THESIS TITLE

by

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B.S., Molecular Biology and Genetics, Canakkale Onsekiz Mart University, 2018

Submitted to the Institute for Graduate Studies in
Science and Engineering in partial fulfillment of
the requirements for the degree of
Master of Science

Graduate Program in Computational Science and Engineering
Boğaziçi University

2020

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1. INTRODUCTION

1.1. Systems Biology

With the increasing availability of the computational tools and the development of high throughput techniques in the omics field, systems biology has shown a strong emergence in the last few years as a key multi-disciplinary field for integrating the multi-layer complexity of biological systems, particularly in the areas of transcriptomics, proteomics, metabolomics and fluxomics [1]. This amount of available data allows researchers to investigate molecular cell processes in a large scales, applying theoretical, experimental and computational methods.

Biological systems based on complex interactions between various molecular components. The relations between these components are often obey nonlinear kinetics, for example, most of the reactions are regulated by one or more feedback or feed-forward loops with incomprehensible behaviours. When considered, cell structure and compartmentalization are also often introduce complexities to the unexpected behavior of the entire biological system [2]. Mathematical modeling with these factors taken into consideration is used as a general approach to encompass existing knowledge in biological systems, and to gather information by analyzing these models to acquire a better understanding [3].

A mathematical model of a cell can be approached by two different approaches in either a bottom-up or top-down directionality (Figure 1.1) [4, 5]. Top-down approach is an experimental oriented approach, it starts from the whole picture and aims to characterize biological mechanisms closer to the smaller parts and their interactions in the network. In the bottom-up approach, collected data from biological knowledge is used as a starting point, a subsystem is generated to deduce the functional properties of smaller points in the network. Combination of the pathway level models (bottom-up) into a model for the entire system level (top-down) is the ultimate goal in the systems biology therefore these approaches are complementary.

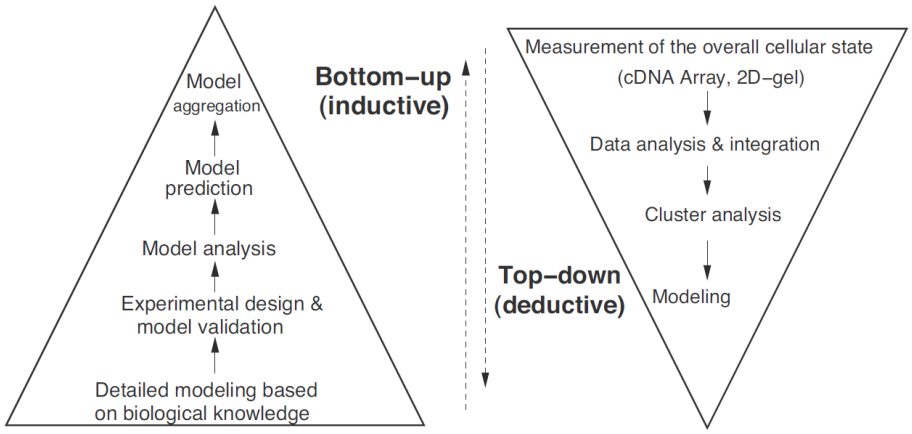

Figure 1.1. Systems biology approaches. Left: Bottom-up approach. Right: Top-down approach. Figure is taken from [3].

1.1.1. Metabolic Networks

In the context of systems biology, metabolic network reconstructions have become a common interest for the researchers over the past 20 years [6]. Organism-specific metabolic network analyses allow scientists to design experiments and even obtain beforehand predictions. These networks are the main sources of the mathematical models which can simulate metabolic fluxes reflecting the experimantal reality [7].

Before the improvement of genome sequencing or annotation technologies, initial core metabolic networks were based on the accessible information of biochemical pathways [8] [9]. In the last decade, larger genome-scale metabolic models (GSMMs) have been able to be developed rapidly with the help of databases for annotated genomes, providing information on substrates and products of each enzyme and each bioreaction [10]. Growing biochemical databases provide automatization processes for the metabolic network reconstructions. As a result, genome-scale metabolic networks are available today for almost all organisms with an annotated genome available in the literature [11, 12]. From the first genome-scale metabolic model of *Escherichia coli* to other organisms, the steps are required for GSMM development remained the same regardless of the biological diversity.

A generally applicable protocol is defined by the Palsson group [6, 10] for the reconstruction of biochemical networks described in the Figure 1.2 [13]. Briefly, genomic data for the biochemical reactions of an organism are identified from the databases, such as NCBI, DDBJ and EMBL-EBI. Extraction and processing of the gene-protein-reaction relationship (GPR) of the genomic data results a draft reconstruction. GPR associations in the draft model should be reviewed by the researchers and manually curated if the identifying process is achieved with the help of automated computational algorithms [11]. Since the genomic data is the least representative of the biological phenotypes, available transcriptomic, proteomic, metabolomic and/or subcellular localization data are also used to further curate the model. Once the final metabolic network is reconstructed with bibliographic information, it is translated into a mathematical model.

Once a metabolic network is reconstructed, a rational link between a genome sequence, the proteins encoded in the genome, and the reactions catalyzed by the proteins allowing to investigate the relationships between genotype and phenotype is achieved [14]. As the final step, GSMM needs to be validated by the new experimental data sets. GSMM validation process for various experimental conditions require detailed cultivation data from experiments. For example, information on the biomass composition of the specific organism leads more accurate biomass equation in the model, that is one of the key factors in the GSMM optimization and validation [15]. Even tough multiple steps in the GSMM reconstruction can be achieved with the automated softwares available, it is usually necessary to curate the obtained model manually.

Approaches for analyzing metabolic networks are mainly categorized as dynamic or structural approaches. Even though the former is promising more realistic approach, its implementation in the literature is obstructed due to the unavailability of kinetic parameters for the majority of enzymes within a metabolic network [16, 17] Because of the lack of kinetic parameters, structural metabolic modeling has been widely used for analyzing cellular metabolism at a steady-state assumption.

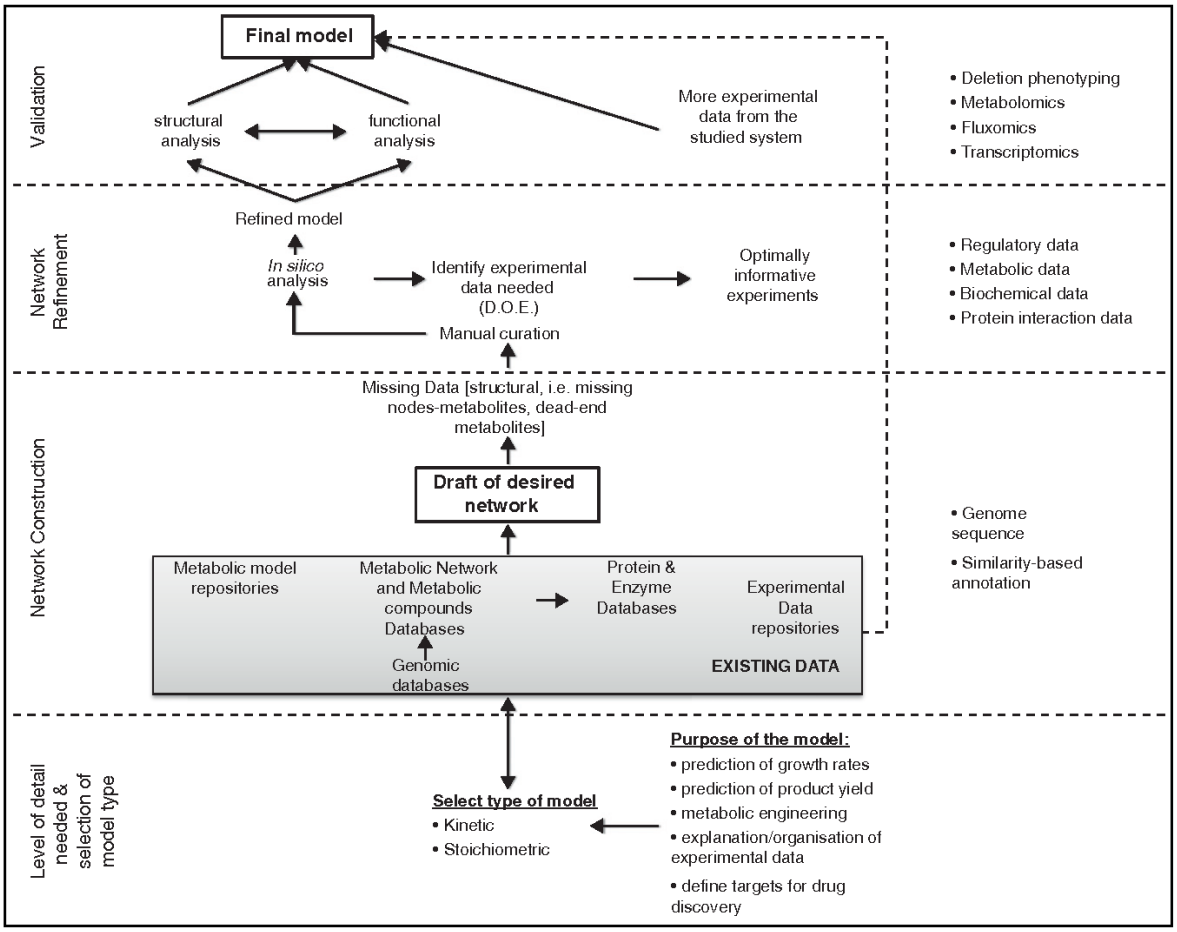

Figure 1.2. Overview of metabolic network reconstruction protocol. Figure is taken from [13].

GSMMs are one of the most useful tools in systems biology, especially in metabolic engineering studies [18]. In 1998, with the publication of *Metabolic Engineering: Principles and Methodologies*, the term metabolic engineering is defined as the optimization of natural processes within cells to increase the production of certain substances [19]. Hence, studies of metabolic engineering can be considered as genetic engineering in strain development. However, while metabolic engineering manipulates strains by altering flux distributions in the pathways; genetic engineering modificates specific genes, proteins and/or enzymes of interest [20]. Although GSMMs are mainly used in metabolic engineering strategies, other applications both for descriptive and predictive purposes can be found in the literature [21].

The ultimate goal of the GSMM reconstruction is to predict flux distribution profiles as close *in silico* as they are *in vivo*. Hence, GSMMs are in continuous research to improve predictability of organism-specific models.

Here, a figure of "GSMM citation numbers vs year" can be added from WOS

1.1.2. Mathematical Representation of Metabolic Networks

In this section, there will be a review on "Mathematical Framework Behind the Reconstruction and Analysis of Genome Scale Metabolic Models" [22], similar to slides that Ive prepared for MFA.

1.1.3. Flux Balance Analysis

In this section, there will be a review on "What is flux balance analysis?" [7]

1.2. *Saccharomyces cerevisiae*

The species "yeast" includes a range of eukaryotic single-celled microorganisms, although it is commonly used to describe *Saccharomyces cerevisiae*. Also known as the baker's yeast, *S. cerevisiae* is one of the extensively used microorganisms for alcoholic fermentation of beverages, bio-ethanol production, and processing various foods since ancient times [23]. It was the first eukaryotic organism whose genome was fully sequenced and annotated [24], and besides its benefits in the industry, it is used as a model system for other eukaryotic cells including humans [25, 26].

1.2.1. Central Carbon Metabolism of *S. cerevisiae*

From the end of the eighteenth century, mainly after the fermentation is defined as "respiration without oxygen", the metabolism of *S. cerevisiae* has been studied extensively [27, 28]. Its capability to produce ethanol is one of the most characterized microbial processes due to industrial utilization.

The set of anabolic and catabolic reactions in the cell are reffered as the metabolism. A shematic representation of the central carbon metabolism in *S. cerevisiae* can be found in Figure 1.3. Glycolysis, pentose-phosphate pathway (PPP), tricarboxylic acid cycle (TCA) or Krebs cycle, the glyoxylate cycle and the electron transport chain are the main pathways in central carbon metabolism.

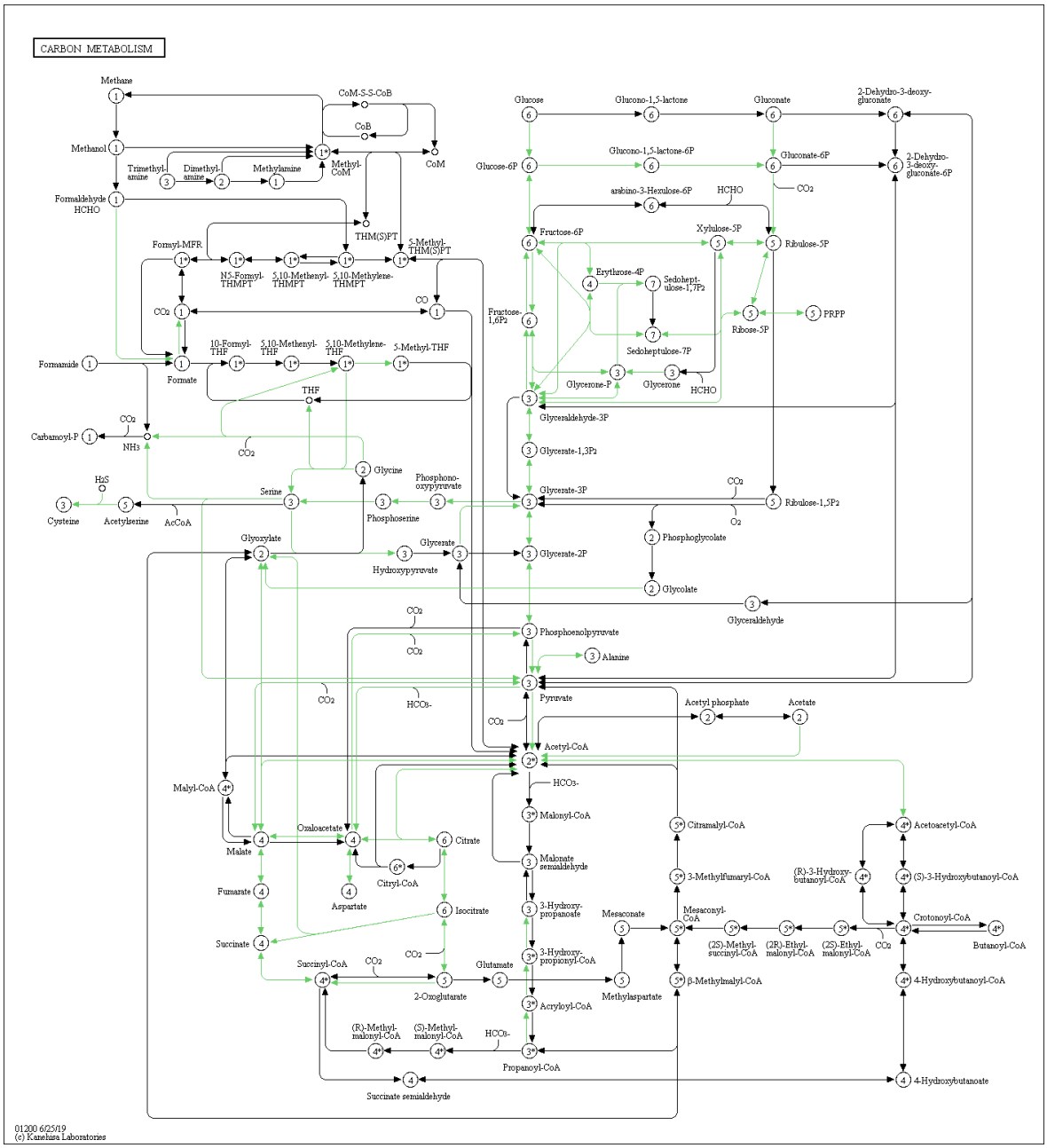

Figure 1.3. Central carbon mechanism of *S. cerevisiae* obtained from KEGG [29].

1.2.2. Metabolic Models of *S. cerevisiae*

After the first *S. cerevisiae* genome sequence is published, the first cDNA spotted microarray exploring metabolic gene regulation in 1997 [30], and the first commercial platform for oligonucleotide microarray data (Affymetrix) to investigate cellular regulations were reported in 1998 [31]. Existing genome data is integrated with the extensive annotation based on microarray data and biochemical knowledge from literature, leading of the publication of the first GSMM of *S. cerevisiae* in 2013 [32]. More in this section, there will be review on: Genome-scale modeling of yeast: chronology, applications and critical perspectives [33]

1.2.3. Applications of *S. cerevisiae* GSMMs

Literature research on the applications will be added.

1.3. Significance of Thesis

The purpose of this master's thesis is to enlighten molecular mechanisms behind the tolerance in yeast. Intracellular flux distributions of resistant strains for various substances such as caffeine, ethanol, iron, phenylethanol, nickel and sodium chloride are going to be analyzed comparetively. This study will also contribute to the global understanding of metabolic regulations in the *S. cerevisiae*, and will be further expandable into metabolic engineering studies.

2. MATERIALS AND METHODS

2.1. Experimental Data Preparation

2.1.1. Data Acquisition

Extracellular metabolomics data is obtained from Cakar's Lab [34].Briefly, they perform ethyl methane sulfonate (EMS) mutagenesis on the prototrophic *Saccharomyces cerevisiae* strain CEN.PK 113-7D (MATa, MAL2-8c, SUC2) to increase the genetic diversity as an evolutionary engineering selection strategy. Cells were inoculated in 2% Yeast Minimal Media (YMM), and the extracellular concentrations of glucose, ethanol, glycerol and acetate were measured at different time points. OD_{600} values were determined by a spectrophotometer. Additionally, cell dry weight analysis was conducted to determine biomass production. Acquired extracellular metabolite concentrations, OD_{600} values and dry weights of the reference strain (without mutagenesis) were used in this study are collected in Table 2.2 and Table 2.1.

Table 2.1. Measured OD_{600} and cell dry weight values of reference strain.

Time (h)	**OD600**	**ln(OD600)**	**Cell DW (g/L)**
0	0.21	-1.560647748	-
3	0.53	-0.634878272	-
6	1.76	0.565313809	0.9
7.5	2.66	0.978326123	-
9	4.46	1.495148766	1.9
12	5.31	1.669591835	-
15	5.88	1.771556762	-
18	5.83	1.763017	2.32
21	6.07	1.803358605	-
24	5.87	1.769854634	-
30	6.14	1.814824742	2.26
40	6.44	1.86252854	-
46	6.36	1.850028377	-
50	6.3	1.840549633	-
54	6.55	1.87946505	-
63	6.54	1.877937165	-
67	6.88	1.928618652	-
72	6.97	1.941615225	2.66

Table 2.2. Measurements of extracellular concentrations.

Time (h)	**Glucose (g/L)**	**Ethanol (g/L)**	**Glycerol (g/L)**	**Acetate (g/L)**
0	19.99	0	0	1.08
3	17.98	0.58	0.02	1.24
6	15.85	1.2	0.06	1.16
9	12.21	3.39	0.18	1.37
12	9.18	7.97	0.61	2.45
15	0.4	8.17	0.69	2.46
27	0	8.28	0.76	2.6
46	0	8	0.77	2.45
50	0	6.62	0.64	2.02
54	0	5.74	0.55	1.73
58	0	5.46	0.54	1.74
72	0	3.72	0.49	1.33

2.1.2. Determination of Rates

As the slope in the curve of $\ln OD_{600}$ as a function of time gives the growth rates of cells, natural logarithm of OD_{600} values were calculated to obtain specific growth rates by using the equation 2.1.

$$\mu = \frac{\Delta \ln OD_{600}}{\Delta t} \tag{2.1}$$

In order to determine uptake and secretion rates of the metabolites, the steady-state assumption is applied in three hours intervals as the shortest measured time-points. Missing data on cell dry weights are estimated from the OD_{600} values, and these cell dry weight data is used to calculate fluxes (in the unit of mmol/gDWh). Measurement of the cell dry weight at the 3rd hour was crucial for the steady-state

assumption, however data was not available from the experiments. Curve trend of the OD_{600} plot is used as a guide to estimate cell dry weight (Figure 2.1) Need a method here: Estimation approach, maybe regression or curve fitting?. Calculated flux values can be found in the Table 2.3.

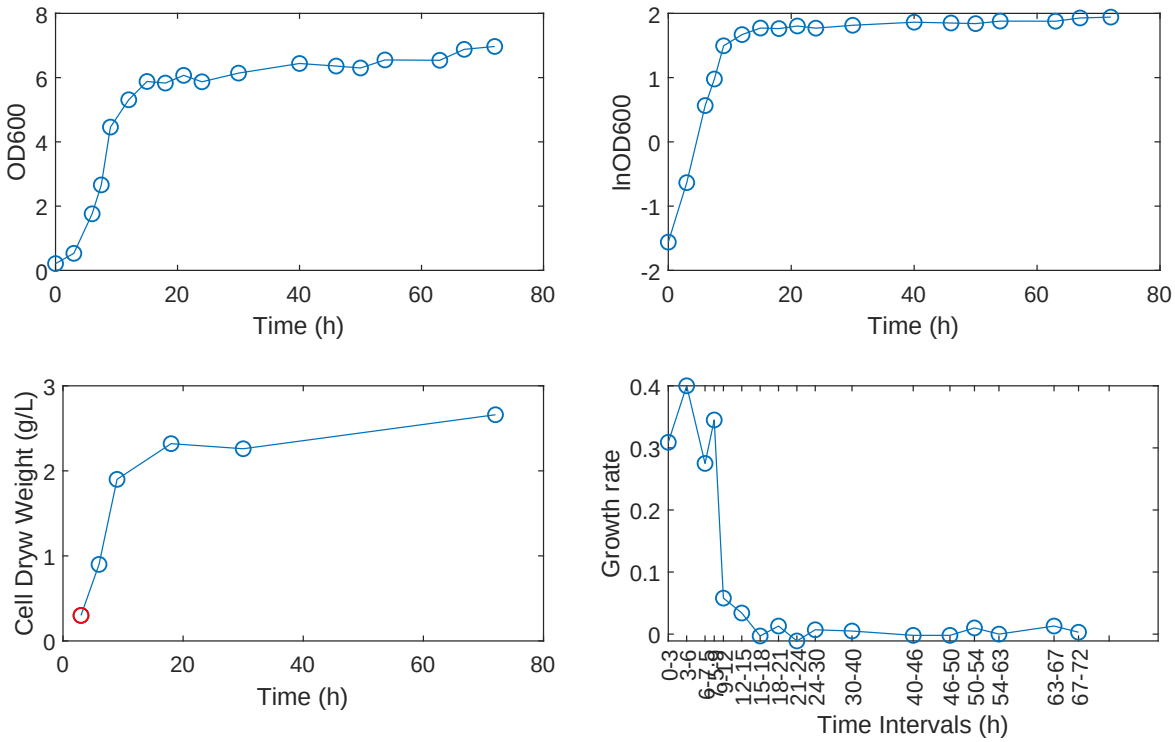

Figure 2.1. OD_{600}, $lnOD_{600}$, cell dry weights and growth rates graphs. Estimated missing cell dry weight data is shown in red color.

Table 2.3. Calculated flux values.

Time	**Metabolite fluxes in mmol/gDWh**				**Growth h-1**
	Glucose	**Ethanol**	**Glycerol**	**Acetate**	**Biomass**
0-3	-12.3963884	13.98837518	0.24131	2.960496	0.30859
3-6	-4.378823762	4.984363569	0.160873	-0.49342	0.400064

2.2. Model Selection

iAN50 [35], a stoichiometric model of intermediary metabolism including glycolysis, the pentose phosphate pathway (PPP), anaerobic excretion, citric acid cycle (TCA cycle), oxidative phosphorylation, and uptake pathways for galactose, ethanol and acetate is used to simulate batch conditions. The model was implemented based on the first GSMM of yeast, iFF708 [32] with the addition of the yeast intermediary metabolism from the RAVEN toolbox [36], and finally curated using the KEGG database [29].

Biomass equation (Eq. 2.2) in the model was left as a function of sub-reactions, for example protein, lipid, DNA synthesis reactions, so that the coefficients of biomass constituents could be optimized for each FBA simulation.

$$\begin{aligned} &0.5185\ \text{Glycogen} + 0.0234\ \text{Trehalose} + 0.8079\ \text{Mannan} + 1.1348\ \text{Glucan} \\ &+ 0.1966\ \text{DNA} + 0.012\ \text{DNA} + 4.14\ \text{Protein} + 0.0269\ \text{Lipid} + 35.3630\ \text{Maintainance} \\ &= \text{BIOMASS} \end{aligned} \quad (2.2)$$

Enzymatic reactions that are found in the model is collected in Table 2.4. In contrast to other GSMM's, each reaction was irreversible in the iAN50. This was achieved by splitting each reversible reaction into two seperate reactions in both directions.

Another reason to select iAN50 is that the total masses of enzymes catalyzing reactions in the GSMM were already estimated, and fluxes through these reactions were constrained to the biologic level, using an approach similar to intracellular crowding method using kinetic parameters [37,38]. To clarify the method briefly, a flux value for each reaction was obtained by applying a standart flux balance analysis, and this value was divided by the maximum *in vitro* activity collected from the enzyme database BRENDA [39], and a saturation factor of 0.5 (half) for simplification. Therefore, the mass of the enzymes required for that particular reaction was estimated and the

constrains are applied to the corresponding enzymatic reactions.

Table 2.4. Reaction list in the iAN50 model.

NAME	EQUATION	GENE ASSOCIATION	EC-NUMBER	SUBSYSTEM
Hexokinase	GLC[c] + ATP[c] =>ADP[c] + G6P[c]	YFR053C; YGL253W; YCL040W	2.7.1.1 OR 2.7.1.2	Glycolysis
Glucose-6-phosphate isomerase	G6P[c] <=>F6P[c]	YBR196C	5.3.1.9	Glycolysis
Phosphofructokinase	ATP[c] + F6P[c] =>ADP[c] + F16P[c]	YGR240C; YMR205C	2.7.1.11	Glycolysis
Fructose-1,6-bisphosphatase	F16P[c] =>F6P[c] + PI[c]	YLR377C	3.1.3.11	Glycolysis
Fructose-bisphosphate aldolase	F16P[c] <=>GA3P[c] + DHAP[c]	YKL060C	4.1.2.13	Glycolysis
Triosephosphate isomerase	DHAP[c] <=>GA3P[c]	YDR050C	5.3.1.1	Glycolysis
Triosephosphate dehydrogenase	GA3P[c] + NAD[c] + PI[c] <=>P13G[c] + NADH[c]	YJL052W; YJR009C; YGR192C	1.2.1.12	Glycolysis
Phosphoglycerate kinase	P13G[c] + ADP[c] <=>P3G[c] + ATP[c]	YCR012W	2.7.2.3	Glycolysis
Phosphoglycerate mutase	P3G[c] <=>P2G[c]	YKL152C; YDL021W; YOL056W	5.4.2.11	Glycolysis
Enolase	P2G[c] <=>PEP[c]	YGR254W; YHR174W; YOR393W; YPL281C; YMR323W	4.2.1.11	Glycolysis
Pyruvate kinase	ADP[c] + PEP[c] =>ATP[c] + PYR[c]	YOR347C; YAL038W	2.7.1.40	Glycolysis
Glucose-6-phosphate 1-dehydrogenase	G6P[c] + NADP[c] =>G15L[c] + NADPH[c]	YNL241C	1.1.1.49	Pentose Phosphate
6-phosphogluconolactonase	G15L[c] =>P6G[c]	YNR034W; YCR073W-A; YHR163W; YGR248W	3.1.1.31	Pentose Phosphate
6-phosphogluconate dehydrogenase, decarboxylating	P6G[c] + NADP[c] =>CO2[c] + RU5P[c] + NADPH[c]	YGR256W; YHR183W	1.1.1.44	Pentose Phosphate
ribose 5-phosphate isomerase	RU5P[c] <=>R5P[c]	YOR095C	5.3.1.6	Pentose Phosphate
Ribulose-phosphate 3-epimerase	RU5P[c] <=>X5P[c]	YJL121C	5.1.3.1	Pentose Phosphate
Transketolase	R5P[c] + X5P[c] <=>GA3P[c] + S7P[c]	YBR117C; YPR074C	2.2.1.1	Pentose Phosphate
Transaldolase	GA3P[c] + S7P[c] <=>F6P[c] + E4P[c]	YLR354C	2.2.1.2	Pentose Phosphate
Transketolase	E4P[c] + X5P[c] <=>F6P[c] + GA3P[c]	YBR117C; YPR074C	2.2.1.1	Pentose Phosphate
Pyruvate carboxylase 1	ATP[c] + CO2[c] + PYR[c] =>ADP[c] + OAA[c] + PI[c]	YGL062W; YBR218C	6.4.1.1	TCA
Citrate synthase, mitochondrial	ACCOA[m] + OAA[m] =>CI[m] + COA[m]	YNR001C; YPR001W	2.3.3.1	TCA
Aconitate hydratase, mitochondrial	CI[m] <=>ICI[m]	YLR304C	4.2.1.3	TCA
Isocitrate dehydrogenase	ICI[m] + NAD[m] =>AKG[m] + CO2[m] + NADH[m]	YNL037C; YOR136W	1.1.1.41	TCA
Isocitrate dehydrogenase [NADP], mitochondrial	ICI[m] + NADP[m] =>AKG[m] + CO2[m] + NADPH[m]	YDL066W; YLR174W	1.1.1.42	TCA
Alpha-ketoglutarate dehydrogenase	AKG[m] + NAD[m] + ADP[m] + PI[m] =>CO2[m] + NADH[m] + ATP[m] + SUC[m]	YDR148C; YIL125W	1.2.4.2	TCA
Succinate dehydrogenase complex	FAD[m] + SUC[m] =>FADH2[m] + FUM[m]	YKL148C; YLL041C	1.3.5.1	TCA
Fumarate reductase	FADH2[m] + FUM[m] =>FAD[m] + SUC[m]	YJR051W; YEL047C	1.3.1.6	TCA
Fumarate hydratase	FUM[m] <=>MAL[m]	YPL262W	4.2.1.2	TCA
Malate dehydrogenase	MAL[m] + NAD[m] <=>NADH[m] + OAA[m]	YKL085W	1.1.1.37	TCA
Galactokinase	GAL[c] + ATP[c] =>GALP[c] + ADP[c]		2.7.1.6	Galactose metabolism
UDP-glucose 4-epimerase	GALUDP[c] <=>GLUUDP[c]		5.1.3.2	Galactose metabolism
Galactose-1-phosphate uridylyltransferase	GLUUDP[c] + GALP[c] <=>G1P[c] + GALUDP[c]		2.7.7.12	Galactose metabolism
Phosphoglucomutase-1	G1P[c] <=>G6P[c]		5.4.2.2	Galactose metabolism
glycerol-3-phosphate dehydrogenase	DHAP[c] + NADH[c] =>GP[c] + NAD[c]	YDL022W; YOL059W	1.1.1.8	Anaerobic excretion
sn-glycerol-3-phosphate phosphohydrolase	GP[c] =>GLY[c] + PI[c]	YER062C; YIL053W	3.1.3.21	Anaerobic excretion
Pyruvate decarboxylase	PYR[c] =>ACA[c] + CO2[c]	YGR087C; YLR134W; YLR044C	4.1.1.1	Anaerobic excretion
Alcohol dehydrogenase	ACA[c] + NADH[c] <=>ETH[c] + NAD[c]	YGL256W; YMR303C; YOL086C	1.1.1.1	Anaerobic excretion
Aldehyde dehydrogenase	ACA[c] + NADP[c] =>AC[c] + NADPH[c]	YPL061W	1.2.1.3	Anaerobic excretion
Aldehyde dehydrogenase [NAD(P)+] 1	ACA[c] + NAD[c] =>NADH[c] + AC[c]	YMR170C; YMR169C; YOR374W; YOR374W; YER073W	1.2.1.5	Aromatic amino acid biosynthesis
Isocitrate lyase	ICI[m] =>Glyoxylate[m] + SUC[m]	YER065C; YPR006C	4.1.3.1	Anaplerotic reactions
Malate synthase 1, glyoxysomal	ACCOA[m] + Glyoxylate[m] =>COA[m] + MAL[m]	YIR031C; YNL117W	2.3.3.9	Anaplerotic reactions
NADH-ubiquinone oxidoreductase, mitochondrial ("Complex 1")	NADH[m] + Ubiquinone-9[m] =>Ubiquinol[m] + NAD[m]	YML120C; YMR145C	1.6.5.3 (1.6.5.9)	Oxidative Phosphorylation
External NADH-ubiquinone oxidoreductase 2, mitochondrial ("Complex 1")	NADH[c] + Ubiquinone-9[m] =>Ubiquinol[m] + NAD[c]	YDL085W	1.6.5.3 (1.6.5.9)	Oxidative Phosphorylation
Succinate dehydrogenase [ubiquinone] cytochrome b subunit, mitochondrial (Complex II)	FADH2[m] + Ubiquinone-9[m] <=>FAD[m] + Ubiquinol[m]	YKL141W; YDR178W	1.3.5.1	Oxidative Phosphorylation
Cytochrome b-c1 complex subunit Rieske, mitochondrial (complex III)	Ubiquinol[m] + 2 Ferricytochrome_c[m] + 1.5 H[m] =>Ubiquinone-9[m] + 2 Ferrocytochrome_c[m] + 1.5 HMit[m]	YEL024W; YBL045C; YHR001W-A; YPR191W; YFR033C; YDR529C; YJL166W; YGR183C	1.10.2.2	Oxidative Phosphorylation
Cytochrome c oxidase subunit 1 (Complex IV)	Ferrocytochrome_c[m] + 0.25 O2[c] + 1.5 H[m] =>Ferricytochrome_c[m] + 1.5 HMit[m]	YNL052W; YIL111W; YLR395C; Q0045; Q0250; YGL187C; YHR051W; YGL191W; YLR038C; YMR256C; YDL067C	1.9.3.1	Oxidative Phosphorylation
ATP synthase subunit alpha, mitochondrial (Complex V)	ADP[m] + PI[m] + 3 HMit[m] =>ATP[m] + 3 H[m]	YBL099W; Q0080; YPL078C; YDR298C; Q0130; Q0085; YJR121W; YKL016C; YDL004W; YDR322C-A; YPL271W; YDR377W; YPR020W; YBR039W; YLR295C; YML081C-A; YOL077W-A; YML042W	3.6.3.14	Oxidative Phosphorylation
ATP hydrolysis	ATP[c] =>ADP[c] + PI[c]			Other
Pyruvate dehydrogenase complex	COA[m] + NAD[m] + PYR[m] =>ACCOA[m] + CO2[m] + NADH[m]	YER178W; YFL018C; YBR221C	1.2.4.1	Other
Acetyl-coenzyme A synthetase 1	AC[c] + 2 ATP[c] + COA[c] =>ACCOA[c] + 2 ADP[c] + 2 PI[c]	YAL054C; YLR153C	6.2.1.1	Other
Phosphoenolpyruvate carboxykinase	ATP[c] + OAA[c] =>ADP[c] + CO2[c] + PEP[c]	YKR097W	4.1.1.49	Other

2.3. Flux Balance Analysis

Uptake reaction of glucose with the secretion reactions of glycerol and acetate were constrained according to the calculated flux values in Table 2.3, for both time intervals seperately. Since the main goal was to validate model for experimental conditions, ethanol was not constrained in regard to be used as the control metabolite. Experiments were done in fully aerobic conditions, therefore oxygen uptake reaction was set unlimited.

Coefficients of the biomass constituents are defined as the same as the batch conditions in the reference article [35], for the reason that detailed knowledge is not available in the acquired experimental data. Coefficients for the final biomass equation can be found in the Table 2.5.

Table 2.5. Biomass coefficients that are used in the simulation.

Constituent	**Coefficient**
Protein	3.703704
RNA	0.37037
DNA	0.018519
Lipid	0.041667
Glycogen	0.030864
Trehalose	0.029214
Mannan	0
Glucan	2.469136
Maintainance	40

For the linear optimization, an implementation of pFBA from the reference publication was performed [35, 40]. After solving the system using a linear solver with the objective maximizing growth, the solution was used as a constraint. From that point, a second optimization was run to minimize the sum of all other fluxes.

2.4. Visualization of the Model

The GSMM was visualized in Cytoscape [41] using the Fluxviz plug-in [42] and the solution fluxes were mapped onto edges. Network was imported in SBML [43] and the flux distributions were imported in csv format.

3. RESULTS

3.1. Intracellular Flux Distributions

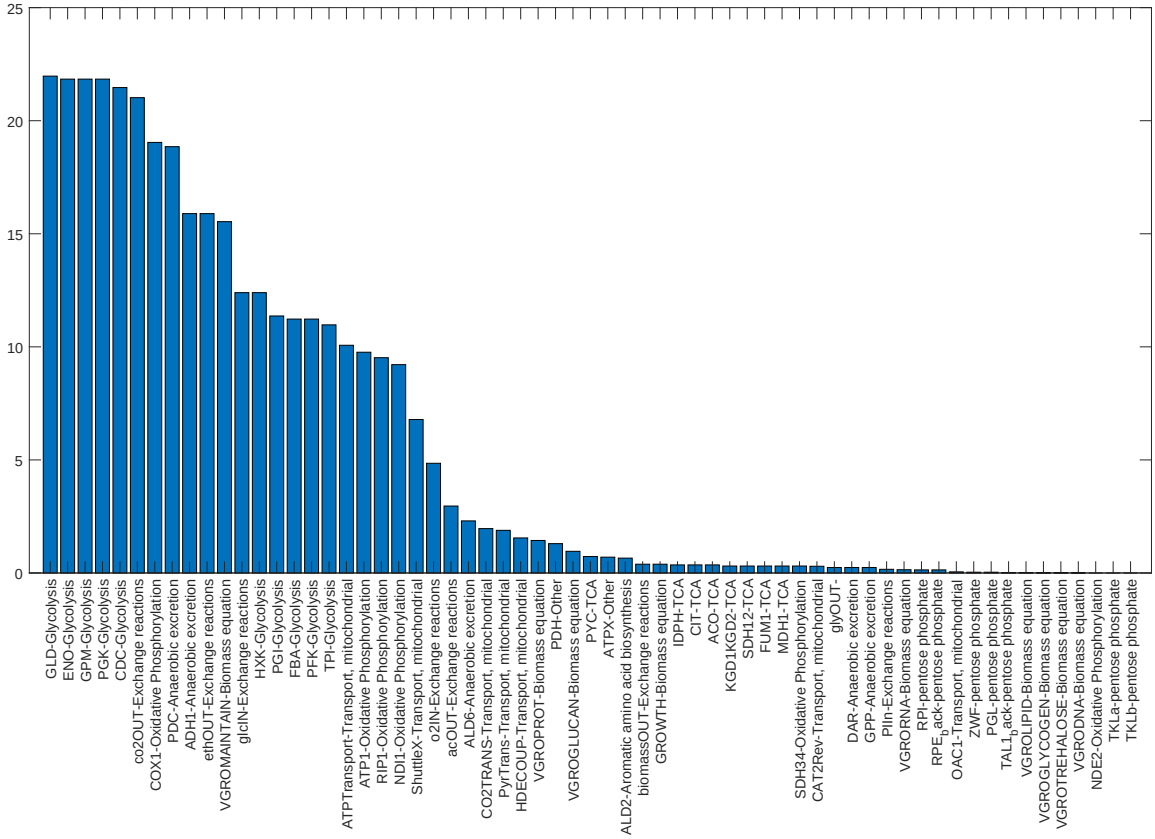

Figure 3.1. Non-zero fluxes in the solution.

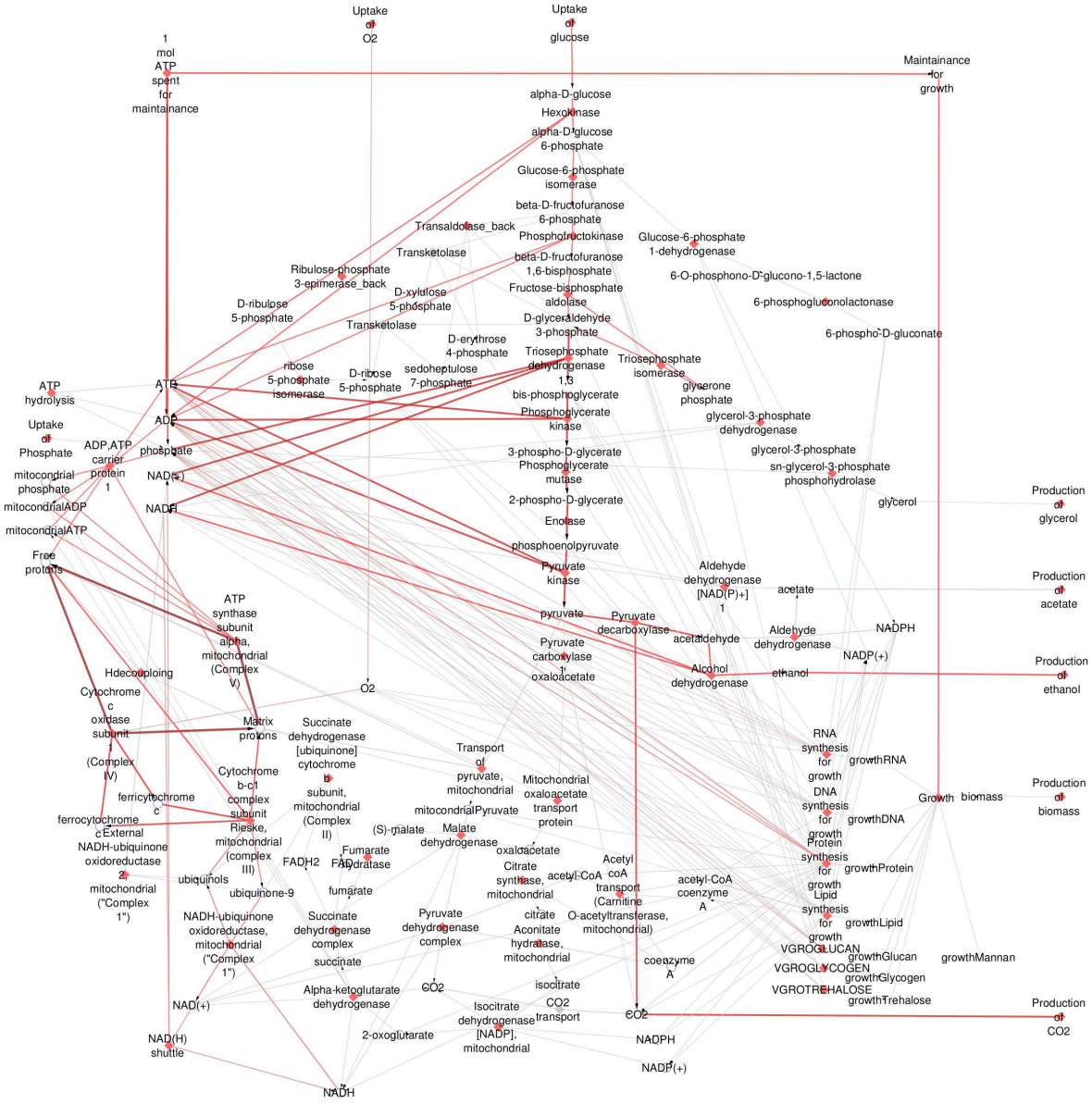

Figure 3.2. Non-zero flux distributions on the map.

4. DISCUSSION

5. EXPANDABLE TOPICS

5.1. Following topics can be added into introduction

Biological pathways (explaining each in detail, NAD regulation etc.), Fermantation (industrial applications, bio-ethanol, shift echanisms in the yeast), Regulation Strategies in Cells (feedback/feedforward loops with figures), Toolboxes (such as COBRA, RAVEN or Other Softwares and Algorithms), FBA Methods (pFBA, dFBA, TFBA etc), Omics Data (subtopics for each omics, Model Integration Methods, maybe Transcriptomics Analysis if it is going to be used), Control analysis...

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