

# Statistics for Human Genetics

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November 22, 2022

# What can we do with Statistics ?

1. Estimation
2. Modelling
3. Hypothesis testing
4. Predicting

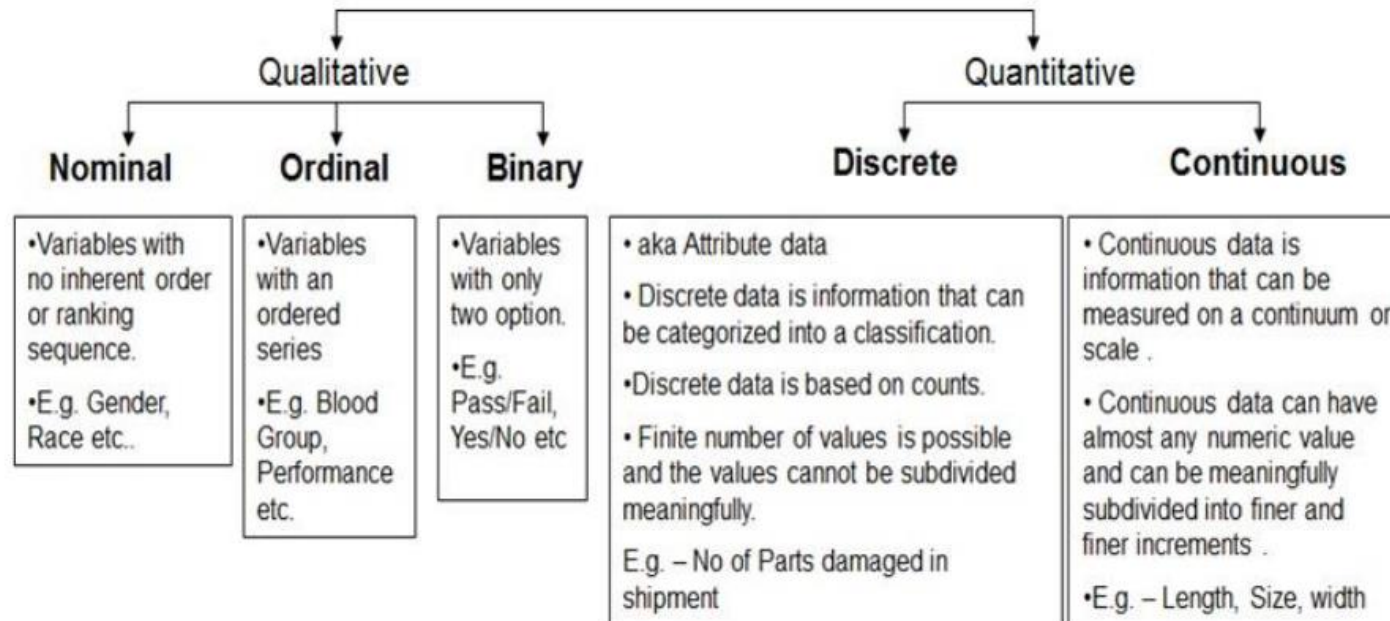
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# Random variables and estimation



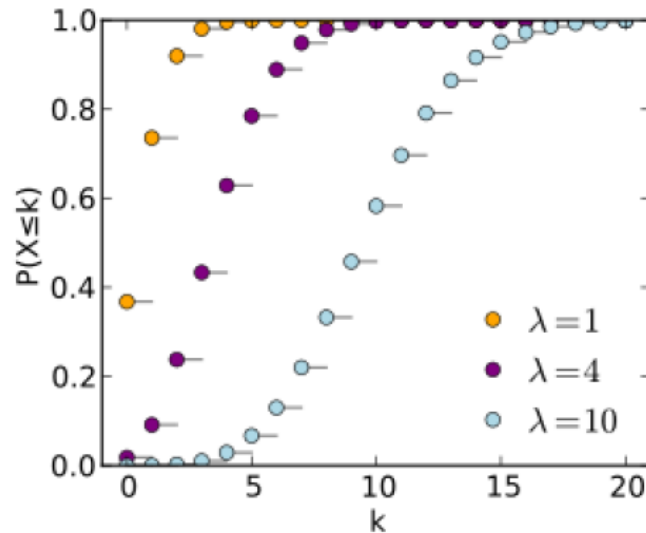
# Random Variables

- In statistics, we measure the realizations/observations of random variables
- Often, these random variables follow a distribution
- They can be qualitative or quantitative (continuous or discrete)



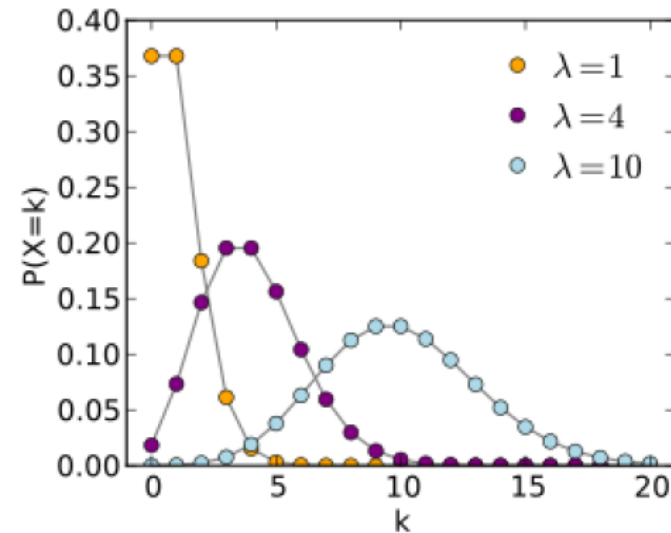
# Distributions

- Two ways to represent them:



## ***Cumulative distribution function (CDF)***

- $y = p(X \leq x)$
- Always growing
- Ideal way to represent but hard to read
- All distributions look the same

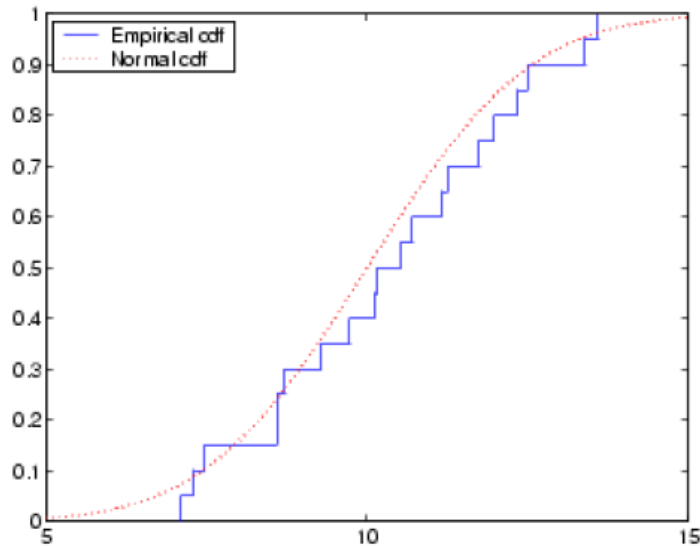


## ***Probability density function (PDF)***

- $y = p(X = x)$  for discrete
- Shows how values are distributed
- Nice visually but hard to deal with

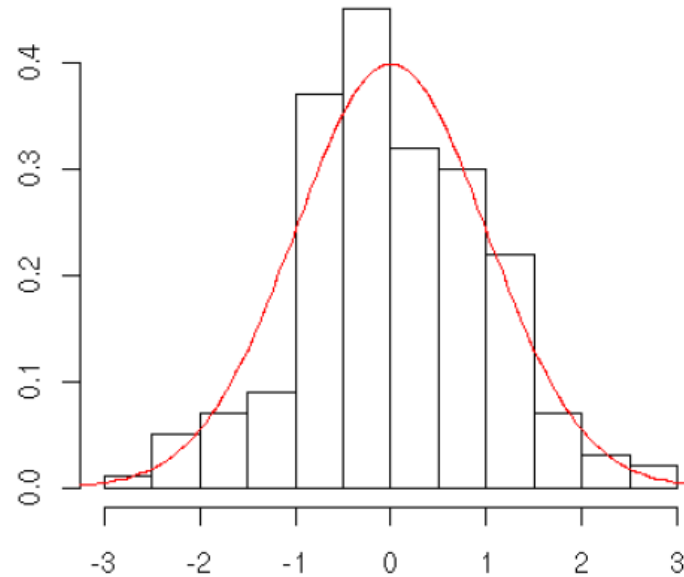
# Distributions

- How to estimate them:



***Empirical CDF***

- Rarely used
- Except when you want to compute empirical quantile functions



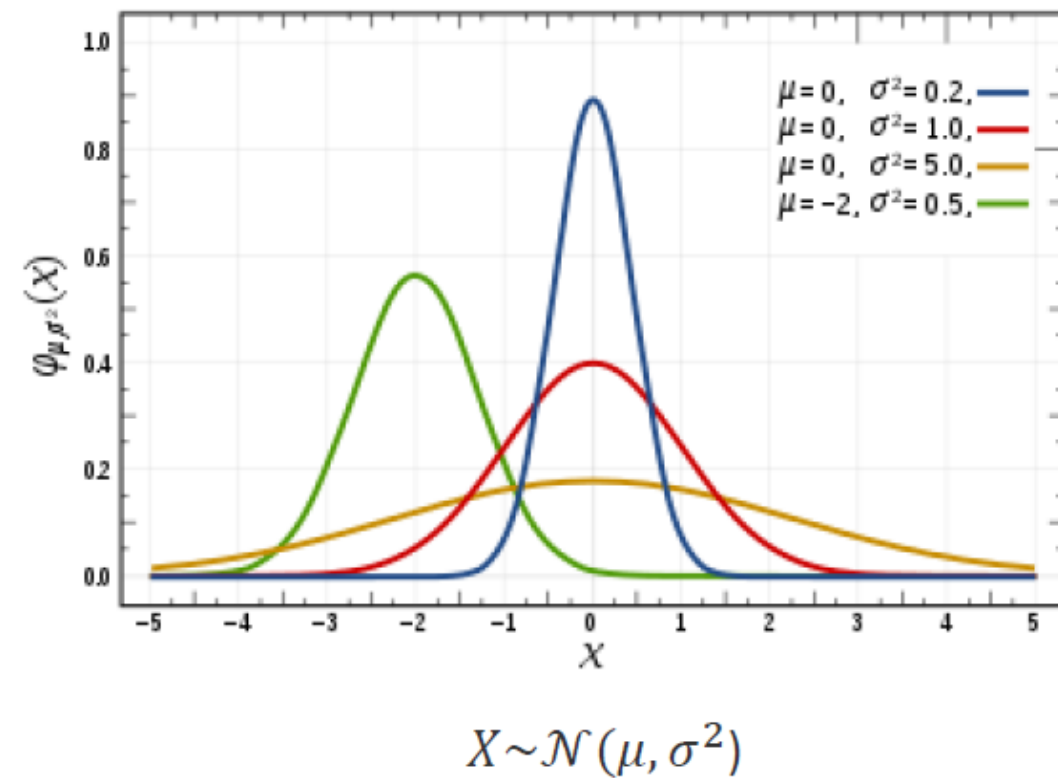
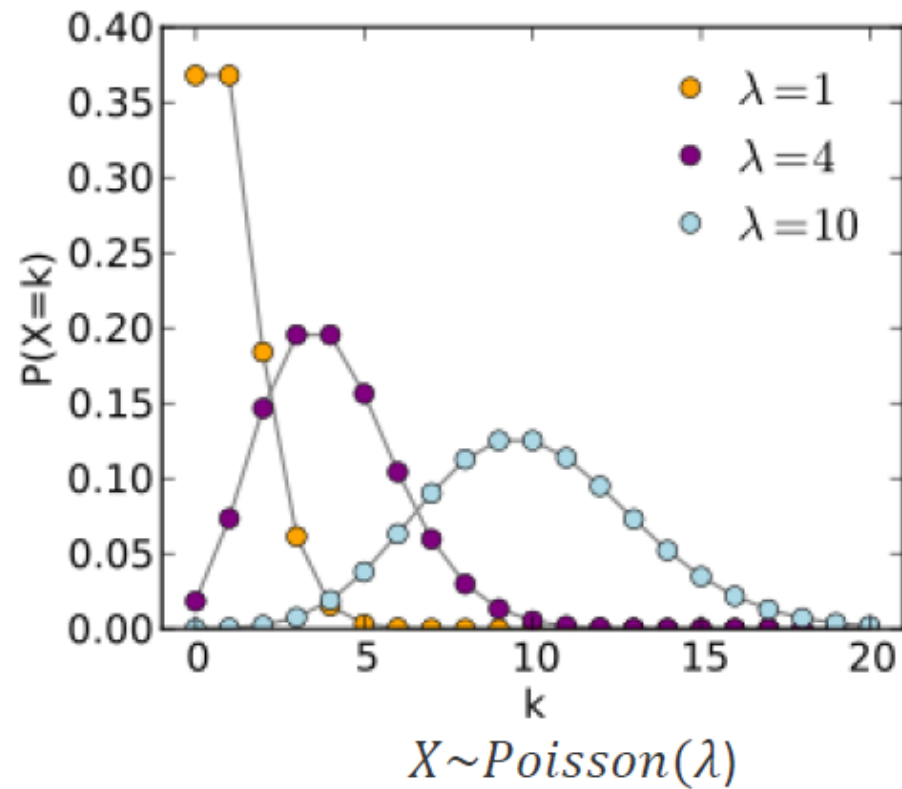
***Barplot (discrete)***

- For every value, count occurrences

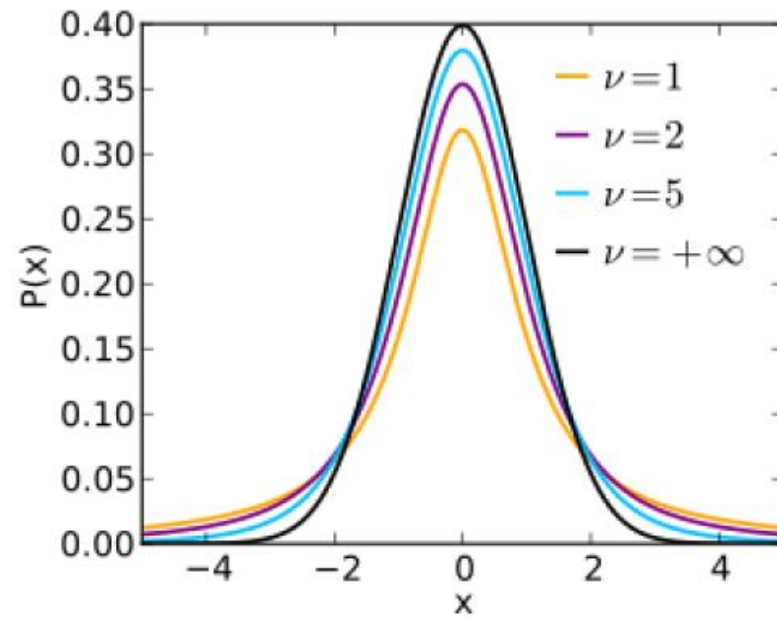
***Histogram (continuous)***

- Cut the interval into bins and count observations within bins

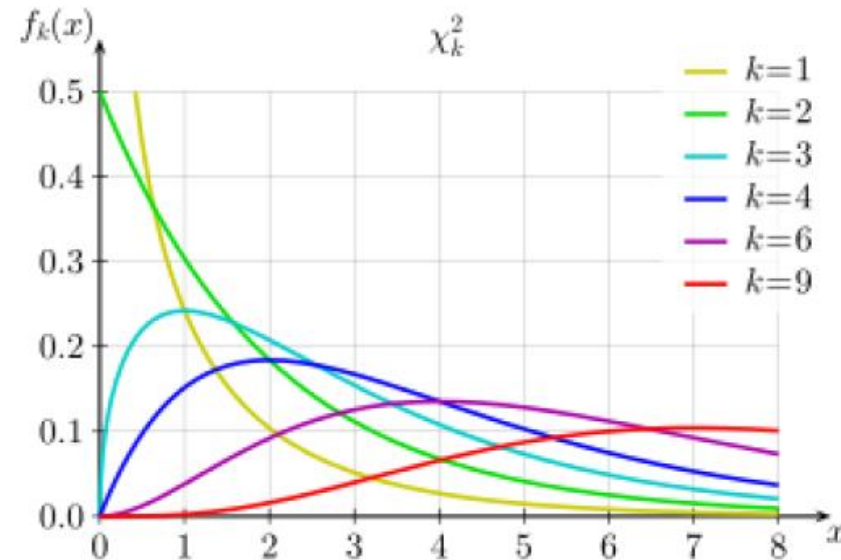
# Distributions – random variables (real world data)



# Distributions – tests statistics



$$X \sim T(\nu)$$



$$X \sim \chi^2(k)$$

$\lambda$ ,  $\mu$ ,  $\sigma$ ,  $\nu$  and  $k$  are the ideal, theoretical parameters  
→ Use estimations from the data to approximate them



# Statistics

- A statistics is a meaningful quantity derived from the data
- Often, estimators are realizations of distribution parameters
- Examples include mean, proportion, ...
- For simple distributions/parameters, there is a formula
- For more complex ones, we have to use other techniques (Monte-Carlo, Permutations, ...)

$$\hat{p} = \frac{x}{n}$$

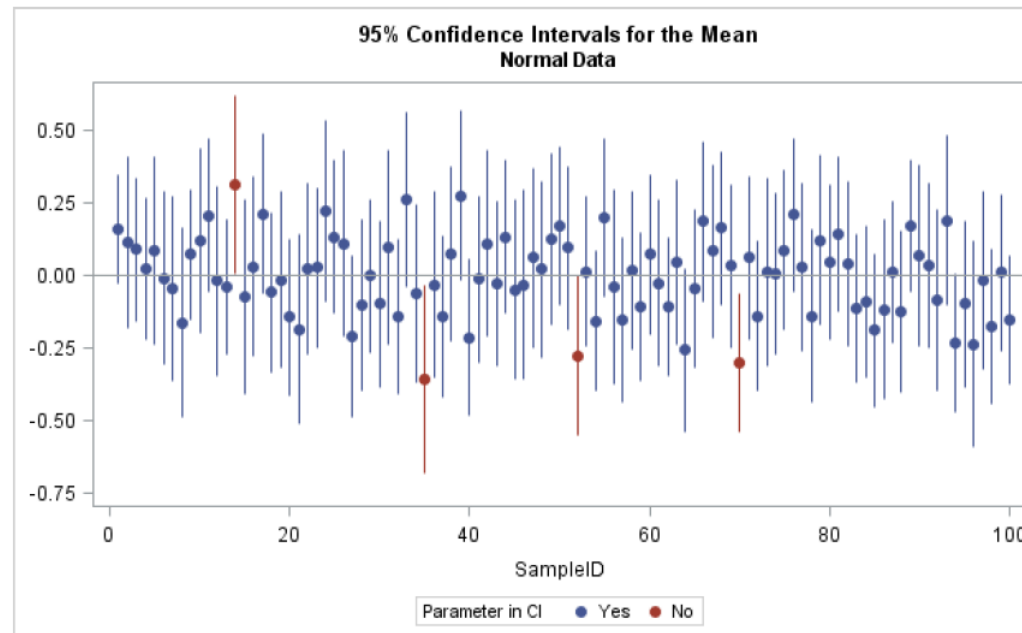
$$(\hat{\mu} =) \bar{x} = \frac{1}{n} \sum_{k=0}^n x_k$$

$$w = \frac{(\hat{\theta} - \theta_0)^2}{se(\hat{\theta})} \sim \mathcal{N}(0,1)$$

$$(\widehat{\sigma^2} =) s^2 = \frac{1}{N-1}$$

# Confidence intervals

- $x\%$  confidence interval ( $x\%C.I.$ ):
  - $x\%$  of the time when this interval is calculated, it will contain the true value of the parameter
  - The true value of the parameter has  $x\%$  chances to be in the  $x\%C.I.$
- Often 95% $C.I.$  used corresponding to the classical  $\alpha$  level



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# Modelling



# Modelling

- Estimate the effect of one variable on another variable

$$phenotype \sim \beta \times genotype + \epsilon$$

$$\begin{bmatrix} pheno_0 \\ \vdots \\ pheno_n \end{bmatrix} \quad \begin{bmatrix} A/T \\ \vdots \\ T/T \end{bmatrix} = \begin{bmatrix} 1 \\ \vdots \\ 2 \end{bmatrix}$$

$= \{0,1\}$  (case-control)  
 $\in \mathbb{R}$  (quantitative)  $\sim \mathcal{N}(0,1)$

$= \{0,1,2\}$  (genotype, directly typed)  
 $\in [0,2]$  (dosage, imputed)

$$\begin{bmatrix} 0.965 \\ \vdots \\ 1.816 \end{bmatrix}$$

- What is the effect of the genotype on the phenotype ?
  - $\beta$  estimations for continuous phenotypes
  - Odds Ratio (OR) for binary phenotypes. Ex: Case/Control studies

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## Modelling

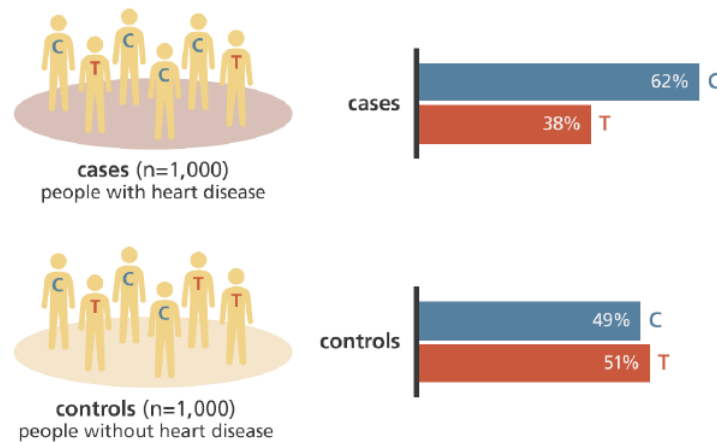
### 3.1 – Case/Control



# Case/control studies

- OR: *how much more likely are you to be a case if you carry the risk allele ?*

➤ Per genotype  $g$  and disease  $Y$ , we compute the odds  $O = \frac{p}{1-p} = \frac{p_{Y=1|g}}{1-p_{Y=1|g}}$



	Cases	Controls	N
T	380	510	890
C	620	490	1100

$$O_T = \frac{380/890}{510/890} \quad O_C = \frac{620/1100}{490/1100}$$

- OR: Ratio of the odds of the two alleles

➤ OR>1: the allele is 'deleterious'

➤ OR<1: the allele is 'protective'

$$OR_{C/T} = \frac{620 * 510}{490 * 380} = 1.70$$

# Case/control studies

## Dominant

Marker allele	Affected	Unaffected
DD+Dd	$n_{2A} + n_{1A}$	$n_{2U} + n_{1U}$
dd	$n_{0A}$	$n_{0U}$

$$OR = \frac{(2 \times n_{2A} + n_{1A}) \times (2 \times n_{0U} + n_{1U})}{(2 \times n_{0A} + n_{1A}) \times (2 \times n_{2U} + n_{1U})}$$

Allelic odds-ratio

## Recessive

Marker allele	Affected	Unaffected
DD	$n_{2A}$	$n_{2U}$
Dd+dd	$n_{1A} + n_{0A}$	$n_{1U} + n_{0U}$

$$OR = \frac{n_{affected\ carriers} \times n_{healthy\ non-carriers}}{n_{healthy\ carriers} \times n_{affected\ non-carriers}}$$

## Additive

Marker genotype	Affected	Unaffected
DD	$n_{2A}$	$n_{2U}$
Dd	$n_{1A}$	$n_{1U}$
dd	$n_{0A}$	$n_{0U}$

	Cases	Controls
T	380	510
C	620	490

$$OR_{C/T} = \frac{620 * 510}{380 * 490}$$

# Case/control studies

- Output: OR and 95% confidence interval of the OR
- Association test: is it significantly different from 1 ?
  - $H_0: OR = 1$
  - $H_1: OR \neq 1$
- Statistics: Fisher's exact test or Chi-squared
- In case of dosages or if covariates are included: logistic regression



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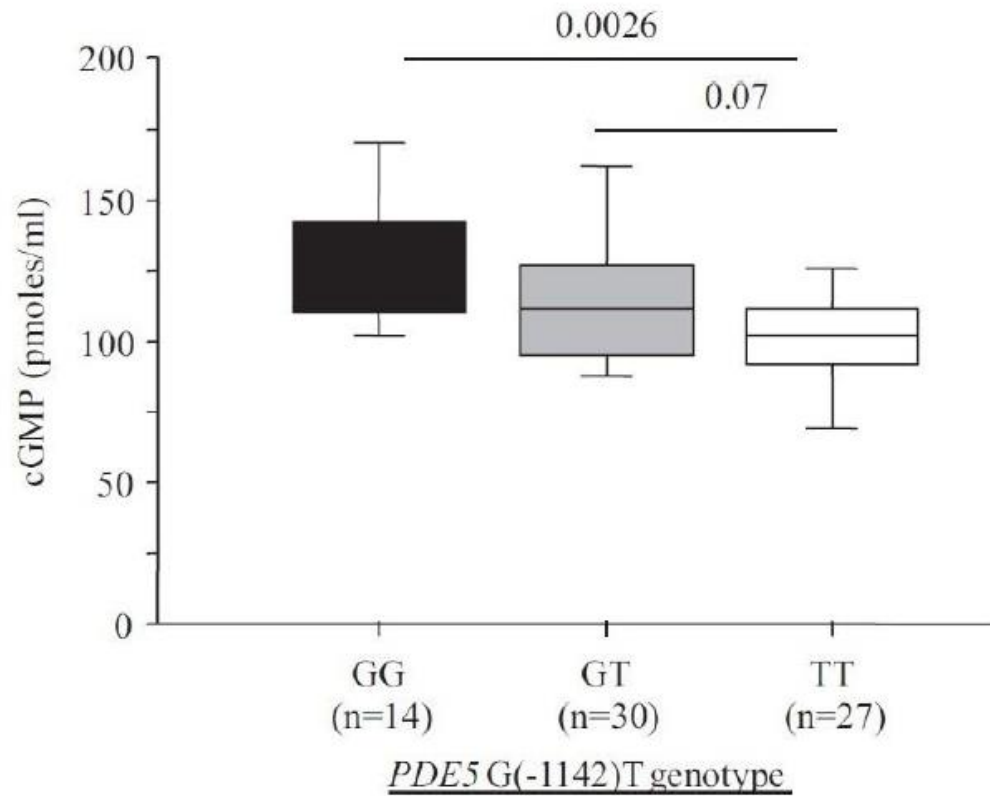
# Modelling

## 3.2 – Continuous traits

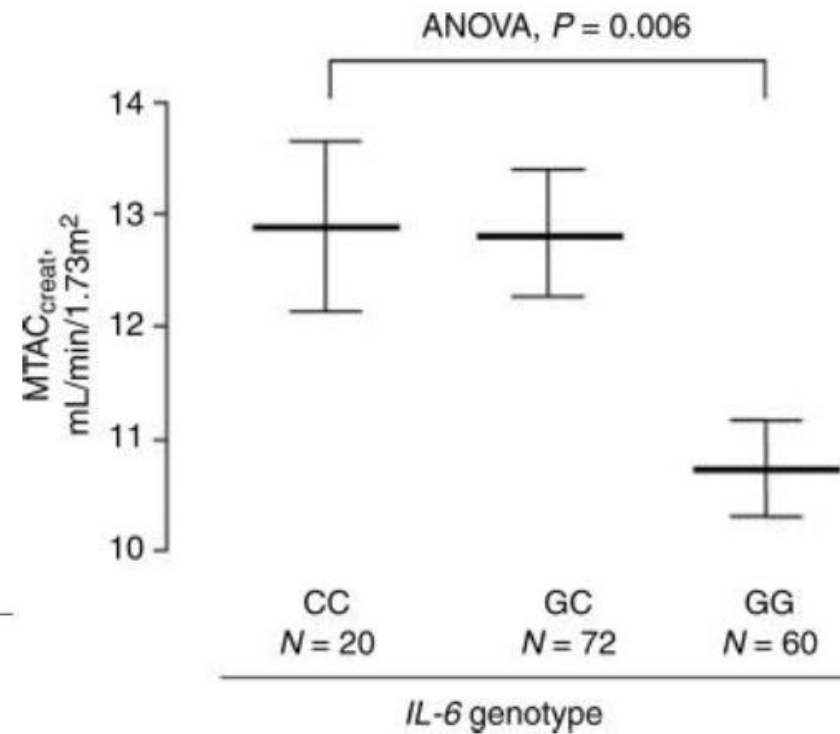


# Continuous traits

- If directly typed genotypes (0, 1, 2) are analyzed: ANOVA



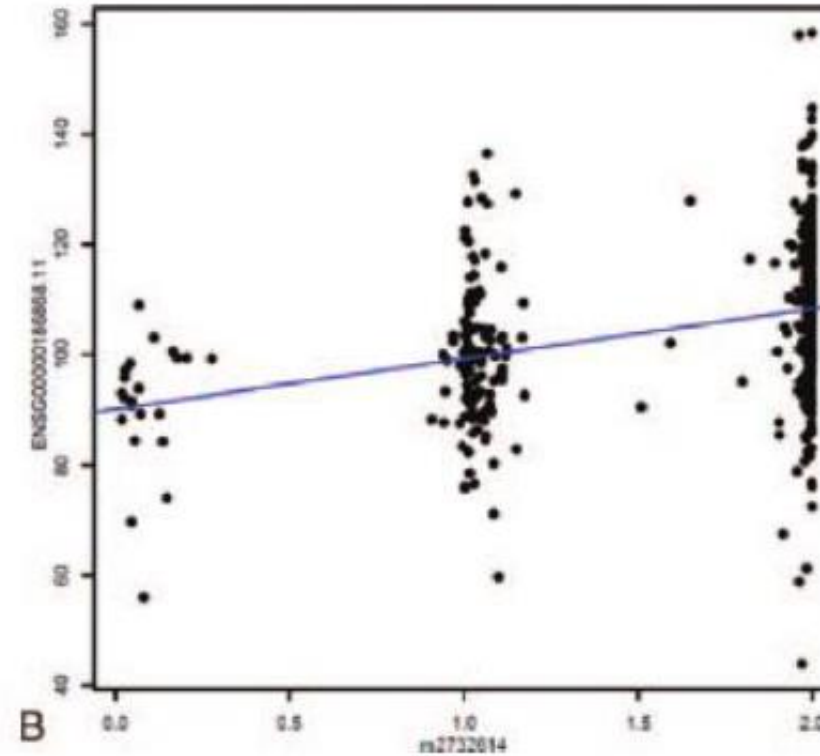
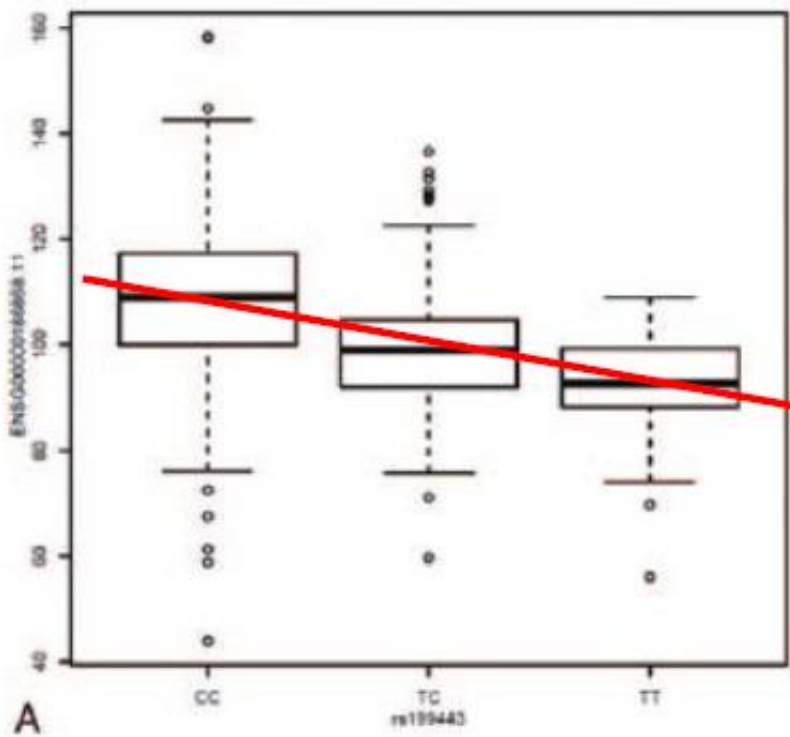
Additive



Recessive

# Continuous traits

- If dosages are analyzed (imputed quantity of minor allele  $d \in [0,1]$ ): linear regression
- In general: generalized linear models



# Continuous traits

- A linear regression model is defined as:

$$y = x\beta_1 + \beta_0 + \varepsilon$$

- Data:

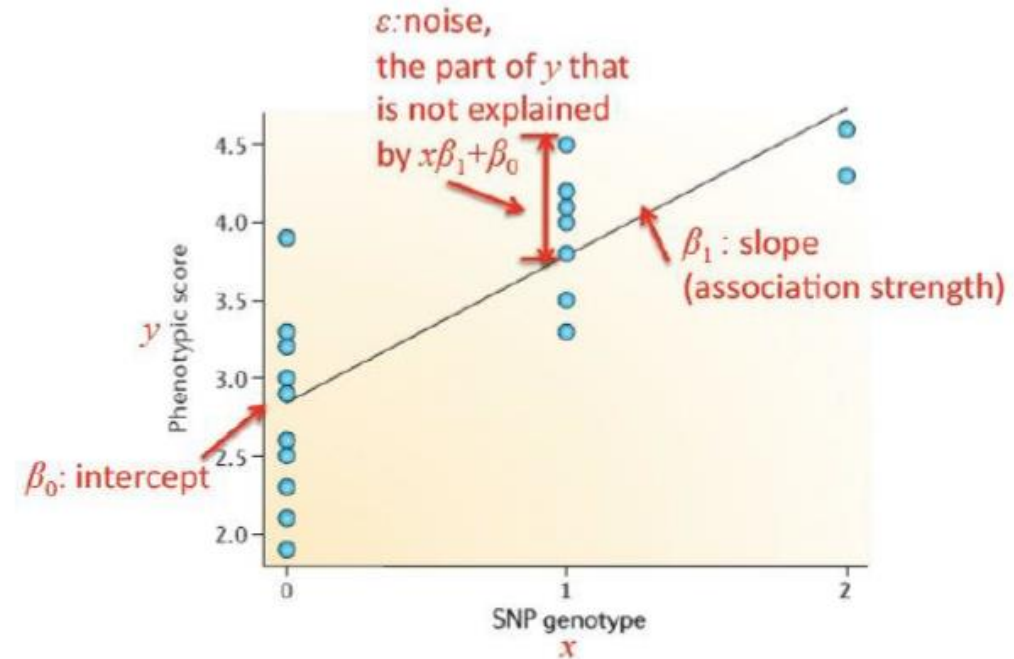
- $y$  is a continuous trait
- $x$  is the SNP genotype at a given locus

- Parameters:

- $\beta_1$  is the regression coefficient, represents the strength of association between  $y$  and  $x$ 
  - $\beta_1 > 0$ : for every supplementary allele, the phenotype will increase by the beta coefficient value
  - $\beta_1 < 0$ : for every supplementary allele, the phenotype will decrease by the beta coefficient value
- $\beta_0$ : intercept term (is often ignored)

- Assumptions:

- The individuals in the study are not related
- The phenotype  $y$  has a normal distribution



# 3

## Hypothesis testing

### 3.1 – Statistical tests



# Hypothesis testing

- Measure whether the data gives sufficient evidence to reject a hypothesis

## Null hypothesis $H_0$ vs Alternative hypothesis $H_A$ ( $H_1$ )

- $H_1$  = Hypothesis of interest
- Use a statistic that follows a certain distribution under  $H_0$ 
  - Name of the test = name of the statistics
  - Can we reject  $H_0$ ? Not rejecting  $H_0$  is different from proving it!
- We calculate the statistics based on our data
- As we know the distribution, we can compute the CDF  $p(X \leq x)$
- Decision based on a significance threshold  $\alpha$  and the p-value = how likely the measurement comes from the null
  - If  $p < \alpha \rightarrow$  we reject  $H_0$  and consider the test significant

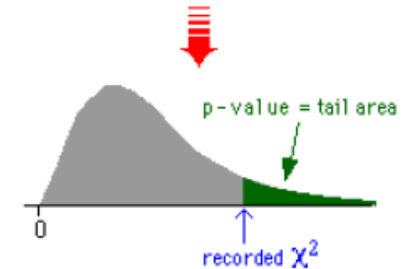
**Summary statistic**  
(helps distinguish  $H_0$  and  $H_A$ )

↓  
**Test statistic**  
(standard distribution with no unknown parameters under  $H_0$ )

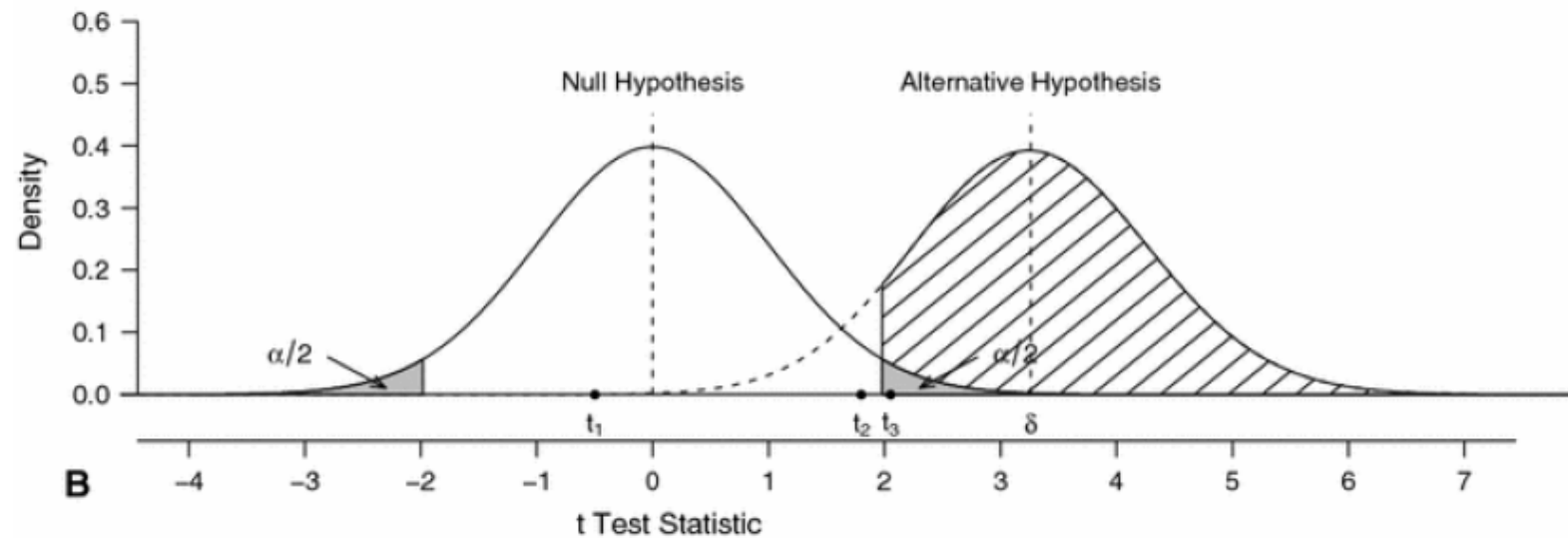
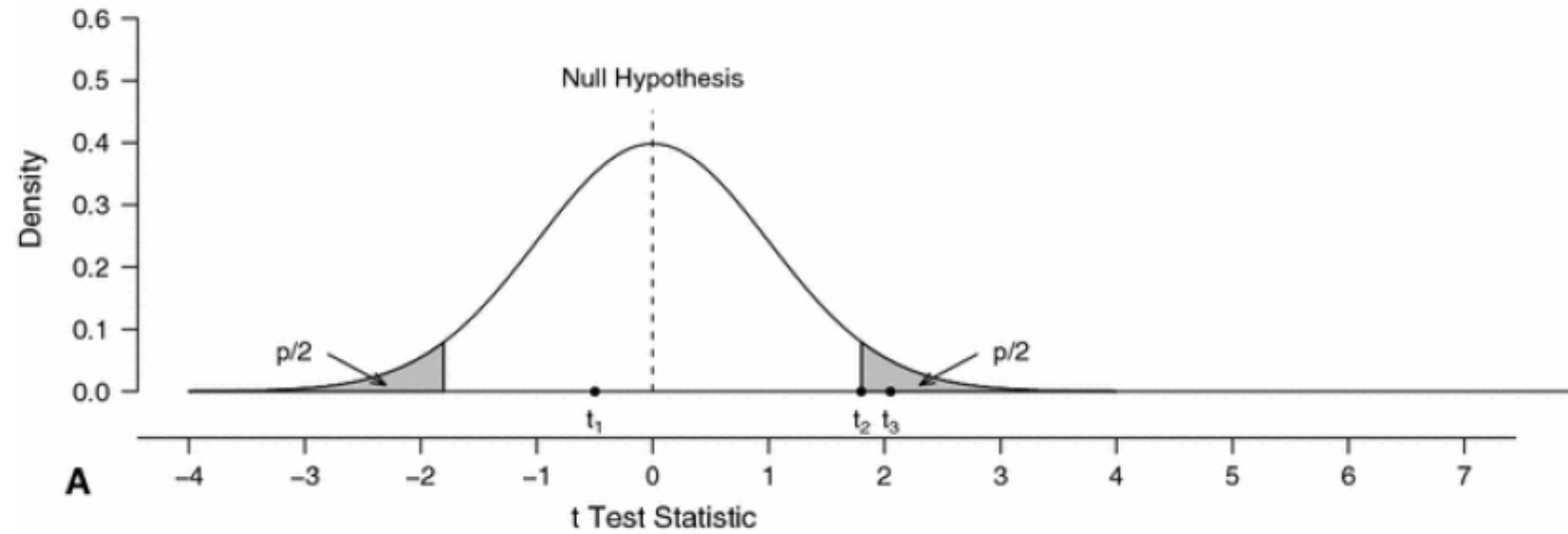
↓  
**P-value**  
(probability of more 'extreme' test statistic)

$$\chi^2 = \sum \frac{(n_{xy} - e_{xy})^2}{e_{xy}}$$

↓  
 $\chi^2 \sim \text{chi-squared } ((r-1)(c-1) \text{ df})$



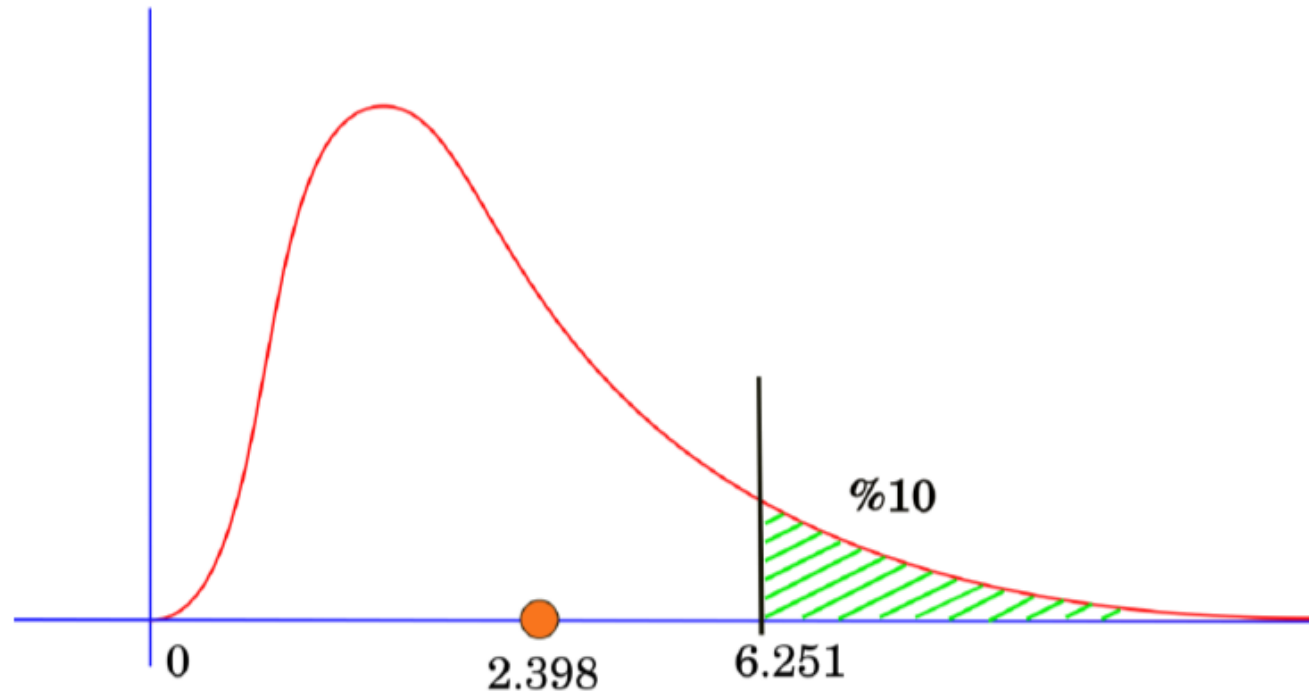
$H_0$  vs  $H_1$



# One-sided vs Two-sided test

- Depends on the hypothesis

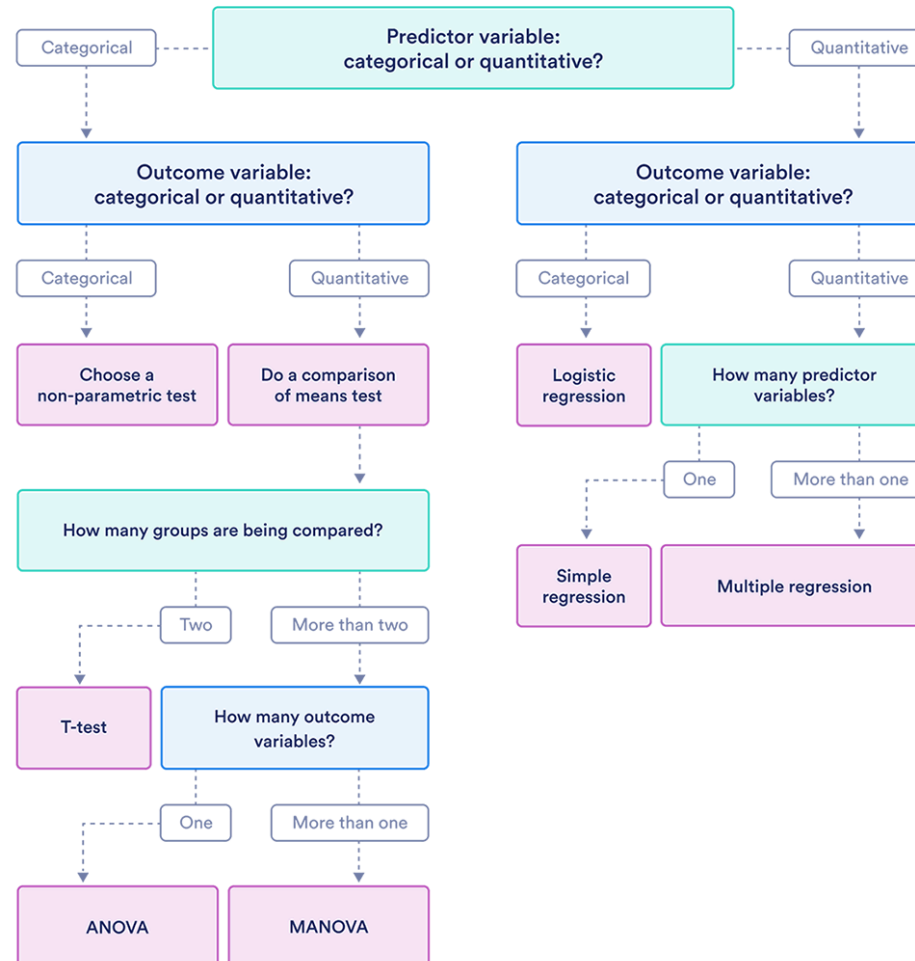
<u>Two-sided:</u>	$H_0: " = "$	vs.	$H_1: " \neq "$
<u>One-sided:</u>	$H_0: " \leq "$	vs.	$H_1: " > "$
	$H_0: " \geq "$	vs.	$H_1: " < "$





# Choosing a statistical test

This flowchart helps you choose among parametric tests



GWAS are performed under an additive model

3

# Hypothesis testing

## 3.2 – Multiple testing

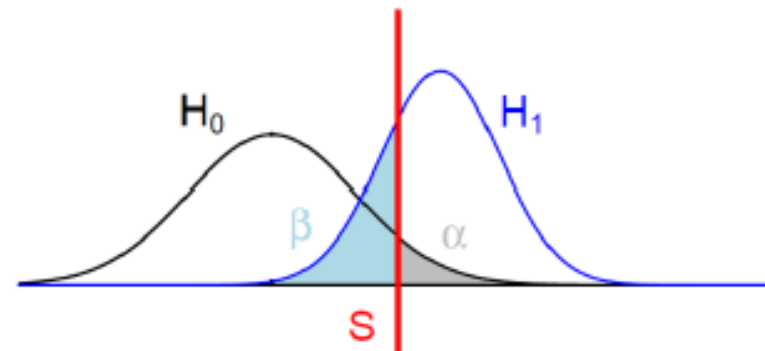


# Multiple testing

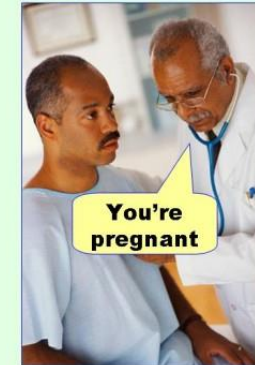
- If  $p < \alpha \rightarrow$  we reject  $H_0$  and consider the test significant
- $\alpha$  corresponds to the type I error risk that we want to control

	$H_0$ is true	$H_1$ is true
$H_0$ not rejected	Correct decision	Type II error
$H_0$ rejected	Type I error = $\alpha$	Correct decision

- If the number of tests increases, the risk increases  
 $\rightarrow$  Need to take into account the multiple tests to maintain  $\alpha$  at the desired level across all tests



**Type I error**  
(false positive)



**Type II error**  
(false negative)



# Multiple testing: m tests

## Family-Wise Error Rate (FWER)

- Bonferroni correction
- Simple to implement, harder to interpret

$$p_{critical} = \frac{0.05}{m}$$

- *“If all tests are under the null, probability that **one or more** of them is a false positive.”*

## False-discovery based (FDR)

- Benjamini Hochberg procedure
- Harder to implement, easy to understand

$$p_{critical} = \operatorname{argmax}(p < \frac{i}{m} Q)$$

- $i = \text{rank}, Q = \text{FDR}$
- *“Proportion of significant tests that are false positives.”*

When to use which depends on

1. Best practices
2. Relative cost of a false negative/positive

# Multiple testing: application in genomic studies

- Statistical significance:

- 5% for one test

- Genome-wide: one test per variant and per phenotype

$$phenotype \sim \beta \times genotype + \epsilon$$

- But all the variants are not independent and in reality we account for LD = correlation between the variants

- $5 \times 10^{-8}$  for GWAS,  $10^{-9}$  for sequencing based

$$p_{critical} = \frac{0.05}{m}$$

## I➔ Exercise : Significance threshold

- If the adjusted genome wide significance threshold is  $5 \times 10^{-8}$  for GWAS, how many “effective” variants are there in a genotyped human genome?  **$10^6$**
- You are writing an article about a GWAS for 16 different traits. What will be your threshold for declaring significance?  **$3.125 \times 10^{-9}$**

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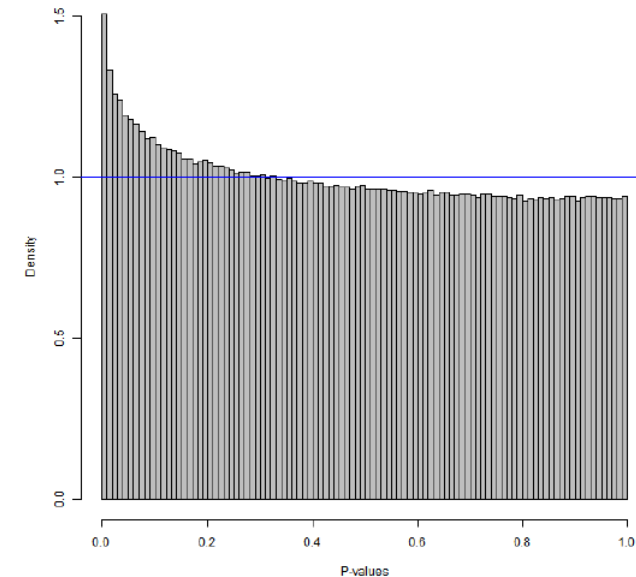
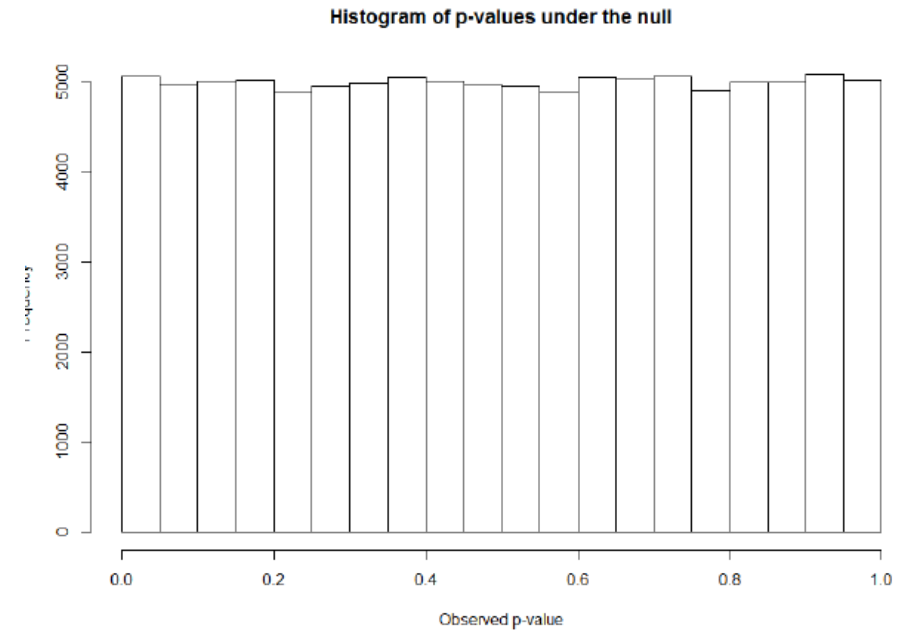
## Hypothesis testing

### 3.3 – Checking the results



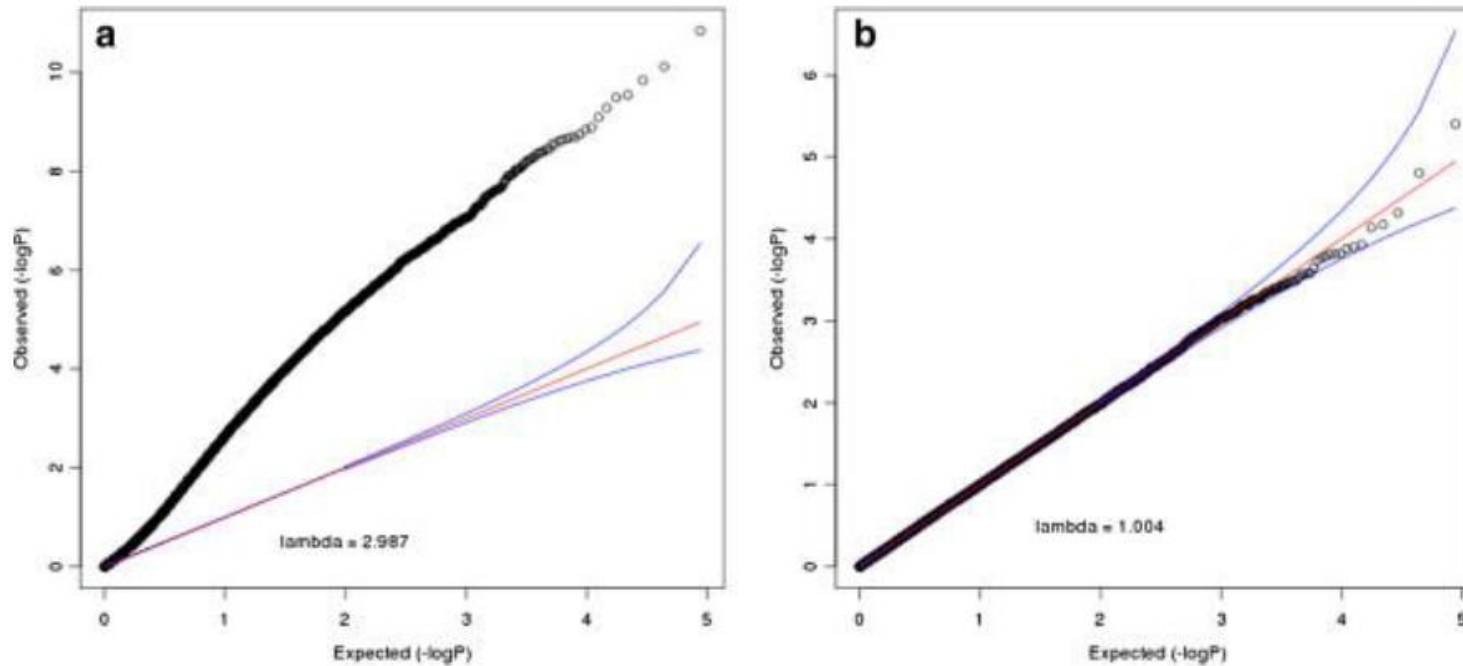
# QQ-plots

- Under  $H_0$ , p-values should be uniform [0,1]
- It is expected that most signals are around  $H_0$   
→ If we have much signal, more around 0
- Compare quantiles with expected ones: QQ-plot
- In R: *qqunif*





# QQ-plots



- Inflation: too much signal
- Visual inspection but also lambda value

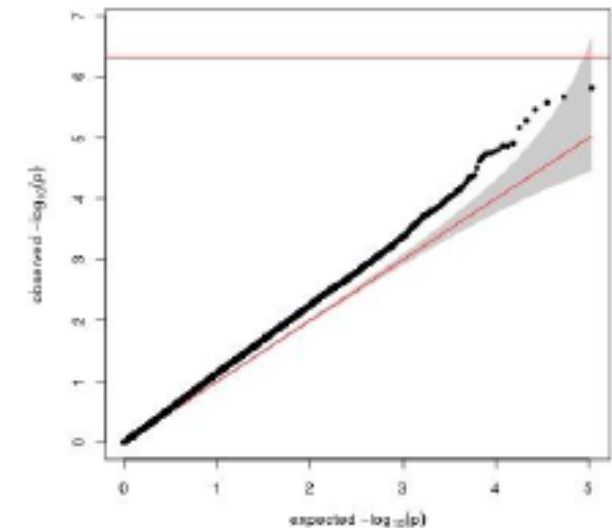
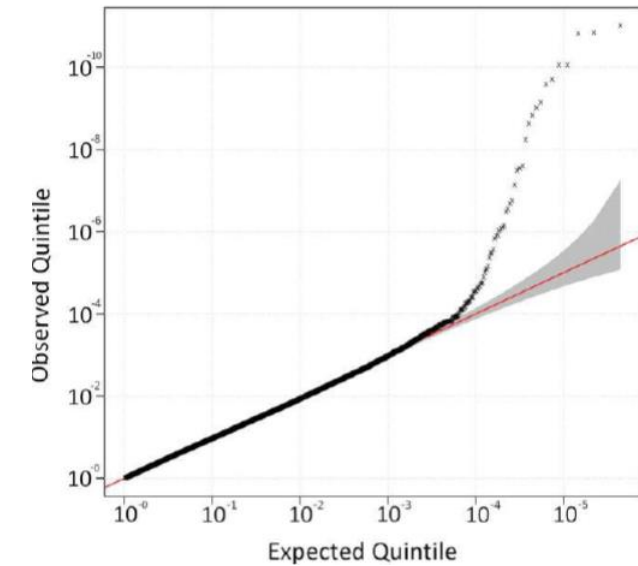
# QQ-plots

- Appearance can be misleading:
  - A QQ-plot can seem inflated but just a lot of signal
  - And conversely...
- Calculation of the genomic inflation factor:

$$\lambda = \frac{\text{median}(Q_{\chi^2}(p))}{0.45}$$

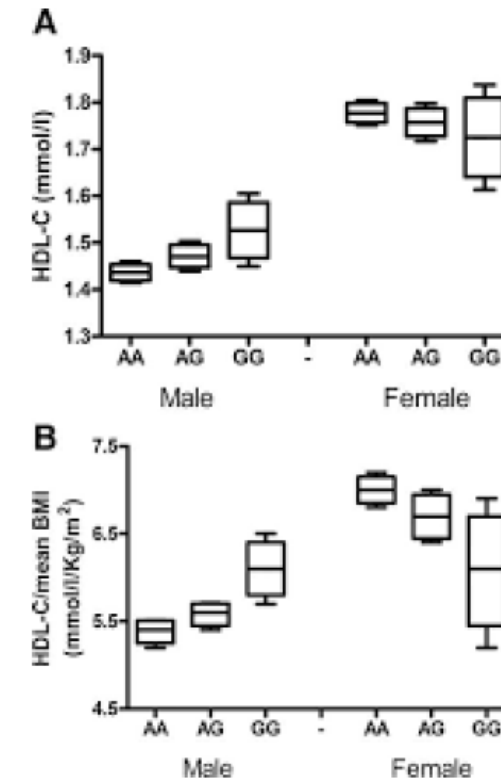
With 0.45 being the median of  $\chi_1^2$

- $\lambda > 1$ : **inflation** (systematic bias)
- $\lambda < 1$ : deflation (potential power issue)
- Ideally, we want to correct the model
- Can also adjust: GC correction: divide by lambda



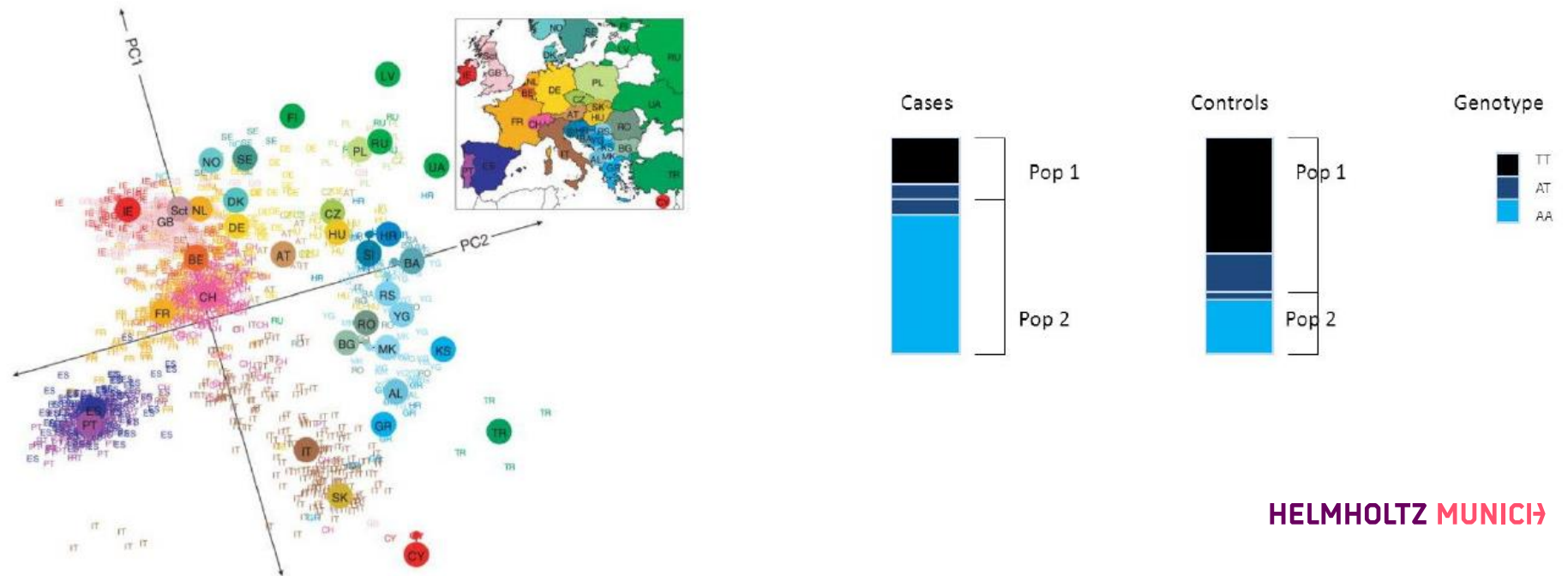
# Problems in the model

- Covariates: sex, batch effects, chip effects  
→ Potential bias if associated to phenotype and genotype



# Problems in the model

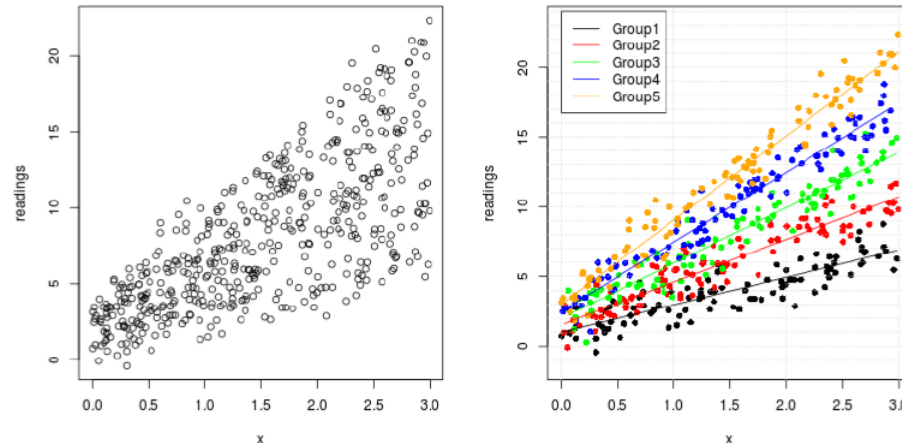
- Covariates: sex, batch effects, chip effects
  - Potential bias if associated to phenotype and genotype
- Structure or subpopulations
  - Allelic frequencies are known to be different from one population to another



# Problems in the model

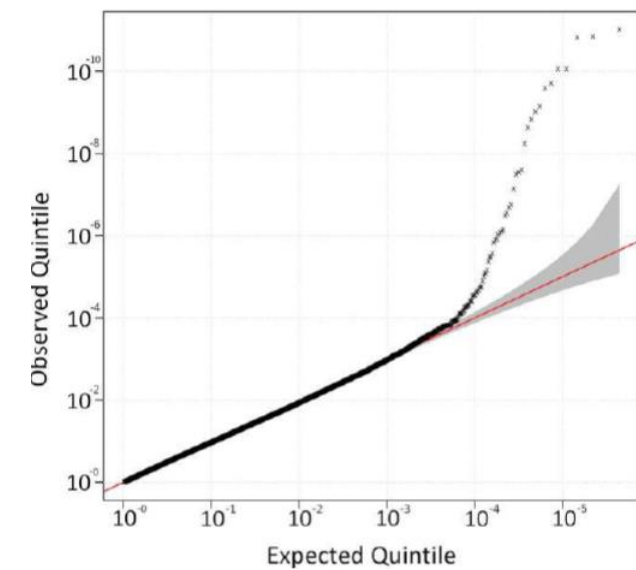
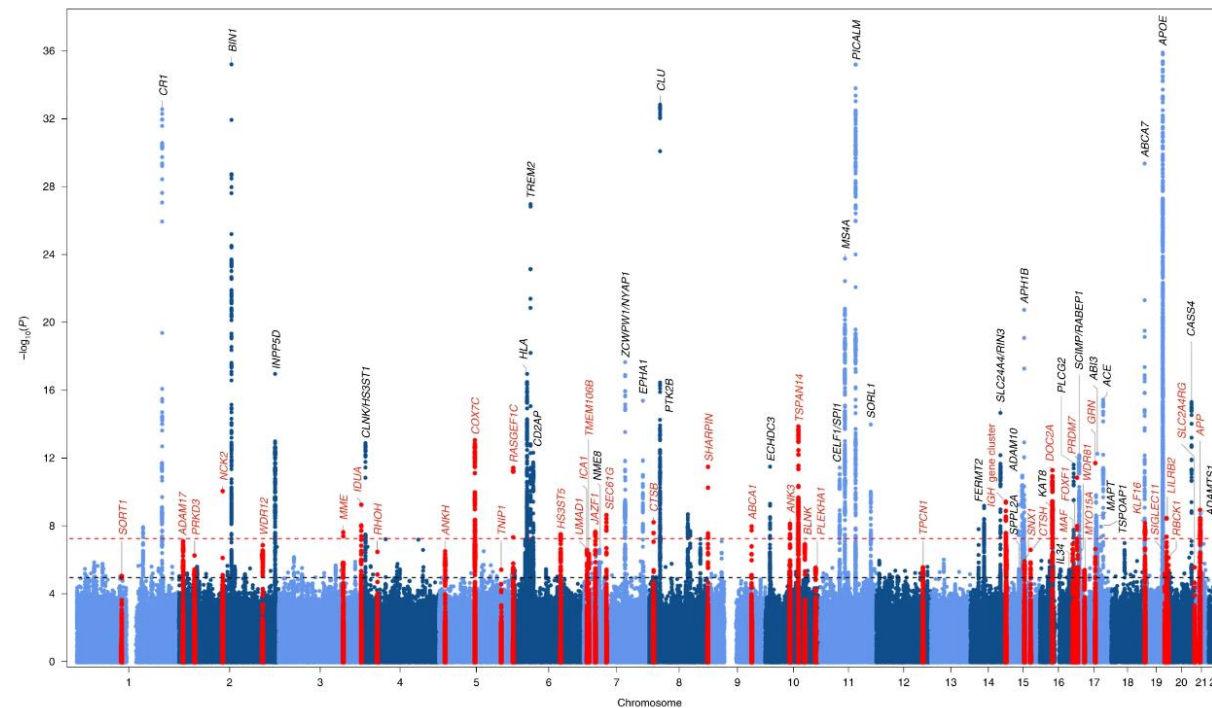
- Covariates: sex, batch effects, chip effects
  - Potential bias if associated to phenotype and genotype
- Structure or subpopulations
  - Allelic frequencies are known to be different from one population to another
  - Linear mixed models can model the intra-group effect
  - Adjust for principal components of a PCA

$$\text{phenotype} \sim \beta \times \text{genotype} + \beta_1 \times \text{covariates} + \beta_2 \times \text{structure} + \epsilon$$



# Manhattan plot: visualization of the results

- Display  $-\log_{10}(p)$  for every position in the genome
- Use a threshold ( $5 \times 10^{-8}$ ) to declare significance



4

Prediction



# Prediction

- When we apply the estimated effects to new observations of the variable
- Suppose underlying assumptions
- Process = machine learning, predictive modelling, predictive analysis
- In human genetics, main task = **model effects of genotypes on phenotypes**
- Usually we do not predict
  - Except PRS (Lecture 5 on 6<sup>th</sup> December)





Thank you.