# March 2022 host and holobiont protein analysis

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## Import data for host protein from

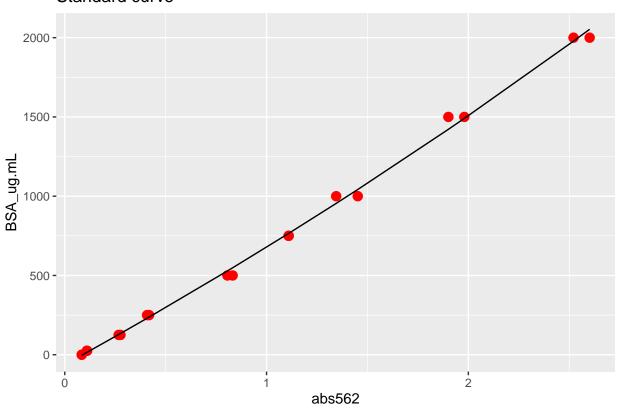
```
# List protein data files
prot_path = "baseline_TPC_physio/data/TPC_total_protein/host/"
all_prot_files <- list.files(path = prot_path, pattern = "*.csv")
                                                                            # List all files in director
prot_platemaps <- list.files(path = prot_path, pattern = "platemap")</pre>
                                                                            # List platemap files
prot_data_files <- setdiff(all_prot_files, prot_platemaps)</pre>
                                                                             # List data files
# Read in all files into tibble
df_host <- tibble(file = prot_data_files) %>%
  separate(file, into = c("trip", "date", "plate"), remove = FALSE) %%
  unite(plate, trip, date, plate) %>%
  dplyr::mutate(platemap = map(plate, ~read_csv(paste0(prot_path, ., "_platemap.csv"))),
         prot_data = map(file, ~read_csv(paste0(prot_path, .)) %>% rename(well = Well)))
# Merge platemap and data for each plate
df host <- df host %>%
 mutate(merged = map2(platemap, prot_data, ~ right_join(.x, .y)))
```

#### Plot standard curve

```
# Create standard curve following kit instructions
standards_host <- tribble(</pre>
  ~std, ~BSA_ug.mL,
 "A",
              2000,
  "B",
              1500,
  "C",
              1000,
  "D",
               750,
  "E",
               500.
  "F",
               250,
  "G",
              125,
  "H",
               25,
  "I",
std_curve_host <- df_host %>%
  unnest(merged) %>%
  filter(grepl("Standard", fragment_ID)) %>%
  select(plate, well, fragment_ID, abs562 = `562:562`) %>%
```

```
rename(std = fragment_ID) %>%
       mutate(std = str_sub(std, 9, 9)) %>%
       #group_by(std) %>%
       #summarise(abs562 = mean(abs562)) %>%
                                                                                                                                                                                                                                      # calculate mean of standard duplicates
       #mutate(abs562.adj = abs562 - abs562[std == "I"]) %>%
                                                                                                                                                                                                                                     # subtract blank absorbace value from all
       left_join(standards_host)
## Fit nonlinear model for standard curve
mod_host \leftarrow nls(formula = BSA_ug.mL \sim z + a * exp(b * abs562), start = list(z = 0, a = 1, b = 1), data = constant = cons
fitted_host <- mod_host %>% broom::augment()
# Plot standard curve
std_curve_plot_host <- std_curve_host %>%
      ggplot(aes(x = abs562, y = BSA_ug.mL)) +
      geom_point(color = "red", size = 3)
std_curve_plot_host +
       geom_line(\frac{data}{data} = fitted_host, aes(x = abs562, y = .fitted)) +
      labs(title = "Standard curve")
```

#### Standard curve



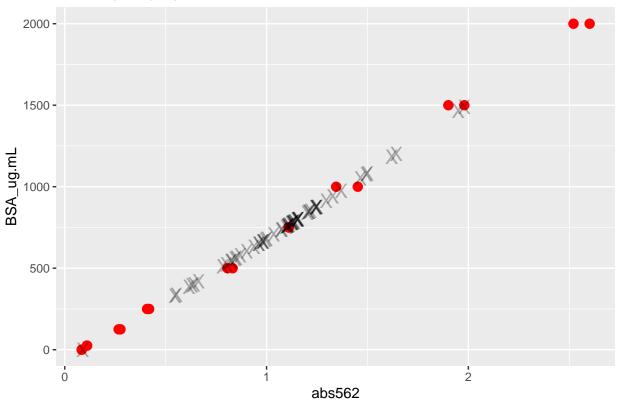
# Calculate protein concentrations

```
# Calculate protein concentration for all samples using standard curve prot_host <- df_host %>%
```

```
unnest(merged) %>%
filter(!grep1("Standard", fragment_ID)) %>%  # Get just samples (not standards)
select(plate, well, fragment_ID, abs562 = `562:562`) %>%  # Select only needed columns
filter(!is.na(fragment_ID)) %>%  # Filter out empty wells
filter(fragment_ID != "BK") %>%  # Filter out blank wells
mutate(prot_ug.mL = map_dbl(abs562, ~ predict(mod_host, newdata = data.frame(abs562 = .))))  # Use

std_curve_plot_host +
geom_point(data = prot_host, aes(x = abs562, y = prot_ug.mL), pch = "X", cex = 5, alpha = 0.3) +
labs(title = "All samples projected on standard curve")
```

#### All samples projected on standard curve



#### Normalize to surface area

```
# Surface area data
sa <- read.csv("baseline_TPC_physio/output/surface.area.calc.csv")

# Tissue homogenate volume data
homog_vols <- read_csv("baseline_TPC_physio/data/TPC_homogenate_vols/homogenate_vols.csv") %>% select(1

# Coral sample metadata
metadata <- read_csv("baseline_TPC_physio/metadata_POC_TPC.csv") %>% select(1:4)

# Join homogenate volumes and surface area with sample metadata
metadata <- full_join(metadata, homog_vols) %>%
```

## Plot results by species and site

```
# Plot all data points with mean \pm se
prot_host %>%
  filter(!is.na(species)) %>%
  ggplot(aes(x = species, y = prot_mg.cm2, color = species)) +
  #facet_wrap(~species) +
  coord_cartesian(ylim = c(0, 1.5)) +
  labs(x = "", y = "Total protein (mg/cm2)") +
  geom_jitter(width = 0.1) +
                                                                            # Plot all points
  stat_summary(fun.data = mean_cl_normal, fun.args = list(mult = 1),
                                                                            # Plot standard error
                geom = "errorbar", color = "black", width = 0.5) +
  stat_summary(fun.y = mean, geom = "point", color = "black")
                                                                            # Plot mean
   1.5 -
Total protein (mg/cm2)
                                                                              species
                                                                                   Pocillopora
   0.0 -
                                    Pocillopora
# Remove outlier data points where absorbance was super low
prot_host <- prot_host %>%
  filter(abs562 > 0.1)
```

### Write data to output file

### Import data for holobiont protein

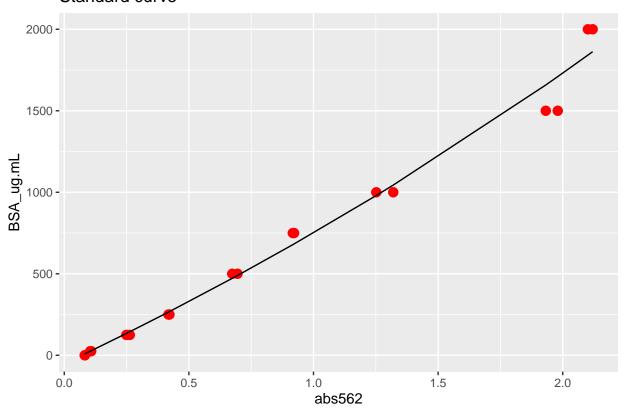
```
# List protein data files
prot_path = "baseline_TPC_physio/data/TPC_total_protein/holobiont/"
all_prot_files <- list.files(path = prot_path, pattern = "*.csv")</pre>
                                                                             # List all files in director
prot_platemaps <- list.files(path = prot_path, pattern = "platemap")</pre>
                                                                             # List platemap files
prot_data_files <- setdiff(all_prot_files, prot_platemaps)</pre>
                                                                             # List data files
# Read in all files into tibble
df_holobiont <- tibble(file = prot_data_files) %>%
  separate(file, into = c("trip", "date", "plate"), remove = FALSE) %>%
  unite(plate, trip, date, plate) %>%
  dplyr::mutate(platemap = map(plate, ~read_csv(paste0(prot_path, ., "_platemap.csv"))),
         prot_data = map(file, ~read_csv(paste0(prot_path, .)) %>% rename(well = Well)))
# Merge platemap and data for each plate
df_holobiont <- df_holobiont %>%
  mutate(merged = map2(platemap, prot_data, ~ right_join(.x, .y)))
```

#### Plot standard curve

```
# Create standard curve following kit instructions
standards_holobiont <- tribble(</pre>
  ~std, ~BSA_ug.mL,
  "A",
              2000,
  "B",
              1500,
  "C",
              1000,
  "D",
               750,
  "E".
               500,
  "F",
               250,
  "G".
               125,
 "H",
               25,
  "I",
                 0
std curve holobiont <- df holobiont %>%
 unnest(merged) %>%
 filter(grepl("Standard", fragment_ID)) %>%
```

```
select(plate, well, fragment_ID, abs562 = `562:562`) %>%
      rename(std = fragment_ID) %>%
      mutate(std = str_sub(std, 9, 9)) %>%
      #group_by(std) %>%
      #summarise(abs562 = mean(abs562)) %>%
                                                                                                                                                                                                                   # calculate mean of standard duplicates
      #mutate(abs562.adj = abs562 - abs562[std == "I"]) %>%
                                                                                                                                                                                                                   # subtract blank absorbace value from all
      left_join(standards_holobiont)
## Fit nonlinear model for standard curve
mod_holobiont \leftarrow nls(formula = BSA_ug.mL \sim z + a * exp(b * abs562), start = list(z = 0, a = 1, b = 1), verified by the start of the st
fitted_holobiont <- mod_holobiont %>% broom::augment()
# Plot standard curve
std_curve_plot_holobiont <- std_curve_holobiont %>%
      ggplot(aes(x = abs562, y = BSA_ug.mL)) +
      geom_point(color = "red", size = 3)
std_curve_plot_holobiont +
      geom_line(\frac{data}{data} = fitted_holobiont, aes(x = abs562, y = .fitted)) +
      labs(title = "Standard curve")
```

#### Standard curve

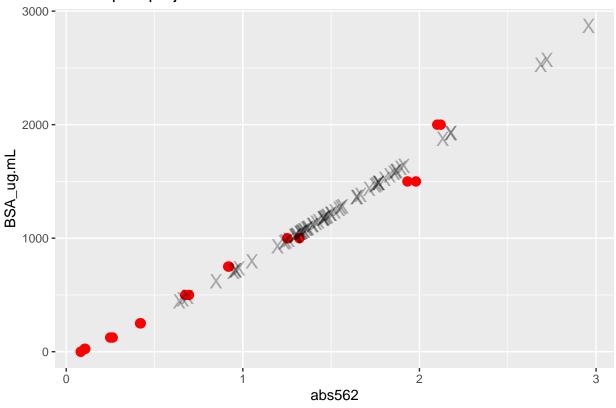


## Calculate protein concentrations

```
# Calculate protein concentration for all samples using standard curve prot_holobiont <- df_holobiont %>%
```

```
unnest(merged) %>%
filter(!grepl("Standard", fragment_ID)) %>%  # Get just samples (not standards)
select(plate, well, fragment_ID, abs562 = `562:562`) %>%  # Select only needed columns
filter(!is.na(fragment_ID)) %>%  # Filter out empty wells
filter(fragment_ID != "BK") %>%  # Filter out blank wells
mutate(prot_ug.mL = map_dbl(abs562, ~ predict(mod_holobiont, newdata = data.frame(abs562 = .))))  #
std_curve_plot_holobiont +
geom_point(data = prot_holobiont, aes(x = abs562, y = prot_ug.mL), pch = "X", cex = 5, alpha = 0.3) +
labs(title = "All samples projected on standard curve")
```

## All samples projected on standard curve



#### Normalize to surface area

```
# Surface area data
sa <- read.csv("baseline_TPC_physio/output/surface.area.calc.csv")

# Tissue homogenate volume data
homog_vols <- read_csv("baseline_TPC_physio/data/TPC_homogenate_vols/homogenate_vols.csv") %>% select(1

# Coral sample metadata
metadata <- read_csv("baseline_TPC_physio/metadata_POC_TPC.csv") %>% select(1:4)

# Join homogenate volumes and surface area with sample metadata
metadata <- full_join(metadata, homog_vols) %>%
```

```
full_join(sa)
# Join prot data with metadata
prot_holobiont <- left_join(prot_holobiont, metadata) %>%
  mutate(prot_ug = prot_ug.mL * homog_vol_ml,
         prot_ug.cm2 = prot_ug / surface.area.cm2,
         prot_mg.cm2 = prot_ug.cm2 / 1000)
```

## Plot results by species and site

```
# Plot all data points with mean \pm se
prot_holobiont %>%
  filter(!is.na(species)) %>%
  ggplot(aes(x = species, y = prot_mg.cm2, color = species)) +
  #facet_wrap(~species) +
  coord_cartesian(ylim = c(0, 2)) +
  labs(x = "", y = "Total protein (mg/cm2)") +
  geom_jitter(width = 0.1) +
                                                                             # Plot all points
  stat_summary(fun.data = mean_cl_normal, fun.args = list(mult = 1),
                                                                             # Plot standard error
                geom = "errorbar", color = "black", width = 0.5) +
  stat_summary(fun.y = mean, geom = "point", color = "black")
                                                                             # Plot mean
   2.0 -
   1.5 -
Total protein (mg/cm2)
                                                                               species
                                                                                    Pocillopora
   0.5 -
   0.0 -
                                     Pocillopora
# Remove outlier data points where absorbance was super low
prot_holobiont <- prot_holobiont %>%
```

```
filter(abs562 > 0.1)
```

# Write data to output file