Bayesian Joint Models, with Application to HIV Data (R Code)

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### 1. Data Processing

```
hiv <- read_csv("~/Desktop/UBC/project/rebound_data.csv")</pre>
dat0 <- hiv %>%
  drop_na(RNA_V, CD4_V) %>%
  group_by(PATIENT) %>%
 mutate(first=as.numeric(row_number()==min(row_number()[RNA_V==-1]))) %>%
 filter(first==1)
dat1 <- hiv %>%
  drop_na(RNA_V, CD4_V) %>%
  filter(RNA_V!=-1) %>%
  full_join(dat0) %>%
  arrange(PATIENT, days_from_seroco)
dat <- dat1 %>%
  mutate(RNA_V=if_else(RNA_V==-1, log10(RNA_L/2), log10(RNA_V)),
         CD4_V=log(CD4_V),
         months_from_seroco=days_from_seroco/30,
         art_start=art_start/30,
         ati start=ati start/30) %>%
  dplyr::select(-RNA_T, -RNA_L, -days_from_seroco, -first) %>%
  filter(is.na(months_from_seroco) == FALSE)
```

### 2. Define Functions

```
# 2.1 Find RNA_V viral rebound
Find_rebound <- function(rna) {
    # Find the index of viral rebound. If there is no rebound yet, then return 0.
    if (length(rna)==1) ind <- 0
    else {
        for (i in 2:length(rna)) {
            if (rna[i] > rna[i-1]) {ind <- i; break}
            else {ind <- 0}
        }
    }
    return(ind)
}</pre>
```

```
# 2.2 Find RNA_V peak points
Find_peak <- function(rna) {</pre>
  # Find the index of viral peak point. If there is no peak yet, then return 0.
  ind <- which.max(rna)</pre>
  if (ind == length(rna)) ind <- 0
  return(ind)
}
# 2.3 Define `status` of viral rebound/viral peak
Define_status <- function(fn, rna) {</pre>
  # Define the status: 1 - there is a rebound or peak; 0 - there is no rebound or peak
  # Returns `status` and `ind`
  ind <- fn(rna)
  if (ind!=0) status <- 1</pre>
  else {status <- 0; ind <- length(rna)}</pre>
  df <- c(status, ind)</pre>
  return(df)
}
```

### 3. Select Eligible Patients

```
# 3.1 LME data
lme.dat <- dat %>%
 mutate(t1=months_from_seroco-art_start) %>%
 filter(treatment==1 & t1<6 & t1!=0) %>%
 group_by(PATIENT) %>%
 filter(length(t1)>1)
pid1 <- unique(lme.dat$PATIENT)</pre>
# 3.2 survival data
## 3.2.1 RNA rebound
surv.dat.rebound <- dat %>%
 filter(months_from_seroco >= ati_start) %>%
 group_by(PATIENT) %>%
  mutate(status=Define_status(Find_rebound, RNA_V)[1],
         t2=months_from_seroco-art_start,
         t3=months_from_seroco-ati_start) %>%
  slice(Define_status(Find_rebound, RNA_V)[2])
pid2 <- surv.dat.rebound$PATIENT</pre>
## 3.2.2 RNA peak point:
surv.dat.peak <- dat %>%
  filter(months_from_seroco >= ati_start) %>%
 group_by(PATIENT) %>%
 mutate(status=Define_status(Find_peak, RNA_V)[1],
         t2=months_from_seroco-art_start,
         t3=months_from_seroco-ati_start) %>%
  slice(Define_status(Find_peak, RNA_V)[2])
pid3 <- surv.dat.peak$PATIENT</pre>
```

```
# 3.3 find common patients
pid <- Reduce(intersect, list(pid1, pid2, pid3))
length(pid)

lme.dat <- lme.dat %>% filter(PATIENT %in% pid)
surv.dat.peak <- surv.dat.peak %>% filter(PATIENT %in% pid)
surv.dat.rebound <- surv.dat.rebound %>% filter(PATIENT %in% pid)

# 3.4 identify outliers
lme.dat %>% filter(PATIENT==23) %>% summarise(m=10^mean(RNA_V))
lme.dat %>% filter(PATIENT!=23) %>% ungroup() %>% summarise(m=10^mean(RNA_V))
lme.dat %>% filter(PATIENT!=50) %>% summarise(m=exp(mean(CD4_V)))
lme.dat %>% filter(PATIENT!=50) %>% ungroup() %>% summarise(m=exp(mean(CD4_V)))

# 3.5 remove outliers
lme.dat.rm <- lme.dat %>% filter(!(PATIENT %in% c(23, 50)))
surv.dat.peak.rm <- surv.dat.peak %>% filter(!(PATIENT %in% c(23, 50)))
surv.dat.rebound.rm <- surv.dat.rebound %>% filter(!(PATIENT %in% c(23, 50)))
```

## 4. Exploratory Analysis

```
# 4.1 RNA viral loads trajectories
dat %>%
  ggplot(aes(x=months_from_seroco-art_start, y=RNA_V, group=PATIENT)) +
   geom line(alpha=0.5, color="dark blue") +
   geom_point(alpha=0.2, size=0.8) +
   xlim(0,100) +
   labs(title="Individual Trajectories of (log10) RNA Viral Loads on Eligible Patients",
         subtitle="For the Duration of the Study",
         x="Months from the Start of ART", y="log10 of RNA Viral Load") +
   theme_bw()
ggsave("EDA_RNA.pdf", width=6, height=5)
# 4.2 CD4 cell counts trajectories
dat %>%
  ggplot(aes(x=months_from_seroco-art_start, y=CD4_V, group=PATIENT)) +
    geom_line(alpha=0.5, color="dark blue") +
   geom_point(alpha=0.2, size=0.8) +
   xlim(0,100) +
   labs(title="Individual Trajectories of (log) CD4 Cell Counts on Eligible Patients",
         subtitle="For the Duration of the Study",
         x="Months from the Start of ART", y="Natural log of CD4 Counts") +
    theme_bw()
ggsave("EDA_CD4.pdf", width=6, height=5)
```

# 5. Survival analysis (Cox Model)

```
# 5.1 Model Selection
## 5.1.1 RNA rebound
rebound.cox1 <- coxph(Surv(t2, status)~GENDER+age,</pre>
                         data=surv.dat.rebound.rm, x=TRUE)
rebound.cox2 <- coxph(Surv(t2, status)~1,
                         data=surv.dat.rebound.rm, x=TRUE)
anova(rebound.cox1, rebound.cox2)
## 5.1.2 RNA peak point
peak.cox1 <- coxph(Surv(t2, status)~GENDER+age,</pre>
                         data=surv.dat.peak.rm, x=TRUE)
peak.cox2 <- coxph(Surv(t2, status)~1,</pre>
                         data=surv.dat.peak.rm, x=TRUE)
anova(peak.cox1, peak.cox2)
# 5.2 COX Model on rebound and peak point
cox.fit.rebound <- coxph(Surv(t2, status)~1, data=surv.dat.rebound.rm, x=TRUE)
cox.fit.peak <- coxph(Surv(t2, status)~1, data=surv.dat.peak.rm, x=TRUE)
# 5.3 Histogram
surv.dat.rebound.rm %>%
  ggplot(aes(x=t3)) +
  geom histogram(color="black", fill="gray", bins=60) +
 theme bw() +
 xlim(0,30) +
 labs(title="Histogram on Time to Viral Load Rebound",
       x="Months since ART Interruption", y="Number of Patients")
ggsave("hist_rebound.pdf", width=6, height=5)
surv.dat.peak.rm %>%
  ggplot(aes(x=t3)) +
  geom_histogram(color="black", fill="gray", bins=60) +
  theme_bw() +
 xlim(0,100) +
  labs(title="Histogram on Time to Viral Load Peak Point",
       x="Months since ART Interruption", y="Number of Patients")
ggsave("hist_peak.pdf", width=6, height=5)
```

### 6. LME

```
# 6.1 Model Selection
## 6.1.1 RNA viral load decay
rna.lme1 <- lme(RNA_V~t1+age+GENDER, random=~t1|PATIENT, data=lme.dat, method="ML")
rna.lme2 <- lme(RNA_V~t1, random=~t1|PATIENT, data=lme.dat, method="ML")
anova(rna.lme1, rna.lme2)
## 6.1.2 CD4 cell counts</pre>
```

```
cd4.lme1 <- lme(CD4_V~t1+age+GENDER, random=~t1|PATIENT, data=lme.dat, method="ML")
cd4.lme2 <- lme(CD4_V~t1, random=~t1 PATIENT, data=lme.dat, method="ML")
anova(cd4.lme1, cd4.lme2)
# 6.2 LME Model
## 6.2.1 RNA viral loads
lme.fit.rna <- lme(RNA V~t1, random=~t1 PATIENT, data=lme.dat)</pre>
lme.fit.rna.rm <- lme(RNA_V~t1, random=~t1|PATIENT, data=lme.dat.rm)</pre>
## 6.2.2 CD4 cell counts
lme.fit.cd4 <- lme(CD4_V~t1, random=~t1|PATIENT, data=lme.dat)</pre>
lme.fit.cd4.rm <- lme(CD4_V~t1, random=~t1|PATIENT, data=lme.dat.rm)</pre>
# 6.3 Individual Trajectories
## 6.3.1 RNA viral loads
lme.dat.rm %>%
  ggplot(aes(x=t1, y=RNA_V, group=PATIENT)) +
   geom_line(alpha=0.5, color="dark blue") +
   geom_point(alpha=0.2, size=0.8) +
   ylim(0,6) +
   labs(title="Individual Trajectories of (log10) RNA Viral Loads on Eligible Patients",
         subtitle="First 6 months of ART Treatment",
         x="Months from the Start of ART", y="RNA Viral Load (log10 scale)") +
   theme bw()
ggsave("long_RNA.pdf", width=6, height=5)
## 6.3.2 CD4 cell counts
lme.dat.rm %>%
  ggplot(aes(x=t1, y=CD4_V, group=PATIENT)) +
   geom_line(alpha=0.5, color="dark blue") +
   geom_point(alpha=0.2, size=0.8) +
   ylim(5,7.5) +
   labs(title="Individual Trajectories of (log) CD4 Cell Counts on Eligible Patients",
         subtitle="First 6 months of ART Treatment",
         x="Months from the Start of ART", y="Natural log of CD4 Counts") +
   theme bw()
ggsave("long_CD4.pdf", width=6, height=5)
# 6.4 Diagnostics
## 6.4.1 residual plot
pdf("lme_residual1.pdf", width=6, height=5)
plot(lme.fit.rna.rm, resid(.,type="p")~fitted(.),id=0.05,adj=-0.3)
dev.off()
pdf("lme_residual2.pdf", width=6, height=5)
plot(lme.fit.cd4.rm, resid(.,type="p")~fitted(.),id=0.05,adj=-0.3)
dev.off()
## 6.4.2 QQ plot for error terms
pdf("lme_error1.pdf", width=6, height=5)
qqnorm(lme.fit.rna.rm, ~resid(.))
```

```
dev.off()
pdf("lme_error2.pdf", width=6, height=5)
qqnorm(lme.fit.cd4.rm, ~resid(.))
dev.off()
## 6.4.3 QQ plot for random effects
pdf("lme_ranef1.pdf", width=6, height=5)
qqnorm(lme.fit.rna.rm, ~ranef(.),id=0.1,cex=0.7)
dev.off()
pdf("lme_ranef2.pdf", width=6, height=5)
qqnorm(lme.fit.cd4.rm, ~ranef(.),id=0.1,cex=0.7)
dev.off()
# 6.5 Goodness of fit
## 6.5.1 RNA viral loads
lme.dat.rm %>%
  add_column(pred1=predict(lme.fit.rna.rm), pred2=predict(lme.fit.cd4.rm)) %>%
  mutate(patid=as.factor(PATIENT)) %>%
  filter(patid %in% c(1, 29, 36, 43, 69, 75)) %>%
  ggplot(aes(x=t1, y=pred1, group=patid)) +
  geom_line() +
  geom_point(aes(x=t1, y=RNA_V, group=patid), size=2, alpha=0.5) +
  facet_wrap(~patid) +
  theme bw() +
 labs(title="LME Prediction Plot for (log10) RNA Viral Loads on Selected Patients",
      x="Time in Month", y="RNA Viral Loads on log10 Scale")
ggsave("rna_pred.pdf", width=7, height=5)
## 6.5.2 CD4 cell counts
lme.dat.rm %>%
  add_column(pred1=predict(lme.fit.rna.rm), pred2=predict(lme.fit.cd4.rm)) %%
  mutate(patid=as.factor(PATIENT)) %>%
  filter(patid %in% c(1, 29, 36, 43, 69, 75)) %>%
  ggplot(aes(x=t1, y=pred2, group=patid)) +
  geom_line() +
  geom_point(aes(x=t1, y=CD4_V, group=patid), size=2, alpha=0.5) +
 facet_wrap(~patid) +
  theme_bw() +
  labs(title="LME Prediction Plot for (log) CD4 Cell Counts on Selected Patients",
      x="Time in Month", y="CD4 Cell Counts on Natural Log Scale")
ggsave("cd4_pred.pdf", width=7, height=5)
```

### 7. Bayesian Joint model (shared random effects)

#### 7.2 Diagnostics for Analysis 1

```
## 7.2.1 Trace plot
pp <- do.call(cbind, joint.fit1$mcmc["alphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Intercept Association Structure $\\alpha^{(r)}_{a_0}$"),
       subtitle="Analysis 1: Joint Model on Viral Load and Viral Rebound
                with Shared Random Effects",
      x="iteration", y=TeX("^{(r)}_{a_0}")) +
  theme_bw()
ggsave("trace1.pdf", width=7, height=3.5)
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,2]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Slope Association Structure $\\alpha^{(r)}_{a_1}$"),
       subtitle="Analysis 1: Joint Model on Viral Load and Viral Rebound
                with Shared Random Effects",
      x="iteration", y=TeX("\\alpha^{(r)} {a 1}$")) +
  theme bw()
ggsave("trace2.pdf", width=7, height=3.5)
## 7.2.2 ACF plot
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))</pre>
pp2 <- with(acf(pp[,2], plot=FALSE), data.frame(lag, acf))
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Intercept Association Structure
                 \alpha^{(r)} \{a 0\}
       subtitle="Analysis 1: Joint Model on Viral Load and Viral Rebound
                with Shared Random Effects",
       x="lag", y="Autocorrelation") +
  theme_bw()
```

### 7.3 Diagnostics for Analysis 2-4

```
### 7.3.2 Analysis 2
### Trace plot
pp <- do.call(cbind, joint.fit2$mcmc["alphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Intercept Association Structure
                 $\\alpha^{(p)}_{a_0}$"),
       subtitle="Analysis 2: Joint Model on Viral Loads and Viral Peak Point
                with Shared Random Effects",
       x="iteration", y=TeX("$\Lambda^{(p)}_{a_0}") +
  theme bw()
ggsave("trace4.pdf", width=7, height=3.5)
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,2]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Slope Association Structure $\\alpha^{(p)}_{a_1}$"),
       subtitle="Analysis 2: Joint Model on Viral Loads and Viral Peak Point
                with Shared Random Effects",
       x="iteration", y=TeX("\$\lambda^{(p)}_{a_1}\$")) +
  theme bw()
ggsave("trace5.pdf", width=7, height=3.5)
### ACF plot
pp <- do.call(cbind, joint.fit2$mcmc["alphas"]) %>% data.frame()
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))</pre>
pp2 <- with(acf(pp[,2], plot=FALSE), data.frame(lag, acf))</pre>
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Intercept Association Structure
                 $\\alpha^{(p)}_{a_0}$"),
       subtitle="Analysis 2: Joint Model on Viral Loads and Viral Peak Point
                with Shared Random Effects",
       x="lag", y="Autocorrelation") +
  theme_bw()
```

```
ggsave("acf4.pdf", width=7, height=3.5)
ggplot(aes(x=lag, y=acf), data=pp2) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Slope Association Structure
                 $\\alpha^{(p)}_{a_1}$"),
       subtitle="Analysis 2: Joint Model on Viral Loads and Viral Peak Point
                with Shared Random Effects",
      x="lag", y="Autocorrelation") +
  theme bw()
ggsave("acf5.pdf", width=7, height=3.5)
### 7.3.3 Analysis 3
### Trace plot
pp <- do.call(cbind, joint.fit3$mcmc["alphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Intercept Association Structure $\\alpha^{(r)}_{b_0}$"),
       subtitle="Analysis 3: Joint Model on CD4 Cell Counts and Viral Rebound
                with Shared Random Effects",
       x="iteration", y=TeX("^{(r)}_{b_0}")) +
  theme_bw()
ggsave("trace6.pdf", width=7, height=3.5)
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,2]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Slope Association Structure $\\alpha^{(r)}_{b_1}$"),
       subtitle="Analysis 3: Joint Model on CD4 Cell Counts and Viral Rebound
                with Shared Random Effects",
      x="iteration", y=TeX("\$\\\lambda^{(r)}_{b_1}\$")) +
  theme bw()
ggsave("trace7.pdf", width=7, height=3.5)
### ACF plot
pp <- do.call(cbind, joint.fit3$mcmc["alphas"]) %>% data.frame()
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))
pp2 <- with(acf(pp[,2], plot=FALSE), data.frame(lag, acf))</pre>
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Intercept Association Structure
                 $\\alpha^{(r)}_{b_0}$"),
       subtitle="Analysis 3: Joint Model on CD4 Cell Counts and Viral Rebound
                with Shared Random Effects",
      x="lag", y="Autocorrelation") +
  theme bw()
ggsave("acf6.pdf", width=7, height=3.5)
ggplot(aes(x=lag, y=acf), data=pp2) +
  geom_hline(aes(yintercept=0)) +
```

```
geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Slope Association Structure $\\alpha^{(r)}_{b_1}$"),
       subtitle="Analysis 3: Joint Model on CD4 Cell Counts and Viral Rebound
                with Shared Random Effects",
       x="lag", y="Autocorrelation") +
  theme bw()
ggsave("acf7.pdf", width=7, height=3.5)
### 7.3.4 Analysis 4
### Trace plot
pp <- do.call(cbind, joint.fit4$mcmc["alphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Intercept Association Structure $\\alpha^{(p)}_{b_0}$"),
       subtitle="Analysis 4: Joint Model on CD4 Cell Counts and Viral Peak Point
                with Shared Random Effects",
       x="iteration", y=TeX("$\Lambda^{(p)}_{b_0}$")) +
  theme_bw()
ggsave("trace8.pdf", width=7, height=3.5)
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,2]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Slope Association Structure $\\alpha^{(p)}_{b_1}$"),
       subtitle="Analysis 4: Joint Model on CD4 Cell Counts and Viral Peak Point
                with Shared Random Effects",
       x="iteration", y=TeX("$\Lambda^{(p)}_{b_1}")) +
  theme bw()
ggsave("trace9.pdf", width=7, height=3.5)
### ACF plot
pp <- do.call(cbind, joint.fit4$mcmc["alphas"]) %>% data.frame()
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))</pre>
pp2 <- with(acf(pp[,2], plot=FALSE), data.frame(lag, acf))</pre>
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Intercept Association Structure
                 $\\alpha^{(p)}_{b_0}$"),
       subtitle="Analysis 4: Joint Model on CD4 Cell Counts and Viral Peak Point
                with Shared Random Effects",
       x="lag", y="Autocorrelation") +
  theme bw()
ggsave("acf8.pdf", width=7, height=3.5)
ggplot(aes(x=lag, y=acf), data=pp2) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Slope Association Structure
                 $\\alpha^{(p)}_{b_1}$"),
       subtitle="Analysis 4: Joint Model on CD4 Cell Counts and Viral Peak Point
                with Shared Random Effects",
```

```
x="lag", y="Autocorrelation") +
theme_bw()
ggsave("acf9.pdf", width=7, height=3.5)
```

#### 7.4 Sensitivity Analysis 1

```
## 7.4.1 RNA viral loads and time to rebound
joint.fit1.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",
                              param="shared-RE", n.iter=50000,
                              control=list(priorVar=10))
summary(joint.fit1.sen)
## 7.4.2 RNA viral loads and time to peak point
joint.fit2.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",
                              param="shared-RE", n.iter=50000,
                              control=list(priorVar=10))
summary(joint.fit2.sen)
## 7.4.3 CD4 cell counts and time to rebound
joint.fit3.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                              param="shared-RE", n.iter=50000,
                              control=list(priorVar=10))
summary(joint.fit3.sen)
## 7.4.4 CD4 cell counts and time to peak point
joint.fit4.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                              param="shared-RE", n.iter=50000,
                              control=list(priorVar=10))
summary(joint.fit4.sen)
```

#### 7.5. Sensitivity Analysis 2

### 8. Bayesian Joint model (shared $\eta_i(t)$ )

```
# 8.1 Model fitting
## 8.1.1 RNA viral load and time to rebound
joint.fit1.eta <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound,</pre>
                                   timeVar="t1", n.iter=50000)
summary(joint.fit1.eta)
## 8.1.2 RNA viral load and time to peak points
joint.fit2.eta <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak,</pre>
                                   timeVar="t1", n.iter=50000)
summary(joint.fit2.eta)
## 8.1.3 CD4 cell counts and time to rebound
joint.fit3.eta <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound,</pre>
                                   timeVar="t1", n.iter=50000)
summary(joint.fit3.eta)
## 8.1.4 CD4 counts and time to peak points
joint.fit4.eta <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak,</pre>
                                   timeVar="t1", n.iter=50000)
summary(joint.fit4.eta)
```

### 8.2 Sensitivity Analysis 3

```
# 8.2.1 RNA viral loads and time to rebound
joint.fit1.eta.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",
                              n.iter=50000, control=list(priorVar=1000))
summary(joint.fit1.eta.sen)
# 8.2.2 RNA viral loads and time to peak point
joint.fit2.eta.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",
                              n.iter=50000, control=list(priorVar=1000))
summary(joint.fit2.eta.sen)
# 8.2.3 CD4 cell counts and time to rebound
joint.fit3.eta.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                              n.iter=50000, control=list(priorVar=1000))
summary(joint.fit3.eta.sen)
# 8.2.4 CD4 cell counts and time to peak point
joint.fit4.eta.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                              n.iter=50000, control=list(priorVar=1000))
summary(joint.fit4.eta.sen)$`CoefTable-Event`[-3, -6] %% xtable(digits=4)
```

### 8.3 Sensitivity Analysis 4

```
# 8.3.1 RNA viral loads and time to rebound
joint.fit1.eta.sen2 <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",
                              n.iter=50000, control=list(priorVar=10))
summary(joint.fit1.eta.sen2)
# 8.3.2 RNA viral loads and time to peak point
joint.fit2.eta.sen2 <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",
                              n.iter=50000, control=list(priorVar=10))
summary(joint.fit2.eta.sen2)
# 8.3.3 CD4 cell counts and time to rebound
joint.fit3.eta.sen2 <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",</pre>
                              n.iter=50000, control=list(priorVar=10))
summary(joint.fit3.eta.sen2)
# 8.3.4 CD4 cell counts and time to peak point
joint.fit4.eta.sen2 <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                              n.iter=50000, control=list(priorVar=10))
summary(joint.fit4.eta.sen2)
```

# 9. Bayesian Joint Model (shared $\eta_i(t)$ and $\eta'_i(t)$ )

```
# 9.1 Model fitting
## 9.1.1 RNA viral loads and time to rebound
dForm <- list(fixed=~1, random=~1, indFixed=2, indRandom=2)
joint.fit1.eta.p <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm)
summary(joint.fit1.eta.p)
## 9.1.2 RNA viral loads and time to peak point
joint.fit2.eta.p <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm)
summary(joint.fit2.eta.p)
## 9.1.3 CD4 cell counts and time to rebound
joint.fit3.eta.p <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm)
summary(joint.fit3.eta.p)
## 9.1.4 CD4 cell counts and time to peak point
joint.fit4.eta.p <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",</pre>
                                    n.iter=50000, param="td-both", extraForm=dForm)
summary(joint.fit4.eta.p)
```

### 9.2 Sensitivity analysis 5

```
## 9.2.1 RNA viral loads and time to rebound
joint.fit1.eta.p.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                    control=list(priorVar=1000))
summary(joint.fit1.eta.p.sen)
## 9.2.2 RNA viral loads and time to peak point
joint.fit2.eta.p.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                    control=list(priorVar=1000))
summary(joint.fit2.eta.p.sen)
## 9.2.3 CD4 cell counts and time to rebound
joint.fit3.eta.p.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                    control=list(priorVar=1000))
summary(joint.fit3.eta.p.sen)
## 9.2.4 CD4 cell counts and time to peak point
joint.fit4.eta.p.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                    control=list(priorVar=1000))
summary(joint.fit4.eta.p.sen)
```

### 9.3 Sensitivity Analysis 6

```
## 9.3.1 RNA viral loads and time to rebound
joint.fit1.eta.p.sen2 <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",</pre>
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                    control=list(priorVar=10))
summary(joint.fit1.eta.p.sen2)
## 9.3.2 RNA viral loads and time to peak point
joint.fit2.eta.p.sen2 <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",</pre>
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                     control=list(priorVar=10))
summary(joint.fit2.eta.p.sen2)
## 9.3.3 CD4 cell counts and time to rebound
joint.fit3.eta.p.sen2 <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                    control=list(priorVar=10))
summary(joint.fit3.eta.p.sen2)
## 9.3.4 CD4 cell counts and time to peak point
joint.fit4.eta.p.sen2 <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                                    n.iter=50000, param="td-both", extraForm=dForm,
                                     control=list(priorVar=10))
summary(joint.fit4.eta.p.sen2)
```

# 10. Bayesian Joint Model (shared cumulative effects)

```
# 10.1 Model fitting
## 10.1.1 RNA viral loads and time to rebound
iForm \leftarrow list(fixed=~0+t1+I(t1^2/2), random=~0+t1+I(t1^2/2), indFixed=1:2, indRandom=1:2)
joint.fit1.cum <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",
                                   n.iter=50000, param="td-extra", extraForm=iForm)
summary(joint.fit1.cum)
## 10.1.2 RNA viral loads and time to peak point
joint.fit2.cum <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",</pre>
                                     n.iter=50000, param="td-extra", extraForm=iForm)
summary(joint.fit2.cum)
## 10.1.3 CD4 cell counts and time to rebound
joint.fit3.cum <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",</pre>
                                     n.iter=50000, param="td-extra", extraForm=iForm)
summary(joint.fit3.cum)
## 10.1.4 CD4 cell counts and time to peak point
joint.fit4.cum <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",</pre>
                                     n.iter=50000, param="td-extra", extraForm=iForm)
summary(joint.fit4.cum)
```

### 10.2 Diagnostics for Analysis 13

```
### 10.2.1 Trace plot
pp <- do.call(cbind, joint.fit1.cum$mcmc["Dalphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Cumulative Association Structure $\\alpha^{(r)}_{y}$"),
       subtitle="Analysis 13: Joint Model on Viral Load and Viral Rebound
                with Shared Cumulative Effects",
       x="iteration", y=TeX("\$\alpha^{(r)}_{y}$")) +
  theme bw()
ggsave("trace3.pdf", width=7, height=3.5)
### 10.2.2 ACF plot
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Cumulative Association Structure
                 \alpha^{(r)}_{y}^{y},
       subtitle="Analysis 13: Joint Model on Viral Load and Viral Rebound
                with Shared Cumulative Effects",
      x="lag", y="Autocorrelation") +
  theme_bw()
ggsave("acf3.pdf", width=7, height=4)
```

### 10.3 Diagnostics for Analysis 14-16

```
## 10.3.1 Analysis 14
### Trace plot
pp <- do.call(cbind, joint.fit2.cum$mcmc["Dalphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Cumulative Association Structure $\\alpha^{(p)}_{y}$"),
       subtitle="Analysis 14: Joint Model on Viral Loads and Viral Peak Point
                with Shared Cumulative Effects",
       x="iteration", y=TeX("\$\alpha^{(p)}_{y}\$")) +
  theme_bw()
ggsave("trace10.pdf", width=7, height=3.5)
### ACF plot
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))</pre>
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Cumulative Association Structure
                 \alpha^{(p)}_{y}^{y},
       subtitle="Analysis 14: Joint Model on Viral Loads and Viral Peak Point
                with Shared Cumulative Effects",
       x="lag", y="Autocorrelation") +
  theme bw()
ggsave("acf10.pdf", width=7, height=4)
## 10.3.2 Analysis 15
### Trace plot
pp <- do.call(cbind, joint.fit3.cum$mcmc["Dalphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom_line() +
  labs(title=TeX("Trace Plot for Cumulative Association Structure $\\alpha^{(r)}_{z}$"),
       subtitle="Analysis 15: Joint Model on CD4 Cell Counts and Viral Rebound
                with Shared Cumulative Effects",
       x="iteration", y=TeX("\$\alpha^{(r)}_{z}\$")) +
  theme_bw()
ggsave("trace11.pdf", width=7, height=3.5)
### ACF plot
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Cumulative Association Structure
                 \alpha^{(r)}_{z}^{,}
       subtitle="Analysis 15: Joint Model on CD4 Cell Counts and Viral Rebound
                with Shared Cumulative Effects",
       x="lag", y="Autocorrelation") +
```

```
theme_bw()
ggsave("acf11.pdf", width=7, height=4)
## 10.3.3 Analysis 16
### Trace plot
pp <- do.call(cbind, joint.fit4.cum$mcmc["Dalphas"]) %>% data.frame()
ggplot(aes(x=seq(1, nrow(pp)), y=pp[,1]), data=pp) +
  geom line() +
  labs(title=TeX("Trace Plot for Cumulative Association Structure $\\alpha^{(p)} {z}$"),
       subtitle="Analysis 16: Joint Model on CD4 Cell Counts and Viral Peak Point
                with Shared Cumulative Effects",
       x="iteration", y=TeX("\$\alpha^{(p)}_{z}\$")) +
  theme bw()
ggsave("trace12.pdf", width=7, height=3.5)
### ACF plot
pp1 <- with(acf(pp[,1], plot=FALSE), data.frame(lag, acf))</pre>
ggplot(aes(x=lag, y=acf), data=pp1) +
  geom_hline(aes(yintercept=0)) +
  geom_segment(aes(xend=lag, yend=0)) +
  labs(title=TeX("Autocorrelation Plot for Cumulative Association Structure
                 $\\alpha^{(p)}_{z}$"),
       subtitle="Analysis 16: Joint Model on CD4 Cell Counts and Viral Peak Point
                with Shared Cumulative Effects",
       x="lag", y="Autocorrelation") +
  theme_bw()
ggsave("acf12.pdf", width=7, height=4)
```

### 10.4 Sensitivity Analysis 7

```
## 10.4.1 RNA viral loads and time to rebound
joint.fit1.cum.sen0 <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",</pre>
                                     n.iter=50000, param="td-extra", extraForm=iForm,
                                     control=list(priorVar=1000))
summary(joint.fit1.cum.sen0)
## 10.4.2 RNA viral loads and time to peak point
joint.fit2.cum.sen0 <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",</pre>
                                     n.iter=50000, param="td-extra", extraForm=iForm,
                                     control=list(priorVar=1000))
summary(joint.fit2.cum.sen0)
## 10.4.3 CD4 cell counts and time to rebound
joint.fit3.cum.sen0 <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",</pre>
                                     n.iter=50000, param="td-extra", extraForm=iForm,
                                     control=list(priorVar=1000))
summary(joint.fit3.cum.sen0)
## 10.4.4 CD4 cell counts and time to peak point
joint.fit4.cum.sen0 <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
```

### 10.5 Sensitivity Analysis 8

```
## 10.5.1 RNA viral loads and time to rebound
joint.fit1.cum.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",</pre>
                                    n.iter=50000, param="td-extra", extraForm=iForm,
                                     control=list(priorVar=1))
summary(joint.fit1.cum.sen)
## 10.5.2 RNA viral loads and time to peak point
joint.fit2.cum.sen <- jointModelBayes(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",</pre>
                                    n.iter=50000, param="td-extra", extraForm=iForm,
                                    control=list(priorVar=1))
summary(joint.fit2.cum.sen)
## 10.5.3 CD4 cell counts and time to rebound
joint.fit3.cum.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                                    n.iter=50000, param="td-extra", extraForm=iForm,
                                     control=list(priorVar=1))
summary(joint.fit3.cum.sen)
## 10.5.4 CD4 cell counts and time to peak point
joint.fit4.cum.sen <- jointModelBayes(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                                    n.iter=50000, param="td-extra", extraForm=iForm,
                                    control=list(priorVar=1))
summary(joint.fit4.cum.sen)
```

#### 10.6 Sensitivity Analysis 9

### 11. Individual Prediction

```
## 11.1 Survival prediction
nd.surv <- dat %>%
 mutate(t1=months_from_seroco-art_start) %>%
 filter(PATIENT==1 & treatment==0 & t1<23.73333)
sfit <- survfitJM(joint.fit1, newdata=nd.surv, idVar="PATIENT")</pre>
pdf("survpred.pdf", width=6, height=4)
plot(sfit, estimator="mean", include.y=TRUE, conf.int=TRUE, fill.area=TRUE, xlim=c(21,31),
     main="Survival Prediction for Patient 1", xlab="Month since the start of ART",
     ylab="Survival Probability",
    ylab2="RNA Viral Load (log10-scale)")
dev.off()
## 11.2 Longitudinal Prediction
nd.long <- dat %>%
 mutate(t1=months_from_seroco-art_start) %>%
 filter(PATIENT==1 & treatment==1)
lfit <- predict(joint.fit1, newdata=nd.long, idVar="PATIENT", type="Subject",</pre>
                interval="confidence", return=TRUE, FtTimes=seq(5.966667, 8, length.out=25))
last.time <- max(with(lfit, t1[is.na(low)]))</pre>
pdf("longpred.pdf", width=6, height=4)
xyplot(pred+low+upp~t1, data=lfit, type="l", lty=c(1,2,2),
       col=c(2,1,1), abline=list(v=last.time, lty=3),
       main="Longitudinal Prediction for Patient 1",
       xlab="Month since the start of ART",
       ylab=("Predicted RNA Viral Load (log10-scale)"))
dev.off()
```

# 12. Joint Models with Likelihood Method (shared $\eta_i(t)$ )

### 13. Joint Model with Likelihood Method (shared $\eta_i(t)$ and $\eta'_i(t)$

```
## 13.1 RNA viral loads and time to rebound
dForm <- list(fixed=~1, random=~1, indFixed=2, indRandom=2)</pre>
jm.fit1.eta.p <- jointModel(lme.fit.rna.rm, cox.fit.rebound, timeVar="t1",</pre>
                            parameterization="both",
                            method="piecewise-PH-GH", derivForm=dForm)
summary(jm.fit1.eta.p)
## 13.2 RNA viral loads and time to peak point
jm.fit2.eta.p <- jointModel(lme.fit.rna.rm, cox.fit.peak, timeVar="t1",</pre>
                            parameterization="both",
                            method="piecewise-PH-GH", derivForm=dForm)
summary(jm.fit2.eta.p)
## 13.3 CD4 cell counts and time to rebound
jm.fit3.eta.p <- jointModel(lme.fit.cd4.rm, cox.fit.rebound, timeVar="t1",
                            parameterization="both",
                            method="piecewise-PH-GH", derivForm=dForm)
summary(jm.fit3.eta.p)
## 13.4 CD4 cell counts and time to peak point
jm.fit4.eta.p <- jointModel(lme.fit.cd4.rm, cox.fit.peak, timeVar="t1",
                            parameterization="both",
                            method="piecewise-PH-GH", derivForm=dForm)
summary(jm.fit4.eta.p)
```

# 14. Model Comparison for Bayesian Joint Models

```
anova(joint.fit1, joint.fit1.eta, joint.fit1.eta.p, joint.fit1.cum)
anova(joint.fit2, joint.fit2.eta, joint.fit2.eta.p, joint.fit2.cum)
anova(joint.fit3, joint.fit3.eta, joint.fit3.eta.p, joint.fit3.cum)
anova(joint.fit4, joint.fit4.eta, joint.fit4.eta.p, joint.fit4.cum)
```