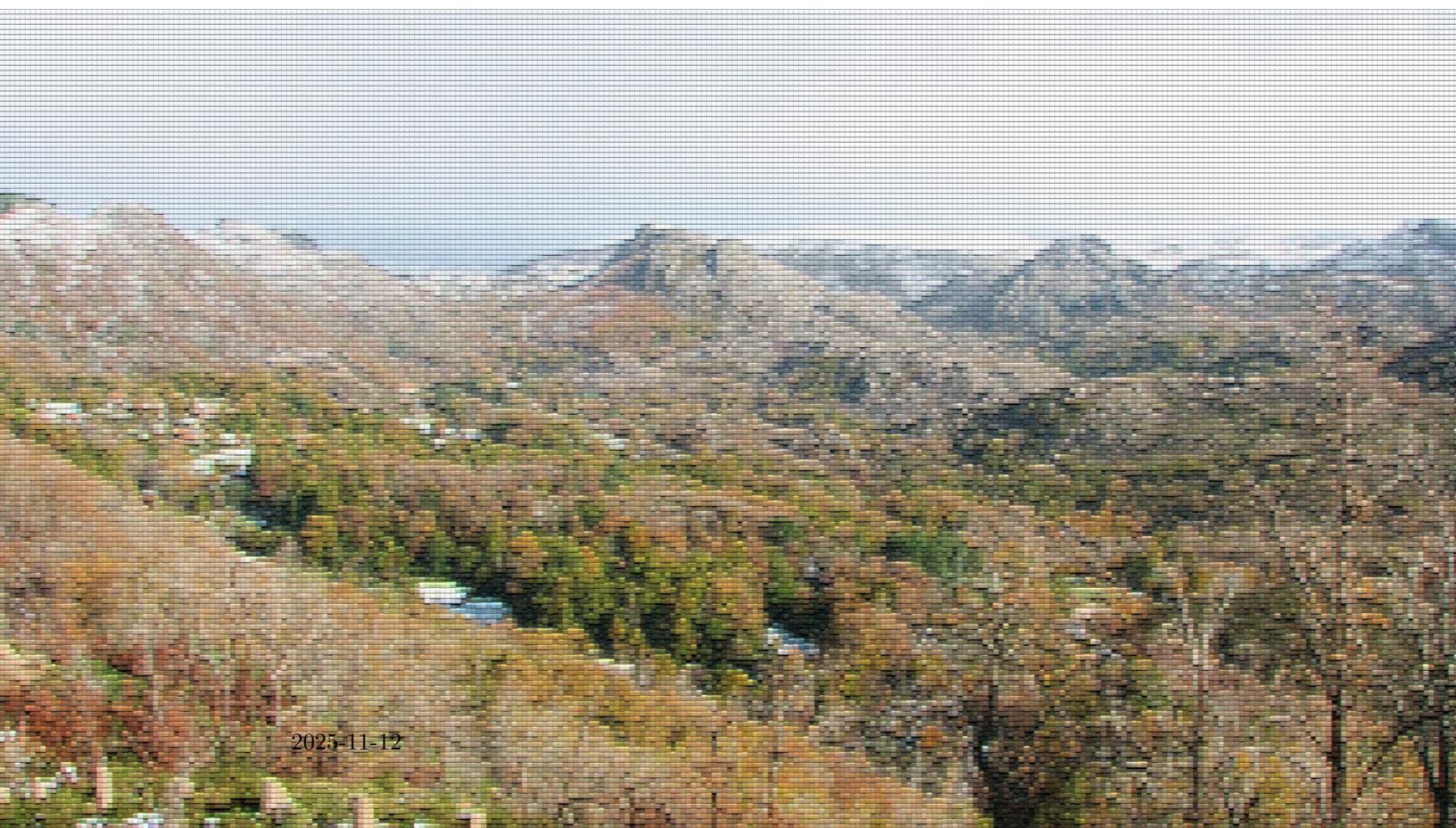


Ecological theory for the biodiversity crisis

Henrique Miguel Pereira



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Preface

We live in the midst of the biodiversity crisis. Biodiversity science, at first mostly a spin off from Ecology, has developed tremendously in the last two decades, becoming a truly interdisciplinary field, with contributions from Geography, Economy, Social Sciences and many other disciplines. Big data and the development of statistical ecology has also been remarkable. As I write these lines, the Global Biodiversity Information Facility or GBIF (<https://gbif.org>) is about to hit 3 billion records. The availability of large amounts of data and the availability of the open source R software (<https://www.r-project.org>) has made statistical approaches take the main stage in much of ecological research. However, at the same time, as a biodiversity researcher, practitioner, and a teacher, I find again and again the need for a good understanding of ecological theory to be able to carry out my work. This book if for all of you that have also felt a similar need.

Programming and mathematics are key tools of ecological theory. The programming language I use in this book is R (version 4, available from <https://www.r-project.org>). If you have familiarly with programming in another computer language, a fast skimming of Appendix A will be enough to get you going with the book. If you are new to programming, I recommend you spend a few hours working through the short course in Appendix A, but you may also wanna consult some web resources (e.g. <https://ourcodingclub.github.io/course>). With the advent of Large Language Models (LLM) such as ChatGPT there may be the temptation to think programming knowledge will no longer be needed. I think LLMs will accelerate our writing of computer code, but they will not replace the need for being able to read and understand the code.

You may wonder why I chose **R**. There are plenty other programming languages out there, several suitable for ecological theory and modelling (e.g. data scientists love Python). However, **R** has become the *de facto* language of ecologists with dozens of packages¹ for a wide range of analysis, from camera trapping to species richness estimation. **R** has also powerful packages for spatial data analysis and can work as a Geographic Information System. So R, grew from a statistics software (the origins of R are the comercial S-PLUS) to a general purpose programming language with dedicated packages for different uses.

Why was R so successful, to the point of making some commercial software companies go out of business? R was open-source and free from its beginning, part of the free and open-source movement that gained popularity in the 1990's, particularly with the development of the operating system Linux. Open-source means that the code behind R (R itself is a computer program written in C, Fortran, and other languages) is available for anyone to study. It also allows for anyone to

¹An R package is a set of functions developed by someone to extend the basic functionality of R.

contribute to improve the code in a collaborative way, including by contributing with packages. R is also free, meaning that anyone can download the software or any of its packages without any cost. Isn't that a wonderful tool for science? Scientists dedicate their life to the pursue of knowledge, and the idea that someone has to pay for accessing that knowledge has always been a bit apocryphal. So R allowed scientists to share their research and code for free in an integrated platform. Another advantage of open-source software, is that we can in theory ran programs that we wrote decades ago by downloading archived versions of the software. At least in theory, the practice of code reproducibility in science is a bit more complex and requires careful data and code management by the researcher². Platforms such as GitHub (<https://github.com>) for sharing code and Zenodo (<https://zenodo.org>) or Dryad (<https://datadryad.org>) for sharing data are a key component of reproducible science.

In contrast a lot of the code I wrote for my PhD was based on a commercial package (Mathematica) and anyone that wants to reuse it a couple decades later has to pay a few hundred Euros for the package (well there are student discounts) and has to find a version of the Mathematica that still runs this code. This latter problem can be insurmountable, as sometimes older versions of the software are no longer commercially available. One can argue that many programming languages are also free and public, however for ecological theory and modelling a high-level language like R provides a nice integrated platform for development and is literally priceless.

But programming is not enough. In order to be able to effectively develop theory and models of biodiversity, a good grasp of mathematics, including of probability and statistics is essential. One challenge is that, and this at least the case in Europe where I have lectured the most, most biology students receive very little mathematical training. This book cannot address by itself such gap and assumes some basic familiarity with probability theory (e.g. what is a probability distribution function), some calculus training (e.g. what area a derivative and an integral), a little algebra (e.g. multiplication of a matrix by a vector). I try to take the reader forward from that level. There are excellent books out there for those that need this basic background (e.g. Otto's and Day's *A Biologist's Guide to Mathematical Modelling* (2007) for math and Mangel's and Hilborn's *Ecological Detective* (Hilborn and Mangel 1997) for statistics). And Wikipedia (<https://www.wikipedia.org>) is often your best friend when you wanna a fast refresh of any mathematical concept or even for many of the models and ecological concepts presented in this book.

This book can be used as the support for a semester course in Ecological Theory or Ecological Modelling. I have used many of these materials over the last decade in a similar course at iDiv/University of Halle-Wittenberg. But the book can also be used as self-learning tool or even as a reference tool. This book takes a lot of inspiration from the Primer of Ecological Theory of Joan Roughgarden (1998) (the first two chapters draw heavily on her first chapters), that I was lucky to have as a mentor. As she used to say, one writes papers for the reviewers and books for the readers. This book is for you.

²For a guide on best practices for reproducible code see <https://www.britishecologicalsociety.org/wp-content/uploads/2019/06/BES-Guide-Reproducible-Code-2019.pdf>

Part I

Ecology of individuals

Chapter 1

Ecophysiology and the climate space: when to bask in the sun?

We are now almost daily bombarded with news about climate change and its impacts on people and ecosystems. But how does climate mechanistically affects organisms? The basic foundations to address this question were laid out by ecophysiology research. Interestingly, at the time that some of seminal research on ecophysiology was carried out, back in the 1940's (see review in (Huey 1982)), climate change was not yet in the radar of most people. The research was driven by the interest in the basic understanding of how biophysical conditions affect organisms. Today the knowledge gained from this research has acquired new importance. This is an area whether the mechanistic knowledge learnt from theory and experiments is now sometimes overlooked because of the massive datasets and machine learning approaches available. But let's start with the basics.

1.1 A model for the body temperature of an animal

Animals can be divided in two big groups in regard to the way they regulate their temperature: ectotherms and endotherms (Figure 1.1). Endotherms such as mammals and birds are able to regulate their temperature by producing heat through metabolism. Ectotherms in contrast must regulate their temperature by obtaining heat from the environment. By obtaining energy from the environment, ectotherms have lower energy demands than endotherms, which have to be burning energy all the time to keep their bodies warm. Both do have to avoid getting too hot because indeed there can be too much from a good thing.

To build a model of the body temperature of an animal, let's consider a lizard that is perched in a rock basking in the sun (Figure 1.2). The lizard receives heat from the direct solar radiation than can be measured in for instance calories per hour. It can also receive indirect solar radiation, reflected for instance by the ground. The lizard also exchanges heat with the surrounding air through convection. If the lizard body temperature is higher than the surrounding air temperature it will lose heat, while if the lizard body temperature is lower than it will gain heat. The rate at which this exchange of heat through convection happens depends on the body of the animal



Figure 1.1: Shrews (left, *Crocidura russula*) are ectotherms and have high energy demands, eating almost their body weight every day. In contrast, vipers (right, *Vipera seonae*) can go days without eating, or even hibernate or aestivate as needed.

and the properties of its skin. These properties are captured in the heat transfer coefficient, which can be measured as calories per hour per degree Celsius. Similarly, the lizard can receive heat by conduction from being in contact with a warm rock, or lose heat to the rock if the rock is colder than the lizard's body temperature. Finally the animal can lose heat through evapotranspiration, this is by transpiring water that has a cooling effect when it evaporates. I also like to think about this model as representing our own experience on the beach. We can be laying on a towel receiving heat by conduction from the sand and solar radiation from the sun. If the air temperature is warm, we can become uncomfortable in the sun and we may seek a shade to reduce the input from solar radiation, but if there is a cool breeze we may be able to stay in the sun a bit longer. If the air temperature is really cold and it's really windy our skin hair may rise to reduce the convection coefficient.

The total flow of energy into the lizard, also known as the heat exchange equation, can be written as

$$f = q - k(b - a) \quad (1.1)$$

where

- f is the energy flow (cal/h),
- q is the quantity of heat in the solar radiation (cal/h),
- k is the convection coefficient (cal/h/ $^{\circ}$ C),
- b is the body temperature ($^{\circ}$ C) and
- a is the air temperature ($^{\circ}$ C).

If the energy flow is positive, then the lizard is warming, while if it is negative then the lizard is

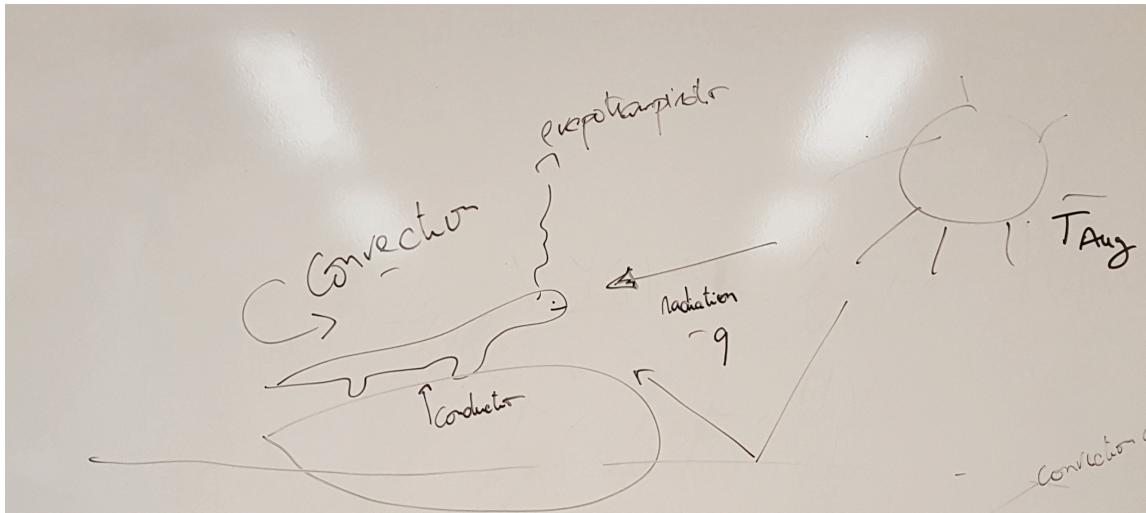


Figure 1.2: The energy flows of a lizard basking in the sun.

cooling. The equilibrium¹ happens when the energy flow is zero and so the lizard is neither cooling nor warming. If we solve Equation 1.1 for equilibrium by replacing f with zero and expanding the right-hand side of the equation,

$$0 = q - k b + k a$$

and rearranging for b we find the equilibrium body temperature that we denote with a hat,

$$\hat{b} = q/k + a. \quad (1.2)$$

Let's see if this equation makes sense. It says that the equilibrium body temperature is the sum of the solar radiation divided by the convection coefficient with the air temperature. So, at minimum the equilibrium body temperature is equal to the air temperature when the solar radiation is zero (e.g. during the night). But when the solar radiation is greater than zero then the equilibrium body temperature is higher than the air temperature as one would expect. How much higher? Well that depends on the convection coefficient. If the convection coefficient is very high then the body temperature is mainly determined by the air temperature. In contrast, if the convection coefficient is very low then the solar radiation can contribute significantly to the body temperature.

It's time to start using **R** to explore this model. For instance, we can use **R** to plot the relationship between the body temperature and the solar radiation. First we create variables for the heat coefficient and the air temperature and assign some values:

¹The concept of equilibrium is very important in ecological theory. It is often introduced in the context of differential equations and corresponds to the point at which the derivative of the variable of interest is zero. The heat equation can also be seen as a differential equation where f corresponds to the derivative of the amount of heat Q in the lizard through time t , i.e. dQ/dt .

```
k = 50 #convection coefficient (cal/h/°C)
a = 18 #air temperature (°C)
```

We want to plot the equilibrium body temperature for a range of solar radiation values. So we create a vector with radiation values, for instance ranging from 0 cal/h to 1500 cal/h in steps of 500. I am going to use big letters to denote vectors in **R** code, in contrast with scalars which I will denote with small letters.

```
Q = seq(0,1500,by=500) #vector with radiation values
```

Now we can write Equation 1.2 in **R** to produce a vector of the equilibrium body temperatures for each value of radiation.

```
B_eq = Q/k+a #vector with equilibrium body temperatures
```

Let's examine the values of the vector, by binding the two vectors as a matrix,

```
rbind(Q,B_eq)
```

	[,1]	[,2]	[,3]	[,4]
Q	0	500	1000	1500
B_eq	18	28	38	48

This is nice as we can see the values of the equilibrium body temperature for each value of solar radiation. But let's visualize this as a graph, by plotting these vectors in **R**,

```
plot(Q,B_eq,type="l")
```

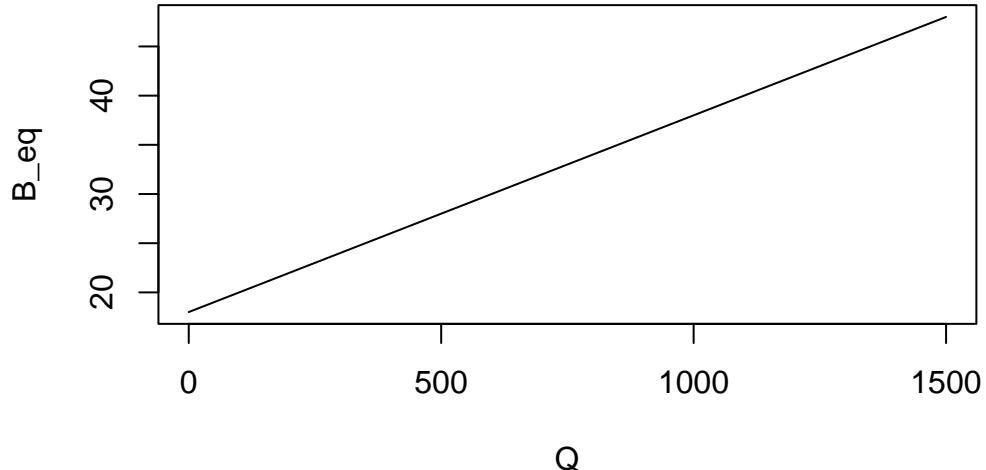


Figure 1.3: Equilibrium body temperature as a function of solar radiation.

Note that we added `type="l"` as a parameter of the plot to have a line drawn instead of a sequence of dots in the plot.

1.2 Understanding the climate space

Another way of looking at the heat exchange equation Equation 1.1 is to look at what combinations of air temperature and solar radiation values are livable for the lizard. This is also known as the climate space of an organism. In order to explore the climate space of the lizard we need to first assess what are the maximum and minimum body temperature that the lizard can experience. Let's assume those are respectively 36 and 24°C and store them in **R**,

```
b_max = 36 #maximum body temperature (°C)
b_min = 24 #minimum body temperature (°C)
```

We now want to solve Equation 1.1 at equilibrium for the air temperature as a function of the solar radiation and body temperature. We can do this by rearranging Equation 1.2,

$$a = b - q/k. \quad (1.3)$$

This equation can be then used in **R** to calculate vectors of the maximum and minimum survivable air temperatures for each value of the solar radiation in vector **Q**,

```
A_max = b_max - Q/k #vector with maximum air temperatures
A_min = b_min - Q/k #vector with minimum air temperatures
```

We can now visualize the climate space, this is, the combination of solar radiation and air temperatures in which the lizard can survive, by plotting these two equations (i.e. by plotting lines with x coordinates given by the vector **Q** and the y coordinates given by the vectors **A_max** and **A_min**),

```
plot(Q, A_max,type="l", #plots the maximum survivable air temperature
      xlab="Solar radiation (cal/h)", #adds x and y axis labels to the plot
      ylab="Air temperature (°C)")
lines(Q,A_min) #adds line for the minimum survivable air temperature
```

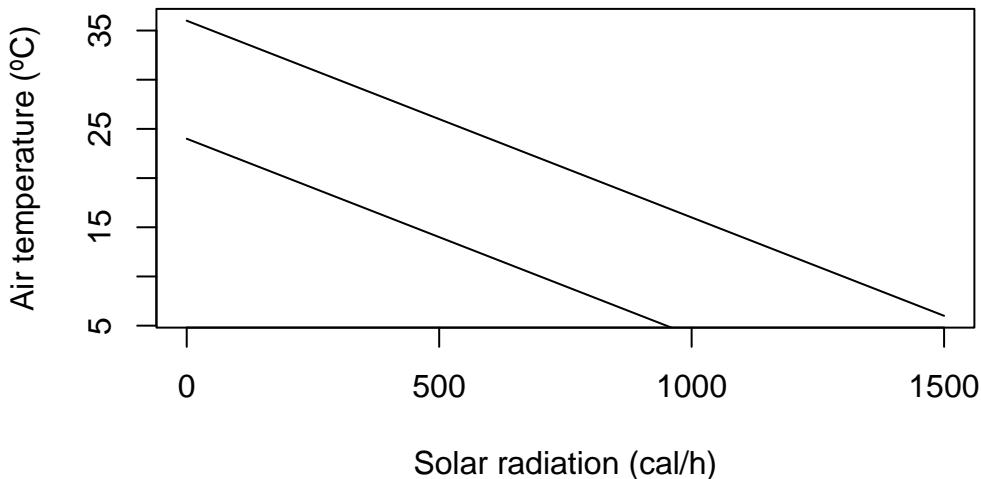


Figure 1.4: The climate space of a lizard.

Let's paint the area between the two lines which corresponds to the climate space of the lizard. We can use the **R** function `polygon`, to which we need to give the set of `x` and `y` coordinates delimiting the climate space as vectors. For instance, the upper lower corner can be the first coordinate, being 0 for the `x` vector and `b_min` for the `y` vector. The right upper corner is 1500 for `x` and `b_max-1500/k` for `y`.

```
X = c(0,0,1500,1500)
Y = c(b_min,b_max,b_max-1500/k,b_min-1500/k)
polygon(X,Y,col="green")
```

Now that we have the climate space we can plot on top of it some empirical data of how the conditions are in the field during the day. We can for instance consider two micro-habitats, a rock (in the sun) and a bush (in the shade). Suppose we obtain some data from temperature loggers that were installed in each micro-habitat , recording every three hours the values of temperature and radiation as tabled below.

Table 1.1: Hypothetical radiation values in two micro-habitats and air temperatures at different times of the day.

Time (hh:mm)	Radiation rock (cal/h)	Radiation bush (cal/h)	Temperature (°C)
00:00	150	150	18
03:00	150	150	13
06:00	800	450	10
09:00	1100	600	14
12:00	1300	650	21
15:00	1200	650	24
18:00	800	350	22
21:00	400	200	20

We can overlay these values of temperatures and radiations on the plot to understand which micro-habitat should the lizard choose at each time of the day. First we create a vector for each column of Table 1.1. Note that we must enclose the times of the day in commas as they are strings. We also append at the end of the vector the values for 0:00 in order to close the lines (otherwise there would be a gap between 21:00 and 0:00).

```
#Times of the day
T = c("00:00","03:00","06:00","09:00",
      "12:00","15:00","18:00","21:00","00:00")

#Solar radiation in the rock habitat
Rock_q = c(150,150,800,1100,1300,1200,800,400,150)

#Air temperature in the rock habitat
Rock_a = c(18,13,10,14,21,24,22,20,18)

#Solar radiation in the bush habitat
```

```
Bush_q = c(150, 150, 450, 600, 650, 650, 350, 200, 150)

# Air temperature in the bush habitat
Bush_a = c(18, 13, 10, 14, 21, 24, 22, 20, 18)
```

We overlay these vectors in the figure by invoking the function `lines(Xcoor,Ycoord)` for each micro-habitat. We add labels to each point to show the correspondence between each point and the time of the day.

```
lines(Rock_q,Rock_a,col="orange")
text(Rock_q,Rock_a,T)

lines(Bush_q,Bush_a,col="blue")
text(Bush_q,Bush_a,T)
```

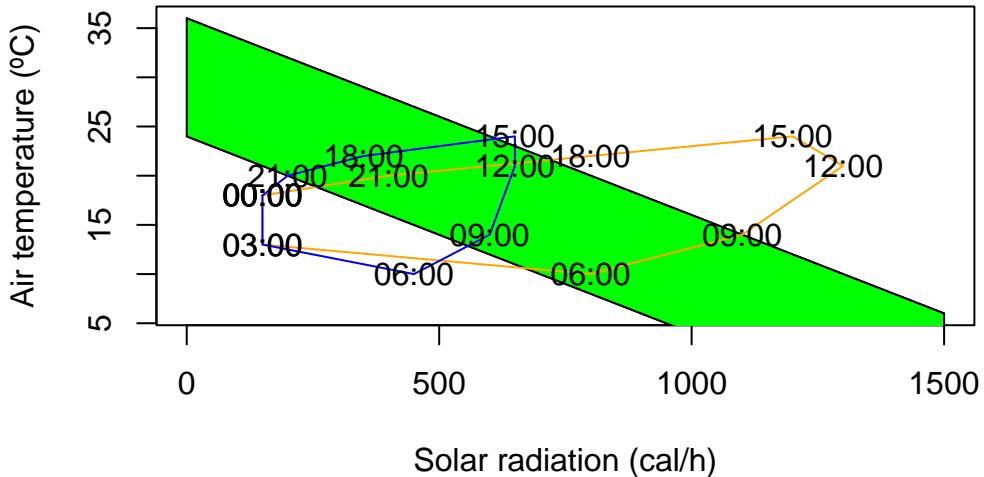


Figure 1.5: The climate space of a lizard with the conditions of two microhabitats plotted at different times of the day: rock (orange), bush (blue).

1.3 From ecophysiological models to species distribution models

In the previous section we developed a simple mechanistic model for the climate space of an organism. This is a micro-habitat level climate space. But one can also infer the climate space from the distribution of a species at the macro-habitat level. They are different but they are conceptually similar. The inference of such macro climate space is an area where species distribution models excel (for more on species distribution models check for example (Guisan, Thuiller, and Zimmermann 2017)). The idea is relatively simple. One starts with a bunch of locations of a species in geographical space. Then, one obtains the climate characteristics for those locations, and then plots those locations in climate space. The convex hull in climate space delimited by those locations is the

macro-habitat climate space. What species distribution models (SDMs) learn to do is to delineate that climate space, this is, SDMs try to predict the probability of a species occurring somewhere in the climate space by learning from presences and absences from species. Then the SDM can extrapolate again what happens in geographical space, by predicting the species occurrence probability for any point based on the climate data for that point. Without getting all the way into a full blown SDM, let's try to follow some of the basics.



Figure 1.6: The Iberian emerald lizard, *Lacerta schreiberi*.

I will illustrate this with one of my favorite species, the Iberian emerald lizard, *Lacerta schreiberi*, a species endemic to the Iberian peninsula, often found near water streams in mountain landscapes, but also in other habitats. First we obtain the presences for a species from GBIF (Global Biodiversity Information Facility, <http://www.gbif.org>). GBIF is a repository for biodiversity observations, and has over three billions of occurrences of over a million species all over the world at the time I am writing these words. Natural history museums, citizen science platforms such as iNaturalist (<http://www.inaturalist.org>) and eBird (<http://www.ebird.org>), and scientists, publish their species records in GBIF using a data format called Darwin Core. There is a R package that allows one to download observations from GBIF and also other relevant geodata, the **geodata** package. We will be doing some geospatial processing, so let's load also the **terra** package.

```
library(geodata)
library(terra)
library(predicts)
library(ggplot2)
```

We download the observations of *Lacerta schreiberi* from GBIF with the function `sp_occurrence(genus, species)`. We also download a world map with the function `world(path)`, where path should point towards a local directory where we want to store the data.

```

occ <- sp_occurrence("Lacerta","schreiberi")
world_data <- world(path = ".") #download world countries limits
ext_eur <- c(-15,45,35,72) # coordinates of European extent

#download bioclim data
bioclim <- worldclim_global(var = 'bioc', res = 10, path = ".")

# plot the map of Europe
plot(world_data,
      xlab = "Longitude", ylab = "Latitude", axes = TRUE, ext=ext_eur)

# add the species occurrences
points(occ$lon,occ$lat,cex=.1,col="blue")

# create pseudo-absences
eumask<-crop(bioclim[[1]], ext(ext_eur))
absences<-backgroundSample(eumask,nrow(occ),vect(occ),tryf=10)
points(absences,cex=.1,col="red")

# plot the climate variables in geographic space
plot(bioclim[[19]],ext=ext_eur)
plot(bioclim[[3]],ext=ext_eur)

# plot the climate space figure
# first merge occurrences and absences into a single matrix
allpts <- rbind(cbind(occ$lon,occ$lat,rep(1,nrow(occ))),
                 cbind(absences,rep(0,nrow(absences))))
colnames(allpts)[3]<-"pres"

# extract the climate variable values for all points
bioclim_sample<-extract(bioclim,allpts[,1:2])
allpts_bioclim <- cbind(allpts,bioclim_sample)

allpts_bioclim_plot<-allpts_bioclim
allpts_bioclim_plot$pres<-as.factor(allpts_bioclim_plot$pres)
ggplot(allpts_bioclim_plot, aes(x=wc2.1_10m_bio_19,y=wc2.1_10m_bio_3,shape=pres, col=pres)) +
  geom_point(size = 3, alpha=0.5) + scale_color_manual(values = c("0" = "red", "1" = "blue"))

```

1.4 Statistical confrontation: regressing the climate space

By Andres Marmol

Why anoles? Anoles are among the most studied reptile organisms. They are vastly diverse with more than 488 species described until today — circa 11% of all the squamates without considering

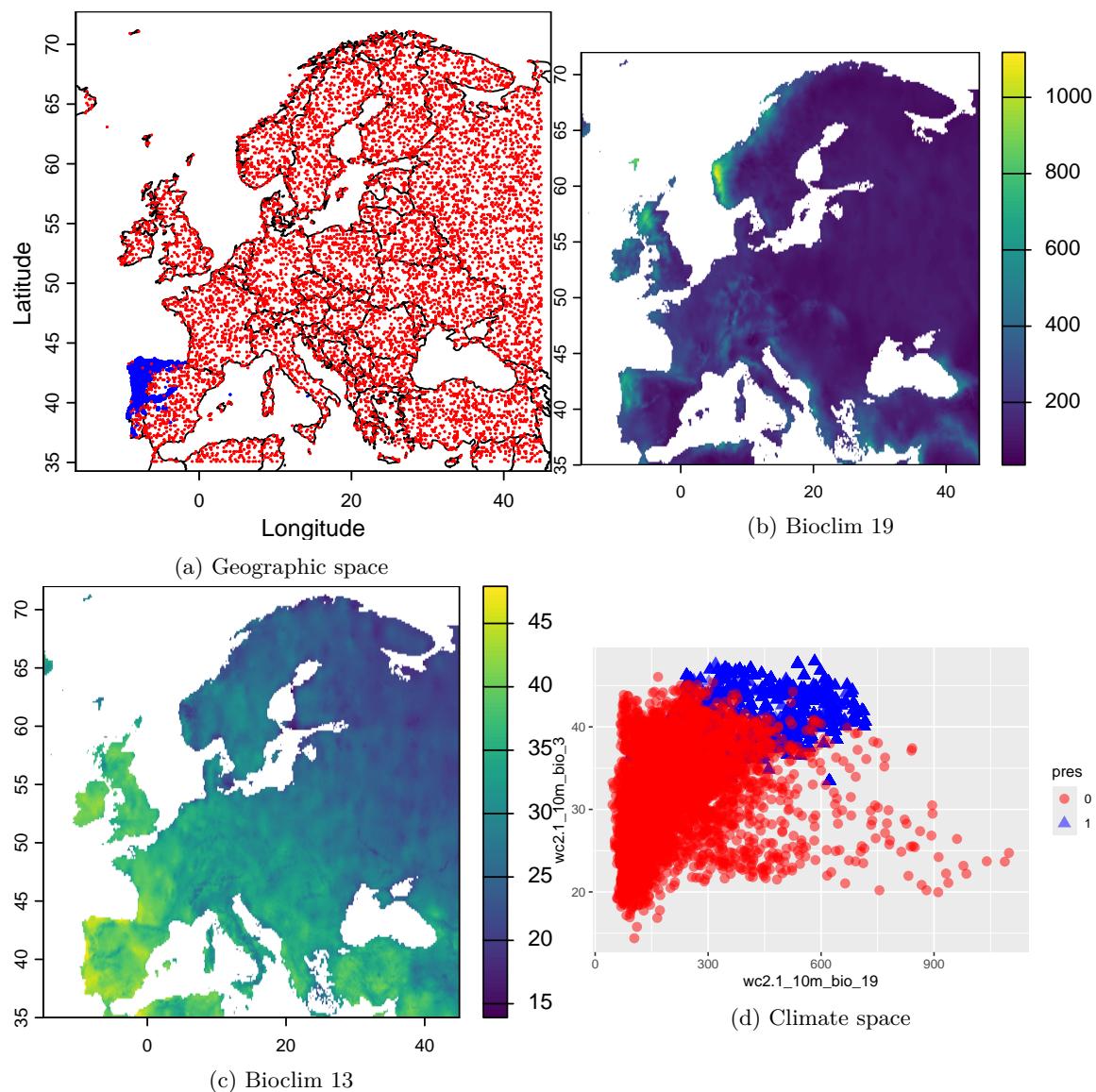


Figure 1.7: The geographic space and climate space of *Lacerta schreiberi*.

snakes — due to their outstanding adaptive radiation, which has inspired many to dive-in-depth into understanding how evolution has operated within this group (Lizards in an Evolutionary Tree by J. B. Losos (2009) is a highly recommended instructive reading for more info). As a result, very detailed data sets on many species of anoles, including information on micro habitat, field body temperature, activity patterns and morphological traits, performance, etc., can be found online in relatively few time.

In the following steps we will be downloading from Dryad and using a data set created by Winchell et al. (2016) on *Anolis cristatellus* making a pair-comparison of several morphological and thermal traits of individuals living in urban and forested areas at Puerto Rico. *A. cristatellus* is widely distributed across Puerto Rico and can be found in natural and human-intervened spaces easily. It can be seen over ground and laying on the trunks of trees in natural settings, and on walls and metal fences in more urban areas.

1.4.1 Calling the dataset from Dryad

To download the dataset, it is necessary to install the R package `rdryad`.

```
install.packages("rdryad")
```

Then we open the newly installed package and download the dataset by specifying the DOI associated to this dataset.

```
#calling rdryad
library(rdryad)

# Specify the DOI
doi <- "10.5061/dryad.h234n"

# Download the dataset using the dryad_download function
downloaded_file <- dryad_download(dois = doi)
```

The data sets are now downloaded on the paths indicated in the box below, although for this exercise only table [2] `winchell_evol_phenshifts.csv` is needed.

```
# The downloaded_file will contain the file path where the dataset is saved
print(downloaded_file)

$`10.5061/dryad.h234n`
[1] "/Users/henrique/Library/Caches/R/rdryad/10_5061_dryad_h234n/winchell_evol(CG).csv"
[2] "/Users/henrique/Library/Caches/R/rdryad/10_5061_dryad_h234n/winchell_evol_phenshifts.csv"
```

Once this is done, it is always good practice to check if the recently open data frame works by checking the columns within the data frame.

```
#calling the dataset of interest as df
df <- read.csv(downloaded_file[[1]][2])

# checking the columns (variables) available in the data frame df using the str function
names(df)
```

```
[1] "ID"                  "date"                 "Site"
[4] "context"              "perch"                "bodytemp.C"
[7] "perch.temp.C"        "ambient.temp.C"    "humidity.percent"
[10] "perch.height.cm"     "perch.diam.cm"     "weight.g"
[13] "head.height.mm"      "svl.mm"               "local.time.decimal"
[16] "flags"                "JL"                  "JW"
[19] "METC"                 "RAD"                 "ULN"
[22] "HUM"                  "FEM"                 "TIB"
[25] "FIB"                  "METT1"               "METT2"
[28] "FL"                   "HL"
```

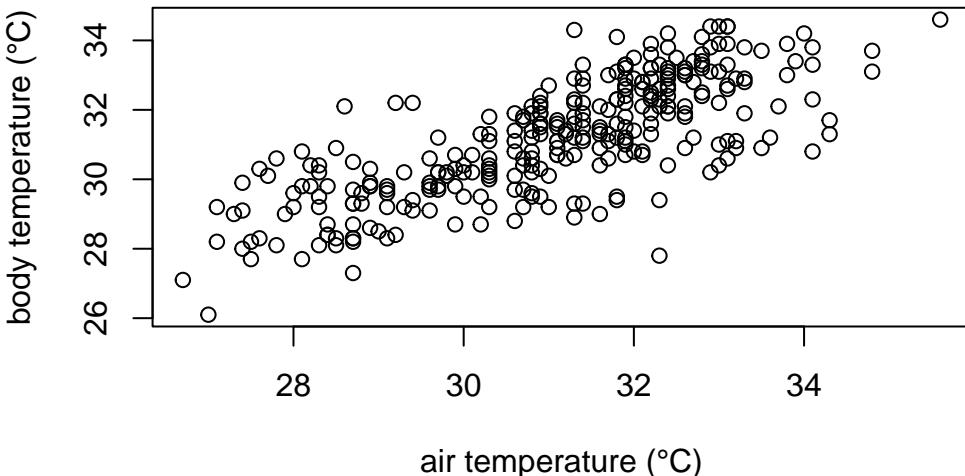
There are 29 different variables within `df`, but this exercise will focus only in a few of them including `context`, `perch`, `bodytemp.C`, `perch.temp.C`, `ambient.temp.C`, and `local.time.decimal` where `context` indicates if the lizard was captured in natural or urban spaces; `perch` refers to the micro-habitat where the lizard was found; `bodytemp.C` is body temperature b ; `perch.temp.C` is substrate temperature T_s ; `ambient.temp.C` is air temperature a ; and `local.time.decimal` is the time of the day in hours when the lizards was captured and measured.

1.4.2 Exploring the thermophysiological data of *A. cristatellus*.

As covered in the first chapter, b increases proportionally to a as indicated in equation (1.2). Let's check if the data confirms this equation.

```
# to call a variable directly from a data frame one must use $, indicating the name of the data frame
```

```
plot(df$ambient.temp.C, df$bodytemp.C,
  xlab="air temperature (°C)",
  ylab="body temperature (°C)")
```



In the plot we can observe that indeed body temperature increases with increasing temperature. However, we cannot quantitatively know how much is this increase. For that, we can model how air temperature affects body temperature modelling them a a line with the equation:

$$\hat{y} = \alpha + \beta * x + e$$

where \hat{y} is the predicted value of y for any given x ; α is point of intersection of the line when $x = 0$, β is the slope of the regression line (i.e. how much increases \hat{y} with a unit increase in x) and e is the random error.

Placing our variables in formula, it would look like this:

$$b = b_0 + \beta * a + e$$

where b is body temperature, a is ambient temperature, and b_0 is the body temperature when a is zero.

Now lets model it using a least-squares linear regression using the `lm` function in R.

```
#define the linear model
#the variable at the left of the tilde (~) is the dependent variable
#data calls the specific dataframe to be used.

model1 <- lm(bodytemp.C ~ ambient.temp.C, data=df)

# print model outputs
print(model1)
```

Call:

```
lm(formula = bodytemp.C ~ ambient.temp.C, data = df)
```

Coefficients:

(Intercept)	ambient.temp.C
8.7004	0.7237

1.4.3 Interpreting the results of our model.

The results of our model indicates that if the air temperature (a) is zero, *A. cristatellus* would show a body temperature (b) of 8.7 °C, which increases in approximately 0.7 °C for each 1 °C increase in air temperature.

Now it is also possible to draw a few predictions

```
# a number of possible ambient temperatures that can be found in Puerto Rico during the day

amb_temps = data.frame(ambient.temp.C = seq(27,38, 1))# defines a vector with values from 27 to 38,
predict(model1, newdata=amb_temps)
```

1	2	3	4	5	6	7	8
28.23967	28.96335	29.68702	30.41070	31.13437	31.85805	32.58172	33.30540

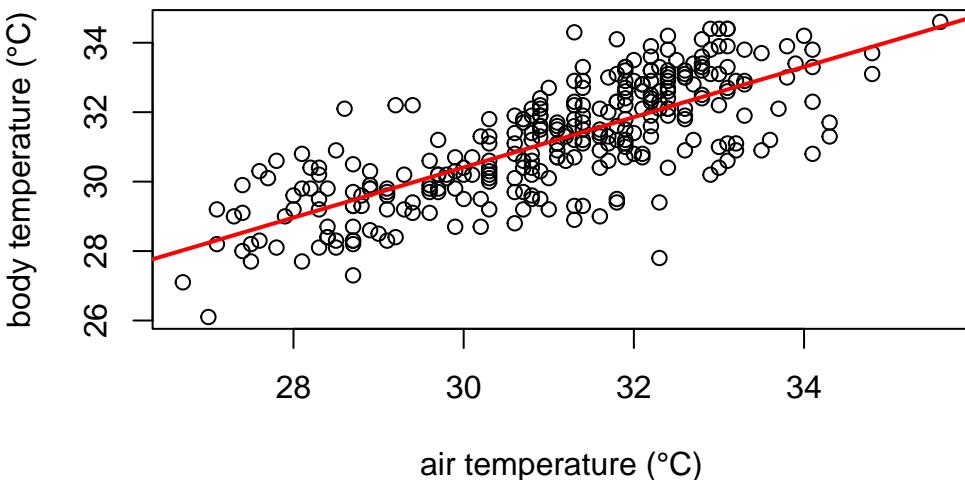
9	10	11	12
34.02907	34.75275	35.47642	36.20010

1.4.4 How good our model is performing?

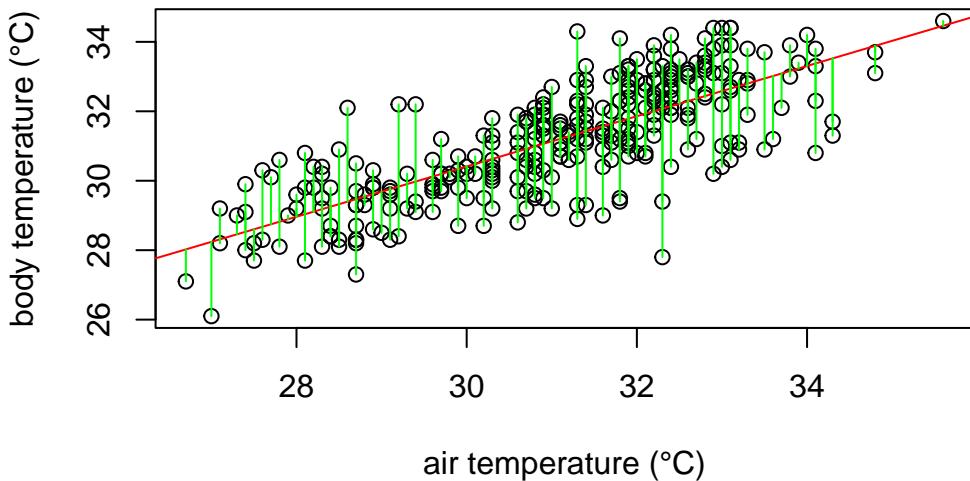
Graphically the linear model fits the data as follows:

```
# Create the scatter plot
plot(df$ambient.temp.C, df$bodytemp.C,
      xlab="air temperature (°C)",
      ylab="body temperature (°C)")

# add the line of fit from our recently created linear model "model1"
abline(model1, col = "red", lwd= 2)
```



The red line in the plot follows very well the pattern of increase shown by our data. It is noticeable, however, that some points fall farther from the line of best fit than others. In other words, many \hat{y} values (body temperature) are predicted with larger errors than others, for a given x value (ambient temperature). These errors are also known as residuals, and are the vertical distance between the observed values and the line of best fit. They are graphically represented in green in our plot:



What a least square linear regression does is to find the lowest sum of squares of the residuals (every green line in the plot) so that the least amount of error is kept in the linear model.

Finally, how well the model fits the data? In our example, the body temperatures do not perfectly fall on the line of best fit, and many of them are scattered around it. A common metric assess this question in a simple regression model is the r^2 . r^2 values indicates how much of the variance of body temperature (our dependent variable) is explained by ambient temperature (our independent variable). A perfect fit is indicated by an r^2 of 1, while no fit would be zero. To check the r^2 in our model we can simply use the `summary` function in R.

```
# obtaining the r-squared of our model
summary(model1)$r.squared
```

[1] 0.5736088

r^2 equals 0.574 indicating that almost 60% of the variance in body temperature is explained by ambient temperature.

In summary, our expectation that lizards body temperature increases depending on how the temperature in their environment changes has been confirmed. Specifically for *A. cristatellus* in this study, body temperature increases ~ 0.7 °C every 1 degree increase in air temperature. A few questions remain. For example, does habitat has an effect in the body temperature in *A. cristatellus*, and if it does, how different could the body temperatures of the lizards in habitat A are from those in habitat B?

Chapter 2

Economic models of behavior: do animals optimize?

Animals have to make decisions all the time: where and when to move, when and what to eat, whom to mate, etc. How do animals make those decisions? One approach to study this question is to assume that animals take decisions that maximize their fitness. This approach is rooted in the idea that natural selection, under certain conditions, maximize the average fitness of individuals in a population. It underpins many of the studies in behavioral ecology, and lead to the development during the 1970's and 1980's of a sub-field dedicated to the study of foraging decisions, known as optimal foraging (see for instance Pyke 1984). The idea of optimal foraging is to develop economic models of foraging behavior, assessing the costs and benefits of different actions, and identifying in any particular circumstance the action that maximizes benefits and minimizes costs. This is, animals are seen as optimizer of decision-making. Here we consider models for two "idealized" types of foragers: searching predator and sit-and-wait predators.

2.1 Searching predator

The searching predator is typified as an animal that actively searches for food. For instance, wolves roam the landscape constantly searching for their prey (Figure 2.1). The question is then, any time that a forager comes across a prey whether to spend the time catching and eating it, or continuing to search for another prey, for instance a larger or easier prey item. Joan Roughgarden also liked this type of foraging to the sushi-bar problem where dishes are coming one after the other on a moving tray, and a person needs to decide whether to pick a given dish. This decision is particularly akin to the searching predator situation when one assumes that a person can have only one dish at a time in her or his table. We consider two types of searching predators: time minimizers and energy maximizers.

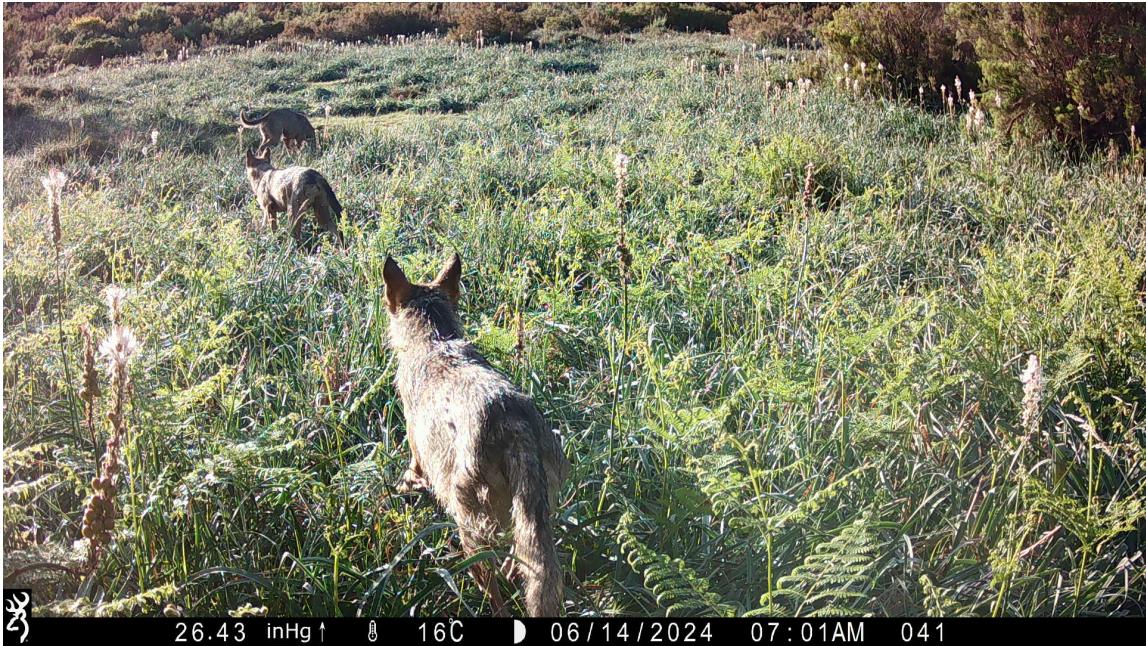


Figure 2.1: A pack of wolf (*Canis lupus*) roaming around the landscape of Peneda-Gerês National Park, Portugal.

2.1.1 Time minimizer

Many searching predators are themselves preys to other animals. For instance shrews search for insects to eat, but can be themselves eaten by carnivores or owls. Therefore a reasonable assumption is that they are making foraging decisions that minimize the time foraging. Let's assume there are two types of prey items, type 1 and 2. These two preys occur at different abundances in the environment, let's name them a_1 and a_2 . These abundances can be measured as encounter rates from the predator perspective, this is the number of prey items found per unit time. The two types of prey also have different handling times, this is the amount of time required to chase and process the prey, h_1 and h_2 . We also convention to call the type 1 prey the prey with the lowest handling time, i.e. $h_1 < h_2$,

A searching predator can adopt one of the following three strategies:

- Strategy 1: to consume only prey items of type 1
- Strategy 2: to consumer only prey items of type 2
- Strategy 1&2: to consumer both types of prey

In order to find out which strategy should be adopted by the forager, one needs to calculate the average spent per food item in each of the strategies. For Strategy 1 the average time per item, T_1 is the sum of the amount of time the predator needs to encounter a prey with the amount of time that it takes to process that prey. As the abundance is measured in encounter rates, i.e. prey items

per unit time, the inverse of that is the waiting time for a prey. Therefore we have for Strategy 1,

$$T_1 = 1/a_1 + h_1 \quad (2.1)$$

Similarly, for Strategy 2, the average time per item T_2 is given by

$$T_2 = 1/a_2 + h_2. \quad (2.2)$$

A more interesting case is Strategy 3. Here the waiting time is the inverse of the sum of the abundances of both times of prey, while the handling time is average of the handling times of both types of prey weighted by their relative abundance,

$$T_3 = \frac{1}{a_1 + a_2} + \frac{a_1 h_1 + a_2 h_2}{a_1 + a_2} \quad (2.3)$$

We can now define the problem that the forager has to solve as to find the strategy i that minimizes T_i :

$$\min(T_i) \quad \text{for } i = 1, 2, 3$$

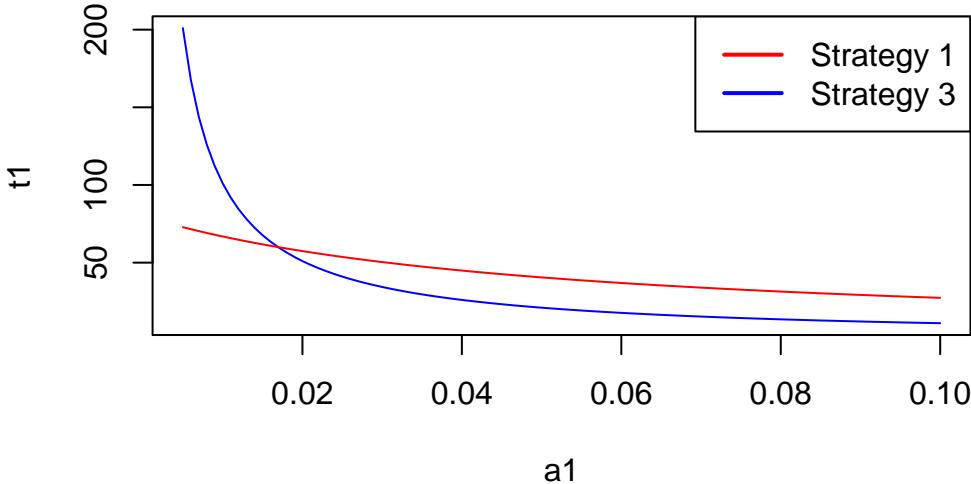
It is easy to demonstrate that Strategy 2 is always worse than Strategy 3, independently of the abundances of the two types of prey. This happens because Strategy 3 always implies a longer waiting time than the two other specialized strategies (i.e. one has to wait less time to find any item of an prey than items of a given prey time, $1/(a_1 + a_2) < 1/a_2$) and the handling time of strategy 3 can never be higher than the handling time of strategy 2 (it's always a value between h_1 and h_2). So we can exclude Strategy 2 from our analysis. The choice is then between Strategy 1, just taking items of the preferred prey item, and 3, taking items of both types of prey. Let's assess this two strategies with a little bit of help from R. We start by assuming that the handling time of prey type 1 is 1 second while prey type 2 takes 60 seconds. Let's also assume that the abundance of prey type 2 is 0.05 individuals per second, i.e. one individuals needs to wait in average 20 seconds to find prey type 2.

```
h1=1          #Handling time of prey type 1 (s)
h2=60         #Handling time of prey type 2 (s)
a2=0.05       #abundance of prey type 2 (ind/s)
```

Let's now plot the time per item of each of the strategies as a function of the abundance of the preferred prey.

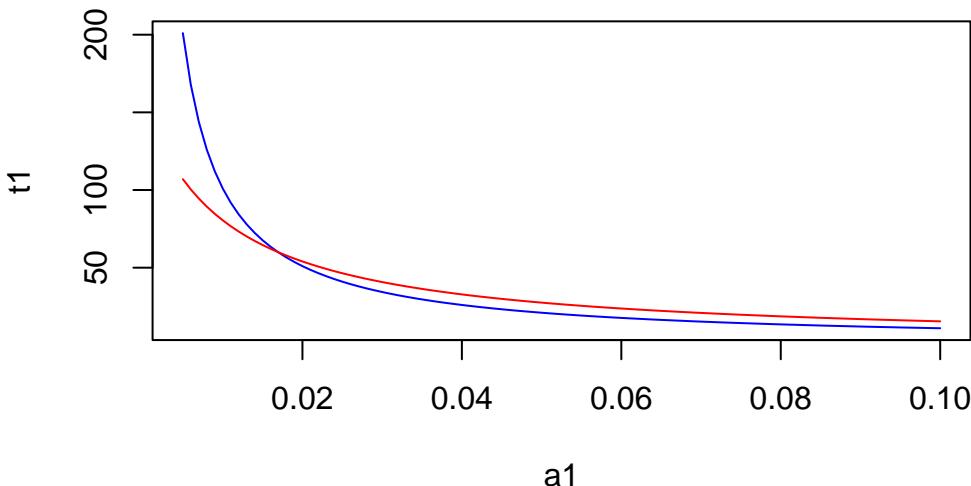
```
a1<-seq(0.005,0.1,0.001)    #abundance of prey time 1 (ind/s)
t1=1/a1+h1                  #time per item of Strategy 1 (s)
t3=1/(a1+a2)+h1*a1/(a1+a2)+h2*a2/(a1+a2) #time per item of Strategy 2 (s)
plot(a1,t1, type="l", col="blue")
lines(a1,t3, type="l", col="red")
legend("topright",
      legend = c("Strategy 1", "Strategy 3"), # Labels
```

```
col = c("red", "blue"),          # Line colors
lwd = 2,                      # Line width
lty = 1)
```



There is a critical threshold of the abundance of prey type 1 above which strategy 1 is preferable, while below that threshold strategy 3 is the best strategy. Interestingly this threshold does not depend on the abundance of the less preferable prey. For instance, if we assume a low abundance of prey time 2 at 0.01, the resulting plot is:

```
a2 = 0.01
t1=1/a1+h1           #time per item of Strategy 1 (s)
t3=1/(a1+a2)+h1*a1/(a1+a2)+h2*a2/(a1+a2) #time per item of Strategy 2 (s)
plot(a1,t1, type="l", col="blue")
lines(a1,t3, type="l", col="red")
```



To determine this critical threshold one can compare the two vectors, T1 and T3, and find the first position at which T1 becomes smaller than T3,

```
pos=which(t1<t3)[1]
a1[pos]
```

[1] 0.017

So the critical threshold for these handling times occurs when $a_1 = 0.017$ individuals per second.

2.1.2 Energy per time maximizer

Perhaps more often, animals try to maximize their energy yield (benefits) while minimizing the time foraging (costs). Or in another way of looking at it, they try to maximize their energy yield per unit time. We already know the time per item associated to each of the three strategies of the searching predator. We now need to calculate the average energy yield per item. Consider now that the energy content of the prey items are e_1 and e_2 for prey of type 1 and 2, respectively. We define prey 1 as the preferred type of prey, so we assume that the ratio of the energetic content (measured for instance in calories) to the handling time is higher for type 1 prey, i.e. $e_1/h_1 > e_2/h_2$. Now we calculate the energy yield per item for each strategy. We start with the energy content of the prey, but need to subtract the energy spent while waiting the prey and the energy spent chasing and processing the prey,

$$E_1 = e_1 - ew * tw_1 - eh * h_1 \quad (2.4)$$

where ew and eh are the energy spent per unit time while waiting for the prey and the energy spent per unit time while handling, respectively. They can both be measured for instance in cal/s. We already know from the time minimizer that the waiting time for the prey is the inverse of the abundance, $tw_1 = 1/a_1$. Therefore substituting in Equation 2.4 we have

$$E_1 = e_1 - \frac{ew}{a_1} - eh * h_1. \quad (2.5)$$

A similar expression can be written for Strategy 2, replacing 1 with 2 in Equation 2.4.

$$E_2 = e_2 - \frac{ew}{a_2} - eh * h_2. \quad (2.6)$$

More interesting is to derive the expression for Strategy 3, where the foragers takes both types of prey. The energy content of the prey is the average of the energetic contents of each prey type, weighted by their abundances, $(e_1 * a_1 + e_2 * a_2)/(a_1 + a_2)$. The waiting time is the inverse of the sums of the abundances of preys of both types, as in Equation 2.3. The handling time is the average of the handling times of each prey type, weighted by their abundances. So we have,

$$E_3 = \frac{e_1 * a_1 + e_2 * a_2}{a_1 + a_2} - \frac{ew}{a_1 + a_2} - eh \frac{h_1 * a_1 + h_2 * a_2}{a_1 + a_2}. \quad (2.7)$$

Finally we can calculate the energy per unit time for each strategy by dividing Equation 2.5 by Equation 2.1 for Strategy 1, dividing Equation 2.6 by Equation 2.2 for Strategy 2, and dividing Equation 2.7 by Equation 2.3 for strategy 3,

$$ET_i = \frac{E_i}{T_i}$$

Similarly to the time minimizer, it's possible to show mathematically that strategy 2 is never an optimal strategy. So the interesting comparison is again between strategy 1 and strategy 3. Let's use R to plot the energy per time yield for both strategies. We start by setting the parameter values of our model with some realistic numbers.

```
e1<-10          #Caloric content of prey 1
e2<-100         #Caloric content of prey 2
h1<-1           #Handling time of prey 1
h2<-60          #Handling time of prey 2
ew<-1           #Energy spend per unit time while waiting (cal/s)
eh<-1           #energy spend per unit time handling the prey (cal/s)
```

We will examine the energy yields for strategy 1 and strategy 3 for a fixed abundance of prey type 2 and a sequence of abundances of prey type 1 from 0.005 individuals per second to 0.5 individuals per second

```
a1<- seq(0.005,0.5,0.001)    #Sequence of abundances of prey 1
a2<-0.05
```

With the parameter and abundance values defined, we can calculate E_1 , E_3 , T_1 , T_3 , and then ET_1 and ET_3 based on the equations above, resulting in vectors for these variables with each entry in the vector corresponding to an abundance value in the vector of abundances `a1`.

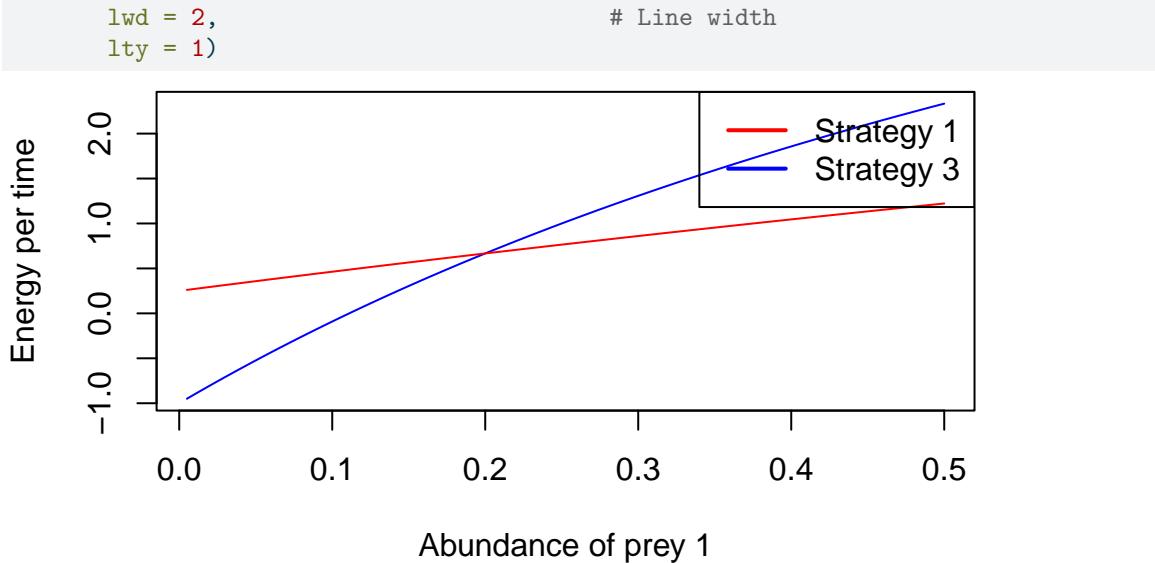
```
E1 = e1 -(eh*h1) - ew/a1    #energy per item of Strategy 1 (s)
E3 = (e1*a1+e2*a2)/(a1+a2)-
      eh*(h1*a1+h2*a2)/(a1+a2)-ew/(a1+a2) #energy per item of Strategy 3 (s)

T1=1/a1+h1                  #time per item of Strategy 1 (s)
T3=1/(a1+a2)+h1*a1/(a1+a2)+h2*a2/(a1+a2) #time per item of Strategy 3 (s)

ET1 = E1/T1 #energy per time of strategy 1
ET3 = E3/T3 #energy per time of strategy 3
```

Next we plot the energy yields against the values of abundance of prey of type 1, and add a nice legend:

```
plot(a1,ET1,type="l",xlab="Abundance of prey 1", ylab="Energy per time", col="blue")
lines(a1,ET3,type="l",col="red")
legend("topright",
       legend = c("Strategy 1", "Strategy 3"), # Labels
       col = c("red", "blue"),                 # Line colors
```



Similarly to the the time minimizer, for the energy maximizer there is also a critical threshold of the abundance of prey type 1 above which strategy 1 is preferable, while below that threshold strategy 3 is the best strategy.

2.2 Sit-and-wait predator

In contrast to the searching predator, the sit-and-wait predator forages by patiently ambushing its prey. For instance, the lizard *Anolis gingivinus* (Figure 2.2) waits in a perch, often a tree trunk, for a prey to come into its reach, sprinting then down the trunk or into the ground to capture its prey, returning then to its perch again. So here the decision that the predator has to take after seeing the prey is whether it should run an capture the prey or if it should ignore it.

We will build on the model we developed for the energy per time maximizer searching predator to develop a model for the decisions of the sit-and-wait predator. The energy per prey item consumed is,

$$E = e - e_p t_p - e_w t_w \quad (2.8)$$

where e_p is the energy per unit time while pursuing the prey, t_p is the average time it takes to sprint to the prey and come back to the perch, e_w is the energy per unit time while waiting for the prey, and t_w is the average time it takes to wait for a prey item to show up within the home-range or territory of the predator. We will use these two terms interchangeably, although they are sometimes used in the literature differently.

Let's consider that the home-range of the lizard is shaped as a semi-circle centered in the perch. This is the lizard has a viewing angle of 180 deg from its perch (Figure 2.3). Then the total abundance of the the prey in the territory is the integral of the prey point density over the territory. So, using polar coordinates for the integral this can be written as,



Figure 2.2: An *Anolis gingivinus*, a sit-and-wait predator, captures a prey after running down from its perch. Photo taken in the island of Saint Martin, Leeward Islands, Caribbean.

$$A = \int_0^{r_c} a\pi r dr$$

The waiting time is just the inverse of the total abundance.

$$t_w = 1/A = \frac{1}{\int_0^{r_c} a\pi r dr}$$

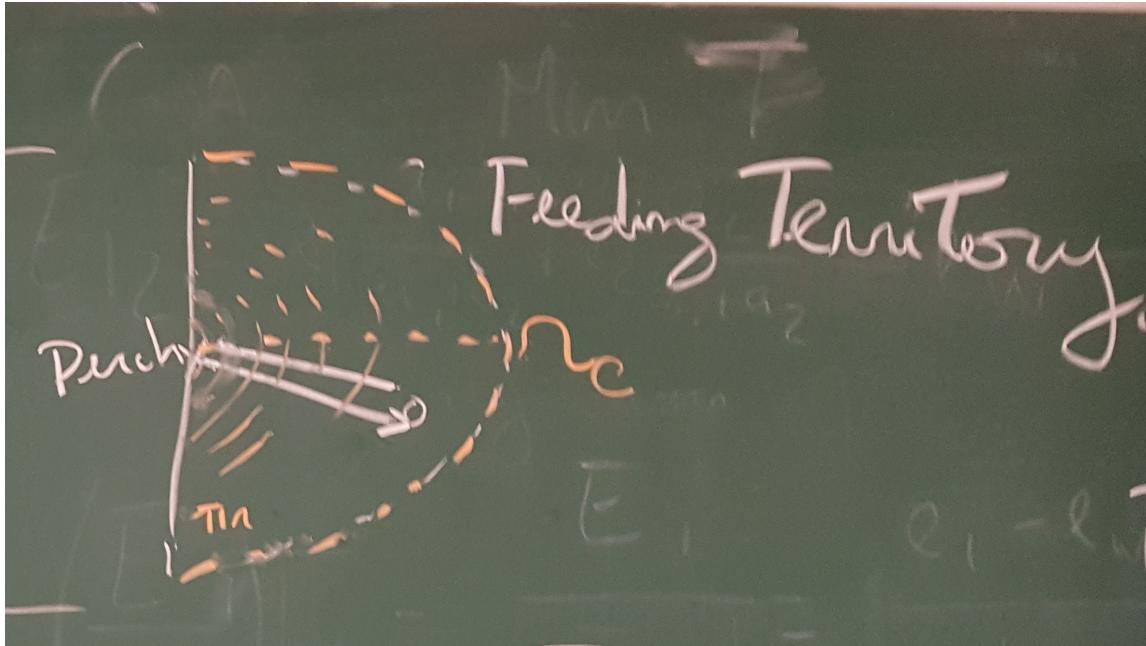


Figure 2.3: Feeding territory of a sit-and-wait predator with radius r_c .

The average pursuit time is a bit more complicated. For a prey landing at distance r from the predator, the pursuit time is the time the lizard needs to run to that location and back. This is calculated by dividing the double of the distance by the speed of the lizard, $2r/v$. We have now to average these sprint times over the range of possible distance that the lizard can run, from 0 to the radius of its territory r_c , weighted by the probability of a prey appearing at each distance. The probability of a prey appearing at distance r is the length of the semi-circle with radius r multiplied by the point density of prey divided by the total density of prey in the territory of the lizard, $a\pi r/A$. So we now need to calculate the integral of the sprint times at each distance times the probability of running to a prey at that distance over the range of possible radius values,

$$t_p = \int_0^{r_c} \frac{2r}{v} \frac{a\pi r}{A} dr$$

The average time per prey item consumed is simply the sum of the average pursuit time, t_p , and the average waiting time t_w ,

$$T = t_p + t_w \quad (2.9)$$

Therefore the energy per time that the optimal sit-and-wait predator wants to maximize is obtained by dividing Equation 2.8 by Equation 2.9,

$$ET(r_c) = \frac{e - e_p t_p(r_c) - e_w t_w(r_c)}{t_p(r_c) + t_w(r_c)}$$

where we highlighted that the waiting time and the pursuit time are functions of the territory size or cut-off radius r_c .

The optimal decision is to find the cut-off radius r_c that maximizes the energy per time,

$$\max_{r_c}(ET)$$

Let's plot the energy per time as a function of the territory size,

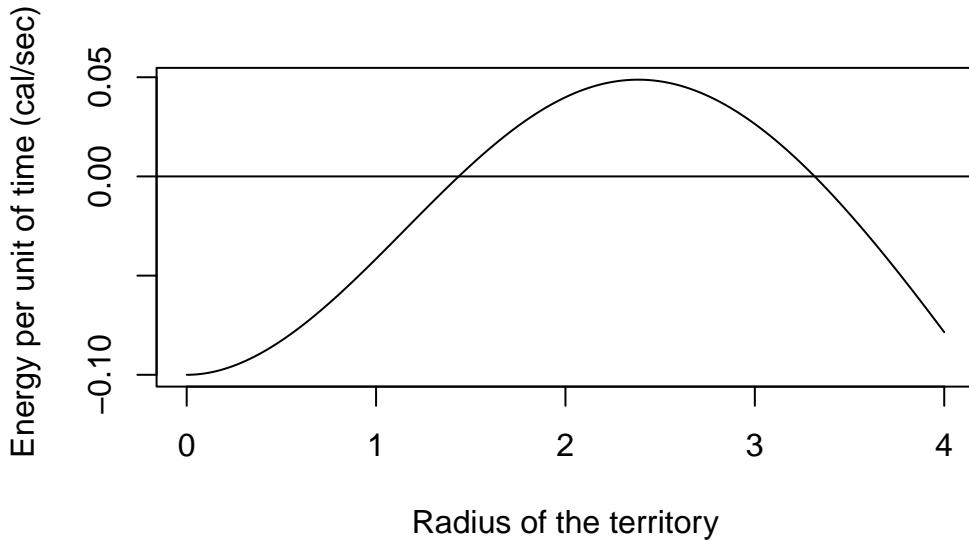
```
ew<-0.1          #Energy per unit of time of waiting
ep<-1            #Energy per unit of time of pursuing the prey
v<-0.5           #Velocity of the predator in chase
a<-0.005         #Abundance of prey
e<-10            #Energy value of the prey

rc<-seq(0.001,4,0.001)  #Radius of the hunting ground

tw <- 1/(a*pi*((rc^2)/2))  #Waiting time
tp<-4/3*rc/v               #Pursuing time
ti<-tw+tp                  #Time per item
ei<-e-ew*tw-ep*tp          #Energy per item
et<-ei/ti                  #Energy per unit of time

et <-function()
{
  (e-ew*(1/(a*pi*((rc^2)/2)))-ep*(4/3*rc/v))/(1/(a*pi*((rc^2)/2))+4/3*rc/v)
}

plot(rc,et(),type="l",
      xlab="Radius of the territory",
      ylab="Energy per unit of time (cal/sec) ")
abline(0,0)
```



If you wanna read about an empirical test of this model, I suggest the paper by Shaffir and Rougharden [[@-shafir1998](#)].

2.3 Does natural selection maximizes fitness?

- Simple models of population genetics for frequency independent selection.
- Something on Tinberger, Konrad Lorenz on the genetic basis of behavior

2.4 Statistical confrontation: finding the maximum

- Explain how to find the maximum of a function with R. Parallel between genetic algorithms and optimization.

2.5

Chapter 3

Conflict resolution in ecology and evolution: war and peace

In the previous chapter we looked at the decisions of individual animals assuming that the decisions were taken by each forager alone. For instance, for the sit-and-wait predator we assumed that the territory could be as large as the predator wanted, and that the optimal territory size was the one where the energy per unit time was maximized. But what happens if the territory is so large that it starts intruding in the territory of the neighbors? Or in other words, what happens if the optimal territory size is for instance 1 m^2 , but the density of the predators is so high that there is more than 1 predator per square meter? Then there is a potential conflict between individuals and it may not be possible to have territory sizes that are optimal. More generally, one can say that the decision of an individual now affects the decisions of its conspecifics.

In evolutionary contexts, where decisions or phenotypes are determined by genes, we enter the so called realm of frequency-dependent selection. In contrasts with frequency independent selection, where the fitness of a genotype depends only on the environment, in frequency-dependent selection the fitness of a genotype depends also on the frequency of the different genotypes in the population.

The analysis of this kind of behavior conflict or evolutionary dynamics is the topic of game theory, particularly non-cooperative game theory. Some of the key concepts of game theory for animal behavior were developed by the biologist John Maynard Smith in the 1970's and beautiful explained in his book "Evolution and the Theory of Games" Maynard Smith (1982). We now know that similar ideas were developed independently by the mathematician John Nash, twenty years earlier in the 1950's, ideas that would lead to Nash being awarded the Nobel Prize of Economics in 1994.

In this chapter we examine three non-cooperative games that model situations of behavior or evolutionary conflict: the prisoner's dilemma, the Hawk-Dove game, and the war of attrition. They are called non-cooperative because each "player" does not know what the other "player" is going to do in the game, and the players cannot discuss the strategy that they are going to take in any cooperative game. They are also sometimes described as closed envelope games. This is, each player should write what "strategy" it adopts in each play without knowing the choice of the other player.

3.1 The prisoner's dilemma and the evolution of cooperation

A lot of the emphasis on evolutionary biology and ecology is placed on competition, on the idea of “survival of the fittest” and that selection operates at the level of individuals favoring selfish behaviors. However, cooperation is super common in nature, from prairie dogs taking turns as sentinels, informing the group of the arrival of a predator, to back scratching in primates. One beautiful explanation for such cooperation is based on the principles of kin selection, developed by Bill Hamilton, or the idea that it makes evolutionary sense to help others if they are genetically related to you, as this increases the probability of the same genes you have to be passed into the next generation. Hamilton’s contribution to this problem was seminal because he noted that genetic relatedness can be particularly high between females in haploid-diploid systems such as bees and ants, leading to a widespread of cooperating in rearing offspring and even the abdication of many individuals from reproducing in favor of assisting the queen. But can cooperation also evolve in the absence of genetic relatedness? This is the problem that can be studied with the prisoner’s dilemma.

In the prisoner’s dilemma game there are two players. It is inspired by a situation where two suspects of being accomplices in a crime are arrested and are being questioned in separate rooms without contact between them. They are faced with the dilemma of cooperating with each other by claiming innocence, or cutting a deal with the investigators by recognizing the responsibility for the crime and by denouncing the other player. If both cooperate they can potentially get both away, but if one accomplice cooperates and the other defects, then the one that cooperated gets the maximum penalty.

We can formalize the prisoner’s dilemma game by stating that each player can choose one of two strategies in each round of the game, defect or cooperate. The pay-off matrix of the Prisoner’s Dilemma, from the perspective of player 1 (rows), playing against player 2 (columns) is

Player 1 \ Player 2		C	D
		C	0
C	3	0	
	5	1	

You can think of the pay-off points as a fitness gain in relation to the fitness before the play. The exact payoffs are not so important, and there are other variants of the game with different pay-offs. Instead, it is the relationship between the different pay-offs that determines the best strategy for the game. If the players knew what the other player would do, the best collective strategy would be to cooperate, as they both would receive 3 points for a total of 6 points. If one of the players defects while the other cooperates they accrue collectively only 5 points, while if both defect they would accrue together 2 points. But they don’t know what the other player is going to do (non-cooperative game) and need to make a decision that maximizes their individual fitness. What should they do?

The general solution that both Maynard Smith and Nash came up for these games is that one needs to identify which strategy is the best response to itself. This is called an Evolutionary Stable Strategy (ESS, also known as Nash Equilibrium¹) because it is the strategy that when dominant in the population cannot be invaded by any other strategy. Mathematically speaking, the ESS is the strategy S^* that has the higher payoff when played against itself,

¹There’s a small difference between the two, see (Dugatkin and Reeve 1998).

$$\forall S_i, \quad E(S^*, S^*) \geq E(S_i, S^*)$$

3.2 The Hawk-Dove game and animal contests

3.3 The war of attrition

Part II

Ecology of populations

Chapter 4

Demography: boom or bust

Part III

Ecology of communities

Chapter 5

Modelling biodiversity change: winners and losers

Every minute about 2 football fields of native forest are cleared somewhere in the world. What are the consequences of land-use change for biodiversity?

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Appendix A

A brief R tutorial

If you are new to **R** you can have a short dive into its main features by working through this tutorial. If you had learnt programming in another computer language, you will be able to skim over this tutorial to find the main differences from what you have learnt to how things are done in **R**.

A.1 Basic data structures

A.1.1 Variables

Variables can be any sequence of letter and numbers, but `#` it cannot start with a number

```
x = 2  
x <- 4  
2+2
```

```
[1] 4  
y <- x^5  
y
```

```
[1] 1024
```

Please note that you can comment code by using the `#` character.

A.1.2 Vectors

Let's create vectors.

```
# Introduction to vectors  
v1 <- c(2,3,6,12)  
v2 <- 1:100  
length(v2)
```

```
[1] 100
```

```
v2
```

```
[1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18
[19] 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36
[37] 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54
[55] 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72
[73] 73 74 75 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
[91] 91 92 93 94 95 96 97 98 99 100
```

```
v3 <- seq(1,100,5) # call without naming arguments
v3
```

```
[1] 1 6 11 16 21 26 31 36 41 46 51 56 61 66 71 76 81 86 91 96
```

```
v3 <- seq(from=1,to=100,by=5) # call with names of arguments
v3
```

```
[1] 1 6 11 16 21 26 31 36 41 46 51 56 61 66 71 76 81 86 91 96
```

```
v3 <- seq(to=100,by=5) # call skipping the first argument
#and using the default value 1 - see help(seq)
v3
```

```
[1] 1 6 11 16 21 26 31 36 41 46 51 56 61 66 71 76 81 86 91 96
```

```
v3 <- seq(by=5,to=100) # call by arguments and change order or arguments
```

How to index vectors?

```
# Indexing vectors
v3[3] #uses square brackets to obtain the third element of the vector
```

```
[1] 11
```

```
v3>20 # produce a vector of boolean values that are TRUE when
```

```
[1] FALSE FALSE FALSE FALSE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
[13] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
      #v3 is greater than 20
v3[v3>20] # select from v3 all the values that are greater than 20
```

```
[1] 21 26 31 36 41 46 51 56 61 66 71 76 81 86 91 96
```

```
v4<-c(1,2,3,4,5)
v4[c(FALSE,TRUE,FALSE,TRUE,FALSE)] #select from v4 the second and fourth element
```

```
[1] 2 4
```

```
v3[1:10] # first ten elements
```

```
[1] 1 6 11 16 21 26 31 36 41 46
```

```
v3[-1] # dropping first element
[1] 6 11 16 21 26 31 36 41 46 51 56 61 66 71 76 81 86 91 96
head(v2) # prints the first few elements of v2
[1] 1 2 3 4 5 6
tail(v2) # prints the last few elements of v2
[1] 95 96 97 98 99 100
which(v3 == 26) # returns the position of v3 that equals 26
[1] 6
```

What kind of numerical operations are possible on vectors?

```
2^v2
[1] 2.000000e+00 4.000000e+00 8.000000e+00 1.600000e+01 3.200000e+01
[6] 6.400000e+01 1.280000e+02 2.560000e+02 5.120000e+02 1.024000e+03
[11] 2.048000e+03 4.096000e+03 8.192000e+03 1.638400e+04 3.276800e+04
[16] 6.553600e+04 1.310720e+05 2.621440e+05 5.242880e+05 1.048576e+06
[21] 2.097152e+06 4.194304e+06 8.388608e+06 1.677722e+07 3.355443e+07
[26] 6.710886e+07 1.342177e+08 2.684355e+08 5.368709e+08 1.073742e+09
[31] 2.147484e+09 4.294967e+09 8.589935e+09 1.717987e+10 3.435974e+10
[36] 6.871948e+10 1.374390e+11 2.748779e+11 5.497558e+11 1.099512e+12
[41] 2.199023e+12 4.398047e+12 8.796093e+12 1.759219e+13 3.518437e+13
[46] 7.036874e+13 1.407375e+14 2.814750e+14 5.629500e+14 1.125900e+15
[51] 2.251800e+15 4.503600e+15 9.007199e+15 1.801440e+16 3.602880e+16
[56] 7.205759e+16 1.441152e+17 2.882304e+17 5.764608e+17 1.152922e+18
[61] 2.305843e+18 4.611686e+18 9.223372e+18 1.844674e+19 3.689349e+19
[66] 7.378698e+19 1.475740e+20 2.951479e+20 5.902958e+20 1.180592e+21
[71] 2.361183e+21 4.722366e+21 9.444733e+21 1.888947e+22 3.777893e+22
[76] 7.555786e+22 1.511157e+23 3.022315e+23 6.044629e+23 1.208926e+24
[81] 2.417852e+24 4.835703e+24 9.671407e+24 1.934281e+25 3.868563e+25
[86] 7.737125e+25 1.547425e+26 3.094850e+26 6.189700e+26 1.237940e+27
[91] 2.475880e+27 4.951760e+27 9.903520e+27 1.980704e+28 3.961408e+28
[96] 7.922816e+28 1.584563e+29 3.169127e+29 6.338253e+29 1.267651e+30
log(v2)
[1] 0.0000000 0.6931472 1.0986123 1.3862944 1.6094379 1.7917595 1.9459101
[8] 2.0794415 2.1972246 2.3025851 2.3978953 2.4849066 2.5649494 2.6390573
[15] 2.7080502 2.7725887 2.8332133 2.8903718 2.9444390 2.9957323 3.0445224
[22] 3.0910425 3.1354942 3.1780538 3.2188758 3.2580965 3.2958369 3.3322045
[29] 3.3672958 3.4011974 3.4339872 3.4657359 3.4965076 3.5263605 3.5553481
[36] 3.5835189 3.6109179 3.6375862 3.6635616 3.6888795 3.7135721 3.7376696
[43] 3.7612001 3.7841896 3.8066625 3.8286414 3.8501476 3.8712010 3.8918203
```

```
[50] 3.9120230 3.9318256 3.9512437 3.9702919 3.9889840 4.0073332 4.0253517
[57] 4.0430513 4.0604430 4.0775374 4.0943446 4.1108739 4.1271344 4.1431347
[64] 4.1588831 4.1743873 4.1896547 4.2046926 4.2195077 4.2341065 4.2484952
[71] 4.2626799 4.2766661 4.2904594 4.3040651 4.3174881 4.3307333 4.3438054
[78] 4.3567088 4.3694479 4.3820266 4.3944492 4.4067192 4.4188406 4.4308168
[85] 4.4426513 4.4543473 4.4659081 4.4773368 4.4886364 4.4998097 4.5108595
[92] 4.5217886 4.5325995 4.5432948 4.5538769 4.5643482 4.5747110 4.5849675
[99] 4.5951199 4.6051702
```

```
v5 <- 101:200
```

```
v5/v2
```

[1]	101.000000	51.000000	34.333333	26.000000	21.000000	17.666667
[7]	15.285714	13.500000	12.111111	11.000000	10.090909	9.333333
[13]	8.692308	8.142857	7.666667	7.250000	6.882353	6.555556
[19]	6.263158	6.000000	5.761905	5.545455	5.347826	5.166667
[25]	5.000000	4.846154	4.703704	4.571429	4.448276	4.333333
[31]	4.225806	4.125000	4.030303	3.941176	3.857143	3.777778
[37]	3.702703	3.631579	3.564103	3.500000	3.439024	3.380952
[43]	3.325581	3.272727	3.222222	3.173913	3.127660	3.083333
[49]	3.040816	3.000000	2.960784	2.923077	2.886792	2.851852
[55]	2.818182	2.785714	2.754386	2.724138	2.694915	2.666667
[61]	2.639344	2.612903	2.587302	2.562500	2.538462	2.515152
[67]	2.492537	2.470588	2.449275	2.428571	2.408451	2.388889
[73]	2.369863	2.351351	2.333333	2.315789	2.298701	2.282051
[79]	2.265823	2.250000	2.234568	2.219512	2.204819	2.190476
[85]	2.176471	2.162791	2.149425	2.136364	2.123596	2.111111
[91]	2.098901	2.086957	2.075269	2.063830	2.052632	2.041667
[97]	2.030928	2.020408	2.010101	2.000000		

A.1.3 Strings

```
#Using strings in R
mystring <- "Ecology"
vstrg <- c("Anna", "Peter", "Xavier")
vstrg[2]
```

```
[1] "Peter"
```

A.1.4 Matrices

```
m <- matrix(5,3,2)
m
```

	[,1]	[,2]
[1,]	5	5
[2,]	5	5

```
[3,]      5      5
m2 <- matrix(1:6,3,2)
m2
 [,1] [,2]
[1,]    1    4
[2,]    2    5
[3,]    3    6
t(m2) # transposes matrix

 [,1] [,2] [,3]
[1,]    1    2    3
[2,]    4    5    6
x <- 1:4
y <- 5:8

m3<-cbind(x,y)
m3

x y
[1,] 1 5
[2,] 2 6
[3,] 3 7
[4,] 4 8
m4<-rbind(x,y)
m4

 [,1] [,2] [,3] [,4]
x    1    2    3    4
y    5    6    7    8
# Indexing matrices
m3[3,2] #element in row 3 and column 2

y
7
m3[1,] #entire first row

x y
1 5
m3[,1] #entire first column

[1] 1 2 3 4
colnames(m3)<-c("col1","col2")
m3
```

```

  col1 col2
[1,]    1    5
[2,]    2    6
[3,]    3    7
[4,]    4    8
m3[, "col2"]
[1] 5 6 7 8

```

A.1.5 Lists

```

# Lists in R
mylist <- list(elem1=m, elem2=v2, elem3="my list")
mylist$elem2

```

```

[1]   1   2   3   4   5   6   7   8   9   10  11  12  13  14  15  16  17  18
[19]  19  20  21  22  23  24  25  26  27  28  29  30  31  32  33  34  35  36
[37]  37  38  39  40  41  42  43  44  45  46  47  48  49  50  51  52  53  54
[55]  55  56  57  58  59  60  61  62  63  64  65  66  67  68  69  70  71  72
[73]  73  74  75  76  77  78  79  80  81  82  83  84  85  86  87  88  89  90
[91]  91  92  93  94  95  96  97  98  99  100

```

A.1.6 Dataframes

```

# Dataframes
df <- as.data.frame(m3)
df$col1

```

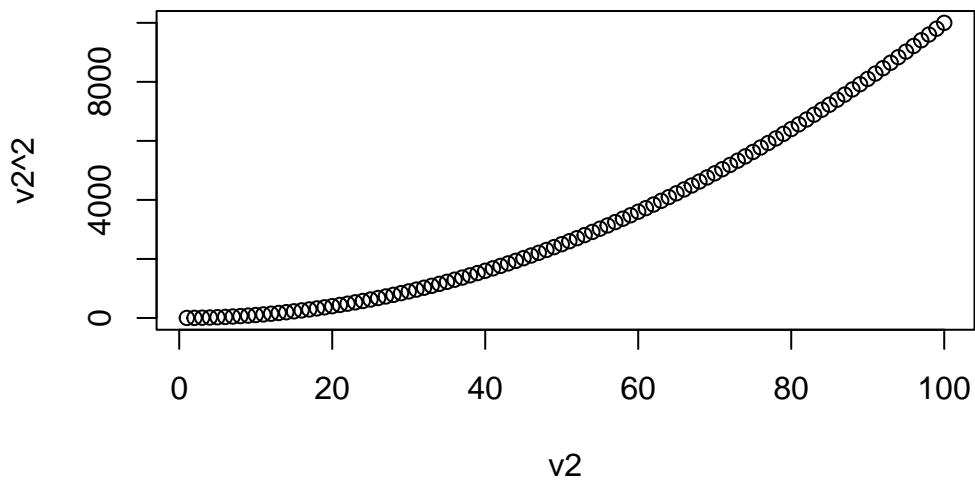
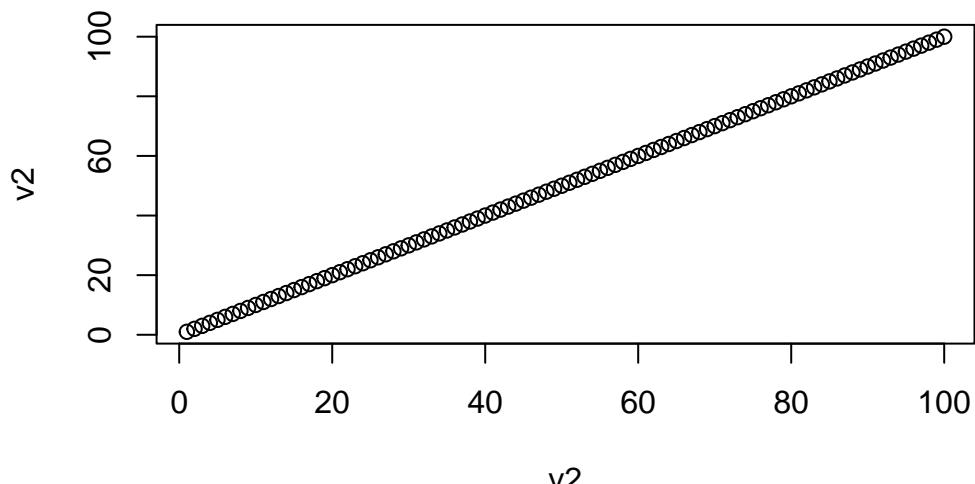
```
[1] 1 2 3 4
```

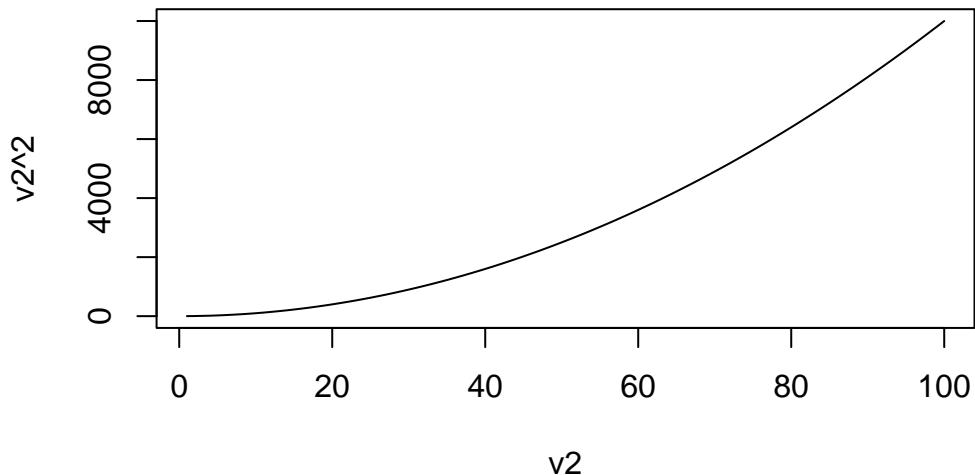
A.2 Basic plots in R

```

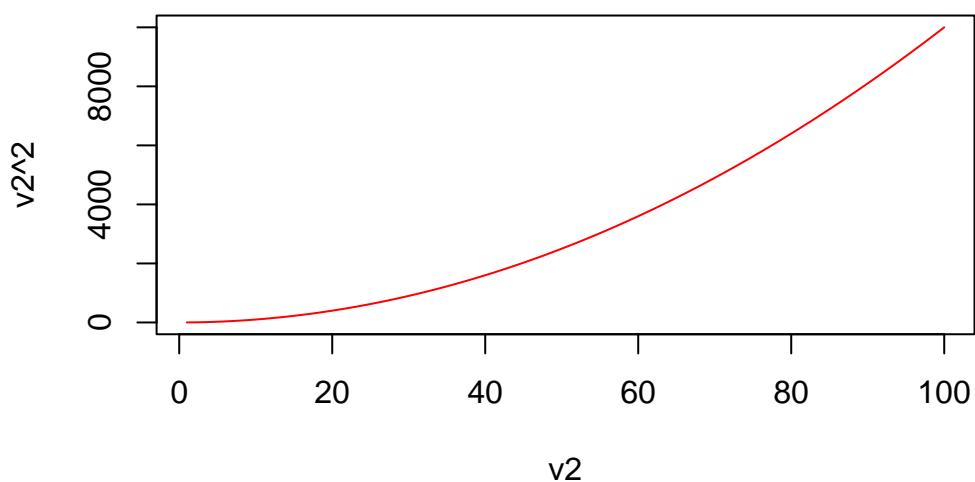
#making plots in R
plot(v2,v2)

```



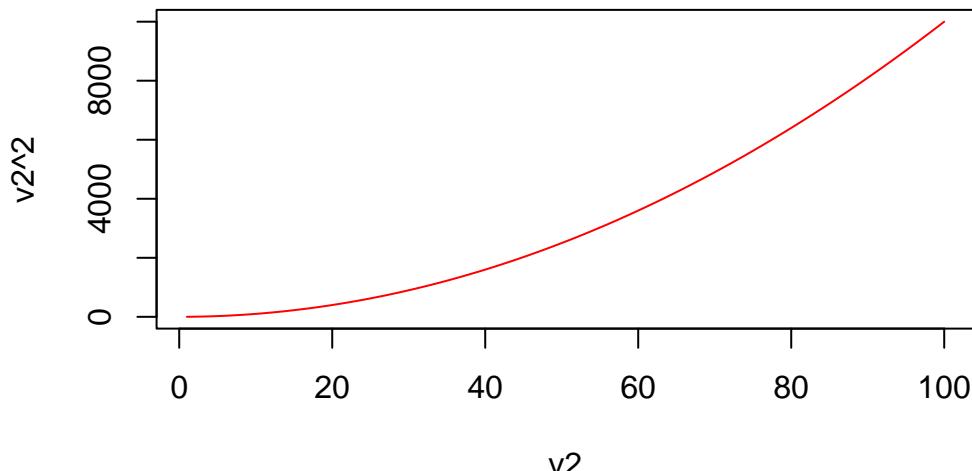


```
plot(v2,v2^2,type="l",col="red")
```



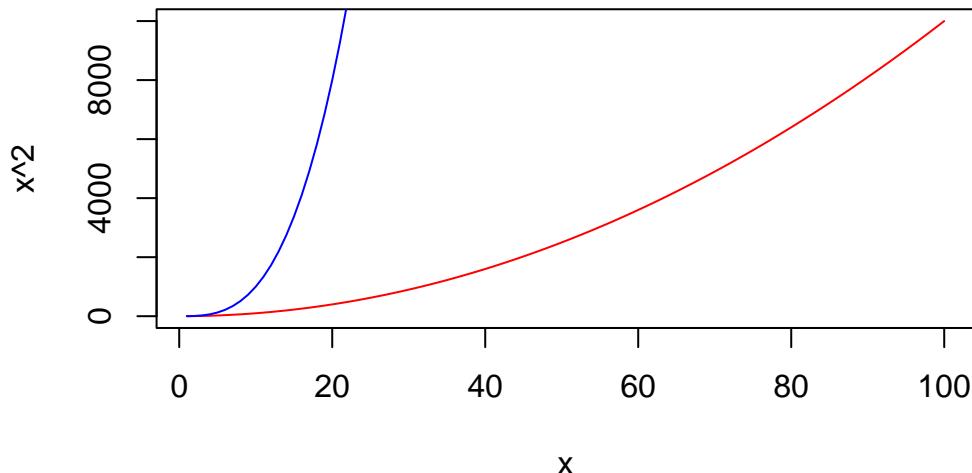
```
plot(v2,v2^2,type="l",col="red",main="My beautiful plot")
```

My beautiful plot



```
plot(v2,v2^2,type="l",col="red",main="My beautiful plot",xlab="x",
      ylab="x^2")
lines(v2,v2^3,col="blue")
```

My beautiful plot



A.3 Iterations and conditional expressions

```
# FOR loops
```

```
for (k in 1:10) # for k =1, 2, 3, 4, 5,...10
  print (k^2)   #do this
```

```
[1] 1
[1] 4
[1] 9
[1] 16
[1] 25
[1] 36
[1] 49
[1] 64
[1] 81
[1] 100
```

```
R <- 1.2
n <- 1
print(n[1])
```

```
[1] 1
for (t in 1:100)
{
  n[t+1] <- R*n[t]
  print(n[t+1])
}
```

```
[1] 1.2
[1] 1.44
[1] 1.728
[1] 2.0736
[1] 2.48832
[1] 2.985984
[1] 3.583181
[1] 4.299817
[1] 5.15978
[1] 6.191736
[1] 7.430084
[1] 8.9161
[1] 10.69932
[1] 12.83918
[1] 15.40702
[1] 18.48843
[1] 22.18611
[1] 26.62333
[1] 31.948
[1] 38.3376
[1] 46.00512
```

```
[1] 55.20614  
[1] 66.24737  
[1] 79.49685  
[1] 95.39622  
[1] 114.4755  
[1] 137.3706  
[1] 164.8447  
[1] 197.8136  
[1] 237.3763  
[1] 284.8516  
[1] 341.8219  
[1] 410.1863  
[1] 492.2235  
[1] 590.6682  
[1] 708.8019  
[1] 850.5622  
[1] 1020.675  
[1] 1224.81  
[1] 1469.772  
[1] 1763.726  
[1] 2116.471  
[1] 2539.765  
[1] 3047.718  
[1] 3657.262  
[1] 4388.714  
[1] 5266.457  
[1] 6319.749  
[1] 7583.698  
[1] 9100.438  
[1] 10920.53  
[1] 13104.63  
[1] 15725.56  
[1] 18870.67  
[1] 22644.8  
[1] 27173.76  
[1] 32608.52  
[1] 39130.22  
[1] 46956.26  
[1] 56347.51  
[1] 67617.02  
[1] 81140.42  
[1] 97368.5  
[1] 116842.2  
[1] 140210.6  
[1] 168252.8  
[1] 201903.3
```

```
[1] 242284  
[1] 290740.8  
[1] 348889  
[1] 418666.7  
[1] 502400.1  
[1] 602880.1  
[1] 723456.1  
[1] 868147.4  
[1] 1041777  
[1] 1250132  
[1] 1500159  
[1] 1800190  
[1] 2160228  
[1] 2592274  
[1] 3110729  
[1] 3732875  
[1] 4479450  
[1] 5375340  
[1] 6450408  
[1] 7740489  
[1] 9288587  
[1] 11146304  
[1] 13375565  
[1] 16050678  
[1] 19260814  
[1] 23112977  
[1] 27735572  
[1] 33282687  
[1] 39939224  
[1] 47927069  
[1] 57512482  
[1] 69014979  
[1] 82817975
```

```
R <- 1.2  
n <- 1  
for (t in 1:100)  
  n[t+1] <- R*n[t]  
  
# IF conditional statement  
  
# logical operators  
# == equal to  
# > greater than  
# < smaller than  
# >= greater or equal  
# <= smaller or equal
```

```
# != different from
# && and
# || or

if (3>2) print ("yes")

[1] "yes"
if (3==2) print ("yes") else print("no")

[1] "no"
if ((3>2)&&(4>5)) print ("yes")

for (k in 1:10) # for k =1, 2, 3, 4, 5,...10
  if (k^2>20) print (k^2)

[1] 25
[1] 36
[1] 49
[1] 64
[1] 81
[1] 100
```

A.4 Writing functions

```
# creating FUNCTIONS in r

pythagoras <- function (c1,c2)
{
  h <- sqrt (c1^2 + c2^2)
  return (h)
}

pythagoras(1,1)

[1] 1.414214
pythagoras(5,5)

[1] 7.071068
pythagoras(10,1)

[1] 10.04988
# regression in R

help(lm)
```

```
x <- c(1,2,3,4)
y <- c(1.1,2.3,2.9,4.1)
plot(x,y)
myreg<-lm(y ~ x)
summary(myreg)
```

Call:
`lm(formula = y ~ x)`

Residuals:

1	2	3	4
-0.06	0.18	-0.18	0.06

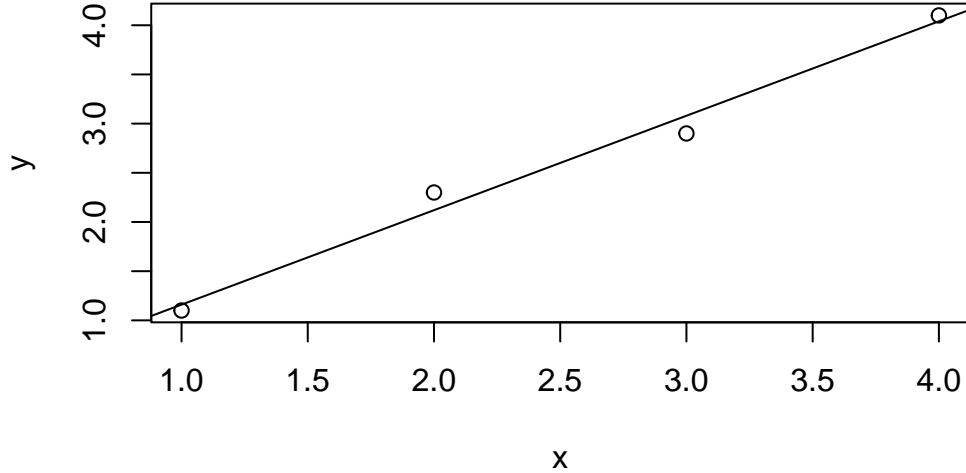
Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	0.20000	0.23238	0.861	0.48012
x	0.96000	0.08485	11.314	0.00772 **

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.1897 on 2 degrees of freedom
Multiple R-squared: 0.9846, Adjusted R-squared: 0.9769
F-statistic: 128 on 1 and 2 DF, p-value: 0.007722

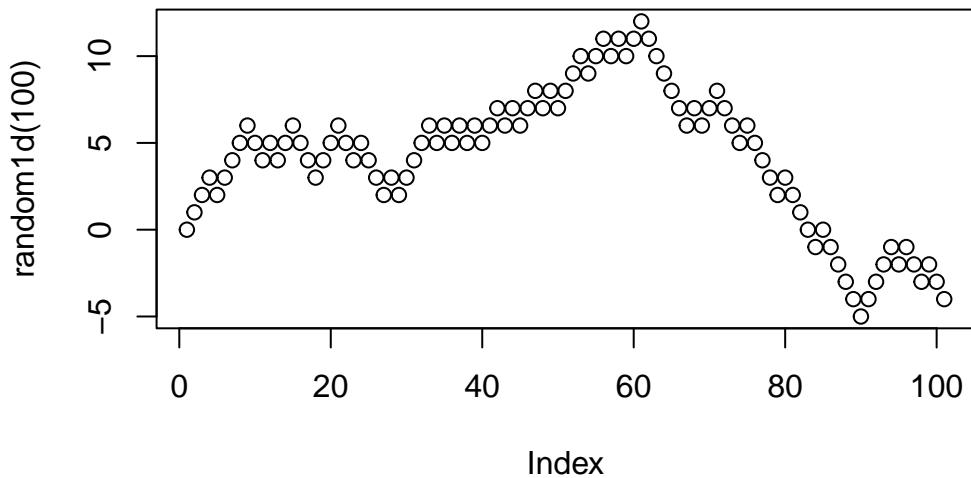
```
abline(myreg)
```



A.5 Random numbers and statistical distributions

```
random1d<-function(tmax)
{
x<-0
for (t in 1:tmax)
{
  r<-runif(1)
  if (r<1/2)
    x[t+1]<-x[t]+1 else
    x[t+1]<-x[t]-1
}
return(x)
}

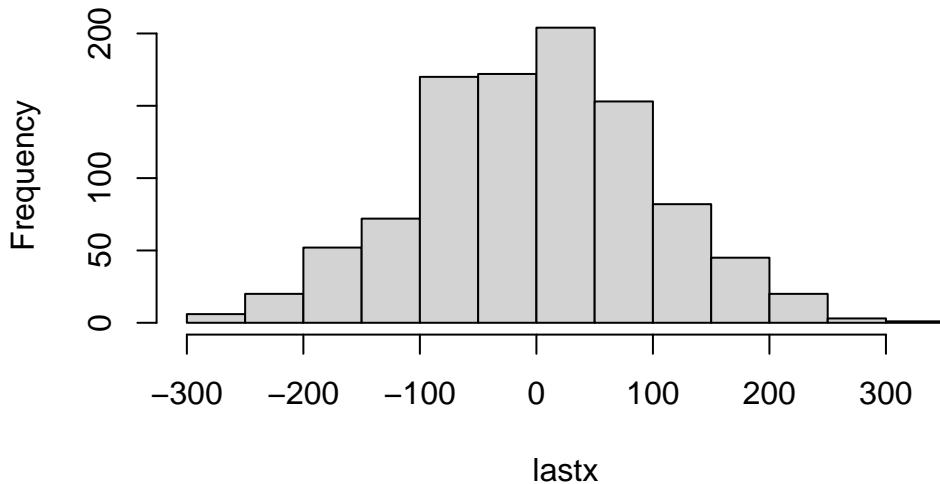
plot(random1d(100))
```



```
tmax<-10000
lastx<-0
for (i in 1:1000)
{
  x<-random1d(tmax)
  lastx[i]<-x[tmax]
}

hist(lastx)
```

Histogram of lastx

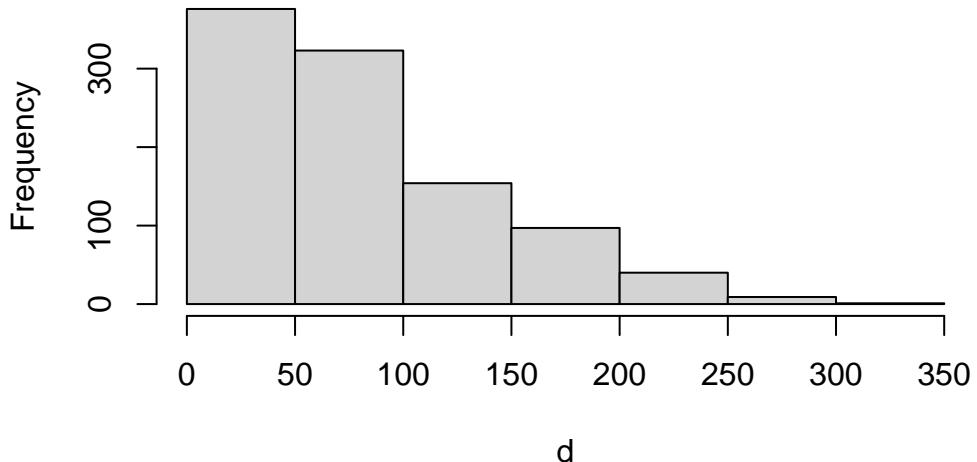


```
mean(lastx)
```

```
[1] -0.618
```

```
d<-sqrt(lastx^2)  
hist(d)
```

Histogram of d

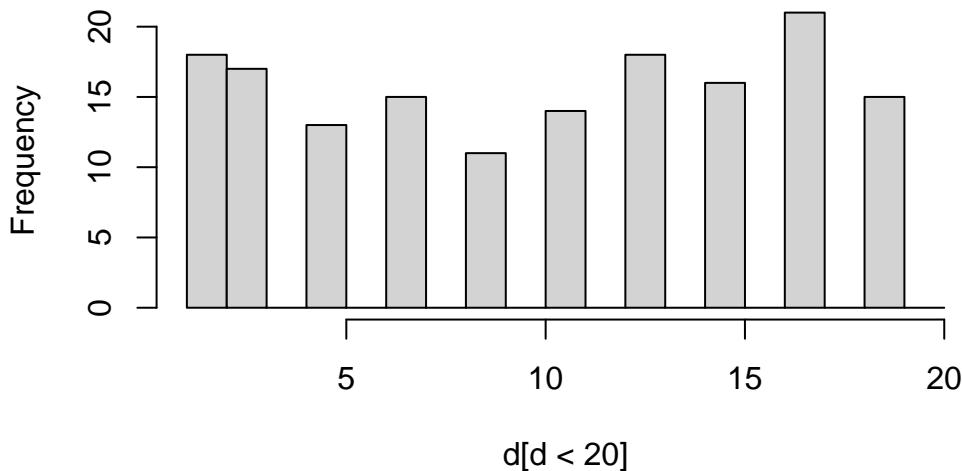


```
mean(d)
```

```
[1] 79.046
```

```
median(d)
[1] 65
max(d)
[1] 323
hist(d[d<20],breaks=c(1:20))
```

Histogram of d[d < 20]



A.6 Intro to apply, pipes, ggplot2, tidyverse

Use of apply instead of for loops.

```
# a recursive function that calculates a factorial
myfun <- function(x)
{
  if (x==1)
    return (1)
  else return(x*myfun(x-1))
}
```

The following does not work

```
myfun(1:10) # does not work
```

One needs to do a for loop or use apply

```
#option1 - with a for loop
start_time <- Sys.time()
```

```
y<-0
for (i in 1:100)
  y[i]<-myfun(i)
end_time <- Sys.time()
end_time-start_time
```

Time difference of 0.01561785 secs

y

[1]	1.000000e+00	2.000000e+00	6.000000e+00	2.400000e+01	1.200000e+02
[6]	7.200000e+02	5.040000e+03	4.032000e+04	3.628800e+05	3.628800e+06
[11]	3.991680e+07	4.790016e+08	6.227021e+09	8.717829e+10	1.307674e+12
[16]	2.092279e+13	3.556874e+14	6.402374e+15	1.216451e+17	2.432902e+18
[21]	5.109094e+19	1.124001e+21	2.585202e+22	6.204484e+23	1.551121e+25
[26]	4.032915e+26	1.088887e+28	3.048883e+29	8.841762e+30	2.652529e+32
[31]	8.222839e+33	2.631308e+35	8.683318e+36	2.952328e+38	1.033315e+40
[36]	3.719933e+41	1.376375e+43	5.230226e+44	2.039788e+46	8.159153e+47
[41]	3.345253e+49	1.405006e+51	6.041526e+52	2.658272e+54	1.196222e+56
[46]	5.502622e+57	2.586232e+59	1.241392e+61	6.082819e+62	3.041409e+64
[51]	1.551119e+66	8.065818e+67	4.274883e+69	2.308437e+71	1.269640e+73
[56]	7.109986e+74	4.052692e+76	2.350561e+78	1.386831e+80	8.320987e+81
[61]	5.075802e+83	3.146997e+85	1.982608e+87	1.268869e+89	8.247651e+90
[66]	5.443449e+92	3.647111e+94	2.480036e+96	1.711225e+98	1.197857e+100
[71]	8.504786e+101	6.123446e+103	4.470115e+105	3.307885e+107	2.480914e+109
[76]	1.885495e+111	1.451831e+113	1.132428e+115	8.946182e+116	7.156946e+118
[81]	5.797126e+120	4.753643e+122	3.945524e+124	3.314240e+126	2.817104e+128
[86]	2.422710e+130	2.107757e+132	1.854826e+134	1.650796e+136	1.485716e+138
[91]	1.352002e+140	1.243841e+142	1.156773e+144	1.087366e+146	1.032998e+148
[96]	9.916779e+149	9.619276e+151	9.426890e+153	9.332622e+155	9.332622e+157

```
#option 2 - with apply
start_time <- Sys.time()
y<-sapply(1:100,myfun)
end_time <- Sys.time()
end_time-start_time
```

Time difference of 0.002145052 secs

y

[1]	1.000000e+00	2.000000e+00	6.000000e+00	2.400000e+01	1.200000e+02
[6]	7.200000e+02	5.040000e+03	4.032000e+04	3.628800e+05	3.628800e+06
[11]	3.991680e+07	4.790016e+08	6.227021e+09	8.717829e+10	1.307674e+12
[16]	2.092279e+13	3.556874e+14	6.402374e+15	1.216451e+17	2.432902e+18
[21]	5.109094e+19	1.124001e+21	2.585202e+22	6.204484e+23	1.551121e+25
[26]	4.032915e+26	1.088887e+28	3.048883e+29	8.841762e+30	2.652529e+32
[31]	8.222839e+33	2.631308e+35	8.683318e+36	2.952328e+38	1.033315e+40
[36]	3.719933e+41	1.376375e+43	5.230226e+44	2.039788e+46	8.159153e+47

```
[41] 3.345253e+49 1.405006e+51 6.041526e+52 2.658272e+54 1.196222e+56
[46] 5.502622e+57 2.586232e+59 1.241392e+61 6.082819e+62 3.041409e+64
[51] 1.551119e+66 8.065818e+67 4.274883e+69 2.308437e+71 1.269640e+73
[56] 7.109986e+74 4.052692e+76 2.350561e+78 1.386831e+80 8.320987e+81
[61] 5.075802e+83 3.146997e+85 1.982608e+87 1.268869e+89 8.247651e+90
[66] 5.443449e+92 3.647111e+94 2.480036e+96 1.711225e+98 1.197857e+100
[71] 8.504786e+101 6.123446e+103 4.470115e+105 3.307885e+107 2.480914e+109
[76] 1.885495e+111 1.451831e+113 1.132428e+115 8.946182e+116 7.156946e+118
[81] 5.797126e+120 4.753643e+122 3.945524e+124 3.314240e+126 2.817104e+128
[86] 2.422710e+130 2.107757e+132 1.854826e+134 1.650796e+136 1.485716e+138
[91] 1.352002e+140 1.243841e+142 1.156773e+144 1.087366e+146 1.032998e+148
[96] 9.916779e+149 9.619276e+151 9.426890e+153 9.332622e+155 9.332622e+157
```

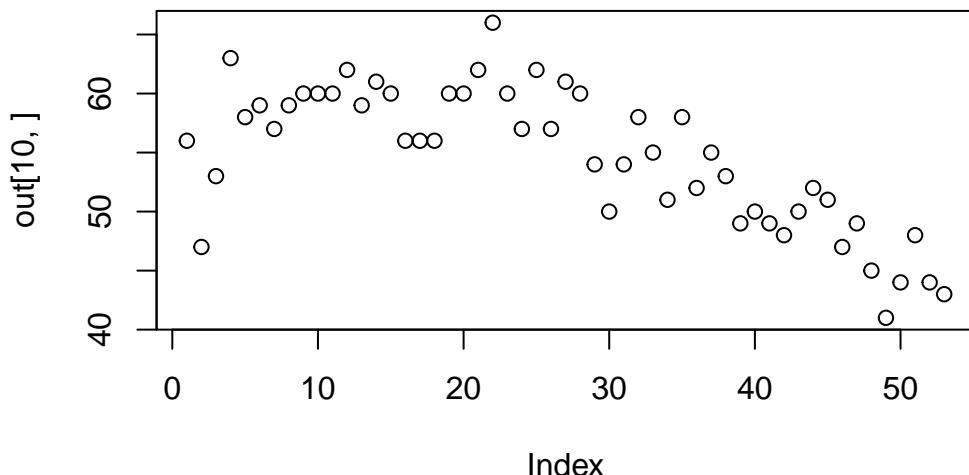
Selecting a subset from a matrix and applying a function to a column of that subset

```
Florida <- read.csv("Labs/Florida.csv")

# number of species for year 1970 and route 20
x=tapply(Florida$Abundance,Florida$Route==20 & Florida$Year==1970, length)
#applies length() to Florida$Abundance but by two levels, when the formula is true
#and when the formula is false

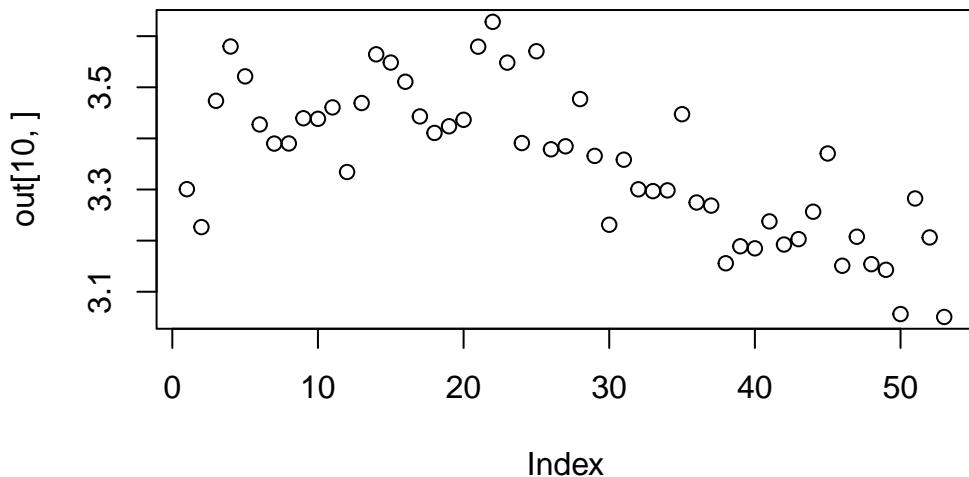
# matrix with number of species per route and per year
out<-tapply(Florida$Abundance,list(Florida$Route,Florida$Year), length)

plot(out[10,])
```



```
shannon<-function(x)
{
  p<-x/sum(x)
  - sum(p*log(p))
}
```

```
out<-tapply(Florida$Abundance,list(Florida$Route,Florida$Year), shannon)
plot(out[10,])
```



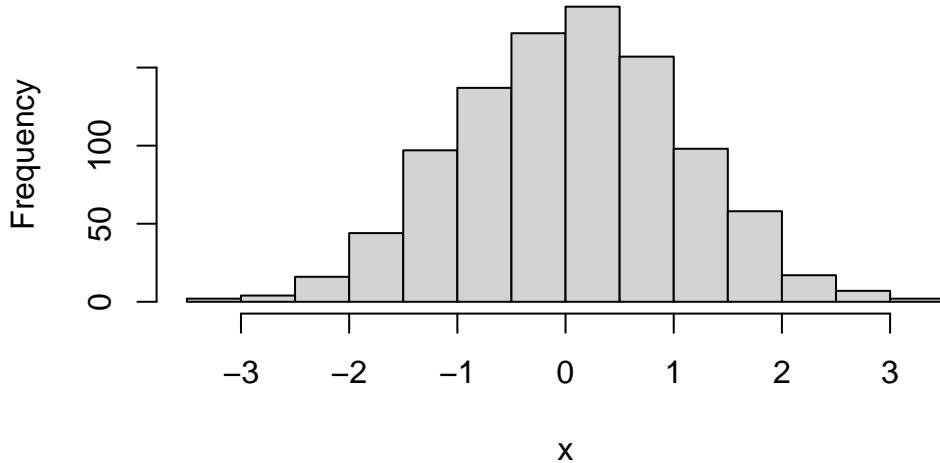
A.6.1 Working with pipes

```
library(tidyverse)

-- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
v dplyr     1.1.4     v readr      2.1.5
v forcats   1.0.1     v stringr    1.5.2
v ggplot2   4.0.0     v tibble     3.3.0
v lubridate 1.9.4     v tidyverse  1.3.1
v purrr     1.2.0
-- Conflicts ----- tidyverse_conflicts() --
x dplyr::filter() masks stats::filter()
x dplyr::lag()   masks stats::lag()
i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

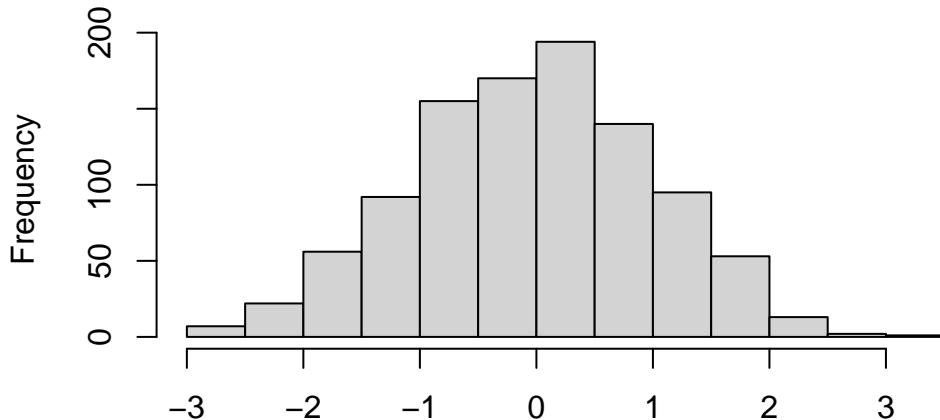
#our first pipe
x<-rnorm(1000)
hist(x)
```

Histogram of x



```
rnorm(1000) %>% hist
```

Histogram of .



Now with the Florida data

```
t<-1:ncol(out)

plot(out[10,])
myreg<-lm(out[10,]~t)
summary(myreg)
```

```

Call:
lm(formula = out[10, ] ~ t)

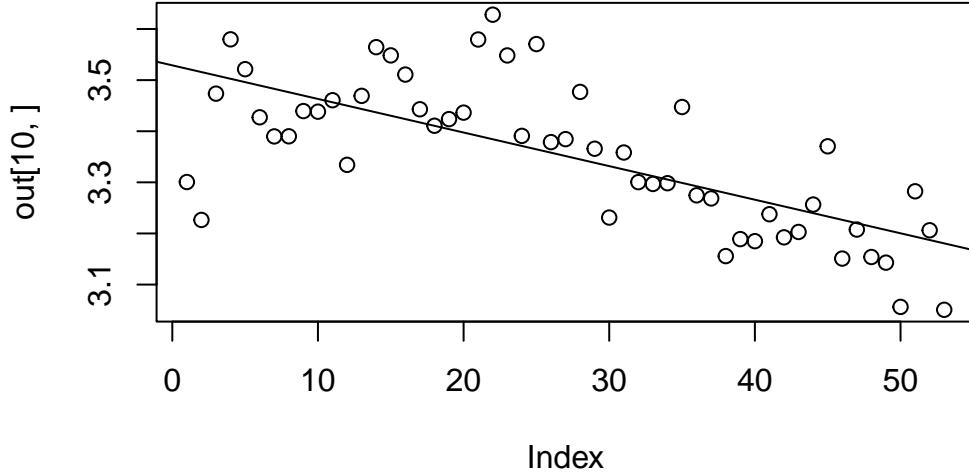
Residuals:
    Min      1Q  Median      3Q     Max 
-0.289301 -0.062193 -0.007159  0.033154  0.243719 

Coefficients:
            Estimate Std. Error t value Pr(>|t|)    
(Intercept) 3.5288806  0.0289650 121.83 < 2e-16 ***
t           -0.0065709  0.0009334   -7.04 4.71e-09 ***  
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.1039 on 51 degrees of freedom
Multiple R-squared:  0.4928, Adjusted R-squared:  0.4829 
F-statistic: 49.56 on 1 and 51 DF, p-value: 4.708e-09

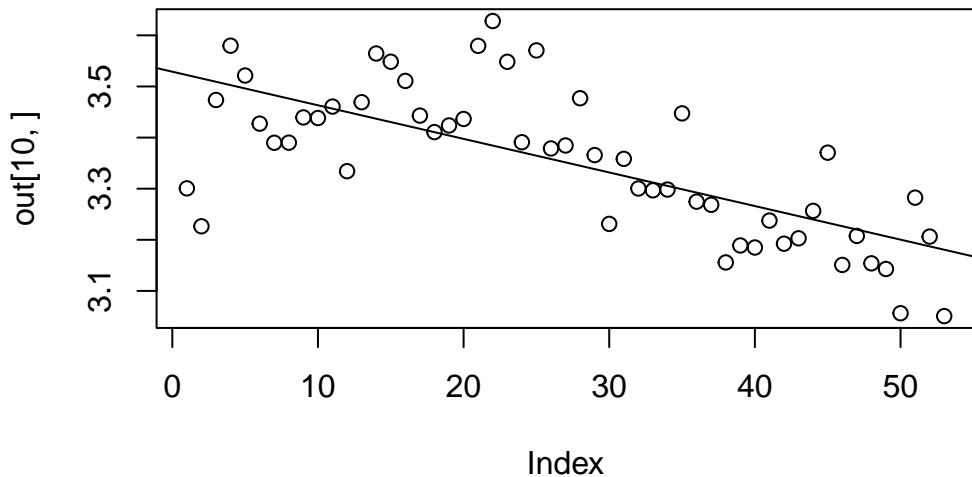
```

```
abline(myreg)
```



```
plot(out[10,])
lm(out[10,]~t) %>% summary %>% abline
```

Warning in abline(.): only using the first two of 8 regression coefficients



A.6.2 Using dplyr (tidyverse)

Base R

```
mtcars
```

	mpg	cyl	disp	hp	drat	wt	qsec	vs	am	gear	carb
Mazda RX4	21.0	6	160.0	110	3.90	2.620	16.46	0	1	4	4
Mazda RX4 Wag	21.0	6	160.0	110	3.90	2.875	17.02	0	1	4	4
Datsun 710	22.8	4	108.0	93	3.85	2.320	18.61	1	1	4	1
Hornet 4 Drive	21.4	6	258.0	110	3.08	3.215	19.44	1	0	3	1
Hornet Sportabout	18.7	8	360.0	175	3.15	3.440	17.02	0	0	3	2
Valiant	18.1	6	225.0	105	2.76	3.460	20.22	1	0	3	1
Duster 360	14.3	8	360.0	245	3.21	3.570	15.84	0	0	3	4
Merc 240D	24.4	4	146.7	62	3.69	3.190	20.00	1	0	4	2
Merc 230	22.8	4	140.8	95	3.92	3.150	22.90	1	0	4	2
Merc 280	19.2	6	167.6	123	3.92	3.440	18.30	1	0	4	4
Merc 280C	17.8	6	167.6	123	3.92	3.440	18.90	1	0	4	4
Merc 450SE	16.4	8	275.8	180	3.07	4.070	17.40	0	0	3	3
Merc 450SL	17.3	8	275.8	180	3.07	3.730	17.60	0	0	3	3
Merc 450SLC	15.2	8	275.8	180	3.07	3.780	18.00	0	0	3	3
Cadillac Fleetwood	10.4	8	472.0	205	2.93	5.250	17.98	0	0	3	4
Lincoln Continental	10.4	8	460.0	215	3.00	5.424	17.82	0	0	3	4
Chrysler Imperial	14.7	8	440.0	230	3.23	5.345	17.42	0	0	3	4
Fiat 128	32.4	4	78.7	66	4.08	2.200	19.47	1	1	4	1
Honda Civic	30.4	4	75.7	52	4.93	1.615	18.52	1	1	4	2
Toyota Corolla	33.9	4	71.1	65	4.22	1.835	19.90	1	1	4	1
Toyota Corona	21.5	4	120.1	97	3.70	2.465	20.01	1	0	3	1
Dodge Challenger	15.5	8	318.0	150	2.76	3.520	16.87	0	0	3	2
AMC Javelin	15.2	8	304.0	150	3.15	3.435	17.30	0	0	3	2
Camaro Z28	13.3	8	350.0	245	3.73	3.840	15.41	0	0	3	4

	mpg	cyl	displ	hp	wt	qsec	vs	am	gear	carb	
Pontiac Firebird	19.2	8	400.0	175	3.08	3.845	17.05	0	0	3	2
Fiat X1-9	27.3	4	79.0	66	4.08	1.935	18.90	1	1	4	1
Porsche 914-2	26.0	4	120.3	91	4.43	2.140	16.70	0	1	5	2
Lotus Europa	30.4	4	95.1	113	3.77	1.513	16.90	1	1	5	2
Ford Pantera L	15.8	8	351.0	264	4.22	3.170	14.50	0	1	5	4
Ferrari Dino	19.7	6	145.0	175	3.62	2.770	15.50	0	1	5	6
Maserati Bora	15.0	8	301.0	335	3.54	3.570	14.60	0	1	5	8
Volvo 142E	21.4	4	121.0	109	4.11	2.780	18.60	1	1	4	2

```
# 1. Filter rows where mpg > 20
subset_data <- mtcars[mtcars$mpg > 20, ]

# 2. Select only mpg, cyl, hp columns
subset_data <- subset_data[, c("mpg", "cyl", "hp")]

# 3. Add new column kpg = mpg * 1.6
subset_data$kpg <- subset_data$mpg * 1.6

# 4. Order by descending mpg
subset_data <- subset_data[order(-subset_data$mpg), ]
```

Or

```
subset_data <- subset(mtcars, mpg > 20, select = c(mpg, cyl, hp))
subset_data <- transform(subset_data, kpg = mpg * 1.6)
subset_data <- subset_data[order(-subset_data$mpg), ]
```

Now with tidyverse

```
library(dplyr)

mtcars |>
  filter(mpg > 20) |>
  dplyr::select(mpg, cyl, hp) |>
  mutate(kpg = mpg * 1.6) |>arrange(desc(mpg))
```

	mpg	cyl	hp	kpg
Toyota Corolla	33.9	4	65	54.24
Fiat 128	32.4	4	66	51.84
Honda Civic	30.4	4	52	48.64
Lotus Europa	30.4	4	113	48.64
Fiat X1-9	27.3	4	66	43.68
Porsche 914-2	26.0	4	91	41.60
Merc 240D	24.4	4	62	39.04
Datsun 710	22.8	4	93	36.48
Merc 230	22.8	4	95	36.48
Toyota Corona	21.5	4	97	34.40
Hornet 4 Drive	21.4	6	110	34.24

Volvo 142E	21.4	4	109	34.24
Mazda RX4	21.0	6	110	33.60
Mazda RX4 Wag	21.0	6	110	33.60

Much better with

```
pak::pak("hadley/genzplyr")
```

```
>Loading metadata database  
Loading metadata database ... done
```

```
No downloads are needed  
1 pkg + 15 deps: kept 16 [5.3s]
```

```
genzplyr loaded fr fr
Your data wrangling is about to be bussin no cap

mtcars |>
  yeet(mpg > 20) |>
  vibe_check(mpg, cyl, hp) |>
  glow_up(kpg = mpg * 1.6) |>
  slay(desc(mpg))
```

		mpg	cyl	hp	kpg
Toyota	Corolla	33.9	4	65	54.24
Fiat	128	32.4	4	66	51.84
Honda	Civic	30.4	4	52	48.64
Lotus	Europa	30.4	4	113	48.64
Fiat	X1-9	27.3	4	66	43.68
Porsche	914-2	26.0	4	91	41.60
Merc	240D	24.4	4	62	39.04
Datsun	710	22.8	4	93	36.48
Merc	230	22.8	4	95	36.48
Toyota	Corona	21.5	4	97	34.40
Hornet	4 Drive	21.4	6	110	34.24
Volvo	142E	21.4	4	109	34.24
Mazda	RX4	21.0	6	110	33.60
Mazda	RX4 Wag	21.0	6	110	33.60

```

hp_per_ton = hp / (wt / 2),
efficiency = mpg / (hp / 100)
) |>
squad_up(cyl) |>                                # Squad up by cylinders
no_cap(
  avg_hp = mean(hp),
  avg_mpg = mean(mpg),
  avg_efficiency = mean(efficiency),
  squad_size = n()
) |>
disband() |>                                    # Disband the squads
slay(desc(avg_efficiency)) |>                  # Sort by slay factor
send_it(10)

# A tibble: 3 x 5
  cyl avg_hp avg_mpg avg_efficiency squad_size
  <dbl>   <dbl>    <dbl>          <dbl>      <int>
1     4     111     25.9        23.3         2
2     6     122.    19.7        16.6         7
3     8     209.    15.1        7.67        14

```

A.6.3 Introduction to ggplot2

ggplot2 is based on ideas from the book “A grammar of graphics” by Leland Wilkinson and developed by Hadley Wickham, which also developed tidyverse.

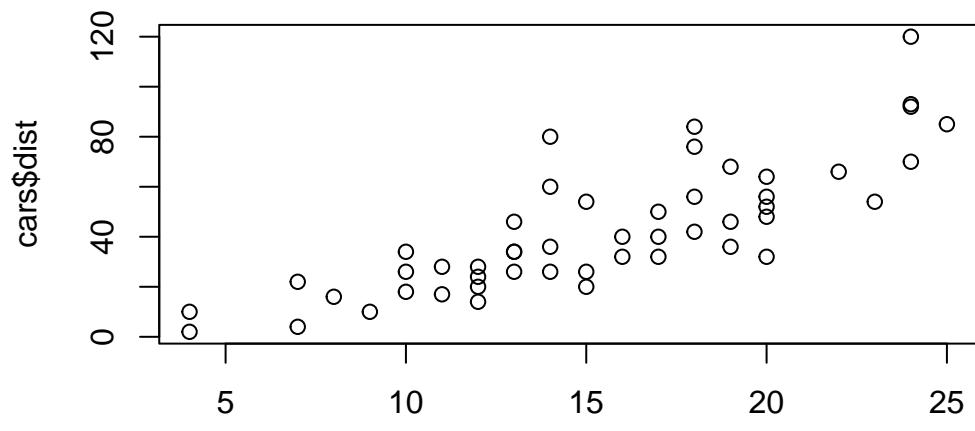
```
library(ggplot2)
```

```
cars
```

	speed	dist
1	4	2
2	4	10
3	7	4
4	7	22
5	8	16
6	9	10
7	10	18
8	10	26
9	10	34
10	11	17
11	11	28
12	12	14
13	12	20
14	12	24
15	12	28
16	13	26

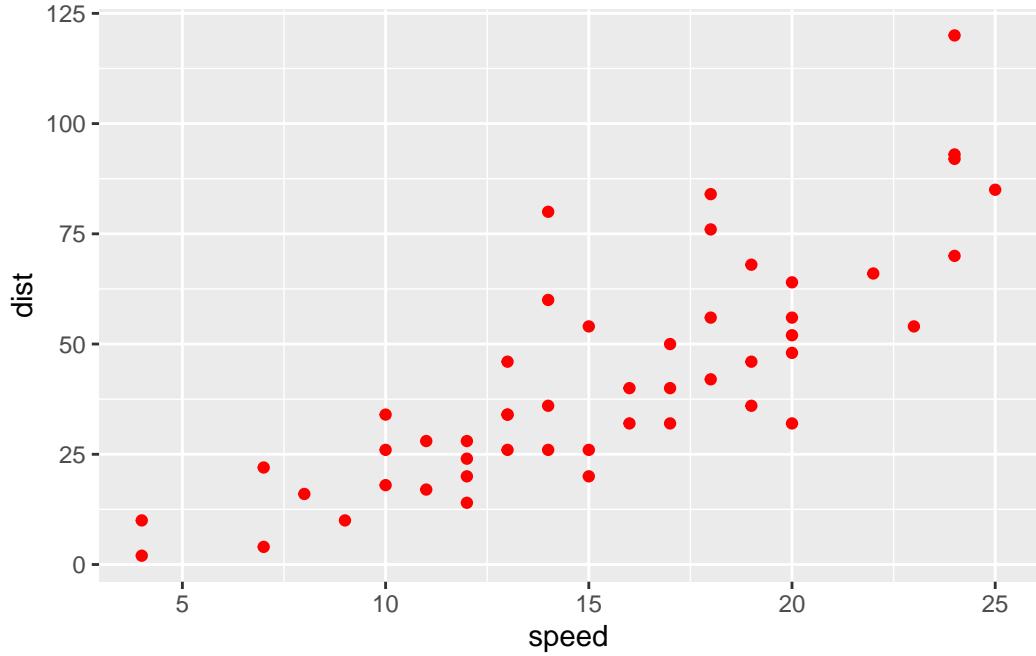
```
17    13   34  
18    13   34  
19    13   46  
20    14   26  
21    14   36  
22    14   60  
23    14   80  
24    15   20  
25    15   26  
26    15   54  
27    16   32  
28    16   40  
29    17   32  
30    17   40  
31    17   50  
32    18   42  
33    18   56  
34    18   76  
35    18   84  
36    19   36  
37    19   46  
38    19   68  
39    20   32  
40    20   48  
41    20   52  
42    20   56  
43    20   64  
44    22   66  
45    23   54  
46    24   70  
47    24   92  
48    24   93  
49    24  120  
50    25   85
```

```
plot(cars$speed,cars$dist)
```



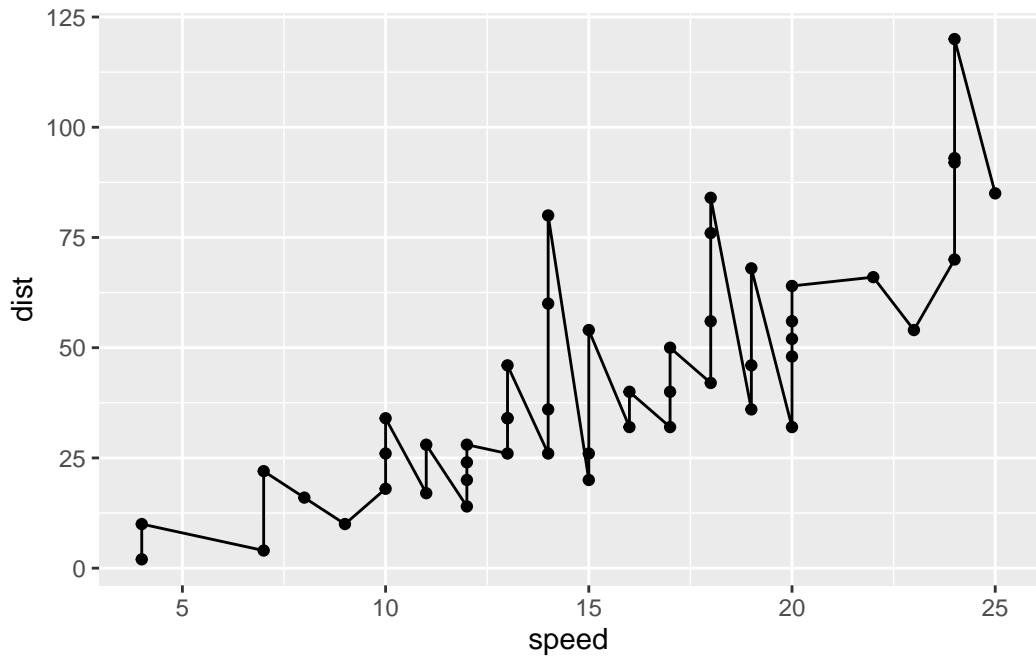
cars\$dist

```
ggplot(data=cars, mapping=aes(x=speed,y=dist)) + geom_point(colour="red")
```



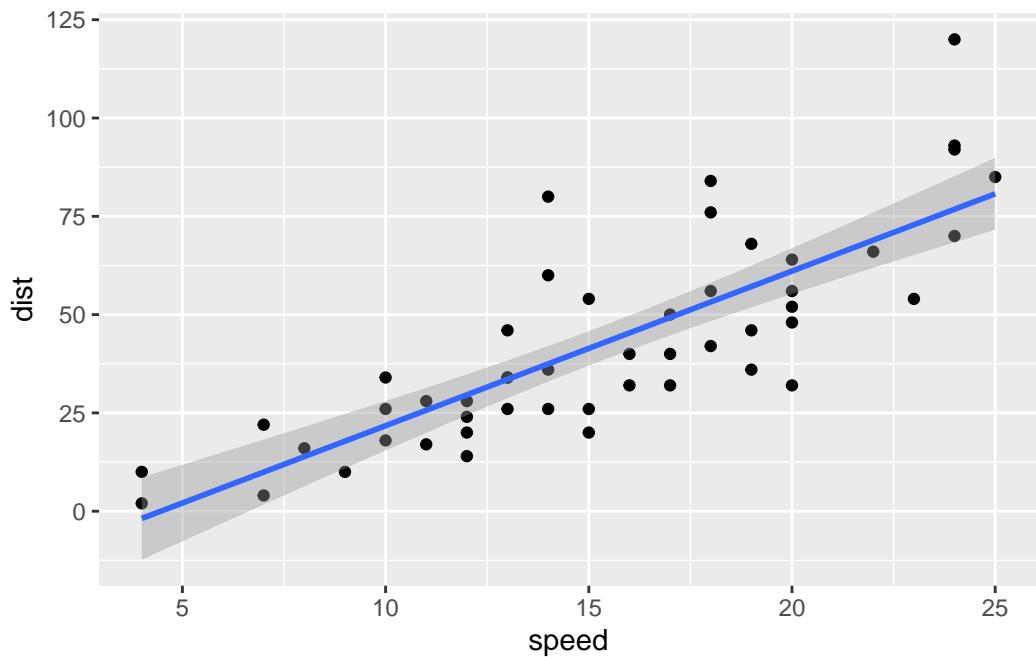
speed

```
myplot <- ggplot(cars, aes(speed,dist))+  
  geom_point()+geom_line()  
myplot
```



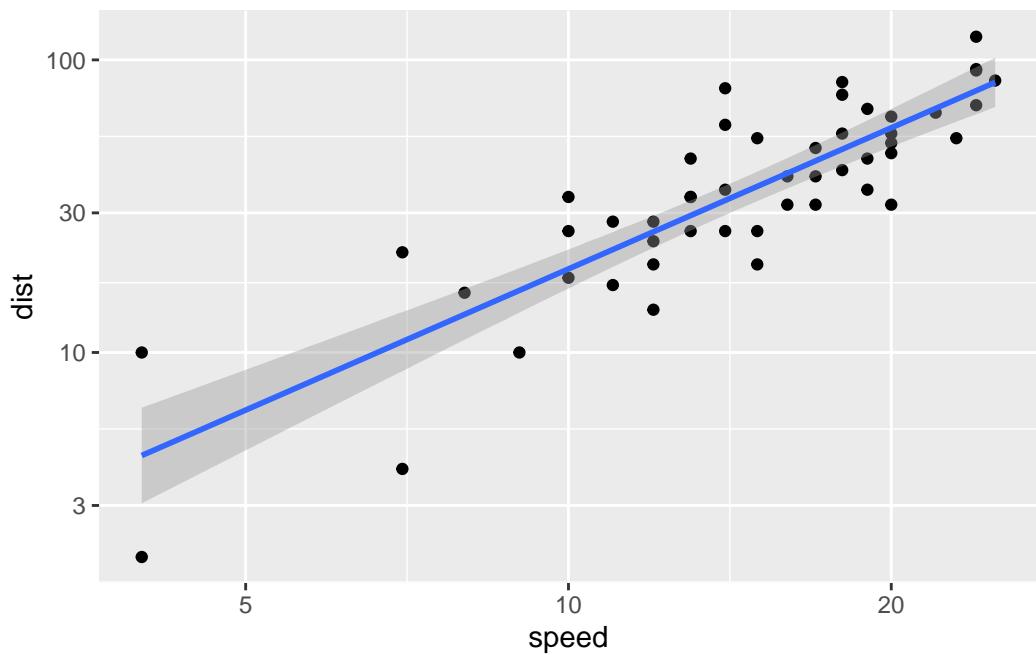
```
data(cars)
myplot <- ggplot(cars, aes(speed,dist))+
  geom_point() + geom_smooth(method="lm")
myplot
```

`geom_smooth()` using formula = 'y ~ x'



```
data(cars)
myplot <- ggplot(cars, aes(speed,dist))+
  geom_point() + geom_smooth(method="lm") + scale_x_log10() + scale_y_log10()
myplot
```

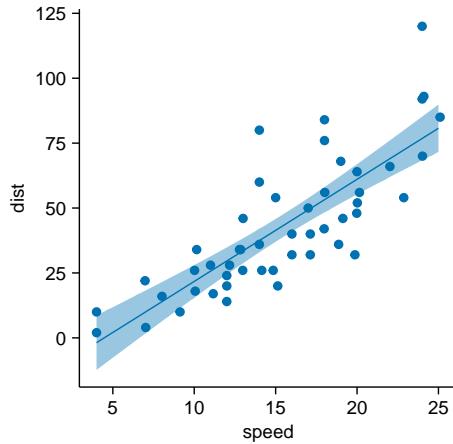
`geom_smooth()` using formula = 'y ~ x'



With tidyplot

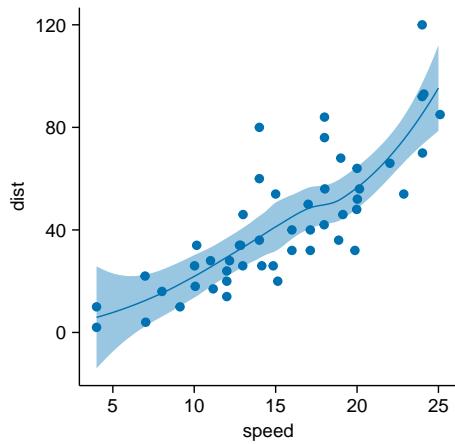
```
library(tidyplots)
cars |> tidyplot(x=speed,y=dist) |>
  add_data_points_beeswarm() |>
  add_curve_fit(method="lm")
```

`geom_smooth()` using formula = 'y ~ x'



```
cars |> tidyplot(x=speed,y=dist) |>
  add_data_points_beeswarm() |>
  add_curve_fit()
```

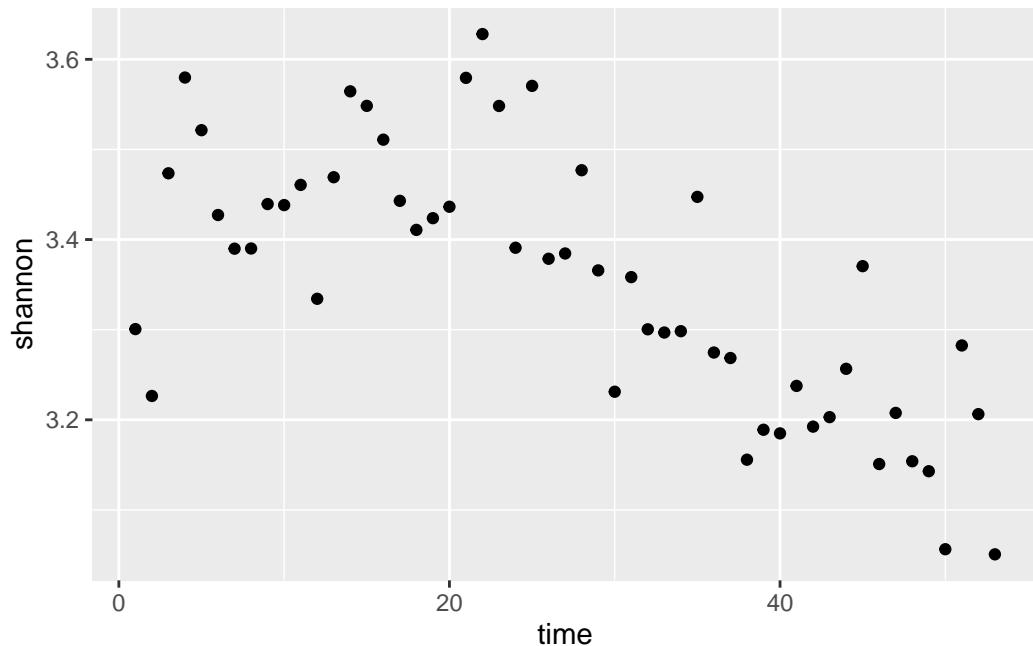
```
`geom_smooth()` using formula = 'y ~ x'
```



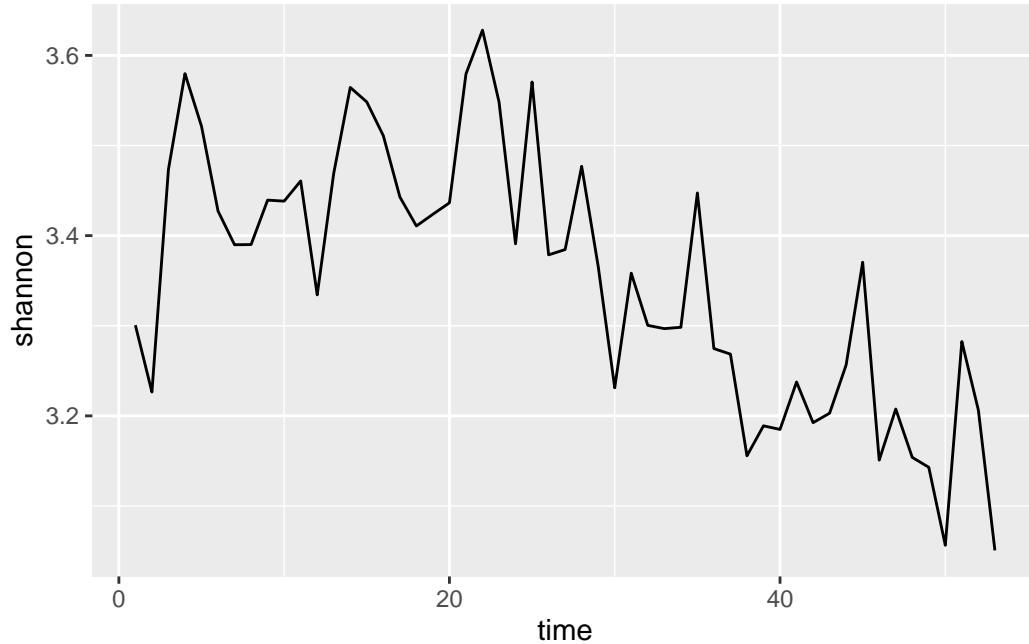
With the florida data

```
#ggplot
mat=cbind(t,out[10,])
colnames(mat)<-c("time","shannon")
mat<-as.data.frame(mat)

myplot <- ggplot(mat, aes(time,shannon))+
  geom_point()
myplot
```



```
myplot <- ggplot(mat, aes(time, shannon))+
  geom_line()
myplot
```



A.7 Working with biodiversity data: GBIF, EBV Portal

Authors: Corey Callaghan, Luise Quoss, Isabel Rosa.

First we load the library rgbif

```
library(rgbif)
library(tidyverse)
```

Now we will download observations of a species. Let's download observations of the common toad *Bufo bufo*.

```
matbufobufo<-occ_search(scientificName="Bufo bufo", limit=500, hasCoordinate = TRUE, hasGeospatialInfo=TRUE)
```

```
Error in occ_search(scientificName = "Bufo bufo", limit = 500, hasCoordinate = TRUE, : could not find function "occ_search"
```

Let's examine the object *matbufobufo*

```
class(matbufobufo)
```

```
Error: object 'matbufobufo' not found
```

```
matbufobufo
```

```
Error: object 'matbufobufo' not found
```

It is a special object of class `gbif` which allows for the metadata and the actual data to all be included, as well as taxonomic hierarchy data, and media metadata. We won't worry too much about the details of this object now.

Let's download data about octupusses. They are in the order "Octopoda". First we need to find the GBIF search key for Octopoda.

```
a<-name_suggest(q="Octopoda",rank="Order")
```

```
Error in name_suggest(q = "Octopoda", rank = "Order"): could not find function "name_suggest"
key<-a$data$key
```

```
Error: object 'a' not found
```

We will only download 2000 observations to keep it simple for now. If you were doing this for real, you would download all data.

```
octopusses<-occ_search(orderKey=key,limit=2000, hasCoordinate = TRUE, hasGeospatialIssue = FALSE)
```

```
Error in occ_search(orderKey = key, limit = 2000, hasCoordinate = TRUE, : could not find function "o
```

Show the result

```
octmat<-octopusses$data
```

```
Error: object 'octopusses' not found
```

```
head(octmat)
```

```
Error: object 'octmat' not found
```

Count the number of observations per species using tidyverse and pipes

```
octmat %>%
  group_by(scientificName) %>%
  summarise(sample_size=n()) %>%
  arrange(desc(sample_size)) %>%
  mutate(sample_size_log=log(sample_size,2)) %>%
  ggplot(aes(x = sample_size_log)) + geom_histogram()
```

```
Error: object 'octmat' not found
```

Plot the records on an interactive map. First load the leaflet package.

```
library(leaflet)
leaflet(data=octmat) %>% addTiles() %>%
  addCircleMarkers(lat= ~decimalLatitude, lng = ~decimalLongitude, popup=~scientificName)
```

A.8 Working with raster and shapefiles

 Summary Table

Feature	raster	terra	stars	
Core data type	Raster*	SpatRaster	stars array	
Modernity	Legacy	Modern replacement	Alternative framework	
Speed/memory	Slower	Fast, efficient	Moderate	
Vector integration	sp	sf	sf	
Dependencies	sp , rgdal , rgeos	none (GDAL/PROJ internal)	sf , units	
Multidimensional (e.g. time)	Limited	Partial	Full	
Recommended for new projects	✗	✓	✓ (if you need multi-dim or tidyverse integration)	

Appendix B

Computational labs

B.1 Climate space of an ectotherm

Solar radiation and convection are the two main pathways for animals such as small lizards. In this case, the total heat flux (f) into the animal is given by the **heat flux equation**:

$$f = q - h * (b - a)$$

- q = solar radiation (cal/h)
- $h * (b - a)$ = heat loss through convection
- b = body temperature of the animal ($^{\circ}$ C)
- a = air temperature ($^{\circ}$ C)
- h = convection heat transfer coefficient (cal/h/ $^{\circ}$ C)

1. *Equilibrium body temperature*

- a. Assuming that air temperature is 18° C, and $h = 50$ cal/h, plot the lizard's equilibrium body temperature as a function of solar radiation. Assume that solar radiation varies between 0 and 1500 cal/h

2. *Climate space of a Lizard*

- a. Draw the climate space of the lizard by drawing the polygon that is limited by the minimum and maximum temperatures that the lizard can support, as a function of solar radiation. Consider that the upper lethal limit for the body temperature (b_{max}) is 36° C and the lower lethal limit for the body temperature (b_{min}) is 24° C.

Hint: `polygon` - draws the polygons whose vertices are given in x and y.

3. *Air Temperature and solar radiation throughout the day in two locations*

- a. Plot the following values of air temperature and solar radiation that are available throughout a day in two locations. For:
 - Times of the day

```
t = c("00:00", "03:00", "06:00", "09:00", "12:00",
  "15:00", "18:00", "21:00", "00:00")
```

- Solar radiation in the rock habitat

```
rock_q = c(150, 150, 800, 1100, 1300, 1200, 800, 400, 150)
```

- Air temperature in the rock habitat

```
rock_a = c(18, 13, 10, 14, 21, 24, 22, 20, 18)
```

- Solar radiation in the bush habitat

```
bush_q = c(150, 150, 450, 600, 650, 650, 350, 200, 150)
```

- Air temperature in the bush habitat

```
bush_a = c(18, 13, 10, 14, 21, 24, 22, 20, 18)
```

- b. At which time of the day is the lizard on the rock, and at which time is it at the bush?
Is there any time when the lizard cannot be at any of the locations?

4. *Body temperature of A. cristatellus in two different habitats*

Section 1.4.2 presents a data frame containing data on individuals *A. cristatellus* living in urban and forested areas in three cities in Puerto Rico, giving us the perfect opportunity to assess the potential thermal effects of habitat upon this species. To do this:

- Make a linear model for individuals of *A. cristatellus* living only in natural habitats (`context = natural`) and another for individuals living in urban habitats (`context = urban`).

Hint: to filter the data you can use the `subset` function in R

```
# example filtering data from anoles only from San Jose sites
df_sanjose <- subset(df, Site == "San Jose")
```

- Is there a habitat where lizards are consistently warmer? If so, provide an ecological explanation (quantitatively) for this observation (assume an ambient temperature of 30 °C).
- Is your model appropriate for explaining your data?

B.2 Optimal foraging theory

1. Predator with a strategy for maximizing energy.

Let ai be the point abundance per unit of time of prey type i , ei be the caloric content of prey type i , hi be the time it takes to consume prey type i , ew is the energy expended per unit of time by the predator while searching for prey, and eh is the energy expended per unit of time by the predator to ingest prey. Consider that there are only two types of prey and that:

Strategy 1: Chosing prey of type 1

Strategy 2: Chosing prey of type 2

Strategy 3: Chosing both preys

- a. Show that if $e1 > e2$ and $h1 < h2$ then the second strategy is never a optimal strategy
- b. Plot and comment the graph of the energy gain per unit of time for the strategy of consuming both preys and for the strategy of consuming only the best prey. Make the abundances vary between 0.005 and 0.5 ind/s for the best prey. Consider the abundance of the worse prey to be 0.05 ind/s and 0.01 ind/s. Consider an active predator with the following physiological parameters:

$$e1 = 10J; e2 = 100J; h1 = 1s; h2 = 60s; ew = 1 J/s; eh = 1 J/S$$

2. Predator with a strategy of sit-and-wait.

Ew is the energy expended per unit of time by the predator while waiting for its prey, ep is the energy expended per unit of time by the predator while chasing the prey, v is the velocity of the predator in chase and a is the point abundance of preys per unit of time.

- a. Produce a graph of energy gain per unit of time in function of the size of the feeding territory. Consider the following values for the parameters of the predator and prey: $ew = 0,1 J/s$; $ep = 1 J/S$; $v = 0,5 m/s$; $a = 0,005 \text{ ind/s/m}^2$; and $e = 10$;

B.3 Evolutionary games

1. **The Prisoners Dilemma Competition.** In this class we will have a competition of algorithms for playing the prisoner's dilemma.

a) Write a function that takes as parameters two vectors that account with the history of the previous plays and returns one play (i.e., Cooperate – “C” or defect – “D”). This play should try to be the best response to the history of past moves. One example of such a function is:

```
strat<-function(own,opponent)
{
  n <- length(opponent)
  if(n==0) "D"
  else
    if(own[n]=="D") opponent[n]
    else "C"
}
```

- b) Use the function pd_sim (see below) to play your strategy with the strategy of the other groups (ask for each group their function(s)) and with itself, in an iterative game with x moves. Compare the results of the cumulative rewards as well as the sequence of moves. Report to the rest of groups the values of the rewards for each of the tournaments. What was the best strategy? What strategies are best responses to themselves?

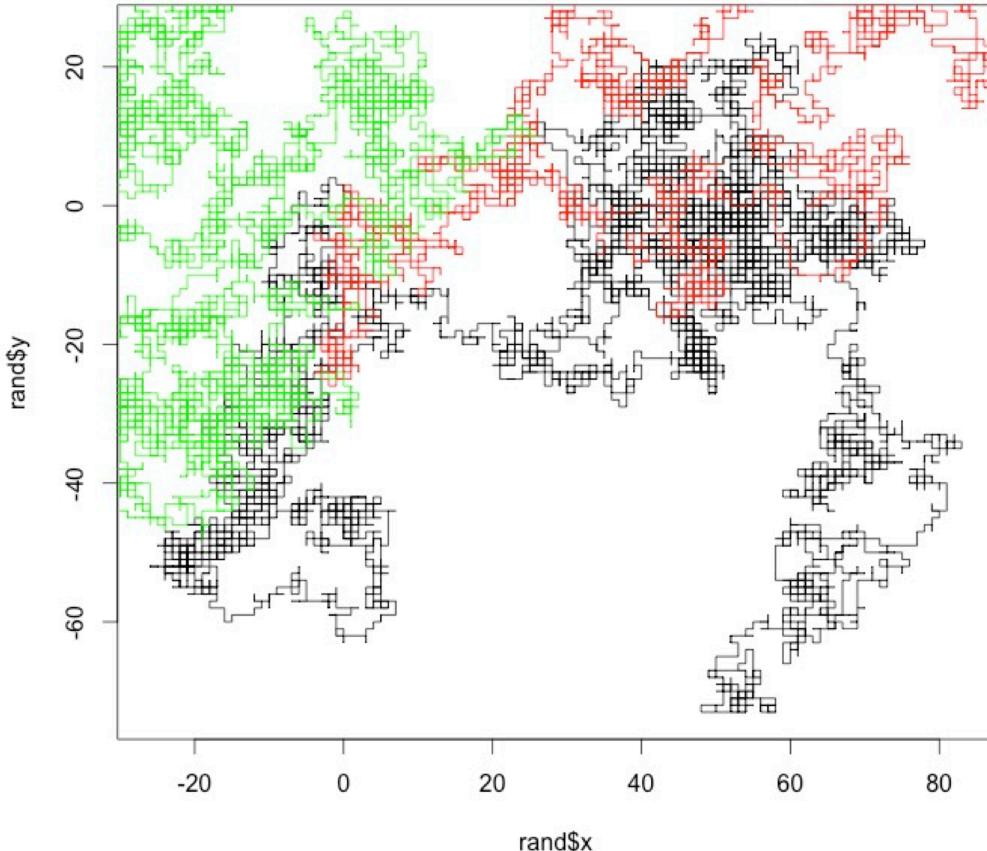
```
pd_sim<-function(p1_strat,p2_strat,n)
{
  w1<-0          #accumulated pay-off (fitness) of player 1
  w2<-0          #accumulated pay-offs of player 2
  h1<-NULL        #history of plays of player 1
  h2<-NULL        #history of plays of player 2

  for (t in 1:n)
  {
    a1<-p1_strat(h1,h2)
    a2<-p2_strat(h2,h1)
    p1<-mat[a1,a2]
    p2<-mat[a2,a1]
    w1<-w1+p1
    w2<-w2+p2
    h1[t]<-a1
    h2[t]<-a2
  }
  list(w1=w1,w2=w2,h1=h1,h2=h2)
}
```

B.4 Dispersal and the random-walk

1. Random walk for 1 individual

- a) Create your own code to simulate an individual random walk. Assume that the starting point is always $x,y = (0,0)$ and the probability of an individual to choose any direction (i.e. left, right, up or down) is the same. It should be a function taking as argument the number of steps and returning a list of two vectors, one with the x positions over time and another with the y positions overtime.
- b) Plot together the random walks (at least 5000 steps) of a few individuals.



2. Random walk for several (n) individuals

- a) Create your own code to simulate random walks by several (n) individuals and returns the last position of each individual. It should be a function that takes as arguments the number of individuals and the numbers of time steps, and returns a list of two vectors, the last x position of each individual, and the last y position of each individual.
- b) Create a histogram showing the distribution of the x and y last positions of 10 000 individuals after 10 time steps, 100 time steps and 1 000 time steps.
- c) Create a function that receives as parameters a vector of values $xlast$, a mean value ($meanx$) and a standard deviation ($stdx$) and returns the log-likelihood of observing those values for those parameters.

- d) Find the value meanx and stdx that maximize the likelihood of the observations. Are they the same as mean(xlast) and std(xlast)? Why?
- e) **Extra credit:** Create a histogram showing the distribution of the distance to the origin ($\sqrt{x^2 + y^2}$) of the last positions of 10 000 individuals for 10 time steps, 100 time steps, 1000 time steps and 10000 time steps. What is the relationship between the length of the randomwalk and the mean distance? Fit the Rayleigh distribution $(2 r / \sigma^2) * \exp(-r^2/2 \sigma^2)$ using non-linear fitting to each of the histograms and overlay it on the graph.

B.5 Pandemic growth

1. **The exponential dynamics of the pandemic.** In early 2020, the first wave of the global pandemic of COVID-19 hit several European countries. Here we will plot the data for those countries and fit an exponential growth model.

- a. Load the cumulative infected individuals time series from the COVID-19 pandemic in five countries. Each time series starts after the first fifteen infections are detected.¹

```
it <- c(17, 79, 132, 229, 322, 400, 650, 888, 1128, 1689, 2036, 2502,
3089, 3858, 4636, 5883, 7375, 9172, 10149, 12462, 15113, 17660, 21157,
23980, 27980, 31506, 35713, 41035, 47021, 53578) #Feb22–March22

es <- c(17, 35, 54, 82, 136, 192, 267, 348, 531, 764, 1094, 1527, 2299,
3274, 4427, 5958, 7641, 9785, 11491, 13994, 17688, 21735, 26304, 31750,
36616, 41262, 48953, 57506, 66460, 75641) #Feb27–March27

fr <- c(17, 38, 57, 100, 130, 178, 212, 285, 423, 613, 716, 1126, 1412,
1784, 2281, 2876, 3661, 4499, 5423, 6633, 7730, 9134, 10995, 12612, 14459,
16018, 19856, 22302, 25233, 29155) #Feb27–March27

uk <- c(18, 22, 30, 42, 47, 69, 109, 164, 220, 271, 352, 412, 469, 617,
876, 1282, 1766, 2244, 2605, 3047, 3658, 4427, 5426, 6481, 7736, 8934,
10312, 12650, 15025, 17717) #Feb27–March27

de <- c(17, 21, 47, 57, 111, 129, 157, 196, 262, 400, 684, 847, 902, 1139,
1296, 1567, 2369, 3062, 3795, 4838, 6012, 7156, 8198, 14138, 18187, 21463,
24774, 29212, 31554, 36508) #Feb26–March26
```

- c. Plot the data in a linear plot, coloring each country with a different colour. What do you observe?
d. Plot the data in a semi-log plot (using the plot option `log="y"`), coloring each country with a different color. What do you observe?
e. Carry out a linear regression with the data of each country (using the log of the n values) and estimate the growth rate R. Are the values the same for all countries? Do they vary over time?
f. Knowing that

$$R_0 = R^\tau$$

and that the infectious period (τ) duration is 10 days, what are the R_0 's in different countries?

- g. Write a for loop to simulate geometric (exponential) growth, going from time t in 1:100, and storing the population size values at each time step t+1 in variable n based on the population values at time t, i.e. `n[t+1] <- R*n[t]`. Don't forget to initialize the population size before the for loop with `n<-1`. Plot a couple of runs of the for loop with different R values (for instance `R=1.01` and `R=1.1`) in linear scale.

¹Data source: <https://github.com/owid/covid-19-data/tree/master/public/data>, accessed 8 Nov 2020

- h. Given that you know the solution of the geometric growth equation to be $n(t) = n_0 R^t$, create a vector of values with this formula using the same R as you used above, but starting with $n_0 < -2$ and overlay them on the plot.

B.6 Population Viability Analysis



1. Consider a population with exponential growth but that exhibits environmental stochasticity, with the logarithm of the population growth rate following a normal distribution with mean $\bar{r} = \log(\bar{R})$ and variance v . Assume that the population has an on-off density dependence and cannot grow above the carrying capacity K . According to Foley (1994)² the expect time to extinction is

$$T = \frac{1}{sr} [e^{s \log(k)} (1 - e^{-s \log(N_0)}) - s \log(N_0)]$$

where $s = 2r/v$.

- a. Plot the mean extinction times, T , as a function of r , v , n_0 , and K . Please comment each plot. r can carry from 0.01 to 0.2, v can vary from 0.05 to 1.0, n_0 can vary from 0 to 10, and K can vary from 10 to 500. Use as base parameters $n0=10$, $K=100$, $v=0.3$ and $r=0.01$.
- b. Write a function that simulates a population with these dynamics numerically. Start by using the for loop that you developed in Lab 2 and modify it to include a carrying capacity and growth rate that is taken every year from a normal distribution. The function should take parameter $n0$, r , v , and K . It should return the vector of the population sizes over time.
- c. Simulate the dynamics with the following parameters:
 1. $n0=3$; $r=0.01$; $v=0.2$; $K=500$
 2. $n0=100$; $r=0.01$; $v=0.2$; $K=500$;

²Foley, P. (1994) Predicting Extinction Times from Environmental Stochasticity and Carrying-Capacity. *Conservation Biology* 8: 124–137.

- d. **Extended credit:** Compare the model predictions with the results from the analytical approximation of Foley. You need to do many simulations to reach the predicted extinction times from Foley. For instance, you can create a function that call the function developed in (b) and executes it 100 times, returning the median time to extinction across the simulations.

B.7 The camera trapper

A researcher place camera traps at a grid of sites during 19 days to observe roe-deer.

1. Load the data on the file roe_deer_2016_r.csv into R. Briefly describe the structure of the dataset.
2. Case 1: Ignoring detection probability.
 - a) Build a vector that for each site takes value 1 when roe-deer is present and 0 when is absent.
 - b) Using maximum-likelihood estimate the occupancy probability (ψ). First create a function that takes as parameters psi and a vector of presence/absences and returns the likelihood. Then plot that function for a range of Ψ values and find the ψ value that maximizes the function.
 - c) Assume that based on previous work we know that occupancy is somewhat between 0.2 and 0.5. Using a Bayesian approach, calculate the posterior probability distribution for the occupancy $P(\psi)$.

Help: A function returning the $P(\psi|data)$. It takes as parameters a vector y of presences/absences, a value for `psi`, and a function for the prior distribution of ψ named `priorpsi`:

```
occupancybayesian <- function(y,psi,priorpsi)
{
  integrand <- function(x)
    occupancylikelihood(y,x)*priorpsi(x)
  occupancylikelihood(y,psi)*priorpsi(psi)/
    integrate(integrand, lower = 0.01, upper = 0.99)$value
}
```

A function returning $P(\psi)$, the prior distribution of ψ values. It takes as parameters a value for `psi`.

```
prior <- function(psi)
{
  dunif(psi,0.2,0.5)
}
```

3. Case 2: Using a hierarchical model with detection probability
 - a. Using maximum-likelihood estimate the occupancy probability (Ψ) and detection probability (p).



B.8 Managing a fishery

1. Consider that you are managing a fishery with the following dynamics:

$$dn/dt = 0.2n(1 - n/10000)$$

- Plot the production function of the fishery, indicating the stock size for the maximum sustainable yield and the corresponding production level.
- Suppose you establish an annual quota (harvest) that is equivalent to 50% of the maximum sustainable yield. What are the two stock size levels that sustainably allow that exploitation level?
- Explore what happens when you manage this fishery for 100 years. Start by writing a function that simulates the logistic growth above with a constant harvest. The function should take as parameters the initial population size n_0 (the stock of the fishery), the annual harvest h , and the number of years for which you simulate the population dynamics and return a vector with the population sizes over time. Discuss the results of the following experiments.
 - Experiment 1:** Set the harvest equal to the production at MSY and initial population size at carrying capacity.
 - Experiment 2.** Set the harvest equal to the production at MSY and initial population size at the stock size of MSY
 - Experiment 3.** Set the harvest equal to 50% of the maximum sustainable yield and initial population size at carrying capacity.
 - Experiment 4.** Set the harvest equal to the production at MSY and initial population size at the stock size of MSY - 1000 individuals.
 - Experiment 5.** Set the harvest equal to the production at MSY and initial population size at the stock size of MSY - 1000 individuals.
- Explain why in a system with constant quotas, MSY is not stable and should not be the management goal.
- Extra credit:** Modify the logistic growth function to have stochastic dynamics so that there is a variance of 10% in the annual productivity. Simulate again experiment 2 a few times. What happens?
- Discuss how these results relate with the paper of Worm et al (2009) about the need to rebuild fisheries.



Figure B.1: Atlantic cod (*Gadus morhua*). Source: Hans-Petter Fjeld, CC-BY-SA, Wikipedia.

B.9 Road Kill

Populations in space and the impacts of roads

The Red fox

...So, while they may be most active at night. Red foxes are especially active during the daytime in spring and summer (April-Sept) as they are foraging for food to feed their young.

Source photo: Wikipedia, CC0.



Goals of the exercise:

- Identify road segments with hotspots of red fox-car-collisions.
 - What impact do activity patterns have on the rate of animal-car-collision hotspots?
 - Use the False Discovery Rate (FDR) method to identify such hotspots.
1. Please install the following R packages


```
library(readxl)
library(dplyr)
library(sf)
library(ggplot2)
library(fuzzySim)
library(grid)
```

 - a. Load the *roadkill* dataset.
 - b. Explain the structure of the dataset.
 - c. Show that differences in seasonal activity patterns impact the FDR hotspot results in red fox-car-collisions.
 - d. How many hotspots were detected for the active season?
 - e. Plot, based on the FDR method, true and “false” hotspots. Explain the general geographic locations of the hotspots and those vary between the two different seasons.
 - f. Think about conservation management implications.

B.10 Simulating a neutral community

The neutral theory in ecology provides a mechanistic explanation for species abundance distributions. It seeks to understand the impact of speciation, extinction, dispersal and ecological drift on the species abundance distribution (SAD), assuming that all species have equal opportunities (Hubbell, 2001).

It is important to note that the neutral theory is a model created to explain a pattern of relative abundance within communities, but it does not necessarily reflect reality (communities' mechanisms). The neutral theory utilizes a model based on the dynamics of a species' population, which is governed by generalized birth and death events, including speciation, immigration, and emigration (Rosindell et al (2011)).

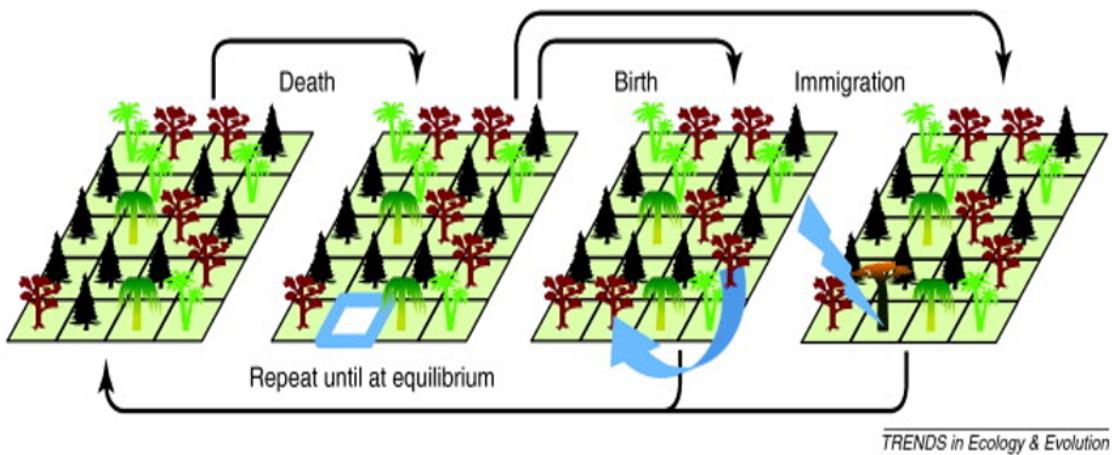


Figure B.2: Diagram of the dynamics of communities in the neutral theory of biodiversity (from Rosindell et al 2011).

The neutral theory paradox is that in absence of migration or mutation diversity gradually declines to zero or monodominance. Let's see what happens to species richness and the species abundance distribution over time for different parameters of the model. Start by installing the package `untb`.

1. **Without mutation or dispersal (pure ecological drift):** We start with a local community with 20 species, each with 25 individuals. The simulation then runs for 2500 generations where 10 individuals die per generation. Mutation rate is zero and immigration rate is zero
 - a. Plot the number of species over time.
 - b. Plot the species abundance distribution at time 1, 100 and 2500
 - c. Plot the map of individuals at those time steps.
2. **With point mutation:** Same parameters as (1) but with speciation rate of 0.1.
 - a. Plot the number of species over time.

- b. Plot the species abundance distribution at time 1, 100 and 2500
 - c. Plot the map of individuals at those time steps.
3. [Extra Credit] **With immigration:** Same parameters as (1) but immigration rate greater than zero. Play with different abundance distributions for the metacommunity.

B.11 Monitoring biodiversity

1. Exploring the BBS dataset
 - a. Load the file Florida.csv into R. Briefly describe the structure of the dataset.
 - b. Map the monitored transects/routes in Florida's map. Use the maps library to plot the counties of Florida as a base map.
 - c. For transect 4 and transect 109 in year 2018 plot calculate the species richness, Shannon diversity index H and evenness J of both transects, with H being

$$H = - \sum_i (p_i \ln p_i)$$

where p_i is defined as the proportion of individuals found in species i . Compare the two transects.

- d. Plot the species abundance distribution and abundance-rank for both transects. What do you observe?
- e. Choose one transect and plot the species richness, Shannon diversity, and geometric mean abundance over time
- f. *Extra credit 1:* Produce a map of the trends of one these metrics across Florida (for instance coloring different points according to the trend)
- g. *Extra credit 2:* Estimate the number of species in Florida by combining the different transects and using one of the estimators. Use bootstrap and/or jackknife to calculate confidence intervals.