Assignment 3: Data Exploration

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

- 1. Rename this file <FirstLast>_A03_DataExploration.Rmd (replacing <FirstLast> with your first and last name).
- 2. Change "Student Name" on line 3 (above) with your name.
- 3. Work through the steps, creating code and output that fulfill each instruction.
- 4. Assign a useful name to each code chunk and include ample comments with your code.
- 5. Be sure to **answer the questions** in this assignment document.
- 6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
- 7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no install.packages() or View() commands exist in your code.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse, lubridate), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets "Neonics" and "Litter", respectively. Be sure to include the subcommand to read strings in as factors.

```
#checking that working directory is correct
getwd()
```

[1] "/Users/hannahnelson/Desktop/env872/EDA-Fall2023"

```
#loading in packages
library(tidyverse)
library(lubridate)
#reading in and viewing each data set
```

```
Neonics <- read.csv(".//Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv", stringsAsFactors = TRUE)
#View(Neonics)
Litter <- read.csv(".//Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv", stringsAsFactors = TRUE)
#View(Litter)</pre>
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Studying the ecotoxicology of neonicotinoids on insects is important because understanding how a widely used class of insecticides impacts different species of insects is necessary knowledge for informed decisions to be made about know to use neonicotinods responsibly. Looking at data on this relationship can show which insect species are being successfully targetted, and if any species are developing resistance. This is helpful because it allows people using neonicotinoids to know which species they will be able to target, and if species they were are intending to target could react lethally to neonicotinoids.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Litter and woody debris are important to study becuase they play an important role in eocsystems. They play a role in nutrient cycling and carbon sequestration, act as a barrier to erosion, and provide habitat building materials for species. Looking at litter and woody debris falling to the forest floor over time can provide insight into the health of forest ecosystems, and the carbon sequestration potential of forests.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON_Litterfall_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Litter and fine woody debris sampling is executed at terrestrial NEON sites that contain woody vegetation over 2 meters tall. 2. An elevated and ground litter trap are deployed for every 400 square meters of of plot area. 3. Target sampling frequency for elevated traps is determined by what vegetation is present at the site. On average, decidious forest sites are sampled every two weeks and evergreen forest sites are sampled once every one to two months.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Neonics)
```

[1] 4623 30

#the data set has 4623 rows and 30 columns

6. Using the summary function on the "Effect" column, determine the most common effects that are studied. Why might these effects specifically be of interest?

summary(Neonics\$Effect) %>% sort(decreasing=TRUE)

##	Population	Mortality	Behavior	Feeding behavior
##	1803	1493	360	255
##	Reproduction	Development	Avoidance	Genetics
##	197	136	102	82
##	Enzyme(s)	Growth	Morphology	Immunological
##	62	38	22	16
##	Accumulation	Intoxication	Biochemistry	Cell(s)
##	12	12	11	9
##	Physiology	Histology	Hormone(s)	
##	7	5	1	

Answer: The most common effects studied are population & mortality. These effects are specifically of interest because the primary purpose of the data is to understand how effective neonicotinoids are on insect species, which can be best understood by looking at population and mortality data.

7. Using the summary function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.[TIP: The sort() command can sort the output of the summary command...]

summary(Neonics\$Species.Common.Name) %>% sort(decreasing=TRUE)

##	(Other)	Honey Bee
##	670	667
##	Parasitic Wasp	Buff Tailed Bumblebee
##	285	183
##	Carniolan Honey Bee	Bumble Bee
##	152	140
##	Italian Honeybee	Japanese Beetle
##	113	94
##	Asian Lady Beetle	Euonymus Scale
##	76	75
##	Wireworm	European Dark Bee
##	69	66
##	Minute Pirate Bug	Asian Citrus Psyllid
##	62	60
##	Parastic Wasp	Colorado Potato Beetle
##	58	57
##	Parasitoid Wasp	Erythrina Gall Wasp
##	51	49
##	Beetle Order	Snout Beetle Family, Weevil
##	47	47
##	Sevenspotted Lady Beetle	True Bug Order

##	46	45
##	Buff-tailed Bumblebee	Aphid Family
##	39	38
##	Cabbage Looper	Sweetpotato Whitefly
##	38	37
##	Braconid Wasp	Cotton Aphid
##	33	33
##	Predatory Mite	Ladybird Beetle Family
##	33	30 Scarab Beetle
## ##	Parasitoid 30	Scarab Beetle
##	Spring Tiphia	Thrip Order
##	29	29
##	Ground Beetle Family	Rove Beetle Family
##	27	27
##	Tobacco Aphid	Chalcid Wasp
##	27	25
##	Convergent Lady Beetle	Stingless Bee
##	25	25
##	Spider/Mite Class	Tobacco Flea Beetle
##	24	24
## ##	Citrus Leafminer 23	Ladybird Beetle 23
##	Mason Bee	Mosquito
##	22	22
##	Argentine Ant	Beetle
##	21	21
##	Flatheaded Appletree Borer	Horned Oak Gall Wasp
##	20	20
##	Leaf Beetle Family	Potato Leafhopper
##	20	20
##	Tooth-necked Fungus Beetle	Codling Moth
## ##	20 Black-spotted Lady Beetle	19 Calico Scale
##	18	18
##	Fairyfly Parasitoid	Lady Beetle
##	18	18
##	Minute Parasitic Wasps	Mirid Bug
##	18	18
##	Mulberry Pyralid	Silkworm
##	18	18
##	Vedalia Beetle	Araneoid Spider Order
##	18	17
## ##	Bee Order 17	Egg Parasitoid 17
##	Insect Class	Moth And Butterfly Order
##	17	17
##		Hemlock Woolly Adelgid Lady Beetle
##	17	16
##	Hemlock Wooly Adelgid	Mite
##	16	16
##	Onion Thrip	Western Flower Thrips
##	16	15
##	Corn Earworm	Green Peach Aphid

```
##
                                      14
                                                                             14
                              House Fly
##
                                                                     Ox Beetle
##
                                      14
                                                                             14
##
                    Red Scale Parasite
                                                           Spined Soldier Bug
##
                                                             Diamondback Moth
##
                 Armoured Scale Family
##
                                                                             13
##
                          Eulophid Wasp
                                                            Monarch Butterfly
##
                                      13
                                                                             13
##
                          Predatory Bug
                                                        Yellow Fever Mosquito
##
                                      13
                                                                             13
                   Braconid Parasitoid
##
                                                                 Common Thrip
##
                                                                             12
         Eastern Subterranean Termite
##
                                                                        Jassid
##
                                      12
                                                                             12
##
                             Mite Order
                                                                     Pea Aphid
##
                                      12
                                                                             12
##
                       Pond Wolf Spider
                                                    Spotless Ladybird Beetle
##
                                                                             11
##
                Glasshouse Potato Wasp
                                                                      Lacewing
##
##
               Southern House Mosquito
                                                     Two Spotted Lady Beetle
##
                                      10
                             Ant Family
##
                                                                 Apple Maggot
                                                                             9
##
                                       9
```

Answer: Six most commonly studied species: 1. Honey Bee 2. Parasitic Wasp 3. Buff Tailed Bumblebee 4. Carniolan Honey Bee 5. Bumble Bee 6. Italian Honeybee These insects are likely of interest over other insects because many bee species are vulnerable or endangered. Understanding if the use of neonicotinoids is harming species of bees is important in the effort to save bee species from extinction and preserve biodiversity. While parasitic wasps are not endangered, they are also a species of insect that is beneficial to the environment and could be harmed by the use of neonicotinoids.

8. Concentrations are always a numeric value. What is the class of Conc.1..Author. column in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

```
## [1] "factor"
```

Answer: The class of "Conc.1..Author.." is factor. It is not numeric because the data is discrete, not continuous.

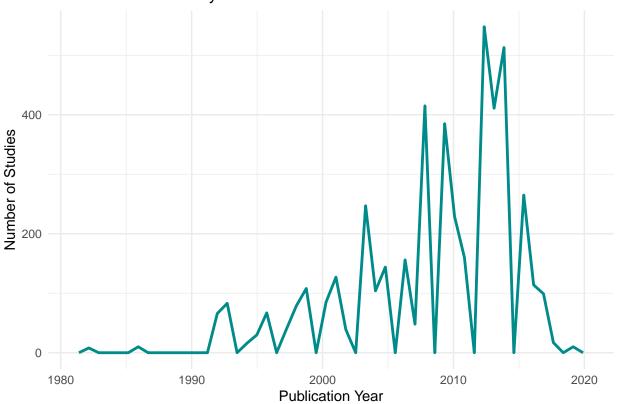
Explore your data graphically (Neonics)

9. Using geom freqpoly, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) +
  geom_freqpoly(aes(x = Publication.Year), bins=50, color="cyan4", size=1)+
  theme_minimal()+
  labs(title="Studies Conducted by Publication Year", x="Publication Year", y="Number of Studies")
```

```
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```

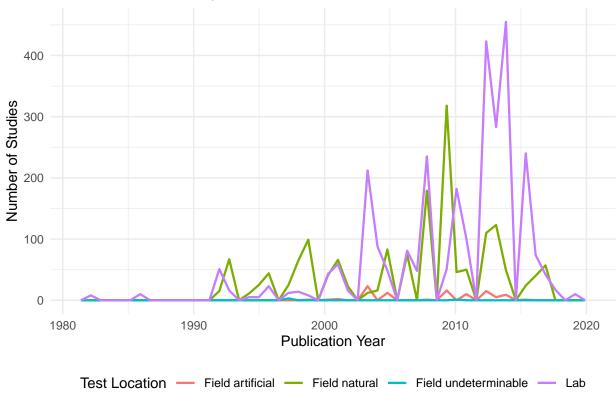
Studies Conducted by Publication Year



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics) +
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins=50, size=0.8)+
  theme_minimal()+
  theme(legend.position = "bottom")+
  scale_color_discrete(name="Test Location")+
  labs(title="Studies Conducted by Test Location and Publication Year", x="Publication Year", y="Number")
```





Interpret this graph. What are the most common test locations, and do they differ over time?

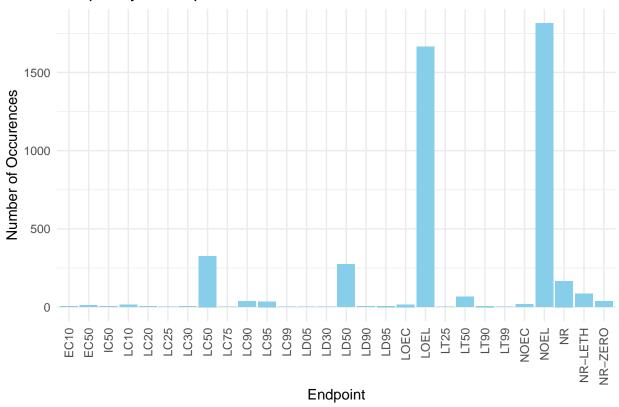
Answer: The most common test locations are lab & field natural. In the 1990s, field natural was the most common test location. Lab because the most common test location in the early 2000s, and has remained in that position except for a brief period around 2008-2009 when field natural overtook lab as the most common. The use of both lab and field natural locations for tests have increased over time with similar patterns.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[TIP: Add theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1)) to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics, aes(x = Endpoint)) +
  geom_bar(fill="skyblue") +
  theme_minimal() +
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))+
  labs(title="Frequency of Endpoint Occurences", x="Endpoint", y="Number of Occurences")
```





Answer: The two most common endpoints are NOEL & LOEL 1. NOEL (no observable effect level): The highest dose produced effects that were not significantly different from responses of controls 2. LOEL (lowest observable effect level): The lowest dose produced effects that were significantly different from the responses of controls

Explore your data (Litter)

12. Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the unique function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate)

## [1] "factor"

#the class of "collectDate" is factor before transforming data

#transforming "collectDate" to date class
Litter$collectDate <- as.Date(Litter$collectDate)

class(Litter$collectDate)</pre>
```

```
#after the transformation the class of "collectDate" is date
unique(Litter$collectDate)

## [1] "2018-08-02" "2018-08-30"

#litter was collected on the 2nd and 30th of August 2018
```

13. Using the unique function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from unique different from that obtained from summary?

```
unique(Litter$plotID)

## [1] NIW0_061 NIW0_064 NIW0_067 NIW0_040 NIW0_041 NIW0_063 NIW0_047 NIW0_051
## [9] NIW0_058 NIW0_046 NIW0_062 NIW0_057
## 12 Levels: NIW0_040 NIW0_041 NIW0_046 NIW0_047 NIW0_051 NIW0_057 ... NIW0_067

#returns list of each unique plot ID

unique(Litter$plotID) %>% length()

## [1] 12
```

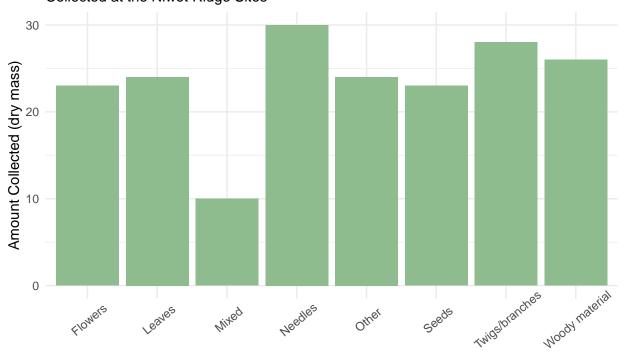
```
#returns the length of the list
#There were 12 plots sampled at Niwot Ridge: NIWO_061, NIWO_064, NIWO_067, NIWO_040, NIWO_041, NIWO_063
```

Answer: Information obtained using unique is different than information obtained using summary because unique removes duplicate values to only give one of each, while summary would include duplicate values. This was important because plot ID and date values repeat multiple times in the data set. Using unique allowed those duplicate values to each only be seen once, which provided clarity in data exploration.

14. Create a bar graph of functional Group counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter, aes(x=functionalGroup))+
  geom_bar(fill="darkseagreen") +
  theme_minimal()+
  theme(axis.text.x = element_text(angle = 40, vjust = .8))+
  labs(title="Type and Quantity of Litter", subtitle="Collected at the Niwot Ridge Sites", x= "Function")
```

Type and Quantity of Litter Collected at the Niwot Ridge Sites



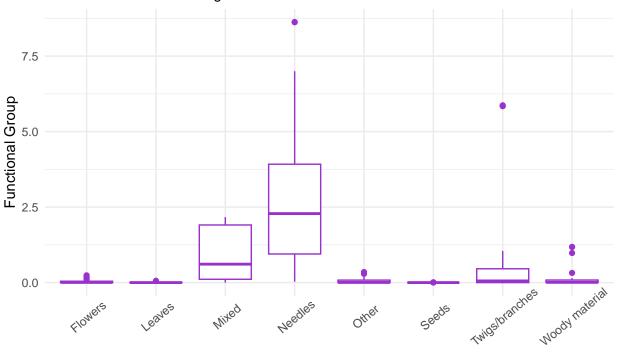
Functional Group

15. Using geom_boxplot and geom_violin, create a boxplot and a violin plot of dryMass by functional-Group.

```
ggplot(Litter)+
  geom_boxplot(aes(x=functionalGroup, y=dryMass), color="darkorchid")+
  theme_minimal()+
  theme(axis.text.x = element_text(angle = 40, vjust = .8))+
  labs(title="Type and Quantity of Litter", subtitle="Collected at the Niwot Ridge Sites", x="Amount Co
```

Type and Quantity of Litter

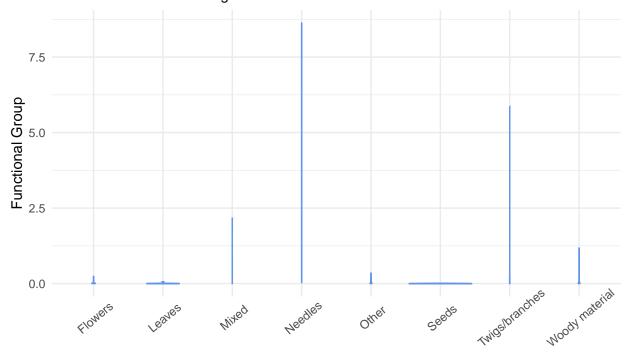
Collected at the Niwot Ridge Sites



Amount Collected (dry mass)

```
ggplot(Litter)+
geom_violin(aes(x=functionalGroup, y=dryMass),
    draw_quantiles = c(0.25, 0.5, 0.75), color="cornflowerblue")+
theme_minimal()+
theme(axis.text.x = element_text(angle = 40, vjust = .8))+
labs(title="Type and Quantity of Litter", subtitle="Collected at the Niwot Ridge Sites", x="Amount Co
```

Type and Quantity of Litter Collected at the Niwot Ridge Sites



Amount Collected (dry mass)

Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: A boxplot is more effective than a violin plot in this case because there is not enough distribution of data points within the interquartile ranges of each litter type for a violin plot to produce a useful visualization.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles & mixed litter types tend to have the highest biomass at the Niwot Ridge sites.