

## **Monitoring of vaccine targets and interventions using global genome data: vaccines.watch**

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on behalf of the NIHR Global Health Research Unit on Genomics and enabling data for  
Surveillance of AMR

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**Keywords:** genomics, vaccines, vaccine development, epidemiology, surveillance, monitoring

46 **Abstract**

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48 The expansion of pathogen genome sequencing into routine disease surveillance  
49 programmes is set to bring rapidly-growing volumes of increasingly structured data on a global  
50 scale. This has the potential to deliver exciting opportunities for accelerating vaccine  
51 development and monitoring. Here we present an interactive platform, vaccines.watch  
52 (<https://vaccines.watch>), which aims to support decision-making around vaccine formulations  
53 and roll-out by enabling interrogation of vaccine target diversity from global genome data. We  
54 have initially focused on targets included in existing or prospective multivalent polysaccharide-  
55 based vaccines for *Streptococcus pneumoniae*, *Klebsiella pneumoniae* (and related species)  
56 and *Acinetobacter baumannii*. The platform currently displays data for >100k high-quality  
57 genomes with geotemporal sampling information (post-2010), with new genomes assembled,  
58 analysed and incorporated on an ongoing basis (every 4 hours) as public data are newly  
59 deposited. Crucially, users can view vaccine target information in the broader context of  
60 genotypic variants (e.g. sequence types) and antimicrobial resistance markers. The platform  
61 also enables users to review the composite serotypes of pneumococcal vaccine formulations  
62 among the available genomes. For example, using data in vaccines.watch from 3 June 2025,  
63 we observed that serotypes included in the PCV13 and PCV21 formulations accounted for  
64 36.2% (11,907/32,918) and 87.4% (28,764/32,918) of global public genomes, respectively.  
65 The platform also enables continuous review of the global genomic landscape of the included  
66 pathogens, enabling identification of gaps (e.g. in geographic coverage) that should be  
67 targeted with increased genomic surveillance. Indeed we demonstrate that substantial  
68 geographic gaps remain in the coverage of available genomes, with over half of countries  
69 contributing no genomes for each of the three pathogens. However, while caution in  
70 interpretation is important, as global representativeness of genome data grows,  
71 vaccines.watch is positioned to support different stages of the vaccine pipeline, from selection  
72 of target antigens to post-rollout monitoring of population changes.

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## 90 Introduction

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92 Vaccines have been a highly effective intervention for decreasing the burden of infectious  
93 diseases. They also have the potential to play an important role in reducing antimicrobial  
94 resistance (AMR), both by preventing infections caused by antimicrobial-resistant (and -  
95 susceptible) strains and by decreasing antimicrobial usage [1]. This has been exemplified by  
96 the recently-introduced typhoid conjugate vaccine (TCV) in Pakistan, which has been shown  
97 to be over 90% effective in protecting young children against extensively drug-resistant (XDR)  
98 typhoid infections [2]. A major advantage of vaccines is that they typically have a sustainable  
99 impact in reducing disease burden, with resistance to vaccines evolving far less readily than  
100 antimicrobial resistance [3]. Yet despite the success of vaccines, we still do not have any  
101 available vaccines for many major pathogens, while others necessitate improved or updated  
102 formulations [4]. Recently, the World Health Organisation (WHO) identified 17 endemic  
103 pathogens, including several bacterial species with a high AMR burden, for which global efforts  
104 in vaccine research and development should be prioritised [5].

105  
106 Genomic information has been used in vaccine development since the availability of the first  
107 bacterial genomes, in particular by enabling high-throughput *in silico* screening of all protein-  
108 encoding genes to identify potential target antigens. This approach, known as reverse  
109 vaccinology [6], has been used in the development of highly effective vaccines against  
110 *Neisseria meningitidis* serogroup B (MenB) [7, 8] as well as in antigen selection for multiple  
111 other vaccines since [9, 10]. An increasing array of bioinformatic tools have also been  
112 developed to further aid antigen selection, which make use of the growing biological databases  
113 (including genomic, transcriptomic and proteomic data) and increasingly sophisticated  
114 analytical and machine-learning methods [11]. In particular, pan-genome methods are  
115 typically used to identify genes that are common (core) to all strains of the target pathogen,  
116 as well as tools that predict protein characteristics (e.g. immunogenicity, subcellular  
117 localisation), epitopes, and the potential toxicity and allergenicity of candidate antigens.

118  
119 Over the coming years, the expansion of pathogen genome sequencing into routine disease  
120 surveillance programmes is set to bring rapidly-growing volumes of increasingly structured  
121 data on a global scale. This ongoing shift towards widespread genomic capacity, supported  
122 by major international public health agencies including the WHO [12], will provide increasing  
123 opportunities to accelerate vaccine development using genomic data. In particular, detailed  
124 genomic insights into contemporary pathogen population dynamics gained from surveillance  
125 data will enable us to better optimise vaccine formulations (including those targeted at specific  
126 populations or regions), improve our understanding of the impact of vaccine rollout on  
127 pathogen diversity, and more precisely monitor and deduce mechanisms of vaccine escape.

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129 However, technical barriers to fully capitalising on the available and upcoming opportunities  
130 remain, both around accessing the growing volumes of genomic data from the public  
131 sequence archives, and processing and translating the data into relevant insights across  
132 different pathogens for relevant stakeholders. We recently described our amr.watch platform  
133 (<https://amr.watch>) that analyses and visualises AMR trends from global public genome data  
134 [13]. For each of the priority bacterial pathogens defined by the WHO [14], amr.watch  
135 incorporates and displays data on an ongoing (“always-on”) basis using relevant analytics,  
136 with the aim of supporting both research and policy. The platform also enables continuous

137 review of the global landscape of pathogen genome sequencing, revealing opportunities  
138 whereby genomic data may already be able to reliably inform public health interventions as  
139 well as identifying gaps (e.g. in geographic coverage) that can be targeted with increased  
140 surveillance efforts.

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142 Using an extension of the “always-on” genome retrieval and analytical pipeline from  
143 amr.watch, here we present a sister platform, vaccines.watch (<https://vaccines.watch>), an  
144 interactive web application for assessing the prevalence and distribution of key vaccine targets  
145 from global genome data. We initially focus on providing support for the development and  
146 monitoring of multivalent vaccines with polysaccharide targets, the high diversity of which can  
147 necessitate choices over the particular polysaccharide types to include within a vaccine.  
148 Currently, vaccines.watch displays data on *Streptococcus pneumoniae* capsular-based  
149 serotypes, which form the targets of all licensed pneumococcal vaccines [15]. It also provides  
150 data on the capsule (K) and lipopolysaccharide (LPS) O-antigen (O) types from *Klebsiella*  
151 *pneumoniae* (and related species), as well as the capsule (K) and lipooligosaccharide (LOS)  
152 outer core (OC) antigen types from *Acinetobacter baumannii*. These are targets for novel  
153 vaccines, monoclonal antibody and phage therapies in both pathogens [16-19]. The  
154 vaccines.watch platform aims to provide scientists and vaccine developers with additional  
155 insights into the pathogen population dynamics around key vaccine targets, thereby  
156 supporting decision-making around vaccine formulations and roll-out.

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## 160 **Methods**

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162 **Overview of the vaccines.watch platform**  
163 Vaccines.watch currently reports genome data from *S. pneumoniae*, *A. baumannii* and the *K.*  
164 *pneumoniae* species complex (SC). The latter includes five related species (see *Input data*).  
165 The platform uses data from genomes that are retrieved from the International Nucleotide  
166 Sequence Database Collaboration (INSDC) databases, curated and processed by  
167 Pathogenwatch (<https://pathogen.watch>) using a previously described workflow [13].

168  
169 **Input data**  
170 The platform accepts data from paired-end Illumina genomes with a taxonomy ID recorded as  
171 1313 (*S. pneumoniae*), 573 (*K. pneumoniae*), 1463165 (*K. quasipneumoniae*), 244366 (*K.*  
172 *variicola*), 2026240 (*K. quasivariicola*), 2489010 (*K. africana*) or 470 (*A. baumannii*) in the  
173 European Nucleotide Archive (ENA) metadata. We also require genomes to have an available  
174 sampling date from 2010 onwards, a sampling location that is decodeable to at least the  
175 country level, and a minimum of 20x coverage. We check for new genomes using the ENA  
176 Portal API every four hours and download genomes meeting the above criteria from the  
177 Sequence Read Archive (SRA) using the SRA-Toolkit fastq-dump v3.1.0  
178 (<https://hpc.nih.gov/apps/sratoolkit.html>). The workflow has been described in detail  
179 previously [13], with specific filtering steps for the pathogens included here also provided at  
180 <https://vaccines.watch/summary>.

181  
182 Sequence reads are assembled with a workflow  
183 (<https://gitlab.com/cgps/ghru/pipelines/assembly>) that uses the SPAdes assembler v3.15.3

184 [20]. We then assess the quality of the resulting assemblies with QUAST v5.0.2 [21] and verify  
185 the species using the Speciator tool (v4.0.0) (<https://cgps.gitbook.io/pathogenwatch/technical-descriptions/species-assignment/speciator>) within Pathogenwatch. For the *K. pneumoniae*  
186 SC, we accept genomes identified as any of the five species listed above, allowing for  
187 inconsistencies with the metadata due to known difficulties with phenotypic identification  
188 methods. Genomes that fail to meet quality criteria, as outlined for each pathogen in  
189 **Supplementary Table 1**, are excluded.  
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## 192 **Variant typing**

193 The genotypic variants of the different pathogens are identified from the genome assemblies  
194 using community-based schemes implemented in Pathogenwatch. This variant typing  
195 comprises multi-locus sequence typing (MLST) for each of *S. pneumoniae* (PubMLST  
196 scheme), *K. pneumoniae* SC (BIGSdb-Pasteur scheme) and *A. baumannii* (Pasteur scheme)  
197 [22]. In addition to MLST, we also use Global Pneumococcal Sequencing Cluster (GPSC)  
198 assignments for *S. pneumoniae* [23] and “clonal group” assignments from the LIN code  
199 nomenclature for *K. pneumoniae* SC [24].  
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## 202 **Vaccine target typing**

203 Kaptive v3.1.0 [25], implemented in Pathogenwatch, is used to identify the K and O loci (and  
204 predicted K and O types) from the *K. pneumoniae* SC genomes and the K and OC loci (and  
205 predicted K and OC types) from *A. baumannii* genomes (using the database described by [26]  
206 for the latter). For both pathogens, we only incorporate data into vaccines.watch from  
207 genomes where Kaptive has indicated that both the K and O/OC loci are “typeable”. SeroBA  
208 v2.0 [27], also implemented in Pathogenwatch, is used to identify the capsular polysaccharide  
209 (cps) loci and predict the resulting serotype from *S. pneumoniae* genomes. The  
210 Pathogenwatch implementation of SeroBA differs from the published method by using  
211 simulated reads generated from assemblies rather than the raw sequence reads, although  
212 inconsistencies between the methods occur rarely (0.14%) (see  
213 <https://cgps.gitbook.io/pathogenwatch/technical-descriptions/typing-methods/seroba>).  
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## 216 **Identification of mechanisms associated with AMR**

217 For each pathogen, we identify genes and mutations associated with AMR for each of the  
218 antimicrobial classes reported in the WHO priority pathogens list [14]. These are identified  
219 using AMRFinderPlus v3.10.23, database version 2021-12-21.1 [28] with a curated database  
220 (**Supplementary Table 2**), as described previously [13].  
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## 223 **vaccines.watch web application**

224 The development of vaccines.watch follows a similar framework to amr.watch, built using  
225 Next.js (<https://nextjs.org/>) and the React library (<https://reactjs.org/>). Geographic data are  
226 represented using Mapbox (<https://www.mapbox.com>) while the Apache ECharts library  
227 (<https://echarts.apache.org/>) is used for visualisation of data in charts (e.g. barplots). The  
228 vaccines.watch website incorporates the processed data from Pathogenwatch on an ongoing  
229 basis following successful implementation of the analytical steps described above. The *S. pneumoniae*  
230 vaccine formulations (up to 2025) available for selection within the vaccines.watch interface were obtained from a recent review [15]. All genomes and associated  
231 metadata visualised in vaccines.watch are also available within Pathogenwatch for further use  
by the community.  
232

232 **Results**

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235 **Monitoring vaccine target diversity from global genomics data - the vaccines.watch  
236 platform**

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238 We have developed vaccines.watch (<https://vaccines.watch>), an interactive platform that  
239 enables monitoring of vaccine target antigens from pathogen genome data. We have initially  
240 focused on targets included in existing or prospective multivalent polysaccharide-based  
241 vaccines. Currently, these include capsular polysaccharide targets of *S. pneumoniae*, which  
242 form the basis of the licensed 23-valent pneumococcal polysaccharide vaccine (PPSV) and  
243 the different pneumococcal conjugate vaccine (PCV) formulations. We have also included the  
244 capsular and LPS O antigens of *K. pneumoniae* SC and the capsular and LOS OC antigens  
245 of *A. baumannii*, which are candidates for inclusion in novel vaccines and therapeutics in both  
246 pathogens.

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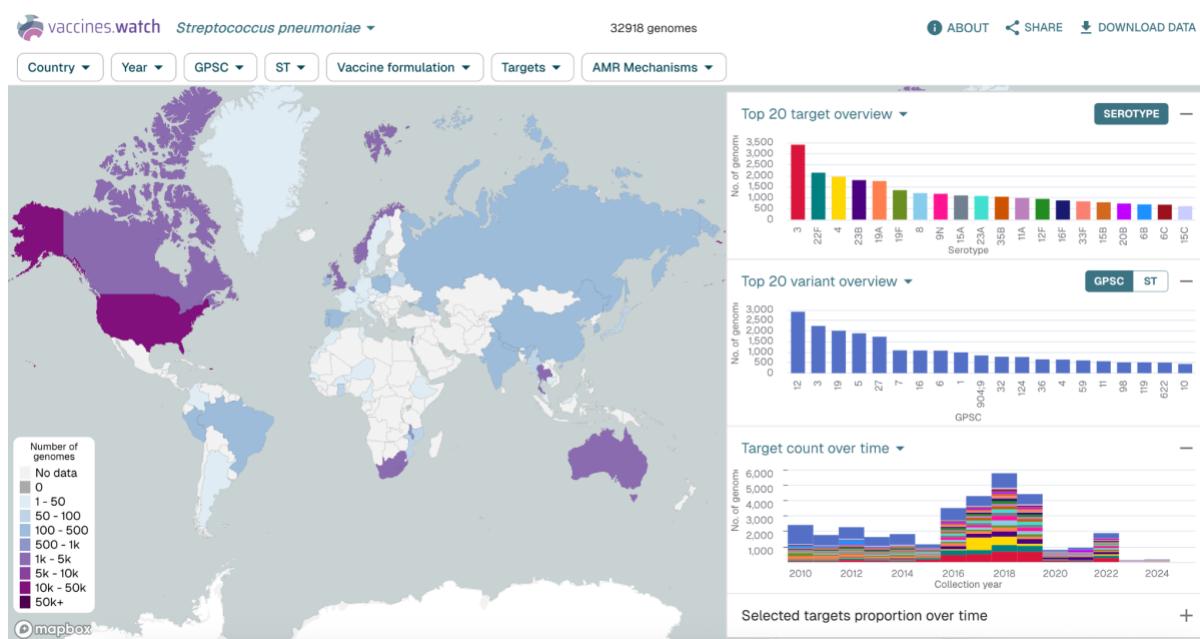
248 As input, the vaccines.watch platform currently incorporates processed genome data from all  
249 high-quality short-read Illumina genomes in the INSDC databases from the relevant pathogens  
250 that have associated geotemporal sampling information and were sampled post-2010 (see  
251 *Methods*). Available public genomes are assembled and processed via Pathogenwatch on an  
252 ongoing basis (every 4h), with the resulting analytics displayed in vaccines.watch in real-time.  
253 This data includes phenotypic predictions of the target types generated from identification of  
254 the relevant genomic loci via the SeroBA [27] and Kaptive [25] tools. Additional information  
255 inferred from the genome assemblies, comprising the variant types (e.g. STs) and AMR  
256 mechanisms, are also incorporated. We provide a live overview of all genomes represented  
257 in vaccines.watch at <https://vaccines.watch/all>, while the filtering processes applied to the  
258 public data can be viewed at <https://vaccines.watch/summary>.

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260 For each pathogen, vaccines.watch displays an interactive visualisation that enables rapid  
261 assessment and exploration of the diversity of vaccine target types and their geotemporal  
262 distribution (**Figure 1**). Vaccine target types are also placed into a broader population context  
263 by enabling assessment of their relationships with associated variant types (e.g. STs) and  
264 AMR markers. Users can survey the most frequent vaccine target types either globally,  
265 regionally or nationally, as well as assess specific target types of interest. In the case of *S.*  
266 *pneumoniae*, users can also select and explore specific sets of serotypes that are included in  
267 already-licensed pneumococcal vaccine formulations and vaccines under development.  
268 Visualisations with selected filters can be saved and/or shared onwards by users via the  
269 generation of URLs. All raw data shown in vaccines.watch can also be downloaded by users  
270 in CSV format.

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275 **Figure 1.** The interactive vaccines.watch platform here shows data for 32,918 *Streptococcus*  
276 *pneumoniae* genomes available as of 3 June 2025. The map shows the number of genomes  
277 sampled in each country. The right-hand figures show the twenty most frequent serotypes  
278 (top), the twenty most frequent variant types (global pneumococcal sequence clusters  
279 (GPSCs)) (middle) and the distribution of serotypes by sampling year (bottom). A similar  
280 visualisation, updated in real-time, can be found at: <https://vaccines.watch/organism/1313>  
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### 283 Contemporary global landscape of *S. pneumoniae*, *K. pneumoniae* SC and *A.* 284 *baumannii* genome sequencing

285 We recently reviewed the available public genome data displayed in our amr.watch platform  
286 for the WHO priority pathogens, which include the three pathogens included within  
287 vaccines.watch (except for the non-*K. pneumoniae* species in the *K. pneumoniae* SC) [13].  
288 Here we provide an updated review for the pathogens included within vaccines.watch with  
289 additional detail. As of 3 June 2025, the vaccines.watch platform showed data for 32,918 *S.*  
290 *pneumoniae*, 49,035 *K. pneumoniae* SC, and 18,428 *A. baumannii* genomes from the public  
291 sequence archives (**Supplementary Table 3**). 94.1% (46,156/49,035) of the *K. pneumoniae*  
292 SC genomes belonged to the *K. pneumoniae* species, with a further 3.0% (1490/49,035) from  
293 *K. variicola*, 2.7% (1342/49,035) from *K. quasipneumoniae*, 0.07% (33/49,035) from *K.*  
294 *quasivariicola* and 0.03% (14/49,035) from *K. africana*. Notably, a large number of public  
295 genomes were excluded from the platform due to the absence of associated geographic  
296 and/or temporal metadata, including 110,361 *S. pneumoniae*, 39,031 *K. pneumoniae* SC and  
297 12,154 *A. baumannii* (see <https://vaccines.watch/summary> for an updated summary). Among  
298 genomes that passed QC criteria and met metadata requirements, we also excluded a further  
299 1576 belonging to the *K. pneumoniae* SC and 283 belonging to *A. baumannii* where the K  
300 and/or O/OC loci were identified as “untypeable” by Kaptive, ensuring the use of high-confident  
301 matches only.  
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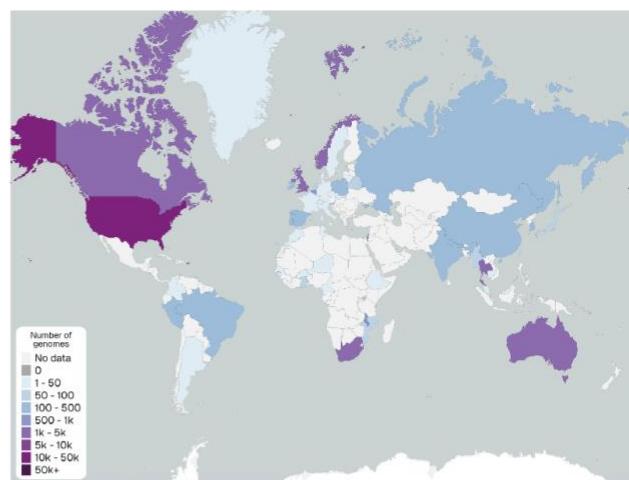
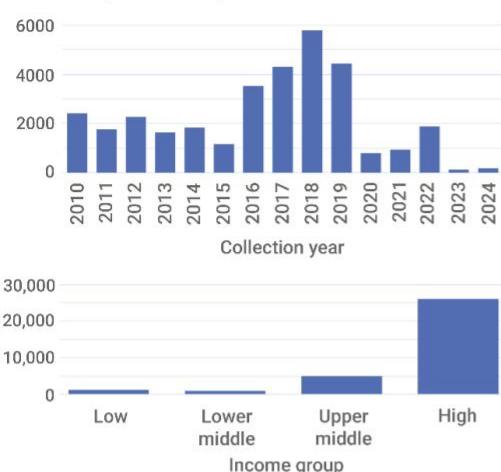
304 For each of the three pathogens, we found that the number of genomes sampled from different  
305 geographic regions and individual countries was highly variable (**Figure 2**). Overall 79.2%  
306 (79,515/100,381) of all genomes were from high-income countries with 14.2%  
307 (14,217/100,381) from upper-middle, 5.1% (5154/100,381) from lower-middle and 1.5%  
308 (1470/100,381) from low-income countries. Almost half (47.1%; 15,519/32,918) of the *S.*  
309 *pneumoniae* genomes were from the USA, with 79.7% (26,250/32,918) of the total genomes  
310 from isolates sampled in seven countries (USA, South Africa, UK, Norway, Australia, Canada  
311 and Thailand). Approximately a third (32.0%; 15,681/49,035) of the *K. pneumoniae* SC  
312 genomes were from the USA with ten countries (USA, UK, China, Norway, Australia, Spain,  
313 Japan, Thailand, Italy and Germany) contributing 71.2% (34,899/49,035) of the total. *A.*  
314 *baumannii* genomes were the most unevenly distributed with approximately two-thirds from  
315 the USA (66.8%; 12,316/18,428) while China contributed an additional 10.3% (1902/18,428).  
316 Notably, we also identified substantial geographic gaps. In particular, over half of countries  
317 (55.8%; 139 of 249 with officially-assigned ISO-3166-1 codes) contributed no genome data  
318 for *K. pneumoniae* SC, which rose to 65.9% (164/249) for *A. baumannii* and 77.9% (194/249)  
319 for *S. pneumoniae*.  
320

321 The temporal distribution of genomes differed across the three pathogens (Figure 2). In  
322 particular, we found the majority of genomes from both *S. pneumoniae* (88.1%;  
323 29,010/32,918) and *K. pneumoniae* SC (75.5%; 36,999/49,035) were sampled prior to 2020,  
324 with a decline observed since 2018. However, the number of *A. baumannii* genomes continued  
325 to increase until 2023, with the majority (59.0%; 10,865/18,428) sampled from 2020 onwards.  
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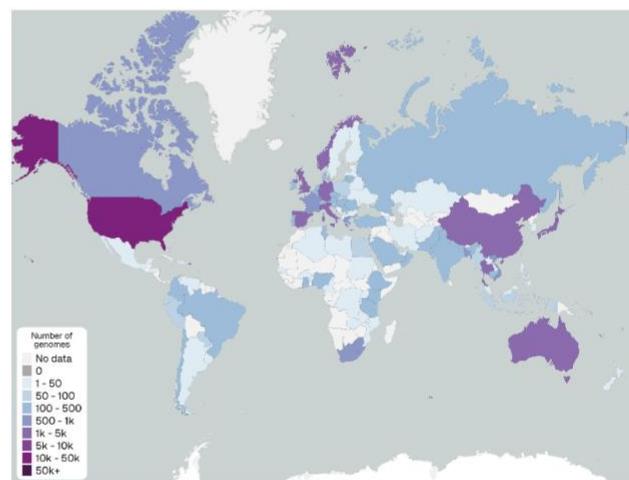
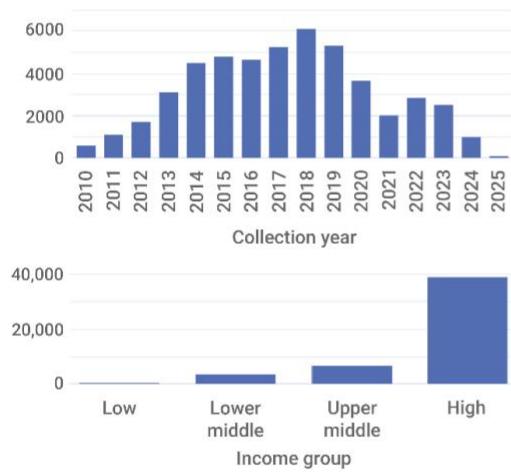
327 As detailed previously [13], we also found that high proportions of the public genomes carried  
328 one or more genes and/or mutations associated with AMR, albeit with the proportions differing  
329 considerably between countries. Overall, 32.7% (10,755/32,918) of *S. pneumoniae* genomes  
330 carried one or more AMR mechanisms associated with beta-lactam resistance. 74.3%  
331 (36,436/49,035) and 57.2% (28,033/49,035) of the *K. pneumoniae* SC genomes carried  
332 mechanisms associated with third-generation cephalosporin and carbapenem resistance,  
333 respectively. 83.5% (15,385/18,428) of the *A. baumannii* genomes carried mechanisms  
334 associated with carbapenems, which rose to >90% for some individual countries including  
335 Brazil (97.9%; 93/95), Vietnam (95.7%; 111/116) and Poland (95.3%; 143/150).  
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337 Altogether these findings continue to show the strong biases that exist among available public  
338 genomes, reflecting the varying availability of genome sequencing worldwide to date and its  
339 use within specific research agendas that often have a major focus on AMR.

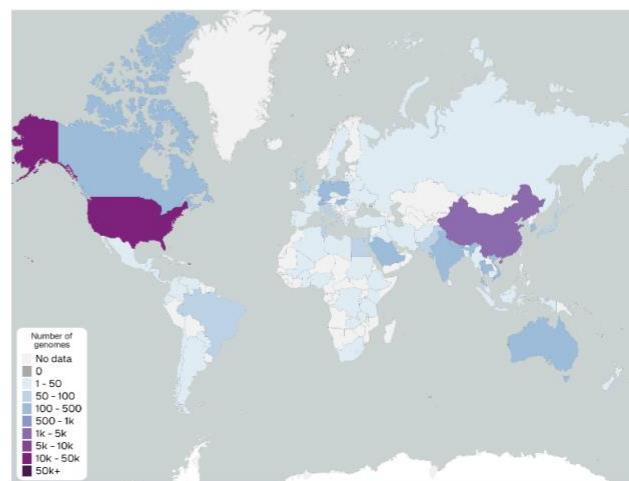
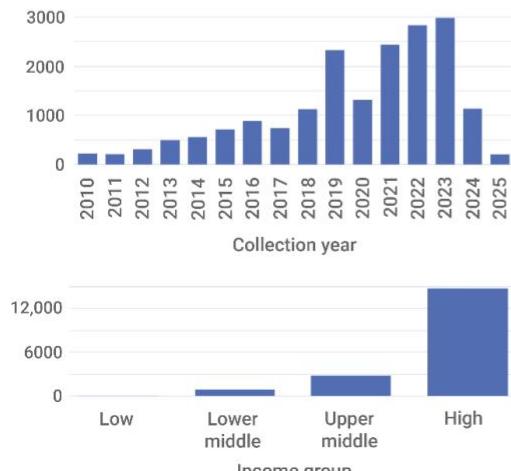
A *Streptococcus pneumoniae*



B *Klebsiella pneumoniae* species complex



C *Acinetobacter baumannii*



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342 **Figure 2.** Distribution of genomes represented in vaccines.watch as of 3 June 2025. Panels  
343 show the number of genomes by collection year, by country income group and by country  
344 (map) for each of the three pathogens, *Streptococcus pneumoniae* (A), *Klebsiella pneumoniae*  
345 species complex (B) and *Acinetobacter baumannii* (C). A live overview of all genomes  
346 represented in vaccines.watch is available at <https://vaccines.watch/all>

347 **Assessing trends in vaccine target diversity from global genome data**

348

349 We next reviewed the vaccine target diversity observed among the global public genomes in  
350 vaccines.watch (available as of 3 June 2025), albeit with awareness of the limitations relating  
351 to the representativeness of currently-available data. Using examples from across the three  
352 different pathogens, we illustrate how vaccines.watch can be used to interrogate trends among  
353 the vaccine targets in the context of available vaccines, their geotemporal dynamics and the  
354 broader population diversity.

355

356

357 *Streptococcus pneumoniae*

358

359 The vaccines.watch platform displays data on the predicted serotypes of *S. pneumoniae*  
360 based on identification of the corresponding *cps* loci by SeroBA. Among the 32,918 *S.  
361 pneumoniae* genomes, we found a total of 89 different serotypes from a total of 102 that can  
362 currently be identified by SeroBA. The top twenty most frequent serotypes accounted for  
363 76.2% (25,075/32,918) of the total genomes while many serotypes were rare (e.g. 38  
364 serotypes accounted for <20 genomes each). Only 0.1% (42/32,918) of genomes were  
365 untypeable by SeroBA, which can reflect either true absence of an intact *cps* locus or poor-  
366 quality data. A further 0.7% (219/32,918) of genomes had matches to “null capsule clade”  
367 (NCC) (i.e. non-encapsulated) variants.

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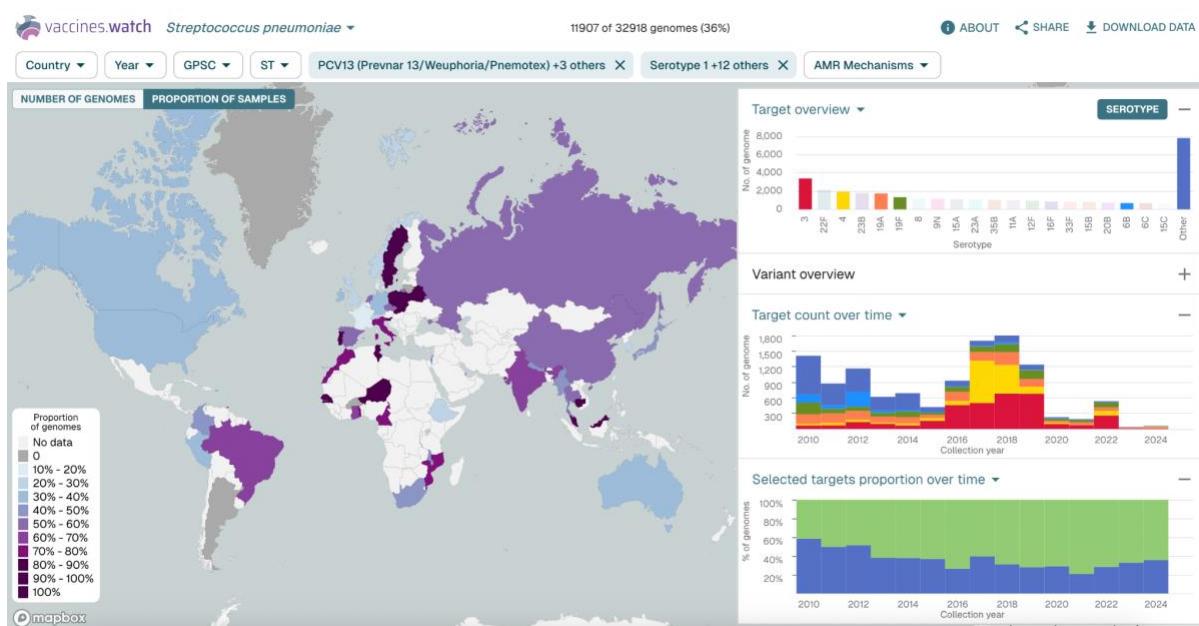
369 As described above, vaccines.watch enables users to select specific pneumococcal vaccine  
370 formulations and review their composite serotypes among all (or selected) genomes. For  
371 example, we observed that serotypes included in the PCV13 formulation accounted for 36.2%  
372 (11,907/32,918) of the global public genomes. Among the highest-sampled countries, this  
373 proportion varied between 20.3% and 55.7% (USA (30.3%), South Africa (49.3%), UK  
374 (20.3%), Norway (28.3%), Australia (36.0%), Canada (35.7%), Thailand (55.7%)) (**Figure 3**).  
375 Serotypes included in the broadest vaccine to date, PCV31 (VAX-31), currently in clinical trials  
376 [29], accounted for 87.4% (28,764/32,918) of global genomes.

377

378 Despite its inclusion within PPSV23 and all PCV formulations since PCV13, serotype 3 was  
379 the most frequent serotype among all genomes, accounting for 10.3% (3400/32,918) of the  
380 total (Figure 3). While this may partially reflect the variable global coverage of pneumococcal  
381 vaccines, studies have also shown that this serotype has continued to be a major cause of  
382 invasive pneumococcal disease even within countries that have achieved high coverage of  
383 PCV13 such as Spain [30], with some evidence of low vaccine effectiveness against this  
384 serotype [31].

385

386 Vaccines.watch can also be used to identify serotypes that are prevalent among public  
387 genomes but not included in vaccine formulations (Figure 3). For example, we observed that  
388 23B, which is not included in PPSV23 or early PCV formulations (e.g. PCV13), was the fourth  
389 most prevalent serotype among the public genomes (accounting for 5.5% (1795/32,918)). This  
390 serotype has been shown to be associated with increasing prevalence and penicillin non-  
391 susceptibility among both carriage and invasive isolates in recent years [32].



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394 **Figure 3.** Assessment of serotypes targeted by the PCV13 pneumococcal vaccine formulation  
395 in the context of the global *S. pneumoniae* genomes included in vaccines.watch (as of 3 June  
396 2025). The map shows the proportion of genomes in each country that belong to one of the  
397 13 serotypes targeted by PCV13. The right-hand panels show the twenty most frequent *S.*  
398 *pneumoniae* serotypes with PCV13 serotypes highlighted (top), the distribution of PCV13  
399 serotypes by sampling year (middle) and the proportion of the total genomes that belong (blue)  
400 or do not belong (green) to one of the PCV13 serotypes by sampling year (bottom). A similar  
401 visualisation can be found at: <https://cgps.dev/qM98oI6>

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#### 404 *Klebsiella pneumoniae* species complex

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406 The vaccines.watch platform displays data from Kaptive on both the best-matching K/O locus  
407 types (genotypes) identified from the *K. pneumoniae* SC genomes and the predicted K/O types  
408 (phenotypes), using the new O serotyping nomenclature proposed recently [33]. We have  
409 included both genotypic and predicted phenotypic types to allow exploration of the  
410 corresponding relationships. In the case of the O loci/types, these do not conform to one-to-  
411 one relationships due to the use of both the O locus and additional genes from outside of the  
412 O locus in the phenotype predictions.

413

414 Among the 49,035 *K. pneumoniae* SC genomes, we identified 139 different K loci from the  
415 186 that are currently defined by Kaptive. 3.2% (1592/49,035) of the total genomes were  
416 predicted to encode an acapsular phenotype (i.e. "capsule null"), based on identification of a  
417 K locus with truncations in one or more essential capsular synthesis genes. The top twenty  
418 most frequent K loci (without essential gene truncations) accounted for 64.7% (31,743/49,035)  
419 of the total genomes. Many of the K loci were found rarely (e.g. 20 accounted for <20 genomes  
420 each). Among genomes with carbapenem resistance markers, the top twenty K loci (without  
421 essential gene truncations) accounted for 79.3% (22,242/28,033). Notably, we also found that  
422 35.0% (17,139/49,035) of all genomes carried a K locus that corresponds to an unknown K  
423 type. These K loci include some of the most frequent in the collection, such as KL107, KL102

424 and KL106, which accounted for the second, third and fifth highest number of genomes,  
425 respectively.

426

427 The diversity of O locus types found among the public genomes was lower, with a total of 12  
428 identified. These correspond to 22 predicted O types (serotypes), the full repertoire currently  
429 defined by Kaptive. Five O types (O2 $\beta$ ; O1 $\alpha\beta,2\alpha$ ; O2 $\alpha$ ; O1 $\alpha\beta,2\beta$ ; O3 $\gamma$ ) accounted for 80.3%  
430 (39,351/49,035) of the genomes. The two O locus types, OL2 $\alpha.1$  or OL2 $\alpha.2$ , which underlie  
431 the different array of O1 and O2 types (together with OL2 $\alpha.3$  which was found rarely),  
432 accounted for 72.0% (35,325/49,035) of all genomes.

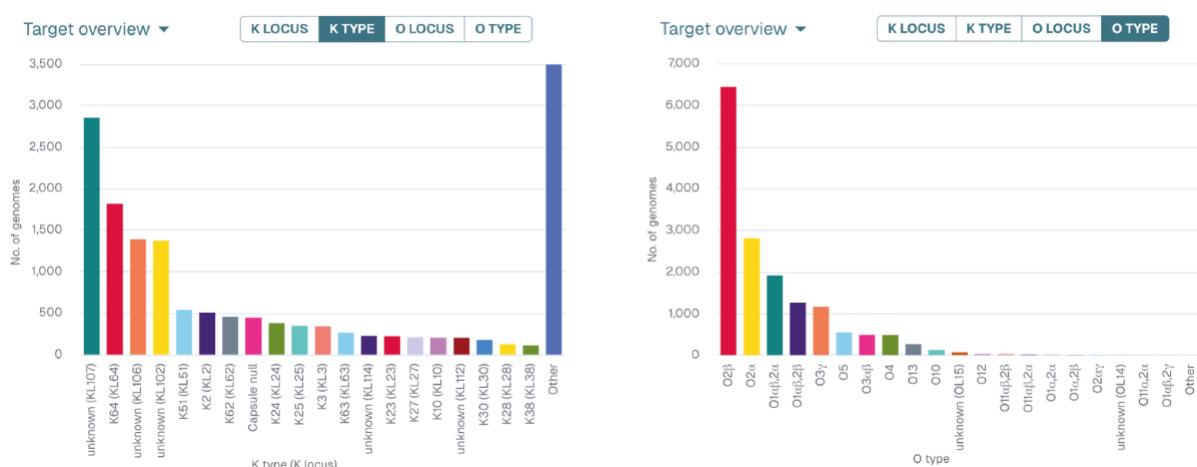
433

434 Using vaccines.watch, we can observe that the distribution of both K and O antigens among  
435 public genomes varies by country (**Figure 4**). This is largely related to the variable geographic  
436 distribution of STs, which are often dominated by single K and O loci/types. Assessment of  
437 the distribution of K antigens over time showed a proportional increase in KL64 (encoding  
438 K64) from around 2014, with it now being the most prevalent K locus type, accounting for  
439 22.5% (219/975) of genomes from 2024. This could be linked to an increase in ST147  
440 genomes, 81.1% (2363/2913) of which possess KL64. We also noted a rise in O2 $\alpha$  since 2020,  
441 accounting for 38.4% (374/975) of genomes from 2024, and mostly encoded by the OL2 $\alpha.1$   
442 variant. We could observe that this rise in O2 $\alpha$  has been driven by a proportional increase in  
443 genomes from both ST147 and ST45, 82.1% (2393/2913) and 92.4% (1112/1203) of which  
444 encode O2 $\alpha$ , respectively.

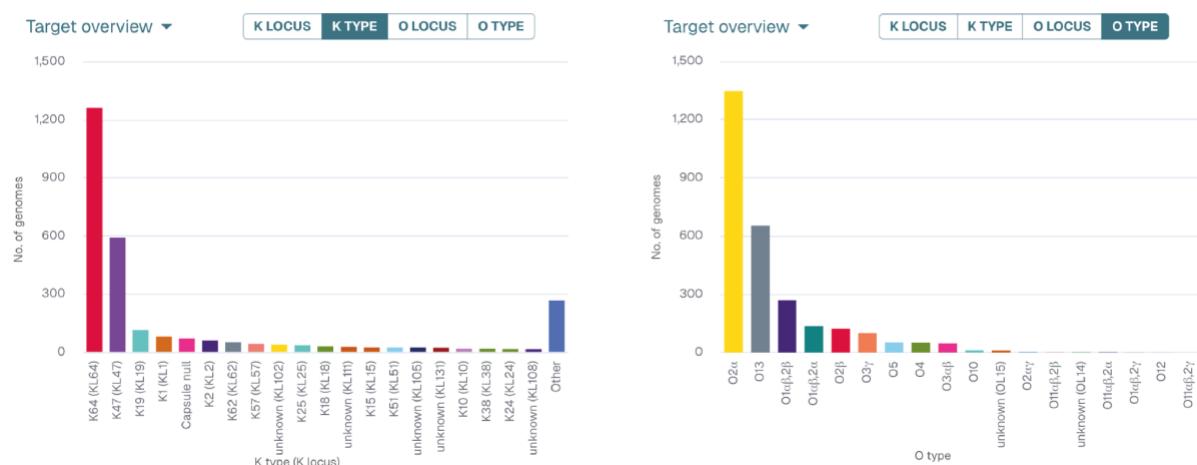
445

446

## A USA



## B China



447  
448

449 **Figure 4.** Number of genomes with predicted K and O types among *K. pneumoniae* species  
450 complex from the USA (A) and China (B), based on genomes represented in vaccines.watch  
451 as of 3 June 2025. Similar visualisations, updated in real-time, are available at:  
452 <https://vaccines.watch/organism/570?Country+Code=US> (A) and  
453 <https://vaccines.watch/organism/570?Country+Code=CN> (B).

454

455

## 456 *Acinetobacter baumannii*

457

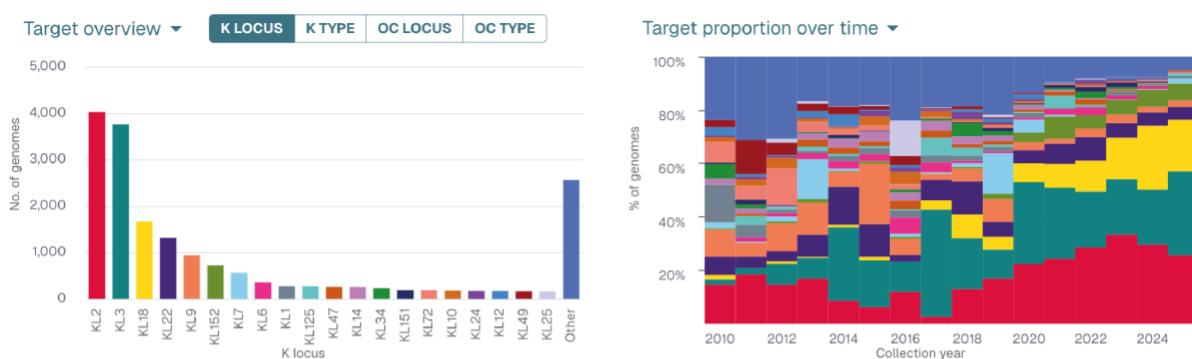
458 Similarly as for *K. pneumoniae* SC, vaccines.watch displays data from Kaptive on both the  
459 best-matching K/OC locus types (genotypes) identified from the *A. baumannii* genomes and  
460 the predicted K/OC types (phenotypes). As with the O loci/types in *K. pneumoniae* SC, the K  
461 loci/types in *A. baumannii* also do not conform to one-to-one relationships due to the use of  
462 both the K locus and additional genes in the phenotype predictions. Notably, the *A. baumannii*  
463 genome collection is highly dominated by a single variant type, ST2, from an internationally-  
464 distributed lineage known as global clone 2 (GC2) [34], which comprises 64.4%  
465 (11,866/18,428) of the total genomes.  
466

467 We investigated the diversity of K loci and predicted types, which are known to be the major  
468 immunodominant antigens in *A. baumannii* [16, 35], as well as the primary determinants of  
469 phage susceptibility [36]. We found 194 K loci among the 18,428 *A. baumannii* genomes, from  
470 a total of 241 currently defined by Kaptive. Despite the high overall diversity, KL2 (encoding  
471 K2) and KL3 (encoding K3 and K3-v1) dominated the public genome collection, accounting  
472 for 21.8% (4018/18,428) and 20.4% (3757/18,428) of the total genomes, respectively. The top  
473 twenty K locus types together accounted for 86.1% (15,867/18,428) of the total genomes,  
474 while 141 of the KL types identified were rare (present in <20 genomes). Of the genomes  
475 possessing KL2, 96.6% (3883/4018) were from ST2. Among those with KL3, 67.4%  
476 (2533/3757) were from ST2 while a further 22.9% (859/3757) were from ST437. Notably, we  
477 also found that 23.5% (4323/18,428) of all genomes possessed a K locus that corresponds to  
478 an unknown K type.  
479

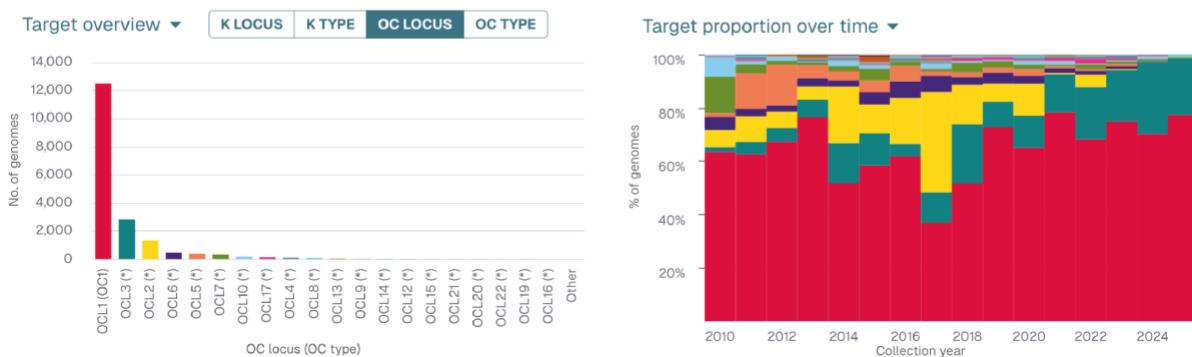
480 Assessment of the diversity of OC loci identified 22 locus types among the public genomes,  
481 corresponding to all of the known OC locus types defined within the Kaptive database.  
482 However, the genomes were highly dominated by OCL1 (encoding OC1) which accounted for  
483 67.8% (12,499/18,428), with OCL3 accounting for a further 15.3% (2816/18,428) and OCL2  
484 accounting for 7.3% (1341/18,428). 87.4% (10,919/12,499) of genomes with OCL1 were from  
485 ST2.  
486

487 Among both K locus and OC locus types, we observed a general trend towards lower diversity  
488 over time (**Figure 5**), which may reflect increased sampling of major multidrug-resistant  
489 lineages in recent years. In particular, we observed an increase in both KL18 and OCL3 from  
490 2020 onwards. This could largely be attributed to a rise in the proportion of ST499, which  
491 accounted for 84.2% (1408/1672) and 50.1% (1412/2816) of genomes with KL18 and OCL3,  
492 respectively. ST499 is reported to have emerged as a dominant non-GC2 carbapenem-  
493 resistant lineage in the USA in recent years [37]. ST499 genomes were primarily obtained  
494 from isolates sampled in the USA (99.2%; 1400/1412), with a further nine genomes from Fiji  
495 and a single genome obtained from each of Canada, Germany and India.

### A K locus



### B OC locus



496  
497

498 **Figure 5.** Distribution of K (A) and OC (B) locus types among *A. baumannii*, based on  
499 genomes represented in vaccines.watch as of 3 June 2025. The plots show the number of  
500 genomes with different locus types, ordered by frequency (left), and the proportion of the  
501 different locus types by sampling year, using the same colouring of types (right). Similar  
502 visualisations, updated in real-time, can be found at <https://vaccines.watch/organism/470>

503  
504  
505

## 506 Discussion

507

508 As pathogen genome sequencing becomes increasingly integrated into routine disease  
509 surveillance, there are growing opportunities for the shared global data to enhance precision  
510 around the targeting of public health interventions. Here we present an interactive platform,  
511 vaccines.watch (<https://vaccines.watch>), which aims to guide efforts in vaccine research and  
512 development by enabling interrogation of vaccine target diversity from genome data within the  
513 context of the pathogen population dynamics. The platform currently displays data from high-  
514 quality public genomes with geotemporal sampling information (post-2010), which are  
515 incorporated on an ongoing basis as genomes are deposited in INSDC databases. We have  
516 initially focused on developing vaccines.watch for polysaccharide targets from major bacterial  
517 pathogens which form the basis of existing or prospective multivalent vaccines. In particular,  
518 we have included the capsular polysaccharide targets from pneumococcal vaccines for which  
519 there remains an ongoing need for improved and/or updated formulations despite their high  
520 efficacy in reducing invasive pneumococcal disease [38]. We have also included capsular and  
521 LPS O antigen-based targets from *K. pneumoniae* SC and capsular and LOS outer core-based  
522 targets from *A. baumannii*. Both *K. pneumoniae* and *A. baumannii* are leading multidrug-

523 resistant pathogens that pose a critical public health threat [39, 40], particularly to vulnerable  
524 patients (including neonates) within hospital settings. There is only a limited vaccine pipeline  
525 for *K. pneumoniae* (one O antigen-based candidate, Kleb4V, completed a phase 1/2 trial in  
526 2022 (NCT04959344)) [41] while there are currently no vaccines in active clinical development  
527 for *A. baumannii* [4].

528  
529 As the global representativeness of genome data grows, we anticipate that vaccines.watch  
530 will aid the selection of vaccine targets for new or updated multivalent vaccine formulations.  
531 The choice of which precise target types to include within vaccine formulations, such as the  
532 *S. pneumoniae* serotypes within pneumococcal vaccines, is critical due to factors such as the  
533 varying frequency among target populations, invasiveness potential and antimicrobial  
534 resistance profiles associated with different types [42, 43]. Due to geographic differences  
535 among pathogen populations, we envisage that new vaccine formulations will be increasingly  
536 tailored to specific localities to enable maximal protection against disease. For example, a 10-  
537 valent PCV (Pneumosil) has already been designed to provide protection against serotypes  
538 causing the highest disease burden in Africa, Asia, Latin America and the Caribbean [44].  
539 Increasing availability of representative genomic data may also be used to assess the likely  
540 efficacy of existing vaccine formulations within different localities and/or target populations,  
541 and inform choice where multiple vaccines with differing compositions exist. This approach  
542 has the potential to expand the use of existing vaccines, including pneumococcal vaccines  
543 which still have highly variable coverage worldwide [45].

544  
545 We also anticipate that, with improving genome representation, vaccines.watch could be used  
546 for monitoring the impact of new vaccines by enabling comparison of target types and the  
547 associated population diversity before and after vaccine introduction. Post-vaccination  
548 monitoring is especially critical with polysaccharide-based vaccines, as these target antigens  
549 are known to exhibit high rates of recombinational exchange and therefore are liable to  
550 "switching" [46, 47]. Numerous studies have demonstrated the value of genomic data for high-  
551 resolution monitoring of vaccine-related population changes that, for example, have led to an  
552 increase in non-vaccine pneumococcal serotypes with higher invasiveness and/or AMR [48,  
553 49]. In the case of pneumococcal disease, the ongoing approach to address these changes is  
554 to develop new PCV formulations with increasing serotype coverage, with licensed vaccines  
555 now including up to 21 serotypes [15].

556  
557 The opportunities described above have the potential to launch an exciting new era of vaccine  
558 development yet remain highly dependent on global efforts to increase and sustain pathogen  
559 genome sequencing capacity for routine surveillance. Our review of global public genomes,  
560 which represent the current data source for vaccines.watch, highlights the substantial gaps in  
561 geographic coverage, paucity of recent data (likely, in part, due to a lag in deposition times),  
562 and ongoing biases in sequencing which remains dominated by specific research agendas  
563 (e.g. relating to AMR). We therefore advise users to remain highly vigilant to these data  
564 limitations and maintain careful consideration of how data shown in vaccines.watch is used  
565 and interpreted. Another important limitation in the current use of genomic data for vaccine  
566 development and monitoring efforts is that there is still an incomplete understanding of how  
567 the genomic information relates to the polysaccharide phenotypes, with many genomic loci  
568 encoding unknown structures [33], including across all three pathogens currently included  
569 within vaccines.watch. However, for each of these three pathogens, there are active efforts to  
570 improve this knowledge base, with regular updates made to the nomenclature databases and

571 phenotype prediction logic provided by the SeroBA [27] and Kaptive tools [25]. In particular,  
572 for *S. pneumoniae*, there is a comprehensive and curated library, SeroBAnk, collating data on  
573 the genetic locus and capsular structure of each known serotype [27].  
574

575 Future developments of vaccines.watch include extending our approach to additional vaccine  
576 targets, starting with polysaccharide targets from other bacterial pathogens with *in silico* typing  
577 schemes and that form the basis of similar multivalent vaccine formulations. We also aim to  
578 develop the ability to display data from bespoke genome collections that may be pre-defined  
579 within vaccines.watch or sourced by users themselves. Finally, we can readily adapt  
580 vaccines.watch to include other data types, such as additional metadata associated with  
581 pathogen genomes (e.g. source, disease type, patient characteristics). This will be important  
582 as available metadata becomes increasingly harmonised for individual pathogens, driven by  
583 recent ongoing curation efforts of the public health and research communities and  
584 development of metadata templates.  
585

586 In conclusion we anticipate that, as genomic surveillance efforts increase, the use of large-  
587 scale genomic data will become an essential component of the vaccine developer's toolkit.  
588 With the development of vaccines.watch, we have demonstrated how global genome data can  
589 be ingested, analysed and delivered in real-time via an accessible platform to provide insights  
590 into the pathogen population dynamics around key vaccine targets. The platform aims to  
591 support scientists and vaccine developers with decision-making around vaccine formulations  
592 and roll-out.  
593  
594  
595

## 596 **Funding**

597  
598 This work was supported by Official Development Assistance (ODA) funding from the National  
599 Institute for Health Research (grant number NIHR133307) with additional funding provided by  
600 the Gates Foundation (grant ref INV-025280).  
601  
602  
603

## 604 **Conflicts of interest**

605  
606 The authors declare no conflicts of interest.  
607  
608  
609

## 610 **Acknowledgements**

611  
612 We would like to thank Joshua Wong for helpful discussions regarding the development of  
613 vaccines.watch.  
614  
615  
616

617 **Data availability**

618

619 All data represented in vaccines.watch are available for download within the application. The  
620 assembled genomes and associated metadata can be accessed via Pathogenwatch  
621 (<https://next.pathogen.watch>), together with additional genotypic data. Raw sequence reads  
622 are available in the ENA/SRA.

623

624

625

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627

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