STAT 640: Homework 11

Due Wednesday, April 27, 11:59pm MT on the course Canvas webpage. Please follow the homework guidelines on the syllabus.

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Problem 1

(This is carry-over from Homework 10)

Consider the data in the file coagulation.csv on Canvas, which comes from an experiment of diet on blood coagulation time. Animals were independently randomized to four diets and the time for blood coagulation was measured.

e. Find the contrast $\mathbf{g}^{\mathsf{T}}\boldsymbol{\beta}$ that results in the largest possible F statistic for $H_0: \mathbf{g}^{\mathsf{T}}\boldsymbol{\beta} = 0$. Report the contrast $(\mathbf{g}^{\mathsf{T}}\boldsymbol{\beta})$, a corresponding \mathbf{d} , and the value of its F statistic.

Answer: The contrast that results in the largest F-test statistic is $g^T \beta = (X^T (P_X - P_1)Y)^T \beta$.

0 -12 12 24 -24

The contrast is

$$\mathbf{g}^{\mathsf{T}}\boldsymbol{\beta} = \begin{bmatrix} 0 \\ -12 \\ 12 \\ 24 \\ -24 \end{bmatrix} \boldsymbol{\beta} = 12(\alpha_2 - \alpha_1) + 24(\alpha_3 - \alpha_4)$$

and d is not unique, so one possibility could be

$$\boldsymbol{d} = \begin{bmatrix} -3 \cdot \mathbf{1}_4 \\ 2 \cdot \mathbf{1}_6 \\ 4 \cdot \mathbf{1}_6 \\ -3 \cdot \mathbf{1}_8 \end{bmatrix}$$

```
d \leftarrow c(rep(c(-3, 2, 4, -3), times = c(4, 6, 6, 8)))
sum(d)
## [1] 0
bbh <- c(0, tapply(coag$time, coag$diet, mean))</pre>
Ginv \leftarrow diag(c(0, 1/diag(t(X2)%*%X2)))
MSE <- ( t(Y) \%*\% (diag(1, 24) - Px) \%*\% Y )/20
F_{\text{stat}} \leftarrow (((t(g) \%*\% bbh)^2 * (t(g) \%*\% Ginv \%*\% g)^(-1))/3)/MSE
cat(F_stat)
## 13.57143
# check
mod <- lm(time ~ diet, coag)</pre>
anova(mod)
## Analysis of Variance Table
##
## Response: time
##
              Df Sum Sq Mean Sq F value
                                              Pr(>F)
## diet
                     228
                            76.0 13.571 4.658e-05 ***
                             5.6
## Residuals 20
                     112
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
```

Problem 2

Summarize the different error rates that can be controlled in multiple comparisons by completing the following table. Where possible use ND/FD/FN/FT/N/D in the formulas, and keep your description to a phrase or two.

Answer:

Error Rate/Quantity	Formula	Description
Simultaneous CI's		
sFWER	$P(FD > 0 \mid \text{ at least } 1 H_0 \text{ true})$	Probability of at least 1 FD.
FDR	$\frac{FD}{D}P(D>0)$	Expected proportion of all D that are FD.
FWER	$P(D > 0 \mid H_0 \text{ all true})$	Probability of at least 1 FD when all H_0 are true.
PCER	$P(FD_i = 1 \mid H_{0i} \text{ true})$	Probability of a FD when H_0 is true.

Problem 3

Suppose we have an ANOVA model with four treatments. Researchers wish to conduct all pairwise comparisons AND test two additional (pre-specified) contrasts. They propose to use Tukey's method for pairwise comparisons at level $\alpha/2$ and then test each of the other two contrasts at level $\alpha/4$. Prove that this approach will control sFWER at level α .

Answer: I'm not sure I understand this problem. When using Tukey's HSD for the pairwise comparisons, they control sFEWER at $P(FD>0) \leq \alpha/2$ for all comparisons. (Notes specify that Tukey Method controls sFEWER). Controlling each of the other contrasts at level $\alpha/4$ means that $P(FD>0) \leq \alpha/4$ for each contrast, which is the same as controlling sFEWER at $\alpha/4$ for each contrast. This means, I think that for all of the contrasts

$$P(FD > 0) = P_{\text{(pairwise)}}(FD > 0) + P_{\text{(contrast 1)}}(FD > 0) + P_{\text{(contrast 2)}}(FD > 0)$$

$$\leq \frac{\alpha}{2} + \frac{\alpha}{4} + \frac{\alpha}{4} = \alpha$$

Problem 4

This continues Problem 2 from Homework 10. The data in ivd.csv on Canvas contain measurements of the in vitro digestibility (IVD) of alfalfa grown at different temperatures. The variable temp has four levels: 17, 22, 27, and 32 degrees Celsius. Each level has four randomly-assigned replicates.

a. Test all pairwise comparisons of mean IVD by treatment group using the Tukey HSD test. For each comparison, provide the estimated difference, a confidence interval, and a *p*-value. Compute the values using your own code (but you can check yourself against the results from TukeyHSD()).

Answer:

```
ivd <- read csv("ivd.csv")</pre>
X \leftarrow cbind(1
                                                                # Design Matrix
            , T17 = as.numeric(ivd$temp == 17)
            , T22 = as.numeric(ivd$temp == 22)
            , T27 = as.numeric(ivd$temp == 27)
             T32 = as.numeric(ivd$temp == 32)
bbh <- c(0, tapply(ivd$ivd, ivd$temp, mean))
                                                                # beta hat
Px \leftarrow X[,-1] \% \%  solve(t(X[,-1])\%*\%X[,-1]) \%*\% t(X[,-1])
                                                               # Projection Matrix
MSE <- (t(ivd$ivd) %*% (diag(1, nrow(X)) - Px) %*% ivd$ivd)/12
# Manual Tukey HSD
diffs <- data.frame(comparison = c("diff17x22", "diff17x27"</pre>
                                      , "diff17x32", "diff22x27"
                                      , "diff22x32", "diff27x32"
                      , difference = c(bbh[2] - bbh[3], bbh[2] - bbh[4]
                                      , bbh[2] - bbh[5], bbh[3] - bbh[4]
                                      , bbh[3] - bbh[5], bbh[4] - bbh[5]
```

```
)
q_05 \leftarrow qtukey(.95, 4, 12)
                                                                     # 95% TukeySR quantile
diffs$lower_95 <- abs(diffs$difference) - q_05*sqrt(MSE/4)</pre>
                                                                     # Lower 95% bound
diffs$upper_95 <- abs(diffs$difference) + q_05*sqrt(MSE/4)</pre>
                                                                     # Upper 95% bound
diffs$pval <- 1 - ptukey(abs(diffs$difference)/sqrt(MSE/4), 4, 12) # P-Value
##
                             lower_95 upper_95
                                                        pval
     comparison difference
## 1
     diff17x22
                     0.325 -0.4920099 1.1420099 0.649409147
## 2
     diff17x27
                    -1.000 0.1829901 1.8170099 0.015654654
## 3
     diff17x32
                    -1.050
                            0.2329901 1.8670099 0.011391656
## 4
     diff22x27
                    -1.325 0.5079901 2.1420099 0.002063035
                    -1.375 0.5579901 2.1920099 0.001528247
## 5
      diff22x32
## 6
     diff27x32
                    -0.050 -0.7670099 0.8670099 0.997749162
# check
TukeyHSD(aov(ivd ~ as.factor(temp), ivd))
##
     Tukey multiple comparisons of means
##
       95% family-wise confidence level
##
## Fit: aov(formula = ivd ~ as.factor(temp), data = ivd)
##
## $'as.factor(temp)'
##
           diff
                       lwr
                                  upr
                                          p adj
## 22-17 -0.325 -1.1420099 0.4920099 0.6494091
## 27-17 1.000 0.1829901 1.8170099 0.0156547
         1.050
                 0.2329901 1.8670099 0.0113917
## 27-22 1.325
                0.5079901 2.1420099 0.0020630
## 32-22 1.375 0.5579901 2.1920099 0.0015282
## 32-27 0.050 -0.7670099 0.8670099 0.9977492
```

Problem 5

(Adapted from Oehlert, Problem 3.2) Scientists are interested in whether the energy costs involved in reproduction affect longevity. In this experiment, 125 male fruit flies were divided at random into five sets of 25. In one group, the males were kept by themselves. In two groups, the males were supplied with one or eight receptive virgin female fruit flies per day. In the final two groups, the males were supplied with one or eight unreceptive (pregnant) female fruit flies per day. Other than the number and type of companions, the males were treated identically. The longevity of the flies was observed.

The data are in FruitFly.csv on Canvas. The treatments are 1 = no companion, 2 = one pregnant, 3 = one virgin, 4 = eight pregnant, and 5 = eight virgin.

a. Does the number and type of companions affect longevity? Conduct your test at level $\alpha = 0.05$. In your response, provide the null hypothesis, alternative hypothesis, test statistic, the distribution of the test statistic under the null, and a brief conclusion.

Answer: This seems to be asking if any of the groups has a mean that is significantly different than the overall mean, so we would conduct a model utility test, considering the 5 treatment levels.

```
fruitfly <- read_csv("FruitFly.csv")</pre>
N <- nrow(fruitfly)</pre>
                                                     # Total Number of Observations
t <- length(unique(fruitfly$trt))
                                                     # Number of Treatment Levels
Y <- fruitfly$days
                                                     # Response
X <- fruitfly %>%
                                                     # Design Matrix: Single Factor
  mutate(mu = 1)
          , NC = as.numeric(trt == 1)
          , P1 = as.numeric(trt == 2)
          , V1 = as.numeric(trt == 3)
          , P8 = as.numeric(trt == 4)
          , V8 = as.numeric(trt == 5)
         ) %>%
  select(-c(days, trt)) %>%
  as.matrix()
X \leftarrow X[,-1]
P_1 \leftarrow matrix(1/N, N, N)
                                                    # Projection matrix onto 1
P_x \leftarrow X_ %*\% solve(t(X_))%*\%X_) %*\% t(X_)
                                                    # Projection matrix onto X
I <- diag(1, N)</pre>
                                                    # Projection matrix onto R^N
F_{num} \leftarrow (t(Y) %*% (P_x - P_1) %*% Y)/(t-1)
                                                    # MSTrt
F_{den} \leftarrow (t(Y) %*% (I - P_x) %*% Y)/(N-t)
                                                    # MSE
pval \leftarrow 1 - pf(F_num/F_den, t-1, N-t)
## F-test Statistic: 2984.82 / 219.2793 = 13.61195
## P-Value: 3.515622e-09 - Reject
# check
anova(lm(days ~ as.factor(trt), fruitfly))
## Analysis of Variance Table
##
## Response: days
                    Df Sum Sq Mean Sq F value
                                                    Pr(>F)
                     4 11939 2984.82 13.612 3.516e-09 ***
## as.factor(trt)
## Residuals
                   120 26314 219.28
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
```

For testing the hypothesis that number and type of companions has no effect on longevity ($H_0: \alpha_1 = \dots \alpha_5 = 0$) versus the alternative hypothesis that at least one combination of receptive/non-receptive and one/eight companions has a significant effect on longevity, we obtained an F-test statistic of 13.612 which, under the null hypothesis, gives a p-value of < 0.001. This is evidence to suggest that there is a difference in average longevity between at least 2 groups which may be caused by the number and type of companions, so we reject H_0 .

b. Design a set of at least three contrasts that seem meaningful to you in this context. For each contrast, describe its purpose (i.e. what it is comparing). Using Bonferroni correction, create *simultaneous* 95% confidence intervals for your contrasts. Provide the point estimates and CI for each contrast.

Answer:

- We can use the contrast $\alpha_1 \frac{1}{4}(\alpha_2 + \alpha_3 + \alpha_4 + \alpha_5) = 0$ to test whether there is a difference in average longevity between fruit flies with no companion and fruit flies with companions.
- We can use the contrast $\frac{1}{2}(\alpha_2 + \alpha_3) \frac{1}{2}(\alpha_4 + \alpha_5) = 0$ to test whether there is a difference in average longevity between flies with one companion and flies with eight companions.
- We can use the contrast $\frac{1}{2}(\alpha_2 + \alpha_4) \frac{1}{2}(\alpha_3 + \alpha_5) = 0$ to test whether there is a difference between average longevity between fruit flies with companions and fruit flies with non-receptive companions.

```
alpha = 0.05
A \leftarrow rbind(c(0, 1, -.25, -.25, -.25, -.25))
                                                         # These are orthogonal contrasts
            , c(0, 0, .5, .5, -.5, -.5)
, c(0, 0, .5, -.5, .5, -.5)
bp \leftarrow alpha/3
                                                        # Bonferroni Adjusted Sig Level
Ginv \leftarrow diag(c(0, diag(solve(t(X_)%*%X_))))
                                                        # Generalized Inverse of XTX
bbh <- c(0, tapply(Y, fruitfly$trt, mean))</pre>
                                                        # beta hat
Abeta <- A %*% bbh
                                                        # Estimates
AXXinvA <- solve(A %*% Ginv %*% t(A))
F_nums <- t(Abeta^2) %*% AXXinvA
F_den <- F_den
pvals <- 1 - pf(c(F_nums/c(F_den)), 1, N-t)</pre>
fq \leftarrow qf(1-bp, 1, N-t)
lower_95sFWER <- Abeta - fq*sqrt(diag(AXXinvA)*F_den)</pre>
upper_95sFWER <- Abeta + fq*sqrt(diag(AXXinvA)*F_den)</pre>
```

```
## # A tibble: 3 x 6
##
     Estimate 'Lower 95% sFWER' 'Upper 95% sFWER' 'F-Statistic'
                                                                      'P-Value' Decision
##
        <dbl>
                           <dbl>
                                               <dbl>
                                                              <dbl>
                                                                          <dbl> <chr>
          7.4
                                                                        2.73e-2 FTR
## 1
                            -383.
                                                398.
                                                               4.99
## 2
         16.4
                            -420.
                                                453.
                                                              30.8
                                                                        1.73e-7 Reject
## 3
                            -428.
                                                445.
                                                               8.04
                                                                        5.36e-3 Reject
          8.4
```

Something is wrong with the confidence bounds and I can't figure out what.