

Predicting Profiles of Cochlear Pathology from Non-Invasive Physiological Biomarkers

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INTRODUCTION

Temporal bone histology shows that damage to outer hair cells, inner hair cells, the stria vascularis, and the auditory nerve contribute to sensorineural hearing loss. Physiological measures such as otoacoustic emissions, the middle ear muscle reflex, and the auditory brainstem response are used in research and clinically to detect these pathologies and to determine a more precise site of lesion. However, it remains difficult to establish a one-to-one correlation between physiological responses and histological profiles, especially when more than one pathology is present.

In this study, we examined the relationship between histological profiles of cochlear pathology and responses on a battery of physiological tests sensitive to both OHC and non-OHC dysfunctions. Our preliminary results focused primarily on inner and outer hair cell counts across chinchillas with a variety of exposure histories.

METHODS

Animal Models and Exposure Conditions

Experimentally-Induced Hearing Loss

Noise Induced Temporary Threshold Shift (TTS) - Cochlear Synaptopathy N = 6 animals	Noise Induced Permanent Threshold Shift (PTS) - Complex SNHL N = 10 animals	Control N = 6 animals
Carboplatin (CA) Induced Inner Hair Cell Dysfunction N = 7 animals	Gentamicin Induced Outer Hair Cell Dysfunction N = 5 animals	Mystery Chinchillas Older/High-Risk N = 5 animals

Young chinchillas (6 months to 3 years) were used in the exposure groups and the controls. "Mystery chinchillas" were older animals (3 to 6 years) who had been used for other experiments in the lab.

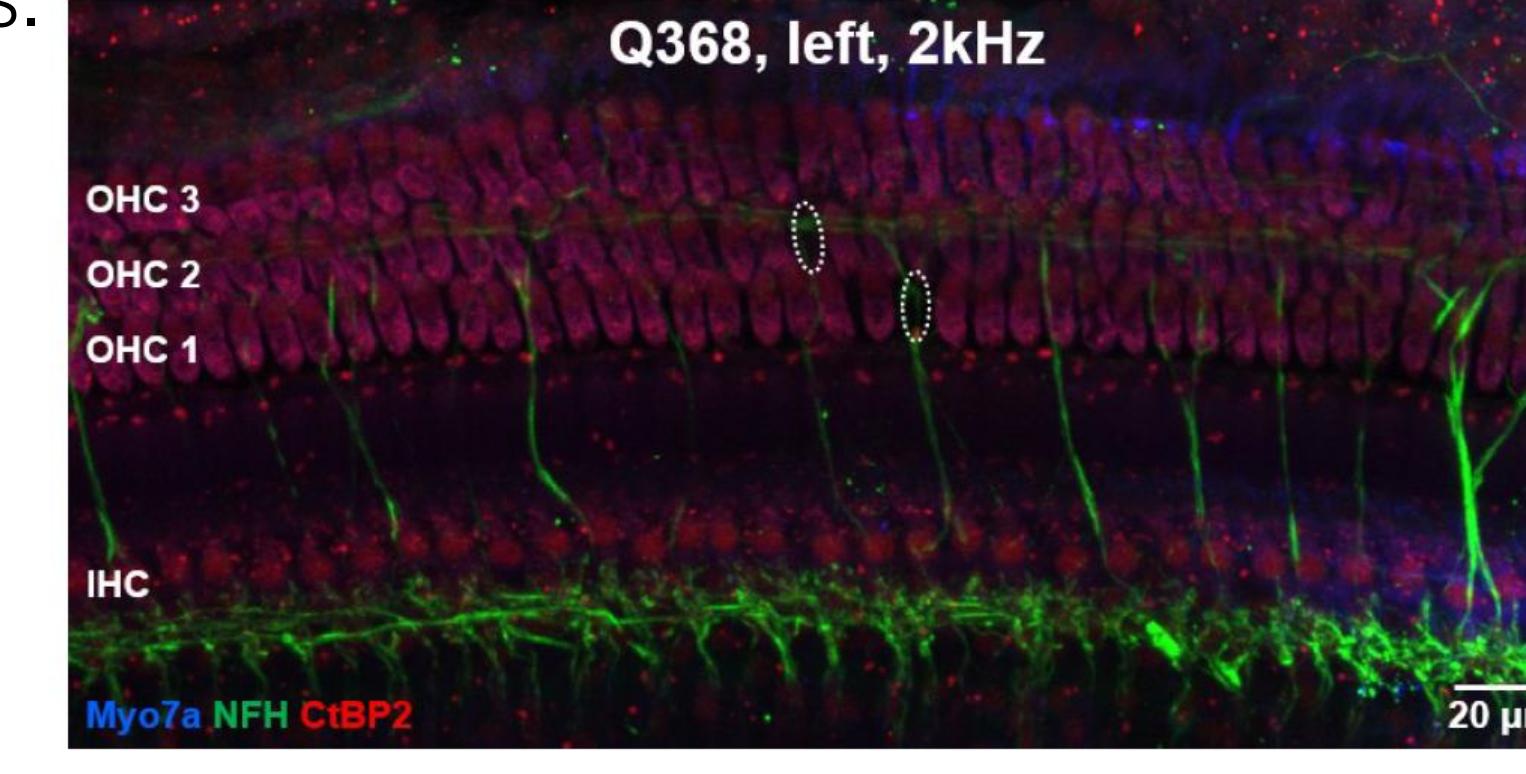
Biomarker Test Battery

ABR Thresholds .5, 1, 2, 4, 8 kHz	Envelope Following Response Rectangular Amplitude Modulated 4kHz tone, 223 Hz modulation	Distortion Product Otoacoustic Emissions
Wideband Middle-Ear Muscle Reflex	Click ABR High level stimulus	Stimulus Frequency Otoacoustic Emissions

Physiological assessment with the above battery was performed pre-exposure (see Control Group physiology) and two weeks after exposure. Physiological results shown here include all animals from a given paradigm whether or not histological data was included. Physiological responses are shown for the tests that contributed most to classifying across exposure groups and are normalized to the mean and standard deviation of the pre-exposure results (37 animals).

Cochlear Histology

After all testing was completed, animals were euthanized and perfused. Cochleas were harvested, post-fixed, and decalcified. Cochleas were then bisected through the modiolus, immunolabeled, and dissected into half turns.

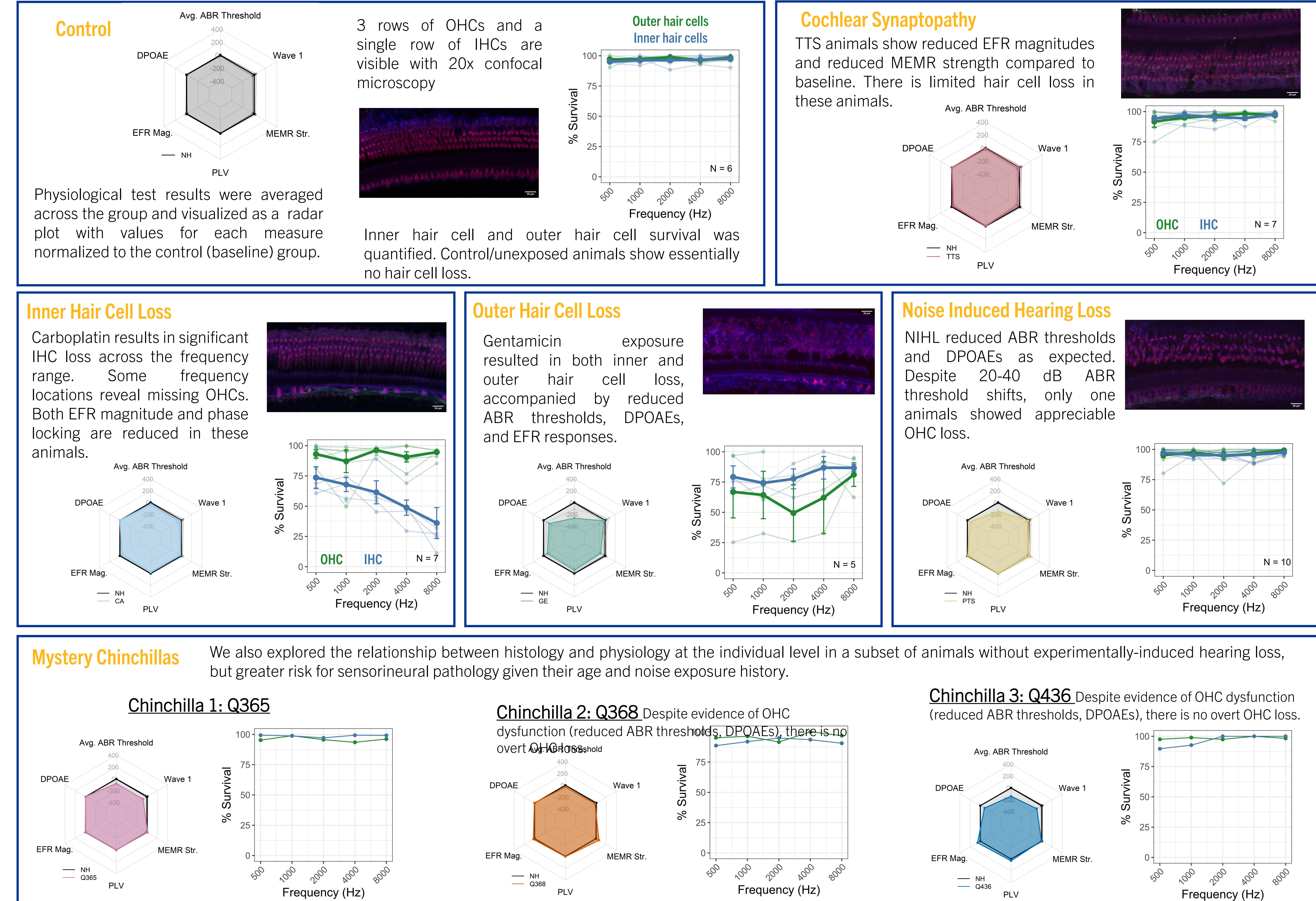


Specimens were labeled for:

- hair cells (*Myo7a*)
- afferent neurons (*NFH*)
- hair cell presynaptic ribbons (*CtBP2*)
- cholinergic efferent neurons (*ChAT*)
- In a subset, stereocilia (*ESPN*)

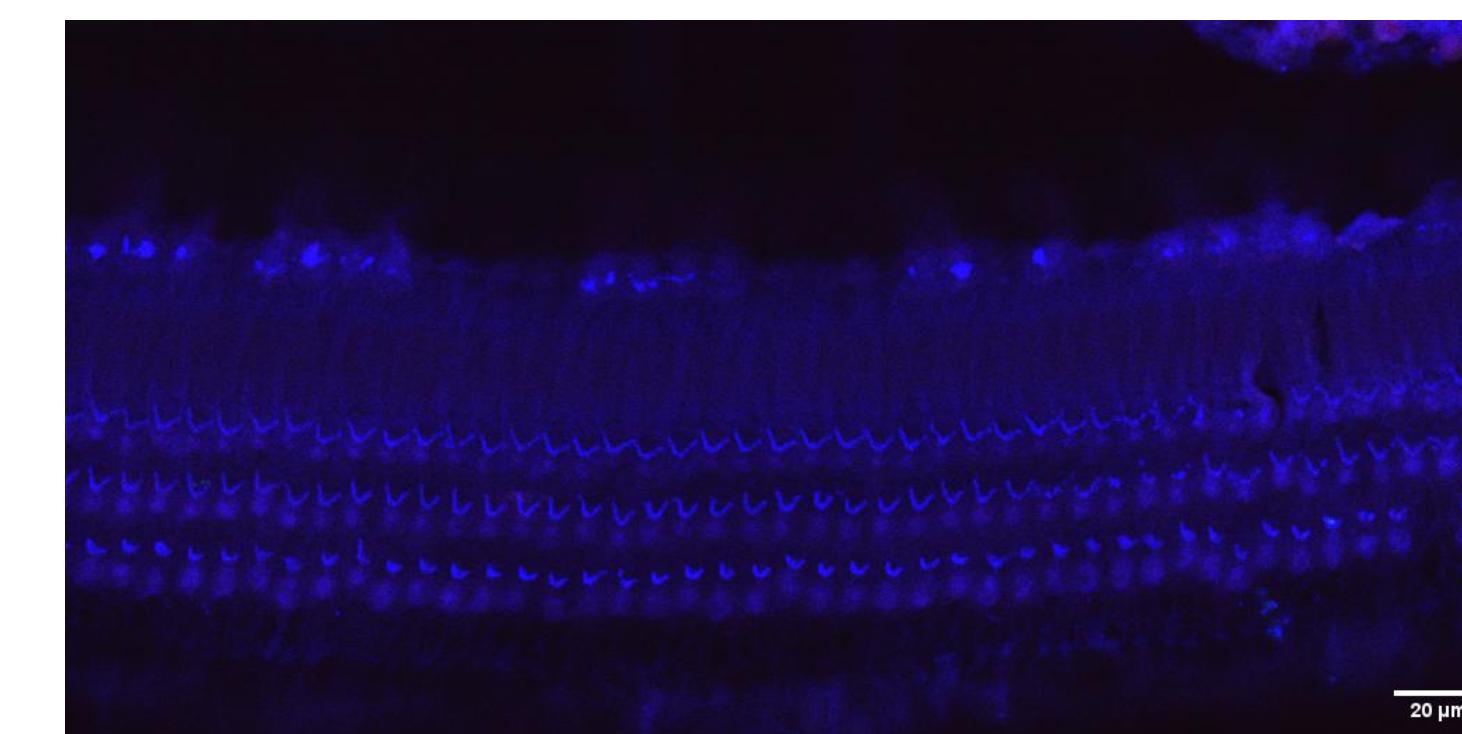
Confocal z-stack images were obtained at .5, 1, 2, 4, and 8 kHz cochlear locations. Hair cell counts were obtained at each of the 5 cochlear locations when dissection and image quality allowed. Hair cell survival is averaged across ears when both ears were available, otherwise, only data from one ear is shown.

RESULTS



FUTURE DIRECTIONS

Future datasets will be analyzed for other features such as synapses, auditory nerve fibers, and stereocilia.



Stereocilia from a carboplatin-exposed animal shows IHC stereocilia disarray in remaining cells.

CONCLUSIONS

- Hair cell counts alone are likely inadequate to account for the full range of pathology possible and more subtle effects such as stereocilia damage.
- These precision biomarkers show promise for non-invasively assessing inner ear health.

REFERENCES

- [1] XXX [2] XXXX

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