

(2) (JOISS)

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Chapter 1

intro

Chapter 2

CTD(SBE 911plus)

2.1

SBE 911plus CTD Sea-Bird Electronics , WOCE(World Ocean Circulation Experiments) (Kim et al,2000). SBE 9plus underwater unit (main housing, pump and sensors) CTD , , SBE 11 deck unit . , deck unit water sampler button , . 24 . SBE 9plus 1 1 .

SBE 5T SBE 5P (30 /s) 0.4 T-C duct . CTD . T-C duct duct 0.073 conductivity cell . (0.073) , SBE 11plus Deck Unit .

2.2

2.2.1

2.2.1.1

(resistance) , thermistor (, signal) . , / / , / thermistor / . time constant(step change , 63%) 70ms(0.070) . SBE9plus underwater unit 24Hz, 0.0417 . Sea bird . Data processing Alignctd .

2.2.1.2

2 cm³, 30 /s 67ms .
 time constant 24ms(0.024s) .
 . , , . SBE11plus deck unit 0.073
 , primary +1.75scans(24Hz 1.75/24=0.073 seconds) factory-set
 , thermocline (-0.08 ~0.08) .
 . alignctd .

2.2.1.3

(piezoresistive effect) .
 . 50 .
 . SBE 9plus , Paroscientific Digiquartz .
 , .

2.2.1.4

SBE 43 membrane , electrode Teflon
 membrane electrode electrolyte . electrode ,
 . 20°C time constant() 2 , sensor membrane
 age .

2.2.2

SBE 9plus SBE 32 Carousel Water Sampler .
 (horizontal mount) (vertical mount) ,
 .

2.2.2.1 vertical mount

13 mm, 19 mm Tygon (Tygon tubing) .
 . DO 13 mm 9.5 mm
 Tygon tubing Tygon tubing(13 mm) . DO
 conductivity cell Y-fitting .

2.2.2.2 horizontal mount

13 mm, 19 mm Tygon tubing . DO conductivity .
 DO . DO 13 mm 9.5 mm Tygon
 tubing Tygon tubing(13 mm) . DO conductivity
 cell .

2.2.3

2.2.3.1 bottom end cap

SBE 9plus bottom end cap pressure port connector 6 (4). pressure port primary temperature primary conductivity connecto(JB1, JB2), connector(JB3), secondary temperature secondary conductivity connector(JB4, JB5) , SBE bottom contact switch connector(JB6) . temperature conductivity pin 3 (3-pin) , 3-pin cable(; 17086, 6) bottom end cap connector(JB1, JB2, JB4, JB5) . JB3 2-pin cable(; 17133, 7) , 2 Y-cable(; 17799, 8) .

2.2.3.2 top end cap

SBE 9plus top end cap 2-pin connector(JT1), 3-pin connector(JT4) 5 6-pin connector(JT2, JT3, JT5, JT6, JT7) (5). 2-pin connector sea cable(; 17027, 17028, 17136 , 9) cable JT2, JT3, JT5, JT6 connector . DO (SBE43) 6-pin-4-pin cable(; 171491, 10) . SBE32 Carousel water sampler JT7 connector 6-pin-6-pin cable(; 17198, 11) , General Oceanics Rosette water sampler JT4 3-pin-3-pin cable(; 17196, 17533 , normal polarity, reverse polarity) . * SBE 9plus 2-pin connector 3 (JT1, JB3, JB6) JT1 sea cable JB6 CTD 2007 2 SBE 9plus JB6 (female) . JB3 sea cable JT1 (male) 2-pin connector .

2.3

2.3.1 SEASAVE

seasave CTD , GPS , CTD , display . seasave setup program setup file (*.psa) (12).

2.3.1.1 (Instrument Configuration) : seasave > configure inputs

configuration file , post-calibration .

seasave , configuration inputs Instrument configuration (3-1), (Open) (Create) (*.con), 'Modify' . 'Create' , '911/917plus CTD' , "Frequency channels suppressed" 0,1,2 (3-2). Frequency . T, C single/dual , dual T, C 0, dual T & single C 1, single T, C 2 . T, C frequency (voltage) T, C . "Voltage words suppressed" 0~4 (3-2), CTD underwater unit (DO, fluorometer, altimeter, nitrate sensor, turbidity-meter) .

JT2(V0, V1), JT3(V2, V3), JT5(V4, V5), JT6(V6, V7) 8 (2 Voltage . voltage) (3-3), voltage Voltage words suppressed . , JT6 altimeter V6 , 0 Voltage 7 . 0 voltage 7 , 1 voltage 5 , 2 voltage 3 , 3 voltage 1 , 4 . 4

“Computer interface” IEEE-448 RS-232 . Deck unit computer RS-232 .

“Scans to average” . CTD full data 1 , 24Hz SBE911 CTD 24 , 1 1 .

“Surface PAR voltage added” underwater unit PAR Surface PAR . Application Note 11s .

“NMEA position data added” NMEA , NMEA . Pressure (m) , configuration inputs “Miscellaneous” .

“Scan time added” data scan (GMT 1970 1 1) . calibration sheet , Free . 3-1 voltage . , Save save as () .

2.3.1.2 ‘Serial Ports’

. Deck unit computer 3-6 deck unit (4) SBE11 interface (RS-232) (7) MODEM CHANNEL(water sampler) Deck unit (3-7) 1 . 9pin 2~3 USB , 9pin USB USB . deck unit (3) . CTD Deck unit underwater unit (8) , NMEA (10) . , port (3-8). “CTD Serial port” deck unit (4) com port , Baud rate(9600 19200), data bits (8), parity (None) . “Water sampling port” deck unit (7) com port . “Serial Data Output data port” “Output data to serial port” . “SBE14 Remote display Serial Port” “Send data to SBE14 remote display” . water sampler . “Water sampler type” SBE carosel , “Number of Water Bottles” Niskin bottle Carosel water sampler trigger part (3-10) Bottle trigger . , bottle 6 , trigger 12 12 . “Firing sequence” sequential/User input , Sequential bottle firing , User input firing . “Enable remote firing” TCP/IP port computer firing .

Seasave window display (depth, average sound velocity, descent rate, acceleration, oxygen, plume anomaly, and potential temperature anomaly) , . “Latitude when NMEA is not available” NMEA navigation Seasave pressure depth depth . NMEA . ‘OK’ configuration file .

2.3.1.3 Water Sampler

Configure Inputs Water Sampler (20). Water sampler type SBE
Carousel . Number of Water Bottles carousel bottle (24
carousel 20 bottle 24). Firing sequence User Input (Sequential
firing bottle firing)

2.3.1.4

Configure Outputs SBE11plus Alarms (21). altimeter Enable
altimeter alarm . Alarm set point(meters) altimeter .

2.3.1.5 (Display)

Window display sea-save . 4-1 Fixed, Scrolled, Plot display
, , . 24Hz 8Hz ()
, 1 , 1 display , 0 .

2.4 4.

46

4.5. CELLTM

$Conductivity = (g + hf^2 + if^3 + jf^4)/[10(1 + \delta t + \varepsilon p)]$ Siemens/meter

Chapter 3

$$(\quad)$$

3.1

1.1 Guildline Autosol
8400B laboratory salinometer, Seabird SBE 4C, YSI/Hydrolab multiprobe
sensor

3.2

$$\begin{aligned} \text{Cox (1967)} \quad & \quad \quad \quad 100 \quad \quad \quad 135 \quad \quad \quad . \quad (1) \\ & (2) \quad \quad \quad (2) \quad \quad \quad (3) \end{aligned}$$

$$\begin{aligned}
S(\%) &= 0.03 + 1.8050 \text{ } Cl(\%) \text{ } \text{-----} \quad (1) \\
S(\%) &= 1.80655 \text{ } Cl(\%) \text{ } \text{-----} \quad (2) \\
S(\%) &= -0.08996 + 28.29720 \text{ } R_{15} + 12.80832 \text{ } R_{15}^2 \\
&\quad - 10.67869 \text{ } R_{15}^3 + 5.98624 R_{15}^4 - 1.32311 R_{15}^5 \text{ } \text{---} \quad (3)
\end{aligned}$$

International Association for Physical Sciences of the Ocean(IAPSO)
(chlorinity)
(Batch P31, chlorinity 19.375, S(‰)=35.002) . 1978 new salinity
scale Practical Salinity Scale (PSS78) . Practical Salinity Scale
(S_p,Practical salinty) 1kg KCl 32.4356g
KCl 15°C, (K₁₅) (4)
KCl 1kg KCl 32.4356 g 15°C, 35 ‰ 1.0000
KCl . IPTS68 (t68 = 15°C) , 1983 ITS-90 . 1983
1983
(T₆₈=1.00024T₉₀, Saunders, 1990).

$$S_p = 0.0080 - 0.1692 K_{15}^{\frac{1}{2}} + 25.3851 K_{15} + 14.0941 K_{15}^{\frac{3}{2}} - 7.0261 K_{15}^2 + 2.7801 K_{15}^{\frac{5}{2}} \text{ --- (4)}$$

(T) (RT) (4) (5) SBE 4C, YSI,
Hydrolab 15 °C (RT) .

$$S_p(\text{‰}) = 0.0080 - 0.1692 R_T^{\frac{1}{2}} + 25.3851 R_T + 14.0941 R_T^{\frac{3}{2}} - 7.0261 R_T^2 + 2.7801 R_T^{\frac{5}{2}} + \frac{(T-15)}{1+k(T-15)} (0.0005 - 0.0056 R_T^{\frac{1}{2}} - 0.0066 R_T - 0.0375 R_T^{\frac{3}{2}} + 0.0636 R_T^2 - 0.0144 R_T^{\frac{5}{2}}) - \text{ --- (5)}$$

k=0.0162 . T: salinometer

3.3

3.1 IAPSO ,

3.2 , , , , ,

3.4

T-S bridge, CTD, Salinometer . T-S bridge CTD
. CTD . (Salinometer)
Guildline Autosol8400B .

3.4.1 (Bentch top Salinometer)

Guildline Autosal 8400B . WOCE

3.4.2 T-S bridge

salinity bridge 5500
2-40 mmho/cm . T-S bridge

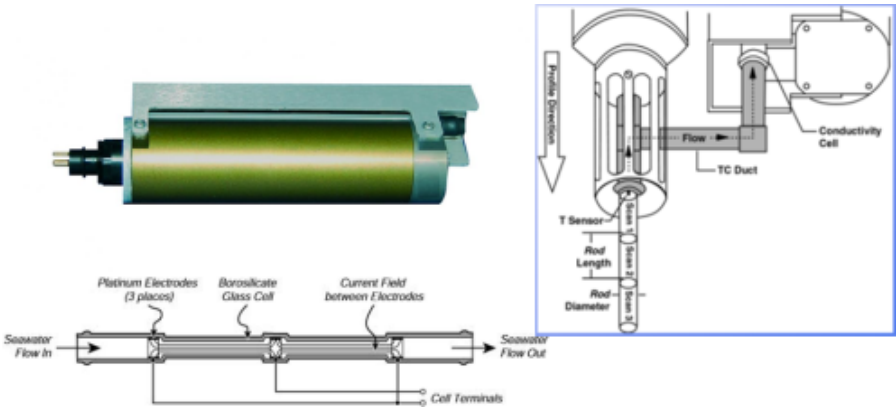


Figure 3.1: Guildline Autosol 8400B



3.4.3 Seabird conductivity sensor (SBE 4C)

SBE 4C 30 cm³/s 0.4 cm T-C duct
duct 0.73 . 0.73 . SBE



9plus data logger

3.4.4 YSI, Hydrolab etc. multiprobe sensor

YSI multiprobe Hydrolab multiprobe





4.5 , , .

3.5 , , ,

T-S bridge, CTD

, , , .

3.5.1

(flint glass) , poly-seal screw
cap . (borosilicate glass)** .
125-250 ml . 0.001 6 (Stalcup,1991). 2
(3) 8 (10) . *Pb K-SiO₂ , ,
, ** (Br)-SiO₂ , , . Pyrex, Boroil,
Simax, Bomex .

3.5.2

4 1 / 10 . 3 .

3.5.3

(Muller, 1999). 15-20 °C 10 (, 2013).

3.5.4

cruise identification, station, cast number, water sampler number, storage case identification, salinity sample bottle number .

3.6

6.1 Autosal 8400B

Autosal

6.2 Seabird 4C

Autosal 8400B

(1)

6.3 YSI, Hydrolab multiparameter conductivity sensor

SBE 4C

(=3%/°C).

mS/cm at 14 °C).

°C (=1.91%=0.0191).

1.0 mS/cm, : 10 mS/cm, : 50 mS/cm). -

Low-End Specific Conductance Check (5 S/cm),

1 Low-End Specific Conductance Check

500 1000 S/cm Mid-Range

Specific Conductance Check

Mid-Range conductivity solution

Mid-Range Specific Conductance Check

10%

3.7 Autosal 8400B

3.7.1

AUTOSAL

AUTOSAL

(standard seawater)

(conductivity ratio)

3.7.2

(1) : 230V AC 115V AC

(2) Bath : 16.8

(3) : 2

Full Calibration Log Sheet

Sonde ID: _____ Date: _____ Time: _____ Calibrator's Initials: _____
 Battery Check (V): _____ Field User's Initials (if different from Calibrator): _____
 Temperature (°C): _____ *Temperature must be recorded for all Calibrations
 (even single probe calibrations)!*

Specific Conductance Probe Calibration

Routine Maintenance Checklist:

1. *For best accuracy, Specific Conductivity should be calibrated according to the expectations of the streams/sites you will be sampling. It may be necessary to recalibrate between streams/sites due to extreme differences between streams/sites (e.g., 1000 vs. 10000 $\mu\text{mhos/cm}$). Was Specific Conductance calibrated accordingly?* ☐ Yes or ☐ No
2. *Be sure to check the age of the Specific Conductivity probe. It should be no more than 3 years old. Is it too old?* ☐ Yes or ☐ No

Conductivity Solution ($\mu\text{mhos/cm}$): _____

Initial Sp Cond ($\mu\text{mhos/cm}$): _____ Final Sp Cond ($\mu\text{mhos/cm}$): _____

Low-End Sp Cond Check (i.e., $<5 \mu\text{mhos/cm}$)

Solution: ☐ Deionized or ☐ Distilled Water Sp Cond ($\mu\text{mhos/cm}$): _____

Monthly Mid-Range Sp Cond Check (e.g., 500 or 1000 $\mu\text{mhos/cm}$)

Conductivity Solution ($\mu\text{mhos/cm}$): _____ Sp Cond ($\mu\text{mhos/cm}$): _____

Figure 3.2:

- (4) : 0.005 42 psu (Conductivity=7.6 S/m Rt=1.15)
- (5) : ±0.002 psu
- (6) : ±0.0002 psu
- (7)Bath :
- setting temp : 18°C, 21°C, 24°C, 27°C, 30°C, 33°C - : ±0.02 °C - : ±0.001 °C/day
- (8)Scale Suppression : 0 2.2 Conductivity ratio (22)



Figure 3.3: UTOSAL(8400B)



Figure 3.4: computer interface

3.7.3

, , , , , , , ()

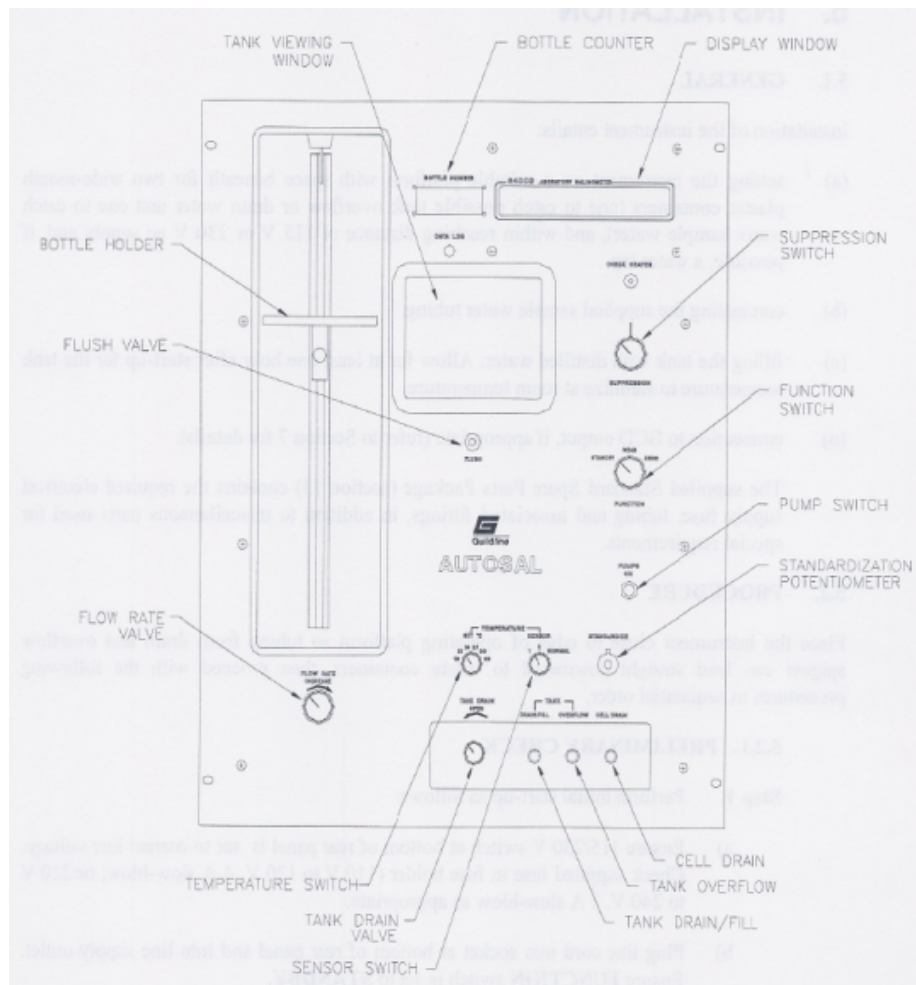


Figure 3.6: Autosol Front Panel

3.7.4 Autosal

- (1) .
- (2) switch valve .
- suppression switch : ()
- function switch : standby
- pump switch : off
- flow rate valve : ()
- temperature set switch : setting
- temperature sensor switch : normal
- tank drain valve :
- (3) tank drain/fill, tank overflow : ()
- (4) .
- (5) cell drain . cell drain
- .
- (6) , , , Autosal .
- (7) .

3.7.5 Autosal

- AUTOSAL 8 . (1) AC
- . - Autosal computer interface . - . - AUTOSAL
 - (AUTOSAL). - FUSE(250V / 2A) . -
 - . (2) temperature bath
 - () .
 - .
 - bath . bath $-2^{\circ}\text{C} + 4^{\circ}\text{C}$. (3) temperature sensor
 - temperature sensor switch normal temperature set switch 33 (heater lamp on-).
 - temperature sensor switch normal temperature set switch 18 (heater lamp off-).
 - (4) temperature sensor check
 - bath 1 , heater lamp .
 - temperature sensor switch normal 1 4 5 heater lamp . -
 - temperature sensor switch 1 2 4 5 heater lamp .
 - temperature sensor switch normal .
 - heater lamp heater lamp (heater lamp).

3.7.6

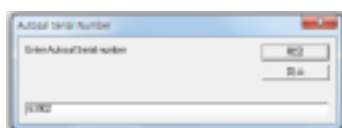
- (1) function switch, standby
- temperature set switch 24°C
- (2) function switch, read
- suppression switch 1.9
- suppression switch 2.0

computer interface — .

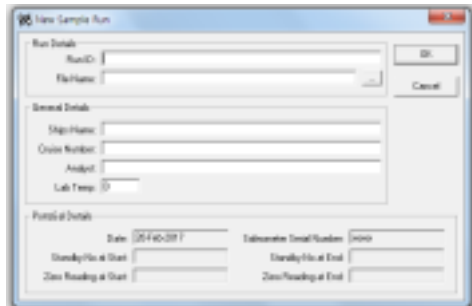
3.7.7 Autosal (warming-up)

, Autosal on , off , Bottle Number, zero

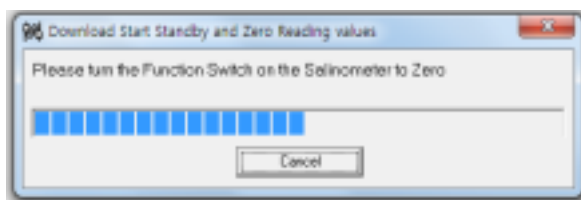
- (1) Salinometer Data Logger
- (2) File → New. Serial Number



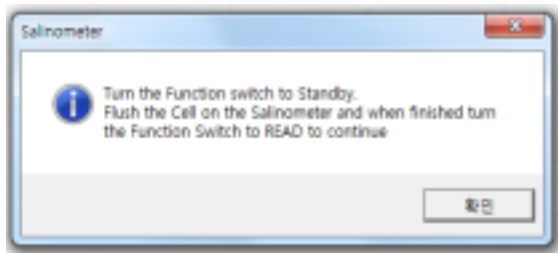
- (3) Run ID File Name OK



- (4) Autosal Function Switch Zero . 0.0+000x : $x=\pm 5$,



- (5) Autosal Function Switch Standby .



(6) 6 .

- Autosal Pump Switch 3 Flushing Pump Switch .
 - Pump(flow rate : 1) Switch Pump Switch .
 - 6 Function Switch Standby Read 10
 - Suppression Switch . display → Suppression Switch 0.0
 - Autosal display (Autosal).
 - Function Switch Read Standby .
- (7) 20 .

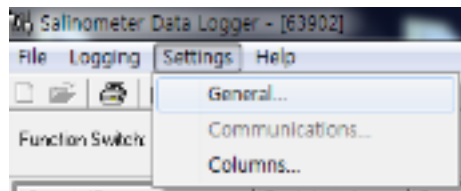


Figure 3.7:

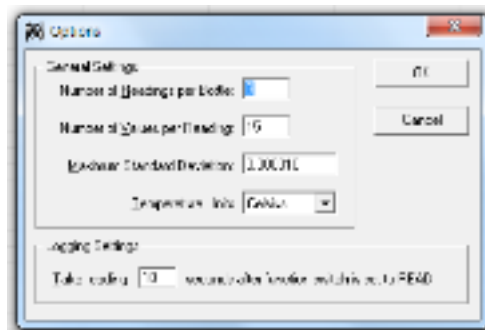


Figure 3.8:

- () pump flushing . 3~4
- Function Switch Read 10 Suppression Switch .
- display . → Sal. 35 Suppression Switch 1.9 → 20 .

- 1200 Reading Function Switch Read Standby .
- Maximum Standard Deviation 0.000010 . Accept Reading



Autosal .

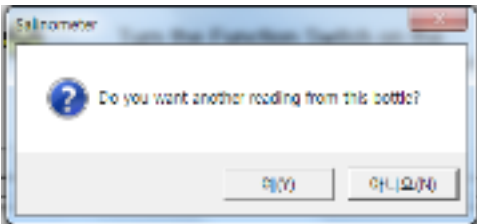


Figure 3.9:

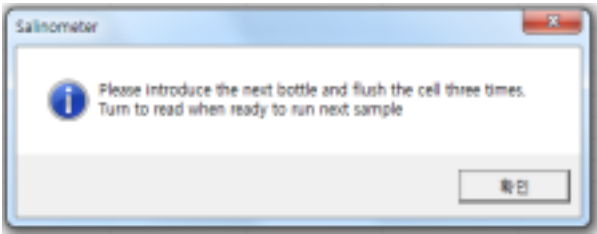


Figure 3.10:

- Bottle Label .
- 63902#3 3 .
1 (15) 1 .
(8) () 3 15 .
- 3 display ± 5 .

Sample ID	Bottle Label	Date/Time	Bath Temp	Uncon. Ratio	Uncon. Ratio SD	Correction	Adj Ratio	Calc Salinity
6399291	물막심(20분)	14-Apr-2017 09:44:00	24C	0.999067	0.029033	0.000000	0.999067	34.9276
6399292	물막심(11분)	14-Apr-2017 10:06:34	24C	0.999639	0.000005	0.000000	0.999639	34.9958
6399293	물막심(15초 38)	14-Apr-2017 10:12:32	24C	0.999817	0.000004	0.000000	0.999817	34.9928
6399294	포만대수 new...	14-Apr-2017 10:20:03	24C	0.999670	0.000004	0.000000	0.999670	34.9870
Calibration#1	cal	14-Apr-2017 10:24:20		0.999674	0.000004	0.000026	0.999700	
6399295	32.5deg T	14-Apr-2017 10:29:56	24C	0.970276	0.000003	0.000026	0.970302	34.1476

Figure 3.11:

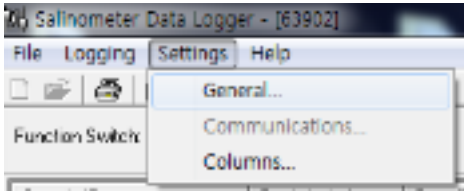


Figure 3.12:

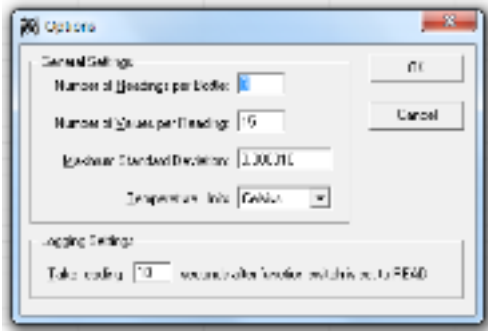


Figure 3.13:

3.7.8 Autosol (standardization potentiometer)

Autosol Warming-up .

(1)

(2)Autosol Pump Switch 3 Flushing Pump Switch .

(3) Pump(flow rate : 1) Switch Pump Switch .

(4) 3~4 .

(5)Function Switch Read 10 Standardization Potentiometer (, Potentiometer K15 2 ()).

$K15 = 0.99996$, $0.99996 \times 2 = 1.99992$.

, 1.99992 display Potentiometer .

(6)30 Reading Function Switch Read Standby .

(7)display K15 2 3 (flushing) (5)~(6) .
4 Bottle Counter . ± 5 Autosol .

3.7.9 Autosol ()

Standardization potentiometer Autosol .

Autosol Warming-up .

(1) . (2)Autosol Pump Switch 3

Flushing Pump Switch .

(3) Pump(flow rate : 1) Switch Pump Switch .

(4) 3~4 .

(5)Function Switch Read 10 Suppression Switch (display

→ Sal. 35 Suppression Switch 1.9) → Reading Function Switch

Read Standby → 3 (flushing)

(6) flushing .

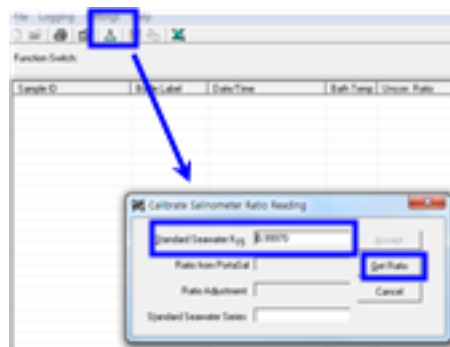


Figure 3.14:

- .
- Standard Seawater K15 21 K15 .

- Get Ratio Function Switch Read 10 Suppression Switch
(display → Sal. 35 Suppression Switch 1.9).
- Reading Function Switch Read Standby .
- Autosal display Accept .
- Ratio Salinity .

Sample ID	Bottle Label	Date/Time	Bath Temp	Uncon. Ratio	Uncon. Ratio SD	Correction	Adj. Ratio	Calc. Salinity
6290281	샘물샘(20분)	14Apr-2017 09:44:00	24C	0.999067	0.000033	0.000000	0.999067	34.9276
6290282	샘물샘(1분)	14Apr-2017 10:06:34	24C	0.999629	0.000005	0.000000	0.999629	34.9958
6290283	샘물샘(15초 3회)	14Apr-2017 10:12:32	24C	0.999817	0.000004	0.000000	0.999817	34.9928
6290284	표준해수 new...	14Apr-2017 10:20:03	24C	0.999670	0.000004	0.000000	0.999670	34.9870
Calibration#1	cal.	14Apr-2017 10:24:20		0.999674	0.000004	0.000026	0.999700	
6290285	32 Secap 1	14Apr-2017 10:28:56	24C	0.978276	0.000003	0.000026	0.978302	34.1476

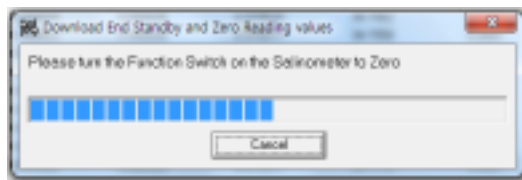
Figure 3.15:

3.7.10

- (1)
- (2)Autosal Pump Switch 3 Flushing Pump
Switch .
- (3) Pump(flow rate : 1) Switch Pump Switch .
- (4) 3~4 .
- (5)Function Switch Read 10 Suppression Switch (display
) → Reading Function Switch Read Standby → 3 (flushing
) .

3.7.11 Autosal

- (1) (6) .
- (2)Function Switch Standby Read 10 Suppression Switch (display
→ Suppression Switch 0.0 → Autosal display
(Autosal).
- (3)Function Switch Read Standby .
- (4)File → Close .



- (5)Function Switch Standby Zero Zero .
- (6)Function Switch Zero Standby .
- (7)AUTOSAL (AUTOSAL).
- (8)AUTOSAL .

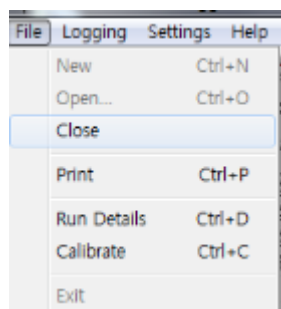


Figure 3.16:

3.7.12 Autosol

(1)OSIL

- 20mL 10mL Decon 90 methanol(95%) 170mL . methanol
- flushing 3~4 20 .
- flushing(10) .
- 400mL flushing .
- drain .
- .

(2)Guidline

- CLR, Isopropyl Alcohol, 200mL .
- CLR flushing 3~4 20 .
- CLR flushing(10) .
- Isopropyl Alcohol flushing 5~7 10~15 .
- Isopropyl Alcohol flushing(10) .
- 400mL flushing .
- drain .
- .

3.7.13

(1)

(2)cell drain . cell drain

(3) . 1~2 , digit . flushing

- flushing . 0.001psu .

(4)

(5) Autosol 4 (bath
) , impeller (O ring) .

(6)30 Autosol 3 ~6 . (7)

4~5

(8) (15%) . 2ppm .

(9)display ± 1 digit 0.0002psu (35psu).(10)bath temperature ± 0.5 mK ± 2 digits .

3.8

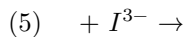
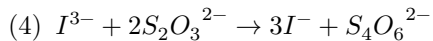
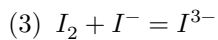
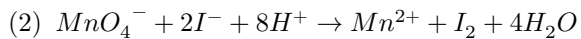
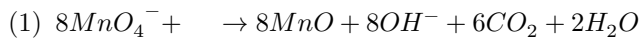
Cox, R.A., Culkin, F., Riley, J.P. (1967) The electrical conductivity/chlorinity relationship in natural sea water. Deep-Sea Research, 14:203-220
 Saunders, P.M. (1990) The international temperature scale of 1990, ITS-90. WOCE Newsl. No. 10
 Stalcup, M.C.(1991) Salinity measurements. WHP Operations and Methods
 Muller, T.J. (1999) Determination of salinity. In Methods of Seawater Analysis. Eds Grasshoff, K., Kremling, K., Ehrhardt, M. Wiley-VCH (2013)
 2013-230

Chapter 4

4.1

Erlenmeyer Flask	60 (2013-230)	Grahm	250ml
Borosilicate Watch Glass	100ml	Borosilicate	
	(KIOST modified MOF 2013-203)		

4.2

$$\begin{array}{c}
\begin{array}{ccccc}
(I_2) & (I^-) & (I^{3-}) & (I^{3-}) & (I_2) \\
(I^-) & (I^-) & (I^{3-}) & (I^{3-}) & (I^{3-})
\end{array}
\end{array}$$


()

(Biochemical Oxygen Demand)

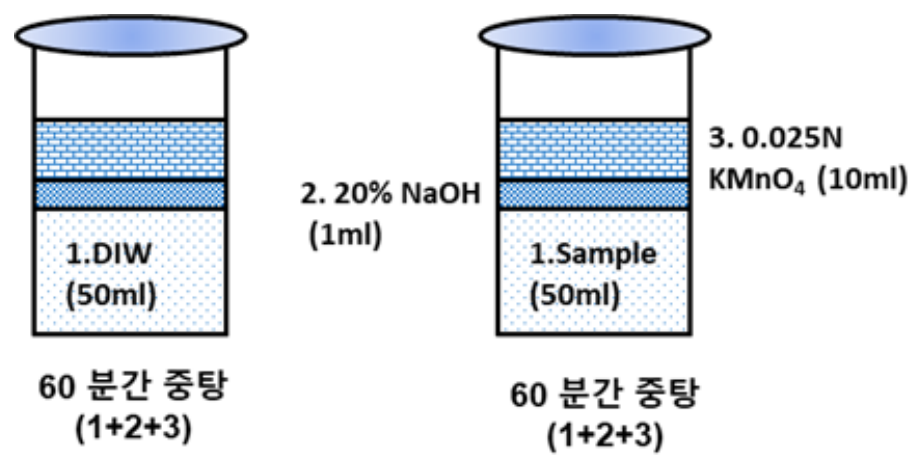


Figure 4.1:

,
(k=2) (MDL) 0.18 mgO₂/L 0.18~8 mgO₂/L , 95%
1/2 0.11 mgO₂/L . 60
8 mg O₂/L 50 mL .

4.3

30 50%

4.4

10% 3 (4.1), (4.2), (4.3), (4.4), (4.5) .

4.4.1

100 ml .

4.4.2

100 °C

4.4.3

100ml

4.4.4

4.5

(; Metrohm 665 Dosimat burette, SCHOTT Instruments TITRONIC universal).

4.6

(5.1), (5.2), (5.3), (5.4), (5.5), (5.6).
(5.7) .

4.6.1 20 % (w/v)

(NaOH) 20 g 100mL .

4.6.2 10% (w/v)

(KI) 5 g 50 mL .

4.6.3 50% (v/v)

(H₂SO₄) 50 mL 40 mL . 100 mL .

4.6.4 1%

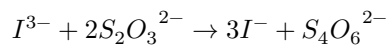
1 g 50 mL 100 mL .
(Hg₂I₂) . 1 .

4.6.5 0.025 N

(KMnO₄) 0.790 g 1000 mL . 1 2

4.6.6 0.01 N

(Na₂S₂O₃ · 5H₂O) 2.48 g 1000 mL .
1 (Na₂S₂O₃ · 5H₂O) (S) 2 1(=0.5 × 2) . $2S_2O_3^{2-} + I^{3-} \rightarrow$
 $S_4O_6^{2-} + 3I^-$;
 $[S^{2+}] \rightarrow [S^{2.5+}]$; $\Delta = 0.5$



(S) ;(-2) \rightarrow (-2.5)

4.6.7 0.001667 M (0.0100 N)

(KIO₃) 120°C 2 0.3567g 1000 mL .
0.3567g , .

$$M(\text{ }) = \frac{1L}{(214.0g)}$$

4.7

1.1. , 0~10°C () .

4.8

4.8.1 ()

7.1.1. 100 ml 10.0 ml (0.00167 M) . 40 mL .

7.1.2. 1 ml 50% 1 ml . (I⁻) (IO₃⁻)
(I₃⁻) (IO₃⁻ + 8I⁻ + 6H⁺ \rightarrow 3I₃⁻ + 3H₂O).

7.1.3. 1 ml .
(I₃⁻ + 2S₂O₃²⁻ \rightarrow 3I⁻ + S₄O₆²⁻). \pm 0.03 ml .

7.1.4. (IO₃⁻) 1 (S₂O₃²⁻) 6 .
3 .

$$C_{Na_2S_2O_3 \cdot 5H_2O} = \frac{C_{KIO_3} \times 60.0}{V_{Na_2S_2O_3 \cdot 5H_2O}} C_{Na_2S_2O_3 \cdot 5H_2O} : Na_2S_2O_3 \cdot 5H_2O \quad (mole/L) C_{KIO_3} : KIO_3 \quad (mole/L)$$

4.8.2

7.2.1. 100 mL 50 mL , 20% 1 mL . 7.2.2. 0.025 N

10.0 mL 60 . 7.2.3.

10% 1 mL , 50% 1 mL . 7.2.4. 1

mL . 7.2.4. (blank)

$$= \frac{C_{Na_2S_2O_3 \cdot 5H_2O} \times (V_{Na_2S_2O_3 \cdot 5H_2O; blank} - V_{Na_2S_2O_3 \cdot 5H_2O; sample}) \times 8000}{V_{sample}}$$

$C_{Na_2S_2O_3 \cdot 5H_2O}$;

$V_{Na_2S_2O_3 \cdot 5H_2O; blank}$; (L)

$V_{Na_2S_2O_3 \cdot 5H_2O; sample}$; (L)

V_{sample} ; (L)

4.9 / (QA/QC)

5 (blank) . 6.0 , .

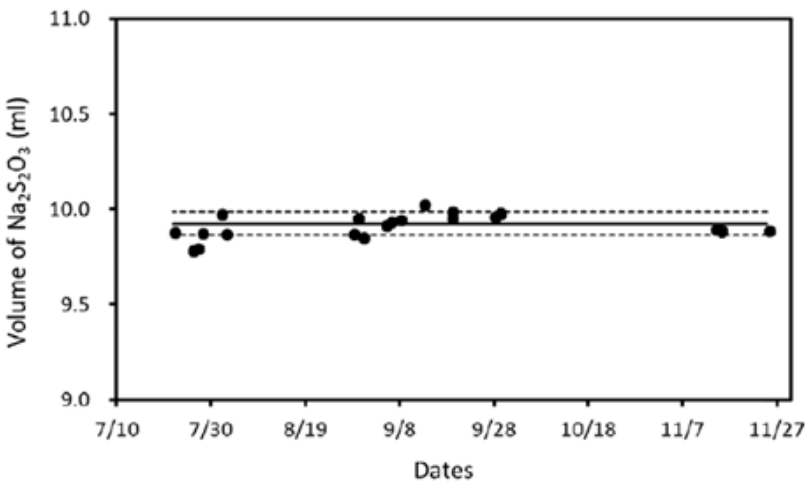


Figure 4.2:

4.10

4.11

(COD) (K₂Cr₂O₇) , pH (COD) ,

Chapter 5

ADCP

5.1

Acoustic Doppler Current Profilers(ADCP) (fixed-mounted)
(vessel-mounted) ADCP , , . ADCP



Figure 5.1: Workhorse Sentinel 300 kHz ADCP

5.2

ADCP , ADCP . ADCP .
ADCP ADCP . ADCP
ADCP . ADCP

5.3

ADCP ADCP PC PC
ADCP ADCP

5.4

ADCP , ADCP
(2.1).
300 kHz ADCP 150m , ADCP (side
lobe interference) ADCP

5.5

3.1. ADCP , ADCP ,
3.2. Teledyne RD Instrument's PlanADCP software
3.3.
3.4. ADCP ,

5.6

4.1. ADCP , ADCP 4
, (tilt) , (fluxgate) () PVC ,
ADCP (acoustic releaser, Benthos Model 875-A or 865-A), (Pinger, RJE
Model ULB-364/37), ADCP (4.1) 4.1 (d) 40 m(,
) ADCP ADCP ,
ADCP (, transducer) 30m
ADCP time-out release
4.1.1. (Acoustic Transducer Head)
4.1.1.1. 3 (u, v, w) 1 (Acoustic Transducer Head)
(transceiver electronic) , 4 piezoceramic acoustic
transducer 4 90° 25° (transducer head)
. 4 transducer() (300 kHz, 600kHz, 1.2 MHz) (acoustic pulse)
4.1.2. sensor tilt(pitch roll) Compass
(heading) . Heading direction . Micro- controller 3
. 3 x, y, z . Tilt-x tilt-y . tilt , heading magnetic
field tilt . RS232 CAN bus . heading pitch, roll
4.1.1. 4.1.2.

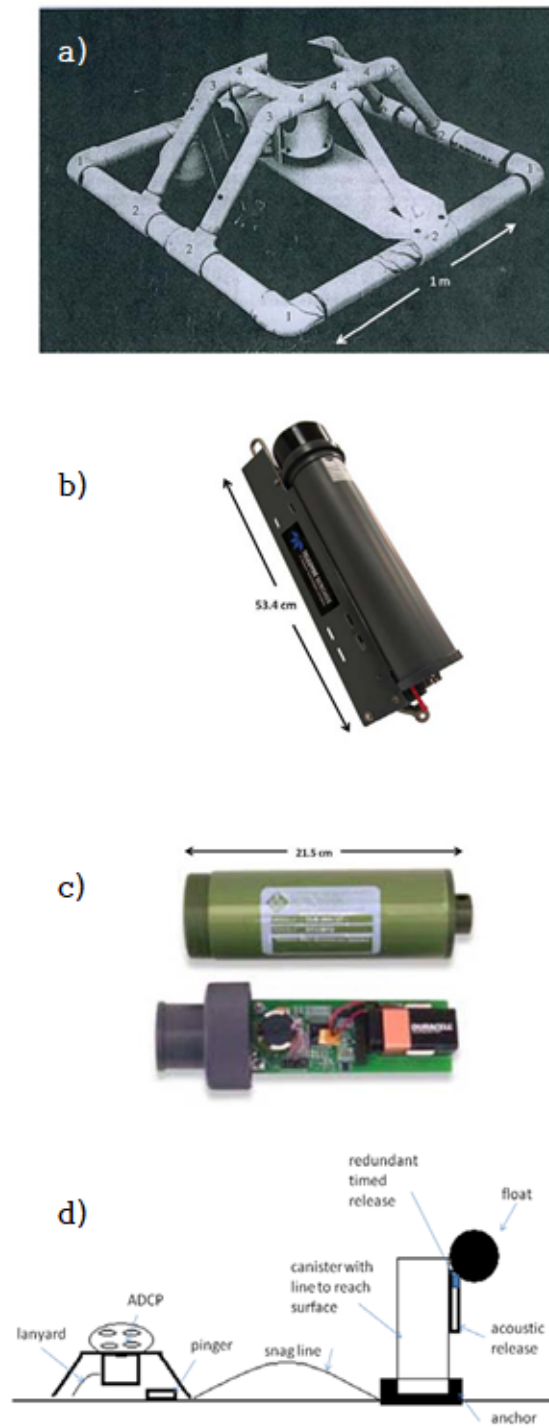


Figure 5.2: ADCP recovery system: a) mount, b) acoustic release, c) pinger, and d) detailed configuration for on-fixed deployment

4.2. Vessel-mounted ADCP , ADCP (ex.) . 4 , , , ADCP 3 . ADCP , ADCP , DGPS GGA-mode(GSA-mode) .



Figure 5.3: vessel-mounted ADCP

5.7

- 5.1. ADCP () 2 .
- 5.2. ADCP .
- 5.3. (ex.) . (ADCP anti-foul paint) .

5.8

- 1.1.
- 1.2. , ,
- 1.3. fixed-mounted (B):
 - 1.3.1. ADCP battery (6.1)
 - 1.3.2. (, , ,)
 - 1.3.3. , mounts, shackles, tackle recovery system
 - 1.3.4. (A)

- 1.3.5. ADCP (B)
- 1.3.6. recovery line tackle
- 1.4. vessel-mounted (B):
- 1.4.1. ADCP
- 1.4.2. DGPS ()
- 1.4.3. ADCP ,
- 1.4.4. (B)
- 1.4.5. ADCP (D, E)



Figure 5.4: fixed-mounted

5.9

5.9.1

- 1.1.1. .(A)
- 1.1.2. ADCP (fixed vessel) B .

5.9.2 ADCP

- 1.2.1. ADCP , ADCP
e, v, w components rotate ADCP beam
- 1.2.2. . 6 . 90 , ADCP 4.5
. PlanADCP software duration, vertical resolution(bin size),
. ADCP 100 , . ADCP 32MB ,
. ADCP (, ,) . Marine
fixed paint (9.0) . ADCP
biofouling .

1.2.3. ADCP .

1.1.1.1. (4). ADCP () . ADCP beam 3

1.1.1.2. ADCP .

1.1.1.3. ADCP ADCP beam 3 . ADCP .

1.1.1.4. . ADCP .

1.1.1.5. O-ring Dow Corning 111 valve lubricant . O-ring

1.1.1.6. . 2 .

1.1.1.2. PlanADCP . (www.rdinstruments.com for latest details).

1.1.1.3. ADCP . , , . ADCP GMT .

(1)

. ADCP 1-2-3-4 .

. Rotate flat, 360 degrees on primary axis.

. Rotate pitch/roll. lift by 10.20 degrees up on side 3 with a non-magnetic tilting block, rotate 360 degrees on primary axis.

. Rotate roll/pitch. lift by 10-20 degrees up on adjacent side, 360 degrees on primary axis.

. Final rotation. not as critical. Rotate somewhere between (and not as much), 360 degrees on primary axis.

1.1.1.1. ErAsE .

1.1.1.2. , .

1.1.1.3. Workhorse Sentinel ADCP 2.2W 5

. ADCP 1mW . ADCP 3
(ADCPlan) .

1.1.1.4. .

7.1 .

1.1.1. (Acoustic Release) .

1.1.1.1. . .

1.1.2.

1.1.2.1.

ADCP . Benthos deck box(7-2) , deck box

, ADCP .

ADCP . ADCP , ADCP

ROV .

1.1.1.1. ADCP WinSC ADCP

PCMIA . ADCP

1.1.1.

1.1.1.1. , . biofouling ,

ADCP

1.1.1.2. . mount

. ADCP .



Figure 5.5: DCP mount



Figure 5.6: hydrophone Benthos-type deck box



Figure 5.7: ADCP mount

5.10

5.10.1

5.10.2

1.2.1. ADCP (A) , , . ADCP , B . Operation Center(OC) , .

1.2.2. ADCP . ()

1.2.3. ADCP . ()

5.10.3

1.3.1. 1.3.1.1. ADCP . ADCP 1 .

1.3.2. ADCP . (1) (2) (3) (4) (5) (6) (7) (8) (9) (10) (11) (12)

1.1.1.

1.1.1.1. , , . (retaining strap) , , . 1.1.1.3. . DC-111 .

1.1.1. 1.1.1.1. (1) (2) 4

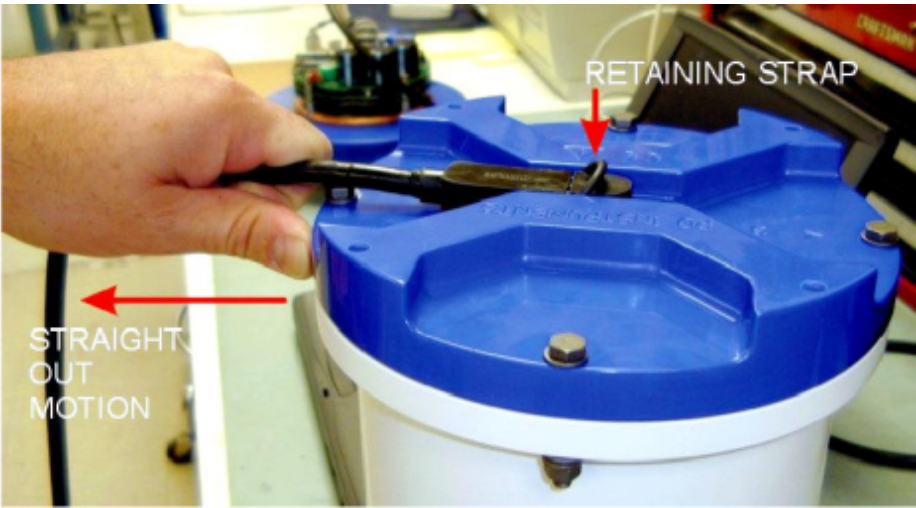
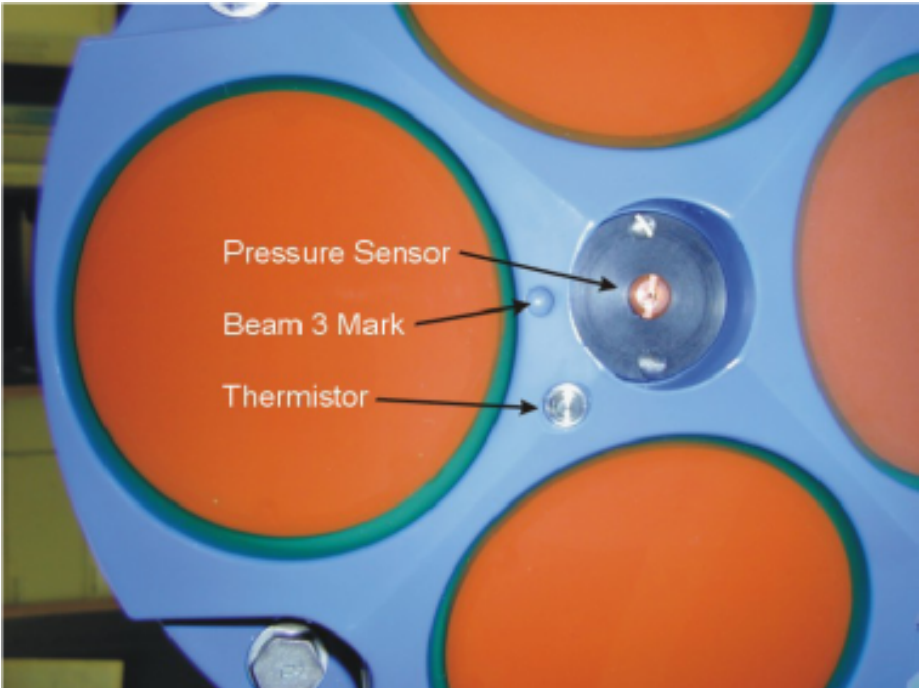


Figure 5.8:

(3) . (4) O . O
 . 1.1.1.2. (1) . (2) .
(3) . (4) . (5)
 . (6) . (7) O .
 .
1.1.2. 1.1.2.1. ADCP . 1.1.2.2. , ,
 . 1.1.2.3. ADCP . 1.1.1.1. O (1) O .
 O . O . (2) O : O
 . O . 0.1 mm . (3) O . , ,
 . (4) O (60)
 . (5) DC-111 O .
 . 1.1.1.1. (1) . (2) O , , . O .
 . (3) . (4) . 3
 . (5) . 1.1.1.2. (1) .
1.1.2. 1.1.2.1. 30 VDC .
1.1.3. 1.1.3.1. .
1.1.4. 1.1.4.1. POI .
1.1.5. 1.1.5.1. . RS-232 RS-422 .
 .
1.1.6. 1.1.6.1. . 7.2 .
 8.4 9 . 250° 14 .
1.1.7. 1.1.7.1. , PC , , .

1.1.8. 1.1.8.1. 100 m .

1.1.9. 1.1.9.1. .



1.1.1. 1.1.1.1. .

1.1.1. ADCP 1.1.1.1. ADCP 8.4 . ADCP
1.1.1.2. BBTalk 1.1.1.3. ADCP
WH_RSLTS.txt WH_RSLTS.tx BBTalk .

1.1.1.1. (B)

1.1.1.2. (B)

5.11

5.11.1

1.1.1. ADCP 4 3 (u, v, w) . (,
20) , / w (u(-), v(-)) . ,
vessel-mounted ADCP . ,

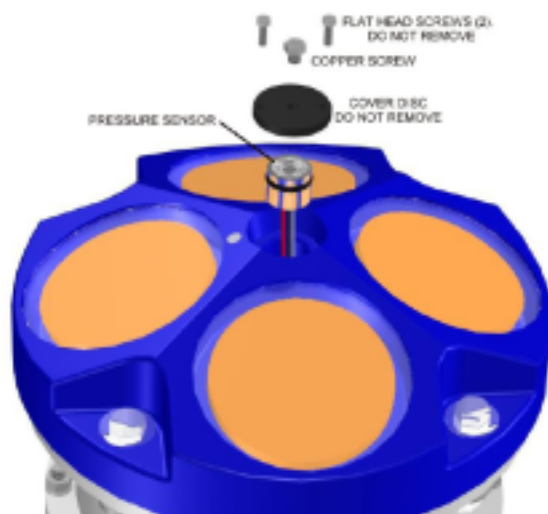


Figure 5.9:

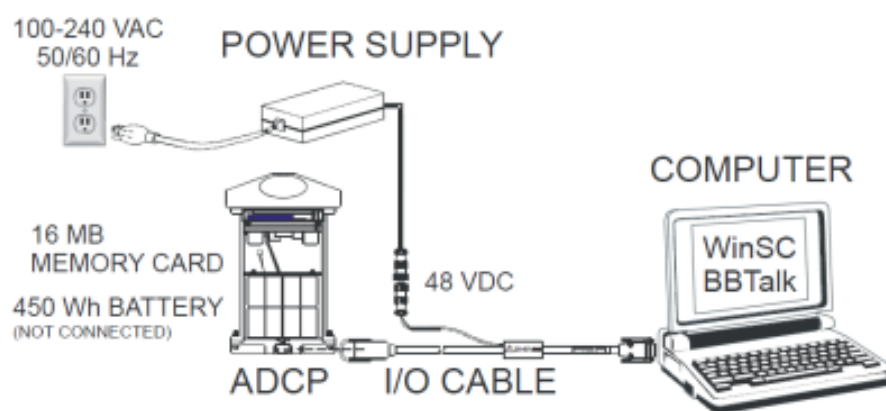


Figure 5.10: ADCP

5.11.2

1.2.1.

ADCP

5.12

1.1. Environmental Assessment Program(EAP) Safety Manual
 , , EAP safety manual 3 . (, ,
)

1.2. MSDS , , , (MSDS Trilux White = http://www.yachtpaint.com/msds_pdf/YBA068_GBR_ENG.pdf). EAP HQ safety Manual .

5.13

1.1. Environmental Assessment Program, 2006. Environmental Assessment Program Safety Manual. March 2006. Washington State Department of Ecology. Olympia, WA.

1.2. WorkHorse Sentinel ADCP User's Guide, P/N 957-6163-00 (January 2001), RD Instruments Acoustic Doppler Solutions (Appendix F).

5.14 1. (Fixed-mounted ADCP)

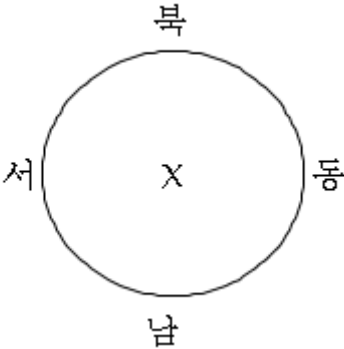
LOG SHEET FOR ADCP BOTTOM DEPLOYMENT

:
 : :
 : :

ADCP :
 ADCP :

: :
 :

ADCP (X)



:
: :

: (m):
Acoustic Release(model 875A) Serial number:
Frequency(Hz):
Release code:
Pinger? /
Snag-line? /
Timed-release? /

5.15 2. (Vessel-mounted ADCP)

LOG SHEET FOR ADCP DATA COLLECTION

:
: :
: :

Transect Number:
Transect Name:
:
Configuration :
: : :

: (m):
Select Bank: Left / Right Transect Direction (heading):
:
:

: (m):

Select Bank: Left / Right Transect Direction (heading):

:
:
:

5.16 3.

(Bottom-mounted ADCP) Office & Electronics

- Laptop Computer (w/ power cord)
- Data stick for backing-up data
- Metal Clipboard
- Float Plan x3 (Section Secretary, Contact, Field)
- Sample Logs
- Tide Table
- Cell Phone (w/ charger)
- Digital Camera
- MSDS

OC Field Supplies

- Data cable and power cord for 110-volt ship power (or battery)
- Life Jackets
- Power winch with footswitch
- Buckets
- Mount with ADCP (and new batteries)
- Acoustic release (batteries), deck unit and hydrophone
- Pinger (batteries)
- Chain, tackle, anchor ball for line canister
- Viny float, and line
- Special line that allows release of float from either acoustic or time signal
- Hefty bag to seal line cylinder
- Buoyant (floating) yellow snag-line (50 m, > ½-inch, 5000# test)
- Hose clamps (or nylon ties), and spares
- Boat hook
- ADCP Rubbermaid Action Packer, Large
- Tool Box (content details listed below)
- Cable to interface with DGPS

From Wet Lab

- *Plastic Bags
 - *Label, Electrical and Duct Tape

- *Sharpies, Pencils and Pens
- *Connectors
- *Bottle Brushes & Paper Clip (straightened)
- *Scissors
- *Dummy plugs for ADCP
- *Nitrile Gloves, Med and Large
- *Extra Plastic Beaker
- *Kimwipes
- Deck unit and hydrophone (for recovery)

Items packed morning of departure

Contents of Bins

Large ADCP Action Packer

- Hose Clamps and/or large nylon ties
- Electrical Tape
- Duct Tape
- Zip-Ties, various sizes
- Spare Parts
- Gloves, Nitrile
- Sandwich Box with Allen Wrench
- Kimwipes
- Misc Hardware (screws, nuts, washers, insulators, Teflon tape)
- Plastic Beaker
- Brushes
- Teflon and Wooden Block

Toolbox

Screwdrivers

- Large flathead
- Small flathead
- Large Philips
- Stubby Philips

Pliers

- Large diagonal cutters (for large zip-ties)
- Small diagonal cutters (for small zip-ties)
- Needle nose
- Locking

Wrenches

- Adjustable
- Ratchet w/ drivers (various)
- Combo wrenches (various)
- Allen wrenches
- Nut drivers (various, 7 mm for heavy duty hose clamps)
- Hammer
- Electrical tape
- Knife
- Extra ADCP plugs
- Spare nuts, bolts, and ties

5.17 4.

(Vessel-mounted ADCP)

Office & Electronics

- Laptop Computer (w/ power cord)
- Data stick for backing-up data
- Metal Clipboard
- Float Plan x3 Section Secretary, Contact, Field)
- Sample Logs
- Tide Table
- Cell Phone (w/ charger)
- Digital Camera
- MSDS

OC Field Supplies

- Data cable and power cord for 110-volt ship power (or battery)
- Life Jackets
- Buckets
- Mount for ADCP to vessel (w/lanyard)
- Boat hook
- ADCP Rubbermaid Action Packer, Large
- Tool Box (content details listed below)
- Cable to interface with DGPS (if desired)

From Wet Lab

- *Plastic Bags
- *Label, Electrical and Duct Tape
- *Sharpies, Pencils and Pens
- *Connectors
- *Bottle Brushes & Paper Clip (straightened)
- *Scissors
- *Dummy plugs for ADCP
- *Nitrile Gloves, Med and Large
- *Extra Plastic Beaker
- *Kimwipes

Items packed morning of departure

Contents of Bins Large ADCP Action Packer (some items duplicative of toolbox)

- Hose Clamps and/or large nylon ties
- Electrical Tape
- Duct Tape
- Zip-Ties, various sizes
- Spare Parts
- Gloves, Nitrile
- Sandwich Box with Allen Wrench
- Kimwipes
- Misc Hardware (screws, nuts, washers, insulators, Teflon tape)
- Plastic Beaker
- Brushes
- Teflon and Wooden Blocks
- Sponges

Toolbox

Screwdrivers

- Large flathead
- Small flathead
- Large Philips
- Stubby Philips

Pliers

- Large diagonal cutters (for large zip-ties)
- Small diagonal cutters (for small zip-ties)
- Needle nose
- Locking

Wrenches

- Adjustable
- Ratchet w/ drivers (various)
- Combo wrenches (various)
- Allen wrenches
- Nut drivers (various, 7 mm for heavy duty hose clamps)
- Hammer
- Electrical tape
- Knife
- Extra ADCP plugs
- Spare nuts, bolts, and ties

Chapter 6

()

6.1

(NO₂-) (Continuous flow analysis, CFA) (NO₃-) 550nm
 Sulfanilamide NEDD Method of Seawater
 Analysis(1999, 3) Mg(OH)₂ NaOH

6.2

20 (, ,) 110°C ()
 sulfanilamide NEDD CFA -
 550nm
 ((1)). - ((2)).
 () ((3)).
 () ((4)). 550 nm

- (1) $NO_2^-N, NO_3^-N, NH_3^-N$, organic- N , particulate form
 + Alkaline oxidizer ($K_2S_2O_8 + H_3BO_3 + NaOH$) $\rightarrow NO_3^-N$
- (2) $NO_3^- + Cd + 2H^+ \rightarrow NO_2^- + Cd^{2+} + H_2O$

- (3)
- (4)

SFA (2, 2006) (Segmented flow analysis, SFA)
 0~6 mg N/L(0~43 mol N/L) , 0.2 mg N/L(1.43 mol N/L)
 7 0.02 mg/L(0.14 mol N/L) . 5 mg N/L(35.7 mol N/L) 10
 (CV) 3%

6.3

U.S. EPA Method 353.2 .

6.4

(4.1), , , (4.2),
(4.3), (4.4), TrAAcs800 system(4.5) . 10%
3 .

6.4.1

NIST Class A 0.05% .
2-3K (Kolthoff et al., 1969; Weast,
1985).
polymethylpentene (PMP),
polypropylene(PP) 3% .

6.4.2

10% (5.1.5.1) 3 (: HDPE, PP)
.

6.4.3

0.01 mg 0.001 g .

6.4.4

0.1%
.
/
/
/

6.4.5

SEAL TrAAcs800 (D2BX-02) ,
(1.3
kgf/cm²(0.13MPa)) .

6.5

(5.1), (5.2), (5.3), (5.4) .
₁ .
 < 1. >

6.5.1

(5.1.1), (5.1.2),
 (5.1.4, 5.1.5) .
 1.1.1. (Ultra Pure Water, UPW): (Reverse Osmosis Membrane, RO
 membrane) 18.2 MΩ .
 18.2 MΩ
 () .
 , (Electrodeionization
 module, EDI module) .
 1.1.2. (Artificial Seawater, ASW): .
 1.1.2.1. Sodium chloride(NaCl) - 35 g 1.1.2.2. Sodium hydrogen carbonate(NaHCO₃)
 - 0.5 g 1.1.2.3. 1000 mL () Sodium chloride 35 g, Sodium
 hydrogen carbonate 0.5 g 800 mL 1000 mL .
 1.1.3. 50% Triton X-100 : Sigma Triton X-100 Isopropanol 50:50 .
 100 mL 1 .
 1.1.3.1. Triton X-100(Sigma) - 50 mL 1.1.3.2. Isopropanol or Ethanol - 50 mL
 1.1.3.3. Triton X-100 50 mL Isopropanol(Ethanol) 50 mL 100 mL .
 1.1.4. Wash-Triton X-100 :
 . 1000 mL 50% Triton X-100 (5.1.3) 2 mL .
 1.1.5. 1N (1N NaOH): (1) , .
 1.1.5.1. Sodium hydroxide(NaOH) - 20 g 1.1.5.2. 300 mL 500 mL Sodium
 hydroxide 20 g 500 mL . 500mL

6.5.2

(, ,) , , , .
 1.1.1.
 1.1.1.1. Potassium peroxodisulfate (K₂SO₈) - 25 g 1.1.1.2. Boric acid (H₃BO₄)
 - 10 g 1.1.1.3. Sodium Chloride (NaCl) - 1 g 1.1.1.4. Sodium Sulfate (Na₂SO₂)
 - 1 g 1.1.1.5. 5N Sodium hydroxide solution (5N NaOH) - 4 mL 1.1.1.6. 1000
 mL 5.2.1.1 ~ 5.2.1.5 800 mL 1000 mL

1.1.2. 5N

1.1.2.1. Sulfuric acid(ACS grade, Analar Pro Analysis) - 70 mL 1.1.2.2. 500 mL 300 mL Sulfuric acid 70 mL 500 mL .

1.1.3. Imidazole buffer

1.1.3.1. Imidazole - 30 g 1.1.3.2. Sulfuric acid(ACS grade, Analar Pro Analysis) - 4 mL 1.1.3.3. Triton X-100, 50% - 1 mL 1.1.3.4. 1000 mL Imidazole 30g 800 mL 4 mL 1000 mL . Triton X-100 50% 1 mL .

1.1.4. Sulfanilamide

1.1.4.1. Sulfanilamide - 5 g 1.1.4.2. Hydrochloric acid (ACS grade, Analar Pro Analysis) - 50 mL 1.1.4.3. 500 mL Sulfanilamide 5g 300 mL 50 mL 500 mL .

1.1.5. NEDD

1.1.5.1. N-(1-naphthyl)ethylenediamine Dihydrochloride ($C_{12}H_{14}N_2 \cdot 2HCl$) - 0.5 g 1.1.5.2. Hydrochloric acid (ACS grade, Analar Pro Analysis) - 5 mL 1.1.5.3. 500 mL NEDD 0.5 g 300 mL 5 mL 500 mL .

6.5.3

, Wood et al.(1967) - (open tube cadmium reactor, OTCR)) - Wood et.al(1967) 0.5-2 mm

(Zhang et. al., 2000).

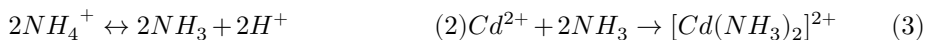
(carry-over effect) Patton (1982) open tube cadmium reactor(OTCR)

(Nydahl, 1976; Olson, 1980; Collos et al., 1992; Garside, 1993). Zhang et.al.(2000)

OTCR OTCR OTCR 40 M 10-15 100% OTCR

(Grasshoff, 1964; Strickland and Parsons,1965) Imidazole(Nydahl, 1976) pH 8.5

pH



(3). (2) (2)
 . Rho et al.(2015) OTCR
 pH .

1.1.1. Stock Copper Sulfate Solution 2% 1.1.1.1. Copper Sulfate($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$)
 - 10 g 1.1.1.2. Copper Sulfate 10 g 500 mL 300 mL 500
 mL .

1.1.2. Daily Working Imidazole buffer(5.2.5)

6.5.4 (Standard Solution)

1.1.1. Stock Nitrate Standard Solution(10,000 M)

1.1.1.1. Potassium Nitrate(KNO_3) 0.255 g 105~110°C
 2 . 1.1.1.2. Potassium Nitrate 0.2550
 g 0.01 mg 0.001 g 0.00001 g .
 () . 1.1.1.3. 250 mL
 0 . 150 mL 150
 mL . Potassium Nitrate 250 mL
 . 1.1.1.4. . ,

1.1.2. Daily NO_3^- (Calibration Standards)

1.1.2.1. Stock nitrate standard solution(5.4.1) 5 ,
 . 1.1.2.2. , (Std.A, 60 M) . 1.1.2.3. 500 mL
 0 . 1.1.2.4. (5.4.1) 3 mL 500 mL (5.1.2)
 500 mL . 500 mL (mol/kg).
 (mol/L, M) (35‰) . 1.1.2.5.
 AACE Method Calibrant .

1.1.2.6. .

6.6

1.1. . 10% (High
 Density Polyethylene, HDPE) (Polypropylene, PP) .
 15 mL Sampler rack .
 . 1.2. ()
)
 CTD .
 . 1.3.
 0.2 m PES(Polyethersulfone)
 membrane . HDPE PP 500 mL 0.2
 m PES . 50 mL 3 . 1.4.
 8 . 8

(-80 °C) .

6.7

(5.4.2) . 2

6.8 (Quality Control and Quality Assessment, QA/QC)

6.8.1

(Accuracy) (Precision) (Quality Control, QC), (Quality Assessment, QA), (Accuracy), (Precision) .
Dickson(2007) “Guide to Best Practices for Ocean CO₂ Measurement”

1.1.1. (Quality Control)

, , , .

1.1.2. (Quality Assessment)

1.1.3. (Accuracy) (Bias) (Precision) .

$$= [- /] \times 100$$

1.1.4. (Precision) (random error) ,

. (absolute standard deviation), (relative standard deviation, RSD), (variance), (coefficient of variation, CV), (relative percent difference), (absolute range of a series of measurement) . RSD

$$\% \text{ RSD} = [(s) /] \times 100$$

1.1.5. (Method Detection Limit, MDL)

. , , . EPA 99%
0 99% 1% 0
(). , , , .

, , .

8.6 .

6.8. (QUALITY CONTROL AND QUALITY ASSESSMENT, QA/QC)65

6.8.2 (Standard Operating Procedures, SOP)

・ ,
・
・
・ , ,
・ ()
(gain)
・ Seal AACE
・ Water check, Reagent check , detector energy
Drift correction, Baseline correction,
Recovery detection

6.8.3 (Internal Checks)

1.3.1.
1.3.2. () (~1000) () ,
10L 1 mL
1 ()
() (1%)
1.3.3. Van Ooijen and Bakker(1992)
CFA
0.1%
60~80% (1
10)

6.8.4 (External Quality Checks)

(Certified
Reference Materials, CRM), (Reference Materials, RM)
(,),
(GO-SHIP,CLIVAR, GEOTRACES)
1.4.1. : KANSO Technos (RMNS)
SCOR Nutrient WG147 JAMSTEC 5 (2 3)

100 mL . Eurofins . 1
 . SCOR-JAMSTEC KANSO (SI) . , ,
 Chemicals Evaluation and Research Institute(CERI) Japan Calibration
 Service System(JCSS), National Metrology Institute of Japan(NMIJ)
 . Merck KGaA National Institute of Standards and
 Technology(NIST) (SRM3150) . GO-SHIP
 SI . 1.4.2. / :

. Scripps
 Institute of Oceanography Royal Netherlands Institute for Sea Research
 2 .

. ()
 . ()
 () .
 / () () , .

6.8.5

, (Quality Control, QC) .
 .
 1.5.1. () , , .
 + () , , , .
 1.5.2. . GO-SHIP (CLIVAR) WOCE
 CCHDO(cchdo.ucsd.edu) . GLODAP .

6.8.6

1.6.1. : .
 .
 1.6.2. : . () .
 . ,
 .
 1.6.3. : 7 .
 1~5 .
 10 1 . , .
 .
 1.6.4. < < 10* .

6.8. (QUALITY CONTROL AND QUALITY ASSESSMENT, QA/QC)67

1.6.5. : 7 . 8 .
 (, ,) . 7 1 .

(,) 7 .
 1.6.6. : 1 .
 . ,

1.6.7. : . batches 7
 , .

1.6.8. : (1) (s) , (2) Students t-value , (3)
 (s) () .

Sample standard deviation (s):

$$s = \sqrt{\frac{\sum (x - \bar{x})^2}{n - 1}}$$

\bar{x} : sample mean; n-1: degree of freedom;

Population standard deviation ():

$$\sigma = \sqrt{\frac{\sum (x - \mu)^2}{n}}$$

μ : population mean; n: number of population

$= t_{(n-1, 1-\infty=0.99)} \times s$
 $t_{(n-1, 1-\infty=0.99)}$: 99% n-1 students' t value
 s :

95% chi (X2/df) .

LCL = 0.64* MDL

UCL = 2.20* MDL

LCL UCL 7 95% .

0.15 0.2 .

1.6.9. S/N : / 2.5-5 . S/N /
 2.5 . / 10 .
 . / 10 .

$$(S/N)_{\text{est}} = X_{\text{ave}}/s$$

X_{ave} :

s :

1.6.10. : (1).

1 . 5 .

(1) 10 ? .

(2) ? .

(3) ?

(4) Signal/Noise (S/N) ?

(5) ?

1.6.11. : 0.05 mg/L .

1, 5, 10 mg/L, . 1 . 0.05 mg/L 5 0.25 mg/L
 . 0.05~0.25 mg/L . MDL_cal_example.xlsx .

6.8.7

1.7.1. : .

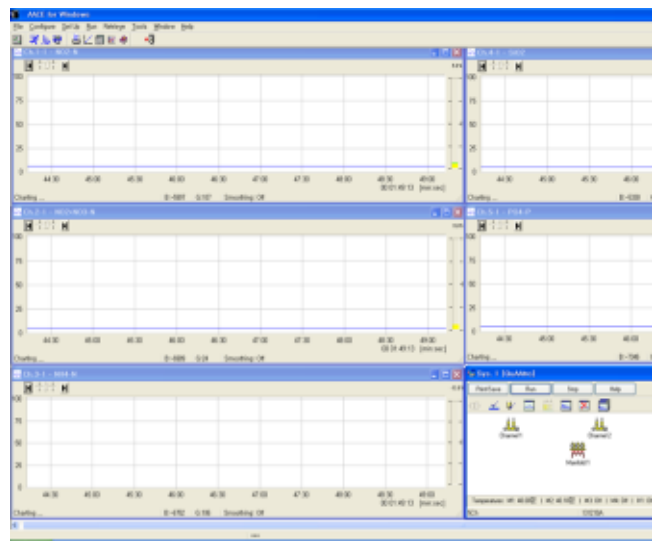
-
-
- , , ,
- , ,
-
- , ,
-
-
-
- /
-
- ()
- ()
- () ,
-

1.7.2. : .

6.9

6.9.1

1.1.1. 1.1.1.1. TrAAcs 800, sampler, , , . 1.1.1.2.
 system wash solution(: wash triton-X, : wash SDS, :) Auto



1.1.2.3. Charting 2 (,) .

1.1.2.4. smoothing 4, smoothing 12 .
smoothing . Smoothing

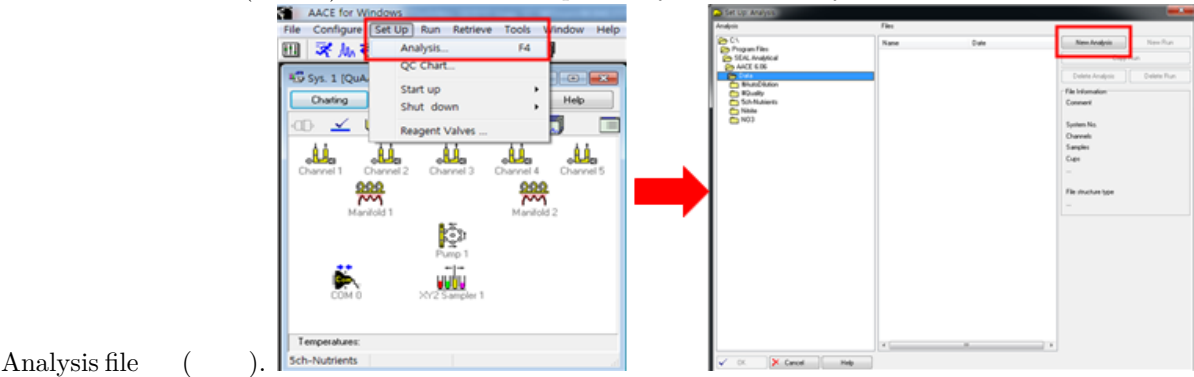
1.1.3. System Wash solution(5.1.4)() sampler sample
wash solution . 110°C ,
baseline $\pm 0.1\%$. 1 . baseline 6~20%
9.2.2.1 baseline .

1.1.4. 1.1.4.1. standard stock(5.4.1) .
. 1.1.4.2. (5.2) , 5.4 5 .

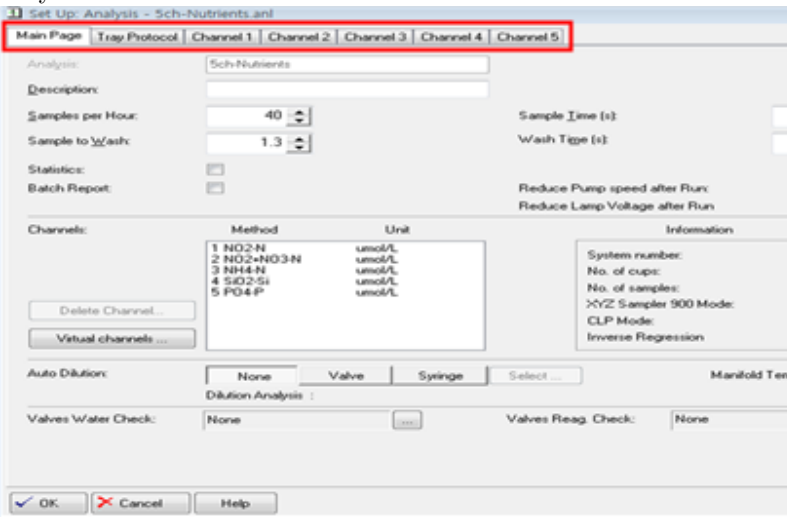
1.1.5. 1.1.5.1. .
. 10 mL 1 50 mL 2~3
. 1.1.5.2. 24 .
24 () . - 15 mL - -80 °C -

6.9.2

1.2.1.1. TrAAcs 800 (AACE) 1.2.1.1. Set up - Analysis - New analysis

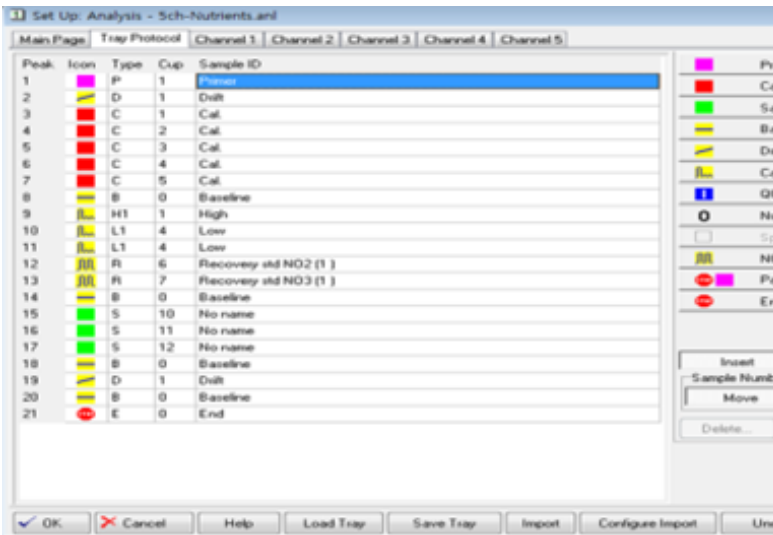


1.2.1.2. New analysis Set Up: Analysis-new****.anl



Main Page, Tray Protocol, Channel 1~2

1.2.1.3. Main Page : Sample/Wash time .
Sample/Wash time 2 /2 . peak ISAC
. Peak ISAC (4) .



1.2.1.4. Tray Protocol :

- primer: primer peak peak tray protocol
- calibrants: Channel (9.2.1.5)
- drift:
- carry over: high-low-low low-low-high
- baseline: baseline peak baseline ,
- Sample: 12 drift
 - MDL : 5 (1.2 M, 0.2 M). 7 (8.7).
 - NO3 Recovery: 5 M NO2, NO3 - 95%
$$= \frac{5\mu M \text{ } NO_3}{5\mu M \text{ } NO_2} \times 100(\%)$$

gain std A peak (%) . peak , gain
 - QC: 3 (8.2.2). 3 (8.2.3).
 - Cup: sampler rack sampler rack . cup sampler

1.2.1.5. Channel : channel channel (method), baseline, drift, carryover correction . (calibrants 1~10),

fitting , (noise) smoothing, peak determination window time
 . Smoothing 4, smoothing 8 .

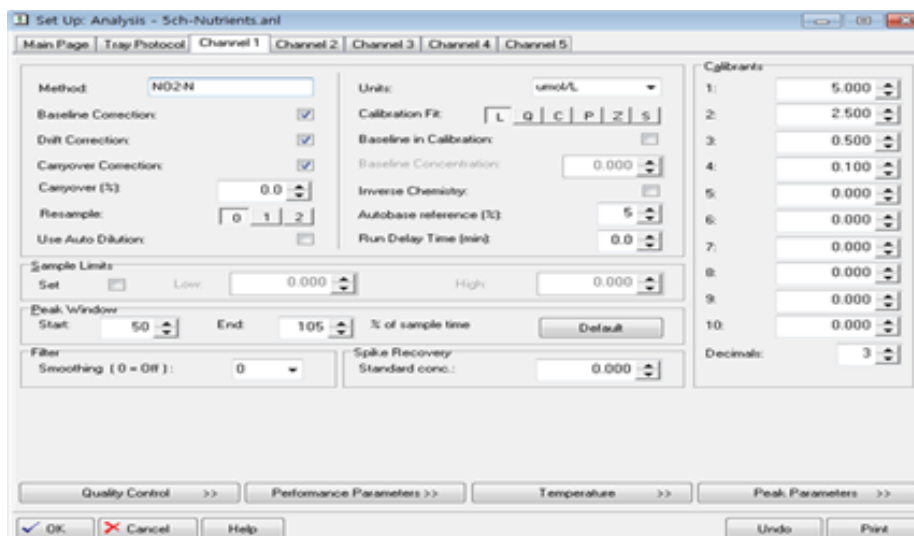


Figure 6.1:

1.2.1.6. OK Main analysis (.anl) . Set
 up - Analysis Analysis file system sample
 table .

1.2.2. baseline

1.2.2.1. 9.1.3 6~20% channel baseline 6~20%
 (UPW baseline). . 1.2.2.2. System wash solution(wash triton-
 X, wash SDS, UPW) . 1.2.2.3. Imidazole
 - valve - . 1.2.2.4. ,
 baseline (Reagent blank, RB). baseline 6%~20% channel 1(),
 channel 2() baseline . 1.2.2.5. Gain : std A (: 60
 M, : 10 M) 2 2 . 20 peak channel
 peak 80~85% gain . baseline gain ,
 baseline gain .

1.2.3.

1.2.3.1. 9.2.2 sample table rack(3) . rack holder rack
 sample probe . 1.2.3.2. run (9.2.1) .
 1.2.3.3. baseline . baseline

1.2.4.

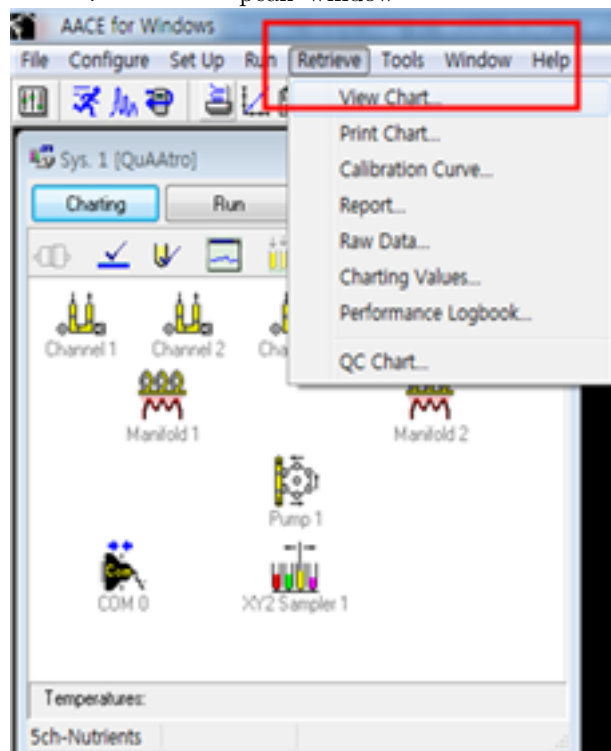
1.2.4.1. - . manifold .

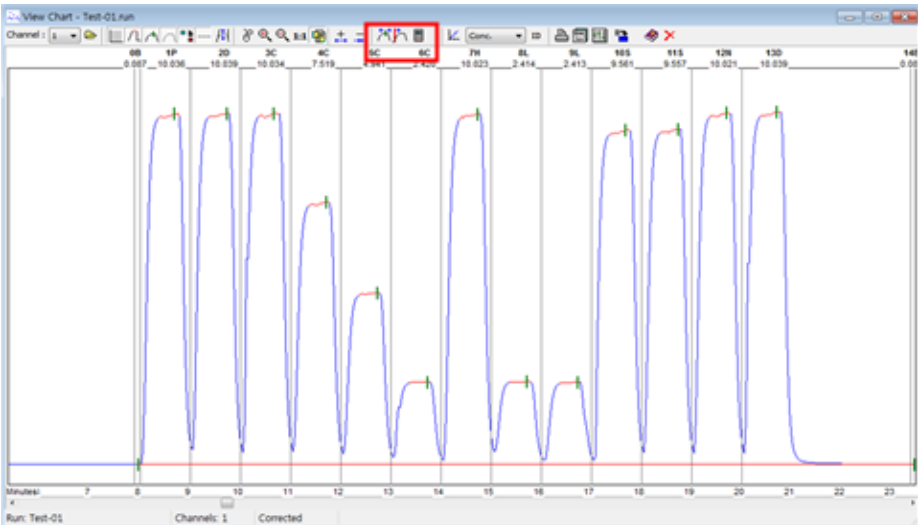
1.2.4.2. 1 manifold . 1.2.4.3. stop
 . 1.2.4.4. / (: -
 10% - 3 -).

6.10

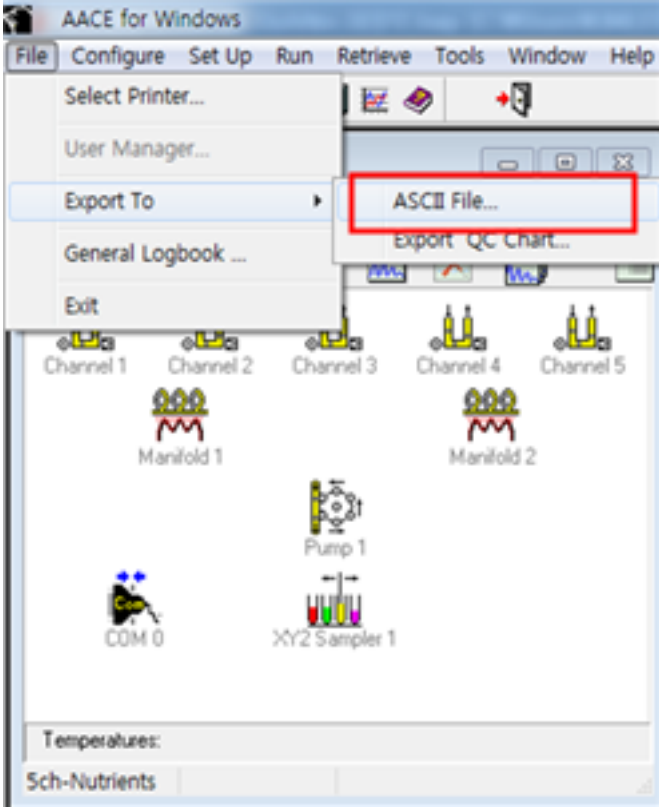
1.1. retrieve - view chart

peak window

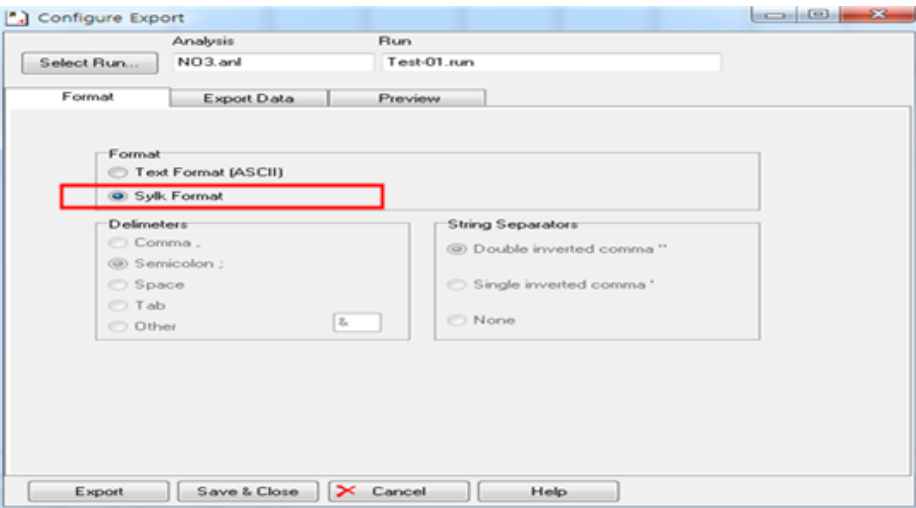




1.1. [file] - [export] - [ASC file] - chart , sylk format .



excel .



1.1. .

6.11

- 1.1. , , , () - , , Vol. 9, No. 2, p.199-206, 2006.
- 1.2. , 2021, p.97~104.

6.12 1. (TrAAcs 800)

6.12.1

(CFA, Continuous Flow Analysis) ,
flowcell .

6.12.2

- 1.2.1 Console(,)
- 1.2.2 2 proportional pump 10 .
platen 12 12 .
- 1.2.3 1.2.3.1 Flow rate . color code
. flow rate color code . 1.2.3.2 Color Code . 1.2.3.3
:
150~200 .
- 1.2.4 (Manifold) , , .
- 1.2.5 1.2.5.1 30mm Flow cell 2 550nm, 800nm , .
- 1.2.5.2 Software (Gain) Baseline .

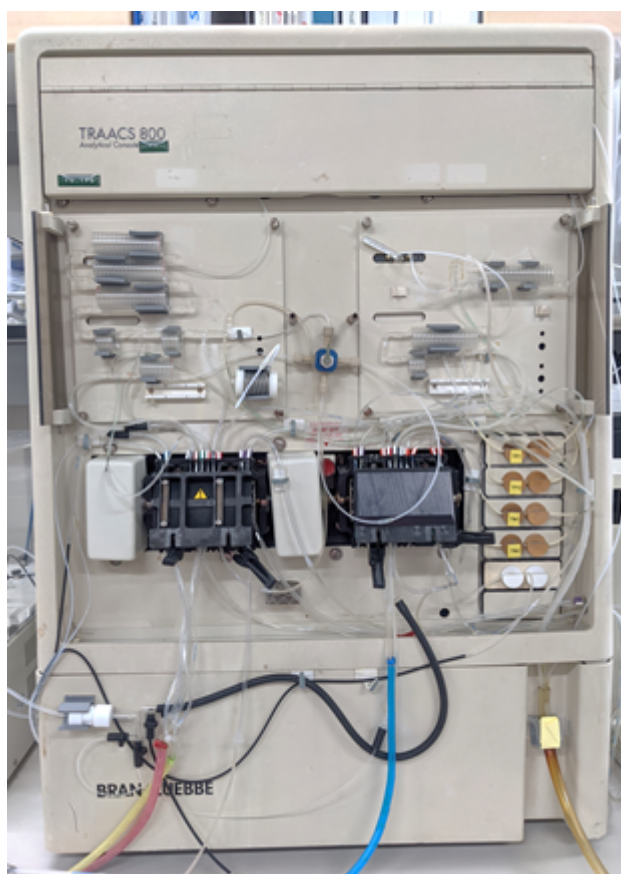


Figure 6.2:

6.12.3 Sampler

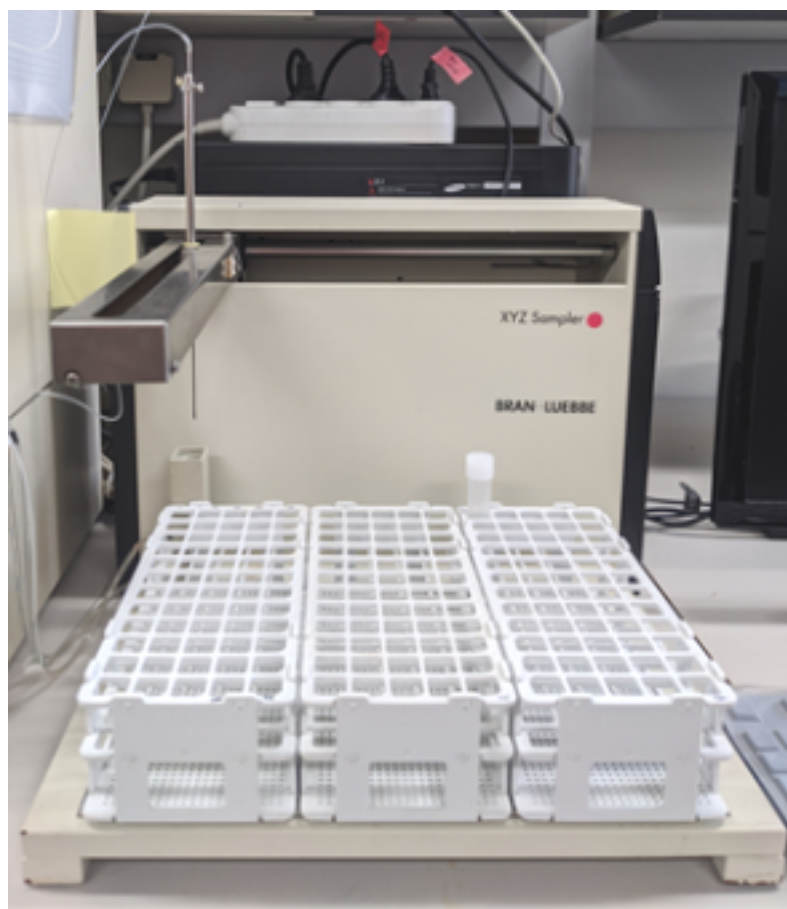


Figure 6.3:

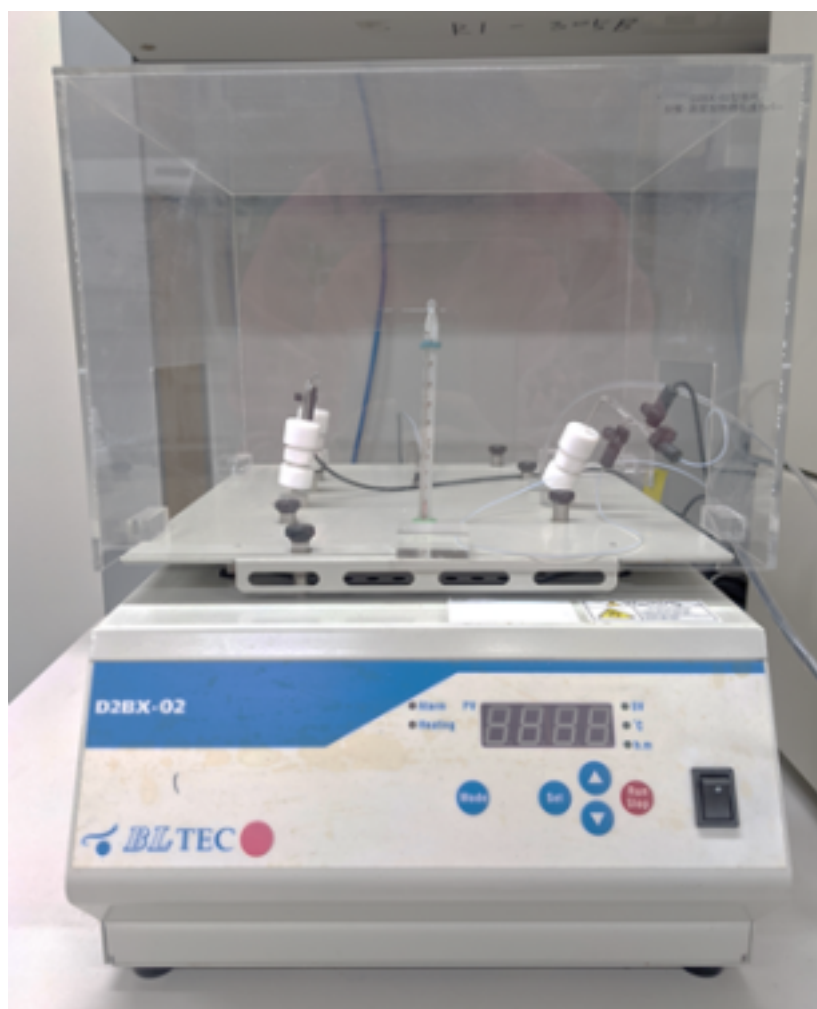


Figure 6.4:



Figure 6.5:

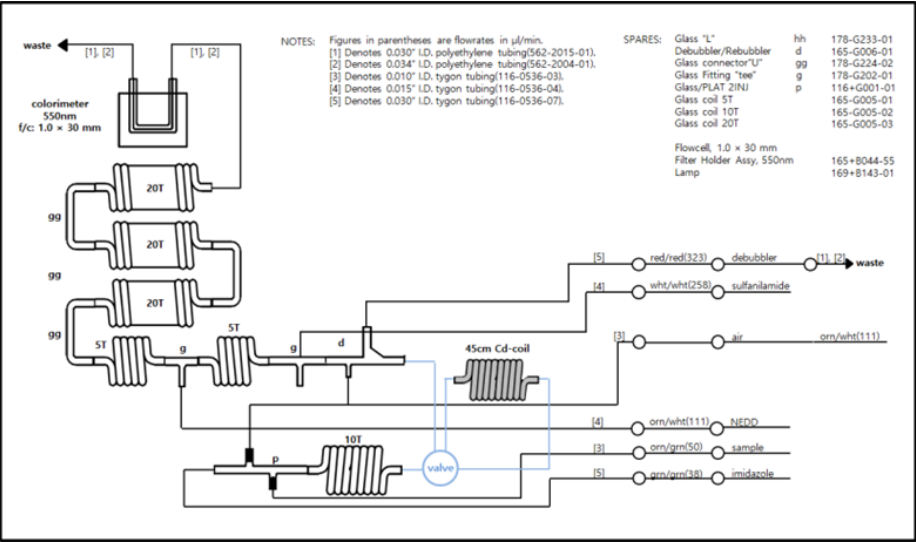


Figure 6.6:

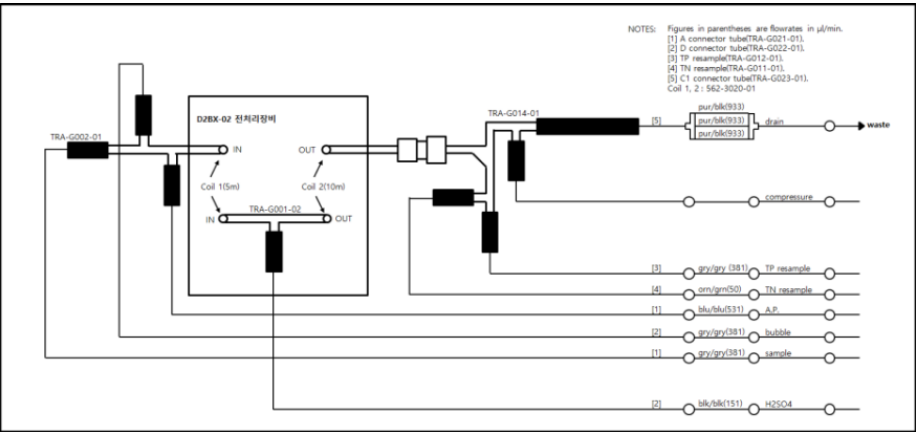
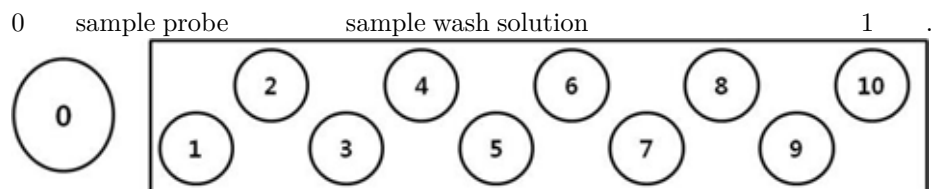


Figure 6.7:

6.12.4 (D2BX-02)**6.12.5 compressure****6.13 2. Flow Diagram****6.13.1****6.13.2****6.14 3. Sample rack position****6.14.1 Sample rack position****6.14.2 Sample rack position****6.15 4. (Inter Sample Air Compression, ISAC)**

(ISAC)

,

ISAC (+) 2~8 (3) . ISAC
 60 , 3:1 . :45 :15 : 60
 52~58 (57) .

2 . (2) .

. Double ISAC = + + = 57 + 45 + 15 = 117

6.16 5. Coating Procedure of Cd coil**6.16.1 (5.3)**

5.1.1 10% Acetone 5.1.2 10% HCl 5.1.3 2% Copper sulfate 5.1.4 Imidazole buffer

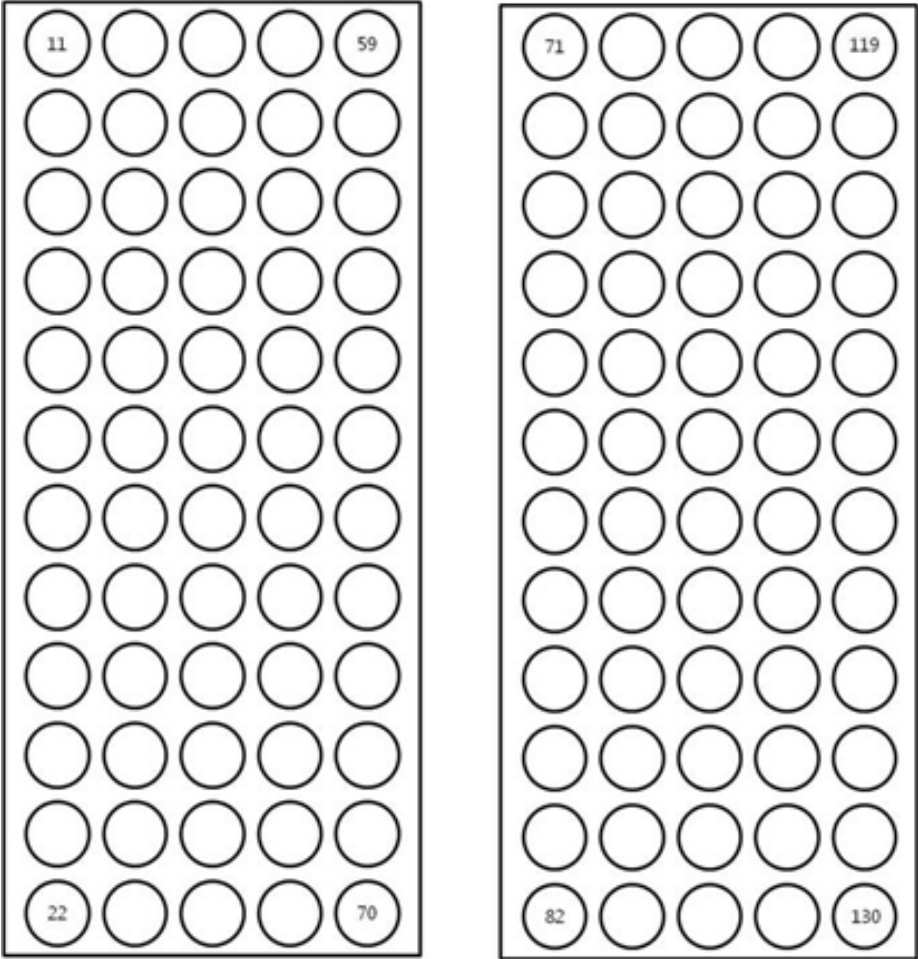


Figure 6.8:



Figure 6.9:

6.16.2

5.2.1 50 mL 10% HCl, 200 mL . 5.2.3 10 mL . 5.2.4.
 10 mL . 5.2.5. 1, 2 .
 . 5.2.6 10 mL Imidazole buffer 10 mL
 . 5.2.7. 10 mL 15 5 mL, 3 mL, 2 mL .
 . 5.2.9. 10 mL Imidazole buffer 10 mL
 . 12 . 5.2.10. 10% 12 mL
 5.2.3 .

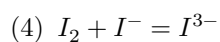
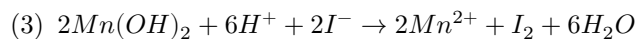
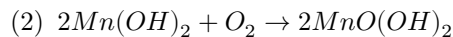
Chapter 7

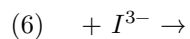
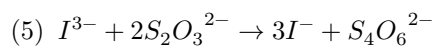
7.1

Winkler(1888) (I^-) (I^{3-}) (I^{3-}) (2013-230)
 , , .

7.2

$(MnCl_2)$ $(NaOH/NaI)$ (Mn^{2+})
 $(Mn(OH)_2)$ (1). 4 $(MnO(OH)_2)$ (I^-)
 (2). 4 $(MnO(OH)_2)$ (I^-) (I^{3-}) (4).
 (I_2) (3). (I_2) (I^-) (I^{3-}) (I^-) .
 (I^{3-}) (I^-) (I^{3-}) (I^{3-}) (5).
 (I^{3-}) (I^-) (I^{3-}) (6).
 (I^{3-}) (I^-) . 2 1 (I^{3-}) $1/2$
 1 2 (I^{3-}) , 4 $(MnO(OH)_2)$ 1 (I_2) . (O_2)
 (I_2) 1 (I_2) (O_2) $1/2$. (O_2) (I^{3-})
 (I^-) 4 .





0-400 mol kg⁻¹ . 0.1% ±0.3 mol kg⁻¹ .

7.3

.

7.4

(4.1), (4.2) . 10% 3 .

7.5

7.5.1

±0.003 ml . 125 ml Pyrex .

7.5.2 /

3 . 1(MnCl₂), 2(NaI/NaOH), 3(Sulphuric Acid) 3 1.0 ml ±0.02
ml . 2(NaI/NaOH) , / .
1(MnCl₂) 2(NaI/NaOH)

.

7.5.3

(Niskin bottle) (Tygon tube) .
Tygon tube .

7.5.4

0.1 oC .
mol L⁻¹ mol kg⁻¹ .

7.5.5

300g 0.001g .

7.6

7.6.1

， ， 。

7.6.2

1 ml, 2 ml, 5 ml 。

。(; Metrohm 665 Dosimat burette, SCHOTT Instruments TITRONIC universal)

7.6.3

(KIO₃) 1.0 ml 10.0 ml 。 SOCOREX

Calibrex 520 bottle-top 。 1-11 ml 0.25 ml 。

±0.002-0.005 ml 。 MetrohmTM 。

7.6.4

25mm 。

7.6.5

250 ml 。

7.7

(5.1), (5.2), (5.3), (5.4), (5.5). (5.6) 。

7.7.1 3M (MnCl₂ · 4H₂O)

(MnCl₂ · 4H₂O) 600 g 500-700 ml 1000 mL 。

(,) 。

(MnSO₄ · 4H₂O) 480 g 1000 ml 。

7.7.2 (NaI/NaOH)

(NaOH) 320 g 500 ml 。

(NaI) 600 g 。

(NaN₃) 10 g 。

1000 ml 。

(,) 。

7.7.3 50% (v/v) (H₂SO₄)

(H₂SO₄) 50 mL 1:1 (50 ml 50 ml) .
 50 ml . 100 mL . (,) .
 , .

7.7.4 1%

1 g 50 mL 100 mL .
 . 1 , (Hg₂I₂) .

7.7.5 0.025 N (Na₂S₂O₃ · 5H₂O)

(Na₂S₂O₃ · 5H₂O) 6.205 g 1000 mL .
 . . 400 mol kg⁻¹
 2 ml .

$$4 \times (C_1 \times V_1) = C_2 \times V_2$$

,
 $C_1 : [O_2] = 400 \mu mol L^{-1}$
 $V_1 : (10\%) = 14 ml$
 $C_2 : [Na_2S_2O_3]$
 $V_1 : (2ml)$
 4:1 (O₂) 4 Na₂S₂O₃

,
 $[Na_2S_2O_3] = \frac{400 \mu mol L^{-1} \times 140 ml \times 4}{2 ml} \simeq 0.11 M$
 $[Na_2S_2O_3] = 400 \times 10^{-6} \times 140 \times 4 / Burette volume(2ml) = 0.11 M$

(Na₂S₂O₃ · 5H₂O) 1 248.17 g 27.4 g 1000 ml .
 (Na₂S₂O₃) 1 158.09 g 17.4 g . 2-5
 1L .

7.7.6 0.001667 M (KIO₃, 0.0100 N)

(KIO₃) 0.5 g 120°C 2 . 0.3567g 1000
 mL . (tp) .
 0.3567g , .

$$M() = \frac{1L}{(214.0g)}$$

7.8

7.9

Noble gases(aragon and zenon), O¹⁷, Oxygen, and pCO₂ . CFCs, Helium,

7.10

1.
2. ()
3.
4.
- 45
- 2-3
5.
- CTD
6. 1 ml 1 ml 90
- NaI/NaOH
7. 2 ml
8.
9.
10. 30
11. 1 30 2

7.11

7.12

1.
2. (KIO₃) 1 ml
3. 50 % 1ml
4. NaI/NaOH 1ml
5. MnCl₂ 1 ml
6.
7. (V1)
- V1 V3
8. (KIO₃) 1 ml (V2)

7.13 () by Carpenter (1965) method

1. .
2. .
3. 10.0 ml (0.00167 M) .
4. 50% 1 ml .
5. (NaI/NaOH) 1 ml . (I⁻) (IO₃⁻)
(I₃⁻) (IO₃⁻ + 8I⁻ + 6H⁺ → 3I₃⁻ + 3H₂O).
6. 1 ml .
7. .
8. 1 ml . ± 0.03 ml .
(I₃⁻ + 2S₂O₃²⁻ → 3I⁻ + S₄O₆²⁻).
9. (IO₃⁻) 1 (S₂O₃²⁻) 6 .
4 0.3% , ml 1 Vstd .

$$C_{Na_2S_2O_3 \cdot 5H_2O} = \frac{C_{KIO_3} \times 10.0 \times 6}{V_{Na_2S_2O_3 \cdot 5H_2O}}$$

$$C_{Na_2S_2O_3 \cdot 5H_2O} : Na_2S_2O_3 \cdot 5H_2O \quad (\text{mole/L})$$

$$C_{KIO_3} : KIO_3 \quad (\text{mole/L})$$

$$V_{Na_2S_2O_3 \cdot 5H_2O} : Na_2S_2O_3 \cdot 5H_2O \quad (\text{mL})$$

7.14 (Standard-curve)

1. (KIO₃) 2, 4, 6, 8, 10 ml 5 .
2. .

7.15

1. .
2. .
3. 50% 1 ml . 50% 1 ml . pH
(I₂) .
4. (t_L) .
5. . ml ul V_{sam} .
6. .

7.16

7.17

$$V_{\text{blk}} = V_1 - V_2 \quad I_2 \quad .$$

$$V_{\text{blk}} = V_1 - (V_2 - (V_3 - 1)) = 2V_1 - V_2 - V_3$$

$$V_1: \quad KIO_3 \text{ 1 ml}$$

$$V_2: \quad \quad KIO_3 \text{ 1 ml}$$

$$V_3: \quad KIO_3 \text{ 1 ml}$$

$$V_{\text{blk}} \quad \text{ml} \quad .$$

7.18 KIO_3

KIO_3 (t_p) 20 °C KIO_3 .

$$M(KIO_3, 20^\circ\text{C}) = \frac{m(KIO_3)/(213.995\text{g} \cdot \text{mol}^{-1})}{V_s} \times \frac{0.998206}{\rho_w(t_p)}$$

$$m(KIO_3): \quad KIO_3$$

$$V_s: KIO_3 \quad (t_p)$$

$$213.995\text{g} \cdot \text{mol}^{-1}: KIO_3 \text{ 1}$$

$$\rho_w(t_p):$$

$$V_s = V_s[1 + \alpha_V(t_L - 20)]$$

$$\alpha_V(\text{Pyrex}): 9.75 \times 10^{-6} \text{ } ^\circ\text{K}^{-1}$$

7.19

(tL) .

$$M(Na_2S_2O_3, t_L) = \frac{6000 \times V(KIO_3, t_L) \times M(KIO_3, t_L)}{V_{std} - V_{blk}}$$

$$V(KIO_3, t_L) = V(KIO_3, 20^\circ\text{C}) \times (1 + 9.75 \times 10^{-6}(t_L - 20))$$

$$M(KIO_3, t_L) = M(KIO_3, 20^\circ\text{C}) \times \frac{\rho_w(t_L)}{0.998206}$$

$$6000 = \frac{6\text{mol } Na_2S_2O_3}{1\text{mol } KIO_3} \times \frac{1000\mu\text{l}}{1\text{ml}}$$

$$V_{std}: KIO_3$$

$$V_{blk}: \quad (\text{reagent blank}) \quad \text{ml} \quad 1 \quad .$$

7.20

(+ O_2) .

$$n(O_2) = (V_{sam} - V_{blk}) \times M(Na_2S_2O_3, t_L) \times \frac{1L}{10^6\mu\text{l}} \times \frac{1\text{mol } O_2}{4\text{mol } Na_2S_2O_3}$$

$$C(O_2) = \frac{[n(O_2) - 7.6 \times 10^{-8}]}{m(sample)}$$

,
7.6 × 10⁸ : (MnCl₂ + NaI/NaOH) 2 ml (O₂)
m(sample) (kg) .
m(sample) = V(O₂ , 20°C) × [1 + 9.75 × 10⁻⁶(t_s - 20)] - 2 × ρ(t_s), S)
,
t_S
2
ρ_{SW}

7.21 / (QA/QC)

5 10 . 0.44 mol/L .
0.16 0.94 mol/L .
±0.1 % ,
±0.45 mol kg⁻¹ .
1.78 mol/L . CTD .
. CTD .

7.22

Chapter 8

chlorophyll a

8.1

chlorophyll a, b, c . chlorophyll a
(,) () .

8.2

8.2.1

90 % 750, 665, 645, 630, 480nm chlorophyll a, b,
c (total carotenoids) . chlorophyll a 0.02 g/L ,
0.04 g/L . chlorophyll a 5 g/L \pm 0.3 g/L, chlorophyll b
0.5 g/L \pm 0.2 g/L .

8.2.2

chlorophyll 5~10 100~200mL
chlorophyll a phaeo-pigment . chlorophyll a 5 g/L \pm
10 % .

8.2.3

chlorophyll (exiting wavelength) 436nm, (emission
wavelength) 670nm chlorophyll a phaeo-pigment . chloro-
phyll a 5 g/L \pm 10 % .

8.3

chlorophyll a, b, c . 750nm

8.4

(4.1), (4.2), (4.3), (15mL)(4.4), (4.5),
(4.6), (4.7), (4.8)
(4.9), filter holder(4.10) .

8.4.1

8.4.2

GF/F, 47mm .

8.4.3

a .

8.4.4

, . 15 mL .

8.4.5

.

8.4.6 (spectrophotometer)

480nm, 630nm, 645nm, 665nm, 750nm 1cm cell .

8.4.7 (Turner 10AU)

F4T4-BL , Wratten 47 B Corning CS 5-60 filter,
Corning CS 2-64 filter .

8.4.8 (spectrofluorometer)

436nm (exciting wavelength), 670nm (emission wavelength)
1cm cell .

8.4.9

a 50mL .

8.4.10 filter holder

a 25mm filter holder .

8.5

a (5.1), (5.2), (5.3, 5.4), (5.5) .

8.5.1 90% (v/v)

1000mL 100mL 900mL .

8.5.2 1%

(MgCO₃) 1g 100mL .

8.5.3 1:1

50 mL 100mL .

8.5.4 0.5 N

(37%) 41mL 1000mL .

8.5.5 100 ppm

100mg (C₂₀H₁₀O₄Na₂, sodium fluorescein) 1L
1mL 100mL .

8.5.6 (Reference Material, RM)

Sigma Aldrich, Fluka

8.6

, 0.3 mm 0.5 5,000 mL()
holder . 100 200 mL filter
(GF/F, 25mm) .
1 % .
15mL .

((-196°C -80°C) , -20°C).

8.7

8.7.1

7.1.1. Cell-to-Cell Blank : cell 90 % Reference cell
cell Cell-to-Cell blank 750 nm, 665 nm, 645 nm, 630 nm, 480 nm
E .

$$E_{665} = OD_{665} - OD_{750} E_{645} = OD_{645} - OD_{750} E_{630} = OD_{630} - OD_{750}$$

OD (optical density) .

7.1.2. Blank : 750 nm E . E ,
Blank ±0.002 , Cell-to- Cell
Blank E .
Membrane E
Blank 750nm Cell-to-Cell Blank E (E_b) f Blank .
f = 1 (, 666, 645, 630nm)
f = 2 (, 510nm)
f = 3 (, 480nm)

, Blank Cell-to-Cell Blank Blank (f x E_b) .

$$Total\ Blank = (Cell - to - CellBlank) + (f \times E_b)$$

7.1.3. 0 : 90 % 0 .

8.7.2

7.2.1. , .
1/2 (20mm-Hg) .
1% (3~5) ().
() , 25mm filter holder 100~200mL
1/2 1% (3~5) ().
7.2.2. chlorophyll . 7.2.3.
90% 15mL() 10mL() 4°C .
7.2.4. 10 (3,000~4,000 rpm) .
7.2.5.
7.2.5.1 : 1cm 750nm, 665nm, 645nm, 630nm, 480nm
.
7.2.5.2 : (R_B). 5% 2 3
2 3 (R_A).
7.2.5.3 : 436nm, 670nm . 1cm

(F_B). 0.5N 2~3 1~2 mL 1~2 (F_A).
.

8.7.3

7.4.1. : (g/L) .

$$\text{Chlorophyll } a, b, c (\mu g/L) = \frac{C_{a,b,c} \times v}{V} \times 1000$$

Chlorophyll a, b, c : a, b, c (g/L)

v : (mL)

V : (mL)

C_a : 11.6 E₆₆₅ - 1.31 E₆₄₅ - 0.14 E₆₃₀ (a)

C_b : 20.7 E₆₄₅ - 4.34 E₆₆₅ - 4.42 E₆₃₀ (b)

C_c : 55 E₆₃₀ - 1.31 E₆₆₅ - 0.14 E₆₄₅ (c)

7.4.2. : .

7.4.3. : (g/L) .

$$\text{Chlorophyll } a (\mu g/L) = F_D \frac{r}{r-1} (R_B - R_A) \times 1000$$

$$\text{Phaeo-pigment} (\mu g/L) = F_D \frac{r}{r-1} (rR_A - R_B) \times 1000$$

F_D : door

r :

R_B :

R_A :

7.4.4. : .

$$\text{Chlorophyll } a (\mu g/L) = \frac{(F_B - F_A)v}{F_{ph}(R-1)V}$$

$$\text{Phaeo-pigment} (\mu g/L) = \frac{R(F_A - F_b)v}{F_{ph}(R-1)V}$$

R : F_{ch}/F_{ph} (F_{ch} F_{ph} chlorophyll a phaeo-pigment) chlorophyll a

F_B/F_A

v : 90% (mL)

V : (mL)

8.7.4

7.5.1. chlorophyll a . chlorophyll a
 chlorophyll . *Skeletonema costatum*
Skeletonema costatum (chlorophyll) *Coccolithus huxleyii* *Peridinium*
trochoidium chlorophyll source . , phaeo-pigment
 . “bloom” () .
 , Sigma Aldrich, Fluka .

7.5.2. 50 mL 90% . door 3 50 (R_3).
 chlorophyll a , door 3 F_3 .

$$F_3 = \frac{C_a}{R_3}$$

chlorophyll a door 10 30 50 90 % .
 F_{10} F_{30} .

7.5.3. Door F_D 7.5.1 7.5.2. , . r phaeo-pigment
 R_B/R_A . 2.2 .

8.7.5

$\times 1$, (exciting wavelength) 436 nm, (emission wavelength) 670
 nm ($C_{20}H_{10}O_4Na_2$, sodium fluorescein) 1 ppm 100 (.
 0.5 ppm 436 nm, 515 nm 100).

8.8 / (QA/QC)

8.9

Chapter 9

9.1

CO₂ (NDIR) (2013-230) ,
99% 0.05 mgC/L(4.16
molC/L) . 5 mgC/L(462.2 M) ±0.08 mg C/L(6.66 M) .

9.2

NDIR 680
(High-Temperature Catalytic Oxidation, HTCO) ,
Shimadzu HTCO .

9.3

NDIR .

9.4

(4.1), (4.2), (4.3) . 10% 3

9.4.1

- 1.1.1. Go-Flo Teflon Teflon Niskin .
O-ring . monofilament nylon Kevlar lanyard .
- 1.1.2. 10ml 40ml . Polycarbonate .

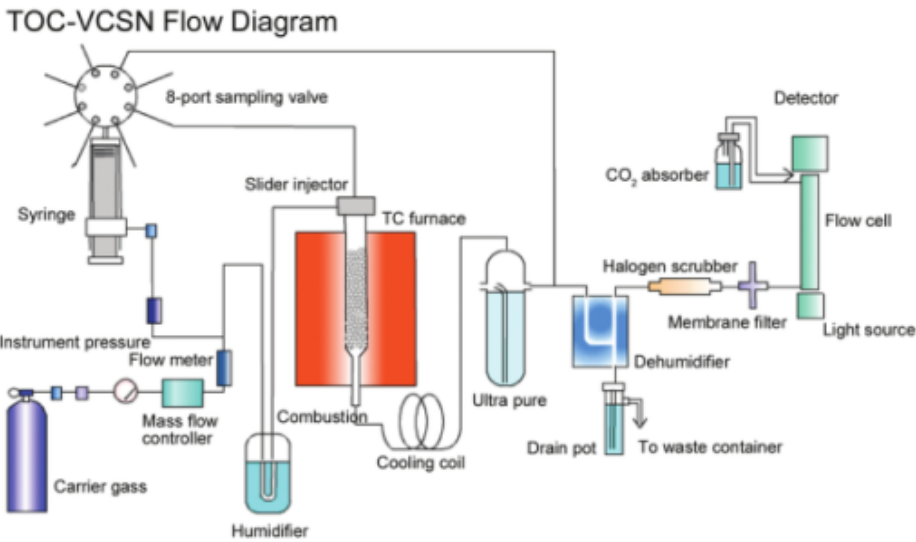


Figure 9.1:

1.1.3. .

9.4.2

PP, PE, Teflon 25/47mm 10%
PP, PE, Teflon .

9.4.3

300g 0.001g .

9.4.4

1.4.1. 24mL vial93 ea, 9ml vial93 ea, 40mL*68 ea

1.4.2. 680

1.4.3.

1.4.4. (NDIR detector)

1.4.5. 250 ml .

9.5

(5.1), (5.2), (5.3), (5.4) .
10% 24 100 .

9.5.1 (Low-Carbon Water, LCW)

Milli-Q . . . , . . . , . . .

9.5.2 (83.3 mmolC/L)

Potassium Hydrogen Phthalate(potassium biphthalate, $\text{C}_8\text{H}_5\text{KO}_4$,
204.22g/mol) 2.5g 100 4 . 2.1254g
700mL LCW 1000mL . LCW (4
l H_3PO_4 /mL) pH 2~3 .

9.5.3 (Oxidation Efficiency Check Stock Solution, OEC)

Sulfathiazole($\text{C}_9\text{H}_9\text{N}_3\text{O}_2\text{S}_2$, 255.31g/mol) 2.8g EDTA($\text{C}_{10}\text{H}_{16}\text{N}_2\text{O}_8$,
292.2438g/mol) 2.8g 100 4 .

1.1.1. _1(OEC_1) Sulfathiazole 2.3641g LCW 700mL
1000 mL (83.34 mmol C/L).

1.1.2. _2(OEC_2) EDTA 2.4354g LCW 700mL 1000
mL (88.33 mmol C/L).

1.1.3. (OEC_1, OEC_2) LCW .
(4 l H_3PO_4 /mL) pH 2~3).

9.5.4 (Carrier Gas)

(99.9995%) .

3

9.5.5

Niskin/Go-Flo 10%
. Niskin .

nylon, PP, Teflon phthalate .
Niskin/Go-Flo .

9.5.6 ()

PP, PE, Teflon 25/47mm 10% .
PP, PE, Teflon .
(GF/F, 0.7 m) . GF/F 500 12
. DOC 10% polycarbonate
. Polycarbonate . 150 mmHg

laminar airflow workbench

9.5.7

10 ml laminar flow workbench
 (ULTREX II, J.T.Baker; SUPRAPUR, Merck) (H₃PO₄) (HCl) (4 l
 H₃PO₄/mL sample, 12 l HCl/mL sample) pH=2~3 . pH
 . pH 2 .
 DOC Ziploc .
 DOC (Wiebinga and Barr, 1998). laminar
 flow workbench HDPE -20°C 5
 (Tupas et al., 1994). -20°C .

9.6

(5.2), (7.1), (7.2), (5.1) (7.3)

9.6.1

3 10% 24 3 500 5 .
 Septum 10% 24 100 . Septum
 3 .

9.6.2

1.2.1. Shimadzu type
 0.5% (Pt) (Al₂O₃) . 0.5%
 24 , LCW .
 250ml (>2500) (Skoog et al., 1997). 1.2.2. 1
 680 . 900 150 mL min⁻¹ ,
 , . Purge gas 50-75 mL min⁻¹ . , , . Shimadzu
 ,
 LCW , , , . LCW 0.5 .

9.7 / (QA/QC)

9.7.1

10 7 3.14 (99%) .

9.7.2

, , .

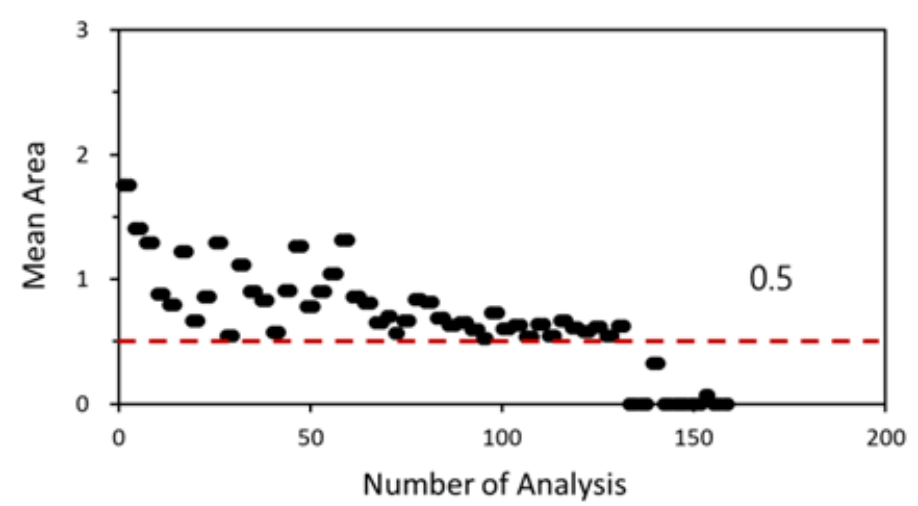


Figure 9.2: LCW (0.5)

9.7.3 laminar airflow workbench LCW (4 , 20) (2).

9.7.4 LCW . .

9.7.5 LCW Niskin () . .

9.7.6 . .

9.7.7 LCW 1 . .

9.7.8 (Certified Reference Material, CRM)
Dr. Dennis Hansell (2 M C, 45 M C) .
(10mL).

9.8

9.9

Turpas, L.M., Popp, B.N., Karl, D.M. (1994) Dissolved organic carbon in oligotrophic waters: experiments on sample preservation, storage and analysis. *Mar.Chem.* 45:207-216.

Wiebinga, C.J., de Baar, H.J.W. (1998) Determination of the distribution of dissolved organic carbon in the Indian sector of the Southern Ocean. *Mar.Chem.* 61:185-201.

Chapter 10

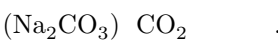
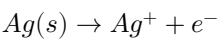
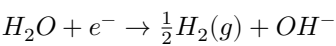
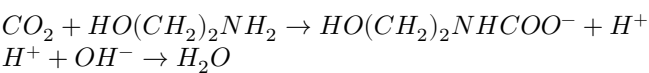
10.1

$$\frac{1\text{kg}}{(3800\sim 4300\text{ mol kg}^{-1})} \cdot (1800\sim 2300\text{ mol kg}^{-1})$$

10.2

CO₂ . . . pH . . . CO₂
610nm

$$CO_2 + HO(CH_2)_2NH_2 \rightarrow HO(CH_2)_2NHCOO^- + H^+$$

$$H^+ + OH^- \rightarrow H_2O$$


10.3

1980 SOMMA (Single Operator Multi-parameter Metabolic Analyzer) 1990 MRIANDA VINDTA 3C.

10.3.1

500mL glass-joint () (CO₂) . VINDTA
 3C(5420150413) , 8 9 19 mL . ±0.4°C
 / .

10.3.2 CO₂

(frit) . CO₂
 .

10.3.3

UIC (UIC CM5017O, 5420160221).
 pH pK ±0.2°C .
 100mL , 1°C 200 . ,
 .

10.3.4 PC

VINDTA 3C UIC CM5017O / VINDTA
 PC

10.4

(4.1), CO₂ (4.2), (4.3), (4.4), (4.5) .

10.4.1

CO₂ CO₂ (>99.999%). 10L , 4~5
 .

10.4.2 CO₂

CO₂ CO₂ (Ascarite).
 .

10.4.3

. 85% 10 8.5% .

10.4.4

CO₂ () CO₂ . Peltier cooler
 0~1°C (condenser) () (Supelco ORBO-53) CO₂
 . H₂S , .

10.4.5

UIC 100 mL (, (DMSO)
). UIC 1 cm (,
DMSO).

10.5

(, ,) . CO₂ .
36 , 3 (, ,) .

10.6

10.6.1

1) , 2) 2 “junk”
, 3) , 4) CO₂ CRM , 5) TCT 24 . CO₂
CRM .

10.6.2

VINDTA “BLANK” . CO₂
10 10 1 25 , 25
25 VINDTA ‘blank’ 50 .

10.6.3

‘junk’ CO₂ CRM, CO₂ .
20 mL , 1.5 mL .

10.6.4

regia . , (frit) . aqua
50°C 12 .

10.7

10.7.1

(counts min⁻¹) .

$$b = \frac{N_b}{10}$$

, $N_b = 10$

10.7.2

$$C_T' = \frac{N_s - b \cdot t - a}{c} \cdot \frac{1}{V_s \cdot \rho}$$

$$\begin{aligned} C_T' &= & (\text{mol kg}^{-1}) \\ N_S &= & (\text{counts}) \\ a &= & (\text{counts}) \\ b &= & (\text{counts min}^{-1}) \\ c &= & (\text{counts mol}^{-1}) \\ t &= & (, \text{ min}) \\ V_S &= & (\text{mL, dm}^3) \\ \rho &= & (\text{g cm}^{-3}) \end{aligned}$$

(headspace) CO_2

$$C_T = 1.0002(C_t' - \Delta C_T)$$

, ΔC_T CO_2 C_T . r 1% 0.5 mol kg⁻¹ .

10.8 /

10.8.1

4 mol kg⁻¹ . CO_2 (1) 1.5 mol kg⁻¹ ,
8.1.1. 25 counts min⁻¹ (0.05 g C min⁻¹) , ± 10 counts min⁻¹
/ .

(\bar{x}) , .

(upper control limit) $UCL = \bar{x} + 3s$
(upper warning limit) $UWL = \bar{x} + 2s$
(lower warning limit) $LCL = \bar{x} - 2s$
(lower control limit) $LCL = \bar{x} - 3s$

8.1.2. CO_2 CRM

Scripps Dr. Dickson CO_2 CRM () . / .

8.1.3.

10 . / .

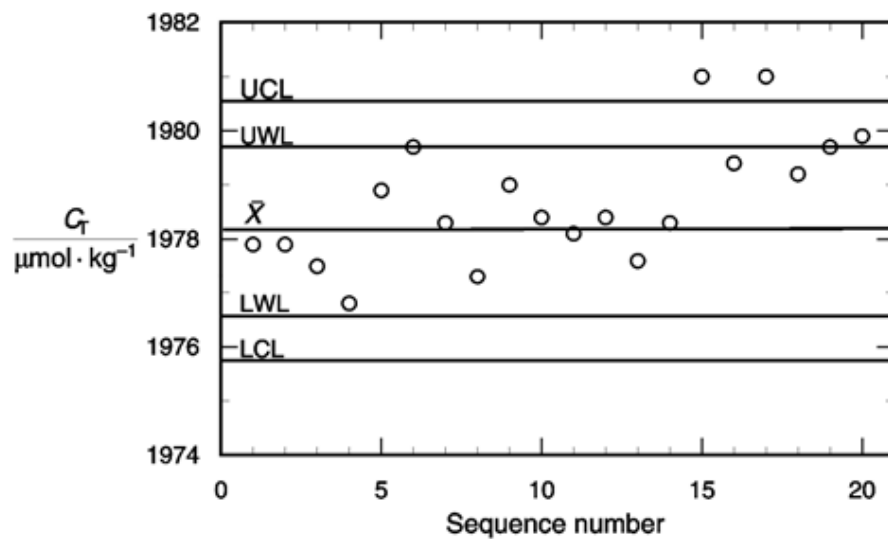


Figure 10.1:

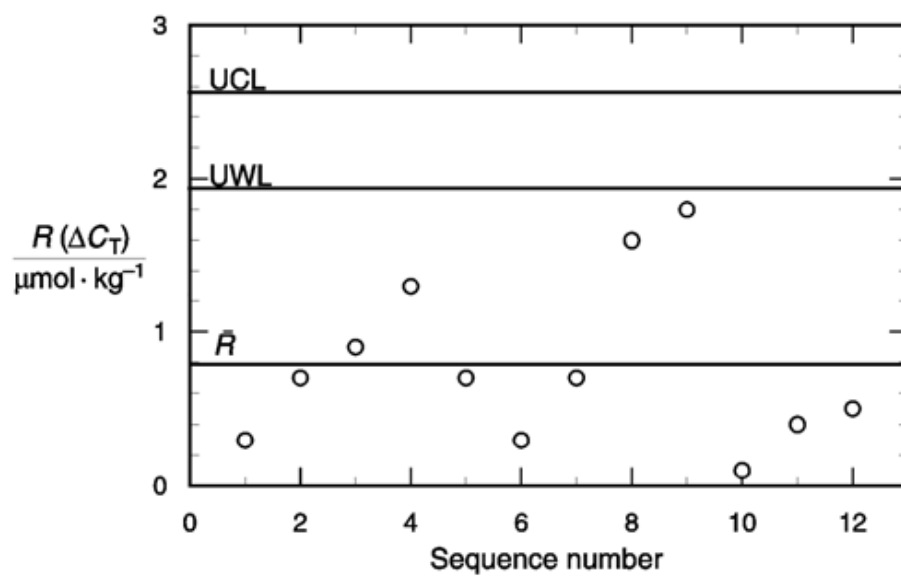


Figure 10.2:

(R)

(upper control limit) $UCL = 3.267\bar{R}$
(upper warning limit) $UWL = 2.512\bar{R}$
(lower warning limit) $LCL = 0$
(lower control limit) $LCL = 0$

10.9

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- Huffman, Jr., E.W.D. 1977. Performance of a new automatic carbon dioxide coulometer. *Microchem. J.* 22: 567–573.
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- Johnson, K.M., Williams, P.J. leB., Brändström, L. and Sieburth, J.M. 1987. Coulometric TCO₂ analysis for marine studies: automation and calibration. *Mar.Chem.* 21: 117–133.
- Johnson, K.M., Wills, K.D., Butler, D.B., Johnson, W.K. and Wong, C.S. 1993. Coulometric total carbon dioxide analysis for marine studies: maximizing the performance of an automated continuous gas extraction system and coulometric detector. *Mar. Chem.* 44: 167–187.
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- Wilke, R.J., Wallace, D.W.R. and Johnson, K.M. 1993. Water-based, gravimetric method for the determination of gas sample loop volume. *Anal. Chem.* 65: 2403–2406.