

Physicochemical Effects on White Wine Quality

STAT632 Section1 Final Project

Hui Huang(hy9489)

Yun Jing(qi2679)

Xuan Zhou(up5758)

1.Introduction

White wine is a wine that is fermented without skin contact. It is produced by the alcoholic fermentation of the non-colored pulp of grapes. It has been existing for at least 2500 years in the world. With the development of technology and the high demand of the white wine all around the world, the white wine industry needs a standard quality control system to test the white wine quality.

In this report, we use the Portuguese “Vinho verde” white wine data set to do the multiple linear regression. Our goal is to find the suitable MLR model to test “Vinho verde” white wine quality using physicochemical factors. The analysis will reveal the important relationship between “Vinho verde” white wine with different physicochemical factors. In order to make sure the model is suitable; we also do the Cross Validation to validate our regression model. Physicochemical factors are very professional laboratory indicator, such as different kinds of acidity, PH value and density. The wine producers and suppliers can use this final model to test the wine quality much easier with standard lab test results. The standardized wine quality control model can also benefit customers when they choose the white wine.

2.Data Description

2.1 Data source

The data sets are public available in <https://www.kaggle.com/danielpanizzo/wine-quality>. We also collect useful information from <http://www.vinhoverde.pt/en/about-vinho-verde> official website. There are two data sets, using red and white wine samples in 2009. We choose the white wine data sets to analyze.

2.2 Data dimension

There are 4898 observations on 12 variables in the white wine data set. The data has no missing attribute value.

2.3 Response and predictor variables

The response variable is Quality which is based on sensory data (median of at least 3 evaluations made by wine experts). The score of each wine quality is between 0(very bad) to 10(very excellent). The quality of the white wine data is approximately normally distributed (see Figure1).

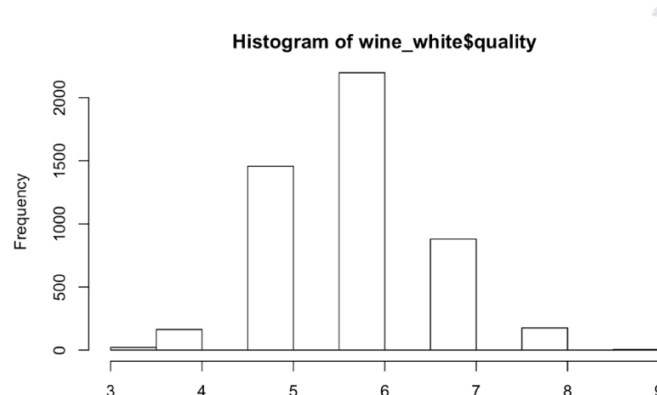


Figure1

The predictors are 11 physicochemical factors. The descriptions are: 1.fixed acidity: most acids involved with wine or fixed or nonvolatile (do not evaporate readily). 2.volatile acidity: the amount of acetic acid in wine, which at too high of levels can lead to an unpleasant, vinegar taste. 3.citric acid: found in small quantities, citric acid can add 'freshness' and flavor to wines. 4.residual sugar: the amount of sugar remaining after fermentation stops, it's rare to find wines with less than 1 gram/liter and wines with greater than 45 grams/liter are considered sweet. 5.chlorides: the amount of salt in the wine. 6.free sulfur dioxide: the free form of SO₂ exists in equilibrium between molecular SO₂ (as a dissolved gas) and bisulfite ion; it prevents microbial growth and the oxidation of wine. 7.total sulfur dioxide: amount of free and bound forms of SO₂; in low concentrations, SO₂ is mostly undetectable in wine, but at free SO₂ concentrations over 50 ppm, SO₂ becomes evident in the nose and taste of wine. 8.density: the density of water is close to that of water depending on the percent alcohol and sugar content. 9.pH: describes how acidic or basic a wine is on a scale from 0 (very acidic) to 14 (very basic); most wines are between 3-4 on the pH scale. 10.sulphates: a wine additive which can contribute to sulfur dioxide gas (SO₂) levels, which acts as an antimicrobial and antioxidant. 11.alcohol: the percent alcohol content of the wine.

2.4 Summary Statistics and graphical displays of data set

fixed.acidity	volatile.acidity	citric.acid	residual.sugar
Min. : 3.800	Min. : 0.0800	Min. : 0.0000	Min. : 0.600
1st Qu.: 6.300	1st Qu.: 0.2100	1st Qu.: 0.2700	1st Qu.: 1.700
Median : 6.800	Median : 0.2600	Median : 0.3200	Median : 5.200
Mean : 6.855	Mean : 0.2782	Mean : 0.3342	Mean : 6.391
3rd Qu.: 7.300	3rd Qu.: 0.3200	3rd Qu.: 0.3900	3rd Qu.: 9.900
Max. : 14.200	Max. : 1.1000	Max. : 1.6600	Max. : 65.800
chlorides	free.sulfur.dioxide	total.sulfur.dioxide	density
Min. : 0.00900	Min. : 2.00	Min. : 9.0	Min. : 0.9871
1st Qu.: 0.03600	1st Qu.: 23.00	1st Qu.: 108.0	1st Qu.: 0.9917
Median : 0.04300	Median : 34.00	Median : 134.0	Median : 0.9937
Mean : 0.04577	Mean : 35.31	Mean : 138.4	Mean : 0.9940
3rd Qu.: 0.05000	3rd Qu.: 46.00	3rd Qu.: 167.0	3rd Qu.: 0.9961
Max. : 0.34600	Max. : 289.00	Max. : 440.0	Max. : 1.0390
pH	sulphates	alcohol	quality
Min. : 2.720	Min. : 0.2200	Min. : 8.00	Min. : 3.000
1st Qu.: 3.090	1st Qu.: 0.4100	1st Qu.: 9.50	1st Qu.: 5.000
Median : 3.180	Median : 0.4700	Median : 10.40	Median : 6.000
Mean : 3.188	Mean : 0.4898	Mean : 10.51	Mean : 5.878
3rd Qu.: 3.280	3rd Qu.: 0.5500	3rd Qu.: 11.40	3rd Qu.: 6.000
Max. : 3.820	Max. : 1.0800	Max. : 14.20	Max. : 9.000

Figure 2

From the Figure 2, the median and mean values are pretty similar for all variables. The first and third quantile are relatively close to the center. We surmise the distributions might be symmetrical and concentrated.

From the data matrix, we can tell there are some relationships between response and predictors. However, due to the data set is large and we have more than 10 predictors it is very hard to see the clear relationships (We do not include the matrix figure here). We need to do further investigation.

3. Methods

For this project, we focus on multiple linear regression model. The methods used in this project include: variable selection, assumptions check, leverage and outliers check, Box-Cox transformation, and cross-validation.

3.1 Original Full Model

We fit a multiple linear regression model with quality as the response and all the other attributes as the predictors, which is the original full model. Then we use variable selection methods to select the best model for our research questions.

3.2 Model Selection

The purpose of the model selection is to avoid that the model with too many predictors. “Over-fit” model performs poorly when making future predictions. In this step, we use four approaches: the Akaike information criterion (AIC), Bayesian information criterion (BIC), adjusted R^2 , and backwards stepwise selection. Since different methods might select different predictors, we used all four approaches to check and guarantee that we can select a best model. The result of the four criteria is shown on the results section. After model selection, we got a reduced model with eight predictors: Fixed.acidity, Volatile.acidity, Residual.sugar, Free.sulfur. Dioxide, Density, pH, Sulphates, and Alcohol.

3.3 Assumptions Check

After we select the most suitable model, we need to use some diagnostic techniques to check assumptions: constant variance, linearity, and normality. Regression diagnostics can also suggest improvements of model, which means model building is an iterative and interactive process. The diagnostic results for our reduced model indicate that we need to do transformation to get a better model to satisfy the assumptions. Diagnostic techniques also help us to check leverage points and outliers.

3.4 Box-Cox Transformation

The assumptions are not satisfied, we use Box-Cox method to estimate transformations. For this data, the quality is an integral variable from 0 to 10, in this case, we do not need to transform response variable. We only consider to transform predictors based on the scatter matrices plot and the summary(powerTransform()) results. After transformation, we also need to check assumptions (residuals vs. fitted value plot and QQ-plot) using our final model. We also compare the adjusted R^2 , and AIC of the reduced model and the final model.

3.5 Cross-validation and Accuracy

Finally, we use cross-validation to validate our model. We split the data into 70% training set and 30% testing set. Cross-validation is a more direct approach is to estimate the test error by holding out a subset of observations from the model fitting process, and then applying the statistical model to make predictions on those withheld observations. Based on the validation set approach, we also fitted ordinary least squares model, backwards stepwise model, ridge model, and Lasso model and compared their RMSE to get the model with best predictive performance.

4. Results

4.1 Models

Original full model: $quality = \beta_0 + \beta_1 fixed.acidity + \beta_2 volatile.acidity + \beta_3 citric.acid + \beta_4 residual.sugar + \beta_5 chlorides + \beta_6 free.sulfur.dioxide + \beta_7 total.sulfur.dioxide + \beta_8 density + \beta_9 pH + \beta_{10} sulphates + \beta_{11} alcohol + e$

Reduced model: $quality = \beta_0 + \beta_1 fixed.acidity + \beta_2 volatile.acidity + \beta_3 residual.sugar + \beta_4 free.sulfur.dioxide + \beta_5 density + \beta_6 pH + \beta_7 sulphates + \beta_8 alcohol + e$

Final model: $quality = \beta_0 + \beta_1 \log(fixed.acidity) + \beta_2 \log(volatile.acidity) + \beta_3 \sqrt{residual.sugar} + \beta_4 \sqrt{free.sulfur.dioxide} + \beta_5 density + \beta_6 1/\sqrt{pH} + \beta_7 \log(sulphates) + \beta_8 \sqrt{alcohol} + e$

4.2 Model Selection

The Figure 3 shows the adjusted R^2 , AIC, and BIC versus the number of predictors for the best subset procedure. The adjusted R^2 , AIC and BIC all select a model with 8 predictors.

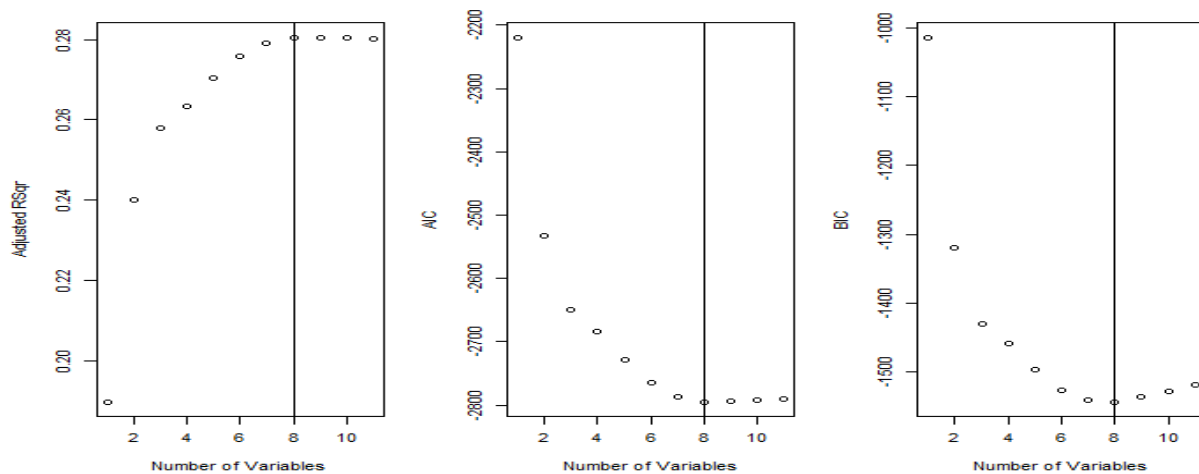


Figure 3

The coefficients result below (Table 1) are the predictors for the best model by using the adjusted R^2 , AIC, and BIC, which shows the same 8 predictors.

```
round(regsub_summ$adjr2, 4)
## [1] 0.1896 0.2399 0.2581 0.2634 0.2703 0.2758 0.2791 0.2806 0.2805 0.2804
## [11] 0.2803

which.max(regsub_summ$adjr2)
## [1] 8

coef(regsub_fit, 8)
```

	(Intercept)	fixed.acidity	volatile.acidity
##	1.541062e+02	6.810394e-02	-1.888140e+00
	residual.sugar	free.sulfur.dioxide	density
##	8.284724e-02	3.349015e-03	-1.542913e+02
	pH	sulphates	alcohol
##	6.942135e-01	6.285081e-01	1.931628e-01

```
which.min(aic_vec)
## [1] 8

coef(regsub_fit, 8)
```

	(Intercept)	fixed.acidity	volatile.acidity
##	1.541062e+02	6.810394e-02	-1.888140e+00
	residual.sugar	free.sulfur.dioxide	density
##	8.284724e-02	3.349015e-03	-1.542913e+02
	pH	sulphates	alcohol
##	6.942135e-01	6.285081e-01	1.931628e-01

```
which.min(regsub_summ$bic)
## [1] 8

coef(regsub_fit, 8)
```

	(Intercept)	fixed.acidity	volatile.acidity
##	1.541062e+02	6.810394e-02	-1.888140e+00
	residual.sugar	free.sulfur.dioxide	density
##	8.284724e-02	3.349015e-03	-1.542913e+02
	pH	sulphates	alcohol
##	6.942135e-01	6.285081e-01	1.931628e-01

Table 1

The summary() results below (Table 2 is AIC, Table 3 is BIC) are the predictors for the best model by using backwards stepwise selection of AIC and BIC respectively, both which show the same 8 predictors.

```
summary(wine_sa)

##
## Call:
## lm(formula = quality ~ fixed.acidity + volatile.acidity + residual.sugar +
##     free.sulfur.dioxide + density + pH + sulphates + alcohol,
##     data = wine_white)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -3.8246 -0.4938 -0.0396  0.4660  3.1208
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.541e+02  1.810e+01  8.514 < 2e-16 ***
## fixed.acidity  6.810e-02  2.043e-02  3.333 0.000864 ***
## volatile.acidity -1.888e+00  1.095e-01 -17.242 < 2e-16 ***
## residual.sugar  8.285e-02  7.287e-03  11.370 < 2e-16 ***
## free.sulfur.dioxide 3.349e-03  6.766e-04  4.950 7.67e-07 ***
## density      -1.543e+02  1.834e+01 -8.411 < 2e-16 ***
## pH           6.942e-01  1.034e-01  6.717 2.07e-11 ***
## sulphates     6.285e-01  9.997e-02  6.287 3.52e-10 ***
## alcohol       1.932e-01  2.408e-02  8.021 1.31e-15 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.7512 on 4889 degrees of freedom
## Multiple R-squared:  0.2818, Adjusted R-squared:  0.2806
## F-statistic: 239.7 on 8 and 4889 DF, p-value: < 2.2e-16
```

Table2

```
summary(wine_sb)

##
## Call:
## lm(formula = quality ~ fixed.acidity + volatile.acidity + residual.sugar +
##     free.sulfur.dioxide + density + pH + sulphates + alcohol,
##     data = wine_white)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -3.8246 -0.4938 -0.0396  0.4660  3.1208
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.541e+02  1.810e+01  8.514 < 2e-16 ***
## fixed.acidity  6.810e-02  2.043e-02  3.333 0.000864 ***
## volatile.acidity -1.888e+00  1.095e-01 -17.242 < 2e-16 ***
## residual.sugar  8.285e-02  7.287e-03  11.370 < 2e-16 ***
## free.sulfur.dioxide 3.349e-03  6.766e-04  4.950 7.67e-07 ***
## density      -1.543e+02  1.834e+01 -8.411 < 2e-16 ***
## pH           6.942e-01  1.034e-01  6.717 2.07e-11 ***
## sulphates     6.285e-01  9.997e-02  6.287 3.52e-10 ***
## alcohol       1.932e-01  2.408e-02  8.021 1.31e-15 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.7512 on 4889 degrees of freedom
## Multiple R-squared:  0.2818, Adjusted R-squared:  0.2806
## F-statistic: 239.7 on 8 and 4889 DF, p-value: < 2.2e-16
```

Table3

4.3 Assumption Check

The Figure 4 below shows the residuals vs. fitted value plot and QQ-plot of the reduced model.

The Figure 5 below shows the residuals vs. fitted value plot and QQ-plot of the final model (after transformation).

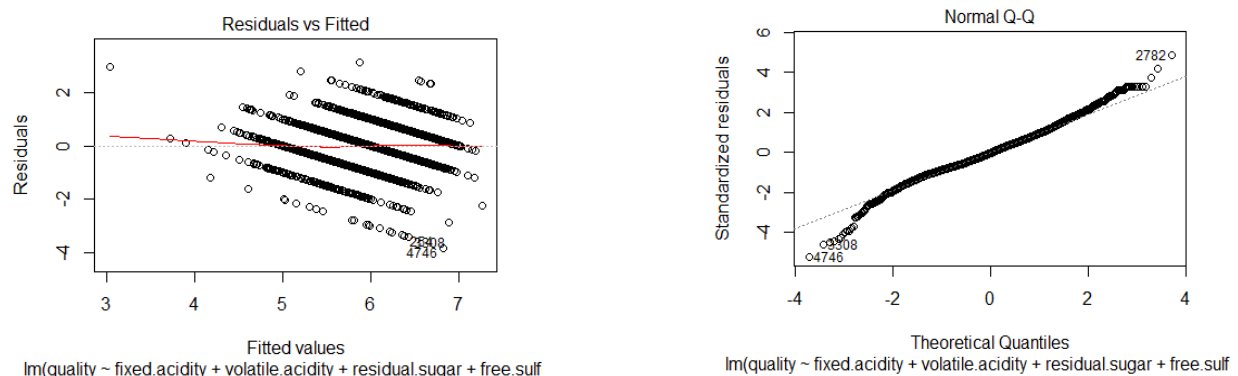


Figure 4

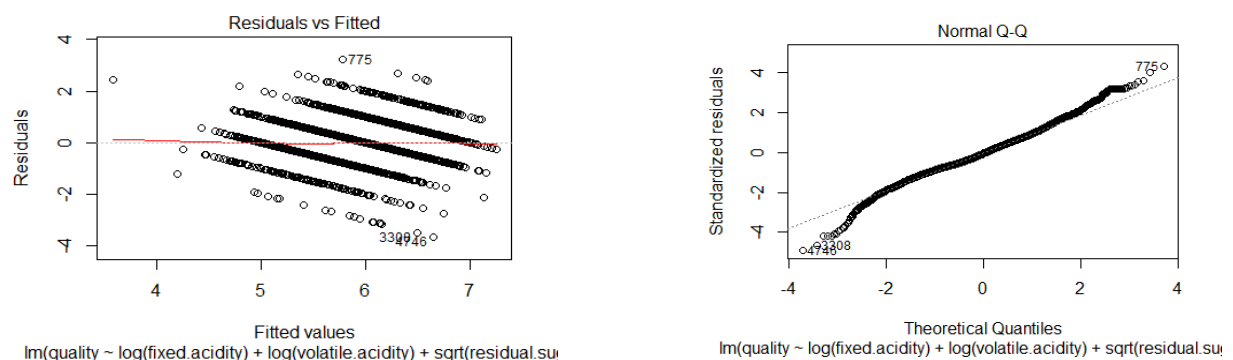


Figure 5

As shown on the Figure 4, the residual plot shows that the red line deviates significantly from the 0 line and the points scatter in a certain pattern, which indicate non-constant variance and non-linearity. The QQ-plot shows that a heavy tails distribution, but most points fall on a straight line. In this case, it might be reasonable to say the residuals follow a normal distribution.

As shown on the Figure 5, the residual plot shows the points scatter more randomly around 0 and the red line is close to 0 line, which indicates that the assumptions of constant variance and linearity are satisfied. The QQ-plot also shows that a heavy tails distribution, but most points still fall on a straight line. After the transformation, the assumptions are satisfied.

4.4 Leverage and Outliers Check

The Figure 6 are the standardized residuals vs. leverage plots. As the plots shown, there are many high leverage points and 4 outliers, 3 of outliers are bad leverage points.

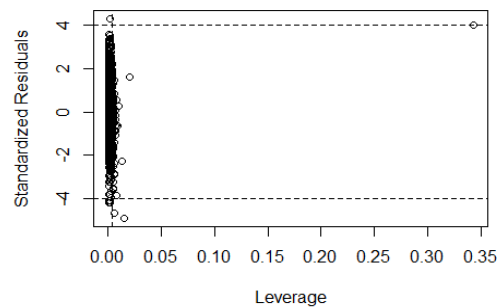


Figure 6

4.5 Box-Cox Transformation

The Figure 7 is the scatter matrices plot of reduced model.

The Table 4 is the result of Box-Cox method to estimate the rounded λ for transformation of predictors.

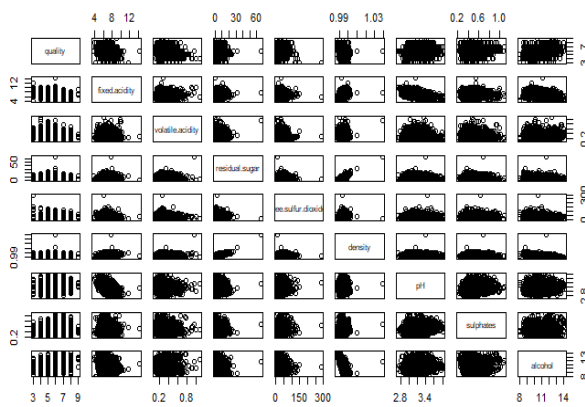


Figure 7

```
summary(powerTransform(cbind(fixed.acidity, volatile.acidity, residual.sugar,
free.sulfur.dioxide, density, pH, sulphates, alcohol) ~ 1, wine_white))
```

## bcPower Transformations to Multinormality				
##	Est Power	Rounded Pwr	Wald Lwr Bnd	Wald Up Bnd
## fixed.acidity	0.0835	0.00	-0.0391	0.2061
## volatile.acidity	-0.1187	-0.12	-0.1816	-0.0558
## residual.sugar	0.5904	0.59	0.5662	0.6146
## free.sulfur.dioxide	0.4816	0.50	0.4428	0.5204
## density	-49.6448	-49.64	-52.4013	-46.8883
## pH	-0.4820	-0.50	-0.8857	-0.0784
## sulphates	-0.3478	-0.33	-0.4476	-0.2481
## alcohol	0.6062	0.50	0.4575	0.7548
## Likelihood ratio test that transformation parameters are equal to 0				
## (all log transformations)				
## LR test, lambda = (0 0 0 0 0 0 0)	5681.753	8	< 2.22e-16	
## Likelihood ratio test that no transformations are needed				
## LR test, lambda = (1 1 1 1 1 1 1)	5725.955	8	< 2.22e-16	

Table 4

As shown on results above, the rounded λ of predictor “density” is -49.64. Since there is usually little justification for making extreme transformations, we do not consider transforming “density”. Thus, the final model can be get through transformation: $quality = \beta_0 + \beta_1 \log(fixed.acidity) +$

$$\beta_2 \log(\text{volatile.acidity}) + \beta_3 \sqrt{\text{residual.sugar}} + \beta_4 \sqrt{\text{free.sulfur.dioxide}} + \beta_5 \text{density} + \beta_6 1/\sqrt{\text{pH}} + \beta_7 \log(\text{sulphates}) + \beta_8 \sqrt{\text{alcohol}} + e.$$

The Table 5 is the comparison of the adjusted R^2 , and AIC of the reduced model and the final model.

	Adjusted R^2	AIC
Reduced model	0.2805767	11108.29
Final model	0.283624	11086.5

Table 5

As shown on the Table 5, the Final model has a smaller AIC (11086.5) than the reduced model and has a larger adjusted R^2 (0.283624). These means that the transformation is helpful to improve the performance of the model.

4.6 Cross-validation

The Table 6 is the comparison of the RMSE of ordinary least squares model of reduced model (OLS_ori) and final model (OLS_final), backwards stepwise model (OLS_step), ridge model (Ridge), and Lasso model (Lasso).

Model	RMSE
OLS_ori	0.7243308
OLS_final	0.7226316
OLS_step	0.7243308
Ridge	0.7410161
Lasso	0.7298092

Table 6

As shown on the Table 6, the RMSE of OLS_final is smallest, which indicates that the accuracy of the OLS_final model. that is the best final model.

6. Discussion

6.1 Discussion summary

From the analysis, we figure out not all 11 predictors in the original data set are useful to the model. After log and sqrt transformation for some predictors, the model satisfies assumptions of MLR.

From summary (lm_wine2) (Table 7), 7 predictors in the final model are significant. The log (fixed. Acidity) is not

significant. We can see that coefficient has relationship with physicochemical factors either negative or positive. From the analysis, we also know not all 11 predictors in the original data set are useful to the

```
Call:
lm(formula = quality ~ log(fixed.acidity) + log(volatile.acidity) +
    sqrt(residual.sugar) + sqrt(free.sulfur.dioxide) + density +
    (1/sqrt(pH)) + log(sulphates) + sqrt(alcohol), data = wine_white)

Residuals:
    Min       1Q   Median       3Q      Max
-3.6470 -0.4888 -0.0382  0.4653  3.2236

Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept)  68.932851   12.267512   5.619 2.03e-08 ***
log(fixed.acidity) -0.136971   0.100982  -1.356   0.175
log(volatile.acidity) -0.608466   0.032325 -18.823 < 2e-16 ***
sqrt(residual.sugar)  0.248908   0.024339  10.227 < 2e-16 ***
sqrt(free.sulfur.dioxide) 0.066818   0.008079   8.271 < 2e-16 ***
density      -71.079757   12.142360  -5.854 5.12e-09 ***
log(sulphates)  0.280385   0.050279   5.577 2.58e-08 ***
sqrt(alcohol)   1.944142   0.117707  16.517 < 2e-16 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.7496 on 4890 degrees of freedom
Multiple R-squared:  0.2846,    Adjusted R-squared:  0.2836
F-statistic: 278 on 7 and 4890 DF,  p-value: < 2.2e-16
```

Table7

model. In the future, we can use the model to predict white wine quality using these 9 physicochemical factors.

6.2 Discussion weakness

we have three weakness points. Firstly, we do not check multicollinearity. The predictors may correlate with each other. Secondly, since we do box-cox transformation, it become hard to interpret the result. Thirdly, we believe there are more factors have effects on white wine quality like climate, temperature and location, etc. We might include some other crucial factors in our model in the future.

7. Reference

Data resource come from website below:

Daniel S. Panizzo. (2017). Wine Quality: Modeling wine preferences by data mining from physicochemical properties, <https://www.kaggle.com/danielpanizzo/red-and-whitewine-quality>.
P. Cortez, A. Cerdeira, F. Almeida, T. Matos and J. Reis. (2009). Modeling wine preferences by data mining from physicochemical properties. In Decision Support Systems, Elsevier, 47(4):547-553. ISSN: 01679236.

Method applied from:

Eric Fox. (2019). Linear and Logistic Regression lecture materials and notes, California State University East Bay.

Code Appendix

```
library(tidyverse)
library(ggplot2)
library(glmnet)

##Input data
wine_white <- read.csv(file = "../Data/wineQualityWhites.csv", header =
TRUE)
head(wine_white)
str(wine_white)

##Checking missing values
library(Amelia)
missmap(wine_white, main = "Missing values vs observed")

sapply(wine_white, function(x) sum(is.na(x)))

##Processing data
wine_white <- wine_white[,-1]
wine_white$quality <- as.numeric(wine_white$quality)
glimpse(wine_white)
```

```

##Data Description
###Dimension
dim(wine_white)

###Attributes
names(wine_white)

###Data displays
hist(wine_white$quality)

summary(wine_white)

###Scatter matrices plot of original full model
pairs(quality ~ ., data = wine_white)

##Model selection

###R squared
library(leaps)
regsub_fit <- regsubsets(quality ~ ., data = wine_white, nvmax=11)
regsub_summ <- summary(regsub_fit)

attributes(regsub_summ)

round(regsub_summ$rsq, 4)

round(regsub_summ$adjr2, 4)

which.max(regsub_summ$adjr2)

coef(regsub_fit, 8)

###Plot of R squared, AIC, BIC
n <- nrow(wine_white)
aic_vec <- n*log(regsub_summ$rss/n) + 2*c(1:11)

par(mfrow=c(1,3), mar=c(4.5, 4.5, 1, 1))
####R squared
plot(c(1:11), regsub_summ$adjr2, xlab="Number of Variables",
ylab="Adjusted RSqr")
abline(v=which.max(regsub_summ$adjr2))

####AIC
plot(c(1:11), aic_vec, xlab="Number of Variables", ylab="AIC")
abline(v=which.min(aic_vec))

####BIC
plot(c(1:11), regsub_summ$bic, xlab="Number of Variables", ylab="BIC")
abline(v=which.min(regsub_summ$bic))

###Predictors selected

```

```

which.min(aic_vec)
coef(regsub_fit, 8)

which.min(regsub_summ$bic)
coef(regsub_fit, 8)

###Backwards stepwise selection of AIC
wine_full <- lm(quality ~ ., data = wine_white)
wine_sa <- step(wine_full)
summary(wine_sa)

###Backwards stepwise selection of BIC
wine_sb <- step(wine_full, k=log(n))
summary(wine_sb)

###Reduced model
lm_wine1 <- lm(quality ~ fixed.acidity + volatile.acidity + residual.sugar
+ free.sulfur.dioxide + density + pH + sulphates + alcohol, data =
wine_white)
summary(lm_wine1)

###Scatter matrices plot of reduced model
pairs(quality ~ fixed.acidity + volatile.acidity + residual.sugar +
free.sulfur.dioxide + density + pH + sulphates + alcohol, data =
wine_white)

###Test relationship
wine_null <- lm(quality ~ 1, data = wine_white)
anova(wine_null ,lm_wine1)

###Check assumptions for reduced model
####QQ-plot
plot(lm_wine1, which = 2)

####Residual plot
plot(lm_wine1, which = 1)

##Box-cox Transformation

###Estimate transformations

library(car)
summary(powerTransform(cbind(fixed.acidity, volatile.acidity,
residual.sugar, free.sulfur.dioxide, density, pH, sulphates, alcohol) ~ 1,
wine_white))

###Final model after transformations
lm_wine2 <- lm(quality ~ log(fixed.acidity) + log(volatile.acidity) +
sqrt(residual.sugar) + sqrt(free.sulfur.dioxide) + density + 1/sqrt(pH) +
log(sulphates) + sqrt(alcohol), data = wine_white)
summary(lm_wine2)

###Check and compare AIC and adjusted R squared

```

```

AIC(lm_wine1)

AIC(lm_wine2)

summary(lm_wine1)$adj.r.squared

summary(lm_wine2)$adj.r.squared

###Scatter matrices plot of final model
pairs(quality ~ log(fixed.acidity) + log(volatile.acidity) +
sqrt(residual.sugar) + sqrt(free.sulfur.dioxide) + density + 1/sqrt(pH) +
log(sulphates) + sqrt(alcobol), data = wine_white)

###Check assumptions final model
####QQ-plot
plot(lm_wine2, which = 1)

####Residual plot
plot(lm_wine2, which = 2)

###Compare the plots of the observed versus predicted values
par(mfrow=c(1,2), mar=c(2.5, 2.5, 2, 2))

plot(predict(lm_wine1), wine_white$quality, xlab="Fitted Values",
ylab="quality")
lines(lowess(predict(lm_wine1), wine_white$quality), col='red')
abline(0,1)

plot(predict(lm_wine2), wine_white$quality, xlab="Fitted Values",
ylab="quality")
lines(lowess(predict(lm_wine2), wine_white$quality), col='red')
abline(0,1)

###Identify leverage points and outliers
plot(lm_wine2, which = 5)

p <- 8
n <- nrow(wine_white)
plot(hatvalues(lm_wine2), rstandard(lm_wine2), xlab= 'Leverage' , ylab=
'Standardized Residuals')
abline(h = c(-4,4), v = 2*(p+1)/n, lty=2)

##Cross-validation
###Split data into 70% for train and 30% for test
set.seed(99)

wine <- model.matrix(quality ~ ., data=wine_white)[, -12]
q <- wine_white$quality

train_idx <- sample(n, size = floor(0.7 * n))

wine_train <- wine[train_idx, ]
nrow(wine_train)

```

```

wine_test <- wine[-train_idx, ]
nrow(wine_test)

q_train <- q[train_idx]

q_test <- q[-train_idx]

####Fit model based on cross validation
####Fit reduced model on training set
lm_wine3 <- lm(quality ~ fixed.acidity + volatile.acidity + residual.sugar
+ free.sulfur.dioxide + density + pH + sulphates + alcohol, data =
wine_white, subset = train_idx)

####Fit final model on training data
lm_wine4 <- lm(quality ~ log(fixed.acidity) + log(volatile.acidity) +
sqrt(residual.sugar) + sqrt(free.sulfur.dioxide) + density + 1/sqrt(pH) +
log(sulphates) + sqrt(alcohol), data = wine_white, subset = train_idx)

####Fit ordinary least squares model w/ stepwise selection on training set
lm_step_wine <- step(lm_wine4, trace=F)

####Fit ridge model on training set
ridge_wine <- cv.glmnet(wine_train, q_train, alpha=0)

####Fit lasso model on training set
lasso_wine <- cv.glmnet(wine_train, q_train, alpha=1)

###Compute RMSE
###Create function
compute_rmse <- function(y, y_pred) {
  n <- length(y)
  sqrt((1 / n) * sum((y - y_pred)^2))
}

####Reduced model
wine_pred1 <- predict(lm_wine3, newdata = wine_white[-train_idx, ])
rmse_ori <- compute_rmse(q_test, wine_pred1)

####Final model
wine_pred2 <- predict(lm_wine4, newdata = wine_white[-train_idx, ])
rmse_fin <- compute_rmse(q_test, wine_pred2)

####Step
wine_step_pred <- predict(lm_step_wine, newdata = wine_white[-
train_idx, ])
rmse_step <- compute_rmse(q_test, wine_step_pred)

####Ridge
wine_ridge_pred <- predict(ridge_wine, newx = wine_test, s = "lambda.min")
wine_ridge_pred <- as.numeric(wine_ridge_pred)
rmse_ridge <- compute_rmse(q_test, wine_ridge_pred)

####Lasso

```

```
wine_lasso_pred <- predict(lasso_wine, newx = wine_test, s = "lambda.min")
wine_lasso_pred <- as.numeric(wine_lasso_pred)
rmse_lasso <- compute_rmse(q_test, wine_lasso_pred)

###Compare RMSE
data.frame(Model = c('OLS_ori', 'OLS_final', 'OLS_step', 'Ridge',
  'Lasso' ), RMSE = c(rmse_ori, rmse_fin, rmse_step, rmse_ridge,
  rmse_lasso))
```