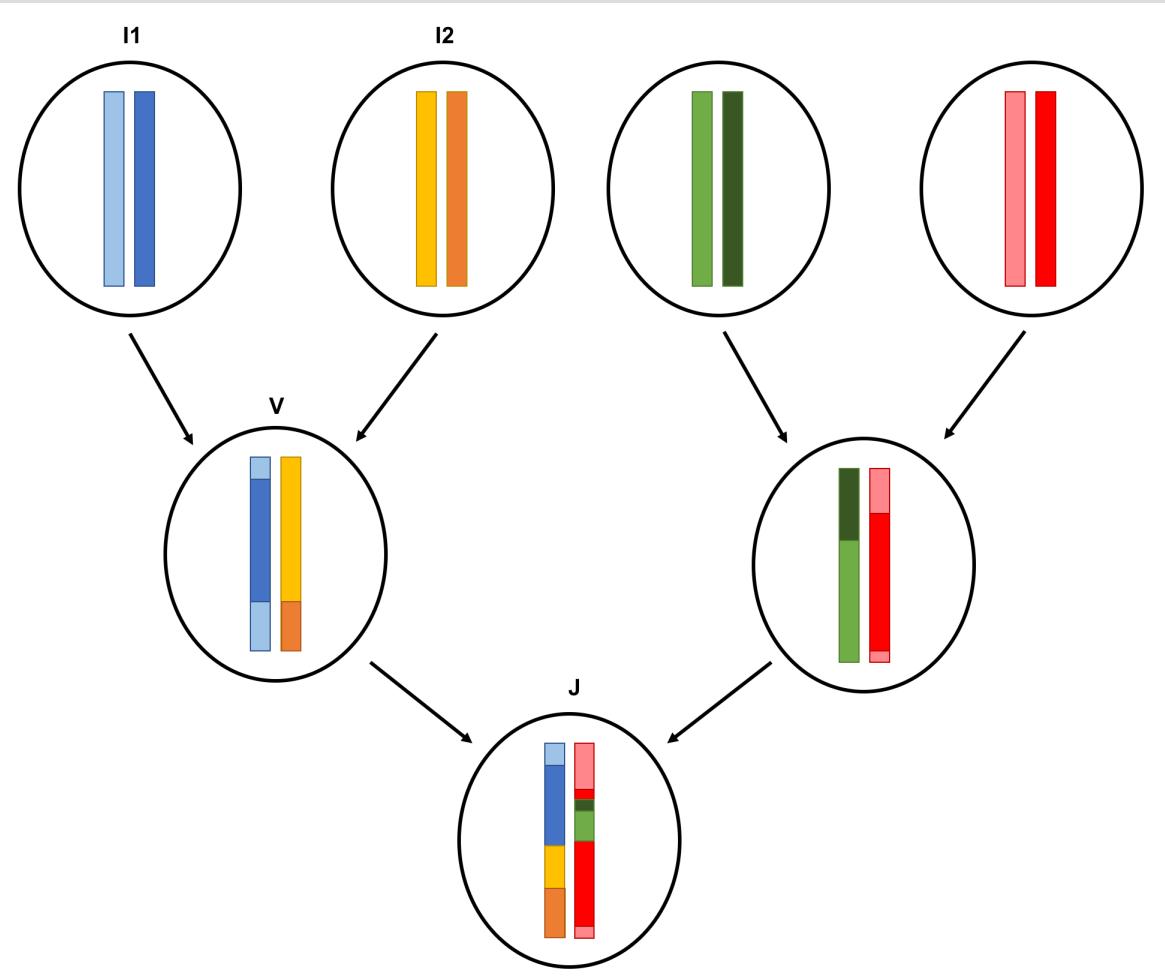


GWAS 7

Matti Pirinen
University of Helsinki
25.11.2020

GENOMIC RECOMBINATION



- Offspring inherits two genomes in continuous segments of the parent's two genomes
- Segments from recent ancestors are longer than from more distant ancestors
- This process determines correlations in GWAS data
 - Concept of genetic relatedness (last week's topic)
 - Sample-by-sample correlation
 - GWAS results at nearby SNPs are correlated (today's topic)
 - SNP-by-SNP correlation

HAPLOTYPES = HAPLOID GENOTYPES

I. True haplotypes

A	T	C
---	---	---

G	C	A
---	---	---

I. Individual has inherited a chromosome with alleles A-T-C from one parent and a chromosome with alleles G-C-A at the same SNPs from the other parent.

These are the two **haplotypes** of the individual at these 3 SNPs.

2. Observed (diploid) genotypes

A/G C/T A/C

3. Possible haplotypes

A	C	A
---	---	---

A	C	C
---	---	---

A	T	A
---	---	---

A	T	C
---	---	---

G	C	A
---	---	---

G	C	C
---	---	---

G	T	A
---	---	---

G	T	C
---	---	---

2. Genotype data does not carry haplotype information for heterozygous loci:
We do not know whether **A** at SNP1 is coming from the same parent as **C** or as **T** at SNP2.

3. **Haplotype phasing** = estimating which are the two haplotypes behind the observed diploid genotypes

EXAMPLE: 2500BPS REGION FROM CHR I

Southern Han Chinese

	Haplotypes			
rs115037027	T	T	T	C
rs12409788	T	C	C	T
rs1576517	C	C	C	T
rs151240271	G	G	G	G
rs12752436	T	T	T	T
rs76864380	T	T	T	A
rs6586443	G	G	A	G
rs35213023	G	G	G	G
rs34910942	C	C	C	C
Counts:	109	54	40	4
Freqs:	0.519	0.2571	0.1905	0.019

Finns

	Haplotypes				
	T	T	T	C	T
	C	C	T	T	T
	C	C	C	T	T
	G	G	G	G	G
	T	T	T	T	C
	T	T	T	A	T
	G	A	G	G	G
	G	G	G	G	A
	C	C	C	C	G
	106	47	34	6	4
	0.5354	0.2374	0.1717	0.0303	0.0202

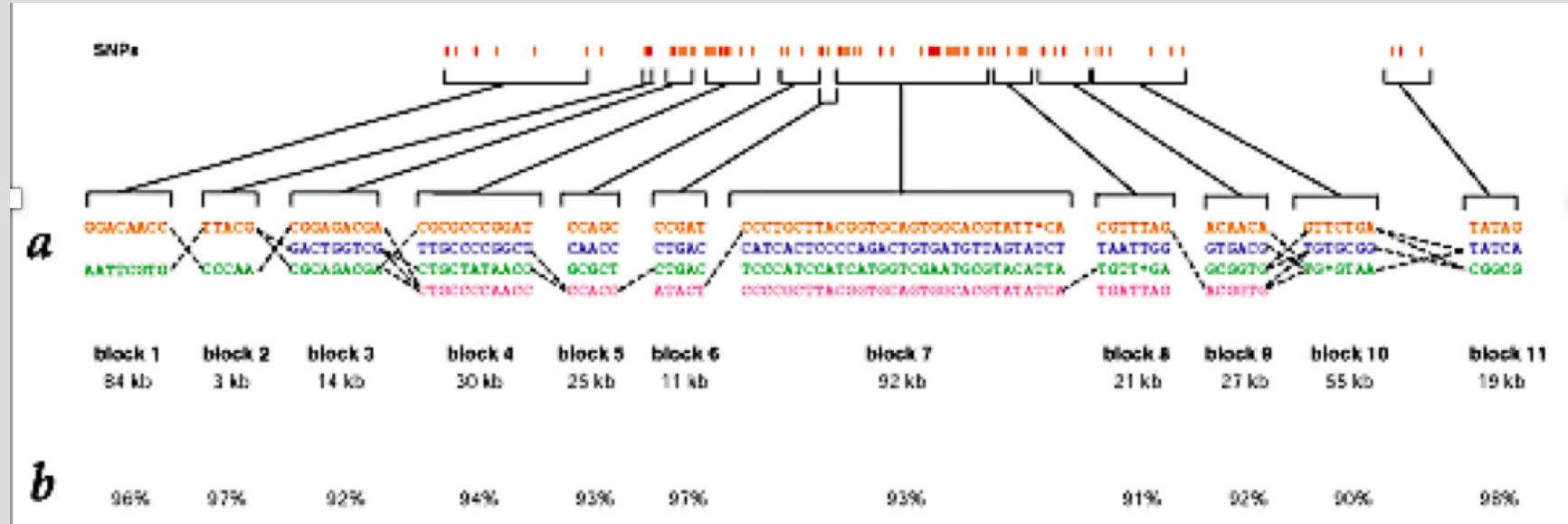
Luhya in Kenya

	Haplotypes								
	T	T	T	T	C	T	T	T	T
	C	T	T	T	T	T	C	T	T
	C	C	T	T	T	C	C	T	T
	G	G	G	G	G	G	T	G	G
	T	T	T	C	T	T	C	T	T
	T	T	T	T	A	T	T	T	A
	G	G	G	G	G	A	G	G	G
	G	G	G	A	G	G	A	A	G
	C	C	C	G	C	C	C	C	C
	65	50	43	15	14	4	4	4	3
	0.3283	0.2525	0.2172	0.0758	0.0707	0.0202	0.0202	0.0152	

From: LDHap

<https://ldlink.nci.nih.gov/?tab=ldhap>

HAPLOTYPE BLOCK STRUCTURE



Haplotype is the sequence of alleles on the same chromosome, or, more generally, sequence of alleles inherited from the same parent.

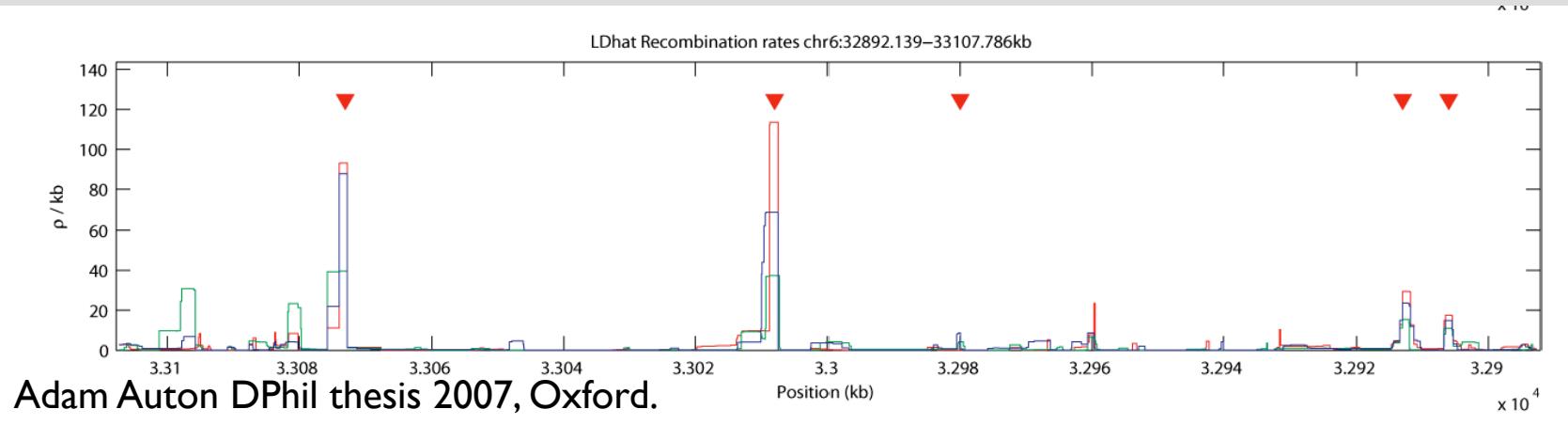
Daly et al. 2001. Nat Gen.

SNP variation in the population is organized as **haplotype blocks**, where recombination seems to happen mainly between the blocks but little within any block.

Consequences to GWAS:

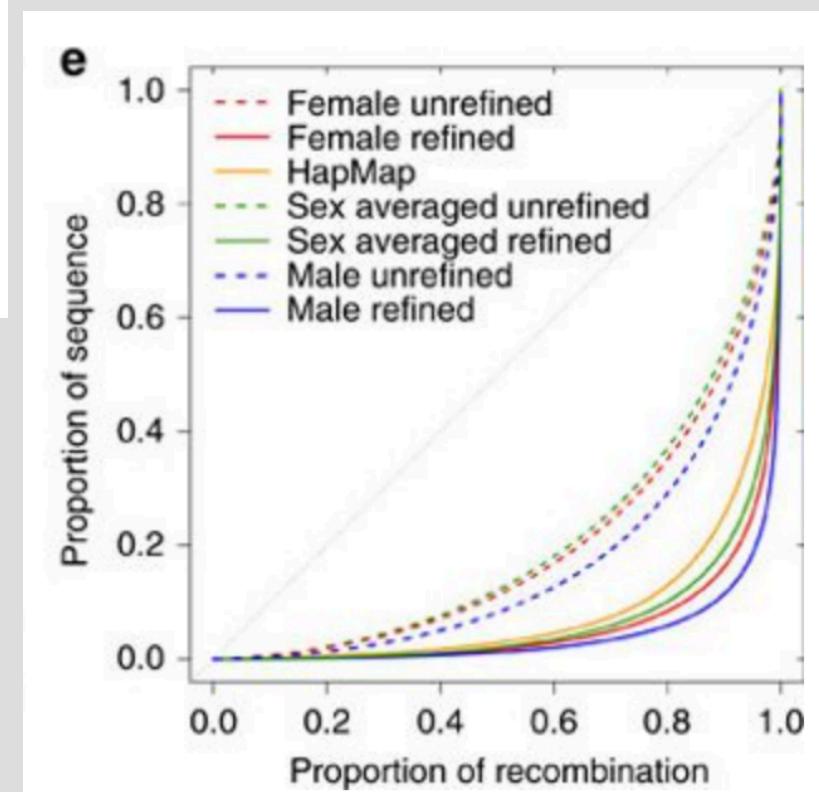
1. A relatively small number of tag-SNPs chosen from the blocks can capture most of (common) variation
2. It may be difficult to know which variant within a block is the causal variant as they are highly correlated

RECOMBINATION HOTSPOTS



Recombination rate as function of physical position on MHC region shows 5 hotspots (triangles). Different colors are rates in different continental populations.

Most recombination events occur in a small proportion genome sequence.



Bherer. et al. 2017 *Nature Communications* 8, 14994

PHASING LEVERAGES SHARING OF HAPLOTYPE BLOCKS

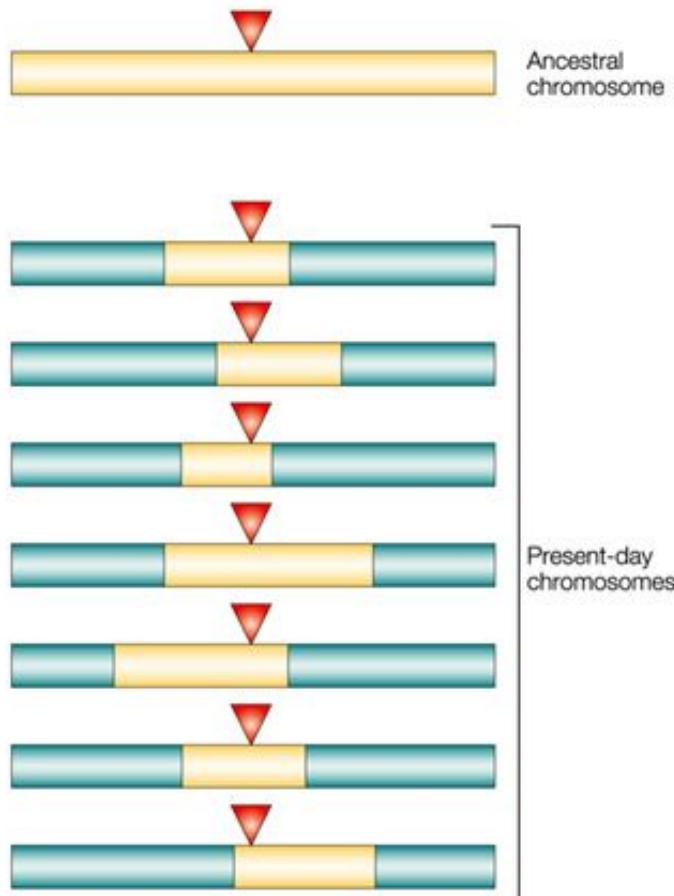
Unphased genotypes	Possible phasing A		Possible phasing B		Possible phasing C		Possible phasing D	
A/C	A	C	A	C	A	C	A	C
G/T	G	T	G	T	T	G	T	G
A/T	A	T	T	A	A	T	T	A
Population haplotype frequency	55%	0%	15%	5%	2%	3%	0%	20%
Population frequency of unordered haplotype pair	0%		$2 \times (15\% \times 5\%) = 1.5\%$		$2 \times (2\% \times 3\%) = 0.12\%$		0%	
Posterior probability of unordered haplotype pair	0%		$1.5\% / (1.5\% + 0.12\%) = 93\%$		$0.12\% / (1.5\% + 0.12\%) = 7\%$		0%	

Browning & Browning. (2011) *Nature Reviews Genetics* **volume 12:** 703–714

Nature Reviews | **Genetics**

Consider one individual with a heterozygous genotype at each of three SNPs in a region. There are four possible haplotype configurations that are consistent with the genotype data (possible phasing patterns A–D). Suppose that haplotype frequencies are available from other individuals in the population at these sites (provided below each phasing pattern). The population frequency of a haplotype pair is obtained using the Hardy–Weinberg principle (independence of the two haplotypes within an individual); the factor of two in the frequency of the haplotype pairs accounts for both possible assignments of maternal and paternal origin to the two haplotypes. The posterior probabilities of the phased data are obtained from the population frequencies of the possible haplotype pairs. In this example, the posterior probability of phasing B (93%) is much greater than that of phasing C (7%).

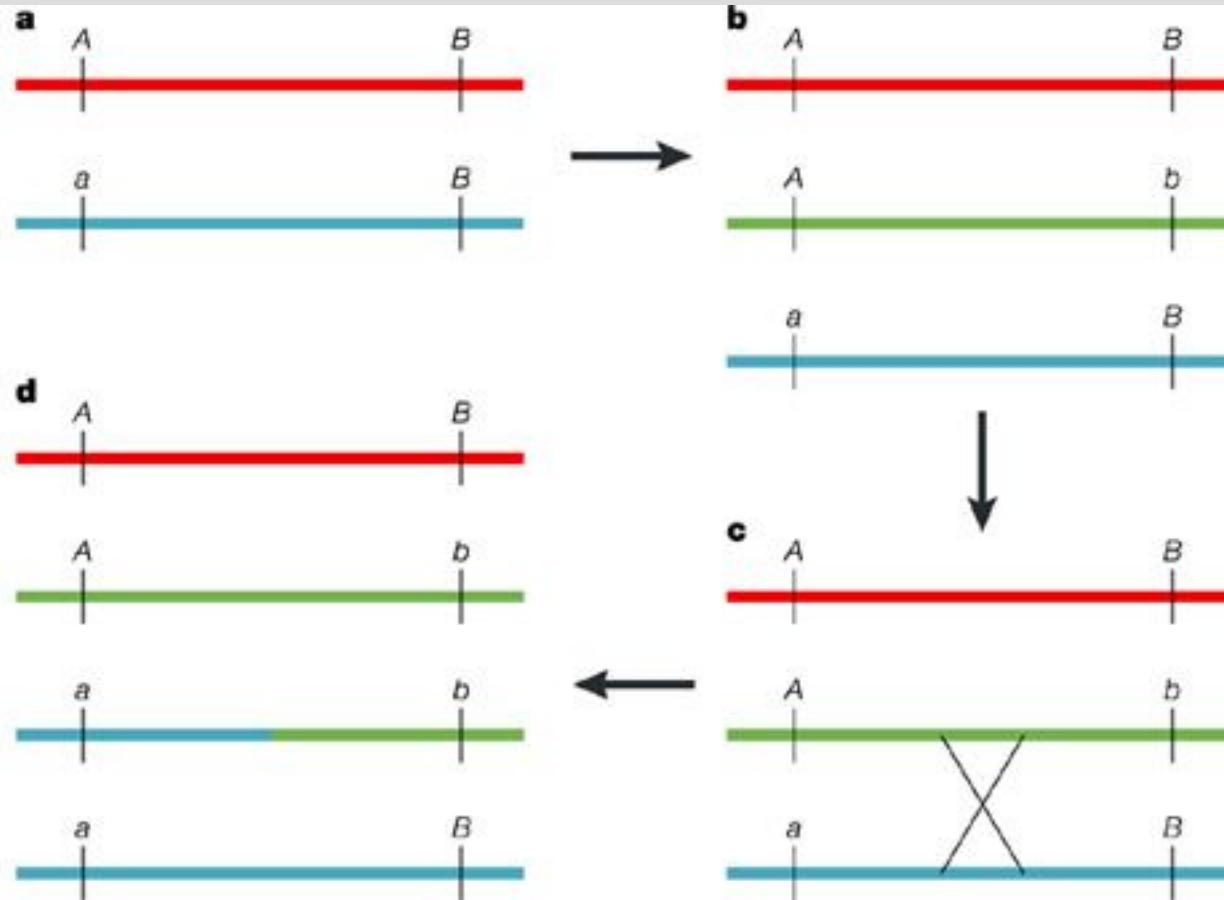
EROSION OF HAPLOTYPE BACKGROUND OF A VARIANT



The mutation is indicated by a red triangle. Chromosomal stretches derived from the common ancestor of all mutant chromosomes are shown in yellow, and new stretches introduced by recombination are shown in blue. Markers that are physically close (that is, in the yellow regions of present-day chromosomes) tend to remain associated with the ancestral mutation even as recombination limits the extent of the region of association over time.

Exactly shared segment around the variant is shrinking over time among the carriers of the variant.

LINKAGE DISEQUILIBRIUM



a | At the outset, there is a polymorphic locus with alleles A and *a*. **b** | When a mutation occurs at a nearby locus, changing an allele B to *b*, this occurs on a single chromosome bearing either allele A or *a* at the first locus (A in this example). So, early in the lifetime of the mutation, only three out of the four possible haplotypes will be observed in the population. The *b* allele will always be found on a chromosome with the A allele at the adjacent locus. **c** | The association between alleles at the two loci will gradually be disrupted by recombination. **d** | This will result in the creation of the fourth possible haplotype and an eventual decline in LD among the markers in the population as the recombinant chromosome (*a*, *b*) increases in frequency.

LDPAIR

CEU
(Central Europe)

rs4242382
chr8:128517573

	A	G		
rs7837688	0	180	180	(0.909)
chr8:128539360	16	2	18	(0.091)
	16	182	198	
	(0.081)	(0.919)		

Haplotypes

G_G: 180 (0.909)
T_A: 16 (0.081)
T_G: 2 (0.01)
G_A: 0 (0.0)

Statistics

D': 1.0
 R^2 : 0.8791
Chi-sq: 174.0659
p-value: <0.0001

rs7837688(G) allele is correlated with rs4242382(G) allele
rs7837688(T) allele is correlated with rs4242382(A) allele

LWK
(Kenya)

rs4242382
chr8:128517573

	A	G		
rs7837688	40	139	179	(0.904)
chr8:128539360	5	14	19	(0.096)
	45	153	198	
	(0.227)	(0.773)		

Haplotypes

G_G: 139 (0.702)
G_A: 40 (0.202)
T_G: 14 (0.071)
T_A: 5 (0.025)

Statistics

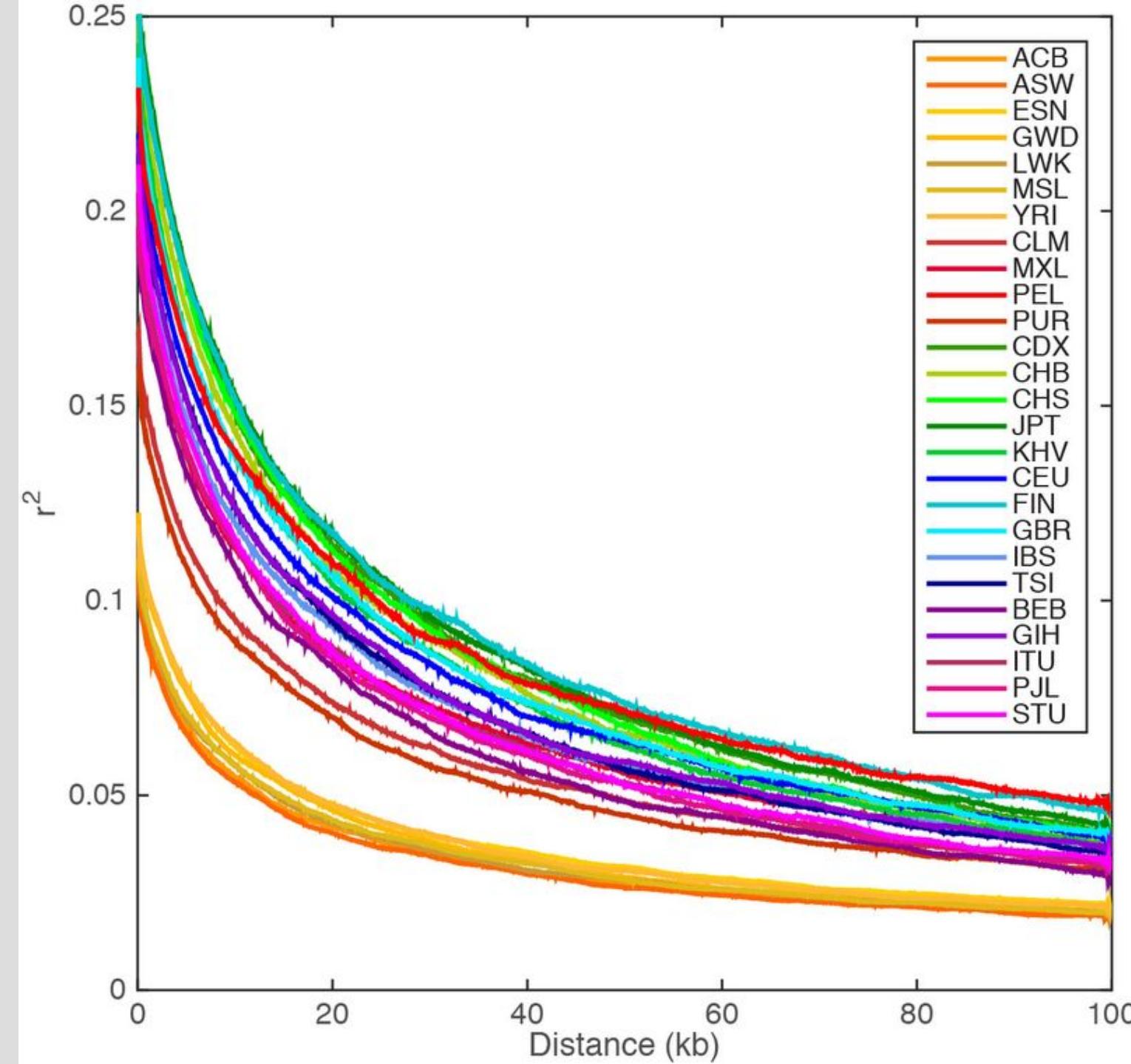
D': 0.0464
 R^2 : 0.0008
Chi-sq: 0.1541
p-value: 0.6946

rs7837688 and rs4242382 are in linkage equilibrium

D' is a normalized version of D that has maximum of 1.

From LDpair
<https://ldlink.nci.nih.gov/>

DECAY OF LD IN HUMANS

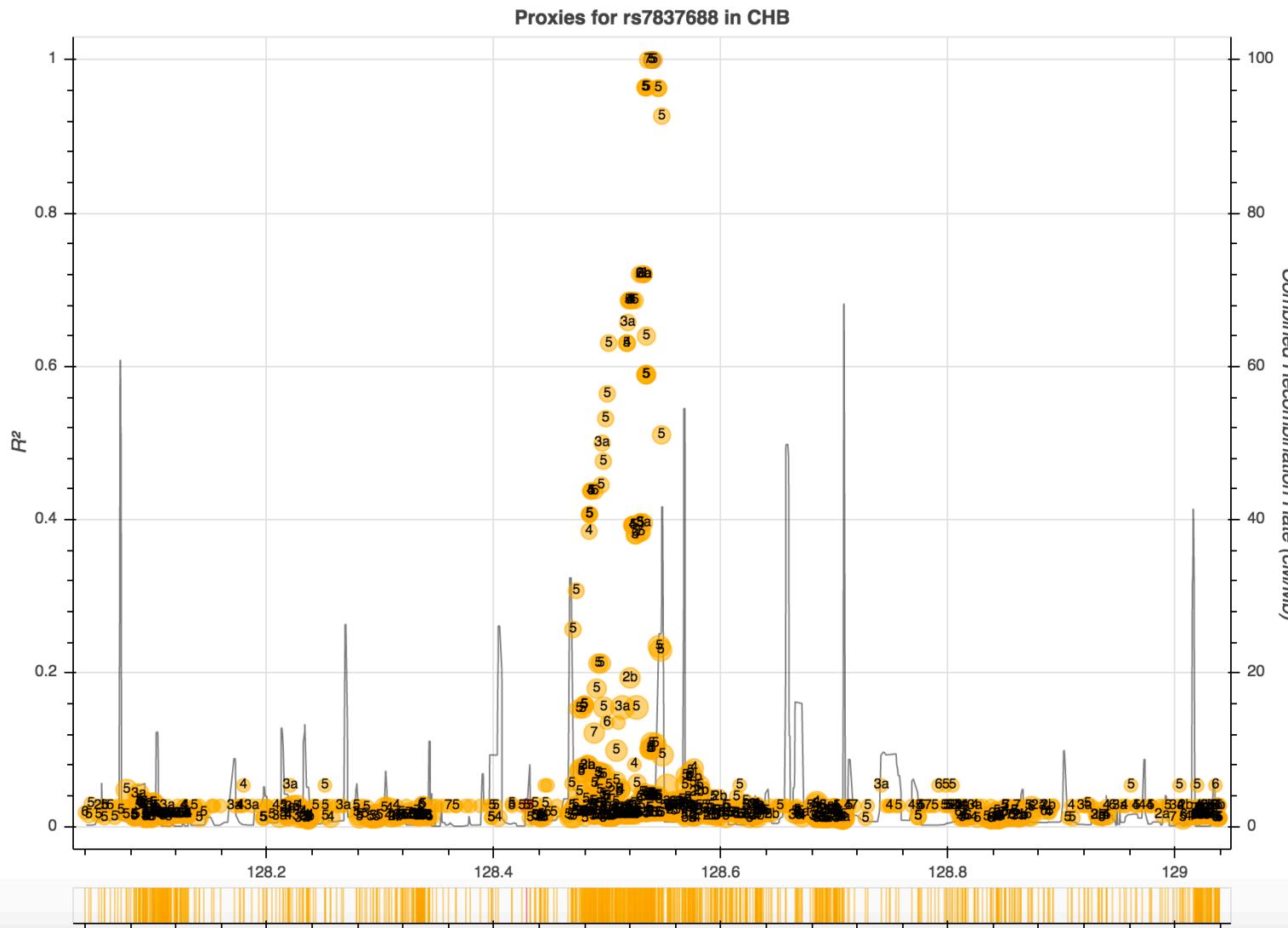


Linkage disequilibrium was calculated around 10,000 randomly selected polymorphic sites in each population, having first thinned each population down to the same sample size (61 individuals). The plotted line represents a 5 kb moving average.

Finns have longest span of LD together with Asian populations followed by Europeans.

Africans have clearly smaller span of LD.

LD FRIENDS



LDproxy gives all other variants that are highly correlated with the target SNP.

Note how r^2 decays after recombination hotspots on either side of the variant.

LDproxy reported 3 variants in perfect LD ($r^2=1$) with this SNP.

We call SNPs in high LD with each other as "LD-friends". Definition of "high" can vary.

rs115037027

rs12409788

rs1576517

rs151240271

rs12752436

rs76864380

rs6586443

rs35213023

rs34910942

rs75047573

rs111549857

rs4847155

rs72683794

rs35748418

rs67818703

rs72683796

rs78376885

rs12760470

rs11185370

rs72683798

rs141938920

rs3811504

rs3811505

rs3811506

rs72685703

rs6664203

rs6698497

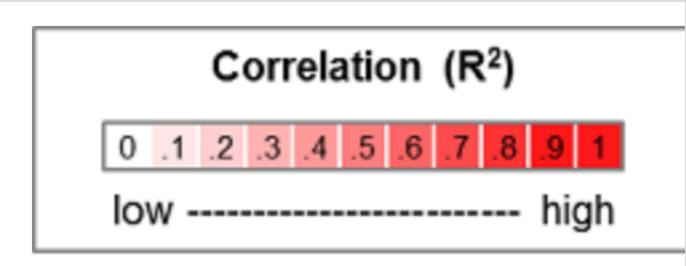
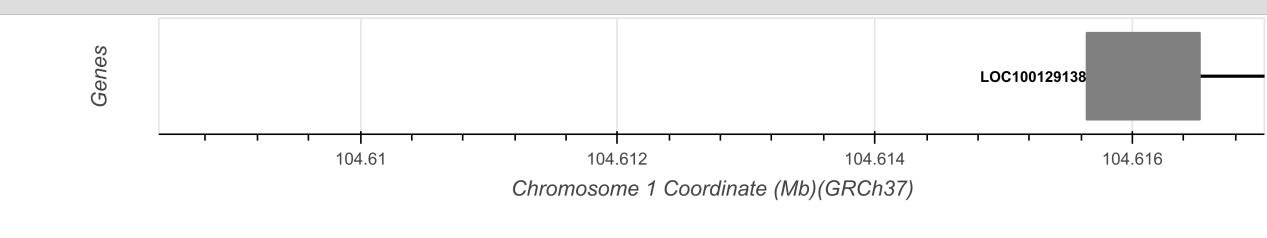
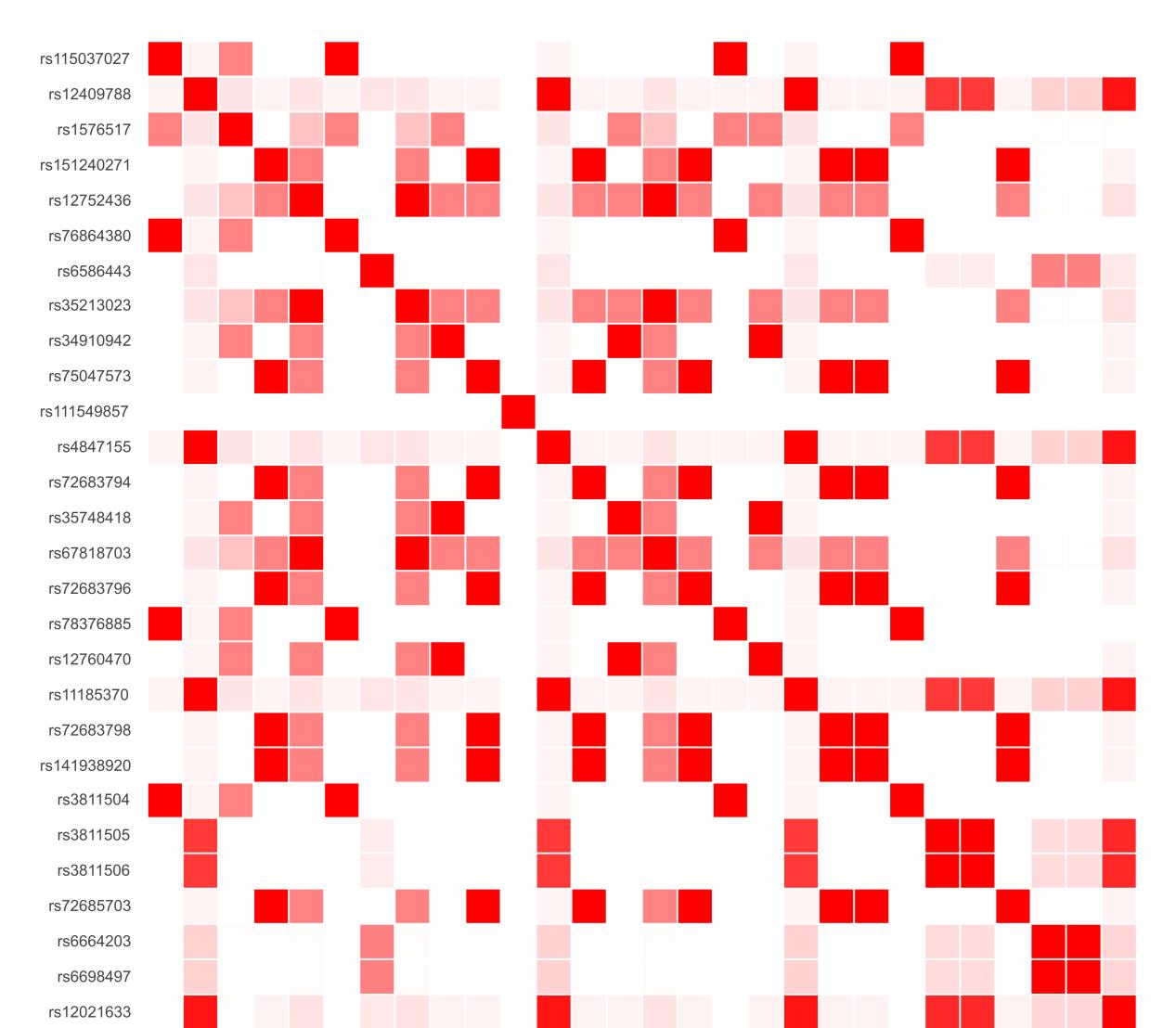
rs12021633

LDMATRIX

<https://ldlink.nci.nih.gov/?tab=ldmatrix>

Makes LD matrix for any 1000G population
For given set of variants.

Example:
CEU population, R² values,
a region from chr 1.



LD IN 4 GENES IN 2 FINNISH COHORTS

Genes:

APOE

CETP

GCKR

PCSK9

Cohorts:

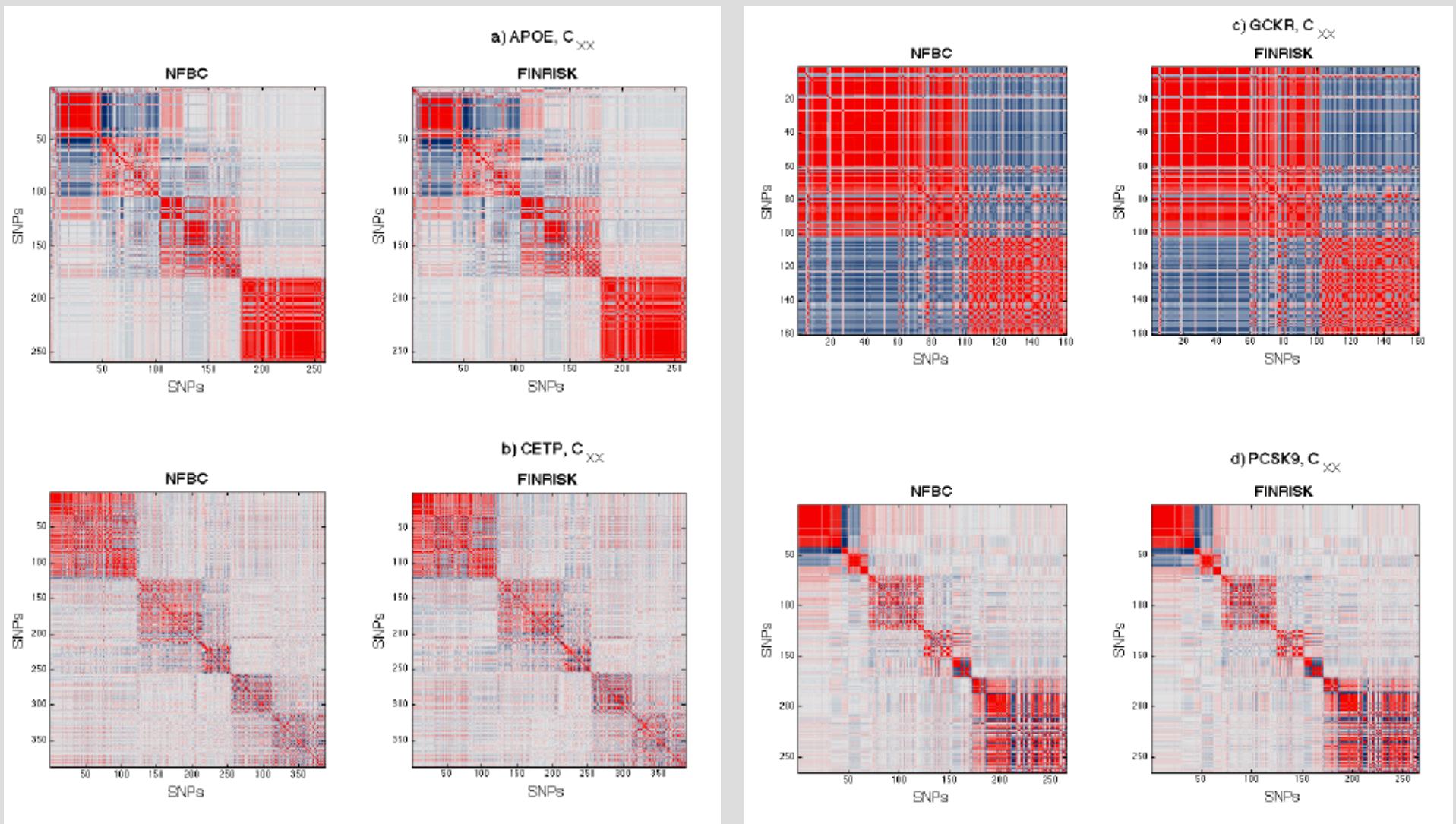
NFBC=

Northern
Finland
Birth Cohort

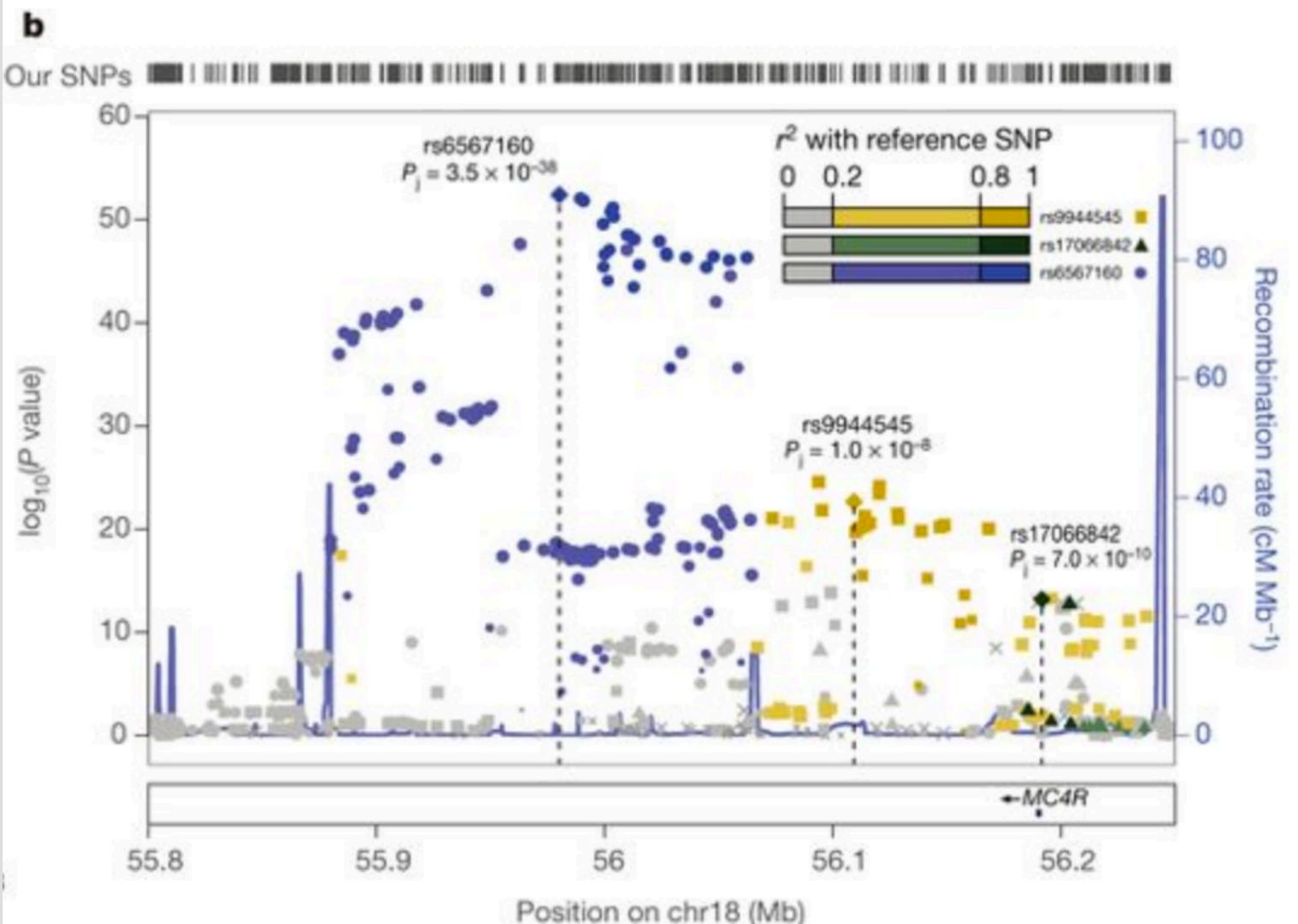
1966 (n~ 4000)

FINRISK (n~3000)

Note how similar
LD is across these
two Finnish
cohorts.



MC4R - BMI REGION (LOCKE ET AL. 2015 NATURE)



SNPs having similar P-values are in high LD with each other. Their genotype data look almost the same and therefore the regression model gives them essentially the same results.

It is not clear based on these marginal P-values alone which (if any of these) variants are the causal ones.

Three lead SNPs have been chosen and LD w.r.t them is shown for their LD-friends

MARGINAL EFFECT AT A NON-CAUSAL SNP

Consider SNPs A and B of which B is causal and its allele I has effect size λ
 What is the marginal effect β at SNP A because of LD with SNP B ?

Haplotype	0 - 0	I - 0	0 - I	I - I
Frequency	f_{00}	f_{I0}	f_{0I}	f_{II}
Effect	0	0	λ	λ

$$\begin{aligned}
 \beta &= E(Y | A = 1) - E(Y | A = 0) \\
 &= \frac{f_{10}}{f_{10} + f_{11}} \times 0 + \frac{f_{11}}{f_{10} + f_{11}} \times \lambda - \frac{f_{00}}{f_{00} + f_{01}} \times 0 - \frac{f_{01}}{f_{00} + f_{01}} \times \lambda \\
 &= \left(\frac{f_{11}}{f_{10} + f_{11}} - \frac{f_{01}}{f_{00} + f_{01}} \right) \times \lambda = \left(\frac{f_{11}}{f_A} - \frac{f_{01}}{1 - f_A} \right) \times \lambda \\
 &= \left(\frac{f_{11}(1 - f_A) - f_{01}f_A}{f_A(1 - f_A)} \right) \times \lambda = \frac{f_{11} - (f_{11} + f_{01})f_A}{f_A(1 - f_A)} \times \lambda \\
 &= \frac{f_{11} - f_B f_A}{\sqrt{f_A(1 - f_A)f_B(1 - f_B)}} \sqrt{\frac{f_B(1 - f_B)}{f_A(1 - f_A)}} \times \lambda = r_{AB} \sqrt{\frac{f_B(1 - f_B)}{f_A(1 - f_A)}} \lambda
 \end{aligned}$$

f_A = allele I frequency at SNP A

f_B = allele I frequency at SNP B

r_{AB} = correlation of allele I at A and B

Conclusion:

The marginal effect at SNP A is shrunk towards 0 by correlation r_{AB} compared to SNP's B causal effect

MARGINAL EFFECT AT A NON-CAUSAL SNP



Marginal effect at SNP A is a linear combination of the causal effects of all variants in LD with A, where the weights are the correlations with A (after scaling the genotypes).

$$\beta_A^* = \lambda_A^* + r_{A1} \lambda_1^* + r_{A2} \lambda_2^* + r_{A3} \lambda_3^* + r_{A4} \lambda_4^* + \dots$$

* denotes **scaled effect**: the allelic effect multiplied by $\sqrt{2f(1 - f)}$, where f is MAF of the SNP

MASKING EFFECT BY LD

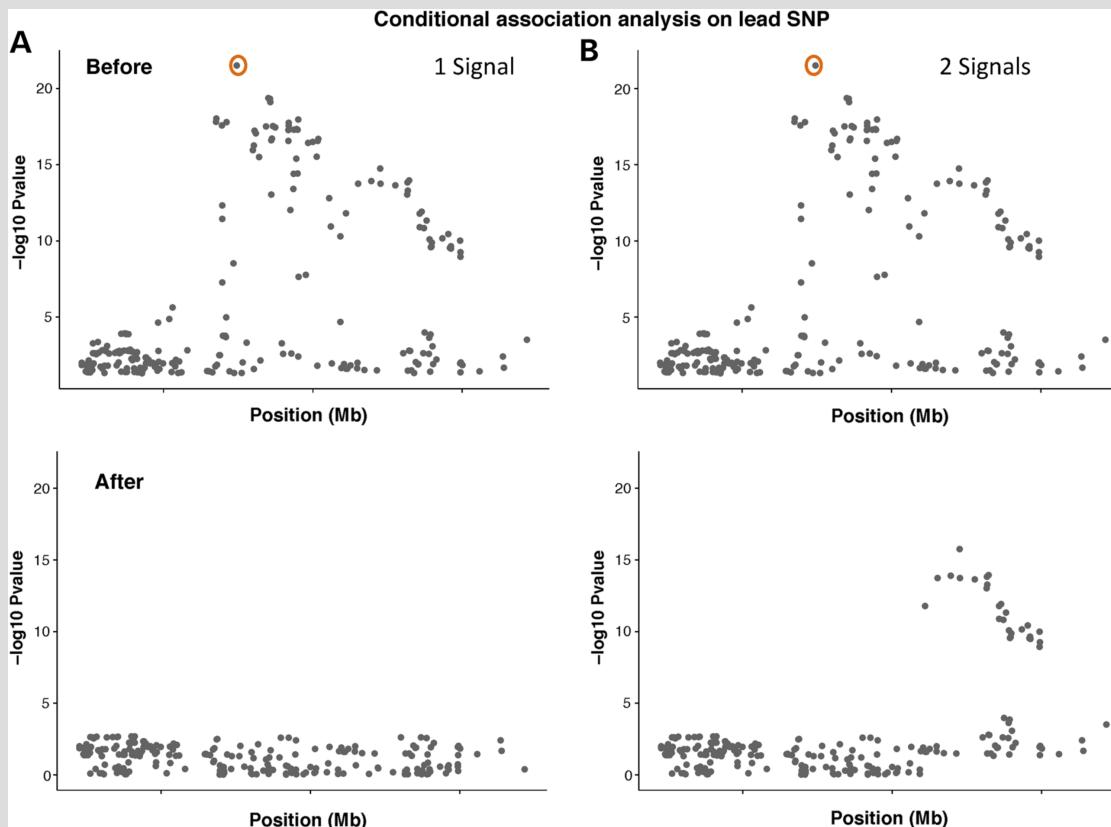
		rs7687945				
		G	A			
rs356220	G	1	1.16 (1.04-1.29)	1.11	1.26 (1.16-1.37)	
		20.8%	41.9%			
	A	1.31 (1.17-1.47)	1.64 (1.41-1.90)	1.39		
		1.18	1.25			
		1.07 (0.98-1.15)				

Viewing the data this way makes clear that the risk allele at the second SNP rs7687945 is more commonly found with the protective allele at rs356220 than would be expected were the SNPs in linkage equilibrium. As a result, the unconditional risk of the rs7687945 A allele (1.25) relative to the G allele (1.18) is 1.07 and not significantly different from 1.0.

Shown in the centre of the table are estimates of odds ratio, 95% confidence limit (in brackets) and percentage frequency of the four haplotypes defined by the alleles at rs356220 and rs7687945. In the margins of the table is the risk of each of the alleles obtained by averaging the odds ratio of two haplotypes on which the allele can be found, weighting by the sample frequency. For example, the risk of carrying the G allele at rs356220 unconditional on the allele carried at rs7687945 is 1.11 (given in the top right) and is calculated as $(1 \times 20.8 + 1.16 \times 41.9)/(20.8 + 41.9)$. By comparing the unconditional risks of the two alleles at each SNP, we recover the odds ratio estimated from a single SNP analysis.

STEPWISE FORWARD SEARCH

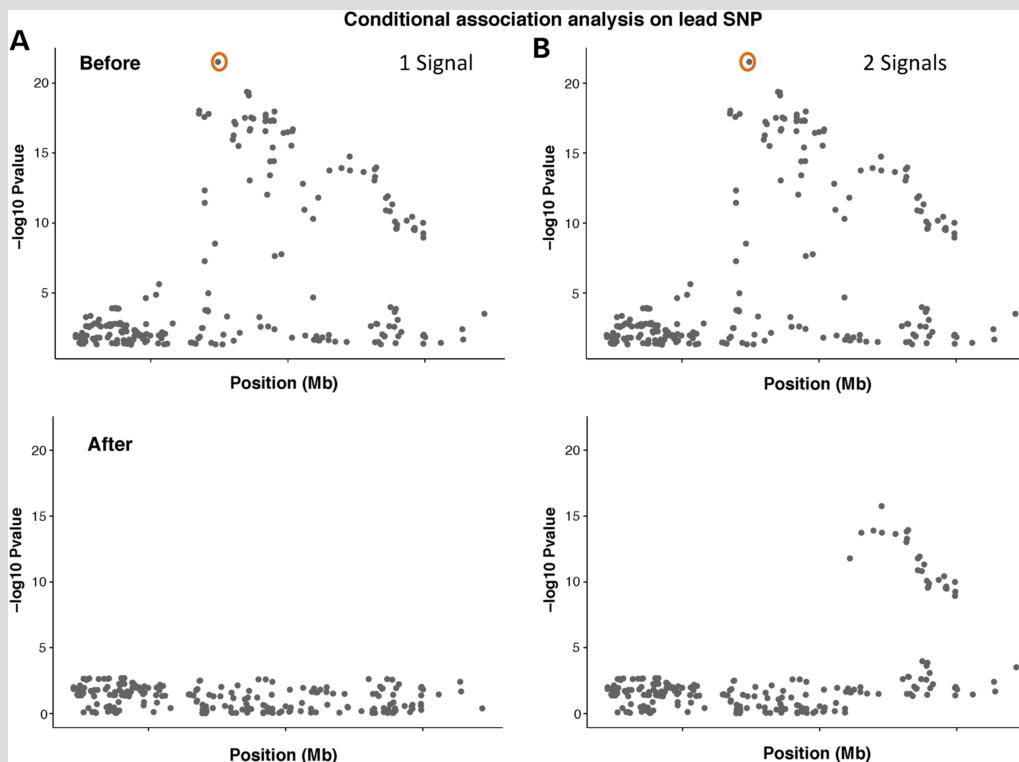
- Starts by conditioning on the lowest P-value
- Continues until no additional variant reaches pre-defined P-value threshold



Spain & Barrett 2016

STEPWISE FORWARD SEARCH

- + Informs about multiple causal variants accounting for LD
- Does not necessarily find the optimal configuration
- Completely ignores the uncertainty of the causal configurations

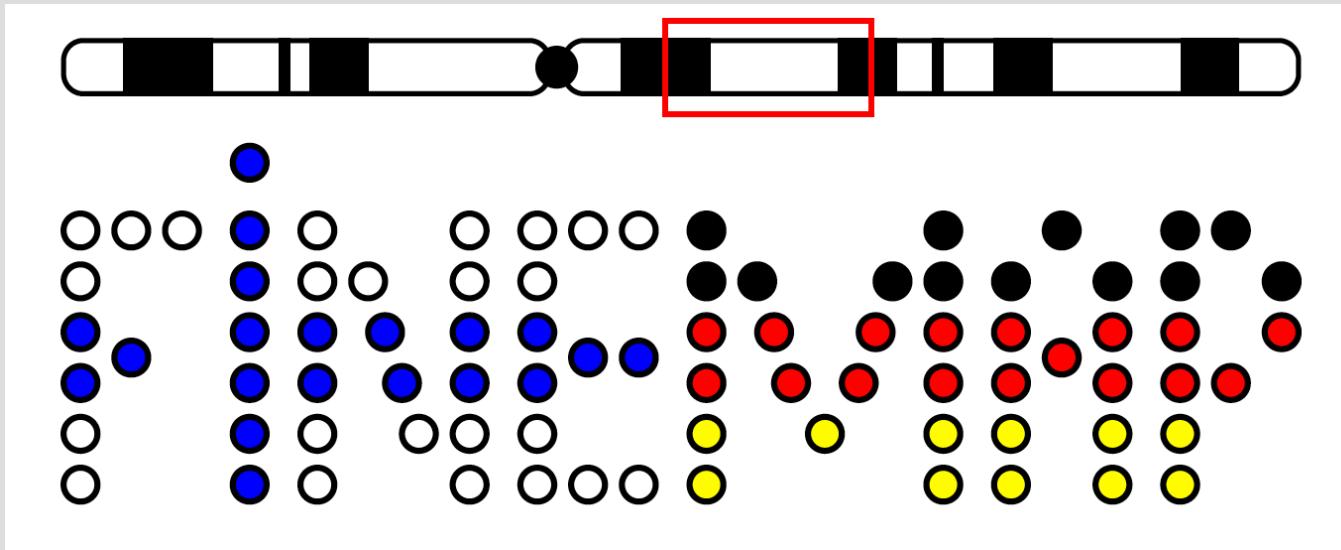


FINE-MAPPING

- Assign probabilities for each variant that the variant is (one of) the causal variant(s)
- Done in Bayesian framework with prior assumptions
 - Prior probability of each variant being a causal variant?
 - Default: Each variant is equally likely to be causal
 - Prior distribution of the non-zero effects of the causal variants?
 - Default: $N(0, \tau^2)$ (See GWAS 4 for how to set tau)

FINE-MAPPING ASSUMING 1 CAUSAL VARIANT

- If there is exactly one causal variant in the region and it is among the genotyped variants, then the posterior probability of being causal is proportional to single-SNP marginal Bayes factor of association (GWAS4)
- This idea can be extended to fine-mapping each independent signal of the region when we have conditioned on the other signals in the region when we have computed the GWAS statistics (betas and SEs)
- For multiple causal variants, we use methods such as FINEMAP



Bioinformatics, 32(10), 2016, 1493–1501

doi: 10.1093/bioinformatics/btw018

Advance Access Publication Date: 14 January 2016

Original Paper

Genetics and population analysis

FINEMAP: efficient variable selection using summary data from genome-wide association studies

Christian Benner^{1,2*}, Chris C.A. Spencer³, Aki S. Havulinna⁴, Veikko Salomaa⁴, Samuli Ripatti^{1,2,5} and Matti Pirinen^{1*}



Christian Benner

Defended PhD thesis on FINEMAP in January 2019

BAYESIAN MODEL FOR FINE-MAPPING

- Define causal configuration γ as a binary vector for variants

$$\gamma = \begin{array}{|c|c|c|c|c|c|c|c|c|} \hline 0 & 0 & 1 & 0 & 0 & 0 & 1 & 0 \\ \hline \end{array}$$

- This configuration says that variants 3 and 7 are causal and the others have no effects.

BAYESIAN MODEL FOR FINE-MAPPING

- Define causal configuration γ as a binary vector for variants

$$\gamma = \begin{array}{cccccccc} 0 & 0 & 1 & 0 & 0 & 0 & 1 & 0 \end{array}$$

- In total there are 2^p configurations on p variants, but we will assume that only **sparse** configurations are plausible, say those with < 10 causal variants per a region.
- Ultimate goal is to compute probability for each configuration, given the observed GWAS data in the region.

BAYESIAN MODEL FOR FINE-MAPPING

- Define causal configuration γ as a binary vector for variants
- Each causal variant picks its effect from $N(0, s^2)$

$$p(\lambda|\gamma) = \mathcal{N} (\mathbf{0}, s^2 \Delta_\gamma)$$

Causal configuration γ

1	0	0	0	0	0	0	0	0	0	0
---	---	---	---	---	---	---	---	---	---	---

$$\Delta_\gamma = \text{diag}(\gamma) = \begin{bmatrix} 1 & & & & & & & & \\ & 0 & & & & & & & \\ & & \ddots & & & & & & \\ & & & 0 & & & & & \end{bmatrix}$$

BAYESIAN MODEL FOR FINE-MAPPING

- Define causal configuration γ as a binary vector for variants
- Each causal variant picks its effect from $N(0, s^2)$
- For each configuration compute the Bayes factor (BF), i.e., how well the configuration explains the data relative to the null model

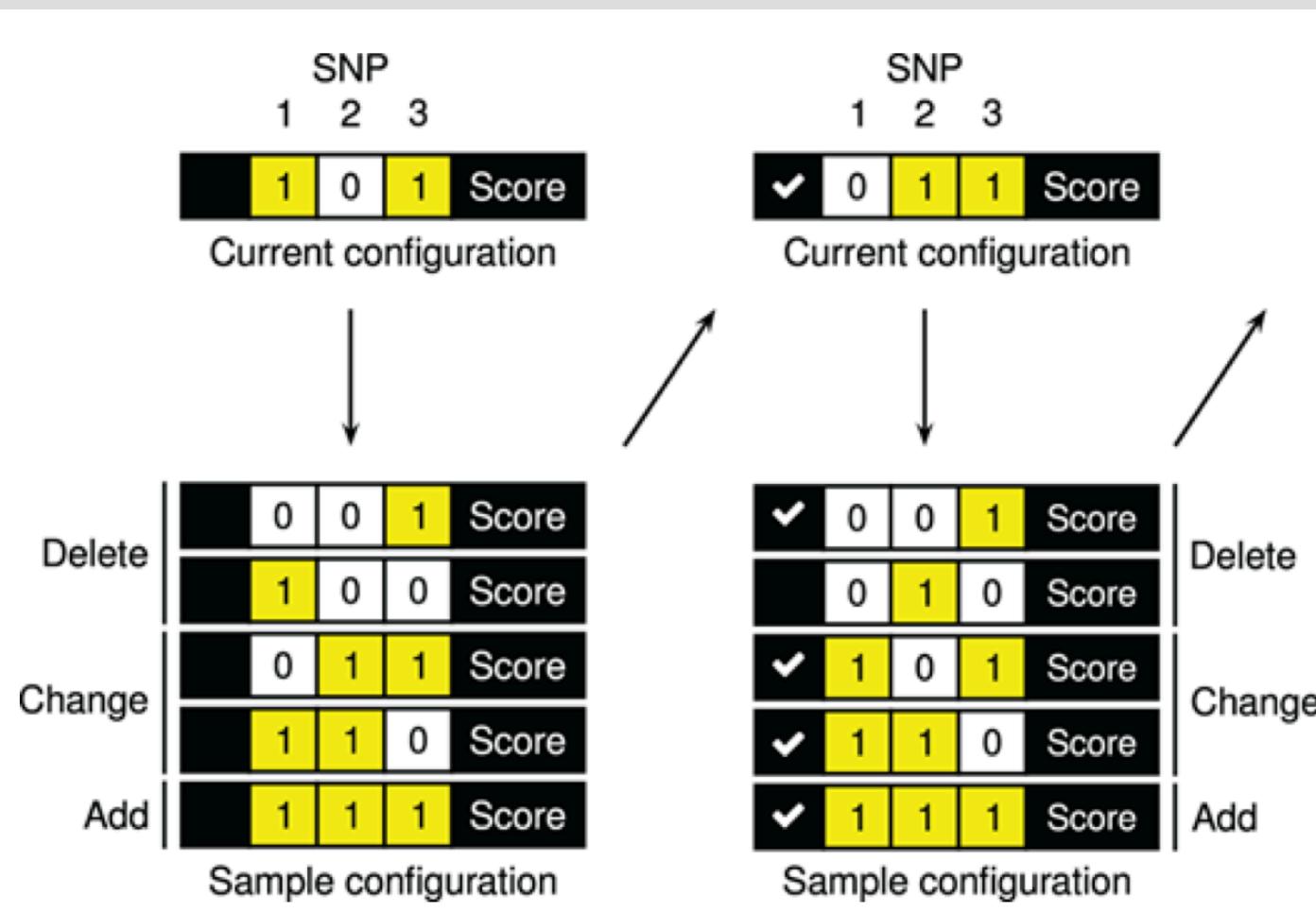
$$BF_{\gamma} = \frac{P(\text{DATA}|\gamma)}{P(\text{DATA}|\text{NULL})} = \frac{\mathcal{N}(\mathbf{z}|0, \mathbf{R} + s^2 \mathbf{R} \Delta_{\gamma} \mathbf{R})}{\mathcal{N}(\mathbf{z}|0, \mathbf{R})}$$

BAYESIAN MODEL FOR FINE-MAPPING

- Define causal configuration γ as a binary vector for variants
- Each causal variant picks its effect from $N(0, s^2)$
- For each configuration compute the Bayes factor (BF), i.e., how well the configuration explains the data relative to the null model
- By combining BFs with prior probabilities of configurations we get the posterior probabilities

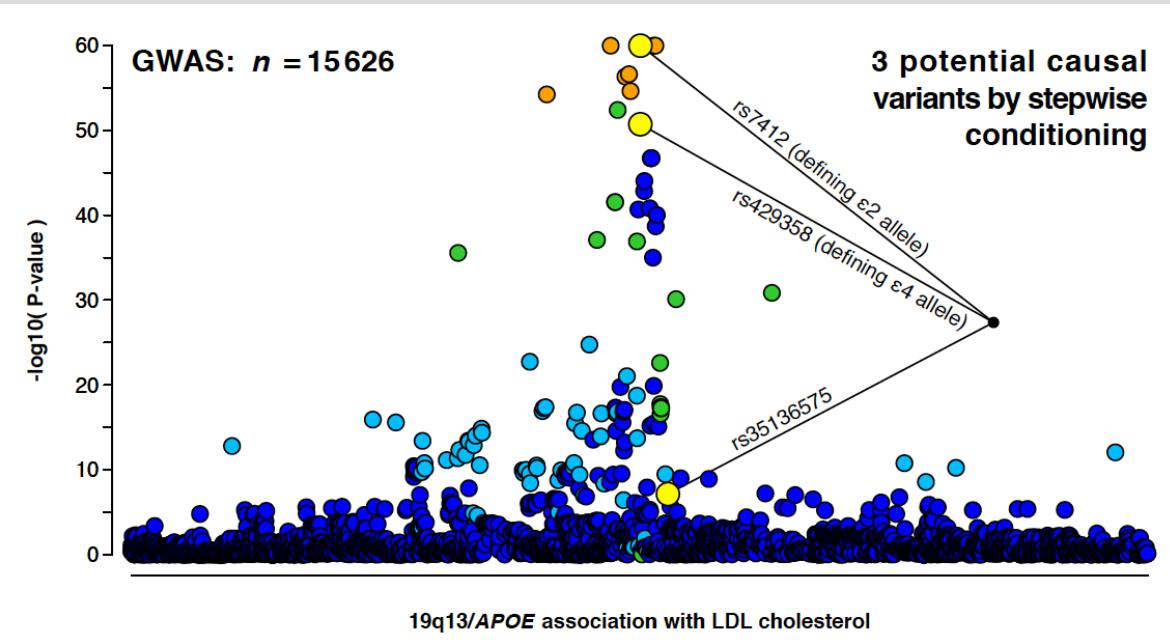
$$p_\gamma = P(\gamma | \text{DATA}) \propto \text{prior}_\gamma \times \text{BF}_\gamma$$

FINEMAP ALGORITHM

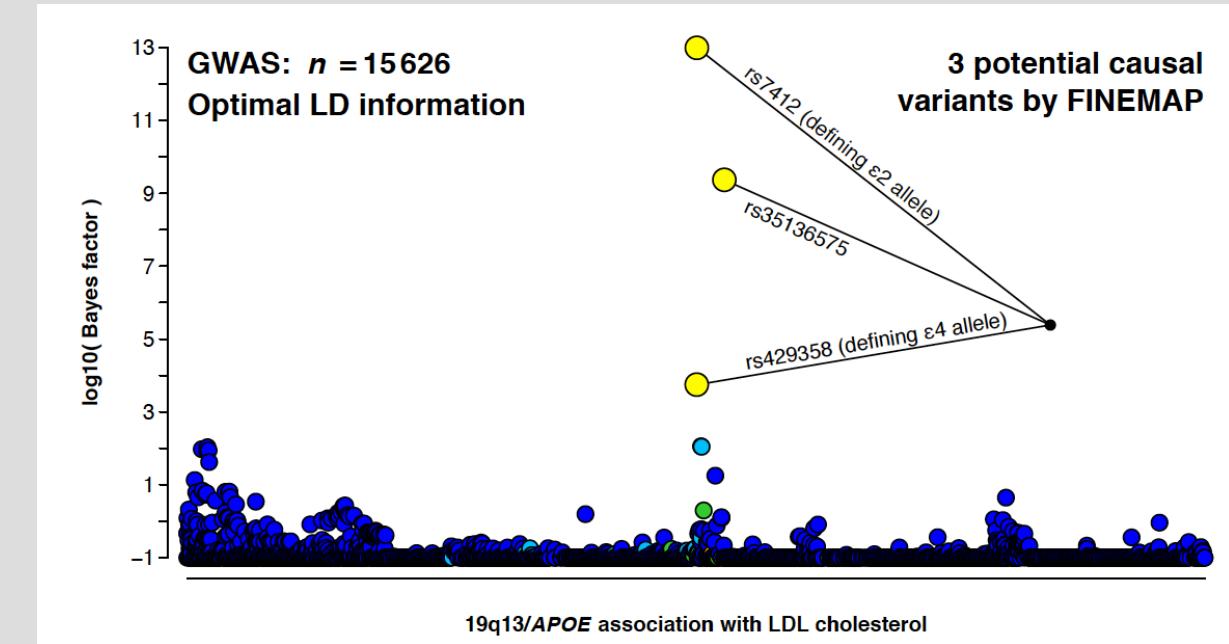


- Collect configurations from a high probability region using Shotgun stochastic search (Hans et al. 2007)
 - Memorize BFs of all those configurations seen during the search
 - Stop once not much new probability mass is found
 - Renormalize posteriors with respect to the configurations visited

FINEMAP RESULTS



GWAS



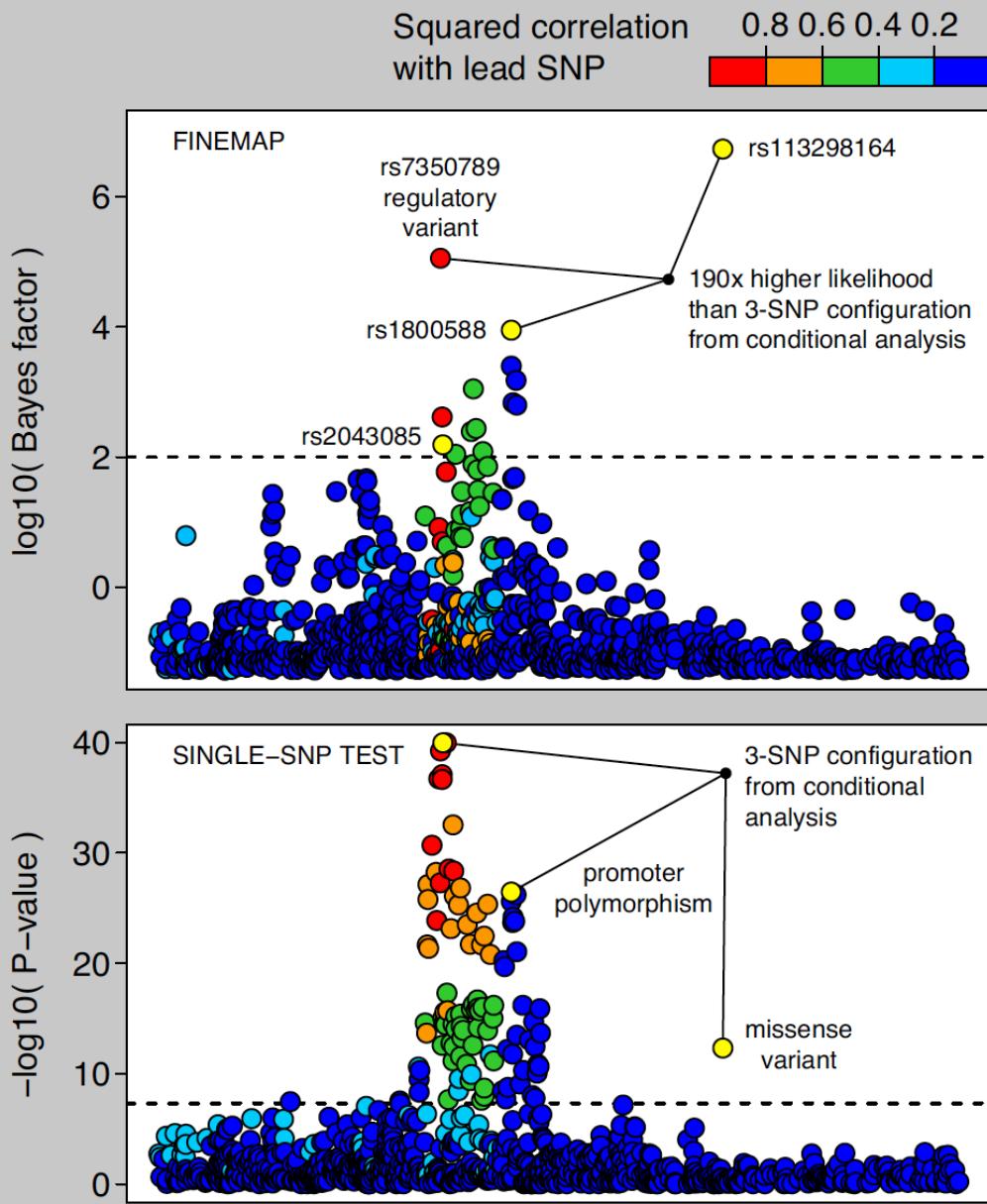
FINEMAP

15q21/*LIPC* association with HDL cholesterol



Christian
Benner

Surakka et al.
Nat. Genet. 2015



NATIONAL INSTITUTE FOR
HEALTH AND WELFARE
FINLAND
FINRISK STUDY
20000 individuals