

1 **Evolution of reproductive isolation in a long-term evolution experiment with *Drosophila***
2 ***melanogaster*: 30 years of divergent life history selection.**
3

4 **ABSTRACT**
5

6 We ask if three decades and over 1 500 generations of divergent life history selection on
7 age at reproduction has resulted in the evolution of reproductive isolation (RI) between
8 laboratory populations of *Drosophila melanogaster*. We tested for premating, postmating-
9 prezygotic and postzygotic reproductive isolation between 3 replicate population pairs. Large
10 evolved differences in body size between selection treatments suggested the potential for
11 prezygotic barriers driven by sexual selection or physical incompatibilities between the sexes.
12 Although a simple prediction would be preference for larger size, creating directional isolation,
13 our results from individual mate choice trials indicate that populations from both selection
14 treatments show a marked bias towards homotypic mate choice; indicative of prezygotic RI
15 driven by sexual selection or sexual conflict. Hybridization between the focal populations
16 resulted in the production of viable adult flies with intermediate size and developmental traits.
17 We observed a suggestive but statistically non-significant trend of fitness decline in the F2
18 generation of hybrids, but no significant evidence suggesting the evolution of postmating-
19 prezygotic or postzygotic RI. Our findings are in accord with extant literature that posits that
20 premating RI evolves before postmating forms of RI.
21
22

INTRODUCTION

Understanding the mechanisms underlying the evolution of reproductive isolation and the timing of their origination is central to the study of speciation. With the fossil record yielding limited information about this process on microevolutionary timescales, there has been a focus on extant incipient species, either where populations naturally intermingle in contact zones or through experimental crosses. Such populations, part way through the process of RI, can help to illuminate the nature of divergence, as indicated by premating isolation or hybrid inferiority due, for example, to antagonistic epistasis between incompatible alleles (Dobzhansky 1937; Muller 1942) and hence offer a snapshot of speciation in action. But what selective agents or historical contingencies have driven the divergence may or may not be apparent, particularly as replication and controls may be absent or limited, and partial isolation and changes in the environment can lead to reversals in the development of RI (Lackey & Boughman 2017), complicating the inference of sequence in the evolution of isolating mechanisms.

Here we focus on early stages of ecological speciation. In this work, we focus on the role of divergent life-history selection in ecological speciation. Ecological speciation is the process through which reproductive isolation evolves as a consequence of local adaptation with limited gene flow. Divergent selection pressures can contribute to this process by acting directly, or through pleiotropy or tight linkage with genes that are directly responsible for reproductive isolation (Rice and Hostert 1993; Rundle and Nosil 2005). Divergent evolved responses between ecotypes may result in (1) changes in morphology, mating behaviours or physiological characters that hinder mating attempts mechanically or via mate choice (prematuring reproductive isolation); (2) changes in gametes/reproductive machinery that inhibit fertilization (postmaturing-prezygotic reproductive isolation); and/or (3) genetic incompatibilities that reduce survival or reproductive success of hybrid individuals (postzygotic reproductive isolation).

In addition to divergent natural selection in allopatric populations, divergent intersexual coevolution via sexual selection has long been recognized to have a potentially significant role in the evolution of reproductive isolation (Lande 1981; Maan and Seehausen 2011; Panhuis *et al.* 2001; Ritchie 2007; Singh and Singh 2014). Likewise, interlocus sexual conflict – where traits that increase the fitness of one sex directly cost the other – can result in rapid coevolution (an “evolutionary chase” or “arms race”) between male and female reproductive traits (Rice & Holland 1997; Parker & Partridge 1998; Rice 1998) and potentially accelerate the process of speciation (Ritchie 2007; Gavrillets 2014; Syed *et al.* 2017); the evolution of female resistance to reduce the direct costs of mating is predicted to contribute to assortative mate choice (Gavrillets *et al.* 2001).

While natural systems such as hybrid zones have advanced our understanding of the evolution of RI immensely, researchers are frequently limited to indirect inferences, such as using estimates of gene flow and clines to infer reproductive isolation. Beyond knowledge of the

61 evolutionary history, these systems limit the number RI barriers that can be assessed, and
62 generally lack replication at the population-pair level. Notable exceptions include stickleback
63 fishes, where repeated invasion of freshwater from marine ancestors, and subsequent
64 differentiation into limnetic-benthic and lake-stream ecotypes provide extensive replication
65 (Lackey & Boughman 2017); and storm-petrels, where repeated separation of populations by
66 breeding time (allochrony) provides replicated evidence of reproductive isolation in sympatry
67 (Wallace *et al.* 2017). Experimental evolution can be a valuable tool for more direct
68 investigation, allowing us to probe potential drivers of speciation between populations with well
69 understood evolutionary histories. Experimental studies in speciation have attempted to test if (1)
70 non-ecological drivers like drift, (2) negative selection against hybridization between ecotypes,
71 (3) divergent selection on populations or (4) sexual conflict produced reproductive isolation
72 (reviewed in Rice and Hostert 1993; Coyne and Orr 2004; Fry 2009; White *et al.* 2020). An
73 obvious limitation to experimental evolutionary studies of speciation is time: research projects
74 rarely exceed granting cycles and few organisms lend themselves to experimental evolution. It is
75 little surprise then that *Drosophila* is far and away the genus most investigated in the laboratory.

76 Early divergent selection studies produced seemingly equivocal results with respect to
77 creating reproductive isolation. A closer examination suggested that divergent selection was
78 highly effective, given a pre-existent tendency to mate assortatively on the basis of the trait
79 selected upon (Rice and Hostert 1993; Fry 2009). Divergent selection upon behavioural traits
80 resulted in the correlated evolution of reproductive isolation (del Solar 1966; Soans *et al.* 1974;
81 Hurd and Eisenberg 1975; Lofdahl *et al.* 1992) while divergent selection on other traits like
82 bristle number (Barker and Cummins 1969; Santibañez and Waddington 1958) produced
83 negative results. More recently, experimental evolution studies have focused on selection
84 regimes that do not directly involve characters that are known to influence intra-population
85 assortment. For example, a number of studies have investigated the importance of local
86 adaptation to distinct nutrition sources for mating assortment. Divergent selection regimes with
87 different nutritional environments result in premating isolation through (1) the evolution of
88 signalling traits and mating preferences in *Drosophila serrata* (Rundle *et al.* 2005), (2) the
89 evolution of symbiotic microbiota in inbred strains of *D. melanogaster* (Najarro *et al.* 2015;
90 Sharon *et al.* 2010), (3) competitive ability differences between controls and selected individuals
91 in *D. melanogaster* (Belkina *et al.* 2018), (4) positive assortative mating preferences ~~(albeit~~
92 ~~unstable)~~ in *D. melanogaster* (albeit unstable) (Nash *et al.* 2019) and in *Saccharomyces*
93 *cerevisiae* (Mahilkar *et al.* 2023); and (5) postzygotic reproductive isolation through genetic
94 incompatibilities in *Saccharomyces cerevisiae* (Dettman *et al.* 2007) and *Neurospora* (Dettman
95 *et al.* 2008).

96 Life history traits are less frequently studied in the context of reproductive isolation, but
97 have also received some attention – predominantly in studies of allochronic speciation. Powell *et*
98 *al.* (2020) note how adaptation to differences in seasonal timing has been shown to contribute to
99 reproductive isolation in a wide variety of organisms including plants (Savolainen *et al.* 2006;

Lowry *et al.* 2008), insects (Tauber *et al.* 1977; Wood & Guttman 1982; Horner *et al.* 1999; Eubanks *et al.* 2003; Abbot & Withgott 2004; Forbes *et al.* 2009; Ordning *et al.* 2009; Wadsworth *et al.* 2013; Hood *et al.* 2015) and vertebrates (Friesen *et al.* 2007). Speciation through other life-history traits and correlated changes in intersexual coevolution has received less attention, due to the myriad challenges associated with studying these systems in nature. For example, divergent selection on pre-adult development time has been shown to produce pre-mating isolation through (1) disturbances in circadian rhythm resulting in changes in phases of mating in the melon fly, *Bactrocera cucurbitae* (Miyatake and Shimizu 1999), and (2) evolved body size differences, and correlated levels of sexual conflict, in *Drosophila melanogaster* (Ghosh and Joshi 2012). Likewise, divergent selection on body size results in premating isolation, through mechanical incompatibilities during attempted copulations between *Columbicola* feather mites (Villa *et al.* 2019).

Age-at-reproduction has been extensively manipulated using experimental evolution in *Drosophila* to investigate correlated changes in longevity, development time, body size and a slew of associated traits (Prasad and Joshi 2003). In this study, we use an exceptionally long term evolution experiment (LTEE) with population pairs separated by over 1,500 generations and three decades under divergent selection in allopatry. The two *D. melanogaster* life-history selection regimes used in this study are selected to reproduce late in life (4 weeks from egg; CO) and extremely early in life (9 days from egg; Accelerated CO / ACO). The ACO populations have evolved small body size, rapid development time, and reduced lifespan (Chippindale *et al.* 1997a; Chippindale *et al.* 2004a). On the other hand, the CO flies are big, develop slowly and likely have an increased ability to resist copulations as females and to inflict harm as males (Chippindale *et al.* 2004b; Chippindale *et al.* 1997b; Mital *et al.* 2021, Verma *et al.* 2022).

In the speciation literature, body size is identified as a crucial trait that can drive the evolution of reproductive isolation (Servedio *et al.* 2011). In *D. melanogaster*, body size is strongly correlated with juvenile development time (Chippindale *et al.* 1997a; Nunney 1996; Prasad and Joshi 2003; Prasad *et al.* 2000; Zwaan *et al.* 1995) and life-history selection on development has been shown to result in the evolution of body size (Chippindale *et al.* 1997a; Prasad *et al.* 2000). An important sexually selected trait for both sexes, large females tend to be more fecund (Roff 2002; Stearns 1992) and attract higher courtship effort from males (Byrne and Rice 2006); while large males have increased competitive abilities (Markow and Ricker 1992; Partridge and Farquhar 1983), and secure more frequent copulations as a consequence of female preference (Markow 1986; Partridge *et al.* 1987, but see Prasad *et al.* 2007). Given the evolved size differences between the ACOs and COs, a simple prediction is that individuals of both population-types display a mating preference for individuals from the larger CO populations, producing conditions of partial, directional, reproductive isolation driven by sexual selection on the basis of body size, along the lines observed by Ghosh and Joshi (2012).

In *D. melanogaster*, divergent reproductive strategies between the sexes have long been recognized (Bateman 1948) potentially resulting in interlocus sexual conflict. Frequent matings

139 increase male fitness but can decrease female fitness as a result of male harassment and the
140 harmful effects of genital wounding and seminal fluid (Chapman et al. 1995; Bonduriansky *et al.*
141 2008; Travers *et al.* 2015). The harm experienced by females as a result of mating, measured by
142 lifespan and egg production rate, is correlated with male body size (Pitnick and García-González
143 2002; Friberg and Arnqvist 2003). In the context of our two selection treatments, we predict that
144 male harm and female resistance traits will be stronger in the CO populations compared to the
145 ACO populations, due to the difference in body size between the large COs and smaller ACOs.
146 In experimental hybridization between CO and ACO lines, this difference could drive premating
147 isolating barriers to evolve in one direction, with CO females resistant to mating with the smaller
148 ACO males, and ACO females vulnerable to unwanted mating with CO males. Indeed, working
149 with a similar system, Ghosh and Joshi (2012) found such one-directional isolation, driven by
150 large females resisting mating attempts by small males, and a preference for (or lack of
151 resistance to) large males in females of both treatments. On the other hand, especially with the
152 large number of generations in allopatry, mating signals and preferences may have diverged
153 between populations in our selection experiment creating the possibility for positive assortment
154 in mate choice experiments.

155 We investigated mate choice through individual female, male, and group mating choice
156 assays in the ACO/CO complex of selected lines. We further looked for potential gametic
157 incompatibilities (postcopulatory prezygotic isolation) by comparing egg hatchability in hybrid
158 matings to the parental populations. Finally we looked for postzygotic effects (hybrid breakdown
159 or vigour) in developmental traits, body size, and adult fitness.

METHODS

Experimental populations

Our experiments employed the accelerated development (ACO) (Chippindale *et al.* 1997a) and control-old (CO) (Rose *et al.* 1992) populations. All populations are maintained under standard laboratory conditions: moderate density (larval density of 80 - 120 larvae / vial), 25°C, 12:12 light:dark cycle on banana/agar/killed-yeast medium, and with a census population size of approximately 2,000 individuals per generation.

The five CO populations (CO₁-CO₅) were derived from like-numbered O populations in 1989 (Rose *et al.* 1992). Each CO population is maintained on a 28-day discrete life cycle, with flies placed into cages at a density of ~1000 flies/cage. On day 26, yeast paste is added to the medium to stimulate egg-laying, and on day 28/day 0, eggs are collected at 80-120 eggs per vial. In 1991, one A population (ACO₁-ACO₅) was derived from each of the five CO populations and initially subjected to direct selection for accelerated development. After generation 175, these accelerated populations were maintained on a discrete life cycle of 9 days.

Experimental populations were derived from three of the five pairs of ACO and CO populations (ACO₁, ACO₃, ACO₅ and CO₁, CO₃, CO₅). To remove environmental effects and synchronise experiments, all populations were maintained over a control 14 day life cycle for two generations prior to any assay. Prior to experiment, life cycles were staggered in light of evolved development time differences (ACO << CO) with a 48h difference in egg collection time to synchronise eclosion time of adults. Each assay was repeated for three replicate population pairs: CO₁-ACO₁, CO₃-ACO₃, and CO₅-ACO₅.

Female mate choice assay

To test for the existence of premating reproductive isolation between populations as mediated by female preference, we evaluated the mate choice of ACO and CO females. Flies were sorted by sex and collected as virgins within 6 hours of eclosion under light CO₂ anaesthesia. A food dye technique was used to differentiate between males from each regime (developed from Verspoor *et al.* 2015). Dye techniques were calibrated based on tests of mortality, and the volume of dye used did not result in detectable harmful side-effects, measured through changes in lifespan. Vials were dyed three days prior to the assay using 6 drops of red or blue food dye. One day prior to the experiment, male flies were added to coloured vials at a density of 10 flies/vial and left to ingest the dye for 24 hours. Females were acclimated to individual vials for at least 24 hours prior to the assay. At the time of the assay one male of each population identity and colour was mouth aspirated into an empty vial, then flipped into a vial containing an acclimatised female. Mating latency (time to amplexus formation), colour of the mated male and mating duration were recorded. Under the conditions we used, copulations lasted on average between 15-20 minutes. A viable mating was considered one that lasted for more than

two minutes, and vials in which mating did not occur within 60 minutes were discarded. This represents a commonly employed design to test mate choice (Dukas 2005, Ghosh & Joshi 2012, Arbuthnott *et al.* 2017). Vials were randomised so that observers were unaware of female identity. Trials were colour balanced so that in half of the trials the ACO male was blue and the CO male red, and in the other half the alternate dye pattern was applied. Flies were 2-3 days old at the time of the assay.

This assay was repeated over two days, with female choice tested in both a morning (9am-12pm) and afternoon (1pm-4pm) trial each day. In each of the 4 trials, we tested the choice of 50 females from both selection regimes for each of the 3 replicate populations. The ratio of homotypic to heterotypic matings was calculated across the 50 replicate vials for each combination of replicate population and selection regime.

Male mate choice assay

It is difficult to distinguish the influence of male competition and courtship effort from female preference and resistance. By testing individual male mate choice in addition to female choice and contrasting the results, we hoped to uncover the source of any observed non-random mating pattern. From the time of virgin collection, males were maintained at 10 males/vial. Two females, one from each selection regime, were acclimatised to the same vial 24 hours prior to the assay. Female flies from ACO and CO selection treatments could be identified reliably based upon size alone. On the day of the experiment, one male was “pootered” (mouth aspirated) into a vial containing two females. As with the female choice assay, observers recorded mating latency, the size of the mated female (large or small) and mating duration and all other procedures were the same.

Group mate choice assay

To test for the existence of premating reproductive isolation between ACO and CO populations when mating occurred in a group environment, we evaluated female mate choice in vials containing 10 females and 12 males from each regime. Flies were handled and marked using the procedures outlined above for individual assays. On the day of the assay, male vials of paired ACO and CO populations were combined and then flipped into a female vial. Vials were observed until at least 8 (out of a possible 10) simultaneously copulating pairs were observed, and were then frozen for 60 minutes without disruption to the copulating pairs. The frozen pairs were observed, and the colour of each mated male was recorded. This methodology was repeated over two trials. During each trial, 10 vials were tested for each regime within each replicate population.

Hatchability

In order to determine the existence of postmating prezygotic isolation we compared the hatch rate of eggs produced from F1 and F2 crosses $CO_i\text{♀} \times ACO_i\text{♂}$ and $ACO_i\text{♀} \times CO_i\text{♂}$ as well as the parental crosses $CO_i\text{♀} \times CO_i\text{♂}$, and $ACO_i\text{♀} \times ACO_i\text{♂}$ ($i = 1,3,5$). Crosses were performed in vials with 12 virgin females and 10 virgin males. A period of at least 4 hours was allowed for mating and egg laying. From each cross, male and female progeny were collected as virgins and maintained in sex specific vials. Hybrid and parental crosses were staggered in an attempt to synchronise time of eclosion.

For tests of hatchability, 100 flies of each sex were deposited in a collector bottle with a plate of yeasted food. After an egg laying period of 3 hours the plate was removed. Eggs were transferred into vials using fine brushes and egg collection solution (Ashburner 2005). Each vial contained 90 eggs arranged in a grid pattern to facilitate easy counting. 9 such vials were created for each cross identity, for each of the three replicate populations. 24 hours after egg collection, the proportion of hatched eggs was determined. Vials continued to be maintained under standard conditions to be used in tests of viability, development time, and size.

Larvae to adult viability

To uncover any survival differences, we compared the larvae to adult viability of hybrids compared to flies from parental crosses. Twelve days after egg collection, the number of eclosed adults in each vial was recorded. The difference between the number of viable eggs and eclosed adults was used as a measure of larvae to adult viability. 9 vials were tested for each cross identity in each replicate population.

Development time

Development time of flies from hybrid and parental crosses was also analysed as an indicator of hybrid viability. Each vial was observed for eclosion over a period of 4 days, beginning 24 hours prior to the time of expected peak eclosion for each line. For the first 2 days, vials were observed every 6 hours. For the following 2 days, observations were made every 12 hours. During each observation, the number of eclosed flies was recorded. An average eclosion time was then calculated for the vial. 9 vials were tested for each cross identity across each replicate population.

Body Size

Body size was measured using dry weight as a proxy. During peak eclosion, 20 female flies were collected from each cross identity. These flies were given 1 hour to physically mature and then were frozen. Flies were divided into 4 groups of 5 females and placed into tinfoil dishes to

be dried in an oven at 70 degrees Celsius for 24 hours. Each dish was measured in a microbalance four times: twice with the flies and twice without. These totals were averaged and divided by 5 to provide an estimate of the dry weight of an individual fly.

Fertility - Competitive reproductive fitness

An adult fitness assay was completed to test for parental line and hybrid fertility differences. In this assay, red-eyed focal flies competed with control flies marked with a recessive brown-eye marker (bw^1) derived from the baseline. The IVb population was selected as these flies are a common ancestor to both the focal populations, also having an intermediate life-history selection protocol. The recessive nature of bw allowed the proportion of red eyed progeny to be used as a metric of paternity share.

Focal F1 and F2 flies of the hybrid crosses $CO_i\text{♀} \times ACO_i\text{♂}$, $ACO_i\text{♀} \times CO_i\text{♂}$ as well as the parental crosses $CO_i\text{♀} \times CO_i\text{♂}$, and $ACO_i\text{♀} \times ACO_i\text{♂}$ ($i = 1,3,5$) were tested. Crosses were performed in vials with 12 virgin females and 10 virgin males. A period of at least 4 hours was allowed for mating and egg laying. From each cross, male and female progeny (focal flies) were collected as virgins and maintained in sex specific vials at a density of 10 flies/vial.

Focal flies were anaesthetized under CO_2 and combined with four IVb flies of the same sex (competitors) and four IVb flies of the opposite sex (potential mates). Vials were supplemented with a sparse amount of yeast. Female focal and IVb flies were acclimatised to vials 24 hours prior to the assay. After 48 hours, flies were flipped into new vials where oviposition occurred. Flies were 5-7 days old at the time of the assay. After 18 hours flies were discarded. At the time of eclosion, the number of red eyed and brown eyed offspring produced from the competition assay were counted. Thirty vials for each sex and cross identity were tested for each of the 3 replicate populations.

Statistical Analysis

Statistical analysis was conducted in R version 4.2.3 -- "Shortstop Beagle" (R Core Team 2023). Data was visually assessed for residual normality and heterogeneity. Where generalised linear models were used, residual dispersion was tested using the DHARMA package (Hartig 2022).

Female, male and group mate choice data was analysed with a repeated G-test for goodness of fit (Ghosh & Joshi 2012, McDonald 2014). In each case, the ratio of homotypic to heterotypic matings was tested for deviation from a 1:1 null expectation separately for each replicate population and overall for each selection regime. To study the effect of the male and female identity on mating latency and duration, we used linear mixed effects models (LMMs) fit

with mated male identity and mated female identity as fixed effects and trial ID and replicate population as crossed random effects. Data from female choice and male choice experiments were analysed independently. In both experiments, mating latency models with raw data produced skewed residual distributions. To correct for this, we used mixed linear models using $\log(\text{mating latency} + 1)$ as the response variable.

To determine the effect of cross identity on fly development time and body size, LMMs were fit with cross identity as a fixed effect and replicate population as a random effect. To determine the effect of cross identity on hatchability, viability and the fertility of flies, binomial generalised linear mixed effects models (GLMMs) were fit with cross identity included as fixed effect and parental replicate included as a random effect. Fertility data was analysed separately for males and females. Due to over-dispersion issues, we remodelled the viability data and the fertility data for both sexes with beta-binomial generalised linear mixed effects models using the 'glmmTMB' package (Brooks *et al.* 2017).

Models were fit using the 'lme4' package (Bates *et al.* 2015). Fixed effects and random effects were analysed using the 'anova()' and 'ranova()', and 'Anova()' or and by backwards model selection for LMMs and GLMMs, respectively (Kuznetsova *et al.* 2017, Fox & Weisberg 2019). Post-hoc comparisons between cross identities were performed using the 'emmeans' package (Lenth 2022). Statistical analysis was conducted in RStudio version 1.3.1073 (RStudio Team 2020) and R version 1.3.1073 (R Core Team 2020). Data was visually assessed for residual normality and heterogeneity. Female, male and group mate choice data was analysed with a repeated G-test for goodness of fit (Ghosh & Joshi 2012, McDonald 2014). In each case, the ratio of homotypic to heterotypic matings was tested for deviation from a 1:1 null expectation separately for each replicate population and overall for each selection regime. To determine the effect of cross identity on fly development time and body size, linear mixed effects models (LMMs) were fit with cross identity as a fixed effect and replicate population as a random effect. To determine the effect of cross identity on hatchability, viability and the fertility of flies, binomial generalised linear mixed effects models (GLMMs) were fit with cross identity included as fixed effect and parental replicate included as a random effect. Fertility data was analysed separately for males and females. Models were fit using the 'lme4' package (Bates *et al.* 2015). Fixed effects and random effects were analysed using the 'anova()' and 'ranova()', and 'Anova()' and by backwards model selection for LMMs and GLMMs, respectively (Kuznetsova *et al.* 2017, Fox & Weisberg 2019). Post-hoc comparisons between cross identities were performed using the 'emmeans' package (Lenth 2022).

RESULTS

Female Mate Choice Assay

Overall, ACO females showed a significantly higher proportion of homotypic (0.57 ± 0.048 , mean \pm 1.96**sed*) compared to heterotypic matings (0.42 ± 0.048 , $P = 0.01$, Figure 1a), with no significant heterogeneity among replicate population pairs ($P = 0.66$, Table S1). The ACO₅ population showed a significant deviation from the null hypothesis, with more homotypic matings compared to heterotypic matings ($P = 0.04$; Figure S1a). Females from the ACO₃ population showed a similar trend towards more homotypic matings with borderline significance ($P = 0.05$). ACO₁ females showed no significant difference between mating types ($P = 0.26$). When data was pooled across trials within each replicate population, both ACO₃ and ACO₅ populations, but not the ACO₁ population, showed a significant deviation from a random mating ratio ($P_1 = 0.21$, $P_3 = 0.02$, $P_5 = 0.02$), with no significant heterogeneity between individual trials for any of the 3 replicate populations ($P_1 = 0.30$, $P_3 = 0.28$, $P_5 = 0.21$).

Overall, CO females showed a significantly higher proportion of homotypic (0.60 ± 0.058) compared to heterotypic matings (0.40 ± 0.058 , $P < 0.0001$, Figure 1a), with no significant heterogeneity among replicates ($P = 0.65$, Table S1). Two CO populations, CO₁ and CO₃, showed an overall significant deviation from the null hypothesis, with more homotypic copulations ($P_1 < 0.01$, $P_3 = 0.01$, Figure S1a). CO₅ females showed a trend towards more homotypic matings, but this was not significant ($P = 0.08$). When data was pooled across trials, all three CO populations showed a significant deviation from the null hypothesis of random mating ($P_1 < 0.001$, $P_3 = 0.01$, $P_5 = 0.04$), with no significant heterogeneity among trials for any of the replicate populations ($P_1 = 0.13$, $P_3 = 0.15$, $P_5 = 0.28$).

Only mated male identity had a significant effect on mating latency ($P < 0.00001$, Table S2[?]). The crossed random effects of the replicate population and trial-ID were significant ($P < 0.0001$), but the cumulative proportional variance attributed to the random effects of replicate population and trial ID was minimal (0.05). Both ACO and CO females displayed significantly shorter latencies in mating with CO males (11.8 ± 1.2 min, 11.6 ± 1.0 min respectively), compared to ACO males (15.3 ± 1.3 min, 15.5 ± 1.5 min respectively, Figure 2a). There was no significant effect of female identity or interaction between male and female identity.

Female identity and interaction between male and female identity had significant effects on mating duration ($P < 0.00001$, Table S2[?]). The crossed random effects of the replicate population and trial ID were significant ($P < 0.0001$), but cumulative proportional variance attributed to the random effects of replicate population and trial-ID was minimal (0.04). In general, ACO females (15.0 ± 3.8 min) displayed shorter mating durations than CO females (16.7 ± 4.2 min), and both females mated longer with homotypic partners (ACO♀-ACO♂ 15.4

+/-0.4 min, ACO♀-CO♂ 14.3 +/- 0.5 min, CO♀-ACO♂ 16.1 +/-0.6 min, CO♀-CO♂ 17.1 +/- 0.4 min). All pairwise contrasts were significant (Figure 2c, $P < 0.05$, Table 2) with the exception of ACO females mated with ACO males vs CO females mated with ACO males. There was no significant effect of mated male identity.

Male Mate Choice Assay

Overall, ACO males showed a significantly higher proportion of matings with ACO females (0.64 +/- 0.058) compared to CO females (0.36 +/- 0.058, $P = < 0.00001$, Figure 1b), with no significant heterogeneity between populations ($P = 0.81$, Table S32). All three A populations showed an overall significant deviation from the null hypothesis, displaying more homotypic compared to heterotypic matings ($P_1 = < 0.0001$, $P_3 = 0.01$, $P_5 = < 0.001$). There was significant heterogeneity observed among trials within the ACO₁ population ($P = 0.01$), but not the ACO₃ ($P = 0.70$) or ACO₅ ($P = 0.38$) populations.

Overall, CO males displayed a nearly equal proportion of matings with ACO (0.51 +/- 0.024) and CO females (0.49 +/- 0.024, $P = 0.95$, Figure 1b), with no heterogeneity among replicate populations ($P = 0.93$, Table S32). All three CO populations showed no significant deviation from the null hypothesis of random mating ($P_1 = 0.99$, $P_3 = 0.65$, $P_5 = 0.96$). There was also no significant heterogeneity among trials within any of the replicate populations ($P_1 = 0.97$, $P_3 = 0.54$, $P_5 = 0.91$).

None of male identity, female identity or interaction between male & female identity influenced mating latency (Table S2). The crossed random effects of the replicate population and trial were significant ($P = 0.01$), but cumulative proportional variance attributed to the random effects of replicate population and trial ID was minimal (0.02). Females from the CO populations displayed a slightly higher mating latency with both ACO and CO males (18.0 +/- 1.7 min, 18.1 +/- 1.6 min respectively), compared to the ACO females (16.6 +/- 1.1 min, 16.9 +/- 1.4 min respectively) but these differences were non-significant (Figure 2b).

Only mated female identity had a significant effect on mating duration ($P = < 0.00001$, Table S2-2). The crossed random effects of the replicate population and trial ID were significant ($P = < 0.0001$), but cumulative proportional variance attributed to the random effects of replicate population and trial ID was minimal (0.04). Males from the both populations displayed a slightly higher mating latency with CO females (16.3 +/- 0.7 min, 17.3 +/- 0.6 min respectively), compared to the ACO females (14.8 +/- 0.5 min, 15.0 +/- 0.7 min respectively) (Figure 2d). There was no significant effect of male identity or interaction between male and female identity.

Group Mate Choice Assay

Overall, ACO females showed a significantly higher proportion of matings with CO males (0.62 +/- 0.0529) compared to ACO males (0.38 +/- 0.0529, $P = <0.00001$, Figure 1c), with significant heterogeneity being observed between replicate populations ($P = 0.01$, Table S43). ACO₃ and ACO₅ females showed an overall significant deviation from the null hypothesis, displaying more heterotypic compared to homotypic matings ($P_3 = 0.02$, $P_5 = <0.0001$). ACO₁ females showed no deviation from the null expectation ($P = 0.78$). There was no significant heterogeneity observed among trials within any of the three replicate ACO populations ($P_1 = 0.49$, $P_3 = 0.45$, $P_5 = 0.50$).

Overall, CO females showed significantly more homotypic (0.61 +/- 0.0417) compared to heterotypic (0.39 +/- 0.0417) copulations ($P = <0.0001$, Figure 1c), with no heterogeneity between replicate populations ($P = 0.12$, Table S43). CO₅ females showed an overall significant deviation from the null hypothesis, with more homotypic compared to heterotypic copulations ($P = <0.001$). CO₁ and CO₃ populations showed no overall deviation from a random mating ratio of 1:1 ($P_1 = 0.10$, $P_3 = 0.07$). When data was pooled across trials, both CO₃ and CO₅ populations showed significant deviation from a random mating ratio ($P_3 = 0.02$, $P_5 = <0.0001$). There was no significant heterogeneity among trials within any of the populations ($P_1 = 0.10$, $P_3 = 0.58$, $P_5 = 0.96$).

Post-mating prezygotic reproductive isolation

Hatchability

Cross identity had a significant effect on egg hatchability ($P = <0.01$, Table S54). The random effect of replicate population was not significant ($P = 0.89$). The only significant differences in hatch rate was the decline of CO_i♀ × A_i♂ F2 hybrids compared to ACO and CO parental lines (Figure 32a). Overall, hatchability was high with average values between 0.96 +/- 0.012 (CO_i♀ × A_i♂ F2 hybrids) and 0.98 +/- 0.01 (ACO parentals).

Postzygotic reproductive isolation

Larvae to adult viability

There was also a significant effect of cross identity ($P = <0.0001$, Table S54), but no random effect of replicate population ($P = <0.01031$) on the viability of flies. On average, larvae to adult viability was lowest for ACO parental (0.78 +/- 0.038) and highest for CO_i♀ × ACO_i♂ F2 flies (0.93 +/- 0.026). Both CO_i♀ × ACO_i♂ hybrids and ACO_i♀ × CO_i♂ F2 hybrids demonstrated significantly higher viability than ACO, CO and ACO_i♀ × CO_i♂ F1 flies and CO flies showed significantly higher viability than ACO flies (Figure 32b).

Development time

There was a significant effect of cross identity ($P = <0.0001$, Table S65), but no ~~random effect of~~ replicate population ($P = 0.51$) on the development time of flies. All four hybrid crosses demonstrated development times that were significantly longer than ACO parental flies (175.76 ± 0.51 hours) and significantly shorter than CO parental flies (218.32 ± 1.23 hours). There were no significant differences in development times among the four hybrid crosses (Figure 32d).

Body size

Cross identity also had a significant effect on the body size of flies ($P = <0.0001$, Table S65). The random effect of the replicate population was not significant ($P = 0.23$). All four hybrid lines showed average individual weights that were significantly higher than parental ACO lines (0.21 ± 0.01 g), and significantly lower than parental CO lines (0.31 ± 0.01 g). Hybrid $ACO_i \times CO_i$ F1 flies were also significantly heavier than $ACO_i \times CO_i$ F2 flies (Figure 32c).

Reproductive fitness

Cross identity had a significant effect on female reproductive fitness ($P = <0.0001$, Table S76). Parental CO lines showed the highest average fitness (0.24 ± 0.12), parental ACO lines the lowest (0.12 ± 0.10) and hybrids displayed intermediate fitness between parental lines (Figure 4-3). All cross identities produced significantly more offspring than parental ACO females and flies from all cross identities, except for $CO_i \times ACO_i$ F1 females, produced significantly fewer offspring than parental CO flies. In both cases, second generation (F2) females displayed lower reproductive fitness than first generation (F1) hybrids, but this effect was only significant for the $CO_i \times ACO_i$ crosses. There was no significant effect of population replicate on female fertility ($P = 1$).

There was a significant fixed effect of cross identity ($P = <0.0001$, Table S76) ~~and random effect of~~ but not replicate population ($P = 0.021$) on the fertility of male flies. $CO_i \times ACO_i$ F1 male hybrids showed the highest average fitness (0.13 ± 0.04) and ACO males the lowest (0.05 ± 0.14). F1 hybrid and CO and $CO_i \times ACO_i$ F1 males demonstrated a significantly higher fitness than F2 and ACO males (Figure 43). ~~F2 hybrid males sired significantly more offspring than A males~~. Similar to that observed in the female assay, male F2 hybrids displayed significantly lower fitness than F1 hybrids, but these differences were not significant. While female flies displayed fitness values around the expected value, males of both parental and hybrid cross identities produced a lower proportion of offspring than expected when in competition with four competitors (Figure 3).

515

516

DISCUSSION

Our experimental assays show the striking differentiation between the ACO and CO populations after over three decades and 1-500 generations of divergent selection on demography. The ACO population adults eclose nearly two days earlier than the CO population adults do. The clear tradeoff between speed and size is evident, as A adults are about 33% lighter than CO adults, and this is likely related to the strong differences in adult fitness we observed between the two selection treatments. Interestingly, the tradeoff between developmental speed and juvenile viability (Chippindale et al. 1997) is not apparent here or in other recent unpublished data on these populations, suggesting compensatory evolution over time. Given these differences in morphology and life history, we asked if we could detect the early stages of the speciation process for these populations in allopatry under different ecological circumstances.

To summarise our results, we found evidence for the evolution of premating barriers to gene flow between these diverged populations. Mate-choice assays generally reflected positive assortative mating patterns driven by female mate-choice. Females from both focal populations showed strong homotypic mate choice in individual tests, but this pattern was less clear in group tests, particularly as the smaller ACO females tended towards heterotypic mates. Under conditions of female choice, mating durations of homotypic pairs were consistently longer than heterotypic pairs. Despite this (and homotypic mate choice), the larger CO males showed lower latencies to mating with both females. In contrast, under conditions of male choice (no male-male competition), ACO and CO males showed no differences in mating latency. Further, under male choice conditions, both males mated for longer durations with the larger CO females, suggesting sperm catering based on body size rather than homotypic preference. We show that post-mating prezygotic barriers have not evolved to the point of detectability between these populations in the form of gamete incompatibility, measured through egg hatchability. We also find no evidence of postzygotic reproductive isolation in the form of survival to reproductive maturity, and measurements of development time and body size show that hybrids display intermediate trait values relative to the parental treatments. We observed only suggestive trends for postzygotic reproductive isolation in the form of hybrid breakdown in competitive reproductive fitness assays. F₁ hybrids display intermediate fitness to that of the two parentals, however, there is a tendency for F₂ hybrid offspring to display lower fitness than the F₁ hybrids; while potentially an indicator of evolving Dobzhansky-Muller incompatibilities, our data are merely suggestive at this juncture, meriting further investigation.

It has been suggested that prezygotic isolating barriers will typically evolve before postzygotic barriers, and *Drosophila* seems to adhere to this rule (Coyne & Orr 2004, Arthur & Dyer 2015). Within the category of prezygotic barriers, premating RI is expected to evolve before postmating-prezygotic RI (Turissini et al. 2018). Our data are generally in accord with

these predictions, as our diverged population pairs show marked premating reproductive isolation, at least under individual choice tests, but clear evidence for later stage RI is lacking.

Strong divergence in body size exists between ACO and CO animals (Xiao et al. 2019; data herein) and are presumed to correlate with differences in persistence/resistance capabilities between the selection treatments. This size difference led us to expect the presence of some premating reproductive isolation. However, we predicted premating isolation in one direction, on the basis of greater resistance from larger (CO) females, greater persistence of CO males, and a possible preference for large mates. That directional RI was noted by Ghosh & Joshi (2012) in a study similar to ours: bigger mates were preferred by both large/slow and small/fast flies. Contrary to that precedent, females of both regimes displayed strong homotypic mate choice and longer mating durations with their homotypic partners in individual tests. This result is especially intriguing for the ACO females, as they displayed a preference for smaller ACO males even though *D. melanogaster* females are expected to prefer larger males (Pitnick 1991, Jagadeeshan et al. 2015, but see Prasad et al. 2007).

Male mate choice tests may present trickier interpretations. ACO selected males largely mated with homotypic females, but male Cos mated nearly randomly. While ACO males appear to be showing positive assortative mating preferences, both types of males were observed making repeated courtship attempts with both homotypic and heterotypic females (unquantified observations) suggesting that the skewed mating pattern could be the result of the increased capability of CO females to resist unwanted mating attempts, particularly from the smaller A males. In alignment with this hypothesis, we observed a non-significant trend of longer mating latencies with CO females. -In populations of flies evolved under a similar pair of life history selection regimes, Ghosh and Joshi (2012) found that small males attempted to court large females, but were unsuccessful, and, as a result, mated more frequently with small females. We therefore cannot easily ascribe mating assortment to the choice of the male, who may be indiscriminating under the test conditions, particularly with no competition for mates. Concurrent with this, we note no differences in the mating latencies of the two male types in the absence of male-male competition. ACO-type females are also under tremendous pressure to mate, as they are selected to lay eggs about 24h after becoming sexually mature. In the absence of competition, many males may have simply taken the most receptive mate first; for the CO males this may have counterbalanced a homotypic preference, creating the ambivalent outcome observed. The results of the mate choice assays taken together suggest that premating isolation in this system is most strongly driven by female choice.

Body size is a likely trait involved in the evolution of reproductive isolation between the population pairs tested. However, it remains possible that the patterns of assortative mating stem from traits that are yet to be characterised. Some possibilities include courtship behaviours (mating call, wing movements) and cuticular hydrocarbon profiles that may have diverged under

the divergent selection regimes. Besides female preference, body size differences also play a clear role in the differential capacities displayed by females to resist mating attempts by males, which creates a further reproductive barrier between large females and small males. Despite large body size differences between the selection treatments, it seems unlikely that the patterns of positive assortative mating can be explained by mechanical incompatibilities in copulation alone; some earlier anecdotal observations suggested they may exist, but we saw no clear evidence in these trials.

In the group mate choice assays, female CO flies continued to show homotypic mating preference, but A females displayed reversed preferences, mating more frequently with heterotypic males than homotypic males. The existence of heterotypic mate choice in groups, in spite of homotypic mate choice in individual assays suggests that the nature of the assay plays a role in the mating patterns observed. In the group mating assays, vials were more crowded (~~32-34~~ individuals vs. 3) - potentially creating circumstances where resisting forcible mating-attempts is more difficult for females. This is a likely explanation given that only the small A females displayed reversed mate choice preferences, while the larger CO females largely reproduced the same mate choice patterns as displayed in individual assays. It is worth testing if similar mate choice assays conducted in larger arenas (such as 90mm petri dishes) would reverse this preference to homotypic choice. At the time these experiments were conducted (since changed) there was an important difference in the mating environment of the ACO and CO selection treatments: whilst the early-reproducing ACO's primarily mated in vials, with only a brief phase in cages for egg-laying, the CO's, with 14 days in cages probably do all mating relevant to fitness in the cage environment. With these differences in the "natural" mating environment of our LTEE, the mate choice patterns found in individual assays seem to show us that A-selected females have a preference for homotypic mating but are unable to exercise it in a group mating setting with larger males.

Asynchronous development time alone can contribute to reproductive isolation, as seen in the experimental work of Miyatake and Shimuzu (1999) on the melon fly. Because the mating times diverged in lines selected for development time, significant RI arose that was evidenced in experimental mate choice trials. Similarly, the extreme early reproduction regime of the A populations means that neither CO individuals nor A/CO hybrids could hybridize, as the latter groups would not even be sexually mature before a generation is completed. This scenario is, of course, artificial because natural populations generally do not grow on synchronised, discrete generations. they are not directly responsible for the assortative mating patterns observed in the controlled mate-choice assays.

Post-mating, we report no evidence of reproductive isolation deriving from gamete incompatibility, as would be evident in reduced egg hatch rates. Similarly, our measurements of larva to adult viability data show that there are no hybrid treatments with lower levels of viability

than the purebred parental populations. In fact, we observed trends of hybrid vigour, with some of the hybrid treatments displaying higher larva to adult viability than the parental treatments. Interestingly, the larva to adult viabilities in F2 treatments were significantly higher than that of the parental selection treatments and one F1 treatment. While the increased viability in hybrids could suggest the existence of homozygous deleterious variation in developmental loci in our selection regimes, this should be consistently seen in both F1 and F2 generations, instead of being more common in the F2 generation. Development time and body size displayed no suggestion of abnormalities indicative of developmental instability, with intermediate hybrid trait values relative to the parental treatments in both sexes.

The competitive reproductive fitness of hybrids was generally intermediate to the parental treatments, and we found no evidence of hybrid sterility. Given that both assays subjected a focal fly to 4 competitors of the same sex, we expect focal flies to sire/dam $\frac{1}{5}$ th of the offspring produced, if all else was equal. In female reproductive fitness assays, our populations did cluster around the predicted threshold, with hybrids scoring intermediate fitness levels relative to the low fitness ACOs and high fitness COs. In male reproductive fitness assays though, fitness fell well below the predicted threshold in all treatments. We rule out the possibility of this being a case of Haldane's rule, because male fitness falls well below predicted levels in parental males as well as hybrids. We believe that this is a consequence of the usage of IVb flies as competitors/mates in these assays. Selected as common ancestors to both selection regimes, the IVbs have evolved in allopatry with both the populations, with a slightly different maintenance regime. We believe that the reduced male fitness estimates may be a consequence of homotypic mate choice by IVb females in male fitness assays. While we do see different levels of competitive reproductive output for the different treatments in the female assay, it is not possible to disentangle male mate choice from innate fecundity and harm resistance differences in the females of different selection regimes. Overall, in spite of the depressed male fitness output relative to predictions, hybrids in both sexes still consistently showed intermediacy in fitness, suggesting no reproductive sterility and a link to an additive trait such as body size.

The comparison between F1 and F2 reproductive fitness revealed an interesting trend: F1 fitness was consistently higher than that of F2 offspring, however, this difference was statistically significant in only 1 out of 4 comparisons ~~2-out-of-12-comparisons~~ (2 reciprocal crosses*~~3-replicate treatment pairs~~*2 sexes). We interpret this with caution as a possible trend of hybrid breakdown as a consequence of incompatible gene complexes evolving in the two selection regimes.

We generally found high repeatability across our three independent replicates, which suggests that the characters measured and pre-mating RI observed stem from consistent treatment differences between the CO and ACO populations. Internal random processes of divergence in allopatry, such as the coevolution of arbitrary signaller-receiver relationships in

676 allopatry may also drive the evolution of RI. A system like ours opens the door to a potential test
677 for such divergence if RI were investigated between replicate populations *within* selection
678 treatment, rather than *between* treatments. Although our three-decade experiment seems too
679 young to see more than hints of post-mating RI, more sensitive queries of characters like
680 developmental stability (e.g., as measured by fluctuating asymmetry) may be of interest. Further
681 investigation of the sources of premating RI and its environmental sensitivity are warranted.
682
683

REFERENCES

- Abbot, P., & Withgott, J. H. 2004. "Phylogenetic and molecular evidence for allochronic speciation in gall-forming aphids (pemphigus)." *Evolution*, **58**, 539–553.
- Arbuthnott, D., T.Y. Fedina, S.D. Pletcher, and D.E.L. Promislow. 2017. "Mate choice in fruit flies is rational and adaptive." *Nature Communications* 8(1):13953.
- Arthur, N.J., Dyer, K.A. 2015. "Asymmetrical sexual isolation but no postmating isolation between the closely related species *Drosophila suboccidentalis* and *Drosophila occidentalis*." *BMC Evolutionary Biology*. 15:38.
- Ashburner, M., Golic, K.G. and Hawley, R.S. 2005. "*Drosophila*: A Laboratory Handbook. Second Edition." Cold Spring Harbor, NY: The University of Chicago Press.
- Barker, J.S.F. and L.J. Cummins. 1969. "The effect of selection for sternopleural bristle number on mating behavior in *Drosophila melanogaster*." *Genetics* 61(3):713.
- Bateman, A.J. 1948. "Intra-sexual selection in *Drosophila*." *Heredity* 2(3):349-368.
- Bates, D., Mächler, M., Bolker, B., Walker, S., 2015. "Fitting linear mixed-effects models using lme4." *Journal of Statistical Software* 67, 1–48
- Belkina, E.G., E.B. Naimark, A.A. Gorshkova, and A.V. Markov. 2018. "Does adaptation to different diets result in assortative mating? Ambiguous results from experiments on *Drosophila*." *Journal of Evolutionary Biology* 31(12):1803-1814.
- Bonduriansky, R., A. Maklakov, F. Zajitschek, and R. Brooks. 2008. "Sexual selection, sexual conflict and the evolution of ageing and life span." *Functional Ecology* 22(3):443-453.
- Brooks, M.E., Kristensen, K., van Benthem K.J., Magnusson A., Berg, C.W., Nielsen, A., Skaug, H.J., Maechler, M., Bolker, B.M. 2017. "glmmTMB Balances Speed and Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling." *The R Journal*, **9**(2), 378–400. doi:10.32614/RJ-2017-066.
- Byrne, P.G. and W.R. Rice. 2006. "Evidence for adaptive male mate choice in the fruit fly *Drosophila melanogaster*." *Proceedings. Biological sciences* 273(1589):917-922.

712 Chippindale, A.K., J.A. Alipaz, H.-W. Chen, and M.R. Rose. 1997a. "Experimental evolution of
713 accelerated development in *Drosophila*. 1. Development speed and larval survival." *Evolution*
714 51(5):1536-1551.

715 Chippindale, A.K., J.A. Alipaz, and M.R. Rose. 2004a. "Experimental Evolution of Accelerated
716 Development in *Drosophila*. 2. Adult Fitness and the Fast Development Syndrome." Pp. 413-435
717 in *Methuselah Flies*: World Scientific.

718 Chippindale, A.K., D.T. Hoang, P.M. Service, and M.R. Rose. 2004b. "The evolution of
719 development in *Drosophila melanogaster* selected for postponed senescence." Pp. 370-389 in
720 *Methuselah Flies*: World Scientific.

721 Chippindale, A.K., A.M. Leroi, H. Saing, D.J. Borash, and M.R. Rose. 1997b. "Phenotypic
722 plasticity and selection in *Drosophila* life history evolution. 2. Diet, mates and the cost of
723 reproduction." *Journal of Evolutionary Biology* 10(3):269-293.

724 Coyne, J.A. and H.A. Orr. 2004. "Speciation": Sinauer.

725 del Solar, E. 1966. "Sexual isolation caused by selection for positive and negative phototaxis and
726 geotaxis in *Drosophila pseudoobscura*." *Proceedings of the National Academy of Sciences*
727 56(2):484.

728 Dettman, J.R., J.B. Anderson, and L.M. Kohn. 2008. "Divergent adaptation promotes
729 reproductive isolation among experimental populations of the filamentous fungus *Neurospora*."
730 *BMC Evolutionary Biology* 8(1):35.

731 Dettman, J.R., C. Sirjusingh, L.M. Kohn, and J.B. Anderson. 2007. "Incipient speciation by
732 divergent adaptation and antagonistic epistasis in yeast." *Nature* 447(7144):585-588.

733 Dobzhansky, T. 1937. "Genetics and the Origin of Species": Columbia Univ. Press, New York;
734 2nd Ed., 1941; 3rd Ed., 1951.

735 Dukas, R. 2005. "Learning affects mate choice in female fruit flies." *Behavioral Ecology*
736 16(4):800-804.

737 Eubanks, M., Blair, C., & Abrahamson, W. 2003. "One host shift leads to another? Evidence of
738 host-race formation in a predaceous gall-boring beetle." *Evolution*, 57, 168–172.

Forbes, A. A., Powell, T. H. Q., Stelinski, L. L., Smith, J. J., & Feder, J. L. 2009. "Sequential sympatric speciation across trophic levels." *Science*, **323**, 776–779.

Fox, J., Weisberg, S., 2019. "An {R} companion to applied regression", third edition. Sage, Thousand Oaks, CA.

Friberg, U. and G. Arnqvist. 2003. "Fitness effects of female mate choice: preferred males are detrimental for *Drosophila melanogaster* females." *J Evol Biol* 16(5):797-811.

Friesen, V. L., Smith, A. L., Gómez-Díaz, E., Bolton, M., Furness, R. W., González-Solís, J., & Monteiro, L. R. 2007. Sympatric speciation by allochrony in a seabird. *Proceedings of the National Academy of Sciences of the United States of America*, **104**, 18589–18594.

Fry, J. 2009. "Laboratory experiments on speciation." Pp. 631-656 in *Experimental Evolution: Concepts, Methods, and Applications of Selection Experiments*, edited by T. Garland and M. Rose.

Gavrilets, S. 2014. "Is sexual conflict an "engine of speciation"?" *Cold Spring Harbor perspectives in biology* 6(12):a017723-a017723.

Gavrilets, S., G. Arnqvist, and U. Friberg. 2001. "The evolution of female mate choice by sexual conflict." *Proceedings of the Royal Society of London. Series B: Biological Sciences* 268(1466):531-539.

Ghosh, S.M. and A. Joshi. 2012. "Evolution of reproductive isolation as a by-product of divergent life-history evolution in laboratory populations of *Drosophila melanogaster*." *Ecology and Evolution* 2(12):3214-3226.

Hartig, F. 2022. "DHARMa: Residual Diagnostics for Hierarchical (Multi-Level / Mixed) Regression Models." R package version 0.4.6

Hood, G. R., Forbes, A. A., Powell, T. H. Q., Egan, S. P., Hamerlinck, G., Smith, J. J., & Feder, J. L. 2015. "Sequential divergence and the multiplicative origin of community diversity." *Proceedings of the National Academy of Sciences of the United States of America*, **112**, E5980–E5989.

Horner, J. D., Craig, T. P., & Itami, J. K. 1999. "The influence of oviposition phenology on survival in host races of *Eurosta solidaginis*." *Entomologia Experimentalis et Applicata*, **93**, 121–129.

768 Hurd, L.E. and R.M. Eisenberg. 1975. "Divergent Selection for Geotactic Response and
769 Evolution of Reproductive Isolation in Sympatric and Allopatric Populations of Houseflies." *The*
770 *American Naturalist* 109(967):353-358.

771 Jagadeeshan, S., Shah, U., Chakrabarti, D., Singh, R.S. 2015 "Female Choice or Male Sex
772 Drive? The Advantages of Male Body Size during Mating in *Drosophila Melanogaster*." *PLOS*
773 *ONE* 10(12):e0144672.

774 Kuznetsova, A., Brockhoff, P.B., Christensen, R.H.B., 2017. "lmerTest package: Tests in linear
775 mixed effects models." *Journal of Statistical Software* 82

776 [Lackey, A.C.R. and Boughman, J.W. 2017. "Evolution of reproductive isolation in stickleback](#)
777 [fish." *Evolution*, 71: 357-372](#)

778 Lande, R. 1981. "Models of speciation by sexual selection on polygenic traits." *Proceedings of*
779 *the National Academy of Sciences* 78(6):3721-3725.

780 Lenth, R. V., 2022. "emmeans: Estimated marginal means, aka least-squares means. R package
781 version 1.7.3." <https://cran.r-project.org/package=emmeans>

782 Lofdahl, K.L., D. Hu, L. Ehrman, J. Hirsch, and L. Skoog. 1992. "Incipient reproductive
783 isolation and evolution in laboratory *Drosophila melanogaster* selected for geotaxis." *Animal*
784 *Behaviour* 44(4):783-786.

785 [Lowry, D. B., Rockwood, R. C., & Willis, J. H. 2008. "Ecological reproductive isolation of coast](#)
786 [and inland races of *Mimulus guttatus*." *Evolution*, 62, 2196–2214.](#)

787 Maan, M.E. and O. Seehausen. 2011. "Ecology, sexual selection and speciation." *Ecology Letters*
788 14(6):591-602.

789 [Mahilkar, A., Nagendra, P., Venkataraman, P., Deshmukh, S. and Saini, S. "Rapid evolution of](#)
790 [pre-zygotic reproductive barriers in allopatric populations" *BiorXiv*:](#)
791 <https://doi.org/10.1101/2023.03.18.533249>

792 Markow, T.A. 1986. "Genetic and sensory basis of sexual selection in *Drosophila*
793 *melanogaster*." Pp. 89-95 in *The evolutionary genetics of invertebrate behavior.*, edited by M.
794 Huettel. New York, NY: Plenum Press.

795 Markow, T.A. and J.P. Ricker. 1992. "Male size, developmental stability, and mating success in
796 natural populations of three *Drosophila* species." *Heredity (Edinb)* 69 (Pt 2):122-127.

- 797 McDonald, J.H. 2014. *Handbook of Biological Statistics*. Baltimore, MD: Sparky House
798 Publishing.
- 799 [Mital, A., Sarangi, M., Nandy, B., Pandey, N., Joshi, A. 2022. "Shorter effective lifespan in](#)
800 [laboratory populations of *D. melanogaster* might reduce sexual selection."](#) *Behav Ecol*
801 [Sociobiol](#) **76**, 52 [Mital, A., Sarangi, M., Joshi, A. 2021. "Evolution of lower levels of inter-locus](#)
802 [sexual conflict in *D. melanogaster* populations under strong selection for rapid development."](#)
803 [BioRxiv](#)
- 804 Miyatake, T. and T. Shimizu. 1999. "Genetic correlations between life-history and behavioral
805 traits can cause reproductive isolation." *Evolution* 53(1):201-208.
- 806 [Muller H.J. 1942. "Isolation mechanisms, evolution and temperature." *Biol. Symp*; 6:71–125](#)
- 807 Najarro, M.A., M. Sumethasorn, A. Lamoureux, and T.L. Turner. 2015. "Choosing mates based
808 on the diet of your ancestors: replication of non-genetic assortative mating in *Drosophila*
809 *melanogaster*." *PeerJ* 3:e1173-e1173.
- 810 Nash, W., I. Mohorianu, and T. Chapman. 2019. "Mate choice and gene expression signatures
811 associated with nutritional adaptation in the medfly (*Ceratitis capitata*)." *Scientific Reports*
812 9(1):6704.
- 813 Nunney, L. 1996. "The response to selection for fast larval development in *Drosophila*
814 *melanogaster* and its effect on adult weight: An example of fitness trade-off." *Evolution*
815 50(3):1193-1204.
- 816 [Ording, G. J., Mercader, R. J., Aardema, M. L., & Scriber, J. M. 2009. "Allochronic isolation](#)
817 [and incipient hybrid speciation in tiger swallowtail butterflies." *Oecologia*, **162**, 523–531.](#)
- 818 Panhuis, T.M., R. Butlin, M. Zuk, and T. Tregenza. 2001. "Sexual selection and speciation."
819 *Trends in Ecology & Evolution* 16(7):364-371.
- 820 Parker, G.A. and L. Partridge. 1998. "Sexual conflict and speciation." *Philosophical transactions*
821 *of the Royal Society of London. Series B, Biological sciences* 353(1366):261-274.
- 822 Partridge, L., A. Ewing, and A. Chandler. 1987. "Male size and mating success in *Drosophila*
823 *melanogaster*: the roles of male and female behaviour." *Animal Behaviour* 35(2):555-562.

Partridge, L. and M. Farquhar. 1983. "Lifetime mating success of male fruitflies (*Drosophila melanogaster*) is related to their size." *Animal Behaviour* 31(3):871-877.

Pitnick, S. and F. García-González. 2002. "Harm to females increases with male body size in *Drosophila melanogaster*." *Proceedings of the Royal Society of London. Series B: Biological Sciences* 269(1502):1821-1828.

Powell, T.H.Q., Nguyen, A., Xia, Q., Feder, J.L., Ragland, G.J. and Hahn, D.A. 2020. "A rapidly evolved shift in life-history timing during ecological speciation is driven by the transition between developmental phases." *J Evol Biol*; 33: 1371– 1386

Prasad, N.G. and A. Joshi. 2003. "What have two decades of laboratory life-history evolution studies on *Drosophila melanogaster* taught us?" *Journal of Genetics* 82(1):45-76.

Prasad, N.G., M. Shakarad, V.M. Gohil, V. Sheeba, M. Rajamani, and A. Joshi. 2000. "Evolution of reduced pre-adult viability and larval growth rate in laboratory populations of *Drosophila melanogaster* selected for shorter development time." *Genet Res* 76(3):249-259.

R Core Team, ~~2020~~2023. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>.

Rice, W.R. and E.E. Hostert. 1993. "Laboratory experiments on speciation: What have we learned in 40 years?" *Evolution* 47(6):1637-1653.

Ritchie, M.G. 2007. "Sexual selection and speciation." *Annual Review of Ecology, Evolution, and Systematics* 38:79-102.

Roff, D. 2002. *Life History Evolution*. Sunderland, MA: Sinauer Associates, Inc.

Rose, M.R., L.N. Vu, S.U. Park, and J.L. Graves. 1992. "Selection on stress resistance increases longevity in *Drosophila melanogaster*." *Experimental Gerontology* 27(2):241-250.

RStudio Team, 2020. RStudio: Integrated development environment for R. Boston, MA. <http://www.rstudio.com/>.

Rundle, H.D., S.F. Chenoweth, P. Doughty, and M.W. Blows. 2005. "Divergent Selection and the Evolution of Signal Traits and Mating Preferences." *PLOS Biology* 3(11):e368.

Rundle, H.D. and P. Nosil. 2005. "Ecological speciation." *Ecology Letters* 8(3):336-352.

851 [Savolainen, V., Anstett, M.C., Lexer, C., Hutton, I., Clarkson, J.J., Norup, M.V., Powell, M.P.,](#)
852 [Springate, D., Salamin, N. and Baker, W.J. 2006 "Sympatric speciation in palms on an oceanic](#)
853 [island." *Nature*:441\(7090\):210-3.](#)

854 Santibañez, S.K. and C.H. Waddington. 1958. "The Origin of Sexual Isolation Between Different
855 Lines Within a Species." *Evolution* 12(4):485-493.

856 Servedio, M.R., G.S.V. Doorn, M. Kopp, A.M. Frame, and P. Nosil. 2011. "Magic traits in
857 speciation: 'magic'; but not rare?" *Trends in Ecology & Evolution* 26(8):389-397.

858 Sharon, G., D. Segal, J.M. Ringo, A. Hefetz, I. Zilber-Rosenberg, and E. Rosenberg. 2010.
859 "Commensal bacteria play a role in mating preference of *Drosophila melanogaster*."
860 *Proceedings of the National Academy of Sciences* 107(46):20051.

861 Singh, A. and Singh, B.N. 2014. "Role of sexual selection in speciation in *Drosophila*."
862 *Genetica* 142(1):23-41.

863 Soans, A.B., D. Pimentel, and J.S. Soans. 1974. "Evolution of Reproductive Isolation in
864 Allopatric and Sympatric Populations." *The American Naturalist* 108(959):117-124.

865 Stearns, S. 1992. ["The evolution of life histories"](#). Oxford, U.K.: Oxford University Press.

866 Syed, Z.A., Chatterjee, M., Samant, M.A. and Prasad, N.G. 2017. "Reproductive Isolation
867 through Experimental Manipulation of Sexually Antagonistic Coevolution in *Drosophila*
868 *melanogaster*." *Scientific Reports* 7, 3330

869 [Tauber, M. J., Tauber, C. A., & Masaki, S. 1986. *Seasonal adaptations of insects*. Oxford, MS:](#)
870 [Oxford University Press.](#)

871 Travers, L.M., F. Garcia-Gonzalez, and L.W. Simmons. 2015. "Live fast die young life history in
872 females: evolutionary trade-off between early life mating and lifespan in female *Drosophila*
873 *melanogaster*." *Scientific Reports* 5(1):15469.

874 Verma, T., Mohapatra, A., Senapati, H.K., Muni, R.K., Dasgupta, P., and Nandy, B. 2022.
875 "Evolution of reduced mate harming tendency of males in *Drosophila melanogaster* populations
876 selected for faster life history". *Behav Ecol Sociobiol* 76, 82.

- Verspoor, R.L., C. Heys, and T.A.R. Price. 2015. "Dyeing insects for behavioral assays: the mating behavior of anesthetized *Drosophila*." *Journal of visualized experiments*: *JoVE*(98):52645.
- Villa, S.M., J.C. Altuna, J.S. Ruff, A.B. Beach, L.I. Mulvey, E.J. Poole, H.E. Campbell, K.P. Johnson, M.D. Shapiro, S.E. Bush, and D.H. Clayton. 2019. "Rapid experimental evolution of reproductive isolation from a single natural population." *Proceedings of the National Academy of Sciences* 116(27):13440.
- Wadsworth, C. B., Woods, W. A., Hahn, D. A., & Dopman, E. B. 2013. "One phase of the dormancy developmental pathway is critical for the evolution of insect seasonality." *Journal of Evolutionary Biology*, **26**, 2359–2368.
- Wallace, S.J., Morris-Pocock, J.A., González-Solís, J., Quillfeldt, P. and Friesen, V.L. 2017. "A phylogenetic test of sympatric speciation in the Hydrobatinae (Aves: Procellariiformes)." *Mol Phylogenet Evol.* Feb;107:39-47.
- White, N.J., Snook, R.R., and I. Eyres. 2020. "The Past and Future of Experimental Speciation." *Trends in Ecology & Evolution* 35(1):10-21.
- Wood, T., & Guttman, S. 1982. "Ecological and behavioral basis for reproductive isolation in the sympatric *Enchenopa-binotata* complex (Homoptera: Membracidae)." *Evolution*, **36**, 233–242.
- Xiao, C., Fard, N.B., Brzezinski, K., Robertson, R.M., Chippindale, A.K. 2019; "Experimental evolution of response to anoxia in *Drosophila melanogaster*: recovery of locomotion following CO₂ or N₂ exposure". *Journal of Experimental Biology* 222 (14):199521.
- Zwaan, B., R. Bijlsma, and R.F. Hoekstra. 1995. "Direct selection on lifespan in *Drosophila melanogaster*" *Evolution* 49(4):649-659.

FIGURE CAPTIONS

Figure 1. The proportion of homotypic compared to heterotypic matings recorded during the female (a), male (b) and group (c) mate choice assay. Female (a,c) and male population identity is shown on the x-axis. An asterisk represents a significant deviation from a random mating ratio as indicated by a repeated G-test for goodness of fit ($p < 0.05$).

Figure 2. The latency (a,b) and duration (c,d) of matings in the choice assays. Female (a,c) and male (b,d) population identity is shown on the x-axis. Letters indicate significant differences between mating types.

Figure 32. The hatchability (a), egg to adult viability (b), body weights (c) and average development time (d) of flies from parental and hybrid crosses. Letters indicate significant differences between cross identities.

Figure 43. The average proportion of red eyed offspring produced by flies from parental and hybrid crosses, in competition with brown eyed competitors. Average proportions were determined for each replicate population and plotted. The dashed line indicates the expected proportion of $\frac{1}{3}$ of the offspring. Letters indicate significant differences between cross identities.