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**Global standardization of micronutrient biomarker statistical analysis: An introduction to the SAMBA R package**[[1]](#footnote-1)**,**[[2]](#footnote-2)**,**[[3]](#footnote-3)

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**Abstract (structured)**

**Background**:

To justify, plan, and manage successful micronutrient intervention programs, micronutrient biomarkers are the most essential. However, to generate accurate results from micronutrient research or surveys, analysts need to understand the biological basis for micronutrient biomarkers (e.g., the mechanism of how inflammation influences micronutrient biomarkers), equip with statistical knowledge, and have familiarity with basic computer code.

**Objectives:** We introduce the Statistical Apparatus of Micronutrient Biomarker Analysis (SAMBA) package, a new tool to increase accessibility of micronutrient biomarker analysis.

**Method:** The freely available and open-access SAMBA package written in R software simplifies estimations of micronutrient biomarker distribution and prevalence of deficiencies, which can drastically shorten the time from data cleaning to presentation of a final results table. We explain the underlying theory behind the tool, include the OSF link where its package and associated documentation can be freely downloaded, and provide examples of potential applications.We used the SAMBA package to analyze various biomarkers, including retinol binding protein, serum retinol, Red Blood Cell and serum folate, serum ferritin, and Soluble transferrin receptor, and apply inflammation adjustment when appropriate among preschool-age children using a single-site observational study in Kenya and among women using data from the US National Health and Examination Survey (NHANES) and compared to results generated by an independent analyst.

**Results:**

The SAMBA package generated identical results vs from the analyst. The examples demonstrate application of the SAMBA package to micronutrient biomarker data with proper inflammation adjustment and statistical implementation.

**Conclusion:** The SAMBA packages may facilitate the analysis of micronutrient biomarker data to inform nutrition research, programs, and policy.

**Word count:** 259

**Keywords:** Micronutrients, biomarkers, prevalence, biochemical, statistical analysis, software, inflammation adjustment

**Introduction**

Vitamin and mineral deficiencies, also known as hidden hunger, has been said to affect more than 2 billion individuals worldwide (1). Deficiencies in these micronutrients can hinder growth and cognitive development and increase risk of morbidity and mortality (2). Micronutrient intervention programs, such as large-scale fortification and supplementation, and food-based interventions are available to address this burden. To justify, plan, and manage successful interventions, a broad range of information is needed, such as data on food availability, food and nutrient intake, micronutrient status, and intervention program coverage. Analysis of micronutrient biomarkers is essential, as these data allow for estimation of the population prevalence and distribution of micronutrient deficiencies (3).

The 2017 Global Nutrition Report (GNR) proposed a “nutrition data revolution”, calling for “better use of data that is collected to create a more responsive information system” for policy decision-making (3). This revolution does not only apply to low- and middle- income countries (LMICs), but is also pertinent to high-income countries, where there is likewise a need to address micronutrient deficiencies in vulnerable groups such as iron deficiencies in adolescent girls and women of reproductive age, particularly pregnant women. During the nutrition data revolution, a “nutrition data value chain” was introduced, which has five critical processes – data prioritization, creation and collection, curation, analysis, interpretation/recommendation, and decision-making (4,5). To date, enormous effort has been made to overcome the early challenges in this chain: data prioritization, collection, curation, and lab analysis (6–8). The recently published Micronutrient Survey Manual & Toolkit provides comprehensive information on planning a national micronutrient survey (6); and the Hemoglobin Measurement Laboratory Protocol outlined the standard laboratory method for obtaining accurate results of hemoglobin in population surveys (7). Even though these guidelines provide recommendations on data analysis and presentation, little has been achieved in *streamlining the analysis* of micronutrient biomarker data. Analysis of micronutrient biomarkers still requires intermediate knowledge of statistics (e.g., analysis of surveys with complex design), high-level of proficiency in computer coding, and a deep understanding of the biological bases of micronutrient biomarkers (e.g., the mechanism of how inflammation influences micronutrient biomarkers) (9). Due to the complex nature of micronutrient data, organizations need to hire senior statisticians with a specialization in micronutrient data, who might not always be available. Alternatively, organizations need to invest in building this capacity in their analysts; however, the time lag may result in lost opportunities to make micronutrient biomarker data available during critical decision-making periods.

To improve the use of micronutrient data, we provide an easy-to-use and “all-in-one” tool, the Statistical Apparatus for Micronutrient Biomarker Analysis (SAMBA) package in R software, which can be used to analyze 12 biomarkers, including 9 micronutrient biomarkers: retinol binding protein, serum retinol, serum b-12, Red Blood Cell (RBC) folate, serum folate, serum zinc, serum ferritin, soluble transferrin receptor (STfR), and serum Vitamin D; 2 inflammation biomarkers: Alpha(1)-acid glycoprotein (AGP) and C-Reactive Protein (CRP); and hemoglobin. Three strengths of this tool are that it 1) standardizes and streamlines analysis of multiple biomarkers and datasets simultaneously, which reduces the time required for analysts to clean and analyze biomarker datasets; 2) allows for customization to analyze additional biomarkers beyond the 12 built-in micronutrient biomarkers; and 3) is built in R, a free and open-access software. The SAMBA package with example codes and example data is available online (Open Science Framework link: XXXX); the corresponding SAMBA package user manual is provided with this article (**supplemental user manual 1**). The detailed user manual and examples will allow analysts to modify the example code for their unique micronutrient biomarker data.

The objectives of this paper are to describe the current challenges in micronutrient biomarker analysis, display the general structure and features of the SAMBA package by explaining the underlying analysis algorithm, present example applications of the SAMBA package for micronutrient biomarker analysis using a single-site study in Kenya (10) and a nationally representative survey, the United States’ National Health And Nutrition Examination Survey (NHANES) 2003 – 2006 (11) and, discuss the strengths and limitations of the SAMBA package.

**Challenges in micronutrient biomarker analysis**

There are numerous challenges to analyzing global micronutrient biomarker data (**Fig. 1**):

* **Datasets stored in different file formats:** It iscommon that survey coordinators who implement surveys are not also data analysts; therefore, analysts might receive data files in a format that is not compatible to the analysis software that they use. Although certain packages were developed to convert between different types of data files (e.g., STATA, SAS, and excel files), analysts still need to learn the related packages to carry out this conversion.
* **Different variable names:** Each biomarker can be stored using different variables names. For example, the biomarker serum ferritin can be stored under the variable name, *serum\_ferritin*, *serum.ferritin*, *sf*, *ferritin*, or *FER*. This can cause confusion for analysts, especially with absence of a comprehensive codebook.
* **Different units:** Biomarkers can be stored in either conventional units or the International System of Units (SI) (12). Most often, analysts will need to use additional information such as molar mass of the biomarker to convert between conventional units and SI and then compare the biomarker with a cutoff established with consistent units.
* **Different methods to adjust for inflammation:** Inflammation is known to affect many micronutrient biomarkers, which can thus lead to an overestimate or underestimate of the prevalence of deficiencies in a population (9). Inflammation adjustment methods, such as Thurnham (13) or Biomarkers Reflecting Inflammation and Nutrition Determinants of Anemia (BRINDA) method (9), have been well-documented; however, analysts still need to learn and apply those methods, which requires additional learning time.
* **Different cutoffs for micronutrient deficiencies:** The diagnosis of micronutrient deficiencies is determined based on varying cutoffs of the biomarkers. Some cutoffs are based on manufactures’ recommendation, and some are based on international consensus. In addition, physiological or environmental factors can also influence the appropriate cutoffs. For example, the hemoglobin concentration cutoff to diagnose anemia can be affected by smoking status and altitude (14,15); and the serum zinc concentration cutoff used to determine zinc deficiency can be influenced by fasting status (16).
* **Different data analysis approaches based on the study design:** Analysts should have the knowledge for analyzing studies with different study design. For a multi-stage complex survey design, analysts should understand how strata, clusters, and survey weights play a role in estimating descriptive statistics (e.g., mean, geometric mean, prevalence of deficiencies).
* **Difficulty in implementing in-country analysis:** In-country analysis of micronutrient data is essential for capacity building in public health nutrition (17), which can facilitate timely reporting and thus lead to effective policy decision. However, such capacity building requires time, technical capacity, and financial commitment as well as multi-sectoral collaboration.
* **Inconsistent and contradictory results:** Inconsistent and contradictory results from analyzing the same study by different analysts continue to emerge (18) and, therefore, compromise the validity of the study and the ability to compare results across countries. Inaccessible analytical methods and code make it difficult to evaluate and diagnose flawed analyses.

**Introduction to the bio\_analysis R package**

The SAMBA R package is an easy-to-use and “all-in-one” tool that facilitates estimation of multiple micronutrient biomarker distributions and prevalence of micronutrient deficiencies for multiple datasets simultaneously. SAMBA also includes additional features, such as the ability to carry out checks on the input datasets and implementation of a variety of specific analyses, including adjusting for inflammation for certain biomarkers (9), while taking into consideration multi-stage complex survey design, if relevant. Prior to this paper, the associated codes and analytical details of those analyses have been published separately in various papers and reports, which requires analysts to learn them individually, and may thus limit the extent to which analysts can then apply these methods to their own data.

An overview of the SAMBA package inputs, core contents, and outputs is illustrated in **Figure 2.** Two input .csv files are required: 1) biomarker dataset template and 2) biomarker cutoff template. For the biomarker dataset template, analysts need to specify the directory for where the dataset will be stored on their computer, survey-related information (if relevant, e.g., strata, cluster, and survey weight variable names), demographic information (e.g., age and sex variable names), and micronutrient biomarker variable names and units. In this template, analysts can include information for multiple datasets (one column per dataset). For the biomarker cutoff template, analysts need to review the existing values of the biomarker cutoffs that vary by age and population groups (men, women, and pregnant women), and then they can make modifications on the cutoff template in case the cutoffs in the template differ from their desired values.

The SAMBA package has multiple steps that will run with simply five lines of codes: users just need to define the directories of the biomarker dataset template and cutoff template in their computer, the number of biomarker datasets users want to analyze, and the directory and name of the output datasets. SAMBA will first load the biomarker dataset template and cutoff template and preload the biomarker dataset, and then use the information to check for dataset specification errors (e.g., if a user-specified variable exists in the dataset) before starting analysis; if any detectable errors are found, SAMBA provides guidance for correcting them. If no errors exist, the biomarker datasets will be officially loaded. Currently, SAMBA supports SAS, SPSS, STATA, CSV, and EXCEL files. SAMBA will change the variable names listed in the biomarker dataset template into standardized variable names (predefined in the biomarker dataset template), as well as change the units of biomarkers into SI. Then, SAMBA will apply the BRINDA method to adjust the effect of inflammation on selected nutrients among preschool-age children 6 mos to 5 y (PSC) and non-pregnant women of reproductive 15 – 49 y (WRA) (9). In brief, the BRINDA method recommends adjustment for retinol binding protein and serum retinol using AGP and CRP among PSC (19), no adjustment for serum or RBC folate or serum B-12 for either PSC or WRA (20), adjustment for serum zinc using AGP and CRP among PSC (21), and adjustment for serum ferritin and sTfR using AGP and CRP among PSC and WRA (22). After adjustment, a cleaned dataset in csv format for each raw dataset analyzed will be exported, which contains standardized variable names and units and adjusted biomarker values for inflammation for all study participants (output 1). SAMBA will also generate a formatted csv file suitable for use in reports or manuscripts containing estimated results of the micronutrient biomarker distribution, such as the mean, geometric, median, IQR, and prevalence of deficiencies with associated standard errors and confidence intervals (output 2).

**Extension of the SAMBA R package**

The SAMBA package was built to be flexible to allow for the analysis of any biomarkers that can be categorized with a binary outcome. Without any modification of the R package, this tool permits analyzing additional biomarkers beyond the 12 default biomarkers built into this package, such as blood pressure and cholesterol. Analysts just need to add the variable name, units, and survey weight variable (if relevant) of the additional biomarkers (e.g., diastolic blood pressure) into the biomarker dataset template and add the cutoffs of the biomarkers (e.g., the cutoff of high blood pressure using diastolic blood pressure) into the cutoff template. A successful run of the SAMBA will then generate a cleaned dataset for all study participants for each raw dataset analyzed (output 1) that includes information on the additional biomarker(s) and a summary result file (output 2) that contains the distribution and prevalence of deficiency (or customized outcome) of this biomarker.

In addition, researchers can merge the outputs of the SAMBA package with other variables, such as underweight and Body Mass Index (BMI) to explore a wide range of research questions. For example, the cleaned dataset (output 1) can be merged with an individual’s weight, height, and other health outcomes to explore the association between micronutrient deficiencies and Non-Communicable Diseases (NCD). As micronutrient deficiencies are an important component of the double-burden malnutrition, the summary result file (output 2) of several nationally representative surveys can be merged with aggregated data on under-nutrition and obesity and NCDs to explore malnutrition in all forms globally (23–25).

**RESULTS**

We used the SAMBA package to analyze biomarker data for PSC using a single-site study in Kenya (10) and for WRA using NHANES 2003 - 2006 (11). We presented the prevalence of deficiencies, mean, geometric mean, and 25th, 50th, and 75th percentiles of retinol binding protein, serum ferritin, and STfR (and their BRINDA adjusted values), and CRP, and AGP concentrations for the Kenya study; and the same statistic parameters of serum retinol, serum ferritin, and sTfR (and their BRINDA adjusted values), serum B-12, RBC and serum folate, CRP, and hemoglobin concentrations for NHANES (**Table 1**). Because NHANES does not collect AGP routinely, and sTfR is adjusted by only AGP among WRA according to the BRINDA adjustment method (26), we were unable to apply the BRINDA adjustment method to sTfR among WRA using NHANES. Thus, the adjusted sTfR is the same as the unadjusted values among WRA. To validate the results generated by the SAMBA package, an independent consultant carried the same analyses, which yielded identical results by the SAMBA package (**Table 1**).

**Discussion**

The SAMBA package provides a streamlined and efficient structure for multiple micronutrient biomarker analysis for multiple datasets. Prior to the SAMBA package, analysts needed to understand the biological basis for micronutrient biomarkers, obtain intermediate statistical knowledge, and have familiarity with basic computer code. The new, freely available, efficient, and open-access tool simplifies estimations of micronutrient distribution and prevalence of deficiencies, which can drastically shorten the time from data cleaning to presentation of a final results table.

Micronutrient deficiencies are an important component of the burden of malnutrition for both LMICs and high-income countries (23), especially when micronutrient deficiencies are also prevalent in normal-weight and overweight population (27). However, micronutrient deficiencies are frequently overlooked when assessing all forms of malnutrition, due to both the scarcity and limited utilization of data (23). The GNR report and other initiatives have emphasized the need to fill in the data gap in a holistic approach by improving all components in the data values chains, from data prioritization, creation and collection, curation, analysis, to interpretation/recommendation (4,5). The SAMBA package is a software tool that can be applied to diverse global contexts to fill the gap in the nutrition data value chain between data collection and interpretation for decision-making. The SAMBA package reduces the time spent on 1) learning basic biological knowledge of micronutrient biomarkers (e.g., how inflammation influences biomarker values), 2) intermediate statistical skills (e.g., analysis of complex survey design), and 3) time to write codes to analyze each biomarker. Researchers and analysts can focus on the actual research questions instead of learning different pieces of micronutrient biomarker analysis from a variety of published papers and reports. The feature of analyzing multiple micronutrient datasets is especially useful for countries that continuously collect micronutrient data in their nutrition surveillance system and organizations that monitor the global micronutrient status. Furthermore, the SAMBA package is built in R, a free and open-access software, so that Institutions and analysts do not need to make additional investment in the software purchase, which can improve the accessibility of the SAMBA package. In addition, there is an increasing push to improve research capacity at institutions in LMICs. The SAMBA package can facilitate these efforts by providing a user-friendly platform for analysts to conduct micronutrient biomarker data analysis, as well as the ability to automatically output a spreadsheet containing the information typically requested for national and regional micronutrient biomarker surveys (e.g., mean, geometric mean, median, and prevalence of micronutrient deficiencies) to local governments and the international community, with substantial reductions in the time and cost required to do so. Moreover, extensions of the tool permit analyses of additional biomarkers beyond the default 12 biomarkers included in the package and connecting the outputs of the SAMBA package (either the summary result file or clean datasets) with other nutrition information (over- and under-nutrition) and health outcomes to explore association between micronutrient deficiencies and NCDs and malnutrition in all forms.

The SAMBA package also has several limitations. First, the SAMBA package can only generate unbiased results when the micronutrient biomarker data are of high quality and represent the population’s micronutrient biomarker distribution. It can neither correct for errors in data collection nor remove extreme or unreasonable values of micronutrient biomarkers. SAMBA will not remove any outliners, because for lots of surveys, such as NHANES, biomarker values are already cleaned and within a reasonable range. However, by default, the SAMBA package will recode zero values to 0.0001 so that log transformation of the biomarker values can be made possible for inflammation adjustment and calculation of the confidence interval of geometric means. To ensure the validity of micronutrient data analysis, besides rigorous, standardized training of laboratory technicians, we recommend analysts carefully clean data, such as changing the zero values of micronutrient biomarker measurements to the minimal detectable values based on manufacturers’ laboratory protocols and remove any outliners before applying the SAMBA package to their own data. Second, the current version of the SAMBA package cannot calculate percent of people with more than one nutrient deficiency, such as the prevalence of individuals with both vitamin A and iron deficiencies. To address this limitation, we recommend analysts use the cleaned dataset (output 1) from the SAMBA package to calculate the overlapping micronutrient deficiencies. Third, because the BRINDA adjustment method is only applicable to selected nutrients (i.e., retinol binding protein, serum retinol, serum ferritin, sTfR, RBC and serum folate, serum B-12, and serum zinc) among WRA and PSC, despite that SAMBA can be used to analyze all types of micronutrients among all population groups, the SAMBA package does not apply inflammation adjustment to nutrients that were not described above or outside of the WRA or PSC population groups. However, to apply inflammation adjustment to other population groups, users are recommended examining the relation between nutrients and inflammation markers (AGP or CRP) and applying the BRINDA adjustment algorithm if an obvious relation exists (9). Fourth, although users can define the cutoffs for micronutrient deficiencies based on age and population groups (man, women, and pregnant women) in the SAMBA package, users cannot define cutoffs based on other factors such as smoking or fasting status and altitude. Instead, users are recommended adjusting the effect of the smoking or fasting status or altitude on biomarker values and then apply the SAMBA package. Fifth, the SAMBA package has specific technical requirements. Analysts need to have R software installed and be equipped with basic R programming skills, which requires time and access to appropriate training materials. These user requirements may block the usage of the SAMBA package from a wider audience.

To facilitate use of the current SAMBA package version, we have designed training materials and a thorough user manual to enhance the usage of the SAMBA package and build the capacity of researchers and policy analysts (training materials are available upon request). Plans to expand the current SAMBA tool include the addition of new functionalities, such as analysis of overlapping micronutrient deficiencies and generating a codebook of datasets based on the information filled into the biomarker dataset template. In the longer term, we envision development of a web-based tool based on the SAMBA package to further increase the accessibility of the method and decrease the time and resources required to make micronutrient biomarker data results available.

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Table 1: Biomarkers estimated by the SAMBA function and an independent analyst in non-pregnant women 15-49 y using National Health and Nutrition Examination Survey 2003-2006 and in children 6 mos - 5 y using a single-site Kenya study.

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Biomarker | n | Deficiency or at risk, % | | mean | | Geometric mean | | 25th percentile | | 50th percentile | | 75th percentile | |
| SAMBA | Independent analyst | SAMBA | Independent analyst | SAMBA | Independent analyst | SAMBA | Independent analyst | SAMBA | Independent analyst | SAMBA | Independent analyst |
| Kenya (children 6 mos - 5 y) |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Retinol binding protein, μmol/L | 896 | 23.0±1.4 | 23.0±1.4 | 0.89±0.01 | 0.89±0.01 | 0.85±0.84 | 0.85±0.84 | 0.71±0.01 | 0.71±0.01 | 0.88±0.01 | 0.88±0.01 | 1.06±0.01 | 1.06±0.01 |
| Adj. retinol binding protein, μmol/L | 896 | 6.3±0.8 | 6.3±0.8 | 1.12±0.01 | 1.12±0.01 | 1.08±1.06 | 1.08±1.06 | 0.91±0.01 | 0.91±0.01 | 1.08±0.01 | 1.08±0.01 | 1.30±0.02 | 1.30±0.02 |
| Serum ferritin, μg/L | 896 | 38.7±1.6 | 38.7±1.6 | 29.91±1.48 | 29.91±1.48 | 16.13±15.01 | 16.13±15.01 | 7.48±0.39 | 7.48±0.39 | 15.58±0.75 | 15.58±0.75 | 31.56±1.64 | 31.56±1.64 |
| Adj. serum ferritin, μg/L | 896 | 72.4±1.5 | 72.4±1.5 | 10.05±0.35 | 10.05±0.35 | 6.57±6.17 | 6.57±6.17 | 3.53±0.15 | 3.53±0.15 | 6.74±0.26 | 6.74±0.26 | 12.97±0.46 | 12.97±0.46 |
| sTfR, mg/L | 896 | 35.5±1.6 | 35.5±1.6 | 8.59±0.19 | 8.59±0.19 | 7.34±7.07 | 7.34±7.07 | 5.65±0.10 | 5.65±0.10 | 7.09±0.10 | 7.09±0.10 | 9.49±0.22 | 9.49±0.22 |
| Adj. sTfR, mg/L | 896 | 27.2±1.5 | 27.2±1.5 | 7.81±0.17 | 7.81±0.17 | 6.69±6.45 | 6.69±6.45 | 5.10±0.10 | 5.10±0.10 | 6.59±0.08 | 6.59±0.08 | 8.64±0.21 | 8.64±0.21 |
| CRP, mg/L | 896 | 27.8±1.5 | 27.8±1.5 | 5.22±0.27 | 5.22±0.27 | 1.38±1.21 | 1.38±1.21 | 0.40±0.03 | 0.40±0.03 | 1.53±0.13 | 1.53±0.13 | 6.15±0.56 | 6.15±0.56 |
| AGP, g/L | 896 | 64.2±1.6 | 64.2±1.6 | 1.26±0.02 | 1.26±0.02 | 1.17±1.14 | 1.17±1.14 | 0.88±0.02 | 0.88±0.02 | 1.16±0.02 | 1.16±0.02 | 1.60±0.04 | 1.60±0.04 |
| NHANES (non-pregnant women 15-49 y) |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Serum retinol, nmol/L | 3145 | 0.3±0.1 | 0.3±0.1 | 1.84±0.01 | 1.84±0.01 | 1.76±1.74 | 1.74±1.79 | 1.46±0.01 | 1.46±0.01 | 1.75±0.01 | 1.75±0.01 | 2.13±0.02 | 2.13±0.02 |
| Adj. serum retinol, μmol/L | 3145 | 0.5±0.2 | 0.5±0.2 | 1.77±0.01 | 1.77±0.01 | 1.70±1.68 | 1.68±1.73 | 1.42±0.01 | 1.42±0.01 | 1.69±0.01 | 1.69±0.01 | 2.06±0.02 | 2.06±0.02 |
| Serum ferritin, μg/L | 3183 | 13.1±0.8 | 13.1±0.8 | 57.29±1.42 | 57.29±1.42 | 38.94±37.27 | 37.27±40.68 | 24.00±0.98 | 24.00±0.98 | 41.00±0.73 | 41.00±0.73 | 70.00±1.71 | 70.00±1.71 |
| Adj. serum ferritin, μg/L | 3183 | 20.7±1.0 | 20.7±1.0 | 41.47±0.98 | 41.47±0.98 | 28.75±27.60 | 27.60±29.96 | 17.51±0.54 | 17.51±0.54 | 30.34±0.68 | 30.34±0.68 | 51.55±0.91 | 51.55±0.91 |
| sTfR, mg/L | 3148 | 9.7±0.8 | 9.7±0.8 | 5.89±0.06 | 5.89±0.06 | 5.54±5.44 | 5.44±5.64 | 4.48±0.04 | 4.48±0.04 | 5.28±0.04 | 5.28±0.04 | 6.56±0.12 | 6.56±0.12 |
| Adj. sTfR, mg/L2 | 3148 | 9.7±0.8 | 9.7±0.8 | 5.89±0.06 | 5.89±0.06 | 5.54±5.44 | 5.44±5.64 | 4.48±0.04 | 4.48±0.04 | 5.28±0.04 | 5.28±0.04 | 6.56±0.12 | 6.56±0.12 |
| Serum B-12, pmol/L | 3166 | 2.8±0.5 | 2.8±0.5 | 383.95±9.61 | 383.95±9.61 | 335.65±325.19 | 325.19±346.45 | 253.13±3.84 | 253.13±3.84 | 328.41±5.24 | 328.41±5.24 | 436.84±6.71 | 436.84±6.71 |
| RBC folate, nmol/L | 3210 | 7.6±0.5 | 7.6±0.5 | 603.60±8.33 | 603.60±8.33 | 565.37±551.43 | 551.43±579.66 | 448.50±6.12 | 448.50±6.12 | 564.00±7.22 | 564.00±7.22 | 707.57±10.60 | 707.57±10.60 |
| Serum folate, nmol/L | 3186 | 2.8±0.4 | 2.8±0.4 | 28.48±0.63 | 28.48±0.63 | 24.87±24.17 | 24.17±25.60 | 18.60±0.27 | 18.60±0.27 | 25.10±0.32 | 25.10±0.32 | 32.80±0.49 | 32.80±0.49 |
| CRP, mg/L | 3197 | 25.6±1.0 | 25.6±1.0 | 4.39±0.18 | 4.39±0.18 | 1.74±1.61 | 1.61±1.89 | 0.60±0.05 | 0.60±0.05 | 1.90±0.10 | 1.90±0.10 | 5.15±0.22 | 5.15±0.22 |
| Hemoglobin, g/L | 3226 | 6.4±0.5 | 6.4±0.5 | 136.10±0.50 | 136.10±0.50 | 135.60±134.63 | 134.63±136.57 | 130.00±0.49 | 130.00±0.49 | 137.00±0.49 | 137.00±0.49 | 143.00±0.51 | 143.00±0.51 |

1 Values are in mean±SE; Adj. Adjusted by the BRINDA method; AGP, Alpha(1)-acid glycoprotein; BRINDA, Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia; CRP, C-reactive protein; NHANES, National Health and Nutrition Examination Survey; RBC, Red Blood Cell, SAMBA, Statistical Apparatus of Micronutrient Biomarker Analysis, sTfR, Soluble Transferrin Receptor. Vitamin A deficiency defined as serum retinol or retinol binding protein < 0.7 μmol/L; B-12 deficiency defined as serum B-12 < 150 pmol/L; folate deficiency defined as RBC folate < 340 nmol/L or serum folate < 10 nmol/L; iron deficiency defined as serum ferritin < 12 μg/L and <15 μg/L for children and non-pregnant women 15-49 y, respectively, or sTfR < 8.3 mg/L for both groups; anemia defined as hemoglobin < 110 g/L and <120 for children and non-pregnant women 15-49 y, respectively; inflammation defined as CRP >5 mg/L and AGP >1 g/L.

2sTfR is adjusted by only AGP. Because AGP is unavailable in NHANES, the results of adjusted sTfR are the same as the unadjusted results.

Figure 1: Challenges in micronutrient biomarker analysis

**Diagram

Description automatically generated**

Figure 2: Overview of the SAMBA package inputs, core contents, and outputs

**Diagram

Description automatically generated**

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1. AGP, Alpha(1)-acid glycoprotein; BRINDA, Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia; CRP, C-reactive protein*;* GNR, Global Nutrition Report; IS, International System of Units; NCD, Non-Communicable Diseases; NHANES, National Health and Nutrition Examination Survey; PSC, Preschool-age children; RBC, Red Blood Cell; SAMBA, Statistical Apparatus of Micronutrient Biomarker Analysis; sTfR, soluble transferrin receptor; WRA, women of reproductive age. [↑](#footnote-ref-1)
2. This analysis was supported by award 7200AA18C00070 [↑](#footnote-ref-2)
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