Evaluation of body composition: Why and how?

Article in Mediterranean Journal of Nutrition and Metabolism · April 2009		
DOI: 10.1007/s12349-009-0042-x - Source: OAI		
CITATIONS		READS
20		272
1 author	:	
0	Hank Lukaski	
	University of North Dakota	
	7 PUBLICATIONS 152 CITATIONS	
	SEE PROFILE	
Some of the authors of this publication are also working on these related projects:		
	la la característica de la	
Project	Muscle assessment by L-BIA measurements View project	

EDITORIAL

Evaluation of body composition: why and how?^{1,2}

H.C. Lukaski

Received: 19 December 2008 / Accepted: 3 February 2009 / Published online: 1 April 2009 © Springer-Verlag 2009

Abstract Evaluation of human body composition *in vivo* remains a critical component in the assessment of nutritional status of an individual. Whereas traditional measurements of standing height and body weight provide information on body mass index and, hence, the risk of some chronic diseases, advanced technologies, such as dual X-ray absorptiometry, air displacement plethysmography and various forms of bioelectrical impedance analysis enable the determination of soft tissue composition (fat and lean) as well as bone. This review summaris-

es the physical bases of these methods and critically evaluates their accuracy in observational and interventional studies. It also discusses a new approach, bioelectrical impedance vector analysis, which assesses the hydration status of an individual, and includes pertinent examples of its novel applications in clinical nutrition.

Keywords Dual X-ray absorptiometry · Body volume · Bioelectrical impedance · Bioelectrical impedance vector analysis · Hydration status

Introduction

Evaluation of human body composition in vivo requires the objective determination of various levels of the chemical composition of the body [1]. This challenge involves the precise and accurate measurement of the soft tissue composition (fat and fat-free or lean) and the bone mineral (mass and quality) of an individual. These measurements may be used to identify differences between groups (static appraisals) or to determine the responses to intervention or treatment (dynamic assessments) for an individual. The overall goals of body composition assessment in vivo for dieticians and clinical nutritionists are to quantitate body energy stores (fat or adipose tissue), structure (bone) and/or functional capacity (muscle mass or body cell mass). More than a century of research has produced diverse methods that utilise specific assumptions and models to accomplish these goals [2, 3].

This presentation concisely reviews and critically evaluates the use of some contemporary methods of evaluating human body composition *in vivo*. It emphasises dual X-ray absorptiometry (DXA), whole-body and segmental bioelectrical impedance techniques, and high-

²US Department of Agriculture, Agricultural Research, Northern Plains Area is an equal opportunity/affirmative action employer and all agency services are available without discrimination.

H.C. Lukaski (☒)
U.S. Department of Agriculture
Agricultural Research Service
Grand Forks Human Nutrition Research Center
2420 2nd Avenue North STOP 9034
Grand Forks, ND 58202-9034, USA
e-mail: henry.lukaski@ars.usda.gov

¹Mention of a trademark or proprietary product does not constitute a guarantee of the product by the United States Department of Agriculture and does not imply its approval to the exclusion of other products that may also be suitable.

lights the novel application of bioelectrical impedance vector analysis (BIVA) to assess the hydration status of individuals in a variety of clinical conditions characterised by disturbances in water balance.

Dual X-ray absorptiometry (DXA)

This radiographic method utilises the graded attenuation of a beam of X-rays by bone, fat-free and bone-free tissue (lean soft tissue), and fat to estimate bone mineral and soft tissue composition. The beam of X-rays is pulsed at two discrete energy levels and administered in a posterior-anterior direction; the attenuation of the Xrays is measured with highly sensitive detectors positioned at a fixed distance from the scan bed and above the anterior surface of the body [4]. DXA overcomes the limitations of traditional methods such as underwater weighing, anthropometry and isotope dilution to estimate body composition because it is independent of assumptions of the constant chemical composition of the fat-free body, which is inherent in the two-component model of body composition [5]. Other beneficial characteristics are the rapidity of the measurement (5–15 min) and the capability to estimate regional and whole-body composition. Concurrent with the determination of soft tissue composition, DXA assesses bone mineral content and areal density. A practical limitation of DXA is exposure of the subject to ionising radiation (<5 μSv), which is low relative to daily radiation exposure (7 µSv) [6].

Figure 1 highlights the value of DXA in assessment of human body composition and shows the compositional variations between two adults with the same body mass index (BMI). Clear differences in the fat-free and bonefree mass (54 vs. 49.8 kg), fat mass (13 vs. 17.6 kg), fatness (19 vs. 25%) and bone mineral content (2696 vs. 2504 g) are evident in these images. This observation is consistent with previous findings that BMI is a non-specific indicator of body composition in healthy adults [7–9].

Alterations in body hydration confound DXA determinations of soft tissue composition. Studies of volunteers before and after ingestion of a standardised volume of water [10], voluntary dehydration [11] or renal dialysis [12] reveal that DXA identifies the increase or decrease in fluid volume as a change in body mass and lean soft tissue mass. Neither fat mass nor bone mineral content is significantly affected by alterations in fluid status up to 3% body weight. Thus, small but systematic and predictable errors in DXA estimates of soft tissue composition can arise with modest changes in hydration.

Body thickness is another factor moderating the validity of DXA determinations of soft tissue composition and bone. The accuracy of DXA determinations of body composition, compared with direct chemical analysis of animal carcasses, decreases significantly when body regions exceed 23 cm in anterior–posterior thickness [13].

The development of DXA for body composition assessment, particularly in multicentre trials, has evolved from pencil-beam to fan-beam instruments. Advantages of the fan-beam devices include a decrease in scan time and improved image resolution. An early validation trial of a fan-beam DXA with magnetic resonance imaging (MRI) found that DXA overestimated fat-free or lean soft tissue mass and underestimated fat mass [14]. A later investigation confirmed these findings and explained the discrepancy based on differences in the chemical and anatomical components measured with each technology

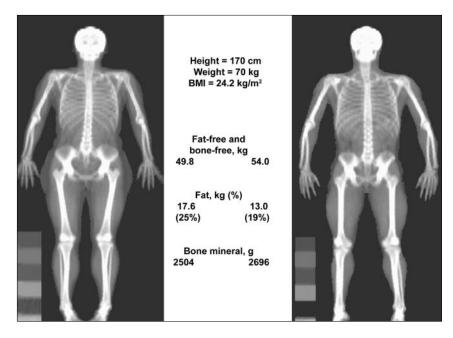


Fig. 1 Differences in the soft tissue composition and the bone mineral content of two adults with the same BMI



[15]. Lean soft tissue composition estimated with DXA includes not only skeletal muscle but also skin, connective tissues, protein matrix of adipose tissue and organs, whereas MRI only determines operator-delineated muscle mass and organs. Similarly, DXA-derived fat mass was less than the corresponding MRI-determined fat because DXA assesses chemical fat and MRI measures operator-defined adipose tissue including protein, mineral and water content.

Comparisons of DXA determinations of soft tissue composition with standard reference methods yield significant correlation coefficients ($r \ge 0.90$) but some discrepancies [16]. Pencil-beam and fan-beam devices produce modest differences in their estimates of body fatness (<4%) [17–19]. Also, differences in software programs to analyse DXA images produce divergent results (~2%) [20]. Thus, it is advised to use the same DXA instrument and analytical software version for any investigation, particularly longitudinal, multicentre interventional studies, and to include an appropriate anthropometric phantom (soft tissue or bone) for quality control and assurance [21].

Air displacement plethysmography (ADP)

Determination of body density, which is inversely related to body fatness, is an established measure in the assessment of human body composition [5, 22]. Although underwater or hydrostatic weighing is the traditional method for measuring body volume and calculating body density (density = weight/volume), this technique and its procedures can be problematic to perform for special populations including children, the elderly and the disabled. A safe and non-invasive alternative is ADP, whose popularity is growing. Body volume and density determined with ADP use a commercial device (Bod Pod, Life Measurement System Inc., Concord, CA) [23]. This technique requires subjects to sit in an enclosed test chamber, wear minimal clothing (e.g., tight-fitting swim suit), a swim cap, have minimal body hair, dry skin and hair, and perform specific breathing manoeuvres to estimate thoracic lung volume, which is used to calculate total body volume [24].

The reproducibility of ADP determinations of body fatness is very high. Mean within-day variability is 1.7–3.7% with one operator [25–28] but increases to ~5% with multiple operators [28]. Average between-day reproducibility over 3–7-day periods is 2–2.3% [28–30]. Thus, the variability of ADP determinations of body fatness exceeds the values (1–2%) reported for underwater weighing [31] and DXA [16].

Reports of the validity of ADP to assess body fatness in healthy subjects are not in agreement. Compared to underwater weighing, ADP either significantly overestimates [32–35] or underestimates [36–39] body fatness by 1–4% in adults. However, other reports indicate no difference between these methods [25, 27, 29, 30, 40]. A limited number of reports find that ADP either significantly overestimates [41] or underestimates [42] the body fatness of children and adolescents whereas other studies show no difference [30, 38].

Validation trials of ADP compared to DXA also describe inconsistent results. More reports find that ADP significantly underestimates [26, 29, 37, 39] than overestimates [33, 43, 44] body fatness in adults, with a growing number of reports indicating no significant difference between the methods [28, 30, 40, 45]. Among children, however, ADP, compared to DXA, significantly underestimates body fatness [41, 42].

Some factors contribute to the lack of consensus of the validity of ADP to estimate body fatness. They include the reliance on the two-component model of body composition (e.g., hydrodensitometry) that is prone to error associated with the between-subject variation in the chemical composition of the fat-free body [5, 22], and the use of different X-ray technologies and software analysis programs [16]. In the few studies that have utilised the four-component model to account for the between-subject differences in hydration and bone mineral density, the errors in estimation of body fatness are 2–3% [46].

Wearing excess clothing during the body volume determination can affect the validity of ADP to determine body density. Among lean adults, excess clothing (e.g., shorts, tshirts, scrubs), as compared to swim suits, worn during the ADP results in a significant underestimation of body volume because the air in contact with the cloth remains isothermal as the pressure changes in the chamber. The more cloth worn, the larger the layer of isothermal air volume between the cloth and body surface. Because isothermal air is very compressible, body volume is underestimated, resulting in an overestimation of body density and decreased body fatness (2–5%) [32]. However, additional clothing (e.g., standardised cloth pants and short-sleeved tops) did not affect ADP estimates of body volume and composition in obese adults because the clothing fit tightly to body surfaces and minimised the isothermal layer as compared to lean individuals on whom the clothing fit loosely [44]. This finding has important practical implications for individuals with negative body image, such as overweight and obese adults and children.

Bioelectrical impedance analysis

Ionic solutions in the intracellular and extracellular spaces oppose the conduction of a low-level, radiofre-



quency, alternating current introduced into the body. This opposition, termed electrical impedance (Z), consists of resistance (R) and reactance (Xc) [47]. Investigators use four surface electrodes placed on the arm (wrist) and leg (ankle) with the subject supine and administer a lowlevel, alternating current at either single (50 kHz) or multiple frequencies (e.g., 1, 5, 50, 200, 500 and 1000 kHz), and then measure the whole-body Z, R and Xc if a phasesensitive device is used. These variables are normalised for standing height (e.g., H²/R) to control for differences in conductor length, and then combined with various physical and demographic variables (e.g., body weight, age, gender, etc) into regression models to predict total body water (TBW) or fat-free mass, assuming a constant hydration level (~73%) [48]. The consensus of evidence is that this 50-kHz, whole-body technique, as compared to reference methods, can lead to inaccurate predictions of body composition variables in some circumstances. Concerns include the use of sample-specific prediction models derived in groups of healthy individuals with relatively homogeneous composition can result in errors and wide ranges of estimates (e.g., large confidence intervals of prediction) when applied to individuals with different characteristics (e.g., body size, sex, ethnicity obesity, and hydration status) compared to the group in which the prediction model was developed [49, 50]. This single-frequency approach, however, yields useful body composition estimates that describe groups in epidemiological surveys [51, 52].

Another four-electrode method uses *a priori* selected pairs or ratios of impedance values derived at multiple frequencies in the prediction of fluid distribution [e.g., TBW, extra- and intracellular water (ECW and ICW)] and body composition of individuals with chronic diseases [53–55]. This approach assumes that low and high frequencies are needed to estimate ECW and TBW, respectively [56]. Overall, the use of multiple frequencies does not improve estimates of body fluid volumes as compared to the results obtained with single-frequency (50 kHz) impedance [57–61].

Another development is the use of mathematical modelling of multiple-frequency impedance data and mixture models of single cells in solution to develop equations to predict TBW and ECW [62]. This method uses empirically derived constants for intra- and extracellular resistivity for which there is wide discrepancy [63, 64]. This problem contributes to inconsistent findings in the application of bioelectrical impedance spectroscopy (BIS) in various clinical conditions (e.g., obesity, renal dialysis, pulmonary disease and muscle wasting). Some reports find that BIS improves accuracy [65, 66], has no improvement [67–69] or increases error [70, 71] as compared to other impedance approaches and tracer dilution

reference methods. A recurrent pattern of BIS errors in the estimation of TBW and ECW water in samples of adolescents and adults [72, 73] has prompted a revision of the constants used in the resistivity calculation models. One successful attempt was to include BMI in the prediction models [74] to account for differences in body shape, which *per se* affects Z values independently of fluid distributions. Additional studies are needed to evaluate the validity of the inclusion of BMI in prediction of fluid distribution and body composition in healthy individuals and patients with altered fluid status and body composition.

Four-electrode, segmental bioelectrical impedance devices

A proposed, practical alternative to the whole-body, fourelectrode method is segmental, four-electrode bioimpedance with contact (tactile) electrodes that measure impedance in the upper (arm-to-arm) or the lower (leg-to-leg) body of a standing subject [75–78]. The attraction of this segmental impedance technique is practicality and convenience for routine, personal monitoring of body composition outside of the laboratory because it eliminates the need for an operator of the device, the placement of adhesive electrodes on the wrist and ankle, and the wait for fluid equilibration with the subject in a supine position. The fundamental assumption of this technique is that conductor volume (e.g., ionic fluids and lean soft tissue) is equally distributed in the upper and lower body, and that segmental impedance is proportional to whole-body impedance [79, 80]. Although segmental and whole-body Z values are highly correlated, segmental impedance values are consistently different to whole-body determinations in adults [75, 76, 81] and children [82, 83].

Comparisons of body fatness values determined with segmental, four-electrode devices and DXA yield inconsistent results. A few reports show good correspondence of body fatness predictions with a foot-to-foot impedance device [75, 76] whereas another report [81] using a similar segmental device finds a significant underestimation of body fatness in men (1.7%) and women (2.6%). A hand-to-hand impedance instrument also significantly underestimates body fatness in men (2.3%) and women (6.3%) [81]. Interestingly, a regression model developed in an independent sample and containing only weight, stature, age and sex provides similar estimates of body fatness as DXA, but significantly overestimates body fatness compared to the segmental impedance devices [81]. A concern of these segmental devices is the validity of the proprietary prediction models and the importance of the measured impedance in the estimation of body fatness.



Eight-electrode, segmental bioelectrical impedance

This approach uses eight electrodes, four of which are embedded in the handles (thumb and palm) and another four in the foot scale pads (ball of foot and heel) of the device. This arrangement permits measurement of the impedance of the upper, lower and whole body at either single or multiple frequencies while the subject stands.

Validation trials of single-frequency (50 kHz), eight-electrode bioelectrical impedance devices find consistent errors. Studies of children and adults report a 2.6% over-estimation of body fatness [84], 2.6% in lean young adults [85], 3.5% in normal-weight and 5% in overweight adults [86], obese women [87] and adults [88]. These findings suggest a trend of larger errors in estimation of body fatness with increasing obesity. Similarly, other validation trials using eight-electrode impedance instruments to estimate TBW and ECW report significant errors of 2–3 l in adults [89–91].

Other eight-electrode, segmental instruments introduce an alternating current (25 μ A) at different frequencies (5, 50 and 250 kHz or 1, 5, 50, 250, 500 and 1000 kHz) into the body. The hypothetical advantage of this multiple frequency technology is to improve the estimation of fat-free mass, and hence body fatness, by determining ECW and ICW. A comparison of body fatness derived with these instruments finds similar values as determined by using the four-component model in men but a significant underestimation (~4%) in women [92]. Other studies report significant underestimation of body fatness (5%–6%), compared to DXA, in adults with a wide range of BMI levels [88, 93]. These findings suggest no improvement in the estimation of body fatness for individuals with eight-electrode, multiple-frequency, segmental impedance devices.

Bioelectrical impedance vector analysis (BIVA)

Altered hydration status or fluid imbalance is present in many physiological and disease states. Assessment of fluid components (e.g., TBW and ECW) requires tracer dilution methods that are impractical because of the need for trained personnel and costly equipment, the inability to provide real-time determinations and the lack of appropriate reference standards for an individual. Also, the use of prediction equations based on weight, height, age and sex, and bioelectrical impedance variables yields poor accuracy (>10%) for an individual assessment [52]. Thus, measurements of fluid volumes are very limited in clinical practice, and indicate the need to assess hydration as a practical alternative.

Whole-body Z, expressed as a vector, is a combination of R (opposition to the flow of an alternating current through intra- and extracellular ionic solutions) and Xc (capacitance produced by tissue interfaces and cell membranes) across tissues. The arc tangent of Xc/R is termed the phase angle (phase difference between voltage and current and determined principally by Xc). Interpretation of these variables suggests that the length of the Z vector is inversely related to fluid volume whereas the phase angle offers insight into the relative distribution of fluids (Fig. 2). Normalisation of Xc and R with the standing height (H) of an individual enables standardisation of these variables for differences in conductor length. This approach, which is termed bioelectrical impedance vector analysis (BIVA), allows for the evaluation of hydration and body composition without the need for prediction models [94].

Applications of BIVA focus on description of vector components in health and disease. Characterisation of a group (e.g., healthy or fluid overloaded) uses the mean group vector that is plotted on the R-Xc mean graph (Fig. 3) and the 95% percentile confidence interval, which is

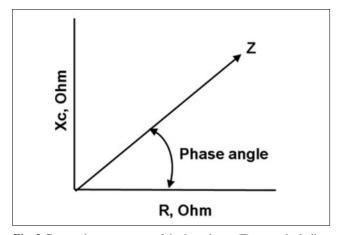


Fig. 2 Geometric components of the impedance (Z) vector including reactance (Xc), resistance (R) and the phase angle

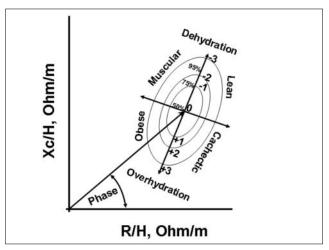


Fig. 3 Conceptual model of bioelectrical impedance vector patterns for clinical use (courtesy of A. Piccoli, University of Padua)



shown as an ellipse. This approach enables a comparison of vectors and tolerance ellipses of groups with different characteristics (e.g., sex, BMI, age, ethnic background, etc.) [95] and changes in these patterns in response to various treatments [96].

Clinical applications of BIVA, however, require a comparison of the affected group or individual vectors with a healthy reference population matched for sex. The mean reference vector is plotted with intervals or prediction ellipses that include 50, 75 and 95% bivariate percentiles [97]. Figure 3 illustrates the use of a general model of BIVA in assessing alterations in fluid status and body composition with the reference intervals derived in a sample of healthy men. For interpretation, the major axis indexes hydration status and the minor axis reflects tissue mass.

The focus of the vast majority of clinical applications of BIVA is characterisation of hydration status in different patient populations. Studies of patients with renal disease, compared to controls, show significantly shorter vectors with lower (smaller) phase angles that lengthen after dialysis [96]. Vectors of unstable (e.g., adverse outcomes), compared to stable, haemodialysis patients are longer with smaller phase angles; these differences between the patient groups persist after haemodialysis. Similar findings occur in peritoneal dialysis patients after fluid removal [98]. A fundamental outcome of these studies is the delineation of the 75% tolerance ellipse as the indicator of the boundary of normal tissue hydration; vectors outside the upper pole of the 75% ellipse indicate dehydration, whereas other vectors outside the 75% ellipse of the lower pole are characteristic of fluid overload or overhydration (Fig. 3).

Bioelectrical impedance vectors can also reflect impaired physiological function. In patients in an intensive care unit, central venous pressure (CVP) was inversely correlated with impedance measurements [99]. Regardless of sex, the vector of patients with low CVP was within the normal hydration range (<75% tolerance ellipse) and vectors from patients with high pressure were outside the 95% reference tolerance ellipse. Vectors from patients with medium CVP were distributed across the lower pole of the 75% ellipse. Thus, progressively increased CVP values are associated with shorter and down-sloping impedance vectors on the R-Xc graph. In contrast, dehydration induced with acute exposure to high altitude results in a significant lengthening of individual impedance vectors from the 50% ellipse at sealevel to the upper pole of the 75% percentile of the reference ellipse [100].

Regional measurements of impedance vectors are more sensitive than whole-body measurements in assessing localised fluid imbalance. Among patients with oedema subsequent to vascular surgery, vectors on the affected leg shortened significantly from pre-surgical condition with no change in the control or non-operated leg [101]. Although a similar response was found on the whole-body side with oedema compared to the control side, the magnitude was smaller. Similarly, BIVA determinations on the abdomen, compared to the limbs of patients undergoing peritoneal dialysis, were significantly more responsive to fluid loss [102].

There is increasing use of BIVA to monitor hydration changes during biological maturation. BIVA discriminates sexual maturity in females with a shorter impedance vector and increased phase angle in postmenarcheal compared to premenarcheal girls [103] and pre-pubescent children compared to adults [104]. In contrast, the vectors of older compared to younger elders were found in the lower right region of the 75% reference ellipse that is indicative of loss of body cell mass [105, 106].

A unique application of BIVA is the discrimination of fat and fluid changes in obesity. Compared to healthy adults with normal BMI (controls), obese individuals have shorter vectors and similar phase angles. However, obese adults with oedema show significantly shorter vector length than the normal weight and otherwise healthy obese adults, and significantly lower phase angle [107]. Vector lengthening occurs with fluid loss (3.2 kg) in the oedematous subjects whereas weight loss (9 kg) in only obese adults does not change vector length. Among healthy women with a wide range of BMI values, mean vector displacement shortens progressively with increasing BMI but phase angle remains constant [108]. The latter finding indicates increased soft tissue mass with normal hydration (e.g., ECW/TBW).

To date, the emphasis in the use of BIVA has been on characterisation of empirical changes in vector displacement with altered hydration states. There are few reports objectively validating BIVA. In response to haemodialysis, changes in the volume of fluid removed are significantly correlated with changes in vector components [96]. Additionally, measurements of TBW made before, during and after pregnancy are significantly correlated with vector length (r = -0.79). Also, changes in vector length are significantly related to changes in TBW (r =-0.599) [109]. Importantly, women whose individual vectors were outside the lower pole of the 75% reference ellipse had significantly greater increases in TBW during the third trimester (10 vs. 5 l) as compared to the other women whose vectors were within the 75% reference ellipse. These findings indicate the validity of BIVA to assess hydration and the sensitivity of this method to identify individuals with fluid overload.

With the increasing use of BIVA there is a need to standardise its application. A critical component is the



use of only phase-sensitive impedance instruments. Reports indicate significant errors in determination of Xc (10–12 ohm) and R (10 ohm) with impedance devices that do not determine Xc directly. Regular calibration of impedance instruments with appropriate resistors and capacitors is needed to assure quality control in studies using BIVA. There is also a need to operationally determine the optimal size of the surface electrodes because wide variance in Xc (20%) can occur with different surface area of the contact electrodes. Because Xc plays a critical role in establishing the position of the individual vector and the phase angle, technical factors that affect the validity of the Xc determination must be understood and controlled.

Conclusions

Measurement of human body composition is a fundamental component of nutritional assessment, with caution advised for the use of newer methods. DXA provides important information regarding soft tissue composition and bone (mass and quality). Practitioners should use a single technology (pencil or fan beam) with the most current software for image analysis. The attraction of bioelectrical impedance analysis should be tempered with the understanding that four-electrode, single-frequency applications are limited to epidemiological or observational studies. Multiple-frequency approaches, particularly impedance spectroscopy, are evolving and require additional validation before routine use can be recommended. Segmental, single- and multiple-frequency (four- and eight-electrode systems) devices yield questionable results and should be considered at a developmental stage. The new technique, BIVA, offers unique opportunities in clinical nutrition for practical assessment of hydration status and body cell mass without the limitation of reliance on assumptions associated with other methods.

Conflict of interest The authors declare that they have no conflict of interest related to the publication of this manuscript.

References

- Wang ZM, Pierson RN, Heymsfield SB (1992) The five level model: a new approach to organizing body composition research. Am J Clin Nutr 56:19–28
- Lukaski HC (1987) Methods for the assessment of human body composition: traditional and new. Am J Clin Nutr 46:537–556
- Heymsfield SB, Lohman TG, Wang ZM, Going SB (2005)
 Human body composition, 2nd Edn. Human Kinetics, Champaign, IL
- Genton L, Hans D, Kyle UG, Pichard C (2002) Dual-energy Xray absorptiometry and body composition: differences between

- devices and comparison with reference methods. Nutrition 18:66-70
- Siri WE (1956) The gross composition of the body. Adv Biol Med Physics 4:239–280
- Laskey MA (1996) Dual-energy X-ray absorptiometry and body composition. Nutrition 12:45–51
- Baumgartner RN, Heymsfield SB, Roche AF (1995) Human body composition and the epidemiology of disease. Obes Res 3:73–95
- Wellens RI, Roche AF, Khamis HJ et al (1996) Relationships between body mass index and body composition. Obes Res 4:35–44
- 9. Prentice AM, Jebb SA (2001) Beyond body mass index. Obes Rev 2:141–147
- Thomsen TK, Jensen VJ, Henriksen MG (1998) In vivo measurement of human body composition by dual-energy x-ray absorptiometry (DXA). Eur J Surg 164:133–137
- Going SB, Massett MP, Hall MC et al (1993) Detection of small changes in body composition by dual-energy x-ray absorptiometry. Am J Clin Nutr 57:845–850
- Horber FF, Thomi F, Casez JP et al (1992) Impact of hydration status on body composition as measured by dual energy X-ray absorptiometry in normal volunteers and patients on haemodialysis. Br J Radiol 65:895–900
- Lukaski HC, Marchello MJ, Hall CB et al (1999) Soft tissue composition of pigs measured with dual X-ray absorptiometry: comparison with chemical analyses and effects of carcass thickness. Nutrition 15:697–703
- Visser M, Fuerst T, Lang T et al (1999) Validity of fan-beam dual-energy x-ray absorptiometry for measuring fat-free mass and leg muscle mass. J Appl Physiol 87:1513–1520
- Chen Z, Wang ZM, Lohman TG et al (2007) Dual-energy X-ray absorptiometry is a valid tool for assessing skeletal muscle mass in older women. J Nutr 137:2775–2780
- Lohman TG, Chen Z (2005) Dual-energy X-ray absorptiometry.
 In: Human body composition, 2nd Edn. Human Kinetics, Champaign, IL, pp 63–78
- Tothill P, Hannan WJ, Wilkinson S (2001) Comparisons between pencil beam and two fan beam dual energy x-ray absorptiometers used for measuring tot body bone and soft tissue. Brit J Radiol 74:166–176
- Oldroyd B, Smith AH, Truscott JG (2003) Cross-calibration of GE/Lunar pencil and fan-beam dual energy densitometers – bone mineral density and body composition studies. Eur J Clin Nutr 57:977–987
- Soriano J-M, Ioannidou E, Wang J et al (2004) Pencil-beam versus fan-beam dual-energy X-ray absorptiometry comparisons across four systems. J Clin Densitom 7:281–289
- Lohman TG (2000) Assessing body composition and changes in body composition. Ann NY Acad Sci 904:45–54
- Lukaski HC (2008) In search of a practical tool to assess regional body composition. Am J Clin Nutr 88:875–876
- 22. Keys A, Brozek J (1953) Body fat in adult men. Physiol Rev 33:245-325
- Dempster P, Aikens S (1995) A new air displacement method for the determination of human body composition. Med Sci Sports Exerc 27:1692–1697
- 24. Fields DA, Goran MI, McCrory MA (2002) Body composition assessment via air displacement plethysmography in adults and children: a review. Am J Clin Nutr 75:453–467
- McCrory MA, Gomez TD, Bernauer EM, Mole PA (1995) Evaluation of a new air displacement plethysmography for estimating human body composition. Med Sci Sports Exerc 27:1686–1691
- Sardinha LB, Lohman TG, Teixeira PJ et al (1998) Comparison of air displacement plethysmography with dual energy X-ray



- absorptiometry and 3 field methods for estimating body composition of middle-aged men. Am J Clin Nutr 68:786–793
- 27. Biaggi RR, Vollman MW, Nies MA et al (1999) Comparison of air displacement plethysmography with hydrostatic weighing and bioelectrical impedance analysis for the assessment of body composition. Am J Clin Nutr 69:898–903
- Miyatake N, Nonaka K, Fujii M (1999) A new air displacement plethysmography for the determination of Japanese body composition. Diabetes Obes Res 1:347–351
- Levenhagen DK, Borel MB, Welch DC et al (1999) A comparison of air displacement plethysmography with three other techniques to determine body fat in healthy adults. J Parenter Enter Nutr 23:293–299
- Nunez C, Korvera AJ, Pietrobelli A et al (1999) Body composition in children and adults by air displacement plethysmography. Eur J Clin Nutr 53:382–387
- Mendez J, Lukaski HC, Buskirk ER (1981) Variability of body density in ambulatory subjects measured at different days. Am J Clin Nutr 34:78–81
- Fields DA, Hunter GA, Goran MI (2000) Validation of the Bod Pod with hydrostatic weighing: influence of body clothing. Int J Obes Relat Metab Disord 24:200–205
- Wagner DR, Heyward VH, Gibson Al (2000) Validation of air displacement plethysmography for assessing body composition. Med Sci Sports Exerc 32:1339–1344
- Vescovi JD, Zimmerman SL, Miller WC et al (2001) Evaluation of the Bod Pod for estimating percentage body fat in a heterogeneous group of adult humans. Eur J Appl Physiol 85:326–332
- 35. Demerath EW, Guo SS, Chumlea WC et al (2002) Comparison of percent body fat estimates using air displacement plethysmography and hydrodensitometry in adults and children. Int J Obes Relat Metab Disord 26:389–397
- 36. Iwaoka H, Yokoyama T, Nakayama T et al (1998) Determination of percent body fat by the newly developed sulfur hexafluoride dilution method and air displacement plethysmography. J Nutr Sci Vitaminol 44:561–568
- 37. Collins MA, Millard-Stafford ML, Sparling PB et al (1999) Evaluation of the Bod Pod for assessing body fat in collegiate football players. Med Sci Sports Exerc 31:1350–1356
- Dewitt O, Fuller NJ, Fewtrell MS et al (2000) Whole body air displacement plethysmography compared with hydrodensitometry for body composition analysis. Arch Dis Child 82:159–164
- Millard-Stafford ML, Collins MA, Evans EM et al (2001) Use of air displacement plethysmography for estimating body fat in a four-component model. Med Sci Sports Exerc 33:1311–1317
- Fields DA, Wilson GD, Gladden LB et al (2001) Comparison of the Bod Pod with the four-compartment model in adult females. Med Sci Sports Exerc 33:1605–1610
- Fields DA, Goran MI (2000) Body composition techniques and the four-compartment model in children. J Appl Physiol 89:613–620
- 42. Lockner DW, Heyward VH, Baumgartner RN, Jenkins KA (2000) Comparison of air displacement plethysmography, hydrodensitometry, and dual X-ray absorptiometry for assessing body composition of children 10 to 18 years of age. Ann NY Acad Sci 904:72–78
- Ball SD, Altena TS (2004) Comparison of the Bod Pod and dual energy X-ray absorptiometry in men. Physiol Meas 25:671–678
- Shafer KJ, Siders WA, Johnson LK, Lukaski HC (2008) Interaction of clothing and body mass index affects validity of airdisplacement plethysmography in adults. Nutrition 24:148–154
- 45. Koda M, Tsuzuku S, Ando F et al (2000) Body composition by air displacement plethysmography in middle-aged and elderly Japanese: comparison with dual-energy absorptiometry. Ann NY Acad Sci 904:484–488
- 46. Fields DA, Higgins PB, Radley D (2005) Air-displacement

- plethysmography: here to stay. Curr Opin Clin Nutr Metab Care 8:624–629
- 47. Kyle U, Boseaus I, De Lorenzo AD et al (2004) Bioelectrical impedance analysis – part I: review of principles and methods. Clin Nutr 23:1226–1243
- Lukaski HC, Johnson PE, Bolonchuk WW, Lykken GI (1985)
 Assessment of fat-free mass using bioelectrical impedance measurements of the human body. Am J Clin Nutr 41:810–817
- 49. Chumlea WC, Sun SS (2005) Bioelectrical impedance analysis. In: Human body composition, 2nd Edn. Human Kinetics, Champaign, IL, pp 79–88
- Kyle U, Boseaus I, De Lorenzo AD et al (2004) Bioelectrical impedance analysis – part II: utilization in clinical practice. Clin Nutr 23:1430–1453
- Chumlea WC, Guo SS, Kuczmarski RJ et al (2002) Body composition estimates from NHANES III bioelectrical impedance data. Int J Obes 26:1596–1609
- 52. Sun SS, Chumlea WC, Heymsfield SB et al (2003) Development of bioelectrical impedance prediction equations for body composition with the use of a multi-component model for use in epidemiologic surveys. Am J Clin Nutr 77:331–340
- Fredrix E, Saris W, Soeters P et al (1990) Estimation of body composition by bioelectrical impedance in cancer patients. Eur J Clin Nutr 44:749–752
- 54. Schols A, Wouters E, Soeters P, Westerterp K (1991) Body composition by bioelectrical impedance analysis compared with deuterium dilution and skinfold anthropometry in patients with chronic obstructive pulmonary disease. Am J Clin Nutr 53:421–424
- 55. Cornish B, Thomas B, Ward L (1993) Improved prediction of extracellular and total body water using impedance loci generated by multiple frequency bioelectrical impedance analysis. Phys Med Biol 38:337–346
- Thomasset A (1962) Bio-electrical properties of tissue impedance measurements. Lyon Med 207:107–118
- 57. Hannan WJ, Cowen SJ, Fearon KC et al (1994) Evaluation of multi-frequency bio-impedance analysis for the assessment of extracellular and total body water in surgical patients. Clin Sci 86:479–485
- Olde Rikkert MGM, Deurenberg P, Jansen RWMW et al (1997) Validation of multifrequency bioelectrical impedance analysis in detecting changes in geriatric patients. J Am Geriatr Soc 45:1345–1351
- Dittmar F, Reber H (2001) New equations for estimating body cell mass from bio-impedance parallel models in healthy older Germans. Am J Physiol 281:E1005–E1014
- 60. Simpson JA, Lobo DN, Anderson JA et al (2001) Body water compartment measurements: a comparison of bioelectrical impedance analysis with tritium and sodium bromide dilution techniques. Clin Nutr 20:339–343
- 61. Martinoli R, Mohamed EI, Mailo C et al (2003) Total body water estimation using bioelectrical impedance: a meta-analysis of the data available in the literature. Acta Diabetol 40:S203–S206
- 62. De Lorenzo A, Andreoli A, Matthie J, Withers P (1997) Predicting body cell mass with bioimpedance by using theoretical methods: a technological review. J Appl Physiol 85:1542–1558
- Ward LC, Elia M, Cornish BH (1998) Potential errors in the application of mixture theory to multifrequency bioelectrical impedance analysis. Physiol Meas 19:53–60
- 64. Schoeller DA (2000) Bioelectrical impedance analysis. What does it measure? Ann NY Acad Sci 904:159–162
- 65. Cox-Reijven PL, Soeters PB (2000) Validation of bio-impedance spectroscopy: effects of degree of obesity and ways of calculating volumes from measured resistance values. Int J Obesity Rel Metab Disord 24:271–280
- 66. Earthman CP, Mathie JR, Reid PM et al (2000) A comparison of



- bioimpedance methods for detection of body cell mass change in HIV infection. J Appl Physiol 88:944–956
- 67. Ho LT, Kushner RF, Schoeller DA et al (1994) Bioimpedance analysis of total body water in hemodialysis patients. Kidney Int 46:1438–1442
- 68. Baarends EM, Van Marken Lichtenbelt WD, Wouters EFM, Schols AMWJ (1998) Body water compartments measured by bio-electrical impedance spectroscopy in patients with chronic obstructive pulmonary disease. Clin Nutr 17:15–22
- 69. Lehnert ME, Clarke DD, Gibbons JG et al (2001) Estimation of body water compartments in cirrhosis by multiple-frequency bioelectrical impedance analysis. Nutrition 17:31–34
- Gudivaka R, Schoeller DA, Kushner RF, Bolt MJ (1999) Singleand multifrequency models for bioelectrical impedance analysis of body water compartments. J Appl Physiol 87:1087–1096
- Ellegård LH, Aåhlen M, Körner U et al (2008) Bioelectric impedance spectroscopy underestimates fat-free mass compared to dual energy X-ray absorptiometry in incurable cancer patients. Eur J Clin Nutr (DOI:10.1308/ejcn.2008.35)
- Ellis KJ, Wong WW (1998) Human hydrometry: comparison of multifrequency bioelectrical impedance with 2H2O and bromine dilution. J Appl Physiol 85:1056–1062
- 73. Mager JR, Sibley SD, Beckman TR et al (2008) Multifrequency bioelectrical impedance analysis and bioimpedance spectroscopy for monitoring fluid and body cell mass changes after gastric bypass surgery. Clin Nutr 27:832–841
- Moissl UM, Wabel P, Chamney PW et al (2006) Body fluid volume determination via body composition spectroscopy in health and disease. Physiol Meas 27:921–933
- Nunez C, Gallagher D, Grammes J et al (1999) Bioimpedance analysis: potential for measuring lower limb skeletal muscle mass. JPEN J Parenter Enteral Nutr 23:96–103
- Utter AC, Nieman DC, Ward AG, Butterworth DE (1999) Use of the leg-to-leg bioelectrical impedance method in assessing body composition change in obese women. Am J Clin Nutr 69:603–607
- Mitayani M, Kanehisa H, Fukunaga T (2000) Validity of bioelectrical impedance and ultrasonographic methods for estimating muscle volume of the upper arm. Eur J Appl Physiol 82:391–396
- Biggs J, Cha K, Horch K (2001) Electrical resistivity of the upper arm and leg yields good estimates of whole body fat. Physiol Meas 22:365–376
- Baumgartner RN, Chumlea WC, Roche AF (1989) Estimation of body composition from segment impedance. Am J Clin Nutr 50:221–226
- Lukaski HC, Scheltinga MRM (1994) Improved sensitivity of tetrapolar bioelectrical impedance estimates of fluid change and fat-free mass with proximal placements of electrodes. Age Nutr 5:123–129
- Lukaski HC, Siders WA (2003) Validity and accuracy of regional bioelectrical impedance devices to determine whole-body fatness. Nutrition 19:851–857
- 82. Rowlands AV, Eston RG (2001) Comparison of arm-to-leg and leg-to-leg (standing) bioelectrical impedance analysis for the estimation of body composition in 8- to 10-year old children. In: Jürimäe T, Hills AP (eds) Body composition assessment in children and adolescents, Vol 44. Karger, Basel, pp 14–22
- 83. Jürimäe J, Leppik A, Jürimäe T (2001) Whole body resistance measured between different limbs and resistance indices in preadolescent children. In: Jürimäe T, Hills AP (eds) Body composition assessment in children and adolescents, Vol 44. Karger, Basel, pp 53–65
- 84. Pietrobelli A, Rubiano F, St-Onge M-P, Heymsfield SB (2004) New bioimpedance analysis system: improved phenotyping with whole-body analysis. Eur J Clin Nutr 58:1479–1484
- 85. Demura S, Sato S, Kitabayashi T (2004) Percentage of total

- body fat as estimated by three automatic bioelectrical impedance analyzers. J Physiol Anthropol Appl Human Sci 23:93–99
- 86. Demura S, Sato S, Kitabayashi T (2005) Estimation accuracy of percent total body fat and percent segmental fat measured by single-frequency bioelectrical impedance analysis with 8 electrodes: the effect of difference in adiposity. J Sports Med Phys Fitness 45:68–76
- 87. Neovius M, Hemmingsson E, Freyschuss B, Udden J (2006) Bioelectrical impedance underestimates total and truncal fatness in abdominally obese women. Obesity 14:1731–1738
- 88. Völgyi E, Tylavsky FA, Lyytikäinen A et al (2008) Assessing body composition with DXA and bioimpedance: effects of obesity, physical activity, and age. Obesity 16:700–705
- 89. Bedogni G, Malavolti M, Severi S et al (2002) Accuracy of an eight-point tactile-electrode impedance method in the assessment of total body water. Eur J Clin Nutr 56:1143–1148
- Sartorio A, Malavolti M, Agosti F et al (2004) Body water distribution in severe obesity and its assessment from eight-polar bioelectrical impedance analysis. Eur J Clin Nutr 59:155–160
- Medici G, Mussi C, Fantuzzi AL et al (2005) Accuracy of eightpolar bioelectrical impedance for the assessment of total and appendicular body composition in peritoneal dialysis patients. Eur J Clin Nutr 59:932–937
- 92. Gibson AL, Holmes JC, Desautels RL et al (2008) Ability of new octapolar bioimpedance spectroscropy analyzers to predict 4-component-model percentage body fat in Hispanic, black, and white adults. Am J Clin Nutr 87:332–338
- 93. Shafer KJ, Siders WA, Johnson LK, Lukaski HC (2009) Validity of segmental multiple-frequency bioelectrical impedance analysis to estimate body composition across a range of body mass indexes. Nutrition 25:25–32
- Piccoli A, Rossi B, Pillon L, Bucciante G (1994) A new method for monitoring body fluid variation: the RXc graph. Kidney Int 46:534–539
- 95. Piccoli A, Pillon L, Dumler F (2002) Impedance vector distribution by sex, race, body mass index, and age in the United States: standard reference intervals as bivariate Z scores. Nutrition 18:153–167
- Piccoli A (1998) Identification of operational clues to dry weight prescription in hemodialysis using bioimpedance vector analysis. Kidney Int 53:1036–1043
- Piccoli A, Nigrelli S, Caberlotto A et al (1995) Bivariate normal values of the bioelectrical impedance vector in adult and elderly populations. Am J Clin Nutr 61:269–270
- Piccoli A (2004) Bioelectrical impedance vector distribution in peritoneal dialysis patients with different hydration status. Kidney Int 65:1050–1063
- Piccoli A, Pittoni G, Facco E et al (2000) Relationship between central venous pressure and bioimpedance vector analysis in critically ill patients. Crit Care Med 28:132–137
- Piccoli A, Piazza P, Noventa D et al (1996) A new method for monitoring hydration at high altitude by bioimpedance analysis. Med Sci Sports Exerc 28:1517–1522
- Codognotto M, Piazza M, Frigatti P, Piccoli A (2008) Influence of localized edema on whole-body and segmental bioelectrical impedance. Nutrition 24:569–574
- Nescolarde L, Donate T, Piccoli A, Rosell J (2008) Comparison of segmental with whole-body impedance measurements in peritoneal dialysis patients. Med Eng Phys 30:817–824
- 103. Buffa R, Floris G, Marini E (2002) Bioelectrical impedance vector in pre- and postmenarcheal females. Nutrition 18:474–478
- 104. De Palo T, Messina G, Edefonti A et al (2000) Normal values of the bioelectrical impedance vector in childhood and puberty. Nutrition 16:417–424
- 105. Guida B, Laccetti R, Gerardi C et al (2007) Bioelectrical impedance analysis and age-related differences in body composition in the elderly. Nutr Metab Cardiovasc Dis 17:175–180



- 106. Buffa R, Floris G, Marini E (2009) Assessment of nutritional status in free-living elderly individuals by bioelectrical impedance vector analysis. Nutrition 25:3–5
- 107. Piccoli A, Brunani A, Savia G et al (1998) Discriminating between body fat and fluid changes in the obese adult using bioimpedance vector analysis. Int J Obesity 22:97–104
- 108. Guida B, Trio R, Pecoraro P et al (2003) Impedance vector distribution by body mass index and conventional bioelectrical impedance analysis in obese women. Nutr Metab Cardiovasc Dis 13:72–79
- 109. Lukaski HC, Hall CB, Siders WA (2007) Assessment of change in hydration in women during pregnancy and postpartum with bioelectrical impedance vectors. Nutrition 23:543–550

