

10 Jan 2019

MITOMI fitting.  
from Ivan's email.

70887

0.020 /  $\lambda$  x 50  $\lambda$   
0.5 1V  
100

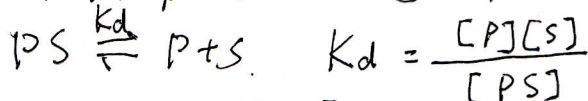
$$[DNA * Pho4] = B_{max} \times \frac{[DNA_f]}{[DNA_f] + K_d}$$

$$\text{Fractional binding} = B_{max} \frac{[DNA_f]}{[DNA_f] + K_d}$$

from my own calc.  $K_d, R$

- ①. Read in protein raw F530. median  $\rightarrow$  protein
  - ②. Read in DNA-bound raw F685. median  $\rightarrow$  DNA
  - ③. Read in DNA free
  - ④. Remove background.
  - ⑤. Remove ~~inter~~ correlation between protein channel and DNA6.
- from an unnamed book's ch 13.

P = TF protein. S = DNA Site



$$[PS] = \frac{[P][S]}{K_d}$$

$$[S]_t = [S] + [PS] = [S] + \frac{[P][S]}{K_d} = \frac{[S]K_d + [P][S]}{K_d}$$

$$\frac{[PS]}{[S]} = \frac{[P]}{K_d}$$

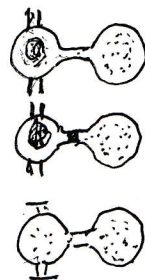
$$= \frac{K_d \cdot [S] + [P] \cdot [S]}{K_d}$$

// I think we are operating in a situation of ~~the~~ "one-component saturating" specifically, we provide sufficient amount of TF s.t.  $[P] \gg K_d$ .

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Steps to perform MITOMI

1. ITT flow into the chip w/ all valves open except for outlet.  
Filling, especially the chamber.
2. Chamber valves closed (protect DNA), keep flushing.
3. Sandwich valves closed, chamber & button open.  
this is the reaction vessel!



4. After 90 min, scan 1.  $\rightarrow$  total DNA conc.  $[S]_t$

note: the concept of "concentration" here refers to total fluorescent intensity



$\Sigma Cy5 / 1 \text{ pixel area.}$   
 $\sim \text{molecule} \sim \text{volume}$

// measured as median of pixel intensity in the chamber? unit  $\frac{F685, \text{median}}{\text{Pixel.}}$

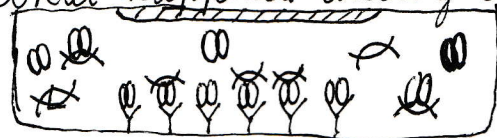
5. button membrane closed.

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MITOMI.

Continue

What happened during the incubation period.



\* surface = on button membrane

Y surface bound  $\alpha$ -His ab.

TF protein

DNA - Cy5.

Free DNA  $[S_f]$

Free protein  $[P_f] = \text{surface bound} + \text{in solution not bound}.$

DNA in complex protein  $[PS] = \text{surface bound} + \text{in solution}.$

Assume  $[PS]$  and  $[P_f]$  has the same distribution between surface & solution, then one can use  $\frac{[PS]_{\text{surface}}}{[P_f]_{\text{surface}}}$  as a proxy for  $\frac{[PS]_{\text{all}}}{[P_f]_{\text{all}}}.$

5. after membrane is closed. keep open sandwich valve, close chamber valve flush

scan 2  $\rightarrow$  surface bound total protein

DNA bound to protein that is bound to surface  $\rightarrow [PS]_{\text{surface}} + [P_f]_{\text{surface}} \rightarrow [PS]_{\text{surface}}$

// note that  $[PS] + [P_f] = [P]_{\text{total}}$  is measured in F530. protein channel

$[PS]$

$\rightarrow$  in DNA units

F685 (Cy5) DNA channel

311-621-5729. David Soll.

What is measured.

$\frac{[PS]_{\text{surface}} \rightarrow \text{Cy5}}{[PS]_{\text{surface}} + [P_f]_{\text{surface}}} \rightarrow \text{fractional occupancy } FO$

$[P]_{\text{total}}$  under button in F530 protein channel

notice that the ratio is not unitless, due to the two different fluorophore being used to estimate  $[PS]$  and  $[P]_t$ .

See Bin He notes on 5<sup>th</sup> mai 2009 (paper)

$[P]_t \cdot C_p$ ,  $[S]_t \cdot C_s$ ,  $[PS] \cdot C_s$

$$y = \frac{[PS] \cdot C_s}{[P]_t \cdot C_p} = \frac{C_s}{C_p} \frac{[PS]}{[P]_t} = \frac{C_s}{C_p} \frac{[S]_f \cdot C_s}{K_d + [S]_f \cdot C_s}$$

$$K_d = \frac{[P_f][S_f]}{[PS]} \text{ all in nm units}$$

$$FO = \frac{[PS]}{[P_f] + [PS]} = \frac{[PS]}{\frac{K_d[PS]}{[S_f]} + [PS]} = \frac{[S_f]}{K_d + [S_f]}$$

During fitting, however,  $[S]_t$  is used in place of  $[S]_f$ .

$\rightarrow$  what's desired?  $[S]_f$   
what's used/measured?  $[S]_t \cdot C_s$ .

Can we use  $[S]_t$  to approximate  $[S]_f$ ?



12 Jan 2019.

\* ~~Various~~ Various approximations

(See also my paper notebook on 5 mai 2009).

□ organize reading list for Zohab / Thomas

1. For strong (e.g. consensus site), we should be able to reach

( $0.1 \mu\text{m} \sim 0.01 \mu\text{m}$  :  $100 \sim 10 \text{ nm}$ .  
if  $K_d$  for CACGTC is  $\sim 10 \text{ nM}$ , and the spotted DNA is ? (can't use the [DNA] in spotting to detect, as volume changes).

ScPhof  $\rightarrow$  CACGTC.  $K_d \sim 11.1 \text{ nM}$  (Maerkle & Quake 2007)

for consensus site, we will reach the plateau - that is,

$[S]_t$  will be greater than  $K_d$  ( $[S]_t \gg K_d$ ), such that the protein is saturated!

$$\frac{[PS]}{[P]_t} \rightarrow 1. \quad \text{or} \quad \frac{C_s \cdot [PS]}{C_p \cdot [P]_t} \rightarrow \frac{C_s}{C_p} = B_{\max}.$$

[PS]

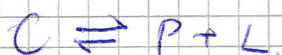
at lower  $[S]_t$ , ( $[S]_t < K_d$ ), if  $[P]_t \ll K_d$ , then  $\frac{[PS]C_s}{[P]_t C_p} = B_{\max} \frac{[S]_f \cdot C_s}{K_d + [S]_f \cdot C_s}$  \*

\*  $\approx B_{\max} \frac{[S]_f \cdot C_s}{K_d}$  (linear) in this case.

DNA $K_d$ / $[S]_t$	low	high
low (consensus)	Fractional Occupancy is low (in linear range) assume $[P] \ll K_d(\text{consensus})$ $\frac{[PS]}{[S]_t} = \frac{[P]_f}{K_d + [P]_f} \approx \frac{[P]_f}{K_d} \ll 1$ $[S]_f \approx [S]_t$	If we assume $[P] \ll K_d(\text{consensus})$ then regardless of $[S]_t$ . $\frac{[PS]}{[S]_t} = \frac{[P]_f}{K_d + [P]_f} \approx \frac{[P]_f}{K_d} \ll 1$ $\Rightarrow [S]_f \approx [S]_t$
high (weak sites)		

Table. 分情况讨论. Can  $[S]_t \approx [S]_f$ .

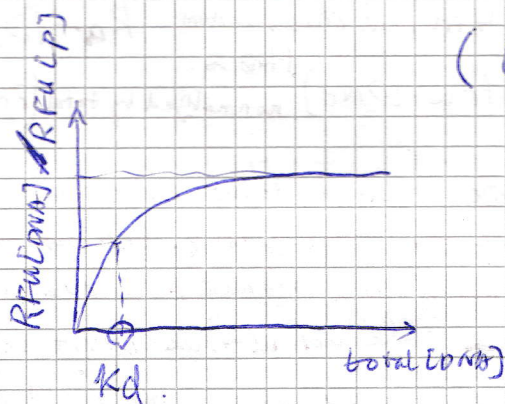
\* Protein - ligand binding affinity [Wiki -  $K_d$  affinity]



$$K_d = \frac{[P][L]}{[PL]}$$

$K_d$  has molar units (M), which corresponds to the [conc] of ligand [L] at which the binding site on a particular protein is half occupied.

$$(K_d [L] = K_d \Rightarrow \frac{[P]}{[P] + [L]} = 1.)$$



~~HT2SDP~~

Saturation : [Protein: Ligand complex] =  $B_{max} [Ligand] / K_d + B_{max}$

where  $B_{max}$  is the maximum

[Theory III - Fraile!]

What I measure

$S_1 [D]_{free} \cdot C_{CONA}$  — before wash, outside the Button.

$S_2 [PD] \cdot C_{CONA}$  — after wash.

$S_2 [P]_t \cdot C_p$  — after wash

where  $S_1, S_2$  are the size of the detection area.

$C_{CONA}$  is the constant that translates  $\frac{[CD]}{[RFU]}$  to RFU. determined by the standard curve



$$K_D = \frac{[P][D]}{[PD]} \Rightarrow [PD] K_D = [P][D] = ([P]_t - [PD])[D]$$

$$\Rightarrow [PD] (K_D + [D]) = [P]_t \cdot [D]$$

$$\Rightarrow \frac{[PD]}{[P]_t} = \frac{[D]}{K_D + [D]}$$

$$Y = B_{max} \frac{X}{X + K_D}$$

$$\frac{1}{Y} = \frac{1}{B_{max}} \left( 1 + \frac{1}{X} K_D \right)$$

$$\frac{S_2 [PD] \cdot C_{DNA}}{S_2 [P]_t \cdot C_p} \cdot \frac{C_p}{C_{DNA}} = \frac{\frac{C_{DNA}}{C_p} \cdot [D]}{K_D + [D]}$$

$\downarrow$   $\uparrow$   
 $Y$   $X$   
 $\downarrow$   $\uparrow$   
 $B_{max}$   $K_D$

Standard RFU curve

$$Y \approx RFU = 28.147 (DNA)_{(nM)}$$

$$\text{or } (DNA)_{(nM)} = 0.0355 RFU$$

der

$$\frac{1}{Y} = \frac{1}{B_{max}} \cdot \frac{1}{X} \cdot K_D + \frac{1}{B_{max}}$$

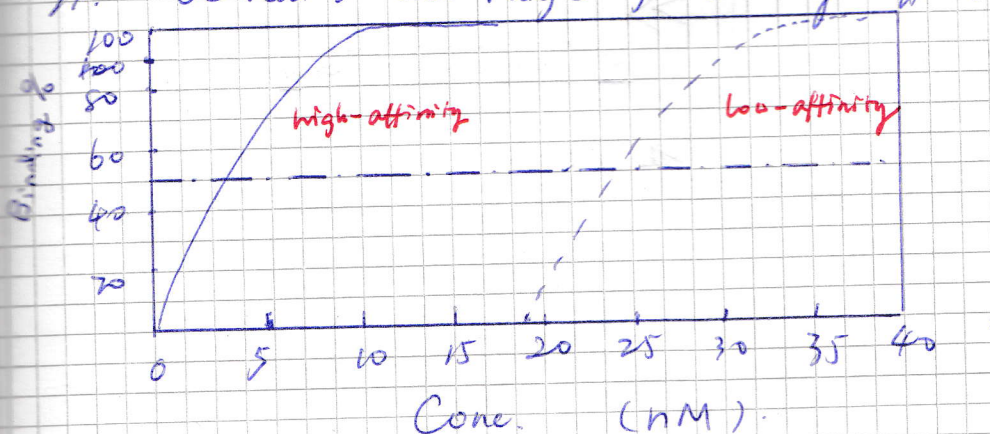
$$\text{let } y = \frac{1}{Y} \quad x = \frac{1}{X}$$

$$y = 470.39x + 0.841 \quad R^2 = 0.9336 \rightarrow \text{for HB consensus}$$

$$B_{max} = 1.189$$

$$K_D = 559.322 (RFU) \approx 19.85 \text{ nM}$$

// What's the range of binding affinities



For hb 18'

$$y = 27979x + 11961$$

$$B_{max} = 0.0836$$

If use consensus  $B_{max}$

$$K_D = 2339.186 \times 0.0355 = 83.041 \text{ nM}$$

$$\text{Another hb seq: } K_D = 4807.2 \approx 170.656 \text{ nM}$$

Get pretty low binding for bcd.