Missing Data

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(N	ote: Updated and modified from the hst953-edx github version.)	

Source MIMIC-III demo version data

In the original version (on the hst953-edx github site), they used the MIMIC-III demo version directly loaded. Now, I have in <code>project_base_dir/database/mimic3.db</code> the SQLite version of the full MIMIC-III v1.4 database loaded. I'll use that in the processing below - with some pre-coded inclusion criteria to extract just the demo data. The following code chunk attaches the database and loads auxiliary functions for extracting database data (<code>db_functions.R</code>) and for doing some MIMIC data interpretation and pre-processing (mimic3_meta_data.R) - including the processing to get just the demo data.

```
base_dir <- here::here("")
db_dir <- fs::path(base_dir, "database")
db_file <- fs::path(db_dir, "mimic3.db")

if(dbCanConnect(RSQLite::SQLite(), db_file)) {
    mimic3 <- dbConnect(RSQLite::SQLite(), db_file)
}

source(fs::path(base_dir, "db_functions.R"))
source(fs::path(base_dir, "mimic3_meta_data.R"))</pre>
```

Extract base data

These are the tibbles used in the following exercises:

```
adm <- db_get_admissions(mimic3, where = demo_subject_ids)

icu <- db_get_icustays(mimic3, where = demo_subject_ids)

icu_adm <- adm %>%
    left_join(icu, by = c("SUBJECT_ID", "HADM_ID"))

vitals <- db_get_chartevents(mimic3, where = demo_subject_ids)</pre>
```

Exercise: Selection of variables

Aim: Select variables to analyze missing values.

Vital Signs

D_ITEMS is sourced from two distinct ICU databases. The main consequence is that there are duplicate ITEMID for each concept. For example, heart rate is captured both as an ITEMID of 211 (CareVue) and as an ITEMID of 220045 (Metavision). As a result, it is necessary to search for multiple ITEMID to capture a single concept across the entire database.

For more information read: https://mimic.mit.edu/docs/iii/tables/d_items/

In GitHub you may find code regarding vital signs and respective items IDs:

https://github.com/MIT-LCP/mimic-code/blob/main/mimic-iii/concepts/firstday/vitals_first_day.sql

We will use data from chartevents, namely heart rate (hr) and pulse oximetry (SpO2) measurements. Attention to the units. Since much of the information is manually typed in the system, human error can always be present.

Heart rate data:

```
SELECT ITEMID, LABEL, ABBREVIATION, DBSOURCE, LINKSTO, CATEGORY, UNITNAME, PARAM_TYPE, CONCEPTID FROM D_ITEMS
WHERE ITEMID in (211, 220045);
```

Table 1: 2 records

ITEMIDLABEL		ABBREVIA TIOS OURCELINKSTO			CATEGORY	UNITNA	T CDE CEPTID	
211	Heart Rate		carevue	chartevents	NA			NA
220045	Heart Rate	HR	metavision	chartevents	Routine Vital Signs	bpm	Numeric	NA

Pulse oximetry data:

```
SELECT ITEMID, LABEL, ABBREVIATION, DBSOURCE, LINKSTO, CATEGORY, UNITNAME, PARAM_TYPE, CONCEPTID FROM D_ITEMS
WHERE ITEMID in (646, 220277);
```

Table 2: 2 records

ITEMILABEL	ABBREVIA	ATDBSOURC	CEINKSTO	CATEGORYUNITN	AMARAM_	TCOPNECEPTID
646 SpO2 220277 O2 saturation pulseoxymetry	SpO2		chartevents chartevents	NA Respiratory %	Numeric	NA NA

Review the amount of missing values in both vitals.

```
hr <- vitals %>%
  filter(((ITEMID == 211) | (ITEMID == 220045)) & (toupper(VALUEUOM) == "BPM"))
summary(hr$VALUENUM)
#>
     Min. 1st Qu. Median
                              Mean 3rd Qu.
                                              Max.
                                                      NA's
#>
      0.00
           75.00
                     87.00
                             87.77 100.00 189.00
                                                         5
length(hr$VALUENUM)
#> [1] 15490
na_hr <- hr %>% filter(is.na(VALUENUM)) %>% count() %>% pull(n)
sp <- vitals %>%
  filter(((ITEMID == 646) | (ITEMID == 220277)) & (VALUEUOM == "%"))
summary(sp$VALUENUM)
     Min. 1st Qu.
                   Median
                              Mean 3rd Qu.
                                              Max.
                                                      NA's
      0.00
           96.00
                     98.00
                             97.01 100.00 100.00
                                                        15
length(sp$VALUENUM)
#> [1] 15315
na_sp <- sp %>% filter(is.na(VALUENUM)) %>% count() %>% pull(n)
```

We observe there are 5 missing values ("NA") for heart rate and 15 "NA" for SpO2. Heart rate has a slightly higher frequency of measurement, (however this is a demo version of the database).

Exercise: Handling missing data in dataframes

Two of the simplest methods to handle missing data are presented below. Recoding a missing value consists of assigning a value to an already existing value (eg. outlier) which we want to recode as missing. Excluding missing values can be performed by excluding objects (patients, rows in the dataframe) or variables (columns in the dataframe) with significant amount of missing data (see this chapter contents for more detailed theory).

Outliers

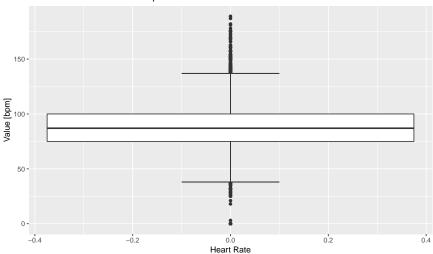
Before handling missing data, we should analyze the presence of outliers (refer to the respective chapter in the course). If we apply an imputation method before processing outliers, our imputation will be based on incorrect data and therefore not valid.

Looking at the distribution of heart rate for all patients, we can assess potential outliers through a box plot.

Observe the distribution of heart rate for all the patients in the dataset.

```
#Preferring ggplot to (arguably easier for more interactive plots) plotly #p <- plot_ly(y = ^hr$VALUENUM, name = "Heart rate", type = "box") %>% #layout(
```

Heart rate distribution for all patients



```
uw <- quantile(hr$VALUENUM, 3/4, na.rm = TRUE) + (1.5 * IQR(hr$VALUENUM, na.rm = TRUE))
lw <- quantile(hr$VALUENUM, 1/4, na.rm = TRUE) - (1.5 * IQR(hr$VALUENUM, na.rm = TRUE))
above <- sum(hr$VALUENUM > uw, na.rm = TRUE)
below <- sum(hr$VALUENUM < lw, na.rm = TRUE)</pre>
```

We observe there are a few outliers. 100 observations lie above the upper whisker (137.5) and 39 observations lie below the lower whisker (37.5).

Missing data recoding

Aim: Recode a value as missing.

All of the heart rate values that we see in these data are within a reasonable range. If there were extreme outlier values (greater than, say, 300, or less than zero), we would want to correct these. We can recode all values outside the physiological ranges [0, 300] bpm as "NA" in the dataset for all patients.

```
hr <- hr %>%
mutate(
    VALUENUM = if_else((VALUENUM < 0) | (VALUENUM > 300), NA_real_, VALUENUM)
)
```

```
summary(hr$VALUENUM)

#> Min. 1st Qu. Median Mean 3rd Qu. Max. NA's

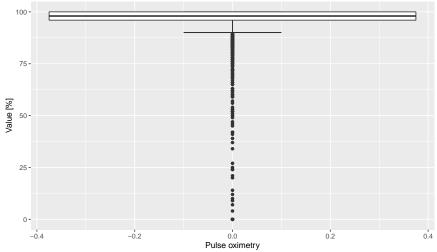
#> 0.00 75.00 87.00 87.77 100.00 189.00 5
```

And now we have a complete dataset with missing data preprocessed for heart rate measurements.

Now repeat the process for pulse oximetry (SpO2).

Observe the distribution of SpO2 for all the patients in the dataset.

Pulse oximetry distribution for all patients



```
uw <- quantile(sp$VALUENUM, 3/4, na.rm = TRUE) + (1.5 * IQR(sp$VALUENUM, na.rm = TRUE))
lw <- quantile(sp$VALUENUM, 1/4, na.rm = TRUE) - (1.5 * IQR(sp$VALUENUM, na.rm = TRUE))
above <- sum(sp$VALUENUM > uw, na.rm = TRUE)
below <- sum(sp$VALUENUM < lw, na.rm = TRUE)</pre>
```

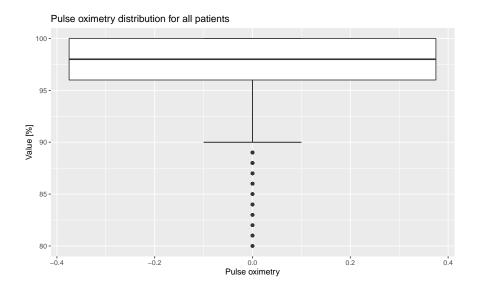
We observe there are a few outliers. 0 observations lie above the upper whisker (106) and 403 observations lie below the lower whisker (90).

As with the heart rate values, All of the pulse oximetry values that we see in these data are within the physiological range. If there were extreme outlier values (the physiological range is [0, 100] %), we would want to correct these. We can recode all values outside the physiological ranges as "NA" in the dataset for all patients.

```
sp <- sp %>%
  mutate(
    VALUENUM = if_else((VALUENUM < 0) | (VALUENUM > 100), NA_real_, VALUENUM)
)
summary(sp$VALUENUM)
#> Min. 1st Qu. Median Mean 3rd Qu. Max. NA's
#> 0.00 96.00 98.00 97.01 100.00 100.00 15
```

While within the physiological range, we observe that there are several values bellow 80%. Some of these values may correspond to other variables, such as respiratory or heart rate that were mistyped in the system. For our analysis we will recoded them as "NA".

```
sp <- sp %>%
 mutate(
    VALUENUM = if_else(VALUENUM < 80, NA_real_, VALUENUM)</pre>
  )
summary(sp$VALUENUM)
      Min. 1st Qu.
                                                       NA's
                   Median
                              Mean 3rd Qu.
                                               Max.
#>
             96.00
                     98.00
                             97.33 100.00 100.00
                                                        131
sp %>%
 ggplot(aes(y = VALUENUM)) +
  stat_boxplot(geom = "errorbar", width = 0.2, na.rm = TRUE) +
  geom_boxplot(na.rm = TRUE) +
  labs(title = "Pulse oximetry distribution for all patients",
       x = "Pulse oximetry",
       y = 'Value [%]')
```



```
uw <- quantile(sp$VALUENUM, 3/4, na.rm = TRUE) + (1.5 * IQR(sp$VALUENUM, na.rm = TRUE))
lw <- quantile(sp$VALUENUM, 1/4, na.rm = TRUE) - (1.5 * IQR(sp$VALUENUM, na.rm = TRUE))
above <- sum(sp$VALUENUM > uw, na.rm = TRUE)
below <- sum(sp$VALUENUM < lw, na.rm = TRUE)</pre>
```

After recoding, 0 observations lie above the upper whisker (106) and 287 observations lie below the lower whisker (90).

Exercise: Plotting missing data

Aim: Analyze and visualize the missing data

Complete cases for heart rate

The function complete.cases returns a logical vector indicating which cases are complete.

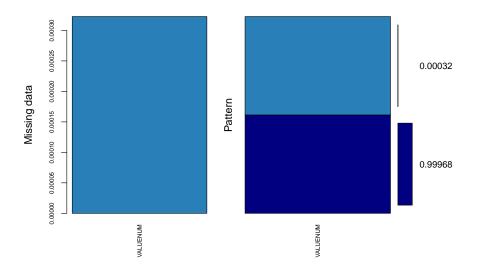
```
hr %>% filter(!complete.cases(SUBJECT ID))
#> # A tibble: 0 x 14
#> # ... with 14 variables: SUBJECT_ID <int>, HADM_ID <int>, ICUSTAY_ID <int>,
#> # ITEMID <int>, CHARTTIME <dttm>, STORETIME <dttm>, CGID <int>, VALUE <chr>,
     VALUENUM <dbl>, VALUEUOM <chr>, WARNING <int>, ERROR <int>,
#> # RESULTSTATUS <chr>, STOPPED <chr>
na_sid <- hr %>% filter(!complete.cases(SUBJECT_ID)) %>% count()
na_sid_pct <- round(100 * na_sid / (hr %>% count()), 2)
hr %>% filter(!complete.cases(VALUEUOM))
#> # A tibble: 0 x 14
#> # ... with 14 variables: SUBJECT_ID <int>, HADM_ID <int>, ICUSTAY_ID <int>,
#> # ITEMID <int>, CHARTTIME <dttm>, STORETIME <dttm>, CGID <int>, VALUE <chr>,
#> # VALUENUM <dbl>, VALUEUOM <chr>, WARNING <int>, ERROR <int>,
#> # RESULTSTATUS <chr>, STOPPED <chr>
na_uom <- hr %>% filter(!complete.cases(VALUEUOM)) %>% count()
na_uom_pct <- round(100 * na_uom / (hr %>% count()), 2)
hr %>% filter(!complete.cases(VALUENUM))
#> # A tibble: 5 x 14
   SUBJECT ID HADM ID ICUSTAY ID ITEMID CHARTTIME
                                                               STORETIME
#>
         \langle int \rangle
                 \langle int \rangle
                            \langle int \rangle \langle int \rangle \langle dttm \rangle
                                                               \langle dttm \rangle
          10069 146672
#> 1
                            290490
                                    211 2188-02-12 18:00:00 2188-02-12 18:36:00
                          246080 211 2132-08-06 10:00:00 2132-08-06 10:23:00
#> 2
         10089 190301
#> 3
         10126 160445
                         249805 211 2171-07-14 12:00:00 2171-07-14 12:28:00
         10126 160445
                            249805 211 2171-07-29 22:00:00 2171-07-29 22:33:00
#> 4
                                    211 2198-07-04 15:45:00 2198-07-04 17:07:00
         10127 182839
#> 5
                            271544
#> # ... with 8 more variables: CGID <int>, VALUE <chr>, VALUENUM <dbl>,
     VALUEUOM <chr>, WARNING <int>, ERROR <int>, RESULTSTATUS <chr>,
#> # STOPPED <chr>
na_val <- hr %>% filter(!complete.cases(VALUENUM)) %>% count()
na_val_pct <- round(100 * na_val / (hr %>% count()), 2)
```

We have 0 observations with missing subjects IDs (0%), 0 observations with missing heart rate units (0%), and we have 5 observations with missing heart rate values (0.03%).

Plotting missing values for heart rate

The aggr function from the VIM package allows one to calculate and/or plot the amount of missing/imputed values in each variable and in certain combinations of variables.

```
na_hr <- hr %>%
select(VALUENUM) %>%
aggr(col = c('navyblue','#2980b9'), numbers = TRUE, sortVars = TRUE,
cex.axis = 0.7, gap = 3, ylab = c("Missing data","Pattern"))
```



- #>
 #> Variables sorted by number of missings:
 #> Variable Count
- #> VALUENUM 0.0003227889

We observe that for the case of heart rate there are 99.97% of the values in the data set or 0.03% missing.

Omit cases for heart rate

The function na.omit returns the input object with listwise deletion of missing values.

```
hr_complete <- na.omit(hr)</pre>
```

And now we have a complete dataset with missing data preprocessed for heart rate measurements.

Now repeat the steps for pulse oximetry.

Complete cases for pulse oximetry

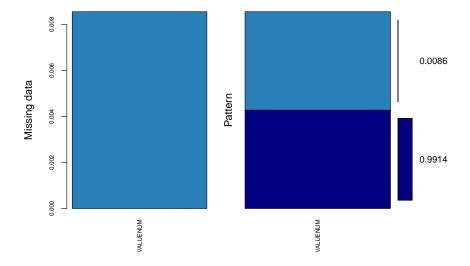
```
sp %>% filter(!complete.cases(SUBJECT_ID))
#> # A tibble: 0 x 14
#> # ... with 14 variables: SUBJECT_ID <int>, HADM_ID <int>, ICUSTAY_ID <int>,
```

```
ITEMID <int>, CHARTTIME <dttm>, STORETIME <dttm>, CGID <int>, VALUE <chr>,
#> #
       VALUENUM <dbl>, VALUEUOM <chr>, WARNING <int>, ERROR <int>,
     RESULTSTATUS <chr>, STOPPED <chr>
na sid <- sp %>% filter(!complete.cases(SUBJECT ID)) %>% count()
na_sid_pct <- round(100 * na_sid / (sp %>% count()), 2)
sp %>% filter(!complete.cases(VALUEUOM))
#> # A tibble: 0 x 14
#> # ... with 14 variables: SUBJECT ID <int>, HADM ID <int>, ICUSTAY ID <int>,
     ITEMID <int>, CHARTTIME <dttm>, STORETIME <dttm>, CGID <int>, VALUE <chr>,
     VALUENUM <dbl>, VALUEUOM <chr>, WARNING <int>, ERROR <int>,
#> # RESULTSTATUS <chr>, STOPPED <chr>
na_uom <- sp %>% filter(!complete.cases(VALUEUOM)) %>% count()
na_uom_pct <- round(100 * na_uom / (sp %>% count()), 2)
sp %>% filter(!complete.cases(VALUENUM))
#> # A tibble: 131 x 14
      SUBJECT_ID HADM_ID ICUSTAY_ID ITEMID CHARTTIME
                                                                STORETIME
#>
#>
           \langle int \rangle
                   \langle int \rangle
                              \langle int \rangle \langle int \rangle \langle dttm \rangle
                                                                \langle dttm \rangle
#> 1
           10013 165520
                             264446 646 2125-10-06 21:30:00 2125-10-06 21:17:00
           10013 165520
#> 2
                             264446 646 2125-10-06 22:00:00 2125-10-06 23:32:00
           10013 165520
#> 3
                             264446
                                       646 2125-10-07 11:15:00 2125-10-07 11:19:00
#> 4
           10013 165520
                             264446
                                       646 2125-10-07 12:10:00 2125-10-07 12:14:00
#> 5
           10019 177759
                             228977
                                       646 2163-05-15 06:30:00 2163-05-15 08:28:00
           10040 157839
#> 6
                                       646 2147-02-23 16:45:00 2147-02-23 16:59:00
                             272047
#> 7
           10040 157839
                             272047
                                       646 2147-02-23 11:50:00 2147-02-23 11:51:00
#> 8
           10040 157839
                             272047
                                       646 2147-02-23 23:30:00 2147-02-23 23:19:00
#> 9
           10045 126949
                             203766
                                       646 2129-11-30 22:35:00 2129-11-30 22:43:00
           10045 126949
#> 10
                             203766
                                       646 2129-11-30 22:40:00 2129-11-30 22:43:00
#> # ... with 121 more rows, and 8 more variables: CGID <int>, VALUE <chr>,
       VALUENUM <dbl>, VALUEUOM <chr>, WARNING <int>, ERROR <int>,
       RESULTSTATUS <chr>, STOPPED <chr>
na_val <- sp %>% filter(!complete.cases(VALUENUM)) %>% count()
na_val_pct <- round(100 * na_val / (sp %>% count()), 2)
```

We have 0 observations with missing subjects IDs (0%), 0 observations with missing pulse oximetry units (0%), and we have 131 observations with missing pulse oximetry values (0.86%).

Plotting missing values for pulse oximetry

```
na_sp <- sp %>%
select(VALUENUM) %>%
aggr(col = c('navyblue','#2980b9'), numbers = TRUE, sortVars = TRUE,
cex.axis = 0.7, gap = 3, ylab = c("Missing data","Pattern"))
```



```
#>
Variables sorted by number of missings:
#> Variable Count
#> VALUENUM 0.008553706
```

We observe that for the case of pulse oximetry there are 99.14% of the values in the data set or 0.86% missing.

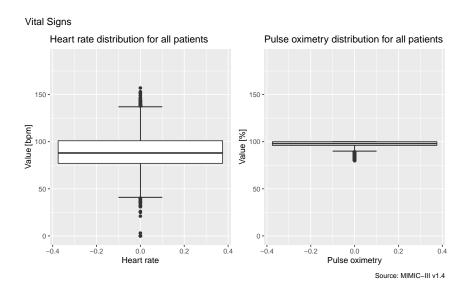
Omit cases for pulse oximetry

```
sp_complete <- na.omit(sp)</pre>
```

And now we have a complete dataset with missing data preprocessed for pulse oximetry measurements.

Attention however, because missing data might not be missing at random (MNAR - see chapter contents for details) and in that case it might be important to identify and handle in a different way the missing values in the dataset. Eg. this can be done through missing data imputation.

We can now see how the distribution looks like for complete and processed data.



Exercise: Missing data imputation

Aim: To impute missing data using several methods.

There are several approaches for missing data imputation. Here we show how to impute missing data with packages available in R Studio.

Before missing data imputation we must first remove outliers, so our imputation is performed based on values within the physiological ranges.

First, select heart rate and pulse oximetry values in the same dataframe.

```
hr_sp <- vitals %>%
filter(
   (((ITEMID == 211) | (ITEMID == 220045)) & (toupper(VALUEUOM) == "BPM")) |
   (((ITEMID == 646) | (ITEMID == 220277)) & (VALUEUOM == "%"))
)
```

And recode as "NA" where values are outside the physiological ranges. We also include a new variable MEASURE which quickly identifies whether the observation is for heart rate or pulse oximetry. (This will be used to pivot the data frame wider - see below.)

Imputation using the population median values

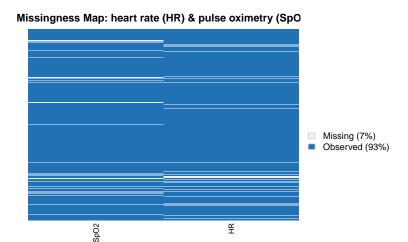
We can directly impute the median of the population for each variable.

```
med_hr_sp <- hr_sp %>%
mutate(
    HR = ifelse(is.na(HR), median(hr_sp$HR, na.rm = TRUE), HR),
    Sp02 = ifelse(is.na(Sp02), median(hr_sp$Sp02, na.rm = TRUE), Sp02)
)
```

Imputation with the Amelia package

Let's visualize and then impute the missing values with functions from the Amelia package - named after Amelia Earhart.

We selected pulse oximetry and heart rate as examples, however this can be performed for all the variables. Select 1 patient ICU stay as example for visualization.



Now, impute missing data using the Amelia package. For the cases (correspondent to rows) where there is no value for heart rate or pulse oximetry, the value is not imputed. With a dataset containing higher amount of variables, we will not have so many of these cases. However, in the cases where we do not have any value we can proceed with eg. case deletion.

The amelia function runs the bootstrap EM algorithm on incomplete data and creates imputed datasets.

```
amelia_fit <- hr_sp %>%
 filter(!(is.na(HR) & is.na(Sp02))) %>%
 select(HR, SpO2) %>%
 as.data.frame() %% # data needs to be in a pure data frame or matrix form
 amelia(parallel = "multicore")
#> -- Imputation 1 --
#>
    1 2 3 4
#>
#>
  -- Imputation 2 --
#>
#>
    1 2 3 4
#>
#>
  -- Imputation 3 --
#>
    1 2 3 4
#>
#>
  -- Imputation 4 --
#>
#>
    1 2 3 4
#>
#> -- Imputation 5 --
    1 2 3 4
```

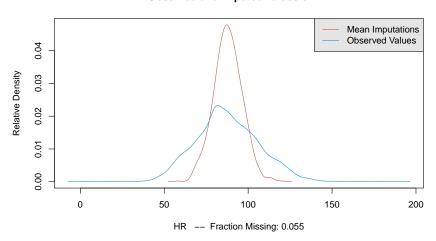
Analyze the imputed values

We will analyze the imputed values created to understand if the imputation method was adequate. A common practice consists in comparing the distribution of the imputed values and of the observed values.

Use the compare.density function from the Amelia package for this.

For heart rate:

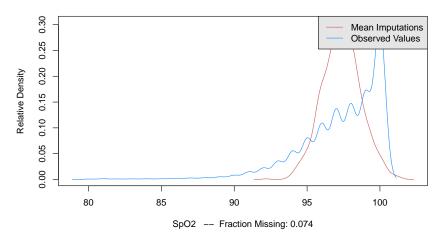
Observed and Imputed values of HR



For pulse oximetry:

compare.density(amelia_fit, var = "Sp02")

Observed and Imputed values of SpO2



We observe that other methods should be applied in order to have a more fitted distribution of the data.