# ORIGINAL RESEARCH PAPER

# Equivalence testing for standardized effect sizes in linear regression

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#### ARTICLE HISTORY

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#### ABSTRACT

Determining a lack of association between an outcome variable and a number of different explanatory variables is frequently necessary in order to disregard a proposed model (i.e., to confirm the lack of a meaningful association between an outcome and predictors). Despite this, the literature rarely offers information about, or technical recommendations concerning, the appropriate statistical methodology to be used to accomplish this task. This paper suggests that when using linear regression, researchers use equivalence testing for standardized effect sizes. A simulation study is conducted to examine the type I error rates and statistical power of the tests, and a comparison is made with an alternative Bayesian testing approach. The results indicate that the proposed equivalence test is a potentially useful tool for "testing the null."

#### **KEYWORDS**

equivalence testing, non-inferiority testing, linear regression, standardized effect sizes  $\,$ 

# 1. Introduction

Let  $\theta$  be the parameter of interest. An equivalence test reverses the question that is asked in a null hypothesis significance test (NHST). Instead of asking whether we can reject the null hypothesis of no effect, e.g.,  $H_0: \theta = 0$ , an equivalence test examines whether the magnitude of  $\theta$  is at all meaningful: Can we reject the possibility that  $\theta$  is as large or larger than our smallest effect size of interest,  $\Delta$ ? The null hypothesis for an equivalence test is defined as  $H_0: \theta \notin [-\Delta, \Delta]$ . In other words, equivalence implies that  $\theta$  is small enough that any non-zero effect would be at most equal to  $\Delta$ . The interval  $[-\Delta, \Delta]$  is known as the equivalence margin and represents a range of values for which  $\theta$  is considered negligible. The value of  $\Delta$  is sometimes known as the "smallest effect size of interest" (Lakens, 2017). Note that the equivalence margin need not necessarily be symmetric, i.e., we could have  $H_0: \theta \notin [\Delta_1, \Delta_2]$ , where  $\Delta_1 \neq -\Delta_2$ .

In order for one to conduct an equivalence test, one must ideally define the equivalence margin prior to observing any data. This can often be challenging; see Campbell and Gustafson (2018b) for details. Indeed, for many researchers, defining and justifying the equivalence margin is one of the "most difficult issues" (Hung et al., 2005). If the margin is too large, then any claim of equivalence will be considered meaningless. If the margin is somehow too small, then the probability of declaring equivalence will be substantially reduced; see Wiens (2002). While the margin is ideally based on some objective criteria, these can be difficult to justify, and there is generally no clear consensus among stakeholders (Keefe et al., 2013).

To make matters worse, in many scenarios (and very often in the social sciences), the effects considered are measured on different and completely arbitrary scales. Without interpretable units of measurement, the task of defining and justifying an appropriate equivalence margin is even more challenging. How can one determine the "smallest effect size of interest" in units that have no particular meaning?

Researchers working with variables measured on arbitrary scales will typically report standardized effect sizes to aid with interpretation. For example, for linear regression analyses, reporting standardized regression coefficients is quite common (West

et al., 2007; Bring, 1994) and can be achieved by normalizing the outcome variable and all the predictors before fitting the regression. There are other reasons besides arbitrary scales for reporting standardized effects. For example, Nieminen et al. (2013) argues that standardized effect sizes might be helpful for the synthesis of epidemiological studies. And standardization can also help with interpretation: subtracting the mean can improve the interpretation of main effects in the presence of interactions, and dividing by the standard deviation will ensure that all predictors are on a common scale.

Unfortunately, equivalence testing of standardized effects is not straightforward. In this paper we introduce equivalence testing procedures for standardized effect sizes in a linear regression. We show how to define valid hypotheses and calculate p-values for these tests for two different cases: (1) with fixed regressors, and (2) with random regressors. In Section 3, we conduct a small simulation study to better understand the test's operating characteristics and in Section 4 we consider how a frequentist testing scheme compares to a Bayesian testing approach based on Bayes Factors. We conclude with practical recommendations in Section 6.

# 2. Equivalence testing for standardized $\beta$ coefficient parameter

Let us begin by defining some notation. All technical details are presented in the Appendix. Let:

- N, be the number of observations in the observed data;
- $\bullet$  K, be the number of explanatory variables in the linear regression model;
- $y_i$ , be the observed value of random variable Y for the ith subject;
- $x_{ki}$ , be the observed value of covariate  $X_k$ , for the *i*th subject, for k in 1, ..., K;
- X, be the N by K+1 covariate matrix (with a column of 1s for the intercept; we use the notation  $X_i$ , to refer to all K+1 values corresponding to the ith subject);

- $R_{Y \cdot X}^2$  is the coefficient of determination from the linear regression where Y is the dependent variable predicted from X;
- $R^2_{X_k \cdot X_{-k}}$  is the coefficient of determination from the linear regression model where  $X_k$  is the dependent variable predicted from the remaining K-1 regressors; and
- $R_{Y \cdot X_{-k}}^2$  is the coefficient of determination from the linear regression where Y is the dependent variable predicted from all but the kth covariate.

We operate under the standard linear regression assumption that observations in the data are independent and normally distributed with:

$$Y_i \sim Normal(X_{i,.}^T \beta, \sigma^2), \quad \forall i = 1, ..., N;$$
 (1)

where  $\beta$  is a parameter vector of regression coefficients, and  $\sigma^2$  is the population variance. Least squares estimates for the linear regression model are denoted with:  $\hat{y}_i$ ,  $\hat{\epsilon}_i$ ,  $\hat{\sigma}$ , and  $\hat{\beta}_k$ , for k in 0,...,K, and for i in 1,...,N; see Appendix for details.

A NHST for a specific variable,  $X_k$ ,  $(H_0: \beta_k = 0, \text{ vs.} \quad H_1: \beta_k \neq 0)$  is typically done with one of two different (yet mathematically identical) tests. Most commonly a t-test is done to calculate a p-value as follows:

$$p - \text{value}_k = 2 \cdot p_t \left( \frac{|\widehat{\beta_k}|}{\widehat{SE(\beta_k)}}, N - K - 1, 0 \right), \text{ for } k \text{ in } 0, \dots, K,$$
 (2)

where we use  $p_t(\cdot; df, ncp)$  to denote the cdf of the non-central t-distribution with df degrees of freedom and non-centrality parameter ncp; and where:  $\widehat{SE(\beta_k)} = \hat{\sigma}\sqrt{[(X^TX)^{-1}]_{kk}}$ . Note that when ncp = 0, the non-central t-distribution is equivalent to the central t-distribution. In R, we can obtain the p-values as follows:

```
## PART 1 ##
lmmod <- summary(lm(y~X[,-1]))
N <- length(y); K <- dim(X[,-1])[2]</pre>
```

```
beta_hat <- lmmod$coef[,1]; SE_beta_hat <- lmmod$coef[,2]
pval <- 2*pt(abs(beta_hat/SE_beta_hat), N-K-1, 0, lower.tail=FALSE)</pre>
```

Alternatively, we can conduct an F-test and, for k in 1,...,K, we will obtain the very same p-values with:

$$p - \text{value}_k = p_F \left( (N - K - 1) \frac{\text{diff} R_k^2}{1 - R_{Y \cdot X}^2}, 1, N - K - 1, 0 \right),$$
 (3)

where  $p_f(\cdot ; df_1, df_2, ncp)$  is the cdf of the non-central F-distribution with  $df_1$  and  $df_2$  degrees of freedom, and non-centrality parameter, ncp (note that ncp = 0 corresponds to the central F-distribution); and where:  $\text{diff} R_k^2 = R_{Y \cdot X}^2 - R_{Y \cdot X_{-k}}^2$ . In R, we can obtain these p-values as follows:

```
## PART 2 ##

R2 <- lmmod$r.squared

diffR2k <- unlist(lapply(c(2:(K+1)), function(k) {R2-summary(lm(y~X[,-k]))$r.squared}))

pval <- pf((N-K-1)*(diffR2k/(1-R2)), 1, N-K-1, 0, lower.tail=FALSE)</pre>
```

Regardless of whether the t-test or the F-test is employed, if p-value<sub>k</sub> <  $\alpha$ , we reject the null hypothesis of  $H_0: \beta_k = 0$  against the alternative  $H_0: \beta_k \neq 0$ .

An equivalence test asks a different question: Can we reject the possibility that  $\beta_k$  is as large or larger than our smallest effect size of interest? See Counsell and Cribbie (2015) who review equivalence testing procedures for linear regression coefficients. Formally, the null and alternative hypotheses for the equivalence test are:

```
H_0: \beta_k \le \Delta_1 \quad \text{or:} \quad \beta_k \ge \Delta_2,

H_1: \beta_k > \Delta_1 \quad \text{and:} \quad \beta_k < \Delta_2,
```

where the equivalence margin is  $[\Delta_1, \Delta_2]$  and defines the range of values considered negligible. Often one has a symmetric margin with  $\Delta_1 = -\Delta_2$ , but this is not necessarily the case.

Recall that there is a one-to-one correspondence be between an equivalence test and a confidence interval Wellek (2017). For example, we will reject the above  $H_0$  at

a  $\alpha = 0.05$  significance level if and only if the 90% (=1-2 $\alpha$ ) CI for  $\beta_k$  fits entirely within  $[\Delta_1, \Delta_2]$ . As such an equivalence test can be constructed by simply inverting a confidence interval. To obtain a p-value for the equivalence test one considers two one-sided t-tests (TOST) and calculates the two following p-values:

$$p_k^{[1]} = p_t \left( \frac{\widehat{\beta_k} - \Delta_1}{\widehat{SE(\beta_k)}}, N - K - 1, 0 \right); \quad \text{and} \quad p_k^{[2]} = p_t \left( \frac{\Delta_2 - \widehat{\beta_k}}{\widehat{SE(\beta_k)}}, N - K - 1, 0 \right), \tag{4}$$

for k in 0,...,K. In order to reject this equivalence test null hypothesis, both p-values,  $p_k^{[1]}$  and  $p_k^{[2]}$ , must be less than  $\alpha$ . As such, a p-value for the equivalence test is calculated as: p-value $_k = max(p_k^{[1]}, p_k^{[2]})$ . In R, we can obtain this p-value as follows:

```
## PART 3 ##
p1[k] <- pt((beta_hat[k] - DELTA[k,1])/SE_beta_hat[k], N-K-1, 0, lower.tail=FALSE)
p2[k] <- pt((-beta_hat[k] + DELTA[k,2])/SE_beta_hat[k], N-K-1, 0, lower.tail=FALSE)
pval[k] <- max(c(p1[k],p2[k]))</pre>
```

# 2.1. An equivalence test for standardized regression coefficients

Unfortunately, in many scenarios (and very often in the social sciences), the variables considered are measured on different and completely arbitrary scales. Without interpretable units of measurement, defining (and justifying) the equivalence margin can be rather challenging. How can one determine the "smallest effect size of interest" (Lakens, 2017) in units that have no particular meaning? In these scenarios, it may be preferable to work with standardized regression coefficients.

The process of standardizing a regression coefficient can proceed by multiplying the unstandardized regression coefficient,  $\beta_k$ , by the ratio of the standard deviation of  $X_k$  to the standard deviation of Y. The population standardized regression coefficient parameter,  $\mathcal{B}_k$ , for k in 1,...,K, is defined as:

$$\mathcal{B}_k = \beta_k \frac{\sigma_{X_k}}{\sigma_Y},\tag{5}$$

and can be estimated by:

$$\widehat{\mathcal{B}_k} = \widehat{\beta_k} \frac{\widehat{\sigma_{X_k}}}{\widehat{\sigma_Y}},\tag{6}$$

where  $\widehat{\sigma_{X_k}}$  and  $\widehat{\sigma_Y}$  are the observed standard deviations of  $X_k$  and Y, respectively. We can obtain the estimated standardized regression coefficients in R as follows:

#### ## PART 4 ##

 $b_{\text{vec}} \leftarrow (\text{beta\_hat*}(\text{apply}(X,2,\text{sd})/\text{sd}(y)))[-1]$ 

An equivalence test for  $\mathcal{B}_k$  can be defined by the following null and alternative hypotheses:

$$H_0: \mathcal{B}_k \leq \Delta_1$$
 or:  $\mathcal{B}_k \geq \Delta_2$ ,

$$H_1: \mathcal{B}_k > \Delta_1$$
 and:  $\mathcal{B}_k < \Delta_2$ .

By inverting a confidence interval for  $\mathcal{B}_k$  (see Kelley et al. (2007) for details), we can obtain the following, for k in 1,...,K:

$$p_k^{[1]} = p_t \left( \frac{\widehat{\mathcal{B}_k}}{\widehat{SE(\mathcal{B}_k)_{FIX}}}; df = N - K - 1, ncp = \Delta_1 \frac{\sqrt{N\left(1 - R_{X_k \cdot X_{-k}}^2\right)}}{\sqrt{\left(1 - R_{Y \cdot X}^2\right)}} \right), \text{ and:}$$

$$(7)$$

$$p_k^{[2]} = p_t \left( \frac{-\widehat{\mathcal{B}}_k}{\widehat{SE(\mathcal{B}_k)}_{FIX}}; df = N - K - 1, ncp = -\Delta_2 \frac{\sqrt{N\left(1 - R_{X_k \cdot X_{-k}}^2\right)}}{\sqrt{\left(1 - R_{Y \cdot X}^2\right)}} \right), \quad (8)$$

where:

$$\widehat{SE(\mathcal{B}_k)}_{FIX} = \sqrt{\frac{(1 - R_{Y \cdot X}^2)}{(1 - R_{X_k \cdot X_{-k}}^2)(N - K - 1)}}.$$
(9)

In order to reject this equivalence test null hypothesis, p-value<sub>k</sub> =  $max(p_k^{[1]}, p_k^{[2]})$  must be less than  $\alpha$ . We can obtain this p-value in  $\mathbb{R}$  with the following code:

This calculation assumes that the covariates, X, are not stochastic, i.e., the covariates are fixed in advance by the researcher. When X is random (i.e., randomly sampled from a larger population of interest) the sampling distribution of  $\mathcal{B}_k$  can be substantially different. In the social sciences, the assumption of fixed regressors is often violated and therefore it is important to consider this possibility (Bentler and Lee, 1983).

Yuan and Chan (2011) derive an estimator for the standard error of  $\mathcal{B}_k$  which takes into account the additional variance in  $\mathcal{B}_k$  that exists as a result of the regressors being random (see Yuan and Chan (2011) eq. 23). This estimator,  $\widehat{SE(\mathcal{B}_k)}_{RDM}$ , is based on central limit theorem and the delta-method (see Appendix for details and derivation). Jones and Waller (2013) suggest (based on a simulation study) using  $\widehat{SE(\mathcal{B}_k)}_{RDM}$  to construct confidence intervals for  $\mathcal{B}_k$ . Following the same logic, we can make use of  $\widehat{SE(\mathcal{B}_k)}_{RDM}$  to calculate a p-value for our equivalence test when regressors are random. We have p-value=  $\max(p_k^{[1]}, p_k^{[2]})$ , where:

$$p_k^{[1]} = p_t \left( \frac{\widehat{\mathcal{B}}_k - \Delta_1}{\widehat{SE(\mathcal{B}_k)}_{RDM}}, df = N - K - 1, ncp = 0 \right), \text{ and:}$$
 (10)

$$p_k^{[2]} = p_t \left( \frac{\Delta_2 - \widehat{\mathcal{B}_k}}{\widehat{SE(\widehat{\mathcal{B}_k})_{RDM}}}, df = N - K - 1, ncp = 0 \right).$$
(11)

We can obtain the above p-value in R with the following code:

```
## PART 6 ##
SE_std_beta_RDM <- DEL(X=X[,-1], y=y)$SEs
p1[k] <- pt((b_vec[k] - DELTA[k,1])/SE_std_beta_RDM[k], N-K-1, 0, lower.tail=FALSE)
p2[k] <- pt((DELTA[k,2] - b_vec[k])/SE_std_beta_RDM[k], N-K-1, 0, lower.tail=FALSE)
pval[k] <- max(c(p1[k], p2[k]))</pre>
```

# 2.2. An equivalence test for the increase in the squared multiple correlation coefficient

The increase in the squared multiple correlation coefficient associated with adding a variable in a linear regression model, diff  $R_k^2$ , is a commonly used measure for establishing the importance the added variable. In a linear regression model, the  $R^2$  is equal to the square of the Pearson correlation coefficient between the observed and predicted outcomes (Nagelkerke et al., 1991; Zou et al., 2003). Despite the  $R^2$  statistic's ubiquitous use, its corresponding population parameter, which we will denote as  $P^2$ , as in Cramer (1987), is rarely discussed. When considered, it is sometimes is known as the "parent multiple correlation coefficient" (Barten, 1962) or the "population proportion of variance accounted for" (Kelley et al., 2007); see Cramer (1987) for a technical discussion. Campbell and Lakens (2020) introduce a non-inferiority test (a one-sided equivalence test) for  $R_{Y \cdot X}^2$  in order to test the null hypotheses  $H_0: P_{Y \cdot X}^2 \geq \Delta$  vs.  $H_1: P_{Y \cdot X}^2 < \Delta$ . Things are slightly different for testing diff  $P_k^2$ .

Note that the diff  $R_k^2$  measure is simply a re-calibration of  $\widehat{\mathcal{B}}_k$ , such that:

$$\operatorname{diff} R_k^2 = \widehat{\mathcal{B}_k}^2 (1 - R_{X_k \cdot X_{-k}}^2). \tag{12}$$

Similarly, we have that, for the corresponding population parameter:  $\operatorname{diff} P_k^2 = \mathcal{B}_k^2 (1 - P_{X_k \cdot X_{-k}}^2)$ . It may be preferable to consider the effect sizes (and what should be considered a "negligible difference") in terms of  $\operatorname{diff} P_k^2$  instead of in terms of  $\mathcal{B}_k$ . If this is the case, one can conduct a non-inferiority test, for k in 1,...,K, with the following hypotheses:

$$H_0: \mathrm{diff} P_k^2 \geq \Delta,$$

$$H_1: 0 \le \operatorname{diff} P_k^2 < \Delta.$$

The *p*-value for this non-inferiority test is obtained by replacing  $\mathcal{B}_k$  with  $\sqrt{\text{diff}P_k^2/(1-P_{X_k\cdot X_{-k}}^2)}$  and can be calculated, for fixed regressors as follows:

$$p - \text{value}_k = 1 - p_t \left( \frac{\sqrt{(N - K - 1) \text{diff} R_k^2}}{\sqrt{(1 - R_{Y \cdot X}^2)}}; df = N - K - 1, ncp = \frac{\sqrt{N\Delta}}{\sqrt{(1 - R_{Y \cdot X}^2)}} \right).$$
(13)

One can calculate the above p-value in R with the following code:

## PART 7 ##

 $pval[k] \leftarrow pt(sqrt((N-K-1)*diffR2k[k])/sqrt(1-R2), N-K-1, sqrt(N*DELTA[k])/sqrt(1-R2), lower.tail=TRUE)$ 

For random regressors we have, for k in 1,...,K:

$$p - \text{value}_k = 1 - p_t \left( \frac{\sqrt{\text{diff}R_k^2} - \sqrt{\Delta}}{\widehat{SE(\mathcal{B}_k)}_{RDM} \sqrt{(1 - R_{X_k \cdot X_{-k}}^2)}}, df = N - K - 1, ncp = 0 \right). \tag{14}$$

The above p-value is calculated in R with the following code:

# 2.3. Simulation Study 1

We conducted a simple simulation study in order to better understand the operating characteristics of the proposed equivalence test for standardized regression coefficients and to confirm that the test has correct type 1 error rates. The test in the simulation study considers the following equivalence test with a symmetric equivalence margin (i.e., equiv. margin =  $[-\Delta, \Delta]$ ):

 $H_0: |\mathcal{B}_1| \geq \Delta,$ 

 $H_1: |\mathcal{B}_1| < \Delta.$ 

We simulated data for each of 48 scenarios  $(2 \times 4 \times 2 \times 3)$ , one for each combination of the following parameters:

- either fixed or random regressors;
- one of four sample sizes: N = 180, N = 540, N = 1,000, or N = 3,500;
- one of two designs with K=2, or K=4 binary covariates; with  $\beta=(-0.2,0.1,0.2)$  or  $\beta=(0.2,0.1,0.14,-0.1,-0.1)$ ; and
- one of three variances:  $\sigma^2=0.05$  ,  $\sigma^2=0.15$ , or  $\sigma^2=0.5$ .

Note that for the scenarios with "fixed regressors," the covariates are set such that we have an orthogonal, balanced design. With "random regressors," each possible covariate value is chosen at random with equal likelihood. For scenarios with "fixed regressors," we calculated the equivalence test p-value using the formula for fixed regressors and for scenarios with "random regressors," we calculated the equivalence test p-value using the formula for random regressors (equation 11).

Depending on the specific value of  $\sigma^2$ , the true population standardized coefficient,  $\mathcal{B}_1$ , for these data is either: 0.07, 0.12, or 0.20. In order to examine situa-

tions with  $\mathcal{B}_1 = 0$ , we also simulated data from an additional 16 scenarios where the regression coefficients were fixed to be  $\beta = (-0.2, 0.0, 0.2)$ , for K = 2, and  $\beta = (0.2, 0.0, 0.14, -0.1, -0.1)$ , for K = 4. For all of these additional scenarios,  $\sigma^2$  was set equal to 0.5.

Parameters for the simulation study were chosen so that we would consider a wide range of values for the sample size (representative of those sample sizes commonly used in the psychology literature; see Kühberger et al. (2014), Fraley and Vazire (2014), and Marszalek et al. (2011)). We also wished to obtain three unique values for  $\mathcal{B}_1$  approximately evenly spaced between 0 and 0.20.

For each of the total 64 (=48+16) configurations, we simulated 50,000 unique datasets and calculated an equivalence p-value with each of 49 different values of  $\Delta$  (ranging from 0.01 to 0.25). We then calculated the proportion of these p-values less than  $\alpha = 0.05$ . We specifically chose to conduct 50,000 simulation runs so as to keep computing time within a reasonable limit while also reducing the amount of Monte Carlo standard error to a negligible amount (for looking at type 1 error with  $\alpha = 0.05$ , Monte Carlo SE will be approximately  $0.001 \approx \sqrt{0.05(1-0.05)/50,000}$ ); see Morris et al. (2019).

## 2.3.1. Simulation Study 1 Results-

Figure 1 and Figure 2 show results for the simulations with fixed regressors. In the Appendix, Figure 3 and Figure 4 show results for the simulations with random regressors. We see no substantial difference between the simulation study results with fixed and random regressors.

When  $\mathcal{B}_1 = 0.20$ , we see that when the equivalence bound  $\Delta$  equals the true effect size (i.e., when  $\mathcal{B}_1 = \Delta = 0.20$ ), the type 1 error rate is exactly 0.05, as it should be, for all N. This situation represents the boundary of the null hypothesis. As the equivalence bound increases beyond the true effect size (i.e.,  $\Delta > \mathcal{B}_1$ ), the alternative hypothesis is then true and it is more and more likely we will correctly conclude equivalence.

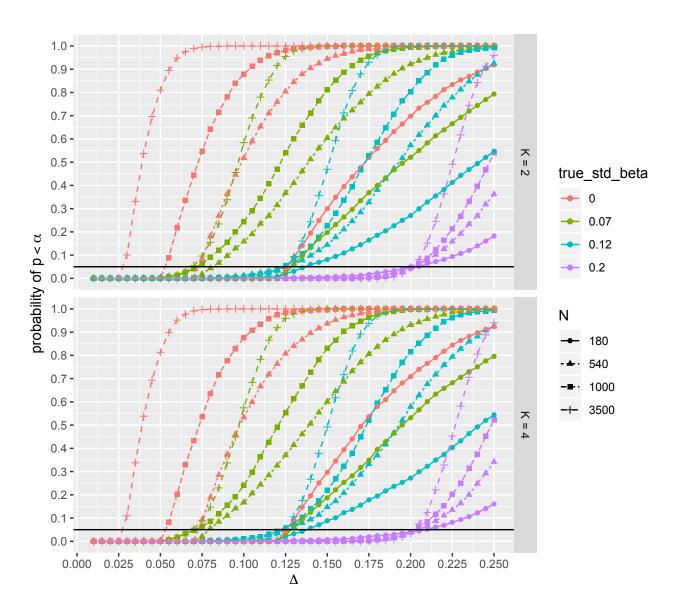


Figure 1. Simulation Study 1 - complete results for "fixed regressors." Upper panel shows results for K=2; Lower panel shows results for K=4. The solid horizontal black line indicates the desired type 1 error of  $\alpha=0.05$ .

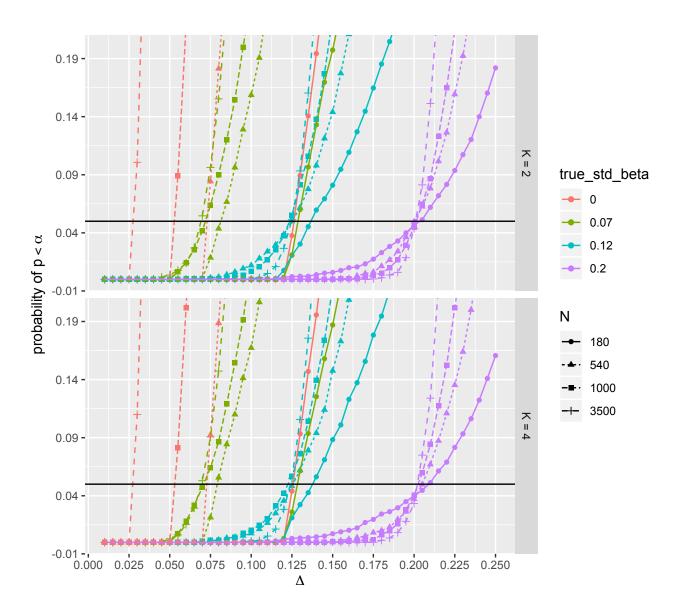


Figure 2. Simulation Study 1 - results for "fixed regressors". Upper panel shows results for K=2; lower panel shows results for K=4. Both plots are presented with a restricted vertical-axis to better show the type 1 error rates. The solid horizontal black line indicates the desired type 1 error of  $\alpha=0.05$ .

For smaller values of  $\mathcal{B}_1$  (i.e., for  $\mathcal{B}_1 = 0.07$  and  $\mathcal{B}_1 = 0.12$ ), when the equivalence bound equals the true effect size (i.e., when  $\mathcal{B}_1 = \Delta$ ), the test is somewhat conservative, particularly for small N. Even when  $\Delta > \mathcal{B}_1$ , the equivalence test may reject the null hypothesis for less than 5% of cases. For example, when N = 180,  $\mathcal{B}_1 = 0.12$  and K = 2, the rejection rate is only 0.025 when  $\Delta = 0.125$ . This is due to the fact that with a small N, the sampling variance of  $\widehat{\mathcal{B}}_1$  will be far too large to reject  $H_0: |\mathcal{B}_1| \geq \Delta$ . Consider that, when N is small and  $\sigma^2$  is relatively large, the 90% CI for  $\mathcal{B}_1$  may be far too wide to fit entirely within the equivalence margin.

In order for the test to have any substantial power,  $\mathcal{B}_1$  must be substantially smaller than  $\Delta$ . Also, as expected, the power of the test increases with larger values of  $\Delta$ , larger values of N, and smaller values of K. In some cases, the power is strictly zero. For example, when  $\mathcal{B}_1 = 0.00$ , N = 180 and K = 2, the  $\Delta$  must be greater or equal to 0.1 for there to any possibility of rejecting  $H_0$ . Otherwise, for  $\Delta < 0.10$ , the power is zero; see Figure 2.

## 3. Comparison to a Bayesian alternative

# 3.1. Conditional equivalence testing

Ideally, a researcher uses the non-inferiority test to examine a preregistered hypothesis concerning the absence of a meaningful effect. However, in practice, one might first conduct a NHST (i.e., calculate a p-value,  $p_1$ , using equation (2) or (3) and only proceed to conduct the equivalence test (i.e., calculate a second p-value,  $p_2$ , using equation (??) or (11)) if the NHST fails to reject the null. Such a two-stage sequential testing scheme has recently been put forward by Campbell and Gustafson (2018a) under the name of "conditional equivalence testing" (CET). Under the proposed CET scheme, if the first p-value,  $p_1$ , is less than the type 1 error  $\alpha$ -threshold (e.g., if  $p_1 < 0.05$ ), one concludes with a "positive" finding:  $\mathcal{B}_k$  is significantly greater than 0. On the other hand, if the first p-value,  $p_1$ , is greater than  $\alpha$  and the second p-value,  $p_2$ , is smaller than  $\alpha$  (e.g., if  $p_1 \ge 0.05$  and  $p_2 < 0.05$ ), one concludes with a "negative" finding: there is evidence of

a statistically significant equivalence, i.e.,  $\mathcal{B}_k$  is at most negligible. If both p-values are large, the result is inconclusive: there are insufficient data to support either finding. In this paper, we are not advocating for (or against) CET but simply use it to facilitate a comparison with Bayes Factor testing (which also categorizes outcomes as either positive, negative or inconclusive).

## 3.2. Bayes Factor testing for linear regression

For linear regression models, based on the work of Liang et al. (2008), Rouder and Morey (2012) propose using Bayes Factors (BFs) to determine whether the data support the inclusion of a particular variable in the model. This is a common approach used in psychology studies (e.g., see the tutorial of Etz (2015)). Here we refer to the null model ("Model 0") and alternative model ("Model 1") as:

Model 
$$0: Y_i \sim Normal(X_{i,-k}^T \beta_{-k}, \sigma^2), \quad \forall i = 1, ..., N;$$
 (15)

Model 
$$1: Y_i \sim Normal(X_{i,\cdot}^T \beta, \sigma^2), \quad \forall i = 1, ..., N;$$
 (16)

where  $\beta_{-k}$  ( $X_{i,-k}$ ) is the vector (matrix) of regression coefficients (covariates), with the kth coefficient (covariate) omitted.

We define the Bayes Factor,  $BF_{10}$ , as the probability of the data under the alternative model relative to the probability of the data under the null:

$$BF_{10} = \frac{Pr(Data \mid Model \ 1)}{Pr(Data \mid Model \ 0)},\tag{17}$$

with the "10" subscript indicating that the alternative model (i.e., "Model 1") is being compared to the null model (i.e., "Model 0"). The BF can be easily interpreted. For example, a  $BF_{10}$  equal to 0.20 indicates that the null model is five times more likely

than the alternative model.

Bayesian methods require one to define appropriate prior distributions for all model parameters. Rouder and Morey (2012) suggest using "objective priors" for linear regression models and explain in detail how one may implement this approach. We will not discuss the issue of prior specification in detail, and instead point interested readers to Consonni et al. (2008) who provide an in-depth overview of how to specify prior distributions for linear models.

Using the BayesFactor package in R (Morey et al., 2015) with the function regressionBF(), one can easily obtain a BF corresponding to Y and X. (See also the baymedr package in R (Linde and van Ravenzwaaij, 2019)). Since we can also calculate frequentist p-values corresponding to values for Y and X, the frequentist and Bayesian approaches can be compared in a relatively straightforward way. We will explore this in Simulation Study 2.

#### 3.3. Simulation Study 2

We wish to compare a frequentist testing scheme based on NHST and equivalence testing to the Bayesian approach based on BFs by means of a simple simulation study. How often will the frequentist and Bayesian approaches arrive at the same conclusion?

Frequentist conclusions were based on the CET procedure and by setting  $\Delta$  equal to either 0.05, or 0.10, or 0.25; and with  $\alpha$ =0.05. Bayesian conclusions were based on an evidence threshold of either 3, 6, or 10. A threshold of 3 can be considered "substantial evidence," a threshold of 6 can be considered "strong evidence," and a threshold of 10 can be considered "very strong evidence" (Wagenmakers et al., 2011). Note that for the simulation study here we examine only the "fixed-n design" for BF testing; see Schönbrodt and Wagenmakers (2016) for details. Also note that, as in Section 3, all priors required for calculating the BF were set by simply selecting the default settings of the regressionBF() function (with rscaleCont = "medium"); see Morey et al. (2015).

We simulated datasets for 48 unique scenarios. For this second simulation study,

we only considered the case of fixed regressors. We varied over the following:

- one of twelve sample sizes: N = 20, N = 33, N = 55, N = 90, N = 149, N = 246, N = 406, N = 671, N = 1,109, N = 1,832, N = 3,027, or N = 5,000;
- one of two designs with K=4 binary covariates (with an orthogonal, balanced design), with either  $\beta=(0.2,0.1,0.14,-0.1,-0.1)$  or  $\beta=(0.2,0.0,0.14,-0.1,-0.1)$ ;
- one of two variances:  $\sigma^2 = 0.5$ , or  $\sigma^2 = 1.0$ .

Note that for the  $\beta = (0.2, 0.0, 0.14, -0.1, -0.1)$  design, we only consider one value for  $\sigma^2 = 1.0$ . Depending on the particular design and  $\sigma^2$ , the true standardized regression coefficient,  $\mathcal{B}_1$ , for these data is either  $\mathcal{B}_1 = 0.00$ ,  $\mathcal{B}_1 = 0.05$ , or  $\mathcal{B}_1 = 0.07$ .

#### 3.3.1. Simulation Study 2 Results-

For each simulated dataset, we obtained frequentist p-values, BFs and declared the result to be positive, negative or inconclusive accordingly. Results are presented in Figures XX, XX and XX and are based on 5,000 distinct simulated datasets per scenario.

We are also interested in how often the two approaches will reach the same overall conclusion. Table 3.3.1 displays the the average rate of agreement between the Bayesian and frequentist methods. Averaging over all 48 scenarios, how often on average will the Bayesian and frequentist approaches reach the same conclusion given the same data?

	BF=3	BF=6	BF=10
$\Delta = 0.25$	0.77	0.54	0.44
$\Delta = 0.10$	0.74	0.82	0.72
$\Delta = 0.05$	0.63	0.79	0.84

**Table 1.** Averaging over all  $\overline{XX}$  scenarios and over all 5,000 Monte Carlo simulations per scenario, how often on average did the Bayesian and frequentist approaches reach the same conclusion? Numbers in the above table represent the average proportion of simulated datasets (averaged over  $XX \times 5,000 = XX0,000$  unique datasets) for which the Bayesian and frequentist methods arrive at the same conclusion.

Three observations merit comment:

• The Bayesian testing scheme is always less likely to deliver a positive conclusion

(see how the dashed blue curve is always higher than the solid blue curve). In the scenarios like the ones we considered, the BF may require larger sample sizes for reaching a positive conclusion and thus may be considered "less powerful" in a traditional frequentist sense.

- With Δ = 0.10 or Δ = 0.25, and a BF threshold of 6 or 10, the BF testing scheme requires substantially more data to reach a negative conclusion than the frequentist scheme. Note that, the probability of reaching a negative result with CET will never exceed 0.95 since the NHST is performed first (before the equivalence test) and will typically reach a positive result with probability of at least 1 α; see dashed orange lines in Figures XX, XX, and XX panels XX, XX, and XX.
- While the JZS-BF requires less data to reach a conclusive result when the sample size is small (see how the solid black curve drops more rapidly than the dashed grey line), there are scenarios in which larger sample sizes will surprisingly reduce the likelihood of the BF obtaining a conclusive result (see how the solid black curve drops abruptly then rises slightly as n increases for  $\mathcal{B}_1 = 0.05$ , and 0.07; see for example, Figure XX ?? panels 4 and 5).

Based on our comparison of BFs and frequentist tests, we can confirm that, given the same data, both approaches will often provide one with the same overall conclusion. The level of agreement however is highly sensitive to the choice of  $\Delta$  and the choice of the BF evidence threshold, see Table 3.3.1. The highest level of agreement, recorded at 0.84, is when  $\Delta = 0.05$  and the BF evidence threshold is equal to 10. In contrast, when  $\Delta = 0.025$  and the BF evidence threshold is 10, the two approaches will deliver the same conclusion less than half of the time. Table 3.3.1 shows that the two approaches rarely arrive at entirely contradictory conclusions. In less than XX% of cases, did we observe one approach arrive at a positive conclusion while the other approach arrived at a negative conclusion when faced with the same exact data.

This simulation study result is reassuring since it suggests that the conclusions obtained from frequentist and Bayesian testing will rarely lead to substantial disagreements.

	BF=3	BF=6	BF=10
$\Delta = 0.25$	0.XX	0.XX	0.XX
$\Delta = 0.10$	0.XX	0.XX	0.XX
$\Delta = 0.05$	0.XX	0.XX	0.XX

**Table 2.** Averaging over all  $\overline{XX}$  scenarios and over all 5,000 Monte Carlo simulations per scenario, how often on average did the Bayesian and frequentist approaches strongly disagree in their conclusion? Numbers in the above table represent the average proportion of simulated datasets (averaged over  $XX \times 5,000 = XX0,000$  unique datasets) for which the Bayesian and frequentist methods arrived at completely opposite (one positive and one negative) conclusions.

## 4. Conclusion

There is a great risk of bias in the scientific literature if researchers only rely on statistical tools that can reject null hypotheses, but do not have access to statistical tools that allow them to reject the presence of meaningful effects; see (Amrhein et al., 2019); see also Altman and Bland (1995). Equivalence tests provide one approach to improve current research practices by allowing researchers to falsify their predictions concerning the presence of an effect.

In this paper we presented an equivalence test for standardized effect sizes in linear regression analyses. We also considered how frequentist equivalence testing offers an attractive alternative to Bayesian methods for "testing the null." We believe that equivalence testing for standardized effect sizes can help researchers conduct equivalence tests by facilitating what is often a very challenging task: defining an appropriate equivalence margin. While the use of "default equivalence margins" based on standardized effect sizes cannot be whole-heartily recommended for all cases, their use is not unlike the use of "default priors" for Bayesian inference which have proven useful for researchers in certain scenarios.

Note that our the non-inferiority test for the increase in the squared multiple correlation coefficient  $(\operatorname{diff} P_k^2)$  in a standard multivariable linear regression is limited to comparing two models for which the difference in degrees of freedom is 1. In other words, the test is not suitable for comparing two nested models where the difference is more than a single variable. For example, we cannot use the test to compare a "smaller model" with only "income" as a covariate, with a "larger model" that includes "income," "age" and "gender," as covariates. A more general equivalence test for

comparing two nested models will be addressed in future work. We also wish to develop equivalence testing for standardized regression coefficients in logistic regression models.

# 5. Appendix

Least squares estimates for the linear regression model are:

- $\widehat{\boldsymbol{\beta}}_k = ((X^TX)^{-1}X^Ty)_k$ , for k in 1,..., K;
- $\hat{y}_i = X_{i,\cdot}^T \hat{\beta}$ , for i in 1,..., N;
- $\hat{\epsilon}_i = \hat{y}_i y_i$ , for i in 1,..., N; and
- $\hat{\sigma} = \sqrt{\sum_{i=1}^{N} (\hat{\epsilon}_i^2)/(N-K-1)}$ .

# 5.1. R| - code

```
library(RCurl)
script <- getURL(</pre>
"https://raw.githubusercontent.com/harlanhappydog/EquivTestStandardReg/master/EquivTestStandardReg.R",
ssl.verifypeer = FALSE)
eval(parse(text = script))
## Example data: ##
set.seed(123)
y <- rnorm(100)
X <- cbind(1, rnorm(100), rpois(100,4))</pre>
## PART 1 ##
lmmod <- summary(lm(y~X[,-1]))</pre>
N \leftarrow length(y); K \leftarrow dim(X[,-1])[2]
beta_hat <- lmmod$coef[,1]; SE_beta_hat <- lmmod$coef[,2]</pre>
pval <- 2*pt(abs(beta_hat/SE_beta_hat), N-K-1, 0, lower.tail=FALSE)</pre>
pval
# alternatively:
```

```
summary(lm(y^X[,-1]))$coef[,4]
## PART 2 ##
R2 <- lmmod$r.squared
\label{eq:diffR2k} $$ $$ \dim(x^* X[-k]) = \min(x^* X[-k]) $$ r.squared) $$ if $R^2 - \sup(x^* X[-k]) $$ r.squared) $$ if $R^2 - \sup(x^* X[-k]) $$ r.squared $$ if $R^2 - \sup(x^* X[-k]) $$ r.squared $$ if $R^2 - \sup(x^* X[-k]) $$ if $R^2 - 
\label{eq:pval} $$\operatorname{pr}((N-K-1)*(diffR2k/(1-R2)), 1, N-K-1, 0, lower.tail=FALSE)$$
pval
# alternatively:
## PART 3 ##
DELTA <- rbind(c(-0.3, 0.25), c(-0.2, 0.2), c(-0.3, 0.1))
pval <- p1 <- p2 <- rep(0, K+1)</pre>
for(k in 1:(K+1)){
p1[k] <- pt((beta_hat[k] - DELTA[k,1])/SE_beta_hat[k], N-K-1, 0, lower.tail=FALSE)</pre>
\label{eq:p2k} \texttt{p2[k]} \ \leftarrow \ \texttt{pt((-beta\_hat[k] + DELTA[k,2])/SE\_beta\_hat[k], N-K-1, 0, lower.tail=FALSE)}
pval[k] <- max(c(p1[k],p2[k]))</pre>
pval
# alternatively:
equivBeta(Y = y, Xmatrix = X[,-1], DELTA = DELTA)$pval
## PART 4 ##
b_vec <- (beta_hat*(apply(X,2,sd)/sd(y)))[-1]</pre>
b_vec
# alternatively:
lm(scale(y) ~ scale(X[,-1]))$coef[-1]
## PART 5 ##
DELTA <- rbind(c(-0.2, 0.2), c(-0.3, 0.1))
SE\_beta\_FIX \leftarrow R2YdotX \leftarrow R2XkdotXminK \leftarrow pval \leftarrow p1 \leftarrow p2 \leftarrow rep(0, K)
for(k in 1:K){
\label{eq:r2XkdotXminK[k] <- (summary(lm(X[,-1][,k]~X[,-1][,-k])))$r.squared} % \begin{center} \begin{center}
R2YdotX[k]
                                                                          <- (summary(lm(y~X[,-1])))$r.squared
SE\_beta\_FIX[k] \  \  <- \  \, sqrt(\  \, (1-R2YdotX[k])/(\  \, (1-R2XkdotXminK[k])*(N-K-1) \  \, ) \, \, )
p1[k] <- pt(b_vec[k]/SE_beta_FIX[k], N-K-1,</pre>
```

```
DELTA[k,1]*sqrt(N*(1-R2XkdotXminK[k]))/sqrt(1-R2YdotX[k]), lower.tail=FALSE)
p2[k] <- pt(-b_vec[k]/SE_beta_FIX[k], N-K-1,</pre>
                -DELTA[k,2]*sqrt(N*(1-R2XkdotXminK[k]))/sqrt(1-R2YdotX[k]), lower.tail=FALSE)
pval[k] <- max(c(p1[k], p2[k]))</pre>
pval
# alternatively:
equivstandardBeta(Y = y, Xmatrix = X[,-1], DELTA = DELTA, random = FALSE)$pval
## PART 6 ##
SE_std_beta_RDM <- DEL(X=X[,-1], y=y)$SEs</pre>
pval <- p1 <- p2 <- rep(0, K)</pre>
for(k in 1:K){
p1[k] \leftarrow pt((b_vec[k] - DELTA[k,1])/SE_std_beta_RDM[k], N-K-1, 0, lower.tail=FALSE)
p2[k] <- pt((DELTA[k,2] - b_vec[k])/SE_std_beta_RDM[k], N-K-1, 0, lower.tail=FALSE)
pval[k] \leftarrow max(c(p1[k], p2[k]))
}
pval
# alternatively:
equivstandardBeta(Y = y, Xmatrix = X[,-1], DELTA = DELTA, random = TRUE)$pval
## PART 7 ##
DELTA <- rep(0.05,2)
pval <- rep(0, K)</pre>
for(k in 1:K){
pval[k] \leftarrow pt(sqrt((N-K-1)*diffR2k[k])/sqrt(1-R2), N-K-1, sqrt(N*DELTA[k])/sqrt(1-R2), lower.tail=TRUE)
}
pval
# alternatively:
equivdiffP2(Y = y, Xmatrix = X[,-1], DELTA = DELTA, random = FALSE)pval
## PART 8 ##
DELTA <- rep(0.05,2)
pval <- rep(0, K)</pre>
for(k in 1:K){
pval[k] \leftarrow pt((sqrt(diffR2k[k]) - sqrt(DELTA[k])) / (SE_std_beta_RDM[k] * sqrt(1-R2XkdotXminK[k])),
N-K-1, lower.tail=TRUE)
```

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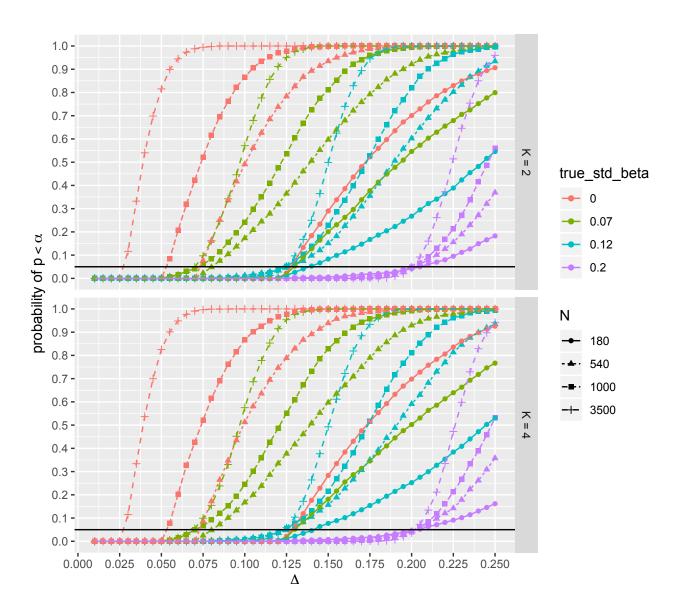


Figure 3. Simulation Study 1 - complete results for "random regressors." Upper panel shows results for K=2; Lower panel shows results for K=4. The solid horizontal black line indicates the desired type 1 error of  $\alpha=0.05$ .

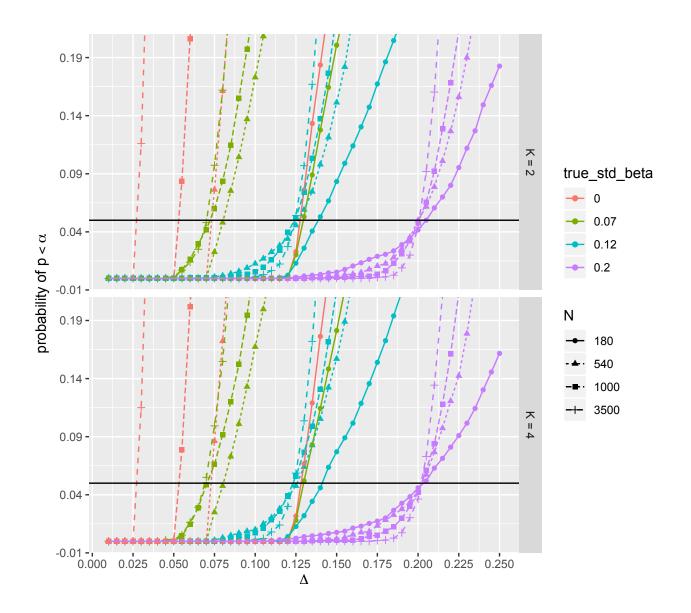


Figure 4. Simulation Study 1 - results for "random regressors". Upper panel shows results for K=2; lower panel shows results for K=4. Both plots are presented with a restricted vertical-axis to better show the type 1 error rates. The solid horizontal black line indicates the desired type 1 error of  $\alpha=0.05$ .

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