

1 Data Descriptor Title (110 character maximum, inc. 2 spaces)

3 Yuhang Lin^{1,2} and Heike Hofmann^{1,2}

4 ¹Iowa State University, Department of Statistics, Ames,

5 ²Center for Statistics and Applications in Forensic Evidence (CSAFE), Iowa State University, Ames,

6 *corresponding author(s): Yuhang Lin (yhlin@iastate.edu) ????????it.corresponding returns nothing

7 *corresponding author(s): Heike Hofmann (hofmann@iastate.edu) ????????it.corresponding returns nothing

8 ABSTRACT

9 (maximum 170 words) This is a manuscript template for Data Descriptor submissions to *Scientific Data* (<http://www.nature.com/scientificdata>). The abstract must be no longer than 170 words, and should succinctly describe the study, the assay(s) performed, the resulting data, and the reuse potential, but should not make any claims regarding new scientific findings. No references are allowed in this section.

10 Please note: Abbreviations should be introduced at the first mention in the main text no abbreviations lists or tables should be
11 included. Structure of the main text is provided below.

12 Background & Summary

13 (unlimited length) An overview of the study design, the assay(s) performed, and the created data, including any background
14 information needed to put this study in the context of previous work and the literature. The section should also briefly outline
15 the broader goals that motivated the creation of this dataset and the potential reuse value. We also encourage authors to include
16 a figure that provides a schematic overview of the study and assay(s) design. The Background & Summary should not include
17 subheadings. This section and the other main body sections of the manuscript should include citations to the literature as
18 needed.

19 Methods

- 20 • Materials used in the study: Aluminum wire (16 Gauge/1.5 mm, anodized).
- 21 • Why aluminum not lead as in the case: to create a best-case scenario where the most amount of information could be
22 transferred from the tool to the substrate

23 Cutting Aluminium Wires

- 24 1. Unspool a piece of aluminum wire approximately 4" long, then use a neutral cutting edge to cut the piece off the spool.
25 Repeat 6 times per wirecutters needing to be tested, in our case, 5 wirecutters, resulting in 30 pieces of wire.
26 <!-- -->
- 27 2. Take the newly cut piece of wire and cut directly in the middle of the piece, ideally making two 2" pieces of wire. It
28 is important to track which end of the wire is the AB cut and which is the CD cut. Therefore, a sharpie line across the
29 wire was made to indicate which end the cut would be on before cutting.
30 <!-- --> <!-- --> <!-- -->
- 33 3. After cutting, take a small Ziploc bag, and label it with Test #, Cut #, Tool #, Edge AB/CD, and Date. Then package the
34 pieces of wire. <!-- -->
- 35 4. Repeat Step 2-3 for each piece of 4" long wire that has been cut.

Scanning Aluminium Wires

1. Remove a packaged piece of wire from the Ziploc bag.
2. Secure the piece of wire on the confocal microscope stage using sticky tac. Ensure the wire is stable before scanning.
<!-- -->
3. Under the 5x optic, begin by leveling the large edge of the cut. Turning the dial to the right or clockwise can reach the lowest point on the wire, while turning it to the left or counter-clockwise will lead to the highest point. The wire can be shifted left or right based on the perceived high or low point. Additionally, tilting the stage forward and backward can help level the wire. The levelness can be determined by observing how much of the cut is visible. Ideally, all pieces of the wire should be visible without any blurry spots. However, it is common to encounter variations in height throughout the scan, where certain areas consistently appear higher or lower than others.
4. Take the overall photo of the cut under the 5x optic.
5. Switch to the 20x optic and adjust the number of cells needed for scanning accordingly. Typically, four cells are sufficient to cover most scans. However, the number of cells needed may vary depending on the specific side of the scanned cut. <!-- -->
<!-- -->
6. Rotate the dial to the right or clockwise to locate the lowest point of the cut when using the 20x optic. Continue rotating slightly past that point, which may cause details in the scan to appear slightly fuzzy. For accuracy, all cells should be checked to confirm that the lowest point of the scan has been reached. It may also be necessary to rotate the dial to the left initially. This adjustment compensates for the possibility of the focus being set beyond the low point when switching to the 20x optic. <!-- -->
7. Plant the flag for the scan, indicating to the microscope that the scan will start from this specific location. <!-- -->
8. Rotate the dial to the left or counter-clockwise to locate the highest point of the cut. Continue rotating slightly past that point, which may cause details in the scan to appear slightly fuzzy. A closer Z-value to 0 in the left column indicates a better result. Typically, if this value exceeds 0.100, it indicates that the cut needs to be leveled further.
9. After identifying and slightly passing the highest point, record the first three digits after the decimal point without leading 0s of the Z-value number into the Z-score box located in the right column of the interface. For example, if the Z-value displays '0.0581', '58' should be input into the Z-score box. Adjusting the Z-score communicates to the microscope the desired scanning distance. <!-- -->
10. Turn the dial to the right or clockwise to reset the Z-value to 0. <!-- -->
11. Adjust the light as necessary with the help of the auto light feature represented by a sun icon on the interface. <!-- -->
12. Click the green 'ACQUIRE' button. <!-- -->
13. Save the scan with the naming protocol: Wire Cutting Project - Scan # - Cut # - Aluminium Wire - (Large/Small) Edge - (Inner/Middle/Outer) Surface - Kaiweets Tool #(1/2/3/4/5) - Edge (A/B/C/D) - 20x - Light
14. Repeat Step 1-13 for all the pieces of wire.

Summary of Data Collection

- 60 = 5 \times 2 \times 2 \times 3 samples
- 5 Kaiweets wire cutters - model KWS-105 (tools)
- 2 wire cuts (replicates) for 2 sides of wire cutter (AB or CD)

- 3 locations along the blade (inner, middle, outer)
- 120 = 60 \times 2 samples
- 2 scans separating sides into cuts by each blade
- Scans are saved at a resolution of 0.645 \times 0.645 per square pixel in an x3p file format

-
- any steps or procedures used in producing the data, experimental design: README for [heike/Wirecuts](#)
 - Transparent Methods Checklist

- Materials & reagents: Identify commercial suppliers of reagents, instrumentation or kits, when the source is critical to the outcome of the experiments. Declare any restrictions on the availability of unique materials (more information here). Provide catalogue or clone numbers for all antibodies (if available). For primary antibodies, provide proof of validation for the relevant species and applications.
- Exclusion criteria: If any data or samples were excluded, explain the exclusion criteria and state in the methods whether the criteria were established before the study was conducted.
- Randomization & blinding: For any studies that involve assigning samples, animals or participants into different groups: State clearly whether randomization methods were used. If randomization was not employed, this should be clearly stated. State clearly whether blinding was employed during data collection. If blinding was not employed, this should be clearly stated.
- Animal & human studies (full journal policies here): Experiments involving human participants must identify the committee approving the experiments, and include a statement confirming that informed consent was obtained from all participants. Studies employing nonhuman animals should ensure that methods descriptions comply with the ARRIVE checklist.
- Cell lines: For each eukaryotic cell line used, state the source and whether the cell line has been authenticated or otherwise tested for integrity. If any commonly misidentified cell lines were used (see ICLAC or NCBI Biosample), justify their use. Report whether the cell lines were tested for mycoplasma contamination.
- Chemistry & materials science: Manuscripts describing chemical syntheses, or characterizing new chemicals or materials should refer to the guidance at Nature Chemistry.

(unlimited length) The Methods should include detailed text describing any steps or procedures used in producing the data, including full descriptions of the experimental design, data acquisition assays, and any computational processing (e.g. normalization, image feature extraction). See the detailed section in our submission guidelines for advice on writing a transparent and reproducible methods section. Related methods should be grouped under corresponding subheadings where possible, and methods should be described in enough detail to allow other researchers to interpret and repeat, if required, the full study. Specific data outputs should be explicitly referenced via data citation (see Data Records and Citing Data, below).

Authors should cite previous descriptions of the methods under use, but ideally the method descriptions should be complete enough for others to understand and reproduce the methods and processing steps without referring to associated publications. There is no limit to the length of the Methods section. Subheadings should not be numbered.

Authors should review the transparent methods checklist below, and ensure that their manuscript complies with any relevant points. Authors are also encouraged to search FAIRsharing.org for community reporting standards that may be relevant to their specific data-type.

Data Records

- [ISU DataShare](#)

(unlimited length) The Data Records section should be used to explain each data record associated with this work, including the repository where this information is stored, and to provide an overview of the data files and their formats. Each external data record should be cited numerically in the text of this section, for example ?, and included in the main reference list as described below. A data citation should also be placed in the subsection of the Methods containing the data-collection

or analytical procedure(s) used to derive the corresponding record. Providing a direct link to the dataset may also be helpful to readers (<https://doi.org/10.6084/m9.figshare.853801>).

Tables should be used to support the data records, and should clearly indicate the samples and subjects (study inputs), their provenance, and the experimental manipulations performed on each (please see 'Tables' below). They should also specify the data output resulting from each data-collection or analytical step, should these form part of the archived record.

Technical Validation

- Measurement of data quality?

- Numeric measurements / tests: ?

- Visualizations: ?

- Check with existing data: ?

- Questionable / slur procedures:

- * **AidDats Geospatial Global Chinese Development Finance Dataset:** Second, all data collected is reviewed by at least two individuals. Although this is not a double-blind review procedure, the use of satellite imagery to verify project locations results in far less uncertainty when compared to previous approaches to geocoding where locations were selected entirely based on text descriptions.

- * **A large open access dataset of brain metastasis 3D segmentations on MRI with clinical and imaging information:** A medical student (D.R.) double checked and adjusted the revised NIFTI segmentation masks and manually counted the number of lesions with contrast-enhancement, necrosis, and peritumoral edema for each patient.

- * **Time series of freshwater macroinvertebrate abundances and site characteristics of European streams and rivers:** Technical validation of the TREAM dataset was achieved through exclusion of time series data that did not match our inclusion criteria and data standardisation steps (outlined in Methods above). Any noted issues that did not adhere to the outlined standardisation within the datasets from the 41 independent projects included in this dataset were checked with data providers and corrected or removed when standardisation was not achievable (e.g., when collection methods changed over the course of the time series).

- * **3D surgical instrument collection for computer vision and extended reality:** The main issue...Since we store our models in a standard format (STL), they are compatible with a large variety of visualisation and processing software.

- * **Three-dimensional reconstruction of high latitude bamboo coral via X-ray microfocus Computed Tomography :** Regular quality assurance inspections are carried out on the µ-CT scanner to verify its metrological and geometrical (alignments) accuracy for conducting the scans. The geometry of source to object and source to detector distances are verified whenever there is any significant physical interaction with the source such as re-alignment, change of filament, or source anode change. This calibration process involves scanning a specially designed phantom known as an hourglass³⁶, which consists of three pairs of high-sphericity spheres. The sphere sizes are as follows: two spheres with a diameter of 3.000 mm, two spheres with a diameter of 6.000 mm, and two spheres with a diameter of 9.525 mm, and each sphere is kept in contact with its size-counterpart. By using this phantom, it becomes possible to accurately determine a known distance, specifically the centre-to-centre distance of the spheres, in a threshold-independent manner. If the measured distance deviates beyond the acceptable limits of metrological accuracy, the systems calibration parameters are adjusted to ensure agreement between the measured distance and the actual distance.

(unlimited length) This section presents any experiments or analyses that are needed to support the technical quality of the dataset. This section may be supported by figures and tables, as needed. This is a required section; authors must present information justifying the reliability of their data.

Usage Notes

- software packages that are suitable for analysing the assay data files:
 - CRAN R package: `x3ptools`
 - Github R package: `wire`
 - GitHub R shiny app: `wireShiny`
- suggested downstream processing steps: scaling resolution by `x3p_scale_unit`?
- tips for integrating or comparing the data records with other datasets: comparing with sheet datasets (if in another data descriptor)?
- data-processing workflows: automatic / manual pipeline?

(unlimited length) The Usage Notes should contain brief instructions to assist other researchers with reuse of the data. This may include discussion of software packages that are suitable for analysing the assay data files, suggested downstream processing steps (e.g. normalization, etc.), or tips for integrating or comparing the data records with other datasets. Authors are encouraged to provide code, programs or data-processing workflows if they may help others understand or use the data. Please see our code availability policy for advice on supplying custom code alongside Data Descriptor manuscripts.

For studies involving privacy or safety controls on public access to the data, this section should describe in detail these controls, including how authors can apply to access the data, what criteria will be used to determine who may access the data, and any limitations on data use.

Code availability

For all studies using custom code in the generation or processing of datasets, a statement must be included under the heading "Code availability", indicating whether and how the code can be accessed, including any restrictions to access. This section should also include information on the versions of any software used, if relevant, and any specific variables or parameters used to generate, test, or process the current dataset.

1 End of Body

Note that the bibliography style and the name of the bib-file are hard coded in the template file right now.

LaTeX formats citations and references automatically using the bibliography records in your .bib file, which you can edit via the project menu. Use the cite command for an inline citation, e.g. `????`. For data citations of datasets uploaded to e.g. *figshare*, please use the `howpublished` option in the bib entry to specify the platform and the link, as in the `Hao:gidmaps:2014` example in the sample bibliography file. For journal articles, DOIs should be included for works in press that do not yet have volume or page numbers. For other journal articles, DOIs should be included uniformly for all articles or not at all. We recommend that you encode all DOIs in your bibtex database as full URLs, e.g. <https://doi.org/10.1007/s12110-009-9068-2>.

Acknowledgements

Acknowledgements should be brief, and should not include thanks to anonymous referees and editors, or effusive comments. Grant or contribution numbers may be acknowledged.

Author contributions statement

Y.L. did all of the work, H.H. made him do the work. But seriously, this paper is the one where we need to cite everybody: Eden Amin, Curtis Mosher, Jeff Salyards. Must include all authors, identified by initials, for example: A.A. conceived the experiment(s), A.A. and B.A. conducted the experiment(s), C.A. and D.A. analysed the results. All authors reviewed the manuscript.

205 **Competing interests**

206 (mandatory statement)

207 The corresponding author is responsible for providing a [competing interests statement](#) on behalf of all authors of the paper.
208 This statement must be included in the submitted article file. H.H. is a technical advisor to AFTE (Association of Firearms and
209 Toolmarks Examiners), fellow of the ASA (American Statistical Association), and committee member of the ASA Forensic
210 Science Committee. H.H. has testified as court witness on behalf of judge April Neubauer, NY State Supreme Court Criminal
211 Term in New York City.

212 **Figures & Tables**

213 Figures, tables, and their legends, should be included at the end of the document. Figures and tables can be referenced in
214 L^AT_EX using the ref command, e.g. Figure 1 and Table 1.

215 Authors are encouraged to provide one or more tables that provide basic information on the main inputs to the study
216 (e.g. samples, participants, or information sources) and the main data outputs of the study. Tables in the manuscript should
217 generally not be used to present primary data (i.e. measurements). Tables containing primary data should be submitted to an
218 appropriate data repository.

219 Tables may be provided within the L^AT_EX document or as separate files (tab-delimited text or Excel files). Legends, where
220 needed, should be included here. Generally, a Data Descriptor should have fewer than ten Tables, but more may be allowed
221 when needed. Tables may be of any size, but only Tables which fit onto a single printed page will be included in the PDF
222 version of the article (up to a maximum of three).

223 Due to typesetting constraints, tables that do not fit onto a single A4 page cannot be included in the PDF version of the
224 article and will be made available in the online version only. Any such tables must be labelled in the text as Online-only tables
225 and numbered separately from the main table list e.g. Table 1, Table 2, Online-only Table 1 etc.



Figure 1. Legend (350 words max). Example legend text.

Condition	n	p
A	5	0.1
B	10	0.01

Table 1. Legend (350 words max). Example legend text.