1	Modeling amino acid requirements in dairy cows
2	
3	Interpretive Summary: Higgs et al. Improved predictions of the net and optimum AA supply
4	to dairy cows in ration formulation systems could provide an opportunity to balance diets closer
5	to animal requirements and improve nutrient utilization. Predictions of AA supply in a dynamic
6	version of the Cornell Net Carbohydrate and Protein System (CNCPS) were refined by
7	modeling endogenous N transactions along the entire gastrointestinal tract including
8	incorporation into microbial N supply. A strong relationship was observed when the efficiency
9	of AA use was regressed against AA supply relative to ME suggesting expressing AA supply
10	relative to energy could improve predictions of AA utilization.
11	
12	Version 7 of the Cornell Net Carbohydrate and Protein System: III. Endogenous nitrogen
13	and amino acid requirements
14	
15	R. J. Higgs,* D. R. Ouellet,† H. Lapierre,† E. A. Collao-Saenz,‡ H. R. Kingi,§ and M. E.
16	Van Amburgh*1

- 18
- †Research and Development Centre, Agriculture and Agri-Food Canada, Sherbrooke, QC,
- 19 Canada J1M 0C8

‡ Department of Animal Science, Universidade Federal de Goiás, Jataí-GO, Brazil 20

*Department of Animal Science, Cornell University, Ithaca, NY 14853

§ Department of Economics, Cornell University, Ithaca, NY 14853 21

22 ABSTRACT

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

Improved predictions of the net and optimum AA supply to dairy cattle in ration formulation models like the Cornell Net Carbohydrate and Protein System (CNCPS) would provide an opportunity to balance diets closer to animal requirements and improve nutrient utilization. Predictions of net AA supply in a dynamic version of the CNCPS were refined by modeling endogenous N (EN) transactions along the entire gastrointestinal tract (GIT) including incorporation of EN into microbial N supply. Studies that used isotopic enrichment of total N from 15N-Leu infusion to mark endogenous components were used to develop the model. Predictions were close to measured data at the duodenum, ileum and in the feces. Incorporation of EN into microbial N and the original source of EN at various points in the GIT and in the feces were also accurately predicted. The optimum AA supply was determined using a dataset of published studies that infused AA post-ruminally. A logistic model was used to estimate AA requirements for the physiological processes quantified by the model. The optimum AA supply to maximize AA use and minimize wastage was determined where the third derivative of the logistic model was 0. The optimum AA supply was described for all essential AA. Requirements for Met (5.7% EAA) and Lys (15.1 % EAA) were similar to previous recommendations, indicating the model was within the range of current data. A log-logistic relationship was observed when the efficiency of AA use was regressed against AA supply relative to ME supply but no relationship was found when AA supply was expressed relative to MP. Considering AA supply relative to energy could improve predictions of AA utilization and indicates the relationships are not separate and linear, but integrated, and depend on the profile of nutrients consumed.

Key Words: CNCPS, amino acid, endogenous nitrogen, requirement, dairy cattle

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

INTRODUCTION

An improved understanding of both the net and optimum supply of AA to a dairy cow can provide an opportunity to supply AA closer to animal requirements and reduce total protein feeding while still maintaining high levels of production (Haque et al., 2012). This strategy can reduce feed costs and lower the environmental impact of dairy production (Higgs et al., 2012). Amino acids flowing to the duodenum encompass proteins originating from three major sources: undegraded feed, microbial and endogenous proteins (Lapierre et al., 2006). Combined, these fractions represent the gross AA supply, potentially available to the animal after digestion. However, quantification of the endogenous N (EN) fraction, and its contribution to the microbial pool of AA, is needed to establish the net AA supply (Ouellet et al., 2002). The contribution of EN to the duodenal AA flow, either as free proteins or incorporated into microbial protein, represents a recycling of previously absorbed AA that cannot be considered new supply, but is significant (Lapierre et al., 2006). Currently, the prediction of AA supply in the Cornell Net Carbohydrate and Protein System v6.5 (CNCPS) is the sum of AA from feed and bacteria that escape the rumen and are digested in the small intestine and does not consider endogenous AA or protozoa (O'Connor et al., 1993). Incorporating both endogenous AA and protozoa into the CNCPS could refine and possibly improve predictions of the net supply of AA to the animal.

63

64

65

66

67

68

Requirements in the CNCPS are first calculated individually by quantifying different physiological processes, including their AA composition, then dividing these by a transfer coefficient (efficiency of use): the results of each physiological process are summed to give total AA requirements (O'Connor et al., 1993). Previous versions of the CNCPS have assumed the protein and AA requirements for maintenance are the sum of scurf, endogenous urinary and

metabolic fecal N (Fox et al., 2004). Metabolic fecal N (MFN) has been estimated using regression techniques in the previous versions of CNCPS and the NRC (2001) using the work of Swanson (1977). Fox et al. (2004) suggested these calculations might have shortcomings due to the contribution of microbial N from hind gut fermentation to total fecal N. The regression techniques used would consider fecal microbial N as EN. Therefore, the protein or AA requirement for maintenance estimated by a model using these predictions might be overestimated. The assumption used when considering MFN in the maintenance requirement of an animal is that for the protein to be a true requirement and net loss, it needs to be excreted. However, considerably more EN is secreted into the rumen of dairy cows than escapes in the free form or is incorporated in bacteria (Ouellet et al., 2002; Marini et al., 2008; Ouellet et al., 2010a). This means the balance is degraded in the rumen and the N absorbed as ammonia. Once degraded, essential AA are lost for the animal and can only be replaced by absorbed AA, originating, on a net basis, from RUP or microbial protein (MCP). Therefore, it makes sense to consider all protein secreted into the gastrointestinal tract (GIT) which is not recovered in the small intestine a maintenance requirement, not just what appears in the feces.

The objectives of this study were to replace current predictions of MFN with estimations of EN transactions (AA, excludes urea) through the entire GIT in a dynamic version of the CNCPS (Higgs and Van Amburgh CHO and PRO and Higgs and Van Amburgh microbial growth). In doing this, the net supply of AA to the small intestine from all sources can be refined and the shortcomings of the current predictions improved. A second objective was to evaluate the efficiency of transfer of AA to milk and maintenance using the predicted net supply and requirements of the new model. Interactions between protein and energy play an important role in determining how an animal will utilize absorbed AA and it has been recommended they be considered together (Hanigan et al., 1998; Lobley, 2007). Current

European models are incorporating energy supply or DMI in the estimation of the efficiency of MP use (NorFor, 2011; Van Duinkerken et al., 2011; Sauvant et al., 2015). These interactions were investigated in determining the optimum AA requirements for this revised version of the model.

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

94

95

96

97

MATERIALS AND METHODS

Modeling endogenous AA losses in the gut

Predictions of EN losses into the GIT were modeled mechanistically to capture the various transactions along the GIT and between microbial pools. Gross EN to the forestomach and intestines were estimated according to Ouellet et al. (2002) and Ouellet et al. (2010a) which were subsequently partitioned into individual components (Table 1) using estimates reported in Egan et al. (1984). The studies by Ouellet and co-workers directly estimate EN using 15N-Leu intra-venous infusion for 8 d, in cows with rumen, duodenal and ileal cannulas. Using this technique, different labelled precursor pools are available to represent the different sites of EN production which have different isotopic enrichments. In dairy cattle, the enrichment of milk probably provides a good representation of protein secretions whereas the rumen or intestinal mucosa represents the pool contributing to EN through cell desquamation (Ouellet et al., 2002). Enrichment of the rumen mucosa was used to estimate the contribution of EN to MCP as EN contributions to the rumen would largely be from desquamation (Egan et al., 1984). Free EN at the duodenum was assumed to be best represented by the 'combined' precursor pool, the average of rumen mucosa and milk (Ouellet et al., 2010a), due to the contribution of both sloughed cells and secretions into the abomasum and duodenum upstream of the cannula. Data using a 'combined' precursor pool are not presented in Ouellet et al. (2002). Therefore, the relative difference between the estimates using 'combined' and 'mucosa' precursor pools

(combined = 60% of mucosa) presented in Ouellet et al. (2010a) were used to calculate estimates of EN flows with a combined pool from Ouellet et al. (2002). Endogenous secretions early in the small intestine were assumed to be largely digested. Therefore, EN measured at the ileum and in the feces would predominantly be from sloughed keratinized cells with poor digestibility and would be best represented by the mucosa precursor pool. Endogenous contributions are reasonably consistent among diets when expressed relative to DMI or OMI (Tamminga et al., 1995; Ouellet et al., 2002; Marini et al., 2008; Ouellet et al., 2010a). Thus, the model expresses each component as g EN per kg DMI. Quantitative estimates of fluxes to and from the various pools in the model were estimated by setting the kinetic parameters and digestibility coefficients in the model to align predictions at various points in the gut to measured data (Ouellet et al., 2002 and 2010a). A summary of the model inputs used to estimate the EN transactions are in Table 1.

Table 1 somewhere here

Endogenous N in the rumen has three potential fates: 1) degradation to ammonia; 2) incorporation into MCP; 3) escape from the rumen. Degradation and passage are estimated using the kinetic relationships described in Higgs and Van Amburgh (CHO and PRO) where free EN is assumed to flow in the liquid phase. Incorporation into MCP is estimated using two extensions of the microbial model described in Higgs and Van Amburgh (microbial growth). The first (Figure 1) is used to replicate the enrichment of 15N in MCP after an 8 d intra-venous infusion of 15N-Leu for the purpose of estimating the kinetic and digestibility coefficients of EN secretions. The studies of Ouellet exclude the transfer of 15N from recycled urea, but it is still possible for 15NH3 to be produced in the rumen by fiber bacteria (**FB**), non-fiber bacteria (**NFB**) and protozoa (**PZ**) and incorporated into microbial protein. The model assumes if EN

is degraded to NH₃, the AA are lost to the animal, and are only recoverable if incorporated into microbial protein intact. Therefore, the second model (Figure 2) uses the inputs estimated from the first model but excludes transfers through the NH₃ pool and is used to estimate true EN AA uptake by the microbes and subsequent endogenous AA recovery.

Transactions in the first model (Figure 1) begin with labeled EN (LEN) that is degraded (LEN to R) and enters the peptide and free AA (PAA) pool in the rumen (LEN PAA R). From there, the LEN can escape (LEN PAA Escape), be degraded to NH3 (LEN PAA Deg) or be taken up as PAA by NFB (LEN PAA Uptake NFB) or PZ (LEN PAA Engulfment). Protozoa either incorporate the LEN (PZ LEN Engulfed Incorporated), excrete it as PAA (PZ LEN Engulfed excreted as PAA), or excrete it as NH3 (PZ LEN Engulfed excreted as NH3). Labelled PZ can escape the rumen (PZ Cell LEN Escape) or lyse (PZ Cell LEN Lysis). Protozoal excretion of PAA, NH3 and lysis has the effect of transferring EN through numerous rumen N pools and also allows FB to be enriched through the labeled NH3 pool (NH3 LEN R) which can also escape (FB Cell LEN Escape). Enrichment of MCP through the NH3 pool is not considered available for recovery given the AA itself has been degraded. Therefore, these same transactions are considered in second model with LEN transfers through the NH3 excluded (LEN*; Figure 2).

Figure 1 somewhere here

Figure 2 somewhere here

Each individual source of EN can be quantified within the model, as either free EN, or incorporated in MCP, from the initial transfer into the gut to its final fate. An AA profile is applied to each component using the profiles in Table 2. Microbial AA of endogenous origin

are not considered new supply and are subtracted off digested microbial AA using the profile of the original source (Table 2). Endogenous AA in MCP are assumed to be evenly distributed through the cell N and digestion is relative to the digestion of total MCP. Free EN can be recovered if digested in the small intestine; otherwise the AA are either fermented in the hindgut or excreted in the feces and are considered lost. Overall, losses occur from degradation and absorption as NH₃ in the rumen and large intestine, or excretion in the feces. The requirement for endogenous AA can, therefore, be calculated as total entry into the gut less recovery in the small intestine.

Table 2 somewhere here

Estimating total AA requirements

Amino acid net requirements (**AAnR**) estimated in the CNCPS v6.5 include those for milk, growth, reserves, pregnancy, scurf, metabolic urinary losses and endogenous losses in the gastrointestinal tract (GIT). Endogenous N losses in the GIT in this model are calculated as described above whereas the other requirements, including AA profiles, are estimated according to Fox et al. (2004). Amino acids used for other processes not accounted for by the model (**AAO**) can be calculated as the difference between predicted AA supply (**AAS**) and AAnR. The term often used to estimate AAnR relative to AAS is 'efficiency of use' which can vary depending on AA supply relative to other nutrients and the physiological state of the animal (Hanigan et al., 1998; Doepel et al., 2004). In order to balance a diet in which individual EAA supply is not excessive, but also not limiting, estimates of AAO relative to AAnR are required. In this model, the approach used to generate these estimates was similar to the study of Doepel et al. (2004). Briefly, a dataset was constructed of studies that infused AA into the abomasum, duodenum, or intravenously (Table 3). Infusion studies were used so that the

addition of AA above the basal diet was known and limited the reliance on model predictions (Doepel et al., 2004). The final dataset included 41 publications, 51 experiments and 218 treatment means. Descriptive statistics for the dataset are in Table 4. Dietary and animal characteristics reported in the publications were entered into the CNCPS. When limited information was reported on the chemical composition of the dietary components, the reported information was used, and uncertain values were predicted using an extension of the method described in Higgs et al. (2015). Briefly, it was assumed that the feeds used in different treatments in the same study had the same chemical composition. The procedure optimized each chemical component in each feed to be within a likely range, to be internally consistent (chemical components sum to 100% DM) and to allow the compiled diet to match the reported composition. As described previously, infused AA were assumed to be 100% digested (Doepel et al., 2004). Once compiled, each treatment was evaluated through the model to estimate AAS and AAnR for each of the 10 EAA.

Table 3 somewhere here

Table 4 somewhere here

A logistic model with three parameters was used to fit the data which was previously shown to give the most appropriate fit (Doepel et al., 2004). The selected model has the form

$$213 y = \frac{\theta_1}{1 + \theta_2 e^{\theta_3(x)}} [1]$$

where y is the AAnR (g/d), x is the predicted AAS (g/d), θ_{1-3} are the model parameters used to described the sigmoidal shape of the curve. The optimum supply of AA was considered to be

the point on the curve where the rate of change in the ratio of AAnR:AAS was the most rapid, or, in other words, the rate at which cows were changing the way they managed additional AAS was most rapid (Figures 3 and 4). This can be calculated when the third derivative of the logistic model is zero. The third derivative has the form

$$222 \quad \frac{d^3y}{dx^3} = -\theta_1\theta_2\theta_3^{\ 3}e^{\theta_3(x)} \frac{1 - 4\theta_2e^{\theta_3(x)} + \theta_2^{\ 2}e^{2\theta_3(x)}}{(1 + \theta_2e^{\theta_3(x)})^4}$$
[2]

and the zero point of interest is calculated using the equation

$$226 x = \frac{1}{\theta_3} \log \left(\frac{2 - \sqrt{3}}{\theta_2} \right) [3]$$

where x is considered the optimum AAS for the dataset used. By substituting x into equation [1], and dividing y (AAnR) by x (AAS) the optimum ratio of AAnR to AAS can be calculated, and indirectly, the optimum level of additional AA for other functions not considered by the model. When balancing a diet, the total required supply (g AA/d) can be calculated by dividing AAnR by the optimum ratio of AAnR to AAS. The same calculations were also performed for MP.

The relationship between ratio of AAnR and AAS and AA supply relative to other nutrient supplies (g AA/ Mcal ME and g AA/ 100g MP) was also investigated. A log-logistic model with three parameters was used to fit this relationship with the form

239
$$y = \theta_1 - \log(1 + \theta_2 e^{-\theta_3(x)})$$
 [4]

where y is the ratio of AAnR to AAS, x is the AA supply expressed relative to Mcal of total ME supply or relative to 100 g MP, and θ_{1-3} are the model parameters used to describe the shape of the curve. The optimum supply of a given EAA relative to ME or MP can then be found by rearranging formula [4] and solving for x using the AAnR:AAS (y) previously calculated.

$$247 x = \frac{-1}{\theta_3} log\left(\frac{e^{\theta_1 - y} - 1}{\theta_2}\right) [5]$$

Given the information presented by studies published in the literature is typically limited compared to the inputs required by the CNCPS, a large number of assumptions had to be made. To limit the influence of potential input errors, points were weighted on the likelihood of being an outlier. The scheme used was the Tukey Biweight and was implemented according to Motulsky and Christopoulos (2004). Data analysis was performed using the non-linear modelling function in SAS (2010).

RESULTS AND DISCUSSION

Endogenous N flows

The mechanistic framework developed in Higgs and Van Amburgh (CHO and PRO) and Higgs and Van Amburgh (microbial growth) enabled EN to be modeled in all parts of the GIT including the microbial transactions in the rumen and large intestine in a manner that reconciled entrance and disposal of both N and AA by compartment. Model estimates compared to measurements taken from multi-cannulated animals in the studies of Ouellet and coworkers are in Table 5. Model predicted flows of EN at the duodenum were similar to measured values.

The greatest difference was observed in the prediction of microbial EN in the 'Inoc' and 'Formic' treatments (Ouellet et al., 2010a). The model assumes microbes do not differentiate between the original source of N in the rumen with uptake being based on the relative availability of each source (Marini et al., 2008). Silages fed in the 'Inoc' and 'Formic' treatments had higher levels of soluble protein than the hay treatment (Martineau et al., 2007) which increased the availability of feed N in the rumen relative to EN and resulted in lower predicted microbial uptake of EN. The rate of CHO digestion in the rumen also impacts predictions of EN uptake through its effect on microbial growth (see Higgs and Van Amburgh microbial growth). Therefore, more accurate estimates of the CHO digestion kinetics could improve the present model predictions. Although differences in EN secretion into the foregut among dietary treatments has been observed (Ouellet et al., 2010a), the mechanism of action is still unclear (Larsen et al., 2000). Therefore, expressing EN secretion relative to DMI seemed appropriate until the factors involved are better understood. Further down the GIT, estimates were similar to measured values at the terminal ileum and in the feces (Table 5).

Total EN transactions through each compartment in the model for the 'Hay' treatment in Ouellet et al. (2010a) are summarized in Figure . These data were generated using the model in Figure 2 where EN transfers through the NH₃ pool were excluded. The 'Hay' treatment was chosen given the close agreement between model and measured values. Total EN secretions into the GIT were 135.4 g/d of which 46.4 g/d was recovered as absorbed AA from intestinal digestion originating from either the forestomach or the small intestine. The EN contribution to total rumen N supply was 90 g/d and demonstrates an additional method by which the cow provides nutrients to support the microbial population. Previous versions of the CNCPS, based on the data from Russell et al. (1992), indicate dietary provision of peptides was necessary to enhance the fermentation and growth of NFB. However, the data from Ouellet et al. (2010a)

as described in this model demonstrate that large quantities of peptides are provided to the microbes in the form of EN reducing the reliance on dietary sources. Endogenous N not recovered (89.0 g/d) was considered lost by the animal and is part of the maintenance requirements for protein. Of the 89.0 g/d lost, 31.8 g/d appeared in the feces and 57.2 g/d was degraded in the GIT to NH₃. The total estimated requirement (89.0 g/d) when expressed relative to DMI is 5.1 g EN/ kg DMI which, interestingly, is similar to current model (CNCPS v6.5, Van Amburgh et al. 2015) estimates of MFN for the same diet (5.0 g MFN/kg DMI).

Figure somewhere here

Table 5 somewhere here

Amino acid requirements

Requirements for each individual EAA in the CNCPS v6.5 are predicted from the net protein required for each physiological process, quantified by the model (maintenance, lactation, pregnancy, growth) multiplied by the AA composition of each process and subsequently divided by the efficiency of transfer to that process to give the total AA requirement (O'Connor et al., 1993; Fox et al., 2004). The efficiency of transfer could also be considered as the additional usage of each AA relative to the requirements quantified by the model. Such processes include oxidation across tissues, anaplerotic requirements, synthesis of non-essential AA, gluconeogenesis etc. (Lapierre et al., 2005; Lapierre et al., 2006; Lemosquet et al., 2010; Lobley, 2007). The apparent efficiency of AA use for any given diet can be calculated by dividing model predicted AAnR by AAS, which can be variable, and typically decreases as AAS increases relative to AAnR (Hanigan et al., 1998; Doepel et al., 2004); the apparent efficiency has also been reported to decrease when the ratio of MP relative to energy supply increases (NorFor, 2011; van Duinkerken et al., 2011). The ability of cows to direct AA

to other uses demonstrates the interactions among different nutrients and is an example of the metabolic flexibility that allows productivity to be maintained across a wide range of nutrient inputs and supply (Lobley, 2007). The pertinent question for diet formulation is: what level of additional AA supply above the predicted requirements for milk protein synthesis, body protein requirements and other metabolic needs will maximize productivity and minimize AA wastage? The answer to this question is going to differ among models as supply and requirements are calculated in different ways. For example, changing the maintenance requirements from using MFN as in previous version of the CNCPS to estimating AA loss through the GIT using isotopic enrichment techniques considers 9 different sources of EN, each with a different AA profile (Table 2), and so it would be expected that AA requirements among models will be different.

The optimum supply of EAA in this study was defined where the logistic curve was approaching a plateau most rapidly. This point was defined by Doepel et al. (2004) as an optimal supply and is similar to the break-point in the segmented linear model used in the NRC (2001). Previous versions of the CNCPS have treated different physiological functions separately with the original values coming from a range of sources outlined in O'Connor et al. (1993). Lapierre et al. (2007) suggested using a single efficiency of use to calculate total AA requirement for maintenance and milk production. Amino acid catabolism is linked to the tissue where enzymes are present to ensure catabolic pathways (Lobley and Lapierre, 2003) and is not linked to the site of protein secretion. For example, there is no catabolism of Phe in the mammary gland, independently of the Phe supply (Lemosquet et al., 2010); Phe is almost exclusively removed by the liver (Lapierre et al., 2005). Recommendations for fixed efficiencies of use of individual AA were presented by Lapierre et al. (2007) and have been implemented in the most recent update of the CNCPS (v6.5; Van Amburgh et al., 2015). Model

parameters and the fit summary for the logistic model used to make the calculations in this study are in Table 6. The variation explained in the present study by the logistic model was similar to Doepel et al. (2004). Examples of model fit and optimum supply for Met and Lys are in Figures 3 and Figure. The optimum ratio of model predicted AAnR to AAS for each AA and MP are in Table 6. As explained, it is difficult to compare the ratio of AAnR:AAS among studies due to the different approach models used to calculate AAnR. However, it is possible to compare the optimum AAS expressed as % EAA and also in g/d relative to the study of Doepel et al. (2004) given the similarities in the datasets. The required supply and balance of EAA in the current study compared with Doepel et al. (2004) are fairly similar despite the differences in the models used to estimate supply.

Table 6 somewhere here

Figure 3 somewhere here

Figure somewhere here

This concept is important when discussing how high the efficiency of use of an AA should be. Does a high efficiency describe efficient use of an AA or does it describe a relative deficiency of this particular AA? On a general basis, the higher the efficiency the more restricted the supply is relative to the requirement. As the AA becomes less 'limiting' up to an optimum, the amount of milk, milk protein or energy corrected milk increases until a plateau is achieved, and any AA supplied beyond that point is not utilized with the same efficiency for the expected outcome (Rulquin et al., 1993; Schwab, 1996). Thus, a high efficiency of use more likely reflects an under-supply of AA relative to the available energy, other AA or both.

Interactions between AA supply and energy

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

The impact of energy supply on the utilization of AA was investigated by regressing the ratio of AAnR and AAS (efficiency of use) against AA supply relative to total ME and total MP supply. No relationship was found when AA were expressed relative to MP, but a loglogistic relationship was observed when AA supply was expressed relative to ME. The optimum supply of each EAA relative to ME was determined by using the optimum ratio of AAnR to AAS calculated in the previous analysis and solving for x using the log-logistic model Eq. [5]. Examples of the log-logistic fit and optimum supply relative to ME for Met and Lys are in Figures 3 and 4, respectively. The model parameters, summary of fit and optimum AA supply relative to ME for all 10 EAA are in Table 7. Using the proportional approach, recommendations for AA balancing are made with the ratio of the AA relative to total MP supply. This approach has been successful in establishing Met and Lys requirements from dose response studies (Rulquin et al., 1993; Schwab, 1996; NRC, 2001). The studies used to estimate these requirements are unique in that they isolate the response to the increased supply of Lys, Met or both, from post-ruminal infusion or rumen-protected AA, but holding all other dietary variables constant. The data used in the present study was different in that 81% of the treatments simultaneously infused more than 1 AA with the average number of AA infused > 8. Interestingly, the optimum supply of Met and Lys estimated in this study was 15.1% and 5.7% of EAA, respectively, which is similar to results found in other studies that used different approaches (Schwab et al., 1992b; Rulquin et al., 1993; Schwab, 1996). Under these circumstances, no relationship was observed between the 'efficiency' of AA use when AA supply was expressed relative to MP supply but a strong relationship was observed when AA were expressed relative to ME supply which is in agreement the findings of Van Straalen et al. (1994), whereas Sauvant et al. (2015) used the ratio of AA supply to DMI. This approach is consistent with the relationship between energy intake and endocrine signals to regulate

nutrient partitioning of absorbed nutrients (Bauman and Currie, 1980, McGuire et al., 1995, Bequette et al., 1998, Arriola Apelo et al., 2014). Protein synthesis within the mammary gland is dependent on the energy supply and the associated signals necessary to direct nutrients to the mammary gland for uptake (Lemosquet et al., 2010, Mackle et al., 2000). Therefore, when balancing rations, it might be more appropriate to consider AA supply relative to ME which is the approach used in swine (NRC, 2012). Establishing requirements for monogastrics is less complicated than in ruminants as the net AA supply is more easily determined (Lapierre et al., 2006). Following the monogastric example, the predicted Lys requirement for a lactating sow in the NRC (2012) model is 2.72 g Lys/Mcal ME which is 11% lower than the 3.03 g Lys/Mcal ME calculated in this study for dairy cows. The recommended ratios for each EAA relative to ME and relative to Lys are compared in Table 7: out of the 10 EAA, 5 differ by more than 25%. As improvements are made to the predictions of net AA supply in dairy cows, consideration of the approach used to balance AA in other species where AA supply is more easily determined could provide opportunities to improve productivity and the efficiency of nutrient use.

Figure 3 somewhere here

Figure 4 somewhere here

Table 7 somewhere here

CONCLUSIONS

Predictions of endogenous N transactions along the entire GIT have been incorporated into a dynamic version of the CNCPS (v7). This has replaced metabolic fecal N used in previous versions of the CNCPS in estimating AA requirements for maintenance. Model predictions for

endogenous N transactions along the GIT are close to measured data and have refined the predictions of net AA supply to the animal. Using a logistic approach, efficiency of utilization of individual AA to support the physiological processes was also quantified. The optimum supply of Met and Lys relative to total EAA were similar to other studies. A log-logistic relationship was also observed when the efficiency of AA use was regressed against AA supply relative to ME. Expressing AA supply relative to energy could improve predictions of AA utilization. Recommendations for each EAA are given in g AA/Mcal ME and also in a ratio with Lys.

ACKNOWLEDGEMENTS

Financial support for R. J. Higgs was provided in partnership by Adisseo (Commentry, France) and DairyNZ (Hamilton, NZ).

REFERENCES

- Aldrich, J., L. Muller, and G. Varga. 1993. Effect of somatotropin administration and duodenal
 infusion of methionine and lysine on lactational performance and nutrient flow to the small
- 428 intestine. Br. J. Nutr. 69:49-58.
- 429 Arriola Apelo, S. I., J. R. Knapp, and M. D. Hanigan. 2014. Invited review: Current
- representation and future trends of predicting amino acid utilization in the lactating dairy cow.
- 431 J. Dairy Sci. 97:4000-4017.

- Bauman, D. E., and W. B. Currie. 1980. Partitioning of nutrients during pregnancy and
- lactation: A review of mechanisms involving homeostasis and homeorhesis. J. Dairy Sci.
- 434 63:1514-1529.
- Bequette, B. J., F. R. C. Backwell, and L. A. Crompton. 1998. Current concepts of amino acid
- and protein metabolism in the mammary gland of the lactating ruminant. J. Dairy Sci. 81:2540-
- 437 2559.
- Bruckental, I., I. Ascarelli, B. Yosif, and E. Alumot. 1991. Effect of duodenal proline infusion
- on milk production and composition in dairy cows. Anim. Prod 53:299-303.
- 440 Cant, J., E. DePeters, and R. Baldwin. 1991. Effect of dietary fat and postruminal casein
- administration on milk composition of lactating dairy cows. J. Dairy Sci. 74:211-219.
- Choung, J.-J., and D. G. Chamberlain. 1992a. Protein nutrition of dairy cows receiving grass
- silage diets. Effects on silage intake and milk production of postruminal supplements of casein
- or soya-protein isolate and the effects of intravenous infusions of a mixture of methionine,
- phenylalanine and tryptophan. J. Sci. Food Agric. 58:307-314.
- Choung, J.-J., and D. G. Chamberlain. 1992b. Protein nutrition of dairy cows receiving grass
- silage diets. The effects of post-ruminal supplements of proteins and amino acids. J. Sci. Food
- 448 Agric. 60:25-30.

- Choung, J.-J., and D. G. Chamberlain. 1993. Effects on milk yield and composition of intra-
- 450 abomasal infusions of sodium caseinate, an enzymic hydrolysate of casein or soya-protein
- isolate in dairy cows. J. Dairy Res. 60:133-138.
- 452 Choung, J.-J., and D. G. Chamberlain. 1995a. Effects of abomasal infusions of sodium
- 453 caseinate and of casein hydrolysates varying in the relative proportions of peptides and free
- amino acids on milk production in dairy cows. J. Dairy Res. 62:423-429.
- 455 Choung, J.-J., and D. G. Chamberlain. 1995b. Effects of abomasal infusions of sodium
- 456 caseinate, a hydrolysate of casein or a corresponding mixture of free amino acids on milk yield
- and composition in dairy cows. J. Dairy Res. 62:29-37.
- 458 Choung, J.-J., and D. G. Chamberlain. 1995c. The effects of intravenous supplements of amino
- acids on the milk production of dairy cows consuming grass silage and a supplement containing
- 460 feather meal. J. Sci. Food Agric. 68:265-270.
- Clark, J. H., H. R. Spires, R. G. Derrig, and M. R. Bennink. 1977. Milk production, nitrogen
- 462 utilization and glucose synthesis in lactating cows infused postruminally with sodium caseinate
- 463 and glucose. J. Nutr. 107:631-644.
- 464 Cohick, W. S., J. L. Vicini, C. R. Staples, J. H. Clark, S. N. McCutcheon, and D. E. Bauman.
- 465 1986. Effects of intake and postruminal casein infusion on performance and concentrations of
- hormones in plasma of lactating cows. J. Dairy Sci. 69:3022-3031.

- Doepel, L., D. Pacheco, J. J. Kennelly, M. D. Hanigan, I. F. Lopez, and H. Lapierre. 2004.
- 468 Milk protein synthesis as a function of amino acid supply. J. Dairy Sci. 87:1279-1297.
- Doepel, L., and H. Lapierre. 2010. Changes in production and mammary metabolism of dairy
- 470 cows in response to essential and nonessential amino acid infusions. J. Dairy Sci. 93:3264-
- 471 3274.
- Doepel, L., and H. Lapierre. 2011. Deletion of arginine from an abomasal infusion of amino
- acids does not decrease milk protein yield in Holstein cows. J. Dairy Sci. 94:864-873.
- Egan, A. R., K. Boda, and J. Varady. 1984. Regulation of nitrogen metabolism and recycling.
- 475 Pages 386-402 in Proc. International Symposium on Ruminant Physiology. Prentice-Hall,
- 476 Banff, Canada.
- 477 Fox, D. G., L. O. Tedeschi, T. P. Tylutki, J. B. Russell, M. E. Van Amburgh, L. E. Chase, A.
- N. Pell, and T. R. Overton. 2004. The Cornell net carbohydrate and protein system model for
- evaluating herd nutrition and nutrient excretion. Anim. Feed Sci. Technol. 112:29-78.
- 480 Griinari, J. M., M. A. McGuire, D. A. Dwyer, D. E. Bauman, D. M. Barbano, and W. A. House.
- 481 1997. The role of insulin in the regulation of milk protein synthesis in dairy cows. J. Dairy Sci.
- 482 80:2361-2371.
- Guinard, J., and H. Rulquin. 1994. Effects of graded amounts of duodenal infusions of lysine
- on the mammary uptake of major milk precursors in dairy cows. J. Dairy Sci. 77:3565-3576.

- Guinard, J., H. Rulquin, and R. Vérité. 1994. Effect of graded levels of duodenal infusions of
- casein on mammary uptake in lactating cows. 1. Major nutrients. J. Dairy Sci. 77:2221-2231.
- 487 Guinard, J., and H. Rulquin. 1995. Effects of graded amounts of duodenal infusions of
- 488 methionine on the mammary uptake of major milk precursors in dairy cows. J. Dairy Sci.
- 489 78:2196-2207.
- Hamza, A. N. 1976. Rate of protein secretion by sheep pancreas and amino-acid composition
- 491 of pancreatic-juice. Nutri. Reports Inter. 14:79-87.
- 492 Hanigan, M., J. Cant, D. Weakley, and J. Beckett. 1998. An evaluation of postabsorptive
- 493 protein and amino acid metabolism in the lactating dairy cow. J. Dairy Sci. 81:3385-3401.
- Haque, M. N., H. Rulquin, A. Andrade, P. Faverdin, J. L. Peyraud, and S. Lemosquet. 2012.
- 495 Milk protein synthesis in response to the provision of an "ideal" amino acid profile at 2 levels
- 496 of metabolizable protein supply in dairy cows. J. Dairy Sci. 95:5876-5887.
- 497 Higgs, R. J., L. E. Chase, and M. E. Van Amburgh. 2012. Application and evaluation of the
- 498 Cornell net carbohydrate and protein system as a tool to improve nitrogen utilization in
- 499 commercial herds. Prof. Anim. Scient. 28:370-378.
- Higgs, R. J., L. E. Chase, D. A. Ross, and M. E. Van Amburgh. 2015. Updating the Cornell net
- carbohydrate and protein system feed library and analyzing model sensitivity to feed inputs. J.
- 502 Dairy Sci. 98:6340-6360.

- Huhtanen, P. J., H. O. Miettinen, and V. F. Toivonen. 1997. Effects of silage fermentation and
- 504 post-ruminal casein supplementation in lactating dairy cows: 1 Diet digestion and milk
- 505 production. J. Sci. Food Agric. 74:450-458.
- Jansman, A. J. M., W. Smink, P. van Leeuwen, and M. Rademacher. 2002. Evaluation through
- literature data of the amount and amino acid composition of basal endogenous crude protein at
- 508 the terminal ileum of pigs. Anim. Feed Sci. Technol. 98:49-60.
- 509 Kim, C.-H., J.-J. Choung, and D. G. Chamberlain. 1999. Determination of the first-limiting
- amino acid for milk production in dairy cows consuming a diet of grass silage and a cereal-
- based supplement containing feather meal. J. Sci. Food Agric. 79:1703-1708.
- Kim, C.-H., J.-J. Choung, and D. G. Chamberlain. 2000. Variability in the ranking of the three
- 513 most-limiting amino acids for milk protein production in dairy cows consuming grass silage
- and a cereal-based supplement containing feather meal. J. Sci. Food Agric. 80:1386-1392.
- King, K., W. Bergen, C. Sniffen, A. Grant, D. Grieve, V. King, and N. Ames. 1991. An
- assessment of absorbable lysine requirements in lactating cows. J. Dairy Sci. 74:2530-2539.
- Köning, B. A., J. Oldham, and D. Parker. 1984. The effect of abomasal infusion of casein on
- acetate, palmitate and glucose kinetics in cows during early lactation. Br. J. Nutr. 52:319-328.
- Lapierre, H., R. Berthiaume, G. Raggio, M. C. Thivierge, L. Doepel, D. Pacheco, P. Dubreuil,
- and G. E. Lobley. 2005. The route of absorbed nitrogen into milk protein. Anim. Sci. 80:10-
- 521 22.

- Lapierre, H., D. Pacheco, R. Berthiaume, D. R. Ouellet, C. G. Schwab, P. Dubreuil, G. Holtrop,
- and G. E. Lobley. 2006. What is the true supply of amino acids for a dairy cow? J. Dairy Sci.
- 524 89:E1-E14.
- 525 Lapierre, H., G. E. Lobley, D. R. Ouellet, L. Doepel, and D. Pacheco. 2007. Amino acid
- requirements for lactating dairy cows: Reconciling predictive models and biology. Pages 39-
- 527 59 in Proc. Cornell Nutrition Conference for Feed Manufacturers. Department of Animal
- 528 Science, Cornell University, Syracuse, NY.
- Lapierre, H., L. Doepel, E. Milne, and G. Lobley. 2009. Responses in mammary and splanchnic
- metabolism to altered lysine supply in dairy cows. Anim. 3:360-371.
- Larsen, M., T. G. Madsen, M. R. Weisbjerg, T. Hvelplund, and J. Madsen. 2000. Endogenous
- amino acid flow in the duodenum of dairy cows. Acta Agric. Scand., Sec. A Anim. Sci.
- 533 50:161-173.
- Lemosquet, S., J. Guinard-Flament, G. Raggio, C. Hurtaud, J. Van Milgen, and H. Lapierre.
- 535 2010. How does increasing protein supply or glucogenic nutrients modify mammary
- metabolism in lactating dairy cows? Pages 175-186 in Proc. Energy and protein metabolism
- and nutrition. Wageningen Academic Publishers, Parma, Italy.
- Lemosquet, S., G. E. Lobley, R. Koopman, L. J. C. van Loon, A. K. Kies, and H. Lapierre.
- 539 2010. A large supply of phenylalanine is not oxidised by the mammary gland of dairy cows
- Pages 139-140 in Energy and Protein Metabolism and Nutrition. EAAP publication No.127.
- Ed. C.M. Crovetto, Wageningen Academic Publishers. The Netherlands.

- Lobley, G. E., and H. Lapierre. 2003. Post-absorptive metabolism of amino acids. Pages 737-
- 544 756 in Progress in research on energy and protein metabolism. EAAP publication No.109. Ed.
- 545 W.B. Souffrant and C.C. Metges.

- Lobley, G. E. 2007. Protein-energy interactions: horizontal aspects. Pages 445-462 in Proc.
- 548 Energy and protein metabolism and nutrition. Butterworths, Vichy, France.
- Lynch, G. L., T. H. Klusmeyer, M. R. Cameron, J. H. Clark, and D. R. Nelson. 1991. Effects
- of somatotropin and duodenal infusion of amino acids on nutrient passage to duodenum and
- performance of dairy cows. J. Dairy Sci. 74:3117-3127.
- Mackle, T., D. Dwyer, and D. Bauman. 1999a. Effects of branched-chain amino acids and
- sodium caseinate on milk protein concentration and yield from dairy cows. J. Dairy Sci.
- 554 82:161-171.
- Mackle, T. R., D. A. Dwyer, K. L. Ingvartsen, P. Y. Chouinard, J. M. Lynch, D. M. Barbano,
- and D. E. Bauman. 1999b. Effects of insulin and amino acids on milk protein concentration
- and yield from dairy cows. J. Dairy Sci. 82:1512-1524.
- Mackle, T. R., D. A. Dwyer, K. L. Ingvartsen, P. Y. Chouinard, D. A. Ross, and D. E. Bauman.
- 559 2000. Effects of insulin and postruminal supply of protein on use of amino acids by the
- mammary gland for milk protein synthesis. J. Dairy Sci. 83:93-105.

- Marini, J. C., D. G. Fox, and M. R. Murphy. 2008. Nitrogen transactions along the
- gastrointestinal tract of cattle: A meta-analytical approach. J. Anim. Sci. 86:660-679.
- Martineau, R., H. Lapierre, D. R. Ouellet, D. Pellerin, and R. Berthiaume. 2007. Effects of the
- method of conservation of timothy on nitrogen metabolism in lactating dairy cows. J. Dairy
- 565 Sci. 90:2870-2882.
- McGuire, M. A., J. M. Griinari, D. A. Dwyer, and D. E. Bauman. 1995. Role of insulin in the
- regulation of mammary synthesis of fat and protein. J. Dairy Sci. 78:816-824.
- Metcalf, J., L. Crompton, D. Wray-Cahen, M. Lomax, J. Sutton, D. Beever, J. MacRae, B.
- Bequette, F. Backwell, and G. Lobley. 1996. Responses in milk constituents to intravascular
- administration of two mixtures of amino acids to dairy cows. J. Dairy Sci. 79:1425-1429.
- Motulsky, H., and A. Christopoulos. 2004. Fitting models to biological data using linear and
- 572 nonlinear regression: A practical guide to curve fitting. Oxford University Press.
- NorFor The Nordic feed evaluation system. 2001. Ed. H. Volden, EAAP Publication No. 130,
- Wageningen Academic Publishers, The Netherlands, 180p.
- 576 NRC. 2001. Nutrient requirements of dairy cattle. 7th revised ed. National Academy Press,
- Washington, DC.

NRC. 2012. Nutrient requirements of swine. National Academy Press, Washington, DC.

- 579 O'Connor, J. D., C. J. Sniffen, D. G. Fox, and W. Chalupa. 1993. A net carbohydrate and
- protein system for evaluating cattle diets: IV. Predicting amino acid adequacy. J. Anim. Sci.
- 581 71:1298-1311.
- 582 Ørskov, E. R., N. A. MacLeod, and D. J. Kyle. 1986. Flow of nitrogen from the rumen and
- abomasum in cattle and sheep given protein-free nutrients by intragastric infusion. Br. J. Nutr.
- 584 56:241-248.
- Ouellet, D. R., M. Demers, G. Zuur, G. E. Lobley, J. R. Seoane, J. V. Nolan, and H. Lapierre.
- 586 2002. Effect of dietary fiber on endogenous nitrogen flows in lactating dairy cows. J. Dairy
- 587 Sci. 85:3013-3025.
- Ouellet, D. R., R. Berthiaume, G. Holtrop, G. E. Lobley, R. Martineau, and H. Lapierre. 2010a.
- 589 Effect of method of conservation of timothy on endogenous nitrogen flows in lactating dairy
- 590 cows. J. Dairy Sci. 93:4252-4261.
- Ouellet, D. R., D. Valkeners, and H. Lapierre. 2010b. Does endogenous nitrogen contribute to
- over-estimate bacterial duodenal flow in ruminant estimated by N dilution technique? Pages
- 593 125-126 in Proc. Energy and protein metabolism and nutrition. Wageningen Academic
- 594 Publishers, Parma, Italy.
- 595 Pisulewski, P. M., H. Rulquin, J. L. Peyraud, and R. Verite. 1996. Lactational and systemic
- responses of dairy cows to postruminal infusions of increasing amounts of methionine. J. Dairy
- 597 Sci. 79:1781-1791.

- Raggio, G., G. E. Lobley, S. Lemosquet, H. Rulquin, and H. Lapierre. 2006. Effect of casein
- and propionate supply on whole body protein metabolism in lactating dairy cows. Can. J. Anim.
- 600 Sci. 86:81-89.
- Relling, A. E., and C. K. Reynolds. 2008. Abomasal infusion of casein, starch and soybean oil
- differentially affect plasma concentrations of gut peptides and feed intake in lactating dairy
- 603 cows. Domest. Anim. Endocrinol. 35:35-45.
- Rius, A. G., J. A. D. R. N. Appuhamy, J. Cyriac, D. Kirovski, O. Becvar, J. Escobar, M. L.
- McGilliard, B. J. Bequette, R. M. Akers, and M. D. Hanigan. 2010. Regulation of protein
- synthesis in mammary glands of lactating dairy cows by starch and amino acids. J. Dairy Sci.
- 607 93:3114-3127.
- Robinson, P., W. Chalupa, C. Sniffen, W. Julien, H. Sato, T. Fujieda, T. Ueda, and H. Suzuki.
- 2000. Influence of abomasal infusion of high levels of lysine or methionine, or both, on ruminal
- 610 fermentation, eating behavior, and performance of lactating dairy cows. J. Anim. Sci. 78:1067-
- 611 1077.
- Rogers, J. A., J. H. Clark, T. R. Drendel, and G. C. Fahey, Jr. 1984. Milk production and
- 613 nitrogen utilization by dairy cows infused postruminally with sodium caseinate, soybean meal,
- or cottonseed meal. J. Dairy Sci. 67:1928-1935.
- Rulquin, H., P. Pisulewski, R. Vérité, and J. Guinard. 1993. Milk production and composition
- as a function of postruminal lysine and methionine supply: a nutrient-response approach.
- 617 Livest. Prod. Sci. 37:69-90.

- Russell, J., and C. Sniffen. 1984. Effect of carbon-4 and carbon-5 volatile fatty acids on growth
- of mixed rumen bacteria in vitro. J. Dairy Sci. 67:987-994.
- Russell, J. B., J. D. O'Connor, D. G. Fox, P. J. Van Soest, and C. J. Sniffen. 1992. A net
- 621 carbohydrate and protein system for evaluating cattle diets: I. Ruminal fermentation. J. Anim.
- 622 Sci. 70:3551-3561.
- 623 SAS. 2010. JMP. SAS Institute Inc., Cary, NC, USA.
- 624 Sauvant, D., G. Cantalapiedra-Hijar, L. Delaby, J. B. Daniel, P. Faverdin, and P. Nozière. 2015.
- Actualisation des besoins protéiques des ruminants et détermination des réponses des femelles
- laitières aux apports de protéines digestibles dans l'intestin. INRA Prod. Anim. 28:347-368.
- 627
- 628 Schwab, C. G., L. Satter, and A. Clay. 1976. Response of lactating dairy cows to abomasal
- infusion of amino acids. J. Dairy Sci. 59:1254-1270.
- 630 Seymour, W. M., C. E. Polan, and J. H. Herbein. 1990. Effects of dietary protein degradability
- and casein or amino acid infusions on production and plasma amino acids in dairy cows. J.
- 632 Dairy Sci. 73:735-748.
- 633 Schwab, C. G., C. K. Bozak, N. L. Whitehouse, and M. M. A. Mesbah. 1992a. Amino acid
- 634 limitation and flow to duodenum at four stages of lactation. 1. Sequence of lysine and
- methionine limitation. J. Dairy Sci. 75:3486-3502.

- 636 Schwab, C. G., C. K. Bozak, N. L. Whitehouse, and V. M. Olson. 1992b. Amino acid limitation
- and flow to the duodenum at four stages of lactation. 2. Extent of lysine limitation. J. Dairy
- 638 Sci. 75:3503-3518.
- 639 Schwab, C. G. 1996. Rumen-protected amino acids for dairy cattle: Progress towards
- determining lysine and methionine requirements. Anim. Feed Sci. Technol. 59:87-101.
- Swanson, E. W. 1977. Factors for computing requirements of protein for maintenance of cattle.
- 642 J. Dairy Sci. 60:1583-1593.
- Tamminga, S., H. Schulze, J. Vanbruchem, and J. Huisman. 1995. Nutritional significance of
- endogenous N-losses along the gastrointestinal-tract of farm-animals. Arch. Anim. Nutr. 48:9-
- 645 22.
- Van Amburgh, M. E., E. A. Collao-Saenz, R. J. Higgs, D. A. Ross, E. B. Recktenwald, E.
- Raffrenato, L. E. Chase, T. R. Overton, J. K. Mills, and A. Foskolos. 2015. The Cornell Net
- 648 Carbohydrate and Protein System: Updates to the model and evaluation of version 6.5. J. Dairy
- 649 Sci. 98:6361-6380.
- Van Duinkerken, G., M. C. Blok, A. Bannink, J. W. Cone, J. Dijkstra, A. M. Van Vuuren, and
- S. Tamminga. 2011. Update of the Dutch protein evaluation system for ruminants: The
- 652 DVE/OEB2010 system. J. Agric. Sci. 149:351-367.

- Van Straalen, W., C. Salaun, W. Veen, Y. Rijpkema, G. Hof, and T. Boxem. 1994. Validation
- of protein evaluation systems by means of milk production experiments with dairy cows.
- 655 Nether. J. Agri. Sci. 42:89-104.
- Vanhatalo, A., P. Huhtanen, V. Toivonen, and T. Varvikko. 1999. Response of dairy cows fed
- grass silage diets to abomasal infusions of histidine alone or in combinations with methionine
- 658 and lysine. J. Dairy Sci. 82:2674-2685.
- 659 Varvikko, T., A. Vanhatalo, T. Jalava, and P. Huhtanen. 1999. Lactation and metabolic
- responses to graded abomasal doses of methionine and lysine in cows fed grass silage diets. J.
- 661 Dairy Sci. 82:2659-2673.
- Vicini, J., J. Clark, W. Hurley, and J. Bahr. 1988. Effects of abomasal or intravenous
- administration of arginine on milk production, milk composition, and concentrations of
- somatotropin and insulin in plasma of dairy cows. J. Dairy Sci. 71:658-665.
- Weekes, T. L., P. H. Luimes, and J. P. Cant. 2006. Responses to amino acid imbalances and
- deficiencies in lactating dairy cows. J. Dairy Sci. 89:2177-2187.
- Yisehak, K., A. Becker, J. M. Rothman, E. S. Dierenfeld, B. Marescau, G. Bosch, W. Hendriks,
- and G. P. J. Janssens. 2012. Amino acid profile of salivary proteins and plasmatic trace mineral
- response to dietary condensed tannins in free-ranging zebu cattle (*Bos indicus*) as a marker of
- habitat degradation. Livest. Sci. 144:275-280.

673

TABLES AND FIGURES

Table 1. Endogenous contributions and digestion coefficients used to predict endogenous AA

flows in the models described in Figures 1 and 21

Endogenous component	Secretion (g N/kg DMI)	Kd (%/hr)3	SID (%)4
Saliva	0.9	150	5
Rumen sloughed cells	4.3	150	5
Omasum/abomasum sloughed cells	0.3	0.0	70
Omasum/abomasum secretions	0.2	0.0	70
Pancreatic secretions	0.4	0.0	70
Bile	0.1	0.0	70
Small intestine sloughed cells ₂	0.7	75	50
Small intestine secretions ₂	0.7	75	50
Large intestine sloughed cells	0.3	150	N/A

- 675 Back calculated to align predictions at various points in the gut to measured data (Ouellet et al.,
- 676 2002; Ouellet et al., 2010a).
- 677 2Includes secretions prior to the terminal ileum (Egan et al., 1984).
- 678 3Rate of microbial degradation in either the rumen or large intestine
- 679 4Small intestinal digestibility

681

Table 2. Profiles of EAA as a percent of EAA-N, total EAA-N as a percent of AA-N, and AA-N as a percent of total N for endogenous
 N components predicted by the model

Endogenous component	Met	Lys	Arg	Thr	Leu	Ile	Val	His	Phe	Trp	EAA-N ₇	AA-N
Salivaı	1.0	12.4	24.7	13.2	13.2	6.5	12.9	8.7	4.6	2.8	48.9	80.0
Rumen, omasum												
/abomasum, large	2.5	18.5	29.2	6.7	12.8	6.3	8.5	8.5	4.8	2.2	56.1	79.0
intestine sloughed cells2												
Omasum/abomasum	1.9	19.4	21.9	10.6	7.1	6.9	10.1	13.5	5.5	3.0	52.2	64.9
secretions3												
Pancreatic secretions4	2.0	16.2	18.0	10.6	13.0	7.7	12.4	12.6	5.0	2.6	51.3	94.3
Bile5	2.6	13.3	13.4	9.8	13.4	7.4	13.3	18.9	4.7	3.0	7.2	51.5
Small intestine sloughed cells and secretions ₆	1.9	14.3	23.7	13.2	9.8	7.5	11.8	9.5	5.4	3.0	39.2	72.9

685 Salivary protein (Yisehak et al., 2012)

686 2Rumen epithelia (Larsen et al., 2000)

687 3Abomasal isolates (Ørskov et al., 1986)

688 4Pancreatic juice from Hamza (1976) reported by Larsen et al. (2000)

689 5Cow bile (Larsen et al., 2000)

690 6Ileal endogenous AA in pigs (Jansman et al., 2002)

7The contribution of NEAA-N to endogenous secretions is represented by the proportion AA-N not accounted for by total EAA-N.

Table 3. Studies included in the dataset to estimate AA requirements by the logistic approach

Aldrich et al., 1993 Bruckental et al., 1991 Cant et al., 1991 Choung and Chamberlain, 1992a Choung and Chamberlain, 1992b Choung and Chamberlain, 1993 Choung and Chamberlain, 1993 Choung and Chamberlain, 1993 Choung and Chamberlain, 1995 Choung and Chamberlain, 1995a Choung and Chamberlain, 1995b Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Koning et al., 1984 Lapierre et al., 2009 Mackle et al., 1996 Mackle et al., 1996 Retal., 1996 Regis et al., 1996 Robinson et al., 2000 Rogers et al., 1984 Schwab et al., 1976 Schwab et al., 1992a Schwab et al., 1992b Guinard and Rulquin, 1995 Seymour et al., 1990 Varvikko et al., 1999 Vicini et al., 1999 Kim et al., 2000 Weekes et al., 2006		
Cant et al., 1991 Choung and Chamberlain, 1992a Choung and Chamberlain, 1992b Choung and Chamberlain, 1993 Choung and Chamberlain, 1993 Choung and Chamberlain, 1995 Choung and Chamberlain, 1995a Choung and Chamberlain, 1995b Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Weekes et al., 1988 Kim et al., 2000 Wackle et al., 1999 Mackle et al., 1996 Neughard et al., 1996 Mackle et al., 1996 Neughard et al., 1996 Mackle et al., 1996 Mackle et al., 1996 Neughard et al., 1996 Mackle et al., 1996 Neughard et al., 1996 Mackle et al., 1996 Neughard et al., 1996 N	Aldrich et al., 1993	Köning et al., 1984
Choung and Chamberlain, 1992a Choung and Chamberlain, 1992b Choung and Chamberlain, 1993 Choung and Chamberlain, 1993 Choung and Chamberlain, 1995 Choung and Chamberlain, 1995a Choung and Chamberlain, 1995b Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1994 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Mackle et al., 1996 Regulary Raggio et al., 1996 Relling and Reynolds, 2008 Relling and Reynolds, 2008 Robinson et al., 2000 Rogers et al., 1984 Schwab et al., 1992a Schwab et al., 1992b Schwab et al., 1992b Guinard et al., 1994 Vanhatalo et al., 1999 Vicini et al., 1999 Kim et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Bruckental et al., 1991	Lapierre et al., 2009
Choung and Chamberlain, 1992b Choung and Chamberlain, 1993 Choung and Chamberlain, 1995a Choung and Chamberlain, 1995a Choung and Chamberlain, 1995b Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1994 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 1999 Kim et al., 2000 Metcalf et al., 1996 Reguste al., 1996 Raggio et al., 1996 Relling and Reynolds, 2008	Cant et al., 1991	Lynch et al., 1991
Choung and Chamberlain, 1993 Choung and Chamberlain, 1995a Choung and Chamberlain, 1995b Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Choung and Lapierre, 2010 Cohick et al., 1986 Choung and Reynolds, 2008 Robinson et al., 2000 Rogers et al., 1984 Choung and Lapierre, 2011 Cohick et al., 1997 Cohick et al., 1997 Cohick et al., 1997 Cohick et al., 1994 County and Chamberlain, 1994 County and Chamberlain, 1994 County and Chamberlain, 1995 County and Chamberlain, 1995 County and Chamberlain, 1995 County and Chamberlain, 1996 County and Chamberlain, 1995 County and Chamberlain, 1996 Choung and Reynolds, 2008 Robinson et al., 1984 County and Reynolds, 2008 Clark et al., 1996 Choung and Chamberlain, 1995 Choung and Reynolds, 2008 Clark et al., 1986 Choung and Reynolds, 2008 Clark et al., 1996 Choung and Chamberlain, 1995 Choung and Chamberlain, 1995 Choung and Reynolds, 2008 Clark et al., 1996 Choung and Chamberlain, 1996 Choung and Chamberlain, 1995 Choung and Chamberlain, 1996 Choung and Reynolds, 2008 Clark et al., 2000 Choung and Reynolds, 2008 Clark et al., 2006 Choung and Reynolds, 2008 Clark et al., 2006 Choung and Reynolds, 2008 Clark et al., 2996 Choung and Reynolds, 2008 Clark et al., 2906 Choung and Reynolds, 2006 Choung and Reynolds, 2008 Clark et al., 2906 Choung and Reynolds, 2008 Choung and Reyno	Choung and Chamberlain, 1992a	Mackle et al., 1999a
Choung and Chamberlain, 1995a Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Pisulewski et al., 1996 Raggio et al., 1990 Relling and Reynolds, 2008 Relling and Reynolds, 2008 Robinson et al., 2000 Robinson et al., 1984 Schwab et al., 1984 Schwab et al., 1992a Schwab et al., 1992b Seymour et al., 1990 Vanhatalo et al., 1999 Vicini et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Choung and Chamberlain, 1992b	Mackle et al., 1999b
Choung and Chamberlain, 1995b Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Rogers et al., 1984 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Raggio et al., 2006 Relling and Reynolds, 2008 Relling and Reynolds, 2008 Relling and Reynolds, 2008 Relling and Reynolds, 2008 Roiling and Reynolds, 2008 Roiling and Reynolds, 2008 Roiling and Reynolds, 2008 Roiling and Reynolds, 2008 Relling and Reynolds, 2008	Choung and Chamberlain, 1993	Metcalf et al., 1996
Choung and Chamberlain, 1995c Clark et al., 1977 Cohick et al., 1986 Doepel and Lapierre, 2010 Rogers et al., 1984 Doepel and Lapierre, 2011 Griinari et al., 1997 Schwab et al., 1992a Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Seymour et al., 1990 Guinard et al., 1994 Huhtanen et al., 1997 Varvikko et al., 1999 Kim et al., 1999 Kim et al., 2000 Rogers et al., 2010 Rogers et al., 1984 Schwab et al., 1992a Schwab et al., 1992b Seymour et al., 1990 Varvikko et al., 1999 Vicini et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Choung and Chamberlain, 1995a	Pisulewski et al., 1996
Clark et al., 1977 Cohick et al., 1986 Robinson et al., 2000 Doepel and Lapierre, 2010 Rogers et al., 1984 Doepel and Lapierre, 2011 Schwab et al., 1976 Griinari et al., 1997 Schwab et al., 1992a Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Seymour et al., 1990 Guinard et al., 1994 Huhtanen et al., 1997 Varvikko et al., 1999 Kim et al., 1999 Kim et al., 2000 Weekes et al., 2006	Choung and Chamberlain, 1995b	Raggio et al., 2006
Cohick et al., 1986 Doepel and Lapierre, 2010 Rogers et al., 1984 Doepel and Lapierre, 2011 Schwab et al., 1976 Griinari et al., 1997 Schwab et al., 1992a Guinard and Rulquin, 1994 Guinard et al., 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Rogers et al., 1984 Schwab et al., 1992a Schwab et al., 1992b Seymour et al., 1990 Vanhatalo et al., 1999 Vicini et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Choung and Chamberlain, 1995c	Relling and Reynolds, 2008
Doepel and Lapierre, 2010 Rogers et al., 1984 Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Rogers et al., 1984 Schwab et al., 1992a Schwab et al., 1992b Seymour et al., 1990 Vanhatalo et al., 1999 Vicini et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Clark et al., 1977	Rius et al., 2010
Doepel and Lapierre, 2011 Griinari et al., 1997 Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Kim et al., 2000 Schwab et al., 1996 Schwab et al., 1992b Seymour et al., 1990 Vanhatalo et al., 1999 Vicini et al., 1999 Weekes et al., 2006	Cohick et al., 1986	Robinson et al., 2000
Griinari et al., 1997 Schwab et al., 1992a Guinard and Rulquin, 1994 Schwab et al., 1992b Guinard and Rulquin, 1995 Seymour et al., 1990 Guinard et al., 1994 Vanhatalo et al., 1999 Huhtanen et al., 1997 Varvikko et al., 1999 Kim et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Doepel and Lapierre, 2010	Rogers et al., 1984
Guinard and Rulquin, 1994 Guinard and Rulquin, 1995 Guinard et al., 1994 Seymour et al., 1990 Vanhatalo et al., 1999 Huhtanen et al., 1997 Vicini et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Doepel and Lapierre, 2011	Schwab et al., 1976
Guinard and Rulquin, 1995 Guinard et al., 1994 Huhtanen et al., 1997 Kim et al., 1999 Vicini et al., 1988 Kim et al., 2000 Vsekes et al., 2006	Griinari et al., 1997	Schwab et al., 1992a
Guinard et al., 1994 Vanhatalo et al., 1999 Huhtanen et al., 1997 Varvikko et al., 1999 Kim et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Guinard and Rulquin, 1994	Schwab et al., 1992b
Huhtanen et al., 1997 Varvikko et al., 1999 Kim et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Guinard and Rulquin, 1995	Seymour et al., 1990
Kim et al., 1999 Vicini et al., 1988 Kim et al., 2000 Weekes et al., 2006	Guinard et al., 1994	Vanhatalo et al., 1999
Kim et al., 2000 Weekes et al., 2006	Huhtanen et al., 1997	Varvikko et al., 1999
, , , , , , , , , , , , , , , , , , ,	Kim et al., 1999	Vicini et al., 1988
***	Kim et al., 2000	Weekes et al., 2006
King et al., 1991	King et al., 1991	

Table 4. Descriptive statistics of the dataset used to estimate optimum AA efficiency

	Mean	SD	Min	Max
Dry matter intake (kg/d)	18.0	3.1	11.0	27.6
Day in milk (d)	107	51	28	240
Body weight (kg)	551	55	487	733
Milk yield (kg/d)	26.3	5.85	10.7	40.0
Milk fat (%)	3.98	2.65	2.37	41.90
Milk true protein (%)	2.88	0.20	2.38	3.52
Fat yield (kg/d)	1.01	0.51	0.53	8.09
Milk true protein yield (kg/d)	0.76	0.16	0.32	1.11

Table 5. Measured and model predicted endogenous N flows along the gut (g EN/kg DMI)

	High f	orageı	Low	forage	Н	ay	For	mic	In	oc	Ave	rage
Endogenous flow	Study	Model	Study	Model	Study	Model	Study	Model	Study	Model	Study	Model
Total duodenum	3.4	3.8	3.7	3.6	4.9	4.8	4.3	4.1	4.7	4.1	4.2	4.1
Microbial	2.0	2.3	2.3	2.1	3.3	3.3	3.1	2.6	3.4	2.5	2.8	2.6
Free ₂	1.3	1.5	1.4	1.5	1.6	1.5	1.2	1.5	1.3	1.5	1.4	1.5
Total ileum3		2.0		2.0	2.1	2.3	2.4	2.2	2.9	2.1	2.5	2.2
Secreted in the forestomach ₃		1.3		1.3	1.3	1.6	1.8	1.5	1.8	1.5	1.6	1.5
Secreted in the small intestine		0.7		0.7	0.8	0.7	0.6	0.7	1.1	0.7	0.8	0.7
Fecal	1.8	2.0	2.0	1.9	2.4	2.3	2.1	2.1	2.5	2.1	2.1	2.1
Secreted in the forestomach	1.4	1.3	1.3	1.3	1.8	1.6	1.5	1.5	1.7	1.5	1.6	1.4
Secreted in the intestine4	0.4	0.7	0.6	0.6	0.6	0.7	0.6	0.7	0.8	0.7	0.6	0.7

700 High forage and Low forage are from Ouellet et al. (2002); Hay, Formic and Inoc are from Ouellet et al. (2010b)

²Estimated using the combined precursor pool. All other data represent the mucosa precursor pool

3Only 2 cows had ileal canula (this is why the contributions from the forestomach are different compared with values reported with

fecal EN)

699

701

702

703

704

705

706

4Includes contributions from the large intestine

Table 6. Model parameters, RMSE, R₂ and model outcomes for the log-logistic model fit between predicted AA requirement and supply

	Model	paramete	ers					
AA	θ_1	θ_2	θ_3	RMSE	R_2	AAnR:AAS1	g/d_2	% EAA
Arg	66.72	3.17	-0.03	3.31	0.79	0.55	96.4	10.2%
His	39.22	2.77	-0.05	2.47	0.76	0.70	43.9	4.5%
Ile	79.32	3.93	-0.03	4.85	0.74	0.61	102.7	10.8%
Leu	135.12	2.81	-0.01	8.52	0.72	0.67	158.3	17.1%
Lys	114.87	3.21	-0.02	7.33	0.72	0.62	145.1	15.1%
Met	39.23	2.49	-0.04	2.40	0.73	0.53	58.2	5.7%
Phe	69.30	3.52	-0.02	4.23	0.74	0.53	103.4	10.7%
Thr	69.54	3.50	-0.02	4.23	0.74	0.53	102.9	10.7%
Trp	20.74	4.42	-0.10	1.04	0.81	0.58	28.1	2.9%
Val	93.80	2.99	-0.02	6.10	0.68	0.62	118.8	12.4%
MP ₃	1625.35	3.67	-0.002	93.35	0.76	0.73	1751.8	N/A

709 Optimum ratio of predicted AA requirement (AAnR) and supply (AAS)

2 Optimum duodenal AA supply for the dataset used and presented in Table 4

3 MP = Metabolizable protein

Table 7. Model parameters and fit summary for the log-logistic relationship between AA
 requirement and supply as well as optimum supply of each EAA relative to ME and relative to
 Lys.

	Mode	el parame	eters					
AA	θ_1	θ_2	θ_3	R_2	RMSE	g AA/ Mcal ME	Lys:AA Dairy	Lys:AA Swine ₂
Arg	0.14	-0.88	0.47	0.80	0.05	2.04	1.49	1.85
His	0.19	-1.01	1.01	0.79	0.07	0.91	3.33	2.50
Ile	-0.53	-0.87	0.12	0.71	0.06	2.16	1.40	1.78
Leu	-0.27	-0.90	0.11	0.79	0.06	3.42	0.89	0.89
Lys	0.02	-0.89	0.23	0.73	0.06	3.03	1.00	1.00
Met	0.16	-0.97	1.01	0.75	0.06	1.14	2.66	3.71
Phe	0.09	-0.81	0.39	0.72	0.05	2.15	1.40	1.82
Thr	-0.53	-0.84	0.12	0.71	0.05	2.14	1.41	1.49
Trp	-0.21	-0.81	0.67	0.68	0.05	0.59	5.16	5.33
Val	-0.09	-0.88	0.22	0.75	0.06	2.48	1.22	1.15

⁷¹⁸ Optimum Lys:EAA ratio for the data set used and presented in Table 4

2 Optimum Lys:EAA ratio for a lactating sow (NRC, 2012)

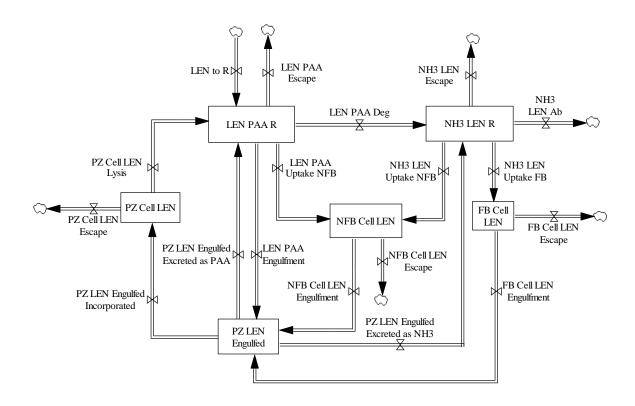


Figure 1. Schematic representation of the model used to predict the incorporation of labelled endogenous N (LEN) into rumen microorganisms (see text for abbreviations)

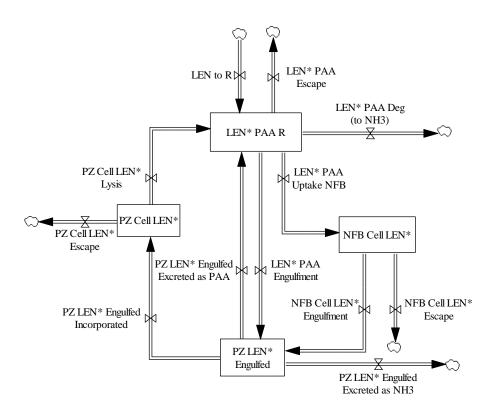


Figure 2. Schematic representation of the model used to predict the incorporation of labelled endogenous N, excluding transaction through the NH₃ pool (LEN*), into rumen microorganisms (see text for abbreviations)

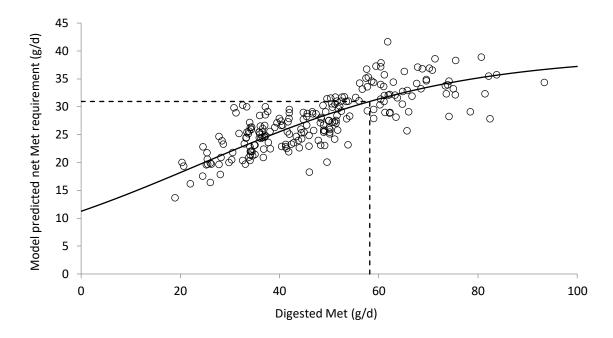


Figure 3. Logistic fit of model predicted net Met requirement and Met supply. The dashed line represents the optimum ratio of Met requirement and Met supply and is calculated by setting the third derivative of the logistic equation to 0. For this dataset, the optimum level of digested Met is 58.2 g/d (X axis) which corresponds to a model predicted requirement of 30.8 g/d (Y axis)

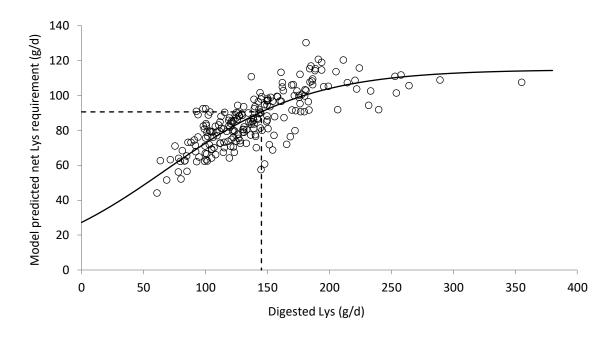


Figure 4. Logistic fit of model predicted net Lys requirement and Lys supply. The dashed line represents the optimum ratio of Lys requirement and Lys supply and is calculated by setting the third derivative of the logistic equation to 0. For this dataset, the optimum level of digested Lys is 145.1 g/d (X axis) which corresponds to a model predicted requirement of 90.0 g/d (Y axis)

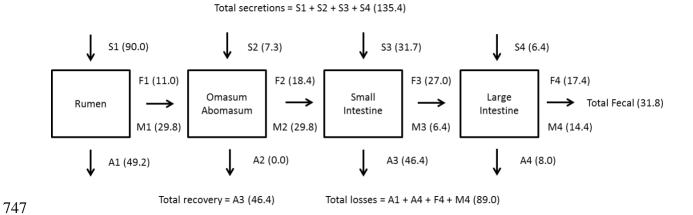


Figure 5. Model predicted endogenous transactions (g endogenous N/d) by compartment for the hay treatment presented in Ouellet et al. (2010a). S1-S4 are the endogenous secretions into the gut; F1-F4 are the flows of free endogenous N; M1-M4 are the flow of endogenous N in bacteria; A1-A4 is the endogenous N absorption at different sites. Recovery is only possible in the small intestine (A3) where the N can be absorbed as AA

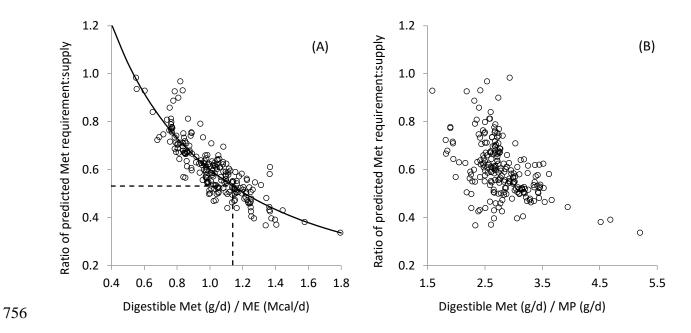


Figure 3. Relationship between model predicted Met net requirement:supply and Met supply relative to ME (A) or MP (B). The dashed line in (A) represents the Met supply at the optimum ratio of model predicted Met requirement and supply and is calculated by rearranging the log-logistic equation and solving for X. The optimum ratio of Met requirement:supply is 0.53 (Y axis) and the optimum supply is 1.14 g/Mcal ME (X axis). No significant relationship was obtained in (B)

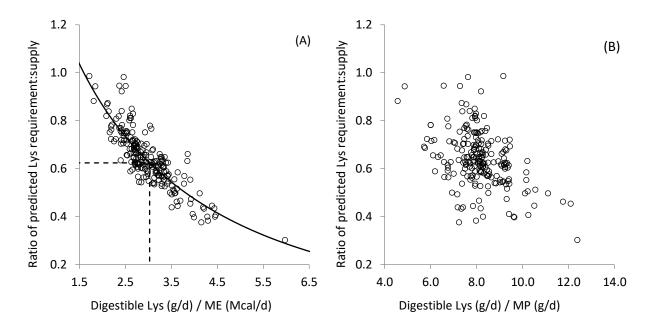


Figure 4. Relationship between model predicted Lys net requirement:supply and Lys supply relative to ME (A) or MP (B). The dashed line in (A) represents the Lys supply at the optimum ratio of model predicted Lys requirement and supply and is calculated by rearranging the log-logistic equation and solving for X. The optimum ratio of Met requirement:supply is 0.62 (Y axis) and the optimum supply is 3.03 g/Mcal ME (X axis). No significant relationship was obtained in (B)