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Supplementary Information

Environmental Science: Processes & Impacts

- 3 Pharmaceuticals and personal care products (PPCPs) in urban and suburban
- 4 rivers of Beijing, China: occurrence, source apportionment and potential
- 5 ecological risk

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Table S1 Information of the study area and WWTPs investigated

Watershed	Area (km²)	Population × 10 ³	WWTPs	Population serviced \times 10 ³ a	Daily flow (10 ³ m ³)	HRT (h)	SRT (d)	Secondary treatment
Qing River	210	3000	QWTP XJWTP	2000 500	400 80	13.5 11	12-15 15	A ² /O A ² /O
Liangshui River	624	4500	XHMWTP BWTP YJWTP	2415 400 	600 80 10	11 10.6	15 12-15 	A ² /O OD

QWTP, XJHWTP, XHMWTP and BWTP employ similar conventional treatment process: primary treatment to remove particles coupled with secondary biological treatment. YJWTP only have primary treatment equipment to remove particles without secondary biological treatment. For the secondary biological treatment process, QWTP, XJHWTP and XHMWTP employ anaerobic/anoxic/oxic (A²/O) activated sludge process, and BWTP employs oxidation ditch (OD). HRT: hydraulic retention time; SRT: solid retention time. ^a Population serviced by QWTP was estimated according to its capacity.

35 Table S2 Optimized ESI-MS/MS parameters for analysis of the analytes by MRM

			Retention time (min)	Precursor ion (m z ⁻¹)	Product	MS/MS parameters				
Compound	Acronym	ESI			ion (m z ⁻¹)	DP (V)	ED (V)	CEP	CE	CXP
					. , ,	DP (V)	EP (V)	(V)	(V)	(V)
Sulpiride	SP	+	2.34	342.0	112.1	65	5	8	22	2
Sulpinde	51	,	2.54	342.0	214.2	65	5	14.6	28	2
Trimethoprim	TP	+	5.23	291.2	230.3	70	8	20	30	2
тинешориш	11	·	3.23	2)1.2	123.2	70	7	13	30	3
Propranolol	PPN	+	5.87	259.8	155.2	65	5	8	22	2
					127.2	65	5	14.6	28	2
Caffeine	CF	+	5.91	195.1	138.1	57	5	11.6	23	2.2
					110.0	55	4	10	32	2.2
Metoprolol	MTP	+	6.75	268.2	116.0	50	4	10	24	2.2
					133.2	60	4	10	30	2
¹³ C-phenacetin		+	8.75	181.1	139	53	4	10	20	4
- p					110	63	4	7	30	4
Nalidixic acid	NA	+	10.3	233.2	215.3	60	5	10	20	4
T WITH WALL		'			187.2	60	5	10	30	4
Carbamazepine	CBZ	+	11.08	237.1	194.2	57	6	14	32	2.5
Curoumuz o pm c			11.00	257.1	192.3	61	8	10	32	2.5
DEET- d_7		+	11.94	199.2	126.2	53	4	8	35	2
DEET wy			11.5	1,,,,=	98.1	55	4	7	28	2
N,N-diethyl-meta	DEET	+	12.06	192.2	119.1	60	8	18	20	2
- toluamide	5221				91.1	60	8	14	40	3
Chloramphenicol-d ₅		_	4.37	326	157.1	-60	-4	-12	-28	-2
emorumphemeor u ₃					262.1	-55	-3	-25	-26	-2
Chloramphenicol	СР	_	4.43	320.8	152.1	-70	-4.9	-20	-30	-2
Cinoramphenicor	Ci		1.15	320.0	257.0	-70	-4.5	-20	-22	-2
Ketoprofen	KP	-	5.60	252.9	209.1	-33	-4	-25	-20	-2
Mecoprop- <i>d</i> ₃		-	5.86	215.7	144	-35	-3.2	-5	-25	-1
Bezafibrate	BF	_	6.34	360.0	274.0	-44.5	-4.5	-20	-25	-2.3
2 Camerate	21		0.5 .	200.0	154.2	-40	-4.1	-20	-25	-1.4
Diclofenac	DF	_	7.12	293.7	250.0	-36	-4.5	-24	-25	-2
2.0101011110	Δ.		7.12	2,5.1	214.2	-33	-4	-20	-33	-2
Indomethacin	IM	_	7.45	355.9	312.1	-35	-3	-26	-22	-3
	-		,	500.7	297.1	-35	-3.5	-25	-35	-3.5
Mefenamic acid	MA	_	7.61	239.9	196.1	-50	-3	-13	-29	-1.5
	1121 2		, <u>.</u>		180.1	-50	-4.5	-14	-40	-1.5
Gemfibrozil-d ₆		_	8.73	255.1	121.1	-40	-2.5	-20	-30	-2.5
			0.,0	200.1	133.2	-40	-2.5	-17	-17	-2
Gemfibrozil	GF	_	8.73	248.9	121.1	-65	-4	-17	-30	-2
- Jimiorozni			0.75		127.1	-65	-4	-20	-17	-2

³⁶ Note: The bold figure of parent ions were used for quantification, other ions were used for confirmation

³⁷ DP: declustering potential; EP: entrance potential; CEP: collision cell entrance potential; CE: collision energy potential; CXP: collision cell

³⁸ exit potential

Table S3 Concentration (ng L-1) and detection frequency (%) of pharmaceuticals and personal care products (PPCPs) in all samples

			Concentr	- Recovery	Matrix effect				
Compounds	River waters		WWTP effluents		Discharged samples		- (%)	(%)	LOQ b
	L-H (M) a	Frequency (%)	L-H (M)	Frequency (%)	L-H (M)	Frequency (%)	(mean±SD)	(mean±SD)	(ng L ⁻¹)
СР	13.4-43.2 (17.0)	100	14.7-48.4 (18.3)	100	nq-73.2 (25.2)	92	90±6.2	35.6±7.1	5.32
NA	nq-231 (89.7)	91	22.7-254 (130)	100	nq-243 (93.1)	75	49±5.3	37.5±6.3	4.98
ГР	nq-125 (51.5)	69	33.0-167 (39.2)	100	nq-862 (63.5)	83	71±5.1	4.3±2.2	3.85
GF	8.24-39.5 (15.5)	100	11.1-19.5 (12.0)	100	8.95-63.5 (13.3)	100	111±2.6	26.7±2.7	4.24
BF	14.4-116 (36.3)	100	14.0-297 (40.4)	100	13.7-148 (31.6)	100	99±2.9	25.6±3.1	4.17
DF	17.5-241 (71.5)	100	75.4-214 (135)	100	14.1-159 (88.5)	100	109±3.1	2.3±1.2	7.62
IM	12.2-91.6 (42.3)	100	30.5-148 (73.8)	100	nq-157 (42.8)	92	96±5.0	1.5±4.5	6.40
MA	8.63-27.4 (13.1)	100	11.2-16.4 (15.7)	100	8.62-20.9 (13.5)	100	90±8.2	12.3±6.2	2.98
KP	17.4-326 (77.6)	100	57.0-178 (114)	100	15.0-241 (124)	100	104±6.9	25.2±5.6	8.24
МТР	32.1-332 (115)	100	99.8-880 (296)	100	nq-284 (165)	92	69±4.5	15.6±4.3	3.31
PPN	nq	0	nq	0	nq	0	56±2.6	27.6±3.2	11.0
CBZ	5.25-55.6 (24.7)	100	22.0-58.7 (30.5)	100	3.64-85.0 (39.4)	100	90±4.2	5.7±1.2	2.34
SP	5.91-187 (90.0)	100	52.8-174 (141)	100	nq-193 (173)	83	45±2.8	49.5±2.1	3.44
DEET	16.5-177 (83.4)	100	57.2-217 (110)	100	29.9-881 (127)	100	89±1.9	24.3±1.4	1.78
CF	19.8-4720 (1870)	100	23.9- 11700 (285)	100	14.5- 11900 (3570)	100	117±2.6	35.9±5.2	4.40
SumPPCP	276-6109 (2780)		685-13560 (1971)		219-13805 (4706)				

⁴¹ nq: not quantified (< LOQ).

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^{42 &}lt;sup>a</sup> L, the lowest concentration; H, the highest concentration; M, median concentration.

^{43 &}lt;sup>b</sup> Limit of quantification (LOQ).

Table S4 Predicted no-effect concentrations (PNECs, ng L⁻¹) and corresponding risk quotients (RQs) for the detected PPCP compounds in the WWTP effluents, directly discharged wastewaters and river waters. Values between 0.1 and 1 (medium risk) are reported in bold

DDCD _a	PPCPs PNEC a (µg L-1)		Effluents		Discharged wastewater		Liangshui River			Qing River					
PPCPS	Fish	Daphnia	Algae	Fish	Daphnia	Algae	Fish	Daphnia	Algae	Fish	Daphnia	Algae	Fish	Daphnia	Algae
TP	100	4.8	2.6	0.002	0.03	0.06	0.01	0.2	0.3	0.001	0.03	0.05	0.0004	0.01	0.02
GF	0.9	6	4	0.02	0.003	0.005	0.07	0.01	0.02	0.04	0.01	0.01	0.02	0.002	0.004
BF	5.3	25	18	0.06	0.01	0.02	0.03	0.006	0.008	0.02	0.005	0.01	0.01	0.001	0.002
DF	532	22.4	16.3	0.0004	0.01	0.01	0.000	0.007	0.010	0.0005	0.01	0.01	0.000	0.004	0.01
IM	3.9	26	18	0.04	0.006	0.008	0.04	0.006	0.009	0.02	0.004	0.01	0.01	0.002	0.003
KP	32	248	164	0.006	0.001	0.001	0.01	0.002	0.003	0.01	0.001	0.002	0.003	0.0004	0.001
MA		0.43			0.04			0.05			0.06			0.04	
MTP	116	8	14	0.008	0.1	0.06	0.002	0.04	0.02	0.003	0.04	0.02	0.001	0.02	0.01
CBZ	35.4	13.8	70	0.002	0.004	0.001	0.002	0.006	0.001	0.002	0.004	0.001	0.001	0.002	0.0003
DEET	71.3	75	388	0.003	0.003	0.001	0.01	0.01	0.002	0.002	0.002	0.0005	0.001	0.001	0.0002
CF	151	46	46	0.08	0.3	0.3	0.08	0.3	0.3	0.03	0.1	0.1	0.01	0.05	0.05

^a PNEC data is assumed as the lowest toxicological value among those available in literature divided per 1000, for the three trophic levels, algae, daphnia and fish, respectively. Detailed information is shown in Table S3.

Table S5 Ecotoxicity data for the selected compounds available in literature, together with the corresponding species assayed, test endpoint, toxicity concentration, references and the resulting Predicted No-Effect Concentrations (PNECs). This is assumed as the lowest toxicological value among those available and here reported divided per 1000

Compounds	Carrier	Test (endpoint)	Toxicity (mg/L)	References	PNEC (μg L-1)
	Fish	EC50 ECOSAR	795	1	
	Daphnia	EC50 ECOSAR	4.8	1	4.8
	Algae	EC50 ECOSAR	2.6	1	2.6
	Bacteria	EC50 (15min)	177	2	
TD	Daphnia	EC50 (96h-immobility)	121	2	
TP	Fish	EC50 ECOSAR 4.8 1 EC50 ECOSAR 2.6 1 EC50 ECOSAR 2.6 1 EC50 (15min) 177 2 EC50 (96h-immobility) 121 2 EC50 (48h) >100 3 Prates LC50 (96h) >100 4 EC50 ECOSAR 0.9 1 EC50 ECOSAR 6 1 EC50 ECOSAR 4 1 EC50 ECOSAR 4 1 EC50 (15 min) 35.3 5 EC50 (15 min) 18.8 6 Prates EC50 (48 h) 10.4 7 Prates EC50 (48 h) 10.4 7 Prates EC50 ECOSAR 5.3 1 EC50 ECOSAR 5.3 1 EC50 ECOSAR 18 1 EC50 ECOSAR 18 1 EC50 ECOSAR 500 4 EC50 ECOSAR 500 50 4 EC50 ECOSAR 500 50 1	3		
	Invertebrates		2		
	Invertebrates		4		
	Algae	EC50	EC50 100 4 EC50 ECOSAR 0.9 1 EC50 ECOSAR 6 1	4	
	Fish	EC50	100	4	100
	Fish	EC50 ECOSAR	0.9	1	0.9
	Daphnia	EC50 ECOSAR	6	1	6
GF	Algae	EC50 ECOSAR	4	1	4
	Bacteria	EC50 (15 min)	35.3	5	
	Bacteria	EC50(15 min)	18.8	6	
	Invertebrates	EC50 (48 h)	10.4	7	
	Invertebrates	tebrates EC50 (48 h) 10.4 7 tebrates EC50 (96 h) 1.18 3	3		
	Fish	EC50 ECOSAR	5.3	1	5.3
DE	Daphnia	EC50 ECOSAR	25	1	25
BF	Algae	EC50 ECOSAR	18	1	18
	Invertebrates	EC50	50	4	
	Fish	EC50 ECOSAR	532	1	532
	Daphnia	EC50 ECOSAR	5057	1	
	Algae	EC50 ECOSAR	2911	1	
	Daphnia	EC50 (48h-mortality)	22.4	8	22.4
DF	Algae	EC50 (96h-growth)	16.3	8	16.3
	Bacteria	EC50(30min-luminescence)	11.4	8	
	Bacteria	EC50 (15min-inhibition)	9.7	9	
	Microtox	EC50 (30min)	11.45	10	
	Daphnia	EC50 (48h)	22.43	10	

Compounds	Species Assayed	Test (endpoint)	Toxicity (mg/L)	References	PNEC (µg L-1)
	C. dubia	EC50 (48h)	22.7	10	
	Algae	EC50 (96h-growth)	14.5	10	
	Invertebrates	EC50	90	4	
	Algae	EC50-inhibition	72	11	
	Daphnia	EC50-immobilization	68	11	
	Fish	EC50 ECOSAR	3.9	1	3.9
IM	Daphnia	EC50 ECOSAR	26	1	26
	Algae	EC50 ECOSAR	18	1	18
	Fish	EC50 ECOSAR	32	1	32
IZ D	Daphnia	EC50 ECOSAR	248	1	248
KP	Algae	EC50 ECOSAR	164	1	164
	Bacteria	EC50 (15min)	15.6	6	
MA	Daphnia	EC50 ECOSAR	0.43	12	0.43
	Fish	EC50 ECOSAR	29.5	1	
	Daphnia	EC50 ECOSAR	2.3	1	2.3
	Algae	EC50 ECOSAR	5.5	1	
	Bacteria	EC50(30min-luminescence)	61	8	
	Algae	EC50 (48 h)	0.7	13	
PPN	Diatoms	EC50 (96 h-growth)	0.244	8	
	Invertebrates	LC50(48 h)	0.8	14	
	Fish	LC50 (48 h)	24.3	14	
	Invertebrates	EC50	11	4	
	Algae	EC50	0.8	4	0.8
	Fish	EC50	20	4	20
	Fish	EC50 ECOSAR	116	1	116
	Daphnia	EC50 ECOSAR	8	1	8
	Algae	EC50 ECOSAR	14	1	14
MTP	Invertebrates	LC50 (48h)	>100	14	
	Invertebrates	LC50 (48h)	8.8	14	
	Invertebrates	LC50 (48h)	63.9	14	
	Fish	LC50 (48h)	>100	14	
	Fish	EC50 ECOSAR	101	1	
CBZ	Daphnia	EC50 ECOSAR	111	1	
	Algae	EC50 ECOSAR	70	1	70

Compounds	Species Assayed	Test (endpoint)	Toxicity (mg/L)	References	PNEC (μg L-1)
	Algae	EC50 (3days)	74	15	
	Bacteria	EC50 (15min)	52.2	2	
	Fish	EC50 (48h)	35.4	2	35.4
	Daphnia	EC50 (48h-mortality)	13.8	8	13.8
	Diatoms	EC50 (96h- growth)	31.6	8	
	C. dubia	EC50 (48h)	77.7	10	
	Algae	EC50 (24 h)	388	16	388
DEET	Daphnia	EC50 (48h-immobilisation)	75	17	75
DEET	Fish	LC50 (96 h)	71.3	17	71.3
	Daphnia	LC50 (96 h)	108	18	
	Fish	EC50 ECOSAR	805	1	
	Daphnia	EC50 ECOSAR	46	1	46
CF	Algae	EC50 ECOSAR	46	1	46
	Cladocerans	LC50	182	4	
	Fish	LC50	151	19	151

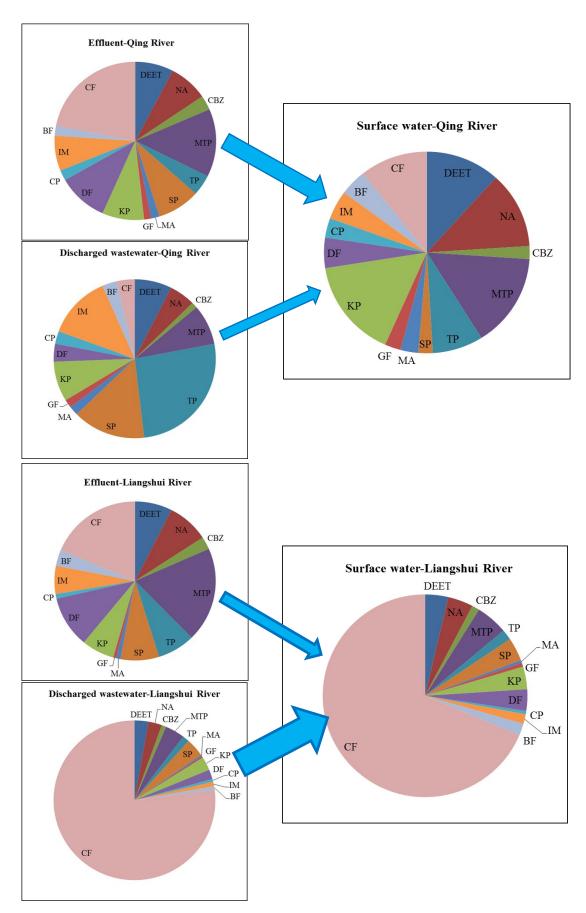


Figure S1: Fraction of PPCPs in effluents, discharged wastewaters and river waters.

Reference

- H. Sanderson, D. J. Johnson, C. J. Wilson, R. A. Brain, K. R. Solomon, *Toxicol. Lett.*, 2003, 144, 383-395.
- Y. Kim, K. Choi, J. Jung, S. Park, P. Kim, J. Park, *Environ. Int.*, 2007, 33, 370-375.
- 3 B. Quinn, F. Gagné, C. Blaise, Sci. Total Environ., 2008, 389, 306-314.
- 4 C. Boillot, PhD Thesis, 2008, Institut National des Sciences Appliquées de Lyon, France, Nd'ordre ISAL 0021.
- 5 R. Rosal, M. S. Gonzalo, K. Boltes, P. Letón, J. J. Vaquero, E. García-Calvo, *J. Hazard. Mater.*, 2009, **172**, 1061-1068.
- M. Farré, I. Ferrer, A. Ginebreda, M. Figueras, L. Olivella, L. Tirapu, M. Vilanova, D. Barceló, J. Chromatogr. A, 2001, 938, 187-197.
- 7 G. H. Han, H. G. Hur, S. D. Kim, *Environ. Toxicol. Chem.*, 2006, **25**, 265-71.
- 8 B. Ferrari, R. Mons, B. Vollat, B. Fraysse, N. Paxeus, R. Lo Giudice, A. Pollio, J. Garric, *Environ. Toxicol. Chem.*, 2004, **23**, 1344-1354.
- 9 J. S. Ra, S. Y. Oh, B. C. Lee, S. D. Kim, Environ. Int., 2008, 34, 184-192.
- 10 B. Ferrari, N. Paxeus, R. Lo Giudice, A. Pollio, J. Garric, *Environ. Toxicol. Chem.*, 2003, **55**, 359-370.
- 11 M. Cleuvers, Ecoto. Environ. Safe, 2004, **59**, 309-315
- 12 O. A. H. Jones, N. Voulvoulis, J. N. Lester, *Water Res.*, 2002, **36**, 5013-5022.
- 13 M. Cleuvers, *Chemosphere*, 2005, **59**, 199-205.
- D. B. Huggett, B. W. Brooks, B. Peterson, C. M. Foran, D. Schlenk, *Bull. Environ. Contam. Toxicol.*, 2002, **43**, 229-235.
- 15 M. Cleuvers,. Toxicol. Lett., 2003, 142, 185-194.
- 16 S. D. Costanzo, A. J. Watkinson, E. J. Murby, D. W. Kolpin, M. W. Sandstrom, Sci. Total Environ., 2007, 384, 214-220.
- 17 Office of Pesticide Programs, Environmental Fate and Effects Division, U.S.EPA, Washington, D.C., 2000.
- 18 J. Seo, Y. G. Lee, S. D. Kim, C. J. Cha, J. H. Ahn, H. G. Hur, Arch. Environ. Contam. Toxicol., 2005, 48, 323-328.

19 C. L. Russom, S. P. Bradbury, S. J. Broderius, D. E. Hammermeister, R. A. Drummond, *Environ. Toxicol. Chem.*, 1997, **16**, 948-967.