

Ligation sequencing amplicons - Native Barcoding Kit 24 V14 (SQK-NBD114.24)

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Flow Cell Number:

DNA Samples:

INSTRUCTIONS	NOTES/OBSERVATIONS
<p>IMPORTANT</p> <p><input type="checkbox"/> Do not vortex the NEBNext Ultra II End Prep Enzyme Mix.</p> <p>IMPORTANT</p> <p><input type="checkbox"/> It is important that the NEBNext Ultra II End Prep Reaction Buffer is mixed well by vortexing.</p> <p><input type="checkbox"/> Dilute your DNA Control Sample (DCS) by adding 105 µl Elution Buffer (EB) directly to one DCS tube. Mix gently by pipetting and spin down.</p> <p>처음 키트 사용시에만 만들고 그 뒤부터는 만들어진 것 사용 35 ul + 105 EB buffer (140회 분량)</p> <p><input type="checkbox"/> In clean 0.2 ml thin-walled PCR tubes (or a clean 96-well plate), aliquot 200 fmol (130 ng for 1 kb amplicons) of DNA per sample.</p> <p><input type="checkbox"/> Make up each sample to 11.5 µl using Nuclease-free water. Mix gently by pipetting and spin down.</p> <p>Combine the following components per tube/well: total(15ul) PCR tube에 넣기</p> <p><input type="checkbox"/> 11.5 µl 200 fmol (130 ng for 1 kb amplicons) amplicon DNA</p> <p><input type="checkbox"/> 1 µl Diluted DNA Control Sample (DCS)</p> <p><input type="checkbox"/> 1.75 µl Ultra II End-prep Reaction Buffer</p> <p><input type="checkbox"/> 0.75 µl Ultra II End-prep Enzyme Mix</p> <p><input type="checkbox"/> Ensure the components are thoroughly mixed by pipetting. Close the tubes (or seal the plate) and spin down in a centrifuge.</p> <p><input type="checkbox"/> Using a thermal cycler, incubate at 20°C for 5 minutes and 65°C for 5 minutes. PCR 기계에서 바로 맞춰하기</p> <p><input type="checkbox"/> Transfer each sample into a clean 1.5 ml Eppendorf DNA LoBind tube. AXP과정 모두 PCR tube 사용</p> <p><input type="checkbox"/> Resuspend the AMPure XP beads (AXP) by vortexing.</p> <p><input type="checkbox"/> Add 15 µl of resuspended AMPure XP Beads (AXP) to each end-prep reaction and mix by flicking the tube. 1st(1X)</p> <p><input type="checkbox"/> Incubate on a Hula mixer (rotator mixer) for 5 minutes at RT.</p> <p><input type="checkbox"/> Prepare 500 µl of fresh 80% ethanol in Nuclease-free water.</p> <p><input type="checkbox"/> Spin down the samples and pellet the beads on a magnet until the eluate is clear and colourless. Keep the tubes on the magnet and pipette off the *supernatant.</p> <p><input type="checkbox"/> Keep the tube on the magnet and wash the beads with 200 µl of freshly prepared 80% ethanol without disturbing the pellet. Remove the ethanol using a pipette and discard.</p> <p><input type="checkbox"/> Repeat the previous step. 크랙이 생기지 않도록 주의!! 긴 단편의 DNA는 손상을 받을 수 있음.</p> <p><input type="checkbox"/> Briefly spin down and place the tubes back on the magnet for the beads to pellet. Pipette off any residual ethanol. Allow to dry for 30 seconds, but do not dry the pellets to the point of cracking.</p> <p><input type="checkbox"/> Remove the tubes from the magnetic rack and resuspend the pellet in 10 µl Nuclease-free water. Spin down and incubate for 2 minutes at RT.</p> <p><input type="checkbox"/> Pellet the beads on a magnet until the eluate is clear and colourless.</p>	<p>DNA 길이 별 fmol 농도 계산 해서 만들기 예) 1.5 Kb 200 fmol 194.7 ng 이 11.5ul 필요하므로 17 ng/ul 농도로 넣어주기 (이때 농도 계산은 나노드롭 사용)</p> <p>*For trouble shooting 1) 1st 상층액 보관 2) bead 버리지 않고 보관 -> 37도 incubation -> supernatant + bead reaction 다시 한번 진행</p> <p>pellet 마르지 않도록(무광상태의 경우, DNA 깨질 수 있음, long read 끊어지지 않도록 주의)</p>