

Protocol to clone amiRNAs or syn-tasiRNAs in *BsaI/ccdB*-based ('B/c') vectors

1. Oligonucleotide annealing

-Dilute sense oligonucleotide and antisense oligonucleotide in sterile H₂O to a final concentration of 100 μ M.

-Prepare Oligo Annealing Buffer:

60 mM Tris-HCl (pH 7.5)
500 mM NaCl
60 mM MgCl₂
10 mM DTT

Note: Prepare 1 ml aliquots of Oligo Annealing Buffer and store at -20°C.

-Assemble the annealing reaction in a PCR tube as described below:

Forward oligonucleotide (100 μ M)	2 μ L
Reverse oligonucleotide (100 μ M)	2 μ L
<u>Oligo Annealing Buffer</u>	<u>46 μL</u>
Total volume	50 μ L

The final concentration of each oligonucleotide is 4 μ M.

-Use a thermocycler to heat the annealing reaction 5 min at 94°C and then cool down (0.05°C/sec) to 20°C.

-Dilute the annealed oligonucleotides just prior to assembling the digestion-ligation reaction as described below:

Annealed oligonucleotides	3 μ L
<u>dH₂O</u>	<u>37 μL</u>
Total volume	40 μ L

The final concentration of each oligonucleotide is 0.15 μ M.

Note: Do not store the diluted oligonucleotides.

2. Digestion-ligation reaction

-Assemble the digestion-ligation reaction as described below:

B/c vector (x ug/uL)	Y μ L (50 ng)
Diluted annealed oligonucleotides	1 μ L
10x T4 DNA ligase buffer	1 μ L
T4 DNA ligase (400 U/ μ L)	1 μ L
<i>Bsa</i> I (10U/ μ L, NEB)	1 μ L
dH ₂ O	to 10 μ L
Total volume	10 μ L

Note: Prepare a negative control reaction lacking *Bsa*I.

-Mix the reactions by pipetting. Incubate the reactions for 5 minutes at 37°C.

3. *E.coli* transformation and analysis of transformants

-Transform 1-5 ul of the digestion-ligation reaction into an *E. coli* strain that doesn't have *ccdB* resistance (e.g. DH10B, TOP10, ...) to do counter-selection.

-Pick two colonies/construct, grow LB-Kan (100 mg/ml) cultures and purify plasmids.

-Sequence 2 clones per construct with appropriate primers: M13-F (CCCAGTCACGACGTTGTAAAACGACGG) and M13-R (CAGAGCTGCCAGGAAACAGCTATGACC) for *pENTR*-based vectors; attB1 (ACAAGTTTGTACAAAAAAGCAGGCT) and attB2 (ACCACTTTGTACAAGAAAGCTGGGT) primers for *pMDC32B*-, *pMDC123SB*-, *pFK210B*- or *pH7WG2B*-based vectors).

Eudicot amiRNA vectors: *Bsal/ccdB*-based ('B/c') vectors for direct cloning of amiRNAs to use in eudicot species.

CaMV, *Cauliflower mosaic virus*; *nos*, nopaline synthase; *rbcS*, Rubisco small subunit.

Vector	Bacterial antibiotic resistance	Plant antibiotic resistance	GATEWAY use	Promoter	Terminator	Plant tested	species	Addgene number
<i>pENTR-AtMIR390a-B/c</i>	Kanamycin	-	Donor	-	-	-		51778
<i>pFK210B-AtMIR390a-B/c</i>	Spectinomycin	BASTA	-	<i>CaMV</i> 35S	<i>rbcS</i>	<i>A.thaliana</i>		51777
<i>pMDC123SB-AtMIR390a-B/c</i>	Kanamycin	BASTA	-	<i>CaMV</i> 2x35S	<i>nos</i>	<i>A. thaliana</i> <i>N. benthamiana</i>		51775
<i>pMDC32B-AtMIR390a-B/c</i>	Kanamycin Hygromycin	Hygromycin	-	<i>CaMV</i> 2x35S	<i>nos</i>	<i>A. thaliana</i> <i>N. benthamiana</i>		51776

Monocot amiRNA vectors: *OsMIR390-Bsal/ccdB* ('B/c') vectors for direct cloning of amiRNAs to use in monocot species.

CaMV, *Cauliflower mosaic virus*; *nos*, nopaline synthase; *Os*, *Oryza sativa*.

Vector	Bacterial antibiotic resistance	Plant antibiotic resistance	GATEWAY use	Promoter	Terminator	Plant tested	species	Addgene number*
<i>pENTR-OsMIR390-B/c</i>	Kanamycin	-	Donor	-	-	-		-
<i>pMDC123SB-OsMIR390-B/c</i>	Kanamycin	BASTA	-	<i>CaMV</i> 2x35S	<i>nos</i>	<i>N. benthamiana</i>		-
<i>pMDC32B-OsMIR390-B/c</i>	Kanamycin Hygromycin	Hygromycin	-	<i>CaMV</i> 2x35S	<i>nos</i>	<i>N. benthamiana</i> <i>B. distachyon</i>		-
<i>pH7WG2-OsMIR390-B/c</i>	Spectinomycin	Hygromycin	-	<i>Os Ubiquitin</i>	<i>CaMV</i>	<i>B. distachyon</i>		-

*These vectors are not yet available at Addgene. Please, contact us at administrator@carringtonlab.org to request any of them.

syn-tasiRNA vectors: *BsaI/ccdB*-based ('B/c') vectors for direct cloning of syn-tasiRNAs to use in *Arabidopsis thaliana* and closely related species*.

CaMV, *Cauliflower mosaic virus*; nos, nopaline synthase.

Vector	Bacterial antibiotic resistance	Plant antibiotic resistance	GATEWAY use	Promoter	Terminator	Plant tested	species	Addgene number
<i>pENTR-AtTAS1c-B/c</i>	Kanamycin	-	Donor	-	-	-		51774
<i>pMDC123SB-AtTAS1c-B/c</i>	Kanamycin	BASTA	-	<i>CaMV</i> 2x35S	<i>nos</i>	<i>A. thaliana</i> <i>N. benthamiana</i> *		51772
<i>pMDC32B-AtTAS1c-B/c</i>	Kanamycin Hygromycin	Hygromycin	-	<i>CaMV</i> 2x35S	<i>nos</i>	<i>A. thaliana</i> <i>N. benthamiana</i> *		51773

*As miR173 is not conserved in *N. benthamiana* and other species not closely related to *Arabidopsis thaliana*, a construct expressing miR173 has to be co-expressed with the syn-tasiRNA construct to trigger syn-tasiRNA biogenesis.