Lecture 3: Statistics for Human Geneticists



Volos Summer School

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Arthur Gilly



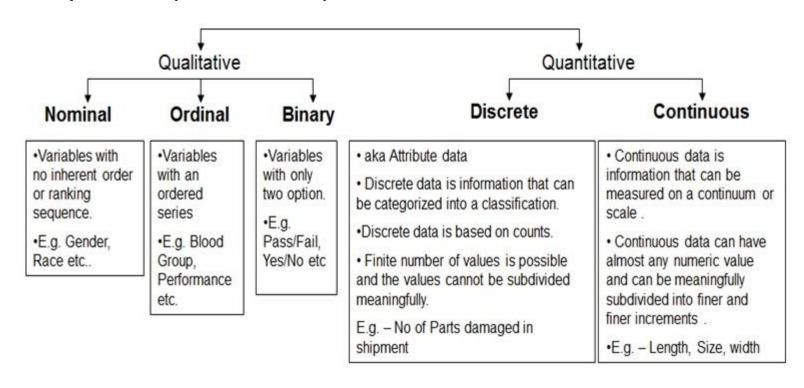
What can we do with statistics?

- Estimation
- Hypothesis testing
- Modelling
- Predicting



Random Variables

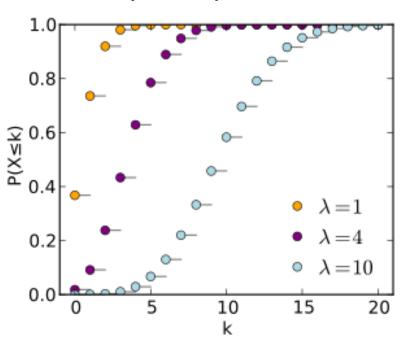
- In statistics, we measure realizations of random variables
- Often, random variables follow a distribution
- They can be qualitative or quantitative, continuous or discrete

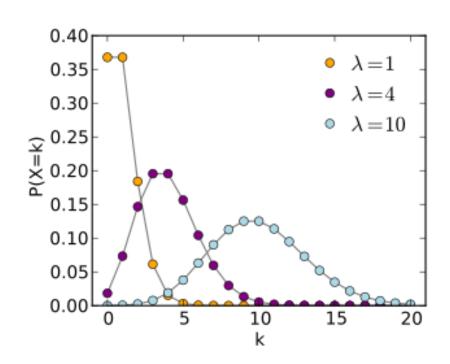




Distributions

Two ways to represent them :





Cumulative distribution function (CDF)

- $y = p(X \le x)$
- Always growing
- Ideal way to represent, but hard to read
- All distributions look the same

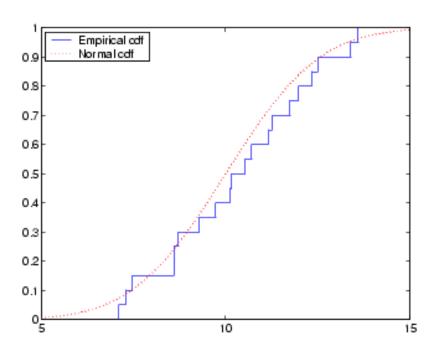
Probability density function (PDF)

- y = p(X = x) for discrete
- Shows how values are distributed
- Nice visually, but mathematically hard to deal with



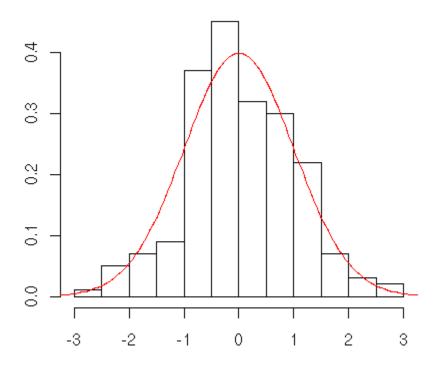
Distributions

How to estimate them:



Empirical CDF

- Rarely used
- Except when you want to compute empirical quantiles



Barplot (discrete)

• For every value, count occurrences

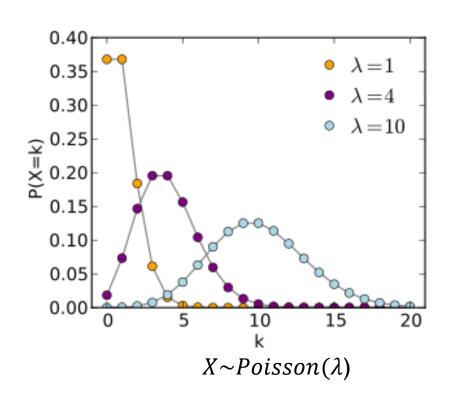
Histogram (continuous)

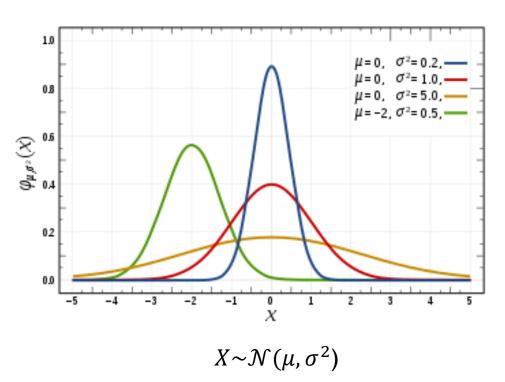
Cut the interval into bins, count observations within bin



Distributions

- Two broad types:
 - those followed by random variables (real world data)

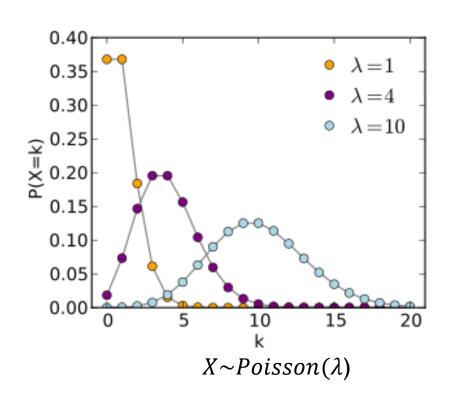


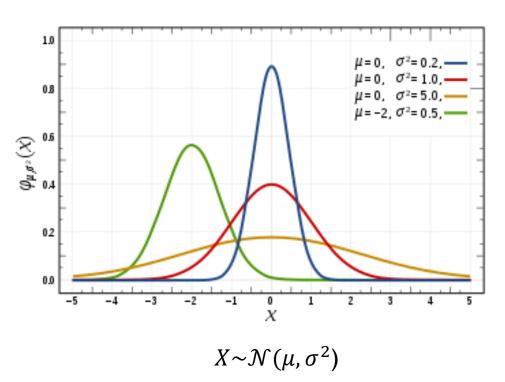




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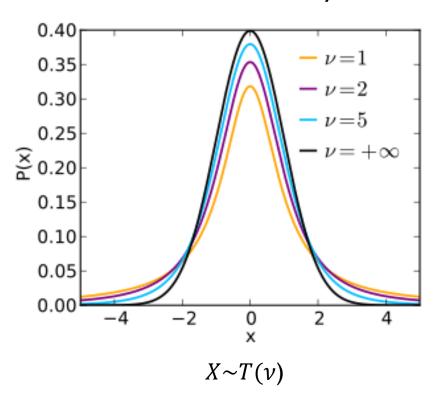


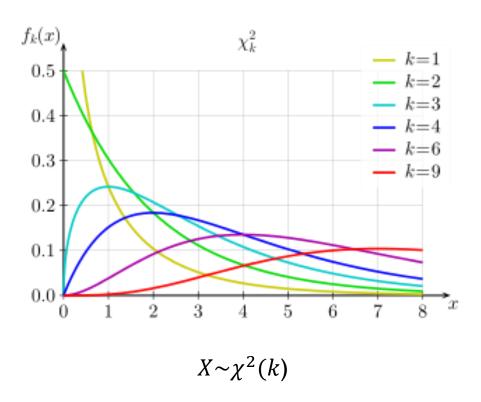




Distributions

- Two broad types:
 - those followed by test statistics





 λ , μ , σ , ν and k are ideal parameters. How to estimate them?



Statistics

- A statistic is a meaningful quantity derived from the data
- Often, estimators are realization of distribution parameters
- Examples? Mean, proportion
- For simple distributions/parameters, there is a formula
- For more complex ones, we have to use other techniques (Monte-Carlo, Permutations...)

$$(\hat{\mu} =) \bar{x} = \frac{1}{n} \sum_{k=0}^{n} x_k$$

$$\hat{p} = \frac{x}{n}$$

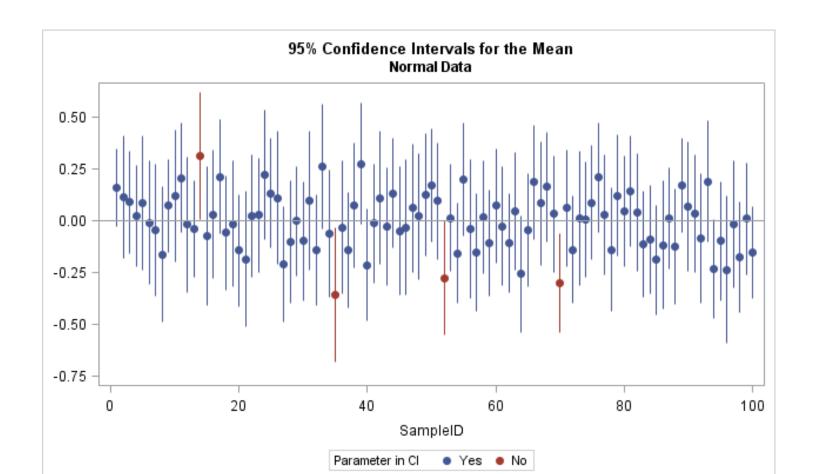
$$(\widehat{\sigma}^2 =) s^2 = \frac{1}{N-1}$$

$$w = \frac{(\widehat{\theta} - \theta_0)^2}{se(\widehat{\theta})} \sim \mathcal{N}(0,1)$$



One particular statistic: Confidence intervals

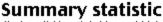
• x% confidence interval (x%C.I.): x% of the time when this interval is calculated, it will contain the true value of the parameter





Hypothesis testing

- We want to measure whether the data gives sufficient evidence to reject a hypothesis
- Null/Alternative hypothesis $(\mathcal{H}_0/\mathcal{H}_A)$
- We prove that we can produce a statistic that follows a certain distribution if the null hypothesis is true = name of the test
- We calculate the statistic based on our data
- Because we know the distribution, we can calculate the CDF $p(X \le x)$
- = how unlikely it is that our measurement comes from the null : p-value
- Example: proportion test, t-test, chisquared test...



(helps distinguish H₀ and H₄)



Test statistic

(standard distribution with no unknown parameters under H₀)

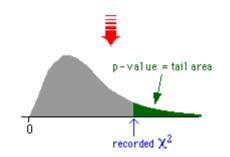


P-value

(probability of more 'extreme' test statistic)

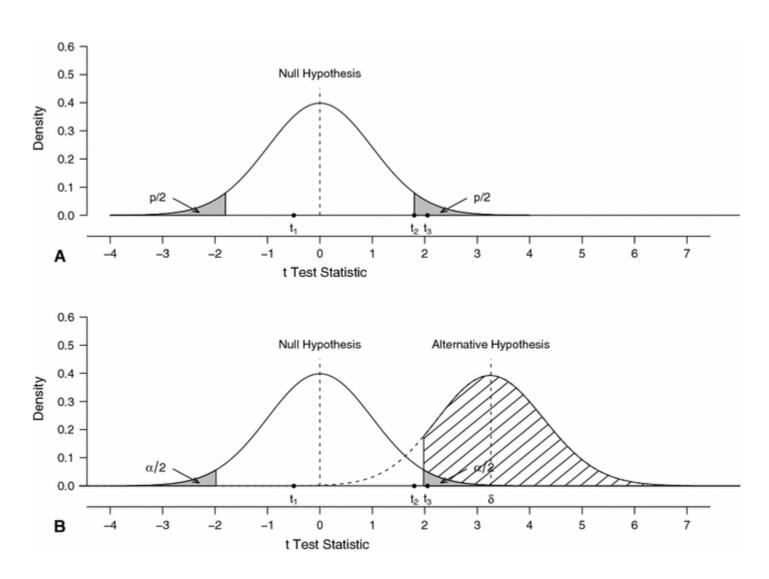
$$\chi^2 = \sum \frac{\left(n_{xy} - e_{xy}\right)^2}{e_{xy}}$$

$$\chi^2 \sim \text{chi-squared } ((r-1)(c-1) \text{ df})$$



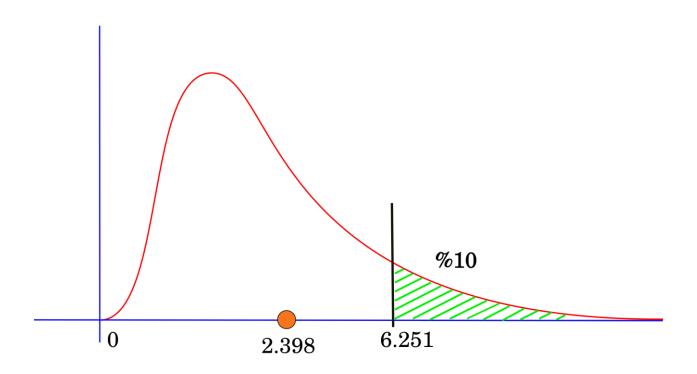


Hypothesis testing





Two tails or one





Exercise I: Proportion test

- In a population, we observe 41,009 potentially damaging variants among 14,281,180 variants
- What is the proportion? In a very large reference population, we observe a proportion of 1.52×10^{-3} . Is it significantly different? (prop.test, binom.test)



Modelling and predicting

- If we estimate the effect of one variable on another variable, we do modelling
- When we apply this effect to new observations of the variable, we do prediction
- Process is called machine learning, predictive modelling or predictive analysis
- In human genetics, main task is to model effect of genotypes on phenotypes

$$phenotype \sim \beta \times genotype + \epsilon$$

$$\begin{bmatrix} pheno_0 \\ \vdots \\ pheno_n \end{bmatrix} \qquad \begin{bmatrix} A/T \\ \vdots \\ T/T \end{bmatrix}$$



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phenotype ~ $\beta \times genotype + \epsilon$

 $= \{0,1\}$ (case-control) $\in \mathbb{R}$ (quantitative) $\sim \mathcal{N}(0,1)$

$$\begin{bmatrix} pheno_0 \\ \vdots \\ pheno_n \end{bmatrix} \qquad \begin{bmatrix} A/T \\ \vdots \\ T/T \end{bmatrix} = \{0,1,2\} \text{ (genotype, directly typed } \\ \in [0,2] \text{ (dosage, imputed)} \\ \mathcal{N}(0,1) \qquad \begin{bmatrix} 0.965 \\ \vdots \\ 1.816 \end{bmatrix}$$



Modelling and predicting

- If we estimate the effect of one variable on another variable, we do modelling
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Usually, we do not predict (except PRS)

phenotype $\sim \beta \times genotype + \epsilon$

$$\begin{bmatrix} pheno_0 \\ \vdots \\ pheno_n \end{bmatrix}$$

$$= \{0,1\}$$
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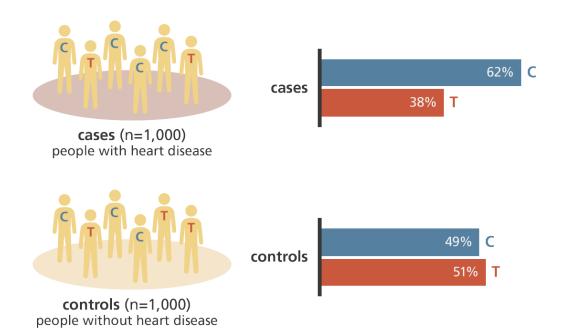
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Case/control

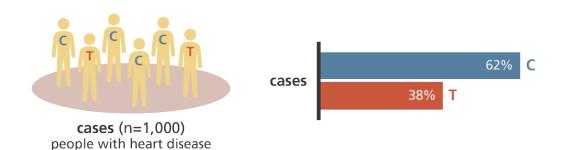
• Estimated effect: odds ratio (OR) "how much more likely are you to be a case if you carry the risk allele?" per genotype, calculate the odds $O = \frac{p}{1-p}$



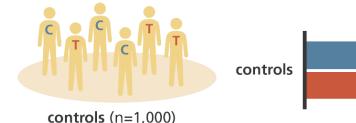


Case/control

• Estimated effect: odds ratio (OR) "how much more likely are you to be a case if you carry the risk allele?" per genotype g and for a disease Y, calculate the odds $O = \frac{p_{Y=1|g}}{1-p_{Y=1|g}}$



	cases	controls
Т	380	490
С	620	510



people without heart disease

$$O_T = \frac{380/n_T}{490/n_T}$$
 $O_C = \frac{620/n_C}{510/n_C}$

$$OR_{C/T} = \frac{620 \times 490}{510 \times 380} = 1.56$$



Case/control

Dominant					
Marker allele	Affected	Unaffected			
DD+Dd	$n_{2A} + n_{1A}$	$n_{2U} + n_{1U}$			
dd	n_{0A}	n_{0U}			

$$OR = \frac{n_{affected \; carriers} \times n_{healthy \; non-carriers}}{n_{healthy \; carriers} \times n_{affected \; non-carriers}}$$

Recessive					
Marker allele	Affected	Unaffected			
DD	n _{2A}	n_{2U}			
Dd+dd	$n_{1A} + n_{0A}$	$n_{1}U + n_{0}U$			

Additive

Marker genotype	Affected	Unaffected
DD	n _{2A}	n _{2U}
Dd	n_{1A}	n_{1U}
dd	n_{0A}	n_{0U}

$$OR = \frac{(2 \times n_{2A} + n_{1A}) \times (2 \times n_{0U} + n_{1U})}{(2 \times n_{0A} + n_{1A}) \times (2 \times n_{2U} + n_{1U})}$$
Allelic odds-ratio



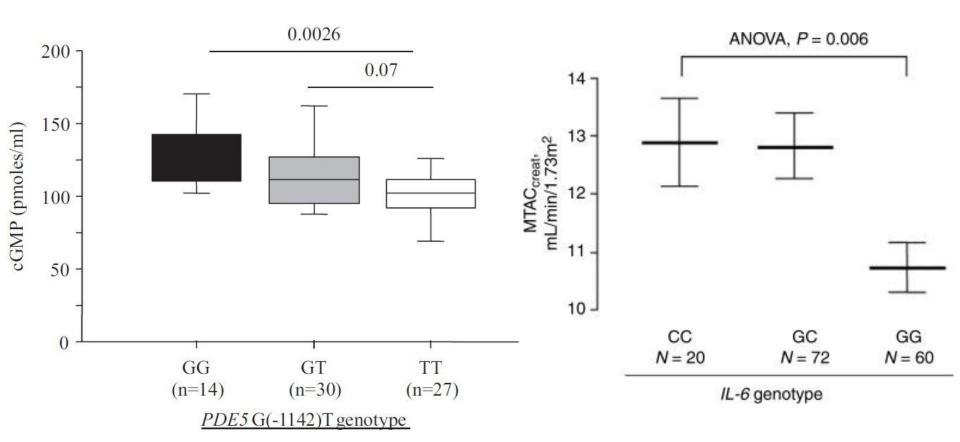
Case/control

- Output: OR and 95% confidence interval of the OR
- Test: is it significantly different from 1?
- Tests: Fisher's exact test or Chi-squared
- In case of dosages or covariates: logistic regression



Continuous trait

For directly typed (0,1,2):ANOVA

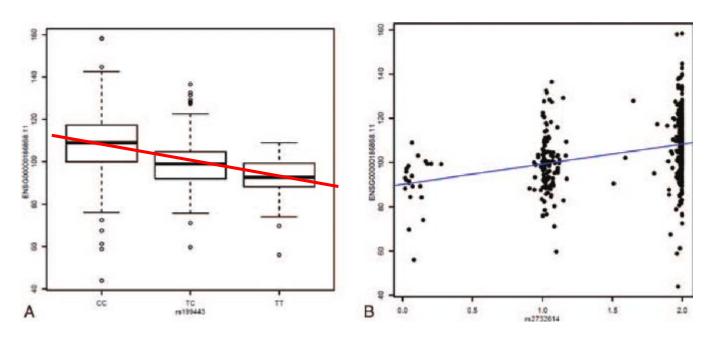


additive recessive



Continuous trait

- For dosages (imputed quantity of minor allele $d \in [0,1]$) : linear regression
- In general: generalized linear model





Continuous trait

A linear regression model is defined as

$$y = x\beta_1 + \beta_0 + \varepsilon$$

Data:

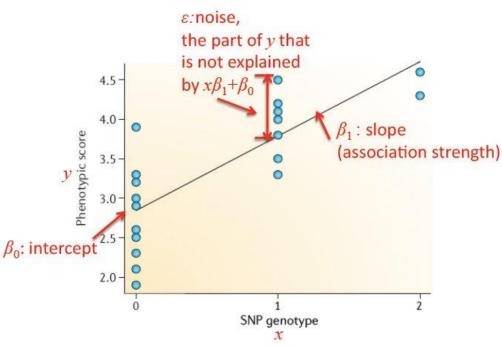
- y: a continuous trait
- x: SNP genotype at a given locus

Parameters:

- β_1 : regression coefficient, represents the strength of association between x and y
- β_0 : intercept term (is 0 or ignored)
- ε: noise or the part of y that is not explained by x (e.g., environmental effect)

Assumptions:

- The individuals in the study are not related
- The phenotype y has a normal distribution

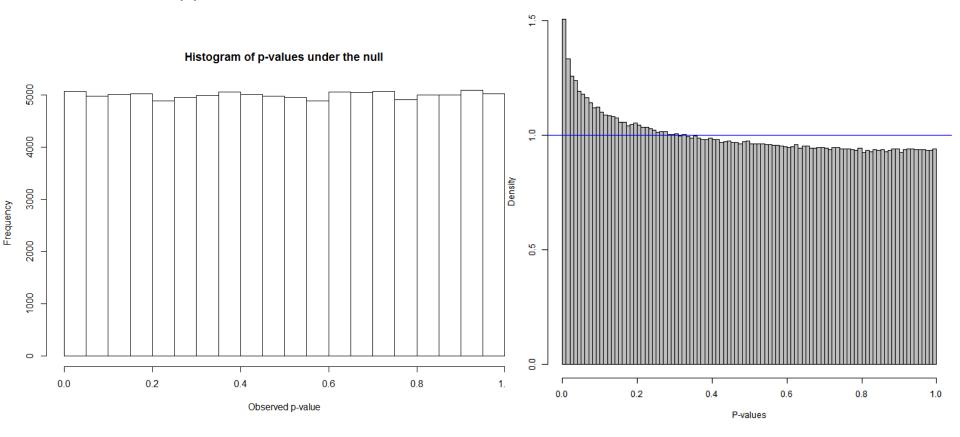




- Statistical significance:
 - One test: p<0.05
 - Genome-wide: one test per variant and per phenotype
 - But all variants are not independent, in reality, we account for LD
 - 5×10-8 for GWAS, 10-9 for sequencing-based

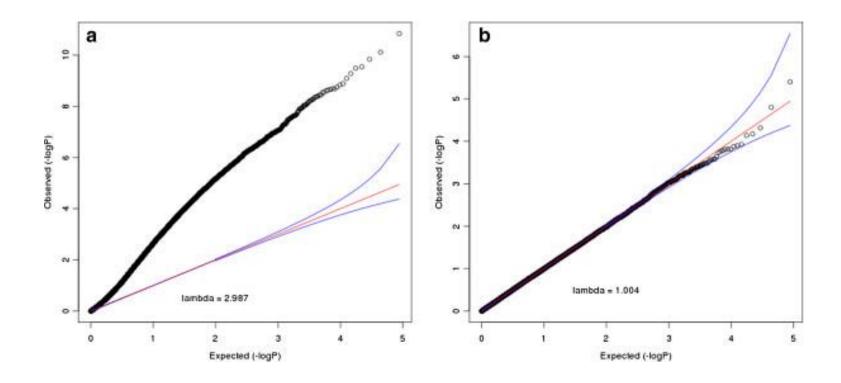


- QQ-plot
 - Distribution of p-values is uniform [0,1] under the null
 - If we have much signal, more around 0
 - Compare quantiles with expected ones: QQ-plot
 - In R: qqunif



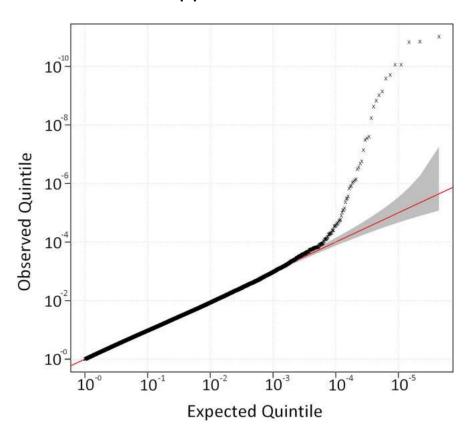


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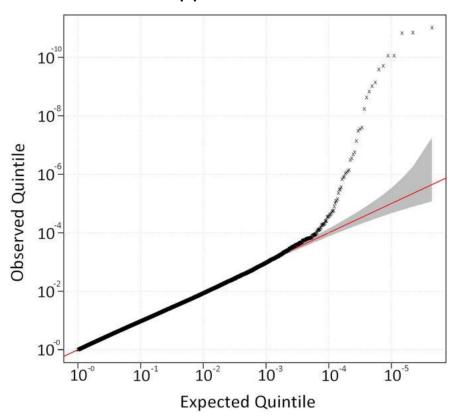


- Inflation: too much signal
- Measured visually, but also lambda



Checking results

- QQ-plot
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Appearances can be deceiving:

- A QQ-plot can look inflated when it isn't (just a lot of signal)
- And conversely
- We calculate the genomic inflation factor

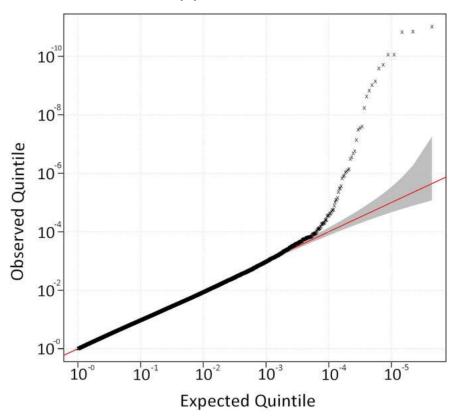
$$\lambda = \frac{median(Q_{\chi^2}(p))}{0.45}$$

(median of χ^2 test statistics divided by median of χ^2_1)



Checking results

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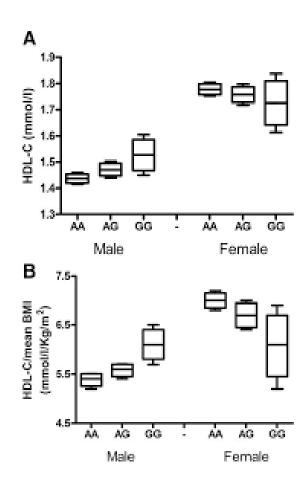
(median of χ^2 test statistics divided by median of χ_1^2)

- Ideally, want to correct in the model
- Can also adjust: GC correction (divide by lambda)



Problems

Covariates: sex, batch effects, chip effects





Genotype

Controls

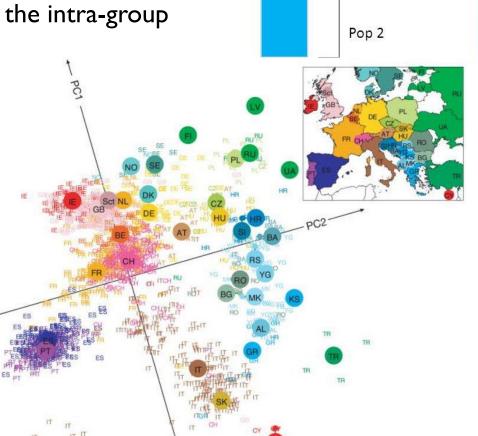
Pop 1

Pop 1

Pop 2

• Covariates: sex, batch effects, chip effects

 Structure: villages or subpopulations: linear mixed models can model the intra-group effect





Problems

- Covariates: sex, batch effects, chip effects
- Structure: villages or subpopulations: linear mixed models can model the intra-group effect

$$phenotype \sim \beta \times genotype + \beta_1 \times covariates + \beta_2 \times structure + \epsilon$$

