

# *Tree House Explorer Documentation*

ANDREW J. HARRIS, NICOLE M. FOLEY, TIFFANI L. WILLIAMS, WILLIAM J. MURPHY

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## Introduction

Genomes are mosaics of evolutionary histories that reflect ancient signatures of species divergence, as well as incomplete lineage sorting (ILS) or gene flow. In recent years, rapid advancements in massively parallel sequencing technologies have enabled researchers to collect large volumes of genome-scale phylogenetic data for many species. Understanding how and why phylogenetic signal varies across species genomes can yield powerful insights into evolutionary histories and adaptive evolution. By integrating diverse data types with local genealogies, users can differentiate genetic variation that is consistent with the species tree from that stemming from natural selection, ILS, or gene flow (1-4). While several tools have been developed for the independent analysis of genomic data (e.g., IGV, UCSC Genome Browser, etc.) and phylogenetic tree visualization (Iroki, FigTree, etc.), a tool that can simultaneously analyze phylogenetic signal variation with other chromosomal and gene-based annotations has yet to be developed for the field of phylogenomics. Tree House Explorer (THEx) is a novel genome browser that allows users to integrate phylogenomic data and genomic annotations into a single interactive platform for combined analysis. THEx allows users to visualize genome-wide variation in evolutionary histories as well as genetic divergence on a chromosome-by-chromosome basis, with continuous sliding window comparisons to gene annotations, recombination rates, GC-content, and other user-specified, highly customizable feature annotations. THEx provides a new resource for interactive data visualization in phylogenomics and a novel approach to analyze and interpret the diverse evolutionary histories woven throughout genomes.

## What Can Tree House Explorer Do?

Tree House Explorer offers a collection of dashboards that visualize phylogenetic and genomic data concurrently. Each dashboard provides highly interactive graphs that allow for smooth browsing through data at chromosome or genome-wide levels. In addition to being interactive, each dashboard provides an array of graph customization that helps create publication-quality figures. Each graph comes with a built-in snapshot feature located in the top right corner in the toolbar that allows users to download a graph with a click of a button. Depending on the graph, users can also find other tools in the toolbar that provide other means of interacting with each graph. For Unix and macOS systems, THEx comes with a command-line suite called THExBUILDER that provides pipelines and tools to generate, modify, and analyze THEx input files, allowing for further customization and control over your data.

## THExBUILDER

### Purpose

THExBUILDER is a suite of pipelines and tools designed to generate, modify, and analyze THEX input files. It provides a customized solution for generating Tree Viewer input files. It also contains a single command pipeline for calculating raw p-distance values from multiple-sequence fasta files in non-overlapping sliding windows for Signal Tracer. THExBUILDER was designed to be flexible and scalable, taking advantage of multiprocessing and partitioned workflows. For example, the Tree Viewer Pipeline is designed to be run either with a configuration file or by command-line arguments and run the pipeline step-by-step or from start to finish with a single command. This flexibility provides the user the freedom to tailor their runs and track the progress through pipelines with ease.

### Usage

In addition to being a novel interactive genome browser, THEX comes with a built-in command-line suite that contains tools and pipelines that help simplify the process of building and manipulating input files for the dashboards. There are two main pipelines, one for Tree Viewer and one for Signal Tracer. To use THExBUILDER, activate the Conda environment that THEX is installed within and enter the command “thexb -h” to begin. This command will bring up the help section, providing an overview of all the options and available tools.

THExBUILDER's output is contained in a single THExBUILDEROutput directory. By default, the output directory is written to the current working directory. If users do not provide an output location, be sure to use the same working directory moving through the pipelines. Within the output directory, users will find the intermediate output files for each stage of a given pipeline and log files to help track performed tasks. This information includes what files have been used/modified, in what order, and input parameters used for a given command.

### Input Files

All of the pipelines for THExBUILDER start with multiple-sequence alignment fasta files organized within a single directory. Each fasta file ideally represents the multiple-sequence alignment of a single chromosome. However, scaffold level assemblies and subsets of sequences (i.e., specific genes) can be used too, with the caveat that each sequence is treated as independent sequences. There is no way to link scaffolds into pseudo-chromosomes within the pipeline or application. Below is a basic example of the directory structure and file structure of the multiple-sequence fasta file.

#### **MultiAlignmentDir/**

| -- chr1.fasta

| -- chr2.fasta

| -- chX.fasta

#### **Example file: (chr1.fasta)**

> Sample 1

AGTGCTAGC...GTTC

> Sample 2

AGTGTCCTA...GCTT

> Sample 3

AGTGTCGTA...GCT

## Tree Viewer Pipeline

The Tree Viewer pipeline is designed to take multi-alignment fasta files, parse them into non-overlapping windows, run the windows through several filtration steps, infer maximum likelihood phylogenies per-window using IQ-Tree2, and then bin identical topologies using topobinner for visualization in the Tree Viewer dashboard. The examples below show how to run each stage with and without a configuration file, providing the user flexibility to run the pipeline with all arguments conveniently located in a single file. It is recommended to use a configuration file as this helps improve reproducibility and also reduces the length of the commands, but the option is left to the user. Since the configuration file has to be built with a specific format, you can generate a blank configuration file by running “thexb --tv\_config\_template”.

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### Start-to-Finish Run

The Tree Viewer pipeline is designed to be flexible, providing the user the ability to run each step independently or continuously one after another. Although the option to run the pipeline start-to-finish is available, it is recommended to run each step individually to inspect and validate the intermediate results. However, if you choose to run it all at once you will need to create a configuration file and pass it to the '-c' argument.

---

#### Command Example

```
$ thexb --tv_all -c config.ini
```

---

### Fasta Windowing

The fasta windowing step takes a directory containing multi-alignment fasta files and returns a directory of subdirectories, labeled by the original sequence names (i.e., chr1.fasta -> chr1/), containing the non-overlapping windowed fasta files. Note that this step can return thousands of files, so be mindful when opening these directories in a file browser. Fasta files can be uncompressed or compressed using bgzip.

---

#### Command Example

```
$ thexb --minifastas -c config.ini
```

**or**

```
$ thexb --minifastas \  
-i ./chromosome_alignments \  
-o ./output \  
--window_size 100kb
```

---

### Trimal Gap Trimming

The Trimal (5) stage of the pipeline uses the gap-threshold feature that removes regions of a window where a taxon has missing data that exceeds the provided threshold for allowed gaps. A second threshold is provided to remove resulting sequences that do not meet a minimum sequence length.

---

### Command Example

```
$ thexb --trimal -c config.ini
```

**or**

```
$ thexb --trimal \  
-i ./output/windowed_fastas \  
-o ./output \  
--trimal_min_seq_len 1000bp \  
--trimal_gap_threshold 0.85
```

---

### Pairwise Distance + Coverage Filter

The pairwise distance and coverage filter walks through each window of the genome and conducts a sliding sub-window analysis with a step size of  $n$ -bp to mask local regions of extreme divergence that may be caused by misalignment or undetected paralogy. It also masks entire windows where a single taxon's base coverage (valid bases include A, T, G, and C) is below the provided threshold. This step was integrated from Foley et al. 2021 (6) and updated to run on Python 3.

---

### Command Example

```
$ thexb --pw_filter -c config.ini
```

**or**

```
$ thexb --pw_filter \  
-i ./output/trimal_filtered_windows \  
-o ./output \  
--reference ReferenceName \  
--pw_subwindow_size 100bp \  
--pw_step 10bp \  
--pw_min_seq_len 5000bp \  
--pw_max_pdist 1.0 \  
--pw_zscore_cutoff 2 \  
--pw_missing_char "-" \  
--pw_seq_coverage 0.8
```

---

### IQ-Tree: Maximum Likelihood Phylogeny Inference

IQ-Tree (7-8) is used to infer maximum likelihood phylogenies providing the basis for the distribution of phylogenetic signal visualized in Tree Viewer. The filtered fasta files generated by the pairwise filtration step is passed to IQ-Tree and the resulting Newick files are collected and organized into the basis of the Tree Viewer input file. IQ-Tree creates several output files per-window, but we are only interested in the trees so the other data can be saved or discarded.

Two approaches are built into the Tree Viewer pipeline for the IQ-Tree analysis. The first is to have THExBuilder call IQ-Tree for each filtered window by using the "--iqtree" command, sequence evolution model, and the number of bootstrap replications. The second approach is to take the filtered fasta files from the pairwise filtration step and run IQ-Tree externally on a different local machine, server, or cluster. If users choose to run IQ-Tree externally, users will need to organize the resulting ".treefile"

Newick trees into a single directory and pass the directory to the "--iqtree\_external" command rather than the "--iqtree" command. Users can also optionally create sub-directories per chromosome, organize the ".treefile" by chromosome, and pass it to the "--iqtree\_external" command, but it is not required.

---

### Command Example

```
$ thexb --iqtree -c config.ini
```

**or**

```
$ thexb --iqtree \  
-i ./output/pairwise_filtered_windows/ \  
-o ./output/ \  
--iqtree_model GTR*H4 \  
--iqtree_bootstrap 1000 \  
--tv_file_name TVinput.xlsx
```

---

### Topobinner

Topobinner is a tool used to organize and label identical tree topologies based on RF-distance. Trees are treated as equal topologies when their RF-distance is equal to 0 (9-10). The trees are binned and then labeled based on their whole genome frequency, meaning the most frequent topology in the genome will be labeled Tree1, then Tree 2 for the second most frequent topology, and so on. Once this step is completed, an updated Tree Viewer file with binned topologies will be generated and placed in the defined output directory. It should also be noted that users can also pass a Tree Viewer input file produced by the File Pruning export option (see Tree Viewer export options section for more details) in Tree Viewer to bin new trees that have been pruned into a subset from a larger Tree Viewer input file. No special steps need to be taken to run this, simply provide the Tree Viewer input file as you would normally.

---

### Command Example

```
$ thexb --topobinner -c config.ini
```

**or**

```
$ thexb --topobinner \  
-i ./output/TVinput.xlsx \  
-o ./output/
```

---

### TOPOBINNER ALTERNATIVE:

An alternative to using topobinner is PhyBin (<https://github.com/rnewton/PhyBin>). PhyBin is not hosted on conda, so users would need to download the program from GitHub and install it externally.

It is important that users use PhyBin's "--bin" option and not its clustering algorithm. When users run PhyBin, it will generate binned results in a new output directory named either phybin\_output or whatever users provide as an output location. Passing the output directory generated by PhyBin and a Tree Viewer file with the TopologyID column blank to THExBuilder's "--phybin\_external" command will produce a complete Tree Viewer input file just like topobinner.

---

### Command Example

```
$ thexb --phybin_external \  
-i phybin_output_directory/\  
--tv_file_name TVinput.xlsx
```

### Signal Tracer Pipeline

The Signal Tracer pipeline is a single script that calculates raw, uncorrected p-distance between a reference and other taxon of one or more multi-alignment fasta files. There are two ways to run the pipeline, you can provide a single multi-alignment fasta file or a directory of multi-alignment fasta files. The directory of files is typically the input directory used for the entire Tree Viewer pipeline that is ideally a multi-alignment fasta file for each chromosome. The script enables users to include or exclude missing data in the p-distance calculation by providing the `--pdist_ignore_missing` argument, and also the `--pdist_missing_character` if missing data is represented by something other than "N".

---

### Command Example

```
$ thexb --pdistance \  
-i ./chromosome_alignments \  
-o ./output/\  
-r Tiger \  
-w 100kb \  
--pdist_threshold 0.75
```



## Tree Viewer

### Purpose

Tree Viewer is a dashboard designed for window-based approaches to visualizing phylogenetic signal across a reference genome alongside additional data types like recombination rate, GC-content, and gene annotations. Through simultaneous visualization of phylogenetic signal and additional data types, users are offered an all-in-one experience for identifying and understanding the implications of phylogenetic signal variation and its underlying genomic context. Through Tree Viewer, users are able to make more impactful findings and gain a better understanding of their data all in one place. Download the example data sets from GitHub and refer to Li et al. 2019 for an example clade that illustrates Tree Viewer (2).

### Input Files

There are two required input files to run Tree Viewer: 1) Tree Viewer input file and 2) chromosome length BED file. These two files provide the required information to visualize phylogenetic signal across the genome in proper scale to the chromosome length. Additional window-based data types can be added to the Tree Viewer input file providing, the ability to visualize a multitude of data types concurrently. These additional data types are added as a new column in the Tree Viewer file keeping your data succinct and organized.

### Tree Viewer Input File

The input file for Tree Viewer is designed to be simple to make and even easier to incorporate new window-based data. Tree Viewer takes a tab or comma delimited file where the first four columns are Chromosome, Window (i.e., 100,000 - which covers bases 1-100,000 for 100kb windows), NewickTree, and TopologyID. The first four columns are required and must have the appropriate headers in the order given in the example input below. Tree Viewer accepts four different file extensions (.csv, .tsv, .txt, .xlsx) for the input file. Note there are column and per-cell limitations to Excel (.xlsx) files, so large datasets may be better off in a flat file format like .csv, .tsv, or .txt.

Chromosome	Window	NewickTree	TopologyID
chr1	1000	(A,(B,C));	Tree1
chr2	2000	(B,(A,C));	Tree2
chr3	3000	(C,(A,B));	Tree3
chr4	4000	(A,(B,C));	Tree1

Additional numeric and categorical data (i.e., GC-content, recombination rate low/high regions, etc.) can be easily incorporated into your Tree Viewer file by adding an additional column on the right-most side of the Tree Viewer input file. When users load your Tree Viewer file into a new session, the additional data features will load as options in the "Additional Data" dropdown in the "Single Chromosome" tab of the toolbar.

Chromosome	Window	NewickTree	TopologyID	GC-content
chr1	1000	(A,(B,C));	Tree1	0.35
chr2	2000	(B,(A,C));	Tree2	0.46
chr3	3000	(C,(A,B));	Tree3	0.33

---

## Chromosome Lengths BED File

The second file required to run Tree Viewer is a BED file (.bed) of chromosome lengths. Ensure that the chromosome length bed file contains all chromosomes that are found in the Tree Viewer main input file, otherwise users will be prompted with an error and asked to update your BED file with the missing data. It is also important that users create your file with headers as shown below.

Chromosome	Start	End
chr1	0	1000
chr2	0	2000
chr3	0	3000
chr4	0	4000

---

## Project File

Project files enable users to load all required and additional input files all at once rather than separately. Users can download a template project.ini file using the "Download project.ini Template" button and fill out with the file path information using any text editor. In future releases, provided a project directory is given within the project file, changes made during the session will be logged to the project directory. This includes information about pruned files, current view export coordinates, and other important changes.

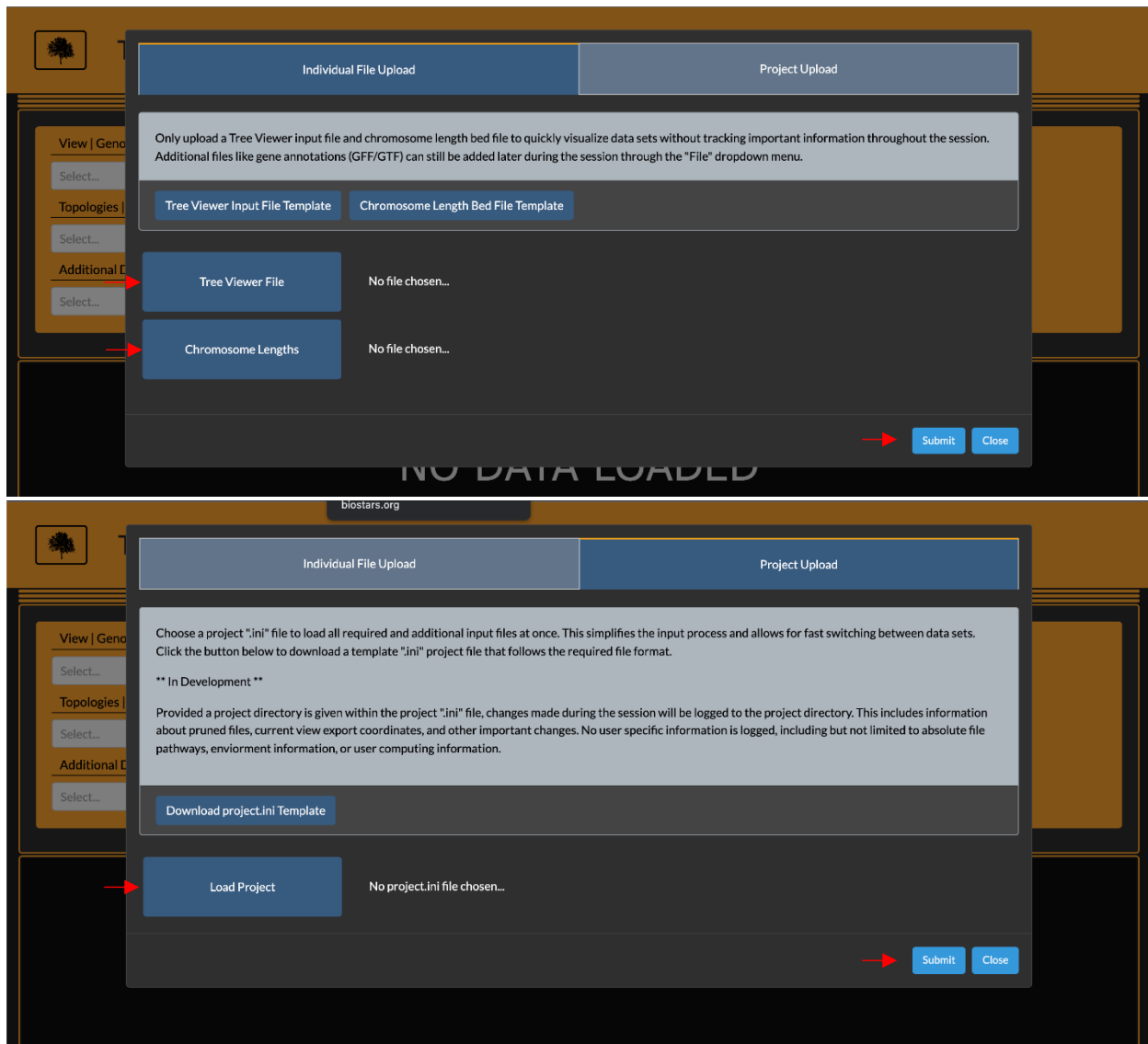
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## Standardized File Types

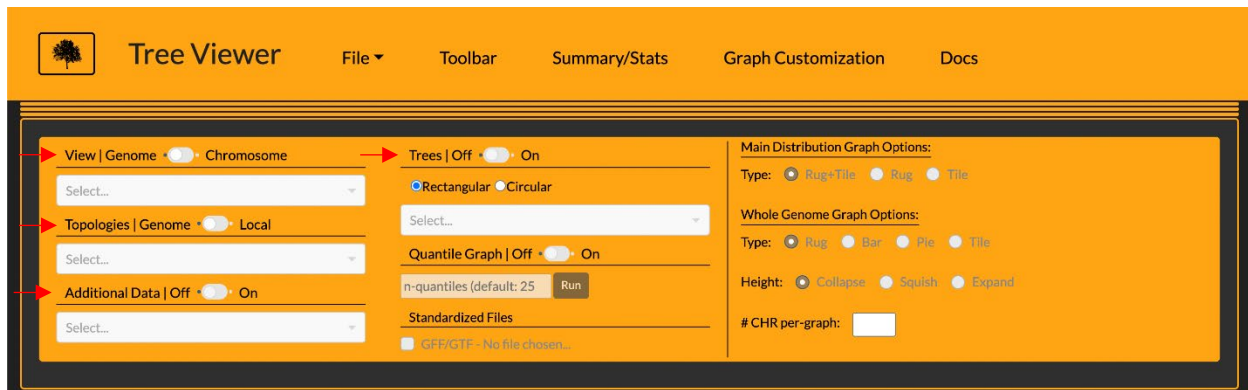
Tree Viewer currently only offers the ability to load GFFv3/GTF gene annotation files. Provided one of these files, users can investigate underlying genes, coding regions, and other annotations concurrently with phylogenetic signal and all other user-provided window-based data types. More standard data types will be added in future updates.

## Usage

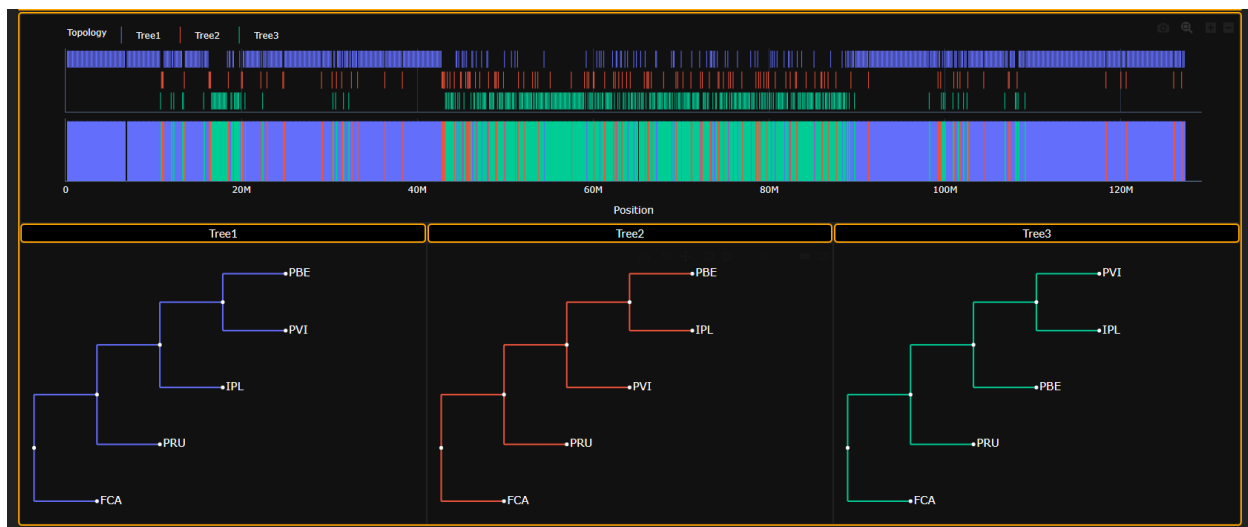
A pop-up will appear upon opening Tree Viewer, prompting users to upload individual input files or a single project file. The two required input files for individual file uploads are a Tree Viewer input file and a chromosome length BED file (formats are described in the input file structure section). Clicking each button will bring up a file system browser allowing users to navigate to the location of your input files. When a file is selected, the upload button will turn green if the file type is valid or red if it is invalid. After selecting both a Tree Viewer input file as well as a chromosome length BED file, users click submit. If users want to upload a project file instead, click the "Project Upload" tab and the "Load Project" button to select the desired project file. Once the upload button has turned green, click submit. Tree Viewer then compares the input files and returns an error message if one or both files are incorrectly formatted or have missing required data. If both files pass the validation checks, they will automatically be uploaded into the session, and the toolbar dropdowns will be populated with the data provided in the input files.



The dashboard functions by two primary viewing levels, per-chromosome/local and whole genome. View type can be changed by toggling the "Genome – Chromosome" switch next to the "View" data entry within the main toolbar. While in single chromosome view, the main distribution graph is the master graph for viewing, meaning all zooming and panning performed on the master graph will update the x-axis range for all other additional data graphs currently being shown. This allows for smooth, simultaneous investigation of phylogenetic signal alongside additional data types like recombination rates, GC-content, and gene annotations without the need to align the data yourself. Whole-genome view allows for a large-scale overview of the phylogenetic signal, making it easy to pick out interesting chromosomal regions to investigate in further detail in the single chromosome view.



Single chromosome view provides the most interactive place to explore your phylogenetic signal. This is where users will add/remove topologies, load additional data types into graphs, and visualize tree topologies. The dropdowns for the topologies, additional data, and tree taxa allow the user to customize which data are being shown at a given time. Users may plot one topology or all topologies, with the caveat that as one increases the number of topologies, visualization and analysis of the signal becomes more complex. By default, Tree Viewer orders the topologies by their genome-wide frequencies. However, it may be useful to look at them by local frequency, and there is a switch next to the "Topologies" header that allows users to change between these alternatives. Turning the Additional Data switch to "On" will load all additional data types that are selected within its respective dropdown, allowing users to load multiple data types at once. Lastly, turning on the Trees switch will load simple representations of the tree topologies selected in the Topologies dropdown. Tree Viewer provides plots of the tree topologies without branch lengths since branch lengths can change from window to window. However, if the user is in single chromosome view and clicks on a window, the tree within the selected window will be drawn with its respective branch lengths. By selecting/removing taxa names from the Tree's dropdown, users can prune the tree topology to zoom in on a specific clade or set of taxa. Note that this function does not re-bin topologies. Instead, it simply visually prunes the trees. If users want to prune and re-bin topologies, see the File Pruning function in the Tree Viewer export options section.



There are a few extra graphing options on the rightmost side of the main toolbar. Under *Main Distribution Graph Options*, users will find options to change the graph type of the master graph to preview the rug plot and tiled histogram chromosome displays independently. Under *Whole Genome*

*Graph Options*, users can choose from a selection of graph types and change the height of the graphs. These previews can be exported using the snapshot button in the top-right of each graph. The "Summary/Stats" tab is home to various summary tables for single chromosome/local regions as well as whole-genome overviews. It also has several tools to compare and explore the relationships between phylogenetic signal and other genomic data. Lastly, to upload a GFF/GTF file, go to "File - > GFF/GTF" and upload the file similarly to the main input files. Once the file is loaded into the session, the GFF/GTF switch will activate, allowing users to load the GFF/GTF graph.

---

#### Walk-Through: Start New Session + Load Individual Input Files

1. Click "*File + New Session*"
2. Click "*Tree Viewer Input*" button – navigate to and select Tree Viewer input file.
3. Click "*Chromosome Lengths*" button – navigate to and select Chromosome Length BED file.
4. Assuming both input buttons have turned green, click "Submit".

---

#### Walk-Through: Start New Session + Load Project File

1. Click "*File + New Session*"
2. Click "*Project Upload*" tab at the top
3. Click "*Load Project*" button – navigate to and select project ".ini" file.
4. Assuming the input button has turned green, click "Submit".

---

#### Walk-Through: Load GFF/GTF Files

1. Click "*File + GFF/GTF*" – A pop-up will appear titled "Alternative Input Files".
2. Click "*GFF/GTF File*" button – navigate to and select GFF/GTF input file.
3. Once the button turns green, click "Submit" – users should see the GFF/GTF switch now active.

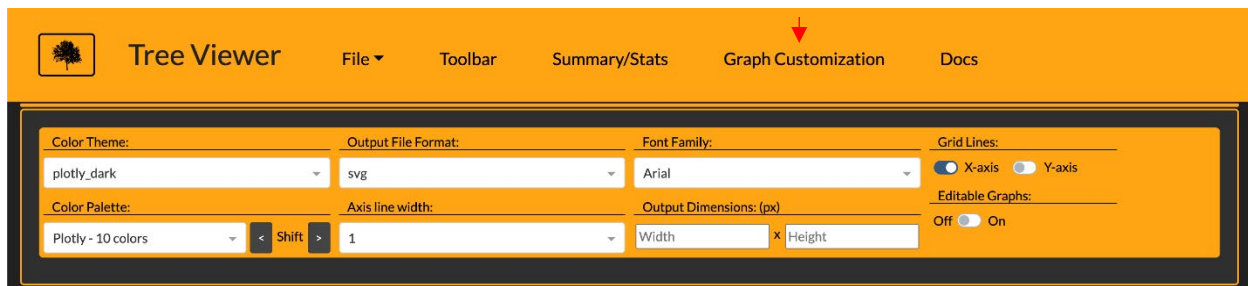
---

#### Walk-Through: Zooming + Panning

1. Zoom in by clicking and dragging across a region of interest.
2. Users can zoom in or out in steps by using the plus and minus buttons in the toolbar at the top right corner of the graph.
3. Reset the x-axis range by double-clicking the graph.
4. Pan by clicking and dragging the x-axis of the graph

### Graph Customization

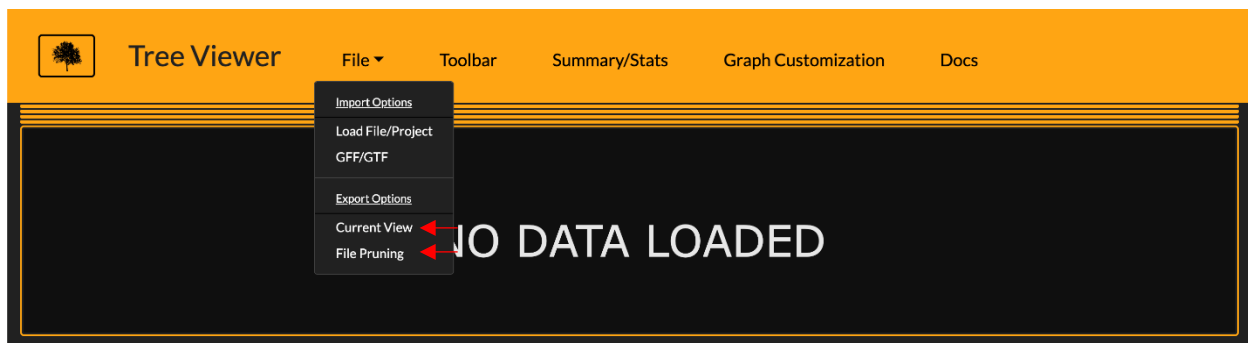
Graph Customization allows users to customize the look of the topology or data distribution graphs. Dropdowns provide lists of themes, color palettes, gridline toggles, and several output formats to choose. Making changes to the graph customizations will update all graphs currently loaded on the browser. Through these customizations, one can create publication-quality graphs that can be dropped directly into your manuscript. We have allowed users to specify the output dimensions in pixels; however, leaving the inputs blank will revert each graph to default dimensions optimized for a word document or PDF file.



## Tree Viewer Export Options

Tree Viewer offers two export options: current view and file pruning. The current view option allows users to extract local information for a chromosomal region. For example, users are in single chromosome view and looking at Chromosome 1, and users zoom into range 1.5Mb-2Mb. If users click the current view export option, a download prompt will appear, allowing users to choose a place to download a new Tree Viewer file that only contains the information for the current range. In addition, if users also have a GFF/GTF gene annotation file loaded into the session, it will also extract the information from the given range.

The file pruning export option allows users to select a subset of taxa and create a new Tree Viewer input file with a pruned set of Newick trees. This process will also clear the TopologyID column, enabling users to re-bin the new tree topologies. There is an option to run the binning process before downloading the new file, but please note that re-binning large trees across large genomes (e.g., >2 Gbp) may require considerable run time, which will lock your session until it completes. In such circumstances, we recommend running the `--topobinner` command in THExBuilder.



## Walk-Through: Current View Export

1. Click "File + Current View".
2. A file-system prompt will appear and ask users to select a location and file name. Once users have selected a location and file name, click "Save".
  - a. If a GFF/GTF file is loaded into the session, then a second file-system prompt will appear to download a subset of the GFF/GTF file too.

## Walk-Through: File Pruning

1. Click "File + File Pruning" – A pop-up will appear titled "Tree Viewer File Pruning".
2. Select taxa to retain in the Tree Viewer input file subset using the dropdown.

3. Check "*run Topobinner*" if users wish to run topobinner prior to downloading the file subset.
4. Finally, click "*Submit*" and wait for a file-system prompt to appear to choose the location to download the new subset file.

## Signal Tracer

### Purpose

Signal Tracer (ST) is a simple dashboard that visualizes window-based values calculated from multiple sequence alignments. It provides an easy way to investigate variation in variables like genetic distance, branch length, or divergence times at single chromosome and whole genome levels. Signal Tracer was initially developed to validate the phasing of F1 hybrid long-reads using the trio-binning approach for single haplotype genome assembly references. By visualizing the genetic distance of reference and non-reference species, we could visually validate that there were no regions that indicate potential improper phasing. If users would like to learn more about this process, refer to our paper (11).

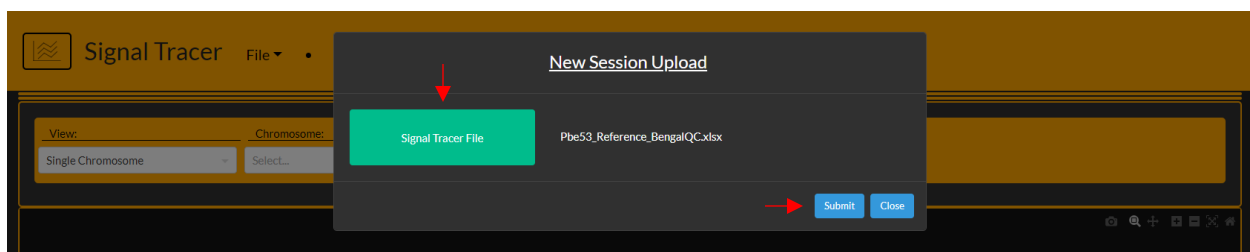
### Input File Structure

The input file for Signal Tracer is a tab or comma-delimited file consisting of four columns: *Chromosome*, *Window*, *Sample*, and *Value*. The *Value* column can only contain numerical values, not categorical values. Ensure the headers of your input file match the headings listed in italics above. An error message will appear if they do not match exactly.

Chromosome	Window	Sample	Value
chr1	100	Tiger1	0.015
chr1	100	Tiger2	0.25
chr2	200	Tiger1	0.011
chr2	200	Tiger2	0.008

### Usage

A new session pop-up will appear upon entering Signal Tracer, prompting users to upload your data. Click submit once the "Load File" button turns green. From there, users can switch between chromosomes or change to whole-genome view and explore your data.



This dashboard was originally designed to visualize raw genetic distances variation across the genome, so THExBuilder comes with a "--pdistance" command to generate input files for Signal Tracer from multi-alignment fasta files. Visit the Signal Tracer Pipeline in the THExBuilder documentation for more information.

---

### Walk-Through: Load Input Files

1. Click "*File + New Session*" – A pop-up will appear titled "New Session Upload".
2. Click "*Signal Tracer File*" button – navigate to and select Tree Viewer input file.
3. Assuming the input button turned green, click "Submit".

---

### Walk-Through: Load GFF/GTF Files

1. Click "*File + GFF/GTF*" – A pop-up will appear titled "Alternative Input Files".
2. Click "*GFF/GTF File*" button – navigate to and select GFF/GTF input file.
3. Once the button turns green, click "*Submit*" – users should see the GFF/GTF switch now active.

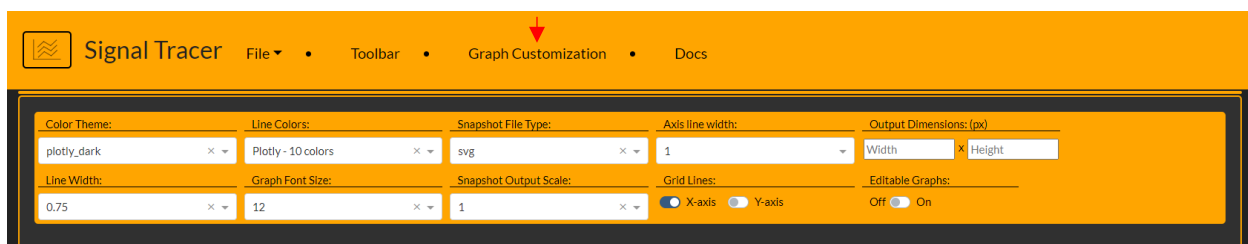
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### Walk-Through: Zooming + Panning

1. Zoom in by clicking and dragging across a region, then release.
2. Users can zoom in or out in steps by using the plus and minus buttons in the top right corner of the graph.
3. Reset the view by double-clicking the graph.
4. Pan by clicking and dragging the x-axis of the graph

## Graph Customization

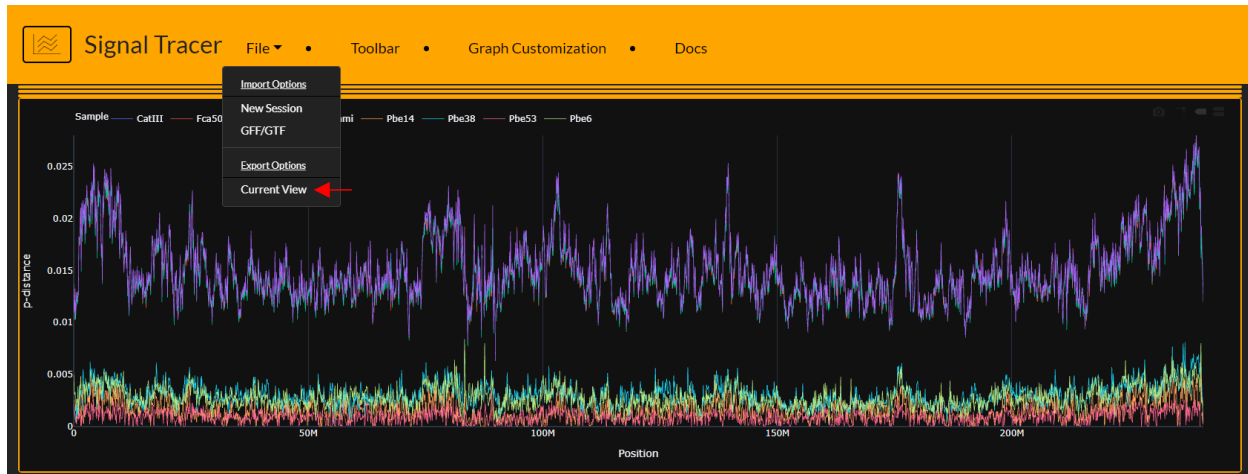
Graph Customization allows users to customize the look of the chromosome graphs in an easy-to-use way. Dropdowns provide lists of themes, color palettes, gridline toggles, and several output formats to choose from. Making changes to the graph customizations will update all graphs currently loaded on the browser. Through these customizations, one can create publication-quality graphs that can be dropped directly into your in-progress manuscript. Although Signal Tracer allows users to specify the output dimensions in pixels, leaving the inputs blank will revert each graph to default dimensions that are optimized for a word document or PDF file.





## Signal Tracer Export Options

Signal Tracer offers a single export option that allows users to extract local information from your input file. Similar to Tree Viewer, zooming into a region of a chromosome in single chromosome view and selecting "File -> Current View" will bring up a download prompt where users can specify the file name and download location.



### Walk-Through: Current View Export

1. Click "*File + Current View*".
2. A file-system prompt will appear and ask users to select a location and file name. Once users have selected a location and file name, click "*Save*".
  - a. If a GFF/GTF file is loaded into the session, then a second file-system prompt will appear to download a subset of the GFF/GTF file too.

## References

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