# Advanced Image Processing and Analysis

ECE 4438B/ECE 9202B/ECE 9022B
BIOMED/BIOPHYS/CAMI 9519B
Winter 2018

Instructor

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Robarts Research Institute

Department of Electrical and Computer Engineering

Biomedical Engineering Graduate Program

Department of Medical Biophysics

# Advanced Image Processing and Analysis (AIPA)

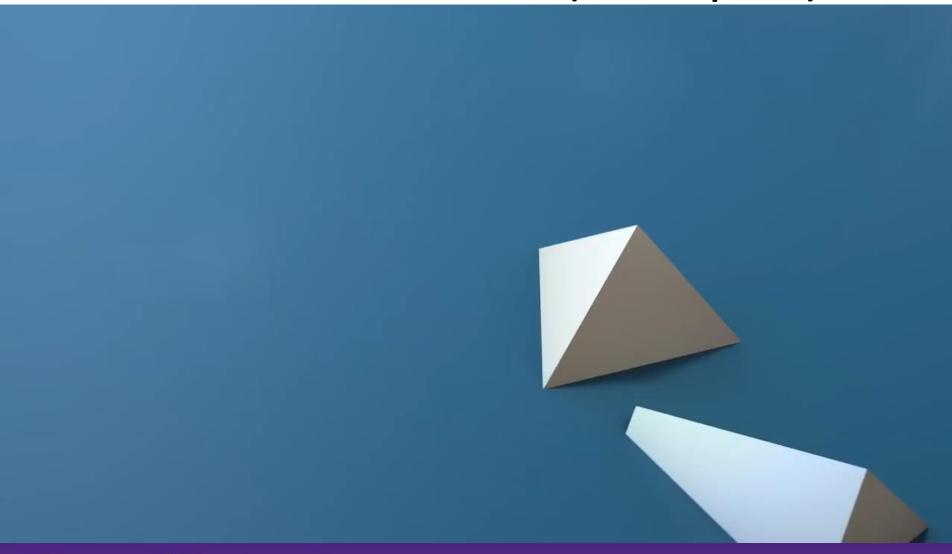
- Offered to 3 departments (undergrad/grad)
  - ECE 4438/ECE 9202/ECE 9022
  - BIOMED 9519
  - BIOPHYS/CAMI 9519
- Time and Place
  - Monday 2:30-3:30pm, AHB-1B08
  - Tuesday 12:30-2:30pm, FNB-3210

- Theme: Digital image analysis with examples drawn from medical imaging
- Questions:
  - How to design and implement algorithms to delineate structures of interests (Segmentation)?
  - How to design and implement algorithms to align different images (Registration)?
  - How to evaluate the performance of these algorithms (Validation)?

## **Applications**

- Entertainment
  - Movies (special effects)
- Informatics
  - Self-driving car
  - Adjuncts to computer vision and computer graphics
- Augmentation
  - Medical interventions

## What/Motivation (Examples)



## Motivation (Special Effects)

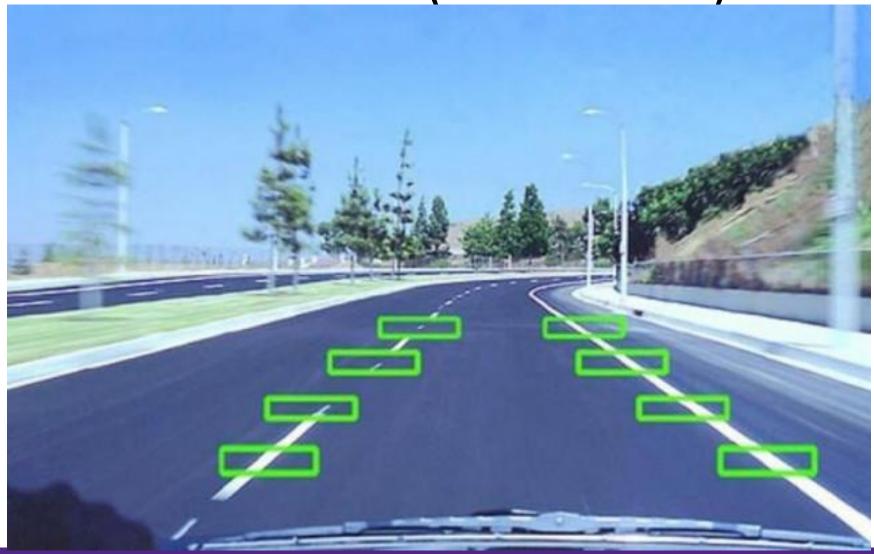
## **Applications**

- Entertainment
  - Movies (special effects)
- Informatics
  - Self-driving car
  - Adjuncts to computer vision and computer graphics
- Augmentation
  - Medical interventions

Motivation (Land Departure Warning)



Motivation (Informatics)





Western Engineering http://g3ict.org/resource\_center/newsletter/news/p/id\_

## **Motivation (Interventions)**





#### Revealing Invisible Changes In The World

Created for the NSF International Science & Engineering
Visualization Challenge 2012

### Motivation



- Theme: Digital image analysis with examples drawn from medical imaging
- Questions:
  - How to design and implement algorithms to delineate structures of interests?
    - Segmentation
      - Manual
      - Semi-automatic
      - Automatic

- Theme: Digital image analysis with examples drawn from medical imaging
- Questions:
  - How to design and implement algorithms to align different images?
    - Registration
      - Rigid registration
      - Deformable registration

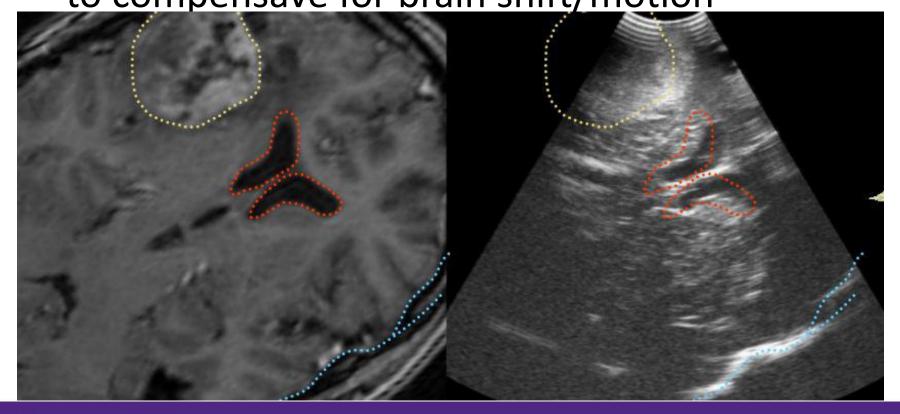
- Theme: Digital image analysis with examples drawn from medical imaging
- Questions:
  - How to evaluate the performance of these algorithms?
    - Evaluation

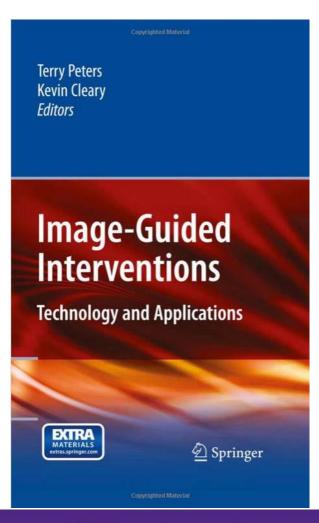
 Medical Images are often acquired preoperatively (MRI/CT), used for diagnostics and surgical planning

However, intra-operatively, organ deforms/moves



 Pre-op MRI registered to intra-op ultrasound to compensave for brain shift/motion





 PDF version available free of charge through Western's library: <a href="https://journals.scholarsportal.info/details/15239829/v12inone/119">https://journals.scholarsportal.info/details/15239829/v12inone/119</a> ii

traca.xml

## Application: Vessel Stenosis

- Segmentation of an MR angiogram allows 3D depiction of vessels and detection of stenosis (vessel narrowing)
- On the basis of segmentation, the diameter of stenosis can accurately be determined





## Application: Orthopaedic Surgery

Virtual representation
 of the ankle determined
 from pre-op CT, volume rendered, and
 registered to patient
 intra-operative to allow
 an augmented reality
 display



#### **Evaluation**

	Assignments	Mid-term	Final Exam	Project
Undergrad	20%	30%	50%	n/a
MEng	20%	30%	50%	n/a
MSc	20%	20%	25%	35%
PhD	20%	20%	25%	35%

- Undergrad (ECE 4438B) and Meng (ECE9022)
  - 4 assignments
  - Closed-book mid-term exam (Feb 27<sup>th</sup>, 2hr)
    - Review session on Feb 12 and 13 (tentative)
  - Closed-book final exam (comprehensive)

#### **Evaluation**

	Assignments	Mid-term	Final Exam	Project
Undergrad	20%	30%	50%	n/a
MEng	20%	30%	50%	n/a
MSc	20%	20%	25%	35%
PhD	20%	20%	25%	35%

#### Graduate students

- 4 assignments
- Closed-book mid-term exam (Feb 27<sup>th</sup>, 2hr)
- Closed-book final exam (comprehensive)
- Project involves implementation of an image processing technique/algorithm
  - Topic chosen with consultation with course instructor
  - With alignment to thesis topic/work

- Late submission policy: penalized at a rate of 20% per 24 hours overdue, NO EXCEPTIONS
- Any reason for late submission must be
  - Brought to the attention to the instructor BEFORE the deadline
  - With documentation
  - No consideration after the assignment deadline

- Use of English: in accordance with Senate and Faculty Policy, students may be penalized up to 10% of the marks on assignments, tests, and examinations for improper use of English
- Poorly written work may be returned without grading (except the final exam)

- Attendance: Any student who is absent too frequently from class will be reported to the Dean (after due warning has been given)
- On the recommendation of the department, and with permission of the Dean, the student will be debarred from taking the regular final examination

- Mid-term and the final examination cannot be missed:
  - Unless
    - Due to illness
    - Other extreme circumstances (death of family member, etc)
  - Should consult with the instructor/Department
     Chair immediately
  - Documentation

#### Course format

- OWL (<a href="https://owl.uwo.ca/">https://owl.uwo.ca/</a>)
  - Course notes and assignments can be downloaded
  - Assignments are submitted via OWL
  - OWL entry is "<u>ECE 4438B 001 FW17"</u>
- Github
  - Additional course materials (codes and examples) are accessible via github
  - Jupyter notebook

## Theme and Topics

- Theme: Digital image processing and analysis, with examples drawn from medical imaging
- Topics:
  - Segmentation
  - Registration
  - Validation

- Python (<a href="https://www.python.org/">https://www.python.org/</a>)
  - Interpreted language
  - Power of C++
  - Ease of Matlab
  - Processing + visualization, large number of algorithms, best suited to 2D

- Insight Segmentation and Registration Toolkit (ITK, <a href="https://itk.org/">https://itk.org/</a>)
  - Library for image processing
  - Open source
  - Implemented in C++, with binding to many other languages include python and java
  - Processing only, vary large number of algorithms, suited to 2D/3D/4D/nD

- SimpleITK (<u>www.simpleitk.org</u>)
  - Open source
  - Simplified layer built on top of ITK
  - Intended to facilitate its use in rapid prototyping, education, interpreted languages
    - Python
    - R

- Jupyter Notebook (<a href="http://jupyter.org/">http://jupyter.org/</a>)
  - Open-source web application that allows one to create and share documents that contain live code, equations, visualizations, and narrative text
  - SimpleITK kernel
  - Executes python/SimpleITK codes within a browser (think Matlab) and visualize the results immediately

#### ITK

```
#include "itkImage.h"
#include "itkImageFileReader.h"
#include "itkRescaleIntensityImageFilter.h"
#include "itkLaplacianSharpeningImageFilter.h"
#include "itkSubtractImageFilter.h"
#include "QuickView.h"
int main(int argc, char * argv[])
 // Verify command line arguments
 if (argc < 2)
    std::cerr << "Usage: " << std::endl;</pre>
    std::cerr << argv[0] << " inputImageFile" << std::endl;</pre>
    return EXIT FAILURE;
 // Parse command line arguments
 std::string inputFilename = argv[1];
```





#### ITK

```
// Setup types
typedef itk::Image< float, 2 >
                                  FloatImageType;
typedef itk::ImageFileReader< FloatImageType > readerType;
readerType::Pointer reader = readerType::New();
reader->SetFileName(inputFilename);
typedef itk::LaplacianSharpeningImageFilter<FloatImageType, FloatImageType >
                                                                              LaplacianSharpeningImageFilterType;
LaplacianSharpeningImageFilterType::Pointer laplacianSharpeningImageFilter =
  LaplacianSharpeningImageFilterType::New();
laplacianSharpeningImageFilter->SetInput( reader->GetOutput() );
typedef itk::SubtractImageFilter<FloatImageType>
                                                           SubtractType;
SubtractType::Pointer diff = SubtractType::New();
diff->SetInput1(reader->GetOutput());
diff->SetInput2(laplacianSharpeningImageFilter->GetOutput());
```





#### ITK

```
QuickView viewer;
viewer.AddImage(
  reader->GetOutput(),true,
  itksys::SystemTools::GetFilenameName(argv[1]));
std::stringstream desc;
desc << "LaplacianSharpeningImageFilter";</pre>
viewer.AddImage(
  laplacianSharpeningImageFilter->GetOutput(),
  true,
  desc.str());
std::stringstream desc2;
desc2 << "Original - LaplacianSharpening";</pre>
viewer.AddImage(
  diff->GetOutput(),
  true.
  desc2.str());
viewer.Visualize();
return EXIT SUCCESS;
```





#### ITK

```
cmake_minimum_required(VERSION 2.8.12)

project(LaplacianSharpeningImageFilter)

find_package(ITK REQUIRED)
  include(${ITK_USE_FILE})
  if (ITKVtkGlue_LOADED)
    find_package(VTK REQUIRED)
    include(${VTK_USE_FILE})
  else()
    find_package(ItkVtkGlue REQUIRED)
    include(${ItkVtkGlue_USE_FILE})
    set(Glue ItkVtkGlue_USE_FILE})
  set(Glue ItkVtkGlue)
  endif()

add_executable(LaplacianSharpeningImageFilter MACOSX_BUNDLE LaplacianSharpeningImageFilter.cxx)
  target_link_libraries(LaplacianSharpeningImageFilter
    ${Glue} ${VTK_LIBRARIES} ${ITK_LIBRARIES})
```





#### SimpleITK Notebooks

#### Image Sharpening Example using SimpleITK

This is a simple demonstration of the power of SimpleITK, based on its C++ equivalent shown here: https://itk.org/Wiki/ITK/Examples/ImageProcessing/LaplacianSharpeningImageFilter

We assume the an image is available and located in the same directory as this Jupyter Notebook. In this case, an image of a woodpicker is copied and saved as 'woodpicker.png" from the <u>URL</u> above.

```
In []: # import SimpleITK library
import SimpleITK as sitk

In []: # Assuming the image is located under the data/image directory, this is how we can load an image

# The input to the function is a string (enclosed in '') of the file name.

# The output is the image/array. Note no explicit typing is needed in Python
img = sitk.ReadImage('...\data\images\woodpicker.png')

In []: # display the image. If we set up our environment properly using the in-class example, ImageJ will be used to display the image.

sitk.Show(img)

# Note there is a slider bar in ImageJ, why?

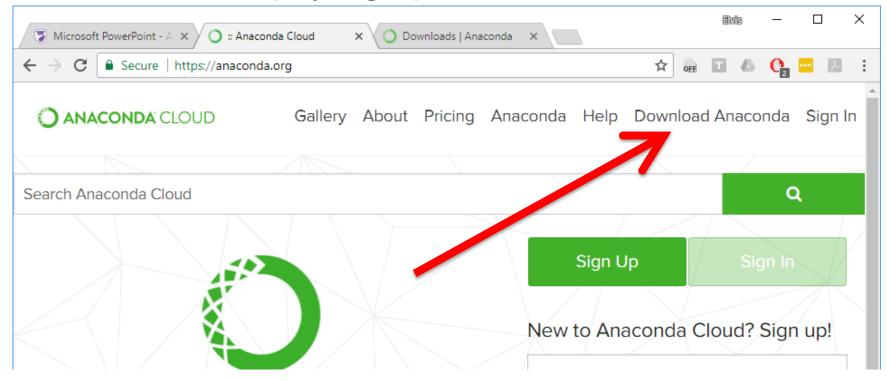
In []: lap = sitk.LaplacianSharpeningImageFilter() # this is an in-line comment

In []: outimg = lap.Execute(img) # input is the original image, output is the sharpened image

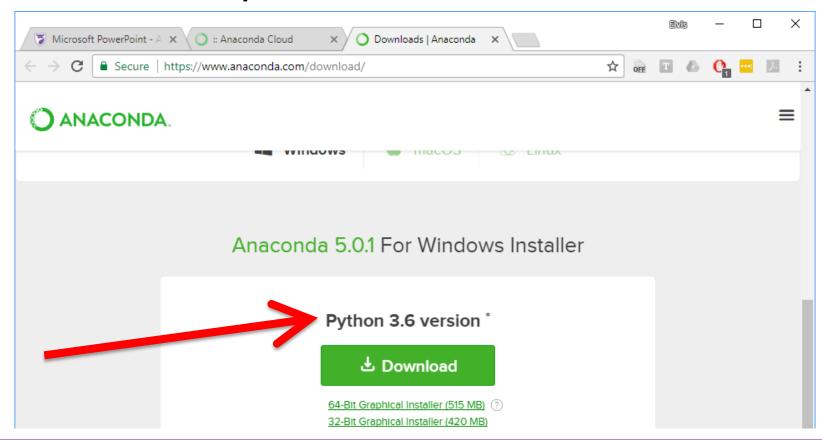
In []: sitk.Show(outimg) # display the result.
```

This is it! Compare this python code in SimpleITK to the C++ equivalent in ITK.

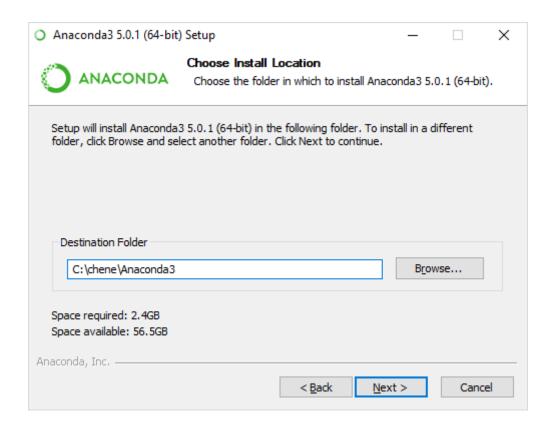
- Install Anaconda (<a href="https://anaconda.org/">https://anaconda.org/</a>)
  - Download (top-right)



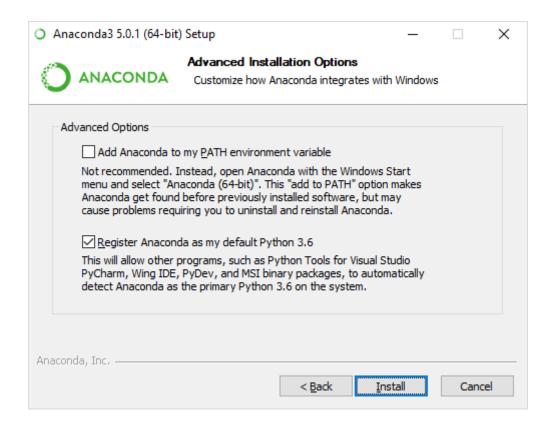
Download Python 3.6



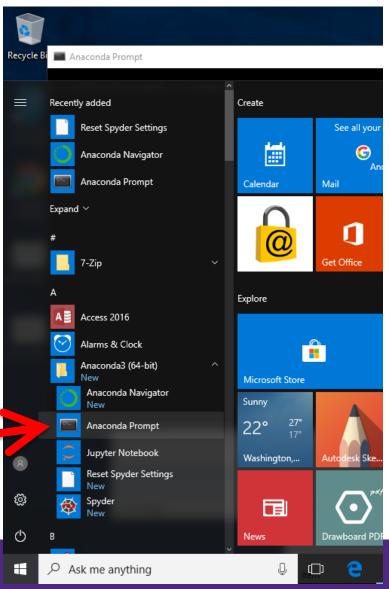
Install



Install



 Anaconda Command Prompt



- Update using the following 2 commands
- 'conda update conda'

```
Select Anaconda Prompt
                                                                                                              (C:\chene\Anaconda3) c:\chene>conda update conda
etching package metadata ......
CondaHTTPError: HTTP 000 CONNECTION FAILED for url <https://repo.continuum.io/pkgs/r/win-64/repodata.json.bz2>
Elapsed: -
An HTTP error occurred when trying to retrieve this URL.
HTTP errors are often
                      termittent, and a simple retry will get you on your way.
ConnectionError(ReadTime
                             ror("HTTPSConnectionPool(host='repo.continuum.io', port=443): Read timed out.",),)
(C:\chene\Anaconda3) c:\chene>conda update conda
 etching package metadata .....
Solving package specifications: .
Package plan for installation in environment C:\chene\Anaconda3:
The following packages will be UPDATED:
   anaconda: 5.0.1-py36h8316230 2 --> custom-py36h363777c 0
   conda: 4.3.30-py36h7e176b0 0 --> 4.4.6-py36 0
   pycosat: 0.6.2-py36hf17546d 1 --> 0.6.3-py36h413d8a4 0
Proceed ([y]/n)? y
anaconda-custo 100% |######################## Time: 0:00:00 192.77 kB/s
pycosat-0.6.3- 100% |##############################
                                                    Time: 0:00:00 166.84 kB/s
conda-4.4.6-py 100% |######################### Time: 0:00:43 22.62 kB/s
(C:\chene\Anaconda3) c:\chene>
```

- Update using the following 2 commands
- 'conda update anaconda'

```
Select Anaconda Prompt - conda update anaconda
                                                                                                         Time: 0:00:00 192.77 kB/s
                                                 Time: 0:00:00 166.84 kB/s
onda-4.4.6-py 100% |#############################
                                                 Time: 0:00:43 22.62 kB/s
(C:\chene\Ana
               c:\chene>conda update anaconda
Solving envir
                 : done
  rackage Pl
 environment location: C:\chene\Anaconda3
 added / updated specs:
   - anaconda
The following packages will be downloaded:
   package
                                        build
                                py36hb8ac631 0
   certifi-2017.11.5
                                                     196 KB
   openssl-1.0.2n
                                                     5.4 MB
                                        Total:
                                                     5.6 MB
The following packages will be UPDATED:
   certifi: 2017.7.27.1-py36h043bc9e_0 --> 2017.11.5-py36hb8ac631_0
   openssl: 1.0.2l-vc14hcac20b0 2 --> 1.0.2n-h74b6da3 0
Proceed ([y]/n)? _
```

- (Optional) install git
- 'conda install git

```
Anaconda Prompt - conda install git

(d:\chene\dl\Continuum\anaconda3) d:\chene\home>conda install git
Fetching package metadata ......

Solving package specifications: .

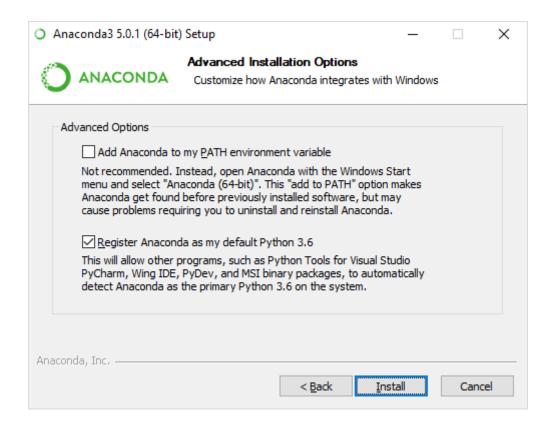
Package plan for installation in environment d:\chene\dl\Continuum\anaconda3:

The following packages will be SUPERSEDED by a higher-priority channel:

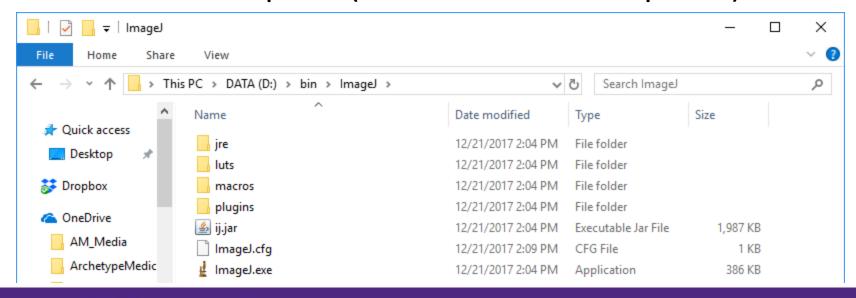
git: 2.15.0-heaa8ca2_0 --> 2.14.2-3 conda-forge

Proceed ([y]/n)? __
```

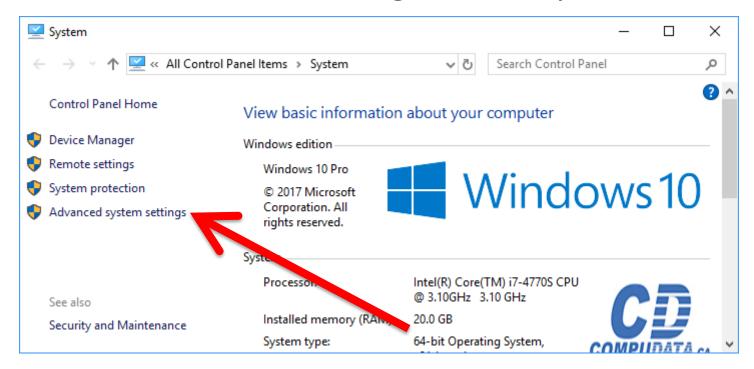
Install



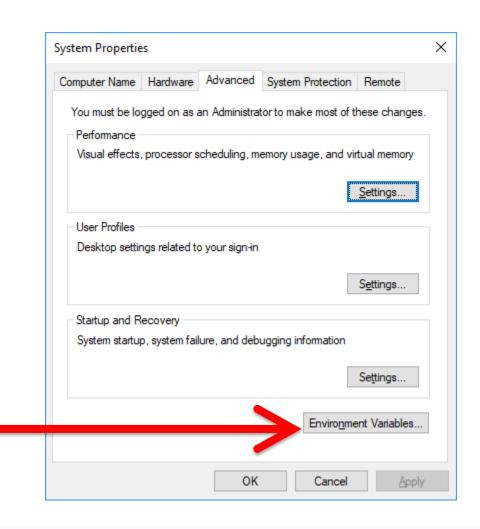
- Install imagej
  - Download it from https://imagej.nih.gov/ij/download.html
  - extract the zip file (no 'installation' required)



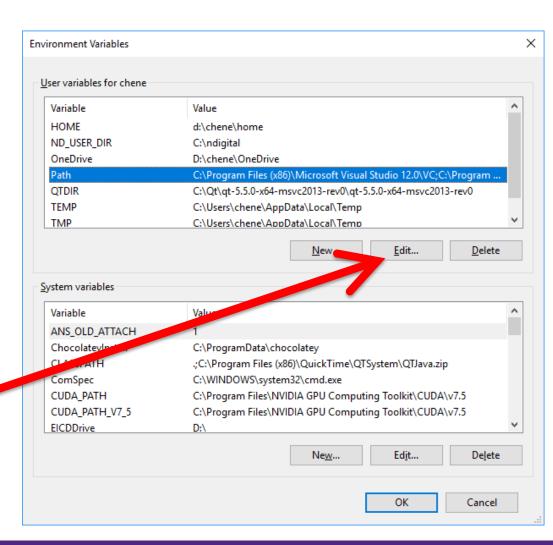
- Install imagej
  - Put the location of 'ImageJ.exe' to path



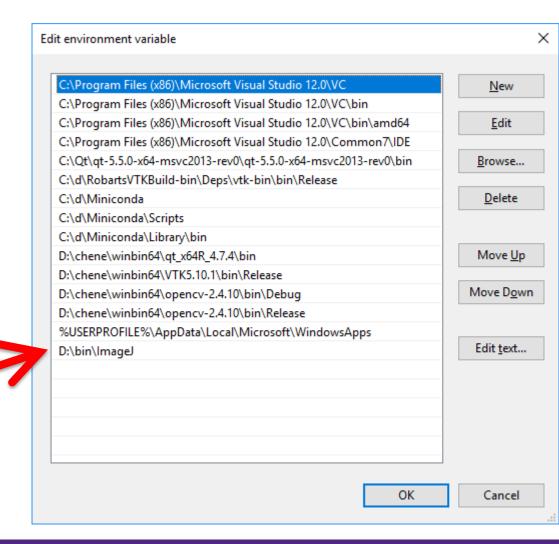
- Install imagej
  - Put the location of 'ImageJ.exe' to path



- Install imagej
  - Put the location of 'ImageJ.exe' to path



- Install imagej
  - Put the location of 'ImageJ.exe' to path



 In an Anaconda prompt, create a directory at a location of choice,

```
AIP_W18_Notebooks

(d:\chene\d1\Continuum\anaconda3) C:\Users\chene\Downloads\temp>c:

(d:\chene\d1\Continuum\anaconda3) C:\Users\chene\Downloads\temp>cd \Users\chene\Downloads\temp

(d:\chene\d1\Continuum\anaconda3) C:\Users\chene\Downloads\temp>git clone https:
//github.com/chene77/AIP_W18_Notebooks.git
fatal: destination path 'AIP_W18_Notebooks' already exists and is not an empty d irectory.

(d:\chene\d1\Continuum\anaconda3) C:\Users\chene\Downloads\temp>
```

 Using the newly install git, clone the following github repository: 'git clone <a href="https://github.com/chene77/AIP">https://github.com/chene77/AIP</a> W18 Noteb ooks.git'

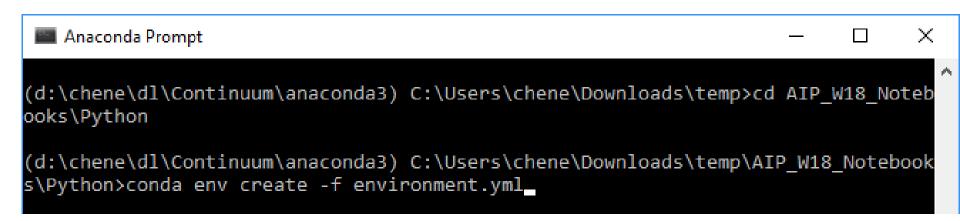
```
Anaconda Prompt

— — — X

(d:\chene\dl\Continuum\anaconda3) C:\Users\chene\Downloads\temp>git clone https:
//github.com/chene77/AIP_W18_Notebooks.git
fatal: destination path 'AIP_W18_Notebooks' already exists and is not an empty d irectory.

(d:\chene\dl\Continuum\anaconda3) C:\Users\chene\Downloads\temp>
```

 'cd' to the Python directory, create an environment for SimpleITK using the following command 'conda env create –f environment.yml'

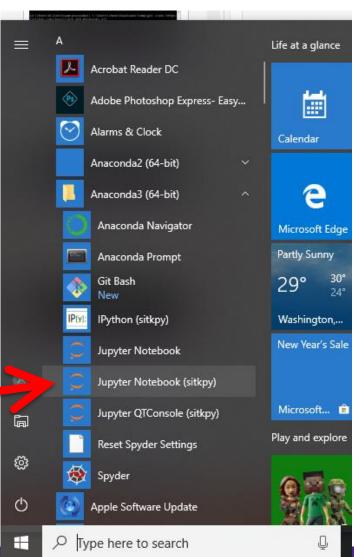


 After the installation is complete, activate SimpleITK environment with the following 'conda activate sitkpy'

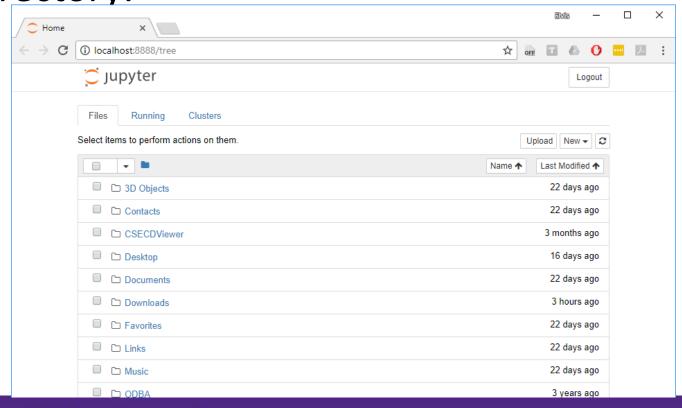
```
\\sitkpy\\Scripts\\jupyter-qtconsole-script.py']
done
#
# To activate this environment, use
#
# $ conda activate sitkpy
#
# To deactivate an active environment, use
#
# To deactivate an active environment, use
#
# $ conda deactivate
```



 Now you should have a Jupyter Nootbook environment for SimpleITK. Execute it will bring up a web page hosted locally



 The 'root' directory is the user's home directory:



#### Conclusions

- Now we have a working python with a visualization environment
- Continued with the python tutorial...

#### Useful links

- Jupyter Notebook tutorial
  - https://www.datacamp.com/community/tutorials/ /tutorial-jupyter-notebook

# Jupyter Notebook

Jupyter Notebook (SITKPY) live demo

# Questions/Comments

- Instructor
  - Elvis Chen, PhD, LEL
  - chene@Robarts.ca
- Teaching Assistant
  - Madeleine Van De Kleut
  - mvandekl@uwo.ca