



Version 2.0

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### **MetumpX 2.0 User Guide**

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## I. INTRODUCTION

**Metabolomics** is the comprehensive (qualitative and quantitative) study of all the small molecules in an organism called **metabolites**. These molecules are smaller than 1500 daltons (Da). Some important metabolites are peptides, oligonucleotides, sugars organic acids ketones, aldehydes, amines and amino acids.

A range of analytical techniques are used to analyze metabolites in different organisms, tissues, or fluids. There are two approaches used in metabolomics:

### 1 *Untargeted Metabolomics*

It involves measurement of as many metabolites as possible from a biological sample to classify phenotypes based on metabolite pattern. It is also known as Metabolic fingerprinting.

### 2 *Targeted Metabolomics*

It involves measurement of metabolites of a focused group from a biological sample. It is also known as Metabolic profiling.

There are several techniques for extraction of metabolic information from samples such as Solid Phase Extraction, Chromatography, Mass Spectrometry (MS) and Nuclear Magnetic Resonance (NMR).

This paper presents **MetumpX**, a software package pertaining to **MS-based Untargeted Metabolomics**.

## II. METABOLOMICS WORKFLOW

Metabolomics Workflow can be divided into many steps as shown in Fig. 1. Details of each step is as follows:

### 1 *Sample Acquisition*

Metabolites are measured in different samples such as tissue, biofluids (blood, urine, feces, seminal fluid, saliva, bile, cerebrospinal fluid) and cell cultures. There are numerous laboratory methods of sample preparation.

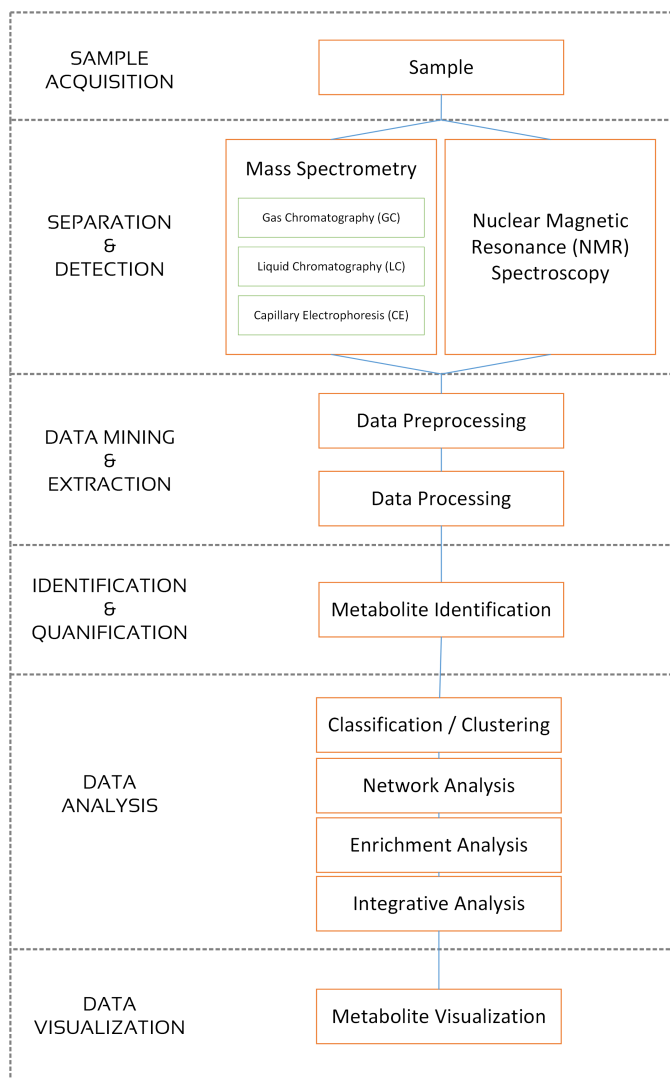


Fig. 1: Metabolomics Analytical Workflow

### 2 *Separation and Detection of metabolites*

The two main analytical methods for separation and detection are Mass Spectrometry (MS) and Nuclear Magnetic Resonance (NMR) Spectroscopy.

#### 1) *Nuclear Magnetic Resonance (NMR) Spectroscopy:*

In NMR spectroscopy, biological sample is placed in a magnetic field. Isotopes in the sample absorb the radiation and resonate at frequencies relative to the size of the molecules. The resultant spectrum shown in Fig. 2 is a collection of peaks at different positions and intensities and each sample has a unique pattern.

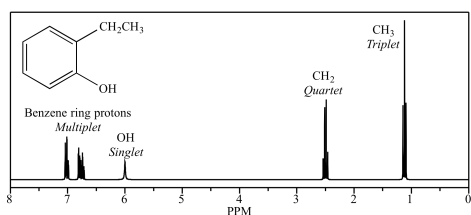


Fig. 2: NMR spectrum for 2-ethyl-phenol

## 2) Mass Spectrometry (MS):

Mass Spectrometry can be used to analyze biological samples by ionizing the sample and then sorting the ions according to their mass-to-charge ( $m/z$ ) ratio. Mass Spectrometer function is shown in Fig. 3. Significant methods for ionization of sample contain Chemical Ionization (CI), Electron Impact Ionization (EI) and Electrospray Ionization (ESI). A resulting MS Spectrum is shown in Fig. 4

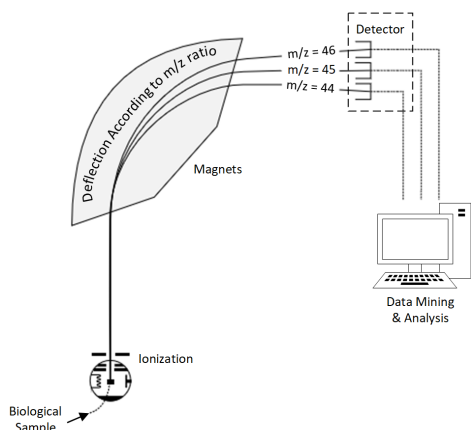


Fig. 3: Standard Mass Spectrometer

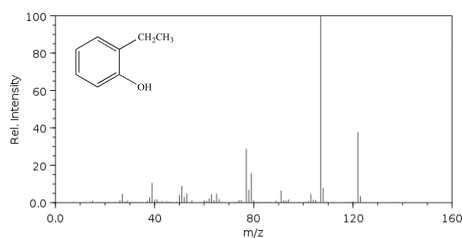


Fig. 4: MS spectrum for 2-ethyl-phenol

An important enhancement to the mass resolving and mass determining capabilities of mass spectrometry is using it in tandem with chromatographic and other separation techniques.

## • Gas Chromatography Mass Spectrometry (GC-MS):

GC-MS is a commonly used in metabolomics for measurement of volatile compounds such

as fatty acids and organic acids. Sample need to be volatile and thermally stable as separation in GC occurs in an oven at high temperatures. Metabolite loss is one of the major drawbacks of GC/MS. Characteristic spectral patterns and extensive libraries are available online for GC-MS. A Gas Chromatograph is shown in Fig. 5

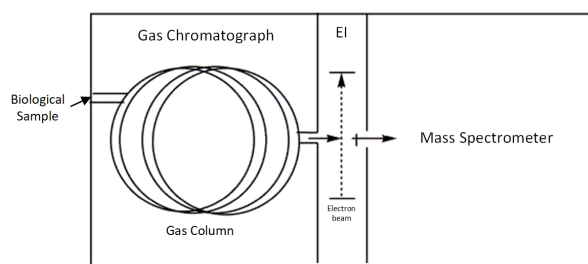


Fig. 5: Gas Chromatograph Mass Spectrometer

## • Liquid Chromatography Mass Spectrometry (LC-MS):

Liquid chromatography (LC) and High-Performance Liquid Chromatography (HPLC) is a technique that has high resolution and analytical flexibility. It can be used for the analysis of a specific metabolite or class of compounds. LCMS has one advantage over GCMS that there is no need for chemical derivatization of metabolites. A Liquid Chromatograph is shown in Fig. 6

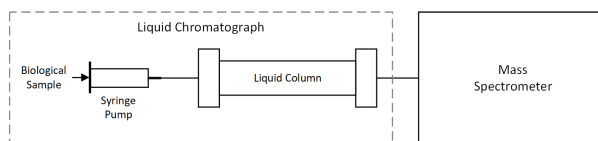


Fig. 6: Liquid Chromatograph Mass Spectrometer

## • Capillary Electrophoresis Mass Spectrometry (CE-MS):

CE-MS provides several advantages over other separation techniques such as high resolving power, very small sample requirement and short analysis time. One of the significant advantages of the CEMS is that it separates cations, anions and uncharged molecules in a single analytical run, and therefore CE can be used for simultaneous profiling of metabolites. Capillary Electrophoresis is shown in Fig. 7

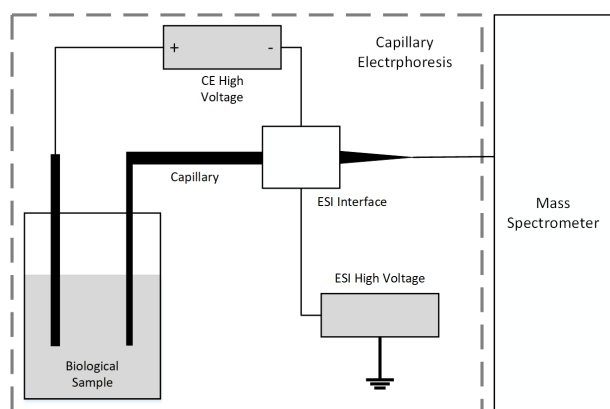


Fig. 7: Capillary Electrophoresis Mass Spectrometer

### 3 Data Mining and Extraction

Untargeted Metabolomic analysis generate large amount of complex data sets that require analysis by specialized software to properly interpret the data. Data is first preprocessed and then processed at this stage. There are several commercial and free software packages available to automate this process.

#### 1) Data Preprocessing:

Data Preprocessing is divided into several sub-steps as follows:

- **Noise Filtering:**

Noise filtering is the process of removing noise from a signal which facilitates further peak detection. It is an optional stage in data processing and can also be left out if the data is not noisy.

- **Chromatogram Alignment:**

Chromatogram alignment is the process of aligning retention time for chromatographic methods with the mass spectrometers.

- **Peak Alignment:**

Peak alignment is a process of correction of samples to point to the same metabolite or component. It is important in metabolomic studies as there is always a difference in the samples due to machine drift.

- **Spectral Deconvolution:**

For GC-MS data, deconvolution is the process of computationally separating co-eluting components and creating a pure spectrum for each component. Deconvolution calculates the contribution of each component from an Extracted Ion Chromatogram that results from two or more components.

- **Retention Time Correction:**

A process of removals of errors in the retention time due to temperature, polarity etc.

#### 2) Data Processing:

Data Processing is divided into several substeps as follows:

- **Peak Detection:**

Peak Detection is a process of identification and quantification of the features present in the spectra acquired from the spectrometer.

- **Data Normalization:**

The total sample amount or concentration of metabolites in metabolomic workflow can be significantly different from one sample to another in each step. Data Normalization is the reduction or elimination of the effect of this variation.

- **Statistical Analysis:**

After pre-processing, the LC-MS raw data is summarized by a peak list. The Statistical Analysis aims to detect those peaks whose intensity levels are significantly altered between distinct biological groups.

- **Quality Control:**

Quality assurance and quality control provides a mechanism to ensure that a scientific process meets the predefined criteria. Quality control (QC) sample should qualitatively and quantitatively represent the entire collection of samples included in the study, providing an average of all of the metabolomes analysed in the study.

- **Metabolite Quantification:**

Metabolite Quantification is method which evaluate changes in metabolic activity in response to disease, treatment, environmental and genetic perturbations.

- **Data Imputation:**

Data Imputation is the process of handling the missing values in mass spectrometry. Typically, there are three types of missing values, missing not at random (MNAR), missing at random (MAR), and missing completely at random (MCAR).

- **In silico fragmentation:**

in silico fragmentation is used to identify unknown compounds outside the database domain by comparing theoretical and experimental data.

#### 4 Metabolite Identification

Metabolite Identification is the main step in which the metabolites are identified, profiled and compared with the chemical database.

#### 5 Data Analysis

Several types of Data Analysis is done as follows:

##### 1) Classification and Clustering Analysis:

Clustering is a well-established technique in the which samples are grouped and visualized according to intrinsic similarities in their measurements, irrespective of sample groupings.

##### 2) Network Analysis:

Network Analysis can be further divided into three types:

##### • Metabolic Network Analysis:

Metabolite datasets are combined through clustering analysis and represented as connection networks between genes and metabolites. The process of analyzing these datasets is called Metabolic Network Analysis.

##### • metabolite Genome-wide Association Study (mGWAS):

Genome-wide association studies with metabolic traits (mGWAS) investigate how genetic variation effect metabolic phenotypes specially metabolism and complex disease. It maps the loci, position of a genes on a chromosome, responsible for natural variations in a target phenotype.

##### • Metabolite Mapping:

Metabolite mapping is a process of Integration of biochemical pathway and chemical relationships to map all detected metabolites in network graphs

##### 3) Enrichment analysis:

Metabolite Enrichment Analysis is statistical analysis of metabolite annotations and/or associated quantitative data.

##### 4) Integrative analysis:

Integrative Analysis is process of linking metabolite data with other types of data (e.g. transcriptomics, proteomics), and incorporating prior knowledge of pathways and molecular interactions.

#### 6 Data Visualization

Visualization of the metabolic spectrum. These are visual graphs with peaks at certain points.

### III. METHODS

The authors of MetumpX researched for all the software tools that are available for the MS-based Untargeted Metabolomics and applied a specific criteria is used for enclosure of tools in MetumpX. Softwares which are included are due to these specific reasons:

- Publication Date (later then 2012)
- Freeware License
- Linux based
- Offline

Table I shows the selection criterion.

TABLE I: MetumpX Selection Criteria

Sr. no	Software Tool Name	Pub. Date (2012-)	Offline	Linux based	Free	Installed
1	ProbMetab[1]	✓	✓	✓	✓	✓
2	intCor[2]	✓	✓	✓	✓	✓
3	CorrectOverloadedPeaks[3]	✓	✓	✓	✓	✓
4	iMet-Q[4]		✓	✓	✓	
5	AnalyzerPro[5]	✓	✓	✓		
6	ADAP-GC[6]		✓	✓	✓	
7	ChromGenius[7]	✓	✓	✓		
8	ChromA[8]	✓			✓	
9	X13CMS[9]	✓	✓	✓	✓	✓
10	MET-COFEA[10]	✓	✓		✓	
11	MET-XAlign[11]	✓	✓		✓	
12	batchCorr[12]	✓	✓	✓		✓
13	MZMine[13]	✓	✓	✓	✓	✓
14	MET-IDEA[14]	✓	✓		✓	
15	PyMS[15]		✓	✓	✓	
16	MassCascade[16]		✓	✓	✓	
17	Mnova-MS[17]	✓	✓	✓		
18	XCMS[18]	✓	✓	✓	✓	✓
19	flagme[19]		✓	✓	✓	✓
20	Elm. Metabolomics[20]	✓	✓		✓	
21	MetCirc[21]	✓	✓	✓	✓	✓
22	mzAccess[22]	✓	✓	✓		
23	SpeckTackle[23]		✓	✓	✓	
24	COMSPAR[24]		✓	✓	✓	
25	DAVE[25]	✓		✓	✓	
26	TargetSearch[26]		✓	✓	✓	
27	HCor[27]	✓		✓	✓	✓
28	MS-DIAL[28]	✓	✓		✓	
29	MaxEnt[29]		✓	✓	✓	
30	RANSY/RAMSY[30]	✓		✓	✓	
31	UniDec[31]	✓	✓		✓	
32	Metab[32]		✓	✓	✓	✓
33	decoMS2[33]	✓	✓	✓	✓	✓
34	GAGdecon[34]		✓	✓	✓	✓
35	TMTc+[35]	✓	✓	✓		
36	BernetAI2018[36]		✓	✓	✓	
37	BUPID-Top-Down[37]	✓		✓	✓	
38	PicaudEtAl2018[38]	✓		✓	✓	✓
39	msXpertSuite[39]	✓	✓	✓	✓	✓
40	proFIA[40]	✓		✓	✓	✓
41	apLCMS[41]	✓	✓	✓	✓	✓
42	yamss[42]	✓	✓	✓	✓	✓
43	cosmiq[43]	✓	✓	✓	✓	✓
44	AMDIS[44]	✓	✓		✓	
45	mzMatch-ISO[45]	✓	✓	✓	✓	✓
46	Elgen-MS[46]		✓	✓	✓	
47	MetaQuant[47]			✓	✓	
48	CAMERA[48]	✓	✓	✓	✓	✓
49	MS-FLQ[49]	✓		✓	✓	
50	JumPm[50]	✓	✓	✓		
51	FastChrom[51]	✓	✓		✓	
52	GridMass[52]		✓	✓	✓	
53	KMMDA[53]	✓	✓	✓	✓	✓
54	HayStack[54]	✓		✓	✓	
55	msPeak[55]		✓	✓	✓	
56	GC-Aanlyzer[56]	✓	✓	✓		
57	MsXclerator[57]		✓	✓		
58	MarkerLynx[58]	✓	✓	✓		
59	GCxGCAnalyzer[59]	✓	✓			
60	MetNorm[60]	✓	✓	✓	✓	✓
61	MetTailor[61]		✓	✓	✓	✓
62	MetaPre[62]	✓		✓	✓	

63	NOREVA[63]	✓		✓	✓	
64	crmn[64]		✓	✓	✓	
65	Normalizer[65]	✓			✓	
66	metaX[66]	✓		✓	✓	
67	MetabR[67]		✓		✓	
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69	MSPrep[69]		✓	✓	✓	✓
70	Aloutput [70]	✓	✓			
71	Ionwinzel[71]	✓	✓		✓	
72	MPP [72]	✓	✓	✓		
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74	mSPA[74]	✓	✓	✓	✓	✓
75	Maui-VIA[75]	✓	✓		✓	
76	SECIMTools[76]	✓	✓	✓	✓	✓
77	BatMass[77]	✓	✓	✓	✓	✓
78	MetaboQC[78]	✓	✓	✓		
79	QCScreen[79]	✓	✓	✓	✓	✓
80	QC-RFSC[80]		✓	✓	✓	✓
81	mscompare[81]	=		✓	✓	
82	PYQUAN[82]	✓	✓			
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196	specmine[196]	✓	✓	✓	✓	✓
197	W4M [197]		✓	✓	✓	✓
198	MeltDB [198]	✓	✓	✓	✓	
199	xMSAnalyzer [199]		✓	✓	✓	✓
200	ChromaTOF[200]	✓	✓	✓	✓	
201	MetabolmeExpress[201]	✓		✓	✓	
202	Metabox [202]		✓	✓	✓	✓
203	PiMP[203]	✓		✓	✓	
204	MET-COFEI[204]		✓	✓	✓	✓
205	MAIT [205]		✓	✓	✓	✓
206	BinVestigate[206]	✓		✓	✓	✓
207	CEU Mass Mediator [207]	✓		✓	✓	✓
208	MAGMa[208]		✓	✓	✓	✓
209	CSI-FingerID [209]	✓		✓	✓	✓
210	MSLDA SUPPORT [210]	✓		✓	✓	✓
211	MetExtract [211]		✓	✓	✓	✓
212	T-BioInfo [212]	✓		✓	✓	
213	MetAlign[213]	✓	✓	✓	✓	✓
214	CFM-ID [214]		✓	✓	✓	✓
215	Ideom [215]		✓	✓	✓	✓
216	ASream [216]		✓	✓	✓	✓
217	PuTMEDID-LCMS[217]		✓	✓	✓	✓
218	DECOMP [218]		✓	✓	✓	✓
219	MetTree[219]	✓		✓	✓	✓
220	MIA [220]	✓	✓	✓	✓	✓
221	MFSearcher[221]	✓	✓	✓	✓	✓
222	ChemDistiller[222]		✓	✓	✓	✓
223	MSeasy[223]	✓	✓	✓	✓	✓
224	SIMPLE[224]		✓	✓	✓	✓
225	MAVEN[225]		✓	✓	✓	✓
226	SpectConnect [226]	✓		✓	✓	✓
227	RAMClustR[227]		✓	✓	✓	✓
228	Molfind[228]	✓	✓	✓	✓	✓
229	MS2Analyzer [229]		✓	✓	✓	✓
230	MS-FINDER [230]	✓	✓	✓	✓	✓
231	geoRge[231]	✓	✓	✓	✓	✓
232	MetFamily[232]	✓	✓	✓	✓	✓
233	eRAH[233]	✓	✓	✓	✓	✓
234	IsoMS [234]	✓	✓	✓	✓	✓
235	MetDIA [235]		✓	✓	✓	✓
236	iMET[236]	✓		✓	✓	✓
237	MIDAS[237]		✓	✓	✓	✓
238	InterpretMSSpectrum [238]	✓		✓	✓	✓
239	AssayR[239]		✓	✓	✓	✓
240	MCID[240]	✓		✓	✓	✓
241	compMS2Miner[241]	✓		✓	✓	✓
242	MetShot [242]		✓	✓	✓	✓
243	MINE[243]	✓		✓	✓	✓
244	NP-StructurePredictor [244]	✓	✓	✓	✓	
245	MetaboSearch[245]	✓	✓	✓	✓	✓
246	ALLocator[246]	✓		✓	✓	✓
247	PROFANCY[247]		✓	✓	✓	✓
248	SpiderMass[248]	✓	✓	✓	✓	✓
249	MZedDB[249]	✓		✓	✓	✓
250	BimBase [250]		✓	✓	✓	✓
251	PowerGet[251]	✓		✓	✓	✓
252	AMDORAP [252]		✓	✓	✓	✓
253	MBIdent [253]		✓	✓	✓	✓
254	peakANOVA[254]		✓	✓	✓	✓
255	MI-Pack[255]		✓	✓	✓	✓
256	SetupX [256]			✓	✓	✓



257	FeatureFinderMetabo [257]		✓	✓	✓	
258	MassMetaSite[258]		✓	✓	✓	
259	MetDNA[259]	✓		✓	✓	
260	DASI[260]	✓	✓	✓	✓	
261	MetaboList[261]	✓	✓	✓	✓	✓
262	MetaMS[262]	✓	✓	✓	✓	✓
263	SIEVE[263]	✓	✓	✓	✓	
264	SimMet[264]	✓		✓	✓	
265	Apex[265]	✓	✓	✓	✓	
266	Nontarget[266]	✓	✓	✓	✓	✓
267	NIST MS Search[267]	✓	✓	✓	✓	✓
268	El-Maven [268]	✓	✓	✓	✓	✓

The authors employed "Zenity" as the front-end GUI for users so that they have an easy and user friendly interface for installation of eighty-eight tools. However, linux bash scripting was employed to carefully handle each installation including necessary dependencies to ensure the users are able to develop workstations quickly and easily. The dependencies which are necessary for the installation of other tools are mentioned in Table II along with their sizes. These dependencies are automatically installed during the execution and Installation of MetumpX.

TABLE II: MetumpX Selection Criteria

Sr. no	Dependencies	Size (MB)
1	Oracle Java JDK	177
2	Libraries	13.8
3	bwidget	167
4	itcl	48.5
5	tk	1.3
6	tdom	0.2
7	tcllib	4
8	Tclx	0.05
9	tkimg	0.1
10	rxvt-uicode	3.4
11	sdcc	3.5
12	doxygen	15.9
13	hunspell	0.04
14	pandas	8.5
15	scipy	31.2
16	numpy	13.9
17	pyomo	1.8
18	decorator	0.01
19	pytz	0.5
20	six	0.01
21	PyUtilib	0.2
22	appdirs	0.01

23	ply	0.05
24	nose	0.1
25	cython	3.4
26	h5py	2.8
27	jupyter client	0.1
28	qtconsole	0.1
29	ipywidgets	0.1
30	pygments	0.8
31	nbconvert	0.2
32	notebook	8.9
33	ipykernel	0.1
34	traitlets	0.1
35	jupyter core	0.1
36	ipython	0.7
37	widetsnbextension	2.2
38	nbformat	0.2
39	prompt toolkit	0.2
40	jinja2	0.1
41	testpath	0.1
42	pyzmq	1.0
43	tornado	0.5
44	pexpect	0.1
45	jedi	0.2
46	html5lib	0.1
47	parso	0.1
48	psutil	0.5
49	matplotlib	0.01
50	pyparsing	0.06
51	kiwisolver	1.0
52	binutils	2.3
53	gcc-avr	14.1
54	avr-libc	4.3
55	uisp	0.5
56	avrdude	0.3
57	flex	0.3
58	byacc	0.5
59	bison	0.5

A Software Categorization Scheme is shown in Table III which shows the the category of each software along with the sizes and versions of each tool included in MetumpX.

One software can be multi-functional and is listed in more then one category but authors have recommended a specific category in which the tools works most efficiently. This categorization is further shown in Fig.8

TABLE III: MetumpX Software Categorization Scheme

Sr. No.	Software	Noise Filtering	Chromatogram Alignment	Peak Alignment	Retention Time Correction	Spectral Deconvolution	Peak Detection	Data Normalization	Statistical Analysis	Quality Control	Metabolite Quantification	Data Imputation	In-silico Fragmentation	Metabolite Identification	Spectral Visualization	Clustering Analysis	mGWAS	Mapping	Network Analysis	Enrichment Analysis	Integrative Analysis	Size (MB)	Version
1	<b>CorrectOverloadedPeaks</b> [3]	✓																				4.8	
2	specmine[196]	✓					✓	✓	✓			✓		✓								15.2	
3	intCor[2]	✓	✓	✓																		3.0	
4	<b>batchCorr</b> [12]		✓																			17.2	
5	<b>mSPA</b> [74]			✓																		0.1	
6	<b>Metab</b> [32]					✓																3.2	
7	decoMS[28]					✓								✓								5.2	
8	GAGdecon[34]					✓																0.1	
9	msXpertSuite[39]					✓																0.7	
10	<b>HCor</b> [27]				✓																	0.1	
11	MetMSLine[193]	✓	✓				✓								✓							1.6	
12	<b>X13CMSX13CMS</b> [9]		✓				✓															0.1	
13	proFIA[40]	✓					✓								✓							2.0	
14	cosmiq[43]		✓				✓				✓											17.5	
15	mzMatch-ISO[45]						✓							✓								0.1	
16	Metabox							✓	✓									✓	✓	✓		53.0	
17	<b>MetNorm</b> [60]	✓						✓														0.5	
18	<b>KMMIDA</b> [53]						✓		✓													0.5	
19	MSPrep[69]							✓	✓													1.0	
20	flagme[19]		✓				✓		✓		✓				✓							22.0	
21	SECIMTools[76]							✓	✓													0.6	
22	<b>QCScreen</b> [79]									✓												12.1	
23	<b>MetTailor</b> [61]		✓								✓											0.9	
24	MetaQuant[47]						✓	✓			✓			✓								12.0	
25	apLCMS[41]		✓				✓				✓			✓								14.1	
26	<b>MINMA</b> [83]											✓										2.6	
27	<b>SIRIUS</b> [85]												✓									37.0	
28	iontree[91]												✓									0.9	
29	MetSho[242]													✓								0.9	
30	MolInd[228]													✓								36.2	
31	MIA[220]													✓								2.7	
32	MetaMS[262]													✓								3.7	
33	MSeasy[223]													✓								5.8	
34	RAMClustR[227]													✓								51.6	
35	MetaboSearch[245]													✓								45.7	
36	El-Maven[268]													✓								91.4	
37	<b>geoRge</b> [231]													✓								13.7	
38	eRAH[233]													✓								3.7	
39	MetaboLis[261]i													✓								0.3	
40	InterpretMSSpectrum[238]													✓								0.2	
41	AssayR[239]													✓								71.6	
42	MS2Analyzer[229]													✓								3.0	
43	Nontarget[266]													✓								3.5	
44	SimEX Targid[194]	✓	✓	✓			✓	✓			✓			✓	✓	✓						49.8	
45	MetaboAnalyst[181]							✓	✓		✓			✓	✓	✓			✓	✓		49.3	
46	CAMERA[48]						✓							✓		✓						2.1	
47	KPIC2		✓				✓				✓					✓						15.7	
48	<b>MWASTools</b> [94]																✓					56.3	
49	<b>InCroMAP</b> [96]																	✓				54.6	
50	PathVisto[97]																	✓				17.6	
51	Mapping Tool[101]																	✓				15.0	
52	PycesToolbox[143]																	✓				5.1	
53	MetaDiff[141]																	✓				6.0	
54	ReactomePA																	✓				11.6	
55	MEBS[144]																	✓				73.2	
56	RxnSim[134]																	✓				21.6	
57	phraSED-ML[162]																	✓				1.8	
58	ScrumPy[164]																	✓				1093.9	
59	Subpathway-GMur[145]																	✓				1.7	
60	Kamneva[152]																	✓				101.8	
61	MetaboSignal[139]																	✓				169.1	
62	ReactPRED[122]																	✓				52.4	
63	Mmine																	✓				52.2	
64	<b>PAP</b> [121]																	✓				0.6	
65	MetaNetSam[147]																	✓				3.8	
66	Fbar[169]																	✓				2.1	
67	SED-ED[132]																	✓				6.4	
68	QSSPN[128]																	✓				2.1	
69	JMassBalance[140]																	✓				3.7	
70	Pyabolism[166]																	✓				0.4	
71	Pybrn[165]																	✓				0.6	
72	MonaLisa[127]																	✓				17.8	
73	MoDitify[138]																	✓				0.6	
74	integrOmics[184]																	✓				0.1	
75	MetScape[172]																	✓		✓	✓	17.2	
76	MetDisease[185]																	✓			✓	15.2	
77	zeroSum[176]																			✓		1.0	
78	FELLA[178]																			✓		3.1	
79	MetaboliteIDConverter[180]																			✓		0.2	
80	MZMine[13]	✓	✓			✓	✓	✓			✓			✓	✓							148.7	
81	XCMS[18]		✓	✓			✓	✓			✓			✓	✓							3.5	
82	yamss[42]		✓				✓															15.5	
83	R2DGC[73]		✓	✓			✓							✓	✓							0.6	
84	<b>BatMass</b> [77]									✓					✓							0.1	
85	MAIT		✓				✓					✓		✓	✓				✓			36.2	
86	ProbMetab[1]	✓	✓				✓		✓					✓	✓							0.2	
87	MetCirc[21]			✓										✓	✓							5.0	
88	xMSAnalyzer[199]		✓			✓	✓			✓				✓	✓							38.9	

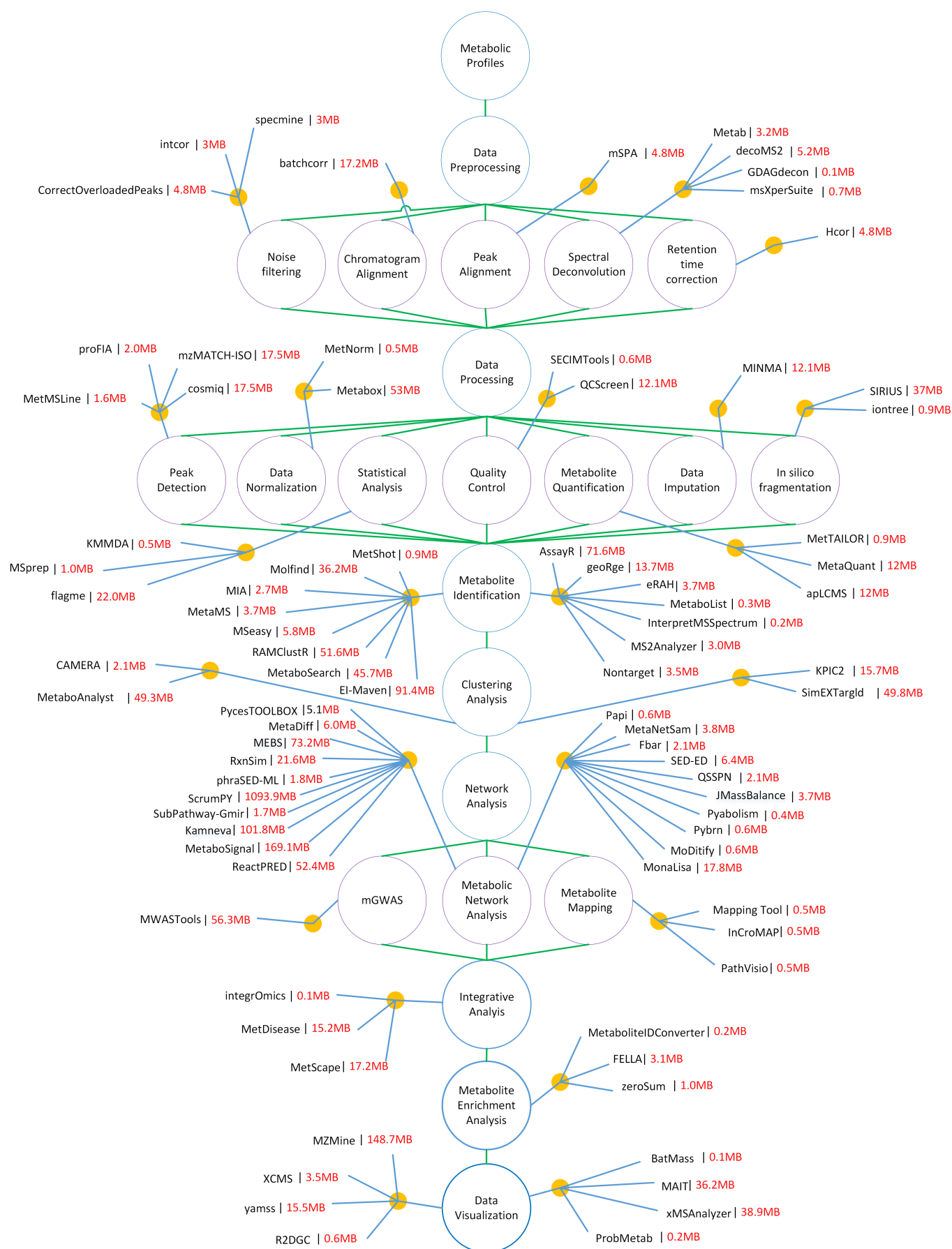


Fig. 8: The figure presents an overview of the Metabolomics software pipeline. The software tools packaged in MetumpX, along with their size are mentioned along each recommended category in the figure.

#### IV. SOFTWARE INSTALLATION PROCESS

For downloading of MetumpX, visit its website:  
<https://github.com/hasaniqbal777/MetumpX>

- 1) MetumpX can also be downloaded from the following command through git:

```
git clone <url>
```

- 2) Now run the following command on terminal:

```
chmod +x MetumpX_setup_enUS.sh
sudo bash MetumpX_setup_enUS.sh
```

- 3) Installation wizard of MetumpX will start

*Note: Installation of MetumpX require a proper internet connection to proceed, otherwise the installation terminates.*

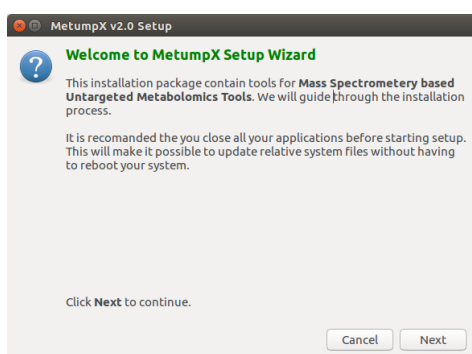


Fig. 9: Installation Welcome Screen

- 4) Click **Next** to proceed and confirm the internet connection.

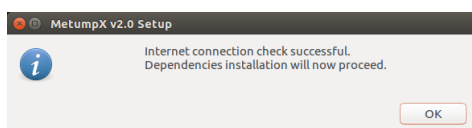


Fig. 10: Internet Check Successful Screen

- 5) Installation is **terminated** if there is no internet.

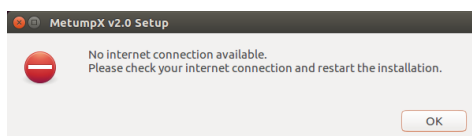


Fig. 11: Internet Check failed screen

- 6) **Dependencies** related to software will start installing.

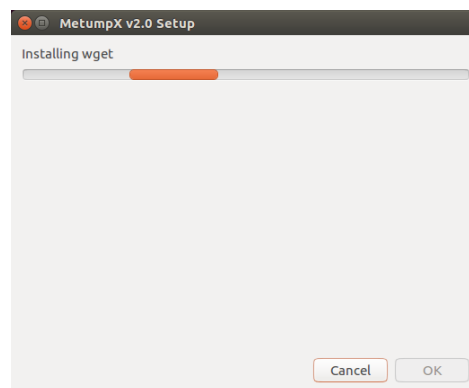


Fig. 12: Installation Screen

- 7) All the dependencies which are installed are displayed at the end of the installation. Click **Ok**.

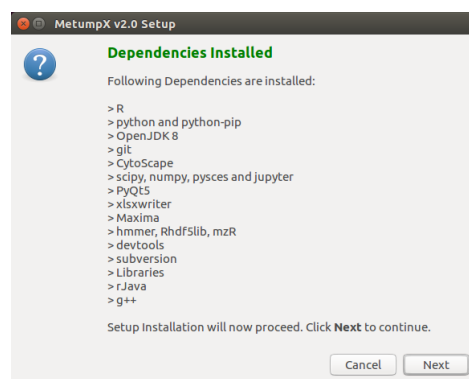


Fig. 13: Dependencies installation finished screen

- 8) Selection screen for Pre-processing Tools is displayed. Select the required tools and Click **Next**.

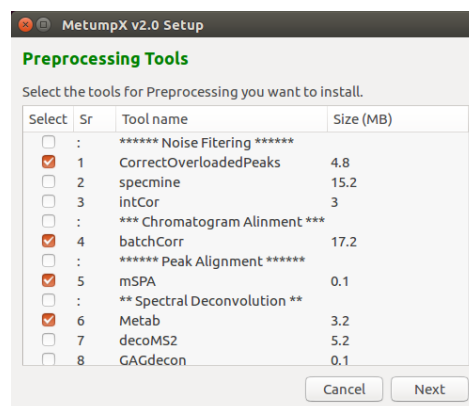


Fig. 14: Preprocessing tools selection screen

- 9) Selection screen for Processing Tools is displayed. Select the required tools and Click **Next**.

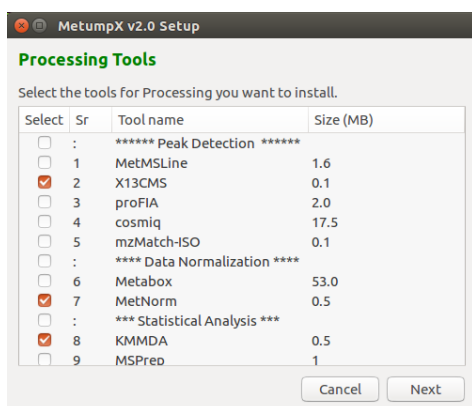


Fig. 15: Processing tools selection screen

- 12) Selection screen for Data Network Analysis tools is displayed. Select the required tools and Click **Next**.

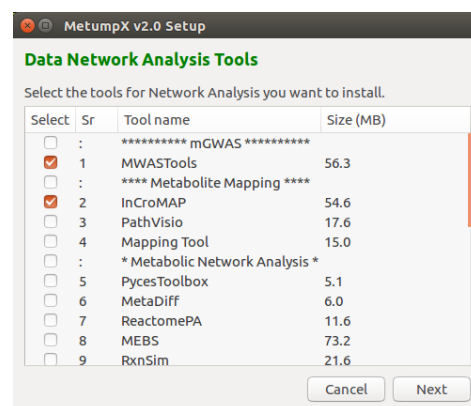


Fig. 18: Data network analysis tools selection screen

- 10) Selection screen for Metabolite Identification tools is displayed. Select the required tools and Click **Next**.

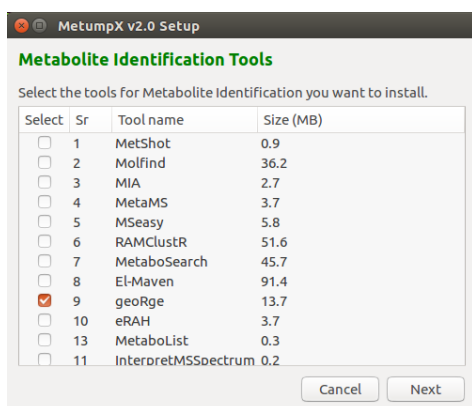


Fig. 16: Metabolite identification tools selection screen

- 13) Selection screen for Enrichment analysis tools is displayed. Select the required tools and Click **Next**.

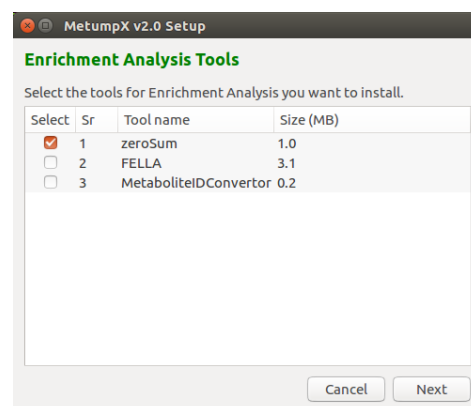


Fig. 19: Enrichment analysis tools selection screen

- 11) Selection screen for Clustering Analysis tools is displayed. Select the required tools and Click **Next**.

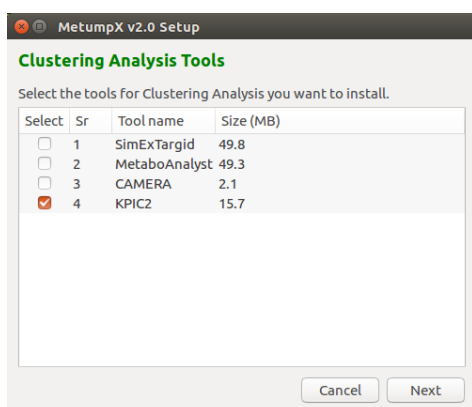


Fig. 17: Clustering analysis tools selection screen

- 14) Selection screen for Integrative analysis tools is displayed. Select the required tools and Click **Next**.

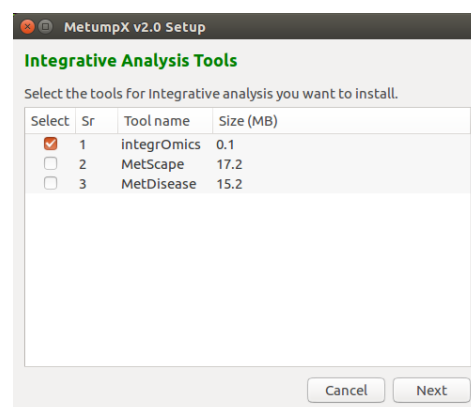


Fig. 20: Integrative analysis tools selection screen

- 15) Selection screen for Data visualization tools is displayed. Select the required tools and Click **Next**.

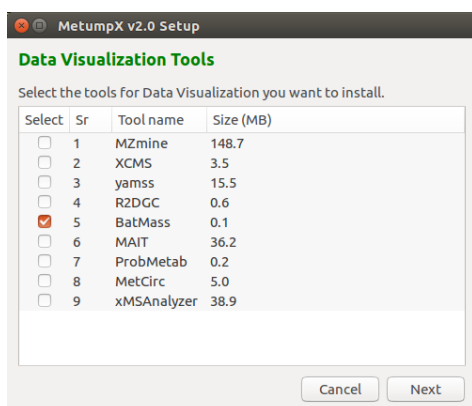


Fig. 21: Data visualization tools selection screen

- 16) Click **Next** to proceed with the installation.

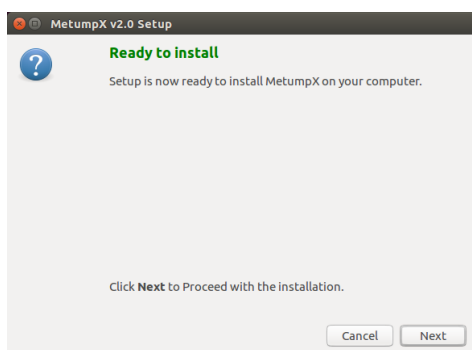


Fig. 22: Ready to Install Screen

- 17) Tool installation will **continue**. Some tools **install** as standalone installation and will be called automatically.

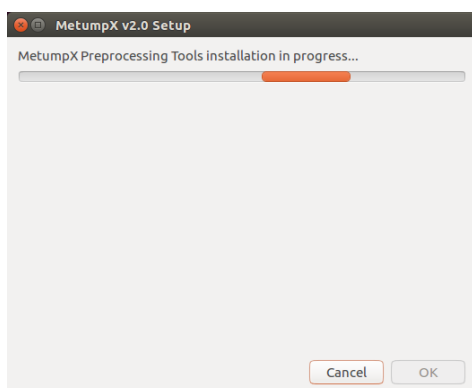


Fig. 23: Tools installation progress screen

- 18) Installation of MetumpX is now finished. Click **Finish** to use the tools.

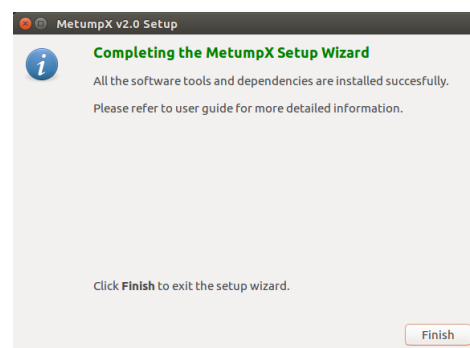


Fig. 24: Finishing Installation Screen

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