# Package 'aMSE'

June 28, 2022

Title A Framework for Abalone Management Strategy Evaluation

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to facilitate the generation of input data file templates, model

**Description** aMSE provides a formal framework for conducting management strategy evaluation for abalone stocks (although, in principle, such analyses for any sedentary organisms could be conducted). There are functions designed

Type Package

**Version** 0.0.7 **Date** 2022-06-22

initiation and conditioning, and a standard template for the generation of new harvest control rules. The vignette Use_aMSE provides details
of its operation. Rcpp is included in readiness of speeding various
vital dynamics but currently no functions are implemented in Rcpp.
The package has now been checked to operate using R4.0.0.
<b>Depends</b> R (>= $3.5.0$ )
License MIT + file LICENSE
Imports hutils, hplot, captioner, makehtml
RoxygenNote 7.1.1
Suggests knitr, rmarkdown
VignetteBuilder knitr
Encoding UTF-8
<pre>URL https://github.com/haddonm/aMSE</pre>
BugReports https://github.com/haddonm/aMSE/issues
NeedsCompilation no
Author Malcolm Haddon [aut, cre]
R topics documented:
addpops
addrecvar
alldirExists
aMSE
asSAU
uozono
1

2

<i>D</i> , –r · · · · · · · · · · · · · · · · · ·	9
blockE13	
calcprojsel	
C	
$\epsilon$	
$\boldsymbol{c}$	
$\epsilon$	
1	
compareCPUE	
1	
compzoneN	
confirmdir	
copyto	
ctrlfiletemplate	
datafiletemplate	
defineBlock	
definepops	
depleteSAU	
diagnosticsproj	
dodepletion	
lohistoricC	
doproduction	
doprojections	
dosau	
dosauplot	
do_condition	
do_MSE	
lriftrec	4
illzoneDef	
indF1	
indlinenumber	6
indmsy	7
ishery_plots	7
getaav	
getavrec	9
getConst	9
getCPUEssq	0
getLFdata	0
getLFlogL	1
getline	2
getlistvar	2
getLogical	3
getmaxCE	4
getnas	4
getprojyrC	5
getrecdevcolumn	
getsauzone	_
getsingleNum	
getssqparts	1
getStr	1
g	-

4 addpops

	poptosauCE	89
	poptozone	90
	prepareDDNt	91
	prepareprojection	91
	preparesizecomp	92
	print.zone	93
	print.zoneDefinition	94
	printline	94
	putNA	95
	readctrlfile	95
	readdatafile	96
	readsaudatafile	97
	replaceVar	97
	resetexB0	98
	resetLML	99
	restart	99
	rewritecontrolfile	100
	rewritedatafile	100
	runthreeH	101
	sauavrecssq	
	sauplots	
	saurecdevs	
	sautopop	
	saveobject	
	save_hsargs	
	scaleto1	
	setupzone	
	STM	
	summarizeprod	
	sumpop2sau	
	sumpops	
	tasab	
	testequil	
	WtatLen	
	zone	
	zonetosau	113
Index		115

addpops

addpops adds the populations from a single replicate together

# Description

addpops adds the populations from a single replicate together to form zone totals. It can only be used on matureB, exploitB, catch, acatch, and recruits. the sum of acatch is already available as TAC in the projection dynamics object 'zoneDP'

```
addpops(invar, nyrs, reps)
```

addrecvar 5

### **Arguments**

invar the summable variable eg catch, recruits, matureB

nyrs the number of years of data reps the number of replicates

# Value

```
a 2D matrix of yrs x reps
```

### **Examples**

```
print("wait on data")
```

addrecvar

addrecvar adds recruitment variation to end of conditioning step

#### **Description**

addrecvar replicates the historical depletion step in zoneDD reps times and copies the first Nyrs - varyrs of values into each replicate. Then it steps through each replicate adding recruitment variation to the dynamics for the last varyrs years. This is so that when the projections begin each replicate will already have recruitment variability fully developed and there will be no time lag on the recruitment and subsequent dynamics. Take especial note that prior to the projections the proposed harvest strategy needs to be applied to the historical fishery data (in Tasmania this is currently catches and CPUE). This application occurs in lines 60 - 65 in the addrecvar function. This WILL NEED ATTENTION with other jurisdictions.

# Usage

```
addrecvar(
  zoneDD,
  zoneC,
  glob,
  condC,
  ctrl,
  varyrs,
  calcpopC,
  sigR = 1e-08,
  sigB = 1e-08,
  lastsigR = 0.3
)
```

# Arguments

zoneDD	the dynamic zone after historical fishery data has been used to finalize the con-
	ditioning #param zoneDDR the empty dynamic zone object ready for expansion

and variation

zoneC the zone's constants object for each population glob the object containing the global constants condC the object containing the historical fishery data

6 alldirExists

ctrl	the control object
varyrs	the number of years at the end of the historical period to which recruitment variation is to be added
calcpopC	a function that takes the output from herfun and generates the actual catch per population expected in the current year.
sigR	the initial recruitment variation default=1e-08
sigB	the initial biomass cpuie variation default = 1e-08
lastsigR	the recruitment variation to be added to the final varyrs

### Value

an initialized dynamics zone object for the projections with the first year populated

### **Examples**

```
print("wait on suitable data-sets")
```

alldirExists alldirExists Checks the existence of both a run and data directory	
---	--

# **Description**

alldirExists answers the questions 'do both a rundir and a datadir directory exist?' It uses dir.exists and reports existence if already present and provides a stop warning if either does not exist. Of course it can also be used to determine whether a single directory exists. By default the second directory, indir2 = indir1. This allows for the datadir = rundir within aMSE, but also allows for a separate datadir.

# Usage

```
alldirExists(indir1, indir2 = indir1, make = FALSE, verbose = TRUE)
```

# Arguments

indir1	a character string containing the name of the first directory whose existence is to be checked before it is created if it does not already exist.
indir2	a character string containing the name of a second directory whose existence is to be checked, by default this has the same value as indir1
make	if the directory does NOT exist should it be created. default = FALSE; if make=FALSE and a directory does not exist a warning will be given to the console.
verbose	default=TRUE, prints directory status to the console, If make is set to FALSE and a directory does not exist a warning will always be given.

# Value

a message to the screen if the directory exists or is created; if make is TRUE then it also creates the directory as listed in 'indir1'.

aMSE 7

#### **Examples**

```
indirect <- getwd()
alldirExists(indirect)</pre>
```

aMSE

aMSE functions for Conditioning and Running an Invertebrate MSE

# **Description**

The aMSE package provides functions to facilitate the conditioning and running of an invertebrate Management Strategy Evaluation system. The Operating model dynamics are based around using size-based dynamics rather than age-based dynamics (because most invertebrates, like abalone, are difficult to age accurately and consistently). The growth dynamics are described using the inverse logistic curve (see Haddon et al. 2008). Earlier versions of this invertebrate MSE are described on A development version is #useDynLib aMSE, registration = TRUE #importom Rcpp evalCpp available on GitHub at github.com/haddonm/aMSE.

#### Data sets

**constants** is conditioning data for 6 populations

ctrl control file for a 2 MSU 6 population MSE run

midg an abalone tagging data-set from the Actaeons

product productivity matrix from a 2 MSU 6 population example

zone1 the constants common to a zone

tasab Abalone maturity data from two sites on the south-west of Tasmania.

taszoneC a zone list made up of 6 equilibrium populations

taszoneD a list of 8 matrices and 2 arrays defining the dynamics of a zone

# References

Haddon, M., Mundy, C., and D. Tarbath (2008) Using an inverse-logistic model to describe growth increments of blacklip abalone (*Haliotis rubra*) in Tasmania. *Fishery Bulletin* **106**: 58-71.

Haddon, M., Mayfield, S., Helidoniotis, F., Chick, R. and C. Mundy (2013) *Identification and Evaluation of Performance Indicators for Abalone Fisheries*. FRDC Final Report 2007/020. CSIRO Oceans and Atmosphere and Fisheries Research Development Corporation. 295 p.

Haddon, M. and C. Mundy (2016) Testing abalone empirical harvest strategies for setting TACs and associated LMLs, which include the use of novel spatially explicit performance measures. FRDC Final Report 2011/028. CSIRO Oceans and Atmosphere and Fisheries Research Development Corporation. Hobart 182 p

8 aszone

asSAU

asSAU generates an output list of SAU values from the in put zoneDP

#### **Description**

asSAU takes the population based results in zoneDP and converts them into a list based around SAU. saucpue is already calculated during the application of the harvest strategy.

# Usage

```
asSAU(projzone, sauindex, saunames, b0, exb0)
```

### **Arguments**

projzone the population based results list zoneDP plus the saucpue all from doprojection

sauindex the SAU index of each population

saunames the names of each SAU

b0 a vector of the B0 for each SAU exb0 a vector of the ExB0 for each SAU

#### Value

a list of results based around SAU

### **Examples**

```
print("wait on new data")
```

aszone

aszone sums the various SAU dynamics and fishery properties

# Description

aszone calculates zone wide totals by summing the mature and exploitable biomass across SAU, it also sums the catches and recruitment levels. It calculates zone wide harvest rates by dividing the catches by the available exploitable biomass. It also calculates the depletion levels of both the mature and exploitable biomass by dividing their zone totals through time by the initial zone wide B0 and ExB0. Finally, it calculates a zone-wide cpue by catch-weighting each of the SAU cpue values and then summing each set of SAU for each year and iteration.

# Usage

```
aszone(sauzone, zoneCP)
```

### **Arguments**

sauzone the output from the applyharvest strategy function, a large list of arrays or results

zoneCP the constant part of the projection zone

biology\_plots 9

### Value

a list of 8 nyrs x reps matrices, summarizing the fishery outputs at the geographical scale of the zone

# **Examples**

```
print("wait on data")
```

biology\_plots

biology\_plots generates a series of stored plots and tables

# Description

biology\_plots generates the yield vs spawning biomass, weight-at-lenght, and emergence plots for all populations. In addition, it also tabulates the biological properties of each population and SAU and the total zone

# Usage

```
biology_plots(rundir, glb, zoneC, matL = c(30, 210), Lwt = c(80, 210))
```

# Arguments

rundir	the results directory
glb	the globals list
zoneC	the zonal constants by population, zoneC
matL	a vector of two containing the left and right hand size classes for use in the maturity-at-length plots
Lwt	a vector of two containing the left and right hand size classes for use in the

# Value

invisibly returns the biological properties of the populations

wight-atlength plots

```
args(biology_plots)
```

10 calcprojsel

blockE13

blockE13 is an abalone data-set for testing performance measures

### **Description**

blockE13 is a fishery data-set for blacklip abalone (*Haliotis rubra*) from block 13 in the eastern zone, this is Tasmania's Block 13. It constitutes three time-series of the same cpue data in different formats (see below). It is for use when testing the performance measures within Tasmania's MCDA, although it could be used for other purposes, such as illustrating the typical linear relationship between catch and CPUE. Note there will only ever be one less PM value than there are cpue data in the time-series.

#### **Format**

A data.frame of abalone fishery data

year the year of fishing

coeff the back-transformed coefficients from a standardization

scaled the coefficients scaled to a mean of one

**cpue** the coefficients scaled to the nominal geometric mean of the time series, to place it on the nominal scale

catch the related catch from the eastern parts of block 13

# **Subjects**

- performance measure estimation
- relationship between catch and CPUE

# Source

Mundy, C. and J.M. McAllister (2020) Tasmanian Abalone Assessment 2019. IMAS, University of Tasmania.

# **Examples**

data(blockE13)
blockE13

calcprojsel

calcprojsel generates the selectivity and selectivity x weight

# **Description**

calcprojsel is used to produce each projection years' selectivity and selectivity x weight-at-size. projSel is an Nclass x projyrs matrix, a separate selectivity for each projection year, which allows for changing the selectivity during the projection. The projSelWt is a Nclass x projyrs x numpop array, required because each population's weight-at-size relationship will be slihtly different.

calcsau 11

# Usage

```
calcprojsel(zoneC, projC, glb)
```

# **Arguments**

zoneC the constants object

projC the projection object from zone1

glb the globals object

### Value

a list of projSel and projSelWt

# **Examples**

```
print("wait on new example data")
```

calcsau

calcsau compares an input variable to a constant for each SAU

# Description

calcsau divides an input variable, such as matureB or exploitB by their respective constant, such as B0 or ExB0, from a vector of values for each SAU. This is used to calculate the depletion levels of mature and exploitable biomass

# Usage

```
calcsau(invar, saunames, ref0)
```

# **Arguments**

invar either zoneDP\$matureB or zoneDP\$exploitB

saunames the names of each SAU

ref0 either a vector of B0 or ExB0 for each SAU

# Value

```
an array of projyrs x nSAU x reps
```

```
print("wait on new data")
```

12 changecolumn

catchweightCE	catchweightCE uses historical catch-by-sau to make weighted zone cpue
---------------	---

### **Description**

catchweightCE is used when characterizing the historical fisheries data. It uses matching catch-by-sau to generate catch-weighted estimates of zone-wide CPUE through time.

# Usage

```
catchweightCE(cedat, cdat, nsau)
```

# Arguments

cedat historical cpue from condC
cdat historical catches from condC
nsau the number of SAU, from glb

#### Value

a vector of catch-weghted cpue for the zone from SAU cpue estimates

# **Examples**

```
print("wait on suitable data sets")
```

changecolumn

changecolumn alters a selected column of recdevs in the control file

# **Description**

changecolumn is designed to be used when conditioning the OM when optimizing ad hoc recruitment deviates by calculating the SSQ between the predicted and observed CPUE for a selected range of years. If the length of new values is not the same as the selected linerange the function will stop with a warning. When a line is changed then all -1 values are changed to +1

# Usage

```
changecolumn(rundir, filename, linerange, column, newvect, verbose = FALSE)
```

# **Arguments**

_	
rundir	the rundir for the scenario
filename	the character name of the control file
linerange	the linerange within the control file containing the rows representing the selected years of recruitment deviates. Obtained by using 'getrecdevcolumn
column	the sau + 1 to account for the initial year in each row of deviates
newvect	the new values to replace those present. If using optim these would be exp(ans\$par), if 'nlm' exp(ans\$estimate)
verbose	should console reports of before and after be made? default = FALSE

changeline 13

### Value

nothing although values within the control file will be changed and, if verbose=TRUE, it will write to the console.

# **Examples**

```
print("wait on suitable internal data sets")
```

changeline	changeline replaces a given line in a given file with new text
------------	--

# Description

changeline enables a text file to be changed line by line. One identifies a given line, perhaps by using 'findlinenumber', and this can then be replaced by an input character string. Obviously this is a fine way to mess up a data file so use with care.

# Usage

```
changeline(indir, filename, linenumber, newline, verbose = FALSE)
```

### **Arguments**

indir	the directory path in which to find the text file. Usually, rundir
filename	the full name of the text file in quotations.
linenumber	either the line number within the text file to be changed, or, the character name of the variable to be changed, e.g. 'AvRec'
newline	the character string with which to replace the line

should confirmation be output to the console. default=FALSE

# Value

verbose

nothing but it does alter a line in a text file. Optionally it may confirm the action to the console

# See Also

```
findlinenumber, changevar
```

```
print("wait on an example")
```

14 changevar

changevar	changevar can alter the value in a single line in, eg, the control file

# **Description**

changevar is a DANGEROUS function for lazy people. I say that because it can damage your control files if not used carefully. It speeds changing a single value within, say, the control.csv file. For example, if one wanted to conduct a retrospective analysis on what would have happened should we have introduced the HS sooner, one could use something like the following to change the value to 44 implying to start projections in 2017. changevar(varname="CATCHES",newvalue=44,filename="controlsau.csv", rundir=rundir) This can only change a single value on a single line but can be used to alter the values of the variables listed in 'goodnames'. Currently, this only works for Windows machine. Let me know if others are required.

# Usage

```
changevar(filename, rundir, varname, newvalue, prompt = FALSE, verbose = TRUE)
```

# Arguments

filename	name of the file to	be changed. e	g 'control.csv'

rundir name of the scenario directory in which the file is to be found

varname the name of the object whose value is to be changed

newvalue the new value of the object

prompt should allowable names first be listed with current values?

verbose should concluding remarks to console be made. default=TRUE

#### Value

nothing but a file is changed - be careful

# See Also

findlinenumber, changeline

```
print("Wait on some time passing")
```

checkmsedata 15

checkmsedata	checkmsedata contains some tests of the niput MSE saudata file
--------------	--

# Description

checkmsedata contains some tests of the niput MSE saudata file

# Usage

```
checkmsedata(intxt, rundir, verbose = TRUE)
```

# **Arguments**

intxt the data file from readLines used in readsaudatafile

rundir the rundir for the scenario

verbose should test results be put to the console as well as filed, default=TRUE

### Value

nothing but will write a file to rundir and may write to the console

### **Examples**

```
print("wait on example data sets")
```

checksizecompdata Title

# **Description**

Title

# Usage

```
checksizecompdata(rundir, controlfile, verbose = TRUE)
```

# Arguments

rundir the directory in which all files relating to a particular run are to be held.

controlfile default="control.csv", the filename of the control file present in rundir contain-

ing information regarding the run.

verbose Should progress comments be printed to console, default=TRUE#'

# Value

nothing but it does plot a graph

```
print("wait on data sets")
```

16 comparevar

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compareCPUE plots the historical cpue against conditioned cpue

# **Description**

compareCPUE generates a plot of the historical cpue and compares it with the predicted cpue from the conditioning. The predicted is black and the observed is green. This is primarily there are an aid to conditioning the operating model. It also calculates the simple sum of squared differences between the observed and predicted, again to aid in the conditioning.

### Usage

```
compareCPUE(histCE, saucpue, glb, rundir, filen = "", obscol = 2)
```

# **Arguments**

histCE the matrix of historical cpue by SAU

saucpue the predicted cpue from the conditioning on historical data

glb the globals object

rundir the rundir for the given scenario

filen the file name of the plot if it is to be saved

obscol the colour of the observed CPUE on the plots, default=2=red

# Value

a vector of length nsau containing the ssq for each SAU

# **Examples**

```
print("wait on suitable internal data sets")
# sauZone=out$condout$sauZone; saucpue=sauZone$cpue; filen="";histCE=out$condC$histCE
# glb=out$glb;obscol=2
```

comparevar

comparevar generates the quantiles for eahc of a set of input scenarios

# **Description**

comparevar generates quantiles for each of a set of input scenarios. It takes the full timeline of dynamics and outputs just the projections and the quantiles of those projections

```
comparevar(dyn, glbc, scenes, var = "cpue")
```

compzoneN 17

### **Arguments**

dyn this is a list of the out\$sauout\$zonePsau produced by each scenario. This has to

be be generated from teh saved RData files from each scenario

glbc a list of the global objects from each scenario being compared

scenes a list of the out\$ctrl\$runlabel from each scenario

var what variable from the dynamics to summarize, valid names include catch,

acatch, cpue, harvestR, desplsB, depleB, recruit, matureB, and exploitB, default

= 'cpue'

#### Value

a list of the quantiles of the input var for all sau, with each scenario being a different component of the quantscen list, and a list of three dimensional arrays of the actual values of var in the varc list.

#### See Also

plotscene

### **Examples**

```
print("wait on internal datasets")
```

compzoneN compares numbers-at-size before/after depletion

# Description

compzoneN generates a plot comparing the unfished numbers-at-size with those for a given level of depletion.

### Usage

```
compzoneN(unfN, curN, glb, yr, depl, ssc = 5, LML = 0, rundir = "")
```

# **Arguments**

unfN	the unfished numbers-at-size from getzoneprops
curN	the current numbers-at-size from getzoneprops
glb	the global object
yr	the year of the dynamics
depl	the depletion level of the current n-a-s

ssc index for starting size class. thus 1 = 2, 2 = 4, 5 = 10, etc. default = 5 for it plots

size classes from 10mm up

LML the legal minimum length in the comparison year

rundir the results directory, default = "" leading to no .png file, just a plot to the screen.

### Value

invisibly the filename ready for logfilename

18 copyto

#### **Examples**

```
print("still to be developed")
# unfN=unfN; curN=depN;glb=glb; yr=1; depl=0.3993; LML=132; rundir=rundir
```

confirmdir

confirmdir checks to see if a directory exists and makes it if not

# **Description**

confirmdir enables one to be sure a selected directory exists. If it has not bee created then confirmdir will create it if it does not already exist. This is useful when defining output directories on the hard drive where large objects may be stored.

### Usage

```
confirmdir(x, make = TRUE, verbose = TRUE)
```

# **Arguments**

x the directory to be checked and created if necessary

make should the directory be created if it does not already exist? default=TRUE

verbose should responses be sent to the console? default=TRUE

#### Value

nothing but it can create a directory

### **Examples**

```
x <- tempdir()
confirmdir(x)</pre>
```

copyto

copyto copies a vector of files from one scenario directory to annother

### **Description**

copyto copies a vector of files (see examples) from one scenario's directory to another. If a filename includes the name of the scenario, eg controlM15h75.csv, is found in the scenario M15h75, and it is to be copied to, say, M15h5, then the filename will be changed automatically. Care should be taken when using this function as it writes files and potentially new directories to your storage drives.

```
copyto(prefixdir, fromdir, todir, filelist, makenew = TRUE, verbose = TRUE)
```

ctrlfiletemplate 19

#### **Arguments**

prefixdir the directory (path) containing the different scenarios

fromdir just the name of the directory from which to source the files listed in filelist (no path)

todir just the name of the new destination directory (no path)

filelist a vector of filenames to be copied, as character.

makenew if the 'todir' does not exist should it be created using dir.create? default = TRUE

verbose should details be printed to the console, default=TRUE

#### Value

Nothing but it will transfer files and change their names. This information will be printed to the console if verbose = TRUE

### **Examples**

ctrlfiletemplate

ctrlfiletemplate generates a template input control file

### **Description**

ctrlfiletemplate generates a standardized control file template. Generate this and then modify the contents to suit the system you are attempting to simulate. Defaults to 100 replicates. There needs to be as many recdevs as there are conditioning catches (in the template = 58 rows. If they are all set to -1, the default, then they will have no effect and recdevs will be taken off the stock-recruitment curve with random deviates defined by withsigR found in the ctrl file. If devrec = 1.0 then the recdevs from 1980 to 2016 will be set to 1.0 which means there will be no random variation and the system is ready to be conditioned on those recdevs to improve the predicted CPUE during the historical conditioning period. If devrec = 0.0, then the recdevs for 1980 - 2016 will be set to values that have already conditioned the model and provides a reasonable fit to the observed CPUE trends.

```
ctrlfiletemplate(indir, filename = "controlsau.csv", devrec = -1)
```

20 datafiletemplate

#### **Arguments**

indir directory in which to place the control.csv file

filename the name for the generated ctrlfile, a character string that defaults to control-

sau.csv.

devrec what form should the recdevs take? valid values are -1, the default (= random

deviates imposed dependent upon withsigR), 0.0, where a set of conditioned values between 1980 and 2016 are imposed that improve the fit to the observed CPUE as inserted by ctrlfiletemplate, or 1.0, which prepares the recdevs for a

round of conditioning ready to improve the observed fit to cpue.

#### Value

invisibly the fill path and name of the control file. More importantly, it write a control file template to that directory.

#### See Also

datafiletemplate, readsaudatafile, readctrlfile

### **Examples**

```
## Not run:
yourdir <- tempdir()
datafiletemplate(nSAU=8,yourdir,"saudata_test.csv")
ctrlfiletemplate(yourdir,filename="testctrl.csv",devrec=-1) #
control <- readctrlfile(yourdir,"testctrl.csv")
str(control,max.level=1)
## End(Not run)</pre>
```

datafiletemplate

datafiletemplate generates a template input datafile akin to M15h75

# **Description**

datafiletemplate generates a standard input datafile to use as a template, go in and edit it appropriately to suit your own needs. It contains the probability distributions that are sampled to provide the necessary biological constants for each population. It also contains the proportional distribution of recruitment levels determined for Tasmania by examining the GPS data-logger data for the zone and allocating recruitment in proportion to the relative yield by area over the last 8 years.

### Usage

```
datafiletemplate(nSAU, indir, filename = "saudata_test.csv")
```

### **Arguments**

nSAU the number of SAU in the zone

indir the directory into which to place the generated data file

filename the name for the generated datafile, a character string, defaults to saudata\_test.csv,

which is the default within the ctrlfiletemplate function

defineBlock 21

#### Value

a standard definition data file, to be read by readdatafile whose name and path is returned invisibly

### See Also

ctrlfiletemplate, readsaudatafile, readctrlfile

# **Examples**

```
## Not run:
   yourdir <- tempdir()
   datafiletemplate(nSAU=8,yourdir,"saudata_test.csv")
   constants <- readsaudatafile(yourdir,"saudata_test.csv")
   str(constants,max.level=1)
   print(constants[,1:10])
## End(Not run)</pre>
```

defineBlock

defineBlock subdivides defined number of populations into blocks

# **Description**

subdivides the defined number of populations into blocks in the numbers defined in the constant blkpop read into 'constants'. It allocates the populations sequentially to numblk blocks; if numblk does not divide into numpop exactly, any remainder is put into the final block. For block can read SAU = spatial assessment unit.

# Usage

```
defineBlock(numblk, blknum, numpop)
```

# **Arguments**

numblk number of blocks in the simulated zone, from 'constants' blknum number of populations in each block, from 'constants' numpop the total number of populations in the zone

# Value

a vector of length numpop, indexing which block each pop is in.

```
nblock <- 4
blkpop <- c(3,4,8,2)
blockI <- defineBlock(nblock,blkpop,numpop=17)
print(blockI)
cat(length(blockI),sum(blkpop),"\n")</pre>
```

22 definepops

definepops	definepops makes a parameter vector, popdef, for each population

# **Description**

definepops makes a parameter vector, popdef, for each population. Each population will have slightly different biological properties sampled from probability density distributions defined in const=condDat\$constants. The input can be either the whole of condDat or just condDat\$constants, which contains the definitions of the probability density functions used to define each population.

# Usage

```
definepops(inSAU, inSAUindex, const, glob)
```

### **Arguments**

inSAU the number of SAUs defined in the zone, a scalar

inSAUindex a vector of the block index for each population

const a matrix with rows containing the PDF parameters for each of the biological parameters. The columns of the matrix reflect values for each populations from readdatafile

glob the globals object from readzonefile

# Value

a matrix with a row for each population and whose columns are parameters defining the biological parameters for each population.

```
## Not run:
    data(zone1)
    glb <- zone1$globals
    data(constants)
    ans <- makezoneC(zone1,constants)
    zoneC <- ans$zoneC
    popdefs <- ans$popdefs
    print(popdefs)
## End(Not run)</pre>
```

depleteSAU 23

depleteSAU	depleteSAU resets zoneD, approximately to an input depletion level

# **Description**

depleteSAU resets the depletion level of the whole zone and does this by searching for the harvest rate that leads to the mature biomass in each population is as close as possible to the desired depletion level. This means the individual populations will likely vary around the target depletion, so the depletion across the zone will only be approximately at the target depletion. The depletion is measured relative to the effective B0 as that takes account of any larval dispersal. This function uses the production curve array to search for harvest rates that bound the target depletion and then re-searches across those bounds using len intervals.

# Usage

```
depleteSAU(zoneC, zoneD, glob, initdepl, product, len = 15)
```

# Arguments

zoneC	the constants components of the simulated zone
zoneD	the dynamic components of the simulated zone
glob	the general global variables
initdepl	a vector of target depletion levels for each SAU. This is found in zone1\$condC, but could obviously be modified in each run.
product	the production curve matrix from doproduction
len	the number of intervals in the trial harvest rates

### Value

a revised zoneD object

# **Examples**

```
## Not run:
    data(zone)
    glb <- zone$glb
    depl <- rep(0.3,glb$nSAU)
    zoneDD <- depleteSAU(zone$zoneC,zone$zoneD,glb,initdepl=depl,zone$product)
    sum((zoneDD$matureB[1,]/sum(zoneDD$matureB[1,]))*zoneDD$deplsB[1,])
    mean(zoneDD$deplsB[1,])</pre>
```

## End(Not run) # zoneC=zone\$zoneC;zoneD=zone\$zoneD;glob=zone\$glb;initdepl=origdepl;product=zone\$product;le

24 dodepletion

diagnosticsproj	diagnosticsproj plots a series of diagnostics to DiagProj	

### **Description**

diagnosticsproj provides a series of plots and results that illustrate the properties of the projections. These include the residuals between SAU actual catches and their predicted catches. But also a series of plots of the projections for only nrep trajectories to illustrate that the dynamic variables are changing through time in a plausible or 'realistic' manner.

### Usage

```
diagnosticsproj(zonePsau, glb, rundir, nrep = 3)
```

### **Arguments**

zonePsau the SAU scale object containing the dynamics results

glb the global constants object rundir the rundir for the scenario

nrep the number of replicate trajectories to plot; default=3

#### Value

Nothing but it does add some plots to rundir

### **Examples**

```
print("wait on suitable data sets")
```

dodepletion	dodepletion resets zoneD to an input depletion level
-------------	--

# **Description**

dodepletion resets the depletion level of the whole zone and does this by searching for the harvest rate that leads to the sum of the mature biomass, across populations, divided by the sum of the effective B0 across populations is as close as possible to the desired depletion level. This means the individual populations will likely vary around the target depletion, but across the zone it will be correct. The depletion is measured relative to the effective B0 as that takes account of any larval dispersal. This function uses the production curve matrix to search for harvest rates that bound the target depletion and then re-searches across those bounds using len intervals.

```
dodepletion(zoneC, zoneD, glob, depl, product, len = 15)
```

dohistoricC 25

### **Arguments**

zoneC	the constants components of the simulated zone
zoneD	the dynamic components of the simulated zone
glob	the general global variables
depl	the target depletion proportion for the whole zone
product	the production curve matrix from doproduction
len	the number of intervals in the trial harvest rates

### Value

a revised zoneD object

# **Examples**

```
## Not run:
  data(zone1)
  glb <- zone1$globals</pre>
  data(constants)
  ans <- makezoneC(zone1,constants)</pre>
  zoneC <- ans$zoneC</pre>
  glb <- ans$glb # now contains the move matrix</pre>
  ans <- makezone(glb,zoneC)</pre>
  zoneC <- ans$zoneC
  zoneD <- ans$zoneD
  ans2 <- modzoneC(zoneC,zoneD,glb)</pre>
  zoneC <- ans2$zoneC # now has MSY and dep1MSY</pre>
  product <- ans2$product</pre>
  zoneDD <- dodepletion(zoneC,zoneD,glb,depl=0.3,product)</pre>
  sum((zoneDD$matureB[1,]/sum(zoneDD$matureB[1,]))*zoneDD$deplsB[1,])
  mean(zoneDD$dep1sB[1,])
## End(Not run)
```

dohistoricC

dohistoricC imposes the historical catches on an unfished zone

# Description

dohistoricC is used during the conditioning of the zone/region and imposes the historical catches, held in the zone data object obtained using readzonefile, onto an unfished initial zone, and it does this by imposing the time-series of catches to each SAU/block. The operation is through the use of oneyearC, which imposes one year's catches, which are a vector of SAU catches for each year of the series.

```
dohistoricC(zoneDD, zoneC, glob, condC, calcpopC, sigR = 1e-08, sigB = 1e-08)
```

26 doproduction

### **Arguments**

zoneDD The input unfished dynamic zone, zoneD, object

zoneC the zone constants object, zoneC

glob the globals object

condC from the zone1 object, contains historical fisheries data plus

calcpopC a function that takes the output from herfun and generates the actual catch per

population expected in the current year.

sigR the recruitment variation included

sigB the variability introduced to the catches by population by fishers not knowing

the distribution of exploitable biomass exactly. What this value should be is unknown, the default=1e-08, is arbitrary but avoids any effective fisher allocation

error between populations.

#### Value

a zoneD object

#### See Also

```
oneyearsauC, oneyearcat, oneyearrec, do_MSE
```

# **Examples**

```
print("wait on some data sets")
# zoneD,zoneC,glob=glb,condC,calcpopC=calcpopC,sigR=1e-08,sigB=1e-08
```

doproduction

doproduction estimates a production curve for each population

# **Description**

doproduction estimates a production curve for each population in the simulated zone. It does this by sequentially applying a series of increasing harvest rates and running the populations to equilibrium to discover, empirically, the yield and other details, such as exploitable biomass, mature biomass, the actual harvest rate, the catch or yield, the mature depletion, and the relative cpue. Keep in mind that the time taken to run this function depends on the number of populations but mainly on the length of the harvest rate sequence, which is determined by lowlim, uplim, and inc. The lonjger the sequence the slower the run. However, the estimates of MSY and related statistics are expected to have better accuracy (resolution) the finer the harvest rate increments. So when initiating the zone it is best to have a long sequence of finely incremented harvest rates that take a long time.

```
doproduction(zoneC, zoneD, glob, lowlim = 0, uplim = 0.35, inc = 0.005)
```

doprojections 27

# **Arguments**

zoneC	the constants components of the simulated zone
zoneD	the dynamic components of the simulated zone
glob	the general global variables
lowlim	the lower limit of harvest rate applied, default=0.0
uplim	the upper limit of harvest rate applied, default=0.35
inc	the harvest rate increment at each step, default=0.005

# Value

an array of the six productivity variables by numpop by the number of harvest rates applied

# **Examples**

```
print("wait") # zoneC=zoneC; zoneD=zoneD; glob=glb; lowlim=0.0;uplim=0.4;inc=0.01
```

doprojections

doprojections conducts the replicate model runs for Tasmania

# Description

doprojections conducts the replicate model runs for Tasmania using the mcdahcr and the input hsargs. Note the use of a list that takes up both the population and catch numbers-at-size and puts that into getdata. By referencing the iteration in the arrays being passed to getdata the object sizes are being reduced and hence, hopefully, the runtime will be reduced accordingly.

```
doprojections(
  ctrl,
  zoneDP,
  zoneCP,
  glb,
  hcrfun,
  hsargs,
  sampleCE,
  sampleFIS,
  sampleNaS,
  getdata,
  calcpopC,
  makehcrout,
  verbose = FALSE,
  ...
)
```

28 doprojections

# Arguments

ctrl	the ctrl object from readctrlfile
zoneDP	the object used to contain the dynamics from the replicate model runs
zoneCP	the object used to contain the constants for each population used in model dynamics
glb	the object containing the global constants for the given run
hcrfun	the name of the harvest control rule that is used to calculate the multiplier for the previous aspirational catches (possibly for each SAU but possibly the TAC for the whole zone) so as to estimate the aspirational catches or TAC or the following year
hsargs	the constants used to define the workings of the hcr
sampleCE	a function that generates the CPUE statistics
sampleFIS	a function that generates the FIS statistics
sampleNaS	a function that generates the Numbers-at-size samples
getdata	a function that gathers all the data required by the hcrfun and combines it into an hcrdata object ready for the hcrfun. It is expected to call sampleCE, sampleFIS, and sampleNAS, even if they only return NULL.
calcpopC	a function that takes the output from herfun and generates the actual catch per population expected in the current year.
makehcrout	is a function from HS.R that produces an object that is updated in each iteration by the hcrfun. If no such object is required then have a function that returns NULL.
verbose	should the iterations be counted on the console?
	the ellipsis used in case any of the functions herfun, sampleCE, sampleFIS, sampleNas, and getdata require extra arguments not included in the default named collection

# Value

a list containing the full dynamics across all years zoneDP, and the final output from the HCR as herout

# See Also

```
oneyearsauC, make_html, do_MSE
```

```
print("wait on suitable internal data sets")
```

dosau 29

dosau

dosau plots the conditioning history for the dynamics

### **Description**

dosau plots the deplsB, cpue, matureB, catch, harvestR, and recruits for a given SAU during the years of conditioning. This aims to assist the conditioning process by illustrating the state of the sau during and at the end of conditioning on the fishery. If extra is TRUE then it also plots the exploitable biomass and the depletion of the exploitable biomass. A loess fit is also plotted onto the recruitments plot to give some insight into the recruitment deviates. Unfortunately, the 'true' predicted recruitment levels (without variation) cannot be easily estimated because the implementation of larval movement disturbs how many recruits are present each year in each population and hence each SAU.

### Usage

```
dosau(inzone, glb, picksau, histCE, yrnames, recdev)
```

# Arguments

inzone the conditioned zone (zoneDD) after it has been converted to sau scale by using

getsauzone

glb the global constants object picksau which sau should be plotted

histCE the historical cpue series from condC

yrnames the years of historical catches eg 1973:2019

recdev the recdevs for a single SAU

#### Value

nothing but it does generate a plot

#### **Examples**

```
print("wait on suitable data-sets")
```

dosauplot

dosauplot generates the plot of the chosen variable for each sau

# **Description**

dosauplot takes the results of the prepareproject and the projection function, it combines the trajectories across the years of conditioning and the projection years, for all replicates, and plots all replicates for the years from 'startyr' to the final year of the projections. It includes the median and inner 90 plot, if the historical conditioning CPUE is included in histCE, it also includes the original cpue series. Beware of trying to plot the historical cpue in years where there are no historical cpue data, although there are error captures at work to limit the pot to available.

30 do\_condition

#### **Usage**

```
dosauplot(
  ylabel,
  postrep,
  glb,
  startyr,
  addCI = FALSE,
  CIprobs = c(0.05, 0.5, 0.95),
  histCE = NULL,
  addCE = FALSE
)
```

### **Arguments**

ylabel the variable name to be plotted, used as a y-axis label

postrep the replicate values from the replicated projection zoneDP for the variable se-

lected taken from the output of 'zonetosau'.

glb the global constants object

startyr the index for the first year of the conditioning dynamics to include

addCI should quantile confidence bounds be included on the plots, default=FALSE

CIprobs the quantiles used to generate the confidence intervals. The default = c(0.05, 0.5, 0.95)

histCE the historical cpue data if included, default = NULL

addCE should historical cpue be included on the plots, default=FALSE

### Value

invisibly, a list of CI and median for each SAU

### **Examples**

```
print("wait on suitable built in data sets")
```

do\_condition

do\_condition is a function to assist with conditioning the model

#### **Description**

do\_condition is a utility function that can be used to speed the conditioning of the operating model on available fishery data. During the generation of the simulated zone the operation that takes the longest is the estimation of the productivity of each population. The productivity is estimated at equilibrium and getting there is what takes the time. When conditioning the model the objective is to match the predicted fishery response to the historical catches to the observed responses. 'do\_condition' generates the simulated zone without running the projections, which simplifies any searches for optimal values of AvRec (average unfished recruitment), and especially for suitable values of recruitment deviates. If any of the initdepl (initial depletion) values, as listed in the control file are less than 1.0 then before applying the historical catches it first depletes each population within each SAU to the initdepl value for each SAU. For this to work doproduct must be TRUE. This will slow down any searches for an optimum set of AvRec and recdevs, so a different strategy for doing that will be needed.

do\_*MSE* 31

#### Usage

```
do_condition(
  rundir,
  controlfile,
  calcpopC,
  verbose = FALSE,
  doproduct = FALSE,
  dohistoric = TRUE,
  mincount = 100
)
```

#### **Arguments**

rundir the full path to the directory in which all files relating to a particular run are to

be held.

controlfile the filename of the control file present in rundir containing information regarding

the particular run.

calcpopC a function that takes the output from herfun (either the aspirational catch x SAU

or TAC x zone) and generates the actual catch per population in each SAU ex-

pected in the current year.

verbose Should progress comments be printed to console, default=FALSE

doproduct should production estimates be made. default=FALSE

dohistoric should the historical catches be applied. Default=TRUE

mincount determines the minimum sample size for a size-composition sample to be in-

cluded in plots and analyses. Default = 100

# Value

a large list containing tottime, projtime, starttime, glb, ctrl, zoneDD, zoneDP, projC, condC, sauout, and outzone

# See Also

makeequilzone, dohistoricC, prepareprojection, doprojections

```
print("wait on suitable data sets in data")
# rundir=rundir; controlfile=controlfile;calcpopC=calcexpectpopC
# verbose=TRUE; doproduct=FALSE; dohistoric=TRUE; mincount=100
```

32 do\_MSE

### **Description**

do\_MSE is a form of meta-function that contains the code and function calls that constitute a single scenario run for aMSE. This is the reason the argument list is as long as it is. The code has been encapsulated like this to simplify development, maintenance, and distribution. Already with the number of variant conditioning examples produced as tests of the code-base, errors have arisen through a failure to propagate all changes to the code now in this function to all the separate MSE run files used to manage each scenario. A potential flaw lies with the need to apply the jurisdictionHS functions to any historical fishery data used to condition the operating model, ready for the first year of the projections. Currently, the code used is unique to the Tasmanian case and this obviously still requires generalization. This may require a new function to be produced in each jurisdictionHS.R file, but that is still under consideration. Part of the development of this meta-function will be to further articulate the generation of results, and the inclusion of error traps on the argument list entries. To aid in the conditioning the option of not calculating the productivity has been added. By default it will occur and when projecting it will also always occur. HSstats is a list that is always saved and contains performance statistics for the HS used (currently contains sum5 and sum10 the cumulative catches to 5 and 10 years into the projections).

# Usage

```
do_MSE(
 rundir,
  controlfile,
 hsargs,
 hcrfun,
  sampleCE,
  sampleFIS,
  sampleNaS,
  getdata,
  calcpopC,
 makeouthcr,
  varyrs = 7,
  startvr = 42.
  verbose = FALSE,
  ndiagprojs = 3,
  savesauout = FALSE,
  cutcatchN = 56,
 matureL = c(70, 200),
 wtatL = c(80, 200),
 mincount = 100
)
```

### **Arguments**

rundir	the full path to the directory in which all files relating to a particular run are to be held.
controlfile	the filename of the control file present in rundir containing information regarding the particular run.
hsargs	the constants used to define the workings of the hcr
hcrfun	the name of the harvest control rule that is used to calculate the aspirational catches by SAU or TAC by zone for the following year
sampleCE	a function from jurisdictionHS.R that generates the CPUE statistics from the

projected data

do\_MSE

sampleFIS	a function from jurisdictionHS.R that generates the FIS statistics
sampleNaS	a function from jurisdictionHS.R that generates the Numbers-at-size samples
getdata	a function that gathers all the data required by the herfun and combines it into an herdata object ready for the herfun
calcpopC	a function that takes the output from herfun (either the aspirational catch x SAU or TAC x zone) and generates the actual catch per population in each SAU expected in the current year.
makeouthcr	Another JurisdictionHS.R function that generates (or not) an object that is continually updated by the herfun. To avoid very inefficient code this object (at least for Tasmania) is cycled through the iterations.
varyrs	how many years at the end of the conditioning on the fishery, data into zoneDD, to which to add recruitment variation, default = 7, which suits the Tasmanian west coast. Used in prepareprojection
startyr	the index for the first year of the conditioning dynamics to include in the plotted cpue trajectories, to give startyr:indexoflastyear. used in sauplots.
verbose	Should progress comments be printed to console, default=FALSE
ndiagprojs	the number of replicate trajectories to plot in the diagnostics tab to illustrate ndiagprojs trajectories to ensure that such projections appear realistic; default=3
savesauout	should the sau dynamics object be saved as an sauoutD.RData file? 100 replicates of 56 populations for 58 years of conditioning and 30 years of projection = about 5.7 Mb. default=FALSE.
cutcatchN	to reduce the size of the final array of numbers-at-size in the catch one can remove all the empty cells below a given size class. In the default there are $105$ 2mm size classes and setting cutcatchN = $56$ removes size classes $1:55$ 2 - $110$ mm; only from catchN
matureL	is a vector of 2, default = $c(70,200)$ , that define the x-axis bounds on the biology maturity-at-Length plots
wtatL	is a vector of 2, default = $c(80,200)$ , that define the x-axis bounds on the biology weight-at-Length plots
mincount	given size-composition data what minimum sample size will be deemed acceptable for inclusion in the plots and conditioning. default=100.

# Value

a large list containing tottime, projtime, starttime, glb, ctrl, zoneDD, zoneDP, projC, condC, sauout, and outzone

# See Also

makeequilzone, dohistoricC, prepareprojection, doprojections, getprojyrC

```
print("wait on suitable data sets in data")
```

34 fillzoneDef

driftrec

driftrec adjusts the recruitment allowing for larval drift

### **Description**

driftree adjusts the recruitment levels in the stock dynamics allowing for larval drift. The input drift levels can be a constant for each population, or a vector, with a value for each population if their rates are assumed to different. If a constant valuer is input it is extended to a vector of the same length as the number of populations. Each population is assumed to connect only to the populations either side of it. The populations on the end only connect into the simulated populations and so only lose half the recp value for that population, which is sent to the population it is beside in the conditioning. If the zone is not conditioned using yield or productivity from spatial data then productivity will be allocated randomly and the recp (recruitment proportion lost) may as well be set to zero.

### Usage

```
driftrec(recs, recp)
```

### **Arguments**

recs the vector of recruits calculated internally to oneyear

recp the recruitment proporiton lost. Either a constant (eg 0.02 for 2 percent) or a

vector, a value for each population.

#### Value

a vector of revised recruitment levels accounting for larval drift.

### **Examples**

```
recs <- c(243699,49747,1285137,1492742,923282,273427)
newrec <- driftrec(recs,0.04)
newrec0 <- driftrec(recs,0.0)
recs
newrec
newrec0
sum(recs)
sum(newrec0)</pre>
```

fillzoneDef

fillzoneDef characterizes simulated Zone; holds zone properties

# Description

fillzoneDef Characterizes the simulated Zone; holds the zone properties. Lists the date and time the zone was made, its size and basic biological properties: MSY, B0, Average bLML, Average SaM, and more details

findF1 35

#### Usage

```
fillzoneDef(zoneC, zoneD, prod)
```

### **Arguments**

zoneC the constnats for the zone to be characterized.

zoneD the dynamic parts of the rgion being characterized prod the production object from doproduction

#### Value

A list of 10 objects containing the properties of the zone; has the class zoneDefinition - for S3 methods

- Production the productivity of each populations separately
- defpop each population's defining parameters
- Struct the zone's hierarchical structure
- nBlock the number of blocks
- numpop the number of populations
- SummaryPop biological properties of populations
- SummaryZone summary of the zone's properties
- BlockProp summary of the block properties
- Date date/time of fillzoneDef activation; if the function is run soon after a zone's creation this can be a proxy for the zone creation.

# **Examples**

```
# needs summaryPop and summaryZone
txt2 <- 'always use examples rather than example'</pre>
```

findF1

findF1 approximates the F0.1 harvest rate for each population

# Description

findF1 generates estimates of the annual harvest rate that would be equivalent to the F0.1 for each population. It does this by calculating the gradient of the production curve for the selected pop number and selecting the best approximation by searching for the gradient that is closest to 0.1.

### Usage

```
findF1(product)
```

# **Arguments**

product

the 3D array from doproduction

36 findlinenumber

#### Value

either the index within the production array or the vector of gradients

# **Examples**

```
## Not run:
# data files changed, no longer works
data(product) #for pop=1, a 28percent drop from Hmsy leads to a
findF1(product=product) # loss of 3 tonnes, 4 percent of MSY
findmsy(product) # compare the AnnH, Deplet, and RelCE levels.
## End(Not run)
```

findlinenumber

findlinenumber prints out the contents of a text file with line numbers

# **Description**

findlinenumber solves the problem of finding the line number in a given text file when one wants to change a specific line using the function changeline. After inc lines are printed the function stops and waits for any character to be input (a space will suffice) before printing the next inc lines

# Usage

```
findlinenumber(rundir, filename, inc = 20)
```

# **Arguments**

rundir the directory path in which to find the text file.

filename the full name of the text file in quotations.

inc the number of lines on screen

# Value

nothing, but it does print the contents of a text file with the respective line numbers on the console.

```
print("wait on a suitable example")
```

findmsy 37

findmsy

findmsy identifies the closest productivity value to MSY

## **Description**

findmsy for each population in the zone, identifies the closest productivity value to MSY and returns the vector of productivity values for the selected harvest rate. The Catch in each populaiton = MSY and the other variables relate to the value at MSY

## Usage

```
findmsy(product)
```

### **Arguments**

product

array of productivity values output from doproduction

### Value

a matrix of numpop rows containing the approximate MSY related productivity values and the index of the approx MSY

### **Examples**

```
## Not run:
               # takes far too long
  data(zone1)
  glb <- zone1$globals</pre>
  data(constants)
  ans <- makezoneC(zone1,constants)</pre>
  zoneC <- ans$zoneC
  glb <- ans$glb
  ans <- makezone(glb,zoneC)</pre>
  zoneC <- ans$zoneC</pre>
  zoneD <- ans$zoneD
  ans2 <- modzoneC(zoneC,zoneD,glb)</pre>
  zoneC <- ans2$zoneC # zone constants</pre>
  product <- ans2$product</pre>
  approxMSY <- findmsy(product)</pre>
  print(approxMSY)
## End(Not run)
```

fishery\_plots

fishery\_plots generates a set of plots relating to the fishery properties

## **Description**

fishery\_plots produces a series of plots relating to the fishery and its properties. The selectivity curves reflect the LML through time and so this needs to be represented.

38 getaav

#### Usage

```
fishery_plots(rundir, glb, select, histyr, projLML, rge = 50:90)
```

### **Arguments**

rundir the rundir for the particular scenario
glb the object contianing the global constants

select the selectivity matrix from zoneCP, with all years of selectivity

histyr the object from condC containing the historical LML projLML the object containing the LML in the projections

rge the range of the midpts for the size classes so that the view can be limited so that

the lower arm set to zero and the upper set to 1 need not all be viewed and to give greater separation to any set of LML, which will usually be close to each

other.

#### Value

invisibly, the matrix of unique selectivities

### **Examples**

```
print("wait on suitable internal data sets")
```

getaav

getaav calculates annual absolute variation in catch

### **Description**

getaav calculates the annual absolute change in catch for an input vector of catches, which could be across a series of years or even across different spatial units for a single year (an unusual use). The equation used is  $aav = 100 \times sum(|Ct - Ct-1|)/(sum(Ct))$ .

## Usage

```
getaav(invect)
```

## **Arguments**

invect a vector of catches

#### Value

a single scalar value the AAV of the input catches

```
catch <- c(1,2,3,4,5,4,3,2,1)
getaav(catch) # should equal 0.32</pre>
```

getavrec 39

getavrec	getavrec pulls out just the AvRec values for each sau for a scenario

## **Description**

getavrec extracts the AvRec values for each SAU for a given scenario. It does this by reading in the saudata file and using grep to search for the correct line and returning the line as is. Care is required to only take the first record of 'AvRec' so as not to consider the variability in sAvRec.

## Usage

```
getavrec(rundir, datafile, nsau)
```

### **Arguments**

rundir the directory in which one finds the saudata file for the given scenario

datafile the exact name of the saudata file nsau the number of SAU to be found

### Value

a numeric vector of the average recruitment for each SAU

## **Examples**

```
## Not run:
    rundir <- "c:/Users/User/DropBox/A_codeUse/aMSEUse/scenarios/MhLML/M1h5/"
    getavrec(rundir, "saudataM1h5.csv", 8)
## End(Not run)</pre>
```

getConst

getConst extracts 'nb' numbers from a line of text

## **Description**

getConst parses a line of text and extracts 'nb' pieces of text as numbers

# Usage

```
getConst(inline, nb, index = 2)
```

## **Arguments**

inline text line to be parsed, usually obtained using readLines

nb the number of numbers to extract

index which non-empty object to begin extracting from?

40 getLFdata

#### Value

```
a vector of length 'nb'
```

#### **Examples**

```
txtline <- "MaxDL , 32,32,32"
getConst(txtline,nb=3,index=2)</pre>
```

getCPUEssq

getCPUEssq calculates ssq between historical and conditioned cpue

### **Description**

getCPUEssq takes in the historical CPUE and the conditioned zone's cpue and calculates the sumof squared differences between them to assist with the conditioning. This is a greatly simplified version of plotcondCPUE, which plots a graph as well as estimating the ssq. For speed it is better to use getCPUEssq.

## Usage

```
getCPUEssq(histCE, saucpue, glb)
```

## **Arguments**

histCE the matrix of historical cpue by SAU

saucpue the predicted cpue from the conditioning on historical data

glb the globals object

## Value

a vector of length nsau containing the ssq for each SAU

## **Examples**

```
print("wait on suitable internal data sets")
```

getLFdata

getLFdata reads the observed LF-composition data

### **Description**

getLFdata reads the observed LF-composition data, which needs to be stored in a csv file with the following format. Each SAU must use the same sequence of length-classes as rows, and the same sequence of years as columns, even if there are empty rows and columns. In addition, the first column must list the length-class and the second column lists the SAU. See the 'data-file-description.docx', and the example data files to see examples of the required format.

```
getLFdata(rundir, filename)
```

getLFlogL 41

### **Arguments**

rundir the directory containing the LF data files

filename the complete filename for the length-composition data file

#### Value

a list containing the lfs array (lengths x years x sau) and palfs matrix of the count of observations in each year for each sau. This is then contained in the condC object.

#### See Also

```
getzoneLF, getnas
```

### **Examples**

```
print("wait on suitable internal data files")
# rundir=rundir; filename="lf_WZ90-20.csv"
```

getLFlogL

getLFlogL calculates the multi-nomial log-likelihood for sizecomp

## **Description**

getLFlogL when comparing the observed size-composition data from commercial catches with those rpedicted during the historical fishing period one can quantify any differences using the multinomial log- likelihood. This function calculates this, and the value can then be combined with am ssq value from a comparison of the predicted and observed CPUE ready to be minimized if conditioning the OM on a fisheries real data. As always, the operating model is running at a population level while data arrives at an SAU level, so this is only an approximate approach, but it should improve over using purely the results from the sizemod R package.

### Usage

```
getLFlogL(catchN, obsLFs, glb, wtsc, sau)
```

#### **Arguments**

catchN the predicted numbers-at-size in the catch for all years

obsLFs the observed NAS, after cleaning by the preparesizecomp function

glb the globals list

wtsc the weighting given to the size-composition data

sau which sau is the subject of analysis

## Value

the weighted ssq for the filtered observed size-composition data

```
print("wait on data sets")
```

42 getlistvar

getline	getline extracts a vector of numbers from a txt or csv files
_	· · · · · · · · · · · · · · · · · · ·

### **Description**

getline can be used to extract the AvRec and MaxCEpars values for each SAU for a given scenario. It does this by reading in the saudata file and using grep to search for the varname, which gives the first linenumber with that name. One can extract from the saudata file as well as from the control file. The idea is to use values from that line when conditioning the operating model.

## Usage

```
getline(rundir, filen, varname, nobs)
```

#### **Arguments**

rundir the directory in which one finds the saudata or control file for the given scenario

filen the exact name of the file being examined

varname what variable name is required? Take care with spelling

nobs how many numbers to return, usually nSAU

### Value

a numeric vector for each SAU or of length nobs

## **Examples**

```
## Not run:
    rundir <- "c:/Users/User/DropBox/A_codeUse/aMSEUse/scenarios/MhLML/M1h5/"
    getline(rundir, "saudataM1h5.csv", "AvRec", 8)
## End(Not run)</pre>
```

getlistvar

getlistvar extracts a vector or matrix from zoneC

## **Description**

getlistvar extracts a vector or matrix from zoneC. If a vector of scalars, the names relate to populations, if a matrix the columns relate to populations. Only Me, R0, B0, effB0, ExB0, effExB0, MSY, MSYDepl, bLML, scalece, SaM, SAU, popdef, LML, Maturity, WtL, Emergent, and MatWt are currently valid choices. The indexvar = popdef would generate a listing of all the constants. If you only want a single constant from popdefs then use indexvar2.

```
getlistvar(zoneC, indexvar, indexvar2 = "")
```

getLogical 43

## **Arguments**

zoneC the constants components of the simulated zone indexvar the name of the variable to be extracted; character indexvar2 the name of the variable within popdef to extract

## Value

either a vector or matrix of values depending on variable

## **Examples**

```
data(zone)
zoneC <- zone$zoneC
getlistvar(zoneC,"MSY")
getlistvar(zoneC,"B0")
getlistvar(zoneC,"popdef","AvRec")</pre>
```

getLogical

getLogical extracts nb logicals from an input line of text

### **Description**

getLogical obtains nb logicals from an input line

## Usage

```
getLogical(inline, nb)
```

## **Arguments**

inline text line to be parsed, usually obtained using readLines

nb the number of logicals to extract, if nb is longer than the number of logicals

within inline the vector will contain NAs

## Value

```
a vector of length nb
```

```
txtline <- "Depleted, TRUE"
aMSE:::getLogical(txtline,nb=1)
txtline2 <- "calcthis, TRUE, FALSE"
aMSE:::getLogical(txtline2,nb=2)</pre>
```

44 getnas

getmaxCE

getmaxCE identifies peak CPUE during projections for each SAU

## **Description**

getmaxCE is one of the HS performance statistics. Because it uses the targetCE from the TasmanianHS it is obviously Tasmanian specific.

## Usage

```
getmaxCE(ceCI, glb, targetCE)
```

## **Arguments**

ceCI is the sauout\$outCI\$cpue object which contains the CI for the replicate projec-

tions

glb the globals object

targetCE the target CE from the hsargs

### Value

a matrix of nSAU x 5: 5p, 50p, 95p year, and difference between the target and the maximum median value, all for each SAU

## **Examples**

```
## Not run:
    ceCI <- out$sauout$outCI$cpue
    glb <- out$glb
    targCE <- hsargs$maxtarg
    getmaxCE(ceCI=ceCI,glb=glb,targetCE=targCE)
## End(Not run)</pre>
```

getnas

getnas gets the numbers-at-size for all populations and the zone

### **Description**

getnas extracts numbers-at-size for all populations, SAUs, and the zone. There will therefore be numpop + 1 columns of numbers-at-size fr the particular year given.

# Usage

```
getnas(zoneD, yr, glob)
```

## **Arguments**

zoneD the dynamic portion of the zone

yr which yr from the range available should be summarized

glob the global variables object

getprojyrC 45

### Value

a matrix of numpop + 1 columns of numbers-at-size for each population and for the zone

### See Also

```
getLFdata, getzoneLF, prepareprojection, doprojections
```

## **Examples**

```
print("wait on suitable data sets")
```

getprojyrC

getprojyrC returns cumulative catch from selected projection years

## Description

getprojyrC is used to calculate the cumulative catch taken in each SAU and across the zone for selected years of each replicate. Thus, the output would be a replicates x nSAU matrix of total catches across the first 'period' years of the projection under the given harvest strategy.

# Usage

```
getprojyrC(catsau, glb, period = 10)
```

# **Arguments**

catsau the time series of catches summed across populations within each SAU. This

array (reps x SAU x Allyears), is found within the 'out' object from the do\_MSE

function (as in out\$catsau).

glb the globals object. Again found in 'out', as in out\$glb

period how many years to cumulate; default = 10

### Value

a reps x SAU matrix of summed catches

```
print("wait on suitable internal data sets")
```

46 getsauzone

getrecdevcolumn	getrecdevcolumn extracts a column of recdevs from a control file

### **Description**

getrecdevcolumn is used when conditioning the operating model by modifying the recruitment deviates. getrecdevcolumn extracts a column of years selected from a selected controlfile. It does this to use those values as the starting parameter vector for optim.

## Usage

```
getrecdevcolumn(rundir, filename, yearrange, sau, verbose = FALSE)
```

## **Arguments**

rundir the rundir for the scenario

filename the character name of the control file
yearrange the range of years of rec devs to be selected

sau which SAU or block is to be fitted?

verbose should details of the run be made. default=FALSE

#### Value

a list of the vewctor of recdevs, the vector of the yearrange, and a vector of the linenumbers within the control file that are to be changed

## **Examples**

```
print("wait on suitable internal data-sets")
```

getsauzone	getsauzone summarizes zoneD into SAU and zone
------------	---

## Description

getsauzone rowsums the matrices of matureB, exploitB, catch, and recruit from the input zoneD into the separate SAUs and the total into the zone. The harvestR is simply the respective catch divided by the exploitB, and the cpue are the individual population cpue weighted relative to the catch taken from each population in either the SAU or the complete zone.

## Usage

```
getsauzone(zoneD, glb, B0, ExB0)
```

## Arguments

zoneD	the zoneD after ny	rs of dynamics
201100	the ZoneD arter hy	is of dynamics

glb the globals object
B0 is the sau based B0
ExB0 is the sau based ExB0

getsingleNum 47

#### Value

a list of six matrices of nSAU columns of SAU summaries, and one column for the zone

#### **Examples**

```
print("wait on an example")
```

getsingleNum

getsingleNum find a line of text and extracts a single number

#### **Description**

getsingleNum uses grep to find an input line. If the variable being searched for fails then NULL is returned

### Usage

```
getsingleNum(varname, intxt)
```

## **Arguments**

varname the name of the variable to get from intxt

intxt text to be parsed, usually obtained using readLines

#### Value

a single number or, if no value is in the data file a NULL

## **Examples**

```
## Not run:
    txtlines <- c("replicates, 100","Some_other_text, 52")
    getsingleNum("replicates",txtlines)
    getsingleNum("eeplicates",txtlines)
    getsingleNum("other",txtlines)
## End(Not run)</pre>
```

getssqparts

getssqparts calculates the ssq for the cpue and size-comp data

## Description

getssqparts is used during the conditioning of the operating model to improve the fit of the model parameters to the observed SAU scale data on cpue and size-composition. It only uses sum-of squared deviations so an arbitrary weight is given to the size-composition data, and that value should be such that changes in the ssq for the cpue are of the same order of magnitude for the size-composition data.

```
getssqparts(rundir, controlfile, calcpopC, mincount = 100, wtsc = 0.02)
```

48 getStr

#### **Arguments**

rundir the directory in which all the files and outputs are kept

controlfile the name of the controlfile for the MSE run

calcpopC the function from the harvest strategy that is used to calculate the expected catch

by population within each say each year. The annual catch by sau is known but in the MSE that needs to be distributed, in this case without error, across each

population.

mincount the minimum count used in the size-composition data selection default = 100,

this can be a vector of length nsau with a value for each sau if you want to give

a different value to each sau.

wtsc the weighting given to the size-composition data ssq, default= 0.02. This can be

a vector of length nsau if you want to give each sau a different weighting.

#### Value

a vector of the total ssq, the cpue ssq, and the size-comp ssq

### **Examples**

```
print("wait on internal data sets")
```

getStr

getStr obtains a string from an input text line

## Description

getStr obtains a string from an input text line in which any parts are separated by ','. Then, after ignoring the first component, assumed to be a label, it returns the first nb parts.

#### Usage

```
getStr(inline, nb)
```

## **Arguments**

inline input text line with components separated by ','

nb number of parts to return

### Value

```
a vector of character string(s)
```

```
txt <- "runlabel, development_run, label for this particular run"
getStr(txt,1)</pre>
```

getsum 49

getsum

getsum sums each of the main dynamics within zoneD

## **Description**

getsum is only used by getsauzone to sum each of the main dynamic variables within zoneD, and hence is not exported.

## Usage

```
getsum(inmat, index)
```

### **Arguments**

inmat what matrix within zoneD to sum into SAU and zone index a vector containing an index of populations within SAU

#### Value

an nSAU+1 column matrix summarizing each SAU and the zone

### **Examples**

```
print("wait on an example")
```

gettasdevssq

gettasdevssq calculates the SSQ between observed and predicted CPUE

### **Description**

gettasdevssq is used when conditioning the aMSE operating model using ad hoc recruitment deviates after conditioning on AvRec. It focuses on a single SAU at a time. log transformed recdevs are used to avoid the possibility of negative recdevs. NOte this does not test for equilibrium of the initial operating model, it just assumes it reaches equilibrium.

```
gettasdevssq(
  param,
  rundir,
  ctrlfile,
  calcpopC,
  locyrs,
  sau,
  obsLFs = obsLFs,
  wtsc = 0.1,
  verbose = FALSE,
  outplot = FALSE,
  console = FALSE,
  full = FALSE
)
```

50 getunFished

### **Arguments**

param	the log transformed sequence of recdevs for the selected years
rundir	the rundir for the scenario
ctrlfile	the character name of the control file
calcpopC	the function used to distribute catches among populations. This is from the external JurisdictionHS.R file that is 'sourced' in.
locyrs	the rows within the matrix of recruitment deviates corresponding to the selected years.
sau	which sau (in TAS 1 - 8) is to be worked on
obsLFs	the observed length-composition of the catches for the sau, usually from cond C $compdat fs[.,sau]$
wtsc	what weighting should the total Multi-Nomial ikelihood be multiplied by to scale to the range of change in thw CPUE ssq? Default=0.1
verbose	should console reports be made? default = FALSE
outplot	should a plot be generated for output to the webpage. default=FALSE
console	should the plot go to the console. If FALSE and outplot is TRUE then a plot will go to rundir. If TRUE theplot will go to the console. DEfault=FALSE
full	should the ssq components be returned together or separately. default=FALSE

### Value

a scalar value which is the SSQ for the selected years of CPUE

## Examples

```
print("wait on suitable internal data sets")
```

getunFished	getunFished - extracts all data relating to year 1; unfished.	

# Description

getunFished - extracts all data relating to year 1; unfished. Of course, this will only work as long as movezoneYear hasn't been called, which would disrupt the contents of the first year. But if applied to zone rather than zone1 etc, this should be fine. The outputs include: R0, B0, B0crypt, popdef, MSY, MSYDepl, LML, ExB0, and Nt

## Usage

```
getunFished(zoneC, zoneD, glb)
```

## **Arguments**

zoneC	the constant part of the zone
zoneD	the dynamic part of the zone

glb contains the global variables used everywhere

getvar 51

### Value

a list of multiple components relating to the unfished stock this includes a list of each of the equilibrium populations and the zone as the last element of the list

# **Examples**

```
## Not run:
    data(zone) # would normally use zone <- makeequilzone(rundir,"control.csv")
    unfish <- getunFished(zone$zoneC,zone$zoneD,zone$glb)
    str(unfish,max.level=1)
## End(Not run)</pre>
```

getvar

getvar a replacement for sapply to obtain scalar constants

## **Description**

getvar is a replacement for sapply to obtain scalar constants from zoneC and is significantly faster. It should be used to obtain things like B0, R0, MSY, scalece, etc. Still need to use sapply to pull out vectors.

## Usage

```
getvar(zoneC, invar)
```

## **Arguments**

zoneC the constants object for the zone invar a character variable eg. "B0" or "R0"

## Value

a numpop vector of the invar constants from zoneC

## See Also

getlistvar

```
data(zone)
zoneC <- zone$zoneC
getvar(zoneC,"MSY")
getvar(zoneC,"B0")</pre>
```

52 getzoneLF

getvect

getvect extracts invar from the popdef vector in zoneC

## **Description**

getvect extracts a numpop vector of invar from the popdef vector in zoneC. Still need to use sapply to pull out complete vectors such as popdef or maturity etc.

### Usage

```
getvect(zoneC, invar)
```

## **Arguments**

zoneC the constants object for the zone

invar a character variable eg. "steeph", "DLMax"

## Value

a numpop vector of invar from the numpop popdefs in zoneC

## See Also

getlistvar

# Examples

```
data(zone)
zoneC <- zone$zoneC
getvect(zoneC, "steeph")</pre>
```

getzoneLF

getzoneLF extracts all LF data from a zone across pops and years

## Description

getzoneLF extracts all LF data from a zone across all populations for each year. Thus an Nclass x Nyrs X numpop matrix is compressed into a Nclass x Nyrs matrix

## Usage

```
getzoneLF(zoneD, glb)
```

## **Arguments**

zoneD the dynamic part of the zone

glb the global constants

### Value

an Nclass x Nyrs matrix containing LF data across all populations by year

getzoneprod 53

### **Examples**

```
print("An example needs to be written")
```

getzoneprod

getzoneprod zone scale summary of product matrix from doproduction

## **Description**

getzoneprod takes in the product matrix from doproduction and sums the mature and exploitable biomassand the total catches. Then it recalculates, for the zone, the approximate annual harvest rate, the depletion level, and the relative catch rate. The annual harvest rate and relative catch rate are simply the arithmetic mean of the values for all populations, which the depletion is the column of mature biomass divided by the first value = B0

## Usage

```
getzoneprod(product)
```

### **Arguments**

product

The productivity array from doproduction containing the range of imposed harvest rates, and the resulting outputs for each population

### Value

a matrix containing the approximate productivity matrix for the zone

### **Examples**

```
data(zone)
zoneprod <- getzoneprod(zone$product)
head(zoneprod,20)</pre>
```

getzoneprops

getzoneprops extracts the depletion level for a given year

### **Description**

getzoneprops extracts a set of properties for each population and summarizes them for the zone as well. These properties include effB0, matureB, legalmatB, propprot, MSY, effexB0, exploitB, SpBDepl, ExBDepl, legalDepl, MSYDepl, LML, and bLML. See the model documentation for the meaning of each of these.

```
getzoneprops(zoneC, zoneD, glb, year = 1)
```

54 historical plots

### **Arguments**

zoneC the constants for the zone being simulated zoneD the dynamic part of the zone being simulated

glb the global constants

year which years information is wanted, default=1

## Value

a matrix of population properties with a column of totals

## **Examples**

```
data(zone)
str(zone,max.level=1)
# round(getzoneprops(zone$zoneC,zone$zoneD,zone$glb),4)
# zoneC=zoneC; zoneD=zoneD;glb=glb;year=1
```

historicalplots

historical plots a wrapper function to call plots of fishery plots

## **Description**

historical plots is a wrapper used by do\_MSE and do\_condition, to plot out details of the historical fishery data used in the conditioning. It currently plots out the historical catches and CPUE.

## Usage

```
historicalplots(rundir, condC, glb, commonscale = FALSE, proportion = FALSE)
```

## **Arguments**

rundir the directory where all results are stored

condC the R object containing the historical fishery data generated by the readctrlfile

function

glb the global variables object

commonscale should the SAU plots share a common y-axis scale, default = FALSE

proportion should the plot be of proportion relative to the maximum catch through each

time-series or as raw tonnes x year, default=FALSE

## Value

nothing but it does add plot files to the rundir

```
print("wait on suitable data sets")
```

imperr 55

imperr	imperr calculates population catches from sau catches with error

## **Description**

imperr converts aspirational sau catches into population level catches while introducing management implementation error. Here this error is implemented as Log-Normal errors on diver intuitions concerning the relative abundance in each population. The error is imposed separately on the populations in each SAU.

## Usage

```
imperr(catchsau, exb, sauindex, sigmab = 1e-08)
```

## **Arguments**

catchsau	the predicted or aspirational catch per SAU from the Harvest control rule
exb	the exploitable biomass at the end of the previous year. In the first year of the projections this would be the last year of the conditioning.
sauindex	the SAU index for each population
sigmab	the Log-Normal standard deviation of implementation error. The default value = 1e-08, which effectively means no errors.

## Value

a vector of population catches for the year to be imposed after the estimation of exploitable biomass

## **Examples**

# Description

lf10 is a data.frame of length-composition of commercial catch from block 10 on the west coast of Tasmania in a longdat format as derived from the commlf function makelongdat. This makes it useful for illustrating the use of makewidedat.

#### **Format**

```
A data.frame of abalone size-composition data

year the year of sampling
sau the block or SAU

length the measured size in mm, with 2mm size classes
counts the counts at each length class
propounts the proportion of each length class of the total
```

56 logistic

### **Subjects**

- · selectivity
- · size-based stock assessment model fitting
- · size-distributions of commercial catches

#### **Source**

Thanks to the Institute of Marine and Antarctic Science, which is part of the University of Tasmania, and especially to Dr Craig Mundy, leader of the Abalone Group, for permission to use this data.

## **Examples**

```
data(1f10)
head(1f10,20)
mids <- seq(138,210,2)
answer <- makewidedat(1f10,mids)
answer[1:20,]</pre>
```

logistic

logistic a Logistic selectivity function

## Description

logistic a logistic selectivity function. This uses the logistic function  $1/(1+\exp(-\log(19.0))*(\text{lens-inL50})/(\text{delta}))$ ), where delta = inL95 - inL50. This explicitly defines the SM50 but uses delta (which is SM95-SM50) as the second. This ensures that when adding variation to parameters, to vary between populations, when SM95 and SM50 are close together it is not possible for SM50 to become larger than SM95. Be careful using the knifeedge option. Strictly knifeedge selectivity would entail the selectivity values being zero up to the knife-edge and then being 1.0. This is not what happens here. Instead the knifeedge option literally sets all values to zero at and below the value of knifeedge but leaves any curve above that value as it is. Hence this is not strict knife-edge selectivity. However, it does provide a selectivity curve that reflects the selectivity of a diver led fleet workingon abalone.

### Usage

```
logistic(inL50, delta, lens, knifeedge = 0)
```

## **Arguments**

inL50	is the length at 50 percent selection
delta	is the difference between the 95 percent selection and the 50 percent selection
lens	a vector of lengths for which the logistic maturity value will be calculated
knifeedge	defaults to 0. If knifeedge is set to a particular length then the selectivity less than the value of knifeedge is set to zero

## Value

A vector of length(lens) containing the predicted selectivity at length values

makeabpop 57

### **Examples**

```
## Not run:
inL50 <- 100.0
delta <- 8.0
lens <- seq(2,210,2)
select <- logistic(inL50,delta,lens)
select <- logistic(inL50,delta,lens,knifeedge=105)
## End(Not run)</pre>
```

makeabpop

makeabpop generates full population structure in an unfished state

## Description

makeabpop generates the full population structure in an unfished state. The structure is pre-determined: Me R0 B0 effB0 ExB0, effExB0 MSY MSYDepl bLML scalece SaM popdef (vector of constants) LML G Maturity WtL Emergent Select SelWt MatWt SAUname. See the AbMSE documentation to see the full definition of a zone, made up of a zoneC and a zoneD. Notice the presence of effB0 and effExB0, these relate to the influence of larval dispersal on each populations productivity. The effective B0 relates to the unfished mature biomass after larval dispersal occurs and the population achieves equilibrium.

## Usage

```
makeabpop(popparam, midpts, projLML)
```

### **Arguments**

popparam the vector of biological parameters values that define the specific properties of

this population. Obtained from popdefs, which is produced by definepops

midpts the center values of the size classes dervied from the zone data file

projLML the LML expected in the projection years 2 - Nyrs; a vector obtained from the

zonedatafile

### Value

a list of numpop lists of 19 objects as detailed above.

```
print("See the code for makezoneC to see usage of makeabpop")
```

58 makemove

makeequilzone

makeequilzone high level function generates equilibrium zone

### **Description**

makeequilzone is a high level function that merely hides the details of generating the original unfished zone after reading in the data files, estimates the productivity, and sets up the results directory, rundir, ready to receive files.

### Usage

```
makeequilzone(
  rundir,
  ctrlfile = "control.csv",
  doproduct = TRUE,
  verbose = TRUE
)
```

## Arguments

rundir the directory containing the control csv files. It can/will also act to store results

in a manner that will allow them to be displayed using makehtml.

ctrlfile the main file that controls the particular run. It contains the name of the data file

that is used to biologically condition the numpop populations

doproduct boolean, should the productivity calculations be made during the conditioning.

Set to FALSE conditionOM

verbose Should progress comments be printed to console, default=TRUE

## Value

```
a list of zoneC, zoneD, glb, constants, saudat,product, ctrl, and zone1
```

#### **Examples**

```
print("wait on datafiles")
```

makemove

makemove produces the movement matrix

## **Description**

makemove produces the movement matrix, which assumes that only adjacent populations contribute to each other. Thus, a central population would lose half its larvae to one side and half to the other side, but would receive half of their larval dispersal each. As larval dispersal is modelled as a proportion those populations that produce more larvae will lose more in absolute terms. Those populations at the edges of the zone only lose half the larval dispersal into the zone and it is assumed that what they lose out of the zone will be matched by what will come into the zone.

makeoutput 59

### Usage

```
makemove(npop, recs, ld, sigmove = 0)
```

## **Arguments**

npop the number of populations

recs the original unfished recruitment levels estimated for each population

1d the larval dispersal rate for all populations

sigmove currently no variation is assumed to occur. This is here in case it needs to get

implemented, but as there is little or no data on actual larval dispersal levels

currently this is set to zero and not used.

## Value

a npop x npop matrix describing movement among populations

## **Examples**

```
recruits <- c(636146,263878,819189,1112513,285025,671573.9) makemove(npop=6,recs=recruits,ld=0.04)
```

makeoutput

makeoutput take the output from do\_MSE and generates the HTML files

# Description

makeoutput simplifies taking the output from do\_MSE and producing the HTML files used to display all the results in rundir. Note that 'runnotes' is now a vector of character strings made up using paste0.

```
makeoutput(
  out,
  rundir,
  postdir,
  controlfile,
  hsfile = NULL,
  doproject = TRUE,
  openfile = TRUE,
  verbose = FALSE
)
```

60 makewidedat

### **Arguments**

out the output from do\_MSE

rundir the full path to the directory holding the all scenario files

postdir the name of the directory holding the results, also used to name the internal

webpage

controlfile the controlfile used to run the MSE

hsfile the name of the harvest strategy file, default=NULL doproject have the projections been run? default = TRUE.

openfile should the website be opened automatically? default=TRUE

verbose should details of producing the HTML files be written to the console? de-

fault=FALSE

#### Value

nothing but it does generate a set of HTML files in rundir. If verbose=TRUE it also writes text to the console

## **Examples**

print("wait on internal data-sets")

makewidedat

makewidedat converts long data to a wide data format

## **Description**

makewidedat takes the output of the commlf function makelongdat and converts the columns of year x length x count into a matrix of counts with axes of lengths x years, ready for comparisons with predicted length composition of catches from the operating model.

### Usage

```
makewidedat(inlong, mids, counts = FALSE)
```

### **Arguments**

inlong the long form data.frame derived for commlf's makelongdat containing at least

'year', 'length', and 'propcounts'

mids the range of lengths to be found within the input data file. These are matched

with the size distribution for each year of data and the appropriate matrix cells

filled with the propounts.

counts default = FALSE, which means that the wide format will contain the proportion

of counts for each year. If TRUE, then the actual counts by size-class will be

output by year

## Value

a matrix of proportional counts for each size-class x year

makezone 61

### **Examples**

```
data(1f10)
mids <- seq(138,210,2)
answer <- makewidedat(1f10,mids)
answer[1:20,]
answerC <- makewidedat(1f10,mids,counts=TRUE)
answerC[1:20,]</pre>
```

makezone

makezone generates the dynamic parts of the simulated zone

## **Description**

makezone generates the dynamics components of the simulated zone and completes the constant components of the simulated zone. The term 'zone' refers to the upper level of geographical detail that is used. Thus, for example, in Tasmania we might simulate a number of statistical blocks with multiple populations. In combination, we refer to the total as a zone. The matrices are Nyrs x numpop and the two arrays are  $N \times N$  x Nyrs x numpop.

## Usage

```
makezone(glob, zoneC)
```

## Arguments

glob the global constants defined for the current simulation. These include numpop,

nblock, midptsd, Nclass, and Nyrs

zoneC the zoneC object from makezoneC

## Value

a list of the dynamics and the constant components of the simulation

```
## Not run:
    data(zone1)
    glb <- zone1$globals
    data(constants)
    ans <- makezoneC(zone1,constants)
    zoneC <- ans$zoneC
    glb <- ans$glb
    ans2 <- makezone(glb,zoneC)
    str(ans2,max.level=2)
## End(Not run)</pre>
```

62 makezoneDP

makezoneC

makezoneC makes the constant part of the simulation

### **Description**

makezoneC makes the constant part of the simulated zone. Once defined this does not change throughout the simulation. Once made it still requires makezone to be run to fill in the B0, ExBo, MSY, MSYDepl, and the scalece values, and to produce zoneD, the dynamic part of the new zone

## Usage

```
makezoneC(zone, const)
```

### **Arguments**

zone the object derived from the readzonefile function const the object derived from the readdatafile function

### Value

a list containing the constant part of the simulated zone

## **Examples**

```
## Not run:
data(zone1)
data(constants)
ans <- makezoneC(zone=zone1,const=constants)
zoneC <- ans$zoneC
popdefs <- ans$popdefs
str(zoneC,max.level=1)
str(zoneC[[1]]) # not complete at this stage
print(popdefs)
## End(Not run)</pre>
```

makezoneDP

makezoneDP generates the container for the projection dynamics

## **Description**

makezoneDP generates an object designed to hold the outputs from each replicate within a set of projections. This is identical to zoneD except it contains a repeat for each iteration and now includes a cesau, which is the population-catch-weighted cpue for each SAU.

```
makezoneDP(allyr, iter, glb)
```

maturity 63

## **Arguments**

allv	r number of	vears of histo	orical catches p	plus pro	iection v	ears
------	-------------	----------------	------------------	----------	-----------	------

iter the number of replicates

glb the object containing the global variables

## Value

a large list containing an object ready for the projection dynamics

## **Examples**

```
print("Could add variation to the harvest rates so that when ")
print("prepareprojection was run the range of initial H values would increase ")
```

maturity

maturity Logistic maturity curve

## **Description**

maturity this uses the logistic function:  $\exp(a+bL)/(1+\exp(a+bL))$ , which has the property that the SM50 = -a/b and the interquartile distance is 2.Ln(3)/b.

### Usage

```
maturity(ina, inb, lens)
```

### **Arguments**

ina is the intercept of the exponential function inb is the gradient of the exponential function

lens a vector of lengths for which the logistic maturity value will be calculated

# Value

A vector of length(lens) containing the predicted maturity at length values

```
## Not run:
a <- -14.383
b <- 0.146017
lens <- seq(2,210,2)
Maturity <- maturity(a,b,lens)
## End(Not run)</pre>
```

64 modprojC

midg

midg is an abalone tagging data-set from the Actaeons

### **Description**

midg is a tagging data-set for blacklip abalone (*Haliotis rubra*) from the middle ground in the Actaeons in Tasmania's Block 13. All individuals were recaptured in 2003, the site number was 478, at Latitude -43.54 longitude 146.99, and there are 347 observations. All Dt = 1 year.

#### **Format**

A data frame of abalone tagging data

RecapL the length at recapture

Lt the length at tagging

Dt The time interval between tagging and recapture, in this instance they are all listed as 1 year

**DL** the growth increment in mm

### **Subjects**

- · growth curves
- inverse logistic, von Bertalanffy, Gompertz
- · Static model fitting

### **Source**

Thanks to the Institute of Marine and Antarctic Science, which is part of the University of Tasmania, and especially to Dr Craig Mundy, leader of the Abalone Group, for permission to use this data collected in 2003.

### **Examples**

```
data(midg)
head(midg,20)
oldpar <- par(no.readonly=TRUE)
plot(midg$Lt,midg$DL,type="p",pch=16,cex=1.0,xlim=c(5,180))
abline(h=0,col=1)
par(oldpar)</pre>
```

modprojC

modprojC produces three objects used to condition the zone

## Description

modprojC produces an object used to condition the zone when projecting it following any conditioning. Once the initial conditions for the projection have been attained through an initial depletion of an unfished equilibrium, or more particularly by conditioning on historical data then the projC object will be used to condition the projections. In essence, this puts the selectivity and selectivity x weight-at-size into projC. It can handle changes in LML through the years of projection.

modzoneC 65

#### Usage

```
modprojC(zoneC, glob, projC)
```

## **Arguments**

zoneC the constant part of the zone
glob the globals for the simulation

projC the project definition object from zone1 from readctrlfile

### Value

an updated projC with projSel and projSelWt completed

## **Examples**

```
print("wait on new example data")
```

modzoneC

modzoneC runs the zone 3 x hyrs to equilibrium

## **Description**

modzoneC runs the doproduction function so it can then appropriately fill in each population's MSY and MSYDepl values. It outputs both the finalized zoneC and the prodution array. Hence we need to push copies of both zoneC and zoneD into modzoneC

### Usage

```
modzoneC(zoneC, zoneD, glob, lowlim = 0, uplim = 0.4, inc = 0.005)
```

## **Arguments**

zoneC	the constant components of the simulated zone
zoneD	the dynamic components of the simulated zone
glob	the general global variables

lowlim the lower limit of harvest rate applied, default=0.0
uplim the upper limit of harvest rate applied, default=0.4
inc the harvest rate increment at each step, default=0.005

## Value

a list containing the updated zoneC and product

66 modzoneCSel

## **Examples**

```
## Not run: # the doproduction part takes too long to run
  data(zone1)
  glb <- zone1$globals
  data(constants)
  ans <- makezoneC(zone1,constants)
  zoneC <- ans$zoneC
  ans <- makezone(glb,zoneC)
  zoneC <- ans$zoneC
  zoneD <- ans$zoneC
  ans2 coneD <- ans$zoneD
  ans2 <- modzoneC(zoneC,zoneD,glb)
  str(ans2,max.level=2)

## End(Not run) # zoneC=zoneC; zoneD=zoneD; glob=glb; lowlim=0.0;uplim=0.4;inc=0.01</pre>
```

modzoneCSel

modzoneCSel changes the selectivity characteristics in zoneC

# Description

modzoneCSel changes the selectivity characteristics in zoneC which is necessary when making a projection under a different LML. This function assumes a constant LML during any projections.

# Usage

```
modzoneCSel(zoneC, sel, selwt, glb)
```

## Arguments

zoneC	the constant zone object from setupzone
sel	the new selectivity as a matrix of selectivity by population
selwt	new selectivity x WtL as a matrix of SelWt x Population
glb	the globals object

## Value

the zoneC object modified ready for projection

```
print("wait on suitable data")
```

numbersatsize 67

numbersatsize

numbersatsize plots details of the numbers-at-size

## **Description**

numbersatsize plots up the initial unfished numbers- at-size distribution, omitting the first four size classes to avoid the recruitment numbers dominating the plot.

### Usage

```
numbersatsize(rundir, glb, zoneD, ssc = 5)
```

## Arguments

rundir the results directory, if set to "" then plot is sent to the console instead

glb the globals list

zoneD the dynamic part of the zone, zoneD

ssc index for starting size class. thus 1 = 2, 2 = 4, 5 = 10, etc. default = 5 for it plots

size classes from 10mm up

#### Value

nothing but it does add one plot to the results directory

## **Examples**

```
print("this will be quite long when I get to it")
```

onesau

onesau plots the dynamics for a single SAU

# Description

onesau plots the details of matureB, exploitB, catch, acatch, harvestR, cpue, recruit, deplsB, depleB for a single SAU on one plot

```
onesau(
  prerep,
  postrep,
  glb,
  startyr,
  picksau,
  addCI = FALSE,
  CIprobs = c(0.05, 0.5, 0.95),
  histCE = NULL
)
```

68 oneyear

## **Arguments**

prerep	the zoneDsau object that represents the SAU data
postrep	the zonePsau object that represents the SAU data
glb	the global constants object
startyr	the year from which to begin the conditioned dynamics
picksau	which sau should be plotted
addCI	should the 90 percent CI be added, default=FALSE
CIprobs	what CI should be fitted, default=c(0.05,0.5,0.95)
histCE	should the historical CPUE be added to the cpue plot

### Value

a list of the CI for each variable

## **Examples**

```
print("Wait on suitable data sets")
```

oneyear	oneyear one year's harvest	dynamics for one abpop

## Description

oneyear do one year's dynamics for one input population. Used to step through populations of a zone. Its dynamics are such that it first undergoes growth, then half natural mortality. This allows an estimate of exploitable biomass before fishing occurs. The remaining dynamics involve the removal of the catch, the application of the last half of natural mortality and the addition of recruits. Which allows the exploitable biomass to be estimated after fishing. The recruitment occurs in oneyearD so that larval dispersal can be accounted for. Thus, oneyear requires oneyearD.

## Usage

```
oneyear(MatWt, SelWt, selyr, Me, G, qest, WtL, inNt, inH, lambda, scalece)
```

# Arguments

MatWt	maturity x Weight-at-size from zoneCC for the population and year
SelWt	selectivity x Weight-at-size from zoneCC for the population and year
selyr	selectivity from zoneC for the population and year
Me	natural morality from zoneC for the population
G	growth transtion matrix from zoneC for the population
qest	the catchability from zoneCC for the population
WtL	the weight-at-alength from zoneC for the population
inNt	the numbers at size for the year previous to the year of dynamics. These are projected into the active year.
inH	a literal annual harvest rate as a proportion to be removed as catch during the year, a scalar
lambda	the non-linearity parameter. if = $1.0$ then cpue linearly related to expB, if < $1$ then hyper-stability of cpue occurs.
scalece	the xoneC'pop'\$scalece used to scale al cpue to the sau nominla scale

oneyearcat 69

#### Value

a list containing ExploitB, MatureB, Catch, Harvest, Nt, ce, CatchN, and midyexpB used to update the given pop in yr + 1

### **Examples**

```
print("need to wait on built in data sets")
#oneyear <- function(MatWt,SelWt,selyr,Me,G,gest,WtL,inNt,inH,lambda,scalece)</pre>
```

oneyearcat

oneyearcat one year's catch dynamics for one abpop

#### **Description**

oneyear does one year's dynamics for one input population. Where the fishing is based on a given catch per population, which would be determined by any harvest control rule. It is used to step through populations of a zone. Its dynamics are such that each population first undergoes growth, then half natural mortality. This allows an estimate of exploitable biomass before fishing occurs. The remaining dynamics involve the removal of the catch, after estimating the harvest rate from catch/exploitb, the application of the last half of natural mortality and the addition of recruits. This allows the exploitable biomass to be estimated after fishing. The cpue uses the average of the pre-fishing and post-fishing exploitB to smooth over any changes brought about by fishing. The recruitment occurs in oneyearC so that larval dispersal can be accounted for oneyearcat is not used independently of oneyearC.

#### Usage

```
oneyearcat(
  MatWt,
  SelWt,
  selyr,
  Me,
  G,
  scalece,
  WtL,
  inNt,
  incat,
  sigce,
  lambda,
  qest
)
```

#### **Arguments**

MatWt	maturity x Weight-at-size from zoneCC for the population and year
SelWt	selectivity x Weight-at-size from zoneCC for the population and year
selyr	selectivity from zoneCC for the population and year
Me	natural morality from zoneCC for the population
G	growth transtion matrix from zoneCC for the population

70 oneyearD

scal	ece	the xoneC'pop'\$scalece used to scale al cpue to the sau nominla scale
WtL		the weight-at-alength from zoneCC for the population
inNt		the numbers at size for the year previous to the year of dynamics. These are projected into the active year.
inca	t	the literal annual catch from the population. Derived from the harvest control rule for each SAU, then allocated to each population with respect to its relative catching performance.
sigc	e	the process error variation associated with the estimates of cpue from the model. Found in the ctrl object as with sigCE. It is implemented as Log-Normal error on the exploitable biomass value from the model
lamb	da	the hyper-stability term from zoneC
qest		the estimated catchability from sizemod from zoneC

## Value

a list containing a vector of ExploitB, MatureB, Catch, and ce, and a matrix NaL containing Nt and CatchN used to update a pop in yr + 1

### See Also

dohistoricC, oneyearcat, oneyearrec

### **Examples**

```
print("need to wait on built in data sets")
```

oneyearD oneyearD conducts one year's dynamics on zoneD in the MSE

## **Description**

oneyearD conducts one year's dynamics on zoneD in the MSE returning the revised zoneD, which will have had a single year of activity included in each of its components. This uses zoneC but always within the environment of another function in which zoneC (as zoneC) can be found. Used in runthreeH, (and hence dodepletion and doproduction) and in testequil.

## Usage

```
oneyearD(zoneC, zoneD, inHt, year, sigmar, Ncl, npop, movem)
```

## **Arguments**

zoneC	the constant portion of the zone with a list of properties for each population
zoneD	the dynamics portion of the zone, with matrices and arrays for the dynamic variables of the dynamics of the operating model
inHt	a vector of harvest rates taken in the year from each population
year	the year of the dynamics, would start in year 2 as year 1 is the year of initiation.
sigmar	the variation in recruitment dynamics, set to 1e-08 when searching for equilibria.
Ncl	the number of size classes used to describe size
npop	the number of populations, the global numpop
movem	the larval dispersal movement matrix

oneyearrec 71

### Value

a list containing a revised dynamics list

### **Examples**

```
print("wait on revised data")
```

oneyearrec

oneyearrec calculates the Beverton-Holt recruitment

## Description

oneyearrec calculates the Beverton-Holt recruitment for the input populations in a single year; parameterized with steepness, R0, and B0. To drop variation to insignificant levels set sigmar-1-08. To allow for inclusion of recruitment deviates, which can be required to explain high catches when stock biomass would otherwise be low, we have introduced the devR argument. As the deviates are all multiplicative and range in value > 0, then the default = -1 so this will be ignored unless a deviate is input.

## Usage

```
oneyearrec(steep, R0, B0, Bsp, sigR, devR = −1)
```

## **Arguments**

steep	the steepness for the population; scalar
RØ	the unfished recruitment levels for the population; scalar
B0	the unfished spawning biomass; scalar
Bsp	the current spawning biomass; scalar
sigR	standard deviation of the recruitment residuals; scalar. set this to 1e-08 to avoid recruitment variability
devR	if recruitment deviates are available then they should be input here. If negative values then a random epsilon is used, otherwise epsilon is given the value of devR. default=-1, so by default fixed recruitment deviates are off.

## Value

an absolute number of recruits from a given spawning biomass

## See Also

oneyearsauC, oneyearcat, dohistoricC

72 oneyearsauC

## **Examples**

```
## Not run:
steep <- 0.707
R0 <- 319971
B0 <- 313
Bsp <- 147
insigmar <- 0.3
oneyearrec(steep,R0,B0,Bsp,insigmar)
## End(Not run)
 \begin{tabular}{ll} \# & steep=steep; R0=r0; B0=b0; Bsp=dyn["matureb",]; sigR=sigmar; devR=rdev \\ \end{tabular}
```

oneyearsauC

oneyearsauC conducts one year's dynamics using catch not harvest

## **Description**

oneyearsauC conducts one year's dynamics in the simulation using historical SAU catches rather than harvest rates. The harvest rates are estimated after first estimating the exploitable biomass. returning the revised zoneD, which will have had a single year of activity included in each of its components. uses the function imperr to introduce implementation error to the actual catches.

# Usage

```
oneyearsauC(
  zoneCC,
  inN,
  popC,
  year,
  Ncl,
  sauindex,
  movem,
  sigmar = 1e-08,
  sigce = 1e-08,
  r0,
  b0,
  exb0,
  rdev = -1
```

## Arguments

zoneCC	the constant portion of the zone with a list of properties for each population ##param exb the vector of exploitable biomass for each population for the ## previous year
inN	the numbers-at-length for each population for the previous year
рорС	a vector of actual catches to be taken in the year from each SAU
year	the year of the dynamics the exb and inN would be from year-1
Ncl	the number of size classes used to describe size, global Nclass
sauindex	the sau index for each population from glb

oneyrgrowth 73

movem	the larval dispersal movement matrix, global move
sigmar	the variation in recruitment dynamics, set to $1\text{e-}08$ when searching for an equilibrium.
sigce	the process error on the relationship between cpue and exploitable biomass, set to 1e-08 when searching for an equilibrium.
r0	the unfished R0 level used by oneyearrec
b0	the unfished mature biomass B0, used in recruitment and depletion
exb0	the unfished exploitable biomass used in depletion
rdev	the recruitment deviates for each SAU for the given year

#### Value

a list containing a revised dynamics list

#### See Also

dohistoricC, oneyearcat, oneyearrec

# **Examples**

```
print("Wait on new data")
```

oneyrgrowth	oneyrgrowth one years' growth for a population and initial size

# Description

oneyrgrowth one years' growth for a given population and initial size. Used to determine the size after two year's growth as a measure of the expected bLML - biological Legal Minimum Length. A reflection of the two year rule in Tasmania. To get this the function should obviously be run twice.

# Usage

```
oneyrgrowth(inpop, startsize = 2)
```

# Arguments

inpop the abpop to be grown forward

startsize default = 2, but often set to the size at 50 percent maturity or the vector of midpts

to determine the growth of Nt

# Value

the expected mean size after one year's growth

74 onezoneplot

# **Examples**

onezoneplot

onezoneplot plots out one variable from the zone

# Description

once the projected zone has been summarized by poptozone one can add plots to the results using plotZone. This in turn uses onezoneplot to plot each variable in turn.

# Usage

```
onezoneplot(invar, rundir, glb, CIprobs, varname, startyr, addfile = TRUE)
```

# **Arguments**

invar	which zone variable to plot
rundir	the run directory for the results
glb	the global constants object
CIprobs	the quantiles to use for the CI.
varname	the name of the variable for use in labels
startyr	in what year number should the plot begin.
addfile	should a png file be added to the results, default = TRUE

# Value

it generates a png file of the plot if addfile remains TRUE

```
print("wait on suitable data")
```

optimizeAvRec 75

optimizeAvRec	optimizeAvRec i	improves OM fit to	CPUE by	adjustnig AvRec
---------------	-----------------	--------------------	---------	-----------------

# Description

optimizeAvRec is used to improve the fit to the time-series of CPUE by adjusting the SAU initial value for AvRec. If one assesses an SAU using the sizemod package, that summarizes across a whole SAU. After population allocation within aMSE, the productivity of an SAU may well have been disturbed away (up or down) from the assessed optimum. Hence, it becomes necessary, still, to optimize both the AvRec and the recdevs during the conditioning.

# Usage

```
optimizeAvRec(
  rundir,
  controlfile,
  datafile,
  calcpopC,
  snames,
  lowmult = 0.7,
  highmult = 1.3,
  linenumber = 29,
  verbose = TRUE
)
```

# Arguments

rundir	the rundir for the scenario
controlfile	the name of the control file
datafile	the name of the datafile from the controlfile
calcpopC	the funciton, from the HS file that allocates catches across each of the populations within each $SAU$
snames	the names given to each SAU
lowmult	the multiplier to place a lower bound when search for the optimum AvRec for each SAU. default = $0.7$ , ie param AvRec * $0.7$
highmult	the multiplier to place an upper bound when search for the optimum AvRec for each SAU. default = $1.3$ , ie param AvRec * $1.3$
linenumber	the linenumber inside the datafile that is to be changed to the optimum AvRec values. Default = $29$ , whic suits Tasmania.
verbose	should updates on progress be sent to the console. default=TRUE

#### Value

a vector of the final optimized AvRec values. This also modifies the datafile - SO BE CAREFUL.

```
print("wait on suitable data files")
```

76 optimizerecdevs

optimizerecdevs optimizerecdevs improves the fit to the recdevs for a given sau

#### **Description**

optimizerecdevs an important part of optimizing the match between the dynamics observed in the fishery and those of the operating model is to adjust the annual recruitment deviates so that they improve the fit to the observed CPUE and observed numbers-at-size. optimizerecdevs attempts to do that.

# Usage

```
optimizerecdevs(
  rundir,
  sau,
  controlfile,
  calcpopC,
  wtsc = 0.02,
  maxiter = 100,
  yearrange = 1980:2016,
  verbose = FALSE,
  mincount = 100,
  plottoconsole = FALSE,
  optimmethod = "Nelder-Mead")
```

# **Arguments**

rundir the rundir for the scenario
sau which sau to apply this to
controlfile the name of the control file

calcpopC the funciton, from the HS file that allocates catches across each of the popula-

tions within each SAU

wtsc what weight to give to the size-composition data, default=0.1

maxiter The maximum number of iterations to run before stopping. Default = 100, one

needs at least 400 to see a real difference

yearrange which recdev years should be 'conditioned'. Default= 1980:2016

verbose text output to console, default=FALSE

mincount the minimum number of iterations in the solver, default=100

plottoconsole should the final plot be sent to the console = TRUE of the rundir = FALSE.

default=FALSE

optimmethod which optim method to use? default='Nelder-Mead'

#### Value

a scalar value which is the total SSQ for the selected years for the CPUE and sizecomps. It also alters the recdevs in the controlfile!

plotCNt 77

#### **Examples**

```
print("wait on data sets")
```

plotCNt

plotCNt plots the historical conditioning Nt or catchN for all years

#### **Description**

plotCNt the historical conditioning on the fisheries data leads to a three dimensional array of numbers-at-size x years x numpop. We use 'prepareDDNt' to convert this to a 3D array by SAU. PlotCNt then plots, for each SAU, numbers-at-size for all years of conditioning with the first year emphasized in black and the last year in red.

#### Usage

```
plotCNt(Nt, glb, vline = NULL, start = 3)
```

# Arguments

Nt the 3D array of numbers-at-size x years x SAU from prepareDDNt

glb the global constants object

vline add a vertical dashed line, perhaps at the LML, default=NULL

start from what size class should the maximum Y value be measured. The default =

3 (= 6 mm)

#### Value

nothing but it does plot a graph

# **Examples**

```
print("wait on suitable built in data-sets")
```

plotcondCPUE

plotcondCPUE plots the historical cpue against conditioned cpue

# **Description**

plotcondCPUE generates a plot of the historical cpue and compares it with the predicted cpue from the conditioning. The predicted is black and the observed is green. This is primarily there are an aid to conditioning the operating model. It also calculates the simple sum of squared differences between the observed and predicted, again to aid in the conditioning.

```
plotcondCPUE(histCE, saucpue, glb, rundir, filen = "")
```

78 plotconditioning

#### **Arguments**

histCE the matrix of historical cpue by SAU

saucpue the predicted cpue from the conditioning on historical data

glb the globals object

rundir the rundir for the given scenario

filen the file name of the plot if it is to be saved

#### Value

a vector of length nsau containing the ssq for each SAU

## **Examples**

```
print("wait on suitable internal data sets")
```

plotconditioning

plotconditioning illustrate the zones dynamics after conditioning

#### **Description**

plotconditioning converts the conditioned dynamic zone object from population scale to sau scale and then plots the various components to depict their state after conditioning. This is designed to place such plots into the rundir to assist in the conditioning process.

# Usage

```
plotconditioning(zoneDD, glb, zoneC, histCE, rundir, recdevs)
```

#### **Arguments**

zoneDD the conditioned dynamic zone object

glb the global constants object
zoneC the constant zone object
histCE the historical CPUE matrix
rundir the rundir for the given scenario

recdevs the recruitment deviates from the control file, condC

# Value

the sau scale zone dynamics, invisibly

```
print("wait on suitable data sets")
```

plothistcatch 79

plothistcatch generates a plot of the historical catches by SAU
plothistcatch generates a plot of the historical catches by SAU

# Description

plothistcatch generates a plot of histircal catches by SAU and by Zone. It can plot can tonnes x year or proportion. All SAU plots can have independent or common y-axis scales.

# Usage

```
plothistcatch(
   yrs,
   histC,
   nsau,
   saunames,
   commonscale = FALSE,
   proportion = FALSE,
   filen = ""
)
```

# **Arguments**

a vector of the years of catch history yrs histC the matrix of catches by years nsau the number of SAU saunames the SAU names as a vector should the SAU plots share a common y-axis scale, default = FALSE commonscale should the plot be of proportion relative to the maximum catch through each proportion time-series or as raw tonnes x year, default=FALSE filen the name of the file used to save the plot, default="", which sends the plot to a separate window

#### Value

nothing but it does generate a plot either to a window or a file

```
print("wait on suitable data sets")
```

80 plothsstats

plothistce generates a plot of the historical cpue by SAU	
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# **Description**

plothistce generates a plot of historical cpue by SAU. It can plot as cpue or proportion of maximum cpue.

# Usage

```
plothistce(rundir, condC, glb, proportion = FALSE, filen = "")
```

# **Arguments**

rundir the directry where all results are stored

condC the R object contiaining the historical fishery data generated by the readctrlfile

function

glb the globol variables object

proportion should the plot be of proportion relative to the maximum catch through each

time-series or as raw tonnes x year, default=FALSE

filen the name of the file used to save the plot, default="", which sends the plot to a

separate window

#### Value

nothing but it does generate a plot either to a window or a file

#### **Examples**

```
print("wait on suitable data sets")
```

plothsstats plots some HS performance statistics
plothsstats plots some HS performance statistics

# Description

plothstats generates plots for the sum of catches for the first 5 and 10 years of the projections. This is really only interesting when compared with the same statistics from a different operating model definition or different harvest strategy. The input data is generated during the running of do\_MSE.

#### Usage

```
plothsstats(rundir, hsstats, glb)
```

# **Arguments**

rundir the scenario's results directory

hsstats the statistics about the harvest strategies performance

glb the global constants object

plotNt 81

#### Value

nothing but it does generate two plots into rundir

# **Examples**

```
print("wait on suitable in ternal data sets")
```

plotNt

plotNt plots the size-composition for each SAU

# **Description**

plotNt accepts size-composition data (either the population Nt, of the catchN), and plots the replicate size-distributions for each SAU. It then puts the first year's size-distribution on top and if the medcol that defines the colour of hte median of the replicates is different from 0 (not plotted) then it plots the median as well. A contrasting colour choice is 4 = Blue.

# Usage

```
plotNt(Nt, year, glb, start = 3, medcol = 0)
```

# **Arguments**

The time is difficult and of multipers at size inclass, years, its auticult	Nt	the 4 dimensional arra	y of numbers-at-size	Nclass, years, nsau, reps
---	----	------------------------	----------------------	---------------------------

year which year from dimension 2 should be plotted?

glb the global constants object

start from what size class should the maximum Y value be measured. The default =

3 (= 6 mm)

medcol colour of the median line; default=0 = not plotted

# Value

invisibly the matrix of median values, Nclass  $\boldsymbol{x}$  nsau

```
print("wait on suitable built in data-sets")
```

82 plotprod

plotpopprops	plotpopprops plots out some properties of input populations	

# **Description**

once the depleted zone (zoneCP and zoneDD) has been generated the component populations have their properties summarized in propD. This is saved as propertyDD.csv and output to the zoneDD tab. To provide a visual consideration of these properties this function generated histograms for an array of input array column names.

# Usage

```
plotpopprops(x, rundir, glb, varnames, startyr, console = TRUE, bins = 25)
```

# **Arguments**

x	the array containing columns to plot
rundir	the run directory for the results, can be set to "" if plotting to the console
glb	the global constants object
varnames	the names of each variable to select the columns and to use in labels
startyr	in what year number should the plot refer to?
console	should plot go to the console or a png be saved? Default=TRUE
bins	the number of breaks to plot in each histogram, default=25

#### Value

it generates a png file of the plot if console = FALSE, otherwise it is drawn to the console

# **Examples**

```
print("wait on suitable data")
# x=propD;rundir=rundir;glb=glb;varnames=columns;startyr=hyrs;bins=21;console=FALSE
```

plotprod	plotprod graphs a selected productivity variable

# Description

plotprod graphs a selected productivity variable from the choice of ExB exploitable biomass, MatB mature of spawning biomass = Bmsy, AnnH the actual annual harvest rate Hmsy, Catch the yield at Bmsy and Hmsy = MSY, Deplet the mature biomass depletion level, and RelCE the relative cpue at MSY

plotprod 83

# Usage

```
plotprod(
  product,
  xname = "MatB",
  yname = "Catch",
  xlimit = NA,
  xlab = "Mature Biomass t",
  ylab = "Production t",
  font = 7,
  filename = "",
  devoff = FALSE
)
```

# **Arguments**

product	the output from doproduction
xname	the name of the productivity variable for the x-axis, defaults to MatB
yname	the name of the y-xis variable, default=Catch=Yield
xlimit default=NA, enables the range of the x-axis to be constrained, for example $xlimit=c(0.15,0.4)$	
xlab the x-axis label, default=""	
ylab the y-axis label, default=""	
font the type of font used in the plot. default = 7, bold Times, 6 is sans-serif, 2 bold sans-serif	
filename the complete path and filename of where to save the png plot file. empty, meaning no file is produced.	
devoff	a boolean to allow the plot device to remain open for the user to add more components. Of course, if this is set to FALSE then it is up to the user to use dev.off

# Value

invisibly a list of the x and y matrices plotted

```
data(zone)
product <- zone$product
plotprod(product)
stat <- findmsy(product)
abline(h=stat[,"Catch"],col=1:6,lwd=2)
abline(v=stat[,"MatB"],col=1:6,lwd=2)
print(stat)</pre>
```

84 plotscene

plotproductivity

plotproductivity characterizes each population's yield curve

# Description

plotproductivity characterizes each population's yield curve, it also describes the total productivity of the zone.

# Usage

```
plotproductivity(rundir, product, glb)
```

# Arguments

rundir the results directory
product the productivity 3-D array

glb the globals list

# Value

nothing but it does place five png files into rundir

# **Examples**

```
print("this will be quite long when I get to it")
```

plotscene

plotscene literally plots up the output from comparevar

# **Description**

plotscene takes the output from comparevar and plots the quantiles relative to each other.

```
plotscene(
   scenquant,
   glbc,
   var = "cpue",
   ymin = 0,
   filen = "",
   legloc = "topleft",
   legplot = 1
)
```

plotsizecomp 85

# **Arguments**

	scenquant	a list of the quantiles for the given variable from each scenario
	glbc	a list of the global objects from each scenario being compared
•		what variable from the dynamics to summarize, valid names include catch, acatch, cpue, harvestR, desplsB, depleB, recruit, matureB, and exploitB, default = 'cpue'
	ymin	allows one to set the lower limit to the yaxis, default = $0$
	filen	a filename for use if saving the output to said file, default = ", which implies the plot goes to the console.
	legloc	legend location, default='topleft'
	legplot	which plot should contain the legend, default=1

#### Value

nothing but it does generate a plot with nsau panels

# **Examples**

```
print("wait on internal datasets")
```

plotsizecomp generates a plot of available size composition data

# Description

plotsizecomp generates a plot of available size composition data with the option of plotting the predicted size-composition of catch on top of it; this latter option is only possible when plotting the distributions as proportions.

```
plotsizecomp(
  rundir,
  incomp,
  SAU = "",
  lml = NULL,
  catchN = NULL,
  start = NA,
  proportion = TRUE,
  console = TRUE,
  width = 10,
  height = 9,
  fnt = 7,
  tabcategory = "predictedcatchN"
)
```

86 plotZone

#### **Arguments**

rundir the directory for all ctrl, data, and output files.
incomp filtered size-composition data from the fishery
SAU the name of the SAU used to label the y-axis

lml a vector of the lml in each year represented by incomp

catchN the predicted size-composition of the catch

start which size-class to start from, default=NA, which means all size-classes in the

samples will be used.

proportion should the plots be as proportions or counts, default=TRUE

console should the plot be sent of the console or saved to rundir. default=TRUE

width the width of the plot, default = 10 height the height of the plot, default = 9

fnt what font to use, default font = 7, bold times

tabcategory what name to give to the webage tab when the plots are saved. Default="predictedcatchN"

#### Value

nothing but it does generate a plot

# See Also

preparesizecomp, popNAStosau

## **Examples**

```
print("wait on data sets")
```

plotZone

plotZone plots the projected TAC across all replicates

# Description

plotZone plots out the projected TAC for all replicates. The first value is the sum of the final catches in the conditioning period.

```
plotZone(
   inzone,
   rundir,
   glb,
   startyr,
   CIprobs = c(0.05, 0.5, 0.95),
   addfile = TRUE
)
```

plotzonesau 87

# **Arguments**

inzone	the dynamic zone projection object
rundir	the rundir for the scenario
glb the global constants object	
startyr	which year number to begin the plot
CIprobs	the quantiles used to generate the confidence intervals. The default = $c(0.05, 0.5, 0.95)$
addfile	should the plots be added to the rundir =TRUE or only sent to the console =

# FALSE. Default = TRUE

#### Value

nothing but it adds a plot to the zonescale tab in the rundir

# **Examples**

```
print("wait on suitable datasets")
# inzone=outzone;rundir=rundir;glb=glb;CIprobs=c(0.05,0.5,0.95);addfile=TRUE; startyr=40
```

plotzonesau

plotzonesau generates a plot of the zone scale and sau scale

#### **Description**

plotsonesau enables the zone catch or catch weighted zone cpue (from catchweightCE) to be compared with the sau catch or the sau cpue within a single plot. This has value for gaining insights into the fishery dynamics of a zone and how it responds to fishing pressure. NOTE WELL: Currently, this function is setup for Tasmania's set of 8 SAU. Assuming other jurisdictions have more or less then the layout of the plots would need to be altered. Until these are known this function will likely only work as intended with Tasmanian data!

# Usage

```
plotzonesau(
  zonetot,
  saudat,
  saunames,
  label,
  labelsau,
  side = 3,
  sauscale = TRUE
)
```

# Arguments

zonetot a vector of either the total catch or zone summary cpue for a zone. Ideally, the

vector should be named using the year in which each value was taken.

saudat either the catch or the cpue for each SAU (year x sau), again this matrix should

have both row and column names

88 popNAStosau

saunames a vector of nSAU names. Ideally very short ones to ensure they fit on each small

plot

label the y-axis label for the zone plot

labelsau the y-axis label for each row of the SAU plots

side should the SAU label be at the top of bottom of each plot. default = 3 (= top of

plot). The main alternative would be 1 (the bottom) but 2 (left side) and 4 (right

side) also work.

sauscale should all SAU plots have the own y-axis scale = TRUE, the default. If you want

each plot to have the same scale sauscale=FALSE.

#### Value

nothing but this does generate a plot.

#### See Also

catchweightCE

#### **Examples**

print("wait on suitable internal dats sets")

popNAStosau

popNAStosau converts NAS data by population to NAS x sau

#### **Description**

popNAStosau converts numbers-at-size data x population into the same by SAU. This is designed to compare ny observed NAS data from a zone with those predicted by the conditioned model, it could also be used to improve the conditioning code to avoid fitting only to CPUE

#### Usage

```
popNAStosau(popNAS, glb)
```

#### Arguments

popNAS either the catchN or the Nt from zoneDD, that is the zone after the equilibrium

zone (zoneD) has had the historical catches applied.

glb the globals object. Needed for the sauindex and various array dimnesion labels.

#### Value

An Nclass x nyrs x nsau array of numbers-at-size data

#### See Also

preparesizecomp, plotsizecomp

# **Examples**

#e.g. popNAStosau(out\$zoneDD\$catchN,out\$glb)

poptosau 89

poptosau	poptosau converts projected population dynamics to SAU scale results

# **Description**

poptosau the MSE dynamics are run at the population level but all management decisions are made at the SAU level, hence the results of the projections need to be translated into results at the SAU level. poptosau uses the sauindex to sum the variables that can be summed, which include matureB, exploitB, catch, and recruit, and combines their population results to form their respective SAU results. Thus, for an input array of dimensions [projyrs,numpop,reps] one receives back an array of [projyrs, nSAU,reps]

# Usage

```
poptosau(invar, glb)
```

# Arguments

invar either zoneDP\$ matureB, exploitB, catch, or recruit

glb the global constants object

#### Value

a results array of dimension [projyrs, nSAU,reps]

# **Examples**

```
print("wait on appropriate internal datasets")
```

poptosauCE	poptosauCE combines population cpue into sau as catch weighted
	sums

# Description

poptosauCE combines cpue from separate populations into their respective sau using a catch-weighted strategy. The sauindex is used to identify which populations to apply the sau total catches to.

# Usage

```
poptosauCE(catvect, cpuevect, sauindex)
```

# Arguments

catvect the vector of catches x population for a given year cpuevect the vector of cpue x population for a given year

sauindex the sau indices of each population

90 poptozone

#### Value

a list of saucpue and saucatch

#### **Examples**

```
print("wait on appropriate built-in data files")
# catvect=zoneDD$catch[1:finalyr,]; cpuevect=zoneDD$cpue[1:finalyr,]
```

poptozone

poptozone translates the zone\_pop objects to a single zone object

# **Description**

poptozone combines the dynamic results for each variable so that results by population become results by zone. matureB, exploitB, catch, recruit, catchN, and Nt are simple summations of the totals for each population into their respective Zone The harvest rate would be end of year or beginning of year estimates derived from dividing the catch x zone by the exploitable biomass x zone. Similarly the deplsB and depleB are the end of year matureB and exploitB divided by their respective unfished estimated by Zone obtained using getvar(zoneC,"B0").

# Usage

```
poptozone(inzone, NAS = NULL, glb, B0, ExB0)
```

# **Arguments**

inzone	one of the zone dynamics objects containing replicates, made up of populations
NAS	the numbers-at-size 4D arrays from doprojection; default=NULL so it can be ignored during conditioning
glb	the object containing the global constants
В0	the sum of B0 across all populations, use getvar(zoneC,"B0")
ExB0	the sum of ExB0 across all populations use getvar(zoneC, "ExB0")

#### Value

a list of dynamics variables by zone

```
print("wait on suitable internal data sets ")
```

prepareDDNt 91

prepareDDNt converts the population based historical Nt to SA based	\ <i>U</i> -
---	--------------

# **Description**

prepareDDNt processes the zoneDD, following the conditioning on the historical fishery data, so that the population numbers-at-size are converted to SAU-based numbers-at-size. It does this for both Nt and catchN. This is to allow analysis, tabulation, and plotting at the SAU scale. Input data is 3D Nclass x years x numpop

#### Usage

```
prepareDDNt(inNt, incatchN, glb)
```

#### **Arguments**

inNt the 3 dimensional array of population numbers-at-size

incatchN the 3 dimensional array of catch numbers-at-size

glb the global constants object

# Value

a list of Nt and catchN x SAU

# **Examples**

```
print("wait on suitable internal data-sets")
```

prepareprojection

prepareprojection high level function that sets up a projection

# **Description**

prepareprojection is a high level function that restructures zoneC by including the projected selectivity (in case the LML is set to change during the projections), and also selectivity x weight-at-size (for computational speed). It then generates a new zoneD with room for all replicates as well as the aspirational catches from each HS. It does this by converting the arrays of year x pop, to year x pop x replicate. It then uses the conditioned data in zoneDep to predict the first aspirational catches for the projections and conducts the initial replicate, thus starting the application of the HS. Finally, it adds recruitment variation to each of the replicates and keeps the last year of each iteration of the addrecvar function as the start of each replicate projection, with zeros in the catch, cpue, and cesau arrays

92 preparesizecomp

#### Usage

```
prepareprojection(
  projC = projC,
  condC = condC,
  zoneC = zoneC,
  glb = glb,
  zoneDD = zoneDD,
  ctrl = ctrl,
  varyrs = varyrs,
  calcpopC = calcpopC,
  lastsigR = 0.3
)
```

#### **Arguments**

projC	the projection object from readctrlfile
condC	historical conditioning data
zoneC	the constant part of the zone
glb	the global variables
zoneDD	the zone after initial depletion through conditioning on the fishery
ctrl	the ctrl object for the scenario run
varyrs	how many years at the end to add recruitment variation
calcpopC	a function that takes the output from herfun and gernerates the actual catch per population expected in the current year.
lastsigR	recruitment variation for when it is applied for varyrs

#### Value

a list of the dynamic zone object as a list of arrays of projyrs x populations x replicates, plus the revised projC and revised zoneC

# **Examples**

```
print("wait on data files")
```

preparesizecomp

preparesizecomp strips out empty columns and identifies samples

# **Description**

preparesizecomp takes in the size composition of catch data and removes empty columns and those with totals of less than the defined mincount. The minimum used depends on sampling routines. In Tasmania, a sample of 100 is the usual minimum taken from individual landings. Single landings may not be representative of much.

```
preparesizecomp(sizecomp, mincount = 100)
```

print.zone 93

# Arguments

sizecomp the size-composition of catch data from the dat object made by readLBMdata.

mincount the minimum number of observations within a year for inclusion in the analysis,

default = 100.

# Value

a cleaned version of the sizecomp matrix

# **Examples**

```
print("wait on data sets")
```

print.zone

print.zone S3 method for printing a summary of a given zone

# Description

print.zone S3 method for printing a summary of a given zone

# Usage

```
## S3 method for class 'zone'
print(x, ...)
```

# Arguments

x a zone object

... in case there are extra parameters

# Value

nothing, but does print out a summary of the zone

```
## Not run:
txt1 <- 'all example code should be able to run'
## End(Not run)</pre>
```

94 printline

```
print.zoneDefinition print.zoneDefinition S3 method for printing zonedef summary
```

# **Description**

print.zoneDefinition S3 method for printing a summary of the zoneDef.

# Usage

```
## S3 method for class 'zoneDefinition' print(x, ...)
```

#### **Arguments**

x a zonedefinition object

... in case there are extra parameters

#### Value

nothing, but does print out a summary of the zeonDef

# **Examples**

```
## Not run:
txt1 <- 'all example code should be able to run'
## End(Not run)</pre>
```

printline

printline literally prints a selected line from a text file

# **Description**

printline is used to check that one has selected the correct line for modification inside a datafile. This is done when conditioning the MSE operating model on the parameter AvRec

#### Usage

```
printline(rundir, datafile, linenumber = 29)
```

# **Arguments**

rundir the scenario directory holding all files for a scenario

datafile the name of the datafile for the scenario, from the ctrl object

linenumber which linenumber to print, default =29

#### Value

nothing but it does print a line to the console

putNA 95

#### **Examples**

```
print("wait on suitable example")
```

putNA

putNA can add NAs to the start and end of a vector

# **Description**

putNA fulfils a common requirement to expand a vector with NAs to assist a plot by ensuring the x and y vectors are the same length. It can accept both character and numeric vectors. This used to be called addNA, but there is a function in base R with that name that does something completely different.

#### Usage

```
putNA(x, pre, post)
```

#### **Arguments**

x the vector to which NAs are to be added pre how many NA to be added the front post how many to be added to the end

#### Value

```
a vector of length(x) + pre + post
```

# **Examples**

```
vect <- rnorm(10,mean=5,sd=1)
putNA(vect,3,5)</pre>
```

readctrlfile

readctrlfile checks rundir contains the required csv files

#### **Description**

readctrlfile checks rundir contains the required csv files including the named control file, which then contains the names of the region data file, and the population data file. The run stops if any are not present or are misnamed.

# Usage

```
readctrlfile(rundir, infile = "control.csv", verbose = TRUE)
```

# Arguments

rundir	the directory in	n which all files	relating to a	narticular run :	are to be held
I Ulluli	the uncetory is	ii willich all liles	i ciuting to u	purticular run	are to be mera.

infile default="control.csv", the filename of the control file present in rundir contain-

ing information regarding the run.

verbose Should progress comments be printed to console, default=TRUE

96 readdatafile

#### Value

the control list for the run

#### **Examples**

```
## Not run:
# this has sinoce been modified and needs updating
rundir <- tempdir()
ctrlzonetemplate(rundir)
datafiletemplate(6,rundir,filename="zone1sau2pop6.csv")
ctrl <- readctrlfile(rundir)
ctrl
## End(Not run)</pre>
```

readdatafile

readdatafile reads in a matrix of data defining each population

# **Description**

readdatafile expects a matrix of probability density function definitions that are used to define the populations used in the simulation. These constitute the definition of popdefs.

#### Usage

```
readdatafile(numpop, indir, infile)
```

#### **Arguments**

numpop the total number of populations across the zone indir directory in which to find the date file infile character string with filename of the data file

#### Value

a matrix of values defining the PDFs used to define the properties of each population. The contents of popdefs

```
## Not run:
data(zone1)
glb <- zone1$globals
glb
data(constants)
constants
ctrlfile <- "control.csv"
ctrl <- readctrlfile(glb$numpop,rundir,ctrlfile)
reg1 <- readzonefile(rundir,ctrl$zonefile)
popdefs <- readdatafile(reg1$globals,rundir,ctrl$datafile)
print(popdefs)
## End(Not run)</pre>
```

readsaudatafile 97

readsaudatafile readsaudatafile generates the constants matrix from sau data	
--	--

#### **Description**

readsaudatafile uses data described at the SAU level to make the constants file, which is then used to generate the population has been implemented to simplify the conditioning of each operating model.

# Usage

```
readsaudatafile(rundir, infile, optpar = NULL)
```

# **Arguments**

rundir	the directory in which the data file is to be found. This will usually be the rundir for the scenario run
infile	the name of the specific datafile used.
optpar	the optimum parameters from sizemod used to replace inputs for the main pa-

# rameters estimated.

#### Value

a list of the constants matrix with values for each population and the original matrix of sau values from readsaudatafile

# **Examples**

```
print("wait on suitable data sets")
# rundir=rundir; infile=ctrl$datafile;optpar=opar
```

replaceVar replaces values of a variable in the input datafile

# Description

replaceVar replaces the values of a variable in the input datafile. TAKE CARE, it overwrites the original file! However, it also saves the original file by adding an '\_old' to the filename. Obviously if the function is used multiple times the original 'original' file will be over-written on the second use. Alternatively, one could read the datafile using readdateFile and then directly alter the condDat, e.g., in the case of a four block zone in which the AvRec vale is changed: cond-Dat\$constants["AvRec",] <- c(12.5,12.2,12.4,12.1)

```
replaceVar(infile, invar, newval)
```

98 resetexB0

# **Arguments**

infile the name, and path, of the data file to be altered invar the text name of the variable to be changed

newval the new value with which to replace the current values

# Value

The function over-writes the original file but saves the original by adding an '\_old' to the end of the original filename.

## **Examples**

```
## Not run:
filename <- datafiletemplate(numblock=1,filename="oneblock.csv")
replace(filename,"AvRec",15.75)
condDat <- readdatafile(filename)
print(round(condDat$constants,4))
## End(Not run)</pre>
```

resetexB0

resetexB0 resets the unfished exploitable biomass at time zero

# **Description**

resetexB0 sets the unfished exploitable biomass at time zero to the average of the exploitable biomass levels before and after any fishing (when there are zero catches). Hence growth and natural mortality are included.

## Usage

```
resetexB0(zoneC, zoneD)
```

# **Arguments**

zoneC the constant components of the simulated zone zoneD the dynamic components of the simulated zone

#### Value

a refreshed zoneC with updated ExB0 values and zoneD\$depleB=1

```
print("wait on data")
```

resetLML 99

resetLML	resetLML sets the LML for a given zone from a given year	

# **Description**

resetLML sets the LML for a given zone from a given year out to the final year. It changes the selectivity, the SelWt values, and the LML value per block.

# Usage

```
resetLML(inzone, inLML, inyear, glob)
```

# **Arguments**

inzone a zone object

inLML a vector of LML for each block to be imposed from inyear on.

inyear the start year for the new LML glob the global variables object

#### Value

A zone object with the LML changed from inyear on

# **Examples**

```
## Not run:
txt1 <- 'all example code should be able to run'
## End(Not run)</pre>
```

restart

restart transfers final year values of zoneD into the first year

# Description

restart transfers the final year values from the dynamics part of the zone (zoneD), into the first year. This is used, for example, when searching for an equilibrium state if there is larval dispersal > 0.0. Of if one sets the initial depletion to anything other than 1.0. Contains the option of setting every other cell to zero, which is the default.

# Usage

```
restart(oldzoneD, hyrs, npop, N, zero = TRUE)
```

# **Arguments**

oldzoneD the old zoneD containing the dynamics as run for Nyrs.

hyrs The number of years of dynamics, the hyrs from glb

npop The number of populations, the global numpop

N the number of size classes, the global Nclass

zero should the arrays be otherwise filled with zeros? The default = TRUE

100 rewritedatafile

#### Value

a list containing a revised dynamics list

#### **Examples**

```
print("wait for built in data sets")
```

rewritecontrolfile

rewritecontrolfile generates a revised control file

#### **Description**

rewritecontrolfile is used to generate a new control file after the option of reading in parameters from sizemod has been used. The rewritten file contains the revised parameter values.

# Usage

```
rewritecontrolfile(indir, zone1, controlfile)
```

# Arguments

indir the directory into which the new file should be written. This would usually be

the same as rundir, the scenario directory

zone1 a large object inside the output object from makeequilzone that contains SAUnames,

SAUpop, minc, cw, larvdisp, randomseed, initLML, condC, projC, globals, ctrl,

catches, and projyrs. Obviously there is some redundency.

controlfile name of the original control file

#### Value

nothing but it does write a file called 'oldcontrolfilename\_new.csv' into rundir

# Examples

```
print("wait on internal data sets")
```

rewritedatafile

rewrwitedatafile generates a revised saudatafile

# **Description**

rewritedatafile is used to generate a new saudata file after the option of reading in parameters from sizemod has been used. The rewritten file contains the revised parameter values.

```
rewritedatafile(indir, glb, zone1, saudat)
```

runthreeH 101

#### **Arguments**

indir the directory into which the new file should be written. This would usually be

the same as rundir, the scenario directory

glb the globals object

zone1 a large object inside the output object from makeequilzone that contains SAUnames,

SAUpop, minc, cw, larvdisp, randomseed, initLML, condC, projC, globals, ctrl,

catches, and projyrs. Obviously there is some redundency.

saudat the main contents of the saudata file

#### Value

nothing but it does write a file called 'oldsaudatafile\_new.csv' into the indir

# **Examples**

```
print("wait on internal data sets")
```

runthreeH

runthreeH conducts the dynamics with constant catch 3 times

#### **Description**

runthreeH is used when searching numerically for an equilibrium and it conducts the hyrs dynamics three times, each time through it replaces year 1 with year hyrs. Thus if hyrs is 40 it conducts 3 \* 39 years of dynamics (117 years). This is not exported. It uses zoneC but always it does this inside the environment of another function where zoneC can be found Used inside dodepletion and doproduction. maxiter may need to be increased when we introduce a larger movement rate between populations for greenlip, or if the number of conditioning years are fewer than 45.

# Usage

```
runthreeH(zoneC, zoneD, inHarv, glob, maxiter = 2)
```

## **Arguments**

zoneC the constants components of the simulated zone

zoneD the dynamics portion of the zone, with matrices and arrays for the dynamic

variables of the dynamics of the operating model

inHarv a vector, length numpop, of annual harvest rates to be held constant across all

years.

glob the globals variable from readzonefile

maxiter default=3; the number of runs through the equilibrium loop.

#### Value

a list containing a revised dynamics list, zoneD

#### See Also

dodepletion, doproduction

102 sauavrecssq

#### **Examples**

```
print("wait on built in data sets")
# zoneC=zoneC; zoneD=zoneD; glob=glob; inHarv=rep(initH[aH],numpop); maxiter=2
```

sauavrecssq

sauavrecssq applies the AvRec and returns the ssq from the cpue

# **Description**

sauavrecssq applies the AvRec vector while adjusting just one for the picksau, and then compares the predicted CPUE with the observed and returns a sum-of-squared differences. This is used by optim while using the Brent option to find an optimum for a single parameter.

# Usage

```
sauavrecssq(
  param,
  rundir,
  controlfile,
  datafile,
  linenum,
  calcpopC,
  extra,
  picksau,
  nsau,
  outplot = FALSE
)
```

## **Arguments**

param is the AvRec for the selected SAU
rundir the rundir for the scenario being considered
controlfile the name of the control file, no path
datafile the name of the data file, no path
linenum the linenumber containing the AvRec vector

calcpopC the HS function that calculates the aspirational catches for each SAU

extra the vector of SAU AvRecs minus the selected SAU value

picksau which SAU is to be worked on as in 1:nsau

nsau the total number of SAU

outplot shoudla plot be generated for output to the webpage. default=FALSE

# Value

the SSQ for the selected SAU in a comparison of predicted and observed CPUE

```
print("Wait on suitable internal data files")
# param=param;rundir=rundir;controlfile=controlfile;datafile=datafile;linenum=29,
# calcpopC=calcexpectpopC;extra=extra;picksau=sau;nsau=nsau
```

sauplots 103

saur	าเก	T S

sauplots generates the dosauplots for the dynamic variables

# Description

sauplots generates the dosauplots for the dynamic variables including cpue, catch, acatch (aspirational catch), matureB, exploitB, recruit, and harvestR

# Usage

```
sauplots(
  zoneDP,
  NAS,
  glb,
  rundir,
  B0,
  ExB0,
  startyr,
  addCI = TRUE,
  histCE = NULL,
  tabcat = "projSAU"
)
```

# **Arguments**

zoneDP	the dynamic object produced by the projections
NAS	the numbers-at-size 4D arrays from doprojection
glb	the object containing the global variables
rundir	the results directory
В0	the B0 values by population use getvar(zoneC,"B0")
ExB0	the ExB0 values by population use getvar(zoneC,"ExB0")
startyr	the index for the first year of the conditioning dynamics to include in the trajectory, to give startyr:indexoflastyear,eg startyr= $40$
addCI	should confidence intervals be added to envelope plots, default=TRUE
histCE	historical CPUE data used in CPUE plots, default=NULL
tabcat	the name of the results website tab for the plots

# Value

a list of lists of CI for each SAU and variable as well as the zoneDsau and zonePsau

```
print("wait on suitable internal data-sets")
```

104 sautopop

saurecdevs	saurecdevs plots the recruitment	t deviates for each SAU
------------	----------------------------------	-------------------------

# **Description**

saurecdevs enables a visual comparison of the 'fitted' recruitment deviates for each SAU.

# Usage

```
saurecdevs(recdevs, glb, rundir, filen = "")
```

# **Arguments**

recdevs the matrix of recdevs (out\$condC\$recdevs)

glb the globals object

rundir the scenario directory for results storage

filen the filename (no path needed) to be used in the rundir

# Value

nothing, but it does generate a plot and save a png file if filen is populated

# **Examples**

```
print("wait on suitable data sets")
```

sautopop	sautopop translates a vector of SAU properties into population properties

# Description

sautopop uses the sauindex object to distribute a set of SAU properties (for example recruitment deviates) into a set of population properties.

# Usage

```
sautopop(x, sauindex)
```

# **Arguments**

x the vector of length nsau, of properties

sauindex the index of which populations are in which SAU

# Value

a vector of length numpop with each population having the property of the respective SAU

saveobject 105

#### **Examples**

```
sauprop <- c(1,2,3,4,5) # 5 SAU each with 3 populations sauind <- c(1,1,1,2,2,2,3,3,3,4,4,4,5,5,5) # the sauindex sautopop(sauprop,sauind)
```

saveobject

saveobject is used to save RData files of particular objects

# **Description**

saveobject is used to save RData files of particular objects. Currently, after projecting 56 populations for 100 replicates the final 'out' object is over 500Mb, and zoneDP, the projection object, makes up 430Mb of that. Save that if you wish, but if, say, one wanted only to save the outzone object (which summarizes outcomes at the zone level), that is only 22kb. This function facilitates the saving process.

# Usage

```
saveobject(obname, object, postfix = "", rundir)
```

# Arguments

obname the name of the object to be saved, as a character string
object the parent object from which the object\$obname is to extracted
any postfix addition you want for the name default=""
rundir the run directory for the scenario

#### Value

nothing but it does save a file to the rundir

#### **Examples**

```
print("wait on tempdir use")
# obname="outzone"; postfix="test"; object=out; rundir=rundir
```

save\_hsargs

save\_hsargs sends a copy of the hs arguments to rundir

# **Description**

save\_hsargs saves a copy of the HS arguments (held in hsargs) to the rundir directory ready to be printed as text into the HSperf tab of the output webpage. They are stored in a file called hsargs.txt. The function expects hsargs to be a list, which it prints component by component to a txt file.

```
save_hsargs(rundir, hsargs)
```

106 scaleto1

# **Arguments**

rundir the data and results directry for the scenario

hsargs the harvest strategy arguments object.

# Value

it saves a text file into the rundir sub-directory and modifies the resultTable.csv file for the web-page summary of scenario results

# **Examples**

```
print("wait on suitable internal data sets")
```

scaleto1

scaleto1 scales an input vector of CPUE to a mean of one x avCE

# Description

scaleto1 scales a vector of CPUE to a mean of one or avCE. The use of a mean of one means that visual comparisons between different time-series becomes visually simplified. The avCE option could be used to scale the CPUE to the average geometric mean - so as to put it on the nominal scale

# Usage

```
scaleto1(invect)
```

# **Arguments**

invect

a vector of linear scale CPUE

# Value

a vector of CPUE re-scaled to a mean of one

```
ce <- c(0.4667187, 1.2628564, 0.8442146, 0.9813531, 0.5554076, 0.7426321) scaleto1(ce)
```

setupzone 107

setupzone

setupzone makes zone's constant, dynamic, and productivity parts

# Description

setupzone makes the zone's constant, dynamic, and productivity parts returning them, along with glb, in a list. The objective of this function is to generate the original unfished equilibrium zone of nSAU SAU, and numpop populations

# Usage

```
setupzone(
  constants,
  zone1,
  doproduct,
  uplim = 0.4,
  inc = 0.005,
  verbose = TRUE
)
```

# Arguments

constants the population constants derived from readdatafile

zone1 the zonal object driving the construction

doproduct boolean, should the productivity calculations be made during the conditioning.

defined in do\_MSE and makeequilzone

uplim the upper limit of harvest rate applied, default=0.4 inc the harvest rate increment at each step, default=0.005

verbose Should progress comments be printed to console, default=TRUE

#### Value

a list of zoneC, zoneD, product, and glb the main components of the zone

```
## Not run:
data(constants)
data(zone1)
out <- setupzone(constants,zone1)
zoneC <- out$zoneC
glb <- out$glb
str(zoneC[[1]])
str(glb)
## End(Not run)</pre>
```

108 STM

STM

STM Generates the Size Transition Matrix for Inverse Logistic

# **Description**

STM With the input of the four parameters inside a vector, and a vector of initial lengths or midpoints of size classes STM generates a square transition matrix with the probabilities of growing from each initial size into the same or larger sizes. Negative growth is disallowed. All columns in the matrix summ to one.

# Usage

```
STM(p, mids)
```

# **Arguments**

р

a vector of four parameters in the following order MaxDL the maximum growth increment of the inverse logistic, L50 the initial length at which the inflexion of the growth increment curve occurs, L95 - the initial length that defines the 95th percentile of the growth increments, SigMax - the maximum standard deviaiton of the normal distribution used to describe the spread of each distribution of growth increments

mids

a vector of initial lengths which also define the width of each size class thus, mids from 2 - 210 woul dimply 2mm size classes 1 - 3 = 2, 3 - 5 = 4, etc

## Value

A square matrix with dimension =the length of the mids vector

#### References

Haddon, M., Mundy, C., and D. Tarbath (2008) Using an inverse-logistic model to describe growth increments of blackip abalone (Haliotis rubra) in Tasmania. Fisheries Bulletin 106: 58-71

```
param <- c(25.0,120.0,170.0,4.0)
midpts <- seq(2,210,2)
G <- STM(param,midpts)
print(round(G[1:30,1:8],4))</pre>
```

summarizeprod 109

summarizeprod

summarizeprod generates a summary of the productivity properties

#### **Description**

summarizeprod generates a summary of the productivity properties by examining the productivity matrix for each SAU/population and extracting the Bmsy, annualH, MSY, Depletion, and RelCE at the maximum catch level (which approximates the MSY). It summarizes the total zone by summing all the productivity matrices and search for the largest catch again. It generates estimates of the annualH, depletion and RelCE by using a weighted average of those values from the separate SAU or populations, where the weighting is the proportion of the sum of the MSYs taken in each sau or population. This latter is only an approximation but provides at least an indication.

# Usage

```
summarizeprod(product, saunames)
```

#### **Arguments**

product

The productivity array from doproduction containing the range of imposed har-

vest rates, and the resulting outputs for each population

saunames the names of the different SAU

# Value

a matrix containing the approximate productivity matrix for the zone

# **Examples**

```
## Not run:
data(zone)
product <- zone$product
zoneprod <- summarizeprod(product,saunames=zone$zone1$SAUnames)
round(zoneprod,3)
## End(Not run)</pre>
```

sumpop2sau

sumpop2sau gathers population data into sau data using sauindex

# **Description**

sumpop2sau gathers population data into sau data using sauindex

```
sumpop2sau(invect, sauindex)
```

110 sumpops

# **Arguments**

invect a vector of population values for a given variable

sauindex the indices of each sau for each population

#### Value

a vector of length nsau containing the sum of population values for each sau

# **Examples**

```
\label{eq:vect} \begin{array}{lll} \text{vect} &<& c(5.8,6.2,13.2,23.8,3.3,3.7,29.7,26.3,38.9,9.1) \\ \text{sauind} &<& c(1,1,2,2,3,3,4,4,5,5) \\ \text{sumpop2sau(vect,sauind)} & \# \text{ should be 12 37 7 56 48} \\ \end{array}
```

sumpops

sumpops takes the zoneDP and sums the populations within each SAU

# Description

sumpops summarizes the matureB, exploitB, catch, recruit variables within the depleted zone that has undergone the replicate application of an HS, and sums the values from each population within each SAU.

# Usage

```
sumpops(invar, sauindex, saunames)
```

# Arguments

invar either zoneDP\$matureB or exploitB, or catch, or recruit

sauindex the SAU index of each population

saunames the names of each SAU

#### Value

```
an array of projyrs x nSAU x reps
```

```
print("wait on new data")
```

tasab 111

tasab

tasab is a matrix of abalone maturity-at-length data

#### **Description**

tasab is a 715 x 4 matrix of maturity-at-length data for blacklip abalone (*Haliotis rubra*) from two sites along the Tasmanian west coast. All data was collected in February 1995, but details, such as site name, accurate location, statistical block, year, month, and other details have been omitted for brevity.

#### **Format**

```
A data.frame of maturity-at-length data 

site an identifier for the two different sites sampled 

sex I = immature, M = male, F = female 

length the shell length in mm 

mature was the animal mature = 1 or not = 0
```

#### **Subjects**

- maturity ogives or logistic curves
- · Binomial likelihoods

#### Source

Thanks to the Institute of Marine and Antarctic Science, which is part of the University of Tasmania, and especially to Dr Craig Mundy, leader of the Abalone Group, for permission to use this data collected in February 1995.

# Examples

```
data(tasab)
head(tasab,20)
table(tasab$site,tasab$sex)
```

testequil

testequil runs a zone for hyrs and determines stability

#### **Description**

testequil runs a given zone for hyrs at the given harvest rate, and then tests that the last values of matureB, exploitB, recruitment, and spawning biomass depletion are the same as the first (to three decimal places). It reports this to the console if verbose=TRUE. This is used with a harvest rate of zero and no variation in recruitment when defining the equilibrium zone under the application of the movement matrix.

```
testequil(zoneC, zoneD, glb, inH = 0, verbose = TRUE)
```

112 WtatLen

#### **Arguments**

zoneC the constants components of the simulated zone zoneD the dynamic components of the simulated zone

glb the global variables

inH a vector of numpop harvest rates

verbose should results go to the console, default=TRUE

#### Value

the dynamics component with hyrs of dynamics

#### **Examples**

```
## Not run: # modzoneC takes too long to run because of doproduction
data(zone)
zoneDe <- testequil(zoneC=zone$zoneC,zoneD=zone$zoneD,glb=zone$glb)
## End(Not run) #zoneC=zoneC; zoneD=zoneD; glb=glb; inH=0.0; verbose=TRUE</pre>
```

WtatLen

WtatLen Power function to describe weight at length relationship

# **Description**

WtatLen This can be used to generate any power function.

# Usage

```
WtatLen(ina, inb, lens)
```

# **Arguments**

ina is the intercept of the power function

inb is the gradient (or the explonent) of the power functionlens a vector of lengths for which the weight will be calculated

#### Value

A vector of length(lens) containing the predicted values

```
## Not run:
a <- 0.0000542856
b <- 3.161415
lens <- seq(2,210,2)
wtL <- WtatLen(a,b,lens)
## End(Not run)</pre>
```

zone 113

zone	zone the primary object obtained from the function makeequilzone

# **Description**

zone contains seven objects, including 5 lists, a matrix, and an array. This is the

#### **Format**

A list of objects plus a matrix and array that make up the initial equilibrium zone

**zoneC** a list of the constants for each population

zoneD a list of the dynamic parts of the populations of a zone

glb a list of global constants, containing numpop,nSAU,midpts, Nclass, Nyrs

**constants** a matrix of biological properties for each population in the zone, derived from the

**product** the productivity array from doproduction

**ctrl** the list containing control information for the run, including the datafile for the constants, the reps, the variation to be included when projecting

zone1 a list of objects used in the MSE

#### **Examples**

```
data(zone)
str(zone,max.level=1)
```

zonetosau

zonetosau translates the zonexpop objects to zonexsau objects

## **Description**

zonetosau combines the dynamic results for each variable so that results by population become results by SAU. matureB, exploitB, catch, recruit, catchN, and Nt are simple summations of the totals for each population into their respective SAU. The harvest rate would be end of year or beginning of year estimates derived from dividing the catchxsau by the exploitable biomass x sau. Similarly the deplsB and depleB are the end of year matureB and exploitB divided by their respective unfished estimated by SAU obtained using getvar(zoneC,"B0").

#### Usage

```
zonetosau(inzone, NAS = NULL, glb, B0, ExB0)
```

# Arguments

inzone	the projected zone dynamics objects made up of populations
NAS	the numbers-at-size 4D arrays from doprojection; default=NULL so it can be ignored during conditioning
glb	the object containing the global constants
В0	the estimate B0 for each population use getvar(zoneC,"B0")
ExB0	the estimate ExB0 for each population use getvar(zoneC, "ExB0")

114 zonetosau

# Value

a list of dynamics variables by SAU

```
print("wait on suitable internal data sets ")
```

# Index

address 4	Ciabana alata 27
addpops, 4	fishery_plots, 37
addrecvar, 5	getaav, 38
alldirExists, 6	getavrec, 39
aMSE, 7	getConst, 39
asSAU, 8	getCPUEssq, 40
aszone, 8	getLFdata, 40, 45
hislam, plata 0	getLFlogL, 41
biology_plots, 9	getline, 42
blockE13, 10	getlistvar, 42
calcprojsel, 10	getLogical, 43
	getmaxCE, 44
calcsau, 11	getnas, 41, 44
catchweightCE, 12, 88	getprojyrC, <i>33</i> , 45
changecolumn, 12	getrecdevcolumn, 46
changeline, 13, 14	getsauzone, 46
changevar, 13, 14	getsingleNum, 47
checkmsedata, 15	getssqparts, 47
checksizecompdata, 15	getStr, 48
compareCPUE, 16	getsum, 49
comparevar, 16	gettasdevssq, 49
compzoneN, 17	getunFished, 50
confirmdir, 18	getvar, 51
copyto, 18	getvect, 52
ctrlfiletemplate, 19, 21	getzeneLF, 41, 45, 52
d-t-C:1-t1-t- 20 20	_
datafiletemplate, 20, 20	getzoneprod, 53
defineBlock, 21	getzoneprops, 53
definepops, 22	historicalplots, 54
depleteSAU, 23	
diagnosticsproj, 24	imperr, 55
do_condition, 30	
do_MSE, 26, 28, 31	lf10, 55
dodepletion, 24, 101	logistic, 56
dohistoricC, 25, 31, 33, 70, 71, 73	
doproduction, 26, 101	make_html, 28
doprojections, 27, 31, 33, 45	makeabpop, 57
dosau, 29	makeequilzone, <i>31</i> , <i>33</i> , 58
dosauplot, 29	makemove, 58
driftrec, 34	makeoutput, 59
	makewidedat, 60
fillzoneDef, 34	makezone, 61
findF1, 35	makezoneC, 62
findlinenumber, 13, 14, 36	makezoneDP, 62
findmsy, 37	maturity, 63

INDEX

midg, 64	runthreeH, 101
modprojC, 64	
modzoneC, 65	sauavrecssq, 102
modzoneCSel, 66	sauplots, 103
numbersatsize, 67	saurecdevs, 104 sautopop, 104 save_hsargs, 105
onesau, 67	save_nsargs, 105
oneyear, 68	scaleto1, <u>106</u>
oneyearcat, 26, 69, 70, 71, 73	setupzone, 107
oneyearD, 70	STM, 108
oneyearrec, 26, 70, 71, 73	summarizeprod, 109
oneyearsauC, 26, 28, 71, 72	sumpop2sau, 109
oneyrgrowth, 73	sumpops, 110
onezoneplot, 74	
optimizeAvRec, 75	tasab, 111
optimizerecdevs, 76	testequil, 111
opermizer educio, 70	
plotCNt, 77	WtatLen, 112
plotcondCPUE, 77	112
plotconditioning, 78	zone, 113
plothistcatch, 79	zonetosau, 113
plothistce, 80	
plothsstats, 80	
plotNt, 81	
plotpopprops, 82	
plotprod, 82	
plotproductivity, 84	
plotscene, <i>17</i> , 84	
plotsizecomp, 85, 88	
plotZone, 86	
plotzonesau, 87	
popNAStosau, 86, 88	
poptosau, 89	
poptosauCE, 89	
poptozone, 90	
prepareDDNt, 91	
prepareprojection, <i>31</i> , <i>33</i> , <i>45</i> , 91	
preparesizecomp, 86, 88, 92	
print.zone, 93	
print.zoneDefinition, 94	
printline, 94	
putNA, 95	
putna, 93	
readctrlfile, 20, 21, 95	
readdatafile, 96	
readsaudatafile, 20, 21, 97	
replaceVar, 97	
resetexB0, 98	
resetLML, 99	
restart, 99	
rewritecontrolfile, 100	
rewritedatafile, 100	
· -	