scientific Advisory Board, NERC Culture Collections Algae Protist (CCAP)

2021 Ad hoc reviewer and panel member for The Research Council of Norway

2020 Ad hoc reviewer and panel member for Wellcome Trust grant review panel

2020-2023 Board of Visitors (Equivalent to Governing Body Member and Trustee) at the

Oxford Natural History Museum Oxford

2019- Governing Body Member and Trustee, Merton College Oxford

2019-23 Member of the Board of Trustee, BBSRC Earlham Institute

Investigator Prize, Meredith Jones

2017-20 Steering group member for the University of Exeter's Translational Research

Exchange

A £3.5M partnership between the University and the Wellcome Trust (through its ISSF mechanism). The steering group provides advice on the strategic direction and makes decisions on projects to be supported across the institution and its wider

network of clinical and academic partners

2017-20 Associate Editor, Environmental Microbiology

2012-2015 Elected as council member to The Linnean Society, London

2011-2015 Associate Editor, Ecology and Evolution

2011-2014 Associate Editor, Frontiers in Microbiology

2010-2013 Associate Editor, BMC Evolutionary Biology

2009-2012	Chair of SynTax and administrator for UK Grant Review Panel
	Joint research initiative funding systematics and taxonomy, supported by the BBSRC, NERC, DEFRA, Linnean Society and Systematics Association, awarding £300,000 annually
2008-11	Chair of The Linnean Society / Systematics Association Systematic Research Fund
	Awarding £30,000 annually
2000 44	Floring to the control of the Contro

2008-11 Elected as council member to the Society for General Microbiology

2007-11 Elected as council member of the Systematics Association and trustee of the

associated Charitable Association

In 2009 I became the Grants and Awards Officer – my administrative work for the BBSRC and NERC raises a £2,500 annual administration fee for the Systematics

Association's charitable trust

2005- Peer reviewer for: BBSRC, NERC, NSF, NSERC, ANR, ERC, NASA postdoctoral

program, Trends in Genetics, Environmental Microbiology, Journal of Eukaryotic Microbiology, ISME Journal, Molecular Ecology, BMC Evolutionary Biology, BMC Genomics, Aquatic Biology, Molecular Biology and Evolution, Eukaryotic Cell, PLoS Genetics, Heredity, PNAS, Current Biology, Nature Microbiology, and Nature

# **Memberships of Professional Bodies**

2012	Fellow of the Linnean Society
2012	Member of the British Mycology Society
2008	Member of Society of General Microbiology
2009	Member of the Systematics Association

#### **Invited Presentations**

# > 50, selection highlighted here:

Aug. 2018	Hutner Award Lecture, International Society of Protistology, UBC, Vancouver,
	Canada
Nov. 2017	2017 Sir Julian Huxley Lecturer, The Systematics Association, The Linnean Society, London
March 2017	Joint Genome Institute, Users Meeting, California, USA
March 2017	Departmental Lecture, Plant & Microbial Biology, University of California Berkeley, California, USA
June 2016	Gordon Research Seminar, Marine Microbes, Girona, Spain
June 2016	Gordon Research Conference, Cellular & Molecular Fungal Biology, New Hampshire, USA
Dec. 2015	Public Bioscience Lecture, University of York, UK
Nov. 2014	Current Trends in Biomedicine Workshop: Comparative and Functional Genomics of fungal pathogens, Baeza, Spain
Oct. 2013	European Molecular Biology Organization (EMBO), Bangalore ambassadorial trip
Oct. 2013	EMBO meeting of Comparative Genomics, San Feliu de Guixols (Costa Brava), Spain
Sep. 2013	Berkeley Lecture, British Society of Mycology, Cardiff, Wales, UK

July 2013	International Congress of Protistology, Vancouver, British Colombia, Canada
May 2013	Society of Molecular Biology and Evolution, University of California Davis, USA
May 2013	Canadian Institute for Advances Research meeting, Whistler, Canada
Dec. 2012	Departmental Seminar, Biochemistry, University of Cambridge
July 2012	American Mycology Society Annual Meeting, Yale, Connecticut, USA
June 2012	American Society of Microbiology, 112 <sup>th</sup> General Meeting, San Francisco, USA
May 2012	Canadian Institute for Advances Research meeting, Quebec, Canada
Feb. 2012	Departmental Seminar Genetics, Ecology and Evolution, University College London
Feb. 2012	Plenary Talk, Status workshop, Robert Koch Institute, Berlin
Oct. 2011	EMBO meeting of Comparative Genomics, San Feliu de Guixols (Costa Brava), Spain
July 2010	Gordon Conference, Marine Microbes, New Hampshire, USA
Feb. 2010	American Society of Limnology and Oceanography: Ocean Sciences Meeting, Portland, Oregon, USA
July 2009	Presidents' Medal Lecture, Society of Experimental Biology, Glasgow, UK
March 2009	Plenary talk, 25th Fungal Genetics Conference, Asilomar, USA
Meetings Organized	
2023	Chair/Organizer: EMBO/EMBL Symposia, The Mechanics of Endosymbiosis, Heidelberg, Germany
Nov. 2018	Chair/Organizer: Single Cell Ecology. Royal Society Hooke Symposium, London/Buckinghamshire, UK
Feb. 2018	Chair/Organizer: Using genomic comparisons to understand cellular complexity in our ancestors, Royal Society International Scientific Seminar, Buckinghamshire, UK.
Sep. 2016	Session Chair/Organizer: Fungal genome biology and evolution, British Mycology Society. Exeter, UK
2014-2018	Organizing committee: EMBO meeting of Comparative Genomics, San Feliu de Guixols (Costa Brava), Spain
Sept. 2011	Chair/Organizer: Horizontal gene flow & evolution, Society of General Microbiology, York, UK
March 2010	Chair/Organizer: Microbiology of Oceans, Society of General Microbiology, Edinburgh, UK
Dec. 2009	Chair/Organizer: Young Systematists' Forum, Systematics Association, Natural History Museum, London, UK
Sept. 2009	Chair/Organizer: Darwin's tree of life, Society of General Microbiology, Edinburgh,

# **Research Visits and Expeditions**

October 2018 Sampling expedition to Panamanian Rain Forests.

Project: developing and testing new tools for disease assessment in captured and wild endangered frog populations (Funded by Royal Society / GCRF challenge grant)

April 2015-19 Sampling expedition to Curação, Dutch Antilles

Project: Sampling diseased soft corals (Funded by CIFAR program grant and NERC

OMICS grant)

Sept. 2011 Research visit to Dr A. Worden, Monterey Bay Aquarium Research Institute, USA

Project: Molecular Adaptations in Deep Sea Fungi

Expedition: deep-sea sampling using remote operated vehicles (Funded by a Royal

Society Small Grant).

Sept.-Oct. 2009 Research visit to Dr F. Not, Station Biologique de Roscoff, France

Project: novel algae in European oceans (Funded by the British Council).

Tadpole Doctor, Royal Society Public Engagement Fund (£6,000), working with

July-Aug. 2006 Sample collection expedition to Peruvian: deserts, mountains and rainforest

#### **Special Editorial Roles**

Sep. 2020

Richards, T. A., Massana, R. Hall, N., Single Cell Ecology, Special Issue, Phil. Trans. Roy. Soc. B. 2019

Wideman, J. G., **Richards, T. A.,** *Current Opinion in Genetics & Development: Evolutionary Genetics.* 2019; Vol. 58-59

# Education, media and public outreach activities (examples)

Зер. 2020	multiple stakeholders to understand the distribution of tadpole pathogens in the UK and imported aquarium animals.
Aug. 2015	Tadpole infection work featured with quotes across several news outlets including New York Times and the Guardian
July 2015	Lecture to School forum 'Britain Needs Biosciences' on microbial 'eyes'
Jan. 2013	Interview for NERC Planet Earth Pod Cast on our environmental DNA work
May 2011	Interview for BBC Radio 4 Material World on our identification of novel fungi
May 2011	Interview for BBC Science news webpage on our identification of novel fungi
May 2011	Interview for National Public Radio USA on our identification of novel fungi
May 2011	Interview for Nature Pod Cast on our identification of novel fungi
Sept. 2009	Feature on Horizontal Gene Transfer in plants for education outreach journal <i>Scope</i>
2009	I took part in the British Council's Science for Schools Initiative in Brittany, France
Annually	Our laboratory regularly hosts school-age and undergraduate work experience students
July 2008	Presentation to Sir David Attenborough part of University of Exeter Honorary Graduands' reception on the tree of life

# **Teaching and Project Supervision**

2006- Supervision of fifteen Postdoctoral Fellows/Scientists

2005- Supervision to completion of nine PhD students

2013-2020	Visiting Lecturer, University College London
2005-10	Supervision of ten M.Sc./M.Res. projects (four to distinction, one to merit)
2005-19	I teach a range of lectures across the University of Oxford undergraduate syllabus on evolution and ecology, genomics and eukaryotic microbiology
2003-06	University of Oxford M.Sc. Integrative Bioscience (Molecular Biology Course Codirector)

### Stewardship

**External Examination.** PhD dissertations examined at the Universities of: Exeter UK, Sheffield UK, University College London UK, Maynooth Ireland, Uppsala Sweden, Barcelona Spain (x 3), Dalhousie Canada, and Oslo Norway.

## Institutional Responsibility.

At Exeter, I have acted as departmental academic lead for evolutionary biology, line managing nine academic staff including "tenure" progression and promotion. I was the lead organiser for Exeter Campus' Bioscience UK Universities Research Excellence Framework (REF) UoA5 submission and undertake several Director of Research roles. I have a formal mentorship for seven junior faculty members. I have sat on the Departmental Research Committee and the Living Systems Institute management committee.

**At Oxford**, I have taken charge of the Zoology Departments representation on the University wide NERC DTP management committee to cover for Covid-19 long term sick leave. I have taken on the joint role of managing the new 4<sup>th</sup> year biology program.

**Mentorship.** I have mentored numerous individuals to award of independent research fellowships (for example three Royal Society University Research fellowships and one CNRS fellowship position).

#### **Grants Awarded**

Grant type	<b>Title</b> (further information [PI – funding])	End date
Wellcome Trust Sanger Institute,	Ciliate and symbiont genome sequencing initiative	Aug. 2023
Moore Foundation, Aquatic Systems Symbiosis Genome Sequencing Initiative	Large-scale genome sequencing initiative to sequence ciliate protist genomes and their endosymbionts (project partner lab, no direct funding)	
DOE JGI, CSP Functional Genomics.	Development of a competitive transporter phenotype barcoding (CTP-Bar) assay for testing nutrient uptake among uncultivated microbes.	Submitted
	This project will provide proof of concept of a methodology that will allow mass comparisons of transporter uptake substrate diversity (project partner lab, no direct funding)	
Marie Curie Fellowship Grant, awarded to Luis Javier Galindo	FungEye: Characterization of the architecture, composition and evolution of a novel light perception organelle in an emerging model fungus (PI, funding = €213,000)	Aug. 2023
Moore Foundation, Aquatic Systems Initiative's symbiosis model systems (SMS) solicitation	Develop new genetic manipulations systems in endosymbiotic algae to track interaction dynamics in host ciliates (PI, funding = \$290,000)	Nov. 2022

Wellcome Trust, Discretionary	Darwin Tree of Life Project (phase 1).	July 2022
Award	Large consortium grant led by the Sanger Institute with the aim of sequencing large representation of all eukaryotic species in the UK.	
	I am responsible along with the Earlham Institute for UK protist sequencing (Co/Associate-PI, funding = £480,000)	
ERC Consolidator Grant	CELL-in-CELL. Understanding host cellular systems that drive an endosymbiotic interaction.	June 2024
	Developing systems biology approaches for understanding the cellular systems that control and allow endosymbiotic interactions (PI, funding = €2,600,000).	
NERC & STFC 'omics workshop grant	Workshop for 'omics methodology development: use of secretome enriched meta-transcriptome sequencing for understanding interactions in diseased corals	Dec. 2019
	Working together in an international community we will develop new methods for understanding how corals interact with their disease microbiome through the diversity of secreted proteins (PI, funding = £57,593).	
Royal Society / GCRF challenge grant	Assessing protist pathogen threats to endangered ecological keystone frog species of Panama.	July 2019
	Developing field diagnostics for tracking protists infections of frogs (PI, funding = $\$82,100$ ).	
Newton Fellowship Grant, awarded to Elisabet Alacid-	'Omics' and environmental approaches to study host-parasite interactions in dinoflagellate blooms.	Mar. 2019
Fernandez	Using multiple 'omics tools to understand complex heterotrophic interactions in the ocean. (Supervisor of fellowship = £113,000).	
Moore Foundation, Marine Microbe initiative, program grant	Transporter function and kinetics in uncultivated marine microbes.	Jan. 2021
	Developing protein functional analysis methods for studying nutrient transporters of uncultivated microbes (PI, Funding = \$1015,000).	
NERC grant	Calibrating eDNA Tools for Biodiversity Monitoring in the Ocean.	Jan.2020
	Developing eDNA techniques to understand ecosystem function and community diversity (Co-PI, Funding = £ $238,948$ ).	
Marie Curie International Training Network Grant	SINGEK: Promoting SINgle cell Genomics to explore the ecology and evolution of hidden microEuKaryotes.	Jan. 2020

	Developing cross European expertise to study microbial eukaryotes directly from the environment using single cell sequencing approaches (Co-PI, Funding = €546,575)	
Marie Curie Fellowship Grant, awarded to Estelle Kilias	Significant or trivial: Fungi in Polar ecosystems (F-POLE).	Sep. 2018
	Using environmental 'omic'-based approaches to investigate the diversity, abundance and role of fungi in the marine environment (Supervisor of fellowship = €183,454.80)	
Philip Leverhulme award	Personal Award for research development.	Nov. 2017
	Develop new tools to link genomic data with phenotype analysis of individual microbes and how they interact in communities (PI, funding = £100,000)	
Royal Society University Research Fellowship (Renewed with	Dissecting a nascent phototrophic endosymbiotic interaction.	Dec. 2023
enhancement awards)	Using transcriptomics, proteomics and reverse genetics to investigate cellular functions tied to early interactions in Paramecium bursaria photosynthetic endosymbiosis (fellowship funding = £1,034,000)	
Department of Energy (DOE) Joint Genome Institute Community Sequencing Program	CSP: Revealing the ecological function of uncultured fungal dark matter in freshwater ecosystems using single cell genomics.	Oct. 2018
	Community sequencing program to conduct single cell genome sequencing of 'chytrid' fungi from natural environmental samples. (Co-PI)	
EMBO Long Term Fellowship, awarded to Jeremy Wideman	Comparative genomics of diatoms and Bolidophyceae: insight into the evolution of one of Earth's most productive phototrophs.	Feb. 2017
	Taking a single cell genome sequencing approach, we explored the genome diversity of marine protists and algae (supervisor of fellowship – funding = £70,000)	
Department of Energy (DOE) Joint	TDP: Life on the Darkside.	June 2017
Genome Institute Technology Development Program	Technology Development Program partnership to develop methods for targeted isolation and sequencing of eukaryotic single cell isolates. (Co-PI)	
Leverhulme Project Grant	Ancestral gene repertoires at the dawn & diversification of the Eukaryotes.	Nov. 2017
	Using ancestral gene compliment reconstruction, we will identify the gene complement of the Last Eukaryotic Common Ancestor (PI, funding = £ 212,986)	

Did horizontal gene transfer 'rewire' ocean microbial metabolic networks?	Mar. 2016
Aim of this project is to look at the impact of gene transfer on the metabolic network encoded by microbial eukaryotes in the marine environment (supervisor of fellowship – funding = £82,875)	
Evolution of endosymbiosis.	Jan. 2016
Using transcriptomics, proteomics and reverse genetics to investigate cellular functions tied to early interactions in Paramecium bursaria photosynthetic endosymbiosis (PI funding = €70,000)	
Life on the dark side: complex trophic interactions of marine microbial eukaryotes.	Apr. 2016
Using meta-transcriptomics and single cell isolation and genome sequencing we will investigate the role of eukaryotic microbes in marine ecosystems (PI funding = \$422,909)	
Emerging Protist Parasites of Frogs: Genome and cellular biology of a previously unrecognized parasitic group.	Aug. 2015
Using transcriptome and genome sequencing methods combined with cell biology with the aim of identifying host pathogen interactions (Supervisor of fellowship – funding = £50,000)	
"PARAFROGS" Emerging Protist Parasites of Frogs: Global Prevalence and Host/Parasite Interaction.	Aug. 2014
Using molecular methods to identify the global prevalence and host range of this parasite group (Supervisor of fellowship – funding = € 209,033.40)	
Global evolutionary complexity of freshwater alveolates: a new threat to frogs?	Jan. 2012
Using environmental DNA methods to explore the diversity and host specificity of novel group of alveolates that infect frogs (PI - funding = £26,500)	
Investigating active eukaryotic microbial communities in deep-sea environments.	Jan. 2012
In collaboration with Monterey Bay Aquarium Research Institute, USA, we have developed and are pilot testing a novel approach to sampling community RNA from deep-sea sediments for meta-transcriptome sequencing (PI – funding = £14,780)	
	Aim of this project is to look at the impact of gene transfer on the metabolic network encoded by microbial eukaryotes in the marine environment (supervisor of fellowship – funding = £82,875)  Evolution of endosymbiosis.  Using transcriptomics, proteomics and reverse genetics to investigate cellular functions tied to early interactions in Paramecium bursaria photosynthetic endosymbiosis (Pl funding = €70,000)  Life on the dark side: complex trophic interactions of marine microbial eukaryotes.  Using meta-transcriptomics and single cell isolation and genome sequencing we will investigate the role of eukaryotic microbes in marine ecosystems (Pl funding = \$422,909)  Emerging Protist Parasites of Frogs: Genome and cellular biology of a previously unrecognized parasitic group.  Using transcriptome and genome sequencing methods combined with cell biology with the aim of identifying host pathogen interactions (Supervisor of fellowship – funding = £50,000)  "PARAFROGS" Emerging Protist Parasites of Frogs: Global Prevalence and Host/Parasite Interaction.  Using molecular methods to identify the global prevalence and host range of this parasite group (Supervisor of fellowship – funding = €209,033.40)  Global evolutionary complexity of freshwater alveolates: a new threat to frogs?  Using environmental DNA methods to explore the diversity and host specificity of novel group of alveolates that infect frogs (PI - funding = £26,500)  Investigating active eukaryotic microbial communities in deep-sea environments.  In collaboration with Monterey Bay Aquarium Research Institute, USA, we have developed and are pilot testing a novel approach to sampling community RNA from deep-sea sediments for meta-transcriptome

FP6 Biodiversa ERA-net	Biodiversity of Marine EuKaryotes (BioMarKs).	Jan. 2013
	European collaboration using 454 diversity tag sequencing to investigate the complexity of marine protist and fungal communities (Co-I funding = €265,000)	
BBSRC New Investigator Grant	The diversity and phylogeny of molecular motor proteins and fungal cell evolution.	Oct. 2012
	Integrating next generation genome sequencing technologies, we use comparative genomics to investigate major events in fungal and eukaryotic cell evolution (PI – funding = £402,281)	
British Academy Franco-British research alliance grant:	Tracking the diversity and abundance of phototrophic life in the oceans.	Dec. 2010
	Collaboration with Fabrice Not to investigate the evolutionary diversity of marine algae using next generation sequencing methodology (PI – funding = £4,000 & €5,000)	
NERC Grant	Diversity, identity and ecological role of a novel fungal super clade.	Nov. 2010
	Using environmental DNA we identified the phylogenetic postion and cell biology of a new highly diverse microbial group branching with the fungi (PI – funding = £143,846)	
Small grants for short studentships	7 x Nuffield Foundation, 4 x Royal Society funded Studenthips, 3 x Systematics Research Fund, 4 x SGM Vacation Studentship Grants.	Ongoing since 2006
	Used in the large part to support summer studentships (PI – funding total = $^{\sim}$ £20,000)	
BBSRC/NERC CoSyst grant	Molecular diversity of microbial eukaryotes using a large-scale parallel tag sequencing strategy.	Dec. 2010
	This project developed the 454-tag sequencing for investigating the diversity of microbial eukaryotes (PI – funding total = £19,608)	
Leverhulme Early Career Fellowship	Comparative genomics and eukaryote cell evolution. $(PI - funding\ total = £50,000)$	Dec2009

# **Publications**

# Refereed Journal Articles (\*corresponding author)

- Milner, D. S., Wideman, J. G.\*, Stairs, C. W., Dunn, C. D., Richards. T. A.\*, A functional bacterial-derived restriction modification system in the mitochondrion of a heterotrophic protist. *Invited resubmission. PLoS Biol.* Available at https://www.biorxiv.org/content/10.1101/2021.02.01.429123v1.abstract
- Jenkins, B. H.\*, Maguire, F., Leonard, G., Eaton, J. D., West, S., Housden, B. E., Milner, D. S., Richards. T. A., Characterisation of the RNA-interference pathway as a tool for genetics in the nascent phototrophic

endosymbiosis, Paramecium bursaria. Invited resubmission, Invited contribution to special collection 'New Talent in Life Sciences'. Roy. Soc. Open Science.

Available at https://www.biorxiv.org/content/10.1101/2020.12.16.423098v2.full

- Smilansky, V., Jirku, M., Milner, D. S., Ibáñez, R., Gratwicke, B., Nicholls, A., Lukeš, J., Chambouvet, A., Richards. T. A.\*, Expanded geographic and host tadpole associations of the Severe Perkinsea infection group. *Invited resubmission, Roy. Soc. Biology Letters*.
- Smilansky, V.\*, Chambouvet, A., Reeves, M., **Richards. T. A.,** Milner, D. S.\*, A novel duplex qPCR assay for stepwise detection of multiple Perkinsea protistan infections of amphibian tissues. *In Press, Roy. Soc. Open Science*.
- Kilias, E. S.\*, Junges, L. Supraha, L., Leonard, G. Metfies, K., **Richards. T. A.**, Chytrid fungi distribution and co-occurrence with diatoms in the Arctic Ocean is correlated with sea ice melt. *In Press, Com. Biol.*
- Rodríguez-Martínez, R\*, Leonard, G., Milner D. S., Sudek S., Conway, M., Moore, K., Hudson, T., Mahé, F., Keeling, P. J., Santoro, A. E., Worden, A. Z., Richards, T. A.\*, Controlled sampling of ribosomally active protistan diversity in sediment-surface layers identifies putative players in the marine carbon sink. *ISME J.* 2020. https://doi.org/10.1038/s41396-019-0581-y
- Wideman, J. G.\*, Monier, A., Rodríguez-Martínez, R., Leonard, G., Cook, E., Poirier, C., Maguire, F., Milner, D., Irwin, N. A. T., Moore, K., Santoro, A. E., Keeling, P. J., Worden, A. Z., Richards, T. A.\*, Unexpected mitochondrial genome diversity revealed by targeted single-cell genomics of heterotrophic flagellated protists. 2020; 5 154-165 Nature Microbiology.
- Chambouvet, A., Monier, A. Maguire, F., Itoïz, S., del Campo, J., Elies, P., Edvardsen, B., Wenche, E., Richards, T. A., Intracellular infection of diverse diatoms by an evolutionary distinct relative of the Fungi. In Press: *Curr. Biol.* 2019: **29**(23):4093-4101 doi.org/10.1016/j.cub.2019.09.074
- Wideman, J. G.\*, Lax, G., Leonard, G., Milner, D. S., Rodríguez-Matinez, R., Simpson, G. B. Simpson, Richards, T. A., A single-cell genome reveals diplonemid-like ancestry of kinetoplastid mitochondria gene structure. *Phil. Trans. Roy. Soc. B.* 2019; 374(1786) doi/10.1098/rstb.2019.0100
- Needham, D. M., Yoshizawa, S., Hosaka, T., Poirier, C., Choi, C. J., Hehenberger, E., Irwin, N. A. T., Wilken, S., Yung, C-M., Bachy, C., Kurihara, R., Nakajima, Y., Kojima, K., Kimura-Someya, T., Leonard, G., Malmstrom, R. R., Mende, D. R., Olson, D. K., Sudo, Y., Sudek, S., Richards, T. A., DeLong, E. F., Keeling, P. J., Santoro, A. E., Shirouzu, M., Iwasaki, W., Worden, A. Z., A distinct lineage of giant viruses brings a rhodopsin photosystem to unicellular marine predators. *Proc. Natl. Acad. Sci. USA.* 2019; 116(41):20574-20583.
- Del Campo, J., Heger, T., Rodríguez-Martínez, R., Worden, A. Z., Richards, T. A., Massana, R., Keeling, P. J., Assessing the diversity and distribution of apicomplexans in host and free-living environments using high-throughput amplicon data and a phylogenetically informed reference framework. Front. Microbiol. doi:10.3389/fmicb.2019.02373.
- Milner, D. S.\*, Attah, V., Cook, E., Maguire, F., Savory, F., Morrison, M., Müller, C. A., Foster, P. G., Talbot, N. J., Leonard, G., & Richards, T. A.\*, Environment-dependent fitness gains can be driven by horizontal gene transfer of transporter-encoding genes in fungi. *Proc. Natl. Acad. Sci. USA*. 2019; 116(12):5613-5622.
- Strassert, J. F., Hehenberger, E., del Campo, J., Okamoto, N., Kolisko, M., Richards, T. A., Worden, A. Z., Santoro, A. E., & Keeling, P. J.\* Phylogeny, evidence for a cryptic plastid, and distribution of *Chytriodinium* parasites (Dinophyceae) infecting copepods. *J. Euk. Micro*; 2018; https://doi.org/10.1111/jeu.12701
- Savory, F. R., Milner, D. S., Miles, D. C., & **Richards, T. A.\***, Ancestral function and diversification of a horizontally acquired oomycete carboxylic acid transporter. *Mol. Biol. and Evol.* 2018; **msy082**.

- Wideman, J. R.\*, Balacco, D. L., Fieblinger, T., **Richards, T. A.** PDZD8 is not the 'functional ortholog' of Mmm1, it is a paralog. *F1000 Research*; 2018; **7**:1088
- Orsi, W. D., Wilken, S., del Campo, J., Heger, T., James, E., **Richards, T. A.**, Keeling, P. J., Worden, A. Z., & Santoro, A. E., Identifying protist consumers of photosynthetic picoeukaryotes in the surface ocean using stable isotope probing. *Environ. Microbiol.* 2018; **20**(2):815-827.
- Orsi, W. D.\*, **Richards, T. A.**, & Francis W. R., Predicted microbial secretomes and their target substrates in marine sediment. *Nature Microbiology*, 2018; **3**(1):32.
- Okamoto, N., Gawryluk, R. M., del Campo, J., Strassert, J. F., Lukeš, J., Richards, T. A., Worden, A. Z., Santoro, A. E., & Keeling, P. J.\*. A revised taxonomy of diplonemids including the eupelagonemidae n. fam. and a type species, *Eupelagonema oceanica* n. gen. & sp. *J. Euk. Micro.*; 2018; https://doi.org/10.1111/jeu.12679
- Leonard, G., Labarre, A., Milner, D. S., Monier, A., Soanes, D., Wideman, J. G., Maguire, F., Stevens, S., Sain, D., Grau-Bové, X., Sebé-Pedrós, A., Stajich. J. E., Paszkiewicz, K., Brown, M. W., Hall, N., Wickstead, B., Richards, T.A.\*, Comparative genomic analysis of the 'pseudofungus' *Hyphochytrium catenoides*. *Open biology*, 2018; 8(1):170184.
- Guo, J., Wilken, S., Jimenez, V., Choi, C. J., Ansong, C., Dannebaum, R., Sudek, L., Milner, D. S., Bachy, C., Reistetter, E. N., Elrod, V. A., Klimov, D., Purvine, S. O., Wei, C. L., Kunde-Ramamoorthy, G., Richards, T. A., Goodenough, U., Smit, R. D., Callister, S. J., Worden, A. Z.\*, Specialized proteomic responses and an ancient photoprotection mechanism sustain marine green algal growth during phosphate limitation. *Nature Microbiology*, 2018; 3(7):781.
- Gomes-Vieira, A. L., Wideman, J. G., Paes-Vieira, L., Gomes, S. L., Richards, T. A., & Meyer-Fernandes, J. R. \*, Evolutionary conservation of a core fungal phosphate homeostasis pathway coupled to development in *Blastocladiella emersonii*. Fungal Genetics and Biology, 2018; 115:20-32.
- Strassert JF, Karnkowska A, Hehenberger E, del Campo J, Kolisko M, Okamoto N, Burki F, Janouškovec J, Poirier C, Leonard G, Hallam. S. J., Richards, T. A., Worden, A. Z., Santoro, A. E., Keeling, P. J.\*. Single cell genomics of uncultured marine alveolates shows paraphyly of basal dinoflagellates. *The ISME journal*, 2017; 12(1):304.
- Monier A.\*, Chambouvet A., Milner D. S., Attah V., Terrado R., Lovejoy C., Moreau H., Santoro A. E.,
  Derelle É., & Richards T. A.\* Host-derived viral transporter protein for nitrogen uptake in infected
  marine phytoplankton. *Proc. Natl. Acad. Sci. USA*, 2017; 114(36):E7489-E7498.
- Grau-Bove, X., Torruella, G., Donachie, S., Suga, H., Leonard, G., **Richards, T. A.**, & Ruiz-Trillo, I.\*. Dynamics of genomic innovation in the unicellular ancestry of animals. *Elife*, 2017; **6**:e26036.
- Gawryluk, R. M. R., del Campo, J., Okamoto, N., Strassert, J. F. H., Lukeš, J., Richards, T. A., Worden, A. Z., Santoro, A. E., Keeling, P. J.\*. Morphological identification and single-cell genomics of marine diplonemids. *Curr. Biol.*, 2016; 26 (22), 3053-3059.
- Forster, D., Dunthorn, M., Mahé, F., Dolan, J. R., Audic, S., Bass, D., Bittner, L., Boutte, C., Christen, R., Claverie, J. M., Decelle, J., Edvardsen, B., Egge, E., Eikrem, W., Gobet, A., Kooistra, W. H., Logares, R., Massana, R., Montresor, M., Not, F., Ogata, H., Pawlowski, J., Pernice, M. C., Romac, S., Shalchian-Tabrizi, K., Simon, N., Richards, T. A., Santini, S., Sarno, D., Siano, R., Vaulot, D., Wincker, P., Zingone, A., de Vargas, C., Stoeck, T. Benthic protists: the under-charted majority. FEMS Microbiology Ecology; 2016; 92 (8): fiw120.
- Monier A., Worden, A. Z., Richards T. A. Phylogenetic diversity and biogeography of the Mamiellophyceae lineage of eukaryotic phytoplankton across the oceans *Environmental Microbiology Reports*; 2016:

- Masachis, S., Segorbe, D., Turrà, D., Leon-Ruiz, M., Fürst, U., El Ghalid, M., Leonard, G., López-Berges, M. S., Richards, T.A., Felix, G., Di Pietro\*, A. A, Fungal pathogen secretes plant alkalinizing peptides to increase infection. *Nature Microbiology*; 2016: 1:16043.
- Orsi, W. D., Smith, J. M., Liu, S., Liu, Z., Sakamoto, C. M., Wilken, S., Poirier, C., **Richards, T. A.**, Keeling, P. J., Worden, A. Z., Santoro, A.E. Diverse, uncultivated bacteria and archaea underlying the cycling of dissolved protein in the ocean. *The ISME Journal*; 2016: **10**, 2158-2173.
- Chambouvet, A.\*, Valigurová, A., Mesquita, L., **Richards, T. A.**, Jirků, M.\* . *Nematopsis temporariae* (Gregarinasina, Apicomplexa, Alveolata) is an intracellular infectious agent of tadpole livers. *Environmental Microbiology Reports*. 2016: doi: 10.1111/1758-2229.12421
- Avelar, G. M., Glaser, T., Leonard, G., Richards, T.A., Ulrich, H., Gomes, S.L.\*. A cyclic GMP-dependent K+ channel in the blastocladiomycete fungus *Blastocladiella emersonii*. *Eukaryotic cell* 2015 14(9): 958-963.
- Massana, R.,... ...25 authors... ...Richards, T.A.\*... ...7 authors... ...De Vargas, C., (2015). Marine protist diversity in European coastal waters and sediments as revealed by high-throughput sequencing." Environmental Microbiology 2015; 17(10): 4035-4049.
- Richards, T.A\*, Leonard, G., Mahé, F., del Campo, J., Romac, S., Jones, M.D.M., Maguire, F., Dunthorn, M., De Vargas, C., Massana, R., Chambouvet, A., Molecular diversity and distribution of marine fungi across 130 European environmental samples. *Proc. Roy. Soc. B.* 2015; DOI: 10.1098/rspb.2015.2243
- Chambouvet, A., Gower, D.J., Jirků, M., Yabsley, M.J., Davis, A.K., Leonard, G., Maguire, F., Doherty-Boner T.M., Bittencourt-Silva, G.B, Wilkinson, W., Richards, T.A\*, Cryptic infection of a broad taxonomic and geographic diversity of tadpoles by Perkinsea protists, *Proc. Natl. Acad. Sci. USA*. 2015; 112(34) E4743-E4751
- Orsi, W.D.\*, **Richards, T.A.,** Santoro, A.E., Cellular maintenance processes that potentially underpin the survival of subseafloor fungi over geological timescales, *Estuarine, Coastal and Shelf Science,* 2015; doi:10.1016/j.ecss.2015.04.009.
- del Campo, J.\*, Mallo, D., Massana, R., Vargas, C., **Richards, T.A.**, Ruiz-Trillo, I., Diversity and distribution of unicellular opisthokonts along the European coast analysed using high-throughput sequencing, *Env. Micro.* 2015; doi: 10.1111/1462-2920.12759.
- Misner, I., Blouin, N., Leonard, G., **Richards, T.A.**, Lane, C.E.\*, The secreted proteins of *Achlya hypogyna* and *Thraustotheca clavata* identify the ancestral oomycete secretome and reveal gene acquisitions by horizontal gene transfer. *Genome Biol. Evol.* 2015; 7(1): 120-135.
- Maguire, F., Henriquez, F.L., Leonard, G., Dacks, J.B., Brown, M.W., **Richards, T.A.\***, Complex patterns of gene fission in the eukaryotic folate biosynthesis pathway. *Genome Biol. Evol.* 2014; 23(10): 2709-2720.
- Avelar, G. A., Schumacher, R. I., Zaini, P. A., Leonard, G., Richards, T. A. \*, Gomes, S. L\*. A rhodopsin-guanylyl cyclase gene fusion functions in visual perception in a fungus. *Curr. Biol.* 2014; 24(11); 1232-1240.
- Chambouvet, A. \*, Berney, C., Romac, S., Audic, S., Maguire, F., De Vargas, C., Richards, T.A., Diverse
  molecular signatures for ribosomally 'active' Perkinsea in marine sediments. *BMC Microbiology* 2014;
  14(1): 110.
- Logares, R.\*,... 25 authors... ... **Richards, T.A.**, de Vargas, C., Massana, R., Patterns of rare and abundant marine microbial eukaryotes. *Curr. Biol.* 2014; 24(8): 813-821 [Cover].
- Sebé-Pedrós, A., Grau-Bové, X., **Richards, T.A.**, Ruiz-Trillo, I.\*, Evolution and classification of myosins, a paneukaryotic whole-genome approach. *Genome Biol. Evol.* 2014; 6 290-305.

- Read, B. A... ...33 authors... ...Richards, T.A.\*... ...2 authors... ...Grigoriev, I. V., Pan genome of the phytoplankton *Emiliania* underpins its global distribution. *Nature* 2013; **499**(7457): 209-213.
- Leonard, G. & Richards, T.A.\*, Genome-scale comparative analysis of gene fusions, gene fissions and the fungal tree of life. *Proc. Natl. Acad. Sci. USA*. 2012; **109**(52) 21402-21407.
- Curtis, B.A... ...47 authors... *Liu, Y.*, **Richards, T. A.** ...25 authors... ...Archibald, J.M.\*, Cryptophyte and chlorarachniophyte nuclear genomes reveal evolutionary mosaicism and fate of nucleomorphs. *Nature*, 2012; **492**(742) 59-65.
- Jones, M.D.M., **Richards, T.A.**, Hawksworth, D., Bass, D.\* Validation and justification of the phylum name *Cryptomycota* phyl. nov. *IMA Fungus*. 2011; **2**(2) 173-175.
- Richards, T.A.\*, Soanes, D., Jones, M., Vasieva, O., Leonard, G., Paszkiewicz, K., Foster, P., Hall, N., Talbot, N., Horizontal gene transfer facilitated the evolution of plant parasitic mechanisms in the oomycetes. *Proc. Natl. Acad. Sci. USA*, 2011; **108**(37): 15258-15263.
- Jones, M.D.M., Forn, I., Gadelha, C., Bass, D., Massana, R., & Richards, T.A.\* Discovery of novel intermediate forms redefines the fungal tree of life. *Nature*. 2011; 474(7350): 200-203.
- Kim, E., Harrison, J.W., Sudek, S., Jones, M.D., Wilcox, H.M., **Richards, T.A.\***, Worden, A.Z.\*, Archibald, J.M.\*, Newly identified and diverse plastid-bearing branch on the eukaryotic tree of life. *Proc. Natl. Acad. Sci. USA*. 2011; **108**: 1496-1500.
- Wickstead, B.\*, Gull, K. & **Richards, T. A.**\*, Patterns of kinesin evolution reveal a complex ancestral eukaryote with a multifunctional cytoskeleton, *BMC Evol. Biol.* 2010; **10**: 110.
- Stoeck, T., Bass, D., Nebel, M., Christen, R., Jones, M.D.M., Hans-Werner Breiner & **Richards, T. A.\***, Parallel tag environmental DNA sequencing reveals a highly complex eukaryote community. *Mol. Ecol.* 2010; **19**: 21-31, Speicial Edition: Next Generation Ecology.
- Richards, T.A.\*, Soanes, D. M., Foster, P. G., Leonard, G., Thornton, C. R., & Talbot, N. J., Phylogenomic analysis demonstrates a pattern of rare and ancient horizontal gene transfer between plants and fungi. *The Plant Cell*. 2009; **21**(7): 1897-1911.
- Leonard, G., Stevens, J. R., & Richards, T. A.\*, REFGEN and TREENAMER: Automated sequence data handling for phylogenetic analysis in the genomic era. *Evolutionary Bioinformatics*. 2009; **5**: 1-4.
- Bass, D., Brown, N., Mackenzie-Dodds, J., Dyal, P., Nierzwicki-Bauer, S. A., Vepritskiy, A. A., & Richards, T.A. \*, A molecular perspective on ecological differentiation and biogeography of cyclotrichiid ciliates. *J. Euk. Micro*. 2009; 56(6): 559-567.
- Liu. L., **Richards, T.A.**, & Aves, S. J.\*, Ancient diversification of eukaryotic MCM DNA replication proteins. *BMC Evolutionary Biology*. 2009; **9**:60.
- Hofmann, W.A., **Richards, T.A.\***, & de Lanerolle, P.\*, Ancient animal ancestry for nuclear myosin. *J. Cell Sci.* 2009; **122**(Pt 5): 636-643.
- Bass, D., Howe, A., Barton, H., Brown, N., Demidova, M., Michelle, H., Li, L., Sanders, H., Watkinson, S., Willcock, S., Richards, T.A.\*, Yeast forms dominate fungal diversity in the deep oceans. *Proc. Roy. Soc. B*. 2007; 274: 3069-3077.
- Bass, D.\*, **Richards, T.A.,** Matthai, L., Marsh, V., Cavalier-Smith, T., Globally dispersed and endemic genotypes in microbial eukaryotes. *BMC Evol. Biol.* 2007; **7:** 162.
- Ferguson, D.J.P.\*, Campbell, S.A., Henriquez, F.L., Phan, L., Mui, E., **Richards, T.A.**, Muench, S.P., Allary, M., Lu, J.Z., Prigge, S.T., Tomley, F., Shirley, M.W., Rice, D.W., McLeod, R., Roberts, C.W., Enzymes of type II fatty acid synthesis and apicoplast differentation and division in *Eimeria tenella*. *Int. J. Parasitol*. 2007; **37**: 33-51.

- **Richards, T.A.**, Dacks, J.B., Jenkinson, J.M., Thornton, C.R., Talbot, N.J.\*, Evolution of filamentous plant pathogens: gene exchange across eukaryotic kingdoms. *Curr. Biol.* 2006; **16**: 1857-1864.
- Richards, T.A., Dacks, J.B., Campbell, S.A., Blanchard, J.L., Foster, P.G., McLeod, R., Roberts, C.W.\*, Evolutionary origins of the eukaryotic shikimate pathway: gene fusions, horizontal gene transfer, and endosymbiotic replacements. *Eukaryot. Cell.* 2006; **5**: 1517-1531 [cover].
- Richards, T.A., van der Giezen, M\*. Evolution of the Isd11-IscS complex reveals a single alphaproteobacterial endosymbiosis for all eukaryotes. *Mol. Biol. Evol.* 2006; **23**: 1341-1344.
- **Richards, T.A.\***, Cavalier-Smith, T., Myosin domain evolution and the primary divergence of eukaryotes. *Nature*. 2005; **436**: 1113-1118.
- **Richards, T.A.**, Vepritskiy, A.A., Gouliamova, D.E., Nierzwicki-Bauer, S.A.\*, The molecular diversity of freshwater picoeukaryotes from an oligotrophic lake reveals diverse, distinctive and globally dispersed lineages. *Environ. Microbiol.* 2005; **7**: 1413-1425.
- Campbell S.A., Richards T.A., Mui E.J., Samuel B.U., Coggins J.R., McLeod R., Roberts C.W.\*, 2004. A complete shikimate pathway in *Toxoplasma gondii*: an ancient eukaryotic innovation. *Int. J. Parasitol.* 34, 5-13.
- **Richards T.A.**, Hirt R.P., Williams B.A., Embley T.M.\*, 2003. Horizontal gene transfer and the evolution of parasitic protozoa. *Protist* **154**, 17-32.
- Baker R.H., Ashwell R.I.S., **Richards T.A**., Fowler K., Chapman T., Pomiankowski A.\*, 2001. Effects of multiple mating and male eye span on female reproductive output in the stalk-eyed fly, *Cyrtodiopsis dalmanni*. *Behav*. *Ecol.* **12**, 732-739.

### **Reviews, Opinions and Commentaries**

- Chambouvet, A.\*, Smilansky, V., Jirků, M., Isidoro-Ayza, M., Itoïz. S., Derelle, E., Monier, A., Gower, D. J., Wilkinson, M., Yabsley, M. J., Lukeš, J., Richards, T. A.\*, Diverse alveolate infections of tadpoles, a new threat to frogs, In Press: PLoS Pathogens.
- **Richards, T. A.\*** A tangled tale of innovation and personalities in the search for a tree of life (Book review). *Curr. Biol.* **30**(1): 5-7
- Brockhurst, M. A., Harrison, E., Hall, J. P. J., **Richards, T. A.,** McNally, A., MacLean, C., The ecology and evolution of pangenomes. *Curr. Biol.* **29**(20): R1094-R1103
- Richards, T. A.\*, Massana, R. Pagliara, S., Hall, N., Single cell Ecology, *Phil. Trans. Roy. Soc. B.* 2019: https://doi.org/10.1098/rstb.2019.0076.
- Richards, T. A.\*, McCutcheon, J. P., Coral symbiosis is a three-player game, *Nature*, 568(7750) 41.
- Jenkins, B., Richards, T. A.\*, Symbiosis: wolf lichens harbour a choir of fungi, Curr. Biol., 29 (3), R88-R90.
- Richards, T. A.\*, & Talbot, N. J., Osmotrophy. Curr. Biol. 2018, 28(20): R1179-R1180.
- Richards, T. A.\*, Leonard G., & Wideman J. G. What Defines the Kingdom Fungi? *Microbiology spectrum*, 2017; **5**(3).
- Richards, T. A.\* & Monier A, Tale of two taridgrades. *Proc. Natl. Acad. Sci. USA.* 2016; **113**(18): 4892-4894
- **Richards, T. A.\*** and Chambouvet, A. A role for fungi as parasites in the black box of marine trophic interactions. *Environ.l Micro. Rep.*, 2016; **8**(4): 429–430
- Richards, T.A.\* & Gomes, S.L.\*, How to build a microbial eye. Nature. 2015; 523: 166-167

- Savory, F., Leonard, G. & **Richards, T.A.\***, The Role of Horizontal Gene Transfer in the Evolution of the Oomycetes. *PLoS Pathogens*. 2015; **5**: e1004805.
- Soanes, D., & **Richards, T.A.\***, Horizontal gene transfer in eukaryotic plant pathogens. *Annu. Rev. Phytopathol.* 2014; 52: 583-614.
- Maguire, F., & **Richards, T.A.\***, Organelle evolution: a mosaic of 'mitochondrial' functions. *Curr. Biol.* 2014; 24(11): R518-20
- **Richards, T.A.\*** & Talbot N. J. Horizontal gene transfer in osmotrophs: playing with public goods. *Nature Reviews Microbiology*. 2013; **11**: 720–727
- **Richards, T.A.\***, Jones, M.D.M., Leonard, G., Bass, D. Marine Fungi: their ecology and molecular diversity. *Annual Review of Marine Science*. 2012; **4**: 495-522
- Bass, D.\*, **Richards, T.A.** Three reasons to re-evaluate fungal diversity 'on Earth and in the ocean' *Fungal Biology reviews*. 2011; **25**: 159-164
- Richards, T.A.\*, Leonard, G., Soanes, D., & Talbot, N. J. Gene transfer into the fungi. *Fungal Biology Reviews*. 2011; **21**(4): 98-110 (includes meta analysis)
- **Richards, T.A.** \* Genome evolution: horizontal movements in the fungi. *Curr. Biol.* 2011 **21**(4): R166-168.
- **Richards, T.A.\*** & Archibald, J.M. Cell evolution: gene transfer agents and the origin of mitochondria. *Curr. Biol.* 2011; **21**(3): R112-114
- Archibald, J.M.\* & Richards, T.A. Gene transfer: anything goes in plant mitochondria. BMC Biol. 2011; 8:
   147
- Soanes, D.M., Richards, T.A., Talbot, N.J.\*, Insights from sequencing fungal and oomycete genomes: what can we learn about plant disease and the evolution of pathogenicity? *Plant Cell*. 2007; 3318-3326.
- **Richards, T.A.** & Talbot, N.J.\*, Plant parasitic oomycetes such as *Phytophthora* species contain genes derived from three eukaryotic lineages. *Plant Sig. & Beh.* 2007; **2**: 112-114.
- **Richards, T.A.** & Bass, D.\*, Molecular screening of free-living microbial eukaryotes: diversity and distribution using a meta-analysis. *Curr. Opin. Microbiol.* 2005; **8**: 240-252.
- Henriquez, F.L., **Richards, T.A.**, Roberts, F., McLeod, R., Roberts, C.W.\*, The unusual mitochondrial compartment of *Cryptosporidium parvum*. *Trends Parasitol*. 2005; **21**: 68-74 (cover).

## **Book chapters**

- Chambouvet, A.\*, **Richards, T.A.**, Bass, D., & Neuhauser, S., Revealing microparasite diversity in aquatic environments using brute force molecular techniques and subtle microscopy. *Chapter 6: Parasite diversity and diversification: evolutionary ecology meets phylogenetics*; 2015; 93-116.
- Aves, S. J.\*, Liu, Y., **Richards, T. A.,** The eukaryotic replisome: a guide to protein structure and function. *Chapter 2: Evolutionary diversification of eukaryotic DNA replication machinery;* 2012; 19-35.
- Jones, M.D.M., **Richards, T.A.**\* Environmental DNA analysis and the expansion of the fungal tree of life. *Chapter 3: The Mycota*, 2011; **14:** 37-57.