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# **Notes on metabolic modelling analysis**

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## CONSTRAINT BASED ANALYSIS

In this notebook, we are going to discuss some basics of CBA from their mathematical, theoretical and biological perspective.

### 3.1 Overflow metabolism is caused by two growth-limiting constraints

#### Literature

- de Groot et al (2020) [DOI](#)
- Basan et. al (2015) [DOI](#)

Overflow metabolism in *Escherichia coli* results from efficient proteome allocation.

The authors assume that the yield of energy per carbon molecule is higher for respiration than for fermentation:  $n_r > n_f$ , but that fermentation is more proteome-efficient:  $\epsilon_f > \epsilon_r$ .



## HOW TO

In this notebook, we will keep track of handy implementations for metabolic modeling related tasks.

### 4.1 Media and environment setup

#### 4.1.1 Extracellular reactions

COMETS includes the capability to simulate reactions happening in the extracellular environment, without association to a specific organism. Users can implement either elementary reactions of arbitrary order based on mass-action kinetics, or enzyme-catalyzed reactions obeying Michaelis–Menten kinetics, e.g., for the simulation of extracellular enzymes.

#### 4.1.2 Get complete medium for ModelSEED

With the term *complete medium*\* we describe an *in silico* object where any compound that could be used as a nutrient, it is available for the model.

To build this object for the case of ModelSEED, we need to first get all the possible compounds. And we can do this by first, getting locally the [ModelSEEDDatabase repo](#).

Then we can explore the `Biochemistry` folder of that to retrieve all possible nutrients that could be imported in our model.

From the `Biochemistry` folder of the dev branch of the ModelSEEDDatabase repository, run:

```
!awk -F"\t" '$6 != 1 && $18==0 {print $5}' reaction_*.tsv > TRANSPORT_REACTIONS.tsv
```

```
awk: fatal: cannot open file `reaction_*.tsv' for reading: No such file or
↳directory
```

Now, with something like the following Python chunk, you can build the complete medium and export in a `.csv` file that with the applied format, could be used for gapfilling with the `fill` command of the `gapseq` tool.

```
def write_to_gapseq_format(all_compounds, cpd2name, output_file):
    """
    Write a 3-col csv file with the compound id, its name and a boundary flux of 1000
    """
    with open(output_file, "w") as f:
        counter = 0
        for compound in all_compounds:
            if compound in cpd2name:
```

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```

        counter += 1
        f.write(f"{compound}\\t{cpd2name[compound]}\\t1000\\n")
    else:
        print(f"Compound {compound} not found in cpd2name dictionary")

print(f"Total compounds written: {counter}")

def process_transport_reactions(input_file, output_file=None):
    """
    Parse the TRANSPORT_REACTIONS.tsv file to export compounds that should be part of
    the complete medium.
    """
    with open(input_file) as f:
        lines = f.readlines()

    ex = [line.strip() for line in lines if len(line.split(";")) == 2]

    cpd2name = {}
    all_compounds = set()

    for reaction in ex:
        compounds = reaction.split(";")
        c1 = compounds[0].split(":")[1]
        c2 = compounds[1].split(":")[1]

        if c1 == c2:
            name = compounds[0].split(":")[-1]
            all_compounds.add(c2)
            if c2 not in cpd2name:
                cpd2name[c2] = name

    if output_file is not None:
        write_to_gapseq_format(all_compounds, cpd2name, output_file)

# Main execution
if __name__ == "__main__":
    process_transport_reactions("TRANSPORT_REACTIONS.tsv", "complete_modelseed_medium.
    ↪Csv")

```

Total compounds written: 0



## SOFTWARE AND RESOURCES

In this page we will keep a list of the software approaches we are aware of for the various metabolic modeling tasks. Apparently, this list can never be complete, but it can be improved with your contribution! So, feel free to make a PR adding something or contact us to do that for you.

### 5.1 Reconstruction, gap-filling, validation

- DEMETER
- 

### 5.2 Further constraints

#### 5.2.1 Thermodynamics

Tool	Description	Ar- chi- tec- ture	Repr	DOI
pyTFA	implementations of the original thermodynamics-based Flux Analysis (TFA) paper. Specifically, they include explicit formulation of Gibbs energies and metabolite concentrations, which enables straightforward integration of metabolite concentration measurements.	stand alone	<a href="#">GitH</a>	<a href="#">OA</a>
Opt-MDF-pathway**	integration of thermodynamic information in metabolic models to assess the feasibility of flux distributions by thermodynamic driving forces. Extends the framework of Max-min Driving Force (MDF) for thermodynamic pathway analysis. Identifies both the optimal MDF for a desired phenotypic behavior and the respective pathway that supports the optimal driving force.	stand alone	<a href="#">GitH</a>	<a href="#">OA</a>

- [matTFA](#) is the MATLAB version of pyTFA \*\* part of the CNApy library

### 5.2.2 Enzymes

Tool	Description	Architecture	Repo	DOI
Au-toPAC-MEN	Retrieves keat data and adds protein allocation constraints to stoichiometric metabolic models according to the sMOMENT method.	stand-alone	<a href="#">GitHub</a>	<a href="#">OA</a>
ET-GEMs	Construction of enzymatic and thermodynamic constrained GEMs in a single <a href="#">Pyomo</a> * modelling framework.	scripts	<a href="#">GitHub</a>	<a href="#">OA</a>

\*Python-based, open-source optimization modeling language

### 5.2.3 Transporter annotations

Tool	Description	Architecture	Repo	DOI
SPO1	machine learning model that can successfully predict specific substrates for arbitrary transport protein	stand-alone	<a href="#">GitHub</a>	<a href="#">OA</a>
TransYT	Identifies transport systems and the compounds carried across membranes, based on the annotations of the Transporter Classification Database (TCDB). Generates the respective transport reactions while providing the respective Gene-Protein-Reaction associations.	<a href="#">web-service</a>	<a href="#">GitHub</a>	<a href="#">OA</a>

- They showed that the majority of TCDB entries are of low UniProt-based annotation scores.

## DATABASES

Tool	Description	Link	DOI
equili- brator	estimated thermodynamic constants	<a href="#">link</a>	OA
SABIO- RK	curated information about biochemical reactions, their kinetic rate equations with parameters and experimental conditions.	<a href="#">link</a>	OA

has also an API library `equilibrator-api` [ReadTheDocs](#) | [GitLab](#)

and a cache one `equilibrator-cache`: A database application for caching data used by eQuilibrator and related projects. Stored data includes compound names, structures, protonation state information, reaction and enzyme info, and cross-references to other databases. All compounds stored in equilibrator-cache are cross-referenced using InChIKey. [GitLab](#)

## 6.1 Topological approaches

- QFCA
- **FluxModeCalculator**

Efficient Elementary flux mode (EFM) calculation

[paper](#) | [GitHub](#) (part of the MCS repo)

## 6.2 Dynamic approaches

- **dfba** [GitLab repo](#) [Documentation](#) paper A recent approach for dynamic FBA that considers the solution non-uniqueness.

-COMETS

-BacArena

## 6.3 Community modelling

- mergem
- PyCoMo
- 

## 6.4 Data integration

- FASTCROMICS
- TRFBA

tellurium:

<https://tellurium.analogmachine.org>

A Python Environment for Reproducible Dynamical Modeling of Biological Networks

## 6.5 Other resources and tools of interest

Tool	Description	Architecture	Repo	DOI
gRodon2	predicts maximal growth rates using genomic data	R-package	<a href="#">GitHub</a>	<a href="#">OA</a>

## BIOLOGICAL INSIGHT

Here I keep track of bio facts that can be of use in my *in silico* explorations.

Apparently, this notebook will make much less sense since it will include pieces of information that do not necessarily lead somewhere :sweat\_smile:

*Pseudomonas aeruginosa*: May rely on glutamine synthetase and nitrate/nitrite reduction for glutamine production.



**BIBLIOGRAPHY**





## BIBLIOGRAPHY

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- [4] Bernhard Ø Palsson. *Systems biology: simulation of dynamic network states*. Cambridge University Press, 2011.
- [5] Neema Jamshidi and Bernhard Ø Palsson. Mass action stoichiometric simulation models: incorporating kinetics and regulation into stoichiometric models. *Biophysical journal*, 98(2):175–185, 2010.



### Balanced growth

Balanced growth (*background*), 5

### M-models

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### MASS

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### flux

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### gradient matrix

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### jacobian matrix

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### kinetic variables

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