In [21]: library(Seurat) library(stringr) library(SeuratDisk) `%notin%` <- Negate(`%in%`)</pre> library(harmony) library(Matrix) library(pheatmap) library(cluster) library(ggpubr) Read in data In [4]: # dir <- '/mnt/storage/projects/huang macrophage/liver/GSE136103/'</pre> # file <- c('GSM4041150_healthy1_cd45+','GSM4041153_healthy2_cd45+', 'GSM4041155_healthy3_cd45+','GSM4041158 healthy4 cd45+', 'GSM4041160 he 'GSM4041161 cirrhotic1 cd45+', 'GSM4041164 cirrhotic2 cd45+', 'GSM4041166 cirrhotic3 cd45+', 'GSM4041168 cirrhotic4 cd45+', 'GSM4041169 # Liver <- list()</pre> # for(i in 1: length(file)) { counts <- ReadMtx(mtx = paste0(dir, file[i], '_matrix.mtx.gz'), cells = paste0(dir, file[i], '_barcodes.tsv.gz'), features = paste0(c Liver[[i]] <- CreateSeuratObject(counts = counts, project = file[i], min.features = 200) Liver[[i]]\$mito.ratio <- PercentageFeatureSet(Liver[[i]], pattern = "^MT-") / 100 Liver[[i]]\$orig.ident <- file[[i]]</pre> Liver[[i]] <- Liver[[i]][, Liver[[i]]\$mito.ratio < .15] rm(counts) # } # memory.profile() # Liver <- merge(Liver[[1]], Liver[2: length(Liver)])</pre> # DefaultAssay(Liver) <- 'RNA'</pre> # Liver\$rpl <- PercentageFeatureSet(Liver, pattern = 'RPL') / 100 - PercentageFeatureSet(Liver, pattern = 'RPL*P') / 100 # Liver\$rps <- PercentageFeatureSet(Liver, pattern = 'RPS') / 100 - PercentageFeatureSet(Liver, pattern = 'RPs*P') / 100 # Liver\$ribo.ratio <- Liver\$rpl + Liver\$rps</pre> # Liver\$heam.ratio <- (PercentageFeatureSet(Liver, pattern = '^HBB') + PercentageFeatureSet(Liver, pattern = '^HBA')) / 100 # Liver <- Liver[, Liver\$mito.ratio < .15 & Liver\$ribo.ratio < .3 & Liver\$heam.ratio < .001 & Liver\$nFeature_RNA < 5000] # Liver <- SplitObject(Liver, split.by = 'orig.ident')</pre> # for(i in 1:length(Liver)) { Liver[[i]] <- NormalizeData(Liver[[i]])</pre> Liver[[i]] <- FindVariableFeatures(Liver[[i]])</pre> # } # anchors <- FindIntegrationAnchors(object.list = Liver, dims = 1:30)</pre> # Liver <- IntegrateData(anchorset = anchors, dims = 1:30)</pre> # Liver <- ScaleData(Liver, verbose = FALSE, vars.to.regress = c('mito.ratio', 'nFeature RNA'))</pre> # rm(anchors) # memory.profile() # Liver <- RunPCA(Liver)</pre> # Liver <- RunUMAP(Liver, dims = 1:30, verbose = F)</pre> # DefaultAssay(Liver) <- 'integrated'</pre> # Liver <- FindNeighbors(Liver, dims = 1:30)</pre> # Liver <- FindClusters(Liver, resolution = .5)</pre> options(repr.plot.width=12, repr.plot.height=6) p1 <- DimPlot(Liver, label = T)</pre> DefaultAssay(Liver) <- 'RNA'</pre> Liver <- NormalizeData(Liver)</pre> p2 <- FeaturePlot(Liver, 'CD68')</pre> CombinePlots(list(p1, p2)) Warning message: "CombinePlots is being deprecated. Plots should now be combined using the patchwork system." **CD68** 10 10 --5 17 -10 -10 --15 -10 10 -10 10 UMAP_1 UMAP_1 In [6]: # Liver <- Liver[, Idents(Liver) %in% c(2, 6, 9, 12)] # Liver <- SplitObject(Liver, split.by = 'orig.ident')</pre> # for(i in 1:length(Liver)) { Liver[[i]] <- NormalizeData(Liver[[i]])</pre> Liver[[i]] <- FindVariableFeatures(Liver[[i]])</pre> # } # anchors <- FindIntegrationAnchors(object.list = Liver, dims = 1:30)</pre> # Liver <- IntegrateData(anchorset = anchors, dims = 1:30)</pre> # Liver <- ScaleData(Liver, verbose = FALSE, vars.to.regress = c('mito.ratio', 'nFeature_RNA'))</pre> # rm(anchors) # memory.profile() # Liver <- RunPCA(Liver)</pre> # Liver <- RunUMAP(Liver, dims = 1:20, verbose = F)</pre> # DefaultAssay(Liver) <- 'integrated'</pre> # Liver <- FindNeighbors(Liver, dims = 1:20)</pre> # Liver <- FindClusters(Liver, resolution = .8)</pre> options(repr.plot.width=8, repr.plot.height=8) DimPlot(Liver, label = T, label.size = 5) DefaultAssay(Liver) <- 'RNA'</pre> Liver <- NormalizeData(Liver)</pre> options(repr.plot.width=24, repr.plot.height=6) FeaturePlot(Liver, label = T, label.size = 5, features = c('CDKN1C', 'CD1C', 'CCL5', 'CCR7'), ncol = 4) UMAP_1 CDKN1C CCL5 CD1C CCR7 In [10]: #Liver <- t options(repr.plot.width=8, repr.plot.height=8) FeaturePlot(Liver, 'SPP1', label = T) SPP1 UMAP_1 In [16]: # Liver <- Liver[, Idents(Liver) %notin% c(3, 4, 7, 9)]</pre> # DefaultAssay(Liver) <- 'RNA'</pre> # Liver <- Liver[rowSums(Liver\$RNA@counts >0) > 10,] # Liver <- SplitObject(Liver, split.by = 'orig.ident')</pre> # for(i in 1:length(Liver)) { DefaultAssay(Liver[[i]]) <- 'RNA'</pre> Liver[[i]] <- NormalizeData(Liver[[i]])</pre> Liver[[i]] <- FindVariableFeatures(Liver[[i]])</pre> # anchors <- FindIntegrationAnchors(object.list = Liver, dims = 1:20)</pre> # Liver <- IntegrateData(anchorset = anchors, dims = 1:20)</pre> # Liver <- ScaleData(Liver, verbose = FALSE, vars.to.regress = c('mito.ratio', 'nFeature_RNA'))</pre> # rm(anchors) # memory.profile() # Liver <- RunPCA(Liver)</pre> # Liver <- RunUMAP(Liver, dims = 1:12, verbose = F, min.dist = .6) # DefaultAssay(Liver) <- 'integrated'</pre> # Liver <- FindNeighbors(Liver, dims = 1:12)</pre> # Liver <- FindClusters(Liver, resolution = .25)</pre> options(repr.plot.width=8, repr.plot.height=8) DimPlot(Liver, label = T, label.size = 5, group.by = 'integrated snn res.0.3') options(repr.plot.width=12, repr.plot.height=6) DefaultAssay(Liver) <- 'RNA'</pre> FeaturePlot(Liver, c('CCR7', 'SPP1'), ncol = 2) integrated_snn_res.0.3 UMAP_2 -10 UMAP_1 CCR7 SPP1 2.0 UMAP UMAP 1.5 1.0 0.5 0.0 -10 UMAP_1 UMAP_1 In [12]: # DefaultAssay(Liver) <- "integrated"</pre> # Liver <- FindNeighbors(Liver, dims = 1:12)</pre> # Liver <- FindClusters(Liver, resolution = seq(from = 0.05, to = 1, by = .05)) # DefaultAssay(Liver) <- 'integrated'</pre> # resolution <- colnames(Liver@meta.data)[startsWith(x =colnames(Liver@meta.data), prefix = 'integrated_snn_res')] # silhouetteee <- list()</pre> # i <- 1 # dist.matrix <- dist(x = Embeddings(object = Liver[['pca']])[,1:12])</pre> # for(sil in sort(resolution)) { $s \leftarrow silhouette(x = as.numeric(x = as.factor(x = Liver@meta.data[, sil])), dist = dist.matrix)$ silhouetteee[[i]] <- summary(s)</pre> i < -i + 1# sil_score <- c()</pre> # for(sil in silhouetteee) { sil_score <- c(sil_score, mean(sil\$clus.avg.widths))</pre> # } # to_plot <- data.frame('Score' = sil_score[1: length(sil_score)], 'Res' = seq(from = 0.05, to = 1, by = .05)) # to_plot\$Score <- round(to plot\$Score,digits = 2)</pre> # p1 <- ggscatter(data = to_plot, x = 'Res', y = 'Score', xlab = 'Resolution', ylab = 'Silhouette Score') + # rotate_x_text(angle = 45) + theme(text = element_text(size = 19), plot.title = element_text(hjust = 0.5)) # p2 <- DimPlot(Liver, group.by = 'integrated_snn_res.0.3')</pre> options(repr.plot.width=12, repr.plot.height=6) CombinePlots(list(p1, p2)) Warning message: "CombinePlots is being deprecated. Plots should now be combined using the patchwork system." Warning message: "Graphs cannot be vertically aligned unless the axis parameter is set. Placing graphs unaligned." "Graphs cannot be horizontally aligned unless the axis parameter is set. Placing graphs unaligned." integrated_snn_res.0.3 0.4 Silhouette Score 0.1 Resolution UMAP_1 Merge clusters with similiar transcriptome In [17]: DefaultAssay(Liver) <- 'RNA'</pre> Liver<- FindVariableFeatures(Liver, nfeatures = 2000)</pre> Endo_bulk <- AverageExpression(Liver, group.by = 'integrated_snn_res.0.3', assays = 'RNA', features = Liver\$RNA@var.features)\$RNA</pre> Endo_bulk <- Endo_bulk / rowSums(Endo_bulk); Endo_bulk[rowSums(is.na(Endo_bulk)) > 0] <- NULL</pre> corr <- cor(x = Endo_bulk, y =Endo_bulk, method = 'spearman')</pre> options(repr.plot.height = 8, repr.plot.width = 8) paletteLength <- 20</pre> myColor <- colorRampPalette(c("blue", "white", "red"))(paletteLength)</pre> myBreaks <- c(seq(-1, 0, length.out=ceiling(paletteLength/2) + 1), seq(max(corr)/paletteLength, max(corr), length.out=floor(paletteLength/2) pheatmap(corr, display_numbers = T, breaks = myBreaks, color=myColor, number_format = '%.02f', border_color = 'white', number_color = 'black fontsize_number = 25, fontsize_row = 25, fontsize_col = 25, angle_col = 45, cluster_rows = T, cluster_cols = T) 1.00 0.52 -0.49-0.25 0.52 -0.36 1.00 -0.50 -0.49-0.50 -0.34 1.00 -0.25-0.36 -0.341.00 ദ 0 **>** Plot marker gene of each clsuter In [23]: # Idents(Liver) <- Liver\$integrated snn res.0.3 # DefaultAssay(Liver) <- 'RNA'</pre> # Liver <- NormalizeData(Liver)</pre> # Liver <- FindVariableFeatures(Liver)</pre> # Liver <- ScaleData(Liver, vars.to.regress = c('mito.ratio', 'nFeature RNA'))</pre> # All Heart.m <- FindAllMarkers(Liver, only.pos = T, min.diff.pct = .1)</pre> # All Heart.m <- All Heart.m[order(All Heart.m\$avg log2FC, decreasing = T),] tmp <- Liver DefaultAssay(tmp) <- 'RNA'</pre> Idents(tmp) <- as.character(tmp\$integrated_snn_res.0.3)</pre> tmp\$try <- factor(tmp\$integrated_snn_res.0.3 , levels = c(3,1, 0, 2)); res <- 'try'</pre> Heart_all_prev_m <- All_Heart.m [All_Heart.m \$pct.1 - All_Heart.m \$pct.2 > .2 & All_Heart.m \$pct.1 > .2,] tmp 8 gene <- c() for(clus in intersect(levels(tmp\$try), unique(Heart_all_prev_m\$cluster))) { tmp_8_gene <- c(tmp_8_gene, Heart_all_prev_m[Heart_all_prev_m\$cluster == clus,]\$gene[1:10])</pre> genes.to.label <- c('SELENOP','C1QA','RNASE1','MARCO','C1QC','FCN1', 'S100A9','S100A8','VCAN', 'APOE','GPNMB','CD9','STAB1','SPP1', 'FABP5' 'CCL20', 'FOLR2', 'INHBA', 'CHI3L1', 'SPP1', 'MATK', 'PTGS2', 'IL1B', 'FABP4', 'LYVE1', 'APOC1', 'CCL4', 'S100A12', 'CCL3', 'DAB2', 'LYVE1', 'CCL4L2', 'PHLDA3', 'EGR1', 'ATF3', 'VEGFA', 'SEMA6B', 'IFI44L', 'IFIT3', 'IFIT1') all.genes <- intersect(tmp 8 gene, rownames(tmp\$RNA@scale.data))</pre> labels <- rep(x = "transparent", times = length(x = all.genes))</pre> labels[match(x = genes.to.label, table = all.genes)] <- "black"</pre> options(repr.plot.height = 16, repr.plot.width = 12) DoHeatmap(tmp, group.colors = c('#F8766D','#7CAE00','#00BFC4', '#b149ff','#c77cff', '#ddafff'), features = tmp_8_gene, slot = 'scale.data', assay = 'RNA', disp.min = -2, disp.max = 2, angle = -30, hjust = 1, group.by = res) + theme(axis.text.y = element_text(color = rev(x = labels), size = 19)) Warning message in DoHeatmap(tmp, group.colors = c("#F8766D", "#7CAE00", "#00BFC4", : "The following features were omitted as they were not found in the scale.data slot for the RNA assay: NA, CSTA, CD55, S100A6, MNDA" Warning message: "Vectorized input to `element_text()` is not officially supported. Results may be unexpected or may change in future versions of ggplot2." 2 ઝ SPP1 APOC1 FABP5 **GPNMB** CD9 TREM2 **APOE** S100A8 Expression S100A9 S100A12 **VCAN** Identity FCN1 • 0 • 2 MARCO C1QA Proportion of SPP1+ cells in disease vs control In [20]: Liver\$cluster_d <- Liver\$integrated_snn_res.0.3</pre> Liver\$cluster_d <- plyr::mapvalues(Liver\$cluster_d, from = c(3, 1, 0, 2), to = c('SPP1', 'FCN1', 'Trans', 'MARCO'))Liver\$Diagnosis <- 'Control'</pre> Liver\$Diagnosis[Liver\$orig.ident %in% c('GSM4041161_cirrhotic1_cd45+', 'GSM4041164_cirrhotic2_cd45+', 'GSM4041166_cirrhotic3_cd45+', 'GSM4041168_cirrhotic4_cd45+', 'GSM4041169_cirrhotic5_cd45+')] <- 'Cirrhosis' options(repr.plot.height = 8, repr.plot.width = 6) SPP1 <- (table(Liver\$orig.ident, Liver\$cluster_d) / rowSums(table(Liver\$orig.ident, Liver\$cluster_d)))[table(Liver\$orig.ident) > 0, 'SPP1'] Diagnosis <- plyr::mapvalues(names(table(Liver\$orig.ident))[table(Liver\$orig.ident) > 0], from = Liver\$orig.ident[!duplicated(Liver\$orig.ident)], to = Liver\$Diagnosis[!duplicated(Liver\$orig.ident)]) tmp <- data.frame('SPP1' = SPP1, 'Diagnosis' = Diagnosis)</pre> ggboxplot(tmp, x = 'Diagnosis', y = 'SPP1', add = "jitter") + stat_compare_means(method = 'wilcox.test', comparisons = list(c('Cirrhosis', 'Control'))) + theme classic(base size = 20) + theme(axis.text.x=element_text(angle = 30, hjust =1, vjust = 1, size = 20)) + ylab('Proportion of SPP1+ cells') options(repr.plot.height = 8, repr.plot.width = 8) DimPlot(Liver, group.by = 'cluster d', label.size = 10, label = T, repel = T) + NoLegend() 0.032 0.06 roportion of SPP1+ cells Д 0.00 Diagnosis cluster_d Trans UMAP_2 FCN1 -10 UMAP_1 In [26]: saveRDS(Liver, file = 'Liver_code_final.RDS')