

Changes in Phytoplankton and Zooplankton Communities

Common Indicator Assessment



OSPAR

QUALITY STATUS REPORT 2023

Changes in Phytoplankton and Zooplankton Communities

OSPAR Convention

The Convention for the Protection of the Marine Environment of the North-East Atlantic (the “OSPAR Convention”) was opened for signature at the Ministerial Meeting of the former Oslo and Paris Commissions in Paris on 22 September 1992. The Convention entered into force on 25 March 1998. The Contracting Parties are Belgium, Denmark, the European Union, Finland, France, Germany, Iceland, Ireland, Luxembourg, the Netherlands, Norway, Portugal, Spain, Sweden, Switzerland and the United Kingdom.

Convention OSPAR

La Convention pour la protection du milieu marin de l’Atlantique du Nord-Est, dite Convention OSPAR, a été ouverte à la signature à la réunion ministérielle des anciennes Commissions d’Oslo et de Paris, à Paris le 22 septembre 1992. La Convention est entrée en vigueur le 25 mars 1998. Les Parties contractantes sont l’Allemagne, la Belgique, le Danemark, l’Espagne, la Finlande, la France, l’Irlande, l’Islande, le Luxembourg, la Norvège, les Pays-Bas, le Portugal, le Royaume-Uni de Grande Bretagne et d’Irlande du Nord, la Suède, la Suisse et l’Union européenne

Contributors

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Delivered by

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Key Message

Between 2015 and 2019 plankton functional groups experienced significant changes in abundance, reflecting a continuation of long-term trends. Larvae of benthic invertebrates increased, while other groups mostly decreased. Observed changes were primarily linked to rising temperatures within offshore areas in the North-East Atlantic, and to nutrients in some coastal areas.

Background

Plankton (microscopic algae and animals) form the base of marine food webs, making them important indicators of ecosystem state. Short generational times, small size and, for phytoplankton, direct dependence on dissolved nutrients, make plankton well-suited for detecting environmental change. Changes in plankton communities can affect higher food web levels, such as shellfish, fish, and seabirds, which are supported either directly or indirectly by plankton. Indicators based on plankton lifeforms, planktonic organisms with the same functional traits (Figure 1), can reveal plankton community responses to factors such as nutrient loading from human activities and climate-driven change. Changes in the abundances of plankton lifeforms over short time scales can have immediate consequences for higher trophic levels, whereas gradual change over decades can indicate long term shifts in important aspects of ecosystem functioning.

At the North-East Atlantic regional scale, plankton community changes are strongly linked to environmental conditions. Pelagic habitats can be defined according to key water column features, such as depth, freshwater influence, stratification, and based on spatially consistent patterns in phytoplankton dynamics. Examining the direction and magnitude of changes in lifeform abundance over time across pelagic habitats can reveal spatial patterns of environmental change. Similarly, trends determined from regular monitoring of fixed-point stations can be compared to trends in surrounding and adjacent pelagic habitats. This assessment addresses plausible links between environmental pressures and long term changes in plankton communities.

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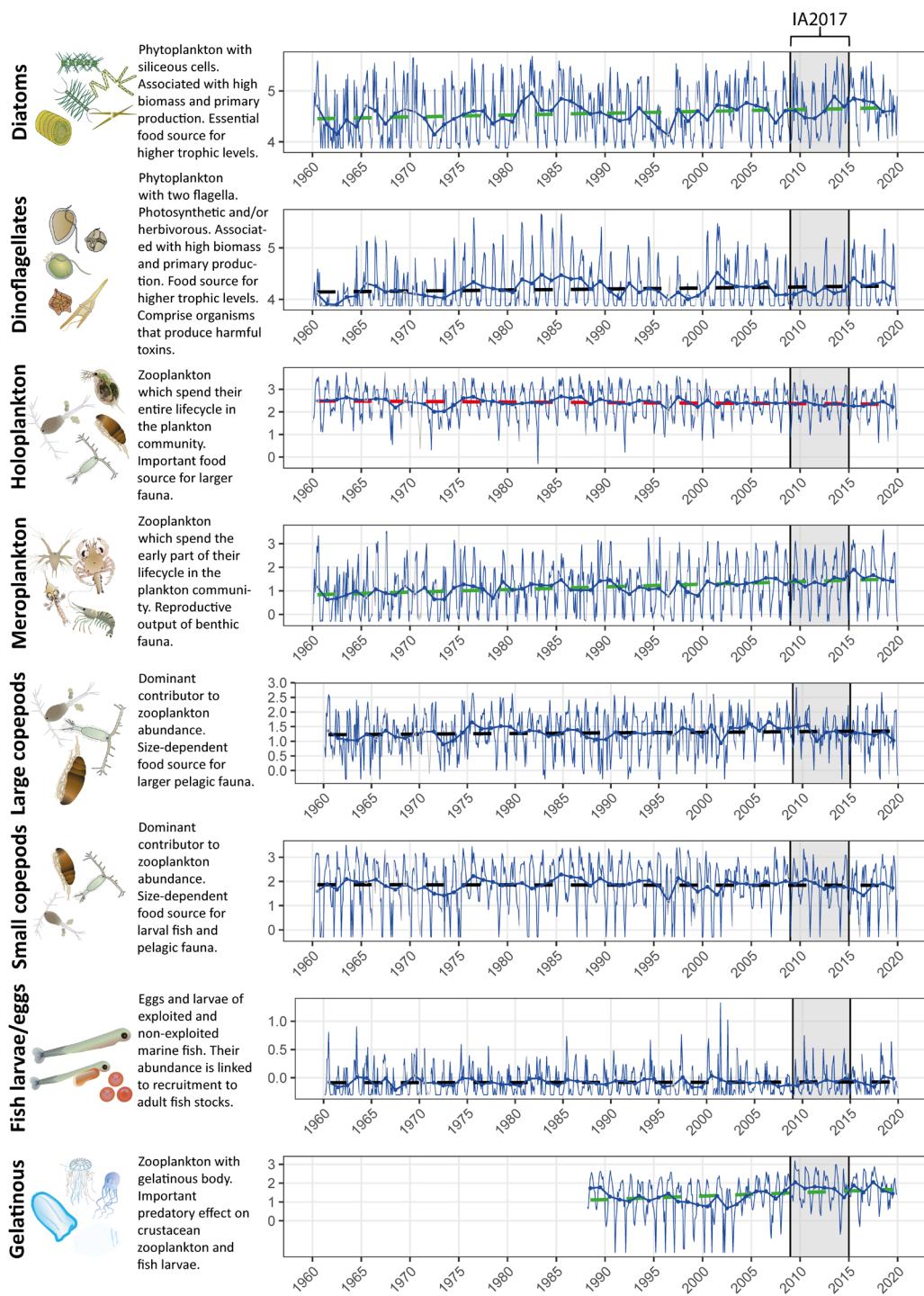


Figure 1: Long term monthly and annual \log_{10} transformed abundance time-series for eight plankton lifeforms in the Western Channel. Blue lines display monthly variability (thinner line), and annual mean abundance (thicker line) values. Dashed lines indicate linear trend lines in annual abundance without any inference on statistical significance. The Kendall trend test is used to infer significance of trends, with red: decreasing trend, green: increasing trend, and black: no trend. Data obtained from the Continuous Plankton Recorder (CPR) survey and Plymouth Marine Laboratory (for gelatinous zooplankton only due to non-quantitative capture of gelatinous taxa by CPR). The shaded region represents the time period of IA2017. Plankton images courtesy of the Integration and Application Network, University of Maryland Center for Environmental Science (ian.umces.edu/symbols/).

Background (Extended)

Indicators based on plankton lifeforms have been used to assess community response to sewage pollution (Charvet *et al* 1998; Tett *et al.*, 2008), **anoxia** (Rakocinski 2012), fishing (Bremner *et al.*, 2004), eutrophication

(HELCOM 2012), climate change (Beaugrand 2005; Bedford *et al.*, 2020a; McQuatters-Gollop *et al.*, 2019), and ocean acidification (Keys *et al.*, 2018). Indicators based on functional groups have been proven relevant for the description of the community's structure and biodiversity and are more easily inter-compared than other indicators based on taxonomy (Estrada *et al.*, 2004; Gallego *et al.*, 2012; Garmendia *et al.*, 2012; Mouillot *et al.*, 2006).

In practice, it is often preferable to aggregate species with similar traits into functional groups, such as lifeforms (**Figure 1**), rather than assessing the dynamics of individual species. Measures of species abundance are frequently subject to large interannual and regional variation, often due to natural physical dynamics and habitat preferences rather than anthropogenic stressors (de Jonge, 2007). Functional group abundance is often less variable because variability in the abundances of the group's constituent species averages out. Cryptic speciation (species with near-identical appearance) within the plankton community, alongside the limitations of identifying plankton using routine light microscopy techniques, make it difficult to generate accurate counts at a species or genus level. Functional group abundance is more reliable as many plankton lifeforms are easily identified, making comparisons between different laboratories and institutes feasible. Both abundance and biomass data can be used to inform lifeform time series, depending on the lifeform in question and data availability from monitoring programmes.

In addition to studying change in individual lifeforms, these concepts can be extended to investigating changes in ecologically relevant pairs of lifeforms in the form of a lifeform pairs index. The precise combination of lifeforms composing the pairs will depend on the habitat and the objective of the indicator, e.g., as a measure for change in pelagic habitats, food webs, seafloor integrity or eutrophication. Change in the abundance of ecologically linked lifeforms over time can also provide an indication of changes in various aspects of ecosystem function. These include, for example: the transfer of energy from primary to secondary producers (changes in phytoplankton and zooplankton); the pathway of energy flow and top predators (changes in gelatinous zooplankton and fish larvae); benthic / pelagic coupling, i.e., changes in holoplankton (fully planktonic) and meroplankton (only part of the lifecycle is planktonic, the remainder is benthic) (see also Gowen *et al.*, 2011; McQuatters-Gollop *et al.*, 2019).

While studying lifeform pairs can be helpful for detecting changes in the annual cycle of ecologically linked lifeforms, assessing lifeforms separately is more suitable for evaluating gradual change over time. It is simpler to interpret results of an analysis of correlation between lifeform abundance and environmental pressures, than between an index of lifeform pairs and environmental pressures (Bedford *et al.*, 2020a). A method has been developed to detect potential links between environmental pressures and change in lifeform abundance over time. The main features of the method are: (i) the grouping of planktonic taxa into functional types or lifeforms; (ii) spatially division of plankton samples to construct distinct time-series of lifeform abundance; (iii) using a robust nonparametric test to quantify long term changes in lifeform abundance; (iv) relating change in lifeform abundance to trends in environmental pressures and climate indices.

The OSPAR Intermediate Assessment 2017 (IA2017) stated that conclusions on the trends observed could not yet be made as additional work was needed to investigate environmental pressures and potential impacts of climate change and other human pressures on plankton communities. This QSR 2023 assessment addresses plausible links between environmental pressures and long-term changes in plankton communities.

Assessment Method

Previous assessment

The previous assessment (Intermediate Assessment 2017; IA2017) was based largely on state space theory (see below) and examined differences in the relative abundances of ecologically relevant lifeform pairs between a past comparison period (2004 to 2008) and a contemporary assessment period (2009 to 2014). While this method is suitable for detecting changes in the annual cycle of ecologically linked lifeforms, it is

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less suited to evaluating gradual change over time. Further, stronger statistical links can be drawn by examining relationships between the abundances of distinct planktonic lifeforms and environmental pressures, than between an index of lifeform pairs and environmental pressures (Bedford *et al.*, 2020a). For the current assessment a simplified approach was developed, based on measuring long-term changes in the abundance of key lifeforms and evaluating how environmental pressures co-varied with changes in lifeform abundance. A long-term perspective helped improve confidence in establishing links to environmental pressures. Short-term changes within the assessment period were also evaluated to identify emerging trends. The PH1 indicator as originally defined in IA2017 was still evaluated for this assessment and is described under the Lifeform pairs indicator approach section of the Assessment Methods and the respective results are described in the Results (extended) section.

Kendall trend test and statistic

Abundance trends for planktonic lifeforms can be evaluated over time by applying the Kendall trend test to annual mean abundance values (Bedford *et al.*, 2020a; Desmit *et al.*, 2020). The test was performed on annual \log_{10} transformed mean abundance values, rather than monthly or seasonal values, to remove the seasonal variation typical of plankton time-series data (**Figure 1**). This nonparametric test generates a statistic which is derived by comparing each value in a time-series with each of the values preceding it. If a latter value is greater than a previous one, the pairwise comparison is assigned a value of 1. If it is lower it is assigned a value of -1, with 0 assigned to cases when there is no difference between values. The sum of the pairwise comparisons for the time series produces Kendall's S-statistic. The variance in the S-statistic is used to derive a Z-score with an approximately normal distribution; thus, confidence in this statistic can be assessed with an associated p-value, with $p \leq 0,05$ generally accepted as statistically significant change. The sign of the test statistic reveals the direction of the trend, with a positive statistic indicating an increasing trend and a negative statistic indicating a decreasing trend. The magnitude of the statistic is proportional to the strength of the trend. A great benefit of this nonparametric test is that it yields identical results irrespective of the data transformation method and is not sensitive either to gaps in data or to non-linear or irregular trends. It also has the advantage that abundance trends are comparable across datasets, lifeforms, and assessment units.

Time-series plots (**Figure 1**) indicate variation in lifeform abundances through time for a single assessment unit, with the blue lines displaying monthly variability (thinner line), and annual mean abundance (thicker line) values used to derive the Kendall statistic. Data correspond to the assessment unit representing the western entrance to the English Channel ('Channel well mixed' see **Figure a**) and contains the fixed-point station 'L4', consistently monitored by the Plymouth Marine Laboratory since 1988.

The ability to aggregate complex plankton data into ecologically relevant lifeforms is a useful tool for assessment because it can help identify where changes are occurring (McQuatters-Gollop *et al.*, 2019). The ability to understand which taxa are driving changes in a lifeform can also be valuable. By calculating the Kendall statistic for each taxon, additional information can be gleaned to assist in interpreting lifeform time-series trends. For example, **Figure a** displays change in the abundance of diatoms for the 'Channel well mixed' COMP4 assessment unit described in **Figure 1**. The abundance of diatoms has been increasing since 1960 ($z = 2,66$, $p \leq 0,05$). Through examining the annual abundance time-series for individual diatom taxa it becomes possible to also conclude that overall trend of increasing diatom abundance is driven by positive abundance trends in 11 out of 54 diatom taxa. Increasing trends and greater absolute abundances for *Proboscia alata* and *Thalassiosira spp* indicate that they are predominantly responsible for the increasing abundance trend in diatoms. Additionally, while the net trend is increasing, four diatom taxa (*Guinardia striata*, *Skeletonema costatum*, *Thalassionema nitzschiooides*, and *Odontella sinensis*) are actually decreasing in abundance.

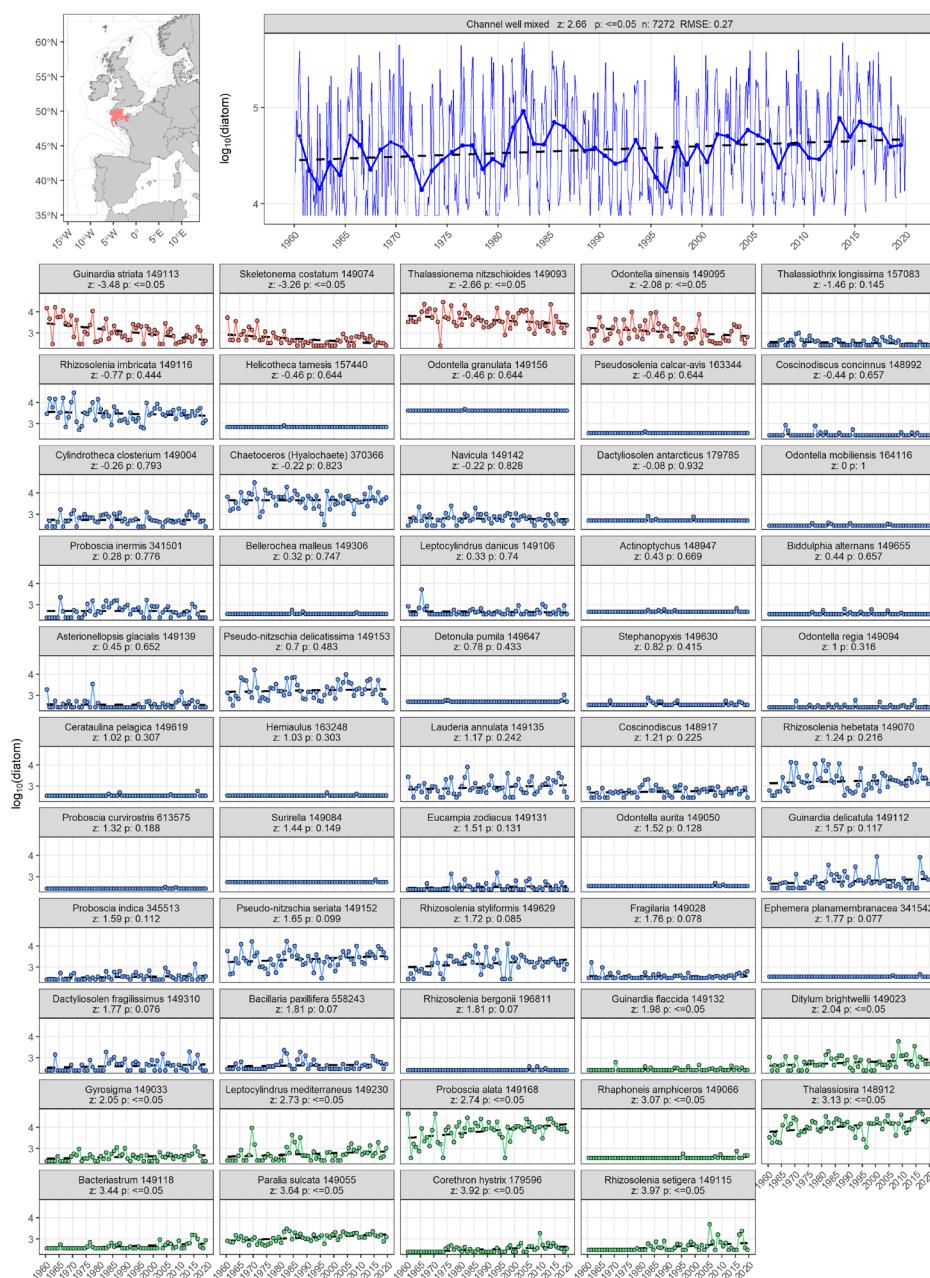


Figure a: Long-term \log_{10} transformed abundance time-series for diatoms in the 'Channel well mixed' COMP4 assessment unit. Data obtained from the Continuous Plankton Recorder (CPR) survey. Individual diatom taxa abundance time series are displayed as facets. Facet labels indicate taxon scientific name and Aphia ID, z : derived from Kendall's S-statistic, and p : Kendall p-value. Taxon time-series are coloured according to the outcome of the Kendall trend test, with red: decreasing trend and $p \leq 0.05$, blue: $p > 0.05$ (no trend), green: increasing trend and $p \leq 0.05$.

Since this assessment was based on data from multiple sources, the Kendall statistic was calculated independently for each combination of dataset, assessment unit, and lifeform. Since the datasets used also varied in duration, to avoid discarding data the full duration of each dataset was assessed up until the end of 2019. For a dataset to be included in this analysis, it needed to contain samples collected within the assessment period (2015-2019) and prior to the assessment period.

Spatial Scale:

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Because plankton community composition, distribution, and dynamics are closely linked to their environment, the analysis was performed at the scale of the ‘COMP4 assessment units’ (COMP4 v8a; **Figure b, Table a**). Assessment units within the Greater North Sea and Celtic Seas (OSPAR Regions II and III, respectively) were initially developed by Deltares and partner institutes as part of the EU ‘Joint Monitoring Programme of the Eutrophication of the North Sea with Satellite data’ (JMP-EUNOSAT; Enserink *et al.*, 2019) and further refined in the revision process of the eutrophication assessment by OSPAR expert groups ICG-EMO and TG-COMP. Assessment units with similar phytoplankton dynamics were derived from cluster analysis of satellite data for chlorophyll *a* and primary production. Boundaries between assessment units were derived by relating clustering results to the best-matching gradients in environmental variables obtained from the three-dimensional hydrodynamic Dutch Continental Shelf model version 6 (DCSMv6 FM). The variables which best matched the divisions highlighted by clustering were depth, salinity, and stratification regime. Additional geographic areas were added such as the Channel, Irish Sea, and Kattegat. These assessment units are a geographical representation of the conditions most likely to drive plankton distribution, dynamics, and community composition. The assessment units are being regularly updated and may not perfectly reflect the true state of the pelagic environment in their present form.

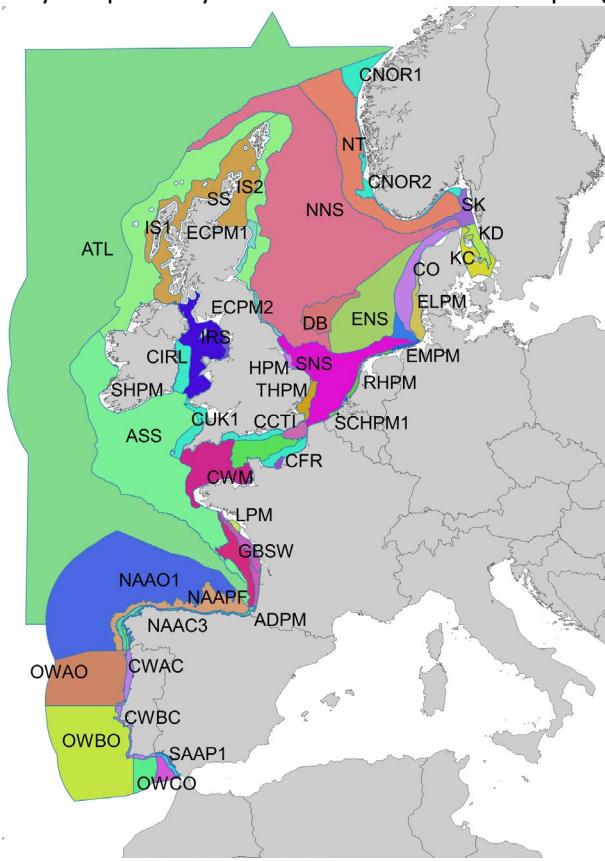


Figure b: COMP4 assessment units developed by JMP-EUNOSAT (Enserink *et al.*, 2019) and OSPAR. Area codes displayed in this figure are referred to in Table a.

Table a: COMP4 assessment unit definitions and categorisation according to pelagic habitat type (variable salinity, coastal, shelf or oceanic / beyond shelf) and OSPAR Region. Additional fixed-point stations which were not part of COMP4 but were evaluated in this indicator analysis are described in **Table b**.

COMP 4 assessment units categories	Area code	Area name	Mean salinity at surface (PSU)	Mean depth (m)	OSPAR Region
Variable salinity assessment units	ADPM	Adour plume	34,4	87	IV
	ELPM	Elbe plume	30,8	18	II
	EMPM	Ems plume	31,4	19	II
	GDPM	Gironde plume	33,5	34	IV
	HPM	Humber plume	33,5	16	II
	LBPM	Liverpool Bay plume	30,6	15	III

LPM	Loire plume	33,8	38	IV	
MPM	Meuse plume	29,3	16	II	
RHPM	Rhine plume	31,0	17	II	
SCHPM1	Scheldt plume 1	31,4	13	II	
SCHPM2	Scheldt plume 2	30,9	15	II	
SHPM	Shannon plume	34,1	61	III	
SPM	Seine plume	31,8	25	II	
THPM	Thames plume	34,4	22	II	
Coastal assessment units	CFR	Coastal FR channel	34,2	33	II
	CIRL	Coastal IRL 3	34,0	65	III
	CNOR1	Coastal NOR 1	34,3	190	II
	CNOR2	Coastal NOR 2	34,0	217	II
	CNOR3	Coastal NOR 3	32,4	171	II
	CUK1	Coastal UK 1	34,5	60	III
	CUKC	Coastal UK channel	34,8	37	II
	CWAC	Coastal Waters AC	No information	No information	IV
	CWBC	Coastal Waters BC	No information	No information	IV
	CWCC	Coastal Waters CC	No information	No information	IV
	ECPM1	East Coast (permanently mixed) 1	34,8	73	II
	ECPM2	East Coast (permanently mixed) 2	34,5	43	II
	GBC	German Bight central	33,4	39	II
	IRS	Irish Sea	33,7	65	III
	KC	Kattegat Coastal	25,7	21	II
	KD	Kattegat Deep	27,6	50	II
	NAAC1A	Noratlantic Area NOR-NorC1	No information	No information	IV
	NAAC1B	Noratlantic Area NOR-NorC1	No information	No information	IV
	NAAC1C	Noratlantic Area NOR-NorC1	No information	No information	IV
	NAAC1D	Noratlantic Area NOR-NorC1	No information	No information	IV
	NAAC2	Noratlantic Area NOR-NorC2	No information	No information	IV
	NAAC3	Noratlantic Area NOR-NorC3	No information	No information	IV
	OC	Outer Coastal DEDK	33,4	27	II
	SAAC1	Sudatlantic Area SUD-C1	No information	No information	IV
	SAAC2	Sudatlantic Area SUD-C2	No information	No information	IV
	SAAP2	Sudatlantic Area SUD-P2	No information	No information	IV
	SNS	Southern North Sea	34,3	32	II
Shelf assessment units	ASS	Atlantic Seasonally Stratified	35,2	134	III, IV
	CCTI	Channel coastal shelf tidal influenced	34,8	40	II
	CWIM	Channel well mixed	35,1	77	II, III
	CWMTI	Channel well mixed tidal influenced	35,0	59	II
	DB	Dogger Bank	35,1	28	II
	ENS	Eastern North Sea	34,8	43	II
	GBCW	Gulf of Biscay coastal waters	34,6	53	IV
	GBSW	Gulf of Biscay shelf waters	34,9	107	IV
	IS1	Intermittently Stratified 1	35,3	138	II, III
	IS2	Intermittently Stratified 2	35,1	102	II
	NAAP2	Noratlantic Area NOR-NorP2	No information	No information	IV
	NAAPF	Noratlantic Area NOR-Plataforma	No information	No information	IV
	NNS	Northern North Sea	35,0	121	II
	NT	Norwegian Trench	34,1	349	II
	SAAP1	Sudatlantic Area SUD-P1	No information	No information	IV

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	SK	Skagerrak	31,8	134	II
	SS	Scottish Sea	35,1	89	II, III
Oceanic / beyond shelf assessment units	ATL	Atlantic	35,3	2291	II, IV, V
	NAAO1	Noratlantic Area NOR-NorO1	No information	No information	IV
	OWAO	Ocean Waters AO	No information	No information	IV
	OWBO	Ocean Waters BO	No information	No information	IV
	OWCO	Ocean Waters CO	No information	No information	IV
	SAAOC	Sudatlantic Area SUD-OCEAN	No information	No information	IV

Table b: Details of fixed-point stations which were not part of COMP4 but were evaluated in this indicator analysis

Fixed-point station categories	Area code	Data provider	Salinity (surface mean)	Depth (mean)	OSPAR Region
Variable salinity fixed-point stations	Norderney	NLWKN	No information	No information	II
	LPO	Scottish Association of Marine Science	No information	No information	III
	Loch Ewe	Marine Scotland Science	No information	No information	III
Coastal fixed-point stations	L4	Plymouth Marine Laboratory	No information	54	II
	NOVANA	Aarhus University	No information	No information	II
	Scalloway	Marine Scotland Science	No information	No information	II
	Scapa	Marine Scotland Science	No information	No information	II
	Stonehaven	Marine Scotland Science	No information	No information	II
	RADIALES	Instituto Espanol de Oceanografia	No information	No information	IV

Because the Bay of Biscay and Iberian Coast (OSPAR Region IV) extended beyond the boundaries of the DCSMv6 FM, assessment units within this Region were developed using a different methodology, based on phytoplankton dynamics (Spain) and salinity dynamics (Portugal). To delineate assessment units for the Spanish coast, a polygon was created to extend from the coast to the exclusive economic zone (EEZ) boundary. Daily NASA Aqua/MODIS Level-2 satellite images

(<https://doi.org/10.5067/AQUA/MODIS/L2/OC/2018>) were used to calculate climatological mean values of chlorophyll *a* for each pixel. K-means clustering was then used to group pixels with similar dynamics, resulting in six distinct groupings within the main Spanish polygon. Portugal's three Water Framework Directive assessment units were extended to the boundaries of the Portuguese EEZ. These assessment units were further divided longitudinally to separate pelagic waters from coastal waters more subject to eutrophication from river influence by applying a salinity threshold of 30,0–34,5 for coastal waters and >34,5 for offshore waters, followed by a bathymetry threshold of 100 m.

COMP4 assessment units and fixed-point stations were categorised according to the OSPAR Regions they intersected as well as the habitat type they represented (**Table a**, **Table b**, **Figure c**). The four habitat types considered were: variable salinity (representing areas of freshwater influence where estuarine plumes extend beyond waters designated as Transitional Waters under Directive 2000/60/EC), coastal (representing areas with mean salinity < 34,5 psu), shelf (representing areas with mean salinity > 34,5 and mean depth < 200 m) and oceanic / beyond shelf (representing areas with mean salinity > 34,5 and mean depth > 200 m).

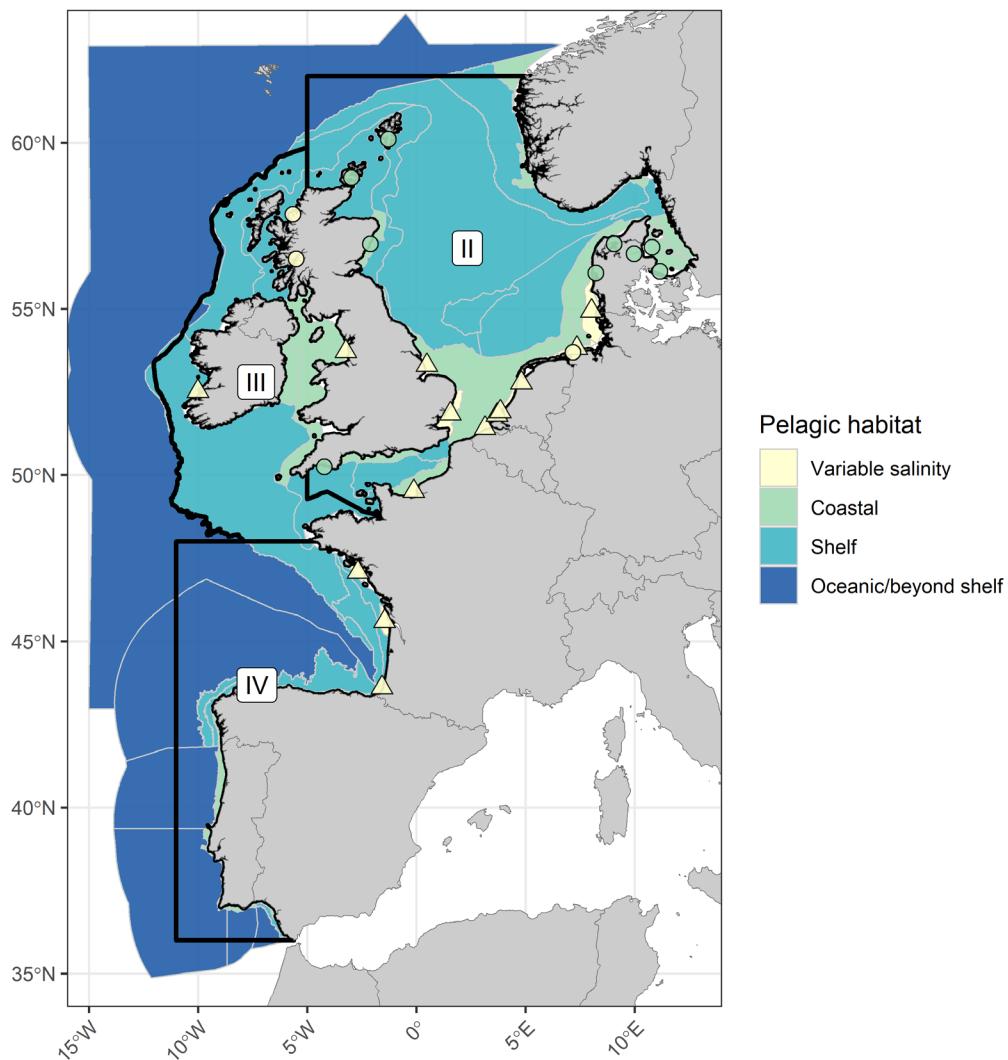


Figure c: The categorisation of COMP4 assessment units and fixed-point stations according to pelagic habitat type. Three assessed OSPAR Regions (I, II, and III) indicated by black outlines. River plumes are indicated by triangle symbols and fixed-point stations are indicated by circles.

Plankton Data:

The assessment has been carried out using 24 phytoplankton and zooplankton datasets from 14 sources (**Table c**, **Figure d**). Other datasets were provided but they were out of the scope of this assessment (e.g., time-series with comparison period duration shorter than assessment period duration, time-series ending prior to the assessment period, time-series commencing during assessment period, distributed sampling locations restricted to transitional waters only). These additional datasets were submitted by: Vlaams Instituut voor de Zee (BE), Bundesamt für Seeschiffahrt und Hydrographie (DE), Centre for Environment, Fisheries and Aquaculture Science (UK), Newcastle University (UK), and Natural Resources Wales (UK).

Table c: Contracting Parties and institutes that provided the datasets used for the indicator assessment.

Contracting Party	Institute	Dataset name	Date range
Belgium (BE)	Vlaams Instituut voor de Zee (VLIZ)	LW_VLIZ_zoo	2014-2020
Denmark (DK)	Aarhus University (AU)	NOVANA phytoplankton	1985-2020
Germany (DE)	Landesamt für Landwirtschaft, Umwelt und ländliche Räume des Landes Schleswig-Holstein (LLUR)	OSPAR_LLUR-SH_2010-2020	2010-2020

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	Niedersächsischer Landesbetrieb für Wasserwirtschaft, Küsten und Naturschutz (NLWKN)	OSPAR_NLWKN_1999-19_phyto	1999-2019
Netherlands (NL)	Rijkswaterstaat (RWS)	RWS_Fpzout_2000-2019_phyto	2000-2019
Portugal (PT)	Instituto Português do Mar e da Atmosfera (IPMA)	Pseudo-nitzschia vs Dinophysis_IPMA	2002-2020
Spain (ES)	Instituto Español de Oceanografía (IEO)	IEO_RADIALES_Photo	1989-2016
		IEO_RADIALES_Zoo	1991-2018
Sweden (SE)	Swedish Meteorological and Hydrological Institute (SMHI)	National data_SMHI_Kattegat-Dnr: S/Gbg-2021_116_phyto	1989-2021
		National data_SMHI_Kattegat-Dnr: S/Gbg-2021_116_zoo	1996-2020
		National data_SMHI_Skagerrak-Dnr: S/Gbg-2021_116_phyto	1986-2020
		National data_SMHI_Skagerrak-Dnr: S/Gbg-2021_116_zoo	1996-2020
United Kingdom (UK)	Centre for Environment, Fisheries and Aquaculture Science (Cefas)	Cefas SmartBuoy Marine Observational Network - UK Waters Phytoplankton Data 2001-2019	2001-2019
	Environment Agency (EA)	EA PHYTO 2000-2020	2000-2020
	Marine Biological Association (MBA)	CPR dataset 1960-2019	1960-2019
	Marine Scotland Science (MSS)	MSS Scalloway Phytoplankton dataset	2000-2018
		MSS Loch Ewe Phytoplankton	2000-2020
		MSS Loch Ewe zooplankton	2002-2017
		MSS Scapa Phytoplankton dataset	2000-2020
		MSS Stonehaven Phytoplankton	2000-2020
		MSS Stonehaven zooplankton	1999-2020
	Plymouth Marine Laboratory (PML)	PML_L4 phytoplankton	1992-2020
		PML_L4 zooplankton	1988-2020
	Scottish Association for Marine Science (SAMS)	SAMS-LPO-Phyto-Dec2021	1970-1981, 2000-2017

The data submitted by AU (DK), PML (UK), MSS (UK), NLWKN (DE) and IEO (ES) were from discrete fixed-point stations which were evaluated independently. The data submitted by SAMS (UK) were from multiple stations within a small area of freshwater influence. These data were aggregated and treated as a single fixed-point time series.

Data from the Continuous Plankton Recorder (CPR) survey, collected by the Marine Biological Association (MBA, UK), consisted of spatially distributed data collected along transects. The CPR survey, provides offshore open ocean data at a broad spatial scale using ships-of-opportunity (Reid *et al.*, 2003). Due to the distributed nature of CPR data, CPR samples are typically aggregated across a grid or a set of polygons at monthly temporal resolution (Bedford *et al.*, 2020a). The remaining fixed-point datasets, including VLIZ (BE), LLUR (DE), RWS (NL), IPMA (PT), SMHI (SE), Cefas (UK), and EA (UK), were also treated in this manner since samples were collected at several distributed stations within close proximity to one another. The dataset from IPMA (PT) only records abundances for the genera *Pseudo-nitzschia* (diatom) and *Dinophysis* (dinoflagellate), thus results from the Portuguese data should only be considered as a proxy for diatom and dinoflagellate lifeform abundances.

Data from the different providers were not combined for analysis due to differences in sampling, plankton analysis and enumeration methods. Instead, the datasets were analysed separately. Each dataset has internal QA/QC procedures to ensure consistency and accuracy of the data. Before total lifeform abundance values were \log_{10} transformed, a nominal value equivalent to half the minimum non-zero observed value for each time-series was added to each sample. All spatially distributed data (e.g., excluding AU, PML, MSS, IEO, NLWKN and SAMS) were averaged per month within each assessment unit. In a few cases, this resulted in localised groupings of samples being extrapolated across much larger assessment units (see distribution of CPR samples in Figure d), as was the case for CPR data along the west of Scotland (Intermittently Stratified 1) and Ireland (Atlantic Seasonally Stratified). For months when there were no samples within an assessment unit, gaps in the time series were filled by extracting a mean value from an inverse distance weighted interpolated surface generated from samples adjacent to the assessment unit (<250 nmi) from the same dataset. Years containing less than eight months of averaged and spatially interpolated values were discarded. Remaining gaps of three months or less were filled via linear interpolation. For the reporting of trends and linking to pressures, in cases where multiple suitable datasets contained samples from within the same assessment unit the trends displayed were based on the dataset with the greatest number of unique months (e.g., June 1978, February 2000, May 2014).

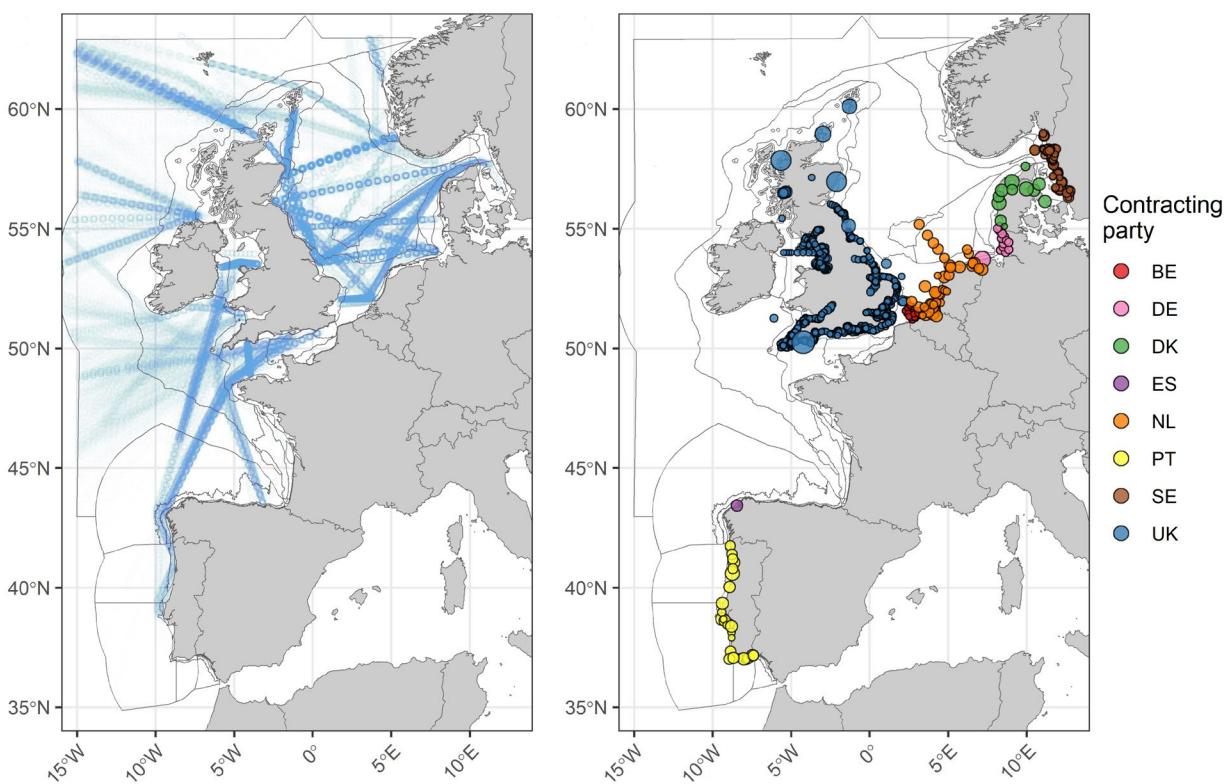


Figure d: CPR tracks and the locations of samples from all other datasets used in the assessment. For the locations, point size is proportional to the number of samples taken at each location. Points are coloured according to Contracting Party.

Confidence scoring:

A confidence scoring methodology, based on an approach developed by ICG-Eut to validate the output from their COMPEAT Tool, was applied to evaluate the robustness of reported trends for each plankton dataset for each assessment unit it intersected. For each assessment unit or fixed-point station temporal confidence

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was evaluated using two metrics, which are referred to here as specific temporal confidence (STC) and general temporal confidence (GTC).

STC was calculated on an annual basis as the proportion of months when samples were reported out of a total of 12 possible months. For example, if a dataset reported samples within an assessment unit only for June, July and August in a particular year, the STC for that year would be 0.25. GTC was calculated as the proportion of unique months with recorded samples out of the total number of potential months across the time-series. For example, a dataset commencing in January 2015 and ending in December 2019 contains a total of 60 potential months. If samples were reported for 30 of those months, the GTC would be 0.5. An overall temporal confidence score was then determined by calculating both the mean STC across all years and subsequently the mean of this value with the GTC.

Spatial confidence was also evaluated for distributed datasets such as the CPR. However, for fixed-point stations spatial confidence was not considered since there was no spatial element to the data. For each COMP4 assessment unit spatial confidence was calculated using two metrics, which similarly are referred to here as specific spatial confidence (SSC) and general spatial confidence (GSC).

SSC was calculated on an annual basis based on the proportion of longitudinal and latitudinal ranges of an assessment unit covered by the spatial extent of samples. For example, if an assessment unit spanned longitude from 1 to 5° E, and the distribution of samples collected within that assessment unit within a particular year ranged from 2 to 4° E, the proportion of longitudinal range covered by samples would be 0.5. The same procedure is also applied to the latitudinal range. The SSC for a particular year would be the mean of the longitudinal and latitudinal proportions covered by the spatial extent of samples.

GSC was derived using the same approach as SSC, but across all samples in a dataset that intersect with an assessment unit. An overall spatial confidence score was then determined by calculating the mean SSC across all years and subsequently calculating the mean of this value with the GSC.

Finally, an overall confidence score was calculated as the mean of temporal and spatial confidence. For fixed-point stations, spatial confidence was not considered, therefore only temporal confidence was used to represent the overall confidence of fixed-point station data.

Lifeform construction:

The eight plankton lifeforms highlighted in this assessment were chosen due to their ecological relevance and owing to the high confidence in their classification (**Table d**). All lifeforms were defined based on common ecological traits. The rationale for selecting the lifeforms and additional criteria containing supplementary information on lifeforms is listed in **Table e**.

Table d: Confidence in the definition of each lifeform and reasoning for low confidence where applicable (Ostle et al., 2021).

Lifeform	Confidence	Reason for confidence (where not 'high')
(micro)Phytoplankton	High	
Large microphytoplankton	Medium	Can reliably identify individual plankton species size class but cannot always reliably assign the size trait if the group counted spans taxa that are both larger and smaller than 20 microns.
Small microphytoplankton	Medium	
Diatoms	High	
Dinoflagellates	High	
Autotrophic and mixotrophic dinoflagellates	Medium	Can identify taxa, but assigning feeding mechanism trait is not always clear (see e.g.: discussion in Flynn et al., (2019))
Pelagic diatoms	High	
Tychopelagic diatoms	High	

Potentially toxic and nuisance diatoms	Low	Designation of some algal blooms as "harmful" (i.e.,: Harmful Algal Blooms, 'HABS'), relates more to societal assessment than plankton traits, these 'lifeforms' are therefore not currently recommended for use though they are defined in the traits list and will be the focus of future development work. Specific issues include: <ul style="list-style-type: none">The toxin producing diatom genus <i>Pseudo-nitzschia</i> contains both amnesic shellfish toxin-producers which can render shellfish unfit for human consumption and potentially negatively impact the health of marine mammals, and non-toxin-producing species/individuals. It is not possible to identify these cells to species level using routine light microscopy; some toxin and non-toxin producing species are morphologically identical.The genus <i>Alexandrium</i> contains both paralytic shellfish toxin- and non-toxin-producing species/strains and it is not possible to distinguish these using routine light microscopy; some toxin and non-toxin producing species are morphologically identical.The taxon <i>Karenia mikimotoi</i> forms high biomass blooms which strips the water column of oxygen which can be fatal to other lifeforms (Barnes et al., 2015).Not all datasets included in PLET reliably record key species (e.g.: CPR does not record <i>Alexandrium</i>)
Potentially toxic and nuisance dinoflagellates	Low	
Ciliates	Low	<ul style="list-style-type: none">Ecological function can be duplicated by mixotrophic dinoflagellates.Ciliates do not preserve well in the standard 0,5% Lugol's iodine preservative used to preserve phytoplankton samples and some (but not all) are too small to be well sampled by many of the datasets currently in PLET.
Holoplankton	Medium	
Meroplankton	Medium	<ul style="list-style-type: none">May not identify taxa specifically enough to determine traits.Some of the rarer species are resuspended from the seabed and definition of their holo- or meroplanktonic status is difficult
Gelatinous zooplankton	High	
Fish larvae/eggs	High	
Carnivorous zooplankton	Medium	Can identify taxa, but assigning diet trait is unclear, especially at different life stages.
Non-carnivorous zooplankton	Medium	
Crustaceans	High	
Small copepods	High	
Large copepods	High	

Table e: Each lifeform comprises organisms with particular ecological traits. A query is then used to assign individual species to lifeforms (McQuatters-Gollop et al., 2019).

Lifeform	Traits	Criteria
Diatoms	'Diatom' only	PhytoplanktonType=Diatom
Dinoflagellates	'Dinoflagellate' only	PhytoplanktonType=Dinoflagellate
Gelatinous zooplankton	'Gelatinous' only	PlanktonType=Zooplankton AND Gelatinous=Y
Fish larvae/eggs	'Fish' only	ZooType=Fish
Carnivorous zooplankton	'Carnivore' only	PlanktonType=Zooplankton AND Diet=Carnivore
Non-carnivorous zooplankton	'Zooplankton' AND either 'Herbivore', 'Omnivore', OR 'Ambiguous'	PlanktonType=Zooplankton AND (Diet=Herbivore OR Omnivore OR Ambiguous)
Crustaceans	'Crustacean' only	Crustacean=Y
Large phytoplankton	'Phytoplankton' AND 'Lg'	PlanktonType=Phytoplankton AND PhytoplanktonSize=Lg
Small phytoplankton	'Phytoplankton' AND 'Sm'	PlanktonType=Phytoplankton AND PhytoplanktonSize=Sm
Phytoplankton	'Phytoplankton' only	PlanktonType=Phytoplankton
Autotrophic and mixotrophic dinoflagellates	'Dinoflagellate' AND either 'Auto' OR 'Auto/Mixo'	PhytoplanktonType=Dinoflagellate AND (FeedingMech=Auto OR Auto/Mixo)
Pelagic diatoms	'Diatom' AND 'Pelagic'	PhytoplanktonType=Diatom AND DiatomDepth=Pelagic
Tychopelagic diatoms	'Diatom' AND 'Tychopelagic'	PhytoplanktonType=Diatom AND DiatomDepth=Tychopelagic
Nuisance and toxin-producing diatoms	'Diatom' AND either 'Toxic' OR 'Nuisance'	PhytoplanktonType=Diatom AND (HAB = Toxic)
Nuisance and toxin-producing dinoflagellates	'Dinoflagellate' AND either 'Toxic' OR 'Nuisance'	PhytoplanktonType=Dinoflagellate AND (HAB = Toxic)
Holoplankton	'Holoplankton' only	Habitat=Holoplankton
Meroplankton	'Meroplankton' only	Habitat=Meroplankton
Large copepods	'Copepod' AND 'Lg'	Copepod=Y AND ZooSize=Lg
Small copepods	'Copepod' AND 'Sm'	Copepod=Y AND ZooSize=Sm
Ciliates	'Ciliate' only	PhytoplanktonType=Ciliate

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Microflagellates	'Dinoflagellate' AND 'Sm'	PhytoplanktonType=Dinoflagellate AND PhytoplanktonSize=Sm
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The database master species list (Ostle *et al.*, 2021) was built by assigning functional traits to each species for a dataset and then adding additional new datasets to expand the master species list (**Figure e**). The species list for each new dataset was assigned a unique Aphia ID via the World Register of Marine Species (WoRMS). The new dataset's species list was then compared with the plankton database's master species list via Aphia IDs and any new species were identified. This process ensures that each species is only entered in the database once. The new species were then manually assigned functional traits by searching the literature; fields were left blank where functional traits for species were unknown. Once traits were assigned, the new species were added to the master species list. Queries were constructed (**Table e**) to build lifeforms from the functional trait information (McQuatters-Gollop *et al.*, 2019).

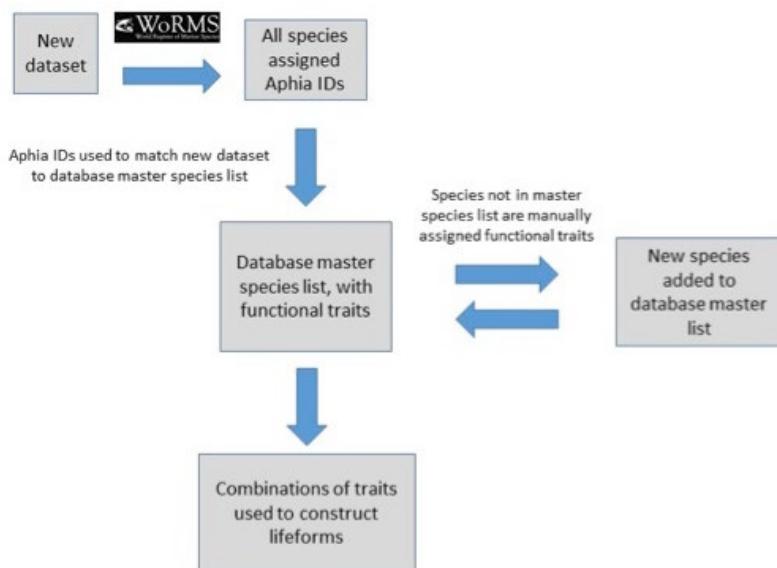


Figure e: Schematic illustrating the process undertaken to assign functional traits to species, then species to lifeforms. Each species must first be assigned a unique Aphia ID to determine whether it is already present in the master species list (McQuatters-Gollop *et al.*, 2019).

A simple method of confidence assessment was applied for each lifeform (**Table f**). Using expert opinion, each lifeform was evaluated on two characteristics: the ability to identify and speciate organisms in that lifeform using light microscopy and the understanding of the accuracy of determining traits assigned to the lifeform. For example, medium confidence is assigned to the lifeforms 'autotrophic dinoflagellates' and 'mixotrophic dinoflagellates' because the mode of nutrition of many dinoflagellate species is currently uncertain (Flynn *et al.*, 2019). Thus, the accuracy of assigning the lifeform category is medium. Likewise, the lifeform 'non-carnivorous zooplankton' has a medium confidence designation since the feeding habits of many abundant and common zooplankton species remain not strictly defined. Further investigation must also be conducted to decide whether both harmful-bloom-forming algae and potentially toxin-producing lifeforms should be considered in future assessments of this indicator. Work is ongoing to increase confidence in lifeforms, however, this work is resource dependent.

Table f: Matrix used to determine the confidence in each lifeform. Only pairs with high confidence were used for this assessment (Ostle *et al.*, 2021).

	Easy to ID/speciate	Difficult to ID/speciate
Known traits	High	Low
Unknown traits	Low	Low

Assessing links to pressures:

The Boruta algorithm (Kursa and Rudnicki, 2010) was applied to evaluate which environmental variables were the best predictors of lifeform abundance. The Boruta algorithm is a tree-based permutational variable

importance method which uses a wrapper around random forest to evaluate the predictive performance of each variable being tested against a shuffled set of the same predictor variables. If a variable achieves better predictive accuracy than the highest-performing shuffled variable, it is determined to be important and is assigned a score based on the mean decrease in predictive accuracy when the variable is excluded from the model.

For each unique combination of dataset, assessment unit, and lifeform whenever a significant Kendall statistic (i.e., a significant change in lifeform abundance over the time-period assessed) was observed, a separate permutational variable importance process based on random forest (Boruta algorithm) was conducted to evaluate relationships between lifeform abundance and a set of 16 environmental variables described in **Table g**. Random forest is robust to detecting separate effects from collinear predictors, since it uses an ensemble approach based on agreement from many trees which each test a different subset of the predictor variables. Based on the results of the previous assessment, a table of implicating factors was developed, linking lifeforms with a set of environmental pressures with plausible links to variation in lifeform abundance (**Table h**). Only variables indicated in **Table h** were tested for each lifeform.

Table g: Environmental variables and their descriptions and sources evaluated in the random forest permutational variable importance process.

Variable name	Variable description	Source
Sea surface temperature	Temperature of surface layer, as measured by satellite	International Comprehensive Ocean-Atmosphere Data Set (ICOADS): https://psl.noaa.gov/data/gridded/data.coads.1deg.html
Salinity	Salinity of the surface layer	European Regional Seas Ecosystem Model (ERSEM; NWSHELF_MULTIYEAR_PHY_004_009): https://doi.org/10.48670/moi-00059);
Total oxidised nitrogen	Total oxidised nitrogen concentration in surface layer	In situ data from Marine Scotland Science (MSS): https://doi.org/10.7489/1881-1 and Aarhus University (Svendsen et al., 2005)
Nitrate	Nitrate concentration in surface layer	European Regional Seas Ecosystem Model (ERSEM; NWSHELF_MULTIYEAR_BGC_004_011): https://doi.org/10.48670/moi-00058); In situ data from Plymouth Marine Laboratory (PML): https://www.westernchannelobservatory.org.uk/l4_nutrients.php
Phosphate	Dissolved inorganic phosphate concentration in surface layer	European Regional Seas Ecosystem Model (ERSEM; NWSHELF_MULTIYEAR_BGC_004_011; https://doi.org/10.48670/moi-00058); In situ data from Plymouth Marine Laboratory (PML): https://www.westernchannelobservatory.org.uk/l4_nutrients.php and Marine Scotland Science (MSS): https://doi.org/10.7489/1881-1
Total phosphorous	Total dissolved phosphorous concentration in surface layer	In situ data from Aarhus University (Svendsen et al., 2005)
N:P ratio	The ratio of molar nitrogen concentration to molar phosphorous concentration	Calculated from European Regional Seas Ecosystem Model (ERSEM; NWSHELF_MULTIYEAR_BGC_004_011): https://doi.org/10.48670/moi-00058); In situ data from Plymouth Marine Laboratory (PML): https://www.westernchannelobservatory.org.uk/l4_nutrients.php , Marine Scotland Science (MSS): https://doi.org/10.7489/1881-1 and Aarhus University (Svendsen et al. 2005)
Silicate	Dissolved silicate concentration in surface layer	In situ data from Marine Scotland Science (MSS): https://doi.org/10.7489/1881-1 , Plymouth Marine Laboratory (PML): https://www.westernchannelobservatory.org.uk/l4_nutrients.php and Aarhus University (Svendsen et al., 2005)
Wind speed	Wind speed (a proxy for turbulence)	International Comprehensive Ocean-Atmosphere Data Set (ICOADS): https://psl.noaa.gov/data/gridded/data.coads.1deg.html
Mixed layer depth	The depth below the surface where the steepest change in density occurs (the thickness of the surface layer in which photosynthesis can occur)	European Regional Seas Ecosystem Model (ERSEM; NWSHELF_MULTIYEAR_PHY_004_009): https://doi.org/10.48670/moi-00059

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Light attenuation	The extinction coefficient for visible light, or the decrease in the intensity of solar radiation with depth (a proxy for the opacity of the water column)	European Regional Seas Ecosystem Model (ERSEM); NWSHELF_MULTIYEAR_BGC_004_011; https://doi.org/10.48670/moi-00058
Precipitation	Rate of precipitation (a proxy for freshwater input)	International Comprehensive Ocean-Atmosphere Data Set (ICOADS); https://psl.noaa.gov/data/gridded/data.coads.1deg.html
Current velocity	Current velocity in surface layer (a proxy for horizontal transport which affects how quickly organisms are advected to adjacent systems)	European Regional Seas Ecosystem Model (ERSEM); NWSHELF_MULTIYEAR_PHY_004_009; https://doi.org/10.48670/moi-00059
Atlantic Multidecadal Oscillation (AMO)	The Atlantic Multidecadal Oscillation, also known as Atlantic Multidecadal Variability, is the theorised variability of the sea surface temperature of the North Atlantic Ocean on the timescale of several decades	National Oceanic and Atmospheric Administration (NOAA); https://psl.noaa.gov/data/timeseries/AMO/
North Atlantic Oscillation (NAO)	The North Atlantic Oscillation is a weather phenomenon over the North Atlantic Ocean of fluctuations in the difference of atmospheric pressure at sea level between the Icelandic Low and the Azores High	National Oceanic and Atmospheric Administration (NOAA); https://www.ncdc.noaa.gov/teleconnections/nao/
pH	Sea water pH reported on total scale	European Regional Seas Ecosystem Model (ERSEM); NWSHELF_MULTIYEAR_BGC_004_011; https://doi.org/10.48670/moi-00058

Table h: The relevant environmental variables (✓) tested for each lifeform where data were available. The categorisation of variables as displayed in Figure 3 is indicated under the column “Variable in Figure 3”.

	North Atlantic Oscillation	✓	✓	✓	✓	✓	✓	✓	✓	✓
pH	pH	✓	✓	✓	✓	✓				

Prior to any manipulation of data, lifeform time-series were divided into separate training and testing sets. The training data were used for variable selection and for generating the random forest models, while the testing data were used to validate model predictive accuracy. For the purposes of this analysis, the training data were limited to all months prior to the assessment period (e.g., 1960 to 2014 for CPR data) and the testing data were restricted to the assessment period itself (e.g., 2015 to 2019).

This analysis covered the full temporal extent of each plankton dataset (1960 to 2019 in the case of CPR data). While gridded data from NOAA were available from 1960 to 2019, the longest duration modelled dataset for nutrients only spanned from 1993 to 2019, with in-situ nutrient datasets commencing as early as 1980. To evaluate long-term links to pressures and avoid excluding the first several decades of many plankton time-series from our analysis the ‘missRanger’ package for R (Mayer and Mayer, 2019) was used to perform multiple imputation by chained random forests. This method uses multiple random forest regressions to impute missing values based on collinearities among observed values in the predictors. For each variable containing missing values the algorithm generates a separate regression model based on all the other predictors. To improve imputation performance, a numeric variable representing ‘month’ was included in this step to better predict the consistent seasonal patterns in some variables. Missing values in the predictors were imputed separately for the training and testing datasets. It is important to note that no imputation of lifeform abundance time-series was performed to avoid introducing greater uncertainty into the process of linking to pressures.

All variables including lifeform abundance were smoothed to remove seasonality and uncover long-term trends by calculating the mean value across a 12 month moving window (Figure f). This step required the first and last six months of data from training and testing sets to be excluded from the analysis.

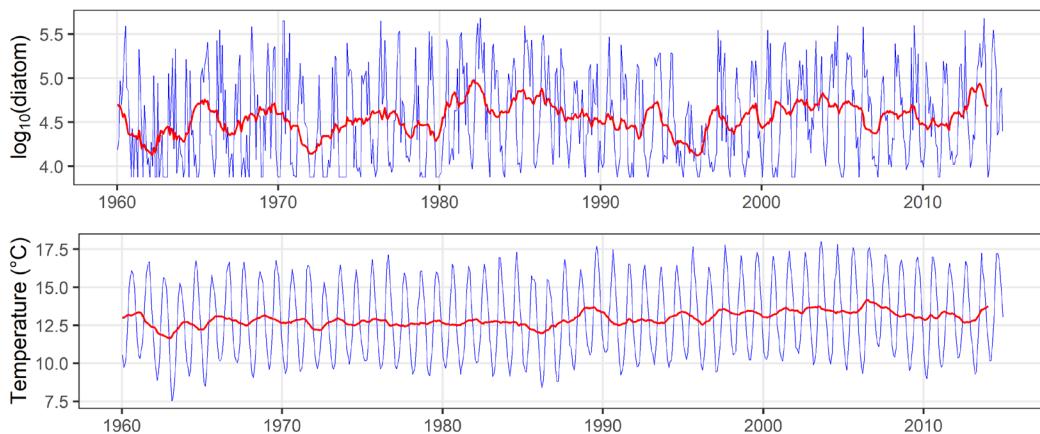


Figure f: Long-term time-series (1960 to 2019) for diatom abundance from CPR, and sea surface temperature from ICOADS, for the ‘Channel well mixed’ COMP4 assessment unit. Each plot displays monthly mean values (blue line) and 12-month moving average or monthly long-term trend (red line) in each time-series.

Values for each variable were calculated as the mean of monthly mean gridded values (modelled and remotely sensed; Table g) intersecting each COMP4 assessment unit. For fixed-point stations, mean values were calculated from all measurements within a 5-nautical mile radius of the station. Where in-situ data were available (total nitrogen, nitrate, phosphate, total phosphorous, silicate) they were evaluated instead of the

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modelled environmental variables. For Atlantic Multidecadal Oscillation (AMO) and North Atlantic Oscillation (NAO), monthly values were applied identically across all assessment units since these variables have basin-scale influence likely to cover most of the assessment region.

Environmental variables were grouped into ten categories to simplify visualisation in map format (**Table h**). Categorisations of the most important predictor variable for each model, or the predictor variable which resulted in the greatest mean decrease in accuracy when it was removed, were reported across a map. Instances when the validation step with the testing data indicated significant positive correlation ($p \leq 0,05$) between predicted and observed lifeform abundance values were also indicated in this map to provide greater confidence in the variable importance results and to avoid reporting spurious relationships.

It is important to note that observing high importance of any environmental variable is simply an indication that it co-varied with lifeform abundance. Our approach cannot imply causation since it is impossible to hold all other potential influences on lifeform abundance constant, as would be the case in a controlled experimental setting.

Integration of indicator results:

A primary objective of this indicator assessment was to integrate results to facilitate an understanding of changes occurring across pelagic habitat types within OSPAR Regions II, III and IV. This required indicator results for each OSPAR Region to be integrated according to the following pelagic habitat categories: variable salinity, coastal, shelf, and oceanic / beyond shelf. This categorisation of COMP4 assessment units and fixed-point stations is described in **Table a** and **Table b**. To meet this objective, the focus was on the primary direction of change detected across assessment units and fixed-point stations within each pelagic habitat category for each of the 8 high confidence plankton lifeforms highlighted in this assessment. The mean confidence, spatial representativeness were then reported, and most likely links to environmental pressures.

As an example, changes in dinoflagellate abundance were assessed across 4 COMP4 assessment units and 4 fixed-point stations representing variable salinity habitats within OSPAR Region II. If 1 decreasing trend, 4 increasing trends, and 3 instances of no trend were detected across these locations, an increasing net trend would be reported and the proportion of assessment units studied where this trend was detected, in this case 0,5.

To report the level of confidence in this result, the mean confidence score for locations where dinoflagellate abundance was increasing, was calculated. COMP4 assessment units and fixed-point stations as equivalent were considered for this integration.

To report the spatial representativeness of the result, the proportion of the total number of COMP4 assessment units considered in the analysis was calculated, in this case 8, out of the total number of possible COMP4 assessment units representing variable salinity habitats within the OSPAR Region, in this case 16. Therefore, the spatial representativeness of the result would be 0,5. Note that fixed-point station datasets do not contribute to this score.

Finally, to report links to environmental pressures which can drive changes in lifeform abundance for the net trend, environmental variables for each location based on their relative variable importance were ranked, with 1 assigned to the variable with highest importance, 2 to the variable with second highest importance and so on. For locations where the net trend was increasing, the mean rank of each environmental variable was calculated and the variable with the lowest mean rank reported.

Assessing the status of pelagic habitats:

To assign a designation of assessment status to pelagic habitats based on the integration of indicator results, a semi-quantitative methodology described in McQuatters-Gollop *et al.*, (2022) was applied, which was developed from the lessons gained from the previous OSPAR assessment (IA 2017). In this case, the status of a habitat type can be designated as either "Good", "Unknown", "Not good", or "Not assessed" (**Table i**).

Following the criteria outlined in this study, if a pelagic habitat has been assessed, it should by default be considered as either “Unknown” or “Not Good”. At this stage it is not realistic to assign “Good” status to pelagic habitats, since it is difficult to develop meaningful assessment thresholds for plankton and generally not possible to determine whether a particular state is desirable or undesirable, except under specific circumstances such as eutrophication. Following this logic, the status of pelagic habitats should be considered “Unknown” by default. In cases when change has been detected and that change can be confidently linked to the impact of an anthropogenic pressure, the status of this habitat is “Not good”.

Table i: Biodiversity status categories and colours used for the interpretation, by expert judgement, of indicator biodiversity state (McQuatters-Gollop *et al.*, 2022).

Not good	Indicator value is below assessment threshold, or change in indicator represents a declining state, or indicator change is linked to increasing impact of anthropogenic pressures (including climate change), or indicator shows no change but state is considered unsatisfactory
Unknown	No assessment threshold and/or unclear if change represents declining or improving state, or indicator shows no change but uncertain if state represented is satisfactory
Good	Indicator value is above assessment threshold, or indicator represents improving state, or indicator shows no change but state is satisfactory
Not assessed	Indicator was not assessed in a region due to lack of data, lack of expert resource, or lack of policy support.

The status at the level of each of the 8 high confidence lifeforms within each pelagic habitat within OSPAR Regions was considered and the results integrated for multiple lifeforms to assign a single quality status designation for the pelagic habitat type. For the status of a lifeform to be shifted from “Unknown” to “Not good” the results of the integration had to meet certain criteria:

- The net trend should be either increasing or decreasing and must be present in at least 0,5 of the locations assessed.
- The mean confidence for locations considered for the determination of the net trend must be at least 0,5.
- The spatial representativeness of the assessed locations out of the total number of possible locations for that habitat type must be at least 0,5.
- The environmental pressure with the greatest mean rank for locations expressing the net trend must represent either sea surface temperature, pH, or nutrients.
- The mean rank of the most important environmental pressure must be ≤ 3 , indicating that across all locations the variable ranks in the top 3 most important for predicting the abundance of the lifeform.

If all the above criteria are met, the lifeform is assigned a status of “Not good”. If 25% or more of assessed lifeforms within a pelagic habitat type are assigned a status of “Not good” then the whole habitat type is also assigned a status of “Not good”. Otherwise, the habitat type is assigned a status of “Unknown”.

Lifeform pairs indicator approach:

Although this assessment focused primarily on long-term trends in lifeform abundance due to their stronger links with environmental pressures at large spatial scales, the lifeform pairs indicator was also evaluated for the current assessment (which constitutes the PH1 indicator as originally defined in IA2017).

Tett *et al.*, (2008) proposed to track changes in the state of the plankton community by means of plots in a state space and calculating a Plankton Community Index, referred to here as a Plankton Index (PI). The conceptual framework is that ecosystems can be viewed as systems with an instantaneous state defined by values of a set of system state variables which are attributes of the system that change with time in response to each other and external conditions. Building on this approach and plotting plankton lifeform abundance

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in a multi-dimensional state space provides a means of monitoring changes in the structure of plankton communities. A state can be defined as a single point in state space, with coordinates provided by the values of the set of state variables, in this case two lifeform abundances, which together are used as a pair.

The plankton community index is calculated by initially mapping relative plankton lifeform abundances from an assessment period in a state space. The distribution of lifeform abundances within the state space is then used to define an envelope, representing the prevailing conditions for the assessment period. Finally, a set of abundance values from a comparison period are mapped onto the state space to evaluate the plankton community index, which represents the proportional similarity of the two periods.

Mapping lifeforms in a state space

In the example illustrated in **Figure g** for diatoms and dinoflagellates, the axes of the two-dimensional space are the abundance values of the two lifeforms. Each point represents the state of the ecosystem in terms of the two lifeforms at the time of sample collection. Subsequent samples yield additional pairs of abundance values that can be mapped onto the lifeform state space.

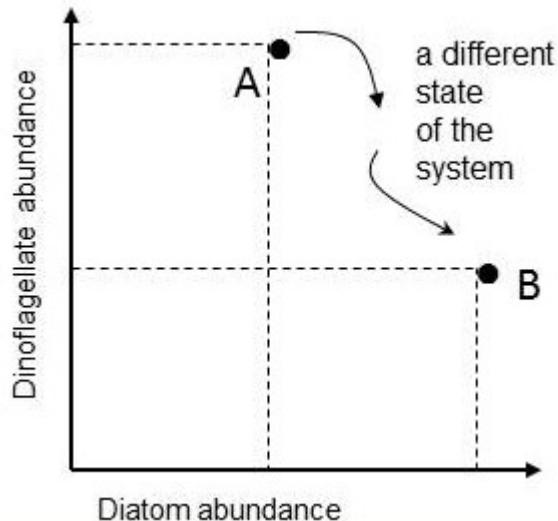


Figure g: Mapping the abundance of two lifeforms in state space

Point A is the ecosystem state at the instant a water sample was taken and is characterised by the abundance values of two lifeforms. Another sample, taken in the same location, yields abundance values that map to a different point in the diatom-dinoflagellate state space (point B).

The path between the two states is called a trajectory, and the plankton condition is defined by the trajectory drawn in the state space by a set of points. Such trajectories reflect: (i) cyclic and medium-term variability (the higher order consistencies in the plankton that result from seasonal cycles, species succession and interannual variability); and (ii) long-term variability that might result from environmental pressure. The seasonal nature of plankton production and the succession of species in seasonally stratifying seas, result in this trajectory tending in a certain direction (as plankton growth increases in spring and declines during autumn), such that the trajectory tends towards its starting point. Given roughly constant external pressures, the data collected from a particular location over a period of years form a cloud of points in state space that can be referred to as a regime. Long-term variability may show a persistent trend of movement away from a starting point in state space.

Approach for defining the envelope

To define a regime, an envelope can be drawn about this group of points using a convex hull method. Because of theoretical arguments that the envelope should be doughnut-shaped with a central hole (Tett *et al.*, 2008), bounding curves can be fitted outside and inside the cloud of points (**Figure h**). The data are from the CPR

dataset for the ‘Channel well mixed’ COMP4 assessment unit. The colour of each point corresponds to the season within which it was sampled; blue = winter, green = spring, yellow = summer, red = autumn.

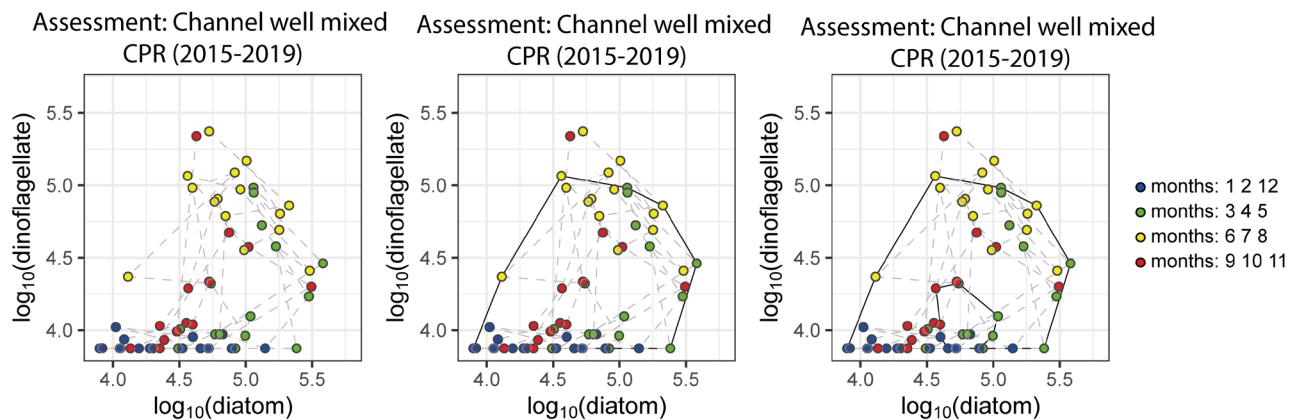


Figure h: Creating the envelope in three steps, from left to right: An example of a regime defined by the envelope drawn by the convex hull method

The size and shape of the envelope are sensitive to sampling frequency and the total number of samples. Envelopes are made larger by including extreme outer or inner points, and the larger the envelope, the less sensitive it will be to change in the distribution of points in state space and therefore to detect a change in condition. Conversely, if too many points are excluded the envelope will be small and even minor changes will result in a statistically significant difference. It is therefore desirable to exclude a proportion (p) of points, to eliminate these extremes, and so the 90-percentile was used. Envelopes are therefore drawn around the cloud of points to include a proportion ($p=0,9$) of the points, excluding the 5% of points that were most distant from the cloud's centre and the 5% of points that were closest to the centre.

For a Plankton Index to be calculated, it is necessary to establish a set of conditions as the basis for making comparisons. For this assessment the Plankton Index was calculated by using the assessment period (2015–2019) to define the envelope. For each dataset, all measurements collected prior to the assessment period were used for comparison. For the CPR data this meant the envelope was defined with samples from 2015–2019 and samples from 1960–2014 were used for comparison. This made it possible to maintain the same range of years to define the envelope across multiple datasets with different temporal coverage. Like the Kendall statistic, the PI was calculated independently for each combination of dataset, assessment unit and lifeform pair.

In the example shown in **Figure i**, CPR data collected from the ‘Channel well mixed’ COMP4 assessment unit during the assessment period (2015–2019), were used to create an envelope. The envelope, thus drawn (**Figure i**) defines a domain in state space that contains a set of trajectories of the diatom-dinoflagellate component of the pelagic ecosystem.

Calculating the plankton community index

The next step is to map a new set of data onto the same state space for comparison (**Figure i**). The value of the PI is the proportion of new points that fall between the inner and outer envelopes. In this example, 30% or 198 of the 660 new points lie outside, and the PI is 0,7. A value of 1,0 would indicate no change, and a value of zero would show complete change. The envelope was made by excluding 10% of points, so 66 (10% of 660) points are expected to fall outside. The exact probability of getting 198 by chance alone when only 66 are expected, can be calculated using a chi-square calculation (with 1 df and a 1-tail test). The value of 0,7 is significantly less than the expected value of 0,9, and so the difference between the two periods is statistically significant ($p < 0,01$).

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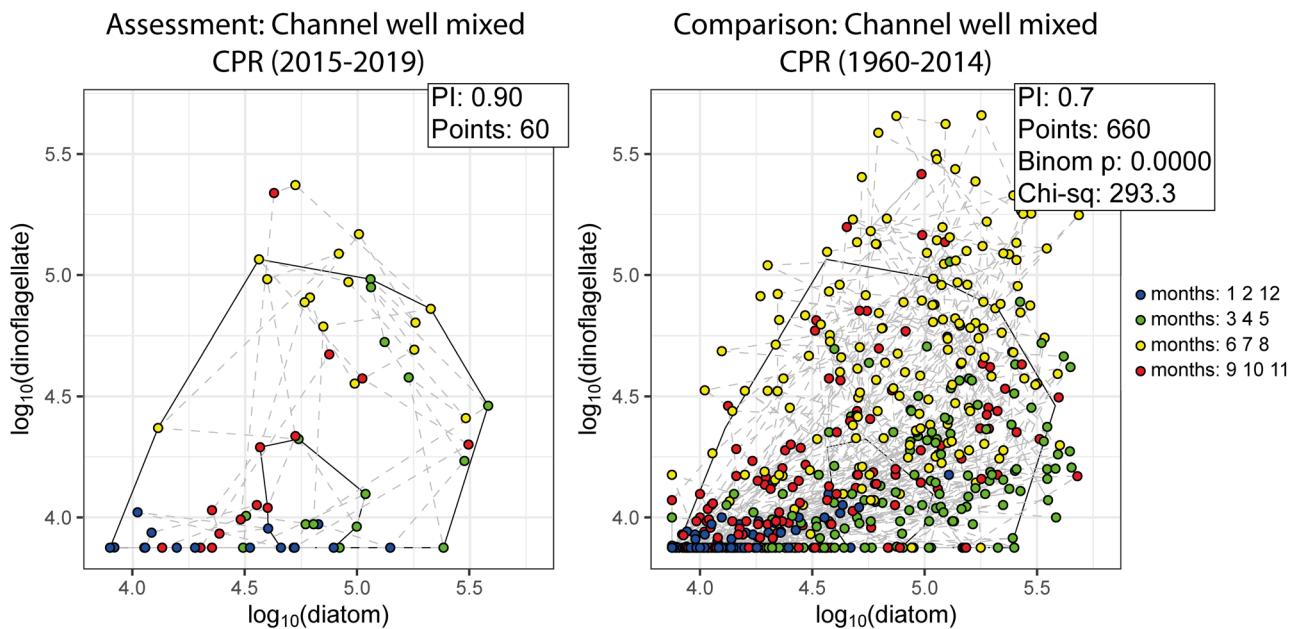


Figure i: An example for the comparison between assessment and comparison periods within the ‘Channel well mixed’ COMP4 assessment unit, with representing the assessment period of 2015-2019 and representing the comparison period of 1960-2014.

Results

Long-term trends (1960-2019; **Figure 2**) indicate that most plankton lifeforms, including diatoms, dinoflagellates, holoplankton, fish larvae/eggs, and large (adult ≥ 2 mm) and small (adult < 2 mm) copepods are declining in abundance throughout the North-East Atlantic. Spatial patterns in the direction of change for both small and large copepods closely match those of the diatoms. Conversely, meroplankton demonstrate a pattern of increasing abundance or no change throughout all assessment units and all but one fixed-point station. Patterns apparent from the widely distributed CPR data are reflected by the fixed-point stations from adjacent transitional waters, except in the case of dinoflagellates, which demonstrate an increasing trend in transitional waters of Scotland (MSS, United Kingdom) and Germany (NLWKN, Germany). This discrepancy may be due to poor preservation of athecate dinoflagellates in CPR samples, which are better preserved in fixed-point station datasets.

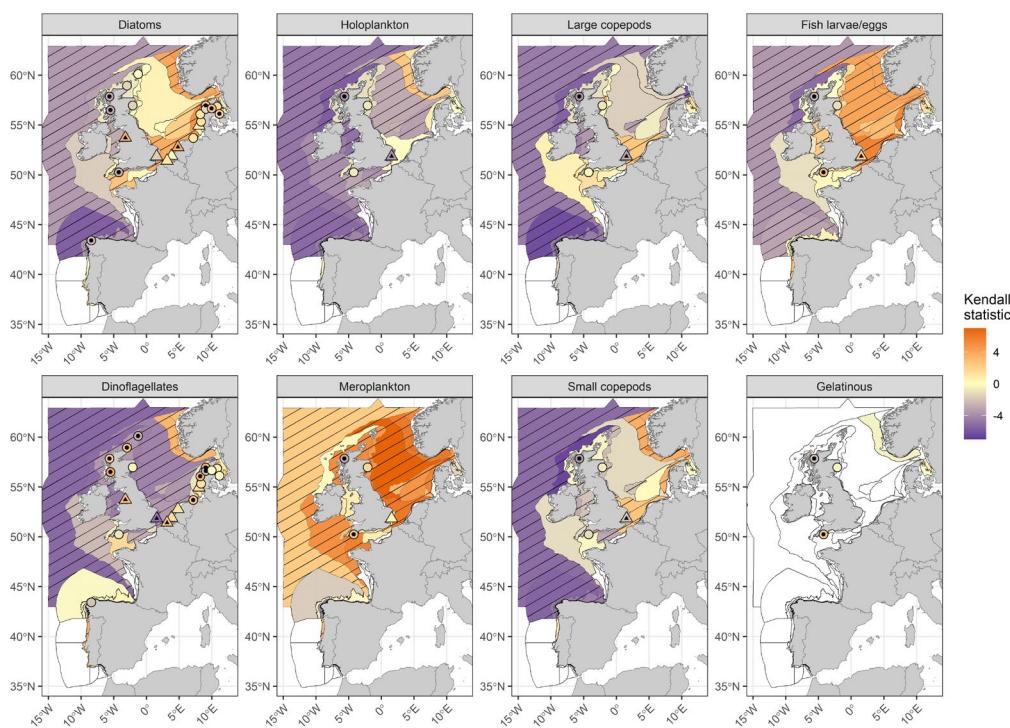


Figure 2: Kendall statistics for eight plankton lifeforms displayed over COMP4 assessment units and fixed-point stations in the Greater North Sea, Celtic Seas, Bay of Biscay and Iberian Coast. River plumes are represented as triangles. Assessment units are coloured according to the results of the Kendall trend test, which indicate the magnitude of long-term increase (> 0) and decrease (< 0) in lifeform abundance from 1960-2019 (or a shorter period for fixed-point stations). Patterned assessment units, fixed-point stations and river plumes with an internal black symbol indicate statistically significant change ($p \leq 0.05$). Assessment units filled in white indicate insufficient data to evaluate a trend.

Over the current assessment period (2015 to 2019), CPR data indicates dinoflagellates have continued to decline in abundance in the Northern North Sea (see Results (Extended); Figure j). Holoplankton also show evidence of continued decline in the Atlantic Seasonally Stratified assessment unit. For other lifeforms and assessment units, changes in abundance in line with long-term trends were not detectable within the five-year assessment period, however, five years is likely insufficient for a robust assessment of environmental change. Newly apparent trends which have emerged within the current assessment period include declining abundance of large copepods in the Atlantic Seasonally Stratified assessment unit, increasing fish larvae/eggs in the Scottish Sea and Coastal UK 1 regions, and declining diatom abundance at Norderney (NLWKN, Germany).

Changes in lifeform abundance were linked to variation in environmental pressures, acting both externally (e.g., precipitation, wind speed) and internally (e.g., pH, water temperature) on the marine environment. Many of these pressures are influenced indirectly by climate change (Figure 3). Modelling results have shown that increased sea surface temperatures were linked to declining abundances of plankton lifeforms, particularly small and large copepods in the Atlantic. Change in water temperature was also strongly linked to increasing meroplankton abundance within the Eastern North Sea. Impacts of increasing temperature were most frequently detected in shelf and oceanic pelagic habitats. While increasing temperatures can have direct impacts, they can also be linked to greater stratification and resulting nutrient limitation.

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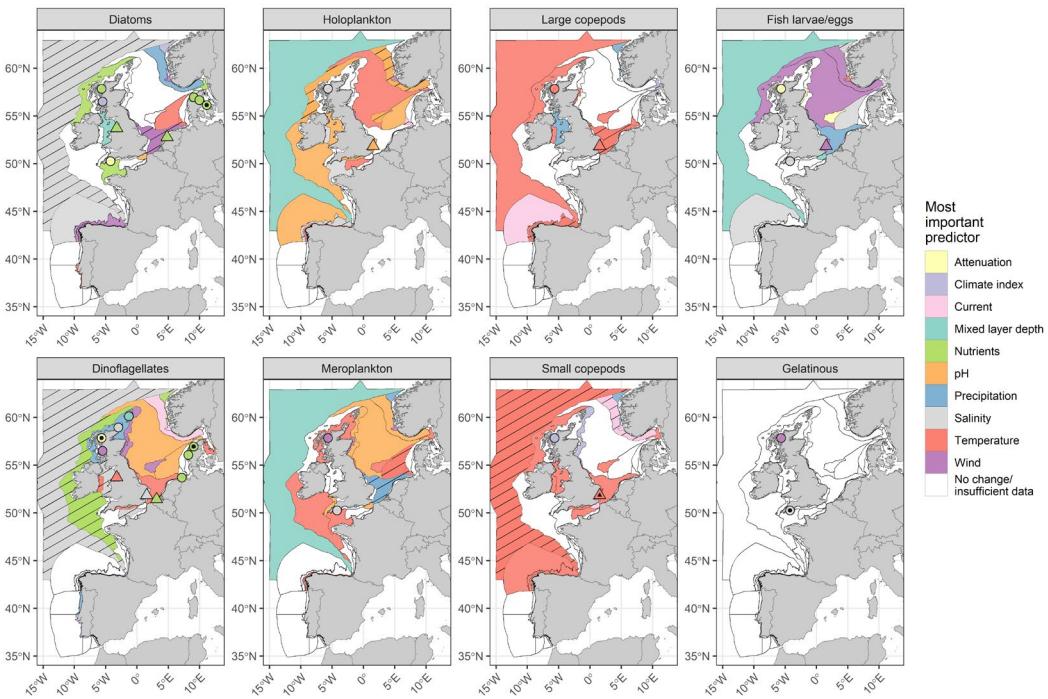


Figure 3: COMP4 assessment units for eight plankton lifeforms, coloured by a categorisation of the most important environmental variable to predict lifeform abundance (see key in Table c). Patterned assessment units, fixed-point stations and river plumes with an internal black symbol indicate statistically significant correlation between predicted and observed testing data, indicating greater reliability of reported results. Assessment units filled in white indicate no change in lifeform abundances or insufficient data to evaluate a trend. Fixed-point stations and river plumes are only displayed where a trend in lifeform abundance is present.

Links to nutrient concentration were less clear since it is difficult to assess the impact of nutrient concentrations on lifeforms due to the lag period between occurrence and phytoplankton uptake and assimilation. However, model results suggest that they were more apparent in coastal regions. Changes in diatom and dinoflagellate abundance in these assessment units were mainly linked to increases in the N:P ratio, driven by reductions in phosphorous concentrations. Of the 182 significant trends in lifeform abundance assessed, 78% did not indicate statistically significant predictive ability of models based on environmental variables (i.e., $p > 0,05$).

Results (Extended)

Statistically significant continuation of long-term trends was only apparent in a few assessment units when the Kendall statistic was calculated for the assessment period only (2015 to 2019; **Figure j**), owing at least partially to the reduction in statistical power associated with detecting trends across such short time periods. Longer assessment periods increase the detection of many of the important changes which are occurring in plankton lifeforms, due to high inter-annual variability in lifeform abundance and gradual rates of change over time. Increasing the temporal extent also increases the detection of change because data from more historic periods are included (Bedford *et al.*, 2020a; Bedford *et al.*, 2020b)

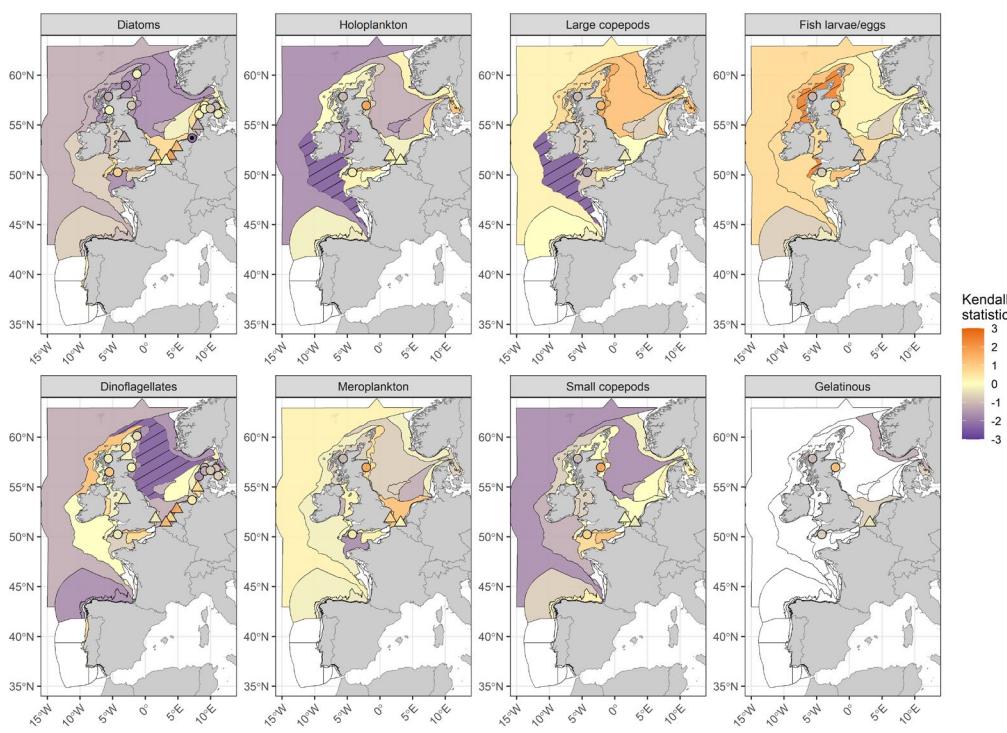


Figure j: Kendall statistics for eight plankton lifeforms displayed over COMP4 assessment units and fixed-point stations in the Greater North Sea, Celtic Seas, Bay of Biscay and Iberian Coast. River plumes are represented as triangles. Assessment units are coloured by the results of the Kendall trend test, which indicate the magnitude of increase (> 0) and decrease (< 0) in lifeform abundance from 2015–2019. Patterned assessment units, fixed-point stations and river plumes with an internal black symbol indicate statistically significant change ($p \leq 0.05$). Assessment units filled in white indicate insufficient data to evaluate a trend.

The lifeform pairs indicator (PI), which indicates the degree of similarity in the co-abundance of lifeform pairs (Figure k) between the assessment period (2015 to 2019) and the comparison period (1960 to 2014) reveals that changes in diatoms and dinoflagellates were greatest in coastal waters surrounding the United Kingdom and within the Norwegian Trench, driven by reduced summer and autumn abundances of dinoflagellates. Diatoms and dinoflagellates showed the most stability within the Channel well mixed, Meuse and Elbe plume assessment units. Fixed-point stations, particularly those around Scotland, did not always reflect patterns in offshore CPR data. For gelatinous zooplankton and fish larvae/eggs there were insufficient records of gelatinous zooplankton to form any conclusions, however there was evidence of important changes in the Norwegian Trench and Kattegat Coastal assessment units. For holoplankton and meroplankton the greatest changes were along the Atlantic on the northern coast of Spain (Noratlantic Area NOR-NorO1) and in the Norwegian Trench. Widespread changes in the Dogger Bank, Eastern and Southern North Sea assessment units were caused by increasing trends in holoplankton winter abundance. Large and small copepods demonstrated a similar pattern to that of holoplankton and meroplankton. The PI also showed poor overlap with patterns in the Kendall statistic for individual lifeform abundances, which indicates there were important changes to seasonal trajectories which did not contribute to detectable trends in mean annual abundance.

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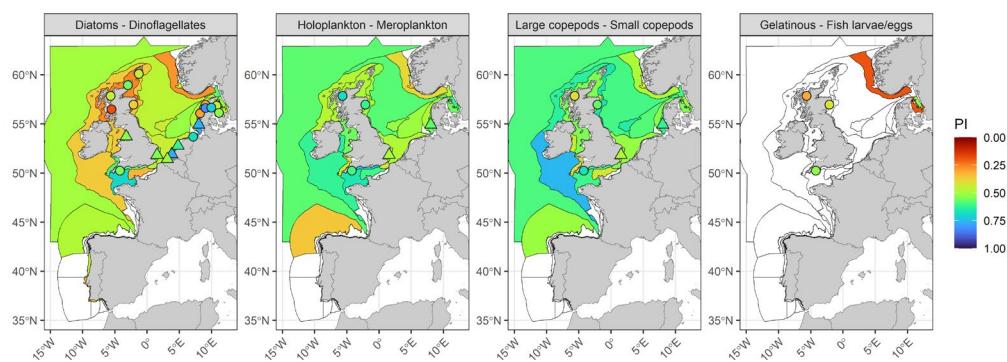


Figure k: Plankton lifeform pairs indicator (PI) for four lifeform pairs displayed over COMP4 assessment units and fixed-point stations in the Greater North Sea, Celtic Seas, Bay of Biscay and Iberian Coast. River plumes are represented as triangles. Assessment units are coloured according to the PI with an assessment period of 2015-2019 and a comparison period ranging from the beginning of each time-series up until the end of 2014. The PI indicates the proportion of months in the comparison period which fall within the assessment envelope. Assessment units filled in white indicate insufficient data to evaluate the PI.

The integration of the indicator results across pelagic habitat categories (**Figure c**) indicate that meroplankton in coastal and shelf areas of the Greater North Sea (**Table j**) are increasing in abundance and the increasing trends are closely linked to rising sea surface temperatures. Simultaneous declines in the abundance of holoplankton in shelf regions have also been linked to temperature increase.

Table j: Integration of the indicator results for OSPAR Region II – Greater North Sea. Column names are described as follows: ↓: the number of COMP4 assessment units and fixed-point stations where decreasing trends have been detected, -: the number of COMP4 assessment units and fixed-point stations where no trends have been detected, ↑: the number of COMP4 assessment units and fixed-point stations where increasing trends have been detected, Dir: the net direction of change in the lifeform (↓: decreasing, ↑: increasing, -: stable), Trend: the percentage of assessment units exhibiting the respective trend, Conf: the mean confidence of datasets considered in the assessment, Change: a logical variable (TRUE/FALSE) to report whether a net trend is likely given the proportion of locations expressing the trend and the confidence and spatial representativeness scores, Press1: the environmental pressure with the greatest mean rank for the respective trend, Rank1: the mean rank of the environmental pressure indicated under Pres1, nStn: the total number of fixed-point stations considered, totAssess: The total number of COMP4 assessment units and fixed-point stations considered, totCOMP4: The total number of potential COMP4 assessment units for the habitat category, spatialRep: the spatial representativeness score of the analysis. The status of the individual lifeforms are indicated by the colours in the Lifeform column, according to the legend in Table i. The overall status of the habitat category is indicated by the colour of the first column, which also identifies pelagic habitat types and follows the same colour legend.

Habitat	Lifeform	↓	-	↑	Dir	Trend	Conf	Change	Press1	Rank1	nStn	totAssess	totCOMP4	SpatialRep
Variable salinity	Diatom	0	4	1		80%	51%	FALSE	np	3,5	1	5	9	44%
	Dinoflagellate	1	2	2	↑	40%	51%	FALSE	np	2,5	1	5	9	44%
	Holoplankton	1	0	0	↓	100%	30%	FALSE	ph	1,0	0	1	9	11%
	Meroplankton	0	1	0	-	100%	30%	FALSE	psal	1,0	0	1	9	11%
	Large copepods	1	0	0	↓	100%	30%	FALSE	sst	1,0	0	1	9	11%

Coastal	Small copepods	1	0	0	↓	100%	30%	FALSE	sst	1,0	0	1	9	11%
	Fish larvae	0	0	1	↑	100%	30%	FALSE	wspd	1,0	0	1	9	11%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	9	0%
	Diatom	2	9	8	-	47%	71%	FALSE	psal	3,2	9	19	12	83%
	Dinoflagellate	5	8	6	-	42%	71%	FALSE	ntot	2,3	9	19	12	83%
	Holoplankton	3	8	1	-	67%	59%	FALSE	psal	2,8	2	12	12	83%
	Meroplankton	0	4	8	↑	67%	59%	TRUE	sst	3,1	2	12	12	83%
	Large copepods	3	7	2	-	58%	59%	FALSE	precip	2,1	2	12	12	83%
	Small copepods	2	6	4	-	50%	59%	FALSE	sst	2,2	2	12	12	83%
Shelf	Fish larvae	0	6	6	↑	50%	59%	TRUE	psal	3,2	2	12	12	83%
	Gelatinous	0	3	1	-	75%	64%	FALSE	psal	1,0	2	4	12	17%
	Diatom	1	5	5	↑	45%	74%	FALSE	phos	3,2	0	11	11	100%
	Dinoflagellate	7	3	1	↓	64%	74%	TRUE	wspd	3,9	0	11	11	100%
	Holoplankton	6	4	1	↓	55%	74%	TRUE	sst	2,0	0	11	11	100%
	Meroplankton	0	2	9	↑	82%	74%	TRUE	sst	2,2	0	11	11	100%
	Large copepods	3	8	0	-	73%	74%	FALSE	sst	2,3	0	11	11	100%
	Small copepods	4	4	3	↓	36%	74%	FALSE	sst	1,3	0	11	11	100%
Gelatinous	Fish larvae	1	4	6	↑	55%	74%	TRUE	attn	2,7	0	11	11	100%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	11	0%

In the Celtic Seas (**Table k**), declining abundance of diatoms in variable salinity habitats have been observed and linked to the increase in the N:P ratio. Declining abundance trends in dinoflagellates, as well as large and small copepods in coastal habitats were linked to temperature increases. Declining abundance of

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holoplankton in coastal areas may also be linked to decreasing pH. Changes in pH can be linked to changes in primary production, since phytoplankton ingest Dissolved Inorganic Carbon (DIC) to fuel growth and reproduction, but also to the concentration of atmospheric carbon dioxide, the main driver of global warming. In shelf areas a similar increase in meroplankton abundance has been observed in response to temperature, while decreasing abundance of holoplankton and dinoflagellates may be linked to reductions in pH and increases in the N:P ratio, respectively.

Table k: Integration of the indicator results for OSPAR Region III – Celtic Seas. An explanation of column names can be found in the caption for Table j.

Habitat	Lifeform	↓	-	↑	Dir	Trend	Conf	Change	Press 1	Rank 1	nStn	totAsses s	totCOMP 4	SpatialRe p
Variable salinity	Diatom	2	0	1	↓	67%	70%	TRUE	np	1,0	2	3	2	50%
	Dinoflagellate	0	0	3	↑	100%	70%	TRUE	attn	2,5	2	3	2	50%
	Holoplankton	1	0	0	↓	100%	99%	TRUE	psal	1,0	1	1	2	0%
	Meroplankton	1	0	0	↓	100%	99%	TRUE	wspd	1,0	1	1	2	0%
	Large copepods	1	0	0	↓	100%	99%	TRUE	sst	1,0	1	1	2	0%
	Small copepods	1	0	0	↓	100%	99%	TRUE	amo	1,0	1	1	2	0%
	Fish larvae	1	0	0	↓	100%	99%	TRUE	attn	1,0	1	1	2	0%
	Gelatinous	1	0	0	↓	100%	99%	TRUE	wspd	1,0	1	1	2	0%
	Diatom	1	2	0	-	67%	63%	FALSE	precip	2,0	0	3	3	100%
Coastal	Dinoflagellate	2	1	0	↓	67%	63%	TRUE	sst	2,0	0	3	3	100%
	Holoplankton	2	1	0	↓	67%	63%	TRUE	ph	1,5	0	3	3	100%
	Meroplankton	0	2	1	-	67%	63%	FALSE	mld	2,5	0	3	3	100%
	Large copepods	2	1	0	↓	67%	63%	TRUE	sst	1,5	0	3	3	100%
	Small copepods	2	1	0	↓	67%	63%	TRUE	sst	1,0	0	3	3	100%
	Fish larvae	0	2	1	-	67%	63%	FALSE	mld	2,5	0	3	3	100%
	Gelatinous	0	0	0	NA	NA	0%	FALSE	NA	NA	0	0	3	0%
	Diatom	1	2	1	-	50%	87%	FALSE	precip	2,5	0	4	4	100%
Shelf	Dinoflagellate	3	1	0	↓	75%	87%	TRUE	np	3,0	0	4	4	100%

	Holoplankton	3	1	0	↓	75%	87%	TRUE	ph	2,7	0	4	4	100%
	Meroplankton	0	1	3	↑	75%	87%	TRUE	sst	1,0	0	4	4	100%
	Large copepods	1	3	0	-	75%	87%	FALSE	sst	1,7	0	4	4	100%
	Small copepods	1	3	0	-	75%	87%	FALSE	precip	1,3	0	4	4	100%
	Fish larvae	1	3	0	-	75%	87%	FALSE	psal	2,7	0	4	4	100%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	4	0%

In the Bay of Biscay and Iberian Coast region (**Table I**), holoplankton, large and small copepods in shelf areas are also exhibiting decreasing abundance trends linked to temperature increase, while large and small copepods in oceanic / beyond shelf habitats are experiencing declines linked to increasing temperatures. Trends for variable salinity habitats were not assessed due to the limited data available for this region.

Table I: Integration of the indicator results for OSPAR Region IV – Bay of Biscay and Iberian Coast. An explanation of column names can be found in the caption for Table j.

Habitat	Lifeform	↓	-	↑	Dir	Trend	Conf	Change	Press1	Rank1	nStn	totAsses	totCOMP4	SpatialRep
Variable salinity	Diatom	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Dinoflagellate	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Holoplankton	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Meroplankton	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Large copepods	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Small copepods	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Fish larvae	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	3	0%
Coastal	Diatom	0	2	1	-	67%	52%	FALSE	mld	1,0	0	3	12	25%
	Dinoflagellate	0	0	3	↑	100%	52%	TRUE	amo	2,3	0	3	12	25%
	Holoplankton	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	12	0%
	Meroplankton	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	12	0%
	Large copepods	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	12	0%
	Small copepods	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	12	0%
	Fish larvae	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	12	0%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	12	0%

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	Diatom	2	1	0	↓	67%	63%	TRUE	wspd	1,0	0	3	6	50%
	Dinoflagellate	1	2	0	-	67%	63%	FALSE	np	2,0	0	3	6	50%
	Holoplankton	3	0	0	↓	100%	63%	TRUE	sst	2,3	0	3	6	50%
	Meroplankton	1	1	1	↓	33%	63%	FALSE	sst	1,0	0	3	6	50%
	Large copepods	2	1	0	↓	67%	63%	TRUE	sst	1,0	0	3	6	50%
	Small copepods	2	1	0	↓	67%	63%	TRUE	sst	1,0	0	3	6	50%
	Fish larvae	0	3	0	-	100%	63%	FALSE	wspd	2,0	0	3	6	50%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	6	0%
Shelf	Diatom	2	0	0	↓	100%	84%	TRUE	psal	1,0	0	2	6	33%
	Dinoflagellate	1	1	0	↓	50%	84%	TRUE	psal	1,0	0	2	6	33%
	Holoplankton	2	0	0	↓	100%	84%	TRUE	mld	1,5	0	2	6	33%
	Meroplankton	0	1	1	↑	50%	84%	TRUE	mld	1,0	0	2	6	33%
	Large copepods	2	0	0	↓	100%	84%	TRUE	sst	1,5	0	2	6	33%
	Small copepods	2	0	0	↓	100%	84%	TRUE	sst	1,0	0	2	6	33%
	Fish larvae	2	0	0	↓	100%	84%	TRUE	mld	1,5	0	2	6	33%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	6	0%
Oceanic / beyond shelf	Diatom	2	0	0	↓	100%	84%	TRUE	psal	1,0	0	2	6	33%
	Dinoflagellate	1	1	0	↓	50%	84%	TRUE	psal	1,0	0	2	6	33%
	Holoplankton	2	0	0	↓	100%	84%	TRUE	mld	1,5	0	2	6	33%
	Meroplankton	0	1	1	↑	50%	84%	TRUE	mld	1,0	0	2	6	33%
	Large copepods	2	0	0	↓	100%	84%	TRUE	sst	1,5	0	2	6	33%
	Small copepods	2	0	0	↓	100%	84%	TRUE	sst	1,0	0	2	6	33%
	Fish larvae	2	0	0	↓	100%	84%	TRUE	mld	1,5	0	2	6	33%
	Gelatinous	0	0	0	N A	NA	0%	FALSE	NA	NA	0	0	6	0%

Conclusion

Pelagic habitats have changed considerably over the past 60 years, with widespread declines in key lifeforms over much of the North-East Atlantic. Within the short assessment period itself (2015 to 2019) only some changes could be detected – long time-series are critical to separating change from background variability. Declining abundances of important planktonic lifeforms are cause for concern for the future of ‘traditional’ pelagic food webs, wherein blooms of phytoplankton feed copepods, which support commercially exploited fish. Conversely, increasing abundances of fish larvae / eggs and of meroplankton throughout the North Sea could benefit fisheries, but only if recruitment increases with larval abundance and if increasing taxa are exploitable.

Environmental pressures with the strongest links to lifeform abundance are driven by climate change, either directly (e.g. change in water temperature) or indirectly (e.g., change in mixed layer depth). While changes in lifeform abundance due to other anthropogenic influences (e.g. nutrient input from rivers) were less clear, results suggest that large changes to pelagic habitats have occurred across coastal, shelf and oceanic areas. Considering the scale at which these pressures operate, regionally targeted management measures are likely to initially only generate noticeable impact in coastal waters and overall will have minimal impact on slowing long-term changes. However, more widespread measures could strengthen ecosystem resilience, even if impacts are not directly measurable at greater distances from the coasts, and thus indirectly contribute to slowing long term changes into the future.

Conclusion (Extended)

This assessment indicates spatial and temporal variability in the plankton community for eight ecologically important planktonic lifeforms. While there were clear increasing and decreasing trends in the abundance of lifeforms across the North-East Atlantic, the overall trend was predominantly decreasing for diatoms, dinoflagellates, holoplankton, large copepods and small copepods, particularly within the Celtic Seas and the Bay of Biscay and Iberian Coast (OSPAR Regions III and IV). Meroplankton were the only clear exception, with evidence of increasing abundance across most of the areas assessed, and particularly the Greater North Sea (OSPAR Region II). Evidence of links between trends in lifeform abundance and anthropogenic climate change mainly occurred in shelf and oceanic pelagic habitats, while changes attributed to variation in nutrients were primarily limited to coastal habitats.

This indicator assessment has been conducted in accordance with expert knowledge on plankton dynamics from the scientific literature (McQuatters-Gollop *et al.*, 2019; McQuatters-Gollop *et al.* 2015). The plankton lifeform indicator has been published in the peer-reviewed literature (McQuatters-Gollop *et al.*, 2019; Tett *et al.*, 2008) and has been developed at the sub-regional scale using data from the pan-European Continuous Plankton Recorder survey which has documented quality assurance procedures and has supported over 1000 scientific publications (McQuatters-Gollop *et al.*, 2015). The methods used here have attempted to improve this established approach, making it more suitable for detecting change linked to environmental pressures.

Links between environmental pressures and plankton lifeforms described here present important areas for further research. There is no single environmental variable linked to change in lifeform abundance, and many of the long-term changes detected in this assessment were unable to be explained by the variables tested. This may be at least partially due to the paucity of robust pressure data available at the spatial and temporal scale and resolution required to conduct a regional assessment of this nature. Local changes may be occurring but are not detectable when considered at a regional scale. It is important to establish clearer causal links by exploring this problem at finer scale than was suitable for a regional assessment, incorporating seasonality and taking better advantage of the high-resolution gridded satellite and other data products currently available, such as abundance of small (nano and pico) plankton which are an important part of the pelagic food web.

As the blue economy continues to develop and industries such as aquaculture are likely to become more important in the coming years, changing plankton communities can impact their sustainability as pelagic habitats change. Further interpretation of results and refinement to the methodology will be essential for understanding the effects these changes can have on ecosystem services, however this will of course be resource dependent.

Knowledge Gaps

It will be important to obtain zooplankton datasets within pelagic habitats lacking sufficient coverage in the current assessment. Further scientific research will be required to explore direct mechanistic links between change in environmental variables and change in lifeform abundance. It may also be useful to examine the rate and direction of change in the Plankton Index with respect to each lifeform pair, as well as the ecological consequences of changing lifeform abundance on higher trophic levels, not considered in the pelagic habitats assessment. This could help link both the pelagic and food webs assessments in terms of identified changes and impacts.

Knowledge Gaps (Extended)

The following work needs to be taken forward to address the knowledge gaps and improve the assessment provided by this indicator:

- Examining national datasets at a finer spatiotemporal scale, which will help establish clearer links between climatic and anthropogenic pressures with changes in pelagic habitats and plankton diversity;
- Introducing lag into the variable selection process to test for delayed effects of environmental pressures on lifeform abundance (e.g., the effects of winter nutrient concentrations on lifeform abundance during the growing season);
- Investigating the impacts of long-term changes in lifeform abundance on pelagic food webs;
- Obtaining pressure datasets that would present an indicator of predation on plankton from higher trophic levels (e.g., forage fish density);
- Obtaining high quality plankton monitoring data for several pelagic habitat types which were underrepresented in this assessment. In particular there is a need for: zooplankton data for variable salinity habitats in the Greater North Sea and Celtic Seas (OSPAR Regions II and III),

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zooplankton data for coastal habitats and both phytoplankton and zooplankton data for variable salinity habitats in the Bay of Biscay and Iberian Coast (OSPAR Region IV);

- Determination of the traits for many plankton species – for some species even information about basic biological characteristics, such as diet, is not yet known;
- Understanding the role of pico and nanoplankton (small plankton composed by cells with sizes between respectively 0,2 µm and 2 µm, and 2 µm and 20 µm) in the ecosystem. These small size categories are difficult to measure routinely and are thus mainly ignored. However, they make up a significant proportion of the plankton biomass, diversity and trophic roles (De Vargas *et al.*, 2015) relevant to marine food webs and carbon export (Leblanc *et al.*, 2018). Their ecosystem role needs further investigation so that they can be included in an appropriate new or existing lifeform;
- Establishment of monthly monitoring programmes for pico and nanoplankton to provide data on a relatively data-sparse component of the plankton community;

Developing a better understanding for the extent of pico and nanoplankton biological interactions in the context of their environments.

References

- Barnes, M. K., G. H. Tilstone, T. J. Smyth, C. E. Widdicombe, J. Gloël, C. Robinson, J. Kaiser, and D. J. Suggett. 2015. Drivers and effects of *Karenia mikimotoi* blooms in the western English Channel. *Progress in Oceanography* **137**: 456-469.
- Beaugrand, G. 2005. Monitoring pelagic ecosystems using plankton indicators. *ICES Journal of Marine Science* **62**: 333-338.
- Bedford, J., C. Ostle, D. G. Johns, A. Atkinson, M. Best, E. Bresnan, M. Machairopoulou, C. A. Graves, M. Devlin, and A. Milligan. 2020a. Lifeform indicators reveal large-scale shifts in plankton across the North-West European shelf. *Global Change Biology* **26**: 3482-3497.
- Bedford, J., C. Ostle, D. G. Johns, A. Budria, and A. McQuatters-Gollop. 2020b. The influence of temporal scale selection on pelagic habitat biodiversity indicators. *Ecological Indicators* **114**: 106311.
- Bremner, J., C. L. J. Frid, and S. I. Rogers. 2004. Biological traits of the North Sea benthos – Does fishing affect benthic ecosystem function? , p. 477-489. In P. Barnes, Thomas, J. [ed.], *Benthic habitats and the effects of fishing*. American Fisheries Society.
- Charvet, S., M. C. Roger, B. Faessel, and M. Lafont. 1998. Biomonitoring of freshwater ecosystems by the use of biological traits. *Annals of Limnology – International Journal of Limnology* **34**: 455-464.
- de Jonge, V. N. 2007. Toward the application of ecological concepts in EU coastal water management. *Marine Pollution Bulletin* **55**: 407-414.
- De Vargas, C., S. Audic, N. Henry, J. Decelle, F. Mahé, R. Logares, E. Lara, C. Berney, N. Le Bescot, and I. Probert. 2015. Eukaryotic plankton diversity in the sunlit ocean. *Science* **348**: 1261605.
- Desmit, X., A. Nohe, A. V. Borges, T. Prins, K. De Cauwer, R. Lagring, D. Van der Zande, and K. Sabbe. 2020. Changes in chlorophyll concentration and phenology in the North Sea in relation to de-eutrophication and sea surface warming. *Limnology and Oceanography* **65**: 828-847.
- Enserink, L., A. Blauw, D. van der Zande, and S. Markager. 2019. Summary report of the EU project 'Joint monitoring programme of the eutrophication of the North Sea with satellite data' (Ref: DG ENV/MSFD Second Cycle/2016), p. 21 pp.
- Estrada, M., P. Henriksen, J. M. Gasol, E. O. Casamayor, and C. Pedrós-Alió. 2004. Diversity of Planktonic Photoautotrophic Microorganisms Along a Salinity Gradient as Depicted by Microscopy, Flow Cytometry, Pigment Analysis and DNA-based Methods. *FEMS Microbiology Ecology* **49**: 281-293.

Flynn, K. J., A. Mitra, K. Anestis, A. A. Anschütz, A. Calbet, G. D. Ferreira, N. Gypens, P. J. Hansen, U. John, and J. L. Martin. 2019. Mixotrophic protists and a new paradigm for marine ecology: where does plankton research go now? *Journal of Plankton Research* **41**: 375-391.

Gallego, I., T. A. Davidson, E. Jeppesen, C. Perez-Martinez, P. Sanchez-Castillo, M. Juan, F. Fuentes-Rodriguez, D. Leon, P. Penalver, J. Toja, and J. J. Casas. 2012. Taxonomic or ecological approaches? Searching for phytoplankton surrogates in the determination of richness and assemblage composition in ponds. *Ecological Indicators* **18**: 575-585.

Garmendia, M., Á. Borja, J. Franco, and M. Revilla. 2012. Phytoplankton composition indicators for the assessment of eutrophication in marine waters: Present state and challenges within the European directives. *Marine Pollution Bulletin* **66**: 7-16.

Gowen, R. J., A. McQuatters-Gollop, P. Tett, M. Best, E. Bresnan, C. Castellani, K. Cook, R. Forster, C. Scherer, and A. McKinney. 2011. The Development of UK Pelagic (Plankton) Indicators and Targets for the MSFD. Advice to Defra, p. 41 pp.

HELCOM. 2012. Development of the HELCOM core-set indicators Part B, GES 8/2012/7b. HELCOM.

Keys, M., G. Tilstone, H. S. Findlay, C. E. Widdicombe, and T. Lawson. 2018. Effects of elevated CO₂ and temperature on phytoplankton community biomass, species composition and photosynthesis during an experimentally induced autumn bloom in the western English Channel. *Biogeosciences* **15**: 3203-3222.

Kursar, M. B., and W. R. Rudnicki. 2010. Feature selection with the Boruta package. *Journal of statistical software* **36**: 1-13.

Leblanc, K., B. Queguiner, F. Diaz, V. Cornet, M. Michel-Rodriguez, X. Durrieu de Madron, C. Bowler, S. Malviya, M. Thyssen, and G. Grégori. 2018. Nanoplanktonic diatoms are globally overlooked but play a role in spring blooms and carbon export. *Nature Communications* **9**: 1-12.

Mayer, M., and M. M. Mayer. 2019. Package 'missRanger'. R Package.

McQuatters-Gollop, A., A. Atkinson, A. Aubert, J. Bedford, M. Best, E. Bresnan, K. Cook, M. Devlin, R. Gowen, and D. G. Johns. 2019. Plankton lifeforms as a biodiversity indicator for regional-scale assessment of pelagic habitats for policy. *Ecological Indicators* **101**: 913-925.

McQuatters-Gollop, A., M. Edwards, P. Helaouët, D. G. Johns, N. J. Owens, D. E. Raitsos, D. Schroeder, J. Skinner, and R. F. Stern. 2015. The Continuous Plankton Recorder survey: How can long-term phytoplankton datasets contribute to the assessment of Good Environmental Status? *Estuarine, Coastal and Shelf Science* **162**: 88-97.

McQuatters-Gollop, A., L. Guérin, N. L. Arroyo, A. Aubert, L. F. Artigas, J. Bedford, E. Corcoran, V. Dierschke, S. A. M. Elliott, S. C. V. Geelhoed, A. Gilles, J. M. González-Irusta, J. Haelters, M. Johansen, F. C. P. Lynam, N. Niquil, B. Meakins, I. Mitchell, B. Padegimas, R. Pesch, I. Preciado, I. Rombouts, G. Safi, P. Schmitt, U. Schückel, A. Serrano, P. Stebbing, A, and C. Vina-Herbon. 2022. Assessing the state of marine biodiversity in the Northeast Atlantic. *Ecological Indicators* **141**: 109148.

Mouillot, D., S. Spatharis, S. Reizopoulou, T. Laugier, L. Sabetta, A. Basset, and T. D. Chi. 2006. Alternatives to taxonomic-based approaches to assess changes in transitional water communities. *Aquatic Conservation-Marine and Freshwater Ecosystems* **16**: 469-482.

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Ostle, C., K. Paxman, C. A. Graves, M. Arnold, L. F. Artigas, A. Atkinson, A. Aubert, M. Baptie, B. Bear, and J. Bedford. 2021. The Plankton Lifeform Extraction Tool: a digital tool to increase the discoverability and usability of plankton time-series data. *Earth System Science Data* **13**: 5617-5642.

Rakocinski, C. F. 2012. Evaluating macrobenthic process indicators in relation to organic enrichment and hypoxia. *Ecological Indicators* **13**: 1-12.

Reid, P. C., J. Colebrook, J. Matthews, J. Aiken, and C. P. R. Team. 2003. The Continuous Plankton Recorder: concepts and history, from Plankton Indicator to undulating recorders. *Progress in Oceanography* **58**: 117-173.

Svendsen, L., L. van der Bijl, S. Boutrup, and B. Norup. 2005. NOVANA. National Monitoring and Assessment Programme for the Aquatic and Terrestrial. and number: NERI Technical Report.

Tett, P., C. Carreira, D. K. Mills, S. van Leeuwen, J. Foden, E. Bresnan, and R. J. Gowen. 2008. Use of a phytoplankton community index to assess the health of coastal waters. *ICES Journal of Marine Science* **65**: 1475-1482.

Assessment Metadata

Field	Data Type	
Assessment type	List	Indicator Assessment
Summary Results (template Addendum 1)	URL	https://odims.ospar.org/en/submissions/ospar_changes_phy_zoo_msfd_2022_06_001/
SDG Indicator	List	14.2: By 2020, sustainably manage and protect marine and coastal ecosystems to avoid significant adverse impacts, including by strengthening their resilience, and take action for their restoration in order to achieve healthy and productive oceans
Thematic Activity	List	Biological Diversity and Ecosystems
Relevant OSPAR Documentation	Text	OSPAR Agreement 2010-03 The North-East Atlantic Environment Strategy. Strategy of the OSPAR Commission for the Protection of the Marine Environment of the North-East Atlantic 2010–2020 OSPAR Agreement 2016-01 OSPAR Coordinated Environmental Monitoring Programme (CEMP), as amended OSPAR Agreement 2021-01 Strategy of the OSPAR Commission for the Protection of the Marine Environment of the North-East Atlantic 2030
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Conditions applying to access and use	URL	https://oap.ospar.org/en/data-policy/
Data Snapshot	URL	https://odims.ospar.org/en/submissions/ospar_changes_phy_zoo_dsnap_2022_06_001/
Data Results	Zip File	https://odims.ospar.org/en/submissions/ospar_changes_phy_zoo_dres_2022_06_001/
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Our vision is a clean, healthy and biologically diverse North-East Atlantic Ocean, which is productive, used sustainably and resilient to climate change and ocean acidification.