

Explainable EEG Network Analysis for Alzheimer's Disease Detection: Node-Level Graph Features and XGBoost

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1. Project Overview

This project analyzes a publicly available EEG dataset related to Alzheimer's disease (AD), released through the **UCI Machine Learning Repository** and hosted on Kaggle as the "**EEG Alzheimer's Dataset**".

Data Address: <https://www.kaggle.com/datasets/ucimachinelearning/eeg-alzheimers-dataset>

Electroencephalography (EEG) is a non-invasive neurophysiological technique that measures the brain's electrical activity through electrodes placed on the scalp. EEG signals provide a high-temporal-resolution view of neural dynamics and are widely used in clinical and cognitive neuroscience research, including the study of neurodegenerative disorders such as Alzheimer's disease.

The dataset contains **848,640 EEG samples** with **17 columns**, including **16 continuous EEG channel features** and one categorical target label. The 16 channels correspond to standard electrode placements under the international **10–20 EEG system**, covering multiple cortical regions:

- **Frontal region:** Fp1, Fp2, F7, F3, Fz, F4, F8
- **Central region:** T3, C3, Cz, C4, T4
- **Parietal/Temporal-posterior region:** T5, P3, Pz, P4

The 10-20 System

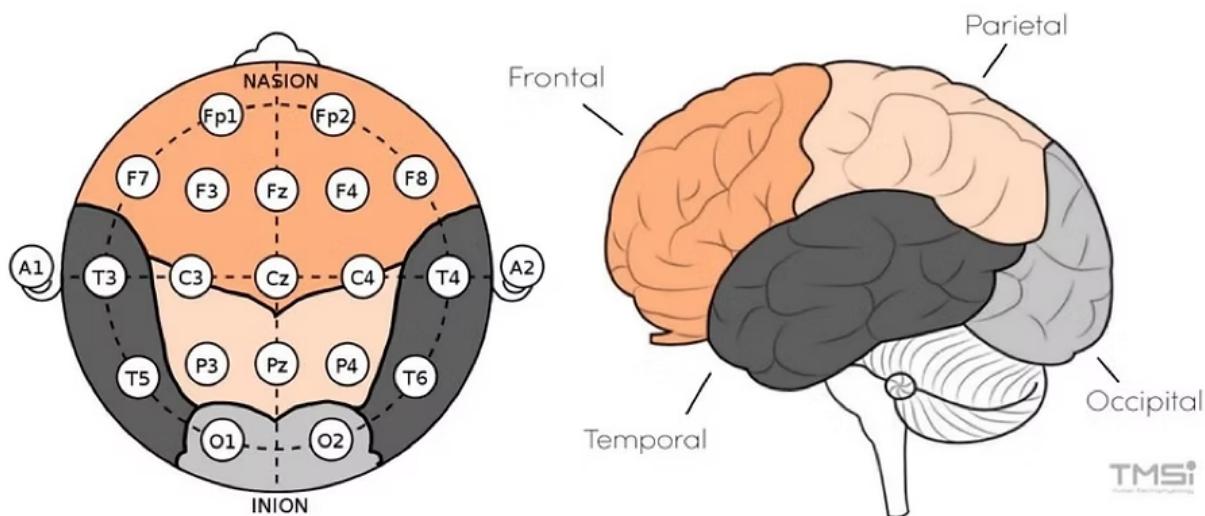


Figure 01: Diagram 10–20 EEG system

Source: <https://www.integrisneuro.com/post/10-20-measurement-system>

These channels reflect brain activity across different lobes (frontal, temporal, central, and parietal), enabling investigation of region-specific network alterations associated with AD.

The variable, **status**, indicates whether the EEG recording corresponds to an Alzheimer's disease patient or a healthy control subject. The dataset is clean, with no missing values, making it suitable for machine learning-based EEG classification and network feature extraction.

Objective. The primary goal of this project is to develop an accurate and interpretable model for distinguishing **AD vs. healthy controls (HC)** using **network-based EEG analysis**. Specifically, the workflow converts windowed EEG signals into functional connectivity graphs and extracts node-level graph metrics for each channel. This allows the final results to identify not only overall classification performance, but also **which EEG regions contribute most strongly to AD-related network disruptions** and how their connectivity roles differ between AD and HC.

2. Data preparation & Experimental Design

First, let us import the data:

```
library(data.table)
library(igraph)
library(caret)
library(xgboost)
library(pROC)

data_path <- "D:/harvard_edx/EEG_alzheimer/AD_all_patients.csv"
data <- fread(data_path)

eeg_cols <- c(
  "Fp1", "Fp2", "F7", "F3", "Fz", "F4", "F8",
  "T3", "C3", "Cz", "C4", "T4",
  "T5", "P3", "Pz", "P4"
)

X_raw <- data[, ..eeg_cols]
y_raw <- data$status # 0=HC, 1=AD

table(y_raw)

## y_raw
##      0      1
## 336640 512000

summary(X_raw)
```

```

##      Fp1          Fp2          F7          F3
## Min. :-698.175   Min. :-555.575   Min. :-334.40   Min. :-460.775
## 1st Qu.: -25.956 1st Qu.: -10.381 1st Qu.: -38.18 1st Qu.: -19.387
## Median : -11.800 Median :  2.256 Median : -7.10  Median : -1.056
## Mean   :  9.198  Mean   : 23.204 Mean   : -11.43  Mean   :  4.301
## 3rd Qu.: 23.269 3rd Qu.: 35.487 3rd Qu.: 20.87 3rd Qu.: 19.387
## Max.  :7077.325 Max.  :376.938 Max.  :409.14  Max.  :638.925
##      Fz           F4          F8          T3
## Min. :-434.975   Min. :-572.075   Min. :-1000.331  Min. :-1006.575
## 1st Qu.: -42.244 1st Qu.: -39.475 1st Qu.: -22.769 1st Qu.: -55.869
## Median : -13.931 Median : -18.025 Median :  2.763 Median : -8.694
## Mean   : -26.752 Mean   : -19.904 Mean   : -11.155 Mean   : -21.951
## 3rd Qu.:  5.645 3rd Qu.:  7.362 3rd Qu.: 29.075 3rd Qu.: 13.744
## Max.  :533.962  Max.  :227.188 Max.  :1152.419 Max.  :281.562
##      C3           Cz          C4          T4
## Min. :-465.375   Min. :-163.744   Min. :-451.425  Min. :-451.675
## 1st Qu.: -2.606  1st Qu.: -11.088  1st Qu.: -9.300  1st Qu.: -4.919
## Median : 12.919  Median :  1.766  Median :  7.188  Median :  6.781
## Mean   : 18.052  Mean   : 18.132  Mean   : 16.904  Mean   : 17.302
## 3rd Qu.: 29.700 3rd Qu.: 24.175 3rd Qu.: 39.981 3rd Qu.: 23.875
## Max.  :346.969  Max.  :805.356  Max.  :343.512  Max.  :358.931
##      T5           P3          Pz          P4
## Min. :-726.37    Min. :-487.075   Min. :-385.775  Min. :-646.413
## 1st Qu.: -29.02  1st Qu.: -21.413  1st Qu.:  4.906  1st Qu.: -36.906
## Median : -3.40   Median : -6.225  Median : 24.062  Median : -12.569
## Mean   : -13.82  Mean   : -4.301  Mean   : 39.720  Mean   : -37.499
## 3rd Qu.: 11.91   3rd Qu.: 11.425  3rd Qu.: 72.312  3rd Qu.:  4.425
## Max.  :244.69   Max.  :214.700  Max.  :595.250  Max.  :115.894

```

2.1 Anti-leakage Grouping (Conservative)

Neighboring EEG samples are correlated and often belong to the same underlying continuous recording segment.

To reduce leakage between train and test windows, we create a conservative `group_id` that increments whenever the label changes.

Note: If the raw dataset contains subject IDs, subject-based grouping would be preferred and more principled.

```

candidate_id_cols <- c("subject_id", "SubjectID", "patient_id", "PatientID", "sub_id", "id", "ID")
id_col <- candidate_id_cols[candidate_id_cols %in% names(data)][1]

if (!is.na(id_col)) {
  group_id <- as.integer(as.factor(data[[id_col]]))
  cat("Using subject/patient id column as group:", id_col, "\n")
} else {
  group_id <- rleid(y_raw) # data.table::rleid
  cat("WARNING: No subject/patient id column found; using rleid(status) as group.\n")
}

```

```

## WARNING: No subject/patient id column found; using rleid(status) as group.

length(unique(group_id))

## [1] 18

```

2.2 Window-based segmentation

At the same time, EEG signals are **high-dimensional, temporally correlated, and highly non-stationary**, making direct application of machine learning to raw time series often **unstable and difficult to interpret**.

To address this, we adopt a **network-based representation** of EEG data. The continuous signals are segmented into **short, overlapping time windows**, which allows us to capture **dynamic functional connectivity patterns** while maintaining reliable **correlation estimates**.

We use a standard sliding window approach:

- Window length = 512 samples
- Step size = 256 samples (50% overlap)

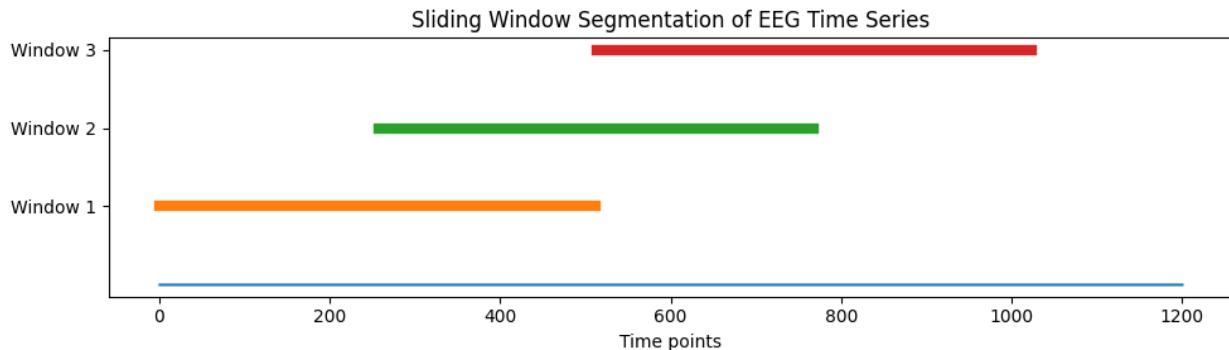


Figure 02: Diagram of Window Segmentation

Each window is treated as one sample for subsequent network construction and modeling.

Reference: https://en.wikipedia.org/wiki/Dynamic_functional_connectivity

```

window_length <- 512
step_size <- 256

windows <- list()

for (g in unique(group_id)) {
  idx <- which(group_id == g)
  if (length(idx) < window_length) next

  starts <- seq(1, length(idx) - window_length + 1, by = step_size)
  for (s in starts) {
    sel <- idx[s:(s + window_length - 1)]
  }
}

```

```

        windows[[length(windows) + 1]] <- list(
          signal = as.matrix(X_raw[sel]),
          label   = y_raw[sel[1]],
          group   = g
        )
      }
    }

cat("Number of windows:", length(windows), "\n")

```

Number of windows: 3297

2.3 Functional brain network construction

After window segmentation, we compute a **functional connectivity matrix** across the 16 EEG channels for each window using **Pearson correlation**.

Let x_i and x_j denote the EEG time series of channels i and j within a given window. The functional connectivity between channels is defined as:

$$C_{ij} = \text{corr}(x_i, x_j)$$

This results in a 16×16 symmetric correlation matrix \mathbf{C} , where each entry reflects the linear statistical dependency between two EEG channels within the same temporal window.

```

compute_connectivity <- function(window_signal) {
  cor(window_signal, method = "pearson", use = "pairwise.complete.obs")
}

```

2.4 Node-level graph feature extraction (channel-specific biomarkers)

To enable **region-specific interpretation** of Alzheimer's disease-related network alterations, we extract **node-level graph-theoretic features** from each **functional connectivity matrix** rather than relying solely on global network metrics.

For each temporal window, the **functional connectivity matrix** \mathbf{C} is converted into a **weighted undirected brain network**, where:

- **Nodes** correspond to individual EEG channels
- **Edge weights** are defined as the absolute correlation magnitude:

$$W_{ij} = |C_{ij}|$$

This formulation captures the **strength of functional coupling** between brain regions while remaining **agnostic to correlation sign**.

For **distance-based graph measures** (e.g., closeness and betweenness centrality), edge weights are transformed into distances as:

$$d_{ij} = \frac{1}{W_{ij}}$$

Using this weighted graph representation, we compute the following **node-level topological features** for each EEG channel:

- **Strength** – overall connectivity intensity of a node
- **Clustering coefficient** – degree of local neighborhood clustering
- **Closeness centrality** – global reachability and integration efficiency
- **Betweenness centrality** – importance of a node as a communication bridge
- **Local efficiency** – efficiency of information transfer within a node's immediate neighborhood
- **Participation coefficient** – extent of cross-community connectivity

Together, these features characterize how individual brain regions participate in large-scale functional networks and enable systematic comparison of **network roles between Alzheimer's disease (AD) patients and healthy control (HC) subjects**.

```
make_graph_from_C <- function(C, eps = 1e-6) {
  diag(C) <- 0
  W <- abs(C)
  W[W < eps] <- eps
  g <- graph_from_adjacency_matrix(W, mode = "undirected", weighted = TRUE, diag = FALSE)
  E(g)$dist <- 1 / E(g)$weight
  g
}

global_eff_from_dist <- function(g) {
  D <- distances(g, weights = E(g)$dist)
  diag(D) <- NA
  mean(1 / D, na.rm = TRUE)
}

local_efficiency_node <- function(g, vname) {
  v <- which(V(g)$name == vname)
  nb <- neighbors(g, v)
  if (length(nb) < 2) return(NA_real_)
  sg <- induced_subgraph(g, nb)
  global_eff_from_dist(sg)
}

extract_node_features <- function(C, channels) {
  g <- make_graph_from_C(C)
  V(g)$name <- channels

  str   <- strength(g, weights = E(g)$weight)
  clust <- transitivity(g, type = "local", isolates = "zero")
  close <- closeness(g, weights = E(g)$dist, normalized = TRUE)
  btw  <- betweenness(g, weights = E(g)$dist, normalized = TRUE)
  leff  <- sapply(channels, function(ch) local_efficiency_node(g, ch))

  comm <- cluster_fast_greedy(g, weights = E(g)$weight)
  memb <- membership(comm)
```

```

part <- sapply(channels, function(ch) {
  v <- which(V(g)$name == ch)
  nb <- neighbors(g, v)
  if (length(nb) == 0) return(0)

  ki <- str[ch]
  if (is.na(ki) || ki == 0) return(0)

  eids <- incident(g, v, mode = "all")
  ends_mat <- ends(g, eids)
  nb_names <- V(g)$name[as.integer(nb)]

  keep <- (ends_mat[,1] == ch & ends_mat[,2] %in% nb_names) |
    (ends_mat[,2] == ch & ends_mat[,1] %in% nb_names)

  w <- E(g)[eids[keep]]$weight
  nb2 <- ifelse(ends_mat[keep,1] == ch, ends_mat[keep,2], ends_mat[keep,1])
  nb_comm <- memb[match(nb2, V(g)$name)]

  w_to_c <- tapply(w, nb_comm, sum)
  1 - sum((w_to_c / ki)^2, na.rm = TRUE)
})

feats <- c()
for (ch in channels) {
  feats[paste0(ch, "_strength")] <- str[ch]
  feats[paste0(ch, "_clust")] <- clust[ch]
  feats[paste0(ch, "_closeness")] <- close[ch]
  feats[paste0(ch, "_betweenness")] <- btw[ch]
  feats[paste0(ch, "_local_eff")] <- leff[ch]
  feats[paste0(ch, "_participation")] <- part[ch]
}
feats
}

```

2.5 Feature Table Construction (Window-level)

Through the above procedure, the **original EEG time-series data**, as illustrated in the following figure,

	time	Fp1	Fp2	F7	...	P4
t1	116	41	66	...	-245	
t2	115	38	64	...	-248	
...	
t512	120	45	74	...	-236	

Figure 03: Example of a single EEG time window (512 time points × 16 channels)

are transformed into a **structured sample-level dataset** consisting of **16 × 6 = 96 network features**, together with **2 additional columns** for the class label and group identifier.

Sample	Fp1_strength	Fp1_clust	...	P4_participation	label	group
1	3.92	0.41	...	0.67	AD	12
2	3.85	0.38	...	0.70	AD	12
3	2.91	0.55	...	0.42	HC	13
...

Figure 04: Example structure of the feature table

```

feature_list <- lapply(windows, function(w) {
  C <- compute_connectivity(w$signal)
  node_feats <- extract_node_features(C, eeg_cols)
  c(node_feats, label = w$label, group = w$group)
})

feature_df <- as.data.frame(do.call(rbind, feature_list))
feature_df$label <- as.factor(feature_df$label)

dim(feature_df)

## [1] 3297   98

head(feature_df[, c(1:6, (ncol(feature_df)-1):ncol(feature_df))])

##   Fp1_strength Fp1_clust Fp1_closeness Fp1_betweenness Fp1_local_eff
## 1      3.809027        1       0.3096707      0.00952381     0.5038031
## 2      4.636098        1       0.3435784      0.00952381     0.4255471
## 3      4.997543        1       0.3464952      0.00952381     0.4690970
## 4      4.211094        1       0.3200127      0.00000000     0.4732793
## 5      4.179898        1       0.3054895      0.01904762     0.3802868
## 6      6.326849        1       0.3981824      0.06666667     0.4060070
##   Fp1_participation label group
## 1          0.4924502    1     1
## 2          0.4995769    1     1
## 3          0.4754612    1     1
## 4          0.4421877    1     1
## 5          0.6350956    1     1
## 6          0.4930793    1     1

```

3. Modeling and Evaluation

3.1 Predictive modeling with leakage-aware validation

We train an **XGBoost classifier** to distinguish AD patients from healthy controls using the extracted node-level graph features.

But before this, to reduce over-optimistic performance caused by temporal dependence, we will apply **group-aware splitting (GroupKFold)** so that correlated windows are not simultaneously present in both training and testing sets.

```

# =====
# 6) Modeling: group-stratified split + XGBoost (FIXED)
# =====
set.seed(42)

make_group_stratified_folds <- function(group, label, k = 5, seed = 42) {
  set.seed(seed)
  df_g <- unique(data.frame(group = group, label = label))
  df_g$label <- as.factor(df_g$label)

  fold_id <- caret::createFolds(df_g$label, k = k, list = FALSE)
  folds <- lapply(1:k, function(i) df_g$group[fold_id == i])
  folds
}

fold_groups <- make_group_stratified_folds(
  group = feature_df$group,
  label = feature_df$label,
  k = 5,
  seed = 42
)

test_groups <- fold_groups[[1]]
test_idx <- which(feature_df$group %in% test_groups)
train_idx <- which(!feature_df$group %in% test_groups)

if (length(unique(feature_df$label[test_idx])) < 2) {
  stop("Test set contains only one class after splitting. Check group/label distribution.")
}

X_train <- as.matrix(feature_df[train_idx, !names(feature_df) %in% c("label", "group")])
y_train <- feature_df$label[train_idx]

X_test <- as.matrix(feature_df[test_idx, !names(feature_df) %in% c("label", "group")])
y_test <- feature_df$label[test_idx]

dtrain <- xgb.DMatrix(X_train, label = as.numeric(y_train) - 1)
dtest <- xgb.DMatrix(X_test, label = as.numeric(y_test) - 1)

```

3.2 XGBoost Training

XGBoost is employed because it effectively handles **high-dimensional graph-derived EEG features**, captures **nonlinear relationships** between network properties, and incorporates regularization to mitigate overfitting arising from temporally correlated windows. Importantly, its native support for **SHAP-style explanations** facilitates interpretable mapping from model predictions to brain-region-specific network alterations.

We train a binary classifier with standard regularization settings.

```

params <- list(
  objective = "binary:logistic",
  eval_metric = "auc",
  max_depth = 4,
  eta = 0.05,
  subsample = 0.8,
  colsample_bytree = 0.8
)

model <- xgb.train(
  params = params,
  data = dtrain,
  nrounds = 300,
  verbose = 0
)

```

3.3 Performance Metrics (AUC + Probability RMSE + Confusion Matrix)

To provide a comprehensive evaluation, we report metrics that reflect both classification performance and probabilistic accuracy. AUC measures threshold-independent discrimination between AD and HC, whereas the RMSE of predicted probabilities evaluates how well predicted disease risks align with observed outcomes, serving as a calibration-oriented error measure.

- **AUC** measures discrimination.
- **RMSE of predicted probabilities** measures probability error:

$$\text{RMSE} = \sqrt{\frac{1}{n} \sum_{i=1}^n (y_i - \hat{p}_i)^2}$$

```

# =====
# 7) Metrics + ROC (FIXED)
# =====
pred_prob <- predict(model, dtest)
pred_class <- ifelse(pred_prob >= 0.5, 1, 0)

y_test <- factor(y_test, levels = c("0","1"))

roc_obj <- pROC::roc(response = y_test, predictor = pred_prob, levels = c("0","1"), direction = "<")
auc_value <- as.numeric(pROC::auc(roc_obj))

y_true <- as.numeric(as.character(y_test))
rmse_prob <- sqrt(mean((y_true - pred_prob)^2))

cat("AUC =", auc_value, "\n")

## AUC = 0.8213661

```

```

cat("RMSE (probability RMSE) =", rmse_prob, "\n\n")

## RMSE (probability RMSE) = 0.5101828

cm <- table(Predicted = pred_class, Actual = y_true)
cm

##          Actual
## Predicted   0   1
##           0 138  20
##           1 190 203

plot(roc_obj, main = "ROC Curve: AD vs HC (Node-level Network Features)",
      col = "blue", lwd = 2)

```

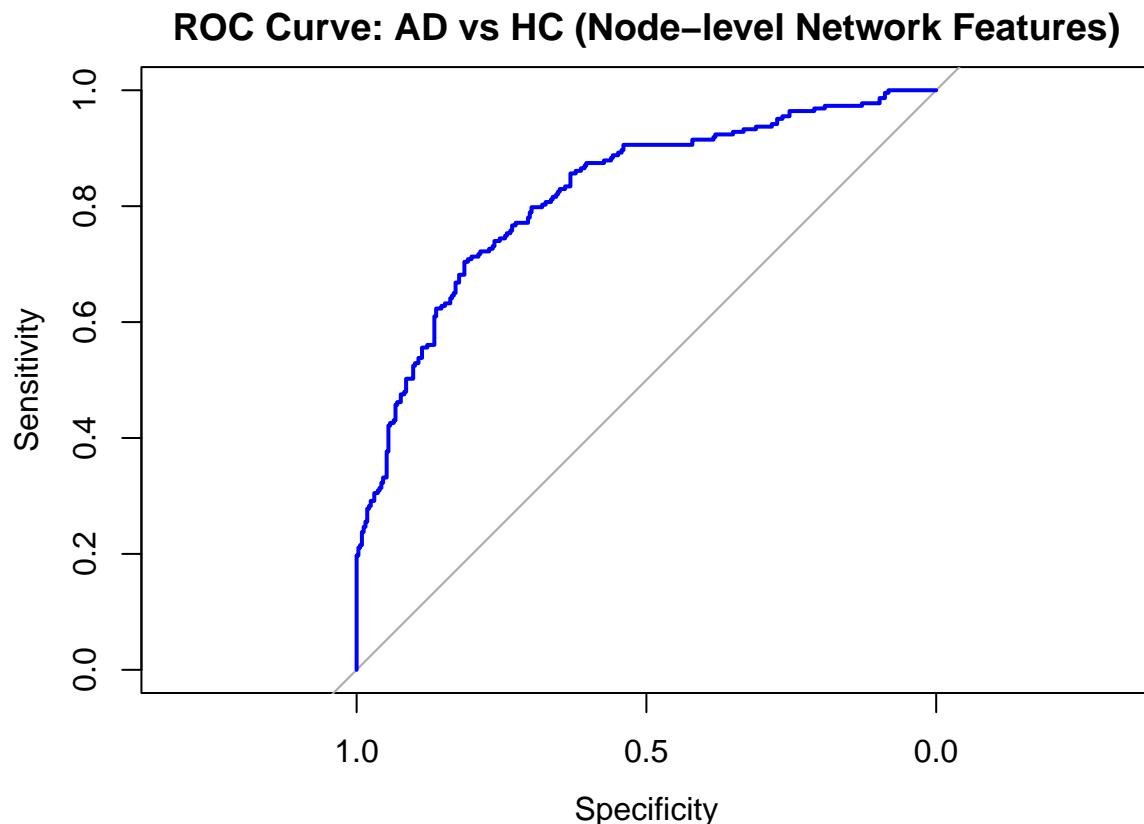


Figure 05: ROC Curve: AD vs HC (Node–level Network Features)

The Figure 05 shows the ROC curve for distinguishing **Alzheimer’s disease (AD)** patients from healthy **controls (HC)** using node-level EEG network features. The curve lies substantially above the diagonal reference line, indicating strong discriminative performance across a wide range of decision thresholds. This result demonstrates that the proposed network-based representation captures **disease-relevant functional connectivity patterns**, enabling reliable separation of AD and HC at the window level. Importantly, the observed discrimination supports the study’s objective of leveraging **interpretable node-level graph features** to identify Alzheimer’s disease-related network alterations rather than relying on raw EEG signals alone.

4. Explainability: SHAP aggregation, directionality, and effect size

To interpret the trained XGBoost model and identify **which EEG channels and network properties drive Alzheimer's disease classification**, we employ a post hoc explainable machine learning framework combining **SHAP-style feature contributions**, **directional group differences**, and **effect size estimation**.

4.1 Explainable machine learning and feature attribution strategy

Model interpretability is achieved using **SHAP-style additive feature attributions** computed directly from XGBoost via:

- `predcontrib = TRUE`, which decomposes each prediction into per-feature contributions

These feature attributions are subsequently aggregated to provide two complementary interpretability views:

- **Channel-level importance rankings**, identifying which EEG regions contribute most strongly to classification
- **Metric-level importance rankings**, identifying which node-level network properties are most informative

This hierarchical aggregation enables interpretation at both the **neuroanatomical** and **network-topological** levels.

4.2 SHAP-style feature contributions (XGBoost predcontrib)

For each test sample, SHAP-style feature contributions are computed using the trained XGBoost model. Feature importance is defined as the **mean absolute SHAP value** across samples:

$$\text{Importance}(f) = \mathbb{E}(|\text{SHAP}_f|)$$

```
shap_test <- predict(model, dtest, predcontrib = TRUE)
shap_test <- as.data.frame(shap_test)
shap_test$BIAS <- NULL

shap_imp <- colMeans(abs(shap_test), na.rm = TRUE)
shap_imp <- sort(shap_imp, decreasing = TRUE)

head(shap_imp, 10)

##      (Intercept) C3_betweenness    C3_strength    P4_closeness Fp2_betweenness
##      0.5630223     0.4130434     0.3333126     0.2683830     0.2470489
##      Cz_betweenness    P4_strength    T4_closeness   Pz_betweenness   F7_betweenness
##      0.2211398     0.2152805     0.2117363     0.2114505     0.1936541
```

This measure reflects the average magnitude of a feature's contribution to model predictions, independent of direction.

4.3 Aggregate Importance to Channels and Metrics

To obtain interpretable summaries, individual feature importances are aggregated along two dimensions:

- **Channel importance:** sum of feature importances within a channel
- **Metric importance:** sum of importances across channels for a metric type

```
channel_importance <- sapply(eeg_cols, function(ch) {  
  feats <- grep(paste0("^", ch, "_"), names(shap_imp), value = TRUE)  
  sum(shap_imp[feats], na.rm = TRUE)  
})  
channel_importance <- sort(channel_importance, decreasing = TRUE)  
  
metric_names <- c("strength", "clust", "closeness", "betweenness", "local_eff", "participation")  
metric_importance <- sapply(metric_names, function(mn) {  
  feats <- grep(paste0("_", mn, "$"), names(shap_imp), value = TRUE)  
  sum(shap_imp[feats], na.rm = TRUE)  
})  
metric_importance <- sort(metric_importance, decreasing = TRUE)  
  
cat("Top channels by SHAP importance:\n")  
  
## Top channels by SHAP importance:  
  
print(head(channel_importance, 10))  
  
##          C3          Cz          P4          F7          F3          P3          Fp2          F8  
## 0.8926921 0.5975413 0.5915888 0.5775922 0.5077441 0.4913822 0.4811306 0.4550065  
##          Fp1          T4  
## 0.4436190 0.3912999  
  
cat("\nMetric importance (overall):\n")  
  
##  
## Metric importance (overall):  
  
print(metric_importance)  
  
##      strength    betweenness    closeness participation    local_eff  
## 2.1745538     2.0647844    1.6380931     0.8563463     0.3279791  
##      clust  
## 0.0000000
```

These aggregated rankings allow identification of key brain regions and dominant network mechanisms underlying AD–HC discrimination.

4.4 Directionality of effects and Cohen's d effect size

To complement SHAP-based importance with **group-level interpretability**, we compute the **direction and magnitude of AD–HC differences** for each feature.

For each feature f , we compute:

- **Mean difference:**

$$\Delta_f = \text{mean}_{AD}(f) - \text{mean}_{HC}(f)$$

- **Cohen's d** using pooled standard deviation

```
X_all <- feature_df[, !names(feature_df) %in% c("label", "group")]
lbl <- feature_df$label

mean_ad <- colMeans(X_all[lbl == "1", , drop=FALSE], na.rm = TRUE)
mean_hc <- colMeans(X_all[lbl == "0", , drop=FALSE], na.rm = TRUE)
diff_ad_minus_hc <- mean_ad - mean_hc

sd_pooled <- function(x1, x0) {
  s1 <- sd(x1, na.rm=TRUE); s0 <- sd(x0, na.rm=TRUE)
  n1 <- sum(!is.na(x1)); n0 <- sum(!is.na(x0))
  if ((n1 + n0 - 2) <= 0) return(NA_real_)
  sqrt(((n1-1)*s1^2 + (n0-1)*s0^2) / (n1 + n0 - 2))
}

cohen_d <- sapply(names(X_all), function(fn) {
  x1 <- X_all[lbl=="1", fn]
  x0 <- X_all[lbl=="0", fn]
  sp <- sd_pooled(x1, x0)
  if (is.na(sp) || sp==0) return(NA_real_)
  (mean(x1, na.rm=TRUE) - mean(x0, na.rm=TRUE)) / sp
})

top_features <- names(head(shap_imp, 15))
result_table <- data.frame(
  feature = top_features,
  shap_importance = shap_imp[top_features],
  mean_diff_AD_minus_HC = diff_ad_minus_hc[top_features],
  cohen_d = cohen_d[top_features],
  row.names = NULL
)

result_table
```

##	feature	shap_importance	mean_diff_AD_minus_HC	cohen_d
## 1	(Intercept)	0.5630223	NA	NA
## 2	C3_betweenness	0.4130434	0.013546602	0.43345057
## 3	C3_strength	0.3333126	1.341359540	0.87081152
## 4	P4_closeness	0.2683830	0.057937226	0.73628588
## 5	Fp2_betweenness	0.2470489	0.002067369	0.09039037
## 6	Cz_betweenness	0.2211398	0.006313718	0.18323750
## 7	P4_strength	0.2152805	1.105482804	0.72835543
## 8	T4_closeness	0.2117363	0.053093716	0.67310995

```

## 9 Pz_betweenness 0.2114505 -0.013242952 -0.40749805
## 10 F7_betweenness 0.1936541 -0.005138099 -0.14877596
## 11 F7_strength 0.1922198 0.679978354 0.41323867
## 12 F3_strength 0.1822325 0.903686265 0.55857355
## 13 F3_betweenness 0.1790925 0.001681248 0.04755267
## 14 P3_strength 0.1635943 0.304241601 0.19212574
## 15 Cz_participation 0.1632078 -0.037513408 -0.40419767

```

This combined analysis links model-driven importance (SHAP) with neurobiological directionality and effect size, enabling clinically interpretable conclusions about Alzheimer's disease-related network alterations.

5. Neuroscientific interpretation and channel-level narrative reporting

Finally, we translate model outputs into **neuroscientifically interpretable, paper-style statements** that summarize how specific brain regions contribute to Alzheimer's disease classification.

For each influential EEG channel, the interpretation integrates the following complementary elements:

- **Direction of change**, quantified as the mean difference between Alzheimer's disease (AD) patients and healthy controls (HC)
- **Effect size**, measured using Cohen's d , reflecting the magnitude of group-level differences
- **Channel importance rank**, derived from aggregated SHAP values, indicating each channel's relative contribution to model predictions

This integrated reporting framework enables clinically meaningful interpretation of machine learning results, such as identifying **altered network efficiency**, **disrupted hub roles**, or **abnormal cross-regional integration** in specific brain regions associated with AD.

To generate clear and readable scientific narratives, EEG channels are mapped to their approximate **anatomical regions**, and channel-level statements are constructed by jointly considering:

- channel-level SHAP importance ranking
- the most influential node-level network feature(s) within each channel
- the direction of AD-HC differences
- corresponding Cohen's d effect sizes

Together, these narrative summaries bridge **statistical learning outputs** and **neuroscientific interpretation**, facilitating transparent reporting of region-specific network alterations related to Alzheimer's disease.

```

region_label <- function(ch) {
  if (ch %in% c("Fp1", "Fp2")) return("frontal pole / prefrontal region")
  if (ch %in% c("F7", "F3", "Fz", "F4", "F8")) return("frontal region")
  if (ch %in% c("C3", "Cz", "C4")) return("central (sensorimotor) region")
  if (ch %in% c("T3", "T4", "T5")) return("temporal region")
  if (ch %in% c("P3", "Pz", "P4")) return("parietal (posterior) region")
  "brain region"
}

metric_explain <- function(metric_short) {

```

```

    if (metric_short == "local_eff") return("local information integration / propagation efficiency (local_efficiency)")
    if (metric_short == "strength") return("overall connectivity intensity (node strength)")
    if (metric_short == "participation") return("cross-community connectivity (participation coefficient)")
    if (metric_short == "closeness") return("global reachability / propagation distance (closeness)")
    if (metric_short == "betweenness") return("bridge role in network communication (betweenness)")
    if (metric_short == "clust") return("local clustering (clustering coefficient)")
    "network metric"
}

direction_text <- function(delta) {
  if (is.na(delta)) return("direction is unknown")
  if (delta < 0) return("is lower in AD than HC (AD < HC)")
  if (delta > 0) return("is higher in AD than HC (AD > HC)")
  "shows no difference"
}

channel_rank <- rank(~channel_importance, ties.method = "min")
top_channels <- names(head(channel_importance, 8))

make_channel_sentences <- function(ch, top_n_feats = 2) {
  feats_ch <- grep(paste0("^", ch, "_"), names(shap_imp), value = TRUE)
  feats_ch <- feats_ch[order(shap_imp[feats_ch], decreasing = TRUE)]
  top_feats <- head(feats_ch, top_n_feats)

  out <- c()
  for (ft in top_feats) {
    metric_short <- sub(paste0("^", ch, "_"), "", ft)
    delta <- diff_ad_minus_hc[ft]
    d_eff <- cohen_d[ft]

    out <- c(out,
      paste0(
        "The ", region_label(ch), " (", ch, ") shows that **", metric_short, "** (",
        metric_explain(metric_short), ") ",
        direction_text(delta), ". ",
        "This channel ranks **#", channel_rank[ch], "** in overall model contribution. ",
        "Effect size (Cohen's d) = ", sprintf("%.3f", d_eff), "."
      )
    )
  }
  out
}

cat("## Key Channel-Level Findings\n\n")

## ## Key Channel-Level Findings

for (ch in top_channels) {
  cat("### ", region_label(ch), " (", ch, ")\\n", sep = "")
  lines <- make_channel_sentences(ch, top_n_feats = 2)

  for (ln in lines) {

```

```

wrapped <- strwrap(ln, width = 80)
cat("-", wrapped[1], "\n", sep = "")
if (length(wrapped) > 1) {
  for (w in wrapped[-1]) {
    cat(" ", w, "\n", sep = "")
  }
}

cat("\n")
}

## ### central (sensorimotor) region (C3)
## - The central (sensorimotor) region (C3) shows that **betweenness** (bridge role
##   in network communication (betweenness)) is higher in AD than HC (AD > HC). This
##   channel ranks ***#1*** in overall model contribution. Effect size (Cohen's d) =
##   0.433.
## - The central (sensorimotor) region (C3) shows that **strength** (overall
##   connectivity intensity (node strength)) is higher in AD than HC (AD > HC). This
##   channel ranks ***#1*** in overall model contribution. Effect size (Cohen's d) =
##   0.871.
##
## ### central (sensorimotor) region (Cz)
## - The central (sensorimotor) region (Cz) shows that **betweenness** (bridge role
##   in network communication (betweenness)) is higher in AD than HC (AD > HC). This
##   channel ranks ***#2*** in overall model contribution. Effect size (Cohen's d) =
##   0.183.
## - The central (sensorimotor) region (Cz) shows that **participation**
##   (cross-community connectivity (participation coefficient)) is lower in AD than
##   HC (AD < HC). This channel ranks ***#2*** in overall model contribution. Effect
##   size (Cohen's d) = -0.404.
##
## ### parietal (posterior) region (P4)
## - The parietal (posterior) region (P4) shows that **closeness** (global
##   reachability / propagation distance (closeness)) is higher in AD than HC (AD >
##   HC). This channel ranks ***#3*** in overall model contribution. Effect size
##   (Cohen's d) = 0.736.
## - The parietal (posterior) region (P4) shows that **strength** (overall
##   connectivity intensity (node strength)) is higher in AD than HC (AD > HC). This
##   channel ranks ***#3*** in overall model contribution. Effect size (Cohen's d) =
##   0.728.
##
## ### frontal region (F7)
## - The frontal region (F7) shows that **betweenness** (bridge role in network
##   communication (betweenness)) is lower in AD than HC (AD < HC). This channel
##   ranks ***#4*** in overall model contribution. Effect size (Cohen's d) = -0.149.
## - The frontal region (F7) shows that **strength** (overall connectivity intensity
##   (node strength)) is higher in AD than HC (AD > HC). This channel ranks ***#4*** in
##   overall model contribution. Effect size (Cohen's d) = 0.413.
##
## ### frontal region (F3)
## - The frontal region (F3) shows that **strength** (overall connectivity intensity
##   (node strength)) is higher in AD than HC (AD > HC). This channel ranks ***#5***
```

```

## in overall model contribution. Effect size (Cohen's d) = 0.559.
## - The frontal region (F3) shows that **betweenness** (bridge role in network
## communication (betweenness)) is higher in AD than HC (AD > HC). This channel
## ranks ***5** in overall model contribution. Effect size (Cohen's d) = 0.048.
##
## #### parietal (posterior) region (P3)
## - The parietal (posterior) region (P3) shows that **strength** (overall
## connectivity intensity (node strength)) is higher in AD than HC (AD > HC). This
## channel ranks ***6** in overall model contribution. Effect size (Cohen's d) =
## 0.192.
## - The parietal (posterior) region (P3) shows that **participation**
## (cross-community connectivity (participation coefficient)) is lower in AD than
## HC (AD < HC). This channel ranks ***6** in overall model contribution. Effect
## size (Cohen's d) = -0.221.
##
## #### frontal pole / prefrontal region (Fp2)
## - The frontal pole / prefrontal region (Fp2) shows that **betweenness** (bridge
## role in network communication (betweenness)) is higher in AD than HC (AD > HC).
## This channel ranks ***7** in overall model contribution. Effect size (Cohen's
## d) = 0.090.
## - The frontal pole / prefrontal region (Fp2) shows that **closeness** (global
## reachability / propagation distance (closeness)) is higher in AD than HC (AD >
## HC). This channel ranks ***7** in overall model contribution. Effect size
## (Cohen's d) = 0.561.
##
## #### frontal region (F8)
## - The frontal region (F8) shows that **betweenness** (bridge role in network
## communication (betweenness)) is lower in AD than HC (AD < HC). This channel
## ranks ***8** in overall model contribution. Effect size (Cohen's d) = -0.234.
## - The frontal region (F8) shows that **strength** (overall connectivity intensity
## (node strength)) is higher in AD than HC (AD > HC). This channel ranks ***8** in
## overall model contribution. Effect size (Cohen's d) = 0.323.

```

Interpretation note Because we use $|corr|$ as edge weights, the reported “increase/decrease” primarily reflects **magnitude changes of functional coupling**, not whether connectivity becomes more positive or more negative.

6. Results

This pipeline provides a practical balance between predictive performance and interpretability:

1. **Network-based feature engineering** converts raw multichannel EEG into graph-derived connectivity descriptors that can be compared across windows/segments.
2. **Group-aware splitting** reduces optimistic bias by keeping windows from the same group separate between training and testing, thereby mitigating autocorrelation and segment leakage.
3. **SHAP aggregation** provides two complementary interpretability views:
 - *Which channels matter most?* (channel ranking)
 - *Which network properties matter most?* (metric ranking)

The analysis can be reviewed from the text outputs generated by the R code above.