

# Machine Learning

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August 11, 2017

# Overview

In this presentation I show some of the ways computers can “learn” or be trained, and then predict classifications based off of training sets.

# Bullet Points

- To begin with machine learning I start with knn prediction. It is a powerful and straight forward algorithm to use in R
- PCA clustering is demonstrated next, and is more complicated because it uses eigen values to project multi-dimensional data into two dimensions
- Next, we go into tree-based classification methods of machine learning by looking at rpart and random forest algorithms
- After an important gene or factor is found it can be used as a measure of prediction -Finally, I end with clustering as a means of prediction by using k-means and PAM clustering

# k-nearest neighbors algorithm (k-NN) is a non-parametric method used for classification and regression (wikipedia)

```
library(HiDimDA)
data("AlonDS")
library(class)

set.seed(1234)
ind <- sample(2, nrow(AlonDS), replace=TRUE, prob=c(.39, 0.61))
apo.training <- AlonDS[ind==1, 2:2001]
apo.test <- AlonDS[ind==2, 2:2001]
apo.trainlabels <- AlonDS[ind==1, 1]
apo.testlabels <- AlonDS[ind==2, 1]
apo_pred <- knn(apo.training, apo.test, apo.trainlabels, k=1)
table(apo_pred, apo.testlabels)
```

```
##          apo.testlabels
## apo_pred  colonc healthy
##   colonc      25      2
##   healthy      2     11
```

## PCA dimension reduction

```
alon.data <- (AlonDS[1:dim(AlonDS)[1], 2:dim(AlonDS)[2]])  
alon.names <- read.csv("data/alon.names.csv", header=F)  
Alon.cl <- c(0,1,0,1,0,1,0,1,0,1,0,1,0,1,0,1,0,1,0,0,0,0,0,0,0,0,0,0,0,0,0,1,0,0,1,1,0,0,0,0,1,0,1,1,0,0,1,1,0,0,0,0,1,0,1)  
AlonFactor <- factor(Alon.cl, levels=0:1, labels=c("colonc", "healthy"))  
talon.data <- t(alon.data)  
eigen(cor(talon.data))$values[1:5]
```

```
## [1] 46.266564 2.831612 2.264533 1.758461 1.151688
```

```
data <- talon.data; p <- ncol(data); n <- nrow(data) ; nboot<-1000
eigenvalues <- array(dim=c(nboot,p))
for (i in 1:nboot) {
  dat.star <- data[sample(1:n,replace=TRUE),]
  eigenvalues[i,] <- eigen(cor(dat.star))$values
}
for (j in 1:5) {
  print(quantile(eigenvalues[,j],c(0.025,0.975)))
}
```

##	2.5%	97.5%
##	44.36836	48.28612
##	2.5%	97.5%
##	2.450823	3.759073
##	2.5%	97.5%
##	1.785127	2.827118
##	2.5%	97.5%
##	1.382515	2.014506
##	2.5%	97.5%
##	1.011605	1.421319

# PCA results

```
sum(eigen(cor(talon.data))$values[1:5])/62*100
```

```
## [1] 87.53687
```

*#Thus, the first five components represent more than 87% of the variance in the data. The data can allow for a reduction in dimensions from  
#sixty two data points to five, while still capturing over 87% of the total variance*

*# to verify that the correlations between the patients are positive*

*# we run;*

```
-eigen(cor(talon.data))$vec[,1:5]
```

```
##           [,1]      [,2]      [,3]      [,4]      [,5]
## [1,] 0.13334450 0.033986091 0.064299240 0.190182448 0.087596270
## [2,] 0.13101243 0.055453409 -0.160080373 0.192870293 0.065330616
## [3,] 0.08123138 0.381194248 0.254511766 -0.158840079 0.066887819
## [4,] 0.12186663 0.187345101 0.024326909 0.076170309 0.227739581
## [5,] 0.13492308 -0.025234418 0.064778140 -0.131490100 0.106656352
## [6,] 0.12573668 -0.042508361 -0.162798987 -0.243283225 0.205817286
## [7,] 0.13142976 -0.067702980 0.063955075 0.139911681 0.108777444
## [8,] 0.11497311 -0.040887571 -0.147787136 -0.086017008 0.171674091
## [9,] 0.12035762 -0.123376691 0.127601183 0.018987419 0.194064424
## [10,] 0.12759766 0.073671501 -0.135938797 -0.112838720 0.114489142
## [11,] 0.11994081 -0.165215734 0.028956883 0.037587940 -0.130940977
## [12,] 0.13114523 0.087751355 -0.077048073 0.079323831 0.095345885
## [13,] 0.13671340 -0.055573830 0.010252231 -0.021086622 0.114873382
## [14,] 0.11569421 0.038688741 -0.049658214 -0.029725807 0.199764625
## [15,] 0.13809274 0.001120874 0.094184776 -0.103703484 0.063806124
## [16,] 0.12442516 -0.025277073 0.134594415 0.034626258 0.109465939
## [17,] 0.12394763 -0.160422616 0.109688079 -0.003395834 0.039133282
## [18,] 0.12247192 -0.115504816 -0.118420118 -0.070117793 0.227967374
## [19,] 0.12982113 -0.162356072 0.079103129 -0.044542409 0.158668333
## [20,] 0.12278348 -0.095501004 -0.195402405 -0.210246203 0.170166072
## [21,] 0.12798672 -0.044834231 0.029938642 0.227943617 0.092220435
## [22,] 0.13003189 0.176046859 -0.094013733 0.070443325 0.068489667
## [23,] 0.11531068 -0.204686374 0.100432637 -0.204685846 0.019681785
## [24,] 0.12168741 -0.095188189 -0.082774295 -0.091211027 0.282243516
## [25,] 0.13290451 0.029251113 0.168042056 -0.160266523 0.014045905
## [26,] 0.13323856 0.019802248 0.106565887 0.165275129 -0.058442060
## [27,] 0.13511859 -0.020930960 0.000432636 0.230172036 -0.041733403
## [28,] 0.13085940 -0.086379885 0.094902516 0.174007957 0.054348256
## [29,] 0.13023929 0.024270947 0.054169492 0.263010813 -0.022021684
## [30,] 0.12650756 -0.107720674 0.015253178 0.206960704 -0.067399870
## [31,] 0.12657652 -0.013398653 0.006917491 0.150793132 0.038063135
## [32,] 0.12781952 -0.152868743 -0.082453566 -0.014880923 -0.164087648
## [33,] 0.13120837 -0.199350101 0.028402632 -0.036243480 -0.119104805
## [34,] 0.13165334 0.095107519 0.173911064 -0.079763756 0.024130942
## [35,] 0.13654398 0.006862681 0.033967618 0.105180091 -0.038677095
```

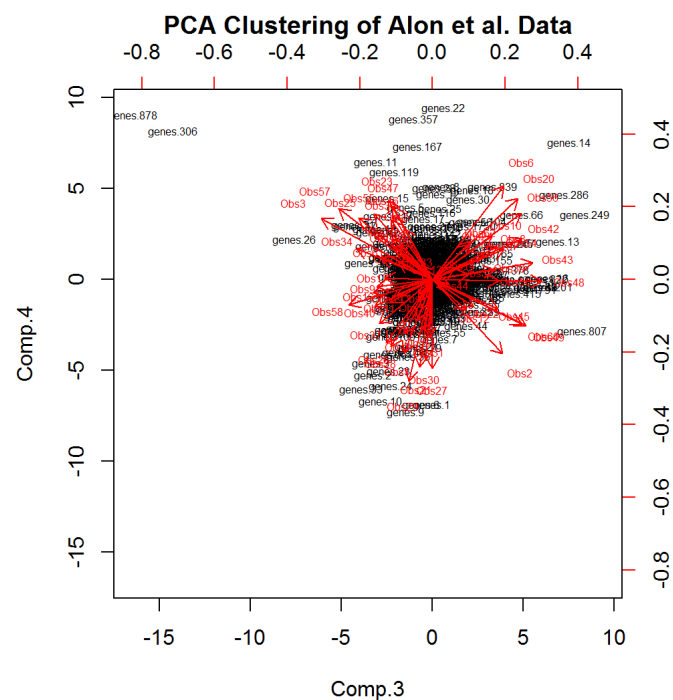
```
## [36,] 0.12753263 -0.176498716 0.094728221 -0.155394342 -0.106052817
## [37,] 0.12499657 0.091633181 0.121564126 0.113979885 0.154206875
## [38,] 0.12142376 -0.247474501 0.086513377 -0.086719621 -0.098649101
## [39,] 0.12972644 0.140905505 -0.141355244 -0.024807465 -0.007195856
## [40,] 0.13562229 -0.011701548 0.132153449 0.068448400 -0.122473596
## [41,] 0.12758597 0.138075514 0.097537111 0.059408314 0.058157902
## [42,] 0.12805261 0.038792278 -0.203808899 -0.105816551 -0.004528208
## [43,] 0.12695986 0.048085391 -0.229061725 -0.042818861 -0.186404835
## [44,] 0.12507601 -0.189008038 -0.037042016 0.011765187 -0.207960639
## [45,] 0.13269938 0.093324863 -0.149496926 0.075425565 -0.122914086
## [46,] 0.13018857 -0.076919772 0.090216978 -0.165045860 -0.215177634
## [47,] 0.12702667 -0.032740376 0.090144639 -0.191153217 -0.166982357
## [48,] 0.12247626 0.110721335 -0.250108174 0.004394967 -0.095453279
## [49,] 0.12597808 0.092051645 -0.214253387 0.119008694 -0.150459561
## [50,] 0.13084612 0.055682312 -0.152138267 0.003286583 -0.100791706
## [51,] 0.13174309 -0.052137569 0.020586878 0.130532967 -0.103712971
## [52,] 0.13318200 -0.110529496 0.075826283 -0.092541590 -0.090512358
## [53,] 0.13503694 -0.023589364 0.032351753 0.056959693 0.039407886
## [54,] 0.12620290 0.081782916 -0.163850480 -0.077543180 0.059139961
## [55,] 0.11426325 0.236062064 0.133687846 -0.170587949 -0.203857916
## [56,] 0.12847582 0.049489774 -0.202409188 -0.171449349 -0.101526188
## [57,] 0.09379796 0.361373049 0.214588704 -0.183748694 -0.034864277
## [58,] 0.12571744 0.153957449 0.191561791 0.065713023 -0.038236070
## [59,] 0.13366113 -0.079822957 0.081939721 0.039871818 -0.090216999
## [60,] 0.12649362 0.126979038 -0.204223030 0.116561202 -0.153176973
## [61,] 0.12977076 0.089854430 0.116352577 -0.056596296 -0.177553088
## [62,] 0.13421142 0.097037064 -0.088053764 -0.098567643 -0.132623999
```

# PCA results in five dimensions

*#From the previous slide we can match up the negative and positive signs with what we actually observed and express as;*  
 Alon.c1

```
## [1] 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 0 0 0 0 0 0 0 0 0
## [36] 0 0 0 1 0 0 1 1 0 0 0 0 1 0 1 1 0 0 1 1 0 0 0 0 1 0 1
```

*#By matching these we find that the 3rd and 4th dimensions matched the real data best. We can plot these with a biplot to show vectors*  
 biplot(princomp(data,cor=TRUE),choices = c(3,4), pc.biplot=TRUE,cex=0.5,expand=0.8, main = "PCA Clustering of Alon et al. Data")



*#There is a clear separation from the left and right in the biplot*



# PCA results; finding the top genes

```
#No we can prints the first ten gene names with respect to the third principal comonent to find out what the main genes of differentiating diseased and healthy state  
pca <- princomp(talon.data, center = TRUE, cor=TRUE, scores=TRUE)  
o <- order(pca$scores[,3])  
alon.names[o[1:10],6]
```

```
## [1] IG GAMMA-1 CHAIN C REGION (HUMAN);.  
## [2] Human Ig gamma3 heavy chain disease OMM protein mRNA.  
## [3] 40S RIBOSOMAL PROTEIN S18 (Homo sapiens)  
## [4] LAMININ RECEPTOR (HUMAN);.  
## [5] 60S RIBOSOMAL PROTEIN L30E (Kluyveromyces lactis)  
## [6] 40S RIBOSOMAL PROTEIN S24 (HUMAN).  
## [7] P24050 40S RIBOSOMAL PROTEIN.  
## [8] EUKARYOTIC INITIATION FACTOR 4A (Oryctolagus cuniculus)  
## [9] EUKARYOTIC INITIATION FACTOR 4A (Oryctolagus cuniculus)  
## [10] 40S RIBOSOMAL PROTEIN S6 (Nicotiana tabacum)  
## 1824 Levels: ...
```

```
pr.out =prcomp (alon.data , scale =TRUE)
```

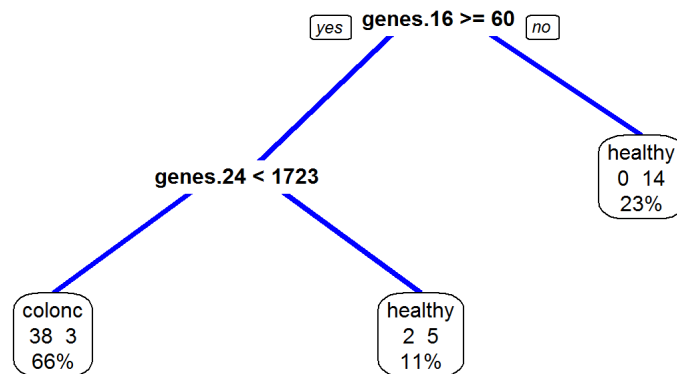
# rpart finds important genes with recursive partitioning and regression to make predictions

```
library(rpart.plot)

row.names(talon.data) <- paste("gene", 1:2000, sep = "")

alonRpart <- rpart(AlonFactor~., data=alon.data, method="class", cp=0.001)
prp(alonRpart, branch.lwd=4, branch.col="blue", extra=101, main="Recursive Partitioning and Regression Tree of Alon et al Data")
```

Recursive Partitioning and Regression Tree of Alon et al Data



```
alon.names[ 16, 6]
```

```
## [1] Human tra1 mRNA for human homologue of murine tumor rejection antigen gp961
## 1824 Levels: ...
```

```
alon.names[ 24, 6]
```

```
## [1] THYMOSIN BETA-4 (HUMAN);.  
## 1824 Levels: ...
```

**Random forests or random decision forests operate by constructing a multitude of decision trees at training time and outputting mean prediction (regression) of the individual trees. (wikipedia)**

```
library(randomForest)
set.seed(1234)
alon.rf <- randomForest(grouping ~ ., data=AlonDS, importance=TRUE,
                        proximity=TRUE)

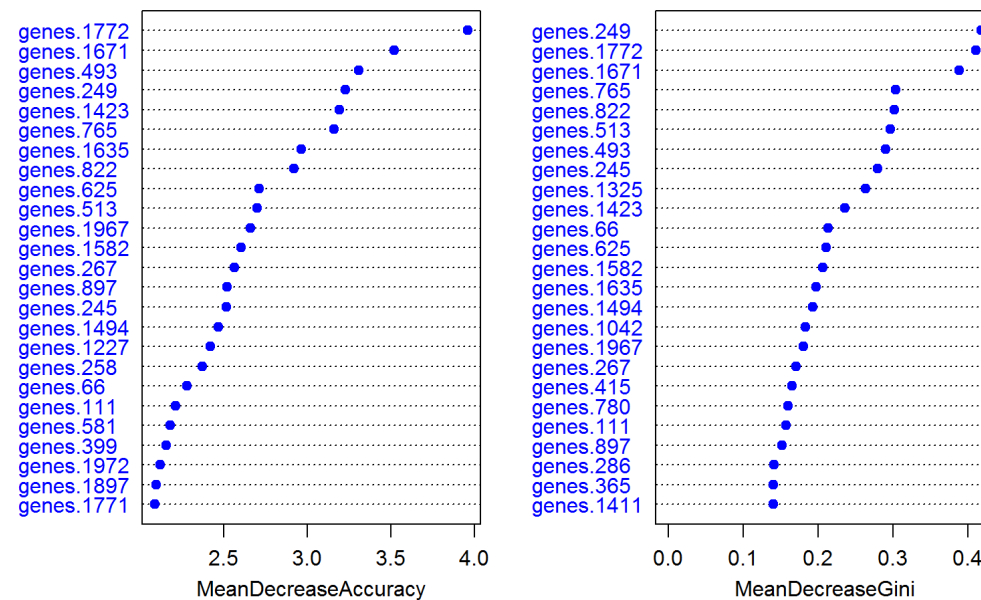
#We can predict the diseased state
print(alon.rf)
```

```
##
## Call:
## randomForest(formula = grouping ~ ., data = AlonDS, importance = TRUE,      proximity = TRUE)
##           Type of random forest: classification
##           Number of trees: 500
## No. of variables tried at each split: 44
##
##           OOB estimate of  error rate: 19.35%
## Confusion matrix:
##           colonc healthy class.error
## colonc      34        6  0.1500000
## healthy      6       16  0.2727273
```

# Random forest accuracy measures by best gene

```
## Random forest plot of gene accuracy
```

```
varImpPlot(alon.rf,
  n.var = 25,
  pch=19,
  main=NULL,
  col="blue",
  gcolor="blue",
  lcolor="black")
```



```
#Next we can find and print the genes that give the most accuracy
alon.names[249,6]
```

```
## [1] Human desmin gene, complete cds.
## 1824 Levels: ...
```

```
alon.names[1772,6]
```

```
## [1] COLLAGEN ALPHA 2(XI) CHAIN (Homo sapiens)
## 1824 Levels: ...
```

```
alon.names[1671,6]
```

```
## [1] Human monocyte-derived neutrophil-activating protein (MONAP) mRNA, complete cds.
## 1824 Levels: ...
```

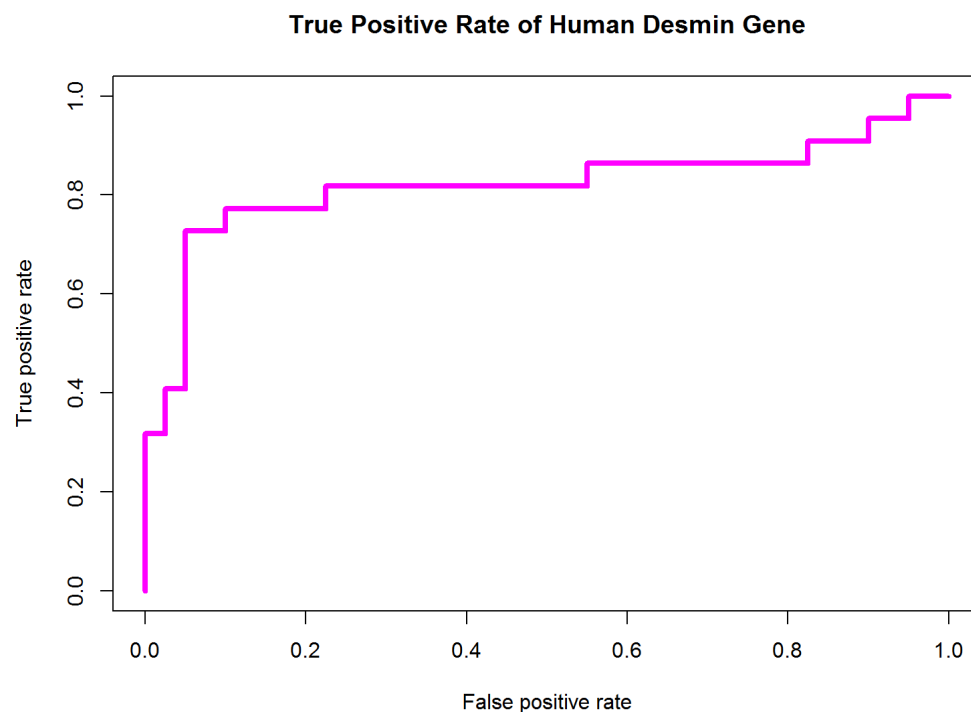
# Desmin gene prediction of diagnosis by expression level

```
colon.data <- read.table("data/AlonGene.txt")
alonLabels<-factor(Alon.c1, levels = 0:1, labels=c("colonc","healthy"));desmin <- grep("Human desmin gene, complete cds",alon.names[,6], ignore.case = TRUE)
alonPredictor <- factor(colon.data[desmin,]> 1700,levels=c("FALSE","TRUE"), labels=c("colonc","healthy"))
table(alonPredictor,alonLabels)
```

```
##           alonLabels
## alonPredictor colonc healthy
##           colonc      38      6
##           healthy      2     16
```

# We can also analyze a specific gene to measure its change of true positive or negative

```
library(ROCR)
alonLabels <- factor(Alon.cl,levels=0:1,labels= c("FALSE","TRUE"))
desmin <- grep("Human desmin gene, complete cds",alon.names[,6], ignore.case = TRUE)
pred <- prediction(talon.data[desmin,], alonLabels)
perf <- performance(pred, "tpr", "fpr" )
plot(perf,main = "True Positive Rate of Human Desmin Gene",
      lwd=4,
      col="magenta")
```



```
performance(pred,"auc")
```

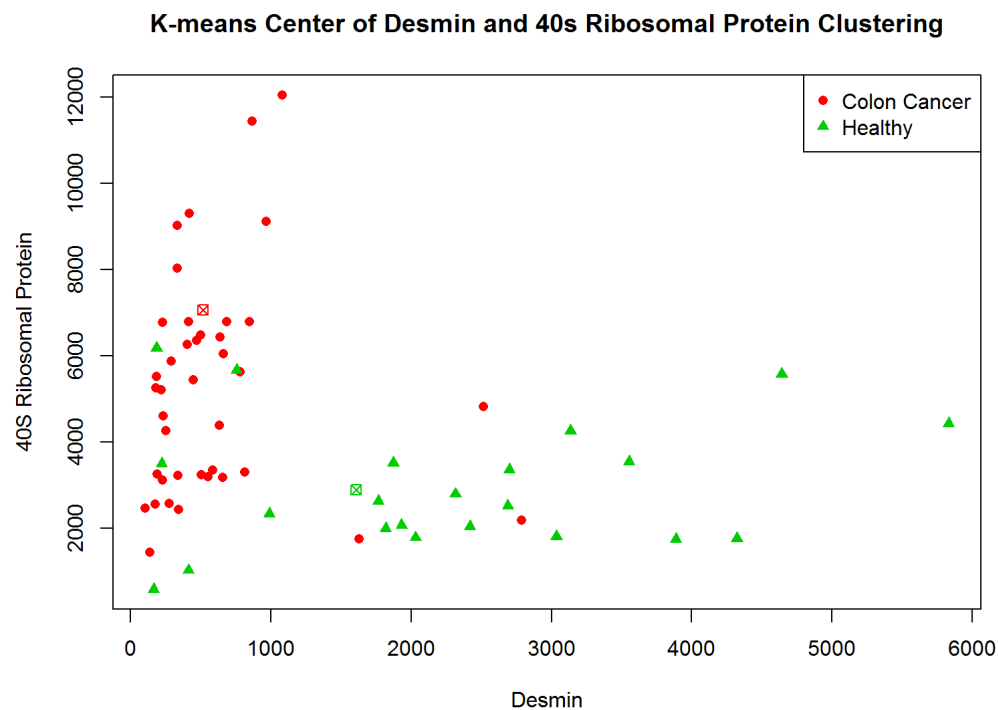
```
## An object of class "performance"
## Slot "x.name":
## [1] "None"
```



```
##  
## Slot "y.name":  
## [1] "Area under the ROC curve"  
##  
## Slot "alpha.name":  
## [1] "none"  
##  
## Slot "x.values":  
## list()  
##  
## Slot "y.values":  
## [[1]]  
## [1] 0.8204545  
##  
##  
## Slot "alpha.values":  
## list()
```

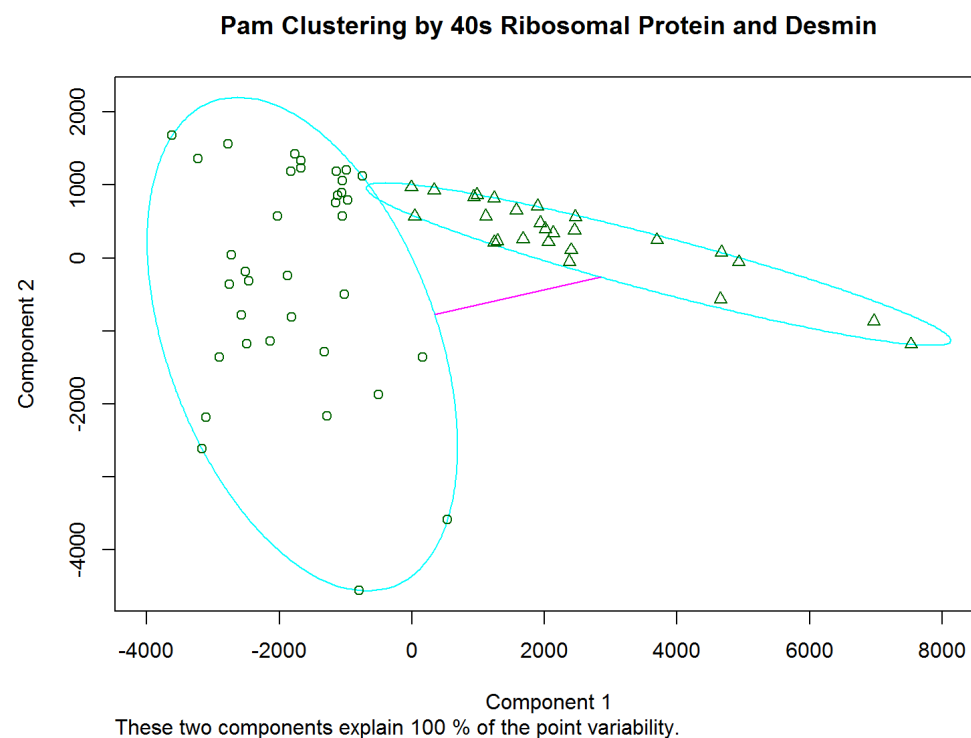
# K-means clustering is used after the top genes are found

```
RibosomalProtein <- grep("40S RIBOSOMAL PROTEIN S18",alon.names[,6], ignore.case = TRUE)
Desmin <- grep("Human desmin gene, complete cds",alon.names[,6], ignore.case = TRUE)
clusdata <- data.frame(talon.data[Desmin,],talon.data[RibosomalProtein,])
colnames(clusdata)<- c("Desmin", "40S Ribosomal Protein")
cl <- kmeans(clusdata, 2, 10)
plot(clusdata, pch=as.numeric(AlonFactor) + 15, col=as.numeric(AlonFactor) + 1,main = "K-means Center of Desmin and 40s Ribosomal Protein Clustering")
legend("topright",
      legend=c("Colon Cancer","Healthy"),
      pch=16:17,
      col=c(2,3))
points(cl$centers, col=c(3,2), pch = 7, lwd=1)
```



# Here I use PAM clustering with the genes 40s Ribosomal Protein and Desmin

```
##Desmin gene prediction of diagnosis by expression level
library(cluster)
pamx <- pam(clusdata, 2)
plot(pamx, main="Pam Clustering by 40s Ribosomal Protein and Desmin")
```



```
legend("topright", legend=c("Colon Cancer", "Healthy"), pch=16:17, col=c(2,3))
```

**Pam Clustering by 40s Ribosomal Protein and Desmin**

n = 62

