### **NAME**

vsearch — dereplicate, filter, sort, search, compare and clusterize amplicons from metagenomic projects

#### **SYNOPSIS**

vsearch [ options ] filename

## DESCRIPTION

Environmental or clinical molecular studies generate large volumes of amplicons (e.g. SSU-rRNA sequences) that need to be filtered, dereplicated, searched, clustered or compared to sequences from other studies. The aim of **vsearch** is to offer a all-in-one open source tool to perform these tasks, using optimized algorithm implementations and harvesting the full potential of modern computers to guarantee a fast and accurate data processing.

Nucleotidic sequence comparisons is at the core of **vsearch**. To speed up comparisons, **vsearch** implements an efficient *k*-mer filtering, and an extremely fast Needleman-Wunsch algorithm making use of the Streaming SIMD Extensions (SSE2) of modern x86-64 CPUs. If SSE2 instructions are not available, **vsearch** exits with an error message.

## Input

**vsearch** input is a fasta file containing one or several nucleotidic sequences. For each sequence, the sequence identifier is defined as the string comprised between the ">" symbol and the first space or the end of the line, whichever comes first. Additionally, if the line ends with the pattern ";size=*integer*;", **vsearch** will interpret *integer* as the abundance of the sequence (in a dereplicated fasta file for instance). The nucleotidic sequence is defined as a string of [acgt] or [acgu] symbols (case insensitive), starting after the end of the identifier line and ending before the next identifier line or the file end; **vsearch** exits with an error message if any other symbol is present in the sequence. Optionally, **vsearch** can be compiled to accepted compressed fasta files as input (gz and bzip2 formats).

# **Options**

**vsearch** recognizes a large number of command-line options. For an easier navigation, options are grouped by theme (dereplication, filtering, sorting, searching, comparison, clustering). We start with general options that apply to all themes.

General options:

- **--help** display a short help and exit.
- --version output version information and exit.
- --fasta\_width positive integer

fasta files produced by **vsearch** are wrapped (sequences written on lines of *integer* nucleotides, 80 by default). Set that value to 0 to eliminate the wrapping.

# --maxseqlength positive integer

all **vsearch** operations will discard sequences of length equal or greater than *integer* (50,000 nucleotides by default).

## --minseqlength positive integer

all **vsearch** operations will discard sequences of length smaller than *integer* (1 nucleotide by default for sorting or shuffling, 32 nucleotides for dereplication, clustering or searching).

### --notrunclabels

do not truncate sequence labels at first space, use the full header.

#### --strand plus/both

when searching or dereplicating, check the *plus* strand only (default) or check *both* strands.

#### --threads positive integer

number of computation threads to use. The number of threads should be lesser or equal to the number of available CPU cores. The default is to launch one thread per available logical core.

## --uc filename

when searching, clustering or dereplicating, output results in *filename* using a uclust-like format.

#### --uc\_allhits

when searching, clustering or dereplicating, and when using the --uc option, show all hits, not just top hit.

# Clustering options:

## --centroids filename

output cluster centroid sequences to *filename* file.

#### -- cluster fast filename

use the fast clustering algorithm and write the results to filename.

# --cluster\_smallmem filename

use a slower clustering algorithm (consumes less memory) and write the results to *file-name*.

#### -- clusters string

output each cluster to a separate fasta file using the prefix *string* and a ticker (0, 1, 2, etc.) to construct the filenames.

#### --usersort

when using --cluster\_smallmem, conserve the initial input order of sequences, do not sort sequences by decreasing length before clustering.

Dereplication, masking, shuffling and sorting options:

# --derep\_fulllength filename

merge strictly identical sequences contained in *filename*. Redundant sequences receive the header of the sequence of their group, and the number of occurrences (abundance) is indicated at the end of the fasta header using the pattern ";size=X;".

# --maskfasta filename

mask sequences contained in filename.

# --maxsize positive integer

when using --sortbysize, discard sequences with an abundance value greater than *integer*.

#### --minsize positive integer

when using --sortbysize, discard sequences with an abundance value smaller than *integer*.

# --minuniquesize positive integer

when dereplicating, discard sequences with an abundance value smaller than integer.

### --output filename

when dereplicating, sorting or shuffling, write the results to filename.

## --relabel string

when sorting, relabel sequence headers using string as suffix.

# --seed positive integer

when shuffling, use integer as seed. Set to 0 to use a pseudo-random seed.

# --sizein read abundance annotation from input

--sizeout add abundance annotation to output

# --shuffle filename

pseudo-randomly shuffle the order of sequences contained in *filename*.

# --sortbylength filename

sort by decreasing length the sequences contained in filename.

## --sortbysize filename

sort by decreasing abundance the sequences contained in *filename*.

# --topn positive integer

when dereplicating, sorting or shuffling, output just the top *integer* sequences.

### Searching options:

## --alnout filename

write pairwise global alignments to filename using a human-readable format.

## --blast6out filename

write search results to *filename* using a blast-like tab-separated format of twelve fields (see below), with one line per query-target matching (or lack of matching if --out-put\_no\_hits is used). A similar output can be obtain with --userout *filename* and --user-fields query+target+id+alnlen+mism+opens+qlo+qhi+tlo+thi+evalue+bits. A complete list and description is available in the section "Fields".

- 1. Query label.
- 2. Target (database sequence or cluster centroid) label.
- 3. Percent identity.
- 4. Alignment length.
- 5. Number of mismatches.
- 6. Number of gap opens.
- 7. 1-based position of start in query.
- 8. 1-based position of end in query.
- 9. 1-based position of start in target.
- 10. 1-based position of end in target.
- 11. E-value (not computed for nucleotidic alignments).
- 12. Bit score (not computed for nucleotidic alignments).

# --db filename

compare query sequences to the fasta-formatted target sequences contained in *filename*, using global pairwise alignment.

## --dbmask none/dust/soft

mask simple repeats and low-complexity regions in target database sequences using the *dust* or the *soft* algorithms, or do not mask (*none*). The default is to mask using *dust*.

## --dbmatched filename

write database target sequences matching at least one query sequence to *filename*, in fasta format.

# --dbnotmatched filename

write database target sequences not matching query sequences to *filename*, in fasta format.

## -- fastapairs filename

write pairwise alignments of query and target sequences to filename, in fasta format.

**--fulldp** dummy option. To maximize search sensitivity, vsearch uses a 8-way SIMD vectorized full dynamic programming algorithm (Needleman-Wunsch), whether or not --fulldp is specified.

# --gapext string

penalties for gap extension (2I/1E)

#### --gapopen string

penalties for gap opening (20I/2E)

#### --hardmask

mask low-complexity regions by replacing them with Ns instead of setting them to lower case.

**--id** real reject the sequence match if the pairwise identity is lower than real (value ranging from 0.0 to 1.0 included). (what is the formula?)

# --idprefix positive integer

reject the target sequence if the first *integer* nucleotides do not match the query sequence.

## --idsuffix positive integer

reject the target sequence if the last *integer* nucleotides do not match the query sequence.

--leftjust reject the target sequence if the alignment begins with gaps.

#### --match integer

score assigned to a match (i.e. identical nucleotides) in the pairwise alignment. The default value is 2.

#### --matched filename

write query sequences matching database target sequences to filename, in fasta format.

## --maxaccepts positive integer

maximum number of hits to accept before stopping the search. The default value is 1. That option works in pair with maxrejects. The search process sorts target sequences by decreasing number of k-mers they have in common with the query sequence, using that information as a proxy for sequence similarity. If the first target sequence passes the acceptation criteria, it is accepted as best hit and the search process stops for that query. If maxaccepts is set to a higher value, more hits are accepted. If maxaccepts and maxrejects are both set to 0, the complete database is searched.

#### --maxdiffs positive integer

reject the target sequence if the alignment contains at least *integer* substitutions, insertions or deletions.

# --maxgaps positive integer

reject the target sequence if the alignment contains at least *integer* insertions or deletions.

# --maxhits positive integer

maximum number of hits to show once the search is terminated (hits are sorted by decreasing identity). The default value is 1. Set to 0 to ignore the option.

#### --maxid real

reject the target sequence if its percentage of identity with the query is equal or greater than *real*.

# --maxqsize positive integer

reject query sequences with an abundance equal or greater than integer.

# --maxqt real

reject if the query/target length ratio is equal or greater than *real*.

## --maxrejects positive integer

maximum number of non-matching target sequences to consider before stopping the search. The default value is 32. That option works in pair with maxaccepts. The search

process sorts target sequences by decreasing number of k-mers they have in common with the query sequence, using that information as a proxy for sequence similarity. If none of the first 32 target sequences pass the acceptation criteria, the search process stops for that query (no hit). If maxrejects is set to a higher value, more target sequences are considered. If maxaccepts and maxrejects are both set to 0, the complete database is searched.

#### --maxsizeratio real

reject if the query/target abundance ratio is equal or greater than real.

#### --maxsl real

reject if the shorter/longer length ratio is equal or greater than real.

#### --maxsubs positive integer

reject the target sequence if the alignment contains at least *integer* substitutions.

#### --mid real

reject the target sequence if its percentage of identity with the query is lower than *real* (ignoring gaps).

### --mincols positive integer

reject the target sequence if the alignment length is shorter than integer.

### --minqt real

reject if the query/target length ratio is lower than real.

#### --minsizeratio real

reject if the query/target abundance ratio is lower than real.

### --minsl real

reject if the shorter/longer length ratio is lower than real.

#### --mintsize positive integer

reject target sequences with an abundance lower than integer.

# --mismatch integer

score assigned to a mismatch (i.e. different nucleotides) in the pairwise alignment. The default value is -4.

## --notmatched filename

write query sequences not matching database target sequences to *filename*, in fasta format.

#### -- output no hits

write both matching and non-matching queries to output files (--alnout, --blast6out, and --userout. Output files --uc and --uc\_allhits always feature non-matching queries). Non-matching queries are labelled "no hit" in --alnout files (**to be verified**).

# --qmask none/dust/soft

mask simple repeats and low-complexity regions in query sequences using the *dust* or the *soft* algorithms, or do not mask (*none*). The default is to mask using *dust*.

# --query\_cov real

reject if the fraction of the query aligned to the target sequence is lower than real.

# --rightjust

reject the target sequence if the alignment ends with gaps.

## --rowlen positive integer

width of alignment lines in almout output. The default value is 64. Set that value to 0 to eliminate the wrapping.

**--self** reject the alignment if the query and target labels are identical.

**--selfid** reject the alignment if the query and target sequences are identical.

### --target\_cov real

reject if the fraction of the target sequence aligned to the query sequence is lower than *real*.

# --top\_hits\_only

output only the hits with the highest percentage of identity with the query.

### --userfields string

when using --userout, select and order the fields written to the output file. See the next section for a complete list of fields.

### --userout filename

write user-defined tab-separated output to *filename*. See "userfields".

## --vsearch\_global filename

filename of queries for global alignment search.

#### --weak\_id real

show hits with percentage of identity of at least *real*, without terminating the search. A normal search stops as soon as enough hits are found (as defined by --maxaccepts, --maxrejects, and --id). As --weak\_id reports weak hits that are not deduced from --maxaccepts, high --id values can be used, hence preserving both speed and sensitivity. Logically, *real* must be smaller than the value indicated by --id.

#### --wordlength positive integer

length of words (i.e. k-mers) for database index. The default value is 8.

#### Fields:

**aln** Print a string of M (match), D (delete, i.e. a gap in the query) and I (insert, i.e. a gap in the target) representing the pairwise alignment. Empty field if there is no alignment.

**alnlen** Print the length of the query-target alignment (number of columns). The field is set to 0 if there is no alignment.

**bits** Bit score (not computed for nucleotidic alignments). Always set to 0.

**caln** Compact representation of the pairwise alignment using the CIGAR format (Compact Idiosyncratic Gapped Alignment Report): M (match), D (deletion) and I (insertion).

**evalue** E-value (not computed for nucleotidic alignments). Always set to 0.

**exts** Number of columns containing a gap extension (zero or positive integer value).

**gaps** Number of columns containing a gap (zero or positive integer value).

id Percentage of identity (real value ranging from 0.0 to 100.0). (what is the formula?).

ids Number of columns containing identical nucleotides (zero or positive integer value).

**mism** Number of columns containing different nucleotides (zero or positive integer value).

**opens** Number of columns containing a gap opening (zero or positive integer value).

**pairs** Number of columns containing only nucleotides. That value corresponds to the length of the alignment minus the gap-containing columns (zero or positive integer value).

**petgaps** Number of columns containing gaps expressed as a percentage of the alignment length (real value ranging from 0.0 to 100.0).

**pctpv** Percentage of positive columns. When working with nucleotidic sequences, this is equivalent to the percentage of matches (real value ranging from 0.0 to 100.0).

**pv** Number of positive columns. When working with nucleotidic sequences, this is equivalent to the number of matches (zero or positive integer value).

**qcov** Fraction of the query sequence that is aligned with the target sequence (real value ranging from 0.0 to 100.0). **(what is the formula?)**.

**qframe** Query frame (-3 to +3). That field only concerns coding sequences and is not computed by vsearch. Always set to +0.

**qhi** First nucleotide of the query aligned with the target. Nucleotide numbering starts from 1. The field is set to 0 if there is no alignment.

**ql** Query sequence length.

**qlo** Last nucleotide of the query aligned with the target. Nucleotide numbering starts from 1. The field is set to 0 if there is no alignment.

**qrow** Print the sequence of the query segment as seen in the pairwise alignment (i.e. with gap insertions if need be). Empty field if there is no alignment.

**qs** Query segment length. Always equal to query sequence length.

**qstrand** Query strand orientation (+ or - for nucleotidic sequences).

query Query label.

raw Raw score (not computed for nucleotidic alignments). Always set to 0.

target Target label. The field is set to "\*" if there is no alignment.

Fraction of the target sequence that is aligned with the query sequence (real value ranging from 0.0 to 100.0). (what is the formula?). The field is set to 0 if there is no alignment

**tframe** Target frame (-3 to +3). That field only concerns coding sequences and is not computed by vsearch. Always set to +0.

thi Last nucleotide of the target aligned with the query. Nucleotide numbering starts from 1. The field is set to 0 if there is no alignment.

tl Target sequence length.

**tlo** First nucleotide of the target aligned with the query. Nucleotide numbering starts from 1. The field is set to 0 if there is no alignment.

**trow** Print the sequence of the target segment as seen in the pairwise alignment (i.e. with gap insertions if need be). Empty field if there is no alignment.

**ts** Target segment length. Always equal to target sequence length. The field is set to 0 if there is no alignment.

**tstrand** Target strand orientation (+ or - for nucleotidic sequences). Always set to "+", so reverse strand matches have tstrand "+" and qstrand "-".

## **DELIBERATE CHANGES**

If you are a usearch user, our objective is to make you feel at home. That's why vsearch was designed to behave like usearch, to some extend. Like any complex software, usearch is not free from quirks and inconsistencies. We decided not to reproduce some of them, and for complete transparency, to document here the deliberate changes we made.

During a search with usearch, when using the options --blast6out and --output\_no\_hits, for queries with no match the number of fields reported is 13, where it should be 12. vsearch outputs 12 fields.

# **EXAMPLES**

(in progress)

Search queries in a reference database, with a 80%-similarity threshold:

vsearch --vsearch\_global queries.fas --db references.fas --alnout results.aln --id 0.8

search a sequence dataset against itself (ignore self hits), get all matches with at least 60% identity, and collect results in a blast-like tab-separated format:

**vsearch** --vsearch\_global *queries.fas* --db *queries.fas* --id 0.6 --alnout *results.aln* --self --blast6out *results.blast6* --maxaccepts 0 --maxrejects 0

clusterize with a 97% similarity threshold, collect cluster centroids, and write cluster descriptions using a uclust-like format:

vsearch --cluster\_fast queries.fas --id 0.97 --centroids centroids.fas --uc clusters.uc

#### **LIMITATIONS**

vsearch does not yet perform chimera detection.

## **AUTHORS**

Implementation by Torbjørn Rognes and Tomas Flouri, documentation by Frédéric Mahé, .

## **REPORTING BUGS**

Submit suggestions and bug-reports at <a href="https://github.com/torognes/vsearch/issues">https://github.com/torognes/vsearch/issues</a>, send a pull request on <a href="https://github.com/torognes/vsearch">https://github.com/torognes/vsearch</a>, or compose a friendly or curmudgeont e-mail to Torbjørn Rognes <a href="https://github.com/torognes/vsearch">torognes@ifi.uio.no></a>.

# **AVAILABILITY**

The software is available from <a href="https://github.com/torognes/vsearch">https://github.com/torognes/vsearch</a>

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### **SEE ALSO**

**swipe**, an extremely fast Smith-Waterman database search tool by Torbjørn Rognes (available from <a href="https://github.com/torognes/swipe">https://github.com/torognes/swipe</a>).

# **VERSION HISTORY**

New features and important modifications of **vsearch** (short lived or minor bug releases are not mentioned):

**v1.0** released November 1st, 2014 First public release