



```
dds <- DESeqDataSetFromMatrix(countData = cts,
                                colData = coldata,
                                design= ~ batch + condition)

dds <- DESeq(dds)
resultsNames(dds) # lists the coefficients
res <- results(dds, name="condition_trt_vs_untrt")
# or to shrink log fold changes association with condition:
res <- lfcShrink(dds, coef="condition_trt_vs_untrt", type="apeglm")
```

Bulk RNA-seq Analysis Part II

Invitation to chat:

What questions are you interested in answering using DGE?

Differential Gene Expression

Harvard Chan Bioinformatics Core

<https://tinyurl.com/hbc-dge-online>

Consulting

- Transcriptomics: bulk, single cell, small RNA
- Epigenomics: ChIP-seq, CUT&RUN, ATAC-seq, DNA methylation
- Variant discovery: WGS, resequencing, exome-seq and CNV
- Multiomics integration
- Spatial biology
- Experimental design and grant support

<http://bioinformatics.sph.harvard.edu/>

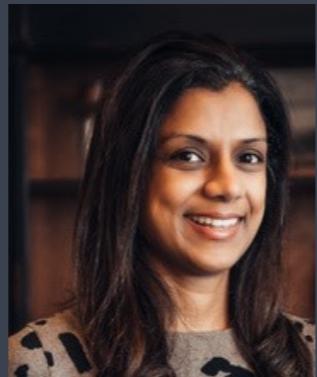


NIEHS

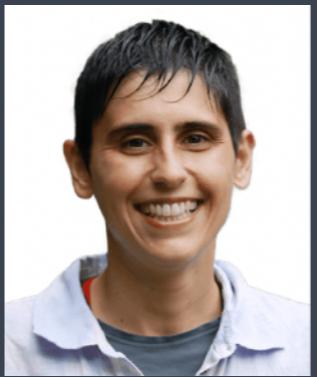




Shannan Ho Sui
Director



Meeta Mistry
Associate Director



Lorena Pantano
Director of Bioinformatics Platform



John Quackenbush
Faculty Advisor



Emma Berdan



Heather Wick



Will Gammerdinger



Noor Sohail



Alex Bartlett



Upen Bhattacharai



Elizabeth Partan



James Billingsley



Zhu Zhuo



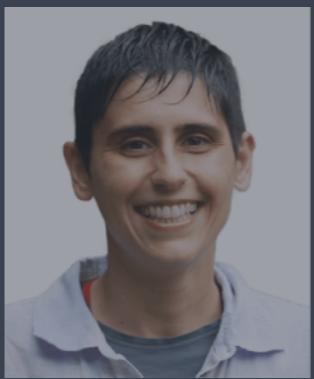
Maria Simoneau



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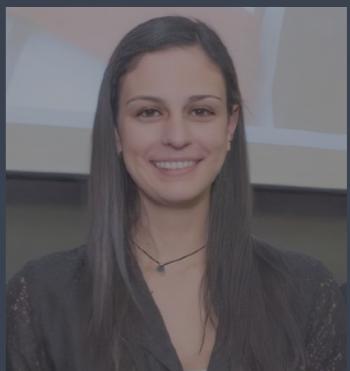
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James Billingsley



Zhu Zhuo



Maria Simoneau

Training

A key component of the HBC's mission is its training initiative. Our dedicated training team holds workshop to help researchers at Harvard better understand analytical methods for NGS data.

HBC's training team is made up of four PhD-level scientists who devote substantial time to material development, training and community building/outreach. All members of the training team also participate in consultations on research projects to ensure they remain up-to-date on current best practices in NGS analysis.

Our hands-on workshops focus on **basic data skills** and **analysis of high-throughput sequencing data**, with an emphasis on **experimental design**, current **best practices** and **reproducibility**. Our workshops are designed for **wet-lab biologists** aiming to independently design sequencing-based experiments and analysing the resulting data.

We offer three types of workshops:

1. Short, 3-hour monthly workshops (*Current topics in bioinformatics*)
2. Basic Data Skills**
3. Advanced Topics: Analysis of high-throughput sequencing (NGS) data**

***The basic data skills workshops serve as the foundation for the advanced workshops.*

<http://bioinformatics.sph.harvard.edu/training/>

<https://hbctraining.github.io/main/>

Training

A key component of the HBC's mission is to train researchers at Harvard and beyond.

HBC's training team is made up of scientists, educators, and community based organizations who work on research projects to ensure the best training for our students.

Our hands-on workshops are designed to provide an emphasis on **experimental design** and **informatics** for **wet-lab biologists** and **bioinformaticians** alike.

We offer three types of workshops:

1. Short, 3-hour monthly workshops
2. Basic Data Skills**
3. Advanced Topics: Analysis of high-throughput sequencing data

**The basic data skills workshop is designed for those new to bioinformatics.



**HARVARD
T.H. CHAN
SCHOOL OF PUBLIC HEALTH**

DF/HCC
DANA-FARBER / HARVARD CANCER CENTER



THE HARVARD CLINICAL
AND TRANSLATIONAL
SCIENCE CENTER



Our dedicated training team holds workshops to help researchers learn how to analyze and interpret NGS data.

In addition to devote substantial time to material development, the training team also participate in consultations on best practices in NGS analysis.

The workshops focus on the analysis of high-throughput sequencing data, with an emphasis on **experimental design**, **informatics**, and **reproducibility**. Our workshops are designed to provide an emphasis on performing wet-lab experiments and analysing the resulting sequencing data.

bioinformatics)

NGS) data**

and **bioinformatics** for the advanced workshops.

<http://bioinformatics.sph.harvard.edu/training/>

<https://hbctraining.github.io/main/>

Training

A key component of the HBC's mission is to provide training for researchers at Harvard and beyond.

HBC's training team is made up of experts in training and community building who work on research projects to ensure the quality of our training.

Our hands-on workshops focus on **bioinformatics**, with an emphasis on **experimental design** and **data analysis**. We also offer **wet-lab biologists** and **clinicians** training in **bioinformatics** and **data analysis**.

We offer three types of workshops:

1. Short, 3-hour monthly workshops
2. Basic Data Skills**
3. Advanced Topics: Analysis of high-throughput sequencing data

**The basic data skills workshop is designed for researchers with no prior experience in bioinformatics.



**HARVARD
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THE HARVARD CLINICAL
AND TRANSLATIONAL
SCIENCE CENTER

**HARVARD
MEDICAL SCHOOL**

Our dedicated training team holds workshops to help researchers learn how to analyze **bioinformatics** or **NGS** data.

The training team also devote substantial time to material development, and the training team also participate in consultations on best practices in NGS analysis.

Analysis of high-throughput sequencing data, with an emphasis on **experimental design**, **data analysis**, and **reproducibility**. Our workshops are designed for **wet-lab biologists** and **clinicians** performing **bioinformatics** experiments and analysing the resulting **bioinformatics** data.

bioinformatics)

bioinformatics (**bio**informatics) **data****

or the advanced workshops.

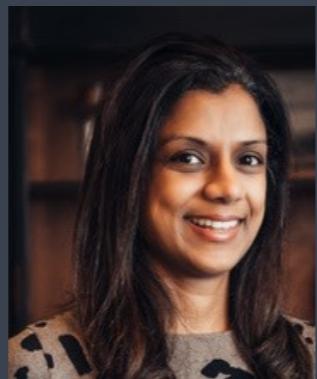
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<https://hbctraining.github.io/main/>

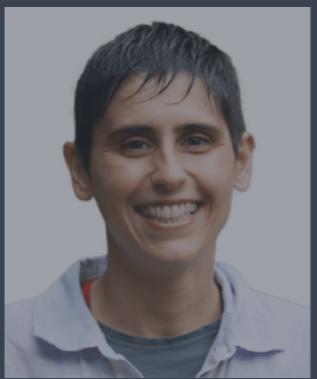
Introductions!



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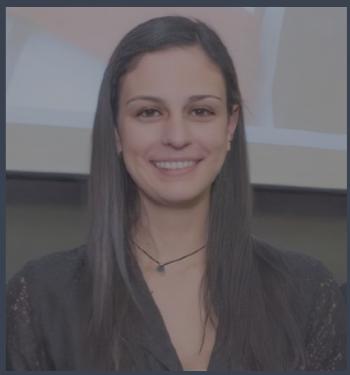
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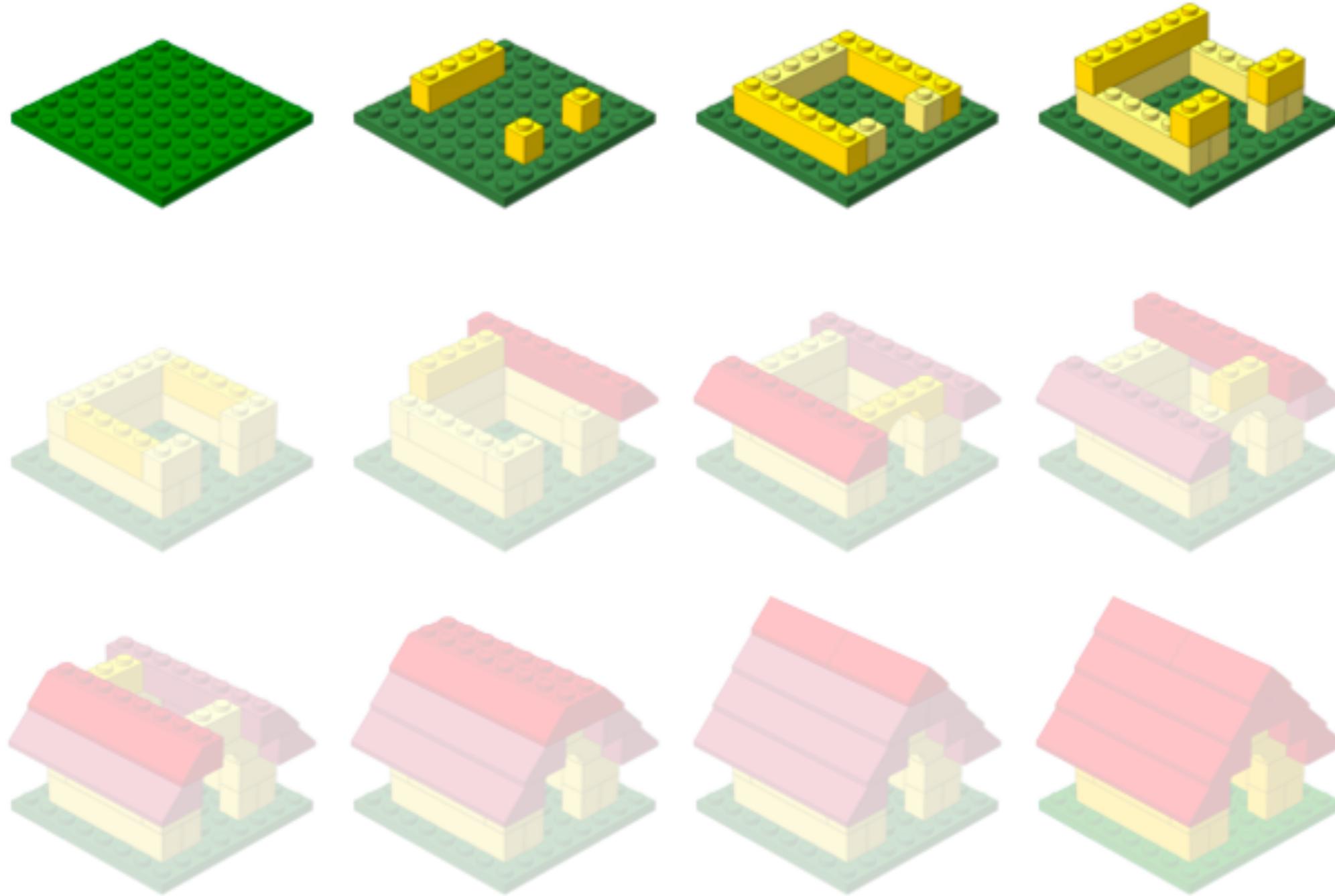


Zhu Zhuo



Maria Simoneau

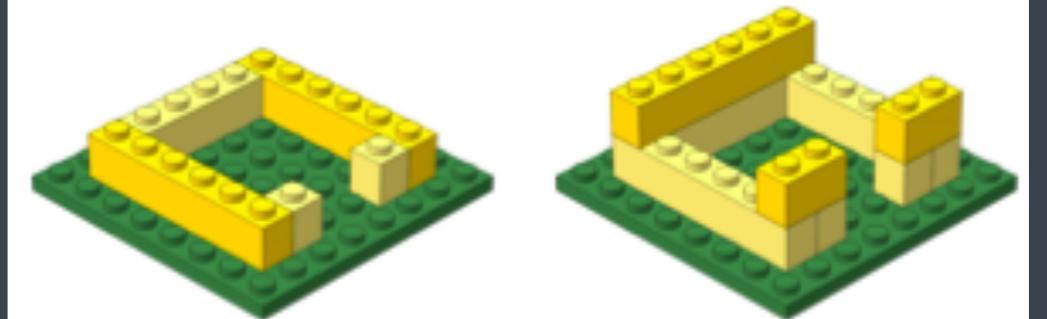
Workshop Scope...



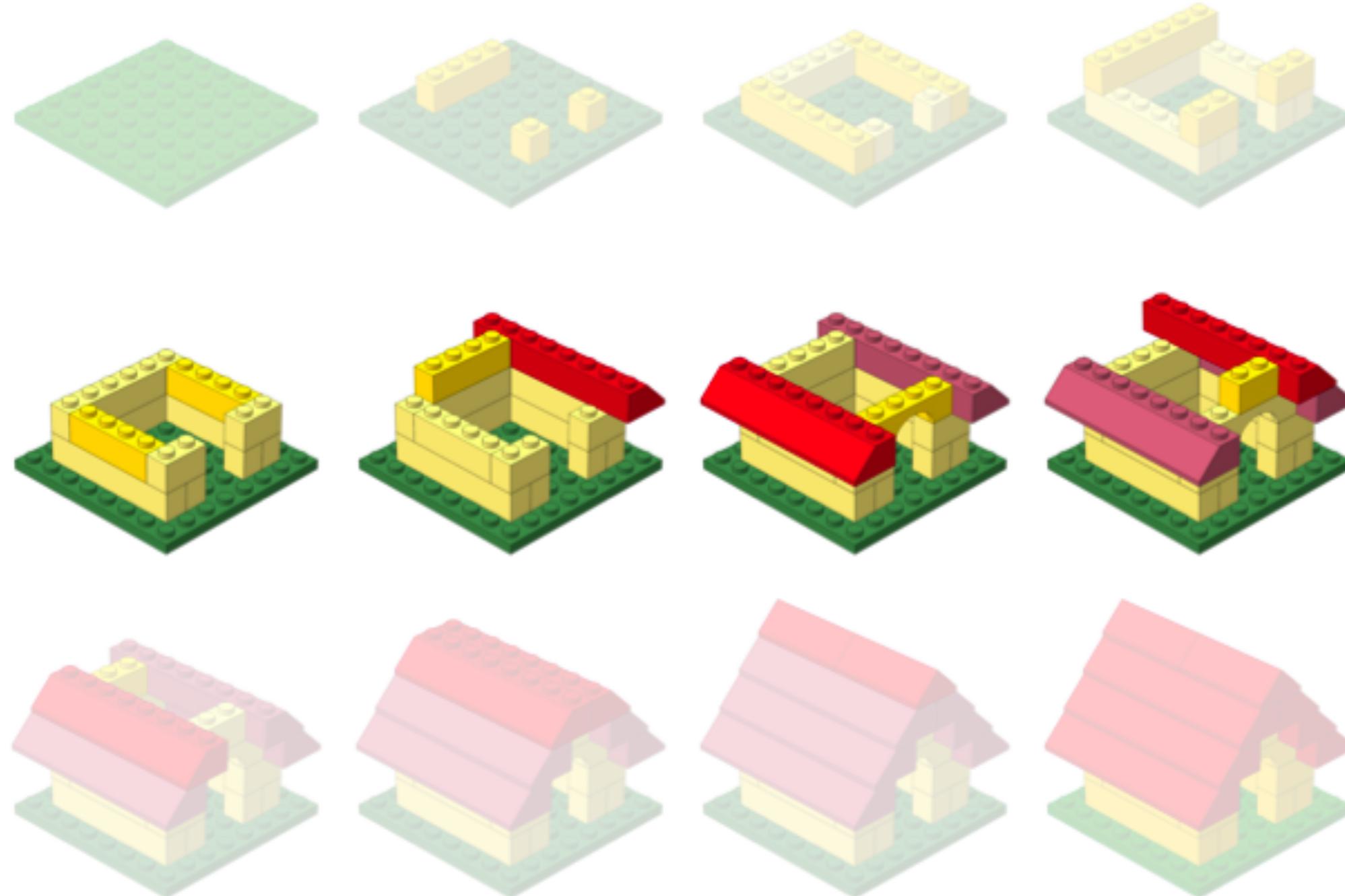
<http://anoved.net/tag/lego/page/3/>

Setting up to perform Bioinformatics analysis

Setting up...



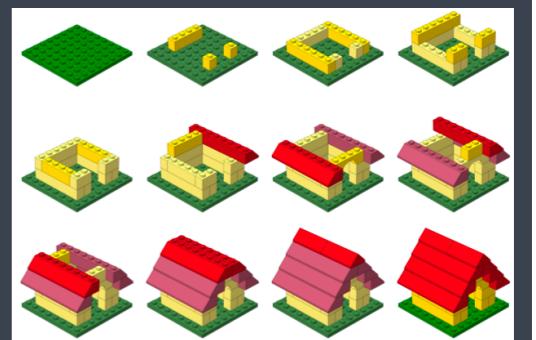
- ✓ Introduction to the command-line interface (shell, Unix, Linux)
 - Dealing with large data files
 - Performing bioinformatics analysis
 - Using tools
 - Accessing and using compute clusters
- ✓ R
 - Parsing and working with smaller results text files
 - Statistical analysis, e.g. differential expression analysis
 - Generating figures from complex data



<http://anoved.net/tag/lego/page/3/>

Bioinformatics data analysis

Workshop Scope



Differential Gene Expression analysis

- ✓ Understand the considerations for performing statistical analysis on RNA-seq data
- ✓ Start with gene counts (after alignment and counting)
- ✓ Perform QC on count data
- ✓ Use DESeq2 to perform differential expression analysis on the count data and obtain a list of significantly different genes
- ✓ Visualize results of the analysis
- ✓ Perform functional analysis on the lists of differentially expressed genes

Logistics

Course webpage

<https://tinyurl.com/hbc-dge-online>

Course schedule online

Workshop Schedule

Pre-reading

1. [Workflow \(raw data to counts\)](#)
2. [Experimental design considerations](#)

Day 1

Time	Topic	Instructor
10:00 - 10:30	Workshop Introduction	Jihe
10:30 - 10:45	R refresher Q & A	Radhika
10:45 - 11:15	RNA-seq pre-reading discussion	Radhika
11:15 - 12:00	Intro to DGE / setting up DGE analysis	Meeta

Before the next class:

1. Please **study the contents** and **work through all the code** within the following lessons:
 - [RNA-seq counts distribution](#)
 - [Count normalization](#)
 - [Sample-level QC \(PCA and hierarchical clustering\)](#)
2. **Complete the exercises:**
 - Each lesson above contain exercises; please go through each of them.
 - **Copy over** your code from the exercises into a text file.
 - **Upload the saved text file to Dropbox** the **day before the next class**.

Questions?

- **If you get stuck due to an error** while running code in the lesson, [email us](#)
- Post any **conceptual questions** that you would like to have **reviewed in class here**.

Course webpage

Introduction to DGE

[View on GitHub](#)

Approximate time: 60 minutes

Learning Objectives

- Explore different types of normalization methods
- Become familiar with the `DESeqDataSet` object
- Understand how to normalize counts using DESeq2

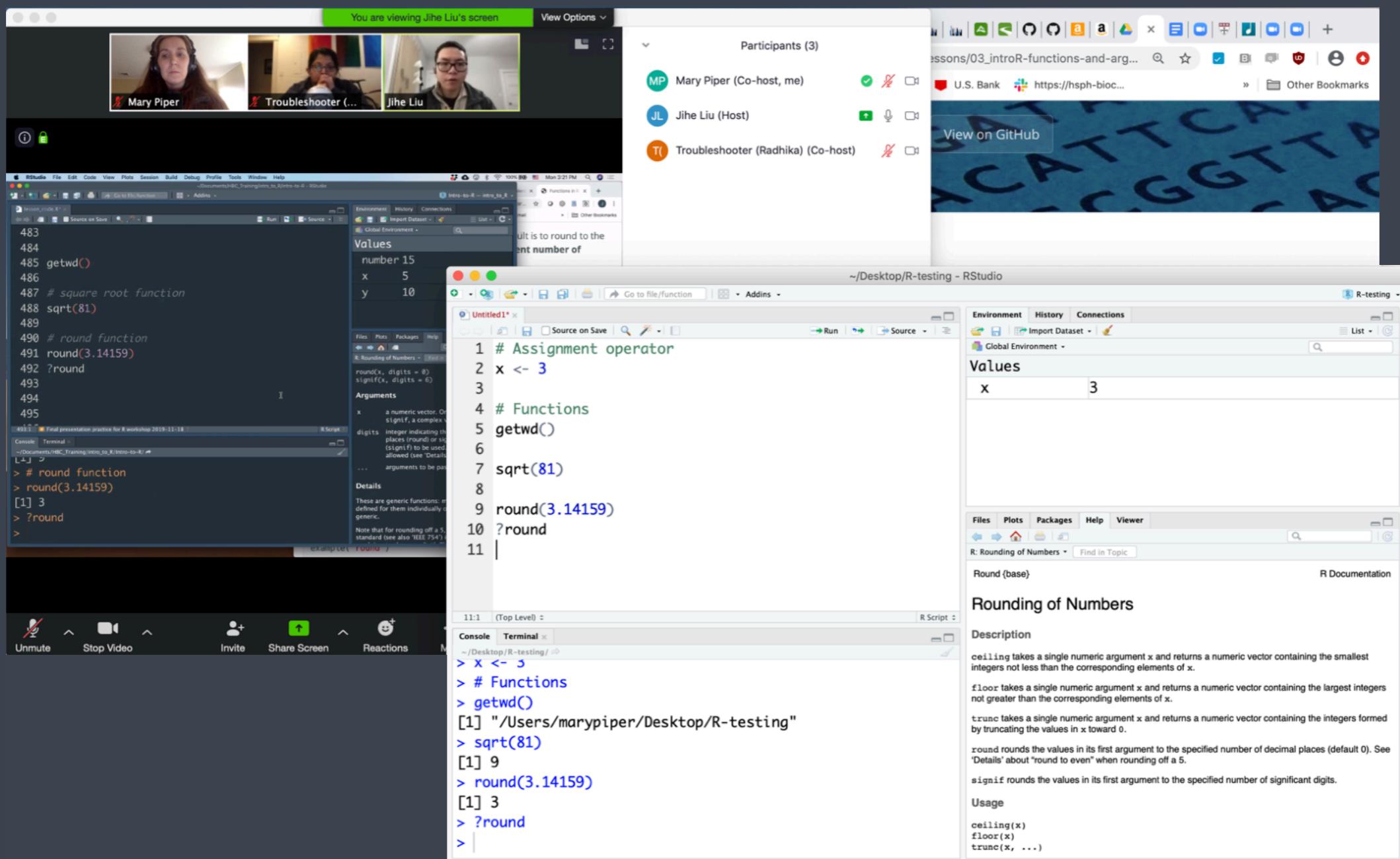
Normalization

The first step in the DE analysis workflow is count normalization, which is necessary to make accurate comparisons of gene expression between samples.

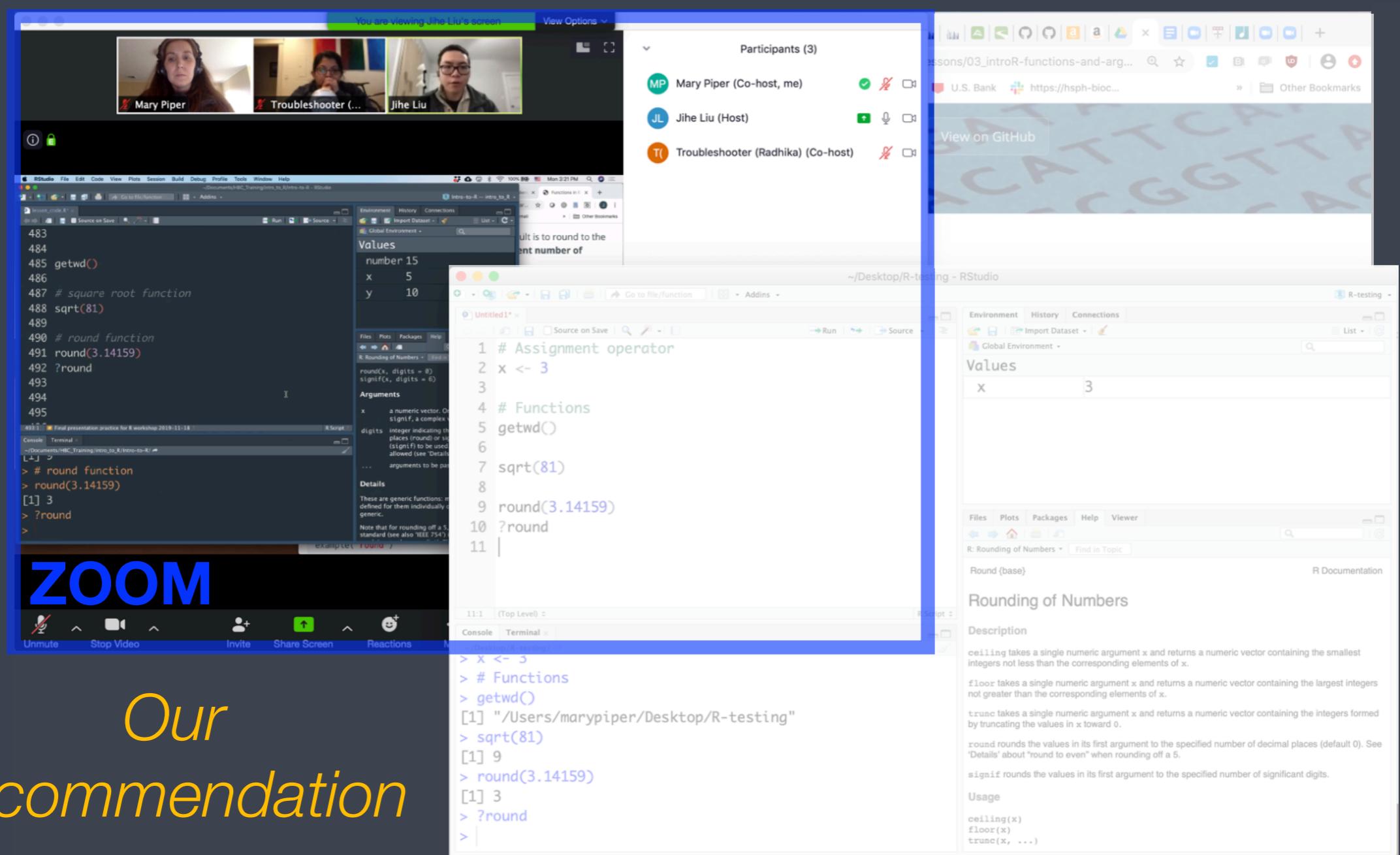
```
graph TD; A["Pseudocounts with  
Kallisto, Sailfish, Salmon"] --> B["Read counts  
associated with genes"]; B --> C["Normalization"]; C --> D["Unsupervised clustering analyses"]; C -.-> E["Quality control"]
```

The flowchart illustrates the DE analysis workflow. It begins with 'Pseudocounts with Kallisto, Sailfish, Salmon', followed by 'Read counts associated with genes'. This leads to 'Normalization', which then leads to 'Unsupervised clustering analyses'. A bracket on the right side groups 'Normalization' and 'Unsupervised clustering analyses' under the heading 'Quality control'.

Single screen & 3 windows?

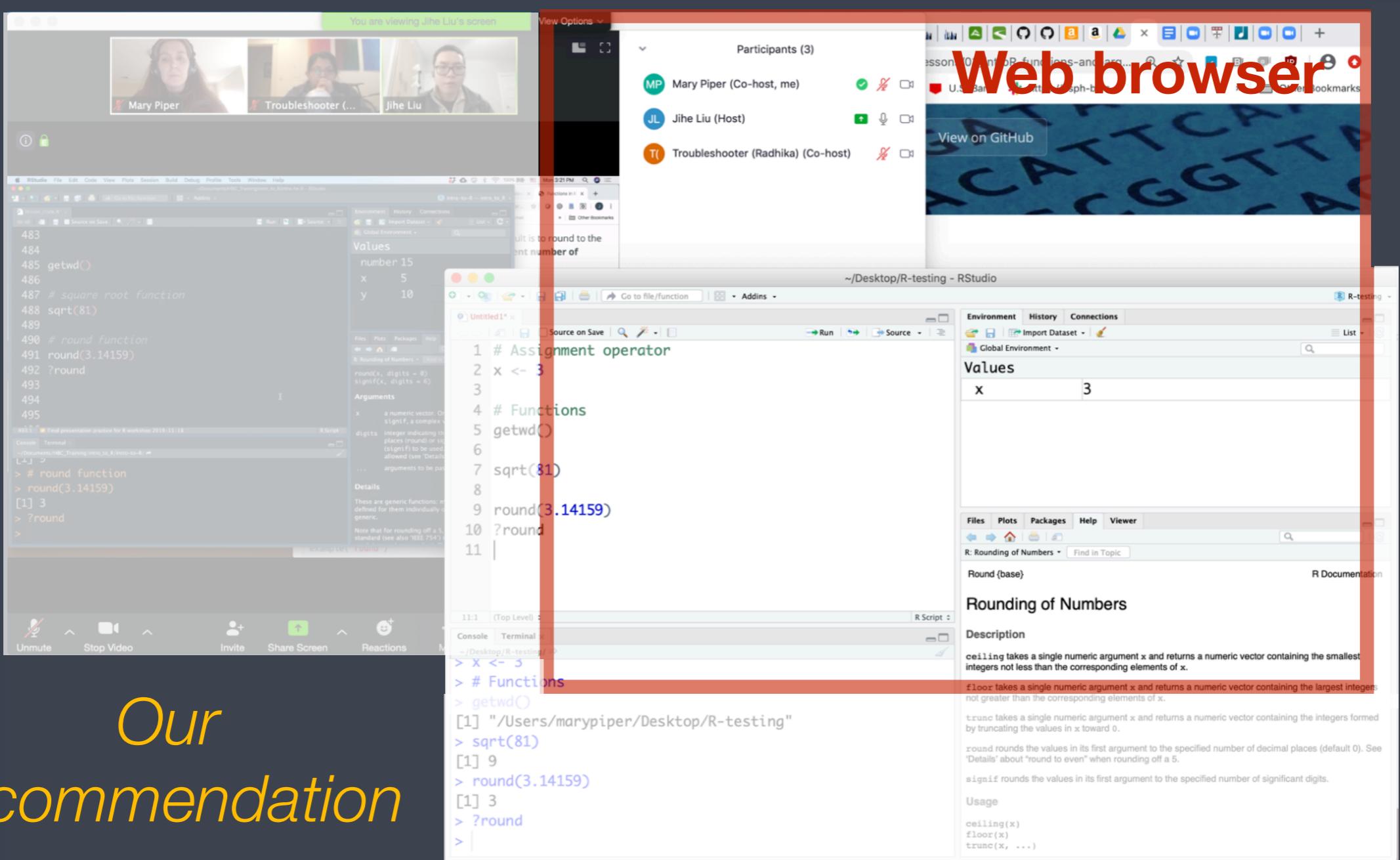


Single screen & 3 windows?



*Our
recommendation*

Single screen & 3 windows?



Single screen & 3 windows?

The image shows a video conference interface with three windows:

- Top Left Window:** A video feed showing three participants: Mary Piper, Troubleshooter (Radhika), and Jihe Liu.
- Middle Left Window:** An RStudio session titled "Untitled1". It contains the following R code:

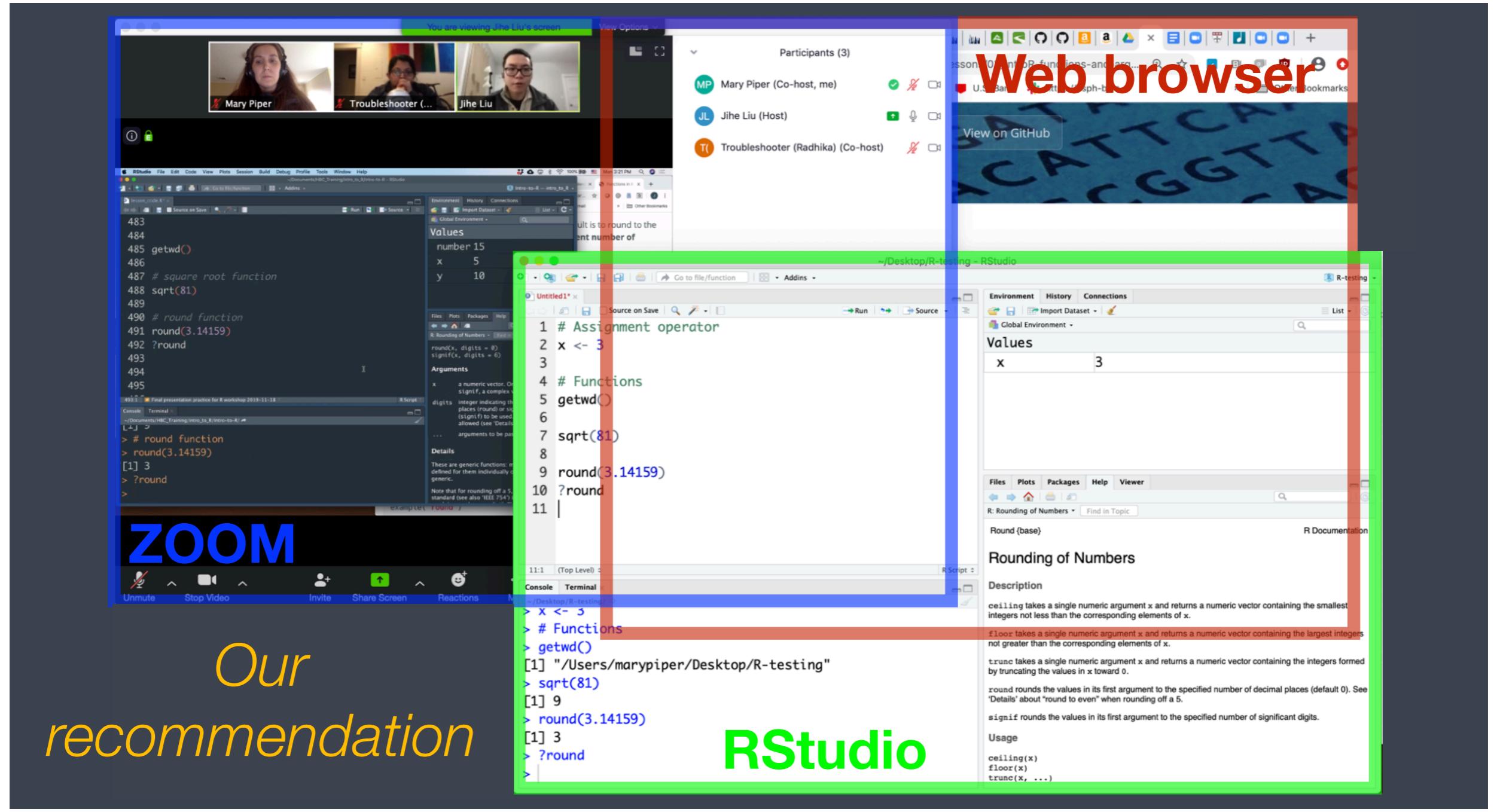
```
1 # Assignment operator
2 x <- 3
3
4 # Functions
5 getwd()
6
7 sqrt(81)
8
9 round(3.14159)
10 ?round
11
```
- Bottom Left Window:** A terminal window showing the same R code being run.

```
> x <- 3
> # Functions
> getwd()
[1] "/Users/marypiper/Desktop/R-testing"
> sqrt(81)
[1] 9
> round(3.14159)
[1] 3
> ?round
>
```
- Top Right Window:** A web browser window showing a GitHub page for a presentation script titled "03_introR-functions-and-args.Rmd".
- Middle Right Window:** Another RStudio session titled "R-testing". It shows the variable `x` assigned the value 3. Below it is a "Rounding of Numbers" documentation page from R Documentation.

Bottom Left Text: Our recommendation

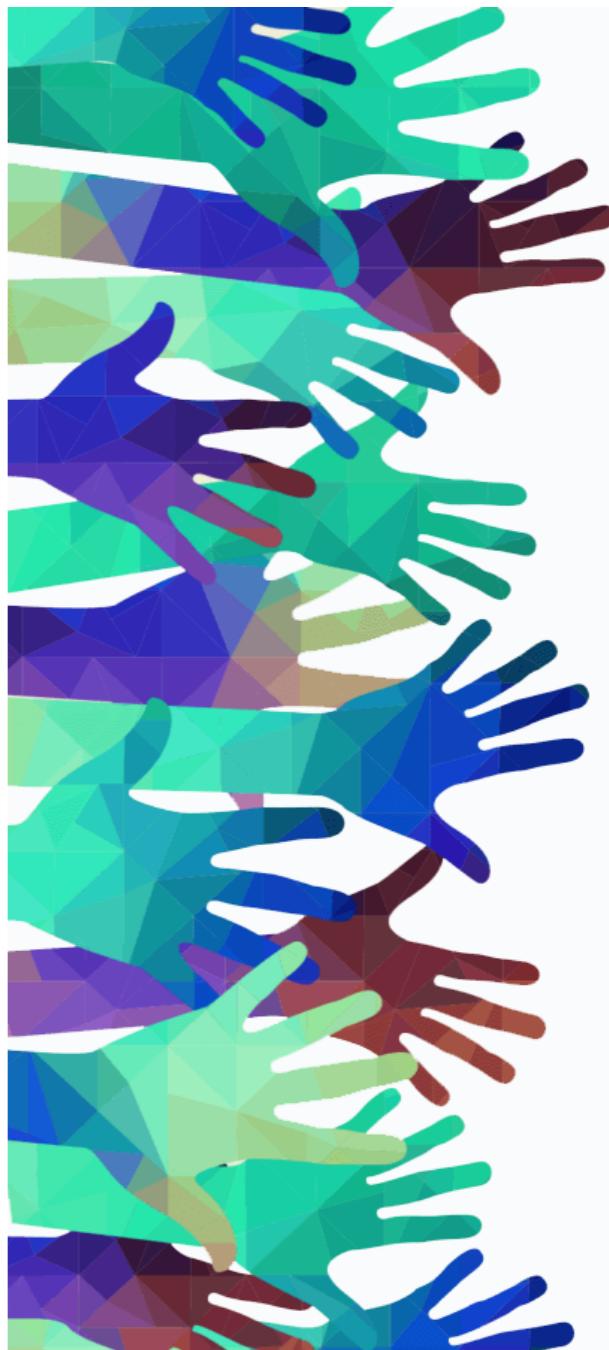
Bottom Right Text: RStudio

Single screen & 3 windows?



Course participation

- ▶ Mandatory review of self-learning lessons and assignments
- ▶ Attendance required for all classes
- ▶ Your questions and active participation drive learning
- ▶ We look forward to all of your questions!



Homework and Expectations

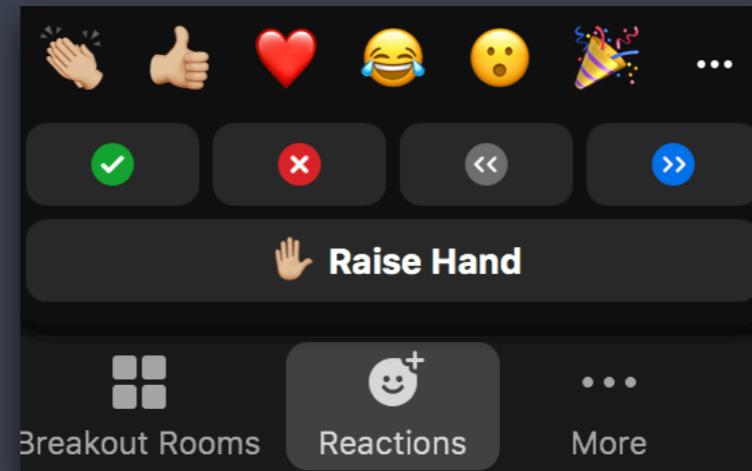
- ❖ At-home lessons and exercises after each session
- ❖ Cover material not previously discussed
- ❖ Provides us feedback to help pace the course appropriately
- ❖ 3-5 hours to complete
- ❖ Homework load can be heavier in the beginning of this workshop series, but it tapers off

Odds and Ends

- ❖ Quit/minimize all applications that are not required for class
- ❖ Captioning is available upon request

Odds and Ends (1/2)

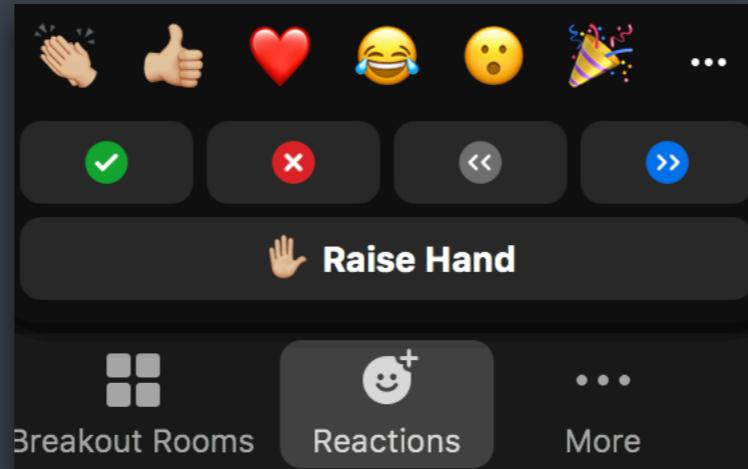
- ❖ Quit/minimize all applications that are not required for class
- ❖ Captioning is available upon request
- ❖ Are you all set?
 - ▶  = "agree", "I'm all set" (equivalent to a **green post-it**)
 - ▶  = "disagree", "I need help" (equivalent to a **red post-it**)



Odds and Ends (2/2)

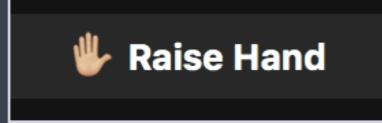
- ❖ Questions for the presenter?

- Post the question in the Chat window OR
-  when the presenter asks for questions
- Let the Moderator know



Odds and Ends (2/2)

- ❖ Questions for the presenter?

- Post the question in the Chat window OR
-  when the presenter asks for questions
- Let the Moderator know

- ❖ Technical difficulties with RStudio or R?

- Start a private chat with the Troubleshooter with a description of the problem.

Contact us!

HBC training team: hbctraining@hsph.harvard.edu

HBC consulting: bioinformatics@hsph.harvard.edu